



Aging and Options to Halt Declining Immunity to Virus Infections

Miguel Ángel Palacios-Pedrero, Albert D. M. E. Osterhaus, Tanja Becker, Husni Elbahesh, Guus F. Rimmelzwaan^{*} and Giulietta Saletti^{*}

Research Center for Emerging Infections and Zoonoses, University of Veterinary Medicine Hannover, Hannover, Germany

Immunosenescence is a process associated with aging that leads to dysregulation of cells of innate and adaptive immunity, which may become dysfunctional. Consequently, older adults show increased severity of viral and bacterial infections and impaired responses to vaccinations. A better understanding of the process of immunosenescence will aid the development of novel strategies to boost the immune system in older adults. In this review, we focus on major alterations of the immune system triggered by aging, and address the effect of chronic viral infections, effectiveness of vaccination of older adults and strategies to improve immune function in this vulnerable age group.

OPEN ACCESS

Edited by:

Mario (Mago) Clerici, University of Milan, Italy

Reviewed by:

Brandt D. Pence, University of Memphis, United States Calogero Caruso, University of Palermo, Italy

*Correspondence:

Guus F. Rimmelzwaan Guus.Rimmelzwaan@tihohannover.de Giulietta Saletti Giulietta.Saletti@tiho-hannover.de

Specialty section:

This article was submitted to Viral Immunology, a section of the journal Frontiers in Immunology

Received: 16 March 2021 Accepted: 26 April 2021 Published: 12 May 2021

Citation:

Palacios-Pedrero MÁ, Osterhaus ADME, Becker T, Elbahesh H, Rimmelzwaan GF and Saletti G (2021) Aging and Options to Halt Declining Immunity to Virus Infections. Front. Immunol. 12:681449. doi: 10.3389/fimmu.2021.681449 Keywords: aging, cell mediated immunity, virus infections, vaccine, immunosenescence

INTRODUCTION

Life expectancy is currently higher than ever and by 2050 the number of individuals over 65 years of age is estimated to be more than 1.6 billion worldwide (https://www.who.int/news-room/fact-sheets/detail/ageing-and-health). Aging is associated with progressive changes involving all organ systems including the immune system, collectively affecting the ability to mount protective immune responses to various infectious pathogens and to vaccination (1–9). Therefore, age-related diseases and conditions are of major public health concern and should prompt the development of more effective therapies and vaccines to prevent or mitigate the impact of infectious diseases that are a major cause of morbidity and mortality in older adults (10–13). For instance, the recent outbreak of COVID-19 caused by SARS CoV-2 has highlighted the increased severity of this virus infection in older adults, resulting in disproportionally high morbidity and mortality rates (14–19).

The term "immunosenescence" refers to alterations and dysregulation of the immune system that take place during aging. It is a multifaceted, biological process causing a progressive malfunctioning of cells of innate and adaptive immunity. Changes in both composition and function of immune cells characterized by up- or down-regulation of surface markers, defects in cell signaling and alterations of cell populations are hallmarks of immunosenescence. On the one hand, genetic background, microbiome, diet, co-morbidities and/or environmental factors are thought to play a role in the aging process (20, 21). Immunosenescence has also been reported in some animal species including non-human primates and dogs (22–24). On the other hand, it should be considered that: (i) many older adults remain healthy until advanced age (>90 years of age); (ii) in some cases "dysfunctional" immune cells can function properly when stimulated adequately, e.g., with the use of vaccines designed for use in older people; (iii) altered response not always translates into being harmful (25, 26).

1

The innate immune system plays a key role as the first line of defense against pathogens and promotes the adaptive immune system to generate long-lasting protective immunity. In case of vaccination, the functionality of innate immune cells at the site of administration (e.g., macrophages, dendritic cells, neutrophils) is of importance and largely dictates the outcome of vaccination. In older adults, it has been reported that antigen processing and presentation capacities are reduced and neutrophil cytokine production is altered, leading to dysfunctional chemotaxis and activation of other immune cells (27-33). Upon infection or vaccination, a small number of naive CD4⁺ and CD8⁺ T cells recognize a cognate peptide/MHC complex presented by antigen presenting cells, which undergo activation, proliferation and differentiation into effector cells. After the contraction phase, the majority of the effector cells (90-95%) die by apoptosis and only a small fraction of the CD4⁺ and CD8⁺ T cell effector population differentiate into memory cells (34-36). These cells are responsible for maintaining long-term immunity and protective recall responses. Older individuals experience a decrease in the number of naive T cells due to reduced hematopoiesis and thymus involution, which impairs the response to novel antigens, including vaccine antigens (37-41). Nonetheless, naive T cells from individuals older than 70 years still exhibit highly diverse T cell receptor (TCR) specificities, albeit reduced compared to those of younger adults (38, 40, 42). Furthermore, immunosenescent T cells are characterized by an impaired proliferation rate, resistance to apoptosis, downregulation of costimulatory surface molecules (e.g., CD27 and CD28), telomere shortening and expression of KLRG-1 and CD57 (43-55). Under normal circumstances, the fine-controlled balance between proinflammatory and anti-inflammatory cytokines maintains the physiologic function of inflammation. In older people, dysregulation of cytokine production with a progressive tendency toward a proinflammatory phenotype (e.g., IL-6, IL-15, IFN- γ), is believed to play a key role in the inability to control systemic inflammation. This chronic low-grade of uncontrolled inflammation is defined as "inflammaging" (56-64). Recent studies have shown that persistence of chronic inflammation is associated with various environmental and metabolic factors (e.g., diet, nutrition and gut microbiota) (65-67). Pro-inflammatory cytokines are also secreted in response to oxidative stress, DNA damage and autophagy [reviewed elsewhere (68, 69)]. The processes of immunosenescence and inflammaging are intertwined with each other. On the one hand, senescent cells, which accumulate during aging, are characterized by a pro-inflammatory cytokine secretion pattern, leading to inflammaging. On the other hand, an increase of such inflammatory mediators drives altered adaptive immune responses, contributing to immunosenescence. Both processes are therefore involved in a vicious cycle that impairs functioning of the immune system in older adults (70-72). In addition, chronic infections (e.g., caused by cytomegalovirus) contribute to the enhancement of both, immunosenescence and inflammaging. Such pathogens trigger a low but continuous inflammatory response in the host, as well as a clonal expansion of differentiated/memory T cells at the cost of naive T cell compartment, contributing to an impaired immune response to novel pathogens (73 - 78).

The second arm of the adaptive immune response is mediated by humoral immunity. Humoral immunity is a key correlate of

protection against infectious pathogens and the induction of specific antibodies and the development of specific B cell memory are pivotal to provide protection against reinfection and contribute to the success of vaccination. Aging has a profound impact on diversity of the B cell repertoire. Impaired somatic hypermutation and isotype switching, together with a reduction in the number of plasma cells, affect the magnitude and quality of the antibody response induced by infection or vaccination (41, 79–82).

Aging also has a major impact on the severity of viral and bacterial infections, resulting in a more severe course and often fatal outcome in the older population. Furthermore, effectiveness and longevity of vaccination, the main preventive measure against infections, are gradually reduced in the aging population. Certain viruses can establish a low level of viral replication and cause persistent and often latent infections. In immunosenescent adults, these viruses can reactivate and cause a severe infection that can lead to recurrent disease of increasing severity. Varicella zoster virus (VZV) infection is a notable example of this. VZV, an α -herpesvirus, usually causes infections during childhood (chickenpox) and then becomes persistent and latent. Virus reactivation leads to shingles, also known as herpes zoster (HZ), and possible further serious complications (83-88). Another example is the human cytomegalovirus (CMV), which is highly prevalent in adults worldwide. CMV, a β-herpesvirus, also establishes life-long latent infection and is generally considered to promote T cell immunosenescence (89-91). CMV infection leads to continuous or chronic antigen presentation, resulting in increased numbers of highly differentiated T cells with a reduced TCR repertoire (62, 92-96).

In addition, other (respiratory) viruses, such as respiratory syncytial virus (RSV) and influenza viruses (IVs), cause more severe respiratory disease with complications and mortality in older adults, especially in association of co-morbidities (97–102). Although effective influenza vaccines are available, their effectiveness is not optimal in older people. Attempts to develop more immunogenic vaccine formulations for this age category have resulted in somewhat more effective influenza vaccines (103–106). In contrast, a registered RSV vaccine is still not available, although several candidate RSV vaccines are in various stages of development (107–110).

In this review, we discuss the current understanding of agerelated changes affecting immune cells and how that influences protective immune response to virus infection and vaccination. Moreover, we will give an insight into strategies that are currently used, or under development, to improve immune cell functions and vaccine efficacy in older adults.

AGE-ASSOCIATED IMMUNE CELL DYSFUNCTIONS

The immune system is complex and involves the interplay of many different cell types that collectively afford protection against infectious pathogens. Aging negatively influences the function of immune cells resulting in increased severity of infections and impaired responses upon vaccination. Although there is an increasing interest in the development of therapeutic and preventive strategies targeting older adults, many molecular processes leading to immunosenescence are still not fully understood. Additionally, a large body of knowledge has been obtained from mouse models; however, it is not clear to what extent this model faithfully represents the mechanisms controlling these processes in humans. Furthermore, most studies have used peripheral blood mononuclear cells (PBMCs) because of ease of accessibility, but these do not account for potential changes of cells residing in tissues. Consequently, more research is required to obtain a better understanding of immunosenescence in aging humans, which will guide the design of improved vaccines tailored for use in the aging population.

Some of the factors that are known to play a role in immunosenescence during aging include genomic instability, telomere shortening, epigenetic modifications, changes of intracellular signal pathways responsible for cell communication and dysfunction of mitochondria [reviewed elsewhere (8, 111)].

Here we will focus on age-related changes of immune cells of both innate and adaptive immunity, along with those induced after infection or vaccination (**Figure 1**).

Innate Immune System

Dendritic cells (DCs) are critical for the induction of protective immune responses against pathogens and represent the link between innate and adaptive immunity. DCs are professional antigen-presenting cells that provide activation signals to T cells and are crucial for mounting a robust humoral response (112-114). DCs can be activated by various stimuli including microbes, apoptotic cells, and inflammatory cytokines. They sense pathogens at the port of entry through pattern recognition receptors (PRRs) (e.g., Toll-like receptors, C-type lectin receptors, intracytoplasmic NOD-like receptors), and then migrate to the lymphoid organs where they prime naive T cells and regulate B cell responses. DCs are comprised of two major subsets of either myeloid (mDCs; conventional/classical DC, cDC) or lymphoid origin (plasmocytoid; pDCs). mDCs regulate pro-inflammatory responses (e.g., via T-helper 1 and cytotoxic T lymphocyte responses) upon bacterial and viral infections, and pDC produce type I interferon, that can directly antagonize viral replication (115-118). It has been shown that magnitude, functionality and signaling of the pDCs are reduced in older people, and that has been correlated with the increased occurrence of severe influenza virus infections in this age group (30, 119, 120). Yet, impaired T cell proliferation and IFN- γ secretion have been observed upon stimulation of aged pDCs with influenza virus (121). Transcriptional analyses have demonstrated that PBMCs from people over 65 years of age display a delayed and atypical response after stimulation via some Toll-like receptors (TLR4 and TLR7/8) compared to those from younger individuals (<40 years old), which has a profound effect on the production of effector molecules involved in T cell activation and proliferation, such as IFN- γ and TNF- α (122). Following TLR7/9 stimulation, pDCs of older subjects produce less type I and III interferons and display a reduced antigen presentation that impairs T cell activation (121). An increased level of IL-6 and TNF- α secreted by "old" mDCs in response to LPS, ssRNA, and self-DNA, has been associated to

alteration in signaling pathways that lead to PI3K, NF-KB, or type I IFN responses (123, 124). A decreased expression and/or function of some TLRs in DCs and monocytes in older adults individuals has also been reported (125-127). Moreover, epigenetic changes (e.g., methylation or phosphorylation) induce up-regulation of immune checkpoint molecules on DCs that reduce the ability to migrate to the secondary lymphoid, leading to an impaired T cell activation (114, 128-132). The development of a robust and protective antibody response with generation of high-affinity antibodies is mediated by the ability of follicular DCs (fDCs) to capture and retain, for extended periods of time, native antigen in the B cell follicles within the germinal center. Studies have documented that age-related defects in the germinal center formation and reaction (e.g., retention of antigen complexes and reduced expression of the FcyRII on fDCs), have a dramatic impact on generation of humoral immunity (133).

Monocytes and tissue-resident macrophages, which can be monocyte-derived or embryo-derived, are phagocytic cells, primarily involved in the innate immune response against infectious pathogens and also involved in the maintenance of tissue homeostasis (134). Age-related defects in the macrophage/ monocyte lineage function are predominantly mediated by the dysregulation of cellular signaling, which affects antigen presentation and response to inflammatory stimuli (135-137). Macrophages are usually divided in two subsets M1 (proinflammatory) and M2 (anti-inflammatory/immune regulatory) and aging alters their polarization and function (137-139). Due to the difficulty in obtaining and studying tissue-resident macrophages and their highly diverse tissue-dependent phenotypes, it is still not completely understood which macrophages/tissue are more affected by aging (140-142). Macrophages may also contribute to the chronic inflammatory state in older adults and lead to the progression of age-associated diseases. Although no differences have been found in the number of peripheral blood monocytes in older adults compared to younger subjects, there is an agedependent upward shift in the proportion of monocyte subsets with an inflammatory profile. For example, 'non-classical' CD14^{low}CD16⁺ monocytes specifically increase with age and display reduced HLA-DR and CX(3)CR1 surface expression (143). Upon TLR stimulation, monocytes display age-dependent differences, both at transcriptional and functional levels, resulting in a differential expression of surface molecules and cytokine production (144). These age-related alterations have been mainly associated to impaired monocyte TLR responses, although few studies show a rather enhanced inflammatory response (125, 144-146). For example, a positive correlation was found between surface expression of B7 co-stimulatory molecules before influenza vaccination and the vaccine induced antibody response, but interestingly, this correlation was weaker for older vaccinees (147-149). Furthermore, expression of MHC class II molecules was lower in aged human monocytes and mouse macrophages (150-153). Moreover, aged human monocytes have shortened telomeres, reduced phagocytic capacity and high level of intracellular TNFα (154).

Neutrophils are phagocytic cells that migrate from blood to infected tissues and rapidly respond to invading pathogens by



FIGURE 1 | Simplified representation of the phenotypical and functional changes affecting cells of the innate and adaptive compartment during aging. ILR, toll like receptor; MHC, major histocompatibility complex; CXCR, C-X-C chemokine receptor; ICAM, intercellular adhesion molecule; LY6C/G, lymphocyte antigen 6 complex (locus C/G); NET, neutrophil extracellular trap; BCR, B cell receptor; TCR, T cell receptor; KLRG-1, killer-cell lectin like receptor G1; CTLA-4, cytotoxic T-lymphocyte antigen 4; PD-1, programmed cell death protein 1; TIM-3, T cell immunoglobulin and mucin domain-containing protein 3; LAG-3, lymphocyte-activation gene 3. The figure was created with BioRender.com.

activating adhesion, chemotaxis, phagocytosis as well as release of oxidants, proteases and other molecules (155–158). However, an excessive accumulation of neutrophils and hyperresponsiveness can be detrimental and cause tissue injury, as recently documented in subjects with lethal COVID-19 (159, 160). Although no difference has been found in the number of neutrophils between young and older adults, aged neutrophils exhibit a dysfunctional phagocytic and chemotactic capacity (28, 161, 162). In aged mice, these dysfunctional cells expand and accumulate in lymph nodes due to impaired apoptosis (163, 164). Phenotypically, up-regulation of CD45, TLR4, CD24, CXCR4, CD11b, CD11c, CD49d, ICAM and down-regulation of CXCR2, CD62L, L-lectin, LY6C/G have been documented in aged human neutrophils (156, 165). In addition, altered

cytokines secretion profiles, generation of reactive oxygen species (ROS) and associated microbial killing have been reported, although these findings might largely be stimulus-dependent (28, 166, 167). Age dependent impaired formation of neutrophil extracellular traps (NETs), structures able to capture and immobilize pathogens, may explain why older adults are more susceptible to invasive bacterial infections (29, 168, 169).

