



# Combination of Cytokine-Induced Killer Cells and Programmed Cell Death-1 Blockade Works Synergistically to Enhance Therapeutic Efficacy in Metastatic Renal Cell Carcinoma and Non-Small Cell Lung Cancer

### Zibing Wang<sup>1</sup>, Xiaoli Liu<sup>1</sup>, Brian Till<sup>2</sup>, Miaomiao Sun<sup>1</sup>, Xiang Li<sup>1</sup> and Quanli Gao<sup>1\*</sup>

<sup>1</sup>Affiliated Hospital of Zhengzhou University, Henan Cancer Hospital, Zhengzhou, China, <sup>2</sup>Fred Hutchinson Cancer Research Center, Seattle, WA, United States

### **OPEN ACCESS**

#### Edited by:

Ignacio Melero, Centro de Investigación Médica Aplicada (CIMA), Spain

#### Reviewed by:

Amorette Barber, Longwood University, United States Krithika Kodumudi, Moffitt Cancer Center, United States

\*Correspondence: Quanli Gao gaoquanli1@aliyun.com

#### Specialty section:

This article was submitted to Cancer Immunity and Immunotherapy, a section of the journal Frontiers in Immunology

Received: 02 April 2018 Accepted: 19 June 2018 Published: 05 July 2018

#### Citation:

Wang Z, Liu X, Till B, Sun M, Li X and Gao Q (2018) Combination of Cytokine-Induced Killer Cells and Programmed Cell Death-1 Blockade Works Synergistically to Enhance Therapeutic Efficacy in Metastatic Renal Cell Carcinoma and Non-Small Cell Lung Cancer. Front. Immunol. 9:1513. doi: 10.3389/fimmu.2018.01513 **Introduction:** Programmed cell death-1 (PD-1) inhibition therapy has changed the treatment paradigm of metastatic renal cell carcinoma (MRCC) and non-small cell lung cancer (NSCLC). However, attempts to use the drug as a single agent have achieved only limited clinical success. To further enhance the clinical benefits of monotherapy, combination therapies will likely be necessary. Cytokine-induced killer (CIK) cells are a heterogeneous subset of *ex vivo* expanded T lymphocytes that have been shown to prolong the survival of cancer patients. We are conducting a study to evaluate the efficacy of PD-1 inhibitor in combination with CIK cells in relapsed/refractory MRCC and NSCLC and to analyze potential biomarkers to predict which patients will benefit most from the combined therapy.

**Case presentation:** The results of two patients treated in an ongoing clinical trial for MRCC and NSCLC are described here. The tumor biopsy from Patient 1 exhibited moderate CD3<sup>+</sup> T cell infiltration, but no PD-1 or PD-L1 expression. The tumor cells from Patient 2 strongly expressed PD-L1, and there was extensive tumor infiltration by CD3<sup>+</sup> T cells; however, no PD-1 staining was seen. Non-synonymous single nucleotide variant (nsSNVs), along with higher indel mutations, in Patient 1 and nsSNVs along with higher tumor mutation burden in Patient 2 correlate with tumor-infiltrating CD3<sup>+</sup> lymphocyte density. Patient 1 achieved a complete response, and Patient 2 achieved a near-complete response.

**Conclusion:** A PD-1 inhibitor in combination with CIK cells led to potent antitumor activity in MRCC and NSCLC; CD3<sup>+</sup> T cell infiltration in baseline tumor biopsies is a potential predictive biomarker. This approach is being further investigated in an ongoing phase I trial.

Keywords: cytokine-induced killer cells, programmed cell death-1, immunotherapy, lung cancer, renal cell carcinoma

Abbreviations: PD-1, programmed cell death-1; MRCC, renal cell carcinoma; NSCLC, non-small cell lung cancer; CIK, cytokine-induced killer; NK, natural killer; TMB, tumor mutation burden; dMME, deficient DNA mismatch repair status; indels, insertions and deletions; nsSNV, non-synonymous single nucleotide variant; PBMCs, peripheral blood mononuclear cells.

