The privileged immunity of immune privileged organs: the case of the eye

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Michal Schwartz, Department of Neurobiology, Weizmann Institute of Science, Rehovot 76100, Israel. e-mail: michal.schwartz@weizmann. ac.il Understanding of ocular diseases and the search for their cure have been based on the common assumption that the eye is an immune privileged site, and the consequent conclusion that entry of immune cells to this organ is forbidden. Accordingly, it was assumed that when immune cell entry does occur, this reflects an undesired outcome of breached barriers. However, studies spanning more than a decade have demonstrated that acute insults to the retina, or chronic conditions resulting in retinal ganglion cell loss, such as in glaucoma, result in an inferior outcome in immunocompromised mice; likewise, steroidal treatment was found to be detrimental under these conditions. Moreover, even conditions that are associated with inflammation, such as age-related macular degeneration, are not currently believed to require immune suppression for treatment, but rather, are thought to benefit from immune modulation. Here, we propose that the immune privilege of the eye is its ability to enable, upon need, the entry of selected immune cells for its repair and healing, rather than to altogether prevent immune cell entry. The implications for acute and chronic degenerative diseases, as well as for infection and inflammatory diseases, are discussed.

Keywords: immune privilege, visual system, immunomodulation, neuroprotection and neuronal repair, inflammation

INTRODUCTION

Over the past decades, the mammalian central nervous system (CNS), including the eye, brain, and spinal cord, were believed to be sealed from the circulation. Thus, immune activity at these sites was considered forbidden, and was collectively assumed to be consistently detrimental. As a consequence, the inflammatory response in the eye or the brain was assessed solely based on counting the number of immune cells, without regard to their phenotype or function. Thus, the poor ability of the optic nerve to regenerate following injury, as well as the poor recovery following acute injury to any other parts of the CNS, were assumed to be an outcome of local detrimental immune activity seen at the lesion site (Fitch et al., 1999; Popovich et al., 1999; Ghirnikar et al., 2001). Such a view was almost universally accepted from the early 1980s and supported the use of anti-inflammatory drugs to treat victims of CNS injuries (Constantini and Young, 1994; Carlson et al., 1998).

With time and the advance of technologies, there was an increase in the understanding of the heterogeneity of innate and adaptive immunity in general, and in the CNS in particular, with respect to both functional cell subsets (Korn et al., 2007; Gee et al., 2008; O'Shea et al., 2008; Auffray et al., 2009; Prinz et al., 2011; Zhu et al., 2011) and origin (Geissmann et al., 2010; Ginhoux et al., 2010; Prinz et al., 2011). As a corollary, it became clear that some of the blanket assumptions regarding the eye and the brain were not accurate, and, accordingly, that some experimental findings had not been properly interpreted. Thus, it became evident that the response to CNS injury, similar to that in other tissues in the body, is a multi-step process that requires a set sequence, and synchrony of events in time and space; many of the steps that

take place in the healing process following "sterile" injuries are similar if not identical to processes occurring outside the CNS with respect to the immune response (Dusart and Schwab, 1994; Frank and Wolburg, 1996; Arnold et al., 2007; Nahrendorf et al., 2007; Rolls et al., 2009; Shechter et al., 2009; Stirling et al., 2009; London et al., 2011). The early innate immune response involves cells that are needed for cleaning the lesion site, yet the activity of these cells must be followed by immune cells that terminate this initial response and subsequently contribute to the repair. Both stages involve innate immune cells of distinct phenotypes; the cells that contribute to the termination of the local early response are largely monocyte-derived macrophages that acquire and exert a local antiinflammatory function (Kigerl et al., 2009; Shechter et al., 2009; London et al., 2011; Zhu et al., 2011). The obvious question is how such a response can be reconciled with the traditional view of the eye as an immune privileged site; do these findings change our understanding of the privilege, or do they require breaking of privilege under severe conditions? Here, focusing on the eye, we will discuss a different view of the physiological meaning of the CNS as an immune privileged site, and its manifestations under pathological conditions.

THE EYE AS AN IMMUNE PRIVILEGED ORGAN

Immune privileged organs were operationally defined as sites in the body where foreign tissue grafts can survive for extended, often indefinite periods of time, whereas similar grafts placed at regular sites in the body are acutely rejected (Medawar, 1948). These organs include the eye and the brain, as well as the pregnant uterus, testis, and several others (Streilein, 2003b; Niederkorn, 2006). Such immune privilege is thought to be an evolutionary adaptation to

protect tissues that are indispensable, yet have limited regeneration capacity, like the brain and the eye, from the potentially damaging effects of an uncontrolled inflammatory immune response. Thus, immune privileged organs were considered as ones to which immune cell entry is forbidden; leukocytes were believed to be excluded from these vital organs by the presence of specialized physical barriers, the blood–tissue barriers.

