



Human V δ 1⁺ T Cells in the Immune Response to *Plasmodium falciparum* Infection

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Naturally acquired protective immunity to *Plasmodium falciparum* malaria is mainly antibody-mediated. However, other cells of the innate and adaptive immune system also play important roles. These include so-called unconventional T cells, which express a $\gamma\delta$ T-cell receptor (TCR) rather than the $\alpha\beta$ TCR expressed by the majority of T cells—the conventional T cells. The $\gamma\delta$ T-cell compartment can be divided into distinct subsets. One expresses a TCR involving V γ 9 and V δ 2, while another major subset uses instead a TCR composed of V δ 1 paired with one of several types of γ chains. The former of these subsets uses a largely semi-invariant TCR repertoire and responds in an innate-like fashion to pyrophosphate antigens generated by various stressed host cells and infectious pathogens, including *P. falciparum*. In this short review, we focus instead on the V δ 1 subset, which appears to have a more adaptive immunobiology, but which has been much less studied in general and in malaria in particular. We discuss the evidence that V δ 1⁺ cells do indeed play a role in malaria and speculate on the function and specificity of this cell type, which is increasingly attracting the attention of immunologists.

Keywords: gamma-delta ($\gamma\delta$) T lymphocytes, Vdelta1 gamma delta T cells, malaria, *Plasmodium falciparum*, innate immunity, acquired immunity, immune regulation

INTRODUCTION

The most serious form of malaria is caused by the hemoprotozoan parasite *Plasmodium falciparum*. The disease is a major humanitarian and economic burden on societies affected by it, mainly in sub-Saharan Africa, and it leads to the death of about half a million children every year (1, 2). Immunity to the disease is gradually acquired after years of exposure and many disease episodes, and is mainly mediated by IgG antibodies targeting the asexual blood stages of the infection, which are responsible for all the clinical symptoms and complications (3–5). T cells are nevertheless also of obvious importance in acquisition of immunity, not least to enable B-cell class switching and affinity maturation.

Most circulating T cells express $\alpha\beta$ type T-cell receptors (TCR- $\alpha\beta$), but a minority of T cells instead expresses the alternative $\gamma\delta$ TCR heterodimer (TCR- $\gamma\delta$). The pivotal role of $\alpha\beta$ T cells in immunity to *P. falciparum* malaria is well-established. The $\alpha\beta$ T cells function both directly as cytotoxic effector cells against infected hepatocytes, and indirectly as CD4⁺ helper cells for a variety

of innate and adaptive immune responses to all stages of the parasite life cycle in the human host. Much less is known about the function and significance of $\gamma\delta$ T cells in this immunity.

The $\alpha\beta$ and $\gamma\delta$ T-cell compartments share several features. In both, the TCR constitutes the antigen recognition element of the multi-molecular TCR complex, which also includes several signal transduction components, such as CD3. TCR diversity is generated by somatic recombination events during T-cell maturation in the thymus. As for $\alpha\beta$ T cells, the TCRs of $\gamma\delta$ T cells are clonally distributed, such that each T-cell clone expresses a single, rearranged TCR variant, which determines the antigen specificity of the clone—at least in the case of $\alpha\beta$ T cells.

The two compartments also exhibit important differences. Thus, $\alpha\beta$ T cells respond predominantly to protein antigens that are processed by antigen-presenting cells (APCs) and subsequently displayed as short peptides bound to major histocompatibility complex (MHC) molecules on the APC surface. In contrast to $\alpha\beta$ T cells, which typically express either CD4 or CD8, $\gamma\delta$ T cells often express neither, in particular in the V γ 9⁺V δ 2⁺ subset. In keeping with this lack of MHC restriction elements, recognition of antigen by “double-negative” $\gamma\delta$ T cells is not MHC-restricted. Furthermore, V γ 9⁺V δ 2⁺ T cells universally respond to non-peptide prenyl pyrophosphate metabolites (termed phospho-antigens, or P-Ag) (6). These antigens, which are produced by a variety of stressed cells (isopentenyl pyrophosphate, IPP, produced via the host mevalonate pathway) and by infectious pathogens, including *P. falciparum* [(E)-4-Hydroxy-3-methyl-but-2-enyl pyrophosphate, HMB-PP, produced via the microbial non-mevalonate pathway] are structurally related. Accordingly, the V γ 9 chains expressed by these cells are relatively invariant (7, 8) due to convergent and recurrent recombinations (9). In addition, the V γ 9⁺V δ 2⁺ TCR repertoire is already restricted from birth, and contains a high proportion of V γ 9 clonotypes that are shared by many clones in a given individual, and conserved between many individuals (i.e., “public” repertoires). Furthermore, the repertoire of these cells does not exhibit dramatic clonotypic focusing in adults relative to neonates (9, 10). The V γ 9⁺V δ 2⁺ T-cell subset, which is usually the dominant $\gamma\delta$ T-cell subset in the peripheral blood of healthy individuals without exposure to *P. falciparum*, can thus be described as an “innate-like” T-cell subset.

