# Natural killer cells and neuroblastoma: tumor recognition, escape mechanisms, and possible novel immunotherapeutic approaches

# Cristina Bottino<sup>1,2</sup>, Alessandra Dondero<sup>1</sup>, Francesca Bellora<sup>1</sup>, Lorenzo Moretta<sup>2</sup>, Franco Locatelli<sup>3,4</sup>, Vito Pistoia<sup>2</sup>, Alessandro Moretta<sup>1,5</sup>\* and Roberta Castriconi<sup>1,5</sup>

<sup>1</sup> Dipartimento di Medicina Sperimentale, Università degli Studi di Genova, Genova, Italy

<sup>2</sup> Istituto Giannina Gaslini, Genova, Italy

<sup>3</sup> Dipartimento di Onco-Ematologia Pediatrica, Ospedale Bambino Gesù, Roma, Italy

<sup>4</sup> Università di Pavia, Pavia, Italy

<sup>5</sup> Centro di Eccellenza per le Ricerche Biomediche, Università degli Studi di Genova, Genova, Italy

#### Edited by:

Daniel Olive, INSERM UMR 891, Institut Paoli Calmettes, France

#### Reviewed by:

Jeroen Van Bergen, Leiden University Medical Center, Netherlands Daniel Olive, INSERM UMR 891, Institut Paoli Calmettes, France

#### \*Correspondence:

Alessandro Moretta, Dipartimento di Medicina Sperimentale, Università degli studi di Genova, Via L.B. Alberti 2, 16132 Genova, Italy e-mail: alemoret@unige.it

Neuroblastoma (NB) is the most common extra-cranial solid tumor of childhood and arises from developing sympathetic nervous system. Most primary tumors localize in the abdomen, the adrenal gland, or lumbar sympathetic ganglia. Amplification in tumor cells of MYCN, the major oncogenic driver, patients' age over 18 months, and the presence at diagnosis of a metastatic disease (stage IV, M) identify NB at high risk of treatment failure. Conventional therapies did not significantly improve the overall survival of these patients. Moreover, the limited landscape of somatic mutations detected in NB is hampering the development of novel pharmacological approaches. Major efforts aim to identify novel NBassociated surface molecules that activate immune responses and/or direct drugs to tumor cells and tumor-associated vessels. PVR (Poliovirus Receptor) and B7-H3 are promising targets, since they are expressed by most high-risk NB, are upregulated in tumor vasculature and are essential for tumor survival/invasiveness. PVR is a ligand of DNAM-1 activating receptor that triggers the cytolytic activity of natural killer (NK) cells against NB. In animal models, targeting of PVR with an attenuated oncolytic poliovirus induced tumor regression and elimination. Also B7-H3 was successfully targeted in preclinical studies and is now being tested in phase I/II clinical trials. B7-H3 down-regulates NK cytotoxicity, providing NB with a mechanism of escape from immune response. The immunosuppressive potential of NB can be enhanced by the release of soluble factors that impair NK cell function and/or recruitment. Among these, TGF-B1 modulates the cytotoxicity receptors and the chemokine receptor repertoire of NK cells. Here, we summarize the current knowledge on the main cell surface molecules and soluble mediators that modulate the function of NK cells in NB, considering the pros and cons that must be taken into account in the design of novel NK cell-based immunotherapeutic approaches.

Keywords: neuroblastoma, natural killer cells, PVR, B7-H3, TGF-beta, tumor escape mechanisms, immunotherapeutic approaches, chemokine receptors

### **NEUROBLASTOMA, WHERE ARE WE?**

Neuroblastoma is a very heterogeneous disease (1) that includes rare familial (<2%) and sporadic forms. It presents as a locoregional (Stage 1, 2, and 3 or L1, L1/L2, and L2) or progressing metastatic disease involving bone, skin, liver, brain, and bone marrow (BM), the latter being frequently refractory to standard therapy (Stage 4 or M). Moreover, a rare metastatic form of spontaneously regressing/maturating disease (Stage 4S or MS) may occur in children below the age of 18 months (2–5). Over the years, major efforts have been focused to unveil the genetic and biological features of the different forms of NB in order to identify novel prognostic factors and druggable targets. To date, stage, patient's age, and presence or absence of the amplification of MYCN, a transcription factor crucial for central nervous system (CNS) development (6), are considered major predictors of patients' clinical outcome. Moreover, risk-group stratification is based upon additional parameters such as loss of chromosome 11q, tumor histology, and ploidy (3, 5). MYCN amplification strongly associates with loss of heterozygosity at chromosome 1p and occurs in approximately 15% of children affected by NB, with increased frequency in stage 4 patients (approximately 50%). A more accurate evaluation of the risk of treatment failure according to these prognostic factors allowed overtime a significant reduction of mortality in patients that at diagnosis presented with localized non-MYCNamplified tumors (low-risk) and non-MYCN-amplified stage 4 under 18 months of age (intermediated-risk), who received less aggressive surgical and chemotherapeutic treatments. In particular, Rubie and coworkers have recently demonstrated that low-dose chemotherapy improved 5 years survival of 9% in infants with lowrisk NB without threatening symptoms compared to cases treated with high dose chemotherapy due to resistance to the low-dose regimen or presence of one threatening symptom (7). A more dramatic scenario exists in children with high-risk NB at diagnosis, which includes stage 4 patients <18 months with MYCN amplification or >18 months with or without MYCN amplification and rare patients with MYCN-amplified localized tumors (3, 5). These patients have a 5-year survival rate <50% although receiving aggressive combination therapies that include intensive chemotherapy, surgery, radiotherapy, autologous stem cell transplantation, and the administration of Retinoids (13-*cis*-retinoic acid) (3, 5), which represent adjuvants for high-risk NB therapy (8) due to their capability of driving neuronal differentiation *in vitro* (9).

Over the years, several studies focused on the identification of new molecular targets. However, although different experimental approaches have been used, including whole-genome sequencing, few somatic mutations in druggable pathways have been identified. Mutations in the anaplastic lymphoma receptor tyrosine kinase (ALK) are infrequent (<10% of sporadic NB). Mutations in the a-thalassemia/mental retardation syndrome X-linked (ATRX) gene are most common. However, they have not been identified in MYCN-amplified NB and the ATRX molecule does not appear to drive tumorigenesis (10, 11). Recurrent genetic alterations that, however, require further investigations, have been reported in the RAC-RHO pathway and in chromatin-remodeling genes At-rich interactive domain 1A (ARID1A) and 1B (ARID1B) (11-13). Due to the difficulty at targeting directly the MYCN transcription factor, the major oncogenic driver identified so far (14), new strategies currently aim to neutralize molecules involved in apoptosis, angiogenesis, invasion, or metastasis (5). Interestingly, the antibodymediated targeting of the oncofetal differentiation antigen GD2 in combination with GM-CSF and IL-2 resulted in an improved outcome of patients and has been recently included in the standard care of high-risk NB (15). Unfortunately, pain, the major side effect of this therapy (16), as well as cytokine release syndrome and iridoplegia, limits its use; moreover, to date, only patients with minimal residual disease can benefit of this therapeutic approach (17). The success of the anti-GD2/GM-CSF/IL-2 therapy might depend on the FcyR-mediated activation of granulocytes (18) as well of other immune cell types such as macrophages and natural killer (NK) lymphocytes.

Among cytotoxic lymphocytes, NK cells represent the most potent anti-tumor effectors and represent promising weapons against aggressive tumors such as NB (19, 20). In addition, as recently demonstrated, they may also attack cancer stem cells (CSC), i.e. tumor cells with stem cell properties (21–23). Thus, in the past decade, several studies have been focused on the identification of the molecular mechanisms involved in the interaction between NB and NK cells in order to establish a convincing biological starting point for a novel NK cell-based immunotherapeutic approach.

## NATURAL KILLER CELLS AND NEUROBLASTOMA RECOGNITION

Tumor cells express surface molecules that either switch off or switch on NK cell-mediated cytotoxicity. HLA class I molecules on tumors negatively regulate NK cell function by engaging immunoreceptor tyrosine-based inhibition motifs (ITIM)-bearing receptors (24) that include the inhibitory killer Ig-like receptors (KIRs, CD158), highly polymorphic clonally distributed receptors able to distinguish among different HLA-A, -B, and -C allotypes (25), and CD94/NKG2A heterodimers, specific for non-classical HLA-E (26). KIR and CD94/NKG2A are differently expressed in CD56<sup>bright</sup> CD16<sup>-</sup> and CD56<sup>dull</sup> CD16<sup>+</sup> NK cell subsets, which represent sequential stages of maturation (27, 28). While high numbers of CD56<sup>bright</sup> CD16<sup>-</sup>, KIR<sup>-</sup>, IFN- $\gamma$  producing cells are found in non-reactive lymph nodes and tumor sites, the majority of NK cells circulating in the blood are mature CD56<sup>dull</sup> CD16<sup>+</sup>, KIR<sup>+</sup>, cytolytic (perforin<sup>high</sup>) cells (29). CD56<sup>bright</sup> can progress to CD56<sup>dull</sup> cells, which upon activation increase the cytotoxic and IFN- $\gamma$  producing capabilities (28, 30).

