



Neutrophils: Orchestrators of the Malignant Phenotype

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Neutrophils are the first leukocytes recruited to sites of inflammation, where they execute anti-microbial functions to eliminate infectious agents. These functions include phagocytosis, release of reactive oxygen species and the formation of neutrophil extracellular traps via NETosis. Neutrophils are receiving increasing attention in the context of cancer, where these same neutrophil-associated functions are also important for modulating tumor growth and metastatic progression. Neutrophils are phenotypically heterogeneous and, depending on the context, exert anti- or pro-tumorigenic functions. Increasing evidence also suggests an important role of neutrophils and their involvement in promoting multiple steps of the metastatic cascade. The steps include: (1) local invasion and intravasation of cancer cells into circulation, (2) survival of cancer cells in the bloodstream and extravasation at a distant site, (3) early cancer cell seeding/survival, and (4) progressive growth of cancer cells to form macroscopic metastases. Although neutrophil functions designed to eliminate infectious agents can also eliminate tumor cells, their dysregulation can promote tumor growth and enable metastasis at multiple steps along the metastatic cascade. In this review, we will provide an overview of the current advances in neutrophil biology in the context of cancer. We also discuss the emerging field of immunometabolism, in which the rewiring of alternative metabolic pathways within neutrophils can impact their pro-tumorigenic/pro-metastatic functions.

Keywords: neutrophils, tumor growth, metastasis, NETosis, immunosuppression, immunometabolism, metabolic plasticity/flexibility

INTRODUCTION

Neutrophils account for 50–70% of circulating leukocytes and are the first immune cells recruited to an inflammatory site. They play an important role in the innate immune response to pathogens, as patients with neutropenia are highly susceptible to bacterial and fungal infections (1). Neutrophils perform numerous functions that target microbes, including phagocytosis, the release of anti-microbial peptides/proteases and NETosis (2). Interestingly, neutrophils have garnered considerable interest for their emerging and prominent roles in modulating cancer growth and metastatic progression (3). The roles played by neutrophils in the cancer setting are diverse and complex, leading to the concept of neutrophil heterogeneity/plasticity and the notion that distinct neutrophil subsets might exist.

Granulopoiesis is a tightly regulated process that involves the differentiation and mobilization of mature segmented neutrophils from the bone marrow into circulation. This process begins with the commitment of granulocyte-monocyte myeloid

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progenitors (GMPs), which progress through a series of neutrophil progenitors (myeloblast, pro-myelocyte, myelocyte, meta-myelocyte, band cell) until they become a mature neutrophil (4).

In cancer, dysregulated granulopoiesis has led to the identification of different neutrophil subsets that play a role in tumor progression. PreNeus comprise a neutrophil precursor population that retain their proliferative capacity and expand in the bone marrow and spleen of tumor bearing mice (5). PreNeus differentiate into immature and mature neutrophils, with the former found to accumulate in growing tumors (5). An early stage committed unipotent neutrophil precursor (NeP) has also been identified and their adoptive transfer into humanized mice promoted solid tumor growth by inhibiting T cell activation (6). Two neutrophil subsets, high-density neutrophils (HDNs), and low-density neutrophils (LDNs) were identified in various tumor models by differential density centrifugation (7). HDNs represent mature, segmented neutrophils whereas LDNs comprise a heterogeneous mixture of mature and immature neutrophils (7). Increasing mobilization of LDNs into the peripheral blood was associated with enhanced tumor growth and metastasis (7-9).

In addition to the identification of distinct neutrophil subsets, neutrophils exhibit plasticity in response to tumor-derived factors in a manner similar to macrophages. Neutrophils have been classified into two categories, N1 and N2, to describe their pro- and anti-tumorigenic functions, respectively, (10, 11). In vivo evidence has shown that tumor-associated neutrophils (TANs) can change their function from a protumor phenotype (N2) to an anti-tumor (N1) phenotype with the addition of a TGF β inhibitor, arguing that TGF β is an important factor driving the N2 phenotype (10). In contrast, signals associated with an anti-tumor (N1) phenotype include type I interferons and those propagated by the MET receptor (12, 13). However, this categorization is likely to represent an oversimplification of neutrophil diversity. Neutrophil polarization, similar to macrophages, could also represent a continuum of different neutrophil phenotypes present in the tumor microenvironment (14). These advances regarding the degree of neutrophil heterogeneity/plasticity observed in the cancer setting have sparked an intense and renewed interest in this cell population. While there are ongoing discussions in the field regarding the relationship between PMN-MDSCs and neutrophil subsets, we direct the reader to excellent reviews that fully discuss these relationships (3, 15). We will briefly discuss anti-tumor neutrophil functions; however, this review will primarily discuss the recent roles of neutrophils and neutrophilassociated functions in promoting tumor growth and metastatic progression.