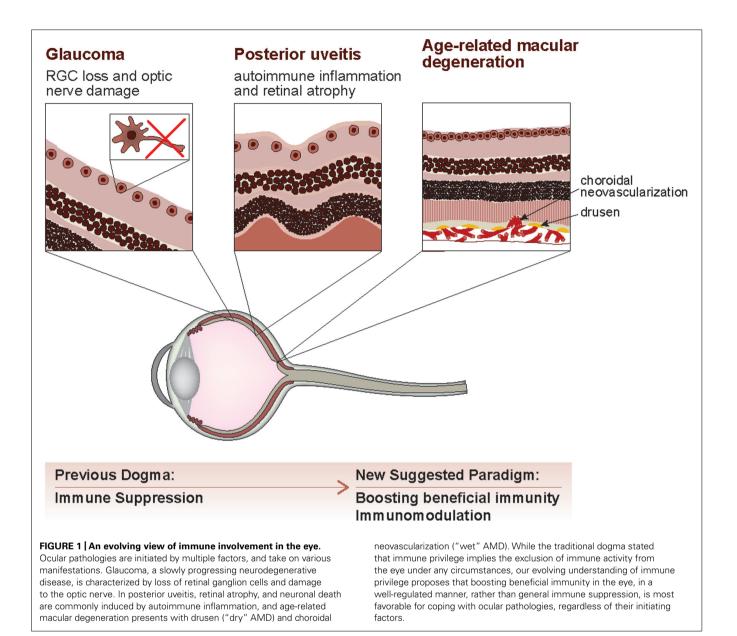
In contrast to the previous view that immune privilege is maintained by immune cell exclusion, it is now increasingly accepted that the privileged status is preserved by local active mechanisms that suppress responses to antigens within the privileged tissues (Niederkorn and Stein-Streilein, 2010). In the eye, one such mechanism is anterior chamber-associated immune deviation (ACAID), referring to a phenomenon in which antigenic material introduced into the anterior chamber of the eye elicits a systemic immune response that results in immune deviation, characterized by the suppression of T cell-mediated immunity, while enabling the production of non-complement-fixing antibodies (Kaplan et al., 1975; Streilein, 2003b; Niederkorn, 2006). ACAID involves the migration of specialized antigen presenting cells from the eye to the thymus and spleen, and is associated with an elevation in regulatory, γδ, and natural killer T cells (Streilein, 2003b; Niederkorn, 2006). Other mechanisms aimed at maintaining the immune privileged state of the eye include the reduced expression of MHC molecules on ocular cells, and the existence of an intraocular anti-inflammatory environment, mediated by resident cells, and various molecules, both surface-bound and soluble, all of which serve to modulate the activity of infiltrating immune cells, in situ (Streilein, 2003b; Schewitz-Bowers et al., 2010; Zhou et al., 2012). These well-orchestrated, multifaceted mechanisms, known to involve numerous pathways, were long thought to be designed to ensure limited infiltration of circulating immune cells to the eye, leaving behind a tissue that was considered autonomous in terms of repair. It is puzzling, however, why a fragile and precious organ such as the eye would evolve such complex tolerance mechanisms, if their sole purpose were to guarantee immune ignorance. Moreover, several studies have shown that immunocompromised mice exhibit worse recovery from optic nerve and retinal insult than do their immunocompetent counterparts (Kipnis et al., 2001; Schori et al., 2001; Yoles et al., 2001), similar to the case in peripheral nerve injury (Serpe et al., 1999). Similarly, recent studies have demonstrated that well-regulated immune responses in the CNS, rather than immune ignorance, are optimal for the recovery of the tissue after insult, whether sterile or immune-induced (Kerr et al., 2008; Shechter et al., 2009; Caspi et al., 2011; London et al., 2011; London et al., under revision). Thus, it is becoming increasingly clear that immune privilege is not aimed at entirely suppressing immune responses in the target organ, but rather at maintaining a specialized, tightly regulated immunological niche to preserve the integrity of especially vulnerable organs, such as the brain and the eye (Streilein, 2003b; Niederkorn, 2006).

