



Bidirectional Crosstalk Between Cancer Stem Cells and Immune Cell Subsets

Luise Müller¹, Antje Tunger^{1,2}, Ioana Plesca¹, Rebekka Wehner^{1,2,3}, Achim Temme^{2,3,4}, Dana Westphal⁵, Friedegund Meier^{2,3,5}, Michael Bachmann^{2,3,6} and Marc Schmitz^{1,2,3*}

¹ Faculty of Medicine Carl Gustav Carus, Institute of Immunology, TU Dresden, Dresden, Germany, ² National Center for Tumor Diseases, Partner Site Dresden, Dresden, Germany, ³ German Cancer Consortium (DKTK), Partner Site Dresden, and German Cancer Research Center, Heidelberg, Germany, ⁴ Department of Neurosurgery, Section Experimental Neurosurgery and Tumor Immunology, University Hospital Carl Gustav Carus, TU Dresden, Dresden, Germany, ⁵ Department of Dermatology, University Hospital Carl Gustav Carus, TU Dresden, Dresden, Germany, ⁶ Department of Radioimmunology, Institute of Radiopharmaceutical Cancer Research, Helmholtz Center Dresden-Rossendorf, Dresden, Germany

OPEN ACCESS

Edited by:

Sergio Rutella, Nottingham Trent University, United Kingdom

Reviewed by:

Pier Paolo Claudio, University of Mississippi, United States Benjamin Bonavida, University of California, Los Angeles, United States

> *Correspondence: Marc Schmitz marc.schmitz@tu-dresden.de

Specialty section:

This article was submitted to Cancer Immunity and Immunotherapy, a section of the journal Frontiers in Immunology

> Received: 26 November 2019 Accepted: 20 January 2020 Published: 05 February 2020

Citation:

Müller L, Tunger A, Plesca I, Wehner R, Temme A, Westphal D, Meier F, Bachmann M and Schmitz M (2020) Bidirectional Crosstalk Between Cancer Stem Cells and Immune Cell Subsets. Front. Immunol. 11:140. doi: 10.3389/fimmu.2020.00140 Cancer stem cells (CSCs), also known as tumor-initiating cells, are characterized by an increased capacity for self-renewal, multipotency, and tumor initiation. While CSCs represent only a small proportion of the tumor mass, they significantly account for metastatic dissemination and tumor recurrence, thus making them attractive targets for therapy. Due to their ability to sustain in dormancy, chemo- and radiotherapy often fail to eliminate cancer cells with stemness properties. Recent advances in the understanding of the tumor microenvironment (TME) illustrated the importance of the immune contexture, determining the response to therapy and clinical outcome of patients. In this context, CSCs exhibit special properties to escape the recognition by innate and adaptive immunity and shape the TME into an immunosuppressive, pro-tumorigenic landscape. As CSCs sculpt the immune contexture, the phenotype and functional properties of the tumor-infiltrating immune cells in turn influence the differentiation and phenotype of tumor cells. In this review, we summarize recent studies investigating main immunomodulatory properties of CSCs and their underlying molecular mechanisms as well as the impact of immune cells on cancer cells with stemness properties. A deeper understanding of this bidirectional crosstalk shaping the immunological landscape and determining therapeutic responses will facilitate the improvement of current treatment modalities and the design of innovative strategies to precisely target CSCs.

Keywords: tumor microenvironment, cancer stem cells, macrophages, T cells, myeloid-derived suppressor cells

INTRODUCTION

In recent years, the role of the tumor microenvironment (TME) has gained an increasing amount of attention, allowing the discovery of new concepts and development of novel therapeutic approaches. Especially immune checkpoint inhibitors emerged as a promising tool to use the power of pre-existing tumor-specific T cells to target cancer cells (1, 2). Despite these advances, a significant fraction of patients fails to respond to those therapies or develops resistance and even patients with a complete response often present with recurrence and metastatic lesions later on. While there are several factors determining the response to therapy and long-term outcome,

1

cancer stem cells (CSCs) are considered to play an important role in the resistance, metastasis, and recurrence of tumors (3). These cells, also known as tumor-initiating cells, exhibit an enhanced capacity for self-renewal, multipotency, and tumorigenicity and were first described in acute myeloid leukemia (4). Subsequently, CSCs were also identified in a multitude of solid cancers and further characterized by various cell surface markers (5). The most common markers are CD44 and CD133, with CD44 being utilized to isolate CSCs from breast, prostate, gastric, as well as head and neck squamous cell cancer (HNSCC). CD133 on the other hand is widely used to identify CSCs in glioblastoma, lung cancer, and sarcomas. Recently, more markers have been established, for example CD90 for breast cancer and glioblastoma, CD117 for breast, ovarian, lung cancer, and glioblastoma, and CD29 for breast and colon cancer (6). Importantly, due to the high plasticity of CSCs and the shared marker expression on other cells, none of these molecules is sufficient to be used as a standalone marker. So far, no consensus regarding marker combinations for the characterization and isolation of CSCs has been reached, complicating the comparison of different studies and partly explaining conflicting results. Other characteristics of CSCs that can be used for their identification are an increased activity of aldehyde dehydrogenase (ALDH) and the high expression of efflux pumps, which allows discrimination based on the exclusion of vital dyes (7, 8). The upregulation of efflux pumps is, together with a decreased sensitivity to apoptosis, an altered cell cycle, and DNA damage repair, associated with an enhanced resistance to chemotherapy (9). Considering all of these characteristics, the interaction between immune cells and CSCs is highly important, as they show distinct properties to shape the TME and represent an attractive target for therapy to improve the long-term outcome of patients.

IMPACT OF CSCs ON THE FUNCTIONAL PROPERTIES OF IMMUNE CELLS

Previous studies revealed the importance of the TME for both prognosis and treatment of malignant diseases and improved the understanding of the highly complex crosstalk between the TME and tumor cells (10, 11). It has been shown that tumor cells themselves can influence the immune contexture by expressing cell membrane-associated coinhibitory receptors or secreting various soluble factors to modulate certain immune subsets, shaping the TME into an immunosuppressive landscape. Due to the outstanding importance of CSCs regarding resistance, metastasis, and recurrence, their role in modulating the TME in general and major immune subsets in particular are discussed below (**Figure 1**).