ANTI-TUMOR NEUTROPHIL FUNCTIONS

Neutrophils can participate in a variety of anti-tumor mechanisms that limit tumor growth or eliminate cancer cells (16). A well-studied neutrophil-associated function is

their ability to generate reactive oxygen species (ROS) to limit tumor progression. Upon tumor cell contact, mousederived neutrophils can release hydrogen peroxide to eliminate metastatic cancer cells in vitro (17). Subsequently, it was demonstrated that expression of TRPM2 (transient receptor potential cation channel, subfamily M2) on tumor cells increased their sensitivity to neutrophil-mediated, H2O2-dependent, cytotoxicity. This occurred through a mechanism that involved a transient increase in Ca²⁺ mobilization within cancer cells (18). TRPM2 upregulation in tumor cells occurred following an epithelial-to-mesenchymal transition (EMT) and cancer cells that have undergone an EMT were more susceptible to neutrophil-mediated killing (19). More recently, an interaction between the receptor for advanced glycation end products (RAGE), which is expressed on tumor cells, and cathepsin G present on murine neutrophils was shown to mediate in vitro tumor cell cytotoxicity in a H₂O₂-dependent manner (20). The release of neutrophil ROS is also dependent on the tumor microenvironment. In hypoxic tumor microenvironments, the ability of murine neutrophils to kill tumor cells in vivo through the release of ROS is greatly diminished (21). Thus, neutrophils have the capacity to mediate ROS-dependent direct tumor cell killing.

The interplay of neutrophils with other immune cell types can also indirectly limit tumor progression. Tumor associated neutrophils suppress the pro-tumorigenic role of IL-17 secreting $\gamma\delta$ T cells by inhibiting their proliferation. Low glutathione levels in $\gamma \delta 17 \text{ T}$ cells rendered them sensitive to neutrophil-derived ROS, causing enhanced oxidative stress, and reduced proliferation (22). In early-stage human lung cancer, a subset of immature neutrophils have been identified as having antigen-presenting functions and act to promote anti-tumor immunity by stimulating the secretion of inflammatory cytokines from T lymphocytes (23). In addition to neutrophil-T cell interactions, communication between neutrophils and monocytes can also elicit anti-tumor effects. Non-metastatic cancer cells can mobilize IFNyproducing monocytes to the lungs. IFNy release activates TMEM173/STING within neutrophils, which stimulates neutrophil-mediated killing of disseminated cancer cells in the lungs (24).

Neutrophils have been shown to infiltrate deposits of prostate cancer cells within bone metastases. Importantly, neutrophils impaired bone metastasis progression by inhibiting STAT5 (signal transducer and activator of transcription 5) function within prostate cancer cells, resulting in their apoptotic cell death (25). Recently, neutrophils have been reported to be involved in antibody-mediated trogocytosis, a process that mechanically disrupts the plasma membrane of antibody-opsinized cancer cells, leading to a lytic/necrotic-type cell death. IgA antibodies against receptors expressed by cancer cells (Her2, EGFR) could enhance neutrophil-mediated trogocytosis of cancer cells if the CD47-SIRPa innate immune cell checkpoint was simultaneously blocked (26, 27). Taken together, these results demonstrate that neutrophils can impair tumor growth and metastasis using a combination of direct and indirect cancer cell killing mechanisms (Supplementary Table 1).



