

# Mixing signals: molecular turn ons and turn offs for innate $\gamma \delta$ T-cells

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INTRODUCTION

The ability to generate antigen receptor diversity by somatic recombination evolved approximately 500 million years ago (1) and became the founding biological property of what we now know as adaptive immunity. This evolutionary milestone provided our immune system with an innate and an adaptive arm that synergized for the fight against infection and the recognition of oncogenesis. Lymphocytes of the gamma delta ( $\gamma\delta$ ) T-cell lineage are evolutionary conserved among species (2) and although they express rearranged antigen-specific receptors, a large proportion display innate properties. In the mouse, where innate  $\gamma\delta$ T-cells have been mostly studied, approximately 25% of lymph node y8 T-cells respond rapidly to cytokine stimuli similar to innate lymphoid cells (ILCs) and appear to have reduced T-cell receptor (TCR) signaling capacity (3). Innate y8 T-cells are characterized by the spontaneous and high expression of interleukin (IL)-17 ( $\gamma \delta^{17}$ ) as well as IL-22 and express functional Toll-like receptors (TLR) (4, 5). Importantly, IL-17 and IL-23 receptor (IL-23R) expression, which is critical for IL-22 induction, are turned on during embryonic development in the thymus strongly pointing toward a bona fide innate nature (6-8). Although a new interferon gamma (IFNγ)-producing innate γδ T-cell subset with no IL-17 potential has recently been described (3), this review will discuss briefly some of the key cytokines, cytokine receptors, and transcription factors (TFs) that regulate the development, activation, and inhibition of mouse innate  $\gamma \delta^{17}$  cells (Figure 1).

Lymphocytes of the gamma delta ( $\gamma\delta$ )T-cell lineage are evolutionary conserved and although they express rearranged antigen-specific receptors, a large proportion respond as innate effectors.  $\gamma\delta$ T-cells are poised to combat infection by responding rapidly to cytokine stimuli similar to innate lymphoid cells. This potential to initiate strong inflammatory responses necessitates that inhibitory signals are balanced with activation signals. Here, we discuss some of the key mechanisms that regulate the development, activation, and inhibition of innate  $\gamma\delta$ T-cells in light of recent evidence that the inhibitory immunoglobulin-superfamily member B and T lymphocyte attenuator restricts their differentiation and effector function.

Keywords: BTLA, dermatitis,  $\gamma\delta$  T-cell, IL-7, lymphotoxin, RORyt

# IL-23 AND IL-1β: KEY PROINFLAMMATORY AND ANTI-BACTERIAL MEDIATORS

Innate  $\gamma\delta^{17}$  cells localize mainly at barrier and mucosal surfaces such as the skin, gut, and lung (9) and within the lymph nodes, they position themselves in close proximity to the subcapsular sinus and interfollicular regions both of which specialize in the capture of antigen (10). Therefore, infectious and inflammatory stimuli can readily activate  $\gamma\delta^{17}$  cells either directly through TLR ligation or through cytokines such as IL-23 and IL-1 $\beta$  that are produced by local innate sensors.

IL-23 induces the expression of IL-17 and IL-22 as well as the transcription factor retinoid-related orphan receptor gamma-t (ROR $\gamma$ t) in T-helper 17 (T<sub>H</sub>17) cells while at the same time promoting survival and cell proliferation (11).  $\gamma \delta^{17}$  cells express functional IL-23R as early as embryonic day E18 in the thymus (7), in contrast to CD4<sup>+</sup> T-cells that upregulate the IL-23R upon T<sub>H</sub>17 differentiation (12). Although IL-23 or IL-23R has not been reported to be important for  $\gamma \delta$  T-cell development, they enhance the production of IL-17 and IL-22 and can promote cellular proliferation (3, 13). *In vivo* infectious and inflammatory models have shown that IL-23 can be important for the activation of the  $\gamma \delta$  T-cell response.

During imiquimod (IMQ)-induced psoriasis, genetic ablation of IL-23 or IL-23R results in a significant reduction of IL-17 production by  $\gamma \delta^{17}$  cells, diminished accumulation of these cells in the skin, and a subsequent decrease in inflammatory symptoms (14– 16). In this model, IL-23 is produced locally in the skin by resident



FIGURE 1 | Major pathways that regulate  $\gamma \delta^{17}$  T-cells. Signals in thymic progenitors (TP): during development, RANK co-ordinates Skint-1 expression, which regulates the fate decision of thymic progenitors into  $\gamma \delta^{17}$  or V $\gamma 5$ /IFNy  $\gamma \delta$  T-cells. IL-7 is critical for the development of  $\gamma \delta^{17}$  cells from thymic progenitors. In the adults, IL-7 is also critical for the normal homeostasis, function, expansion, and survival of adult  $\gamma \delta^{17}$ . BTLA (and perhaps other inhibitory receptors) suppress overt homeostatic proliferation and hyperactivation in part by regulating IL-7 reells in the adult, likely through differentiation of the microenvironment. IL-23 and IL-1 $\beta$  are both critical cytokines that initiate inflammatory  $\gamma \delta^{17}$  responses.

macrophage and dendritic cell (DC) populations that receive a combination of TLR and neuronal signals (15, 17, 18). The onset of experimental autoimmune encephalomyelitis (EAE), which is often used to model human multiple sclerosis, also depends to a certain extent on IL-23-driven IL-17 production by γδ T-cells (5, 19). More specifically, it has been shown that IL-23-activated  $\gamma \delta^{17}$  cells are important for optimal T<sub>H</sub>17 polarization (5) and the suppression of regulatory T-cell responses (19). In a mouse model of brain ischemic injury, absence of IL-23 also abrogated  $\gamma \delta^{17}$ -induced inflammation (20). In addition to regulating inflammatory reactions,  $\gamma \delta^{17}$  cells and IL-23 have been linked with protection from a number of bacterial infections. Thus, cutaneous infection with Staphylococcus aureus triggers a y8 T-cell orchestrated IL-17 response that depends on the combined effects of IL-23 and IL-1β (21). Furthermore, infection with Listeria monocytogenes elicits an IL-23-driven  $\gamma \delta^{17}$  response that is important for bacterial clearance (22, 23), and the IL-23 pathway appears also to operate during  $\gamma \delta^{17}$  activation by Mycobacterium tuberculosis (24). Together, these data highlight the role of IL-23 in activating  $\gamma \delta^{17}$  cell-induced inflammatory responses, both to pathogens and in driving autoimmune disease.