Natural killer (NK) cells are cytotoxic innate cells that eliminate infected, transformed and senescent cells (170–175). In addition, they also exert immunoregulatory activities by secreting cytokines and chemokines, which can activate and modulate the adaptive immune responses (176–178). Like other cells, NK cells are subject to age-related changes in, for instance, number, function, phenotype and redistribution (179, 180). As an example, there is an increased number of highly differentiated mature CD56^{dim} cells accompanied with a decline of immature CD56^{bright} subset as well as reduced NK cell activity, which may lead to impaired immune regulation (181–184).

Adaptive Immune System

Profound age-related changes in the immune system are observed in cells of adaptive immunity. In this section, we will describe major alterations affecting aged T and B lymphocytes.

T Lymphocytes

T cells recognize antigens derived from pathogens or tumors via their T cell receptor and develop antigen-specific memory responses or tolerance (185). Upon activation, naive T cells proliferate, differentiate and generate effector T cells that can help to kill infected cells and/or to activate other cells (e.g., macrophages), and B cells. T cell precursors are generated within the bone marrow whereas their maturation and selection take place in the thymus. The thymus undergoes an involution process starting after puberty that gradually induces its atrophy (186, 187). Epithelial cell structure and cell turnover are altered and there is a shift toward adipose tissue that results into a reduced thymopoiesis, as well as altered transcription factors and cytokine production (40, 188). As a consequence, the output of newly generated naive T cells (CCR7⁺, CD62L⁺, CD45RO⁻, CD45RA⁺, CD27⁺, CD28⁺), and therefore the possibility to respond to novel pathogens, is impaired (38, 189, 190). Despite the compensatory mechanism that promotes homeostatic proliferation of existing naive cells, their frequency is reduced in the periphery and in lymphoid organs (38, 42, 191-195). Coinciding with the loss of naive T cells, highly differentiated effector/memory T cells accumulate during aging, especially CD8⁺ T cells, many of which are dysfunctional. In particular, altered cytokine production profiles, reduced TCR clonal diversity and more self-reactive T cells have been observed (196-200) together with a general decline of the proliferative capacity in response to TCR stimulation, with CD4⁺ memory T cells more prone to this loss of function (201, 202). An increased frequency of a subset of memory CD8⁺ T cells with a naive phenotype (TMNP) that secretes effector molecules (e.g., IFNy, Granzyme B) in response to chronic viral stimulation has been associated with aging. The frequency of these cells positively correlates with the severity of West Nile and influenza virus infections (203). Furthermore, regulatory T cells (Tregs) with upregulated check point molecules, such as cytotoxic T lymphocyte

antigen 4 (CTLA-4), are more abundant in older adults than in young individuals and that may affect the crosstalk between T cells and DCs, since Tregs can prevent the maturation of DCs (9, 204-206). Age-dependent changes in T cells have, directly or indirectly, an effect on their effector functions (201). For example, the frequency of CD8⁺ T cells expressing perforin and granzyme B is reduced in older individuals, which correlated with an increased risk of severe influenza (207-209). Aged T cells can be phenotypically identified based on the expression of surface markers and intracellular molecules (e.g., transcription factors and cytokines). The co-stimulatory molecule CD28 is decreased and subsets of CD4⁺CD28⁻ and CD8⁺CD28⁻ T cells emerge in older adults (46, 210). Chronic activation of T cells also induces downregulation of CD28 expression and that has been associated with impaired vaccine responses (207, 211-214). Loss of CD27, upregulation of CD57 and KLRG-1, reduced expression of granzyme B together with telomere shortening and expression of a senescence-associated secretory phenotype (SASP), are the major hallmarks of senescent T cells (43, 46, 51, 211, 215-218). The immunosenescent T cells with a SASP phenotype play an autocrine role and promote the recruitment of pro-inflammatory innate cells that, due to aging, are not very efficient at eliminating the senescent cells. These concomitant phenomena also contribute to the establishment of inflammaging.

Dysfunction of T cells caused by T cell exhaustion has been reported in various chronic viral infections and cancers [reviewed in (219)] (75, 220–223). Chronic antigenic stimulation induces upregulation of inhibitory receptors such as programmed cell death 1 (PD-1), cytotoxic T lymphocyte antigen 4 (CTLA-4), T cell immunoglobulin and mucin domain-containing protein 3 (TIM-3), and lymphocyte activation gene 3 (LAG-3), that impair TCR signaling pathways and therefore immune T cell functions (e.g., proliferation, transcriptional signature). SAT-B1, a chromatin organizer, is downregulated in aged T cells and its expression negatively correlated with PD-1 expression in virus-specific CD8⁺ T cells (224).

Although exhaustion and senescence are two distinct phenomena that differ phenotypically and functionally, both contribute to the decline of T cell functionality during aging. Therefore, both of these processes should be considered when developing potential novel strategies to overcome dysfunctional immune responses in this age group (225).

B Lymphocytes

B cells undergo profound changes during aging leading to a reduced protective vaccine efficacy and reduced control of infections. They are responsible for antibody production and have effector as well as regulatory functions. Although total B cell counts remain relatively stable in the adult population, a reduced output of naive B cells from the bone marrow (BM) has been reported in mouse and human studies (226–229). Decreased production of IL-7 by stromal cells in the BM, reduces the size of the B cell progenitor population and affects B lymphopoiesis (230–232). The function of hematopoietic stem cells (HSCs) declines with age and shifts toward the generation of non-lymphoid cells, therefore reducing the source of B cell progenitors. Recently, a population of old age-associated B cells (ABC) has been identified

in mice (233–237). These cells produce pro-inflammatory cytokines, in particular TNF- α , that affect the generation of mature B cells (233). In humans, accumulation of B cells with similar characteristics has been described in the peripheral blood of older individuals (238, 239). The reduced generation of naive B cells together with a progressive accumulation of dysfunctional memory B cells in the periphery, contributes to a contraction of the B cell repertoire diversity that limits the ability to recognize and respond to novel antigens.

Alterations of the immunoglobulin (Ig) class-switch recombination (CSR) and somatic hypermutation (SHM), may negatively affect the generation of high-affinity antibodies and germinal center formation, both of which are very important mechanisms for the induction of protective and long-lasting immune responses. Age-related reduction and instability of the transcription factor E47 affects the enzyme activation-induced cytidine deaminase (AID) that is involved in the regulation of CSR and SHM (62, 240). Downregulation of AID and E47, caused by the pro-inflammatory microRNAs (miR-155 and miR-16), has been observed in B cells from aging individuals (241). Reduction in size and output of the germinal centers, as a consequence of a sub-optimal T cell help to B cells, have been reported upon infection and vaccination (79, 133, 242-244). As demonstrated in older adults and aged mice, inflammation also leads to a decrease of chemokine CXCL12 production, which may impair the recruitment and accumulation of plasma cells in the bone marrow, the major site for antibody production (245, 246). The altered architecture of the spleen in older age negatively influences the humoral responses. For example, marginal zone of old mouse spleens showed a dysfunctional antigen recognition and migration of B cells due to an impaired functionality of splenic stromal cells (247). Decrease of IgM and IgD serum levels, probably due to the shift to a more differentiated and effector B cell population has been observed in humans (248-250). Finally, it has been reported that senescent B cells can produce auto-antibodies, which may lead to the development of autoimmune diseases (229, 251-253).

ROLE OF VIRAL INFECTIONS IN OLDER ADULTS

Immunosenescence accounts for an increased susceptibility to viral infections such as those caused by influenza viruses and RSV and reduced vaccine efficacy in this age group. In addition, chronic stimulation of the immune system operated by viruses establishing latent infections, that can re-activate from time to time, may further impair the overall immune status in older individuals.

Human Cytomegalovirus

Human cytomegalovirus (HCMV) is a β -herpes virus that causes lifelong latent infections in a large proportion of the human population. The mechanisms involved in HCMV latency are still poorly understood and despite extensive literature on how HCMV infection influences the adaptive immune response in older adults, this is still a matter of debate (254). Immunologically, HCMV infection is characterized by inflation of HCMV-specific memory T cells, mostly CD8⁺, with proliferation of oligoclonal T cells (96, 255-262). The accumulation of these terminally differentiated apoptosis-resistant CMV-specific T cells clones is believed to compromise the overall T cell repertoire diversity. Chronic HCMV infection also triggers an increased secretion of proinflammatory cytokines that favors cell damage and contributing to inflammaging, although its impact in older adults is still unclear (263-266). CMV-seropositivity has been associated with reduced immune response to (influenza) vaccination in older adults, but also in younger subjects (267, 268). Increased cardiovascular problems and mortality rate have been also reported in CMV⁺ older individuals. However, data are often inconclusive and associations are still controversial, not confirmed and do not need to have a causal relationship (267, 269-274). In contrast, CMV positivity may exert a beneficial effect by improving CD8⁺ T cell polyfunctionality, as demonstrated in healthy young individuals (263, 275, 276). Thus, chronic HCMV infections may have an impact on immune responses induced in older people after infection or vaccination. More studies are warranted to further investigate this association, which may guide the development of effective therapeutic strategies.

Varicella Zoster Virus

Varicella zoster virus (VZV) belongs to the α -herpesvirus family (277). This virus is highly infectious (about 90% of adults are infected) and produces "varicella", also known as "chickenpox", a self-limiting disease which is commonly experienced during childhood and is characterized by widespread vesicular rash and fever, that usually resolves in 1-2 weeks (278-280). Despite a robust primary response, VZV is not eliminated from the body and can remain in the spinal and cranial sensory ganglia in a lifelong latent state (84, 281). The mechanisms leading to latency and subsequent reactivation are still to be unraveled. However, a recent study has suggested VLT-ORF63 transcripts from VZV as potential inducers of reactivation (282). In addition, cellmediated immunity appears to play an important role in the immune response to VZV and also latency (283-285). Reactivation occurs in 10-20% of seropositive individuals and causes the so-called "herpes zoster" (HZ) or "shingles", a painful rash that mainly affects older adults and immunocompromised individuals, in which VZV-specific immune control has declined below a critical level (286). Even if VZV primary infection could lead to complications (e.g., bacterial infections, encephalitis, hemorrhage and pneumonia), a later reactivation process may trigger severe neurological complications, including post-herpetic neuralgia (PHN) (84, 286-289). PHN can last for years, or become permanent, with age being the most important risk factor correlating with its development. Importantly, the severity of HZ has been associated with a reduced frequency of VZVspecific effector memory T cells, highlighting the crucial role of cell-mediated immunity (290).

Three vaccines are currently licensed for preventing VZV associated disease. The most widely used VZV vaccine is a live attenuated vaccine based on the OKA-strain that can be administered in one- or two-dose regimens during childhood (286, 291–293). Although this vaccine is very effective (up to 98%)

in protecting from severe VZV infection, it does not prevent latency and reactivation of the virus. Moreover, it is not suitable for immunocompromised individuals, due to a less favorable safety profile (294).

Two vaccines, ZostavaxTM and ShingrixTM, have been developed to prevent HZ (shingles). ZostavaxTM is a live attenuated vaccine licensed in 2006 for individuals >50 years of age and contains 14fold more of the OKA-strain than needed to prevent varicella. Several studies have indicated that the efficacy afforded by this vaccine wanes with age and appears to be inferior to that obtained with ShingrixTM (295–297). For this reason, the ZostavaxTM is, effective July 2020, no longer available for use in the US. A new ZostavaxTM-based vaccine inactivated by gamma irradiation is under clinical investigation. Data obtained during the phase III clinical trial indicated that the vaccine is well tolerated with no significant safety issues. However, the immune response is not as strong as those induced with other vaccines and it should also be administered in a four-dose regimen (298–300).

ShingrixTM is a recombinant adjuvanted vaccine licensed in 2017 for people >50 years of age. The vaccine contains the recombinant glycoprotein E (gE) formulated with the AS01B adjuvant system. The gE is involved in viral replication, cell to cell virus transfer and is highly expressed in VZV infected cells, being also the primary target of the T cell response (279, 301-303). This vaccine is able to generate strong humoral and cellmediated immunity, therefore overcoming the decline of the VZV-specific response observed in older individuals. The efficacy of this vaccine for preventing HZ and possible complications, such as PHN, is high irrespective of age and, importantly, the vaccine-induced immunity is long lasting and can persist up to 9 years (25, 304-306). When compared to the live-attenuated vaccine, ShingrixTM induced higher frequency of gE-specific CD4⁺ and CD8⁺ memory T cells in older adults (307). VZV-specific CD4⁺ T cells have been associated with a positive vaccine outcome (308). ShingrixTM, unlike ZostavaxTM, has shown a favorable safety profile in immunocompromised and transplant recipients (25, 305, 306, 309, 310). However, ShingrixTM reactogenicity, due to the adjuvant, appeared to be higher compared to other vaccines used for older individuals, although serious adverse events and immune-mediated diseases were not increased in the vaccinees. The success of this vaccine for older adults may reside in the use of an adjuvant that provides a proper activation of the innate immune system, which is pivotal for the induction of an effective and longlasting adaptive immune response.

SARS CoV-2

Since the first reported case of COVID-19 in December 2019, the severe acute respiratory syndrome coronavirus 2 (SARS CoV-2) has infected more than 130 million people and caused over 3 million deaths worldwide (https://covid19.who.int/). SARS CoV-2 is a positive sense single stranded RNA virus that uses its S protein to bind and infect respiratory epithelial cells that express its receptor angiotensin converting enzyme 2 (ACE-2), including type II pneumocytes in the lung (311). Although the majority of infected people only develop mild disease, some develop severe disease often with a fatal outcome. Especially older people have been affected disproportionally with high case fatality rates.

In the US, for example, 80% of all fatal cases were older than 65 years of age (CDC - https://www.cdc.gov/coronavirus/2019ncov/need-extra-precautions/older-adults.html). Furthermore, the more severe course of infection observed in older people often requires hospitalization and treatment in intensive care units (312-314). Studies to elucidate the molecular and cellular mechanisms responsible for the worse outcome of SARS CoV-2 infection in aged people are ongoing. However, the strong association between age and severity of infection seems to be a common feature to other coronavirus infections (e.g., SARS CoV-1 and MERS) and it has also been observed for other respiratory viral pathogens, such as influenza viruses (315-318). Underlying co-morbidities associated with aging (e.g., cancer, hypertension, cardiovascular diseases diabetes or autoimmunity), genetic factors and reduced ability to mount adequate adaptive immune responses due to presence of dysfunctional aged immune cells, may account for the severe clinical outcome (319, 320). Preexisting SARS CoV-2 cross-reactive T cells induced by previous exposures to seasonal human common cold coronavirus are readily detected in younger individuals, but are virtually absent in older people, suggesting that this age group may not benefit from a potential protective effect of these pre-existing crossreactive T cells (19). A chronic, low-grade inflammation (inflammaging), the main hallmark of aging, has been suggested to play a critical role in promoting "cytokine storm" and consequent acute respiratory distress syndrome (ARDS), often observed in older individuals (321-325). The downregulation of the ACE-2 receptor together with a dysregulated angiotensin-II pathways (renin-angiotensin-aldosterone system) may foster the uncontrolled and exaggerated inflammatory response leading to pulmonary damages, multi-organ dysfunction and ultimately, death (326, 327).

