



Triple-Negative Breast Cancer Cells Recruit Neutrophils by Secreting TGF- β and CXCR2 Ligands

Shuvasree SenGupta¹, Lauren E. Hein^{2,3}, Yang Xu¹, Jason Zhang⁴, Jamie R. Konwerski⁴, Ye Li⁵, Craig Johnson⁵, Dawen Cai⁵, Janet L. Smith^{4,6} and Carole A. Parent^{1,2,3,5,6*}

¹ Department of Pharmacology, University of Michigan Medical School, Ann Arbor, MI, United States, ² Cancer Biology Graduate Program, University of Michigan Medical School, Ann Arbor, MI, United States, ³ Rogel Cancer Center, University of Michigan, Ann Arbor, MI, United States, ⁴ Department of Biological Chemistry, University of Michigan Medical School, Ann Arbor, MI, United States, ⁵ Department of Cell and Developmental Biology, University of Michigan Medical School, Ann Arbor, MI, United States, ⁶ Life Sciences Institute, University of Michigan, Ann Arbor, MI, United States

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*Correspondence:

Carole A. Parent
parentc@umich.edu

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Tumor associated neutrophils (TANs) are frequently detected in triple-negative breast cancer (TNBC). Recent studies also reveal the importance of neutrophils in promoting tumor progression and metastasis during breast cancer. However, the mechanisms regulating neutrophil trafficking to breast tumors are less clear. We sought to determine whether neutrophil trafficking to breast tumors is determined directly by the malignant potential of cancer cells. We found that tumor conditioned media (TCM) harvested from highly aggressive, metastatic TNBC cells induced a polarized morphology and robust neutrophil migration, while TCM derived from poorly aggressive estrogen receptor positive (ER+) breast cancer cells had no activity. In a three-dimensional (3D) type-I collagen matrix, neutrophils migrated toward TCM from aggressive breast cancer cells with increased velocity and directionality. Moreover, in a neutrophil-tumor spheroid co-culture system, neutrophils migrated with increased directionality towards spheroids generated from TNBC cells compared to ER+ cells. Based on these findings, we next sought to characterize the active factors secreted by TNBC cell lines. We found that TCM-induced neutrophil migration is dependent on tumor-derived chemokines, and screening TCM elution fractions based on their ability to induce polarized neutrophil morphology revealed the molecular weight of the active factors to be around 12 kDa. TCM from TNBC cell lines contained copious amounts of GRO (CXCL1/2/3) chemokines and TGF- β cytokines compared to ER+ cell-derived TCM. TCM activity was inhibited by simultaneously blocking receptors specific to GRO chemokines and TGF- β , while the activity remained intact in the presence of either single receptor inhibitor. Together, our findings establish a direct link between the malignant potential of breast cancer cells and their ability to induce neutrophil migration. Our study also uncovers a novel coordinated function of TGF- β and GRO chemokines responsible for guiding neutrophil trafficking to the breast tumor.

Keywords: breast cancer, TNBC, tumor spheroid, neutrophil, chemotaxis, CXCL1/2/3, TGF- β

INTRODUCTION

Neutrophils are increasingly recognized as critical tumor-infiltrating immune cells (1). As part of the innate immune system, they rapidly navigate to sites of infection or tissue injury. Upon arrival to the infected/injured sites, the primary functions of neutrophils are to eliminate invading pathogens and mount an inflammatory response using diverse mechanisms, ranging from phagocytosis and respiratory burst, to the release of toxic granule contents and DNA-containing neutrophil extracellular traps (NETs) (2). Apart from these defense functions, neutrophils have gained attention for their newly discovered role at either promoting (3–5) or hindering (6–8) tumor progression, depending on the types of cancer. The metastasis promoting function of neutrophils during breast cancer has been attributed to the suppression of anti-cancer adaptive immunity (9) and the stimulation of tumor cell growth (3, 10, 11), migration, and invasion (4, 12–14). Tumor-associated neutrophils (TANs) have been detected in varying degrees in the tumor niche of various kinds of malignancies (15), including breast cancer (16, 17). Yet, cancer-associated guidance cues that drive neutrophil migration to the breast tumor niche are not well defined, partly because of the molecular heterogeneity among different breast cancer subtypes (18).

The major molecular subtypes of breast cancer are luminal A, luminal B, HER2 positive, and triple-negative breast cancer (TNBC), which widely differ in the expression of estrogen hormone receptor (ER), progesterone hormone receptor (PR), and overexpression of human epidermal growth factor receptor-2 (Her-2/neu) (18). The predominant luminal subtypes are characterized by the presence of hormone receptors (HR+), and exhibit a relatively less aggressive clinical outcome (19). In contrast, TNBC tumors, which comprise ~15–20% of all breast cancers, are notoriously aggressive with poor clinical outcome, and the occurrence of early distal metastasis is common (20). Because of the lack of the receptors commonly targeted by existing targeted therapies, the primary therapeutic options for TNBC patients are currently restricted to chemotherapy. However, recent encouraging outcomes with a subset of TNBC subtypes (21) using immune-based strategies have prompted focus on characterizing the immune landscape of the breast tumor niche more comprehensively. Importantly, accumulating evidence has recently highlighted the importance of myeloid cells in regulating the responses of lymphoid populations (22, 23). Moreover, recent studies have reported the frequent presence of TANs in the tumor niche of TNBC subtypes (16, 17). In contrast, relatively less aggressive, hormone receptor-positive (HR+) breast tumors have fewer TANs (16), pointing toward an association between the malignant potential of breast cancer cells and the degree of neutrophil recruitment to tumors.

Abbreviations: TNBC, Triple negative breast cancer; ER, Estrogen receptor; HR, Hormone receptor; TAN, Tumor associated neutrophil; TCM, Tumor conditioned media; f-MLF, Formyl-methionyl-leucyl-phenylalanine; TGF- β , Transforming growth factor- β ; GRO, Growth related oncogene; G-CSF, Granulocyte colony stimulating factor; GM-CSF, Granulocyte macrophage colony stimulating factor; EMT, Epithelial-to-mesenchymal transition; PT, Pertussis toxin.

Neutrophils are specially equipped with the ability to directionally migrate toward chemical guidance cues, a process referred to as chemotaxis. They undergo chemotaxis by sensing gradients of chemical cues that bind and activate their cognate receptors, which induce polarized morphology and directed migration (24). Both tumor and stromal cells are capable of secreting chemokines, cytokines, and growth factors, which in turn facilitate neutrophil mobilization to the tumor niche (25–27). However, it remains unknown whether tumor cell intrinsic differences in the expression pattern of guidance cues are the sole determinant of the differential presence of TANs in different molecular subtypes of breast cancers.

In the present study, we compared the neutrophil recruitment abilities of tumor-conditioned media (TCM) derived from breast cancer cell lines representing either TNBCs or estrogen receptor positive (ER+) subtypes that constitute the majority of HR+ breast cancer (28, 29). We followed neutrophil migration in real time in 3 dimensional (3D) collagen matrices towards TCM and determined how tumor-secreted factors modulate neutrophil velocity and directionality. In addition, by establishing a neutrophil-tumor spheroid co-culture system, we observed the ability of breast cancer cells to directly recruit neutrophils. Most importantly, we characterized the nature of the active factors secreted from breast cancer cells using a number of pharmacological, biochemical, and immunological approaches, and identified the factors responsible for neutrophil recruitment by breast tumor.

MATERIALS AND METHODS

Materials

Formyl-methionyl-leucyl-phenylalanine or fMLF (F3506), DMSO, and Pertussis toxin (PT, P7208) were purchased from Sigma-Aldrich. Human recombinant CXCL8 (CHM349), CXCL1 or GRO α (CHM329) and GM-CSF (CYT-838) were purchased from ProSpec.

Cell Cultures

We used a panel of 13 different cell lines of breast epithelial origin in the study. Cell lines of different receptor expression status and malignant potential were cultured using medium and serum described in **Table 1**. All cell lines were maintained at 37°C with 5% CO₂ except SUM149 cells, which were maintained with 10% CO₂ (30). All cell lines tested negative for mycoplasma contamination using the Mycoalert detection kit (Lonza).

Isolation of Human Neutrophils

Blood was obtained from healthy human male and female subjects aged 19–65 years, through the Platelet Pharmacology and Physiology Core at the University of Michigan. The Core maintains a blanket IRB for basic science studies, where HIPAA information is not required. Therefore, while the IRB-approved Core enrolls healthy subjects that conform to the protection of human subject standards, we did not have access to this information. The samples that we received were fully de-

TABLE 1 | Characteristics and culture conditions of the cell lines used in this study.

Cell lines	ER	PR	HER2	Malignant potential	Culture Media	Serum
MCF10A (M1)	–	–		Benign	*DMEM F12	5% HS
MCF10CA1a (M4)	–	–		Invasive	DMEM F12	5% HS
MDA-MB-231	–	–		More Aggressive	DMEM	10% FBS
MDA-MB-231 TGL	–	–		More Aggressive	DMEM	10% FBS
MDA-MB-231 BrM2-831	–	–		More Aggressive	DMEM	10% FBS
MDA-MB-231 BoM 1833	–	–		More Aggressive	DMEM	10% FBS
MDA-MB-231 LM2-4175	–	–		More Aggressive	DMEM	10% FBS
SUM149	–	–		More Aggressive	**Ham's F12	5% FBS
BT549	–	–		More Aggressive	RPMI	10% FBS
Hs578T	–	–		More Aggressive	DMEM	10% FBS
MCF-7	–	–		Less Aggressive	DMEM	5% FBS
BT474	+	+	+	Less Aggressive	RPMI	10% FBS
HCC1428	+	+		Less Aggressive	RPMI	10% FBS
T47D	+	+		Less Aggressive	RPMI	10% FBS

ER, estrogen receptor; PR, progesteron receptor; HER2, human epidermal growth factor receptor 2. Media conditions: FBS, fetal bovine serum; HS, horse serum.* Media supplemented with 10 µg/ml insulin (Invitrogen), 10 ng/ml EGF (Peprotech), 0.5 mg/ml hydrocortisone and 100 ng/ml cholera toxin (both from Sigma Aldrich). ** Media supplemented with 5 µg/mL insulin, and 1 µg/mL hydrocortisone (Sigma-Aldrich). MCF10A and MCF10CA1a were obtained from Karmanos Research Institute; MDA 231-TGL, MDA 231-BrM2-831, MDA 231-BoM-1833, MDA 231-LM2-4175 were purchased from Memorial Sloan Kettering Cancer Center; SUM-149 was from Dr. Sofia Merajver, University of Michigan; BT549, Hs578T and BT474 were purchased from ATCC; MCF-7, HCC1428, and T47D were kindly gifted from Dr. James Rae, University of Michigan.

identified. Neutrophils were purified using dextran-based sedimentation followed by histopaque-based density gradient centrifugation, adapted from the method described previously (31, 32). Endotoxin free (< 0.005 EU/mL) cell culture grade water (SH30529, Cytiva) was used to dilute any reagents used for neutrophil isolation from the blood. All tubes used were coated with 0.5% tissue culture grade BSA (A9576, Sigma-Aldrich) diluted in DPBS in order to prevent non-specific cell activation by adhesion to the tube wall. Isolated neutrophils were cleared of residual red blood cells by ACK lysis buffer (10-548E, Lonza) and resuspended in mHBSS buffer (20mM HEPES, pH 7.4, 150mM NaCl, 4 mM KCl, 1.2mM MgCl₂, and 10 mg/ml glucose). Viable cells were counted using 0.1% trypan blue (SV30084.01, Cytiva). Cells were more than 99% viable immediately following isolation. In order to address donor-to-donor variability of neutrophil response, cells were routinely tested for minimum basal activity and a robust response to fMLF stimulation before proceeding with daily studies. A dose dependent conversion from basal circular to elongated polarized morphology of fMLF stimulated neutrophils were confirmed by bright-field imaging of two randomly selected fields using 40x objective lens on a Zeiss Axiovert microscope.

Tumor Spheroid Generation

Tumor spheroids were generated using the hanging drop method (33). Briefly, 1000 cancer cells resuspended in full serum media were seeded in 20 µl drops on the inside surface of petri dish lids. Lids were then inverted and placed onto media filled dishes. Spheroids were grown for 4 days in the incubator at 37°C. Day-4 spheroids, visible as aggregates at the bottom of each hanging drop, were harvested and used either for imaging or to generate TCM.

Spheroid Imaging

Tumor spheroids were imaged using 2-photon microscopy. Briefly, day-4 spheroids were fixed with 4% PFA for 1 hour at room temperature. Spheroids were stained with DRAQ5

(108410, Abcam) (1:1000) overnight at 4°C and then mounted onto poly-L-lysine coated coverslips. To reduce scattered light and improve image quality, the spheroids were optically cleared using the ultrafast optical clearing method (FOCM) (34). Lastly, the coverslips were mounted onto glass slides using ImmuMount (9990402, Thermo Scientific) with small spacers between the slides and the coverslips to prevent the spheroids from being flattened. The spheroids were imaged at 1040 nm using a C-Plan Apochromat 40x/1.3x oil objective on a Zeiss LSM 780. Image stacks were first processed by Histogram Matching with nTracer-AlignMaster (35) to equate brightness differences among slices within the same stack. A median filter with a radius of 0.5px was then applied to suppress the background noise. Center slices of each image stacks were used to representatively demonstrate the spheroids' structure from each cell type. Movies traversing the image stacks from top to bottom were also generated.

Harvesting Conditioned Media From 2D Culture

To generate conditioned media (CM or TCM), cell lines were seeded at a density of 0.15X10⁶/ml in 2 ml (6-well tissue culture plate), 16 ml (T75 tissue culture flask), or 32 ml (T150 tissue culture dish) volume of complete medium for 24 hrs, at which point they reached ~ 70% confluence. The culture medium was then removed and cells were gently washed twice with calcium and magnesium free sterile DPBS (Gibco) to remove left over serum containing media. Cells were then incubated with fresh medium supplemented with reduced serum (1%) (**Figure 1A**) or without serum (**Figure 2A**) and incubated for an additional 24 or 48 hrs, respectively. The media was then harvested and filtered through 0.22 µm membrane filter to remove dead cell debris. Aliquots were frozen at -30°C until analyzed.