Similar to IL-23, IL-1 $\beta$  has also been linked with IL-17-related immunity both in CD4<sup>+</sup> T as well as in innate  $\gamma\delta$  T-cells.  $\gamma\delta^{17}$  cells constitutively express the IL-1 receptor and respond to *in vitro* IL-1 $\beta$  stimulation by rapid proliferation and upregulation of IL-17 (3, 5, 13). Interestingly, IL-1 $\beta$  appears to be important for IL-23-mediated  $\gamma\delta$  T-cell expansion and IL-17 production although the molecular mechanism is not yet understood (5, 13). Effective IL-1 $\beta$  signaling was critical for  $\gamma\delta$  T-cell activation and disease progression in the EAE model (5). However, during IMQinduced psoriasis, usage of *Ilr*1<sup>-/-</sup> mice has resulted in conflicting conclusions. Whereas an earlier report presented no impact of IL-1 $\beta$  on either dermatitis or  $\gamma\delta^{17}$  activation (25), a more recent study showed that *Ilr*1<sup>-/-</sup> mice were consistently protected with severely compromised  $\gamma\delta$  T-cell responses (13). A key difference in the two studies was the site of inflammation: ear (no IL-1 $\beta$  effect) (25) versus dorsal epidermis (strong IL-1 $\beta$  effect) (13), suggesting that IL-1 $\beta$  may have site-specific regulatory roles, such as differential effects on resident stromal and epithelial cells or due to differences in lymphatic drainage.

# IL-7: KEEPING THE BALANCE BETWEEN HOMEOSTASIS AND INFLAMMATION

IL-7 is one of the best-studied T-cell homeostatic cytokines. IL-7 deficiency is associated with lymphopenia and dysfunction of naïve and memory T-cell subsets (26). IL-7 is essential for the development of  $\gamma\delta$  T-cells (27, 28) by regulating the survival of early thymic progenitors and by inducing V(D)J recombination within the TCR- $\gamma$  locus (29, 30). Further experiments have shown that in addition to its developmental role, IL-7 supports the homeostatic proliferation of γδ T-cells (31). Although IL-7 is strongly associated with signaling via the signal transducer and activator of transcription 5 (STAT5) (32), it has been shown to induce STAT3 phosphorylation in diverse lymphocyte populations such as thymocytes (33), B-cell progenitors (34), and y8 T-cells (35). STAT3 is a critical component of the IL-23 and IL-6 signaling pathways, which are important for the differentiation of CD4<sup>+</sup> T-cells into the  $T_H 17$  lineage (11, 36), in part by antagonizing STAT5 (37). Of the γδ T subsets, IL-7 was found to preferentially expand and activate innate  $\gamma \delta^{17}$  cells in a STAT3-dependent manner (35), although it sustained survival of all  $\gamma\delta$  T-cells (38).

We have recently demonstrated that in  $\gamma \delta^{17}$  cells, STAT5mediated IL-7 signaling induces surface expression of the checkpoint receptor B and T lymphocyte attenuator (BTLA), which is necessary for their normal homeostasis and activation during skin inflammation (38). Blockade of IL-7 signaling itself has been shown to acutely diminish  $\gamma \delta^{17}$ -driven dermatitis (35) while during viral hepatitis IL-7 co-operates with IL-23 to rapidly activate intrahepatic  $\gamma \delta^{17}$  cells and initiate inflammation (39). Whether IL-7-induced STAT5 and STAT3 phosphorylation operate in parallel, sequentially, or as mutually exclusive processes within the  $\gamma \delta^{17}$  population is unknown. However,  $\gamma \delta$  T-cells deficient in STAT3 display normal homeostatic responses (40) suggesting that at steady state STAT5 may have a dominant role.

In addition to its direct effects on  $\gamma\delta$  T-cells, IL-7 indirectly influences innate  $\gamma\delta$  T-cell development by promoting the generation of lymphoid tissues in part by inducing the expression of tumor necrosis factor (TNF) superfamily members. IL-7 is produced homeostatically in the developing thymus and lymph node anlagen (41) and has been shown to induce the expression of surface lymphotoxin- $\alpha\beta$  (LT $\alpha\beta$ ) on resident embryonic lymphoid tissue inducer (LTi) cells (42). LT $\alpha\beta$  expressed by LTi interacts with the LT $\beta$  receptor (LT $\beta$ R) in order to initiate lymph node development and organization (43, 44). Genetic ablation of LT $\beta$ R results in the absence of all secondary lymphoid tissues in addition to disorganized splenic and thymic architecture (45, 46). Several members of the TNF superfamily have been shown to directly regulate  $\gamma\delta$  T-cell development, homeostasis, and function, as outlined below.

### LYMPHOTOXIN AND THE TNF NETWORK: CRITICAL REGULATORS OF INNATE γδ T-CELLS

Innate IL-17 producing y8 T-cells as well as Vy5 (Vy3 in Garman nomenclature) expressing cells that colonize the skin as resident dendritic epidermal T-cells (DETCs) are strictly dependent on the embryonic microenvironment (8, 47). Thus, adult progenitors cannot reconstitute either of the aforementioned populations even if they are provided with a fetal thymus suggesting the need for embryonic-only progenitors (8). Thus, the fetal thymus contains fully functional  $\gamma \delta^{17}$  cells that develop between E15-18 (8). The development of these cells is intimately associated with the TNF superfamily since as early as E15  $V\gamma 5^+$ progenitors express the TNF ligand RANKL (receptor activator of NF-kB ligand) and condition the thymic medulla to upregulate Skint-1 (48), an immunoglobulin (Ig) superfamily protein that is necessary for the development of V $\gamma$ 5 cells (49–51). Interestingly, in Skint-1 deficient animals, Vy5 cells are reprogramed into a  $\gamma \delta^{17}$ -like phenotype with severely reduced IFN $\gamma$  production (52). This suggests that innate  $\gamma \delta^{17}$  T-cells are likely to represent the default differentiation pathway of most γδ T-cell progenitors pre-Skint-1 selection. This is in line with the evolutionary evidence that IL-17-producing γδ T-cells are conserved between non-jawed vertebrates and human beings (2) while Skint-1 and related genes (e.g., Btn1a1) are highly restricted to mammals (www.ensembl.org).