# INTRODUCTION

Although blocking of programmed cell death-1 (PD-1) alone has shown great promise in the treatment of advanced solid tumors, combined immunotherapies will be required to efficiently treat a wider range of cancer patients (1). Cytokine-induced killer (CIK) cells are a heterogeneous subset of *ex vivo* expanded T lymphocytes, comprising CD3<sup>+</sup>CD56<sup>+</sup> cells, CD3<sup>-</sup>CD56<sup>+</sup> natural killer (NK) cells, and CD3<sup>+</sup>CD56<sup>-</sup> cytotoxic T cells (2). CIK cells have been shown to prolong the survival of patients in metastatic renal cell carcinoma (MRCC) and non-small cell lung cancer (NSCLC) with minimal side effects (3, 4). Here, we show two representative cases from an ongoing clinical trial of patients with MRCC and NSCLC that were successfully treated with the combination therapy. Written informed consent was obtained for the use of anti-PD-1 antibodies and CIK cells in both patients.

## **CASE PRESENTATION**

An 80-year-old man (Patient 1) received a diagnosis of clear cell carcinoma of the right kidney after partial nephrectomy. One year later, computed tomography showed a right lumbar mass, and surgical specimens revealed clear cell carcinoma. The patient subsequently underwent afatinib treatment. However, therapy was discontinued as a result of intolerable adverse effects. Eighteen months later, PET-CT showed multiple metastases, including to the right pleural tubercle, thoracic vertebra, lumbar vertebra, left ilium, and humerus. In consultation with the radiation therapist, he received stereotactic radiotherapy to the right lung; however, a new left upper gingival soft tissue mass was found during the radiotherapy, and tumor metastasis was confirmed by biopsy. He was then treated with pembrolizumab combined with CIK cell transfer. CIK cells were prepared as described previously (3). Briefly, peripheral blood mononuclear cells (PBMC) were separated and cultured under sterile conditions in 1640 medium containing anti-CD3 monoclonal antibody, interferon  $\gamma$ , interleukin-2, and RetroNectin. After culturing the cells for 10–14 days, a target dose of about  $6 \times 10^9$  CIK cells with over 95% viability was obtained and tested for biological contaminants. Cells were then prepared in sodium chloride solution containing 2% albumin before transfusion. He achieved a complete response following treatment with four cycles of pembrolizumab combined with eight cycles of CIK cell transfer (Table S1 in Supplementary Material) and continues to be in remission on day 537 of the first dose of pembrolizumab treatment (Figure 1A). The patient had gingivitis after first cycle of pembrolizumab and pneumonia after second cycle of pembrolizumab, for which he received systematic antibiotic treatment. He did not receive glucocorticoids or other immunomodulating agents during his treatment with pembrolizumab and CIK cells.

A 63-year-old man (Patient 2) received a diagnosis of squamous cell carcinoma after biopsy of a right lower lobe lung mass. CT scans showed that this patient had developed multiple metastases, including mediastinal, right hilar, and anterior superior phrenic lymph nodes, and also to the sternum. His disease progressed following first-line platinum-based doublet chemotherapy and second-line S-1, a second-generation oral fluoropyrimidine composed of tegafur, gimeracil, and oteracil (5). He was then

treated with pembrolizumab combined with CIK cell transfer as third-line therapy. Follow-up imaging showed a near-complete response after treatment with eight cycles of pembrolizumab in combination with seven cycles of CIK cell transfer (Table S1 in Supplementary Material), and the patient continues to be in remission 185 days posttreatment (**Figure 1B**). No adverse events occurred during treatment in this patient.