To date, the V γ 9⁺V δ 2⁺ cells are the $\gamma\delta$ T cells that have attracted by far the most attention in relation to malaria (11, 12). However, we focus here instead on a largely complementary subset that is characterized by a TCR composed of V δ 1 paired with a variety of γ -chains, and that appears to adopt a distinct immunobiology relative to the innate-like V γ 9⁺V δ 2⁺ subset (13). Unlike V γ 9⁺V δ 2⁺ T cells, V δ 1⁺ T cells typically constitute a minority ($\leq 20\%$) of adult peripheral blood $\gamma\delta$ T cells. However, the subset is enriched relative to the V γ 9⁺V δ 2⁺ T-cell subset in tissues, where they have been reported to recognize a variety of host and microbial antigens (14–16). Also in marked contrast, the TCR repertoire of V δ 1⁺ T cells—and of V γ 9^{neg}V δ 2⁺ T cells (17)—is highly diverse at birth, and largely non-overlapping between individuals (i.e., “private” repertoires). Furthermore, the TCR repertoire of this $\gamma\delta$ T-cell subset becomes increasingly focused over time as a result of selective expansion of specific clonotypes, most likely following antigenic stimulation

(9, 18–20). The V δ 1 subset therefore appears to be much more “adaptive-like” than the V γ 9⁺V δ 2⁺ subset (21), and it bears substantial similarities to conventional $\alpha\beta$ T cells. Nevertheless, there is certainly evidence that V δ 1⁺ T cells play a distinct role from $\alpha\beta$ T cells in the immune response to several infections—including *P. falciparum* malaria.

Increased Proportions and Numbers of V δ 1⁺ T Cells in Malaria Patients and Healthy Residents From Malaria-Endemic Areas

Within a few years of the discovery of the $\gamma\delta$ TCR, several groups reported modest but protracted expansions of $\gamma\delta$ T cells in adult *P. falciparum* and *P. vivax* patients with little or no previous malaria parasite exposure (22–24). A later study of malarious children from a highly malaria-endemic area and employing a pan- $\gamma\delta$ TCR-specific antibody reported similar findings, and did not find significant differences in peripheral blood $\gamma\delta$ T-cell frequencies between children with uncomplicated and severe malaria, respectively (25). The authors also reported significantly decreased absolute numbers of $\gamma\delta$ T cells at the time of admission to hospital with malaria (regardless of severity), followed by a transient increase to numbers above normal during convalescence. This was also observed among the few adult first-time malaria patients included in the study (25). Overall, the $\gamma\delta$ T cell-specific findings appeared similar in patients with or without prior exposure to malaria, and also resembled earlier reports regarding the $\alpha\beta$ T-cell response to malaria, namely an inflammation-induced withdrawal of these cells from the peripheral circulation, followed by their release back into the peripheral blood after successful chemotherapy [reviewed in Hviid (26)].

Substantial $\gamma\delta$ T-cell subset heterogeneity was also reported (27–30). These early papers indicated that the $\gamma\delta$ T-cell response to *P. falciparum* malaria extends beyond V γ 9⁺V δ 2⁺ cells, although that subset remained the dominant one among the non-immune patients that were studied. However, it was reported shortly after that in semi-immune African children and adults with acute *P. falciparum* malaria, the $\gamma\delta$ T cells responding *in vivo* are completely dominated by cells expressing V δ 1, with little contribution from V γ 9⁺V δ 2⁺ T cells (31, 32). A study of children and adults from *P. falciparum*-endemic Lao People's Democratic Republic very recently reported similar findings (33). The expanded V δ 1⁺ subset had an activated phenotype, produced pro-inflammatory cytokines, used a diversity of V γ chains, and showed spectratyping evidence of clonal focusing (31–33). In fact, the V δ 1⁺ subset appeared to dominate even among healthy *P. falciparum*-exposed individuals living in areas with stable transmission of these parasites (20, 34). In the absence of acute malaria, these cells were CD45RA⁺, resting (CD69^{neg} and HLA-DR^{neg}), and about half of them were CD8⁺ (in contrast to the majority of V γ 9⁺V δ 2⁺ cells, which are double-negative). They were clonally restricted in most adults, but less so in children (20). They thus appear phenotypically similar to the V δ 1⁺ cells found in epithelia (35). While V γ 9⁺V δ 2⁺ cells from such individuals could respond when stimulated *in vitro* by *P. falciparum* pyrophosphate antigens (34)—similar to

V γ 9⁺V δ 2⁺ cells from donors without previous malaria exposure [reviewed in Howard et al. (11)]—this response did not appear very prominent *in vivo*.