Harvesting Conditioned Media From 3D Spheroid Culture

To generate TCM from cells grown in 3D spheroids, approximately 30 day-4 spheroids were placed in each well of

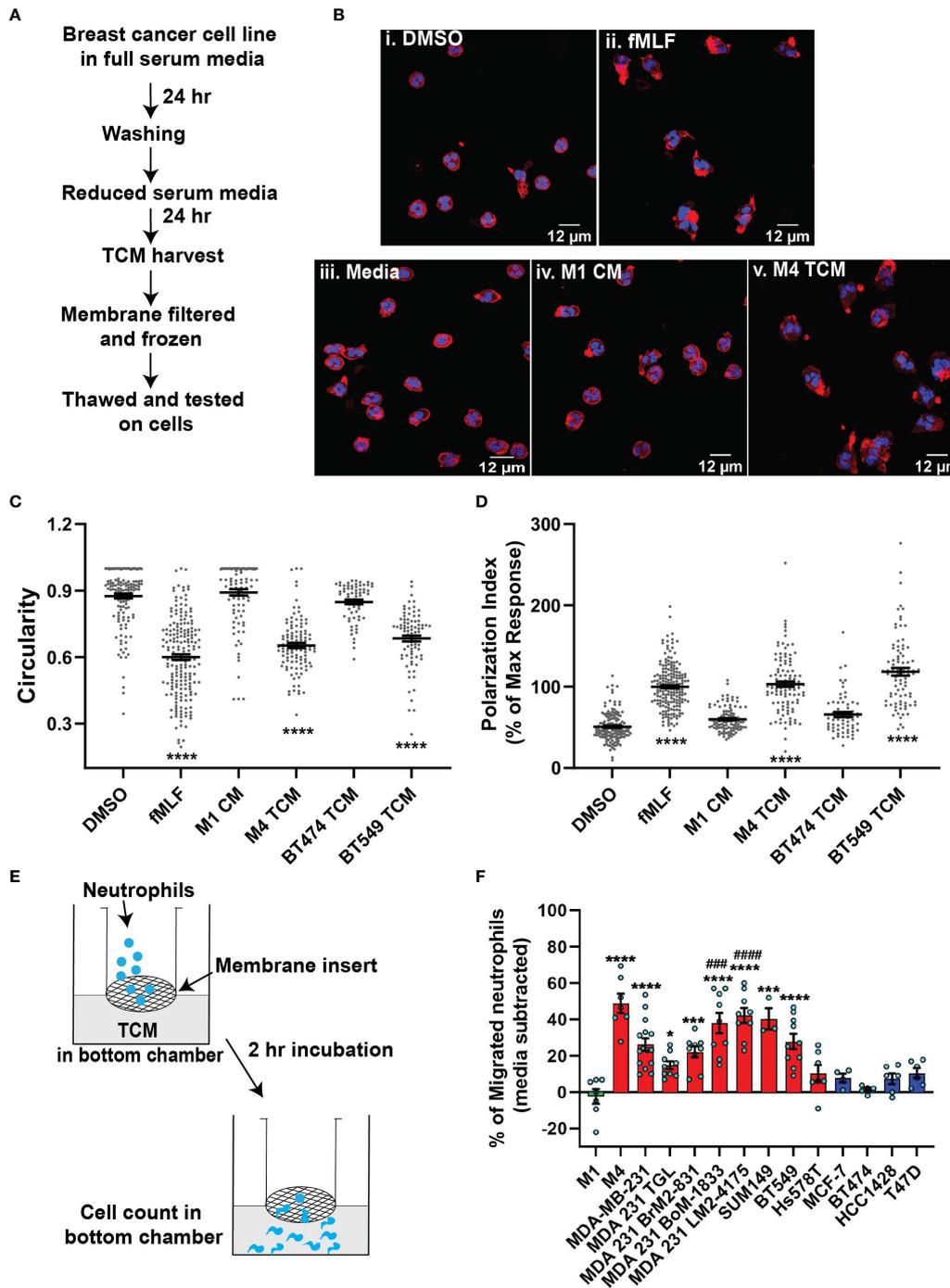


FIGURE 1 | TCM derived from TNBC cell lines induce neutrophil migration. **(A)** Flowchart describing how TCM was generated and harvested from cell lines grown under serum reduced conditions. **(B)** Representative maximum intensity projection confocal images showing fixed neutrophils stimulated with (i), DMSO, (ii) 100 nM fMLF, equal volume of (iii) media, (iv) M1 CM or (v) M4 TCM, and stained for F-actin with phalloidin-TRITC (red) and nuclei with DAPI (blue). Scale bars are 12 μ m. **(C, D)** Graphs depicting the circularity **(C)** and polarization **(D)** index of neutrophils stimulated with controls or equal volume of TCM derived from different breast cancer cell lines. Each dot represents corresponding value of a single neutrophil for each condition. **(E)** Cartoon depicting the neutrophil transwell migration assay. **(F)** Graph depicting the percentage of neutrophils that migrated into the bottom chamber of the transwells containing equal volume of CM from M1 cells or TCM from different breast cancer cell lines. Red and blue bars represent values of neutrophil migration obtained with TCM harvested from TNBC and ER+ cancer cell lines, respectively. Data shown are mean \pm SEM from N=4 **(C, D)** or 3-15 **(F)** independent donors represented by individual dots. Each donor (N) represents an independent experiment. ****P \leq 0.0001, ***P \leq 0.001, *P \leq 0.05 when compared with M1 CM **(C, D, F)** or #####P \leq 0.0001 and ###P \leq 0.001 when compared with MDA 231 TGL (one-way ANOVA with Dunnett’s multiple comparisons test).

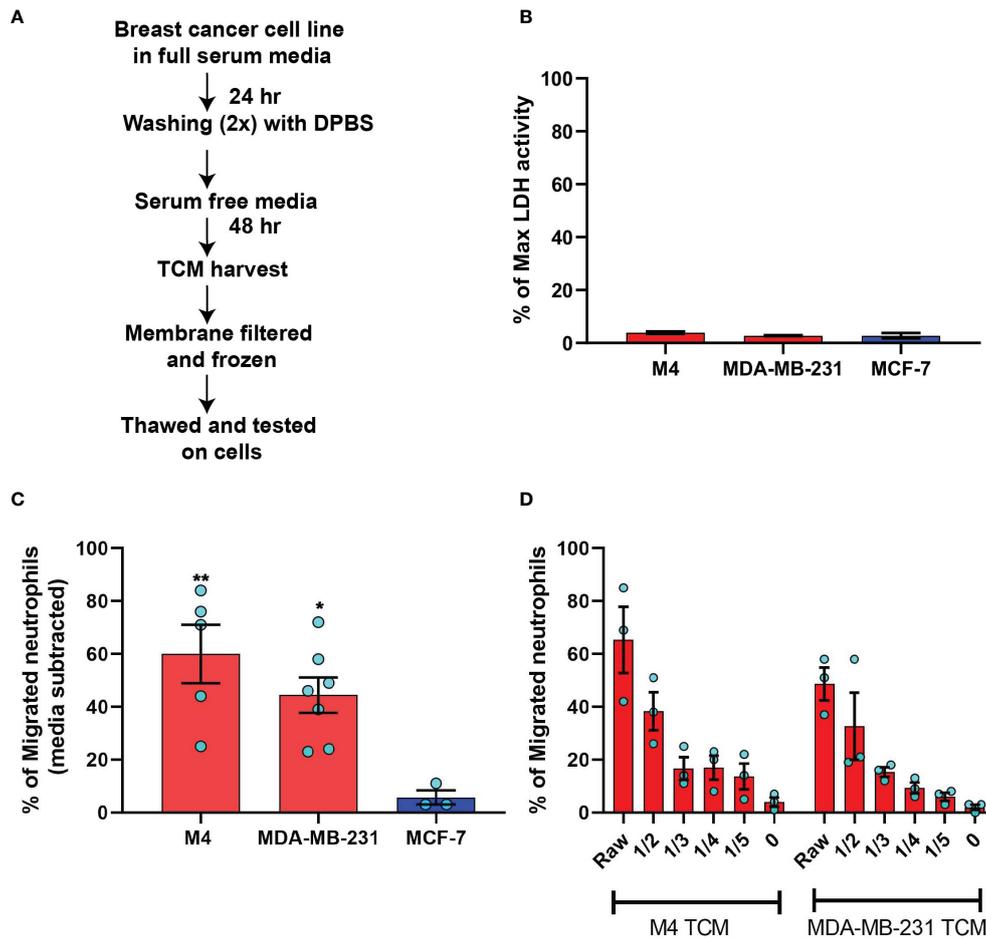


FIGURE 2 | TCM derived from serum deprived TNBC cell lines retain neutrophil recruiting ability. **(A)** Flowchart describing how TCM was generated and harvested from cell lines grown under serum free conditions. **(B)** Graph depicting spontaneous LDH activity in the supernatant of cancer cell lines grown under serum free conditions measured and normalized with respect to maximum LDH activity. Bars represent mean percentage of max LDH activity \pm SEM from 3 independent experiments. **(C)** Graph depicting the percentage of neutrophils that migrated into the bottom chamber of the transwells containing equal volume of TCM from different breast cancer cell lines cultured in serum free condition. Red and blue bars represent values of neutrophil migration obtained with TCM harvested from TNBC and ER+ cancer cell lines, respectively. **(D)** Graph depicting the percent of neutrophils migrating into the bottom chamber of the transwells containing serially diluted equal volume of TCM from two different TNBC cell lines. Results are presented as mean \pm SEM from N=3-7 **(C)** or N=3 **(D)** independent donors represented by individual dots. Each donor (N) represents an independent experiment. ** $P \leq 0.005$, * $P \leq 0.05$ when compared with MCF-7 (one-way ANOVA with Dunnett's multiple comparisons test).

poly-HEMA-coated, 96-well, V-bottom plates. Once settled to the bottom, spheroids were gently rinsed with DPBS and incubated in 200 μ l serum-free media for 48 hrs. The media was then harvested and filtered through a 0.22 μ m membrane filter. Aliquots were frozen at -30°C until analyzed.

Neutrophil Polarized Morphology Analysis by Immunofluorescence

For the analysis of actin polymerization and polarized morphology, neutrophils were seeded at $1-2 \times 10^6$ cells/ml in 1 ml volume in a 35 mm dish with 20 mm #1.5 coverslip (MatTek). The coverslips were pre-coated with 25 μ g/ml fibrinogen (F4883, Sigma-Aldrich) in DPBS for 1 hr at 37°C . Neutrophils were incubated 5-10 min in the incubator at 37°C to

allow floating cells to settle down on the coverslip. Cells were then stimulated with an equal volume of TCM, media control, positive control fMLF or vehicle control DMSO and were further incubated for 15-20 min at 37°C . Following stimulation, the entire liquid medium was gently discarded from each dish and the adherent cells were immediately fixed with 4% PFA (Electron Microscopy Sciences), 0.1% glutaraldehyde (G5882, Sigma-Aldrich), 5% sucrose (BP220, Fisher Scientific), and 0.1M cacodylate (11652, Electron Microscopy Sciences), for 15 min at room temperature (RT). Once fixed, cells were washed with 0.1M glycine (G8898, Sigma Aldrich) in DPBS to quench the autofluorescence of glutaraldehyde. Washed cells were next permeabilized with 0.05% saponin (84510, Sigma Aldrich) in 3% BSA-containing DPBS for 15 min at RT and stained with

phalloidin-TRITC (P1951, Sigma Aldrich) and DAPI (D9542, Sigma Aldrich). Cells were imaged using a 63x objective lens on a Zeiss LSM880 confocal microscope. Cells in three to five different fields across the coverslip were captured randomly per condition in each experiment. F-actin polymerization and cell circularity were measured using the ImageJ image analysis software. Briefly, integrated density, which is a product of cell area and mean fluorescence intensity, and circularity were obtained from ImageJ after manually identifying the boundary of each cell in a given field. Polarization index was calculated by normalizing the integrated density of each cells with respect to the positive control (fMLF) in each experiment.

Neutrophil Polarized Morphology Analysis by Bright-Field Microscopy

To analyze cell circularity in response to TCM fractions or TCM in the presence of inhibitors, 0.15×10^6 neutrophils were seeded in each well of a fibrinogen coated 8 well chambered cover glass with #1.5 cover glass (Cellvis). Cells were allowed to adhere to the cover glass by incubating at $37^\circ\text{C}/5\% \text{CO}_2$ for 5 min followed by stimulation with an equal volume of TCM fractions, controls or TCM with or without the inhibitor for an additional 15 min at $37^\circ\text{C}/5\% \text{CO}_2$. Bright-field images of the cells in two to three randomly selected fields per condition were captured using a 40x objective lens on a Zeiss Colibri microscope. Circularity of each cell was measured by identifying cell boundary manually using ImageJ software.

Transwell Assay

To quantify the ability of TCM to induce neutrophil migration, transwell migration chambers with membrane inserts of 3 μm diameter pore size (Corning and Greiner Bio-One) were used. Both the inserts and the bottom wells were coated with 2% tissue culture grade BSA for 1 hr at 37°C to prevent strong neutrophil adhesion. Coated inserts and wells were rinsed with DPBS twice to remove residual BSA. Freshly isolated neutrophils at a density of $4 \times 10^6/\text{ml}$ were seeded onto the inserts (100 or 200 μl), and placed in 24-well or 12-well plates, respectively. Control chemoattractants or TCM were gently added in 600 or 1200 μl volume to the bottom wells of 24-well or 12-well plates, respectively, avoiding any bubble formation between the insert and the liquid in the well. Migration was allowed to take place in $37^\circ\text{C}/5\% \text{CO}_2$ for 2 hrs. The percentage of neutrophils migrated to the bottom chamber was calculated from the cell counts obtained using a hemocytometer.

Under Agarose Assay

Under agarose migration assay was performed following the protocol described elsewhere (36). Briefly, 0.5% SeaKem ME agarose (Lonza) in 50% each of DPBS and mHBSS was poured and allowed to solidify in 1% BSA-coated 35 mm glass bottom dish with 20 mm micro-well #1.5 cover glass (Cellvis). Using a metallic hole punching tool, three 1 mm diameter wells were carved out at a 2 mm distance from each other. Neutrophils were stained with 0.5 μM CellTracker Red CMPTX dye (Invitrogen) for 15 min at 37°C in rotation and re-suspended in mHBSS. In some experiments, cells were pre-treated with pertussis toxin

(PT) for 2 hrs followed by staining in PT supplemented buffer. 7 μl (5×10^4 cells) of stained cells were added to the wells at the edges and cells were allowed to migrate towards the center well containing 7 μl of either fMLF (100 nM) or 50 $\mu\text{g}/\text{ml}$ of GM-CSF for 2 hrs at $37^\circ\text{C}/5\% \text{CO}_2$. End point images of cells migrating towards the chemoattractant were acquired using a 10x objective lens (Zeiss Axiovert microscope).

3D Collagen Matrix Assay

Migration in 3D matrix of type I collagen was performed as previously described (37). Briefly, neutrophils were fluorescently labeled by staining with 0.5 μM CellTracker Red CMTPX dye (Invitrogen) for 15 min at 37°C in rotating condition. Collagen mix was prepared on ice by adding sodium bicarbonate (S8761, Sigma Aldrich), MEM (10X, M3024, Sigma Aldrich), and Purecol[®] (3 mg/mL, 5005, Advanced Biomatrix) in a 1:2:15 ratio. Labeled cells were washed and resuspended in 1x mHBSS. 0.5×10^6 labeled cells were gently added to the collagen mix such that the final concentration of collagen is 0.2%. The collagen-cell mix was then immediately added to fill two-thirds of a migration chamber made of coverslip and glass slide. The slide with the chamber was then placed in upright position in the incubator at $37^\circ\text{C}/5\% \text{CO}_2$ for at least 20 min to allow for the gel to polymerize. The upper one-third empty space of the chamber was filled with chemoattractant, either TCM or controls such as media, CXCL8 or buffer. The chamber was immediately sealed with paraffin mix. Time-lapse video was captured for 40 mins at different positions (see **Figure 3A**) across the boundary of the chemoattractant and collagen-cell mix using an environmental controlled Zeiss LSM 880 at 10x magnification with the pinhole open. The Manual Tracking plugin in ImageJ software was used to analyze the migration data, where about 70-100 cells were manually tracked over the 40 min time course in each of three randomly selected positions per condition in each experiment. For CXCL8 and buffer control, cells located 1 to 1.5 mm below the matrix boundary were tracked as cells near the boundary mostly exhibited random slow migration, possibly because high concentration of CXCL8 in the boundary saturates the receptors (CXCR1/2). In contrast, cells in the immediate vicinity of the boundary ($\sim 500 \mu\text{m}$) showed maximum directional movement for TCM and were tracked for both TCM and media control. The software quantified the velocity and directionality of migration, where the directionality was represented by X-FMI, which reflects the efficiency of forward migration of the cells parallel to the gradient (38).