The ongoing COVID-19 vaccination campaign in older subjects has already been shown very successful and significantly reduced hospitalizations and mortality among people of this age group. It also demonstrates that despite high age and associated immunosenescence, successful vaccination is still possible in this age group.

Influenza Virus

Seasonal influenza viruses are among the leading causes of respiratory infections and responsible for 290,000-650,000 deaths annually worldwide (100). Despite the high vaccine coverage among individuals >65 years of age in some countries, this age group accounts for the vast majority of deaths and hospitalizations (CDC - https://www.cdc.gov/flu/index.htm). Although the mechanisms underlying increased severity of influenza virus infection in older individuals are still not fully understood, comorbidities such chronic and metabolic diseases, obesity, immunosuppression, and frailty represent a risk for disease severity and development of complications (328–330).

Age-related immunosenescence affects the functionality of B and T cells, hindering an optimal protective response upon seasonal influenza vaccination. Current influenza vaccines mainly provide protection through the induction of antibodies that antigenically match the epidemic strains (331, 332). Several studies have indicated that influenza-specific antibodies in older adults decline

faster, failing to maintain long-lasting protective levels (333–336). Moreover, these individuals exhibit a reduced accumulation of "*de novo*" mutations in the Ig variable gene affecting the adaptability of their antibody responses to influenza virus (337, 338). Therefore, this age group mainly rely on cross-reactive memory B cells generated early in life (339). Interestingly, a recent study has shown that influenza virus infection predominantly recalls pre-existing memory B cells against non-neutralizing epitopes in contrast to vaccination that mainly targets memory B cells to protective HA epitopes. One could speculate that boosting pre-existing immunity may play a key role in susceptibility versus protection upon influenza virus infection (339, 340). Additional defects influencing the B cell responses to influenza virus infection or vaccination are reviewed elsewhere (99).

T cells, especially CD8⁺, are an important correlate of protection against influenza virus infections also providing heterosubtypic immunity (341–344). In older adults, phenotypic and functional defects in the virus-specific CD8⁺ responses upon influenza virus infection have been demonstrated (214, 345, 346). In older adults the number of IL-7R α^{low} effector memory CD8⁺ T cells, a subset accumulating during aging, correlated with the vaccine-induced immune response determined by antibody production. This suggests a possible implication of the IL-7R α^{low} effector memory CD8⁺ T cell population in the aging related alterations (346).

Several studies have indicated that the functionality of the CD4⁺ T cells becomes compromised, including T follicular helper cells, which are essential for an optimal B cell response. Consequently, these functional defects impair the humoral response induced by vaccination and the formation of B cell memory (347-349). It was shown that repeated vaccinations may lead to a reduced CD4⁺ T cell response, which correlates with a reduced antibody response (350).

Strategies to increase vaccine efficacy in older adults have been developed and include the use of a high dose antigen or adjuvant and alternative administration routes and will be discussed in the next section. These improved vaccines boost not only the humoral but also cell-mediated immune responses, in contrast to the standard vaccine (351). However, the mechanisms underlying the success or failure of a vaccination strategy is still not completely understood. It will also be important to gain more insights into new correlates of protection, other than virus neutralizing antibodies, that may be more suitable for predicting influenza vaccine outcome in older adults and would guide the design of new generation influenza vaccines (208).

Respiratory Syncytial Virus

Respiratory syncytial virus (RSV) belongs to the family of Pneumoviridae and is the leading cause of lower respiratory tract infection in infants and young children. RSV was not recognized as a potential problem in the adult population until outbreaks occurred in long-term care nursing facilities (352, 353). RSV causes significant burden of severe disease in older adults with 2-5% mortality rate (354). A relatively short-lived immunity after natural infection together with suboptimal/dysfunctional response of the "aged" immune system might be at the basis of the poor outcome of infections later in life. Despite the large number of candidate antivirals and vaccines against RSV in the pipeline, only a single antiviral treatment is approved, albeit with limited effectiveness and no vaccine has been licensed yet (355, 356). The correlates of immunological protection remain poorly understood which represents a major obstacle in RSV vaccine development, but antibodies to the F and G envelope proteins contribute to protective immunity (357–360). Both systemic and mucosal humoral immunity have been associated with protection (361–363). The frequency of RSV-specific CD4⁺ and CD8⁺ T cells is reduced in older adults compared to that in young adults (364–366). Furthermore, severity of the disease has been shown to correlate with reduced RSV neutralizing antibody titers and low numbers of CD8⁺ memory T cells (367, 368).

THERAPEUTIC APPROACHES TO INCREASE IMMUNITY IN OLDER ADULTS

The immune system is subject to several changes throughout life, and protects against infectious pathogens, but ultimately lose functionality by e.g., immunosenescence. Immunosenescence is partly influenced by external factors such as diet and infections, but also has a genetic component (369–375). Centenarians, for instance, seem to have gene variants that allow an optimized balance between pro- and anti-inflammatory molecules (376). The genetic study of this variation could reveal information about the immunosenescence process and how it is triggered, opening a window of opportunity for potential new therapeutic treatments (53). In addition, a personalized genetic analysis could help identify which treatment will be more effective at overcoming immunosenescence on an individual basis. These aging-related hallmarks are independent phenomena but causally connected.

Several studies have provided important insights into the biological mechanisms underlying the causes of immunosenescence and have identified possible therapeutic targets (**Figure 2**) (377–379).

Senolytic Drugs

Senescent cells are induced by cellular stress, which can occur at any age. Therefore, they can be found at any stage of life and play a beneficial role in embryogenesis, tissue repair/remodeling and tumor suppression in adulthood (380-382). Yet, secretion of soluble mediators, such as chemokines, favors the removal of senescent cells by other immune cells (e.g., macrophages and NKs) (383, 384). This process becomes less efficient with aging and results in accumulation of senescent, apoptosis-resistant cells that, together with a pro-inflammatory phenotype, contribute to tissue dysfunction and pathological manifestations (385). Chemotherapy/radiotherapy, atherosclerosis, organ transplantations or smoking, are factors among others, that accelerate immunosenescence, mainly by inducing DNA and telomere damage (386-389). In this regard, stimulation of cell death by using pharmacologically active small molecules, also known as senolytic drugs, has been identified as a promising anti-aging treatment (390). Some of these drugs inhibit the pro-survival pathways, involving the BCL-2 family members, p53 and PI3K/AKT pathways, leading to selective apoptosis of senescent cells (225, 391, 392). Dasatinib, a kinase inhibitor, and quercetin, a flavonoid targeting the PI3K/AKT, have been proven effective in eliminating senescent cells both *in vivo* and *in vitro* (393). Fisetin, another natural molecule, also appears to have senolytic properties (394). In addition, inhibitors of the chaperon HSP90, downregulate the anti-apoptotic PI3K/AKT pathway whereas a peptide altering the interaction between the transcription factor FOXO and p53, induces apoptosis of senescent cells (391, 395–398).

NF-KB Regulators

The SASP is heterogeneous, depending on the senescent cell type or mechanism that have induced senescence. Yet, some SASPassociated cytokines and growth factors are commonly secreted by different senescent cells (e.g., IL-6, IL-8, TGF- β or activin A) (71, 399-402). Interestingly, the SASP has also been shown to include secretion of extracellular vesicles, which seem to be involved in intercellular communication and inflammatory exacerbation (403-405). In this regard, another promising therapeutic approach for targeting cellular senescence is the modulation of the NF-KB regulators, such as p53 and p38 MAPK pathways and mTOR (mammalian target of rapamycin), that can mitigate the detrimental effect of the SASP by dampening the production of pro-inflammatory cytokines (e.g., IL-1, IL-6 and IL-8) (406–408). For example, the p38 MAPK pathway has been suggested to boost immunosenescence by positively regulating IFN-γ production in T cells and by altering autophagy, which causes an increase of dysfunctional cells (409, 410). First and second generation p38 inhibitors suppress SASP expression in senescent cells (411, 412). Inhibition of the sestrin-dependent MAPK activation complex has been shown to improve T cell activity in older adults, which could

be associated with an increased vaccine efficacy (413). Caloric restriction can also dampen the production of pro-inflammatory cytokines by regulating the MAPK and NF-KB pathways (414, 415). Inhibitors targeting the mTOR complex, such as the immunosuppressant drug Rapamycin and its analogues, are effective in improving the outcome of several diseases affecting older individuals (e.g., respiratory viral infections), as wells as vaccination (e.g., influenza vaccine) (407, 416). These drugs reduce the SASP while improving the innate immune response. For instance, both Rapamycin and a second-generation mTOR inhibitor RTB101, are currently under clinical investigation as possible COVID-19 treatment for adults aged >60 years of age (NCT04584710 and NCT04409327). Metformin, a widely used treatment for type II diabetes, also has been evaluated for its inhibitory effect on the SASP and as immunomodulatory agent (417). Although these drugs might offer the unique opportunity to dampen detrimental effects of aging on the immune cells or induce apoptosis of the senescent cells, it should be pointed out that their safety profile is often a matter of concern.

Improved Vaccines

Vaccination is the most effective measure to prevent infections and reduce the disease severity but unfortunately, their effectiveness appears to be lower in older adults.

The use of adjuvants as component of improved vaccines for use in older people has proven to be another strategy to partially overcome immunosenescence by increasing immunogenicity and durability of vaccine induced immune responses. A licensed



influenza vaccine containing MF59®, an oil-in-water emulsion, elicited higher antibody response in older individuals compared to the standard influenza vaccine and, interestingly, a broader antibody response also directed toward heterologous vaccine strains (103, 418, 419). Although the mode of action is not completely understood, MF59® is believed to activate innate immune cells and favor the germinal center reaction (420, 421). Another emulsion-based adjuvant, AS03, has been licensed and used during the 2009 H1N1 influenza pandemic (422). The adjuvant system AS01, component of the herpes zoster vaccine, induces antibody and cytotoxic CD8⁺ T cell responses and also activates innate immune cells (423, 424). Several other adjuvants, including Toll-like receptors agonists, are currently in the pre-clinical phase or undergoing clinical testing and have been reviewed elsewhere (425). Additional strategies, including the use of higher dose of vaccine antigen or alternative routes of vaccine administration (e.g., intradermal instead of intramuscular), have been successfully developed for influenza vaccines targeting the older population (426, 427). The improved humoral and cell-mediated immune response achieved with such vaccines has been recently confirmed by a meta-analyses conducted on 39 clinical trials, and in a head-to-head comparison clinical trial (351, 428). Promising data have been reported in subjects of >60 years of age receiving a candidate vaccine containing a recombinant HA nanoparticle produced in insect cells and formulated with a saponin-based Matrix-M adjuvant. This vaccine was well-tolerated and induced a potent immune responses when compared to the standard highdose inactivated influenza vaccine (429).

A messenger RNA (mRNA) vaccine against SARS-CoV-2 has been recently licensed and, interestingly, the immune responses induced by this vaccine in older adults, who are most at risk for developing severe disease, was similar to that obtained in younger people with over 90% protective efficacy in adults of >65 years old. Several factors may account for the excellent efficacy including an adequate involvement and stimulation of the innate immune system and increased vaccine uptake due to its formulation (430). More studies are warranted to unravel the immunological mechanisms governing this outstanding protective efficacy. These findings may be important for the future use of this vaccine platform to protect old individuals from other viral and maybe also bacterial pathogens.

The data obtained with the "improved" and also with the mRNA vaccines suggest that reduced vaccine efficacy might be just a limitation of currently used vaccination strategies and that intrinsic features of aging can be overcome with better vaccines. A more detailed understanding of changes affecting the immune system over time will provide fundamental insights into the biology of immunosenescence and would greatly facilitate the rational design of tailor-made vaccines for older adults. Moreover, the presence of chronic diseases and frailty, defined as a decline of function across multiple organ systems, may be better predictors of poor vaccine immunogenicity than age alone (431).

"Trained immunity" or "innate immune memory" is a recent and interesting concept that defines the ability of innate cells to form immune memory and therefore displaying characteristics of adaptive immunity (64, 432, 433). Epigenetic and metabolic mechanisms are thought to be responsible for this phenomenon. An increasing number of studies have shown that several vaccines such as those against smallpox, oral polio, measles and tuberculosis (Bacillus Calmette-Guérin, BCG), induce non-specific protective effects also against heterologous, "off-target", infectious diseases (434, 435). Interestingly, BCG vaccination reduces respiratory infections in older adults and recent data have demonstrated that the innate immune function is less affected by aging than adaptive immunity, suggesting that trained immunity may be functional in this age group (436, 437). Moreover, BCG vaccination has been shown to increase the immunogenicity of subsequent influenza vaccination, highlighting its adjuvant-like property (438). In a recent review, "trained immunity" was proposed as a new target to enhance immune responses in older adults (439). Although data are still scarce and more insights are needed, it may represent a valuable target also for designing better vaccines. However, it cannot be excluded that the excess of cytokines induced by activation of innate cells might have negative effects on the immune system of older adults.

CONCLUSIONS

The ongoing COVID-19 pandemic and its tremendous impact on older individuals highlights the need for the development of therapeutic and preventive measures to protect this vulnerable age group. Therefore, it is of utmost importance to advance our understanding of the mechanisms (molecular, cellular, genetic, environmental, etc.) responsible for aging and the development of immunosenescence, which will aid developing novel medical interventions. Intrinsic and extrinsic cell stress factors induce cellular senescence, which is mainly characterized by cell cycle arrest (G_0/G_1) and secretion of soluble factors (SASP) in the extracellular environment (440). Despite recent technological advances, phenotypical characterization of senescent cells is still challenging due to a lack of defined and univocal/universal biomarkers but also for their heterogeneous nature and dynamic evolution. It is still not clear how many different subsets of senescent cells can be identified and their distribution in tissues and periphery. Although some interesting animal models to study cellular senescence have been developed, their results might not be easily translated to the human situation (441, 442). To obtain a better insight into the complexity of immune ageing/immunosenescence and thus the possibility to decipher and dissect mechanisms and cells involved, requires the use of multiple scientific approaches and disciplines (e.g., high-resolution omicstechnologies, multiparametric analyses, system biology and big data analyses). With the information obtained, better prophylactic and therapeutic interventions can be developed to prevent deterioration of the health status, often associated with aging and immunosenescence. This way, the quality of life in this age group can be maintained and mortality, including that caused by infectious agents, reduced.

AUTHOR CONTRIBUTIONS

MP-P, AO, TB, HE, GR, and GS conceptualized and composed the manuscript. GR and GS supervised all aspects of the manuscript preparation. All authors contributed to the article and approved the submitted version.