V δ 1⁺ T Cells in Malaria: What Do They See and What Do They Do?

Essentially nothing is known about the function or antigen specificity/specificities of the dominant V δ 1⁺ $\gamma\delta$ T-cell subset in *P. falciparum*-exposed individuals (12). A few studies have indicated that these cells might recognize, respond to, and have a direct effector function against infected erythrocytes in a manner resembling V γ 9⁺V δ 2⁺ cells (11, 33, 36). However, already early on May Ho and colleagues speculated that the expansion of V δ 1⁺ T cells in *P. falciparum* malaria might instead involve “unidentified host factors” (29). Their prediction is supported by the findings that V δ 1⁺ cells from parasite-exposed individuals do not respond markedly to *P. falciparum* antigens *in vitro* (34), including the parasite-derived pyrophosphate antigens recognized by V γ 9⁺V δ 2⁺ cells (37, 38).

Although it is not known what drives the expansion and differentiation of the adaptive-like V δ 1⁺ subset in malaria, V δ 1⁺ T-cell expansion has been observed in several other pathological conditions (16). Examples include infections with human immunodeficiency virus (HIV) (39–42), cytomegalovirus (CMV) and other herpes viruses (43–45), *Onchocerca volvulus* parasites (46), as well as autoimmune diseases such as Takayasu arteritis (47), inflammatory bowel disease and Crohn’s disease (48, 49). The possibility that the V δ 1⁺ T-cell response in these diseases involves recognition of host-encoded components is supported by studies of CMV. In that infection, V δ 2^{neg} T cells display shared reactivity against both CMV-infected target cells and uninfected epithelial cells, consistent with recognition of host-encoded antigens (50). Moreover, endothelial protein C receptor (EPCR) has been identified as an antigenic target for a V δ 2^{neg} $\gamma\delta$ TCR expressed by a clonotype heavily expanded after infection with CMV (51), which is known to infect endothelial tissues. T-cell activation was dependent on integration of TCR/EPCR-mediated signals with a TCR-extrinsic “multi-molecular stress signature” induced upon infection of target cells that included CMV-mediated increases in ICAM-1 and LFA-1 expression. Conceivably, this may represent one route for V δ 2^{neg} $\gamma\delta$ T-cell recognition of “stressed self.” It may be of interest in the context of malaria that EPCR has been identified as a clinically important receptor for *P. falciparum*-infected erythrocytes (52, 53).

Dysregulation of the B-cell compartment might constitute another pathogen-induced change that could be sensed by “adaptive-like” $\gamma\delta$ T cells. Of relevance, *P. falciparum* malaria, and indeed a number of other diseases associated with V δ 1⁺ T-cell expansions, is characterized by massive B-cell activation, both of B cells that are specific for the infection causing the disease and B cells that are not (54, 55). This often leads to reactivation of latent EBV (and CMV) infection, and further B-cell proliferation (56–58). From this perspective, it is tempting to speculate that the selective expansion of V δ 1⁺ T cells observed in individuals living in areas with stable transmission of *P. falciparum* occurs in response to antigens expressed by activated B cells, perhaps serving as part of an auto-regulatory response to curb excessive B-cell activation and proliferation. In addition, V δ 1⁺ cells

can recognize EBV-transformed B-cell lines (59, 60), and EBV infection can result in expansion of clonally restricted V δ 1⁺ cell populations after stem cell transplantation (61, 62). Conceivably, CD1c/TCT.1/Blast-1 might be an antigen recognized by these cells. Thus, CD8⁺ V δ 1⁺ cells heavily expanded *in vitro* have been shown to recognize this antigen (63, 64), which is expressed/upregulated on some activated and transformed B cells (65, 66). This is not least the case in the spleen, where V δ 1⁺ cells are also abundant (67), and further increase in numbers in response to *P. falciparum* malaria (68). In addition, V δ 1⁺ T-cell reactivity to CD1c tetramers has been demonstrated (69), although to date only involving a low percentage of the V δ 1 T-cell repertoire. It therefore remains unclear whether CD1c-specific cells overlap with *in vivo* expanded clonotypes (21). In summary, while other possibilities cannot be discounted, responses to “stressed self” via recognition of host antigens may contribute to V δ 1-mediated adaptive surveillance in the context of malaria, which could be linked to immune, stress-linked, or EBV/CMV-related sequelae of parasite infection. Such adaptive surveillance of stressed self has strong relevance for the proposed role of V δ 1⁺ T cells in cancer (16, 70–72).