FIGURE 1 Neutrophil functions that promote tumor growth and metastasis. To support primary tumor growth, neutrophils can mediate T cell suppression and alter macrophage differentiation. Neutrophil release of TIMP-1 enhances tumor cell invasion by inducing epithelial-to-mesenchymal transition. Once in circulating, circulating tumor cells interact with neutrophils, which enables tumor cell proliferation. Secretion of various pro-inflammatory markers such as IL-8, IL-1β, or MMPs can mediate increased tumor cell extravasation. In addition, neutrophils can inhibit intraluminal NK-mediated killing of circulating cancer cells, leading to increased extravasation. At the metastatic site, various systemic, and microenvironmental factors can promote neutrophil infiltration. Neutrophils can awaken dormant cancer cells by promoting ECM remodeling and angiogenesis. Lastly, continued growth of the metastatic lesion is facilitated by key neutrophil-dependent mechanisms, which include angiogenesis, proliferation, immune suppression, and immune exclusion. CSF-1, colony stimulating factor 1; TIMP1, tissue inhibitor of matrix metalloprotease; PD-L1, programmed death ligand 1; TGFβ, transforming growth factor β; ROS, reactive oxygen species; MMP, matrix metalloproteinases; GM-CSF, granulocyte macrophage colony stimulating factor; ANGPTL2, angiopoetin like-2; FGF2, fibroblast growth factor 2; LTB₄, leukotriene B₄; iNOS, inducible nitric oxide synthase; NET, neutrophil extracellular trap; CAF, cancer-associated fibroblast.

NEUTROPHIL FUNCTIONS THAT PROMOTE PRIMARY TUMOR GROWTH

Neutrophils promote primary tumor growth by various mechanisms (**Figure 1**). NETosis is a process that involves the extrusion of neutrophil-derived chromatin structures that are decorated with neutrophil granule constituents, which form extracellular structures called neutrophil extracellular traps (NETs) (28). Normally, NETosis and NET production have been described in the context of a neutrophil's ability to capture and kill bacteria extracellularly (29). However, NETs have been shown to play an important role in the growth of a primary tumor. Tumor microenvironmental changes, including

tumor-associated coagulation and enhanced thrombosis, have been linked to enhanced tumor growth. Several recent studies suggest that NETosis may play an important role in these processes. LPS stimulation was shown to increase C3aR expression within neutrophils, enhance NETosis and increase coagulation. These events were correlated with N2 neutrophil polarization and increased tumor growth (30). Interestingly, it has recently been shown that immature neutrophils preferentially respond to cancer cell derived C3a to promote their migration (31). Subsequently, it was shown that breast cancer cells that expressed high levels of G-CSF and IL-1 β exhibited high neutrophil counts and tumor-associated thrombosis, which was dependent on NET formation (32). Pharmacological blockade of IL-1 receptor signaling reduced NET formation, attenuated tumor-associated thrombosis and impaired tumor growth (32). NETs can also directly influence cancer cell proliferation. Neutrophil elastase (NE) present within NETs activates tumor cells to increase mitochondria biogenesis and ATP production; thereby, further enhancing the growth of cancer cells (33).

In addition to the impact of NETs, neutrophils can also interact with other immune cells through additional mechanisms to promote tumor growth. Neutrophil-derived ROS can inhibit T cell proliferation, creating an immunosuppressive environment that is supportive of tumor growth (34). Phenotypic characterization and single-cell RNA sequencing identified a neutrophil subset that is CD84^{hi}, which exhibited potent T cell suppressive activity and increased ROS production (35). In a model of gastric cancer, neutrophils were activated by tumorderived GM-CSF that resulted in elevated programmed death ligand 1 (PD-L1) expression. These PD-L1⁺ neutrophils were able to suppress T cell function and promote tumor growth (36). Secretion of MMP9 (matrix metalloproteinase 9) from infiltrating neutrophils activates latent TGF-B and induces T cell suppression and tumor growth in a colorectal cancer model (37). SiglecF^{high} neutrophils in lung adenocarcinoma created an immunosuppressive environment by promoting macrophage differentiation, causing the release of high levels of ROS and enabling tumor progression (38). Together, these findings indicate that neutrophils that infiltrate diverse primary tumors can modify the local environment in different ways to favor tumor growth.

NEUTROPHIL FUNCTIONS THAT PROMOTE METASTASIS

The ability of cancer cells to leave the primary tumor and disseminate to distant organs represents the deadliest aspect of cancer progression. Indeed, the emergence of metastatic cancer accounts for \sim 90% of cancer related deaths (39). The metastatic cascade represents a series of barriers to cancer cells and neutrophils have been found to assist cancer cells in successfully navigating several of these distinct steps (**Figure 1**; **Supplementary Table 1**).

Local Invasion/Intravasation

Infiltrating neutrophils within primary tumors are associated with an increase in EMT, enhanced metastasis and poor outcomes. Mechanistically, tissue inhibitor of matrix metalloprotease (TIMP-1) secreted by neutrophils induced an EMT and consequently increased the migration and invasion of tumor cells. Cancer cells that had undergone an EMT expressed CD90, which enhanced TIMP-1 secretion by neutrophils in a contact-dependent manner (40).