Tumor cells switch on NK cell function by expressing at the cell surface non-MHC class I, "danger" molecules that are recognized by an array of activating NK receptors. These include both the FcyRIII (CD16) that mediates the antibody-dependent cytotoxicity (ADCC) and receptors that function in the absence of antibodies. These latter molecules mainly consist of the NKp46, NKp30, and NKp44 receptors, collectively termed natural cytotoxicity receptors (NCR), which are mostly NK-restricted, and NKG2D and DNAM-1 that are also expressed by T cell subsets (31). In the last decade, several groups dedicated many efforts to unveil the nature of the surface tumor ligands specifically recognized by the activating receptors. NKp46 still remains a receptor with "orphan ligand," while NKp44 has been shown to recognize a novel isoform of the mixed-lineage leukemia (MLL5) protein (32) and NKp30 binds to B7-H6 molecules (33). The NKG2D and DNAM-1 (CD226) receptors recognize MICA/B and ULBPs or, PVR (CD155) and Nectin-2 (CD112), respectively (31). DNAM-1 shares the capability of recognizing PVR with tactile (CD96) (34) and TIGIT (35). All the ligands identified so far represent de novo expressed danger signals or molecules that are expressed in healthy cells and over-expressed upon tumor transformation. In most cases, the expression on tumors of multiple ligands leads to the engagement of different activating NK receptors that cooperate in triggering the anti-tumor cytotoxicity (31).

When both inhibitory and activating receptors are engaged by the specific ligands on potential targets, the function of ITIMbearing inhibitory receptors dominates over activation. Thus, in an autologous setting NK lymphocytes spare healthy cells that express high, "protective" levels of HLA class I molecules while they kill tumors, such as NB, in which HLA class I expression is downregulated (36, 37). Moreover, NK cells kill allogeneic targets that express non-self MHC class I alleles. This might occur when two individuals are "KIR/KIR-ligand mismatched," i.e., individual A is characterized by fully functional, educated NK cell subset(s) expressing KIR specific for HLA class I alleles that are missing in individual B. In the context of HLA-haploidentical hematopoietic stem cell transplantation (haplo-HSCT), the presence of a KIR/KIR-ligand mismatch in the donor versus recipient direction allows donor's NK cell subsets to kill recipient's (HLA class I<sup>high</sup>) residual leukemic blasts thus resulting into a graft-versusleukemia effect (GvL), as well as mature DC and activated T lymphocytes, thus preventing graft-versus-host-disease (GvHD) and graft-rejection, respectively (38).

In NB, initial data were obtained by the use, as target cells, of in vitro established NB cell lines (39). NK cells displayed strong cytolytic activity against different NB cells and NCR were involved in the mechanisms leading to killing (39). The susceptibility of human NB cell lines to NK-mediated killing was validated in the context of a metastatic model set up in NOD/SCID mice. In this experimental setting, repeated infusions of IL-2 or IL-15 activated NK cells resulted in both an increased mean survival time of HTLA-230-bearing mice and reduced BM infiltration (40). Considering that long term cultured cell lines might be poorly representative of original tumors, more interesting data originated from in vitro studies that use neuroblasts, freshly purified from BM aspirate of stage 4 patients (36) (Figures 1 and 2). In this study, allogeneic activated NK cells killed neuroblasts isolated from patients although at a lesser extent as compared to the NB cell lines used as control. According to the absence or negligible expression of HLA class I molecules, the NK-mediated lysis of neuroblasts did not increase in the presence of anti-HLA class I mAb. Different molecular mechanisms responsible for the reduced HLA class I expression in NB cells have been elucidated. The immunohistochemical analysis of high-risk human NB showed different abnormalities in the antigen processing machinery, which include defects in the expression of immunoproteasomal subunits LMP2 and LMP7 and of transporters of antigen processing (TAP) (37). In vitro treatment of NB cells with IFN-y induced up-regulation of HLA class I expression (37). Although decreasing their susceptibility to autologous NK cells, this up-regulation of HLA class I molecules could enhance T cell-mediated recognition. In this context, a restoration of killing mediated by antigen specific (MAGE3) cytolytic T cells was observed upon cotransfection of NB cell lines with IRF1 and NF-kB p65, HLA class I transcriptional activators that are also induced by INF $\gamma$  and TNF $\alpha$ , respectively (41). It cannot be excluded that also in vivo neuroblasts could acquire/upregulate HLA class I expression. For example, this might occur in the context of transplantation or following the anti-GD2 antibody-mediated therapy that engages  $Fc\gamma R^+$  immune cells such as NK cells, which are capable of releasing high amounts of IFN-y upon activation (Figure 1). This phenomenon was observed in a murine NB model, where recurrent tumors developed after an NK-dependent anti-tumor response induced by a humanized IL-2 immunocytokine targeted to GD2. In these mice, NB cells showed markedly enhanced MHC class I expression as compared with tumors growing in controls (42). The possible in vivo increase of HLA class I expression in NB cells could explain the benefit of a KIR/KIR-L mismatch in the NK versus NB direction (43-45).

Bone marrow-purified neuroblasts from different patients, although expressing in all instances low levels of HLA class I, displayed a great heterogeneity in terms of susceptibility to lysis by NK cells. The good or poor susceptibility to NK-mediated killing correlated with the presence or the absence of PVR expression on neuroblasts (**Figures 1** and **2A**)(36). Interestingly, neuroblasts lacking PVR expression were from patients in relapse, whereas those expressing PVR were from children at the onset of the disease, suggesting a possible correlation between PVR expression and NB status. The predominant role of DNAM-1/PVR interaction in NK-mediated killing is striking considering that, in most cases, killing of human tumors depends on the cooperation between



ligands for inhibitory KIR

various activating receptors (31). In this context, it is of note that none of the BM-purified neuroblast fractions expressed major adhesion molecules such as ICAM1 or LFA-3 (36). Thus, in NB the presence of PVR, originally described as an adhesion molecule, could become critical in rendering tumor cells susceptible to NKmediated killing. Although PVR and nectin-2 are closely related molecules of the Nectin family, the presence/absence of nectin-2 on NB cells did not influence the susceptibility to NK-mediated killing. Nectin-2, which displays binding affinity to DNAM-1 similar to PVR (46), also shows homophilic recognition. This, by hampering its recognition by DNAM-1, may force the receptor to preferentially bind PVR (46). In line with the increased killing of PVR<sup>+</sup> neuroblasts, mAb mediated masking of DNAM-1 significantly reduced the NK-mediated lysis of neuroblasts showing a pivotal role of DNAM-1 in NB cell killing. While a minor contribution of NKp46 and NKp30 activating receptors could be appreciated (36), consistent with the low expression of the specific ligands, NKG2D did not play a significant role in killing of BM-purified neuroblasts (Figure 1). The expression of NKG2D ligands has been also investigated in human primary NB and cell lines (47). MICA, MICB, and ULBPs transcripts were found in most tumors and cell lines. However, MICA surface expression was absent in primary tumors and present only in some NB cell lines. Importantly, the soluble form of MICA (sMICA) was detected in sera of most patients and in the culture supernatant of some cell lines and was shown to downregulate NKG2D surface expression



in peripheral-blood T cells and decrease NK cell-mediated killing of MICA<sup>+</sup> NB cells (47) (**Figure 2B**). MICB was detected exclusively in the cytosol of NB cells, either primary tumors or cell lines. ULBP-1 was rarely detected, ULBP-2 was expressed by approximately 50% of primary tumors and cell lines, and ULBP-3 was absent in primary tumors but was expressed in most NB cell lines (47).

A crucial role of the DNAM-1 receptor has also been reported in myeloid and lymphoblastic leukemia (48), ovarian tumors (49), and different murine tumor models (50, 51). In particular, it has been shown that in response to chemical carcinogens, DNAM-1-deficient mice develop more DNAM-1 ligand<sup>+</sup> fibrosarcoma and papilloma cells than wt mice. Thus, the study of the molecular mechanisms responsible for the modulation of DNAM-1 ligands appears to be highly relevant. In this context, it has been shown that malignant RAJI lymphoma cells present hypermethylated PVR promoter-associated CpG islands. The epigenetic status was partially reversed by hypomethylating agents that led to upregulation of both transcription and surface expression of PVR (52). PVR up-regulation has been shown to be dependent on DNAdamage response (DDR)-dependent pathways that are activated by oxidative stress (53). Moreover, modulation of PVR surface expression might occur during infections. Different pathogenderived stimuli such as LPS, poly I:C, and flagellin upregulated the surface expression of PVR (and nectin-2) in human DC (54) and in murine antigen-presenting cells (55) via the MYD88 and TRIF pathways. It has also been shown that expression of the human

immunodeficiency virus type 1 (HIV-1) Vpr protein increases PVR levels in Jurkat T cells. This is due to activation of ATR kinase that triggers the DDR pathway and G2 arrest. Moreover, Vpr induces a PVR upregulation in HIV-infected CD4<sup>+</sup> T lymphocytes that overcomes the PVR downregulation induced by the HIV Nef protein (56). A negative regulation of PVR surface expression has been observed during cytomegalovirus (CMV) infection resulting in ineffective NK cell-mediated killing of infected cells (57). The UL141 viral glycoprotein is responsible for PVR downregulation, since it retains the immature form of PVR in the endoplasmic reticulum (57). Recently, it has been shown that UL141 binds to PVR with high affinity (58). Considering the role of PVR in determining high susceptibility to the NK-mediated attack, up-regulation of PVR surface expression might represent an appealing strategy to strengthen the immune responses against tumors. However, the dark side of PVR function cannot be disregarded. Indeed, although data in NB are missing, PVR has been described as a positive regulator of cancer progression, since it is capable of enhancing tumor cell invasiveness and migration (59). Thus, further studies are required to understand whether the immunological favorable effects of PVR up-regulation might balance the induction of a more aggressive tumor phenotype.