Tumor Spheroid-Neutrophil Co-Culture Assay

To evaluate neutrophil recruitment toward cancer cells in real time, a co-culture model system was developed (**Figure 5A**). Wells of a 24 well glass-bottom plate (P24-1.5H-N, Cellvis) were coated with 200 $\mu\text{g}/\text{ml}$ fibronectin (F1141, Sigma-Aldrich) for at least 1 hr at 37°C . Approximately 20 spheroids resuspended in 250 μl fresh media (**Table 1**) supplemented with reduced serum (1%) were plated in each well. Spheroids were allowed to attach to the plate for about 4 hrs in the incubator at $37^\circ\text{C}/5\% \text{CO}_2$.

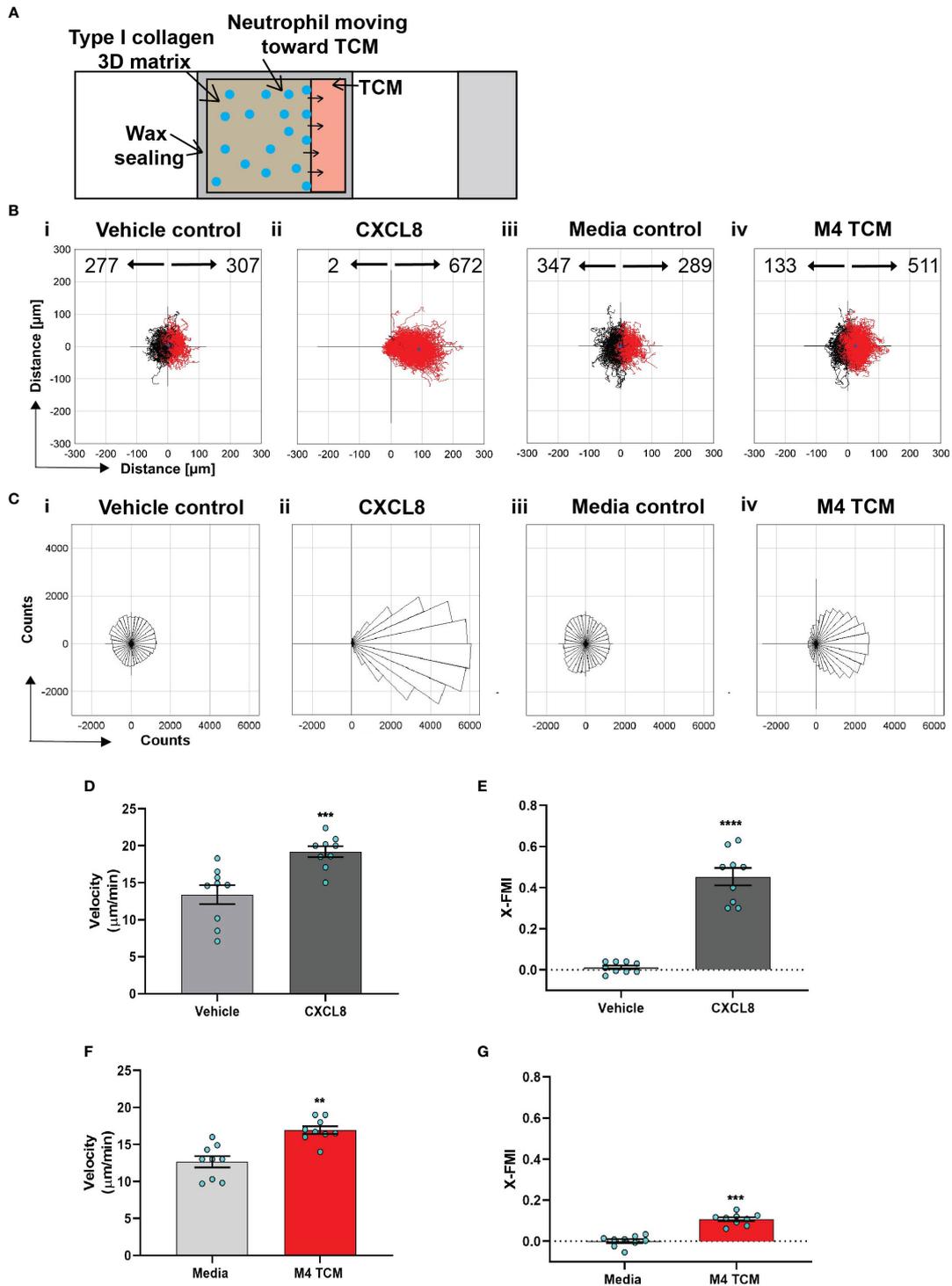


FIGURE 3 | Neutrophils directionally migrate towards secreted factors harvested from TNBC cell line. **(A)** Cartoon depicting the migration chamber used to assess the chemotaxis of neutrophils in 3D collagen matrix towards TCM. **(B, C)** Plots showing individual cell tracks **(B)** and rose plots showing angle histograms of the migrating cells **(C)** generated from the combined tracks from N=3 independent donors in response to vehicle control (Bi, Ci), 500 nM CXCL8 (Bii, Cii), media control (Biii, Ciii), and M4 TCM (Biv, Civ). Numbers with left and right arrowhead in B represent number of neutrophils moving away and towards different conditions, respectively. **(D–G)** Graph depicting velocity **(D, F)** and X-FMI **(E, G)** of neutrophils migrating towards 500 nM CXCL8 and vehicle control **(D, E)** or M4 TCM and media **(F, G)**. Each bar represents mean \pm SEM from N=3 independent donors with each dot representing average values from independent positions. ****P < 0.0001, ***P \leq 0.0005, **P \leq 0.005 when compared with corresponding control (paired sample t-test).

Towards the end of incubation, neutrophils were primed with 5 ng/ml GM-CSF for 30 mins. 5×10^4 or 1×10^5 primed neutrophils were seeded to each well in 250 μ l of 1% serum media. Multiple spheroids in each well were imaged for 24 hrs using an environmental controlled Zeiss Colibri microscope at 10x magnification. To analyze the migration of the neutrophils, the paths of at least 40 neutrophils per video, spaced out both spatially and temporally, were manually tracked using the Manual Tracking plugin in ImageJ software. The position data acquired from the tracking were used in a custom MATLAB script to quantify X-FMI.

Treatment With Pertussis Toxin

Neutrophils were treated with pertussis toxin as previously described (39). Briefly, 4×10^6 neutrophils/ml were pre-incubated in mHBSS with 10% heat inactivated fetal bovine serum without or with 500 ng/ml pertussis toxin for 2 hours at 37°C with gentle rotation. After 2 hrs, the cells were centrifuged and the pellet was resuspended in mHBSS with 0.5% BSA with or without pertussis toxin. The cells were then loaded into transwell inserts and their migration was assessed as described above.

TCM Fractionation by Size Exclusion Chromatography

30 ml serum free TCM was collected from M4 and MDA-MB-231 cell lines as described above. TCM was concentrated 100-120 fold using Amicon Ultra-15 centrifugal tubes (Millipore Sigma) of 3 kDa molecular weight cut-off. Briefly, 15 ml TCM was centrifuged at 4000 rpm for 40 min at RT. Filtrates were discarded and an additional 15 ml TCM was added to the resulting concentrate, mixed well and centrifuged as before. The final concentrate was buffer exchanged twice with sterile DPBS to get rid of the residual media components and frozen at -30°C until use. Concentrated TCM was applied to a 30 cm x 10 mm Superdex 75 10/300 column (GE Life Sciences) equilibrated with 30 ml DPBS. The column was calibrated using four standards: aprotinin MW 6.5 kDa (A3886, Sigma-Aldrich), cytochrome c MW 12.4 kDa (C7150, Sigma Aldrich), carbonic anhydrase MW 29 kDa (C7025, Sigma-Aldrich), and bovine serum albumin MW 66 kDa (A8531, Sigma Aldrich). Sample was run at a rate of 0.2 ml/min and fractions of 150 μ l were collected using DPBS as an eluent. Fractions were stored at 4°C until use.

Multiplex ELISA Array

TCM concentrates were sent to RayBiotech, Inc. (GA) to screen for a panel of potential neutrophil chemoattracting factors encompassing CXCL chemokine family members CXCL1/2/3 aka GRO, CXCL5, CXCL6, CXCL8, CXCL12, CCL family members CCL2, CCL3, CCL5, growth factors G-CSF, GM-CSF, and cytokine TGF- β . The presence of the analytes was detected and quantified using a high throughput custom Quantibody multiplex enzyme-linked immunosorbent assay (ELISA) (RayBiotech) and the data were analyzed by RayBiotech, Inc. Briefly, concentrates were prepared as described above using 60 ml serum free TCM harvested from the respected cell lines such that the protein concentration

approximated 1-2 mg/ml, as estimated using the Pierce BCA protein assay kit (Thermo Fisher). Concentrated samples were tested without dilution in quadruplicate, as per the manufacturer instructions. The protein concentration for each target analyte was quantified by comparing the fluorescence signal from the sample with the individual standard curve generated by known concentrations of each protein.

Western Blotting Analysis

ERK phosphorylation was assessed using western blotting analysis. In brief, neutrophils were pretreated with 2 mM pepabloc protease inhibitor (Sigma Aldrich) for 15 min at 37°C with gentle rotation. 4×10^6 neutrophils/ml were seeded in 500 μ l in each well of a 12 well tissue-culture plate pre-coated with 25 μ g/ml fibrinogen. Cells were allowed to settle down in the well by incubating the plate for 5 min at 37°C. Cells were then exposed to an equal volume of TCM, media control, positive controls including fMLF and CXCL1 or the corresponding vehicle controls. After 5 min stimulation at 37°C/5% CO₂, the plate was placed on ice and the cells were lysed in 1X Laemmli buffer (Biorad) containing 5% β -mercaptoethanol (VWR), phoSTOP (Millipore Sigma) and HaltTM protease and phosphatase inhibitor cocktails (Thermo Scientific) at 95°C for 10 min. An equal volume of each was resolved by 10% SDS-PAGE, transferred onto PVDF membranes (Millipore), blocked with 5% nonfat dry milk for 1 hr and probed with primary Abs: phospho-p44/42 MAPK (ERK1/2) (Thr202/Tyr204, dilution 1:2000, 4370, Cell Signaling Technology) and alpha tubulin (dilution 1:2000, 66031-1-Ig, Proteintech). Bands were visualized using HRP-conjugated secondary antibodies (dilution 1:8000, Jackson ImmunoResearch), SuperSignalTM West Pico PLUS Chemiluminescent Substrate, and a C600 digital imaging system (Azure). Integrated density for each band was measured using ImageJ software.

Neutrophil Inhibition

Neutrophils were pre-treated with the CXCR2 receptor antagonist AZD5069 (Cayman Chemical), the TGF- β 1 receptor ALK5 antagonist SB431542 (Cayman Chemical), or the SMAD3 phosphorylation inhibitor SIS3 (Cayman Chemical) at 1 μ M, 1 μ M and 3 μ M, respectively, or with vehicle control DMSO for 30 min at 37°C with gentle rotation before they were added to the polarization or transwell migration assay.

Lactate Dehydrogenase (LDH) Assay

Cancer cell membrane integrity was analyzed for the detection of cell death using the LDH cytotoxicity assay kit (Thermo Scientific) based on the manufacturer instructions. Briefly, cancer cells were seeded at 0.15×10^6 cells/ml in 100 μ l volume in a 96-well tissue culture dish in two sets of triplicate wells. The control set was for maximum LDH release by subjecting cells to undergo complete lysis, and the test set was for spontaneous LDH release. Cells were subjected to incubation and subsequent media change as described for generating serum free TCM. Three days after initial seeding, 10 μ l of 10x lysis buffer and 10 μ l of sterile water were added to the control and test set,

respectively. After incubating the plate for 45 min in the incubator at 37°C/5% CO₂, 50 µl from each well was transferred to a fresh 96 well plate, mixed with 50 µl LDH reaction mix, and incubated for additional 30 min at RT protected from light. Finally, the reaction was stopped by adding the stop solution and absorbance was measured at 490nm and 680nm by a SpectraMax M5 Multi-Mode Microplate Reader (Molecular Devices). Absorbance was corrected by subtracting background signal of the instrument followed by correcting for the absorbance values of corresponding media. The percentage of maximum LDH release was calculated with respect to corresponding control set.

Cancer Cell Migration and Invasion Assay

Endpoint migration and invasion assays were performed using a transwell system in 24-well plates (353504, Falcon). FluoroBlock™ filter inserts (351152, Corning) with 8 µm pore size were used that block light transmission from 400-700 nm and allow fluorescence detection only from the cells that migrated/invaded to the bottom side of the inserts with a bottom-reading fluorescence plate reader. The inserts were uncoated for the migration assays, and coated for the invasion assays with 0.2 mg/ml type I bovine collagen PureCol® of which the pH was neutralized to 7.2 – 7.4 using sodium bicarbonate. Cancer cells that were serum starved for 24 hrs were seeded onto the inserts at 0.5x10⁵ cells/ml in 100 µl of serum free media. Cells were allowed to migrate toward the bottom chamber containing 500 µl of full-serum media as the chemoattractant or serum-free media as the negative control. After a 24 hr incubation at 37°C/5% CO₂, the inserts were transferred to a fresh 24-well plate with black walls containing 500 µl of 4 µM Calcein AM (Biotium) in mHBSS per well. Cells were incubated for 1 hr at 37°C, and the fluorescence reading of migrated/invaded cells was measured from the bottom at wavelengths of 495/515 nm (Excitation/Emission) by a SpectraMax M5 Multi-Mode Microplate Reader (Molecular Devices).

Bioinformatics

The transcript expression levels of different CXCL chemokine family members in breast cancers were analyzed using datasets identified from the cancer microarray database Oncomine (<http://www.oncomine.org>). A total of 7 datasets from independent studies were selected from a list of studies obtained by setting the search thresholds as follows: Data type, mRNA; gene rank, top 10%; fold change, 2; P-value, 0.0001. Studies that used Affymetrix platforms other than hgu133plus2 and hgu133a were excluded from further analysis. For each of the studies analyzed, data were downloaded from GEO per instructions of the respective publications. Farmer (40), Minn (41), Wang (42), Ivshina (43) used the Affymetrix hgu133A array while Miyake (44), Schuetz (45), Kao (46) used the Affymetrix hgu133plus2.0 array. For the Schuetz dataset (45), we could not get the raw CEL files and subsequently started from the expression matrix provided with the GEO download. CEL files for all other projects were processed using the Robust Multi-array Average (RMA) technique to obtain normalized expression values (47). Estrogen receptor annotation was obtained from the

data download or from Ivshina (43), Wang (42), Minn (41), Miyake (44), Schuetz (45), and Farmer (40) datasets. Estrogen receptor annotation was not provided for Kao (46). Instead, using simple histograms, the cutoff for ER status was set by visually selecting a point between the bimodal distribution of ER expression; that cutoff was 10. The Limma package of Bioconductor was used to fit linear models to each dataset comparing ER- samples to ER+ (48). For Schuetz (45), the model controlled for variation from ductal carcinoma *in situ* (DCIS) and invasive ductal carcinoma (IDC) status. No other study had a co-variate considered. Data for CXCL chemokines were extracted and the log₂ fold change between ER- and ER+ were plotted in a heatmap along with the fold change for the datasets respective estrogen receptor 1 (ESR1) gene (probeset 205225_at). Samples (columns) were sorted by ESR1 expression. From the TCGA data portal, we used the search “Project id” IS “TCGA-BRCA” AND “Workflow Type” IS “HTSEQ-FPKM” AND “Data Category” IS “Transcriptome Profiling” to find 1,222 FPKM (fragments per kilobase of exon model per million reads mapped) data files. The log₂ of the FPKM values were plotted for our chemokines of interest and samples were ordered by the FPKM of ESR1.