CONCLUDING REMARKS

There is an increasing interest in the role of $\gamma\delta$ T cells and other similar cells, such as NK cells, in the immune response to malaria (11, 73, 74). However, the V δ 1⁺ subset has attracted only limited attention so far. Based on the ideas and studies highlighted in this review, we believe that there is a strong case for extending the focus of $\gamma\delta$ T-cell studies in malaria beyond the innate-like V γ 9⁺V δ 2⁺ subset, to include adaptive-like $\gamma\delta$ T cells. Although, we have focused here on V δ 1⁺ T cells, it is worth noting that clonal expansion of $\gamma\delta$ T cells that express V δ 2 chains paired with γ -chains other than V γ 9 has been described in a variety of conditions. Those cell populations also appear to display an “adaptive-like” immunobiology, positioning them functionally much closer to V δ 1⁺ cells than to the innate-like V γ 9⁺V δ 2⁺ cell subset [reviewed in Davey et al. (17)]. Moreover, recent data further suggest that $\gamma\delta$ T cells that express neither V δ 1 nor V δ 2 (e.g., V δ 3⁺ cells) exhibit features of such adaptive immune subsets (13, 75). In light of this emerging adaptive immunobiological human $\gamma\delta$ T-cell paradigm, examining the contributions of $\gamma\delta$ T-cell subsets other than V γ 9⁺V δ 2⁺ in the immune response to malaria is an underexplored and important avenue for investigation.

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All authors listed have made a substantial, direct and intellectual contribution to the work, and approved it for publication.