Macrophages

Tumor-associated macrophages (TAMs) can exhibit both proand antitumor properties depending on the microenvironment and their polarization. The current understanding of macrophage polarization includes the classically activated, pro-inflammatory M1 phenotype and the alternatively activated, anti-inflammatory M2 phenotype (12-14). In general, M1 macrophages exhibit anti-tumor immunity, while M2 macrophages display immunosuppressive properties and contribute to tumor progression. Therefore, a high M1/M2 ratio is associated with improved survival in multiple cancer entities (15-17). Tumor cells are known to influence the composition of intratumoral macrophages by directly recruiting M2 macrophages or driving the polarization of both tissue-resident macrophages and recruited macrophages toward a M2 phenotype (18, 19). Several studies pointed out a particular importance of CSCs in driving the recruitment of macrophages and subsequent M2 polarization. For example, Yi et al. showed that gliomaderived CSCs are associated with an increased density of intratumoral macrophages and exhibit a stronger capacity to recruit macrophages into the TME (20). Periostin, a secreted extracellular matrix protein, produced by glioma stem cells was shown to attract M2 TAMs, promoting tumor growth (21). Another study also proposed that M2 macrophages are recruited by tumor-initiating cells already at the single cell stage and that the elimination of these macrophages may abolish early tumorigenesis (22). Besides the recruitment of macrophages, CSCs are able to promote the macrophage polarization toward the M2 phenotype via secretion of various cytokines and growth factors (23). For example, the CSC-mediated M2 polarization of macrophages was shown to be driven by cyclooxygenase-2 and CCL2 in ovarian cancer (24). Similarly, glioma CSCs polarize macrophages and microglia by the production of colony-stimulating factors (CSFs), transforming growth factor β (TGF- β), and macrophage inhibitory cytokine 1 (MIC-1), and glioma CSC-conditioned medium promoted the immunosuppressive properties of macrophages (25-27). In particular, chemoresistant CSCs have a unique capacity to produce proinflammatory cytokines and soluble factors such as CCL2 and CSF-1, known to recruit and polarize macrophages (28). Altogether, this data suggests that the CSC-mediated accumulation of M2 macrophages may contribute to the generation of an immunosuppressive, pro-tumorigenic TME.

T Cells

CSCs are also able to alter the composition and functional properties of tumor-specific effector T cells and promote the expansion of immunosuppressive, pro-tumorigenic regulatory T cells (Tregs). While a dense infiltration of CD3⁺ T cells generally correlates with a good clinical outcome in many cancers, Tregs in particular are mainly associated with a poor prognosis, with the exception of colorectal and gastric cancers (11). For example, glioblastoma CSCs were shown to produce TGF-β, promoting the induction of Tregs, and CCL2, a chemokine involved in the recruitment of Tregs (29). Furthermore, CSCconditioned medium inhibited T cell proliferation and this effect could be reversed by blocking the signal transducer and activator of transcription 3 (STAT3) pathway. Similarly, CD44⁺ CSCs from HNSCC exhibited a significantly increased secretion of interleukin (IL)-8, granulocyte-CSF, and TGF- β compared to their CD44 $^-$ counterparts, suppressing the proliferation of T cells and Th1 responses while supporting the generation of Tregs (30). Another study revealed that the levels



FIGURE 1 | CSCs alter the composition and functional properties of immune cell subsets, which results in an impaired CSC recognition and elimination by the immune system. CSCs produce soluble molecules such as periostin, CCL2, MIC-1, CSFs, and TGF-β to attract macrophages and drive their M2 polarization. The secretion of CCL2 and CCL5 recruits Tregs into the TME, and IDO and TGF-β facilitate the generation of Tregs from naïve CD4⁺ T cells. The extracellular matrix protein tenascin-C inhibits T cell proliferation by interfering with the T cell receptor signaling pathway. Additionally, the immunomodulatory enzyme IDO impairs the expansion of effector T cells through accumulation of tryptophan catabolites, which also increases the generation of Tregs. VEGF produced by CSCs supports the induction of angiogenesis, which is critical for tumor growth, and promotes the expression of PD-1 by CD8⁺ T cells. Additionally, CSCs avoid the recognition by CD8⁺ T cells and NK cells by reducing the surface expression of HLA class I molecules and NKG2DL, respectively. The ligation of PD-L1 expressed by CSCs to PD-1 on effector T cells decreases their proliferation and IFN-γ production or leads to apoptosis. Furthermore, CSCs express the coinhibitory molecule CD47, which inhibits phagocytosis by macrophages due to repositioning of its ligand SIRPα.

of indoleamine-2,3-dioxygenase (IDO) were particularly elevated in CSCs from breast cancer, prostate cancer, and mesothelioma cell lines, as well as primary human glioblastoma cells (31). The immunomodulatory enzyme IDO substantially suppresses T cell expansion and is involved in the generation and activation of Tregs (32). Recently, exosomes containing the extracellular matrix component tenascin-C were shown to be secreted by brain tumor CSCs, inhibiting T cell activity (33). Furthermore, You et al. proposed that ovarian CD133⁺ CSCs recruit Tregs via CCL5 and additionally enhance their immunosuppressive properties, namely the secretion of IL-10 (34). Moreover, CSCs produced elevated levels of vascular endothelial growth factor (VEGF), which is known to promote angiogenesis and increase the expression of programmed cell death 1 (PD-1) on CD8⁺ T cells (35-37). Importantly, emerging evidence suggests that the immune checkpoint molecule programmed cell death ligand 1 (PD-L1) is expressed by CSCs from glioblastoma (38), HNSCC (39), breast, and colon cancer (40, 41).

IMMUNE EVASION BY CSCs

In addition to the modulation of the immune contexture, CSCs exhibit various properties to directly evade effector mechanisms of immune cells. The prerequisite for the elimination of tumor cells by activated CD8⁺ T cells is the presentation of peptides on the cell surface via human leukocyte antigen (HLA) class I molecules. Various molecules are involved in this process, creating a multitude of possibilities to alter the HLA class I peptide complex expression to evade the recognition by the adaptive immune system. While tumor cells in general are well-known to downregulate components of the antigen processing and presenting machinery, for example, the transporter associated with antigen processing (TAP), CSCs appear particularly specialized in this mechanism. Several groups reported a reduced expression of HLA class I or TAP molecules of CSCs in HNSCC (42), melanoma (43), glioblastoma (38), lung cancer (44), and colorectal cancer (45) in comparison to their non-stem-cell counterparts. However, Chikamatsu et al. showed a decreased expression of TAP2 in HNSCC CSCs, but were not able to find a significant difference between CD44⁺ and CD44⁻ cells with regard to other TAP molecules or the overall HLA I expression (30). Another study also failed to show a difference in the HLA class I expression of CSCs and non-CSCs in colon cancer (46). These discrepancies are likely to arise from strongly differing protocols for the isolation and culture of CSCs and also the heterogeneity of the CSC population.

In theory, missing HLA class I expression makes cancer cells more susceptible to the recognition and elimination by natural killer (NK) cells. However, conflicting results have been published, either arguing that NK cells display an enhanced recognition of CSCs, or reporting an increased evasion of NK cell-mediated killing by CSCs for example due to downregulation of activating NKG2D ligands (NKG2DL) (38, 47–50). A recent study reported that CD34⁺CD38⁻ leukemic stem cells express lower levels of NKG2DL compared to their CD34⁻ counterparts (51). Further analysis revealed that only NKG2DL⁻ AML cells exhibited chemoresistance toward cytarabine and patients with a higher proportion of NKG2DL⁺ AML cells showed a better response to chemotherapy and an improved overall survival.

Moreover, CSCs were shown to express CD47 which engages with signal regulatory protein α (SIRP α) on macrophages, inhibiting phagocytosis. Several groups reported an increased expression of CD47 on leukemic stem cells compared to non-CSCs and that the blocking of CD47 induced efficient phagocytosis (52–54). Although this mechanism seems to be predominantly found in hematologic malignancies, CD47 overexpression was also reported in lung, pancreatic, and hepatocellular carcinoma (HCC) (54–56).