In addition to RANK, LTBR has also been linked with the development and functional maturation of γδ T-cells. Early reports showed that yo T-cells can acquire LTBR expression in the thymus, and that activation of these receptors by LTaβ- and LIGHTexpressing double-positive (DP) thymocytes drives maturation of y8 T-cells assessed by the production of IFNy (53). However, the expression of IL-17 or other  $\gamma \delta^{17}$ -related properties was not evaluated. The authors suggested that LTBR-induced maturation likely occurred at the late stages of thymic development when DP cells predominate. Given that  $\gamma \delta^{17}$  T-cells develop during early embryonic life (8), one scenario to explain these findings is that during thymic development the LTBR pathway in part regulates the IFNy potential of y8 T-cells, presumably following Skint-1 selection. In agreement with this argument, the TNF receptor CD27 is required by thymic progenitors to induce the innate IFNy-related differentiation program and to sustain expression of LT $\beta$ R (7). Thus, while CD27 deficient animals retain an intact  $\gamma \delta^{17}$  compartment, they showed a marked reduction in IFNy and LTBR expression (54). These results predict that LT $\beta$ R signaling is not absolutely necessary for  $\gamma \delta^{17}$  development and function, although mice deficient in LTBR or its ligands had very few IL-17-producing γδ T-cells in the spleen and thymus (55). Mice lacking the NF-KB TFs RelA and RelB also showed reduced IL-17-producing γδ T-cells (55). Since the NF-κB pathway is central to TCR signaling and T-cell development (56), low

IL-17 production might be reflective of impaired TCR stimulation rather than loss of LT $\beta$ R signals. Furthermore, lack of lymph nodes in LT $\beta$ R deficient mice (45) may relocate  $\gamma \delta^{17}$  cells to the skin or intestine and thus explain their reduced numbers in the spleens. Importantly, loss of LT $\beta$ R results in abnormal thymic organization and maturation of the medullary epithelium (46, 57), which may negatively affect  $\gamma \delta^{17}$  T-cell development. Alternatively, organized secondary lymphoid tissues may be important for the survival and steady-state turnover of  $\gamma \delta^{17}$  cells. Of note, LT $\beta$ R has been shown to participate in the production of IL-7 by fibroreticular stromal cells in the lymph node (58), which might explain why deficiency in LT $\beta$ R can result in reduced  $\gamma \delta^{17}$ responses.

In addition to its involvement in stromal cell development, LT\betaR is expressed on tissue resident DCs and macrophages (59) both of which have been linked with the IL-23-mediated activation of  $\gamma \delta^{17}$  T-cells, whether this is in the context of skin (15, 17) or brain inflammation (5). Notably, LTβR regulates the homeostasis of DCs (60, 61) and can directly induce their production of IL-23 (62). Interestingly, an LTβR-LTαβ interaction has been linked with the production of IL-22 by intestinal ILCs (63, 64) raising the possibility that a similar mechanism may be in place at sites where  $\gamma \delta^{17}$  cells preferentially localize, such as the skin.

# BTLA AND INHIBITORY RECEPTORS: PUTTING THE BRAKES ON

In human beings, herpesvirus entry mediator (HVEM) interacts with the two TNF ligands LIGHT (shared with LT $\beta$ R) and soluble LT $\alpha$ , and the Ig superfamily members CD160 and BTLA. BTLA is an inhibitory receptor with an immunoreceptor tyrosine inhibitory motif (ITIM) that has been shown to interact with the Src homology 2 (SH2)-domain containing protein tyrosine phosphatase 1 (SHP1) and SHP2 and to inhibit T-cell activation (65–67) upon interacting with HVEM, its only identified ligand thus far (66, 68, 69). In addition to its inhibitory role in T-cell responses, BTLA was shown to prevent overt TLR stimulation in DCs (70) and to diminish cytokine production by natural killer T (NKT) (71) and follicular T-cells (72) suggesting a regulatory role both in adaptive and innate immunity.

BTLA and HVEM signal bi-directionally providing inhibitory signals in T-cells and survival signals in cells expressing HVEM (68). BTLA expression varies  $\sim 10^3$  fold among hematopoietic lineages, and co-expressed with HVEM forming a complex in cis that may contribute to homeostatic signaling (73). Constitutive surface expression of BTLA (74) implicates a unique ability among inhibitory receptors to sustain the homeostatic balance of T-cells (75) and DCs (61). Similarly, our recent data showed that BTLA is necessary to inhibit homeostatic expansion and activation of lymph node and skin resident  $\gamma \delta^{17}$  T-cells (38).  $\gamma \delta^{17}$  but not other γδ T-cell subsets deficient in Btla were hyperresponsive to IL-7 stimulation suggesting that BTLA diminishes IL-7 receptor (IL-7R) signaling. Interestingly, IL-7 increased surface BTLA on  $\gamma \delta^{17}$ cells in a STAT5-dependent way revealing the presence of a negative feedback loop between IL-7 and BTLA (38) (Figure 2). Given the broad range of SHP1 and SHP2 targets (76), it is likely that these phosphatases can inactivate both STAT3 and STAT5 in response to IL-7. However, the exact molecular details of BTLA-mediated



suppression of IL-7R or other  $\gamma \delta^{17}$ -expressed cytokine receptors are currently not known.

Although there are numerous functional inhibitory receptors that have been reported on the surface of lymphocyte subsets either at steady state or after activation, there is little information regarding their role on innate or non-innate yo T-cells. Several reports have mapped the expression of inhibitory molecules like programed death-1 (PD-1) (77, 78), lymphocyte activation gene-3 (LAG-3) (79), CD200 (80), Tim-3 (81), CD160 (82), and cytotoxic T lymphocyte antigen-4 (CTLA-4) (83) on human or murine γδ T-cells but the capacity to target these receptors using agonistic or antagonistic manipulation has in general not been addressed. Notably, we found that activating BTLA receptors using an agonistic antibody limited pathology in mice (38). Additionally, blockade of BTLA signaling enhanced activation of lymphomaspecific human Vy9V82 T-cells (84). Thorough investigation of the expression patterns and function of the different inhibitory receptors on innate  $\gamma\delta$  T-cells may provide promising targets for intervening when these lymphocytes need to be turned on or off. Currently, and in combination with its suppressive activity, BTLA appears to be a key targetable pathway for regulating innate  $\gamma\delta$ T-cells.