A tumor-promoting role has been shown also for B7-H3, a surface molecule originally identified on NB thanks to the generation of a specific monoclonal antibody (M5B14) (60). Studies on different tumor histotype indicate that B7-H3 might drive tumor cell development by different molecular mechanisms. These

include the capability of B7-H3 of reducing the effect of the chemotherapeutic-induced apoptosis in breast (61) and pancreatic carcinoma (62), as well as that of promoting migration and invasiveness (63, 64). The latter properties have been confirmed in vivo in an orthotopic model that showed that xenografted B7-H3 silenced glioma cells invaded significantly less into the surrounding brain tissue as compared to wt tumors. Moreover, B7-H3 expressed on glioblastoma cells negatively regulated NK cell-mediated cytotoxicity (64). These observations are consistent with previous data demonstrating that B7-H3 expression on BM-purified neuroblasts decreased NK cell-mediated killing (60) (Figure 1). Thus, different experimental approaches demonstrated that, in humans, B7-H3 might block the NK-mediated attack by interacting with a still unknown inhibitory NK receptor. A soluble form of B7-H3 (sB7-H3) that results from MMPs cleavage of the surface protein was detected in sera of tumor patients and appeared to correlate with tumor burden (65, 66). Moreover, B7-H3 is present in NB cell lines-derived exosomes (67). The exosomeassociated form could inhibit NK cell function by engaging the B7-H3-specific inhibitory receptor (Figure 2B). Thus, B7-H3 is an interesting NB-associated molecule that combines immuneevading and tumor progression properties. This correlates with the negative prognostic value associated to high expression of B7-H3 in several tumor types (64, 66, 68–72). In particular, in primary NB (73), high B7-H3 surface intensity and percentage of positive cells correlated with poor event-free survival. Interestingly, the differences in event-free survival were observed also in patients with localized disease (stage 1-3), suggesting the need of a more careful follow-up/aggressive treatment of B7-H3<sup>+</sup> low-risk patients. It should be mentioned that, in the mouse system, B7-H3 has been described as a "friend" in tumor immunology (74). In particular, intratumoral injection of an expression plasmid encoding mouse B7-H3 led to complete regression of 50% of tumors, which was mediated by NK and CD8<sup>+</sup> T cells (75). It is to note that the mouse B7-H3 gene codes for a molecule characterized by two Ig-like domains (2Ig-B7-H3) in the order IgV and IgC, while the predominant isoform in human tissues and cell lines is a longer molecule (4Ig-B7-H3) characterized by four Ig-like domains (IgV-IgC-IgV-IgC), which results from duplication of the exons encoding the IgV and IgC domains (76). The mouse B7-H3 has been shown to bind the TREM-like transcript 2 (TREML2, TLT-2) triggering receptor, which is expressed by myeloid cells, CD8<sup>+</sup> T cells, and activated CD4<sup>+</sup> T cells (77, 78). In humans, TREML2 does not appear to be a receptor of B7-H3 (79) and the predominant inhibitory role of B7-H3 strongly suggests the existence of a (still undefined) receptor that down-regulates NK cell function. However, the existence in the human system of specific receptors with opposite functions cannot be excluded. Indeed, human B7-H3 belongs to the B7 family that includes members such as B7-1 (CD80), B7-2 (CD86), B7-H2 (ICOS-L), B7-H1 (PD-L1), B7-DC (PD-L2), B7-H4 (B7S1, B7x), and BT3 (80), some of which represent specific ligands of receptors with either activating or inhibitory function. The scenario should become even more complex when considering that the B7-H3 inhibitory and activating receptors might be co-expressed in immune cells rather than expressed in different cell types/subsets.

## TUMOR ESCAPE MECHANISMS IN NEUROBLASTOMA AND POSSIBLE NOVEL NK CELL-BASED IMMUNOTHERAPEUTIC APPROACHES

In principle, downregulation of HLA class I surface expression allows NB to evade T cell-mediated attack operated by the host immune system (36, 37), while rendering tumor cells susceptible to NK cell-mediated recognition and killing (31). However, both the fast tumor progression in high-risk NB patients and the in vitro data highlighted the existence of additional escape mechanisms that dampen the NK cell-mediated anti-tumor activity as well. As discussed above, the lack of ligands for activating receptors such as PVR (36) (Figure 2A), MICA/ULBPs (36, 37), or adhesion molecules (36, 81) represents a strategy allowing NB cells to strongly reduce their susceptibility to NK-mediated killing. Thus, one could set up immunotherapeutic interventions aimed at upregulating the expression of these ligands. However, the role of PVR in tumor progression (59), as well as its expression in normal cells, should not be underestimated. In particular, DNAM-1 co-operates with NKp30 or NKp46 in the NK-mediated killing of autologous immature DC and unpolarized (M0) or M2-polarized macrophages, which express low, "non-protective" amounts of HLA class I molecules. Conversely, mature DC and M1-polarized macrophages express high levels of HLA class I molecules and are protected by the NK cell-mediated attack (54, 82). The NK-mediated selection of DC and macrophages with optimal antigen-presentation properties has been named "immunoediting" and results from the fine-tuning of inhibitory and activating signals provided by the ligands physiologically expressed by DC and macrophages. The forced over-expression of ligands for activating receptors, such as PVR, might dangerously alter the immunoediting process. Moreover, it should be taken into account that PVR is expressed in normal endothelium, where it plays a role in leukocyte extravasation (83, 84). Also in this case, up-regulation of PVR expression might overcome the HLA class I-mediated inhibitory signals and result in loss of self-tolerance toward autologous endothelium. On the other hand, it has been shown in primary glioblastoma that endothelial cells of proliferating tumor vessels express PVR at higher surface densities as compared to normal vessels (22). Moreover, several studied indicated that most tumor cells including CSC constitutively express levels of PVR higher than the normal counterparts (21). Although further studies are needed to dissect the possible side effects, therapeutic approaches focused on PVR might represent a chance in the treatment of highly aggressive tumors such as NB. Up-regulation of PVR in NB patients might increase the susceptibility to NK-mediated lysis of PVR+ NB and restored that of PVR<sup>-</sup> NB. In this context, it is of note that the lack of PVR expression was detected in given BM metastatic neuroblasts (36) while no data are available on PVR expression in primary tumors. Thus, it cannot be excluded that PVR<sup>-</sup> NB cells might originate because of the selection pressure occurring during the dissemination process and that primary tumors could contain PVR<sup>+</sup> targetable neuroblasts. If so, a larger cohort of patients might benefit from PVR up-regulation therapies. Importantly, PVR might represent a powerful therapeutic target on both neuroblasts and NB-associated vessels. PVR has been originally identified as the receptor for the poliovirus, a highly contagious

virus that only affects humans. As already mentioned in a study performed in a poliovirus-susceptible animal model, the oncolytic treatment by a novel attenuated poliovirus eradicated PVR<sup>+</sup> NB cells without signs of paralysis (85).

B7-H3 might represent an additional attractive molecular target in NB. Beside its tumor-promoting properties, human B7-H3 represents a shield protecting HLA class I low/- neuroblasts from the NK cell-mediated attack (60, 64) (Figure 1). B7-H3 is expressed in normal endothelium, but is strongly upregulated in tumorassociated vasculature (86). Encouraging results derived from the first in-human intrathecal injection of radioiodinated anti-B7-H3 Ab (following surgery, craniospinal irradiation, and chemotherapy) in 21 NB patients with recurrent CNS metastasis (87). Seventeen patients remained CNS disease free and had a median survival time (33 months) significantly better than patients treated with standard protocols (6.6 months) (88). Anti-B7-H3 antibodies, possibly delivered using lipid-based (liposome) formulations (89–91), might represent a suitable option to target tumoricidal compounds to NB and to increase immune responses (Figure 3). Indeed, acting on different cell types, it might simultaneously block invasiveness/migration of tumor cells, destroy tumor-associated vessels, and strength the activity of NK cells. Moreover, anti-B7-H3 antibodies might induce NK cell-mediated ADCC against NB and destroy the B7-H3/inhibitory receptor interactions. An alternative approach to trigger NK cell function might consist in the use of bi-specific antibodies reacting with B7-H3 (on tumor cells) and activating receptors (on NK cells). In addition B7-H3 might be considered as a molecule alternative to GD2 in CAR-based immunotherapy (Figure 3). CAR are chimeric antigen receptors, formed by tumor-specific Ab single chain Fv fragments (scFvs) genetically fused through a transmembrane domain to the CD3 $\zeta$ chain, which are transfected into cytolytic lymphocytes (92, 93). The use of CD19-specific CARs resulted in a significant clinical benefit in two children with B-cell precursor acute lymphoblastic leukemia (94). The NK92 cell line engineered to express the GD2-specific CAR showed increased cytolytic activity against NB cell lines and primary NB cells (95). Moreover, T lymphocytes equipped with GD2-specific CAR are being used in preclinical and phase I studies with clinical benefit in NB patients (96, 97). Essential requirement in CAR-based immunotherapy is the selection of an adequate tumor surface antigen with a documented tumor-promoting role (92). B7-H3 satisfies this criterion, thus representing a suitable candidate in CAR-based immunotherapy. Moreover, while GD2-specific CAR target a molecule that is easily shed by the NB cell surface (98), B7-H3-specific CAR would target a stable type I transmembrane surface molecule expressed by NB cells and NB-associated vessels. Infusion of engineered and/or activated (by cytokine or DC) (99) NK cells might be helpful in adoptive immunotherapeutic approaches of high-risk NB patients receiving autologous (5) or haplo-HSCT. Indeed, a gap of several weeks exists between the infusion of CD34<sup>+</sup> stem cells and the appearance in peripheral blood of fully "armed" mature NK cells (38). During the early post-transplant period, lack of NK-mediated immunosurveillance could favor re-growth of residual NB cells escaping the preparative regimen to the allograft, this causing tumor relapses. Haplo-HSCT, which represents a new standard therapy in the cure of adult and pediatric leukemic