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REFERENCES

- World Health Organization. *World Malaria Report 2017* (2017).
- Phillips MA, Burrows JN, Manyando C, Van Huijsduijnen RH, Van Voorhis WC, Wells TNC. Malaria. *Nat Rev Dis Primers* (2017) 3:17050. doi: 10.1038/nrdp.2017.50
- Hviid L. Naturally acquired immunity to *Plasmodium falciparum* malaria in Africa. *Acta Trop.* (2005) 95:270–5. doi: 10.1016/j.actatropica.2005.06.012
- Langhorne J, Ndungu FM, Sponaas AM, Marsh K. Immunity to malaria: more questions than answers. *Nat Immunol.* (2008) 9:725–32. doi: 10.1038/ni.f.205
- Cowman AF, Healer J, Marapana D, Marsh K. Malaria: biology and disease. *Cell* (2016) 167:610–24. doi: 10.1016/j.cell.2016.07.055
- Morita CT, Jin C, Sarikonda G, Wang H. Nonpeptide antigens, presentation mechanisms, and immunological memory of human V γ 9V δ 2 T cells: discriminating friend from foe through the recognition of prenyl pyrophosphate antigens. *Immunol Rev.* (2007) 215:59–76. doi: 10.1111/j.1600-065X.2006.00479.x
- Sherwood AM, Desmarais C, Livingston RJ, Andriesen J, Haussler M, Carlson CS, et al. Deep sequencing of the human TCR γ and TCR β repertoires suggests that TCR β rearranges after $\alpha\beta$ and $\gamma\delta$ T cell commitment. *Sci Transl Med.* (2011) 3:90ra61. doi: 10.1126/scitranslmed.3002536
- Dimova T, Brouwer M, Gosselin F, Tassinon J, Leo O, Donner C, et al. Effector V γ 9V δ 2 cells dominate the human fetal $\gamma\delta$ T-cell repertoire. *Proc Natl Acad Sci USA.* (2015) 112:E556–65. doi: 10.1073/pnas.1412058112
- Davey MS, Willcox CR, Hunter S, Kasatskaya SA, Remmerswaal EBM, Salim M, et al. The human V δ 2⁺ T-cell compartment comprises distinct innate-like V γ 9⁺ and adaptive V γ 9⁻ subsets. *Nat Commun.* (2018) 9:1760. doi: 10.1038/s41467-018-04076-0
- Willcox CR, Davey MS, Willcox BE. Development and selection of the human V γ 9V δ 2⁺ T-cell repertoire. *Front Immunol.* (2018) 9:1501. doi: 10.3389/fimmu.2018.01501
- Howard J, Zaidi I, Loizon S, Mercereau-Puijalon O, Dechanet-Merville J, Mamani-Matsuda M. Human V γ 9V δ 2 T lymphocytes in the immune response to *P. falciparum* infection. *Front Immunol.* (2018) 9:2760. doi: 10.3389/fimmu.2018.02760
- Deroost K, Langhorne J. Gamma/delta T cells and their role in protection against malaria. *Front Immunol.* (2018) 9:2973. doi: 10.3389/fimmu.2018.02973
- Davey MS, Willcox CR, Joyce SP, Ladell K, Kasatskaya SA, McLaren JE, et al. Clonal selection in the human V δ 1 T cell repertoire indicates $\gamma\delta$ TCR-dependent adaptive immune surveillance. *Nat Commun.* (2017) 8:14760. doi: 10.1038/ncomms14760
- Pang DJ, Neves JF, Sumaria N, Pennington DJ. Understanding the complexity of $\gamma\delta$ T-cell subsets in mouse and human. *Immunology* (2012) 136:283–90. doi: 10.1111/j.1365-2567.2012.03582.x
- Vantourout P, Hayday A. Six-of-the-best: unique contributions of $\gamma\delta$ T cells to immunology. *Nat Rev Immunol.* (2013) 13:88–100. doi: 10.1038/nri3384
- Siegers GM, Lamb LS Jr. Cytotoxic and regulatory properties of circulating V δ 1⁺ $\gamma\delta$ T cells: a new player on the cell therapy field? *Mol Ther.* (2014) 22:1416–22. doi: 10.1038/mt.2014.104
- Davey MS, Willcox CR, Hunter S, Oo YH, Willcox BE. V δ 2⁺ T cells - two subsets for the price of one. *Front Immunol.* (2018) 9:2106. doi: 10.3389/fimmu.2018.02106
- Beldjord K, Beldjord C, Macintyre E, Even P, Sigaux F. Peripheral selection of V δ 1⁺ cells with restricted T cell receptor delta gene junctional repertoire in the peripheral blood of healthy donors. *J Exp Med.* (1993) 178:121–7. doi: 10.1084/jem.178.1.121
- Giachino C, Granziero L, Modena V, Maiocco V, Lomater C, Fantini F, et al. Clonal expansions of V δ 1⁺ and V δ 2⁺ cells increase with age and limit the repertoire of human $\gamma\delta$ T cells. *Eur J Immunol.* (1994) 24:1914–8. doi: 10.1002/eji.1830240830
- Hviid L, Akanmori BD, Loizon S, Kurtzhals JA, Ricke CH, Lim A, et al. High frequency of circulating $\gamma\delta$ T cells with dominance of the V δ 1 subset in a healthy population. *Int Immunol.* (2000) 12:797–805. doi: 10.1093/intimm/12.6.797
- Davey MS, Willcox CR, Baker AT, Hunter S, Willcox BE. Recasting human V δ 1 lymphocytes in an adaptive role. *Trends Immunol.* (2018) 39:446–59. doi: 10.1016/j.it.2018.03.003
- Ho M, Webster HK, Tongtawe P, Pattanapanyasat K, Weidanz WP. Increased $\gamma\delta$ T cells in acute *Plasmodium falciparum* malaria. *Immunol Lett.* (1990) 25:139–42. doi: 10.1016/0165-2478(90)90105-Y