# TRANSCRIPTIONAL CONTROL: IS EVERYTHING PRE-PROGRAMED?

It is now well-appreciated that there is an extensive network of TFs that are expressed early in pre-committed progenitors and are necessary for the development, functional differentiation, and survival

of all innate cells including  $\gamma \delta^{17}$  T-cells. A subset of these TFs control lineage specification, either through activating or repressing gene transcription. A number of TF mouse knockout lines result in the complete abolishment or severe reduction in the numbers of the  $\gamma \delta^{17}$  subset in the periphery and in the thymus. Thus, mice deficient for the high-mobility group (HMG) box TFs Sox13 and Sox4 show severe reduction of IL-17-producing γδ T-cells due to a differentiation block early on during development (85, 86), which correlates with high expression levels of Sox13 and Sox4 in  $v\delta^{17}$ -comitted T-cell progenitors (86–88). Interestingly, the function of Sox13 can be counteracted embryonically by Egr3, which drives the DETC differentiation program and IFNy expression (52), while TCF1, another HMG box TF, suppresses  $\gamma \delta^{17}$  differentiation (86). Notch signaling turns on TCF1 (89), which can also induce expression of Hes1, another TF critical for the generation of  $\gamma \delta^{17}$  cells during embryonic differentiation (40). Interestingly, a subset of innate y8 T-cells has been shown to depend on the expression of promyelocytic zinc finger (PLZF), which is also required for the development of ILCs (90, 91). It remains to be seen whether PLZF is specifically required for the development of  $\gamma \delta^{17}$  cells.

Although, RORyt is necessary for the differentiation of T<sub>H</sub>17 cells (36), it is not essential for the development of  $\gamma \delta^{17}$  progenitors in the fetal thymus (40). However, consistent with its ability to bind to and transactivate the Il17 promoter (92), RORyt is important for optimal IL-17 production (40). Interestingly, despite being developed, RORyt deficient y8 T-cells cannot persist in the periphery (40), suggesting a potentially critical role for RORyt in the homeostasis of adult  $\gamma \delta^{17}$  T-cells. This could be either cellextrinsic or cell-intrinsic. RORyt is necessary for the development of all secondary lymphoid tissues (93). Thus, upon export in the periphery,  $\gamma \delta^{17}$  T-cells may not have the appropriate microenvironment in order to sustain homeostasis (cell-extrinsic). In the cell-intrinsic scenario, RORyt may be important for the survival of  $\gamma \delta^{17}$  cells by regulating the levels of the anti-apoptotic protein Bcl-xL (93). Our data have demonstrated that via its interaction with LxxLL containing nuclear co-factors RORyt can function as a transcriptional repressor and suppress expression of BTLA (38) (Figure 2). Therefore, an alternative cell-intrinsic hypothesis is that loss of RORyt results in aberrant expression of BTLA and perhaps other co-inhibitory receptors (such as LAG-3; Bekiaris/Ware, unpublished observations) leading to a sustained inhibition of homeostatic expansion.

### **CONCLUSION**

 $\gamma \delta^{17}$  and other  $\gamma \delta$  T-cell subsets comprise a unique family of lymphocytes that provides an innate powerhouse to the immune system. The innate nature of  $\gamma \delta^{17}$  cells is demonstrable by a number of key biological properties including rapid response to cytokines, functional maturation during embryogenesis, largely TCR-independent responses, and TF-dependent lineage commitment. Resolving the complex and fascinating biology of these cells has been breaking the Frontiers of Immunology for a number of years and has taught us a great deal about how lymphocytes develop and function. The continued knowledge of how all innate  $\gamma \delta$  T-cells work will certainly push forward these frontiers and perhaps allow us to develop tools in order to manipulate them for the treatment of human disease.

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### **REFERENCES**

- Flajnik MF, Kasahara M. Origin and evolution of the adaptive immune system: genetic events and selective pressures. *Nat Rev Genet* (2010) 11:47–59. doi:10.1038/nrg2703
- Hirano M, Guo P, Mccurley N, Schorpp M, Das S, Boehm T, et al. Evolutionary implications of a third lymphocyte lineage in lampreys. *Nature* (2013) 501:435–8. doi:10.1038/nature12467
- Wencker M, Turchinovich G, Di Marco Barros R, Deban L, Jandke A, Cope A, et al. Innate-like T cells straddle innate and adaptive immunity by altering antigen-receptor responsiveness. *Nat Immunol* (2014) 15:80–7. doi:10.1038/ ni.2773
- Martin B, Hirota K, Cua DJ, Stockinger B, Veldhoen M. Interleukin-17producing gammadelta T cells selectively expand in response to pathogen products and environmental signals. *Immunity* (2009) 31:321–30. doi:10.1016/j. immuni.2009.06.020
- Sutton CE, Lalor SJ, Sweeney CM, Brereton CF, Lavelle EC, Mills KH. Interleukin-1 and IL-23 induce innate IL-17 production from gammadelta T cells, amplifying Th17 responses and autoimmunity. *Immunity* (2009) **31**:331–41. doi:10.1016/j. immuni.2009.08.001
- Jensen KD, Su X, Shin S, Li L, Youssef S, Yamasaki S, et al. Thymic selection determines gammadelta T cell effector fate: antigen-naive cells make interleukin-17 and antigen-experienced cells make interferon gamma. *Immunity* (2008) 29:90–100. doi:10.1016/j.immuni.2008.04.022
- Ribot JC, Debarros A, Pang DJ, Neves JF, Peperzak V, Roberts SJ, et al. CD27 is a thymic determinant of the balance between interferon-gamma- and interleukin 17-producing gammadelta T cell subsets. *Nat Immunol* (2009) 10:427–36. doi:10.1038/ni.1717
- Haas JD, Ravens S, Duber S, Sandrock I, Oberdorfer L, Kashani E, et al. Development of interleukin-17-producing gammadelta T cells is restricted to a functional embryonic wave. *Immunity* (2012) 37:48–59. doi:10.1016/j.immuni.2012. 06.003
- 9. Vantourout P, Hayday A. Six-of-the-best: unique contributions of gammadelta T cells to immunology. *Nat Rev Immunol* (2013) **13**:88–100. doi:10.1038/nri3384
- Kastenmuller W, Torabi-Parizi P, Subramanian N, Lammermann T, Germain RN. A spatially-organized multicellular innate immune response in lymph nodes limits systemic pathogen spread. *Cell* (2012) 150:1235–48. doi:10.1016/j.cell. 2012.07.021
- 11. Bettelli E, Carrier Y, Gao W, Korn T, Strom TB, Oukka M, et al. Reciprocal developmental pathways for the generation of pathogenic effector TH17 and regulatory T cells. *Nature* (2006) **441**:235–8. doi:10.1038/nature04753
- Yosef N, Shalek AK, Gaublomme JT, Jin H, Lee Y, Awasthi A, et al. Dynamic regulatory network controlling TH17 cell differentiation. *Nature* (2013) 496:461–8. doi:10.1038/nature11981
- Cai Y, Xue F, Fleming C, Yang J, Ding C, Ma Y, et al. Differential developmental requirement and peripheral regulation for dermal Vgamma4 and Vgamma6T17 cells in health and inflammation. *Nat Commun* (2014) 5:3986. doi:10.1038/ncomms4986
- 14. van der Fits L, Mourits S, Voerman JS, Kant M, Boon L, Laman JD, et al. Imiquimod-induced psoriasis-like skin inflammation in mice is mediated via the IL-23/IL-17 axis. J Immunol (2009) 182:5836–45. doi:10.4049/jimmunol. 0802999
- Cai Y, Shen X, Ding C, Qi C, Li K, Li X, et al. Pivotal role of dermal IL-17-producing gammadelta T cells in skin inflammation. *Immunity* (2011) 35:596–610. doi:10.1016/j.immuni.2011.08.001
- 16. Pantelyushin S, Haak S, Ingold B, Kulig P, Heppner FL, Navarini AA, et al. Rorgammat+ innate lymphocytes and gammadelta T cells initiate psoriasiform plaque formation in mice. *J Clin Invest* (2012) **122**:2252–6. doi:10.1172/ JCI61862