Given that PD-1 pathway blockade induces a clinical response in subsets of cancer patients, defining biomarkers that predict therapeutic effects and adverse events is crucial. Recent studies have suggested an association between clinical responses and several promising biomarkers, including PD-1, PD-L1, CD3, tumor mutation burden (TMB), deficient DNA mismatch repair status (dMMR), and insertions and deletions (indels) (6). To examine the relationship between PD-1, PD-L1, or CD3 expression in the tumors and antitumor responses elicited by anti-PD-1 antibodies combined with CIK cell transfer, we evaluated pretreatment tumor biopsies from both patients using immunohistochemistry and found that the tumor biopsy from Patient 1 exhibited moderate CD3<sup>+</sup> T cell infiltration, but no PD-1 or PD-L1 expression (Figure 2A). The tumor cells from Patient 2 strongly expressed PD-L1, and there was extensive tumor infiltration by CD3+ T cells; however, no PD-1 staining was seen (Figure 2B). It has been reported that frameshift indels are more prevalent in renal cell carcinoma compared with other tumor types and are associated with enhanced T-cell activation (7). Across all tumor types, clear cell renal carcinoma was found to have the highest proportion of coding indels, 0.12, a 2.4-fold increase when compared with the pan-cancer average, 0.05 (7). We analyzed the whole-exome sequences of pretreatment tumors and patient-matched blood samples. We observed an indel proportion value of 0.08 and an indel count of 4 in patient 1, while these values were 0.04 and 5, respectively, in patient 2. We also found that tumors from both patients harbored non-synonymous single nucleotide variant (nsSNV) mutations. The TMB in patient 2 was higher than that in patient 1 (2.4 versus 0.8/Mb, respectively); however, no mutations in MMR-associated genes occurred in either patient (Figure 3). Based on these data, we hypothesize that nsSNVs, along with higher indel mutations, in patient 1 with renal cell carcinoma and nsSNVs along with higher TMB in patient 2 with lung cancer correlate with tumor-infiltrating CD3<sup>+</sup> lymphocyte density. Therefore, CD3<sup>+</sup> T cell infiltration in baseline tumor biopsies may be a potential biomarker to predict which patients will benefit most from PD-1 blockade in combination with CIK cell therapy, although this will need to be confirmed in larger numbers of patients.

## DISCUSSION

Immune checkpoint inhibitors, specifically PD-1-directed agents, have been approved for the treatment of advanced MRCC and NSCLC. Landmark trials have demonstrated a survival advantage with these agents compared to standard-of-care first-line and second-line therapies, along with greater durability of response in some patients. However, the majority of patients still do not respond to PD-1 inhibitors. Therefore, new therapies and novel combinations will be necessary.

One such novel therapy might be to combine adoptive cell transfer with PD-1 inhibitors. Among various adoptive



FIGURE 1 | (A) Patient 1 exhibited a complete response after three cycles of treatment with pembrolizumab plus cytokine-induced killer (CIK) cell transfer and continues to be in remission 537 days posttreatment (as of 12/02/2017). (B) Patient 2 exhibited a partial response after two cycles of treatment with pembrolizumab plus CIK cell transfer and continues to be in remission on day 185 after treatment (as of 12/02/2017).

lymphocyte therapies, CIK cells have a particularly advantageous profile as these cells are easily obtained, have a high proliferation rate, and exhibit a potent antitumor activity (8). In previous studies, we showed that CIK cells alone induced regression of MRCC with no major side effects after repeated transfer (3), and the efficacy was significantly improved when CIK cells were combined with chemotherapy (9). Similar results were reproduced in advanced pancreatic cancer (9, 10). Based on these promising

findings, we sought to investigate the potential clinical efficacy of the combination of PD-1 inhibitor with CIK cells in MRCC and NSCLC.

Programmed cell death-1 pathway blockade in combination with CIK cell transfusion elicited durable clinical responses in these patients with a reasonable toxicity profile. The precise mechanisms responsible for the observed antitumor responses are not clear, but we speculate that the several potential factors may play a role.



**FIGURE 2** | (A) Immunohistochemical analysis in Patient 1 shows no membranous expression of programmed cell death-1 or PD-L1 in the tumor cells and the small background cells surrounding the malignant tumor cells. CD3<sup>+</sup> cells are observed among the tumor cells. (B) Immunohistochemical analysis of a pretreatment tumor in Patient 2 shows focal membranous expression of PD-L1 in the tumor cells, with additional expression seen on the small background cells surrounding the malignant tumor cells. Abundant CD3<sup>+</sup> cells are identified among the tumor cells (magnification: left panel, 10x; right panel, 40x).

