



Impact of *N*-Acyl-Homoserine Lactones, Quorum Sensing Molecules, on Gut Immunity

Garance Coquant¹, Jean-Pierre Grill¹ and Philippe Seksik^{1,2*}

¹ Sorbonne Université, Inserm, Centre de Recherche Saint-Antoine, CRSA, Hôpital Saint Antoine, Paris, France, ² Department of Gastroenterology, Saint Antoine Hospital, Assistance Publique-Hôpitaux de Paris (APHP), Paris, France

Among numerous molecules found in the gut ecosystem, guorum sensing (QS) molecules represent an overlooked part that warrants highlighting. QS relies on the release of small molecules (auto-inducers) by bacteria that accumulate in the environment depending on bacterial cell density. These molecules not only are sensed by the microbial community but also interact with host cells and contribute to gut homeostasis. It therefore appears entirely appropriate to highlight the role of these molecules on the immune system in dysbiosis-associated inflammatory conditions where the bacterial populations are imbalanced. Here, we intent to focus on one of the most studied QS molecule family, namely, the type I auto-inducers represented by N-acyl-homoserine lactones (AHL). First described in pathogens such as Pseudomonas aeruginosa, these molecules have also been found in commensals and have been recently described within the complex microbial communities of the mammalian intestinal tract. In this mini-review, we will expound on this emergent field of research. We will first recall evidence on AHL structure, synthesis, receptors, and functions regarding interbacterial communication. Then, we will discuss their interactions with the host and particularly with agents of the innate and adaptive gut mucosa immunity. This will reveal how this new set of molecules, driven by microbial imbalance, can interact with inflammation pathways and could be a potential target in inflammatory bowel disease (IBD). The discovery of the general impact of these compounds on the detection of the bacterial quorum and on the dynamic and immune responses of eukaryotic cells opens up a new field of pathophysiology.

Keywords: quorum sensing, gut microbiota, interkingdom communication, inflammatory bowel disease, gut inflammation

INTRODUCTION

Dysbiosis in inflammatory bowel disease (IBD) is characterized by a reduction in bacterial biodiversity. This change in biodiversity is associated with a reduction in the bacterial load of various bacterial groups and expansion of others (1). Bacteria are capable of exchanging small signaling molecules depending on the bacterial density, a process called quorum sensing (QS). QS coordinates gene expression and physiology of bacterial populations. There are several families of molecule used by microbes to communicate. A universal system relying on type II auto-inducers (AI-2) can be used by all bacteria (2). Gram-positive bacteria use oligopeptides, although Gram-negative QS display various molecules (3). Among them, the most studied system is represented by the type I auto-inducers based on *N*-Acyl-homoserines lactones (AHL). Beyond

OPEN ACCESS

Edited by:

Gislane Lelis Vilela de Oliveira, São Paulo State University, Brazil

Reviewed by:

Elena Vikström, Linköping University, Sweden Anton Hartmann, Ludwig Maximilian University of Munich, Germany

> *Correspondence: Philippe Seksik philippe.seksik@aphp.fr

Specialty section:

This article was submitted to Mucosal Immunity, a section of the journal Frontiers in Immunology

Received: 14 May 2020 **Accepted:** 08 July 2020 **Published:** 28 August 2020

Citation:

Coquant G, Grill J-P and Seksik P (2020) Impact of N-Acyl-Homoserine Lactones, Quorum Sensing Molecules, on Gut Immunity. Front. Immunol. 11:1827. doi: 10.3389/fimmu.2020.01827

1

bacteria-bacteria communication, QS molecules are also involved in interkingdom interplay between gut bacteria and host cells. In the early 2000s, researchers investigated the impact of AHL on plant physiology (4) showing that the presence of these molecules produced by soil bacteria can induce plant benefits (5) such as induction of the immune system to resist pathogens (6, 7). More recently, underlying potential on health and disease, AHL have been described in mammals' gut lumen and lately in humans. It is likely that IBD dysbiosis is a condition that cannot only modify QS but also be maintained by QS. Therefore, a better knowledge of this largely overlooked metabolite component appears important to improve our understanding of the molecular mechanisms implicated in the immune gut responses in IBD.

N-Acyl-Homoserine Lactones

Quorum Sensing Molecules for Bacteria

Bacteria are not single cells living independently. They have "social" interactions called QS. QS was first studied in the 1970s through the bioluminescence emitted by a marine bacterium, *Vibrio fischeri*. These bacteria live in symbiosis with marine animals. *V. fischeri* does not emit light at low concentrations in the water; however, when the bacterial population is growing in symbiosis with a squid, within a rich environment, the bacteria emits light (8). This phenomenon is possible, thanks to QS. However, the term was used for the first time two decades later, by Fuqua et al. (9).

This system relies on the release, by bacteria, of small molecules (auto-inducers) that accumulate in the environment depending on bacterial density and are sensed by the bacteria community. Once a threshold concentration is reached, this triggers the expression of certain bacterial genes, encoding for virulence factors, biofilm formation, etc. (Figure 1). Gramnegative bacteria use AHL as one of the QS molecules (10). AHL is a family of molecules composed by a unique lactone ring, an acyl chain of various lengths, and substitutions at the acyl C3 position that define the molecular species. The integrity of the lactone ring is important for AHL immunoactivity (11). Quorum quenching is the mechanisms by which QS signal is degraded, and it can occur enzymatically, through acylases (cleavage of the AHL amid bond, releasing a fatty acid and homoserine lactone) and lactonases (hydrolysis of the HSL ring) (12).