- Riol-Blanco L, Ordovas-Montanes J, Perro M, Naval E, Thiriot A, Alvarez D, et al. Nociceptive sensory neurons drive interleukin-23-mediated psoriasiform skin inflammation. *Nature* (2014) 510:157–61. doi:10.1038/nature13199
- 18. Yoshiki R, Kabashima K, Honda T, Nakamizo S, Sawada Y, Sugita K, et al. IL-23 from Langerhans cells is required for the development of imiquimod-induced psoriasis-like dermatitis by induction of IL-17A-producing gammadelta T cells. *J Invest Dermatol* (2014) **134**:1912–21. doi:10.1038/jid.2014.98
- Petermann F, Rothhammer V, Claussen MC, Haas JD, Blanco LR, Heink S, et al. Gammadelta T cells enhance autoimmunity by restraining regulatory T cell responses via an interleukin-23-dependent mechanism. *Immunity* (2010) 33:351–63. doi:10.1016/j.immuni.2010.08.013
- Shichita T, Sugiyama Y, Ooboshi H, Sugimori H, Nakagawa R, Takada I, et al. Pivotal role of cerebral interleukin-17-producing gammadelta T cells in the delayed phase of ischemic brain injury. *Nat Med* (2009) 15:946–50. doi:10.1038/ nm.1999
- Cho JS, Pietras EM, Garcia NC, Ramos RI, Farzam DM, Monroe HR, et al. IL-17 is essential for host defense against cutaneous *Staphylococcus aureus* infection in mice. J Clin Invest (2010) 120:1762–73. doi:10.1172/JCI40891
- 22. Hamada S, Umemura M, Shiono T, Tanaka K, Yahagi A, Begum MD, et al. IL-17A produced by gammadelta T cells plays a critical role in innate immunity against *Listeria monocytogenes* infection in the liver. *J Immunol* (2008) **181**:3456–63. doi:10.4049/jimmunol.181.5.3456
- Meeks KD, Sieve AN, Kolls JK, Ghilardi N, Berg RE. IL-23 is required for protection against systemic infection with *Listeria monocytogenes*. J Immunol (2009) 183:8026–34. doi:10.4049/jimmunol.0901588
- Lockhart E, Green AM, Flynn JL. IL-17 production is dominated by gammadelta T cells rather than CD4 T cells during *Mycobacterium tuberculosis* infection. *J Immunol* (2006) 177:4662–9. doi:10.4049/jimmunol.177.7.4662
- Tortola L, Rosenwald E, Abel B, Blumberg H, Schafer M, Coyle AJ, et al. Psoriasiform dermatitis is driven by IL-36-mediated DC-keratinocyte crosstalk. J Clin Invest (2012) 122:3965–76. doi:10.1172/JCI63451
- Mazzucchelli R, Durum SK. Interleukin-7 receptor expression: intelligent design. Nat Rev Immunol (2007) 7:144–54. doi:10.1038/nri2023
- He YW, Malek TR. Interleukin-7 receptor alpha is essential for the development of gamma delta + T cells, but not natural killer cells. *J Exp Med* (1996) 184:289–93. doi:10.1084/jem.184.1.289
- Maki K, Sunaga S, Komagata Y, Kodaira Y, Mabuchi A, Karasuyama H, et al. Interleukin 7 receptor-deficient mice lack gammadelta T cells. *Proc Natl Acad Sci U S A* (1996) 93:7172–7. doi:10.1073/pnas.93.14.7172
- Malissen M, Pereira P, Gerber DJ, Malissen B, Disanto JP. The common cytokine receptor gamma chain controls survival of gamma/delta T cells. J Exp Med (1997) 186:1277–85. doi:10.1084/jem.186.8.1277
- Schlissel MS, Durum SD, Muegge K. The interleukin 7 receptor is required for T cell receptor gamma locus accessibility to the V(D)J recombinase. J Exp Med (2000) 191:1045–50. doi:10.1084/jem.191.6.1045
- 31. Baccala R, Witherden D, Gonzalez-Quintial R, Dummer W, Surh CD, Havran WL, et al. Gamma delta T cell homeostasis is controlled by IL-7 and IL-15 together with subset-specific factors. *J Immunol* (2005) **174**:4606–12. doi:10. 4049/jimmunol.174.8.4606
- Palmer MJ, Mahajan VS, Trajman LC, Irvine DJ, Lauffenburger DA, Chen J. Interleukin-7 receptor signaling network: an integrated systems perspective. *Cell Mol Immunol* (2008) 5:79–89. doi:10.1038/cmi.2008.10
- 33. Takeda K, Kaisho T, Yoshida N, Takeda J, Kishimoto T, Akira S. Stat3 activation is responsible for IL-6-dependent T cell proliferation through preventing apoptosis: generation and characterization of T cell-specific Stat3-deficient mice. *J Immunol* (1998) 161:4652–60.
- Chou WC, Levy DE, Lee CK. STAT3 positively regulates an early step in B-cell development. *Blood* (2006) 108:3005–11. doi:10.1182/blood-2006-05-024430
- Michel ML, Pang DJ, Haque SF, Potocnik AJ, Pennington DJ, Hayday AC. Interleukin 7 (IL-7) selectively promotes mouse and human IL-17-producing gammadelta cells. *Proc Natl Acad Sci U S A* (2012) 109:17549–54. doi:10.1073/pnas. 1204327109
- 36. Ivanov II, Mckenzie BS, Zhou L, Tadokoro CE, Lepelley A, Lafaille JJ, et al. The orphan nuclear receptor RORgammat directs the differentiation program of proinflammatory IL-17+ T helper cells. *Cell* (2006) 126:1121–33. doi:10.1016/j.cell.2006.07.035
- Laurence A, Tato CM, Davidson TS, Kanno Y, Chen Z, Yao Z, et al. Interleukin-2 signaling via STAT5 constrains T helper 17 cell generation. *Immunity* (2007) 26:371–81. doi:10.1016/j.immuni.2007.02.009