Statistical Analysis

GraphPad Prism software was used for data plotting and conducting statistical analysis by tests that are described in the respective figure legends along with the size of the samples. Tests used included two-tailed paired t test, unpaired t test, 1-way ANOVA with Dunnett’s multiple comparisons test or 2-way ANOVA with Sidak’s multiple comparisons test.

RESULTS

Conditioned Media Derived From TNBC Cell Lines Induce Neutrophil Migration

Numerous studies have suggested that soluble factors, such as CXCL chemokine family members, promote neutrophil infiltration in a variety of tumors. For example, CXCL members CXCL8 (aka IL-8) and CXCL1/2 have been implicated in driving neutrophil recruitment in breast cancer (49–52), and overexpression of these chemokines is associated with breast tumor samples and breast cancer cell lines that have negative estrogen receptor expression status and higher aggressiveness (50, 53–55). We confirmed these observations by comparing the expression pattern of all known CXCL chemokines across breast cancer subtypes that differ in ER expression status by analyzing transcriptome-based expression data identified from Oncomine and TCGA databases (**Figure S1**). Out of 12 CXCL chemokines, we found that the expression of CXCL8, CXCL1, and CXCL2 transcripts was significantly elevated in ER- breast cancer specimens compared to ER+ samples in seven, four and three independent studies, respectively (**Figure S1C**). Similarly, TCGA data analysis revealed elevated expression patterns of CXCL8, CXCL1, and CXCL5 in ER- breast cancer specimens (**Figure S1D**).

The differential chemokine expression pattern detected among breast cancer subtypes prompted us to investigate the neutrophil recruiting ability of a panel of breast cancer cell lines that differ in ER expression status and malignant potential. TNBCs and ER+ breast cancer cell lines along with non-malignant M1 and metastatic derivative M4 cells from the MCF10A breast cancer progression model (56–59) (**Table 1**) were cultured *in vitro* under identical seeding cell density. The higher malignant potential of the TNBC cell lines compared to the ER+ breast cancer cell lines was confirmed by assessing their migration and invasion abilities (**Figures S2A, B**). We harvested TCM after growing cells in serum reduced media for 24 hrs (**Figure 1A**), and tested the ability of the TCM to activate neutrophils. Neutrophils stimulated with fMLF, which is a formylated bacterial tripeptide and a potent neutrophil chemoattractant (32), exhibited an elongated morphology with strong F-actin accumulation in the protrusions, detected using fluorescently tagged phalloidin (32) (**Figure 1Bii**; quantified in **Figures 1C, D**). In contrast, the majority of neutrophils exposed to DMSO control maintained a circular shape similar to resting neutrophils, and a uniform distribution of cortical actin (**Figure 1Bi**). Similar to fMLF, TCM derived from metastatic M4 cells induced strong polarized morphology and F-actin depositions at the poles (**Figure 1Bv**; quantified in **Figures 1C, D**), while most of the cells stimulated with conditioned media (CM) collected from M1 cells remained circular (**Figure 1Biv**; quantified in **Figures 1C, D**). In addition, we found a significant decrease in circularity and increase in F-actin intensity of neutrophils stimulated with TNBC BT549 cell-derived TCM, while TCM derived from ER+ BT474 cells failed to induce such changes (**Figures 1C, D**). Next, we assessed the ability of the TCM to induce neutrophil migration using a transwell setup (**Figure 1E**). Similar to the polarization response, we found that a significant proportion of neutrophils migrated toward TCM derived from metastatic M4 and TNBC cells, but not from ER+ cells (**Figure 1F**). Because each breast cancer cell line has a unique genetic background, we also included in our panel bone-, lung- and brain-specific highly metastatic MDA-MB-231 cells selected from the same parental MDA-MB-231 TGL cells (41, 60, 61). Interestingly, neutrophil migration towards TCM derived from lung- and bone-specific MDA-MB-231 cells was significantly higher than towards TCM from the relatively less metastatic MDA-MB-231 TGL parental line (**Figure 1F**).

To minimize neutrophil activation by the presence of serum in the TCM and to mimic the lack of nutrient availability as encountered by cancer cells in an *in vivo* tumor mass, we harvested TCM from cells cultured in 0% serum for 48 hrs (**Figure 2A**) – conditions that lead to minimal cell death (**Figure 2B**). We found that serum free TCM derived from M4 and MDA-MB-231 cells induced strong neutrophil migration in transwell assays compared to the TCM harvested from MCF-7 cells cultured under the same conditions (**Figure 2C**). Notably, a greater percentage of neutrophils migrated toward serum free TCM (**Figure 2C**) than to serum reduced TCM (**Figure 1F**) derived from either M4 (57% vs. 49%) or MDA-MB-231 (42% vs. 26%) cells. Importantly, a gradual decrease in neutrophil

recruitment with serially diluted serum free TCM showed that TCM triggers a dose-dependent response (**Figure 2D**). Together, these findings establish that TCM derived from breast cancer cells with triple negative receptor expression status and aggressive malignant potential induce neutrophil polarization and robust migration by secreting specific chemotactic factors. For subsequent studies, we used TCM generated under serum starved conditions from all cell lines.

Neutrophils Directionally Migrate Towards Secreted Factors Harvested From TNBC Cell Line

We next assessed the ability of TCM to induce neutrophil migration in a more physiologically relevant interstitial tissue-like microenvironment by monitoring neutrophil motility through 3D matrices of type I collagen and time-lapse imaging (**Figure 3A**). By tracking the movement of neutrophils, we generated trace plots (**Figure 3B**) and rose plots (**Figure 3C**) that depict the behavior of cells in response to M4 TCM or media control. CXCL8 was used as a positive control. We found that neutrophils migrated directionally towards M4 TCM compared to media control (**Figure 3Biii, 3Biv, 3Ciii, 3Civ**; see also **Movies S1, S2**). We also found that neutrophils migrated towards M4 TCM with a significantly elevated velocity and directionality, compared to media control (**Figures 3F, G**). These increases in cell velocity and directionality also occurred in response to CXCL8 (**Figure 3Bi, 3Bii, 3Ci, 3Cii, 3D&3E**; see also **Movie S3, S4**). However, a lower directionality was measured in response to M4 TCM (0.1) compared to CXCL8 (0.45), which could reflect the complexity of TCM composition. Indeed, unlike a single chemoattractant like CXCL8, the TCM includes multiple active factors that can potentially modulate their effect and have different diffusibility through the collagen. Taken together, these results indicate that active factors secreted from TNBC cells induce directional migration of neutrophils.

TNBC Spheroids Secrete Factors That Recruit Neutrophils

We investigated the neutrophil recruitment ability of 3D tumor spheroids generated with breast cancer cell lines. First, we generated tumor spheroids with diameters ranging from 200 to 350 μm using the hanging drop method. Using 2 photon microscopy, we assessed the morphology of the spheroids, which differ in appearance, ranging from irregular, loose aggregates (MDA-MB-231 and BT549) to tight, symmetrical spheres (BT474 and M4) (**Figure 4A, Movies S5–S8**). We also determined that under our growth conditions, the spheroids did not harbor a hollow or necrotic core at their center (**Figure 4A**), ruling out the possible release of damage-associated molecular patterns (DAMPs) from damaged or dying cells at the spheroid core that may impact neutrophil recruitment neutrophil recruitment (62) (**Figure 4A, Movies S5–S8**).

To evaluate whether TNBC spheroids secrete neutrophil recruiting factors similarly to TNBC cells grown in monolayers, we assessed neutrophil migration toward spheroid-derived TCM and found that spheroid-derived TCM

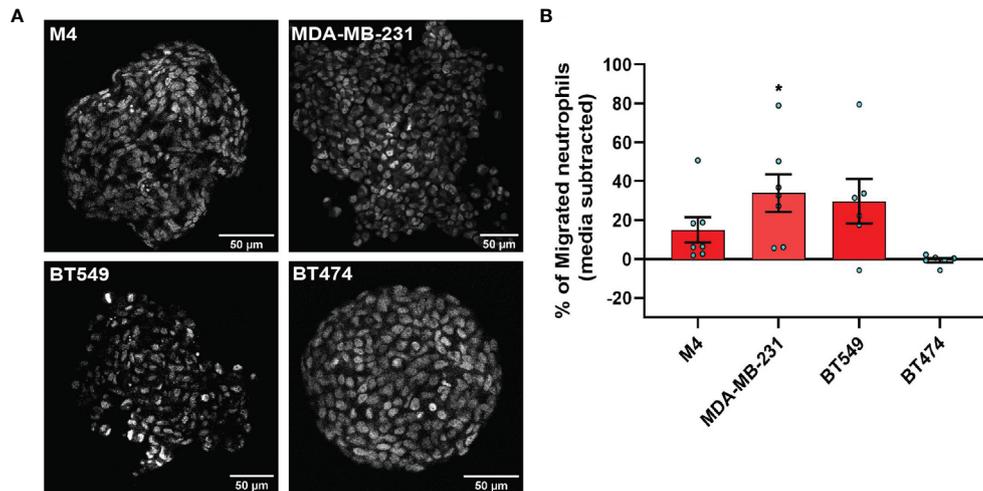


FIGURE 4 | TNBC spheroids secrete factors that induce neutrophil migration. **(A)** Representative 2-photon images of day-4 spheroids stained with the nuclear dye DRAQ5. Each image is from the field of view located at the midpoint between the top and bottom of the z-stack imaging range. **(B)** Graph depicting the percentage of neutrophils that migrated into the bottom chamber of the transwells containing equal volume of TCM derived from M4, MDA-MB-231, BT549, or BT474 spheroids. Data shown are mean \pm SEM from N=6-7 independent donors represented by individual dots. Each donor (N) represents an independent experiment. * $P \leq 0.05$ when compared with BT474 TCM (one-way ANOVA with Dunnett's multiple comparisons test).

from all three TNBC cell lines (M4, MDA-MB-231, and BT549) had neutrophil recruiting activity, while spheroid-derived TCM from the ER+ cell line (BT474) was inactive (**Figure 4B**). These results establish that TNBC cells secrete neutrophil-recruiting factors in either 2D or 3D environments.

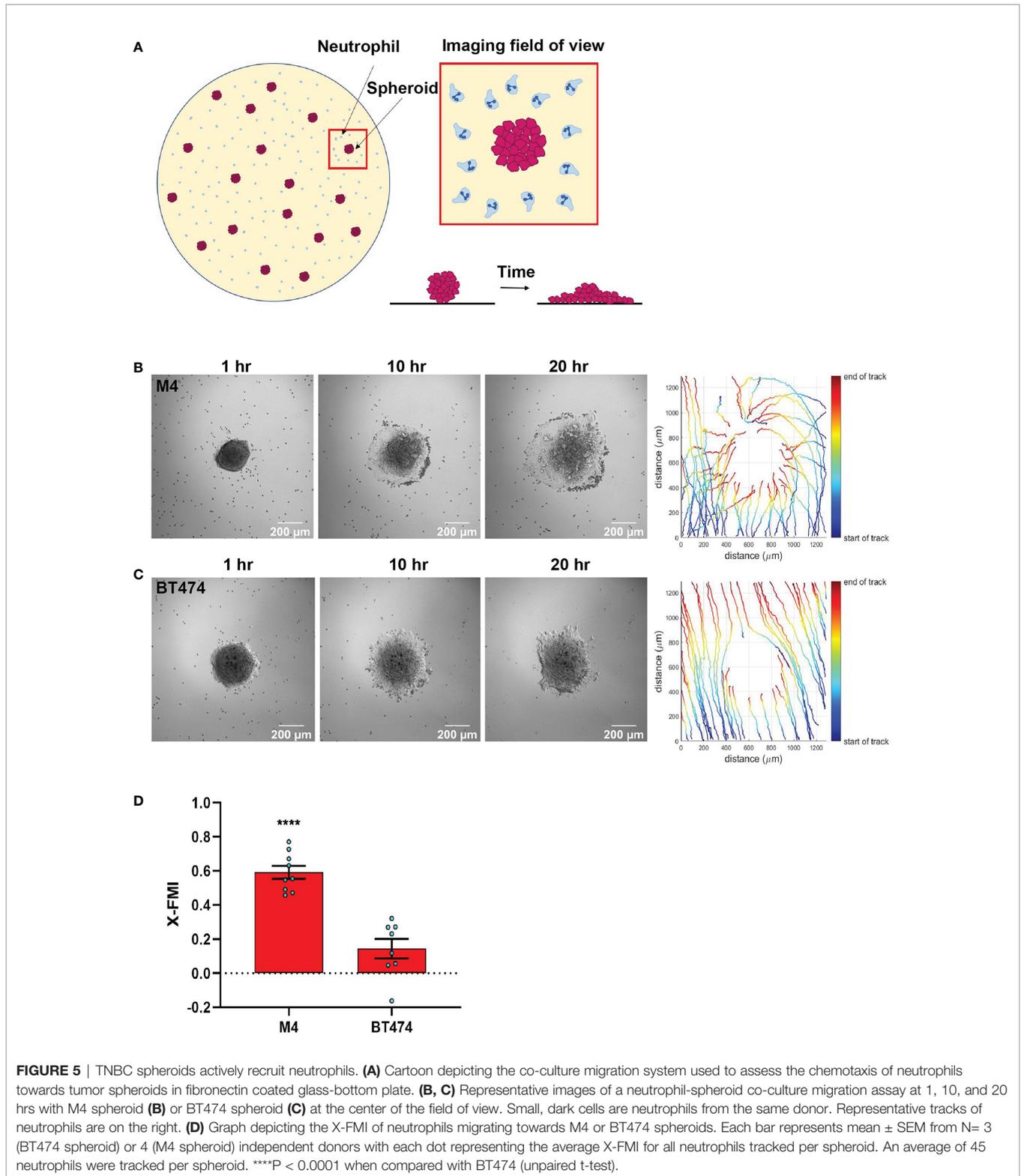
Next, we developed a neutrophil-tumor spheroid co-culture system to monitor the migration of neutrophils towards tumor spheroids by time-lapse imaging (**Figure 5A**). During the early hours of imaging, when neutrophils are not activated and therefore not attached to the surface, we noted that the neutrophils followed the fluid flow and some of them non-specifically got trapped at the boundary of the tumor spheroids (**Movies S9, S10**). However, over time, activated neutrophils clearly attached to the surface, actively migrated towards the M4 spheroids, and accumulated around the spheroid boundary (**Figures 5B, Movie S9**). In contrast, we found that neutrophils never strongly adhered at the boundary or directionally migrated towards BT474 spheroids. Instead, they followed the direction of the flow in the system (**Figure 5C, Movie S10**). Indeed, quantification of the directionality of neutrophil migration towards the spheroid revealed a higher directionality towards M4 spheroids compared to BT474 spheroids (**Figure 5D**). Taken together, these findings further affirm that TNBC cell lines induce directional migration of neutrophils by actively secreting specific chemotactic factors.