REGULATED IMMUNE RESPONSES ARE BENEFICIAL IN MITIGATING EYE PATHOLOGIES

Inflammation is the body's adaptive response to any insult, be it mechanical, biochemical, or immune-mediated. However,

inflammation is beneficial only on the condition that it ends in active resolution (Gronert, 2010). Studies on wound healing outside the CNS have characterized distinct subsets of macrophages that infiltrate the site of injury and display different functions corresponding to the changing needs of the tissue along the course of healing; these include the clearing of dead cells and tissue debris at the first stage, and the secretion of anti-inflammatory cytokines and growth factors at the later stage, to aid tissue regrowth and restoration of immune homeostasis (Arnold et al., 2007; Nahrendorf et al., 2007). Recently, our team demonstrated that a subset of monocyte-derived macrophages, which manifests an immune-resolving phenotype, is essential for the resolution of inflammation after sterile insults, in models of spinal cord injury and retinal glutamate intoxication (Shechter et al., 2009; London et al., 2011). In both of these cases, such macrophages were found to be crucial for recovery, as was measured by a functional motor scale after spinal cord injury, and directly in terms of cell survival in the retina. Thus, despite the classification of these organs as immune privileged, they nevertheless derive benefit from the controlled recruitment of innate immune cells from the circulation, to assist in their healing. Notably, while the CNS contains its own population of immune cells, the resident microglia, we have shown that infiltrating bloodderived macrophages are nonetheless crucial for neuroprotective and anti-inflammatory activities at the injury site; we have therefore proposed that the infiltrating cells fulfill specialized functions in the recovery process, which the resident immune cells either fail to display, or at least do not manifest at the right time or at sufficient levels (Shechter et al., 2009). In animal models of optic nerve injury, it was found that macrophages can modify the non-permissive nature of the optic nerve for regeneration in vitro (David et al., 1990), and that transplantation of activated macrophages into the injured optic nerve can facilitate regrowth in vivo (Lazarov-Spiegler et al., 1996). In line with these observations, the important contribution of a macrophagederived molecule, oncomodulin, to the regeneration of the optic nerve, was identified by Benowitz and colleagues (Yin et al., 2006, 2009; Cui et al., 2009), who coined the term "inflammationinduced regeneration." Collectively, these results attribute to innate immunity an important role in eye repair, and reveal the ability of macrophages to orchestrate neuroprotection and axonal regeneration.

The beneficial role of adaptive immunity in neuroprotection was initially observed in animal models simulating different aspects of glaucoma, where it was found that the extent of retinal ganglion cell loss is increased in immunocompromised animals relative to immunocompetent ones (Kipnis et al., 2001; Schori et al., 2001; Yoles et al., 2001; Bakalash et al., 2002). Moreover, T cell-based vaccinations, both passive and active, promote neuroprotection after optic nerve crush (Moalem et al., 1999; Fisher et al., 2001). Importantly, the potential benefit derived from T cells in these systems relies, at least in part, on a delicate balance between effector and regulatory subsets of these cells (Kipnis et al., 2002, 2004). More recently, results obtained in different models of CNS insult suggested that the beneficial effects of T cells might be mediated in part by controlling the recruitment of monocyte-derived macrophages from the circulation (Butovsky et al., 2007; Schwartz



et al., 2009; Shechter et al., 2009). Thus, the well-orchestrated collaboration between the innate and adaptive arms of the immune system appears to be optimal for achieving neuroprotection.

The beneficial involvement of immune cells in the eye is also observed in diseases that are immune-induced, such as autoimmune posterior uveitis, a potentially blinding inflammatory condition affecting the retina and the choroid of the eye. Studies in experimental autoimmune uveitis (EAU), an animal model of human posterior uveitis, demonstrate the heterogeneity of immune cells along this disease. Beside the well-characterized pro-inflammatory cells known to initiate EAU, the uveitic eye is also endowed with regulatory immune populations (Robertson et al., 2002; Kerr et al., 2008; Caspi et al., 2011; London et al., under revision). These cells, including subsets of macrophages and T cells, act to limit inflammation, presumably bringing the disease to a state of equilibrium and remission.