AHL Genes Induction

AHL diffuse freely in bacteria and recognize the receptor LuxR, allowing them to act as a transcription factor. Once activated, QS induces the expression of a series of bacterial genes contributing to bacterial virulence and/or adaptation such





as toxins, motility, enzymes, secretion systems, iron uptake, metabolism, and biofilms (**Figure 1**) (3). AHL also regulate their own synthase and receptor genes, in a positive retro-control loop. This system has been mostly described in pathogens, while the impact of these factors on gut immunity has not been directly investigated. However, few reports showed that AHL found in gut ecosystem could benefit host physiology (13). Thus, there is still a gap in research on AHL derived from commensals and their impact on host.

Most studies have focused on the impact of auto-inducers from the opportunistic bacterium Pseudomonas aeruginosa, N-(3-oxododecanoyl)-L-homoserine lactone (3-oxo-C12-HSL) and N-butyryl-homoserine lactone (C4-HSL). Signaling works through the LuxI/LuxR homolog system. LuxI synthesizes the AHL, the molecule freely diffuses in the environment, and LuxR detects the auto-inducer, which acts as a transcription factor (14) (Figure 1). Bacteria can use different ways to communicate: (i) crosstalk, different species talking to each other; (ii) self-talk, bacteria from the same species talk with its own auto-inducer and regulate their gene expression; and (iii) eavesdropping, when bacteria from one species can intercept the signal of another species without creating a signal by itself (15). Indeed, some bacteria express a LuxR homolog without its partner LuxI. LuxR is then labeled LuxR solo, and the receptor is called SdiA (16). For instance, Escherichia coli has the SdiA receptor without being

Abbreviations: AHL, *N*-acyl-homoserine lactones; 3-oxo-C12-HSL, *N*-(3-oxododecanoyl)-L-homoserine lactone; AI, auto-inducer; C4-HSL, *N*-butyryl-homoserine lactone; DCs, dendritic cells; IBD, inflammatory bowel disease; IFN γ : interferon- γ ; IL-10, interleukine 10; IQGAP1, IQ motif containing GTPase-activating protein 1; MAPK, mitogen-activated protein kinase; PON, paraoxonase; PPAR, peroxisome proliferator-activated receptor; PRR, pattern recognition receptor; QS, quorum sensing; TGF β , transforming growth factor- β ; TLR-2, Toll-like receptor-2; TNF α , tumor necrosis factor- α ; UPR, unfolded protein response.

able to produce an auto-inducer (17). This ability to "listen" to other bacterial species is essential in complex ecosystem like the gut microbiota, where hundreds of different bacteria coexist. However, these ways of communicating bypass the bacterial world, and those small molecules are part of an interkingdom signaling, allowing cross-talk between gut microorganisms and the host.

AHL Interkingdom Signaling

Since eukaryotic and prokaryotic cells have coevolved for thousands of years, the signals from one have adapted to the other's. In this setting, bacterial QS molecules have been shown to have an effect on eukaryotic cells, a phenomenon called "interkingdom signaling" (18). To date, most studies on the effects of AHL on mammal cells have focused not only on the well-known 3-oxo-C12-HSL and C4-HSL from P. aeruginosa but also on N-(3-oxohexanoyl)-L-homoserine lactone (3-oxo-C6-HSL) from V. fischeri. AHL are amphiphilic molecules, as lactone ring remains hydrophilic and the acyl chain is hydrophobic. It has been shown that these properties allow the 3-oxo-C12-HSL to diffuse freely through the cell membrane of eukaryotic cells, similar to bacteria, while longchained versions are mostly actively transported (19). AHL are chemically analogous to many lipid-based hormones such as the eicosanoid family of lipidic and steroid hormones involved in hundreds of biological functions in eukaryote. It is also thought that AHL can enter the host cell, bind to intracellular receptors, and regulate gene transcription (18). Using labeled molecules, it has been shown that, depending on cell lines, they can indeed enter cells and be detected in the nucleus or the cvtoplasm (19, 20).

The AHL perception mechanism in mammalian cells is still fairly unclear. To date, no mammalian receptor has been clearly demonstrated; only certain hypotheses have been proposed. It has been shown that the AHL were not recognized by the classical pattern recognition receptors (PRRs) from the innate immune system like other microbial molecules (21). The studies done on the 3-oxo-C12-HSL produced by P. aeruginosa have identified different receptors based on the cell types and methods used. Proinflammatory effects have been associated with receptors such as nuclear factor kappa B (NF-κB) or activator protein-2 (AP-2) (21). Using other cellular models, mitogen-activated protein kinases (MAPKs) have been proposed as potential receptors (22). In 2008, Jahoor et al. identified the peroxisome proliferatoractivated (PPAR) receptors PPARß/8 and PPARy as potential AHL receptors (23). The interaction between 3-oxo-C12-HSL and PPAR γ has also been reported by another group, at very low concentration (1 nM) of AHL (24).

Other candidate mammalian AHL receptors are the G-protein-coupled receptors and, among these, member 38 of the bitter taste receptor family (T2R38), one of the most studied bitter taste receptors. It is widely expressed in the human digestive tract from the tongue to the colon (25). In the lower gastrointestinal tract, T2R38 is suspected to play a role in eliciting immune responses to toxic compounds or pathogens in digestive diseases and metabolic conditions. Indeed, *TAS2R38* polymorphisms have been linked to increased

susceptibility to infections and colorectal cancer (26–28). Moreover, reports shows that both 3-oxo-C12-HSL and C4-HSL can activate T2R38 in pulmonary epithelium (28). The interaction between AHL and this receptor was described in neutrophils, by immunofluorescence and pull-down assays (29, 30).