and trigger NK cell function. (A) Anti-B7-H3 antibodies induce ADCC of NK cells against NB. (B) Bi-specific antibodies that react with B7-H3 on NB and trigger activating receptors on NK cells. (C) NK cells engineered with B7-H3-specific CAR that transduce activating signals via CD3 chain.

patients, could also increase the event-free survival of NB patients (45). Recently, a novel transplant strategy based on negative depletion of both  $\alpha/\beta$  T lymphocytes and of CD19+ B-cells has been proposed for pediatric hematological disorders. This approach leaves in the graft an array of immune cells with anti-tumor properties, including mature armed donor-derived alloreactive NK cells and  $\gamma/\delta$  T cells (38). Also in this type of transplant that preserves donor mature NK cells in the graft, engineered NK cells might support the anti-tumor responses. In the perspective of *in vivo* NK-based adoptive immunotherapies as adjuvant in different transplant strategies, recent studies optimized the *in vitro* expansion and activation of NK cells that preserved the anti-NB activity (100–103).

In addition to the escape mechanisms described above (47, 60, 104), NB cells produce soluble factors such as MIF (105, 106) and TGF- $\beta$  (107) capable of dampening NK cell activity (108, 109) (Figure 2B). Initial data showed that in vitro conditioning of NK cells with rTGF-B1 resulted in downregulation of NKp30 and NKG2D activating receptors and in significant reduction of NK-mediated killing (109). Different studies confirmed in vivo the role of TGF-β in regulating NKG2D expression and function in NK cells (110, 111). Although knockout mice lacking TGF-β1 or TGF-β2 showed distinct phenotypes suggesting that each isoform could also have specific, non-overlapping functions (112), in vitro high concentrations of both rTGF- $\beta$ 1 and rTGF- $\beta$ 2 (>10 ng/ml) showed similar capability of inhibiting the expression of the activating NK receptors (113). It has been shown that NB cell lines released low/medium amounts of TGF- $\beta$ 1 (<3 ng/ml), which were unable to significantly downregulate NKp30 and NKG2D expression while clearly modulated the chemokine receptor repertoire of NK cells (113). Interestingly, non-MYCN-amplified NB cells produced amounts of TGF-\u00c61 (and several other soluble factors)

significantly higher than MYCN amplified cells (113). NB-derived TGF-B1 upregulated the expression of CXCR4 and CXCR3 in all NK cells and downregulated that of CX3CR1 in the CD56<sup>dim</sup> subset. Notably, a similar altered chemokine receptor repertoire was observed in peripheral-blood NK cells of stage 4 NB patients resulting in the appearance of a CX3CR1<sup>low</sup> CD56<sup>dim</sup> NK cell population. In patients, also a tendency to CXCR3 upregulation on CD56<sup>bright</sup> NK cells was appreciated. No significant differences in CXCR4 expression were detected in NK cells from NB patients as compared to healthy donors. This might be due to a general difficulty in detecting in vivo CXCR4 expression, which in particular in the BM, might be fully occupied by the CXCL12 (SDF1) ligand (114). In NK cells from NB patients, also a significant CXCR1 down-modulation was observed, that, however, was TGF-B1-independent (113). Thus, in vivo, NB cells can release TGF-β1 and other, still unknown soluble factor(s) that profoundly affect the expression of a number of chemokine receptors that play pivotal roles in NK cell BM homing, egress, interaction with endothelium, and recruitment into peripheral tissues. High concentrations of TGF-\u00b31 might locally exert a paracrine effect on tumor-associated NK cells that might decrease their anti-tumor activity by downregulating NKp30 and NKG2D expression. In this context, it has been shown that CD56<sup>bright</sup>, immature, poor cytolytic NK cells represent the major subset present in tumor tissues (115, 116). The above data suggest that TGF-B1 antagonists and/or proinflammatory cytokines capable of overcoming the modulatory effect of TGF-β might represent a reasonable adjuvant therapy in the cure of different tumors, including NB. In this context, mouse models showed that use of IL-2 targeted to GD2 is associated with increased infiltration of NK (and CD8+ T cells) in subcutaneous NB (117).

Novel immunotherapeutic approaches should also consider the complexity of tumor microenvironment that is populated by different immune cell types that could be endowed with immunomodulatory functions. These cells include fibroblasts (118), regulatory T cells (Treg), myeloid derived suppressor cells (MDSC) (119), and tumor-associated macrophages (TAM) (120). Different NB mouse models demonstrated the immunosuppressive and tumor-promoting role of Treg (120) and MDSC (121). To date, however, data on their presence within the human NB tumor and on their clinical relevance are still scarce. A recent study showed that metastatic NB have higher infiltration of TAM than locoregional tumors, and that metastatic tumors in patients >18 months have higher expression of inflammation-related genes than those in patients <18 months (122). TAM are characterized by an "M2-like" functional phenotype and exert tumor-promoting and immunoregulatory properties, including the release of TGFβ1 (120) (Figure 2). M2-polarized macrophages still present a functional plasticity and they can be recommitted toward an M1-like tumor suppressing, immunostimulatory phenotype. In particular, it has been shown that microbial products such as LPS or BCG reverted their functional phenotype toward M1, which released immunostimulatory cytokines (IL-12, IL-18) and induced strong activation of autologous NK cells. NK cells upregulated CD69 and CD25 (that associates with CD122 to form the high-affinity receptor for IL-2), expressed CCR7 (a chemokine receptor involved in homing of NK cells to secondary lymphoid

organs), increased the anti-tumor cytolytic activity, and released high amounts of IFN- $\gamma$  (82). Interestingly, it has been shown that a subset (30–40%) of M2 express a membrane-bound form of IL-18 (mIL-18) that is released upon TLR stimulation (123, 124). This soluble form of IL-18 (sIL-18) by acting in close cellto-cell contact is crucial for both IFN- $\gamma$  release and expression of CCR7 by NK cells. These data suggest that reconverting the immunosuppressive TAM phenotype using apathogenic TLR ligands might represent an additional immunotherapeutic approach to fully activate immature tumor-associated NK cells.

## **CONCLUSION**

Progress in the understanding of NB cell biology will allow a more accurate stratification of patients, thus reducing toxic side effects of aggressive therapy in low-risk patients. High-risk patients who currently have a dismal prognosis could benefit from multidisciplinary therapeutic protocols that include novel NK cell-based immunotherapeutic strategies. The latter will take advantage of our knowledge about the presence/absence of NB-associated ligands interacting with activating/inhibitory receptors expressed by NK cells. Moreover, it should also take into account the multiple immunomodulatory strategies set up by NB and various immune cell types to impair the recruitment and activation of NK cells in the tumor microenvironment. The encouraging results emerged from haploidentical hematopoietic cell transplantation in pediatric hematological malignancies, might strongly motivate a re-evaluation of transplant approaches in the therapy of high-risk NB patients.

### **ACKNOWLEDGMENTS**

This work was supported by Investigator Grants (10643 and 11650) and special project 5x1000 (9962) from Associazione Italiana per la Ricerca sul Cancro (A.I.R.C.), by Ministero dell'Istruzione, dell'Università e della Ricerca (M.I.U.R), and by Ministero della Salute 5 per mille and Ricerca Corrente, and Progetto di Ricerca Finalizzata RF-IGG-2008-12006685. Francesca Bellora is recipient of a fellowship awarded by A.I.R.C. (special project 5x1000, 9962). We apologize to the colleagues whose work could not be cited because of space constraints.