An additional pathology in which the immune system has been shown to fulfill various, perhaps opposing functions, is age-related macular degeneration (AMD), the leading cause of blindness in the elderly. Naturally, the etiology of AMD is very diverse; the disease is associated with numerous immune-related factors. Here too, the role of macrophages has been a matter of debate; on the one hand, it was found that aging is accompanied by a pathological shift to M2 macrophages, which are known to promote angiogenesis, and would therefore seem likely candidates for promoting choroidal neovascularization (CNV), the process by which abnormal blood vessels develop beneath the retina (Espinosa-Heidmann et al., 2003; Sakurai et al., 2003; Cao et al., 2011). On the other hand, studies have also shown that prevention of macrophage entry into the eye promotes CNV, whereas injection of macrophages inhibits it (Apte et al., 2006). Patel and Chan (2008) reviewed the seemingly contradictory

functions of macrophages in AMD, and proposed that these conflicting findings reflect a dual role of macrophages in this pathology, where the uncontrolled pro-inflammatory M1 macrophages induce tissue damage, and the M2 macrophages, which are recruited to terminate the M1 response and to clear drusen and other age-related deposits, could also adversely affect disease progression by displaying pro-angiogenic activity. Among the additional factors associated with AMD pathogenesis and progression, a pivotal role has been attributed over the past several years to the complement system and its dysregulation (Klein et al., 2005; Patel and Chan, 2008; Anderson et al., 2010). These findings emphasize the need for a regulated immune response, in terms of timing, duration, and phenotype, and further support the argument that there are no "good" or "bad" immune cells; it is all a matter of their control and coordination. Moreover, the accumulating evidence on beneficial immune involvement in AMD and in the other ocular pathologies mentioned above give further reinforcement to the current contention that although the eye is an immune privileged site, it can enjoy the benefits of immune support, and thus immune regulation, rather than immune suppression, is the key to disease resolution, as in other parts of the body (Figure 1).

A DIFFERENT VIEW OF IMMUNE PRIVILEGE

Immune privilege is an evolutionary adaptation aimed at protecting especially vulnerable organs from overwhelming inflammation that could abolish their functions and jeopardize the well-being of the individual. As vision is crucial for survival, it is understandable why the eye would be particularly protected from these risks (Streilein, 2003a). However, we propose that the immune privileged designation of the eye means that it has the privilege to enable selective immune responses most suitable and effective for its proper function in health and pathology. We contend that this is true for all other parts of the CNS, as well.

REFERENCES

Anderson, D. H., Radeke, M. J., Gallo,
N. B., Chapin, E. A., Johnson, P. T.,
Curletti, C. R., Hancox, L. S., Hu, J.,
Ebright, J. N., Malek, G., Hauser, M.
A., Rickman, C. B., Bok, D., Hageman, G. S., and Johnson, L. V. (2010).
The pivotal role of the complement system in aging and age-related macular degeneration: hypothesis revisited. *Prog. Retin. Eye Res.* 29, 95–112.

Apte, R. S., Richter, J., Herndon, J., and Ferguson, T. A. (2006). Macrophages inhibit neovascularization in a murine model of agerelated macular degeneration. *PLoS Med.* 3, e310. doi: 10.1371/journal.pmed.0030310

Arnold, L., Henry, A., Poron, F., Baba-Amer, Y., Van Rooijen, N., Plonquet, A., Gherardi, R. K., and Chazaud, B. (2007). Inflammatory monocytes recruited after skeletal muscle injury switch into antiinflammatory macrophages to support myogenesis. *J. Exp. Med.* 204, 1057–1069.

Auffray, C., Sieweke, M. H., and Geissmann, F. (2009). Blood monocytes: development, heterogeneity, and relationship with dendritic cells. *Annu. Rev. Immunol.* 27, 669–692.

Bakalash, S., Kessler, A., Mizrahi, T., Nussenblatt, R., and Schwartz, M. (2003). Antigenic specificity of immunoprotective therapeutic vaccination for glaucoma. *Invest. Ophthal*mol. Vis. Sci. 44, 3374–3381.

Bakalash, S., Kipnis, J., Yoles, E., and Schwartz, M. (2002). Resistance of retinal ganglion cells to an increase in intraocular pressure is immunedependent. *Invest. Ophthalmol. Vis.* Sci. 43, 2648–2653.