Another potential AHL receptor is the IQ-motif-containing GTPase-activating protein (IQGAP1) (31). IQGAP1 is a scaffolding protein, participating in cytoskeleton organization (32). It has been shown that IQGAP1 plays a role in tight junction assembly (32), and as 3-oxo-C12-HSL is known to disrupt junction integrity (10), the interaction between IQGAP1 and AHL seems rational.

Finally, according to the cell type, the receptors and their localization can be different (33, 34) but mainly involves inflammatory pathways. Host cells have developed the abilities to disrupt QS signaling, by degrading them through the production of paraoxonases (PON), which can be expressed by intestinal epithelial cells and macrophages (35). Regulation of this communication between host cells and bacteria can occur in gut ecosystem. Moreover, it has been shown that PON1 polymorphisms may confer protection against the development of IBD (36). Therefore, AHL interkingdom signaling can be viewed as potent axis of the gut microbiota–host crosstalk in chronic inflammation and especially in IBD.

Impact of AHL on Innate Immunity

It has been shown, on cell lines and primary cell cultures, that AHL, depending on their acyl chain length, double bounds, and concentrations, may differently affect innate immune system.

Epithelial cells provide a physical barrier between the host and the lumen of the intestine, relying on tight junctions. This prevents the luminal content to harm the host's integrity. Epithelial cells contribute to the innate immune response, as they keep foreign particles from spreading into the host mucosa. Vikström et al. showed, on an intestinal epithelial cell line Caco-2, that P. aeruginosa 3-oxo-C12-HSL alters intestinal barrier, disrupting protein junctions integrity (37-40). When exposed to the AHL, permeability to macromolecules and ions is increased, and the expression and localization of junction proteins such as occludin, E-cadherin, and zonula occludens-1 is modified (37-39). This alteration process not only involves the MAPK, notably p38 and p42/44, but also needs the phosphorylation of junction proteins leading to disturbance of junction integrity (37-39). Alteration of calcium signaling is also part of the response to 3-oxo-C12-HSL (39, 41). Interestingly, C4-HSL does not disturb barrier integrity like 3-oxo-C12-HSL (40). Moreover, an AHL identified recently in the gut, 3-oxo-C12:2-HSL, exerts anti-inflammatory effects on Caco-2/TC7 cell line stimulated by interleukine-1ß (IL-1ß), as shown on IL-8-reduced secretion (13).

Some groups identified AHL as chemoattractant to neutrophils, as it is the case for 3-oxo-C12-HSL and 3oxo-C10-HSL, but not C4-HSL. AHL attract neutrophils in a dose-dependent fashion, through actin remodeling and calcium mobilization (42, 43). Moreover, 3-oxo-C12-HSL exerts proapoptotic function on neutrophils, by targeting mitochondria and their calcium balance (43).

Macrophages are one of the most studied cell type regarding 3-oxo-C12-HSL impacts. Overall, the observed effects aim at decreasing the inflammatory response, allowing the set-up of a chronic P. aeruginosa infection. The effects are various, from cell-volume increase through water flux (44), unfold protein response (UPR) (45) to apoptosis (46) and also on immune functions. Indeed, P. aeruginosa 3-oxo-C12-HSL exerts anti-inflammatory responses in macrophages, as was reported by Glucksam-Galnoy et al. on RAW264.7 murine macrophages, in a dose-dependent fashion. Notably, a decrease in tumor necrosis factor-a production and increase in IL-10 secretion was observed (47). As mentioned earlier, when macrophages are in a proinflammatory context, 3-oxo-C12-HSL can modulate the NF-KB pathway resulting in a decrease in the expression of proinflammatory cytokines such as tumor necrosis factor-α (TNFα), regulated upon activation, normal T cell expressed, and secreted (RANTES), or monocyte chemoattractant protein-1 (MCP-1) (48). The involvement of MAPK p38 has also been reported several times (21, 47, 49) but needs further studies to determine its role in the signaling. In addition, it has been shown that macrophages in the presence of 3-oxo-C12-HSL have a higher phagocytic activity (49, 50), and those effects are abolished when MAPK p38 is inhibited (49).

Dendritic cells (DCs) stimulated by lipopolysaccharides and in the presence of 3-oxo-C12-HSL show a decrease in their proinflammatory cytokine such as IL-12 and interferon- γ (IFN γ) (51–53). However, reports on anti-inflammatory IL-10 are contradictory. In both studies, the cells were stimulated by lipopolysaccharides and exposed to the same dose of 3-oxo-C12-HSL; one team reported an increase in IL-10 secretion by human DCs (53), while another group did not see any change in IL-10 production by mouse DCs (52). By modulating DCs activation through its QS molecules, *P. aeruginosa* suppress the adaptive immune response, favoring the establishment of chronic infection (52, 53). 3-oxo-C12-HSL has also proapoptotic effects on humans DCs, as is the case for several cell types (54).

AHL have cell-type-specific impacts, but overall, the effects tend to be anti-inflammatory. Besides, 3-oxo-C12-HSL effects on apoptosis are dependent on the cell types, as multiple studies do not report toxicity on differentiated epithelial cells or fibroblasts (31, 55, 56). The effects of 3-oxo-C12-HSL on immune cells are compiled in the **Table S1**. These pathogen-related AHL also display multiple disruptions on several cell functions from innate immune cells.