First, anti-PD-1 therapy may increase the absolute number of effector T cells in the tumor microenvironment. It is known that anti-PD-1 therapy largely relies on efficient T cell infiltration into tumors and effector T cell function in the tumor microenvironment. Most CIK cells are CD3+CD56- T cells and are thought to have the ability to migrate to tumor sites, based on the observation that a sustained strong bioluminescent imaging signal from the tumor site was still evident 3 weeks after adoptive transfer of luciferase-positive CIK cells in murine model of A20 B-cell lymphoma (11). We hypothesize that in the clinical setting, adoptively transferred CIK cells encounter anti-PD-1 antibodies, neutralizing an inhibitory signal generated by binding of PD-1 on these cells to anti-PD-1 antibodies. This assumption is supported by our finding that percentage of the PD-1<sup>+</sup> subpopulation among circulating CD3+ T cells substantially decreased after treatment compared to baseline levels in both patients (Tables S2 and S3 in Supplementary Material). However, it is unclear whether CIK cell activity is boosted in the periphery by anti-PD-1 antibodies, and then they migrate into the tumor and kill tumor cells; or if the cells are already infiltrating the tumors but become exhausted or inhibited by PD-1 signaling, and their activity is subsequently restored by the PD-1 blockade.

Second, CIK cells may have a direct cytotoxic effect against tumor cells. It has been shown that during CIK cell expansion, NKG2D is upregulated, which is an activating receptor expressed on NK cells and plays a crucial role in tumor cell killing (11, 12). Interestingly, a recent study reported an increase of an unusual population of peripheral blood cells expressing CD56 in patients with regressing melanoma following PD-1 blockade treatment (13). Thus, an increase in the total number of CD56<sup>+</sup> cells might contribute to the efficacy of the combination therapy of CIK cells and PD-1 blockade. To assess this, we measured CD56<sup>+</sup> NK cells in the PBMC before and at various time points after treatment and found that this cell population approximately doubled after treatment in both patients (Tables S2 and S3 in Supplementary Material).

Third, CIK cells secrete high amounts of IFN- $\gamma$ . Thus, in addition to increasing the number of T lymphocytes in the tumor microenvironment, the provision of cytokine help to existing tumor-infiltrating T cells may augment their function and overcome mechanisms of immune escape such as MHC downregulation, ultimately shifting the balance in the tumor microenvironment in favor of an antitumor response.

	TMB ( mutations /Mb )	MMR genes	Driver mutation	nsSNVs Mutation (n)	fs-indels mutation ( n )	Indel proportion
Patient 1	0.8	no mutation	No specific mutation	44	4	0.0833
Patient 2	2.43	no mutation	No specific mutation	144	5	0.0366

FIGURE 3 | Whole exome sequencing analysis revealed a low tumor mutation burden, lack of mutation in MMR or driver genes, and presence of non-synonymous single nucleotide variants or fs-indels.

The feasibility of generating CIK cells is a potential limitation of the combined treatment. The laboratories producing CIK cells must be specialized and have highly trained personnel. Additionally, there are differences in the techniques used by different laboratories in manufacturing CIK cells. One difference between ours and that reported by others was that we added RetroNectin to the traditional CIK cell culture system. The CIK cells prepared using this method secreted more Th1 cell-associated cytokines compared to traditional CIK cells (14). However, whether there is a difference in efficacy between traditional CIK cells and RetroNectin-activated CIK cells when they are combined with PD-1 inhibitor is unclear.

# **CONCLUDING REMARKS**

Programmed cell death-1 blockade in combination with CIK cells showed promising clinical responses in two patients with MRCC and NSCLC and is being further investigated in an ongoing clinical trial.

# ETHICS STATEMENT

No investigations or interventions were performed outside routine clinical care for these patients.