### **REFERENCES**

- Marusyk A, Almendro V, Polyak K. Intra-tumour heterogeneity: a looking glass for cancer? Nat Rev Cancer (2012) 12:323–34. doi:10.1038/nrc3261
- Brodeur GM. Neuroblastoma: biological insights into a clinical enigma. Nat Rev Cancer (2003) 3:203–16. doi:10.1038/nrc1014
- Cohn SL, Pearson AD, London WB, Monclair T, Ambros PF, Brodeur GM, et al. The International Neuroblastoma Risk Group (INRG) classification system: an INRG Task Force report. *J Clin Oncol* (2009) 27:289–97. doi:10.1200/JCO. 2008.16.6785
- Maris JM. Recent advances in neuroblastoma. N Engl J Med (2010) 362:2202–11. doi:10.1056/NEJMra0804577
- Cheung NK, Dyer MA. Neuroblastoma: developmental biology, cancer genomics and immunotherapy. *Nat Rev Cancer* (2013) 13:397–411. doi:10. 1038/nrc3526
- Grimmer MR, Weiss WA. Childhood tumors of the nervous system as disorders of normal development. *Curr Opin Pediatr* (2006) 18:634–8. doi:10.1097/ MOP.0b013e32801080fe
- 7. Rubie H, De Bernardi B, Gerrard M, Canete A, Ladenstein R, Couturier J, et al. Excellent outcome with reduced treatment in infants with nonmetastatic and unresectable neuroblastoma without MYCN amplification: results of the

prospective INES 99.1. *J Clin Oncol* (2011) **29**:449–55. doi:10.1200/JCO.2010. 29.5196

- Matthay KK, Reynolds CP, Seeger RC, Shimada H, Adkins ES, Haas-Kogan D, et al. Long-term results for children with high-risk neuroblastoma treated on a randomized trial of myeloablative therapy followed by 13-cis-retinoic acid: a children's oncology group study. J Clin Oncol (2009) 27:1007–13. doi:10.1200/JCO.2007.13.8925
- Sidell N. Retinoic acid-induced growth inhibition and morphologic differentiation of human neuroblastoma cells in vitro. J Natl Cancer Inst (1982) 68:589–96.
- Cheung NK, Zhang J, Lu C, Parker M, Bahrami A, Tickoo SK, et al. Association of age at diagnosis and genetic mutations in patients with neuroblastoma. *JAMA* (2012) **307**:1062–71. doi:10.1001/jama.2012.228
- Pugh TJ, Morozova O, Attiyeh EF, Asgharzadeh S, Wei JS, Auclair D, et al. The genetic landscape of high-risk neuroblastoma. *Nat Genet* (2013) 45:279–84. doi:10.1038/ng.2529
- Molenaar JJ, Koster J, Zwijnenburg DA, Van Sluis P, Valentijn LJ, Van Der Ploeg I, et al. Sequencing of neuroblastoma identifies chromothripsis and defects in neuritogenesis genes. *Nature* (2012) 483:589–93. doi:10.1038/nature10910
- Sausen M, Leary RJ, Jones S, Wu J, Reynolds CP, Liu X, et al. Integrated genomic analyses identify ARID1A and ARID1B alterations in the childhood cancer neuroblastoma. *Nat Genet* (2013) 45:12–7. doi:10.1038/ng.2493
- Weiss WA, Aldape K, Mohapatra G, Feuerstein BG, Bishop JM. Targeted expression of MYCN causes neuroblastoma in transgenic mice. *EMBO J* (1997) 16:2985–95. doi:10.1093/emboj/16.11.2985
- Yu AL, Gilman AL, Ozkaynak MF, London WB, Kreissman SG, Chen HX, et al. Anti-GD2 antibody with GM-CSF, interleukin-2, and isotretinoin for neuroblastoma. N Engl J Med (2010) 363:1324–34. doi:10.1056/NEJMoa0911123
- Cheung NK, Lazarus H, Miraldi FD, Abramowsky CR, Kallick S, Saarinen UM, et al. Ganglioside GD2 specific monoclonal antibody 3F8: a phase I study in patients with neuroblastoma and malignant melanoma. *J Clin Oncol* (1987) 5:1430–40.
- Cheung NK, Cheung IY, Kushner BH, Ostrovnaya I, Chamberlain E, Kramer K, et al. Murine anti-GD2 monoclonal antibody 3F8 combined with granulocytemacrophage colony-stimulating factor and 13-cis-retinoic acid in high-risk patients with stage 4 neuroblastoma in first remission. *J Clin Oncol* (2012) 30:3264–70. doi:10.1200/JCO.2011.41.3807
- Cheung IY, Hsu K, Cheung NK. Activation of peripheral-blood granulocytes is strongly correlated with patient outcome after immunotherapy with anti-GD2 monoclonal antibody and granulocyte-macrophage colony-stimulating factor. *J Clin Oncol* (2012) **30**:426–32. doi:10.1200/JCO.2011.37.6236
- 19. Ljunggren HG, Malmberg KJ. Prospects for the use of NK cells in immunotherapy of human cancer. *Nat Rev Immunol* (2007) **7**:329–39. doi:10.1038/nri2073
- Moretta L, Locatelli F, Pende D, Sivori S, Falco M, Bottino C, et al. Human NK receptors: from the molecules to the therapy of high risk leukemias. *FEBS Lett* (2011) 585:1563–7. doi:10.1016/j.febslet.2011.04.061
- Castriconi R, Dondero A, Negri F, Bellora F, Nozza P, Carnemolla B, et al. Both CD133+ and CD133- medulloblastoma cell lines express ligands for triggering NK receptors and are susceptible to NK-mediated cytotoxicity. *Eur J Immunol* (2007) **37**:3190–6. doi:10.1002/eji.200737546
- 22. Castriconi R, Daga A, Dondero A, Zona G, Poliani PL, Melotti A, et al. NK cells recognize and kill human glioblastoma cells with stem cell-like properties. *J Immunol* (2009) **182**:3530–9. doi:10.4049/jimmunol.0802845
- Tallerico R, Todaro M, Di Franco S, MacCalli C, Garofalo C, Sottile R, et al. Human NK cells selective targeting of colon cancer-initiating cells: a role for natural cytotoxicity receptors and MHC class I molecules. *J Immunol* (2013) 190:2381–90. doi:10.4049/jimmunol.1201542
- Long EO, Kim HS, Liu D, Peterson ME, Rajagopalan S. Controlling natural killer cell responses: integration of signals for activation and inhibition. *Annu Rev Immunol* (2013) 31:227–58. doi:10.1146/annurev-immunol-020711-075005
- Parham P. MHC class I molecules and KIRs in human history, health and survival. Nat Rev Immunol (2005) 5:201–14. doi:10.1038/nri1570
- Braud VM, Allan DS, O'Callaghan CA, Soderstrom K, D'Andrea A, Ogg GS, et al. HLA-E binds to natural killer cell receptors CD94/NKG2A, B and C. *Nature* (1998) **391**:795–9. doi:10.1038/35869
- 27. Freud AG, Becknell B, Roychowdhury S, Mao HC, Ferketich AK, Nuovo GJ, et al. A human CD34(+) subset resides in lymph nodes and differentiates into CD56bright natural killer cells. *Immunity* (2005) **22**:295–304. doi:10.1016/j.immuni.2005.01.013