Butovsky, O., Kunis, G., Koronyo-Hamaoui, M., and Schwartz, M. (2007). Selective ablation of bone marrow-derived dendritic cells increases amyloid plaques in a mouse

As many CNS pathologies are associated with local inflammation, they are generally treated with anti-inflammatory and immunosuppressive drugs. However, this treatment approach has shown limited success in animal models of ocular pathologies and other neurodegenerative disorders, as well as in the clinic, and in some cases was even found to exacerbate disease (Levin et al., 1999; Solberg et al., 1999; Bakalash et al., 2003; Ohlsson et al., 2004; Dimitriu et al., 2008; Schwartz and Shechter, 2010). The benefits of those drugs, if any, are often temporary, as they help relieve some of the symptoms but do not address the underlying pathological processes (Gronert, 2010). Bearing in mind the heterogeneity of immune cells and their changing functions along the course of disease, together with the delicate balance of counter-regulatory signals required for effective resolution of inflammation (Gronert, 2010), we suggest that a more efficient approach to treating such disorders would be to manipulate specific immune subsets in a timely manner, rather than to globally inhibit the immune response (**Figure 1**).

Finally, our interpretation of the privilege of immunity in immune privileged sites does not negate the possibility that under certain conditions, immune privilege is breached in order to preserve the life of the individual, at the expense of local loss of function; this is the case in certain microbial infections, or in the presence of highly immunogenic tumors (Morrison et al., 1989; Niederkorn, 1991; Li and Niederkorn, 1997; Streilein et al., 1997; Saint Andre et al., 2002; Niederkorn and Stein-Streilein, 2010), in which a powerful immune response is essential, and the risk of blindness is accepted for the sake of survival (Niederkorn and Stein-Streilein, 2010).

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Alzheimer's disease model. Eur. J. Neurosci. 26, 413–416.

Cao, X., Shen, D., Patel, M. M., Tuo, J., Johnson, T. M., Olsen, T. W., and Chan, C. C. (2011). Macrophage polarization in the maculae of agerelated macular degeneration: a pilot study. *Pathol. Int.* 61, 528–535.

Carlson, S. L., Parrish, M. E., Springer, J. E., Doty, K., and Dossett, L. (1998). Acute inflammatory response in spinal cord following impact injury. Exp. Neurol. 151, 77–88.

Caspi, R. R., Horai, R., Chen, J., Hansen, A., Mcmanigle, W., Villasmil, R., and Silver, P. (2011). Retina-specific T regulatory cells accumulate in the eye during autoimmune uveitis and act to control inflammation. *J. Immunol.* 186, 115.22.

Constantini, S., and Young, W. (1994). The effects of methylprednisolone and the ganglioside GM1 on acute spinal cord injury in rats. *J. Neurosurg.* 80, 97–111.

Cui, Q., Yin, Y., and Benowitz, L. I. (2009). The role of macrophages in optic nerve regeneration. *Neuro-science* 158, 1039–1048.

David, S., Bouchard, C., Tsatas, O., and Giftochristos, N. (1990). Macrophages can modify the non-permissive nature of the adult mammalian central nervous system. *Neuron* 5, 463–469.

Dimitriu, C., Bach, M., Lagreze, W. A., and Jehle, T. (2008). Methyl-prednisolone fails to preserve retinal ganglion cells and visual function after ocular ischemia in rats. *Invest. Ophthalmol. Vis. Sci.* 49, 5003–5007.

Dusart, I., and Schwab, M. E. (1994). Secondary cell death and the inflammatory reaction after dorsal hemisection of the rat spinal cord. Eur. J. Neurosci. 6, 712–724.

Espinosa-Heidmann, D. G., Suner, I. J., Hernandez, E. P., Monroy, D., Csaky, K. G., and Cousins, S. W. (2003). Macrophage depletion

diminishes lesion size and severity in experimental choroidal neovascularization. *Invest. Ophthalmol. Vis. Sci.* 44, 3586–3592.