Survival in Circulation/Extravasation

The ability of circulating tumor cells (CTCs) to survive is critical for metastasis formation (41). The formation of heterotypic cancer cell—neutrophil clusters was found to greatly increase metastatic fitness. Using a 4T1 breast cancer model, it was demonstrated that CTC-neutrophil interactions relied on VCAM-1 dependent adhesion, which enhanced cancer cell proliferation and increased metastasis (42). Indirectly, neutrophils can also inhibit NK cell-mediated tumor clearance in circulation; thereby increasing the intraluminal survival of disseminated tumor cells. In this study, 4T1 breast cancer cells were injected subcutaneously to mobilize murine neutrophils (Ly6G⁺), following which D2A1 breast cancer cells were injected intravenously. Mice bearing 4T1 cells exhibited reduced clearance of D2A1 cells from the lungs when compared to mice that were not injected with 4T1 cells (43). Depletion of NK cells resulted in enhanced D2A1 cancer cell accumulation in the lungs while neutrophil depletion had the opposite effect (43).

Cancer cells that have survived in circulation must exit the bloodstream and extravasate into tissue parenchyma (41). Neutrophils have been shown to regulate the extravasation process through several mechanisms. Neutrophil-derived factors can diminish the integrity of the endothelial barrier, permitting cancer cells to extravasate more easily. IL-8, IL-1 β , and matrix metalloproteases (MMP8 and MMP9) released from neutrophils activated endothelial cells, reduced endothelial barrier function, increased transendothelial migration and accelerated the rate of cancer cell extravasation (43, 44).

NETosis, and NET constituents, can support cancer cell extravasation through enhanced trapping of CTCs within metastatic sites (45–48). Importantly, blocking NETosis decreases cancer cell adhesion and inhibits metastatic spread to the lung and liver (49, 50). Furthermore, changes within specific metastatic microenvironments, such as exposure to ozone or redox imbalance, triggered NETosis and led to increased entrapment of cancer cells in the lung and enhanced metastasis (51, 52). Collectively, these studies show that neutrophils play an important role in enhancing tumor cell survival and increased extravasation, which promote cancer metastasis.

Early Seeding/Survival

Systemic and tumor-derived factors have been implicated in neutrophil recruitment in the pre-metastatic niche. Tumor-derived IL-1 β induces $\gamma\delta$ T cells to produce IL-17A and granulocyte-colony stimulating factor (G-CSF), which results in the recruitment of immunosuppressive neutrophils to the lung (8). GM-CSF and IL-5 have been shown to promote the expansion and recruitment of pro-metastatic neutrophils in the lungs of obese mice, which promotes lung metastasis (53). Angiopoetin-like-2 (ANGPTL2), secreted by osteosarcoma cells implanted in the tibia, stimulates lung epithelial cells, which led to the accumulation of neutrophils in the lung, and enhanced lung metastatic burden (54). In the lung, neutrophils secrete LTB₄ that increases the proliferation of LTB4R-positive metastasis initiating cells (55). Activation of NOTCH1 in colorectal cancer cells drives TGF_{β2}-dependent recruitment of immunosuppressive neutrophils within the liver, which enabled the formation of liver metastases (56).

NETs also support early cancer cell seeding and colonization of metastases. Induction of NETs by ovarian tumor-derived factors has been shown to be important in promoting metastasis to the omentum (57). In the liver, NETs have also been shown to promote metastasis by activating cancer-associated fibroblasts (58).

Growth in the Metastatic Site

Neutrophils have been shown to promote the growth of metastases after seeding. Minor subclones of breast cancer cells that secrete IL-11 and FIGF (C-fos-induced growth factor) can support the formation of polyclonal metastases composed of driver and passenger sub-populations. These IL-11 producing sub-clones activated IL-11 responsive mesenchymal stromal cells, which induced chemokine secretion and subsequent recruitment of pro-metastatic neutrophils (59). Tumor cell-derived GM-CSF was shown to stimulate neutrophils to synthesize and secrete transferrin, an iron transport protein, which has mitogenic activity that promotes lung metastatic growth when taken up by cancer cells (60).