TNBC Cell Lines Secrete Neutrophil Activating Factors More Abundantly Than ER+ Cancer Cell Lines

One of the major signaling pathways triggered in neutrophils by a wide range of activating factors, including chemoattractants, is the extracellular signal-regulated kinase (ERK) pathway (63, 64).

ERK pathway is also important for activating the migration machinery of neutrophils (65). We therefore compared the ability of the secreted factors from different breast cancer cell lines to activate ERK by monitoring its phosphorylation status. We found that TCM derived from either M4 or MDA-MB-231 cells induced ERK phosphorylation, with M4 showing a more dramatic response (fMLF and CXCL1 were used as positive controls) (**Figures S3 and 6A**). Importantly, we measured no stimulation with TCM derived from the ER+ MCF-7 cells (**Figures S3 and 6A**).

To identify the nature of the activating factors driving neutrophil migration, we first used multiplex ELISA to quantify the relative presence of potential neutrophil chemoattractants/activators associated with cancer, including a number of CXCL and CCL chemokines, cytokines, and growth factors (1, 66) in TCM derived from M4, MDA-MB-231, and ER+ MCF-7 cell lines (**Figure 6B**). Among the CXCL chemokines associated with breast cancer of ER- status (GRO, CXCL5, CXCL6, and CXCL8) and breast cancer metastasis (GRO, CXCL5, CXCL6, CXCL8, and CXCL12) (67, 68), we found a greater than 500-fold increase in GRO chemokines (CXCL1/2/3) present in the TCM harvested from TNBC cells, compared to TCM harvested from ER+ cells. This high amount of GRO secretion by TNBC cells is in agreement with the robust expression of CXCL1 mRNA in ER- breast tumor specimen compared to ER+ ones [(55) and **Figure S1**]. We detected a greater amount of CXCL8 present in the TCM from TNBC cells compared to the TCM from ER+ cells, which is in accordance with previous reports (53, 54). However, compared with GRO chemokines, the fold change in CXCL8 expression was more modest, suggesting that CXCL8 is not the key factor involved in neutrophil activation. In addition, we detected very little to no changes for CXCL5 and CXCL6 in any of the TCM. We also



measured > 9-fold increase in CXCL12 in the TCM derived from M4 and MDA-MB-231 cells, compared to MCF-7 TCM. In contrast, the CCL family members were in general less abundantly secreted by breast cancer cells. We detected a 5-fold

greater abundance of CCL2 in TCM from TNBC cells than ER+ cells, while very little to no CCL3 in any TCM. In contrast, CCL5 was secreted in equivalent amounts by all breast cancer cells, irrespective of the receptor expression status or malignant

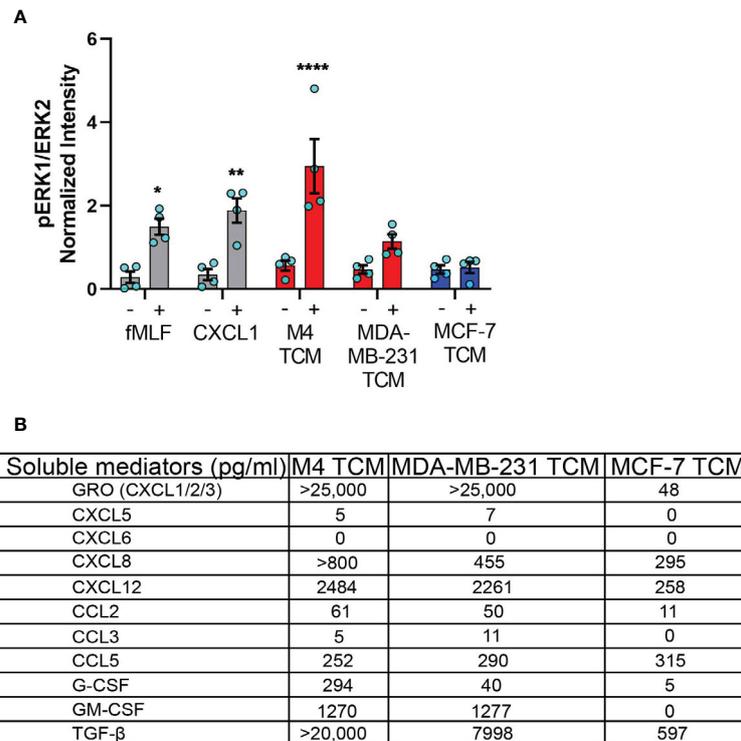


FIGURE 6 | TNBC cell lines secrete neutrophil activating factors more abundantly than ER⁺ cancer cell lines. **(A)** Graph depicting pERK1/ERK2 intensity in neutrophils stimulated with controls (fMLF, CXCL1, media) or equal volume of TCM derived from M4, MDA-MB-231, or MCF-7 cells. The quantified pERK1/2 band intensity was normalized to α -tubulin and is shown as mean \pm SEM from N=4 individual donors represented by each dot. ****P < 0.0001, **P \leq 0.01, *P \leq 0.05 when compared with corresponding vehicle controls (two-way ANOVA with Sidak multiple comparisons test). **(B)** Table showing the amount (pg/ml) of chemokines, growth factors and cytokines secreted by M4, MDA-MB-231 and MCF-7 cells determined by multiplex ELISA assay.

potential. Other than chemokines, we detected G-CSF and GM-CSF more abundantly in TCM harvested from TNBC cells compared to ER⁺ cells. Finally, we found that TGF- β 1 was 12-30 fold more abundant in TCM harvested from TNBC compared to ER⁺ cells. Taken together, these data show that compared to ER⁺ breast cancer cells, TNBC cells overexpress a number of potential neutrophil recruiting factors, the most abundant of which being the CXCR2 specific GRO chemokines and TGF- β 1.

Tumor Derived Active Factors Are of ~12 kDa Molecular Size

To narrow down our search for tumor-secreted active factors, we used a combination of biochemical and pharmacological approaches. We set out to fractionate the TCM based on the molecular weight of their components and identify the fraction with optimum activity. We subjected the concentrates (>3 kDa) of MDA-MB-231 TCM to size exclusion chromatography (Figure 7A) and tested the resulting fractions for their ability to induce polarized morphology in neutrophils. We found that the fractions carrying proteins corresponding to ~12 kDa molecular weight (fraction “g” in Figure 7A) induced maximum polarization as indicated by the elongated shape of the cells (Figure 7Biv). In contrast, most of the cells retained a

circular shape when stimulated with fractions containing proteins that were either larger (Figure 7Bii&iii) or smaller than ~12 kDa (Figure 7Bv&vi). Indeed, when we quantified the extent of polarization induced by a series of fractions, we found that circularity was significantly reduced when exposed to MDA-MB-231 TCM itself or TCM-derived fractions corresponding to 12 kDa sized proteins (Figure 7C, fractions e,f,g,h). A similar pattern of polarized morphology was also induced by M4 TCM derived fractions containing ~12 kDa sized proteins (Figure S5). Importantly, both GRO chemokines (69) and monomeric TGF- β (70) have a molecular weight of 12 kDa.

Chemokines activate G protein-coupled receptors (GPCR) by signaling through G_i heterotrimeric G proteins, which are sensitive to pertussis toxin (PT) inhibition (71). To determine the role of chemokines in TCM induced neutrophil migration, we therefore, targeted chemokine specific GPCR G_i signaling by pretreating neutrophils with PT and measured the ability of cells to migrate towards TCM or CXCL8 as a positive control. Using transwell assays, we found that significantly fewer neutrophils migrated towards TCM derived from either M4 or MDA-MB-231 cells in the presence of PT (Figure 7D), indicating a role for chemokines in TCM-induced neutrophil recruitment. PT

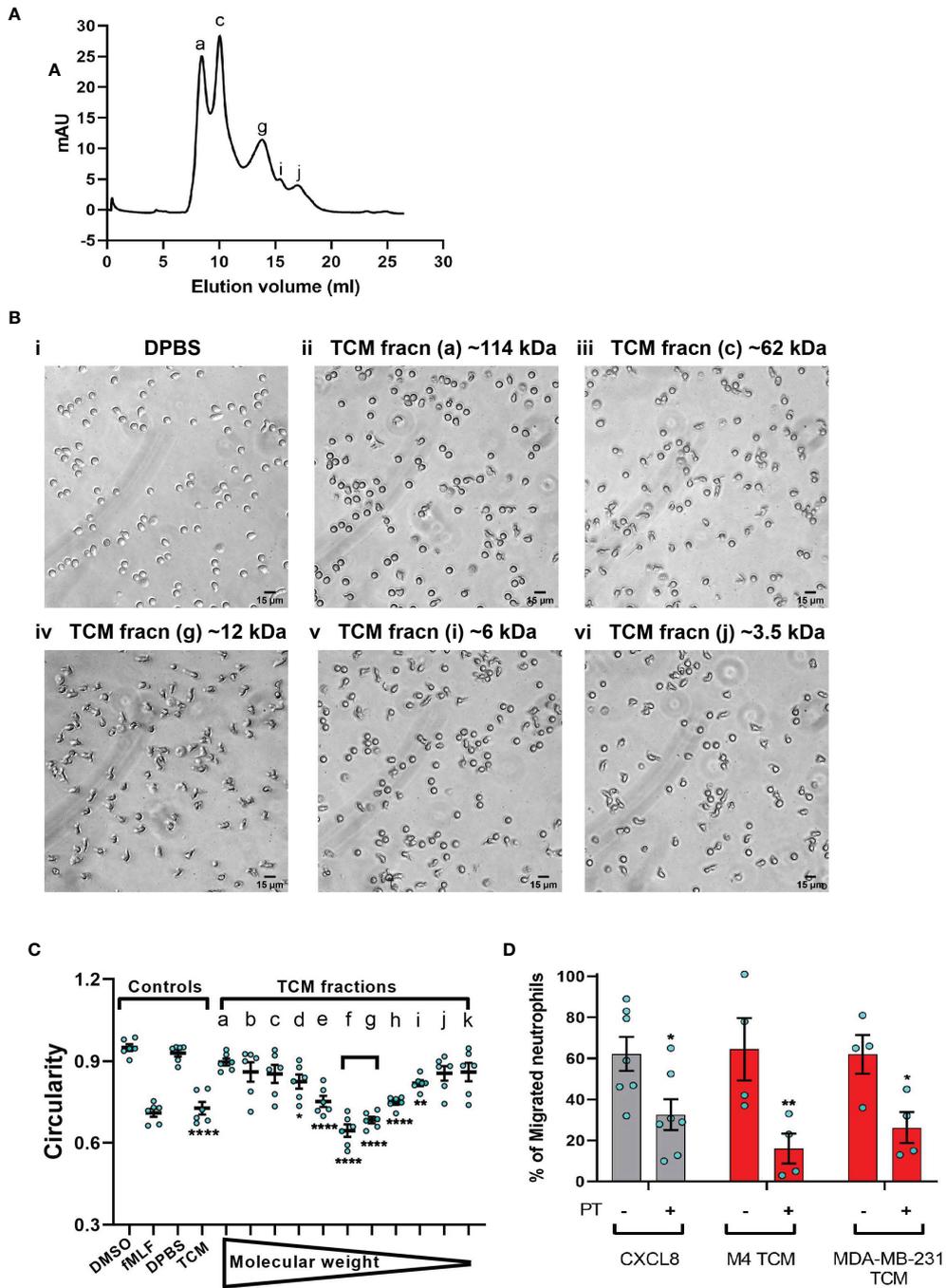


FIGURE 7 | MDA-MB-231-derived active factors are ~12 kDa. **(A)** Representative size exclusion chromatography elution profile of TCM from MDA-MB-231 cells. **(B)** Representative bright field images of neutrophils exposed to vehicle control DPBS (Bi) or TCM peak fractions (fracn (Bii-vi) as labeled in **(A)**) and corresponding to different molecular weights. **(C)** Graph depicting the circularity of neutrophils stimulated with controls (DMSO, fMLF, DPBS, TCM) or equal volume of TCM fractions with gradually decreasing molecular weight. Data show mean \pm SEM from N=3 individual donors with two dots for each donor representing mean circularity of all neutrophils in two randomly selected imaging fields. **** $P < 0.0001$, ** $P \leq 0.01$, * $P \leq 0.05$ when compared with DPBS (one-way ANOVA with Dunnett's multiple comparisons test). **(D)** Graph depicting the percentage of neutrophils that migrated into the bottom chamber of the transwells containing equal volume of TCM or 20 nM CXCL8 in the presence or absence of PT. Bars show mean \pm SEM from N=4-7 individual donors represented by each dot. ** $P \leq 0.01$, * $P \leq 0.05$ when compared with corresponding PT- values (two-way ANOVA with Sidak multiple comparisons test).

treatment was not toxic, as it did not alter the migration of neutrophils towards the growth factor GM-CSF (Figure S4). Taken together, these data suggest that chemokines actively contribute to TCM induced neutrophil migration.