- Bekiaris V, Sedy JR, Macauley MG, Rhode-Kurnow A, Ware CF. The inhibitory receptor BTLA controls gammadelta T cell homeostasis and inflammatory responses. *Immunity* (2013) 39:1082–94. doi:10.1016/j.immuni.2013.10.017
- Hou L, Jie Z, Desai M, Liang Y, Soong L, Wang T, et al. Early IL-17 production by intrahepatic T cells is important for adaptive immune responses in viral hepatitis. J Immunol (2013) 190:621–9. doi:10.4049/jimmunol.1201970
- Shibata K, Yamada H, Sato T, Dejima T, Nakamura M, Ikawa T, et al. Notch-Hes1 pathway is required for the development of IL-17-producing gammadelta T cells. *Blood* (2011) 118:586–93. doi:10.1182/blood-2011-02-334995
- Hara T, Shitara S, Imai K, Miyachi H, Kitano S, Yao H, et al. Identification of IL-7-producing cells in primary and secondary lymphoid organs using IL-7-GFP knock-in mice. *J Immunol* (2012) 189:1577–84. doi:10.4049/jimmunol.1200586
- Yoshida H, Naito A, Inoue J, Satoh M, Santee-Cooper SM, Ware CF, et al. Different cytokines induce surface lymphotoxin-alphabeta on IL-7 receptor-alpha cells that differentially engender lymph nodes and Peyer's patches. *Immunity* (2002) 17:823–33. doi:10.1016/S1074-7613(02)00479-X
- Rennert PD, Browning JL, Mebius R, Mackay F, Hochman PS. Surface lymphotoxin alpha/beta complex is required for the development of peripheral lymphoid organs. J Exp Med (1996) 184:1999–2006. doi:10.1084/jem.184.5.1999
- Mebius RE. Organogenesis of lymphoid tissues. Nat Rev Immunol (2003) 3:292–303. doi:10.1038/nri1054
- Futterer A, Mink K, Luz A, Kosco-Vilbois MH, Pfeffer K. The lymphotoxin beta receptor controls organogenesis and affinity maturation in peripheral lymphoid tissues. *Immunity* (1998) 9:59–70. doi:10.1016/S1074-7613(00)80588-9
- Boehm T, Scheu S, Pfeffer K, Bleul CC. Thymic medullary epithelial cell differentiation, thymocyte emigration, and the control of autoimmunity require lympho-epithelial cross talk via LTbetaR. J Exp Med (2003) 198:757–69. doi:10.1084/jem.20030794
- Havran WL, Allison JP. Origin of Thy-1+ dendritic epidermal cells of adult mice from fetal thymic precursors. *Nature* (1990) 344:68–70. doi:10.1038/344068a0
- Roberts NA, White AJ, Jenkinson WE, Turchinovich G, Nakamura K, Withers DR, et al. Rank signaling links the development of invariant gammadelta T cell progenitors and Aire(+) medullary epithelium. *Immunity* (2012) 36:427–37. doi:10.1016/j.immuni.2012.01.016
- 49. Lewis JM, Girardi M, Roberts SJ, Barbee SD, Hayday AC, Tigelaar RE. Selection of the cutaneous intraepithelial gammadelta+ T cell repertoire by a thymic stromal determinant. *Nat Immunol* (2006) **7**:843–50. doi:10.1038/ni1363
- Boyden LM, Lewis JM, Barbee SD, Bas A, Girardi M, Hayday AC, et al. Skint1, the prototype of a newly identified immunoglobulin superfamily gene cluster, positively selects epidermal gammadelta T cells. *Nat Genet* (2008) 40:656–62. doi:10.1038/ng.108
- 51. Barbee SD, Woodward MJ, Turchinovich G, Mention JJ, Lewis JM, Boyden LM, et al. Skint-1 is a highly specific, unique selecting component for epidermal T cells. *Proc Natl Acad Sci U S A* (2011) **108**:3330–5. doi:10.1073/pnas.1010890108
- Turchinovich G, Hayday AC. Skint-1 identifies a common molecular mechanism for the development of interferon-gamma-secreting versus interleukin-17-secreting gammadelta T cells. *Immunity* (2011) 35:59–68. doi:10.1016/j. immuni.2011.04.018
- Silva-Santos B, Pennington DJ, Hayday AC. Lymphotoxin-mediated regulation of gammadelta cell differentiation by alphabeta T cell progenitors. *Science* (2005) 307:925–8. doi:10.1126/science.1103978
- 54. Ribot JC, Chaves-Ferreira M, D'Orey F, Wencker M, Goncalves-Sousa N, Decalf J, et al. Cutting edge: adaptive versus innate receptor signals selectively control the pool sizes of murine IFN-gamma- or IL-17-producing gammadelta T cells upon infection. *J Immunol* (2010) 185:6421–5. doi:10.4049/jimmunol.1002283
- 55. Powolny-Budnicka I, Riemann M, Tanzer S, Schmid RM, Hehlgans T, Weih F. RelA and RelB transcription factors in distinct thymocyte populations control lymphotoxin-dependent interleukin-17 production in gammadelta T cells. *Immunity* (2011) 34:364–74. doi:10.1016/j.immuni.2011.02.019
- Gerondakis S, Fulford TS, Messina NL, Grumont RJ. NF-kappaB control of T cell development. Nat Immunol (2014) 15:15–25. doi:10.1038/ni.2785
- 57. White AJ, Nakamura K, Jenkinson WE, Saini M, Sinclair C, Seddon B, et al. Lymphotoxin signals from positively selected thymocytes regulate the terminal differentiation of medullary thymic epithelial cells. *J Immunol* (2010) 185:4769–76. doi:10.4049/jimmunol.1002151
- Chai Q, Onder L, Scandella E, Gil-Cruz C, Perez-Shibayama C, Cupovic J, et al. Maturation of lymph node fibroblastic reticular cells from myofibroblastic precursors is critical for antiviral immunity. *Immunity* (2013) 38:1013–24. doi:10.1016/j.immuni.2013.03.012