A recurring function of pro-metastatic neutrophils is their ability to create an immunosuppressive microenvironment that support metastasis. Within lung metastases, inducible nitric oxide synthase (iNOS) producing neutrophils have been shown to limit CD8⁺ T cell dependent anti-tumor responses by promoting immune suppression (8). Recently, p53deficient cancer cells were found to increase the expression of Wnt ligands, which in turn upregulated IL-1ß production from tumor-associated macrophages (61) High IL-1β levels engaged $\gamma \delta 17 T$ cells, which subsequently enhanced neutrophil recruitment that promoted the formation of lung metastases (61). Furthermore, loss of Elf5 (E74-like transcription factor) expression in triple-negative breast cancer led to increased IFN- γ signaling resulting in the expansion of immunosuppressive neutrophils (62). In addition to tumor-derived factors, a lack of systemic testosterone levels can lead to an impairment of anti-tumor neutrophil functions. A shift toward immature neutrophils was observed in castrated male mice, leading to increased neutrophil-derived ROS and suppression of NK cell activation that promoted increased lung metastatic burden in two melanoma models (63). Recently, a role for NET formation has been described for the continued growth of established metastases (64). NETs released during cancer progression was shown to limit the ability of NK and cytotoxic T cells to eliminate cancer cells. Specifically, NET formation impaired direct contact between the cancer cells and cytotoxic immune cells (NK and T cells). Inhibition of NETosis with a protein arginine deiminase 4 (PAD4) inhibitor synergized with immune checkpoint inhibitors to control tumor growth and metastasis (64).

Pro-angiogenic functions have long been ascribed for neutrophils, which revealed that neutrophil-derived proteases (such as MMP9) could release stored angiogenic factors (VEGF, FGFs) that were stored in the local environment to enable blood vessel formation (65, 66). Recently, a different mechanism by which neutrophils enhance angiogenesis has been described. The synthesis and secretion of fibroblast growth factor 2 (FGF2) by neutrophils in the liver microenvironment drives angiogenesis and growth of nascent colorectal cancer-derived hepatic metastases (67).

DORMANT/RESIDUAL DISEASE AND THERAPY RESISTANCE

Neutrophils have also been implicated in awakening dormant cancer cells. LPS-induced tissue inflammation led to metastatic outgrowth of dormant tumor cells in a neutrophil-dependent manner (68). MMP-9 produced by neutrophils can trigger the growth of dormant cancer cells by remodeling extracellular matrix and releasing potent angiogenic factors (69). NE and MMP-9, which are enzymes associated with NETs, can cleave the extracellular matrix (ECM) leading to integrin-mediated signaling, which awakens dormant cancer cells and promotes cancer cell growth (70).

Several studies have shown that neutrophils promote resistance to therapy. Doxorubicin and paclitaxel resistant breast cancer cells express more IL-17 and CXCR2 ligands, which increases neutrophil recruitment (71). A neutrophil-enriched subtype characterized in triple negative breast cancer (TNBC) determined that neutrophils were largely immunosuppressive, rendering these tumors resistant to immune checkpoint blockade therapy (72). In a genetically engineered mouse model of sarcoma, neutrophils promote resistance to radiation therapy by activating mitogen-activated protein kinase (MAPK) pathway (73). In addition, CD177⁺ neutrophil infiltrates in colorectal cancer patients are associated with adverse outcome in patients receiving bevacizumab [anti-vascular endothelial growth factor A (VEGF- A)] (74). Furthermore, Lysyl oxidase-like 4 (LOXL4) expressing neutrophils that infiltrated colorectal cancer liver metastases were found to identify patients that were resistant to anti-angiogenic therapy (75).

METABOLIC PROGRAMMING IN NEUTROPHILS

There has been recent interest in the concept of immunometabolism and the realization that altered cellular metabolism in infiltrating immune cells can have a significant impact on tumor growth and metastasis (76). Neutrophils are typically viewed as a cell type that is heavily reliant on glycolysis to perform their effector functions (77). Consistent with this notion, neutrophils have very few mitochondria and inhibitors of oxidative phosphorylation (OXPHOS) do not alter their rates of oxygen consumption (77, 78). However, during tumor progression, neutrophils have been shown to undergo a metabolic switch, which involves the upregulation of genes associated with OXPHOS, fatty acid metabolism, and glycolysis (Figure 2) (38). Neutrophils isolated from Lewis lung carcinoma exhibit increased flux through OXPHOS, glycolysis, and increased ATP production compared to naïve neutrophils, suggesting that multiple metabolic strategies are engaged in tumor infiltrating neutrophils (79). Recently, upregulation of FATP2 (fatty acid transport protein 2) in neutrophils was shown to increase lipid accumulation in these cells. FATP2 regulated the uptake of arachidonic acid, which was subsequently converted to prostaglandin E2. Neutrophil-derived prostaglandin E2 was



found to be important or neutrophil-mediated $CD8^+$ T cell suppression and tumor growth (80).