IMMUNE CELLS DRIVE THE FORMATION AND MAINTENANCE OF CSCs

As CSCs shape the TME, infiltrating immune cells can in turn influence the characteristics of CSCs. In this chapter, the impact of major immune subsets on stemness properties, metastatic potential, and tumorigenicity of CSCs is summarized (**Figure 2**).

Macrophages

While CSCs are able to recruit macrophages and promote their M2 polarization, TAMs seem to play a predominant role in the maintenance of CSCs within their niche (57). IL-6, which is mainly produced by M1 macrophages, but also by subtypes of M2 macrophages, is upregulated in breast, ovarian, prostate, pancreatic, and colorectal cancer and can confer resistance against apoptosis as well as promote proliferation, invasion, metastasis, and angiogenesis (58–64). Emerging evidence also suggests an important role for IL-6 in the induction and maintenance of CSCs. For example, IL-6 produced by TAMs supported the expansion and drug resistance of CSCs through STAT3 signaling in non-small-cell lung cancer (NSCLC) and HCC (65, 66). Moreover, multiple studies showed an IL-6-STAT3-dependent conversion of non-stem-cell breast cancer cells into CSCs, partially by promoting epithelial to mesenchymal

transition (EMT) (67-69). Similarly, TGF-B secreted by M2polarized TAMs promoted EMT and acquisition of stem-like properties in HCC (70). These findings are in line with data from Wu et al. indicating that chronic TGF- β stimulation in the course of liver cirrhosis induces expression of CSC-associated genes in liver progenitor cells and therefore promotes the development of cancer (71). Moreover, especially M2 macrophages are known to secrete VEGF, which is also produced by CSCs themselves and promotes angiogenesis, tumorigenicity, and their stem-like phenotype (72-74). Additionally, the secretion of pleiotrophin by CD163⁺ TAMs in glioma fostered CSC-mediated tumor growth and the inhibition of this pathway led to decreased tumor growth and prolonged survival in mouse xenografts (75). Besides the paracrine interaction via soluble molecules, M2 macrophages in breast cancer interact with CSCs in a cell-cellcontact dependent manner (76). The ligation of CD90 to ephrin type-A receptor 4 (EphA4) on cancer cells induced downstream signaling that resulted in the production of IL-6, IL-8, and granulocyte-macrophage colony-stimulating factor (GM-CSF), thus facilitating the maintenance of the stem-cell-like niche. In summary, these findings indicate a predominant role of macrophages in driving the induction and maintenance of CSCs.

Myeloid-Derived Suppressor Cells

Myeloid-derived suppressor cells (MDSCs) play an important role in cancer-related immune suppression and are shown to significantly contribute to tumor progression, angiogenesis, and metastasis (77). Recently, several findings indicated that MDSCs are also involved in supporting the stem-cell features of CSCs. For example, MDSCs induce the expression of microRNA101 in ovarian cancer cells, promoting their stemness-properties and increasing the tumorigenic and metastatic potential (78). Additionally, MDSCs fostered the stemness of cervical cancer cells via the secretion of prostaglandin E2 (PGE2) (79). In breast cancer, MDSCs were shown to enhance the stem-like qualities of tumor cells by secretion of IL-6 and nitric oxide (NO) in a STAT3-dependent manner (80). Immunohistochemical analysis revealed that the presence of MDSCs positively correlated with the density of CSCs in breast cancer tissues and a high infiltration of MDSCs was associated with shorter overall survival, indicating their clinical relevance. Interestingly, STAT3 is also involved in the induction of monocytic-MDSCs in a mouse model of pancreatic cancer and monocytic-MDSCs subsequently increased the frequency of ALDH1^{bright} CSCs (81).

T Cells

In general, tumors with a dense infiltration of $CD3^+$ T cells are associated with a good clinical outcome, yet recent advances in the understanding of the immune contexture revealed various mechanisms by which infiltrating T cells can act in an immunosuppressive manner, promoting tumor progression. The most prominent mediators of immunosuppression within the T cell compartment are Tregs, exhibiting both contactdependent and cytokine-mediated actions to inhibit effector T cells and promote tumor progression (82, 83). While CSCs are able to support Treg accumulation in the tumor, in turn, Tregs can influence the CSC-niche. For example, Treg-conditioned



FIGURE 2 Immune cell subsets promote stemness properties in CSCs. IL-6 secreted by TAMs can convert non-stem cancer cells into CSCs and foster their drug resistance. The secretion of TGF-β by M2 macrophages can support the EMT process and the acquisition of stem-cell properties. M2 macrophages also produce VEGF, which promotes angiogenesis and the tumorigenicity of CSCs. Furthermore, the secretion of pleiotrophin by TAMs supports the CSC-driven tumor growth through activation of the Akt pathway. Besides soluble molecules, macrophages can interact with CSCs via CD90 and EphA4 in a cell-cell-contact-dependent manner, inducing the production of IL-6, IL-8, and GM-CSF. MDSCs can facilitate the expression of microRNA101 in CSCs, which increases their tumorigenicity and metastatic potential. The secretion of NO and IL-6 by MDSCs leads to a STAT3-dependent increase of the stemness properties of CSCs. Furthermore, Tregs can enhance the expression of genes, which are associated with CSCs and secrete TGF-β, which promotes the EMT and dedifferentiation of cancer cells. Moreover, Th17 cells contribute to the formation of CSCs by secretion of IL-17. Low IFN-γ levels produced by Th1 cells and CD8⁺ T cells enhance the stemness of tumor cells by activation of the Akt pathway. Non-lytic interactions of CD8⁺ T cells further promote this process by inducing the expression of genes that are associated with cancer cell dedifferentiation.

medium increased the side population of mouse breast cancer cell lines, supported their sphere forming capability and enhanced the expression of Sox2, Nanog, and Oct4 genes, which are associated with stemness-properties (84). Additionally, TGF- β , a hallmark cytokine of Tregs, is well-known to mediate the EMT process and therefore support the generation of CSCs (85). Besides the mechanism of EMT, a recent study described the acquisition of stem cell properties due to a TGF- β driven dedifferentiation process in colorectal cancer cells (86).

In the past years, the T helper cell subset Th17 has also been implicated in mediating not only anti-tumor activity, but also immunosuppression (87, 88). The characteristic cytokine of this subset IL-17 promoted the self-renewal of ovarian CSCs and induced stem cell features in pancreatic cancer cells (89, 90). Similarly, IL-17 was associated with an increased capacity for invasion, migration, and tumorigenicity in both *in-vitro* and *in-vivo* studies of gastric cancer cells (91). Further experiments

revealed that these effects are accompanied by an increase of phosphorylated STAT3, while the results significantly decreased upon blocking the STAT3 pathway, suggesting that IL-17 acts in a STAT3-dependent manner. Importantly, these studies were conducted irrespective of the source of IL-17. Even though Th17 cells are thought to be the main producers of IL-17, some studies suggest that innate immune cells account for the majority of IL-17⁺ cells (92, 93). Additionally, hypoxia-induced expression of IL-17 by FoxP3⁺ Tregs fostered the development of CSCs in colorectal cancer, although all of these findings emerged from in-vitro experiments (94). Besides immunosuppressive T cell subsets and cytokines, also low doses of interferon (IFN)-y, which is mainly produced by activated Th1 cells or CD8⁺ T cells, can increase the stemness of tumor cells in NSCLC (95). Furthermore, Stein and colleagues demonstrated that ineffective, non-lytic interactions of CD8⁺ T cells with breast cancer cells induced the expression of genes associated with stemness and dedifferentiation (96). Subsequent analysis of the generated tumors showed an increased proliferation, tumorigenicity, and capacity for metastasis.