- Roussilhon C, Agrapart M, Ballet J-J, Bensussan A. T lymphocytes bearing the $\gamma\delta$ T cell receptor in patients with acute *Plasmodium falciparum* malaria. *J Infect Dis.* (1990) 162:283–5. doi: 10.1093/infdis/162.1.283-a
- Perera MK, Carter R, Goonewardene R, Mendis KN. Transient increase in circulating $\gamma\delta$ T cells during *Plasmodium vivax* malarial paroxysms. *J Exp Med.* (1994) 179:311–5. doi: 10.1084/jem.179.1.311
- Hviid L, Kurtzhals JA, Dodoo D, Rodrigues O, Ronn A, Commey JOO, et al. The $\gamma\delta$ T-cell response to *Plasmodium falciparum* malaria in a population in which malaria is endemic. *Infect Immun.* (1996) 64:4359–62.
- Hviid L. Peripheral T-cell non-responsiveness in individuals exposed to *Plasmodium falciparum* malaria. *APMIS* (1995) 103 (Suppl. 53):1–46.
- Chang W-L, Van Der Heyde H, Maki DG, Malkovsky M, Weidanz W. Subset heterogeneity among $\gamma\delta$ T cells found in peripheral blood during *Plasmodium falciparum* malaria. *Immunol Lett.* (1992) 32:273–4.
- Schwartz E, Shapiro R, Shina S, Bank I. Delayed expansion of V δ 2⁺ and V δ 1⁺ $\gamma\delta$ T cells after acute *Plasmodium falciparum* and *Plasmodium vivax* malaria. *J Allergy Clin Immunol.* (1996) 97:1387–92. doi: 10.1016/S0091-6749(96)70208-7
- Ho M, Tongtawe P, Kriangkum J, Wimonwattawatee T, Pattanapanyasat K, Bryant L, et al. Polyclonal expansion of peripheral $\gamma\delta$ T cells in human *Plasmodium falciparum* malaria. *Infect Immun.* (1994) 62:855–62.
- Roussilhon C, Agrapart M, Guglielmi P, Bensussan A, Brasseur P, Ballet JJ. Human TcR $\gamma\delta$ ⁺ lymphocyte response on primary exposure to *Plasmodium falciparum*. *Clin Exp Immunol.* (1994) 95:91–7.
- Hviid L, Kurtzhals JA, Adabayeri V, Loizon S, Kemp K, Goka BQ, et al. Perturbation and proinflammatory type activation of V δ 1⁺ $\gamma\delta$ T cells in African children with *Plasmodium falciparum* malaria. *Infect Immun.* (2001) 69:3190–6. doi: 10.1128/IAI.69.5.3190-3196.2001
- Worku S, Björkman A, Troye-Blomberg M, Jemaneh L, Färnert A, Christensson B. Lymphocyte activation and subset redistribution in the peripheral blood in acute malaria illness: distinct $\gamma\delta$ ⁺ T cell patterns in *Plasmodium falciparum* and *P. vivax* infections. *Clin Exp Immunol.* (1997) 108:34–41.
- Taniguchi T, Mannoor KM, Nonaka D, Toma H, Li C, Narita M, et al. A unique subset of $\gamma\delta$ T cells expands and produces IL-10 in patients with naturally acquired immunity against *falciparum* malaria. *Front Microbiol.* (2017) 8:1288. doi: 10.3389/fmicb.2017.01288
- Goodier M, Krause-Jauer M, Sanni A, Massougbdji A, Sadeler B-C, Mitchell GH, et al. $\gamma\delta$ T cells in the peripheral blood of individuals from an area of holoendemic *Plasmodium falciparum* transmission. *Trans R Soc Trop Med Hyg.* (1993) 87:692–6. doi: 10.1016/0035-9203(93)90299-6
- Deusch K, Luling F, Reich K, Classen M, Wagner H, Pfeffer K. A major fraction of human intraepithelial lymphocytes simultaneously expresses the g/d T cell receptor, the CD8 accessory molecule and preferentially uses the Vd1 gene segment. *Eur J Immunol.* (1991) 21:1053–9. doi: 10.1002/eji.1830210429
- Troye-Blomberg M, Worku S, Tangteerawatana P, Jamshaid R, Söderström K, Elghazali G, et al. Human $\gamma\delta$ T cells that inhibit the *in vitro* growth of the asexual blood stages of the *Plasmodium falciparum* parasite express cytolytic and proinflammatory molecules. *Scand J Immunol.* (1999) 50:642–50.
- Behr C, Dubois P. Preferential expansion of V γ 9V δ 2 T cells following stimulation of peripheral blood lymphocytes with extracts of *Plasmodium falciparum*. *Int Immunol.* (1992) 4:361–6.
- Goodier M, Fey P, Eichmann K, Langhorne J. Human peripheral blood $\gamma\delta$ T cells respond to antigens of *Plasmodium falciparum*. *Int Immunol.* (1992) 4:33–41. doi: 10.1093/intimm/4.1.33
- Autran B, Triebel F, Katlama C, Rozenbaum W, Hercend T, Debre P. T cell receptor $\gamma\delta$ ⁺ lymphocyte subsets during HIV infection. *Clin Exp Immunol.* (1989) 75:206–10.
- De Maria A, Ferrazin A, Ferrini S, Ciccone E, Terragna A, Moretta L. Selective increase of a subset of T cell receptor $\gamma\delta$ T lymphocytes in the peripheral blood of patients with human immunodeficiency virus type 1 infection. *J Infect Dis.* (1992) 165:917–9. doi: 10.1093/infdis/165.5.917
- Boullier S, Cochet M, Poccia F, Gougeon M-L. CDR3-independent $\gamma\delta$ V δ 1⁺ T cell expansion in the peripheral blood of HIV-infected persons. *J Immunol.* (1995) 154:1418–31.