The Neutrophil Recruiting Activity of the TCM Depends on the Combined Action of GRO Chemokines and TGF-β

In order to specifically determine the identity of the mediators, we studied TCM induced neutrophil migration in the presence of different inhibitors targeting either the GRO receptor CXCR2 [AZD 5069 (72)] or the TGF-β1 receptor ALK5 [SB431542 (73)],

or both. The presence of both inhibitors did not alter neutrophil migration towards fMLF, showing that the inhibitors are receptor specific (Figure 8A). We found that neutrophil migration induced by M4 TCM remained intact when cells were pretreated with either inhibitor alone (Figure 8A). In contrast, we noted a significant decrease (~38%) in the percentage of migrating neutrophils towards M4 TCM when cells were pretreated with both inhibitors, indicating that CXCR2 ligands and TGF-β mediate the effects of the TCM (Figure 8A). Similarly, we found a significant inhibition of M4 TCM induced neutrophil polarization as reflected by the increase in circularity in the presence of both the inhibitors compared to TCM alone

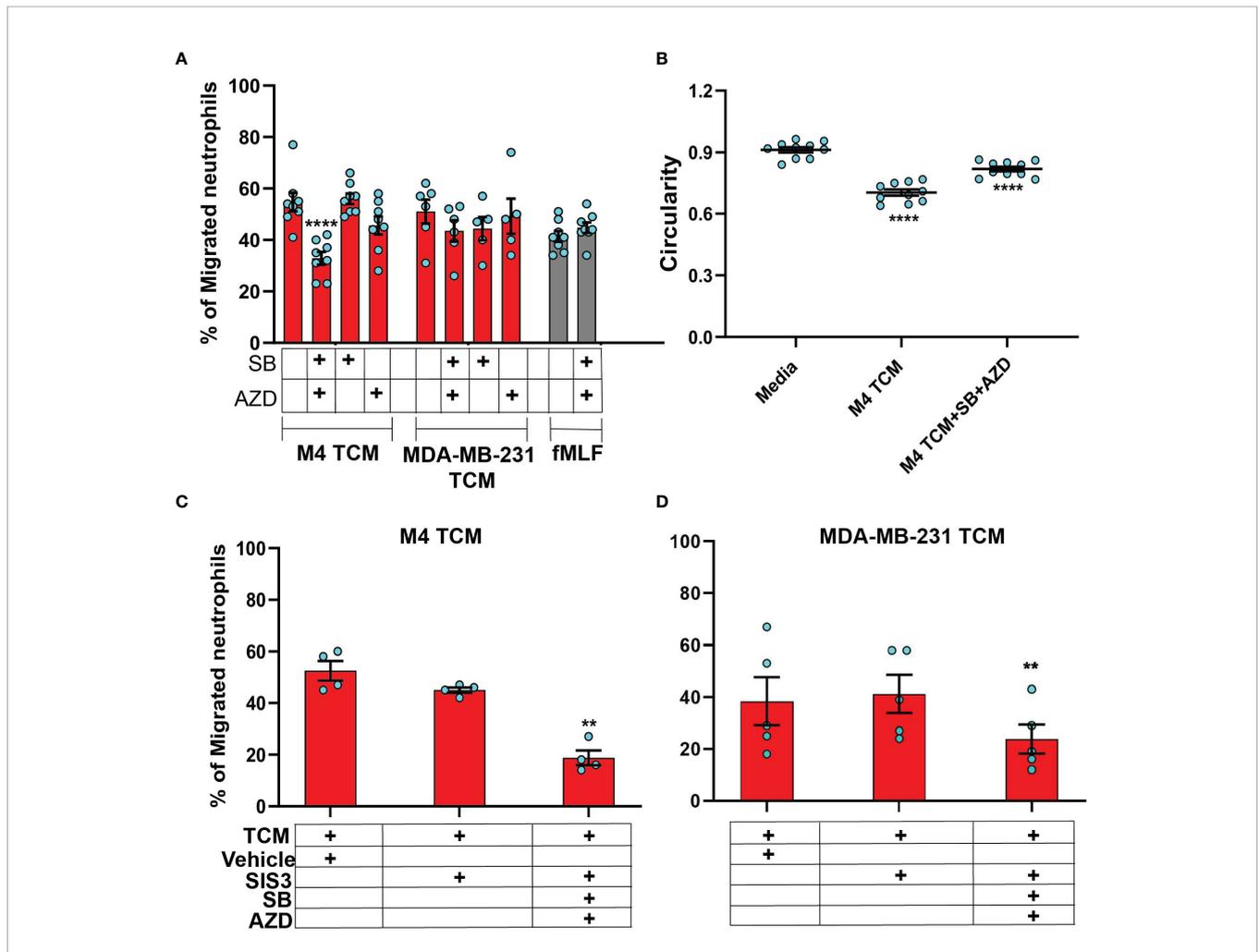


FIGURE 8 | The neutrophil recruiting activity of the TCM depends on the combined action of GRO chemokines and TGF-β. **(A)** Graph depicting the percentage of neutrophils either untreated or pretreated with 1 μM SB, 1 μM AZD, or a combination of 1 μM of SB and 1 μM AZD that migrated into the bottom chamber of the transwells containing equal volume of M4 TCM, MDA-MB-231 TCM, or 100 nM fMLF. Bars show mean ± SEM from N=5-8 donors with each dot representing individual donor. **(B)** Graph depicting the circularity of neutrophils exposed to M4 TCM in the presence or absence of a combination of SB and AZD, or with media control. Bars show mean ± SEM from N=3 donors with 3-4 dots for each donor representing mean circularity of all neutrophils from at least 3 randomly selected imaging fields. **(C, D)** Graphs depicting the percentage of neutrophils migrated into the bottom chamber of the transwells containing equal volume of M4 TCM **(C)** or MDA-MB-231 TCM **(D)** in the presence of 3 μM SIS3 alone or a combination of 1 μM each of SB, AZD and 3 μM SIS3, or vehicle control. Bars show mean ± SEM from N=4 **(C)** or 5 **(D)** donors. ****P < 0.0001 when compared with untreated **(A)**, media control **(B)** (one-way ANOVA with Dunnett’s multiple comparisons test). **P ≤ 0.01 when compared with vehicle control **(C, D)** (paired sample t-test).

(**Figure 8B**). However, neutrophil migration induced by MDA-MB-231 derived TCM was not significantly decreased by the inhibitors (**Figure 8A**).

To further elucidate the role of TGF- β signaling in mediating the effect of TCM on neutrophils, we targeted SMAD3, a downstream signaling component of TGF- β signal transduction pathway (74). Neutrophils pretreated with SIS3 [an inhibitor that specifically inhibits SMAD3 phosphorylation (75)] showed intact migration in response to M4 TCM (**Figure 8C**). However, when cells were pretreated with inhibitors targeting CXCR2, TGF- β receptor, and SMAD3 phosphorylation, we observed a remarkably stronger inhibition (~60%) of neutrophil migration towards M4 TCM (**Figure 8C**) than the inhibition (~38%) measured with CXCR2 and TGF- β receptor inhibitors (**Figure 8A**). Importantly, we also found a significant inhibition (~40%) of neutrophil migration towards MDA-MB-231 TCM in the presence of all three inhibitors, compared to vehicle control (**Figure 8D**). Together, these results indicate that a concerted action of secreted TGF- β and GRO specific chemokines plays a key role in TCM induced neutrophil recruitment.

DISCUSSION

The purpose of this study was to determine whether neutrophil recruitment in breast tumors is dictated by the malignant potential of breast cancer cells. A number of retrospective studies have reported a unique association of neutrophils with TNBC compared to HR+ subtypes (16, 76, 77). Using a panel of breast cancer cell lines representing highly aggressive TNBC or poorly aggressive ER+ breast cancer subtypes, we studied tumor driven neutrophil migration in three distinct migration assay systems. We showed a direct causal relationship between the metastatic potential of human breast cancer cells and their ability to recruit neutrophils, and conclude that TNBC subtypes induce robust neutrophil migration by secreting soluble chemical cues. More importantly, we profiled tumor secreted chemical cues with a number of biochemical, pharmacological and immunological approaches and conclude that a unique combination of GRO chemokines and TGF- β are responsible for driving neutrophil recruitment to breast tumor.

The primary function of GRO is to guide neutrophil to infection or tissue injury sites. The GRO member CXCL1 has also been implicated to recruit neutrophils in tumor mouse models (52, 78). We measured massive amounts of GRO family chemokines in TCM derived from TNBC cells compared to ER+ breast cancer cells. Yet, surprisingly, we found that TCM activity remains intact after targeting the GRO receptor CXCR2. While GRO chemokines are potent agonists of CXCR2, they can also bind and activate CXCR1, although they have lower affinity towards CXCR1 (79). Furthermore, the rate of ligand induced CXCR1 internalization from the cell surface is relatively slower than that for CXCR2 (80). Therefore, a potential role of GRO-CXCR1 signaling in

mediating the neutrophil recruitment activity of TCM derived from TNBC cell lines cannot be excluded.

Factor/s other than chemokines may also play a key role to mediate neutrophil migration. In addition to GRO chemokines, we detected considerable amount of TGF- β 1 cytokine in the TCM from both M4 and TNBC cell lines. The elevated presence of TGF- β in the tumor niche has been shown to facilitate tumor progression and metastasis in different cancer types (81, 82). However, TGF- β lacks direct chemoattracting ability for neutrophils (83), which we also verified using recombinant TGF- β in a transwell setup (data not shown). Interestingly, TGF- β was recently shown to promote tumor-permissive phenotypes in TANs (84). The same study also reported that TGF- β inhibition leads to massive infiltration of cytotoxic neutrophils in tumor-bearing mice. When we inhibited the TGF- β receptor T β RI/type I receptor ALK5, neutrophil migration induced by TCM remained intact. However, targeting both T β RI and CXCR2 led to a significant defect in neutrophil recruitment by M4 TCM, but not MDA-MB-231 TCM. Earlier studies identified a novel role of tumor-derived TGF- β in modulating chemokine receptor expression on immune cells (85). Notably, tumor secreted factors were recently shown to increase expression of T β RI on neutrophils (86). In line with these previous findings, we envision that TGF- β indirectly promotes TCM induced neutrophil migration through amplification of chemokine signaling. A key mediator in the canonical TGF- β signaling pathway that is critical for TGF- β 1-triggered transcriptional regulation is Smad3 (74). We found that addition of the Smad3 inhibitor disrupted the activity of both M4 and MDA-MB-231 TCM, supporting the notion that TGF- β signaling indirectly promotes neutrophil migration induced by chemokines isolated from TNBC cell lines. We envision that the different amounts of GRO chemokines and TGF- β in M4 and MDA-MB-231 TCM is responsible for the distinct sensitivity between the two TCM. It remains to be determined how TGF- β and chemokine signaling pathways regulate neutrophil migration.

While the transwell migration assay is ideal to quantify chemoattracting ability of multiple conditions simultaneously, it lacks the scope to directly monitor migrating cells and assess directionality of cell movement- a true indicator of chemotaxis. The neutrophil recruiting ability of tumor secreted factors collected from lung cancer cell lines has been reported using the underagarose assay (87), which allows to visualize and quantify cell migration (36). However, we observed no neutrophil migration in response to M4 and MDA-MB-231-derived TCM using the underagarose setup (unpublished). The varying TCM content and diffusability of the secreted factors through the agarose could explain this discrepancy. We did find that the M4 TCM induced neutrophil chemotaxis in 3D collagen matrices that closely mimics interstitial space *in vivo*. By tracking migrating neutrophils, we showed that M4 TCM induced neutrophils to migrate with increased velocity and directionality, compared to media control. Using a co-culture system, we also showed that neutrophils migrate directionally toward M4 spheroids, while we measured no directional

migration toward spheroids in a neutrophil-BT474 spheroid co-culture system. Cancer cells grown as 3D spheroids closely mimic the complexity of cell-cell, cell-matrix interactions, and gene expression profiles of the 3D tumor mass *in vivo* (88). Others have shown neutrophil infiltration into lung tumor spheroids using endpoint imaging of tumor sections (89). To our knowledge, our study is the first to demonstrate in real time that tumor spheroids actively recruit neutrophils. Whether neutrophils further amplify their chemotactic migration towards tumor spheroids through the secretion of secondary chemoattractants such as leukotriene B₄ or other chemokines in response to tumor-derived factors remains to be addressed (24, 90). Notably, we found that more neutrophils were present at the 20 hr endpoint in the vicinity of M4 spheroids, compared to BT474 spheroids. Neutrophils have very short life spans, however recent studies point to the role of pro-inflammatory soluble mediators in extending neutrophil survival (64, 91). Indeed, many of the soluble factors including CXCL8, G-CSF, and GM-CSF, secreted abundantly from the M4 and MDA-MB-231 metastatic breast cancer cells, are known to have pro-survival effect on neutrophils (64, 91). Hence, we envisage that the pro-survival effect of M4 secreted factors also underlie the persistence of more neutrophils near M4 spheroids.

Much remains to be determined about the mechanisms underlying the secretion of active factors by highly malignant breast cancer cells. Others have shown a critical role of the epithelial-to-mesenchymal transition (EMT) program in promoting tumor metastasis (92, 93). Through EMT epithelial cancer cells acquire mesenchymal features with enhanced invading properties that aid in cancer cell dissemination. Notably, myeloid cell recruitment was recently detected in EMT marker positive TNBC breast cancer specimen indicating possible role of EMT in tumor-driven neutrophil recruitment (94). Moreover, EMT associated changes were connected to increased secretion of inflammatory soluble factors including CXCL8 from breast cancer cell lines (94). Future studies will determine whether turning on EMT also enables breast cancer cells to secrete neutrophil recruiting GRO chemokines and TGF- β . Recently, the release of tumor-derived extracellular vesicles (EVs) have been suggested as a mechanism for the secretion of diverse soluble factors including chemokines and TGF- β (95, 96). Whether similar mechanisms underlie the secretion of active factors by highly malignant breast cancer cells is the topic of future studies.

In conclusion, findings of this study complement previously established role of CXCL chemokines in mediating neutrophil recruitment to tumors, and expand our current understanding by directly linking malignant potential of breast cancer cells with their neutrophil recruiting ability. It is also the first study to unveil a novel role of tumor-derived TGF- β to cooperate with GRO chemokines and orchestrate neutrophil recruitment to TNBC tumors. Future studies will aim to understand how TCM impact the previously described pro-metastasis phenotypes of neutrophils (3, 9–14, 17, 97). Knowledge from this study may help develop novel therapeutic strategies to target neutrophil recruitment to tumors and TAN function, which

would restrict metastatic spread without perturbing protective functions of neutrophils against infection and injury.

DATA AVAILABILITY STATEMENT

The original contributions presented in the study are included in the article/**Supplementary Material**. Further inquiries can be directed to the corresponding author.

AUTHOR CONTRIBUTIONS

SS, LH, and CP conceptualized and designed the study. SS, LH, YX, JZ, JK, and YL performed the experiments, analyzed, and interpreted the data. CJ analyzed and interpreted the data. SS and LH drafted the manuscript. SS, LH, and CP critically revised the manuscript. DC, JS, and CP supervised the study. All authors reviewed the manuscript. All authors contributed to the article and approved the submitted version.

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SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: <https://www.frontiersin.org/articles/10.3389/fimmu.2021.659996/full#supplementary-material>

Figure S1 | The expression of CXCL1, CXCL2, CXCL5, and CXCL8 is elevated in breast cancer specimens with low ESR1 levels. **(A)** Representative heatmap from the Farmer study [40] with ESR1 and CXCL chemokines annotated with specific probeset ID. Datasets identified from the OncoPrint platform were used to compare CXCL transcript expression between ER+ (blue) and ER- (red) breast cancer

specimens sorted by ESR1 expression levels. The heatmap was generated by plotting log₂ fold change of expression values relative to averaged ER+ for ESR1 and each of the CXCL chemokines. Blue and red indicate high and low expression, respectively. **(B)** Heatmap generated by plotting log₂ fold change of averaged ER- relative to averaged ER+ for ESR1 and each of the CXCL chemokines calculated from the datasets of N=7 independent studies (40–46). **(C)**, Summary of fold changes for CXCL1, CXCL2, and CXCL8 between ER- and ER+ with the adjusted P value from each of the 7 studies. A fold change of 1.5 or greater and an adjusted P-value of 0.05 or less was considered statistically significant. **(D)**, Datasets from TCGA-BRCA were analyzed for ESR1 and CXCL chemokine expression. The log₂ of the FPKM values were plotted for the chemokines and ordered by the FPKM of ESR1.

Figure S2 | TNBC cell lines have greater migration and invasion ability than ER+ breast cancer cells. Graph depicting the migration **(A)** or invasion **(B)** potential of a panel of TNBC and ER+ cell lines. Results are depicted as mean ± SEM from N=3-5 independent experiments represented by each dot. ****P<0.0001, **P<0.01 when compared with values of MCF-7 cell line (one-way ANOVA with Dunnett's multiple comparisons test).