Impact of AHL on Adaptive Immunity

Once again, most of the knowledge in that field relies on 3-oxo-C12-HSL from *P. aeruginosa*. It had been first shown that this AHL could inhibit T-cell proliferation (57). This was confirmed by later reports showing that the same AHL could inhibit the proliferation and function (cytokine production)

of both mitogen-stimulated (11, 58) and antigen-stimulated (59) T lymphocytes and modulate antibody production by B lymphocytes (57, 58). Overall, AHL from P. aeruginosa tend to have a less effective antibody-mediated, rather than a more effective cell-mediated, adaptive immune response to the bacteria, and could thus facilitate persistence of the pathogen. A structure-activity relationship study of 3-oxo-C12-HSL indicated that, like QS activity, immune modulatory activity requires an intact HSL ring, L-configuration at the chiral center, and an acyl chain of 11-13 carbons (11). Moreover, 3-oxo-C12-HSL can rapidly induce apoptosis via mitochondrial pathway on Jurkat cell line (60) and can inhibit DCs and T-cell activation and proliferation, and downregulate the expression of costimulatory molecules on DCs (54, 61, 62). This results in shifting immune responses away from host-protective Th1 responses to pathogen-protective Th2 responses (62). More precisely, 3-oxo-C12-HSL is able to promote the induction of regulatory T cells such as CD4+CD25+Foxp3+ induced regulatory T cells and to enhance their IL-10 and transforming growth factor- β (TGF β) production associated with reduced IFNy and IL-12p70 production (53). At the molecular level, 3-oxo-C12-HSL prevents human DCs maturation by blocking the upregulation of surface molecules, including CD11c, HLA-DR, CD40, and CD80, and DCs switched to an interleukin IL-10 (high), IL-12p70 (low) phenotype (53).

In addition, it has been reported that 3-oxo-C12-HSL increases the expression of Toll-like receptor-2 (TLR2), in a dosedependent fashion, in lymphocytes from peripheral blood (63). Those observations are interesting because it has been shown that this AHL signaling does not rely on TLR recognition (21).

Taking together, there are limited observations showing T-cell inhibition and Treg induction by one AHL from a pathogen. It is crucial to look at the effect of other natural AHL especially those coming from commensal gut bacteria.

AHL Within Intestinal Communities Clues on the Presence of AHL in the Gut

The existence of AHL in the gut has been subject to questioning. An article from 2013 entitled "Are There Acyl-Homoserine Lactones within Mammalian Intestines?" examine the question (64). Mammalian pathogens such as P. aeruginosa or Yersinia enterocolitica are well known for their ability to produce AHL, but what about commensal bacteria? As mentioned by the authors, previous studies relied on the use of LuxR-based biosensors, where the detection limit may be too high to detect AHL from the gut, and there is thus a need for the development of new more sensitive tools (64). Moreover, the Human Microbiome Project, aiming at sequencing and analyzing the microbiome of several cohorts, gives clues about the presence of AHL in the intestines. Indeed, the LuxI/LuxR homolog has been found in three strains from the gastrointestinal tract: Hafnia alvei, Edwardsiella tarda, and Ralstonia sp. strain 5_7_47FAA (64). In addition, bacteria like E. coli, Enterobacter, or Klebsiella are part of the normal gut microbiota and express, as mentioned above, the receptor SdiA and therefore can sense AHL (17). One must highlight the fragility of small molecules like AHL. Indeed, they are pH sensitive and can also be inactivated by the



presence of ont only host lactonases like PON but also bacterial enzymes. Those degradations can decrease the concentration of the molecules, making it harder for biosensors to reach the threshold detection (64). Moreover, it is also possible that auto-inducers are present in the mammalian gut, but with a yet unknown structure, like aryl-homoserine lactones, with an aromatic side chain instead of an acyl chain and thus not detectable by biosensors (64). It is also interesting to note that AHL have been found in the rumen of cattle by several teams (65, 66).

Our team investigated the question of AHL in the human gut, in the context of IBD. By using mass spectrometry, we were able to detect 14 different AHLs in the feces of IBD patients and healthy subject, and the distribution of the AHL were correlated to the disease state (13). Indeed, one of the AHL was prominent: 3-oxo-C12:2-HSL. This molecule was, until then, undescribed and carries two insaturations on its acyl chain. 3-Oxo-C12:2-HSL was highly decreased in fecal samples of IBD patients in flare (16%) compared to remission patients (37.5%) and to healthy subjects (64.5%) (13). The absence of this AHL was correlated with dysbiosis and a decrease in Firmicutes, considered as beneficial bacteria. In addition, 3-oxo-C12:2-HSL exerts anti-inflammatory properties on intestinal cell lines Caco-2/TC7 (13).

Toward the Use of AHL to Modulate Microbiota Composition and Gut Inflammation

The study of QS molecules is mainly done on single species bacteria or *in vitro* cells. As emphasized by Karina Bivar Xavier, QS is also an interspecies communication network, and more studies on the impact of QS on multi-communities ecosystems should be done (67). *In vivo* studies are crucial to understanding relationships in an environment as complex as the intestine. Kumari et al. have developed a whole-cell

sensing systems for the detection of AHL and have shown that these signaling molecules detected in saliva and stool may be potential non-invasive biomarkers of gastrointestinal inflammatory disease (68). To emphasize this point, given that AHL profile relies on bacterial dynamics, it could be considered as an indicator of dysbiosis, opening new perspectives in managing chronical diseases such as IBD. Understanding the interactions between bacteria in high concentrations and high diversity can help us decipher what species are most beneficial to mammalian gut. To note, another QS type of molecule, AI-2, has been used to modulate gut microbiota composition and dysbiosis (69). As illustrated in **Figure 2**, AHLs remain good candidates in the strategy to use natural molecules from QS to modulate microbiota composition and gut inflammation.