- Romagnani C, Juelke K, Falco M, Morandi B, D'Agostino A, Costa R, et al. CD56brightCD16- killer Ig-like receptor-NK cells display longer telomeres and acquire features of CD56dim NK cells upon activation. *J Immunol* (2007) 178:4947–55.
- 29. Caligiuri MA. Human natural killer cells. *Blood* (2008) **112**:461–9. doi:10.1182/ blood-2007-09-077438
- 30. De Maria A, Bozzano F, Cantoni C, Moretta L. Revisiting human natural killer cell subset function revealed cytolytic CD56(dim)CD16+ NK cells as rapid producers of abundant IFN-gamma on activation. *Proc Natl Acad Sci U S A* (2011) 108:728–32. doi:10.1073/pnas.1012356108
- Bottino C, Castriconi R, Moretta L, Moretta A. Cellular ligands of activating NK receptors. *Trends Immunol* (2005) 26:221–6. doi:10.1016/j.it.2005.02.007
- Baychelier F, Sennepin A, Ermonval M, Dorgham K, Debre P, Vieillard V. Identification of a cellular ligand for the natural cytotoxicity receptor NKp44. *Blood* (2013) 122:2935–42. doi:10.1182/blood-2013-03-489054
- 33. Brandt CS, Baratin M, Yi EC, Kennedy J, Gao Z, Fox B, et al. The B7 family member B7-H6 is a tumor cell ligand for the activating natural killer cell receptor NKp30 in humans. *J Exp Med* (2009) 206:1495–503. doi:10.1084/jem. 20090681
- Fuchs A, Cella M, Giurisato E, Shaw AS, Colonna M. Cutting edge: CD96 (tactile) promotes NK cell-target cell adhesion by interacting with the poliovirus receptor (CD155). *J Immunol* (2004) 172:3994–8.
- 35. Stanietsky N, Simic H, Arapovic J, Toporik A, Levy O, Novik A, et al. The interaction of TIGIT with PVR and PVRL2 inhibits human NK cell cytotoxicity. *Proc Natl Acad Sci U S A* (2009) **106**:17858–63. doi:10.1073/pnas. 0903474106
- 36. Castriconi R, Dondero A, Corrias MV, Lanino E, Pende D, Moretta L, et al. Natural killer cell-mediated killing of freshly isolated neuroblastoma cells: critical role of DNAX accessory molecule-1-poliovirus receptor interaction. *Cancer Res* (2004) 64:9180–4. doi:10.1158/0008-5472.CAN-04-2682
- Raffaghello L, Prigione I, Bocca P, Morandi F, Camoriano M, Gambini C, et al. Multiple defects of the antigen-processing machinery components in human neuroblastoma: immunotherapeutic implications. *Oncogene* (2005) 24:4634–44. doi:10.1038/sj.onc.1208594
- 38. Locatelli F, Pende D, Mingari MC, Bertaina A, Falco M, Moretta A, et al. Cellular and molecular basis of haploidentical hematopoietic stem cell transplantation in the successful treatment of high-risk leukemias: role of alloreactive NK cells. *Front Immunol* (2013) 4:15. doi:10.3389/fimmu.2013.00015
- Sivori S, Parolini S, Marcenaro E, Castriconi R, Pende D, Millo R, et al. Involvement of natural cytotoxicity receptors in human natural killer cell-mediated lysis of neuroblastoma and glioblastoma cell lines. *J Neuroimmunol* (2000) 107:220–5. doi:10.1016/S0165-5728(00)00221-6
- Castriconi R, Dondero A, Cilli M, Ognio E, Pezzolo A, De Giovanni B, et al. Human NK cell infusions prolong survival of metastatic human neuroblastoma-bearing NOD/scid mice. *Cancer Immunol Immunother* (2007) 56:1733–42. doi:10.1007/s00262-007-0317-0
- Lorenzi S, Forloni M, Cifaldi L, Antonucci C, Citti A, Boldrini R, et al. IRF1 and NF-kB restore MHC class I-restricted tumor antigen processing and presentation to cytotoxic T cells in aggressive neuroblastoma. *PLoS One* (2012) 7:e46928. doi:10.1371/journal.pone.0046928
- 42. Neal ZC, Imboden M, Rakhmilevich AL, Kim KM, Hank JA, Surfus J, et al. NXS2 murine neuroblastomas express increased levels of MHC class I antigens upon recurrence following NK-dependent immunotherapy. *Cancer Immunol Immunother* (2004) 53:41–52. doi:10.1007/s00262-003-0435-2
- 43. Venstrom JM, Zheng J, Noor N, Danis KE, Yeh AW, Cheung IY, et al. KIR and HLA genotypes are associated with disease progression and survival following autologous hematopoietic stem cell transplantation for high-risk neuroblastoma. *Clin Cancer Res* (2009) 15:7330–4. doi:10.1158/1078-0432.CCR-09-1720
- 44. Delgado DC, Hank JA, Kolesar J, Lorentzen D, Gan J, Seo S, et al. Genotypes of NK cell KIR receptors, their ligands, and Fcgamma receptors in the response of neuroblastoma patients to Hu14.18-IL2 immunotherapy. *Cancer Res* (2010) 70:9554–61. doi:10.1158/0008-5472.CAN-10-2211
- 45. Perez-Martinez A, De Prada Vicente I, Fernandez L, Gonzalez-Vicent M, Valentin J, Martin R, et al. Natural killer cells can exert a graft-vs-tumor effect in haploidentical stem cell transplantation for pediatric solid tumors. *Exp Hematol* (2012) 40(882-891):e881. doi:10.1016/j.exphem.2012.07.004
- 46. Tahara-Hanaoka S, Shibuya K, Onoda Y, Zhang H, Yamazaki S, Miyamoto A, et al. Functional characterization of DNAM-1 (CD226) interaction with

its ligands PVR (CD155) and nectin-2 (PRR-2/CD112). Int Immunol (2004) 16:533–8. doi:10.1093/intimm/dxh059

- Raffaghello L, Prigione I, Airoldi I, Camoriano M, Levreri I, Gambini C, et al. Downregulation and/or release of NKG2D ligands as immune evasion strategy of human neuroblastoma. *Neoplasia* (2004) 6:558–68. doi:10.1593/neo.04316
- 48. Pende D, Spaggiari GM, Marcenaro S, Martini S, Rivera P, Capobianco A, et al. Analysis of the receptor-ligand interactions in the natural killer-mediated lysis of freshly isolated myeloid or lymphoblastic leukemias: evidence for the involvement of the Poliovirus receptor (CD155) and Nectin-2 (CD112). *Blood* (2005) 105(5):2066–73. doi:10.1182/blood-2004-09-3548
- 49. Carlsten M, Bjorkstrom NK, Norell H, Bryceson Y, Van Hall T, Baumann BC, et al. DNAX accessory molecule-1 mediated recognition of freshly isolated ovarian carcinoma by resting natural killer cells. *Cancer Res* (2007) 67:1317–25. doi:10.1158/0008-5472.CAN-06-2264
- Iguchi-Manaka A, Kai H, Yamashita Y, Shibata K, Tahara-Hanaoka S, Honda S, et al. Accelerated tumor growth in mice deficient in DNAM-1 receptor. *J Exp Med* (2008) 205:2959–64. doi:10.1084/jem.20081611
- Chan CJ, Andrews DM, McLaughlin NM, Yagita H, Gilfillan S, Colonna M, et al. DNAM-1/CD155 interactions promote cytokine and NK cell-mediated suppression of poorly immunogenic melanoma metastases. *J Immunol* (2010) 184:902–11. doi:10.4049/jimmunol.0903225
- Wang W, Gao L, Wang X, Kang H, Li Y, Wang L, et al. Modulation of the poliovirus receptor expression in malignant lymphocytes by epigenetic alterations. J Immunother (2011) 34:353–61. doi:10.1097/CJI.0b013e3182188017
- 53. Ardolino M, Zingoni A, Cerboni C, Cecere F, Soriani A, Iannitto ML, et al. DNAM-1 ligand expression on Ag-stimulated T lymphocytes is mediated by ROS-dependent activation of DNA-damage response: relevance for NK-T cell interaction. *Blood* (2011) 117:4778–86. doi:10.1182/blood-2010-08-300954
- 54. Pende D, Castriconi R, Romagnani P, Spaggiari GM, Marcenaro S, Dondero A, et al. Expression of the DNAM-1 ligands, Nectin-2 (CD112) and poliovirus receptor (CD155), on dendritic cells: relevance for natural killer-dendritic cell interaction. *Blood* (2006) 107:2030–6. doi:10.1182/blood-2005-07-2696
- 55. Kamran N, Takai Y, Miyoshi J, Biswas SK, Wong JS, Gasser S. Toll-like receptor ligands induce expression of the costimulatory molecule CD155 on antigenpresenting cells. *PLoS One* (2013) 8:e54406. doi:10.1371/journal.pone.0054406
- Matusali G, Potesta M, Santoni A, Cerboni C, Doria M. The human immunodeficiency virus type 1 Nef and Vpu proteins downregulate the natural killer cellactivating ligand PVR. J Virol (2012) 86:4496–504. doi:10.1128/JVI.05788-11
- 57. Tomasec P, Wang EC, Davison AJ, Vojtesek B, Armstrong M, Griffin C, et al. Downregulation of natural killer cell-activating ligand CD155 by human cytomegalovirus UL141. *Nat Immunol* (2005) **6**:181–8. doi:10.1038/ni1156
- Nemcovicova I, Benedict CA, Zajonc DM. Structure of human cytomegalovirus UL141 binding to TRAIL-R2 reveals novel, non-canonical death receptor interactions. *PLoS Pathog* (2013) 9:e1003224. doi:10.1371/journal.ppat.1003224
- Sloan KE, Eustace BK, Stewart JK, Zehetmeier C, Torella C, Simeone M, et al. CD155/PVR plays a key role in cell motility during tumor cell invasion and migration. *BMC Cancer* (2004) 4:73. doi:10.1186/1471-2407-4-73
- 60. Castriconi R, Dondero A, Augugliaro R, Cantoni C, Carnemolla B, Sementa AR, et al. Identification of 4Ig-B7-H3 as a neuroblastoma-associated molecule that exerts a protective role from an NK cell-mediated lysis. *Proc Natl Acad Sci U S A* (2004) **101**:12640–5. doi:10.1073/pnas.0405025101
- Liu H, Tekle C, Chen YW, Kristian A, Zhao Y, Zhou M, et al. B7-H3 silencing increases paclitaxel sensitivity by abrogating Jak2/Stat3 phosphorylation. *Mol Cancer Ther* (2011) 10:960–71. doi:10.1158/1535-7163.MCT-11-0072
- 62. Zhao X, Zhang GB, Gan WJ, Xiong F, Li Z, Zhao H, et al. Silencing of B7-H3 increases gemcitabine sensitivity by promoting apoptosis in pancreatic carcinoma. *Oncol Lett* (2013) 5:805–12. doi:10.3892/ol.2013.1118
- Yuan H, Wei X, Zhang G, Li C, Zhang X, Hou J. B7-H3 over expression in prostate cancer promotes tumor cell progression. J Urol (2011) 186:1093–9. doi:10.1016/j.juro.2011.04.103
- 64. Lemke D, Pfenning PN, Sahm F, Klein AC, Kempf T, Warnken U, et al. Costimulatory protein 4IgB7H3 drives the malignant phenotype of glioblastoma by mediating immune escape and invasiveness. *Clin Cancer Res* (2012) 18:105–17. doi:10.1158/1078-0432.CCR-11-0880
- 65. Zhang G, Xu Y, Lu X, Huang H, Zhou Y, Lu B, et al. Diagnosis value of serum B7-H3 expression in non-small cell lung cancer. *Lung Cancer* (2009) **66**:245–9. doi:10.1016/j.lungcan.2009.01.017