FUNDING

This work was supported by the Alexander von Humboldt Foundation in the framework of the Alexander von Humboldt Professorship endowed by the German Federal Ministry of

REFERENCES

- 1. Walford RL. Immunologic Aspects of Aging. Klin Wochenschr (1969) 47 (11):599–605. doi: 10.1007/BF01876949
- Hainz U, Jenewein B, Asch E, Pfeiffer KP, Berger P, Grubeck-Loebenstein B. Insufficient Protection for Healthy Elderly Adults by Tetanus and TBE Vaccines. Vaccine (2005) 23(25):3232–5. doi: 10.1016/j.vaccine.2005.01.085
- Song JY, Cheong HJ, Hwang IS, Choi WS, Jo YM, Park DW, et al. Long-Term Immunogenicity of Influenza Vaccine Among the Elderly: Risk Factors for Poor Immune Response and Persistence. *Vaccine* (2010) 28 (23):3929–35. doi: 10.1016/j.vaccine.2010.03.067
- Tohme RA, Awosika-Olumo D, Nielsen C, Khuwaja S, Scott J, Xing J, et al. Evaluation of Hepatitis B Vaccine Immunogenicity Among Older Adults During an Outbreak Response in Assisted Living Facilities. *Vaccine* (2011) 29(50):9316–20. doi: 10.1016/j.vaccine.2011.10.011
- Van Der Meeren O, Crasta P, Cheuvart B, De Ridder M. Characterization of an Age-Response Relationship to GSK's Recombinant Hepatitis B Vaccine in Healthy Adults: An Integrated Analysis. *Hum Vaccin Immunother* (2015) 11(7):1726–9. doi: 10.1080/21645515.2015.1039758
- Yang S, Tian G, Cui Y, Ding C, Deng M, Yu C, et al. Factors Influencing Immunologic Response to Hepatitis B Vaccine in Adults. *Sci Rep* (2016) 6:27251. doi: 10.1038/srep27251
- 7. Pawelec G. Age and Immunity: What is "Immunosenescence"? *Exp Gerontol* (2018) 105:4–9. doi: 10.1016/j.exger.2017.10.024
- Dodig S, Cepelak I, Pavic I. Hallmarks of Senescence and Aging. Biochem Med (Zagreb) (2019) 29(3):030501. doi: 10.11613/BM.2019.030501
- Weinberg A, Pang L, Johnson MJ, Caldas Y, Cho A, Tovar-Salazar A, et al. The Effect of Age on the Immunogenicity of the Live Attenuated Zoster Vaccine Is Predicted by Baseline Regulatory T Cells and Varicella-Zoster Virus-Specific T Cell Immunity. *J Virol* (2019) 93(15):e00305–19. doi: 10.1128/JVI.00305-19
- Roebuck J. When Does Old-Age Begin Evolution of the English Definition. J Soc History (1979) 12(3):416–28. doi: 10.1353/jsh/12.3.416
- Steens A, Eriksen HM, Blystad H. What are the Most Important Infectious Diseases Among Those >/=65 Years: A Comprehensive Analysis on Notifiable Diseases, Norway, 1993-2011. BMC Infect Dis (2014) 14:57. doi: 10.1186/1471-2334-14-57
- Beard JR, Officer A, de Carvalho IA, Sadana R, Pot AM, Michel JP, et al. The World Report on Ageing and Health: A Policy Framework for Healthy Ageing. *Lancet* (2016) 387(10033):2145–54. doi: 10.1016/S0140-6736(15)00516-4
- Demicheli V, Jefferson T, Di Pietrantonj C, Ferroni E, Thorning S, Thomas RE, et al. Vaccines for Preventing Influenza in the Elderly. *Cochrane Database Syst Rev* (2018) 2:CD004876. doi: 10.1002/14651858.CD004876.pub4
- Akbar AN, Gilroy DW. Aging Immunity may Exacerbate COVID-19. Science (2020) 369(6501):256–7. doi: 10.1126/science.abb0762
- Fried MW, Crawford JM, Mospan AR, Watkins SE, Munoz Hernandez B, Zink RC, et al. Patient Characteristics and Outcomes of 11,721 Patients With COVID19 Hospitalized Across the United States. *Clin Infect Dis* (2020) ciaa1268. doi: 10.1093/cid/ciaa1268
- Lee JY, Kim HA, Huh K, Hyun M, Rhee JY, Jang S, et al. Risk Factors for Mortality and Respiratory Support in Elderly Patients Hospitalized With COVID-19 in Korea. J Korean Med Sci (2020) 35(23):e223. doi: 10.3346/ jkms.2020.35.e223
- Wang W, Xu Y, Gao R, Lu R, Han K, Wu G, et al. Detection of SARS-CoV-2 in Different Types of Clinical Specimens. *JAMA* (2020) 323(18):1843–4. doi: 10.1001/jama.2020.3786
- Channappanavar R, Perlman S. Age-Related Susceptibility to Coronavirus Infections: Role of Impaired and Dysregulated Host Immunity. J Clin Invest (2020) 130(12):6204–13. doi: 10.1172/JCI144115

Education and Research; the European Union's Horizon 2020 research and innovation program under grant agreement No 848166 (ISOLDA) and the German Research Foundation, Excellence Strategy program-EXC 2155 Resolving Infection Susceptibility, "RESIST".

- Saletti G, Gerlach T, Jansen JM, Molle A, Elbahesh H, Ludlow M, et al. Older Adults Lack SARS CoV-2 Cross-Reactive T Lymphocytes Directed to Human Coronaviruses OC43 and NL63. *Sci Rep* (2020) 10(1):21447. doi: 10.1038/s41598-020-78506-9
- Amsterdam D, Ostrov BE. The Impact of the Microbiome on Immunosenescence. Immunol Invest (2018) 47(8):801–11. doi: 10.1080/08820139.2018.1537570
- Del Giudice G, Goronzy JJ, Grubeck-Loebenstein B, Lambert PH, Mrkvan T, Stoddard JJ, et al. Fighting Against a Protean Enemy: Immunosenescence, Vaccines, and Healthy Aging. *NPJ Aging Mech Dis* (2018) 4:1. doi: 10.1038/ s41514-017-0020-0
- 22. Muller L, Fulop T, Pawelec G. Immunosenescence in Vertebrates and Invertebrates. *Immun Ageing* (2013) 10(1):12. doi: 10.1186/1742-4933-10-12
- Holder A, Mirczuk SM, Fowkes RC, Palmer DB, Aspinall R, Catchpole B. Perturbation of the T Cell Receptor Repertoire Occurs With Increasing Age in Dogs. *Dev Comp Immunol* (2018) 79:150–7. doi: 10.1016/j.dci.2017. 10.020
- Mietsch M, Paque K, Drummer C, Stahl-Hennig C, Roshani B. The Aging Common Marmoset's Immune System: From Junior to Senior. Am J Primatol (2020) 82(6):e23128. doi: 10.1002/ajp.23128
- Schwarz TF, Volpe S, Catteau G, Chlibek R, David MP, Richardus JH, et al. Persistence of Immune Response to an Adjuvanted Varicella-Zoster Virus Subunit Vaccine for Up to Year Nine in Older Adults. *Hum Vaccin Immunother* (2018) 14(6):1370–7. doi: 10.1080/21645515.2018.1442162
- 26. Arai Y, Martin-Ruiz CM, Takayama M, Abe Y, Takebayashi T, Koyasu S, et al. Inflammation, But Not Telomere Length, Predicts Successful Ageing At Extreme Old Age: A Longitudinal Study of Semi-Supercentenarians. *EBioMedicine* (2015) 2(10):1549–58. doi: 10.1016/j.ebiom.2015.07.029
- Niwa Y, Kasama T, Miyachi Y, Kanoh T. Neutrophil Chemotaxis, Phagocytosis and Parameters of Reactive Oxygen Species in Human Aging: Cross-Sectional and Longitudinal Studies. *Life Sci* (1989) 44 (22):1655–64. doi: 10.1016/0024-3205(89)90482-7
- Wenisch C, Patruta S, Daxbock F, Krause R, Horl W. Effect of Age on Human Neutrophil Function. J Leukoc Biol (2000) 67(1):40–5. doi: 10.1002/ jlb.67.1.40
- Hazeldine J, Harris P, Chapple IL, Grant M, Greenwood H, Livesey A, et al. Impaired Neutrophil Extracellular Trap Formation: A Novel Defect in the Innate Immune System of Aged Individuals. *Aging Cell* (2014) 13(4):690–8. doi: 10.1111/acel.12222
- Agrawal A, Agrawal S, Gupta S. Role of Dendritic Cells in Inflammation and Loss of Tolerance in the Elderly. *Front Immunol* (2017) 8:896. doi: 10.3389/ fimmu.2017.00896
- Drew W, Wilson DV, Sapey E. Inflammation and Neutrophil Immunosenescence in Health and Disease: Targeted Treatments to Improve Clinical Outcomes in the Elderly. *Exp Gerontol* (2018) 105:70–7. doi: 10.1016/j.exger.2017.12.020
- 32. Sapey E, Greenwood H, Walton G, Mann E, Love A, Aaronson N, et al. Phosphoinositide 3-Kinase Inhibition Restores Neutrophil Accuracy in the Elderly: Toward Targeted Treatments for Immunosenescence. *Blood* (2014) 123(2):239–48. doi: 10.1182/blood-2013-08-519520
- Ward JR, Heath PR, Catto JW, Whyte MK, Milo M, Renshaw SA. Regulation of Neutrophil Senescence by Micrornas. *PLoS One* (2011) 6(1):e15810. doi: 10.1371/journal.pone.0015810
- Pepper M, Jenkins MK. Origins of CD4(+) Effector and Central Memory T Cells. Nat Immunol (2011) 12(6):467–71. doi: 10.1038/ni.2038
- Samji T, Khanna KM. Understanding Memory CD8(+) T Cells. Immunol Lett (2017) 185:32–9. doi: 10.1016/j.imlet.2017.02.012
- Johnson ED, Cole GA. Functional Heterogeneity of Lymphocytic Choriomeningitis Virus-Specfic T Lymphocytes. I. Identification of Effector Amd Memory Subsets. J Exp Med (1975) 141(4):866–81. doi: 10.1084/jem.141.4.866

- Haynes L, Eaton SM, Burns EM, Randall TD, Swain SL. Cd4 T Cell Memory Derived From Young Naive Cells Functions Well Into Old Age, But Memory Generated From Aged Naive Cells Functions Poorly. *Proc Natl Acad Sci USA* (2003) 100(25):15053–8. doi: 10.1073/pnas.2433717100
- Goronzy JJ, Lee WW, Weyand CM. Aging and T-cell Diversity. *Exp Gerontol* (2007) 42(5):400–6. doi: 10.1016/j.exger.2006.11.016
- Chidrawar S, Khan N, Wei W, McLarnon A, Smith N, Nayak L, et al. Cytomegalovirus-Seropositivity has a Profound Influence on the Magnitude of Major Lymphoid Subsets Within Healthy Individuals. *Clin Exp Immunol* (2009) 155(3):423–32. doi: 10.1111/j.1365-2249.2008.03785.x
- Nikolich-Zugich J. Aging of the T Cell Compartment in Mice and Humans: From No Naive Expectations to Foggy Memories. *J Immunol* (2014) 193 (6):2622–9. doi: 10.4049/jimmunol.1401174
- Dugan HL, Henry C, Wilson PC. Aging and Influenza Vaccine-Induced Immunity. Cell Immunol (2020) 348:103998. doi: 10.1016/j.cellimm.2019.103998
- Qi Q, Zhang DW, Weyand CM, Goronzy JJ. Mechanisms Shaping the Naive T Cell Repertoire in the Elderly - Thymic Involution or Peripheral Homeostatic Proliferation? *Exp Gerontol* (2014) 54:71–4. doi: 10.1016/ j.exger.2014.01.005
- Boucher N, Dufeu-Duchesne T, Vicaut E, Farge D, Effros RB, Schachter F. CD28 Expression in T Cell Aging and Human Longevity. *Exp Gerontol* (1998) 33(3):267–82. doi: 10.1016/S0531-5565(97)00132-0
- Marcotte R, Lacelle C, Wang E. Senescent Fibroblasts Resist Apoptosis by Downregulating Caspase-3. *Mech Ageing Dev* (2004) 125(10-11):777–83. doi: 10.1016/j.mad.2004.07.007
- Shay JW, Wright WE. Senescence and Immortalization: Role of Telomeres and Telomerase. *Carcinogenesis* (2005) 26(5):867–74. doi: 10.1093/carcin/ bgh296
- Vallejo AN. CD28 Extinction in Human T Cells: Altered Functions and the Program of T-cell Senescence. *Immunol Rev* (2005) 205:158–69. doi: 10.1111/j.0105-2896.2005.00256.x
- Murata Y, Wakoh T, Uekawa N, Sugimoto M, Asai A, Miyazaki T, et al. Death-Associated Protein 3 Regulates Cellular Senescence Through Oxidative Stress Response. *FEBS Lett* (2006) 580(26):6093–9. doi: 10.1016/ j.febslet.2006.10.004
- Takahashi A, Ohtani N, Yamakoshi K, Iida S, Tahara H, Nakayama K, et al. Mitogenic Signalling and the P16ink4a-Rb Pathway Cooperate to Enforce Irreversible Cellular Senescence. *Nat Cell Biol* (2006) 8(11):1291–7. doi: 10.1038/ncb1491
- Demidenko ZN, Blagosklonny MV. Growth Stimulation Leads to Cellular Senescence When the Cell Cycle is Blocked. *Cell Cycle* (2008) 7(21):3355–61. doi: 10.4161/cc.7.21.6919
- Henson SM, Akbar AN. KLRG1–More Than a Marker for T Cell Senescence. Age (Dordr) (2009) 31(4):285–91. doi: 10.1007/s11357-009-9100-9
- Pangrazzi L, Weinberger B. T Cells, Aging and Senescence. Exp Gerontol (2020) 134:110887. doi: 10.1016/j.exger.2020.110887
- Zhao Y, Shao Q, Peng G. Exhaustion and Senescence: Two Crucial Dysfunctional States of T Cells in the Tumor Microenvironment. *Cell Mol Immunol* (2020) 17(1):27–35. doi: 10.1038/s41423-019-0344-8
- He S, Sharpless NE. Senescence in Health and Disease. Cell (2017) 169 (6):1000–11. doi: 10.1016/j.cell.2017.05.015
- Decman V, Laidlaw BJ, Dimenna LJ, Abdulla S, Mozdzanowska K, Erikson J, et al. Cell-Intrinsic Defects in the Proliferative Response of Antiviral Memory CD8 T Cells in Aged Mice Upon Secondary Infection. *J Immunol* (2010) 184(9):5151–9. doi: 10.4049/jimmunol.0902063
- Tahir S, Fukushima Y, Sakamoto K, Sato K, Fujita H, Inoue J, et al. A Cd153 +Cd4+ T Follicular Cell Population With Cell-Senescence Features Plays a Crucial Role in Lupus Pathogenesis Via Osteopontin Production. *J Immunol* (2015) 194(12):5725–35. doi: 10.4049/jimmunol.1500319
- Ferrucci L, Fabbri E. Inflammageing: Chronic Inflammation in Ageing, Cardiovascular Disease, and Frailty. *Nat Rev Cardiol* (2018) 15(9):505–22. doi: 10.1038/s41569-018-0064-2
- Franceschi C, Bonafe M, Valensin S, Olivieri F, De Luca M, Ottaviani E, et al. Inflamm-Aging. An Evolutionary Perspective on Immunosenescence. *Ann* N Y Acad Sci (2000) 908:244–54. doi: 10.1111/j.1749-6632.2000.tb06651.x
- Chaleckis R, Murakami I, Takada J, Kondoh H, Yanagida M. Individual Variability in Human Blood Metabolites Identifies Age-Related Differences. *Proc Natl Acad Sci USA* (2016) 113(16):4252–9. doi: 10.1073/pnas.1603023113