Taken together, different T cell subsets, in addition to macrophages and MDSCs, assist CSCs to maintain their stemcell-like state. The finding that CSCs themselves facilitate the recruitment or induction of Tregs within the tumor illustrates the strong bidirectional crosstalk between CSCs and various immune cell subsets which shapes both the TME and the CSC niche.

CONCLUSION

Emerging evidence suggests that not only genetic alterations determine the development and fate of the tumor, but also the phenotype and functional properties of infiltrating immune cells. As discussed in this review, CSCs are able to shape the TME by attracting immunosuppressive cell subsets and inhibiting effector T cells. Vice versa, infiltrating immune cells interact with CSCs in various ways to promote their self-renewal, tumorigenicity, and metastasis. These findings emphasize the unique role of CSCs and the immense potential that lies in targeting them. Consequently, therapeutic strategies leading to the elimination of CSCs in addition to non-stem cancer cells may further improve the clinical outcome for tumor patients. Many of the aforementioned CSC-immune cell interactions, including the generation of M2 macrophages and MDSCs, the CSC-dependent T cell suppression, the effect of IL-6 and IL-17 on the stemness properties of CSCs, and the expression of PD-L1 are dependent on active STAT3 signaling in CSCs or immune cells. Many of these effects could be reversed by inhibition of STAT3, rendering this molecule an attractive therapeutic target to tackle both the induction of an immunosuppressive TME and the emerging consolidation of the CSC-niche (25, 29, 39, 91). For example, the STAT3 inhibitor napabucasin was shown to reduce stemness gene expression and sphere formation in different entities (97–99). Furthermore, the SIRP α ligand CD47 is overexpressed by CSCs and represents another target structure for therapy. Several studies showed an increased phagocytosis of CSCs by macrophages upon blocking of CD47 and multiple CD47 inhibitors are tested in ongoing clinical trials (53-55, 100, 101). Additionally, CSCs were shown to express increased levels of the immune checkpoint PD-L1 and PD-L1

REFERENCES

- Ribas A, Wolchok JD. Cancer immunotherapy using checkpoint blockade. Science. (2018) 359:1350–5. doi: 10.1126/science.aar4060
- Postow MA, Callahan MK, Wolchok JD. Immune checkpoint blockade in cancer therapy. J Clin Oncol. (2015) 33:1974– 82. doi: 10.1200/JCO.2014.59.4358
- Kreso A, Dick JE. Evolution of the cancer stem cell model. Cell Stem Cell. (2014) 14:275–91. doi: 10.1016/j.stem.2014.02.006
- Lapidot T, Sirard C, Vormoor J, Murdoch B, Hoang T, Caceres-Cortes J, et al. A cell initiating human acute myeloid leukaemia after transplantation into SCID mice. *Nature*. (1994) 367:645–8. doi: 10.1038/367645a0
- 5. Tirino V, Desiderio V, Paino F, De Rosa A, Papaccio F, La Noce M, et al. Cancer stem cells in solid tumors: an overview and new

in turn promoted the generation of CSCs, creating a rationale for combination therapies with checkpoint inhibitors (1, 102). Furthermore, TGF- β secreted by Tregs and M2 macrophages or CSCs themselves is a crucial mediator of immunosuppression that can be targeted by neutralizing antibodies or receptor kinase inhibitors (103). The inhibition of the pro-angiogenic molecule VEGF has also been proven beneficial as combinational therapy in multiple entities and could be used to disrupt both the CSC-mediated angiogenesis and the induction of stemnessproperties by macrophages (104, 105). In addition to targeting the crosstalk between CSCs and the TME, CSCs can be eliminated by using specific immunotherapeutic approaches, such as drugconjugated monoclonal antibodies, bispecific antibodies, and chimeric antigen receptor- or T cell receptor-engineered T cells, targeting antigens that are characteristically expressed by CSCs (106-109).

The described studies exploring important immunmodulatory capabilities of CSCs and the impact of various immune cell subsets on cancer cells with stemness properties led to a deeper understanding of the bidirectional crosstalk between CSCs and the immune system. However, many studies used isolated CSCs to determine their phenotype and properties, which has several limitations and can significantly influence the results (110, 111). For example, the dissociation of solid tumors usually requires enzymatic treatment, which may result in the reduction or elimination of phenotypically and/or functionally relevant CSC-associated surface molecules. The different CSC isolation procedures may also reduce the viability of the purified cells. In addition, the characteristics of CSCs in solid tumors rely on the direct interaction of CSCs with various cellular components of the TME and the extracellular matrix, which is not appropriately considered, when utilizing isolated CSCs. To circumvent these limitations, advanced technologies to explore CSCs in intact tumors, such as lineage tracing approaches, may help to gain novel insights into the phenotype and properties of CSCs and may enable the design of improved therapies to target CSCs.

AUTHOR CONTRIBUTIONS

LM and ATu drafted the manuscript. IP, RW, ATe, DW, FM, MB, and MS reviewed and edited the manuscript.

approaches for their isolation and characterization. FASEB J. (2013) 27:13–24. doi: 10.1096/fj.12-218222

- Karsten U, Goletz S. What makes cancer stem cell markers different? Springerplus. (2013) 2:301. doi: 10.1186/2193-1801-2-301
- Moitra K. Overcoming multidrug resistance in cancer stem cells. *Biomed Res Int.* (2015) 2015:635745. doi: 10.1155/2015/63 5745
- Vassalli G. Aldehyde dehydrogenases: not just markers, but functional regulators of stem cells. Stem Cells Int. (2019) 2019:3904645. doi: 10.1155/2019/3904645
- Izumiya M, Kabashima A, Higuchi H, Igarashi T, Sakai G, Iizuka H, et al. Chemoresistance is associated with cancer stem cell-like properties and epithelial-to-mesenchymal transition in pancreatic cancer cells. *Anticancer Res.* (2012) 32:3847–53.