- 66. Sun J, Chen LJ, Zhang GB, Jiang JT, Zhu M, Tan Y, et al. Clinical significance and regulation of the costimulatory molecule B7-H3 in human colorectal carcinoma. *Cancer Immunol Immunother* (2010) **59**:1163–71. doi:10.1007/s00262-010-0841-1
- Marimpietri D, Petretto A, Raffaghello L, Pezzolo A, Gagliani C, Tacchetti C, et al. Proteome profiling of neuroblastoma-derived exosomes reveal the expression of proteins potentially involved in tumor progression. *PLoS One* (2013) 8:e75054. doi:10.1371/journal.pone.0075054
- Roth TJ, Sheinin Y, Lohse CM, Kuntz SM, Frigola X, Inman BA, et al. B7-H3 ligand expression by prostate cancer: a novel marker of prognosis and potential target for therapy. *Cancer Res* (2007) 67:7893–900. doi:10.1158/0008-5472. CAN-07-1068
- 69. Zang X, Thompson RH, Al-Ahmadie HA, Serio AM, Reuter VE, Eastham JA, et al. B7-H3 and B7x are highly expressed in human prostate cancer and associated with disease spread and poor outcome. *Proc Natl Acad Sci U S A* (2007) 104:19458–63. doi:10.1073/pnas.0709802104
- Crispen PL, Sheinin Y, Roth TJ, Lohse CM, Kuntz SM, Frigola X, et al. Tumor cell and tumor vasculature expression of B7-H3 predict survival in clear cell renal cell carcinoma. *Clin Cancer Res* (2008) 14:5150–7. doi:10.1158/1078-0432.CCR-08-0536
- Sun TW, Gao Q, Qiu SJ, Zhou J, Wang XY, Yi Y, et al. B7-H3 is expressed in human hepatocellular carcinoma and is associated with tumor aggressiveness and postoperative recurrence. *Cancer Immunol Immunother* (2012) 61:2171–82. doi:10.1007/s00262-012-1278-5
- 72. Wang L, Zhang Q, Chen W, Shan B, Ding Y, Zhang G, et al. B7-H3 is overexpressed in patients suffering osteosarcoma and associated with tumor aggressiveness and metastasis. *PLoS One* (2013) 8:e70689. doi:10.1371/journal.pone. 0070689
- Gregorio A, Corrias MV, Castriconi R, Dondero A, Mosconi M, Gambini C, et al. Small round blue cell tumours: diagnostic and prognostic usefulness of the expression of B7-H3 surface molecule. *Histopathology* (2008) 53:73–80. doi:10.1111/j.1365-2559.2008.03070.x
- 74. Wang L, Kang FB, Shan BE. B7-H3-mediated tumor immunology: friend or foe? *Int J Cancer* (2013). doi:10.1002/ijc.28474
- Sun X, Vale M, Leung E, Kanwar JR, Gupta R, Krissansen GW. Mouse B7-H3 induces antitumor immunity. *Gene Ther* (2003) 10:1728–34. doi:10.1038/sj.gt. 3302070
- Chapoval AI, Ni J, Lau JS, Wilcox RA, Flies DB, Liu D, et al. B7-H3: a costimulatory molecule for T cell activation and IFN-gamma production. *Nat Immunol* (2001) 2:269–74. doi:10.1038/85339
- 77. Hashiguchi M, Kobori H, Ritprajak P, Kamimura Y, Kozono H, Azuma M. Triggering receptor expressed on myeloid cell-like transcript 2 (TLT-2) is a counter-receptor for B7-H3 and enhances T cell responses. *Proc Natl Acad Sci* U S A (2008) 105:10495–500. doi:10.1073/pnas.0802423105
- Kobori H, Hashiguchi M, Piao J, Kato M, Ritprajak P, Azuma M. Enhancement of effector CD8+ T-cell function by tumour-associated B7-H3 and modulation of its counter-receptor triggering receptor expressed on myeloid cell-like transcript 2 at tumour sites. *Immunology* (2010) 130:363–73. doi:10.1111/j.1365-2567.2009.03236.x
- 79. Leitner J, Klauser C, Pickl WF, Stockl J, Majdic O, Bardet AF, et al. B7-H3 is a potent inhibitor of human T-cell activation: no evidence for B7-H3 and TREML2 interaction. *Eur J Immunol* (2009) **39**:1754–64. doi:10.1002/eji. 200839028
- Coyle AJ, Gutierrez-Ramos JC. The expanding B7 superfamily: increasing complexity in costimulatory signals regulating T cell function. *Nat Immunol* (2001) 2:203–9. doi:10.1038/85251
- Carlson LM, Pahlman S, De Geer A, Kogner P, Levitskaya J. Differentiation induced by physiological and pharmacological stimuli leads to increased antigenicity of human neuroblastoma cells. *Cell Res* (2008) 18:398–411. doi:10.1038/cr.2008.27
- 82. Bellora F, Castriconi R, Dondero A, Reggiardo G, Moretta L, Mantovani A, et al. The interaction of human natural killer cells with either unpolarized or polarized macrophages results in different functional outcomes. *Proc Natl Acad Sci U S A* (2010) **107**:21659–64. doi:10.1073/pnas.1007654108
- Reymond N, Imbert AM, Devilard E, Fabre S, Chabannon C, Xerri L, et al. DNAM-1 and PVR regulate monocyte migration through endothelial junctions. J Exp Med (2004) 199:1331–41. doi:10.1084/jem.20032206

- Sullivan DP, Seidman MA, Muller WA. Poliovirus receptor (CD155) regulates a step in transendothelial migration between PECAM and CD99. *Am J Pathol* (2013) 182:1031–42. doi:10.1016/j.ajpath.2012.11.037
- Toyoda H, Yin J, Mueller S, Wimmer E, Cello J. Oncolytic treatment and cure of neuroblastoma by a novel attenuated poliovirus in a novel poliovirussusceptible animal model. *Cancer Res* (2007) 67:2857–64. doi:10.1158/0008-5472.CAN-06-3713
- Seaman S, Stevens J, Yang MY, Logsdon D, Graff-Cherry C, St Croix B. Genes that distinguish physiological and pathological angiogenesis. *Cancer Cell* (2007) 11:539–54. doi:10.1016/j.ccr.2007.04.017
- Kramer K, Kushner BH, Modak S, Pandit-Taskar N, Smith-Jones P, Zanzonico P, et al. Compartmental intrathecal radioimmunotherapy: results for treatment for metastatic CNS neuroblastoma. *J Neurooncol* (2010) **97**:409–18. doi:10.1007/s11060-009-0038-7
- Kramer K, Kushner B, Heller G, Cheung NK. Neuroblastoma metastatic to the central nervous system. The Memorial Sloan-Kettering Cancer Center Experience and A Literature Review. *Cancer* (2001) **91**:1510–9. doi:10.1002/1097-0142(20010415)91:8<1510::AID-CNCR1159>E3.3.CO;2-9
- Kawano K, Hattori Y, Iwakura H, Akamizu T, Maitani Y. Combination therapy with gefitinib and doxorubicin inhibits tumor growth in transgenic mice with adrenal neuroblastoma. *Cancer Med* (2013) 2:286–95. doi:10.1002/cam4.76
- 90. Loi M, Di Paolo D, Soster M, Brignole C, Bartolini A, Emionite L, et al. Novel phage display-derived neuroblastoma-targeting peptides potentiate the effect of drug nanocarriers in preclinical settings. *J Control Release* (2013) 170:233–41. doi:10.1016/j.jconrel.2013.04.029
- 91. Pastorino F, Brignole C, Loi M, Di Paolo D, Di Fiore A, Perri P, et al. Nanocarrier-mediated targeting of tumor and tumor vascular cells improves uptake and penetration of drugs into neuroblastoma. *Front Oncol* (2013) 3:190. doi:10.3389/fonc.2013.00190
- Rossig C. Extending the chimeric receptor-based T-cell targeting strategy to solid tumors. Oncoimmunology (2013) 2:e26091. doi:10.4161/onci.26091
- Savoldo B, Dotti G. Chimeric antigen receptors (CARs) from bench-to-bedside. Immunol Lett (2013) 155:40–2. doi:10.1016/j.imlet.2013.09.014
- 94. Grupp SA, Kalos M, Barrett D, Aplenc R, Porter DL, Rheingold SR, et al. Chimeric antigen receptor-modified T cells for acute lymphoid leukemia. N Engl J Med (2013) 368:1509–18. doi:10.1056/NEJMoa1215134
- 95. Esser R, Muller T, Stefes D, Kloess S, Seidel D, Gillies SD, et al. NK cells engineered to express a GD2-specific antigen receptor display built-in ADCC-like activity against tumour cells of neuroectodermal origin. *J Cell Mol Med* (2012) 16:569–81. doi:10.1111/j.1582-4934.2011.01343.x
- Park JR, Digiusto DL, Slovak M, Wright C, Naranjo A, Wagner J, et al. Adoptive transfer of chimeric antigen receptor re-directed cytolytic T lymphocyte clones in patients with neuroblastoma. *Mol Ther* (2007) 15:825–33. doi:10.1038/sj.mt.6300104
- Pule MA, Savoldo B, Myers GD, Rossig C, Russell HV, Dotti G, et al. Virusspecific T cells engineered to coexpress tumor-specific receptors: persistence and antitumor activity in individuals with neuroblastoma. *Nat Med* (2008) 14:1264–70. doi:10.1038/nm.1882
- Valentino L, Moss T, Olson E, Wang HJ, Elashoff R, Ladisch S. Shed tumor gangliosides and progression of human neuroblastoma. *Blood* (1990) 75: 1564–7.
- Valteau-Couanet D, Leboulaire C, Maincent K, Tournier M, Hartmann O, Benard J, et al. Dendritic cells for NK/LAK activation: rationale for multicellular immunotherapy in neuroblastoma patients. *Blood* (2002) 100:2554–61. doi:10.1182/blood.V100.7.2554
- 100. Cho D, Shook DR, Shimasaki N, Chang YH, Fujisaki H, Campana D. Cytotoxicity of activated natural killer cells against pediatric solid tumors. *Clin Cancer Res* (2010) 16:3901–9. doi:10.1158/1078-0432.CCR-10-0735
- 101. Liu Y, Wu HW, Sheard MA, Sposto R, Somanchi SS, Cooper LJ, et al. Growth and activation of natural killer cells ex vivo from children with neuroblastoma for adoptive cell therapy. *Clin Cancer Res* (2013) 19:2132–43. doi:10.1158/1078-0432.CCR-12-1243
- 102. Rujkijyanont P, Chan WK, Eldridge PW, Lockey T, Holladay M, Rooney B, et al. Ex vivo activation of CD56(+) immune cells that eradicate neuroblastoma. *Cancer Res* (2013) **73**:2608–18. doi:10.1158/0008-5472.CAN-12-3322
- 103. Wang X, Lee DA, Wang Y, Wang L, Yao Y, Lin Z, et al. Membrane-bound interleukin-21 and CD137 ligand induce functional human natural killer cells