Metabolic flexibility refers to the ability of a cell to shift between one metabolic program to another in response to changing metabolic demands or nutrient supply. High metabolic flexibility increases the cell's ability to survive various and everchanging metabolic microenvironments (81). Neutrophil sub-populations can also exhibit metabolic flexibility (**Figure 2**). In breast cancer, splenic neutrophils can engage mitochondrialdependent fatty acid oxidation as a predominate fuel source to support ROS production and maintain T cell suppression (82). Under glucose-limiting conditions, similar to certain tumor microenvironments, immature LDNs have been shown to utilize OXPHOS to generate ATP that is required to support their pro-tumorigenic functions. Indeed, immature LDNs can support NETosis under nutrient limiting conditions via mitochondrialdependent amino acid catabolism, which is important for efficient breast cancer liver metastasis (9). In addition, the longevity of neutrophils could also be altered due to the enhanced metabolic flexibility. The *ex vivo* half-life of mouse circulating HDNs and LDNs was 4 and 12 h, respectively (7, 9). Such observations raise the intriguing possibility that, under certain conditions, distinct neutrophil subsets may not be as short-lived as previously thought. These studies argue that increased metabolic flexibility in distinct neutrophil populations may be important for cellular functions that can influence tumor growth and metastatic progression.

CLINICAL IMPORTANCE: FUTURE PERSPECTIVES ON TREATMENT

In keeping with their pro-tumorigenic/metastatic functions, the presence of neutrophils across 25 different cancers was shown to be strongly associated with adverse patient outcomes (83).

Among certain subtypes of breast cancer (ER-), the presence of a neutrophil infiltrate in the primary tumor is also indicative of worse patient outcomes (84). Furthermore, in patients with advanced cancers, serum IL-8 levels, and neutrophil infiltration are associated with worse overall survival and diminished response to immune checkpoint inhibitors (85).

The mobilization of neutrophils into circulation also has prognostic significance. The neutrophil-to-lymphocyte ratio (NLR) is an important risk stratification and treatment selection diagnostic tool for cancer patients. A high NLR is associated with poor prognosis in many solid human cancers (86–96). A high NLR is also associated with decreased overall survival in patients with TNBC or metastatic breast cancer (97, 98).

An important and unanswered question with respect to the NLR is the type of neutrophil that is being detected in these patients, are they high- or low-density neutrophils? Interestingly, LDNs have been identified in patients with breast cancer, lung cancer, head and neck cancers, urologic cancers, and lymphoma (7, 99-101). In patients with advanced lung cancer, it was reported that higher proportion of LDNs (>10%) predicted poorer survival (102). These observations are in keeping with the pro-tumorigenic and pro-metastatic functions associated with LDN/N2 neutrophils. While most studies reveal a negative prognostic impact of neutrophils in cancer, there was one study that associated the presence of a CD16^{high} CD62^{dim} neutrophil subset with increased survival of head and neck squamous cell carcinoma patients (103). These observations highlight the need for better markers that are capable of discriminating between neutrophils that exert anti-tumor vs. those that mediate protumor/metastatic effects.

Mechanistic insights have greatly advanced our knowledge of tumor-derived factors that impact tumor growth and metastasis in a neutrophil-dependent manner. Additional studies

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focused on characterizing the phenotypic and functional role of neutrophils in cancer, it may be possible to develop strategies that specifically target those neutrophil subsets that actively promote tumor growth and metastasis, while sparing those neutrophils that possess anti-tumor and anti-microbial functions. Finally, the emerging concept of metabolic flexibility that is exhibited by certain neutrophil subsets may afford new ways of targeting these pro-tumorigenic/metastatic neutrophils.

AUTHOR CONTRIBUTIONS

BH, YS, and PS wrote the review and prepared the figures. All authors contributed to the article and approved the submitted version.

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SUPPLEMENTARY MATERIAL

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Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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