- Binnewies M, Roberts EW, Kersten K, Chan V, Fearon DF, Merad M, et al. Understanding the tumor immune microenvironment (TIME) for effective therapy. *Nat Med.* (2018) 24:541–50. doi: 10.1038/s41591-018-0014-x
- Fridman WH, Zitvogel L, Sautès-Fridman C, Kroemer G. The immune contexture in cancer prognosis and treatment. *Nat Rev Clin Oncol.* (2017) 14:717–34. doi: 10.1038/nrclinonc.2017.101
- Cassetta L, Cassol E, Poli G. Macrophage polarization in health and disease. Sci World J. (2011) 11:2391–402. doi: 10.1100/2011/213962
- Murray PJ, Allen JE, Biswas SK, Fisher EA, Gilroy DW, Goerdt S, et al. Macrophage activation and polarization: nomenclature and experimental guidelines. *Immunity*. (2014) 41:14–20. doi: 10.1016/j.immuni.2014.06.008
- Murray PJ. Macrophage polarization. Annu Rev Physiol. (2017) 79:541– 66. doi: 10.1146/annurev-physiol-022516-034339
- Pantano F, Berti P, Guida FM, Perrone G, Vincenzi B, Amato MMC, et al. The role of macrophages polarization in predicting prognosis of radically resected gastric cancer patients. J Cell Mol Med. (2013) 17:1415– 21. doi: 10.1111/jcmm.12109
- Zhang M, He Y, Sun X, Li Q, Wang W, Zhao A, et al. A high M1/M2 ratio of tumor-associated macrophages is associated with extended survival in ovarian cancer patients. *J Ovarian Res.* (2014) 7:19. doi: 10.1186/1757-2215-7-19
- 17. Zhang S, Zhang E, Long J, Hu Z, Peng J, Liu L, et al. Immune infiltration in renal cell carcinoma. *Cancer Sci.* (2019) 110:1564–72. doi: 10.1111/cas.13996
- Mantovani A, Sozzani S, Locati M, Allavena P, Sica A. Macrophage polarization: tumor-associated macrophages as a paradigm for polarized M2 mononuclear phagocytes. *Trends Immunol.* (2002) 23:549–55. doi: 10.1016/S1471-4906(02)02302-5
- Rhee I. Diverse macrophages polarization in tumor microenvironment. Arch Pharm Res. (2016) 39:1588–96. doi: 10.1007/s12272-016-0820-y
- Yi L, Xiao H, Xu M, Ye X, Hu J, Li F, et al. Glioma-initiating cells: a predominant role in microglia/macrophages tropism to glioma. J Neuroimmunol. (2011) 232:75–82. doi: 10.1016/j.jneuroim.2010. 10.011
- Zhou W, Ke SQ, Huang Z, Flavahan W, Fang X, Paul J, et al. Periostin secreted by glioblastoma stem cells recruits M2 tumour-associated macrophages and promotes malignant growth. *Nat Cell Biol.* (2015) 17:170– 82. doi: 10.1038/ncb3090
- Guo X, Zhao Y, Yan H, Yang Y, Shen S, Dai X, et al. Single tumor-initiating cells evade immune clearance by recruiting type II macrophages. *Genes Dev.* (2017) 31:247–59. doi: 10.1101/gad.294348.116
- Jinushi M. Role of cancer stem cell-associated inflammation in creating proinflammatory tumorigenic microenvironments. *Oncoimmunology*. (2014) 3:e28862. doi: 10.4161/onci.28862
- 24. Zhang Q, Cai D-J, Li B. Ovarian cancer stem-like cells elicit the polarization of M2 macrophages. *Mol Med Rep.* (2015) 11:4685–93. doi: 10.3892/mmr.2015.3323
- Wu A, Wei J, Kong L-Y, Wang Y, Priebe W, Qiao W, et al. Glioma cancer stem cells induce immunosuppressive macrophages/microglia. *Neuro Oncol.* (2010) 12:1113–25. doi: 10.1093/neuonc/noq082
- Kokubu Y, Tabu K, Muramatsu N, Wang W, Murota Y, Nobuhisa I, et al. Induction of protumoral CD11c(high) macrophages by glioma cancer stem cells through GM-CSF. *Genes Cells*. (2016) 21:241–51. doi: 10.1111/gtc.12333
- 27. Nusblat LM, Carroll MJ, Roth CM. Crosstalk between M2 macrophages and glioma stem cells. *Cell Oncol.* (2017) 40:471–82. doi: 10.1007/s13402-017-0337-5
- Yamashina T, Baghdadi M, Yoneda A, Kinoshita I, Suzu S, Dosaka-Akita H, et al. Cancer stem-like cells derived from chemoresistant tumors have a unique capacity to prime tumorigenic myeloid cells. *Cancer Res.* (2014) 74:2698–709. doi: 10.1158/0008-5472.CAN-13-2169
- Wei J, Barr J, Kong L-Y, Wang Y, Wu A, Sharma AK, et al. Glioblastoma cancer-initiating cells inhibit T-cell proliferation and effector responses by the signal transducers and activators of transcription 3 pathway. *Mol Cancer Ther.* (2010) 9:67–78. doi: 10.1158/1535-7163.MCT-09-0734
- Chikamatsu K, Takahashi G, Sakakura K, Ferrone S, Masuyama K. Immunoregulatory properties of CD44+ cancer stem-like cells in squamous cell carcinoma of the head and neck. *Head Neck.* (2011) 33:208– 15. doi: 10.1002/hed.21420