42. Rossol R, Dobmeyer JM, Dobmeyer TS, Klein SA, Rossol S, Wesch D, et al. Increase in V δ 1⁺ γ δ T cells in the peripheral blood and bone marrow as a selective feature of HIV-1 but not other virus infections. *Br J Haematol.* (1998) 100:728–34. doi: 10.1046/j.1365-2141.1998.00630.x
43. Déchanet J, Merville P, Bergé F, Bone-Mane G, Taupin J-L, Michel P, et al. Major expansion of γ δ T lymphocytes following cytomegalovirus infection in kidney allograft recipients. *J Infect Dis.* (1999) 179:1–8.
44. Barcy S, De Rosa SC, Vieira J, Diem K, Ikoma M, Casper C, et al. γ δ T cells involvement in viral immune control of chronic human herpesvirus 8 infection. *J Immunol.* (2008) 180:3417–25. doi: 10.4049/jimmunol.180.5.3417
45. Knight A, Madrigal AJ, Grace S, Sivakumaran J, Kottaridis P, Mackinnon S, et al. The role of V δ 2-negative γ δ T cells during cytomegalovirus reactivation in recipients of allogeneic stem cell transplantation. *Blood* (2010) 116:2164–72. doi: 10.1182/blood-2010-01-255166
46. Munk ME, Soboslay PT, Arnoldi J, Brattig N, Schulz Key H, Kaufmann SH. *Onchocerca volvulus* provides ligands for the stimulation of human γ δ T lymphocytes expressing V δ 1 chains. *J Infect Dis.* (1993) 168:1241–7. doi: 10.1093/infdis/168.5.1241
47. Chauhan SK, Tripathy NK, Sinha N, Nityanand S. T-cell receptor repertoire of circulating gamma delta T-cells in Takayasu's arteritis. *Clin Immunol.* (2006) 118:243–9. doi: 10.1016/j.clim.2005.10.010
48. Giacomelli R, Parzanese I, Frieri G, Passacantando A, Pizzuto F, Pimpo T, et al. Increase of circulating γ δ T lymphocytes in the peripheral blood of patients affected by active inflammatory bowel disease. *Clin Exp Immunol.* (1994) 98:83–8. doi: 10.1111/j.1365-2249.1994.tb06611.x
49. Kadivar M, Petersson J, Svensson L, Marsal J. CD8 $\alpha\beta$ ⁺ γ δ T cells: a novel T cell subset with a potential role in inflammatory bowel disease. *J Immunol.* (2016) 197:4584–92. doi: 10.4049/jimmunol.1601146
50. Halary F, Pitard V, Dlubek D, Krzysiek R, De La Salle H, Merville P, et al. Shared reactivity of V δ 2^{neg} γ δ T cells against cytomegalovirus-infected cells and tumor intestinal epithelial cells. *J Exp Med.* (2005) 201:1567–78. doi: 10.1084/jem.20041851
51. Willcox CR, Pitard V, Netzer S, Couzi L, Salim M, Silberzahn T, et al. Cytomegalovirus and tumor stress surveillance by binding of a human γ δ T cell antigen receptor to endothelial protein C receptor. *Nat Immunol.* (2012) 13:872–9. doi: 10.1038/ni.2394
52. Turner L, Lavstsen T, Berger SS, Wang CW, Petersen JE, Avril M, et al. Severe malaria is associated with parasite binding to endothelial protein C receptor. *Nature* (2013) 498:502–5. doi: 10.1038/nature12216
53. Lennartz F, Adams Y, Bengtsson A, Olsen RW, Turner L, Ndam NT, et al. Structure-guided identification of a family of dual receptor-binding PfEMP1 that is associated with cerebral malaria. *Cell Host Microbe* (2017) 21:403–14. doi: 10.1016/j.chom.2017.02.009
54. McGregor IA, Gilles HM, Walters JH, Davies AH, Pearson FA. Effects of heavy and repeated malarial infections on Gambian infants and children. Effects of erythrocytic parasitization. *Br Med J.* (1956) 32:686–92. doi: 10.1136/bmj.2.4994.686
55. Donati D, Zhang LP, Chene A, Cheng Q, Flick K, Nystrom M, et al. Identification of a polyclonal B-cell activator in *Plasmodium falciparum*. *Infect Immun.* (2004) 72:5412–8. doi: 10.1128/IAI.72.9.5412-5418.2004
56. Whittle HC, Brown J, Marsh K, Blackman M, Jobe O, Shenton F. The effects of *Plasmodium falciparum* malaria on immune control of B lymphocytes in Gambian children. *Clin Exp Immunol.* (1990) 80:213–8. doi: 10.1111/j.1365-2249.1990.tb05236.x
57. Lam KMC, Syed N, Whittle H, Crawford DH. Circulating Epstein-Barr virus-carrying B cells in acute malaria. *Lancet* (1991) 337:876–8. doi: 10.1016/0140-6736(91)90203-2
58. Moormann AM, Chelimo K, Sumba OP, Lutzke ML, Ploutz-Snyder R, Newton D, et al. Exposure to holoendemic malaria results in elevated Epstein-Barr virus loads in children. *J Infect Dis.* (2005) 191:1233–8. doi: 10.1086/428910