- Milan-Mattos JC, Anibal FF, Perseguini NM, Minatel V, Rehder-Santos P, Castro CA, et al. Effects of Natural Aging and Gender on Pro-Inflammatory Markers. *Braz J Med Biol Res* (2019) 52(9):e8392. doi: 10.1590/1414-431x20198392
- Franceschi C, Campisi J. Chronic Inflammation (Inflammaging) and its Potential Contribution to Age-Associated Diseases. J Gerontol A Biol Sci Med Sci (2014) 69(Suppl 1):S4–9. doi: 10.1093/gerona/glu057
- Mejias NH, Martinez CC, Stephens ME, de Rivero Vaccari JP. Contribution of the Inflammasome to Inflammaging. J Inflammation (Lond) (2018) 15:23. doi: 10.1186/s12950-018-0198-3
- Ciabattini A, Nardini C, Santoro F, Garagnani P, Franceschi C, Medaglini D. Vaccination in the Elderly: The Challenge of Immune Changes With Aging. Semin Immunol (2018) 40:83–94. doi: 10.1016/j.smim.2018.10.010
- Chung HY, Cesari M, Anton S, Marzetti E, Giovannini S, Seo AY, et al. Molecular Inflammation: Underpinnings of Aging and Age-Related Diseases. Ageing Res Rev (2009) 8(1):18–30. doi: 10.1016/j.arr.2008.07.002
- 64. Franceschi C, Salvioli S, Garagnani P, de Eguileor M, Monti D, Capri M. Immunobiography and the Heterogeneity of Immune Responses in the Elderly: A Focus on Inflammaging and Trained Immunity. *Front Immunol* (2017) 8:982. doi: 10.3389/fimmu.2017.00982
- 65. Franceschi C, Garagnani P, Morsiani C, Conte M, Santoro A, Grignolio A, et al. The Continuum of Aging and Age-Related Diseases: Common Mechanisms But Different Rates. *Front Med (Lausanne)* (2018) 5:61. doi: 10.3389/fmed.2018.00061
- Calder PC, Bosco N, Bourdet-Sicard R, Capuron L, Delzenne N, Dore J, et al. Health Relevance of the Modification of Low Grade Inflammation in Ageing (Inflammageing) and the Role of Nutrition. *Ageing Res Rev* (2017) 40:95– 119. doi: 10.1016/j.arr.2017.09.001
- Franceschi C, Garagnani P, Parini P, Giuliani C, Santoro A. Inflammaging: A New Immune-Metabolic Viewpoint for Age-Related Diseases. *Nat Rev Endocrinol* (2018) 14(10):576–90. doi: 10.1038/s41574-018-0059-4
- Furman D, Campisi J, Verdin E, Carrera-Bastos P, Targ S, Franceschi C, et al. Chronic Inflammation in the Etiology of Disease Across the Life Span. *Nat Med* (2019) 25(12):1822–32. doi: 10.1038/s41591-019-0675-0
- Rea IM, Gibson DS, McGilligan V, McNerlan SE, Alexander HD, Ross OA. Age and Age-Related Diseases: Role of Inflammation Triggers and Cytokines. *Front Immunol* (2018) 9:586. doi: 10.3389/fimmu.2018.00586
- Bektas A, Schurman SH, Sen R, Ferrucci L. Human T Cell Immunosenescence and Inflammation in Aging. *J Leukoc Biol* (2017) 102(4):977–88. doi: 10.1189/ jlb.3RI0716-335R
- Coppe JP, Patil CK, Rodier F, Sun Y, Munoz DP, Goldstein J, et al. Senescence-Associated Secretory Phenotypes Reveal Cell-Nonautonomous Functions of Oncogenic RAS and the p53 Tumor Suppressor. *PLoS Biol* (2008) 6(12):2853–68. doi: 10.1371/journal.pbio.0060301
- Fulop T, Larbi A, Dupuis G, Le Page A, Frost EH, Cohen AA, et al. Immunosenescence and Inflamm-Aging as Two Sides of the Same Coin: Friends or Foes? *Front Immunol* (2017) 8:1960. doi: 10.3389/fimmu.2017.01960
- 73. Wikby A, Nilsson BO, Forsey R, Thompson J, Strindhall J, Lofgren S, et al. The Immune Risk Phenotype is Associated With IL-6 in the Terminal Decline Stage: Findings From the Swedish NONA Immune Longitudinal Study of Very Late Life Functioning. *Mech Ageing Dev* (2006) 127(8):695– 704. doi: 10.1016/j.mad.2006.04.003
- Klenerman P, Oxenius A. T Cell Responses to Cytomegalovirus. Nat Rev Immunol (2016) 16(6):367–77. doi: 10.1038/nri.2016.38
- 75. Di Benedetto S, Derhovanessian E, Steinhagen-Thiessen E, Goldeck D, Muller L, Pawelec G. Impact of Age, Sex and CMV-infection on Peripheral T Cell Phenotypes: Results From the Berlin Base-II Study. *Biogerontology* (2015) 16(5):631–43. doi: 10.1007/s10522-015-9563-2
- 76. Spyridopoulos I, Martin-Ruiz C, Hilkens C, Yadegarfar ME, Isaacs J, Jagger C, et al. CMV Seropositivity and T-cell Senescence Predict Increased Cardiovascular Mortality in Octogenarians: Results From the Newcastle 85+ Study. Aging Cell (2016) 15(2):389–92. doi: 10.1111/acel.12430
- Ouyang Q, Wagner WM, Wikby A, Walter S, Aubert G, Dodi AI, et al. Large Numbers of Dysfunctional CD8+ T Lymphocytes Bearing Receptors for a Single Dominant CMV Epitope in the Very Old. J Clin Immunol (2003) 23 (4):247–57. doi: 10.1023/A:1024580531705
- Bauer ME, Fuente Mde L. The Role of Oxidative and Inflammatory Stress and Persistent Viral Infections in Immunosenescence. *Mech Ageing Dev* (2016) 158:27–37. doi: 10.1016/j.mad.2016.01.001

- Goronzy JJ, Weyand CM. Understanding Immunosenescence to Improve Responses to Vaccines. *Nat Immunol* (2013) 14(5):428–36. doi: 10.1038/ ni.2588
- Gibson KL, Wu YC, Barnett Y, Duggan O, Vaughan R, Kondeatis E, et al. B-Cell Diversity Decreases in Old Age and is Correlated With Poor Health Status. Aging Cell (2009) 8(1):18–25. doi: 10.1111/j.1474-9726.2008.00443.x
- Frasca D, Diaz A, Romero M, Landin AM, Blomberg BB. High TNF-alpha Levels in Resting B Cells Negatively Correlate With Their Response. *Exp Gerontol* (2014) 54:116–22. doi: 10.1016/j.exger.2014.01.004
- Frasca D, Blomberg BB, Garcia D, Keilich SR, Haynes L. Age-Related Factors That Affect B Cell Responses to Vaccination in Mice and Humans. *Immunol Rev* (2020) 296(1):142–54. doi: 10.1111/imr.12864
- Hope-Simpson RE. The Nature of Herpes Zoster: A Long-Term Study and a New Hypothesis. Proc R Soc Med (1965) 58:9–20. doi: 10.1177/ 003591576505800106
- Gershon AA, Breuer J, Cohen JI, Cohrs RJ, Gershon MD, Gilden D, et al. Varicella Zoster Virus Infection. *Nat Rev Dis Primers* (2015) 1:15016. doi: 10.1038/nrdp.2015.16
- Markus A, Lebenthal-Loinger I, Yang IH, Kinchington PR, Goldstein RS. An In Vitro Model of Latency and Reactivation of Varicella Zoster Virus in Human Stem Cell-Derived Neurons. *PLoS Pathog* (2015) 11(6):e1004885. doi: 10.1371/journal.ppat.1004885
- Depledge DP, Sadaoka T, Ouwendijk WJD. Molecular Aspects of Varicella-Zoster Virus Latency. Viruses (2018) 10(7):349. doi: 10.3390/v10070349
- Ouwendijk WJD, van Veen S, Mehraban T, Mahalingam R, Verjans G. Simian Varicella Virus Infects Enteric Neurons and Alpha4beta7 Integrin-Expressing Gut-Tropic T-Cells in Nonhuman Primates. *Viruses* (2018) 10 (4):156. doi: 10.3390/v10040156
- Huning S, von Ducker L, Kohl WK, Nashan D. Therapy of Herpes Zoster and Postherpetic Neuralgia. *Hautarzt* (2019) 70(8):645–56. doi: 10.1007/ s00105-019-4446-0
- Looney RJ, Falsey A, Campbell D, Torres A, Kolassa J, Brower C, et al. Role of Cytomegalovirus in the T Cell Changes Seen in Elderly Individuals. *Clin Immunol* (1999) 90(2):213–9. doi: 10.1006/clim.1998.4638
- Cannon MJ, Schmid DS, Hyde TB. Review of Cytomegalovirus Seroprevalence and Demographic Characteristics Associated With Infection. *Rev Med Virol* (2010) 20(4):202–13. doi: 10.1002/rmv.655
- Nikolich-Zugich J, Goodrum F, Knox K, Smithey MJ. Known Unknowns: How Might the Persistent Herpesvirome Shape Immunity and Aging? *Curr Opin Immunol* (2017) 48:23–30. doi: 10.1016/j.coi.2017.07.011
- Komatsu H, Sierro S, A VC, Klenerman P. Population Analysis of Antiviral T Cell Responses Using MHC Class I-peptide Tetramers. *Clin Exp Immunol* (2003) 134(1):9–12. doi: 10.1046/j.1365-2249.2003.02266.x
- Schober K, Voit F, Grassmann S, Muller TR, Eggert J, Jarosch S, et al. Reverse TCR Repertoire Evolution Toward Dominant Low-Affinity Clones During Chronic CMV Infection. *Nat Immunol* (2020) 21(4):434–41. doi: 10.1038/ s41590-020-0628-2
- 94. Pawelec G, Akbar A, Beverley P, Caruso C, Derhovanessian E, Fulop T, et al. Immunosenescence and Cytomegalovirus: Where do We Stand After a Decade? *Immun Ageing* (2010) 7:13. doi: 10.1186/1742-4933-7-13
- Moss P. The Emerging Role of Cytomegalovirus in Driving Immune Senescence: A Novel Therapeutic Opportunity for Improving Health in the Elderly. *Curr Opin Immunol* (2010) 22(4):529–34. doi: 10.1016/j.coi.2010.07.001
- 96. Khan N, Shariff N, Cobbold M, Bruton R, Ainsworth JA, Sinclair AJ, et al. Cytomegalovirus Seropositivity Drives the CD8 T Cell Repertoire Toward Greater Clonality in Healthy Elderly Individuals. *J Immunol* (2002) 169 (4):1984–92. doi: 10.4049/jimmunol.169.4.1984
- Thompson WW, Shay DK, Weintraub E, Brammer L, Cox N, Anderson LJ, et al. Mortality Associated With Influenza and Respiratory Syncytial Virus in the United States. *JAMA* (2003) 289(2):179–86. doi: 10.1001/jama.289.2.179
- Falsey AR, Hennessey PA, Formica MA, Cox C, Walsh EE. Respiratory Syncytial Virus Infection in Elderly and High-Risk Adults. N Engl J Med (2005) 352(17):1749–59. doi: 10.1056/NEJMoa043951
- Frasca D, Blomberg BB. Aging Induces B Cell Defects and Decreased Antibody Responses to Influenza Infection and Vaccination. *Immun* Ageing (2020) 17(1):37. doi: 10.1186/s12979-020-00210-z
- Iuliano AD, Roguski KM, Chang HH, Muscatello DJ, Palekar R, Tempia S, et al. Estimates of Global Seasonal Influenza-Associated Respiratory

Mortality: A Modelling Study. Lancet (2018) 391(10127):1285-300. doi: 10.1016/S0140-6736(17)33293-2

- 101. Wong CK, Smith CA, Sakamoto K, Kaminski N, Koff JL, Goldstein DR. Aging Impairs Alveolar Macrophage Phagocytosis and Increases Influenza-Induced Mortality in Mice. *J Immunol* (2017) 199(3):1060–8. doi: 10.4049/ jimmunol.1700397
- 102. Ackerson B, Tseng HF, Sy LS, Solano Z, Slezak J, Luo Y, et al. Severe Morbidity and Mortality Associated With Respiratory Syncytial Virus Versus Influenza Infection in Hospitalized Older Adults. *Clin Infect Dis* (2019) 69(2):197–203. doi: 10.1093/cid/ciy991
- 103. Domnich A, Arata L, Amicizia D, Puig-Barbera J, Gasparini R, Panatto D. Effectiveness of MF59-adjuvanted Seasonal Influenza Vaccine in the Elderly: A Systematic Review and Meta-Analysis. *Vaccine* (2017) 35(4):513–20. doi: 10.1016/j.vaccine.2016.12.011
- Dunkle LM, Izikson R, Patriarca P, Goldenthal KL, Muse D, Callahan J, et al. Efficacy of Recombinant Influenza Vaccine in Adults 50 Years of Age or Older. N Engl J Med (2017) 376(25):2427–36. doi: 10.1056/NEJMoa1608862
- Chang LJ, Meng Y, Janosczyk H, Landolfi V, Talbot HK, Group QHDS. Safety and Immunogenicity of High-Dose Quadrivalent Influenza Vaccine in Adults >/=65years of Age: A Phase 3 Randomized Clinical Trial. Vaccine (2019) 37(39):5825–34. doi: 10.1016/j.vaccine.2019.08.016
- 106. Smetana J, Chlibek R, Shaw J, Splino M, Prymula R. Influenza Vaccination in the Elderly. *Hum Vaccin Immunother* (2018) 14(3):540–9. doi: 10.1080/ 21645515.2017.1343226
- 107. Williams K, Bastian AR, Feldman RA, Omoruyi E, de Paepe E, Hendriks J, et al. Phase 1 Safety and Immunogenicity Study of a Respiratory Syncytial Virus Vaccine With an Adenovirus 26 Vector Encoding Prefusion F (Ad26.RSV.preF) in Adults Aged >/=60 Years. J Infect Dis (2020) 222 (6):979–88. doi: 10.1093/infdis/jiaa193
- 108. Cicconi P, Jones C, Sarkar E, Silva-Reyes L, Klenerman P, de Lara C, et al. First-in-Human Randomized Study to Assess the Safety and Immunogenicity of an Investigational Respiratory Syncytial Virus (Rsv) Vaccine Based on Chimpanzee-Adenovirus-155 Viral Vector-Expressing Rsv Fusion, Nucleocapsid, and Antitermination Viral Proteins in Healthy Adults. *Clin Infect Dis* (2020) 70(10):2073–81. doi: 10.1093/cid/ciz653
- 109. Langley JM, MacDonald LD, Weir GM, MacKinnon-Cameron D, Ye L, McNeil S, et al. A Respiratory Syncytial Virus Vaccine Based on the Small Hydrophobic Protein Ectodomain Presented With a Novel Lipid-Based Formulation is Highly Immunogenic and Safe in Adults: A First-in-Humans Study. J Infect Dis (2018) 218(3):378–87. doi: 10.1093/infdis/jiy177
- 110. Mazur NI, Higgins D, Nunes MC, Melero JA, Langedijk AC, Horsley N, et al. The Respiratory Syncytial Virus Vaccine Landscape: Lessons From the Graveyard and Promising Candidates. *Lancet Infect Dis* (2018) 18(10): e295–311. doi: 10.1016/S1473-3099(18)30292-5
- 111. Aiello A, Farzaneh F, Candore G, Caruso C, Davinelli S, Gambino CM, et al. Immunosenescence and Its Hallmarks: How to Oppose Aging Strategically? A Review of Potential Options for Therapeutic Intervention. *Front Immunol* (2019) 10:2247. doi: 10.3389/fimmu.2019.02247
- Mildner A, Jung S. Development and Function of Dendritic Cell Subsets. *Immunity* (2014) 40(5):642–56. doi: 10.1016/j.immuni.2014.04.016
- Balan S, Saxena M, Bhardwaj N. Dendritic Cell Subsets and Locations. Int Rev Cell Mol Biol (2019) 348:1–68. doi: 10.1016/bs.ircmb.2019.07.004
- 114. Qian C, Cao X. Dendritic Cells in the Regulation of Immunity and Inflammation. Semin Immunol (2018) 35:3–11. doi: 10.1016/j.smim.2017. 12.002
- 115. Basit F, Mathan T, Sancho D, de Vries IJM. Human Dendritic Cell Subsets Undergo Distinct Metabolic Reprogramming for Immune Response. Front Immunol (2018) 9:2489. doi: 10.3389/fimmu.2018.02489
- Swiecki M, Colonna M. The Multifaceted Biology of Plasmacytoid Dendritic Cells. Nat Rev Immunol (2015) 15(8):471–85. doi: 10.1038/nri3865
- 117. Merad M, Sathe P, Helft J, Miller J, Mortha A. The Dendritic Cell Lineage: Ontogeny and Function of Dendritic Cells and Their Subsets in the Steady State and the Inflamed Setting. *Annu Rev Immunol* (2013) 31:563–604. doi: 10.1146/annurev-immunol-020711-074950
- 118. Hildner K, Edelson BT, Purtha WE, Diamond M, Matsushita H, Kohyama M, et al. Batf3 Deficiency Reveals a Critical Role for CD8alpha+ Dendritic Cells in Cytotoxic T Cell Immunity. *Science* (2008) 322(5904):1097–100. doi: 10.1126/science.1164206