CONCLUDING REMARKS

Gut microbiota mutually interacts with coevolved host epithelial and immune cells in a beneficial reciprocal relationship. QS signaling of bacteria probably contributes substantially to establishing symbiotic interactions in some cross-kingdom interactive dynamics. In IBD, where, host-microbiota interactions drive inflammatory response, looking at quorum sensing changes and impact on immunity appears as a completely novel and original approach. The mechanisms, including the regulation of synthesis and degradation of these diffusible signaling molecules are still not completely understood. However, the discovery of the general importance of auto-inducer signaling molecules involved in quorum or efficiency sensing of bacteria and the dynamic and immune responses by eukaryotes toward them opens up a new field of pathophysiology.

AUTHOR CONTRIBUTIONS

GC and PS drew the outline of the manuscript. GC, J-PG, and PS wrote and proofread the manuscript. All authors contributed to the article and approved the submitted version.

FUNDING

We disclose Financial support from Fondation pour la Recherche Médical that ensures salary fees of GC as a PhD student.

REFERENCES

- Maloy KJ, Powrie F. Intestinal homeostasis and its breakdown in inflammatory bowel disease. *Nature*. (2011) 474:298–306. doi: 10.1038/nature10208
- Bassler BL, Wright M, Silverman MR. Multiple signalling systems controlling expression of luminescence in *Vibrio harveyi* : sequence and function of genes encoding a second sensory pathway. *Mol Microbiol.* (1994) 13:273–86. doi: 10.1111/j.1365-2958.1994.tb00422.x
- Abisado RG, Benomar S, Klaus JR, Dandekar AA, Chandler JR. Bacterial quorum sensing and microbial community interactions. *mBio*. (2018) 9:14. doi: 10.1128/mBio.02331-17
- Mathesius U, Mulders S, Gao M, Teplitski M, Caetano-Anolles G, Rolfe BG, et al. Extensive and specific responses of a eukaryote to bacterial quorum-sensing signals. *Proc Natl Acad Sci USA*. (2003) 100:1444–9. doi: 10.1073/pnas.262672599
- Schikora A, Schenk ST, Hartmann A. Beneficial effects of bacteriaplant communication based on quorum sensing molecules of the N -acyl homoserine lactone group. *Plant Mol Biol.* (2016) 90:605–12. doi: 10.1007/s11103-016-0457-8
- Liu F, Zhao Q, Jia Z, Song C, Huang Y, Ma H, et al. N-3-oxo-octanoylhomoserine lactone-mediated priming of resistance to *Pseudomonas syringae* requires the salicylic acid signaling pathway in *Arabidopsis thaliana*. *BMC Plant Biol.* (2020) 20:38. doi: 10.21203/rs.2.11827/v2
- Hu Z, Shao S, Zheng C, Sun Z, Shi J, Yu J, et al. Induction of systemic resistance in tomato against *Botrytis cinerea* by N-decanoylhomoserine lactone via jasmonic acid signaling. *Planta*. (2018) 247:1217–27. doi: 10.1007/s00425-018-2860-7
- Nealson KH, Platt T, Hastings JW. Cellular control of the synthesis and activity of the bacterial luminescent system1. *J Bacteriol*. (1970) 104:313–22. doi: 10.1128/JB.104.1.313-322.1970
- Fuqua WC, Winans SC, Greenberg EP. Quorum sensing in bacteria: the LuxR-LuxI family of cell density-responsive transcriptional regulators. *J Bacteriol.* (1994) 176:269–75. doi: 10.1128/JB.176.2.269-275.1994
- Holm A, Vikström E. Quorum sensing communication between bacteria and human cells: signals, targets, and functions. *Front Plant Sci.* (2014) 5:309. doi: 10.3389/fpls.2014.00309
- Chhabra SR, Harty C, Hooi DSW, Daykin M, Williams P, Telford G, et al. Synthetic analogues of the bacterial signal (quorum sensing) molecule N -(3-Oxododecanoyl)- L -homoserine lactone as immune modulators. *J Med Chem.* (2003) 46:97–104. doi: 10.1021/jm020909n
- Whiteley M, Diggle SP, Greenberg EP. Progress in and promise of bacterial quorum sensing research. *Nature*. (2017) 551:313–20. doi: 10.1038/nature24624
- Landman C, Grill J-P, Mallet J-M, Marteau P, Humbert L, Balc'h EL, et al. Inter-kingdom effect on epithelial cells of the N-Acyl homoserine lactone 3-oxo-C12:2, a major quorum-sensing molecule from gut microbiota. *PLoS ONE*. (2018) 13:e0202587. doi: 10.1371/journal.pone.0202587
- Parsek MR, Val DL, Hanzelka BL, Cronan JE, Greenberg EP. Acyl homoserinelactone quorum-sensing signal generation. *Proc Natl Acad Sci USA*. (1999) 96:4360–5. doi: 10.1073/pnas.96.8.4360
- Prescott RD, Decho AW. Flexibility and adaptability of quorum sensing in nature. *Trends Microbiol.* (2020) 28:436–44. doi: 10.1016/j.tim.2019.12.004

ACKNOWLEDGMENTS

We thank Ilias Theodorou, PhD, for his careful proofreading of the manuscript.

SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fimmu. 2020.01827/full#supplementary-material

- Hudaiberdiev S, Choudhary KS, Vera Alvarez R, Gelencsér Z, Ligeti B, Lamba D, et al. Census of solo LuxR genes in prokaryotic genomes. *Front Cell Infect Microbiol.* (2015) 5:20. doi: 10.3389/fcimb.2015.00020
- Tobias NJ, Brehm J, Kresovic D, Brameyer S, Bode HB, Heermann R. New vocabulary for bacterial communication. *ChemBioChem.* (2020) 21:759–68. doi: 10.1002/cbic.201900580
- Shiner EK, Rumbaugh KP, Williams SC. Interkingdom signaling: deciphering the language of acyl homoserine lactones. *FEMS Microbiol Rev.* (2005) 29:935– 47. doi: 10.1016/j.femsre.2005.03.001
- Ritchie AJ, Whittall C, Lazenby JJ, Chhabra SR, Pritchard DI, Cooley MA. The immunomodulatory *Pseudomonas aeruginosa* signalling molecule N -(3-oxododecanoyl)- L -homoserine lactone enters mammalian cells in an unregulated fashion. *Immunol Cell Biol.* (2007) 85:596–602. doi: 10.1038/sj.icb.7100090
- Williams P, Cámara M. Quorum sensing and environmental adaptation in *Pseudomonas aeruginosa*: a tale of regulatory networks and multifunctional signal molecules. *Curr Opin Microbiol.* (2009) 12:182–91. doi: 10.1016/j.mib.2009.01.005
- Kravchenko VV, Kaufmann GF, Mathison JC, Scott DA, Katz AZ, Wood MR, et al. N -(3-Oxo-acyl)homoserine lactones signal cell activation through a mechanism distinct from the canonical pathogen-associated molecular pattern recognition receptor pathways. J Biol Chem. (2006) 281:28822–30. doi: 10.1074/jbc.M606613200
- Li L, Hooi D, Chhabra SR, Pritchard D, Shaw PE. Bacterial Nacylhomoserine lactone-induced apoptosis in breast carcinoma cells correlated with down-modulation of STAT3. *Oncogene*. (2004) 23:4894–902. doi: 10.1038/sj.onc.1207612
- Jahoor A, Patel R, Bryan A, Do C, Krier J, Watters C, et al. Peroxisome proliferator-activated receptors mediate host cell proinflammatory responses to *Pseudomonas aeruginosa* autoinducer. *J Bacteriol.* (2008) 190:4408–15. doi: 10.1128/JB.01444-07
- Cooley MA, Whittall C, Rolph MS. Pseudomonas signal molecule 3-oxo-C12homoserine lactone interferes with binding of rosiglitazone to human PPARy. *Microbes Infect.* (2010) 12:231–7. doi: 10.1016/j.micinf.2009.12.009
- Behrens M, Meyerhof W. Gustatory and extragustatory functions of mammalian taste receptors. *Physiol Behav.* (2011) 105:4–13. doi: 10.1016/j.physbeh.2011.02.010
- Rozengurt E. Taste receptors in the gastrointestinal tract. I. Bitter taste receptors and alpha-gustducin in the mammalian gut. Am J Physiol Gastrointest Liver Physiol. (2006) 291:G171–7. doi: 10.1152/ajpgi.00073.2006
- Carrai M, Steinke V, Vodicka P, Pardini B, Rahner N, Holinski-Feder E, et al. Association between TAS2R38 gene polymorphisms and colorectal cancer risk: a case-control study in two independent populations of Caucasian origin. *PLoS ONE.* (2011) 6:e20464. doi: 10.1371/journal.pone.0020464
- Lee RJ, Xiong G, Kofonow JM, Chen B, Lysenko A, Jiang P, et al. T2R38 taste receptor polymorphisms underlie susceptibility to upper respiratory infection. *J Clin Invest.* (2012) 122:4145–59. doi: 10.1172/JCI64240
- Gaida MM, Mayer C, Dapunt U, Stegmaier S, Schirmacher P, Wabnitz GH, et al. Expression of the bitter receptor T2R38 in pancreatic cancer: localization in lipid droplets and activation by a bacteria-derived quorum-sensing molecule. Oncotarget. (2016) 7:12623–32. doi: 10.18632/oncotarget.7206
- 30. Maurer S, Wabnitz GH, Kahle NA, Stegmaier S, Prior B, Giese T, et al. Tasting *Pseudomonas aeruginosa* biofilms: human neutrophils express the

bitter receptor T2R38 as sensor for the quorum sensing molecule N-(3-oxododecanoyl)-l-homoserine lactone. *Front Immunol.* (2015) 6:369. doi: 10.3389/fimmu.2015.00369