- Fisher, J., Levkovitch-Verbin, H., Schori, H., Yoles, E., Butovsky, O., Kaye, J. F., Ben-Nun, A., and Schwartz, M. (2001). Vaccination for neuroprotection in the mouse optic nerve: implications for optic neuropathies. *J. Neurosci.* 21, 136–142.
- Fitch, M. T., Doller, C., Combs, C. K., Landreth, G. E., and Silver, J. (1999). Cellular and molecular mechanisms of glial scarring and progressive cavitation: in vivo and in vitro analysis of inflammation-induced secondary injury after CNS trauma. *J. Neurosci.* 19, 8182–8198.
- Frank, M., and Wolburg, H. (1996). Cellular reactions at the lesion site after crushing of the rat optic nerve. *Glia* 16, 227–240.
- Gee, J. M., Kalil, A., Thullbery, M., and Becker, K. J. (2008). Induction of immunologic tolerance to myelin basic protein prevents central nervous system autoimmunity and improves outcome after stroke. Stroke 39. 1575–1582.
- Geissmann, F., Manz, M. G., Jung, S., Sieweke, M. H., Merad, M., and Ley, K. (2010). Development of monocytes, macrophages, and dendritic cells. *Science* 327, 656–661.
- Ghirnikar, R. S., Lee, Y. L., and Eng, L. F. (2001). Chemokine antagonist infusion promotes axonal sparing after spinal cord contusion injury in rat. *J. Neurosci. Res.* 64, 582–589.
- Ginhoux, F., Greter, M., Leboeuf, M., Nandi, S., See, P., Gokhan, S., Mehler, M. F., Conway, S. J., Ng, L. G., Stanley, E. R., Samokhvalov, I. M., and Merad, M. (2010). Fate mapping analysis reveals that adult microglia derive from primitive macrophages. *Science* 330, 841–845.
- Gronert, K. (2010). Resolution, the grail for healthy ocular inflammation. *Exp. Eye Res.* 91, 478–485.
- Kaplan, H. J., Streilein, J. W., and Stevens, T. R. (1975). Transplantation immunology of the anterior chamber of the eye. II. Immune response to allogeneic cells. *J. Immunol.* 115, 805–810.
- Kerr, E. C., Raveney, B. J., Copland, D. A., Dick, A. D., and Nicholson, L. B. (2008). Analysis of retinal cellular infiltrate in experimental autoimmune uveoretinitis reveals multiple regulatory cell populations. *J. Autoimmun.* 31, 354–361.
- Kigerl, K. A., Gensel, J. C., Ankeny, D. P., Alexander, J. K., Donnelly, D. J., and Popovich, P. G. (2009). Identification of two distinct macrophage

- subsets with divergent effects causing either neurotoxicity or regeneration in the injured mouse spinal cord. *J. Neurosci.* 29, 13435–13444.
- Kipnis, J., Cardon, M., Avidan, H., Lewitus, G. M., Mordechay, S., Rolls, A., Shani, Y., and Schwartz, M. (2004). Dopamine, through the extracellular signal-regulated kinase pathway, downregulates CD4+CD25+ regulatory T-cell activity: implications for neurodegeneration. *J. Neurosci.* 24, 6133–6143.
- Kipnis, J., Mizrahi, T., Hauben, E., Shaked, I., Shevach, E., and Schwartz, M. (2002). Neuroprotective autoimmunity: naturally occurring CD4+CD25+ regulatory T cells suppress the ability to withstand injury to the central nervous system. *Proc. Natl. Acad. Sci. U.S.A.* 99, 15620–15625.
- Kipnis, J., Yoles, E., Schori, H., Hauben, E., Shaked, I., and Schwartz, M. (2001). Neuronal survival after CNS insult is determined by a genetically encoded autoimmune response. J. Neurosci. 21, 4564–4571.
- Klein, R. J., Zeiss, C., Chew, E. Y., Tsai, J. Y., Sackler, R. S., Haynes, C., Henning, A. K., Sangiovanni, J. P., Mane, S. M., Mayne, S. T., Bracken, M. B., Ferris, F. L., Ott, J., Barnstable, C., and Hoh, J. (2005). Complement factor H polymorphism in age-related macular degeneration. *Science* 308, 385–389.
- Korn, T., Reddy, J., Gao, W., Bettelli, E., Awasthi, A., Petersen, T. R., Backstrom, B. T., Sobel, R. A., Wucherpfennig, K. W., Strom, T. B., Oukka, M., and Kuchroo, V. K. (2007). Myelin-specific regulatory T cells accumulate in the CNS but fail to control autoimmune inflammation. *Nat. Med.* 13, 423–431.
- Lazarov-Spiegler, O., Solomon, A. S., Zeev-Brann, A. B., Hirschberg, D. L., Lavie, V., and Schwartz, M. (1996). Transplantation of activated macrophages overcomes central nervous system regrowth failure. *FASEB J.* 10, 1296–1302.
- Levin, L. A., Beck, R. W., Joseph, M. P., Seiff, S., and Kraker, R. (1999). The treatment of traumatic optic neuropathy: the International Optic Nerve Trauma Study. *Ophthalmology* 106, 1268–1277.
- Li, X. Y., and Niederkorn, J. Y. (1997). Immune privilege in the anterior chamber of the eye is not extended to intraocular *Listeria monocytogenes*. Ocul. Immunol. Inflamm. 5, 245–257.
- London, A., Itskovich, E., Benhar, I., Kalchenko, V., Mack, M., Jung, S., and Schwartz, M. (2011). Neuroprotection and progenitor cell renewal