Figure S3 | Active factors secreted from TNBC cell lines induce robust ERK phosphorylation. Representative immunoblots depicting the levels of phospho-ERK and α-tubulin in response to fMLF, CXCL1, and TCM derived from M4, MDA-MB-231, and MCF-7 cells. Blot is representatives of four independent experiments.

Figure S4 | Treatment with PT specifically inhibits Gai dependent signaling. **(A, B)** Representative fluorescent endpoint images of neutrophils migrating under agarose towards a gradient 50 µg/ml GM-CSF in the absence or presence of pre-treatment with 500 ng/ml PT for 2 hrs. **(C, D)** Representative fluorescent endpoint images of neutrophils migrating towards a gradient 100 nM fMLF in the absence or presence of pre-treatment with 500 ng/ml PT for 2 hrs. Images are representative of at least

N=3 independent donors and acquired using an Axiovert fluorescence microscope from the side of the wells facing the chemoattractants (see *Methods*).

Figure S5 | M4-derived active factors are ~12 kDa. **(A)** Representative size exclusion chromatography elution profile of TCM from M4 cells. **(B)** Representative bright field images of neutrophils exposed to vehicle control DPBS **(Bi)** or TCM peak fractions (fracn) **(Bii-vi)** as labeled in **(A)** and corresponding to different molecular weights. **(C)** Graph depicting the circularity of neutrophils stimulated with controls (fMLF, DPBS, TCM) or TCM fractions with gradually decreasing molecular weight. Data show mean ± SD from N=1 donor with two dots representing mean circularity of all neutrophils from two randomly selected imaging fields.

Movies S1 and S2 | Videos showing time-lapse imaging of labeled neutrophils migrating in 3D collagen matrices in response to M4 TCM (S1) or media (S2) placed on the right. Images were taken every 25 sec over 40 min. Individual cell tracks in the immediate vicinity of TCM-collagen boundary are shown.

Movies S3 and S4 | Videos showing time-lapse imaging of labeled neutrophils migrating in 3D collagen matrices in response to 500 nM CXCL8 (S3) or buffer (S4) placed on the right. Images were taken every 25 sec over 50 min. Individual cell tracks in the vicinity of TCM-collagen boundary are shown.

Movies S5–S8 | Videos showing z-stacks of 2-photon images of M4 (Total slice no. 38; S5), MDA-MB-231 (Total slice no. 23; S6), BT549 (Total slice no. 31; S7), and BT474 (Total slice no. 147; S8) spheroids stained with a nuclear dye DRAQ5.

Movies S9 and S10 | Videos showing time-lapse imaging of the neutrophil-M4 (S9) and -BT474 (S10) spheroid co-culture system. Live-imaging was initiated one hour after the addition of neutrophils to the spheroids. Images were taken every 3 min for 20 hrs. Individual neutrophil tracks are shown.

REFERENCES

- SenGupta S, Subramanian BC, Parent CA. Getting TANNed: How the tumor microenvironment drives neutrophil recruitment. *J Leukocyte Biol* (2019) 105 (3):449–62. doi: 10.1002/JLB.3RI0718-282R
- Liew PX, Kubers P. The Neutrophil's Role During Health and Disease. *Physiol Rev* (2019) 99(2):1223–48. doi: 10.1152/physrev.00012.2018
- Wculek SK, Malanchi I. Neutrophils support lung colonization of metastasis-initiating breast cancer cells. *Nature* (2015) 528(7582):413–7. doi: 10.1038/nature16140
- Park J, Wysocki RW, Amoozgar Z, Maiorino L, Fein MR, Jorns J, et al. Cancer cells induce metastasis-supporting neutrophil extracellular DNA traps. *Sci Trans Med* (2016) 8(361):361ra138. doi: 10.1126/scitranslmed.aag1711
- Zhou SL, Zhou ZJ, Hu ZQ, Huang XW, Wang Z, Chen EB, et al. Tumor-Associated Neutrophils Recruit Macrophages and T-Regulatory Cells to Promote Progression of Hepatocellular Carcinoma and Resistance to Sorafenib. *Gastroenterology* (2016) 150(7):1646–58. doi: 10.1053/j.gastro.2016.02.040
- Singhal S, Bhojnagarwala PS, O'Brien S, Moon EK, Garfall AL, Rao AS, et al. Origin and Role of a Subset of Tumor-Associated Neutrophils with Antigen-Presenting Cell Features in Early-Stage Human Lung Cancer. *Cancer Cell* (2016) 30(1):120–35. doi: 10.1016/j.ccell.2016.06.001
- Berry RS, Xiong MJ, Greenbaum A, Mortaji P, Nofchissey RA, Schultz F, et al. High levels of tumor-associated neutrophils are associated with improved overall survival in patients with stage II colorectal cancer. *PLoS One* (2017) 12 (12):e0188799. doi: 10.1371/journal.pone.0188799
- Granot Z, Henke E, Comen EA, King TA, Norton L, Benezra R. Tumor entrained neutrophils inhibit seeding in the premetastatic lung. *Cancer Cell* (2011) 20(3):300–14. doi: 10.1016/j.ccr.2011.08.012
- Coffelt SB, Kersten K, Doornebal CW, Weiden J, Vrijland K, Hau CS, et al. IL-17-producing γδ T cells and neutrophils conspire to promote breast cancer metastasis. *Nature* (2015) 522(7556):345–8. doi: 10.1038/nature14282
- Liang W, Li Q, Ferrara N. Metastatic growth instructed by neutrophil-derived transferrin. *Proc Natl Acad Sci USA* (2018) 115(43):11060–5. doi: 10.1073/pnas.1811717115
- Sasaki S, Baba T, Muranaka H, Tanabe Y, Takahashi C, Matsugo S, et al. Involvement of Prokineticin 2-expressing Neutrophil Infiltration in 5-Fluorouracil-induced Aggravation of Breast Cancer Metastasis to Lung. *Mol Cancer Ther* (2018) 17(7):1515–25. doi: 10.1158/1535-7163.MCT-17-0845
- Donati K, Sépult C, Rocks N, Blacher S, Gérard C, Noel A, et al. Neutrophil-Derived Interleukin 16 in Premetastatic Lungs Promotes Breast Tumor Cell Seeding. *Cancer Growth Metastasis* (2017) 10:1179064417738513. doi: 10.1177/1179064417738513
- Queen MM, Ryan RE, Holzer RG, Keller-Peck CR, Jorczyk CL. Breast cancer cells stimulate neutrophils to produce oncostatin M: potential implications for tumor progression. *Cancer Res* (2005) 65(19):8896–904. doi: 10.1158/0008-5472.CAN-05-1734
- Kowanetz M, Wu X, Lee J, Tan M, Hagenbeek T, Qu X, et al. Granulocyte-colony stimulating factor promotes lung metastasis through mobilization of Ly6G+Ly6C+ granulocytes. *Proc Natl Acad Sci USA* (2010) 107(50):21248–55. doi: 10.1073/pnas.1015855107
- Shen M, Hu P, Donskov F, Wang G, Liu Q, Du J. Tumor-associated neutrophils as a new prognostic factor in cancer: a systematic review and meta-analysis. *PLoS One* (2014) 9(6):e98259. doi: 10.1371/journal.pone.0098259
- Soto-Perez-de-Celis E, Chavarri-Guerra Y, Leon-Rodriguez E, Gamboa-Dominguez A. Tumor-Associated Neutrophils in Breast Cancer Subtypes. *Asian Pac J Cancer Prev APJCP* (2017) 18(10):2689–93. doi: 10.22034/APJCP.2017.18.10.2689
- Wang Y, Chen J, Yang L, Li J, Wu W, Huang M, et al. Tumor-Contacted Neutrophils Promote Metastasis by a CD90-TIMP-1 Juxtacrine-Paracrine Loop. *Clin Cancer Res an Off J Am Assoc Cancer Res* (2019) 25(6):1957–69. doi: 10.1158/1078-0432.CCR-18-2544
- Fragomeni SM, Sciallis A, Jeruss JS. Molecular Subtypes and Local-Regional Control of Breast Cancer. *Surg Oncol Clinics North A* (2018) 27(1):95–120. doi: 10.1016/j.soc.2017.08.005
- Nasrazadani A, Thomas RA, Oesterreich S, Lee AV. Precision Medicine in Hormone Receptor-Positive Breast Cancer. *Front Oncol* (2018) 8:144:144. doi: 10.3389/fonc.2018.00144
- Yin L, Duan JJ, Bian XW, Yu SC. Triple-negative breast cancer molecular subtyping and treatment progress. *Breast Cancer Res BCR* (2020) 22(1):61. doi: 10.1186/s13058-020-01296-5

21. Schmid P, Adams S, Rugo HS, Schneeweiss A, Barrios CH, Iwata H, et al. Atezolizumab and Nab-Paclitaxel in Advanced Triple-Negative Breast Cancer. *New Engl J Med* (2018) 379(22):2108–21. doi: 10.1056/NEJMoa1809615
22. Shaul ME, Fridlender ZG. Neutrophils as active regulators of the immune system in the tumor microenvironment. *J Leukocyte Biol* (2017) 102(2):343–9. doi: 10.1189/jlb.5MR1216-508R
23. Yang Q, Guo N, Zhou Y, Chen J, Wei Q, Han M. The role of tumor-associated macrophages (TAMs) in tumor progression and relevant advance in targeted therapy. *Acta Pharm Sin B* (2020) 10(11):2156–70. doi: 10.1016/j.apsb.2020.04.004
24. Subramanian BC, Majumdar R, Parent CA. The role of the LTB(4)-BLT1 axis in chemotactic gradient sensing and directed leukocyte migration. *Semin Immunol* (2017) 33:16–29. doi: 10.1016/j.smim.2017.07.002
25. Ali S, Lazennec G. Chemokines: novel targets for breast cancer metastasis. *Cancer Metastasis Rev* (2007) 26(3–4):401–20. doi: 10.1007/s10555-007-9073-z
26. Yu PF, Huang Y, Han YY, Lin LY, Sun WH, Rabson AB, et al. TNF α -activated mesenchymal stromal cells promote breast cancer metastasis by recruiting CXCR2 (+) neutrophils. *Oncogene* (2017) 36(4):482–90. doi: 10.1038/ncr.2016.217
27. Zhou SL, Dai Z, Zhou ZJ, Chen Q, Wang Z, Xiao YS, et al. CXCL5 contributes to tumor metastasis and recurrence of intrahepatic cholangiocarcinoma by recruiting infiltrative intratumoral neutrophils. *Carcinogenesis* (2014) 35(3):597–605. doi: 10.1093/carcin/bgt397
28. Li Y, Yang D, Yin X, Zhang X, Huang J, Wu Y, et al. Clinicopathological Characteristics and Breast Cancer-Specific Survival of Patients With Single Hormone Receptor-Positive Breast Cancer. *JAMA Network Open* (2020) 3(1):e1918160–e. doi: 10.1001/jamanetworkopen.2019.18160
29. Cui X, Schiff R, Arpino G, Osborne CK, Lee AV. Biology of Progesterone Receptor Loss in Breast Cancer and Its Implications for Endocrine Therapy. *J Clin Oncol* (2005) Oct 2023(30):7721–35. doi: 10.1200/JCO.2005.09.004
30. Allen SG, Chen YC, Madden JM, Fournier CL, Altemus MA, Hiziroglu AB, et al. Macrophages Enhance Migration in Inflammatory Breast Cancer Cells via RhoC GTPase Signaling. *Sci Rep* (2016) 6:39190. doi: 10.1038/srep39190
31. Boyum A. Separation of blood leukocytes, granulocytes and lymphocytes. *Tissue Antigens* (1974) 4(4):269–74. doi: 10.1111/j.1399-0039.1974.tb00252.x
32. Subramanian BC, Moissoglu K, Parent CA. The LTB(4)-BLT1 axis regulates the polarized trafficking of chemoattractant GPCRs during neutrophil chemotaxis. *J Cell Sci* (2018) 131(18):jcs217422. doi: 10.1242/jcs.217422
33. Berens EB, Holy JM, Riegel AT, Wellstein A. A Cancer Cell Spheroid Assay to Assess Invasion in a 3D Setting. *J Visualized Exp JoVE* (2015), 53409. doi: 10.3791/53409
34. Zhu X, Huang L, Zheng Y, Song Y, Xu Q, Wang J, et al. Ultrafast optical clearing method for three-dimensional imaging with cellular resolution. *Proc Natl Acad Sci USA* (2019) 116(23):11480–9. doi: 10.1073/pnas.1819583116
35. Roossien DH, Sadis BV, Yan Y, Webb JM, Min LY, Dizaji AS, et al. Multispectral tracing in densely labeled mouse brain with nTracer. *Bioinf (Oxford England)* (2019) 35(18):3544–6. doi: 10.1093/bioinformatics/bt084
36. Saunders CA, Majumdar R, Molina Y, Subramanian BC, Parent CA. Genetic manipulation of PLB-985 cells and quantification of chemotaxis using the underagarose assay. *Methods Cell Biol* (2019) 149:31–56. doi: 10.1016/b.smb.2018.09.002
37. Sixt M, Lämmermann T. In vitro analysis of chemotactic leukocyte migration in 3D environments. *Methods Mol Biol (Clifton NJ)* (2011) 769:149–65. doi: 10.1007/978-1-61779-207-6_11
38. Zengel P, Nguyen-Hoang A, Schildhammer C, Zantl R, Kahl V, Horn E. μ -Slide Chemotaxis: a new chamber for long-term chemotaxis studies. *BMC Cell Biol* (2011) 12:21. doi: 10.1186/1471-2121-12-21
39. Niggli V. Microtubule-disruption-induced and chemotactic-peptide-induced migration of human neutrophils: implications for differential sets of signalling pathways. *J Cell Sci* (2003) 116(Pt 5):813–22. doi: 10.1242/jcs.00306
40. Farmer P, Bonnefoi H, Becette V, Tubiana-Hulin M, Fumoleau P, Larsimont D, et al. Identification of molecular apocrine breast tumours by microarray analysis. *Oncogene* (2005) 24(29):4660–71. doi: 10.1038/sj.onc.1208561
41. Minn AJ, Gupta GP, Siegel PM, Bos PD, Shu W, Giri DD, et al. Genes that mediate breast cancer metastasis to lung. *Nature* (2005) 436(7050):518–24. doi: 10.1038/nature03799
42. Wang Y, Klijn JG, Zhang Y, Sieuwerts AM, Look MP, Yang F, et al. Gene-expression profiles to predict distant metastasis of lymph-node-negative primary breast cancer. *Lancet (London England)* (2005) 365(9460):671–9. doi: 10.1016/S0140-6736(05)17947-1
43. Ivshina AV, George J, Senko O, Mow B, Putti TC, Smeds J, et al. Genetic reclassification of histologic grade delineates new clinical subtypes of breast cancer. *Cancer Res* (2006) 66(21):10292–301. doi: 10.1158/0008-5472.CAN-05-4414
44. Miyake T, Nakayama T, Naoi Y, Yamamoto N, Otani Y, Kim SJ, et al. GSTP1 expression predicts poor pathological complete response to neoadjuvant chemotherapy in ER-negative breast cancer. *Cancer Sci* (2012) 103(5):913–20. doi: 10.1111/j.1349-7006.2012.02231.x
45. Schuetz CS, Bonin M, Clare SE, Nieselt K, Sotlar K, Walter M, et al. Progression-specific genes identified by expression profiling of matched ductal carcinomas in situ and invasive breast tumors, combining laser capture microdissection and oligonucleotide microarray analysis. *Cancer Res* (2006) 66(10):5278–86. doi: 10.1158/0008-5472.CAN-05-4610
46. Kao KJ, Chang KM, Hsu HC, Huang AT. Correlation of microarray-based breast cancer molecular subtypes and clinical outcomes: implications for treatment optimization. *BMC Cancer* (2011) 11:143. doi: 10.1186/1471-2407-11-143
47. Irizarry RA, Hobbs B, Collin F, Beazer-Barclay YD, Antonellis KJ, Scherf U, et al. Exploration, normalization, and summaries of high density oligonucleotide array probe level data. *Biostatistics* (2003) 4(2):249–64. doi: 10.1093/biostatistics/4.2.249
48. Ritchie ME, Phipson B, Wu D, Hu Y, Law CW, Shi W, et al. limma powers differential expression analyses for RNA-sequencing and microarray studies. *Nucleic Acids Res* (2015) 43(7):e47. doi: 10.1093/nar/gkv007
49. Kaunisto A, Henry WS, Montaser-Kouhsari L, Jaminet SC, Oh EY, Zhao L, et al. NFAT1 promotes intratumoral neutrophil infiltration by regulating IL8 expression in breast cancer. *Mol Oncol* (2015) 9(6):1140–54. doi: 10.1016/j.molonc.2015.02.004
50. Dominguez C, McCampbell KK, David JM, Palena C. Neutralization of IL-8 decreases tumor PMN-MDSCs and reduces mesenchymalization of claudin-low triple-negative breast cancer. *JCI Insight* (2017) 2(21):e94296. doi: 10.1172/jci.insight.94296
51. Yu PF, Huang Y, Han YY, Lin LY, Sun WH, Rabson AB, et al. TNF α -activated mesenchymal stromal cells promote breast cancer metastasis by recruiting CXCR2+ neutrophils. *Oncogene* (2017) 36(4):482–90. doi: 10.1038/ncr.2016.217
52. Acharyya S, Oskarsson T, Vanharanta S, Malladi S, Kim J, Morris PG, et al. A CXCL1 paracrine network links cancer chemoresistance and metastasis. *Cell* (2012) 150(1):165–78. doi: 10.1016/j.cell.2012.04.042
53. Lin Y, Huang R, Chen L, Li S, Shi Q, Jordan C, et al. Identification of interleukin-8 as estrogen receptor-regulated factor involved in breast cancer invasion and angiogenesis by protein arrays. *Int J Cancer* (2004) 109(4):507–15. doi: 10.1002/ijc.11724
54. Freund A, Chauveau C, Brouillet J-P, Lucas A, Lacroix M, Licznar A, et al. IL-8 expression and its possible relationship with estrogen-receptor-negative status of breast cancer cells. *Oncogene* (2003) 22(2):256–65. doi: 10.1038/sj.onc.1206113
55. Yang C, Yu H, Chen R, Tao K, Jian L, Peng M, et al. CXCL1 stimulates migration and invasion in ER-negative breast cancer cells via activation of the ERK/MMP2/9 signaling axis. *Int J Oncol* (2019) 55(3):684–96. doi: 10.3892/ijo.2019.4840
56. Dawson PJ, Wolman SR, Tait L, Heppner GH, Miller FR. MCF10AT: a model for the evolution of cancer from proliferative breast disease. *Am J Pathol* (1996) 148(1):313–9.
57. Santner SJ, Dawson PJ, Tait L, Soule HD, Eliason J, Mohamed AN, et al. Malignant MCF10CA1 cell lines derived from premalignant human breast epithelial MCF10AT cells. *Breast Cancer Res Treat* (2001) 65(2):101–10. doi: 10.1023/A:1006461422273
58. Neve RM, Chin K, Fridlyand J, Yeh J, Baehner FL, Fevr T, et al. A collection of breast cancer cell lines for the study of functionally distinct cancer subtypes. *Cancer Cell* (2006) 10(6):515–27. doi: 10.1016/j.ccr.2006.10.008
59. Weiger MC, Vedham V, Stuelten CH, Shou K, Herrera M, Sato M, et al. Real-time motion analysis reveals cell directionality as an indicator of breast cancer progression. *PLoS One* (2013) 8(3):e58859. doi: 10.1371/journal.pone.0058859
60. Bos PD, Zhang XH, Nadal C, Shu W, Gomis RR, Nguyen DX, et al. Genes that mediate breast cancer metastasis to the brain. *Nature* (2009) 459(7249):1005–9. doi: 10.1038/nature08021
61. Kang Y, Siegel PM, Shu W, Drobnjak M, Kakonen SM, Cordon-Cardo C, et al. A multigenic program mediating breast cancer metastasis to bone. *Cancer Cell* (2003) 3(6):537–49. doi: 10.1016/S1535-6108(03)00132-6
62. de Oliveira S, Rosowski EE, Huttenlocher A. Neutrophil migration in infection and wound repair: going forward in reverse. *Nat Rev Immunol* (2016) 16(6):378–91. doi: 10.1038/nri.2016.49