- Karlsson T, Turkina MV, Yakymenko O, Magnusson K-E, Vikström E. The *Pseudomonas aeruginosa* N-acylhomoserine lactone quorum sensing molecules target IQGAP1 and modulate epithelial cell migration. *PLoS Pathog.* (2012) 8:e1002953. doi: 10.1371/journal.ppat.1002953
- Tanos BE, Yeaman C, Rodriguez-Boulan E. An emerging role for IQGAP1 in tight junction control. *Small GTPases.* (2018) 9:375–83. doi: 10.1080/21541248.2016.1244440
- Bedi B, Maurice NM, Ciavatta VT, Lynn KS, Yuan Z, Molina SA, et al. Peroxisome proliferator-activated receptor-γ agonists attenuate biofilm formation by *Pseudomonas aeruginosa*. *FASEB J.* (2017) 31:3608–21. doi: 10.1096/fj.201700075R
- Cohen LJ, Esterhazy D, Kim S-H, Lemetre C, Aguilar RR, Gordon EA, et al. Commensal bacteria make GPCR ligands that mimic human signalling molecules. *Nature*. (2017) 549:48–53. doi: 10.1038/nature23874
- Grandclément C, Tannières M, Moréra S, Dessaux Y, Faure D. Quorum quenching: role in nature and applied developments. *FEMS Microbiol Rev.* (2016) 40:86–116. doi: 10.1093/femsre/fuv038
- 36. Karban A, Hartman C, Eliakim R, Waterman M, Nesher S, Barnett-Griness O, et al. Paraoxonase (PON)1 192R allele carriage is associated with reduced risk of inflammatory bowel disease. *Dig Dis Sci.* (2007) 52:2707–15. doi: 10.1007/s10620-006-9700-5
- Vikström E, Tafazoli F, Magnusson K-E. Pseudomonas aeruginosa quorum sensing molecule N -(3 oxododecanoyl)- L -homoserine lactone disrupts epithelial barrier integrity of Caco-2 cells. *FEBS Lett.* (2006) 580:6921–8. doi: 10.1016/j.febslet.2006.11.057
- Vikström E, Bui L, Konradsson P, Magnusson K-E. The junctional integrity of epithelial cells is modulated by *Pseudomonas aeruginosa* quorum sensing molecule through phosphorylation-dependent mechanisms. *Exp Cell Res.* (2009) 315:313–26. doi: 10.1016/j.yexcr.2008.10.044
- Vikström E, Bui L, Konradsson P, Magnusson K-E. Role of calcium signalling and phosphorylations in disruption of the epithelial junctions by *Pseudomonas aeruginosa* quorum sensing molecule. *Eur J Cell Biol.* (2010) 89:584–97. doi: 10.1016/j.ejcb.2010.03.002
- Eum SY, Jaraki D, Bertrand L, Andras IE, Toborek M. Disruption of epithelial barrier by quorum-sensing N-3-(oxododecanoyl)-homoserine lactone is mediated by matrix metalloproteinases. *AJP Gastrointest Liver Physiol.* (2014) 306:G992–1001. doi: 10.1152/ajpgi.00016.2014
- 41. Schwarzer C, Wong S, Shi J, Matthes E, Illek B, Ianowski JP, et al. Pseudomonas aeruginosa homoserine lactone activates store-operated cAMP and cystic fibrosis transmembrane regulator-dependent Cl ⁻ secretion by human airway epithelia. J Biol Chem. (2010) 285:34850–63. doi: 10.1074/jbc.M110.167668
- Karlsson T, Musse F, Magnusson K-E, Vikstrom E. N-Acylhomoserine lactones are potent neutrophil chemoattractants that act via calcium mobilization and actin remodeling. *J Leukoc Biol.* (2012) 91:15–26. doi: 10.1189/jlb.0111034
- Singh PK, Yadav VK, Kalia M, Sharma D, Pandey D, Agarwal V. *Pseudomonas aeruginosa* quorum-sensing molecule N-(3-oxo-dodecanoyl)-l-homoserine lactone triggers mitochondrial dysfunction and apoptosis in neutrophils through calcium signaling. *Med Microbiol Immunol.* (2019) 208:855–68. doi: 10.1007/s00430-019-00631-8
- Holm A, Magnusson K-E, Vikström E. Pseudomonas aeruginosa N-3-oxododecanoyl-homoserine lactone elicits changes in cell volume, morphology, and AQP9 characteristics in macrophages. Front Cell Infect Microbiol. (2016) 6:32. doi: 10.3389/fcimb.2016.00032
- Zhang J, Gong F, Li L, Zhao M, Song J. *Pseudomonas aeruginosa* quorumsensing molecule N-(3-oxododecanoyl) homoserine lactone attenuates lipopolysaccharide-induced inflammation by activating the unfolded protein response. *Biomed Rep.* (2014) 2:233–8. doi: 10.3892/br.2014.225
- Tateda K, Ishii Y, Horikawa M, Matsumoto T, Miyairi S, Pechere JC, et al. The *Pseudomonas aeruginosa* autoinducer N-3-oxododecanoyl homoserine lactone accelerates apoptosis in macrophages and neutrophils. *Infect Immun.* (2003) 71:5785–93. doi: 10.1128/IAI.71.10.5785-5793.2003
- Glucksam-Galnoy Y, Sananes R, Silberstein N, Krief P, Kravchenko VV, Meijler MM, et al. The bacterial quorum-sensing signal molecule N-3-oxo-dodecanoyl-L-homoserine lactone reciprocally modulates pro- and

anti-inflammatory cytokines in activated macrophages. J Immunol. (2013) 191:337-44. doi: 10.4049/jimmunol.1300368