- in the injured adult murine retina requires healing monocyte-derived macrophages. *J. Exp. Med.* 208, 23–39.
- Medawar, P. B. (1948). Immunity to homologous grafted skin; the fate of skin homografts transplanted to the brain, to subcutaneous tissue, and to the anterior chamber of the eye. *Br. J. Exp. Pathol.* 29, 58–69.
- Moalem, G., Leibowitz-Amit, R., Yoles, E., Mor, F., Cohen, I. R., and Schwartz, M. (1999). Autoimmune T cells protect neurons from secondary degeneration after central nervous system axotomy. *Nat. Med.* 5, 49–55.
- Morrison, R. P., Lyng, K., and Caldwell, H. D. (1989). Chlamydial disease pathogenesis. Ocular hypersensitivity elicited by a genus-specific 57-kD protein. J. Exp. Med. 169, 663–675.
- Nahrendorf, M., Swirski, F. K., Aikawa, E., Stangenberg, L., Wurdinger, T., Figueiredo, J. L., Libby, P., Weissleder, R., and Pittet, M. J. (2007). The healing myocardium sequentially mobilizes two monocyte subsets with divergent and complementary functions. J. Exp. Med. 204, 3037–3047.
- Niederkorn, J. Y. (1991). The immunopathology of intraocular tumour rejection. *Eye (Lond.)* 5(Pt 2), 186–192.
- Niederkorn, J. Y. (2006). See no evil, hear no evil, do no evil: the lessons of immune privilege. *Nat. Immunol.* 7, 354–359.
- Niederkorn, J. Y., and Stein-Streilein, J. (2010). History and physiology of immune privilege. *Ocul. Immunol. Inflamm.* 18, 19–23.
- Ohlsson, M., Westerlund, U., Langmoen, I. A., and Svensson, M. (2004). Methylprednisolone treatment does not influence axonal regeneration or degeneration following optic nerve injury in the adult rat. *J. Neuroophthalmol.* 24, 11–18.
- O'Shea, J. J., Hunter, C. A., and Germain, R. N. (2008). T cell heterogeneity: firmly fixed, predominantly plastic or merely malleable? *Nat. Immunol.* 9, 450–453.
- Patel, M., and Chan, C. C. (2008). Immunopathological aspects of agerelated macular degeneration. Semin. Immunopathol. 30, 97–110.
- Popovich, P. G., Guan, Z., Wei, P., Huitinga, I., Van Rooijen, N., and Stokes, B. T. (1999). Depletion of hematogenous macrophages promotes partial hindlimb recovery and neuroanatomical repair after experimental spinal cord injury. Exp. Neurol. 158, 351–365.
- Prinz, M., Priller, J., Sisodia, S. S., and Ransohoff, R. M. (2011). Heterogeneity of CNS myeloid cells