- 119. Della Bella S, Bierti L, Presicce P, Arienti R, Valenti M, Saresella M, et al. Peripheral Blood Dendritic Cells and Monocytes are Differently Regulated in the Elderly. *Clin Immunol* (2007) 122(2):220–8. doi: 10.1016/j.clim.2006.09.012
- 120. Jing Y, Shaheen E, Drake RR, Chen N, Gravenstein S, Deng Y. Aging is Associated With a Numerical and Functional Decline in Plasmacytoid Dendritic Cells, Whereas Myeloid Dendritic Cells are Relatively Unaltered in Human Peripheral Blood. *Hum Immunol* (2009) 70(10):777–84. doi: 10.1016/j.humimm.2009.07.005
- 121. Sridharan A, Esposo M, Kaushal K, Tay J, Osann K, Agrawal S, et al. Age-Associated Impaired Plasmacytoid Dendritic Cell Functions Lead to Decreased CD4 and CD8 T Cell Immunity. Age (Dordr) (2011) 33(3):363– 76. doi: 10.1007/s11357-010-9191-3
- 122. Metcalf TU, Cubas RA, Ghneim K, Cartwright MJ, Grevenynghe JV, Richner JM, et al. Global Analyses Revealed Age-Related Alterations in Innate Immune Responses After Stimulation of Pathogen Recognition Receptors. *Aging Cell* (2015) 14(3):421–32. doi: 10.1111/acel.12320
- 123. Agrawal A, Agrawal S, Cao JN, Su H, Osann K, Gupta S. Altered Innate Immune Functioning of Dendritic Cells in Elderly Humans: A Role of Phosphoinositide 3-Kinase-Signaling Pathway. J Immunol (2007) 178 (11):6912–22. doi: 10.4049/jimmunol.178.11.6912
- 124. Agrawal A, Tay J, Ton S, Agrawal S, Gupta S. Increased Reactivity of Dendritic Cells From Aged Subjects to Self-Antigen, the Human DNA. *J Immunol* (2009) 182(2):1138–45. doi: 10.4049/jimmunol.182.2.1138
- 125. van Duin D, Mohanty S, Thomas V, Ginter S, Montgomery RR, Fikrig E, et al. Age-Associated Defect in Human TLR-1/2 Function. *J Immunol* (2007) 178(2):970–5. doi: 10.4049/jimmunol.178.2.970
- 126. Panda A, Qian F, Mohanty S, van Duin D, Newman FK, Zhang L, et al. Age-Associated Decrease in TLR Function in Primary Human Dendritic Cells Predicts Influenza Vaccine Response. *J Immunol* (2010) 184(5):2518–27. doi: 10.4049/jimmunol.0901022
- 127. Rosenstiel P, Derer S, Till A, Hasler R, Eberstein H, Bewig B, et al. Systematic Expression Profiling of Innate Immune Genes Defines a Complex Pattern of Immunosenescence in Peripheral and Intestinal Leukocytes. *Genes Immun* (2008) 9(2):103–14. doi: 10.1038/sj.gene.6364454
- Gardner JK, Mamotte CDS, Jackaman C, Nelson DJ. Modulation of Dendritic Cell and T Cell Cross-Talk During Aging: The Potential Role of Checkpoint Inhibitory Molecules. *Ageing Res Rev* (2017) 38:40–51. doi: 10.1016/j.arr.2017.07.002
- 129. Cao JN, Agrawal A, Sharman E, Jia Z, Gupta S. Alterations in Gene Array Patterns in Dendritic Cells From Aged Humans. *PLoS One* (2014) 9(9): e106471. doi: 10.1371/journal.pone.0106471
- Wong C, Goldstein DR. Impact of Aging on Antigen Presentation Cell Function of Dendritic Cells. *Curr Opin Immunol* (2013) 25(4):535–41. doi: 10.1016/j.coi.2013.05.016
- 131. Qu C, Brinck-Jensen NS, Zang M, Chen K. Monocyte-Derived Dendritic Cells: Targets as Potent Antigen-Presenting Cells for the Design of Vaccines Against Infectious Diseases. *Int J Infect Dis* (2014) 19:1–5. doi: 10.1016/ j.ijid.2013.09.023
- Worbs T, Hammerschmidt SI, Forster R. Dendritic Cell Migration in Health and Disease. Nat Rev Immunol (2017) 17(1):30–48. doi: 10.1038/nri.2016.116
- 133. Aydar Y, Balogh P, Tew JG, Szakal AK. Altered Regulation of Fc Gamma RII on Aged Follicular Dendritic Cells Correlates With Immunoreceptor Tyrosine-Based Inhibition Motif Signaling in B Cells and Reduced Germinal Center Formation. J Immunol (2003) 171(11):5975–87. doi: 10.4049/jimmunol.171.11.5975
- 134. Zhang L, Wang CC. Inflammatory Response of Macrophages in Infection. Hepatobiliary Pancreat Dis Int (2014) 13(2):138–52. doi: 10.1016/S1499-3872(14)60024-2
- 135. Vetvicka V, Tlaskalova-Hogenova H, Pospisil M. Impaired Antigen Presenting Function of Macrophages From Aged Mice. *Immunol Invest* (1985) 14(2):105–14. doi: 10.3109/08820138509042005
- 136. Plowden J, Renshaw-Hoelscher M, Gangappa S, Engleman C, Katz JM, Sambhara S. Impaired Antigen-Induced CD8+ T Cell Clonal Expansion in Aging is Due to Defects in Antigen Presenting Cell Function. *Cell Immunol* (2004) 229(2):86–92. doi: 10.1016/j.cellimm.2004.07.001
- 137. Mahbub S, Deburghgraeve CR, Kovacs EJ. Advanced Age Impairs Macrophage Polarization. J Interferon Cytokine Res (2012) 32(1):18–26. doi: 10.1089/jir.2011.0058

- 138. Becker L, Nguyen L, Gill J, Kulkarni S, Pasricha PJ, Habtezion A. Age-Dependent Shift in Macrophage Polarisation Causes Inflammation-Mediated Degeneration of Enteric Nervous System. *Gut* (2018) 67(5):827– 36. doi: 10.1136/gutjnl-2016-312940
- 139. Cui CY, Driscoll RK, Piao Y, Chia CW, Gorospe M, Ferrucci L. Skewed Macrophage Polarization in Aging Skeletal Muscle. *Aging Cell* (2019) 18(6): e13032. doi: 10.1111/acel.13032
- 140. Lumeng CN, Liu J, Geletka L, Delaney C, Delproposto J, Desai A, et al. Aging is Associated With an Increase in T Cells and Inflammatory Macrophages in Visceral Adipose Tissue. J Immunol (2011) 187(12):6208–16. doi: 10.4049/ jimmunol.1102188
- 141. Wang Y, Wehling-Henricks M, Samengo G, Tidball JG. Increases of M2a Macrophages and Fibrosis in Aging Muscle are Influenced by Bone Marrow Aging and Negatively Regulated by Muscle-Derived Nitric Oxide. Aging Cell (2015) 14(4):678–88. doi: 10.1111/acel.12350
- 142. Linehan E, Dombrowski Y, Snoddy R, Fallon PG, Kissenpfennig A, Fitzgerald DC. Aging Impairs Peritoneal But Not Bone Marrow-Derived Macrophage Phagocytosis. Aging Cell (2014) 13(4):699–708. doi: 10.1111/acel.12223
- 143. Seidler S, Zimmermann HW, Bartneck M, Trautwein C, Tacke F. Age-Dependent Alterations of Monocyte Subsets and Monocyte-Related Chemokine Pathways in Healthy Adults. *BMC Immunol* (2010) 11:30. doi: 10.1186/1471-2172-11-30
- 144. Metcalf TU, Wilkinson PA, Cameron MJ, Ghneim K, Chiang C, Wertheimer AM, et al. Human Monocyte Subsets Are Transcriptionally and Functionally Altered in Aging in Response to Pattern Recognition Receptor Agonists. *J Immunol* (2017) 199(4):1405–17. doi: 10.4049/jimmunol.1700148
- 145. Qian F, Wang X, Zhang L, Chen S, Piecychna M, Allore H, et al. Age-Associated Elevation in TLR5 Leads to Increased Inflammatory Responses in the Elderly. *Aging Cell* (2012) 11(1):104–10. doi: 10.1111/j.1474-9726.2011.00759.x
- 146. Nyugen J, Agrawal S, Gollapudi S, Gupta S. Impaired Functions of Peripheral Blood Monocyte Subpopulations in Aged Humans. J Clin Immunol (2010) 30 (6):806–13. doi: 10.1007/s10875-010-9448-8
- 147. van Duin D, Allore HG, Mohanty S, Ginter S, Newman FK, Belshe RB, et al. Prevaccine Determination of the Expression of Costimulatory B7 Molecules in Activated Monocytes Predicts Influenza Vaccine Responses in Young and Older Adults. J Infect Dis (2007) 195(11):1590–7. doi: 10.1086/516788
- 148. Strohacker K, Breslin WL, Carpenter KC, McFarlin BK. Aged Mice Have Increased Inflammatory Monocyte Concentration and Altered Expression of Cell-Surface Functional Receptors. *J Biosci* (2012) 37(1):55–62. doi: 10.1007/ s12038-011-9169-z
- 149. Busse S, Steiner J, Alter J, Dobrowolny H, Mawrin C, Bogerts B, et al. Expression of HLA-DR, CD80, and CD86 in Healthy Aging and Alzheimer's Disease. J Alzheimers Dis (2015) 47(1):177–84. doi: 10.3233/JAD-150217
- 150. Herrero C, Sebastian C, Marques L, Comalada M, Xaus J, Valledor AF, et al. Immunosenescence of Macrophages: Reduced MHC Class II Gene Expression. *Exp Gerontol* (2002) 37(2-3):389–94. doi: 10.1016/S0531-5565 (01)00205-4
- Villanueva JL, Solana R, Alonso MC, Pena J. Changes in the Expression of HLA-class II Antigens on Peripheral Blood Monocytes From Aged Humans. *Dis Markers* (1990) 8(2):85–91.
- Herrero C, Marques L, Lloberas J, Celada A. IFN-Gamma-Dependent Transcription of MHC Class II IA is Impaired in Macrophages From Aged Mice. J Clin Invest (2001) 107(4):485–93. doi: 10.1172/JCI11696
- 153. Renshaw M, Rockwell J, Engleman C, Gewirtz A, Katz J, Sambhara S. Cutting Edge: Impaired Toll-like Receptor Expression and Function in Aging. *J Immunol* (2002) 169(9):4697–701. doi: 10.4049/jimmunol.169.9.4697
- 154. Hearps AC, Martin GE, Angelovich TA, Cheng WJ, Maisa A, Landay AL, et al. Aging is Associated With Chronic Innate Immune Activation and Dysregulation of Monocyte Phenotype and Function. *Aging Cell* (2012) 11 (5):867–75. doi: 10.1111/j.1474-9726.2012.00851.x
- 155. Amulic B, Cazalet C, Hayes GL, Metzler KD, Zychlinsky A. Neutrophil Function: From Mechanisms to Disease. *Annu Rev Immunol* (2012) 30:459– 89. doi: 10.1146/annurev-immunol-020711-074942
- Adrover JM, Nicolas-Avila JA, Hidalgo A. Aging: A Temporal Dimension for Neutrophils. *Trends Immunol* (2016) 37(5):334–45. doi: 10.1016/j.it.2016.03.005
- Ley K, Hoffman HM, Kubes P, Cassatella MA, Zychlinsky A, Hedrick CC, et al. Neutrophils: New Insights and Open Questions. *Sci Immunol* (2018) 3 (30):eaat4579. doi: 10.1126/sciimmunol.aat4579

- Cossio I, Lucas D, Hidalgo A. Neutrophils as Regulators of the Hematopoietic Niche. Blood (2019) 133(20):2140–8. doi: 10.1182/blood-2018-10-844571
- 159. Schurink B, Roos E, Radonic T, Barbe E, Bouman CSC, de Boer HH, et al. Viral Presence and Immunopathology in Patients With Lethal COVID-19: A Prospective Autopsy Cohort Study. *Lancet Microbe* (2020) 1(7):e290–9. doi: 10.1016/S2666-5247(20)30144-0
- 160. Veras FP, Pontelli MC, Silva CM, Toller-Kawahisa JE, de Lima M, Nascimento DC, et al. Sars-CoV-2-triggered Neutrophil Extracellular Traps Mediate COVID-19 Pathology. J Exp Med (2020) 217(12): e20201129. doi: 10.1084/jem.20201129
- 161. Simell B, Vuorela A, Ekstrom N, Palmu A, Reunanen A, Meri S, et al. Aging Reduces the Functionality of Anti-Pneumococcal Antibodies and the Killing of Streptococcus Pneumoniae by Neutrophil Phagocytosis. *Vaccine* (2011) 29 (10):1929–34. doi: 10.1016/j.vaccine.2010.12.121
- 162. Lopes AB, Lopes LB, da Silveira Antunes RN, Fukasawa JT, de ACD, Calamita Z. Effects of Immunosenescence on the Lower Expression of Surface Molecules in Neutrophils and Lymphocytes. *Curr Aging Sci* (2018) 11(2):118–25. doi: 10.2174/1874609811666180605092234
- 163. Whyte MK, Meagher LC, MacDermot J, Haslett C. Impairment of Function in Aging Neutrophils is Associated With Apoptosis. *J Immunol* (1993) 150 (11):5124–34.
- 164. Tomay F, Wells K, Duong L, Tsu JW, Dye DE, Radley-Crabb HG, et al. Aged Neutrophils Accumulate in Lymphoid Tissues From Healthy Elderly Mice and Infiltrate T- and B-cell Zones. *Immunol Cell Biol* (2018) 96(8):831–40. doi: 10.1111/imcb.12046
- 165. Zhang D, Chen G, Manwani D, Mortha A, Xu C, Faith JJ, et al. Neutrophil Ageing is Regulated by the Microbiome. *Nature* (2015) 525(7570):528–32. doi: 10.1038/nature15367
- 166. Kovalenko EI, Boyko AA, Semenkov VF, Lutsenko GV, Grechikhina MV, Kanevskiy LM, et al. ROS Production, Intracellular HSP70 Levels and Their Relationship in Human Neutrophils: Effects of Age. *Oncotarget* (2014) 5 (23):11800–12. doi: 10.18632/oncotarget.2856
- 167. Nogueira-Neto J, Cardoso AS, Monteiro HP, Fonseca FL, Ramos LR, Junqueira VB, et al. Basal Neutrophil Function in Human Aging: Implications in Endothelial Cell Adhesion. *Cell Biol Int* (2016) 40(7):796– 802. doi: 10.1002/cbin.10618
- 168. Tseng CW, Kyme PA, Arruda A, Ramanujan VK, Tawackoli W, Liu GY. Innate Immune Dysfunctions in Aged Mice Facilitate the Systemic Dissemination of Methicillin-Resistant S. Aureus. *PLoS One* (2012) 7(7): e41454. doi: 10.1371/journal.pone.0041454
- 169. Xu F, Zhang C, Zou Z, Fan EKY, Chen L, Li Y, et al. Aging-Related Atg5 Defect Impairs Neutrophil Extracellular Traps Formation. *Immunology* (2017) 151(4):417–32. doi: 10.1111/imm.12740
- van Deursen JM. The Role of Senescent Cells in Ageing. Nature (2014) 509 (7501):439–46. doi: 10.1038/nature13193
- 171. Antonangeli F, Zingoni A, Soriani A, Santoni A. Senescent Cells: Living or Dying is a Matter of NK Cells. J Leukoc Biol (2019) 105(6):1275–83. doi: 10.1002/JLB.MR0718-299R
- 172. Campos C, Pera A, Lopez-Fernandez I, Alonso C, Tarazona R, Solana R. Proinflammatory Status Influences NK Cells Subsets in the Elderly. *Immunol Lett* (2014) 162(1 Pt B):298–302. doi: 10.1016/j.imlet.2014.06.015
- 173. Brighton PJ, Maruyama Y, Fishwick K, Vrljicak P, Tewary S, Fujihara R, et al. Clearance of Senescent Decidual Cells by Uterine Natural Killer Cells in Cycling Human Endometrium. *Elife* (2017) 6:e31274. doi: 10.7554/ eLife.31274
- 174. Mace EM, Orange JS. Emerging Insights Into Human Health and NK Cell Biology From the Study of NK Cell Deficiencies. *Immunol Rev* (2019) 287 (1):202–25. doi: 10.1111/imr.12725
- 175. Cook KD, Waggoner SN, Whitmire JK. NK Cells and Their Ability to Modulate T Cells During Virus Infections. *Crit Rev Immunol* (2014) 34 (5):359–88. doi: 10.1615/CritRevImmunol.2014010604
- 176. Vivier E, Tomasello E, Baratin M, Walzer T, Ugolini S. Functions of Natural Killer Cells. Nat Immunol (2008) 9(5):503–10. doi: 10.1038/ni1582
- Sun JC, Ugolini S, Vivier E. Immunological Memory Within the Innate Immune System. EMBO J (2014) 33(12):1295–303. doi: 10.1002/embj.201387651
- Cording S, Medvedovic J, Aychek T, Eberl G. Innate Lymphoid Cells in Defense, Immunopathology and Immunotherapy. *Nat Immunol* (2016) 17 (7):755–7. doi: 10.1038/ni.3448