- Kravchenko VV, Kaufmann GF, Mathison JC, Scott DA, Katz AZ, Grauer DC, et al. Modulation of gene expression via disruption of NF-B signaling by a bacterial small molecule. *Science.* (2008) 321:259–63. doi: 10.1126/science.1156499
- Vikström E, Magnusson K-E, Pivoriunas A. The *Pseudomonas aeruginosa* quorum-sensing molecule N-(3-oxododecanoyl)-l-homoserine lactone stimulates phagocytic activity in human macrophages through the p38 MAPK pathway. *Microbes Infect.* (2005) 7:1512–8. doi: 10.1016/j.micinf.2005.05.012
- Holm A, Karlsson T, Vikström E. Pseudomonas aeruginosa lasI/rhll quorum sensing genes promote phagocytosis and aquaporin 9 redistribution to the leading and trailing regions in macrophages. Front Microbiol. (2015) 6:915. doi: 10.3389/fmicb.2015.00915
- Guo J, Yoshida K, Ikegame M, Okamura H. Quorum sensing molecule N-(3-oxododecanoyl)-l-homoserine lactone: an all-rounder in mammalian cell modification. J Oral Biosci. (2020) 62:16–29. doi: 10.1016/j.job.2020.01.001
- Skindersoe ME, Zeuthen LH, Brix S, Fink LN, Lazenby J, Whittall C, et al. *Pseudomonas aeruginosa* quorum-sensing signal molecules interfere with dendritic cell-induced T-cell proliferation. *FEMS Immunol Med Microbiol.* (2009) 55:335–45. doi: 10.1111/j.1574-695X.2008.00533.x
- Li Y, Zhou H, Zhang Y, Chen C, Huang B, Qu P, et al. N-3-(oxododecanoyl)-L-homoserine lactone promotes the induction of regulatory T-cells by preventing human dendritic cell maturation. *Exp Biol Med.* (2015) 240:896– 903. doi: 10.1177/1535370214564742
- Boontham P, Robins A, Chandran P, Pritchard D, Cámara M, Williams P, et al. Significant immunomodulatory effects of *Pseudomonas aeruginosa* quorumsensing signal molecules: possible link in human sepsis. *Clin Sci.* (2008) 115:343–51. doi: 10.1042/CS20080018
- Josephson H, Ntzouni M, Skoglund C, Linder S, Turkina MV, Vikström E. *Pseudomonas aeruginosa* N-3-oxo-dodecanoyl-homoserine lactone impacts mitochondrial networks morphology, energetics, and proteome in host cells. *Front Microbiol.* (2020) 11:1069. doi: 10.3389/fmicb.2020.01069
- Losa D, Köhler T, Bacchetta M, Saab JB, Frieden M, van Delden C, et al. Airway epithelial cell integrity protects from cytotoxicity of *Pseudomonas* aeruginosa quorum-sensing signals. Am J Respir Cell Mol Biol. (2015) 53:265– 75. doi: 10.1165/rcmb.2014-0405OC
- Telford G, Wheeler D, Williams P, Tomkins PT, Appleby P, Sewell H, et al. The *Pseudomonas aeruginosa* quorum-sensing signal molecule N-(3oxododecanoyl)-l-homoserine lactone has immunomodulatory activity. *Infect Immun.* (1998) 66:36–42. doi: 10.1128/IAI.66.1.36-42.1998
- Ritchie AJ, Yam AOW, Tanabe KM, Rice SA, Cooley MA. Modification of *in vivo* and *in vitro* T- and B-cell-mediated immune responses by the *Pseudomonas aeruginosa* quorum-sensing molecule N-(3oxododecanoyl)-l-homoserine lactone. *Infect Immun.* (2003) 71:4421–31. doi: 10.1128/IAI.71.8.4421-4431.2003
- Ritchie AJ, Jansson A, Stallberg J, Nilsson P, Lysaght P, Cooley MA. The *Pseudomonas aeruginosa* quorum-sensing molecule N-3-(oxododecanoyl)l-homoserine lactone inhibits T-cell differentiation and cytokine production by a mechanism involving an early step in T-cell activation. *Infect Immun.* (2005) 73:1648–55. doi: 10.1128/IAI.73.3.1648-165 5.2005
- 60. Jacobi CA, Schiffner F, Henkel M, Waibel M, Stork B, Daubrawa M, et al. Effects of bacterial N-acyl homoserine lactones on human Jurkat T lymphocytes-OdDHL induces apoptosis via the mitochondrial pathway. *Int J Med Microbiol.* (2009) 299:509–19. doi: 10.1016/j.ijmm.2009. 03.005
- Hooi DSW, Bycroft BW, Chhabra SR, Williams P, Pritchard DI. Differential immune modulatory activity of pseudomonas aeruginosa quorum-sensing signal molecules. *Infect Immun.* (2004) 72:6463–70. doi: 10.1128/IAI.72.11.6463-6470.2004
- Gupta RK, Chhibber S, Harjai K. Acyl homoserine lactones from culture supernatants of *Pseudomonas aeruginosa* accelerate host immunomodulation. *PLoS ONE.* (2011) 6:e20860. doi: 10.1371/journal.pone.0020860
- Bao L, Yu J, Zhong H, Huang D, Lu Q. Expression of toll-like receptors in T lymphocytes stimulated with N -(3-oxododecanoyl)-Lhomoserine lactone from *Pseudomonas aeruginosa*. *APMIS*. (2017) 125:553–7. doi: 10.1111/apm.12690

- Swearingen MC, Sabag-Daigle A, Ahmer BMM. Are there acyl-homoserine lactones within mammalian intestines? J Bacteriol. (2013) 195:173–9. doi: 10.1128/JB.01341-12
- Erickson DL, Nsereko VL, Morgavi DP, Selinger LB, Rode LM, Beauchemin KA. Evidence of quorum sensing in the rumen ecosystem: detection of N acyl homoserine lactone autoinducers in ruminal contents. *Can J Microbiol.* (2002) 48:374–8. doi: 10.1139/w02-022
- Won M-Y, Oyama LB, Courtney SJ, Creevey CJ, Huws SA. Can rumen bacteria communicate to each other? *Microbiome.* (2020) 8:23. doi: 10.1186/s40168-020-00796-y
- Bivar Xavier K. Bacterial interspecies quorum sensing in the mammalian gut microbiota. C R Biol. (2018) 341:297–9. doi: 10.1016/j.crvi.2018.03.006
- Kumari A, Pasini P, Deo SK, Flomenhoft D, Shashidhar H, Daunert S. Biosensing systems for the detection of bacterial quorum signaling molecules. *Anal Chem.* (2006) 78:7603–9. doi: 10.1021/ac061421n

 Thompson JA, Oliveira RA, Djukovic A, Ubeda C, Xavier KB. Manipulation of the quorum sensing signal AI-2 affects the antibiotic-treated gut microbiota. *Cell Rep.* (2015) 10:1861–71. doi: 10.1016/j.celrep.2015.02.049

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2020 Coquant, Grill and Seksik. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.