- and their roles in neurodegeneration. *Nat. Neurosci.* 14, 1227–1235.
- Robertson, M. J., Erwig, L. P., Liversidge, J., Forrester, J. V., Rees, A. J., and Dick, A. D. (2002). Retinal microenvironment controls resident and infiltrating macrophage function during uveoretinitis. *Invest. Ophthalmol. Vis. Sci.* 43, 2250–2257.
- Rolls, A., Shechter, R., and Schwartz, M. (2009). The bright side of the glial scar in CNS repair. *Nat. Rev. Neurosci.* 10, 235–241.
- Saint Andre, A., Blackwell, N. M., Hall, L. R., Hoerauf, A., Brattig, N. W., Volkmann, L., Taylor, M. J., Ford, L., Hise, A. G., Lass, J. H., Diaconu, E., and Pearlman, E. (2002). The role of endosymbiotic *Wolbachia* bacteria in the pathogenesis of river blindness. *Science* 295, 1892–1895.
- Sakurai, E., Anand, A., Ambati, B. K., Van Rooijen, N., and Ambati, J. (2003). Macrophage depletion inhibits experimental choroidal neovascularization. *Invest. Ophthalmol. Vis. Sci.* 44, 3578–3585.
- Schewitz-Bowers, L. P., Lee, R. W. J., and Dick, A. D. (2010). Immune mechanisms of intraocular inflammation. *Expert Rev. Ophthalmol.* 5, 43–58.
- Schori, H., Yoles, E., and Schwartz, M. (2001). T-cell-based immunity counteracts the potential toxicity of glutamate in the central nervous system. J. Neuroimmunol. 119, 199–204.
- Schwartz, M., London, A., and Shechter, R. (2009). Boosting T-cell immunity as a therapeutic approach for neurodegenerative conditions: the role of innate immunity. *Neuroscience* 158, 1133–1142.
- Schwartz, M., and Shechter, R. (2010). Systemic inflammatory cells fight off neurodegenerative disease. *Nat. Rev. Neurol.* 6, 405–410.
- Serpe, C. J., Kohm, A. P., Huppenbauer, C. B., Sanders, V. M., and Jones, K. J. (1999). Exacerbation of facial motoneuron loss after facial nerve transection in severe combined immunodeficient (scid) mice. J. Neurosci. 19, RC7.
- Shechter, R., London, A., Varol, C., Raposo, C., Cusimano, M., Yovel, G., Rolls, A., Mack, M., Pluchino, S., Martino, G., Jung, S., and Schwartz, M. (2009). Infiltrating blood-derived macrophages are vital cells playing an anti-inflammatory role in recovery from spinal cord injury in mice. *PLoS Med.* 6, e1000113. doi: 10.1371/journal.pmed.1000113
- Solberg, Y., Dubinski, G., Tchirkov, M., Belkin, M., and Rosner, M. (1999). Methylprednisolone therapy for retinal laser injury. Surv. Ophthalmol. 44(Suppl. 1), S85–S92.

Stirling, D. P., Liu, S., Kubes, P., and Yong, V. W. (2009). Depletion of Ly6G/Gr-1 leukocytes after spinal cord injury in mice alters wound healing and worsens neurological outcome. *I. Neurosci.* 29, 753–764.

- Streilein, J. W. (2003a). Ocular immune privilege: the eye takes a dim but practical view of immunity and inflammation. J. Leukoc. Biol. 74, 179–185.
- Streilein, J. W. (2003b). Ocular immune privilege: therapeutic opportunities from an experiment of nature. *Nat. Rev. Immunol.* 3, 879–889.
- Streilein, J. W., Dana, M. R., and Ksander, B. R. (1997). Immunity causing blindness: five different paths to herpes stromal keratitis. *Immunol. Today* 18, 443–449.
- Yin, Y., Cui, Q., Gilbert, H. Y., Yang, Y., Yang, Z., Berlinicke, C., Li,

- Z., Zaverucha-Do-Valle, C., He, H., Petkova, V., Zack, D. J., and Benowitz, L. I. (2009). Oncomodulin links inflammation to optic nerve regeneration. *Proc. Natl. Acad. Sci. U.S.A.* 106, 19587–19592.
- Yin, Y., Henzl, M. T., Lorber, B., Nakazawa, T., Thomas, T. T., Jiang, F., Langer, R., and Benowitz, L. I. (2006). Oncomodulin is a macrophage-derived signal for axon regeneration in retinal ganglion cells. *Nat. Neurosci.* 9, 843–852.
- Yoles, E., Hauben, E., Palgi, O., Agranov, E., Gothilf, A., Cohen, A., Kuchroo, V., Cohen, I. R., Weiner, H., and Schwartz, M. (2001). Protective autoimmunity is a physiological response to CNS trauma. *J. Neurosci.* 21, 3740–3748.
- Zhou, R., Horai, R., Silver, P. B., Mattapallil, M. J., Zarate-Blades, C. R.,

- Chong, W. P., Chen, J., Rigden, R. C., Villasmil, R., and Caspi, R. R. (2012). The living eye "disarms" uncommitted autoreactive T cells by converting them to Foxp3(+) regulatory cells following local antigen recognition. *J. Immunol.* 188, 1742–1750.
- Zhu, B., Kennedy, J. K., Wang, Y., Sandoval-Garcia, C., Cao, L., Xiao, S., Wu, C., Elyaman, W., and Khoury, S. J. (2011). Plasticity of Ly-6C(hi) myeloid cells in T cell regulation. *J. Immunol.* 187, 2418–2432.
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