from peripheral blood mononuclear cells through STAT-3 activation. *Clin Exp Immunol* (2013) **172**:104–12. doi:10.1111/cei.12034

- 104. Morandi F, Scaruffi P, Gallo F, Stigliani S, Moretti S, Bonassi S, et al. Bone marrow-infiltrating human neuroblastoma cells express high levels of calprotectin and HLA-G proteins. *PLoS One* (2012) 7:e29922. doi:10.1371/journal. pone.0029922
- 105. Bin Q, Johnson BD, Schauer DW, Casper JT, Orentas RJ. Production of macrophage migration inhibitory factor by human and murine neuroblastoma. *Tumour Biol* (2002) 23:123–9. doi:10.1159/000064028
- 106. Ren Y, Chan HM, Fan J, Xie Y, Chen YX, Li W, et al. Inhibition of tumor growth and metastasis in vitro and in vivo by targeting macrophage migration inhibitory factor in human neuroblastoma. *Oncogene* (2006) 25:3501–8. doi:10.1038/sj.onc.1209395
- 107. Cohen PS, Letterio JJ, Gaetano C, Chan J, Matsumoto K, Sporn MB, et al. Induction of transforming growth factor beta 1 and its receptors during alltrans-retinoic acid (RA) treatment of RA-responsive human neuroblastoma cell lines. *Cancer Res* (1995) 55:2380–6.
- Repp AC, Mayhew ES, Apte S, Niederkorn JY. Human uveal melanoma cells produce macrophage migration-inhibitory factor to prevent lysis by NK cells. *J Immunol* (2000) 165:710–5.
- 109. Castriconi R, Cantoni C, Della Chiesa M, Vitale M, Marcenaro E, Conte R, et al. Transforming growth factor beta 1 inhibits expression of NKp30 and NKG2D receptors: consequences for the NK-mediated killing of dendritic cells. *Proc Natl Acad Sci U S A* (2003) **100**:4120–5. doi:10.1073/pnas. 0730640100
- 110. Friese MA, Wischhusen J, Wick W, Weiler M, Eisele G, Steinle A, et al. RNA interference targeting transforming growth factor-beta enhances NKG2D-mediated antiglioma immune response, inhibits glioma cell migration and invasiveness, and abrogates tumorigenicity in vivo. *Cancer Res* (2004) 64:7596–603. doi:10.1158/0008-5472.CAN-04-1627
- 111. Lee JC, Lee KM, Kim DW, Heo DS. Elevated TGF-beta1 secretion and downmodulation of NKG2D underlies impaired NK cytotoxicity in cancer patients. *J Immunol* (2004) **172**:7335–40.
- 112. Shi Y, Massague J. Mechanisms of TGF-beta signaling from cell membrane to the nucleus. *Cell* (2003) 113:685–700. doi:10.1016/S0092-8674(03)00432-X
- 113. Castriconi R, Dondero A, Bellora F, Moretta L, Castellano A, Locatelli F, et al. Neuroblastoma-derived TGF-beta1 modulates the chemokine receptor repertoire of human resting NK cells. *J Immunol* (2013) **190**:5321–8. doi:10.4049/jimmunol.1202693
- 114. Domanska UM, Kruizinga RC, Nagengast WB, Timmer-Bosscha H, Huls G, De Vries EG, et al. A review on CXCR4/CXCL12 axis in oncology: no place to hide. *Eur J Cancer* (2013) **49**:219–30. doi:10.1016/j.ejca.2012.05.005
- Walzer T, Vivier E. G-protein-coupled receptors in control of natural killer cell migration. *Trends Immunol* (2011) 32:486–92. doi:10.1016/j.it.2011. 05.002
- Carrega P, Ferlazzo G. Natural killer cell distribution and trafficking in human tissues. *Front Immunol* (2012) 3:347. doi:10.3389/fimmu.2012.00347
- 117. Yang RK, Kalogriopoulos NA, Rakhmilevich AL, Ranheim EA, Seo S, Kim K, et al. Intratumoral treatment of smaller mouse neuroblastoma tumors with a recombinant protein consisting of IL-2 linked to the hu14.18 antibody increases intratumoral CD8+ T and NK cells and improves survival. *Cancer Immunol Immunother* (2013) **62**:1303–13. doi:10.1007/s00262-013-1430-x
- 118. Balsamo M, Scordamaglia F, Pietra G, Manzini C, Cantoni C, Boitano M, et al. Melanoma-associated fibroblasts modulate NK cell phenotype and antitumor cytotoxicity. *Proc Natl Acad Sci U S A* (2009) **106**:20847–52. doi:10.1073/pnas. 0906481106
- Pistoia V, Morandi F, Bianchi G, Pezzolo A, Prigione I, Raffaghello L. Immunosuppressive microenvironment in neuroblastoma. *Front Oncol* (2013) 3:167. doi:10.3389/fonc.2013.00167
- 120. Mantovani A, Allavena P, Sica A, Balkwill F. Cancer-related inflammation. Nature (2008) 454:436–44. doi:10.1038/nature07205
- 121. Santilli G, Piotrowska I, Cantilena S, Chayka O, D'Alicarnasso M, Morgenstern DA, et al. Polyphenon [corrected] E enhances the antitumor immune response in neuroblastoma by inactivating myeloid suppressor cells. *Clin Cancer Res* (2013) 19:1116–25. doi:10.1158/1078-0432.CCR-12-2528
- 122. Asgharzadeh S, Salo JA, Ji L, Oberthuer A, Fischer M, Berthold F, et al. Clinical significance of tumor-associated inflammatory cells in metastatic neuroblastoma. J Clin Oncol (2012) 30:3525–32. doi:10.1200/JCO.2011.40.9169

- 123. Bellora F, Castriconi R, Doni A, Cantoni C, Moretta L, Mantovani A, et al. M-CSF induces the expression of a membrane-bound form of IL-18 in a subset of human monocytes differentiating in vitro toward macrophages. *Eur J Immunol* (2012) **42**:1618–26. doi:10.1002/eji.201142173
- 124. Dinarello CA, Novick D, Kim S, Kaplanski G. Interleukin-18 and IL-18 binding protein. *Front Immunol* (2013) **4**:289. doi:10.3389/fimmu.2013.00289

**Conflict of Interest Statement:** Alessandro Moretta is a founder and shareholder of Innate Pharma (Marseille, France). The remaining authors declare no conflicts of interest.

Received: 20 December 2013; paper pending published: 14 January 2014; accepted: 30 January 2014; published online: 12 February 2014.

Citation: Bottino C, Dondero A, Bellora F, Moretta L, Locatelli F, Pistoia V, Moretta A and Castriconi R (2014) Natural killer cells and neuroblastoma: tumor recognition, escape mechanisms, and possible novel immunotherapeutic approaches. Front. Immunol. 5:56. doi: 10.3389/fimmu.2014.00056

This article was submitted to NK Cell Biology, a section of the journal Frontiers in Immunology.

Copyright © 2014 Bottino, Dondero, Bellora, Moretta, Locatelli, Pistoia, Moretta and Castriconi. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) or licensor are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.