59. Hacker G, Kromer S, Falk M, Heeg K, Wagner H, Pfeffer K. V δ 1⁺ subset of human γ δ T cells responds to ligands expressed by EBV-infected Burkitt lymphoma cells and transformed B lymphocytes. *J Immunol.* (1992) 149:3984–9.
60. Siegers GM, Dhamko H, Wang XH, Mathieson AM, Kosaka Y, Felizardo TC, et al. Human V δ 1 γ δ T cells expanded from peripheral blood exhibit specific cytotoxicity against B-cell chronic lymphocytic leukemia-derived cells. *Cytotherapy* (2011) 13:753–64. doi: 10.3109/14653249.2011.553595
61. Fujishima N, Hirokawa M, Fujishima M, Yamashita J, Saitoh H, Ichikawa Y, et al. Skewed T cell receptor repertoire of V δ 1⁺ γ δ T lymphocytes after human allogeneic haematopoietic stem cell transplantation and the potential role for Epstein-Barr virus-infected B cells in clonal restriction. *Clin Exp Immunol.* (2007) 149:70–9. doi: 10.1111/j.1365-2249.2007.03388.x
62. Farnault L, Gertner-Dardenne J, Gondois-Rey F, Michel G, Chambost H, Hirsch I, et al. Clinical evidence implicating gamma-delta T cells in EBV control following cord blood transplantation. *Bone Marrow Transplant.* (2013) 48:1478–9. doi: 10.1038/bmt.2013.75
63. Spada FM, Grant EP, Peters PJ, Sugita M, Melián A, Leslie DS, et al. Self-recognition of CD1 by γ δ T cells: implications for innate immunity. *J Exp Med.* (2000) 191:937–48. doi: 10.1084/jem.191.6.937
64. Del Porto P, Mami-Chouaib F, Bruneau JM, Jitsukawa S, Dumas J, Harnois M, et al. TCT.1, a target molecule for γ δ T cells, is encoded by an immunoglobulin superfamily gene (Blast-1) located in the CD1 region of human chromosome 1. *J Exp Med.* (1991) 173:1339–44.
65. Delia D, Cattoretto G, Polli N, Fontanella E, Aiello A, Giardini R, et al. CD1c but neither CD1a nor CD1b molecules are expressed on normal, activated, and malignant human B cells: identification of a new B-cell subset. *Blood* (1988) 72:241–7.
66. Allan LL, Stax AM, Zheng DJ, Chung BK, Kozak FK, Tan R, et al. CD1d and CD1c expression in human B cells is regulated by activation and retinoic acid receptor signaling. *J Immunol.* (2011) 186:5261–72. doi: 10.4049/jimmunol.1003615
67. Falini B, Flenghi L, Pileri S, Pelicci P, Fagioli M, Martelli MF, et al. Distribution of T cells bearing different forms of the T cell receptor γ δ in normal and pathological human tissues. *J Immunol.* (1989) 143:2480–8.
68. Bordessoule D, Gaulard P, Mason DY. Preferential localisation of human lymphocytes bearing γ δ T cell receptors to the red pulp of the spleen. *J Clin Pathol.* (1990) 43:461–4. doi: 10.1136/jcp.43.6.461
69. Roy S, Ly D, Castro CD, Li NS, Hawk AJ, Altman JD, et al. Molecular analysis of lipid-reactive V δ 1 γ δ T cells identified by CD1c tetramers. *J Immunol.* (2016) 196:1933–42. doi: 10.4049/jimmunol.1502202
70. Halary F, Fournie JJ, Bonneville M. Activation and control of self-reactive γ δ T cells. *Microb Infect.* (1999) 1:247–53. doi: 10.1016/s1286-4579(99)80041-0
71. Born W, Cady C, Jones-Carson J, Mukasa A, Lahn M, O'Brien R. Immunoregulatory functions of γ δ T cells. *Adv Immunol.* (1999) 71:77–144.
72. Wesch D, Peters C, Siegers GM. Human gamma delta T regulatory cells in cancer: fact or fiction? *Front Immunol.* (2014) 5:598. doi: 10.3389/fimmu.2014.00598
73. Arora G, Hart GT, Manzella-Lapeira J, Doritchamou JY, Narum DL, Thomas LM, et al. NK cells inhibit *Plasmodium falciparum* growth in red blood cells via antibody-dependent cellular cytotoxicity. *Elife* (2018) 7:e36806. doi: 10.7554/eLife.36806
74. Wolf AS, Sherratt S, Riley EM. NK cells: uncertain allies against malaria. *Front Immunol.* (2017) 8:212. doi: 10.3389/fimmu.2017.00212
75. Hunter S, Willcox CR, Davey MS, Kasatskaya SA, Jeffery HC, Chudakov DM, et al. Human liver infiltrating γ δ T cells are composed of clonally expanded circulating and tissue-resident populations. *J Hepatol.* (2018) 69:654–65. doi: 10.1016/j.jhep.2018.05.007

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