- Stapelberg M, Zobalova R, Nguyen MN, Walker T, Stantic M, Goodwin J, et al. Indoleamine-2,3-dioxygenase elevated in tumorinitiating cells is suppressed by mitocans. *Free Radic Biol Med.* (2014) 67:41–50. doi: 10.1016/j.freeradbiomed.2013.10.003
- Prendergast GC, Smith C, Thomas S, Mandik-Nayak L, Laury-Kleintop L, Metz R, et al. Indoleamine 2,3-dioxygenase pathways of pathgenic inflammation and immune escape in cancer. *Cancer Immunol Immunother*. (2014) 63:721–35. doi: 10.1007/s00262-014-1549-4
- 33. Mirzaei R, Sarkar S, Dzikowski L, Rawji KS, Khan L, Faissner A, et al. Brain tumor-initiating cells export tenascin-C associated with exosomes to suppress T cell activity. *Oncoimmunology*. (2018) 7:e1478647. doi: 10.1080/2162402X.2018.1478647
- 34. You Y, Li Y, Li M, Lei M, Wu M, Qu Y, et al. Ovarian cancer stem cells promote tumour immune privilege and invasion via CCL5 and regulatory T cells. *Clin Exp Immunol.* (2018) 191:60–73. doi: 10.1111/cei.13044
- 35. Bao S, Wu Q, Sathornsumetee S, Hao Y, Li Z, Hjelmeland AB, et al. Stem cell–like glioma cells promote tumor angiogenesis through vascular endothelial growth factor. *Cancer Res.* (2006) 66:7843–8. doi: 10.1158/0008-5472.CAN-06-1010
- Folkins C, Shaked Y, Man S, Tang T, Lee CR, Zhu Z, et al. Glioma tumor stem-like cells promote tumor angiogenesis and vasculogenesis via vascular endothelial growth factor and stromal-derived factor 1. *Cancer Res.* (2009) 69:7243–51. doi: 10.1158/0008-5472.CAN-09-0167
- Voron T, Colussi O, Marcheteau E, Pernot S, Nizard M, Pointet A-L, et al. VEGF-A modulates expression of inhibitory checkpoints on CD8+ T cells in tumors. J Exp Med. (2015) 212:139–48. doi: 10.1084/jem.20140559
- Tomaso TD, Mazzoleni S, Wang E, Sovena G, Clavenna D, Franzin A, et al. Immunobiological characterization of cancer stem cells isolated from glioblastoma patients. *Clin Cancer Res.* (2010) 16:800–13. doi: 10.1158/1078-0432.CCR-09-2730
- Lee Y, Shin JH, Longmire M, Wang H, Kohrt HE, Chang HY, et al. CD44+ cells in head and neck squamous cell carcinoma suppress T-cell-mediated immunity by selective constitutive and inducible expression of PD-L1. *Clin Cancer Res.* (2016) 22:3571–81. doi: 10.1158/1078-0432.CCR-15-2665
- Wu Y, Chen M, Wu P, Chen C, Xu ZP, Gu W. Increased PD-L1 expression in breast and colon cancer stem cells. *Clin Exp Pharmacol Physiol.* (2017) 44:602–4. doi: 10.1111/1440-1681.12732
- Zhi Y, Mou Z, Chen J, He Y, Dong H, Fu X, et al. B7H1 expression and epithelial-to-mesenchymal transition phenotypes on colorectal cancer stemlike cells. *PLoS ONE*. (2015) 10:e0135528. doi: 10.1371/journal.pone.0135528
- 42. Grau JJ, Mesía R, Iglesia-Vicente M de la, Williams ES, Taberna M, Caballero M, et al. Enrichment of cells with cancer stem cell-like markers in relapses of chemoresistant patients with locally advanced head and neck squamous cell carcinoma. OCL. (2016) 90:267–72. doi: 10.1159/000445118
- Schatton T, Schütte U, Frank NY, Zhan Q, Hoerning A, Robles SC, et al. Modulation of T cell activation by malignant melanoma initiating cells. *Cancer Res.* (2010) 70:697–708. doi: 10.1158/0008-5472.CAN-09-1592
- Morrison BJ, Steel JC, Morris JC. Reduction of MHC-I expression limits Tlymphocyte-mediated killing of Cancer-initiating cells. *BMC Cancer*. (2018) 18:469. doi: 10.1186/s12885-018-4389-3
- Volonté A, Tomaso TD, Spinelli M, Todaro M, Sanvito F, Albarello L, et al. Cancer-initiating cells from colorectal cancer patients escape from T cell– mediated immunosurveillance *in vitro* through membrane-bound IL-4. J Immunol. (2014) 192:523–32. doi: 10.4049/jimmunol.1301342
- Inoda S, Hirohashi Y, Torigoe T, Morita R, Takahashi A, Asanuma H, et al. Cytotoxic T lymphocytes efficiently recognize human colon cancer stem-like cells. *Am J Pathol.* (2011) 178:1805–13. doi: 10.1016/j.ajpath.2011.01.004
- 47. Tseng H-C, Arasteh A, Paranjpe A, Teruel A, Yang W, Behel A, et al. Increased lysis of stem cells but not their differentiated cells by natural killer cells; de-differentiation or reprogramming activates NK cells. *PLoS ONE.* (2010) 5:e11590. doi: 10.1371/journal.pone.0011590
- Ames E, Canter RJ, Grossenbacher SK, Mac S, Chen M, Smith RC, et al. NK cells preferentially target tumor cells with a cancer stem cell phenotype. J Immunol. (2015) 195:4010–9. doi: 10.4049/jimmunol.1500447
- 49. Wang B, Wang Q, Wang Z, Jiang J, Yu S-C, Ping Y-F, et al. Metastatic consequences of immune escape from NK cell cytotoxicity by human breast cancer stem cells. *Cancer Res.* (2014) 74:5746– 57. doi: 10.1158/0008-5472.CAN-13-2563

- She M, Niu X, Chen X, Li J, Zhou M, He Y, et al. Resistance of leukemic stemlike cells in AML cell line KG1a to natural killer cell-mediated cytotoxicity. *Cancer Lett.* (2012) 318:173–9. doi: 10.1016/j.canlet.2011.12.017
- Paczulla AM, Rothfelder K, Raffel S, Konantz M, Steinbacher J, Wang H, et al. Absence of NKG2D ligands defines leukaemia stem cells and mediates their immune evasion. *Nature*. (2019) 572:254–9. doi: 10.1038/s41586-019-1410-1
- Majeti R, Chao MP, Alizadeh AA, Pang WW, Jaiswal S, Gibbs KD, et al. CD47 is an adverse prognostic factor and therapeutic antibody target on human acute myeloid leukemia stem cells. *Cell.* (2009) 138:286– 99. doi: 10.1016/j.cell.2009.05.045
- Theocharides APA, Jin L, Cheng P-Y, Prasolava TK, Malko AV, Ho JM, et al. Disruption of SIRPα signaling in macrophages eliminates human acute myeloid leukemia stem cells in xenografts. *J Exp Med.* (2012) 209:1883– 99. doi: 10.1084/jem.20120502
- Liu L, Zhang L, Yang L, Li H, Li R, Yu J, et al. Anti-CD47 antibody as a targeted therapeutic agent for human lung cancer and cancer stem cells. *Front Immunol.* (2017) 8:404. doi: 10.3389/fimmu.2017.00404
- Cioffi M, Trabulo S, Hidalgo M, Costello E, Greenhalf W, Erkan M, et al. Inhibition of CD47 effectively targets pancreatic cancer stem cells via dual mechanisms. *Clin Cancer Res.* (2015) 21:2325– 37. doi: 10.1158/1078-0432.CCR-14-1399
- Lee TK-W, Cheung VC-H, Lu P, Lau EYT, Ma S, Tang KH, et al. Blockade of CD47-mediated cathepsin S/protease-activated receptor 2 signaling provides a therapeutic target for hepatocellular carcinoma. *Hepatology*. (2014) 60:179– 91. doi: 10.1002/hep.27070
- Sainz B, Carron E, Vallespinós M, Machado HL. Cancer stem cells and macrophages: implications in tumor biology and therapeutic strategies. *Mediators Inflamm*. (2016) 2016:9012369. doi: 10.1155/2016/9012369
- Culig Z, Puhr M. Interleukin-6: a multifunctional targetable cytokine in human prostate cancer. *Mol Cell Endocrinol.* (2012) 360:52–8. doi: 10.1016/j.mce.2011.05.033
- Waldner MJ, Foersch S, Neurath MF. Interleukin-6 a key regulator of colorectal cancer development. *Int J Biol Sci.* (2012) 8:1248–53. doi: 10.7150/ijbs.4614
- 60. Chang CH, Hsiao CF, Yeh YM, Chang GC, Tsai YH, Chen YM, et al. Circulating interleukin-6 level is a prognostic marker for survival in advanced nonsmall cell lung cancer patients treated with chemotherapy. *Int J Cancer*. (2013) 132:1977–85. doi: 10.1002/ijc.27892
- Dethlefsen C, Højfeldt G, Hojman P. The role of intratumoral and systemic IL-6 in breast cancer. Breast Cancer Res Treat. (2013) 138:657– 64. doi: 10.1007/s10549-013-2488-z
- Macciò A, Madeddu C. The role of interleukin-6 in the evolution of ovarian cancer: clinical and prognostic implications–a review. J Mol Med. (2013) 91:1355–68. doi: 10.1007/s00109-013-1080-7
- Miura T, Mitsunaga S, Ikeda M, Shimizu S, Ohno I, Takahashi H, et al. Characterization of patients with advanced pancreatic cancer and high serum interleukin-6 levels. *Pancreas.* (2015) 44:756–63. doi: 10.1097/MPA.00000000000335
- Kumari N, Dwarakanath BS, Das A, Bhatt AN. Role of interleukin-6 in cancer progression and therapeutic resistance. *Tumor Biol.* (2016) 37:11553– 72. doi: 10.1007/s13277-016-5098-7
- Jinushi M, Chiba S, Yoshiyama H, Masutomi K, Kinoshita I, Dosaka-Akita H, et al. Tumor-associated macrophages regulate tumorigenicity and anticancer drug responses of cancer stem/initiating cells. *PNAS.* (2011) 108:12425– 30. doi: 10.1073/pnas.1106645108
- 66. Wan S, Zhao E, Kryczek I, Vatan L, Sadovskaya A, Ludema G, et al. Tumor-associated macrophages produce interleukin 6 and signal via STAT3 to promote expansion of human hepatocellular carcinoma stem cells. *Gastroenterology*. (2014) 147:1393–404. doi: 10.1053/j.gastro.2014. 08.039
- Iliopoulos D, Hirsch HA, Wang G, Struhl K. Inducible formation of breast cancer stem cells and their dynamic equilibrium with non-stem cancer cells via IL6 secretion. *Proc Natl Acad Sci USA*. (2011) 108:1397– 402. doi: 10.1073/pnas.1018898108
- Xie G, Yao Q, Liu Y, Du S, Liu A, Guo Z, et al. IL-6-induced epithelialmesenchymal transition promotes the generation of breast cancer stem-like cells analogous to mammosphere cultures. *Int J Oncol.* (2012) 40:1171–9. doi: 10.3892/ijo.2011.1275