- 179. Shehata HM, Hoebe K, Chougnet CA. The Aged Nonhematopoietic Environment Impairs Natural Killer Cell Maturation and Function. Aging Cell (2015) 14(2):191–9. doi: 10.1111/acel.12303
- Hazeldine J, Lord JM. The Impact of Ageing on Natural Killer Cell Function and Potential Consequences for Health in Older Adults. *Ageing Res Rev* (2013) 12(4):1069–78. doi: 10.1016/j.arr.2013.04.003
- 181. Le Garff-Tavernier M, Beziat V, Decocq J, Siguret V, Gandjbakhch F, Pautas E, et al. Human NK Cells Display Major Phenotypic and Functional Changes Over the Life Span. *Aging Cell* (2010) 9(4):527–35. doi: 10.1111/j.1474-9726.2010.00584.x
- 182. Chidrawar SM, Khan N, Chan YL, Nayak L, Moss PA. Ageing is Associated With a Decline in Peripheral Blood CD56bright NK Cells. *Immun Ageing* (2006) 3:10. doi: 10.1186/1742-4933-3-10
- 183. Hazeldine J, Hampson P, Lord JM. Reduced Release and Binding of Perforin At the Immunological Synapse Underlies the Age-Related Decline in Natural Killer Cell Cytotoxicity. *Aging Cell* (2012) 11(5):751–9. doi: 10.1111/j.1474-9726.2012.00839.x
- 184. Solana C, Tarazona R, Solana R. Immunosenescence of Natural Killer Cells, Inflammation, and Alzheimer's Disease. Int J Alzheimers Dis 2018 (2018) p:3128758. doi: 10.1155/2018/3128758
- Kumar BV, Connors TJ, Farber DL. Human T Cell Development, Localization, and Function Throughout Life. *Immunity* (2018) 48(2):202– 13. doi: 10.1016/j.immuni.2018.01.007
- 186. George AJ, Ritter MA. Thymic Involution With Ageing: Obsolescence or Good Housekeeping? *Immunol Today* (1996) 17(6):267–72. doi: 10.1016/ 0167-5699(96)80543-3
- Aw D, Palmer DB. The Origin and Implication of Thymic Involution. Aging Dis (2011) 2(5):437–43.
- 188. Thapa P, Farber DL. The Role of the Thymus in the Immune Response. Thorac Surg Clin (2019) 29(2):123–31. doi: 10.1016/j.thorsurg.2018.12.001
- 189. Halnon NJ, Jamieson B, Plunkett M, Kitchen CM, Pham T, Krogstad P. Thymic Function and Impaired Maintenance of Peripheral T Cell Populations in Children With Congenital Heart Disease and Surgical Thymectomy. *Pediatr Res* (2005) 57(1):42–8. doi: 10.1203/01.PDR.0000147735.19342.DE
- 190. Sauce D, Larsen M, Fastenackels S, Duperrier A, Keller M, Grubeck-Loebenstein B, et al. Evidence of Premature Immune Aging in Patients Thymectomized During Early Childhood. J Clin Invest (2009) 119(10):3070– 8. doi: 10.1172/JCI39269
- 191. Murray JM, Kaufmann GR, Hodgkin PD, Lewin SR, Kelleher AD, Davenport MP, et al. Naive T Cells are Maintained by Thymic Output in Early Ages But by Proliferation Without Phenotypic Change After Age Twenty. *Immunol Cell Biol* (2003) 81(6):487–95. doi: 10.1046/j.1440-1711.2003.01191.x
- 192. den Braber I, Mugwagwa T, Vrisekoop N, Westera L, Mogling R, de Boer AB, et al. Maintenance of Peripheral Naive T Cells is Sustained by Thymus Output in Mice But Not Humans. *Immunity* (2012) 36(2):288–97. doi: 10.1016/j.immuni.2012.02.006
- 193. Goronzy JJ, Weyand CM. Mechanisms Underlying T Cell Ageing. Nat Rev Immunol (2019) 19(9):573–83. doi: 10.1038/s41577-019-0180-1
- 194. Cicin-Sain L, Messaoudi I, Park B, Currier N, Planer S, Fischer M, et al. Dramatic Increase in Naive T Cell Turnover is Linked to Loss of Naive T Cells From Old Primates. *Proc Natl Acad Sci USA* (2007) 104(50):19960–5. doi: 10.1073/pnas.0705905104
- 195. Thome JJ, Yudanin N, Ohmura Y, Kubota M, Grinshpun B, Sathaliyawala T, et al. Spatial Map of Human T Cell Compartmentalization and Maintenance Over Decades of Life. *Cell* (2014) 159(4):814–28. doi: 10.1016/j.cell.2014.10.026
- 196. Johnson PL, Yates AJ, Goronzy JJ, Antia R. Peripheral Selection Rather Than Thymic Involution Explains Sudden Contraction in Naive CD4 T-Cell Diversity With Age. Proc Natl Acad Sci USA (2012) 109(52):21432–7. doi: 10.1073/pnas.1209283110
- 197. Kohlmeier JE, Ely KH, Connor LM, Roberts AD, Yager EJ, Woodland DL, et al. Beneficial and Detrimental Manifestations of Age on CD8 + T Cell Memory to Respiratory Pathogens, in Handbook of Immunosenescence: Basic Understanding and Clinical Implications. Cham: Springer International Publishing (2017). p. 1–21. doi: 10.1007/978-3-319-64597-1_49-1
- 198. Alves AS, Bueno V. Immunosenescence: Participation of T Lymphocytes and Myeloid-Derived Suppressor Cells in Aging-Related Immune Response Changes. *Einstein (Sao Paulo)* (2019) 17(2):eRB4733. doi: 10.31744/ einstein_journal/2019RB4733

- 199. Drobek A, Moudra A, Mueller D, Huranova M, Horkova V, Pribikova M, et al. Strong Homeostatic TCR Signals Induce Formation of Self-Tolerant Virtual Memory CD8 T Cells. *EMBO J* (2018) 37(14):e98518. doi: 10.15252/ embj.201798518
- 200. Quinn KM, Zaloumis SG, Cukalac T, Kan WT, Sng XY, Mirams M, et al. Heightened Self-Reactivity Associated With Selective Survival, But Not Expansion, of Naive Virus-Specific CD8+ T Cells in Aged Mice. Proc Natl Acad Sci USA (2016) 113(5):1333–8. doi: 10.1073/pnas.1525167113
- Minato N, Hattori M, Hamazaki Y. Physiology and Pathology of T-cell Aging. Int Immunol (2020) 32(4):223–31. doi: 10.1093/intimm/dxaa006
- 202. Sato K, Kato A, Sekai M, Hamazaki Y, Minato N. Physiologic Thymic Involution Underlies Age-Dependent Accumulation of Senescence-Associated Cd4(+) T Cells. J Immunol (2017) 199(1):138–48. doi: 10.4049/jimmunol.1602005
- 203. Pulko V, Davies JS, Martinez C, Lanteri MC, Busch MP, Diamond MS, et al. Human Memory T Cells With a Naive Phenotype Accumulate With Aging and Respond to Persistent Viruses. *Nat Immunol* (2016) 17(8):966–75. doi: 10.1038/ni.3483
- 204. Gregg R, Smith CM, Clark FJ, Dunnion D, Khan N, Chakraverty R, et al. The Number of Human Peripheral Blood CD4+ CD25high Regulatory T Cells Increases With Age. *Clin Exp Immunol* (2005) 140(3):540–6. doi: 10.1111/ j.1365-2249.2005.02798.x
- Walker LS. Treg and CTLA-4: Two Intertwining Pathways to Immune Tolerance. J Autoimmun (2013) 45:49–57. doi: 10.1016/j.jaut.2013.06.006
- 206. Mahnke K, Johnson TS, Ring S, Enk AH. Tolerogenic Dendritic Cells and Regulatory T Cells: A Two-Way Relationship. J Dermatol Sci (2007) 46 (3):159–67. doi: 10.1016/j.jdermsci.2007.03.002
- 207. Shahid Z, Kleppinger A, Gentleman B, Falsey AR, McElhaney JE. Clinical and Immunologic Predictors of Influenza Illness Among Vaccinated Older Adults. Vaccine (2010) 28(38):6145–51. doi: 10.1016/j.vaccine.2010.07.036
- McElhaney JE, Ewen C, Zhou X, Kane KP, Xie D, Hager WD, et al. Granzyme B: Correlates With Protection and Enhanced CTL Response to Influenza Vaccination in Older Adults. *Vaccine* (2009) 27(18):2418–25. doi: 10.1016/j.vaccine.2009.01.136
- 209. Thiery J, Keefe D, Boulant S, Boucrot E, Walch M, Martinvalet D, et al. Perforin Pores in the Endosomal Membrane Trigger the Release of Endocytosed Granzyme B Into the Cytosol of Target Cells. *Nat Immunol* (2011) 12(8):770–7. doi: 10.1038/ni.2050
- 210. Boon AC, Fringuelli E, Graus YM, Fouchier RA, Sintnicolaas K, Iorio AM, et al. Influenza A Virus Specific T Cell Immunity in Humans During Aging. *Virology* (2002) 299(1):100–8. doi: 10.1006/viro.2002.1491
- 211. Brenchley JM, Karandikar NJ, Betts MR, Ambrozak DR, Hill BJ, Crotty LE, et al. Expression of CD57 Defines Replicative Senescence and Antigen-Induced Apoptotic Death of CD8+ T Cells. *Blood* (2003) 101(7):2711–20. doi: 10.1182/blood-2002-07-2103
- Czesnikiewicz-Guzik M, Lee WW, Cui D, Hiruma Y, Lamar DL, Yang ZZ, et al. T Cell Subset-Specific Susceptibility to Aging. *Clin Immunol* (2008) 127 (1):107–18. doi: 10.1016/j.clim.2007.12.002
- Crooke SN, Ovsyannikova IG, Poland GA, Kennedy RB. Immunosenescence and Human Vaccine Immune Responses. *Immun Ageing* (2019) 16:25. doi: 10.1186/s12979-019-0164-9
- 214. Saurwein-Teissl M, Lung TL, Marx F, Gschosser C, Asch E, Blasko I, et al. Lack of Antibody Production Following Immunization in Old Age: Association With CD8(+)CD28(-) T Cell Clonal Expansions and an Imbalance in the Production of Th1 and Th2 Cytokines. *J Immunol* (2002) 168(11):5893–9. doi: 10.4049/jimmunol.168.11.5893
- Hendriks J, Gravestein LA, Tesselaar K, van Lier RA, Schumacher TN, Borst J. CD27 is Required for Generation and Long-Term Maintenance of T Cell Immunity. *Nat Immunol* (2000) 1(5):433–40. doi: 10.1038/80877
- 216. Voehringer D, Koschella M, Pircher H. Lack of Proliferative Capacity of Human Effector and Memory T Cells Expressing Killer Cell Lectinlike Receptor G1 (Klrg1). Blood (2002) 100(10):3698–702. doi: 10.1182/blood-2002-02-0657
- 217. Yang OO, Lin H, Dagarag M, Ng HL, Effros RB, Uittenbogaart CH. Decreased Perforin and Granzyme B Expression in Senescent HIV-1specific Cytotoxic T Lymphocytes. *Virology* (2005) 332(1):16–9. doi: 10.1016/j.virol.2004.11.028
- Coppe JP, Desprez PY, Krtolica A, Campisi J. The Senescence-Associated Secretory Phenotype: The Dark Side of Tumor Suppression. *Annu Rev Pathol* (2010) 5:99–118. doi: 10.1146/annurev-pathol-121808-102144