## REFERENCES

- Melero I, Berman DM, Aznar MA, Korman AJ, Pérez Gracia JL, Haanen J. Evolving synergistic combinations of targeted immunotherapies to combat cancer. *Nat Rev Cancer* (2015) 15:457–72. doi:10.1038/nrc3973
- Schmidt-Wolf IG, Lefterova P, Mehta BA, Fernandez LP, Huhn D, Blume KG, et al. Phenotypic characterization and identification of effector cells involved in tumor cell recognition of cytokine-induced killer cells. *Exp Hematol* (1993) 21:1673–9.
- Wang Z, Zhang Y, Liu Y, Wang L, Zhao L, Yang T, et al. Association of myeloid-derived suppressor cells and efficacy of cytokine-induced killer cell immunotherapy in metastatic renal cell carcinoma patients. *J Immunother* (2014) 37:43–50. doi:10.1097/CJI.000000000000005
- Li R, Wang C, Liu L, Du C, Cao S, Yu J, et al. Autologous cytokineinduced killer cell immunotherapy in lung cancer: a phase II clinical study. *Cancer Immunol Immunother* (2012) 61:2125–33. doi:10.1007/s00262-012-1260-2
- Shirasaka T, Nakano K, Takechi T, Satake H, Uchida J, Fujioka A, et al. Antitumor activity of 1 M tegafur-0.4 M 5-chloro-2,4-dihydroxypyridine-1 M potassium oxonate (S-1) against human colon carcinoma orthotopically implanted into nude rats. *Cancer Res* (1996) 56:2602–6.
- Nishino M, Ramaiya NH, Hatabu H, Hodi FS. Monitoring immune-checkpoint blockade: response evaluation and biomarker development. *Nat Rev Clin Oncol* (2017) 14:655–8. doi:10.1038/nrclinonc.2017.88
- Turajlic S, Litchfield K, Xu H, Rosenthal R, McGranahan N, Reading JL, et al. Insertion-and-deletion-derived tumour-specific neoantigens and the immunogenic phenotype: a pan-cancer analysis. *Lancet Oncol* (2017) 18:1009–21. doi:10.1016/S1470-2045(17)30516-8
- Jäkel CE, Hauser S, Rogenhofer S, Müller SC, Brossart P, Schmidt-Wolf IG. Clinical studies applying cytokine-induced killer cells for the treatment of renal cell carcinoma. *Clin Dev Immunol* (2012) 2012:473245. doi:10.1155/ 2012/473245

# **AUTHOR CONTRIBUTIONS**

ZW and QG have designed the paper; MS and XL provided technical or material support; ZW, XL, and BT wrote, reviewed, and revised the paper; all the coauthors revised paper critically and gave final approval of this version for publishing.

# ACKNOWLEDGMENTS

The authors thank all of the patients and families that enrolled in this study. The authors thank Dr. Torsten Juelich for linguistic assistance during the preparation of this manuscript.

# FUNDING

This study was supported by National Natural Science Foundation of China (81000914, 81272526) and Foundation of He'nan Health Committee (2011010011).

# SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at https://www.frontiersin.org/articles/10.3389/fimmu.2018.01513/ full#supplementary-material.

- Wang Z, Liu Y, Zhang Y, Shang Y, Gao Q. MDSC-decreasing chemotherapy increases the efficacy of cytokine-induced killer cell immunotherapy in metastatic renal cell carcinoma and pancreatic cancer. *Oncotarget* (2016) 7:4760–9. doi:10.18632/oncotarget.6734
- Wang Z, Liu Y, Li R, Shang Y, Zhao Y, Zhao L, et al. Autologous cytokineinduced killer cell transfusion increases overall survival in advanced pancreatic cancer. J Hematol Oncol (2016) 9:6. doi:10.1186/s13045-016-0237-6
- Nishimura R, Baker J, Beilhack A, Zeiser R, Olson JA, Sega EI, et al. In vivo trafficking and survival of cytokine-induced killer cells resulting in minimal GVHD with retention of antitumor activity. *Blood* (2008) 112:2563–74. doi:10.1182/blood-2007-06-092817
- Verneris MR, Karami M, Baker J, Jayaswal A, Negrin RS. Role of NKG2D signaling in the cytotoxicity of activated and expanded CD8+ T cells. *Blood* (2004) 103:3065–72. doi:10.1182/blood-2003-06-2125
- Ribas A, Shin DS, Zaretsky J, Frederiksen J, Cornish A, Avramis E, et al. PD-1 blockade expands intratumoral memory T cells. *Cancer Immunol Res* (2016) 4:194–203. doi:10.1158/2326-6066.CIR-15-0210
- Han L, Shang YM, Song YP, Gao QL. Biological character of RetroNectin activated cytokine-induced killer cells. *J Immunol Res* (2016) 2016:5706814. doi:10.1155/2016/5706814

**Conflict of Interest Statement:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2018 Wang, Liu, Till, Sun, Li and Gao. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.