- Kim S-Y, Kang JW, Song X, Kim BK, Yoo YD, Kwon YT, et al. Role of the IL-6-JAK1-STAT3-Oct-4 pathway in the conversion of nonstem cancer cells into cancer stem-like cells. *Cell Signal.* (2013) 25:961– 9. doi: 10.1016/j.cellsig.2013.01.007
- 70. Fan Q-M, Jing Y-Y, Yu G-F, Kou X-R, Ye F, Gao L, et al. Tumor-associated macrophages promote cancer stem cell-like properties via transforming growth factor-beta1-induced epithelialmesenchymal transition in hepatocellular carcinoma. *Cancer Lett.* (2014) 352:160–8. doi: 10.1016/j.canlet.2014.05.008
- Wu K, Ding J, Chen C, Sun W, Ning B-F, Wen W, et al. Hepatic transforming growth factor beta gives rise to tumor-initiating cells and promotes liver cancer development. *Hepatology*. (2012) 56:2255–67. doi: 10.1002/hep.26007
- Oka N, Soeda A, Inagaki A, Onodera M, Maruyama H, Hara A, et al. VEGF promotes tumorigenesis and angiogenesis of human glioblastoma stem cells. *Biochem Biophys Res Commun.* (2007) 360:553–9. doi: 10.1016/j.bbrc.2007.06.094
- Elaimy AL, Guru S, Chang C, Ou J, Amante JJ, Zhu LJ, et al. VEGFneuropilin-2 signaling promotes stem-like traits in breast cancer cells by TAZ-mediated repression of the Rac GAP β2-chimaerin. *Sci Signal.* (2018) 11:eaao6897. doi: 10.1126/scisignal.aao6897
- Mercurio AM. VEGF/Neuropilin signaling in cancer stem cells. Int J Mol Sci. (2019) 20:490. doi: 10.3390/ijms20030490
- Shi Y, Ping Y-F, Zhou W, He Z-C, Chen C, Bian B-S-J, et al. Tumourassociated macrophages secrete pleiotrophin to promote PTPRZ1 signalling in glioblastoma stem cells for tumour growth. *Nat Commun.* (2017) 8:15080. doi: 10.1038/ncomms15080
- Lu H, Clauser KR, Tam WL, Fröse J, Ye X, Eaton EN, et al. A breast cancer stem cell niche supported by juxtacrine signalling from monocytes and macrophages. *Nat Cell Biol.* (2014) 16:1105–17. doi: 10.1038/ncb3041
- Gabrilovich DI. Myeloid-derived suppressor cells. Cancer Immunol Res. (2017) 5:3–8. doi: 10.1158/2326-6066.CIR-16-0297
- Cui TX, Kryczek I, Zhao L, Zhao E, Kuick R, Roh MH, et al. Myeloidderived suppressor cells enhance stemness of cancer cells by inducing microRNA101 and suppressing the corepressor CtBP2. *Immunity*. (2013) 39:611–21. doi: 10.1016/j.immuni.2013.08.025
- Kuroda H, Mabuchi S, Yokoi E, Komura N, Kozasa K, Matsumoto Y, et al. Prostaglandin E2 produced by myeloid-derived suppressive cells induces cancer stem cells in uterine cervical cancer. *Oncotarget.* (2018) 9:36317– 30. doi: 10.18632/oncotarget.26347
- Peng D, Tanikawa T, Li W, Zhao L, Vatan L, Szeliga W, et al. Myeloidderived suppressor cells endow stem-like qualities to breast cancer cells through IL6/STAT3 and NO/NOTCH cross-talk signaling. *Cancer Res.* (2016) 76:3156–65. doi: 10.1158/0008-5472.CAN-15-2528
- Panni RZ, Sanford DE, Belt BA, Mitchem JB, Worley LA, Goetz BD, et al. Tumor-induced STAT3 activation in monocytic myeloid-derived suppressor cells enhances stemness and mesenchymal properties in human pancreatic cancer. *Cancer Immunol Immunother*. (2014) 63:513– 28. doi: 10.1007/s00262-014-1527-x
- Takeuchi Y, Nishikawa H. Roles of regulatory T cells in cancer immunity. Int Immunol. (2016) 28:401–9. doi: 10.1093/intimm/dxw025
- Sakaguchi S, Miyara M, Costantino CM, Hafler DA. FOXP3+ regulatory T cells in the human immune system. *Nat Rev Immunol.* (2010) 10:490– 500. doi: 10.1038/nri2785
- Xu Y, Dong X, Qi P, Ye Y, Shen W, Leng L, et al. Sox2 communicates with tregs through CCL1 to promote the stemness property of breast cancer cells. *Stem Cells*. (2017) 35:2351–65. doi: 10.1002/stem.2720
- Xu J, Lamouille S, Derynck R. TGF-beta-induced epithelial to mesenchymal transition. *Cell Res.* (2009) 19:156–72. doi: 10.1038/cr.2009.5
- Nakano M, Kikushige Y, Miyawaki K, Kunisaki Y, Mizuno S, Takenaka K, et al. Dedifferentiation process driven by TGF-beta signaling enhances stem cell properties in human colorectal cancer. *Oncogene*. (2019) 38:780–93. doi: 10.1038/s41388-018-0480-0
- Tosolini M, Kirilovsky A, Mlecnik B, Fredriksen T, Mauger S, Bindea G, et al. Clinical impact of different classes of infiltrating T cytotoxic and helper cells (Th1, th2, treg, th17) in patients with colorectal cancer. *Cancer Res.* (2011) 71:1263–71. doi: 10.1158/0008-5472.CAN-10-2907
- Guéry L, Hugues S. Th17 cell plasticity and functions in cancer immunity. Biomed Res Int. (2015) 2015:314620. doi: 10.1155/2015/314620