- Upadhyay V, Fu YX. Lymphotoxin signalling in immune homeostasis and the control of microorganisms. *Nat Rev Immunol* (2013) 13:270–9. doi:10.1038/ nri3406
- Kabashima K, Banks TA, Ansel KM, Lu TT, Ware CF, Cyster JG. Intrinsic lymphotoxin-beta receptor requirement for homeostasis of lymphoid tissue dendritic cells. *Immunity* (2005) 22:439–50. doi:10.1016/j.immuni.2005.02.007
- De Trez C, Schneider K, Potter K, Droin N, Fulton J, Norris PS, et al. The inhibitory HVEM-BTLA pathway counter regulates lymphotoxin receptor signaling to achieve homeostasis of dendritic cells. *J Immunol* (2008) 180:238–48. doi:10.4049/jimmunol.180.1.238
- 62. Tumanov AV, Koroleva EP, Guo X, Wang Y, Kruglov A, Nedospasov S, et al. Lymphotoxin controls the IL-22 protection pathway in gut innate lymphoid cells during mucosal pathogen challenge. *Cell Host Microbe* (2011) 10:44–53. doi:10.1016/j.chom.2011.06.002
- Wang Y, Koroleva EP, Kruglov AA, Kuprash DV, Nedospasov SA, Fu YX, et al. Lymphotoxin beta receptor signaling in intestinal epithelial cells orchestrates innate immune responses against mucosal bacterial infection. *Immunity* (2010) 32:403–13. doi:10.1016/j.immuni.2010.02.011
- 64. Ota N, Wong K, Valdez PA, Zheng Y, Crellin NK, Diehl L, et al. IL-22 bridges the lymphotoxin pathway with the maintenance of colonic lymphoid structures during infection with *Citrobacter rodentium*. *Nat Immunol* (2011) **12**:941–8. doi:10.1038/ni.2089
- 65. Watanabe N, Gavrieli M, Sedy JR, Yang J, Fallarino F, Loftin SK, et al. BTLA is a lymphocyte inhibitory receptor with similarities to CTLA-4 and PD-1. *Nat Immunol* (2003) **4**:670–9. doi:10.1038/ni944
- 66. Sedy JR, Gavrieli M, Potter KG, Hurchla MA, Lindsley RC, Hildner K, et al. B and T lymphocyte attenuator regulates T cell activation through interaction with herpesvirus entry mediator. *Nat Immunol* (2005) **6**:90–8. doi:10.1038/ni1144
- Murphy KM, Nelson CA, Sedy JR. Balancing co-stimulation and inhibition with BTLA and HVEM. *Nat Rev Immunol* (2006) 6:671–81. doi:10.1038/nri1917
- Cheung TC, Steinberg MW, Oborne LM, Macauley MG, Fukuyama S, Sanjo H, et al. Unconventional ligand activation of herpesvirus entry mediator signals cell survival. *Proc Natl Acad Sci U S A* (2009) **106**:6244–9. doi:10.1073/pnas. 0902115106
- 69. Yang X, Zhang X, Sun Y, Tu T, Fu ML, Miller M, et al. A BTLA-mediated bait and switch strategy permits *Listeria* expansion in CD8alpha(+) DCs to promote long-term T cell responses. *Cell Host Microbe* (2014) 16:68–80. doi:10.1016/j.chom.2014.05.021
- 70. Kobayashi Y, Iwata A, Suzuki K, Suto A, Kawashima S, Saito Y, et al. B and T lymphocyte attenuator inhibits LPS-induced endotoxic shock by suppressing toll-like receptor 4 signaling in innate immune cells. *Proc Natl Acad Sci U S A* (2013) **110**:5121–6. doi:10.1073/pnas.1222093110
- Miller ML, Sun Y, Fu YX. Cutting edge: B and T lymphocyte attenuator signaling on NKT cells inhibits cytokine release and tissue injury in early immune responses. *J Immunol* (2009) 183:32–6. doi:10.4049/jimmunol.0900690
- 72. Kashiwakuma D, Suto A, Hiramatsu Y, Ikeda K, Takatori H, Suzuki K, et al. B and T lymphocyte attenuator suppresses IL-21 production from follicular Th cells and subsequent humoral immune responses. *J Immunol* (2010) **185**:2730–6. doi:10.4049/jimmunol.0903839
- Cheung TC, Oborne LM, Steinberg MW, Macauley MG, Fukuyama S, Sanjo H, et al. T cell intrinsic heterodimeric complexes between HVEM and BTLA determine receptivity to the surrounding microenvironment. *J Immunol* (2009) 183:7286–96. doi:10.4049/jimmunol.0902490
- 74. Hurchla MA, Sedy JR, Gavrieli M, Drake CG, Murphy TL, Murphy KM. B and T lymphocyte attenuator exhibits structural and expression polymorphisms and is highly Induced in anergic CD4+ T cells. *J Immunol* (2005) **174**:3377–85. doi:10.4049/jimmunol.174.6.3377
- 75. Krieg C, Boyman O, Fu YX, Kaye J. B and T lymphocyte attenuator regulates CD8+ T cell-intrinsic homeostasis and memory cell generation. *Nat Immunol* (2007) 8:162–71. doi:10.1038/ni1418
- 76. Mazharian A, Mori J, Wang YJ, Heising S, Neel BG, Watson SP, et al. Megakaryocyte-specific deletion of the protein-tyrosine phosphatases Shp1 and Shp2 causes abnormal megakaryocyte development, platelet production, and function. *Blood* (2013) 121:4205–20. doi:10.1182/blood-2012-08-449272
- 77. Castella B, Riganti C, Fiore F, Pantaleoni F, Canepari ME, Peola S, et al. Immune modulation by zoledronic acid in human myeloma: an advantageous crosstalk between Vgamma9Vdelta2 T cells, alphabeta CD8+ T cells, regulatory T cells, and dendritic cells. *J Immunol* (2011) 187:1578–90. doi:10.4049/jimmunol. 1002514