63. Afonso PV, Janka-Junttila M, Lee YJ, McCann CP, Oliver CM, Aamer KA, et al. LTB4 is a signal-relay molecule during neutrophil chemotaxis. *Dev Cell* (2012) 22(5):1079–91. doi: 10.1016/j.devcel.2012.02.003
64. SenGupta S, Rane MJ, Uriarte SM, Woolley C, Mitchell TC. Human neutrophils depend on extrinsic factors produced by monocytes for their survival response to TLR4 stimulation. *Innate Immun* (2019) 25(8):473–86. doi: 10.1177/1753425919871994
65. Zhang ER, Liu S, Wu LF, Altschuler SJ, Cobb MH. Chemoattractant concentration-dependent tuning of ERK signaling dynamics in migrating neutrophils. *Sci Signaling* (2016) 9(458):ra122. doi: 10.1126/scisignal.aag0486
66. Nagarsheth N, Wicha MS, Zou W. Chemokines in the cancer microenvironment and their relevance in cancer immunotherapy. *Nat Rev Immunol* (2017) 17(9):559–72. doi: 10.1038/nri.2017.49
67. Bieche I, Chavey C, Andrieu C, Busson M, Vacher S, Le Corre L, et al. CXC chemokines located in the 4q21 region are up-regulated in breast cancer. *Endocrine-related Cancer* (2007) 14(4):1039–52. doi: 10.1677/erc.1.01301
68. Kang H, Watkins G, Parr C, Douglas-Jones A, Mansel RE, Jiang WG. Stromal cell derived factor-1: its influence on invasiveness and migration of breast cancer cells in vitro, and its association with prognosis and survival in human breast cancer. *Breast Cancer Res BCR* (2005) 7(4):R402–10. doi: 10.1186/bcr1022
69. Mukaida N. Chemokines. In: L Martini, editor. *Encyclopedia of Endocrine Diseases*. New York: Elsevier (2004). p. 490–4.
70. Kim YV, Gasparian ME, Bocharov EV, Chertkova RV, Tkach EN, Dolgikh DA, et al. A New Strategy for High-Level Expression and Purification of Biologically Active Monomeric TGF- β 1/C77S in *Escherichia coli*. *Mol Biotechnol* (2015) 57(2):160–71. doi: 10.1007/s12033-014-9812-7
71. Mahadeo DC, Janka-Junttila M, Smoot RL, Roselova P, Parent CA. A chemoattractant-mediated Gi-coupled pathway activates adenylyl cyclase in human neutrophils. *Mol Biol Cell* (2007) 18(2):512–22. doi: 10.1091/mbc.e06-05-0418
72. Nicholls DJ, Wiley K, Dainty I, MacIntosh F, Phillips C, Gaw A, et al. Pharmacological characterization of AZD5069, a slowly reversible CXC chemokine receptor 2 antagonist. *J Pharmacol Exp Ther* (2015) 353(2):340–50. doi: 10.1124/jpet.114.221358
73. Halder SK, Beauchamp RD, Datta PK. A specific inhibitor of TGF-beta receptor kinase, SB-431542, as a potent antitumor agent for human cancers. *Neoplasia* (2005) 7(5):509–21. doi: 10.1593/neo.04640
74. Bierie B, Moses HL. Tumour microenvironment: TGFbeta: the molecular Jekyll and Hyde of cancer. *Nat Rev Cancer* (2006) 6(7):506–20. doi: 10.1038/nrc1926
75. Jinnin M, Ihn H, Tamaki K. Characterization of SIS3, a novel specific inhibitor of Smad3, and its effect on transforming growth factor-beta1-induced extracellular matrix expression. *Mol Pharmacol* (2006) 69(2):597–607. doi: 10.1124/mol.105.017483
76. Liu C, Huang Z, Wang Q, Sun B, Ding L, Meng X, et al. Usefulness of neutrophil-to-lymphocyte ratio and platelet-to-lymphocyte ratio in hormone-receptor-negative breast cancer. *Oncotargets Ther* (2016) 9:4653–60. doi: 10.2147/OTT.S106017
77. Pistelli M, De Lisa M, Ballatore Z, Caramanti M, Pagliacci A, Battelli N, et al. Pre-treatment neutrophil to lymphocyte ratio may be a useful tool in predicting survival in early triple negative breast cancer patients. *BMC Cancer* (2015) 15:195. doi: 10.1186/s12885-015-1204-2
78. Yuan M, Zhu H, Xu J, Zheng Y, Cao X, Liu Q. Tumor-Derived CXCL1 Promotes Lung Cancer Growth via Recruitment of Tumor-Associated Neutrophils. *J Immunol Res* (2016) 2016:6530410. doi: 10.1155/2016/6530410
79. Ahuja SK, Murphy PM. The CXC chemokines growth-regulated oncogene (GRO) alpha, GRObeta, GROgamma, neutrophil-activating peptide-2, and epithelial cell-derived neutrophil-activating peptide-78 are potent agonists for the type B, but not the type A, human interleukin-8 receptor. *J Biol Chem* (1996) 271(34):20545–50. doi: 10.1074/jbc.271.34.20545
80. Nasser MW, Raghuvanshi SK, Malloy KM, Gangavarapu P, Shim JY, Rajarathnam K, et al. CXCR1 and CXCR2 activation and regulation. Role of aspartate 199 of the second extracellular loop of CXCR2 in CXCL8-mediated rapid receptor internalization. *J Biol Chem* (2007) 282(9):6906–15. doi: 10.1074/jbc.M610289200
81. Yeh HW, Lee SS, Chang CY, Lang YD, Jou YS. A New Switch for TGF β in Cancer. *Cancer Res* (2019) 79(15):3797–805. doi: 10.1158/0008-5472.CAN-18-2019
82. Luo J, Chen X-Q, Li P. The Role of TGF- β and Its Receptors in Gastrointestinal Cancers. *Trans Oncol* (2019) 12(3):475–84. doi: 10.1016/j.tranon.2018.11.010
83. Parekh T, Saxena B, Reibman J, Cronstein BN, Gold LI. Neutrophil chemotaxis in response to TGF-beta isoforms (TGF-beta 1, TGF-beta 2, TGF-beta 3) is mediated by fibronectin. *J Immunol (Baltimore Md 1950)* (1994) 152(5):2456–66.
84. Fridlender ZG, Sun J, Kim S, Kapoor V, Cheng G, Ling L, et al. Polarization of tumor-associated neutrophil phenotype by TGF-beta: “N1” versus “N2” TAN. *Cancer Cell* (2009) 16(3):183–94. doi: 10.1016/j.ccr.2009.06.017
85. Castriconi R, Dondero A, Bellora F, Moretta L, Castellano A, Locatelli F, et al. Neuroblastoma-derived TGF- β 1 modulates the chemokine receptor repertoire of human resting NK cells. *J Immunol (Baltimore Md 1950)* (2013) 190(10):5321–8. doi: 10.10049/jimmunol.1202693
86. Costanzo-Garvey DL, Keeley T, Case AJ, Watson GF, Alsamrae M, Yu Y, et al. Neutrophils are mediators of metastatic prostate cancer progression in bone. *Cancer Immunol Immunother CII* (2020) 69(6):1113–30. doi: 10.1007/s00262-020-02527-6
87. Sun B, Qin W, Song M, Liu L, Yu Y, Qi X, et al. Neutrophil Suppresses Tumor Cell Proliferation via Fas /Fas Ligand Pathway Mediated Cell Cycle Arrested. *Int J Biol Sci* (2018) 14(14):2103–13. doi: 10.7150/ijbs.29297
88. Katt ME, Placena AL, Wong AD, Xu ZS, Searson PC. In Vitro Tumor Models: Advantages, Disadvantages, Variables, and Selecting the Right Platform. *Front Bioeng Biotechnol* (2016) 4:12. doi: 10.3389/fbioe.2016.00012
89. Tazzyman S, Barry ST, Ashton S, Wood P, Blakey D, Lewis CE, et al. Inhibition of neutrophil infiltration into A549 lung tumors in vitro and in vivo using a CXCR2-specific antagonist is associated with reduced tumor growth. *Int J Cancer* (2011) 129(4):847–58. doi: 10.1002/ijc.25987
90. Lämmermann T, Afonso PV, Angermann BR, Wang JM, Kastenmüller W, Parent CA, et al. Neutrophil swarms require LTB4 and integrins at sites of cell death in vivo. *Nature* (2013) 498(7454):371–5. doi: 10.1038/nature12175
91. Colotta F, Re F, Polentarutti N, Sozzani S, Mantovani A. Modulation of granulocyte survival and programmed cell death by cytokines and bacterial products. *Blood* (1992) 80(8):2012–20. doi: 10.1182/blood.V80.8.2012.2012
92. Tsai JH, Yang J. Epithelial-mesenchymal plasticity in carcinoma metastasis. *Genes Dev* (2013) 27(20):2192–206. doi: 10.1101/gad.225334.113
93. Ye X, Brabletz T, Kang Y, Longmore GD, Nieto MA, Stanger BZ, et al. Upholding a role for EMT in breast cancer metastasis. *Nature* (2017) 547(7661):E1–e3. doi: 10.1038/nature22816
94. Suarez-Carmona M, Bourcy M, Lesage J, Leroi N, Syne L, Blacher S, et al. Soluble factors regulated by epithelial-mesenchymal transition mediate tumour angiogenesis and myeloid cell recruitment. *J Pathol* (2015) 236(4):491–504. doi: 10.1002/path.4546
95. Chen T, Guo J, Yang M, Zhu X, Cao X. Chemokine-containing exosomes are released from heat-stressed tumor cells via lipid raft-dependent pathway and act as efficient tumor vaccine. *J Immunol (Baltimore Md 1950)* (2011) 186(4):2219–28. doi: 10.4049/jimmunol.1002991
96. Szajnik M, Czysztowska M, Szczepanski MJ, Mandapathil M, Whiteside TL. Tumor-derived microvesicles induce, expand and up-regulate biological activities of human regulatory T cells (Treg). *PLoS One* (2010) 5(7):e11469. doi: 10.1371/journal.pone.0011469
97. Huo X, Li H, Li Z, Yan C, Agrawal I, Mathavan S, et al. Transcriptomic profiles of tumor-associated neutrophils reveal prominent roles in enhancing angiogenesis in liver tumorigenesis in zebrafish. *Sci Rep* (2019) 9(1):1509. doi: 10.1038/s41598-018-36605-8

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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