- Zhang Y, Zoltan M, Riquelme E, Xu H, Sahin I, Castro-Pando S, et al. Immune cell production of interleukin 17 induces stem cell features of pancreatic intraepithelial neoplasia cells. *Gastroenterology*. (2018) 155:210– 23.e3. doi: 10.1053/j.gastro.2018.03.041
- Xiang T, Long H, He L, Han X, Lin K, Liang Z, et al. Interleukin-17 produced by tumor microenvironment promotes self-renewal of CD133⁺ cancer stem-like cells in ovarian cancer. Oncogene. (2015) 34:165– 76. doi: 10.1038/onc.2013.537
- Jiang Y-X, Yang S-W, Li P-A, Luo X, Li Z-Y, Hao Y-X, et al. The promotion of the transformation of quiescent gastric cancer stem cells by IL-17 and the underlying mechanisms. *Oncogene*. (2017) 36:1256– 64. doi: 10.1038/onc.2016.291
- 92. Punt S, Fleuren GJ, Kritikou E, Lubberts E, Trimbos JB, Jordanova ES, et al. Angels and demons: Th17 cells represent a beneficial response, while neutrophil IL-17 is associated with poor prognosis in squamous cervical cancer. Oncoimmunology. (2015) 4:e984539. doi: 10.4161/2162402X.2014.984539
- Cua DJ, Tato CM. Innate IL-17-producing cells: the sentinels of the immune system. Nat Rev Immunol. (2010) 10:479–89. doi: 10.1038/nri2800
- 94. Yang S, Wang B, Guan C, Wu B, Cai C, Wang M, et al. Foxp3+IL-17+ T cells promote development of cancer-initiating cells in colorectal cancer. J Leukocyte Biol. (2011) 89:85–91. doi: 10.1189/jlb.0910506
- 95. Song M, Ping Y, Zhang K, Yang L, Li F, Zhang C, et al. Low-dose IFNγ induces tumor cell stemness in tumor microenvironment of non-small cell lung cancer. *Cancer Res.* (2019) 79:3737– 48. doi: 10.1158/0008-5472.CAN-19-0596
- 96. Stein RG, Ebert S, Schlahsa L, Scholz CJ, Braun M, Hauck P, et al. Cognate nonlytic interactions between CD8+ T cells and breast cancer cells induce cancer stem cell-like properties. *Cancer Res.* (2019) 79:1507– 19. doi: 10.1002/9781119645214
- Han D, Yu T, Dong N, Wang B, Sun F, Jiang D. Napabucasin, a novel STAT3 inhibitor suppresses proliferation, invasion and stemness of glioblastoma cells. J Exp Clin Cancer Res. (2019) 38:289. doi: 10.1186/s13046-019-1289-6
- MacDonagh L, Gray SG, Breen E, Cuffe S, Finn SP, O'Byrne KJ, et al. BBI608 inhibits cancer stemness and reverses cisplatin resistance in NSCLC. *Cancer Lett.* (2018) 428:117–26. doi: 10.1016/j.canlet.2018.04.008
- Li Y, Rogoff HA, Keates S, Gao Y, Murikipudi S, Mikule K, et al. Suppression of cancer relapse and metastasis by inhibiting cancer stemness. *Proc Natl Acad Sci USA*. (2015) 112:1839–44. doi: 10.1073/pnas.1424171112
- 100. Sikic BI, Lakhani N, Patnaik A, Shah SA, Chandana SR, Rasco D, et al. First-in-human, first-in-class phase i trial of the anti-CD47 antibody Hu5F9-G4 in patients with advanced cancers. J Clin Oncol. (2019) 37:946– 53. doi: 10.1200/JCO.18.02018

- 101. Uger R, Johnson L. Blockade of the CD47-SIRPα axis: a promising approach for cancer immunotherapy. *Expert Opin Biol Th.* (2019) 20:5– 8. doi: 10.1080/14712598.2020.1685976
- 102. Gupta HB, Clark CA, Yuan B, Sareddy G, Pandeswara S, Padron AS, et al. Tumor cell-intrinsic PD-L1 promotes tumor-initiating cell generation and functions in melanoma and ovarian cancer. *Sig Transduct Target Ther.* (2016) 1:1–9. doi: 10.1038/sigtrans.2016.30
- 103. Chen Y, Di C, Zhang X, Wang J, Wang F, Yan J-F, et al. Transforming growth factor β signaling pathway: a promising therapeutic target for cancer. *J Cell Physiol.* (2020) 235:1903–14. doi: 10.1002/jcp.29108
- 104. Cardones AR, Banez LL. VEGF inhibitors in cancer therapy. Curr Pharm Des. (2006) 12:387–94. doi: 10.2174/138161206775201910
- 105. Chen DS, Hurwitz H. Combinations of bevacizumab with cancer immunotherapy. *Cancer J.* (2018) 24:193– 204. doi: 10.1097/PPO.00000000000327
- Weiner GJ. Building better monoclonal antibody-based therapeutics. Nat Rev Cancer. (2015) 15:361–70. doi: 10.1038/nrc3930
- Trabolsi A, Arumov A, Schatz JH. T cell-activating bispecific antibodies in cancer therapy. J Immunol. (2019) 203:585– 92. doi: 10.4049/jimmunol.1900496
- Fesnak AD, June CH, Levine BL. Engineered T cells: the promise and challenges of cancer immunotherapy. *Nat Rev Cancer.* (2016) 16:566– 81. doi: 10.1038/nrc.2016.97
- 109. Guo Y, Feng K, Wang Y, Han W. Targeting cancer stem cells by using chimeric antigen receptor-modified T cells: a potential and curable approach for cancer treatment. *Protein Cell.* (2018) 9:516– 26. doi: 10.1007/s13238-017-0394-6
- Batlle E, Clevers H. Cancer stem cells revisited. Nat Med. (2017) 23:1124– 34. doi: 10.1038/nm.4409
- 111. Abbaszadegan MR, Bagheri V, Razavi MS, Momtazi AA, Sahebkar A, Gholamin M. Isolation, identification, and characterization of cancer stem cells: a review. J Cell Physiol. (2017) 232:2008–18. doi: 10.1002/jcp.25759

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2020 Müller, Tunger, Plesca, Wehner, Temme, Westphal, Meier, Bachmann and Schmitz. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.