- Iwasaki M, Tanaka Y, Kobayashi H, Murata-Hirai K, Miyabe H, Sugie T, et al. Expression and function of PD-1 in human gammadelta T cells that recognize phosphoantigens. *Eur J Immunol* (2011) 41:345–55. doi:10.1002/eji.201040959
- Workman CJ, Rice DS, Dugger KJ, Kurschner C, Vignali DA. Phenotypic analysis of the murine CD4-related glycoprotein, CD223 (LAG-3). *Eur J Immunol* (2002) 32:2255–63. doi:10.1002/1521-4141(200208)32:8<2255::AID-IMMU2255>3. 0.CO;2-A
- Rosenblum MD, Woodliff JE, Madsen NA, Mcolash LJ, Keller MR, Truitt RL. Characterization of CD 200-receptor expression in the murine epidermis. *J Invest Dermatol* (2005) 125:1130–8. doi:10.1111/j.0022-202X.2005.23948.x
- Finney CA, Ayi K, Wasmuth JD, Sheth PM, Kaul R, Loutfy M, et al. HIV infection deregulates Tim-3 expression on innate cells: combination antiretroviral therapy results in partial restoration. *J Acquir Immune Defic Syndr* (2013) 63:161–7. doi:10.1097/QAI.0b013e318285cf13
- Nikolova M, Marie-Cardine A, Boumsell L, Bensussan A. BY55/CD160 acts as a co-receptor in TCR signal transduction of a human circulating cytotoxic effector T lymphocyte subset lacking CD28 expression. *Int Immunol* (2002) 14:445–51. doi:10.1093/intimm/14.5.445
- 83. Li J, Zhu H, Wang S, Ye P, Liu C, Wu J, et al. Blockade of NKG2D synergized with CTLA4-Ig in promoting long-term graft survival in murine models of cardiac transplantation. *Transplantation* (2012) **93**:356–63. doi:10.1097/TP. 0b013e31823ffce7
- Gertner-Dardenne J, Fauriat C, Orlanducci F, Thibult ML, Pastor S, Fitzgibbon J, et al. The co-receptor BTLA negatively regulates human Vgamma9Vdelta2 T-cell proliferation: a potential way of immune escape for lymphoma cells. *Blood* (2013) 122:922–31. doi:10.1182/blood-2012-11-464685
- 85. Gray EE, Ramirez-Valle F, Xu Y, Wu S, Wu Z, Karjalainen KE, et al. Deficiency in IL-17-committed Vgamma4(+) gammadelta T cells in a spontaneous Sox13mutant CD45.1(+) congenic mouse substrain provides protection from dermatitis. *Nat Immunol* (2013) 14:584–92. doi:10.1038/ni.2585
- Malhotra N, Narayan K, Cho OH, Sylvia KE, Yin C, Melichar H, et al. A network of high-mobility group box transcription factors programs innate interleukin-17 production. *Immunity* (2013) 38:681–93. doi:10.1016/j.immuni.2013.01.010
- Melichar HJ, Narayan K, Der SD, Hiraoka Y, Gardiol N, Jeannet G, et al. Regulation of gammadelta versus alphabeta T lymphocyte differentiation by the transcription factor SOX13. *Science* (2007) 315:230–3. doi:10.1126/science. 1135344

- Narayan K, Sylvia KE, Malhotra N, Yin CC, Martens G, Vallerskog T, et al. Intrathymic programming of effector fates in three molecularly distinct gammadelta T cell subtypes. *Nat Immunol* (2012) 13:511–8. doi:10.1038/ni.2247
- Weber BN, Chi AW, Chavez A, Yashiro-Ohtani Y, Yang Q, Shestova O, et al. A critical role for TCF-1 in T-lineage specification and differentiation. *Nature* (2011) 476:63–8. doi:10.1038/nature10279
- 90. Alonzo ES, Gottschalk RA, Das J, Egawa T, Hobbs RM, Pandolfi PP, et al. Development of promyelocytic zinc finger and ThPOK-expressing innate gamma delta T cells is controlled by strength of TCR signaling and Id3. *J Immunol* (2010) 184:1268–79. doi:10.4049/jimmunol.0903218
- Constantinides MG, Mcdonald BD, Verhoef PA, Bendelac A. A committed precursor to innate lymphoid cells. *Nature* (2014) 508:397–401. doi:10.1038/ nature13047
- 92. Zhang F, Meng G, Strober W. Interactions among the transcription factors Runx1, RORgammat and Foxp3 regulate the differentiation of interleukin 17producing T cells. *Nat Immunol* (2008) **9**:1297–306. doi:10.1038/ni.1663
- Sun Z, Unutmaz D, Zou YR, Sunshine MJ, Pierani A, Brenner-Morton S, et al. Requirement for RORgamma in thymocyte survival and lymphoid organ development. *Science* (2000) 288:2369–73. doi:10.1126/science.288.5475.2369

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