- Wherry EJ, Kurachi M. Molecular and Cellular Insights Into T Cell Exhaustion. Nat Rev Immunol (2015) 15(8):486–99. doi: 10.1038/nri3862
- 220. Vescovini R, Telera A, Fagnoni FF, Biasini C, Medici MC, Valcavi P, et al. Different Contribution of EBV and CMV Infections in Very Long-Term Carriers to Age-Related Alterations of CD8+ T Cells. *Exp Gerontol* (2004) 39 (8):1233–43. doi: 10.1016/j.exger.2004.04.004
- 221. Brunner S, Herndler-Brandstetter D, Weinberger B, Grubeck-Loebenstein B. Persistent Viral Infections and Immune Aging. Ageing Res Rev (2011) 10 (3):362–9. doi: 10.1016/j.arr.2010.08.003
- 222. Fulop T, Larbi A, Pawelec G. Human T Cell Aging and the Impact of Persistent Viral Infections. *Front Immunol* (2013) 4:271. doi: 10.3389/fimmu.2013.00271
- 223. Forte E, Zhang Z, Thorp EB, Hummel M. Cytomegalovirus Latency and Reactivation: An Intricate Interplay With the Host Immune Response. *Front Cell Infect Microbiol* (2020) 10:130. doi: 10.3389/fcimb.2020.00130
- 224. Nussing S, Koay HF, Sant S, Loudovaris T, Mannering SI, Lappas M, et al. Divergent SATB1 Expression Across Human Life Span and Tissue Compartments. *Immunol Cell Biol* (2019) 97(5):498–511. doi: 10.1111/imcb.12233
- 225. Paez-Ribes M, Gonzalez-Gualda E, Doherty GJ, Munoz-Espin D. Targeting Senescent Cells in Translational Medicine. *EMBO Mol Med* (2019) 11(12): e10234. doi: 10.15252/emmm.201810234
- 226. Alter-Wolf S, Blomberg BB, Riley RL. Deviation of the B Cell Pathway in Senescent Mice is Associated With Reduced Surrogate Light Chain Expression and Altered Immature B Cell Generation, Phenotype, and Light Chain Expression. J Immunol (2009) 182(1):138–47. doi: 10.4049/ jimmunol.182.1.138
- 227. Ricci C, Cova M, Kang YS, Yang A, Rahmouni A, Scott WWJr., et al. Normal Age-Related Patterns of Cellular and Fatty Bone Marrow Distribution in the Axial Skeleton: MR Imaging Study. *Radiology* (1990) 177(1):83–8. doi: 10.1148/radiology.177.1.2399343
- 228. Chambers SM, Shaw CA, Gatza C, Fisk CJ, Donehower LA, Goodell MA. Aging Hematopoietic Stem Cells Decline in Function and Exhibit Epigenetic Dysregulation. *PLoS Biol* (2007) 5(8):e201. doi: 10.1371/journal.pbio.0050201
- 229. Bulati M, Buffa S, Candore G, Caruso C, Dunn-Walters DK, Pellicano M, et al. B Cells and Immunosenescence: A Focus on IgG+IgD-CD27- (Dn) B Cells in Aged Humans. Ageing Res Rev (2011) 10(2):274–84. doi: 10.1016/ j.arr.2010.12.002
- 230. Henry CJ, Casas-Selves M, Kim J, Zaberezhnyy V, Aghili L, Daniel AE, et al. Aging-Associated Inflammation Promotes Selection for Adaptive Oncogenic Events in B Cell Progenitors. J Clin Invest (2015) 125(12):4666–80. doi: 10.1172/JCI83024
- 231. Stephan RP, Lill-Elghanian DA, Witte PL. Development of B Cells in Aged Mice: Decline in the Ability of Pro-B Cells to Respond to IL-7 But Not to Other Growth Factors. *J Immunol* (1997) 158(4):1598–609.
- 232. Stephan RP, Reilly CR, Witte PL. Impaired Ability of Bone Marrow Stromal Cells to Support B-lymphopoiesis With Age. *Blood* (1998) 91(1):75–88. doi: 10.1182/blood.V91.1.75.75_75_88
- 233. Ratliff M, Alter S, Frasca D, Blomberg BB, Riley RL. In Senescence, Age-Associated B Cells Secrete TNFalpha and Inhibit Survival of B-cell Precursors. Aging Cell (2013) 12(2):303–11. doi: 10.1111/acel.12055
- 234. Cancro MP. Age-Associated B Cells. Annu Rev Immunol (2020) 38:315–40. doi: 10.1146/annurev-immunol-092419-031130
- 235. Rubtsov AV, Rubtsova K, Fischer A, Meehan RT, Gillis JZ, Kappler JW, et al. Toll-Like Receptor 7 (TLR7)-Driven Accumulation of a Novel CD11c(+) B-Cell Population is Important for the Development of Autoimmunity. *Blood* (2011) 118(5):1305–15. doi: 10.1182/blood-2011-01-331462
- Hao Y, O'Neill P, Naradikian MS, Scholz JL, Cancro MP. A B-cell Subset Uniquely Responsive to Innate Stimuli Accumulates in Aged Mice. *Blood* (2011) 118(5):1294–304. doi: 10.1182/blood-2011-01-330530
- 237. Riley RL, Khomtchouk K, Blomberg BB. Age-Associated B Cells (ABC) Inhibit B Lymphopoiesis and Alter Antibody Repertoires in Old Age. Cell Immunol (2017) 321:61–7. doi: 10.1016/j.cellimm.2017.04.008
- Chang LY, Li Y, Kaplan DE. Hepatitis C Viraemia Reversibly Maintains Subset of Antigen-Specific T-bet+ Tissue-Like Memory B Cells. J Viral Hepat (2017) 24(5):389–96. doi: 10.1111/jvh.12659
- 239. Adlowitz DG, Barnard J, Biear JN, Cistrone C, Owen T, Wang W, et al. Expansion of Activated Peripheral Blood Memory B Cells in Rheumatoid Arthritis, Impact of B Cell Depletion Therapy, and Biomarkers of Response. *PLoS One* (2015) 10(6):e0128269. doi: 10.1371/journal.pone.0128269

- 240. Sayegh CE, Quong MW, Agata Y, Murre C. E-Proteins Directly Regulate Expression of Activation-Induced Deaminase in Mature B Cells. Nat Immunol (2003) 4(6):586–93. doi: 10.1038/ni923
- 241. Frasca D, Diaz A, Romero M, Ferracci F, Blomberg BB. MicroRNAs miR-155 and Mir-16 Decrease AID and E47 in B Cells From Elderly Individuals. *J Immunol* (2015) 195(5):2134–40. doi: 10.4049/jimmunol.1500520
- 242. Kosco MH, Burton GF, Kapasi ZF, Szakal AK, Tew JG. Antibody-Forming Cell Induction During an Early Phase of Germinal Centre Development and its Delay With Ageing. *Immunology* (1989) 68(3):312–8.
- Frasca D. Senescent B Cells in Aging and Age-Related Diseases: Their Role in the Regulation of Antibody Responses. *Exp Gerontol* (2018) 107:55–8. doi: 10.1016/j.exger.2017.07.002
- 244. Shankwitz K, Pallikkuth S, Sirupangi T, Kirk Kvistad D, Russel KB, Pahwa R, et al. Compromised Steady-State Germinal Center Activity With Age in Nonhuman Primates. *Aging Cell* (2020) 19(2):e13087. doi: 10.1111/ acel.13087
- 245. Pritz T, Lair J, Ban M, Keller M, Weinberger B, Krismer M, et al. Plasma Cell Numbers Decrease in Bone Marrow of Old Patients. *Eur J Immunol* (2015) 45(3):738–46. doi: 10.1002/eji.201444878
- 246. Han S, Yang K, Ozen Z, Peng W, Marinova E, Kelsoe G, et al. Enhanced Differentiation of Splenic Plasma Cells But Diminished Long-Lived High-Affinity Bone Marrow Plasma Cells in Aged Mice. *J Immunol* (2003) 170 (3):1267–73. doi: 10.4049/jimmunol.170.3.1267
- 247. Turner VM, Mabbott NA. Ageing Adversely Affects the Migration and Function of Marginal Zone B Cells. *Immunology* (2017) 151(3):349–62. doi: 10.1111/imm.12737
- 248. Weksler ME, Szabo P. The Effect of Age on the B-cell Repertoire. J Clin Immunol (2000) 20(4):240–9. doi: 10.1023/a:1006659401385
- Ventura MT, Casciaro M, Gangemi S, Buquicchio R. Immunosenescence in Aging: Between Immune Cells Depletion and Cytokines Up-Regulation. *Clin Mol Allergy* (2017) 15:21. doi: 10.1186/s12948-017-0077-0
- 250. Listi F, Candore G, Modica MA, Russo M, Di Lorenzo G, Esposito-Pellitteri M, et al. A Study of Serum Immunoglobulin Levels in Elderly Persons That Provides New Insights Into B Cell Immunosenescence. *Ann N Y Acad Sci* (2006) 1089:487–95. doi: 10.1196/annals.1386.013
- 251. Ma S, Wang C, Mao X, Hao Y. B Cell Dysfunction Associated With Aging and Autoimmune Diseases. *Front Immunol* (2019) 10:318. doi: 10.3389/ fimmu.2019.00318
- 252. Weksler ME. Changes in the B-cell Repertoire With Age. Vaccine (2000) 18 (16):1624–8. doi: 10.1016/S0264-410X(99)00497-1
- 253. Ahmed R, Omidian Z, Donner T, Hamad ARA. Hiding in Plain Sight: Time to Unlock Autoimmune Clues in Human CD5+ B Cells by Using Nextgen Technology. *Discov Med* (2018) 26(142):79–83.
- Jergovic M, Contreras NA, Nikolich-Zugich J. Impact of CMV Upon Immune Aging: Facts and Fiction. *Med Microbiol Immunol* (2019) 208(3-4):263–9. doi: 10.1007/s00430-019-00605-w
- 255. Marchi E, Lee LN, Klenerman P. Inflation vs. Exhaustion of Antiviral Cd8+ T-Cell Populations in Persistent Infections: Two Sides of the Same Coin? *Front Immunol* (2019) 10:197. doi: 10.3389/fimmu.2019.00197
- 256. Wertheimer AM, Bennett MS, Park B, Uhrlaub JL, Martinez C, Pulko V, et al. Aging and Cytomegalovirus Infection Differentially and Jointly Affect Distinct Circulating T Cell Subsets in Humans. J Immunol (2014) 192 (5):2143–55. doi: 10.4049/jimmunol.1301721
- 257. Karrer U, Sierro S, Wagner M, Oxenius A, Hengel H, Koszinowski UH, et al. Memory Inflation: Continuous Accumulation of Antiviral CD8+ T Cells Over Time. J Immunol (2003) 170(4):2022–9. doi: 10.4049/jimmunol.170.4.2022
- 258. Sylwester AW, Mitchell BL, Edgar JB, Taormina C, Pelte C, Ruchti F, et al. Broadly Targeted Human Cytomegalovirus-Specific CD4+ and CD8+ T Cells Dominate the Memory Compartments of Exposed Subjects. J Exp Med (2005) 202(5):673–85. doi: 10.1084/jem.20050882
- 259. Pourgheysari B, Khan N, Best D, Bruton R, Nayak L, Moss PA. The Cytomegalovirus-Specific CD4+ T-Cell Response Expands With Age and Markedly Alters the CD4+ T-Cell Repertoire. J Virol (2007) 81(14):7759–65. doi: 10.1128/JVI.01262-06
- 260. Arens R, Wang P, Sidney J, Loewendorf A, Sette A, Schoenberger SP, et al. Cutting Edge: Murine Cytomegalovirus Induces a Polyfunctional CD4 T Cell Response. J Immunol (2008) 180(10):6472–6. doi: 10.4049/jimmunol. 180.10.6472

- 261. Strindhall J, Lofgren S, Framsth C, Matussek A, Bengner M, Ernerudh J, et al. CD4/CD8 Ratio<1 is Associated With Lymphocyte Subsets, CMV and Gender in 71-Year Old Individuals: 5-Year Follow-Up of the Swedish Hexa Immune Longitudinal Study. *Exp Gerontol* (2017) 95:82–7. doi: 10.1016/j.exger.2017.03.020
- 262. Alonso-Arias R, Moro-Garcia MA, Echeverria A, Solano-Jaurrieta JJ, Suarez-Garcia FM, Lopez-Larrea C. Intensity of the Humoral Response to Cytomegalovirus is Associated With the Phenotypic and Functional Status of the Immune System (Vol 87, Pg 4486, 2013). *J Virol* (2013) 87(15):8816–6. doi: 10.1128/JVI.01378-13
- 263. Sansoni P, Vescovini R, Fagnoni FF, Akbar A, Arens R, Chiu YL, et al. New Advances in CMV and Immunosenescence. *Exp Gerontol* (2014) 55:54–62. doi: 10.1016/j.exger.2014.03.020
- 264. Bryl E, Vallejo AN, Weyand CM, Goronzy JJ. Down-Regulation of CD28 Expression by TNF-Alpha. J Immunol (2001) 167(6):3231–8. doi: 10.4049/ jimmunol.167.6.3231
- 265. Bartlett DB, Firth CM, Phillips AC, Moss P, Baylis D, Syddall H, et al. The Age-Related Increase in Low-Grade Systemic Inflammation (Inflammaging) is Not Driven by Cytomegalovirus Infection. *Aging Cell* (2012) 11(5):912–5. doi: 10.1111/j.1474-9726.2012.00849.x
- 266. Bennett JM, Glaser R, Malarkey WB, Beversdorf DQ, Peng J, Kiecolt-Glaser JK. Inflammation and Reactivation of Latent Herpesviruses in Older Adults. *Brain Behav Immun* (2012) 26(5):739–46. doi: 10.1016/j.bbi.2011.11.007
- 267. Frasca D, Blomberg BB. Aging, Cytomegalovirus (CMV) and Influenza Vaccine Responses. *Hum Vaccin Immunother* (2016) 12(3):682–90. doi: 10.1080/21645515.2015.1105413
- 268. Bowyer G, Sharpe H, Venkatraman N, Ndiaye PB, Wade D, Brenner N, et al. Reduced Ebola Vaccine Responses in CMV+ Young Adults is Associated With Expansion of CD57+KLRG1+ T Cells. J Exp Med (2020) 217(7): e20200004. doi: 10.1084/jem.20200004
- 269. Trzonkowski P, Mysliwska J, Szmit E, Wieckiewicz J, Lukaszuk K, Brydak LB, et al. Association Between Cytomegalovirus Infection, Enhanced Proinflammatory Response and Low Level of Anti-Hemagglutinins During the Anti-Influenza Vaccination–an Impact of Immunosenescence. *Vaccine* (2003) 21(25-26):3826–36. doi: 10.1016/S0264-410X(03)00309-8
- 270. Clement M, Marsden M, Stacey MA, Abdul-Karim J, Gimeno Brias S, Costa Bento D, et al. Cytomegalovirus-Specific IL-10-Producing Cd4+ T Cells Are Governed by Type-I Ifn-Induced IL-27 and Promote Virus Persistence. *PLoS Pathog* (2016) 12(12):e1006050. doi: 10.1371/journal.ppat.1006050
- 271. Merani S, Kuchel GA, Kleppinger A, McElhaney JE. Influenza Vaccine-Mediated Protection in Older Adults: Impact of Influenza Infection, Cytomegalovirus Serostatus and Vaccine Dosage. *Exp Gerontol* (2018) 107:116–25. doi: 10.1016/j.exger.2017.09.015
- 272. van den Berg SPH, Wong A, Hendriks M, Jacobi RHJ, van Baarle D, van Beek J. Negative Effect of Age, But Not of Latent Cytomegalovirus Infection on the Antibody Response to a Novel Influenza Vaccine Strain in Healthy Adults. *Front Immunol* (2018) 9:82. doi: 10.3389/fimmu.2018.00082
- 273. Enani S, Przemska-Kosicka A, Childs CE, Maidens C, Dong H, Conterno L, et al. Impact of Ageing and a Synbiotic on the Immune Response to Seasonal Influenza Vaccination; a Randomised Controlled Trial. *Clin Nutr* (2018) 37 (2):443–51. doi: 10.1016/j.clnu.2017.01.011
- 274. Savva GM, Pachnio A, Kaul B, Morgan K, Huppert FA, Brayne C, et al. Cytomegalovirus Infection is Associated With Increased Mortality in the Older Population. Aging Cell (2013) 12(3):381–7. doi: 10.1111/acel.12059
- 275. van den Berg SPH, Warmink K, Borghans JAM, Knol MJ, van Baarle D. Effect of Latent Cytomegalovirus Infection on the Antibody Response to Influenza Vaccination: A Systematic Review and Meta-Analysis. Med Microbiol Immunol (2019) 208(3-4):305–21. doi: 10.1007/s00430-019-00602-z
- 276. Furman D, Jojic V, Sharma S, Shen-Orr SS, Angel CJ, Onengut-Gumuscu S, et al. Cytomegalovirus Infection Enhances the Immune Response to Influenza. *Sci Transl Med* (2015) 7(281):281ra43. doi: 10.1126/scitranslmed.aaa2293
- 277. Bharucha T, Houlihan CF, Breuer J. Herpesvirus Infections of the Central Nervous System. Semin Neurol (2019) 39(3):369–82. doi: 10.1055/s-0039-1687837
- Cocchio S, Gallo T, Baldo V. Herpes Zoster: Vaccination Status and Virtuous Experiences. *Minerva Med* (2020) 111(1):4–8. doi: 10.23736/S0026-4806.19.06412-7

- Shah RA, Limmer AL, Nwannunu CE, Patel RR, Mui UN, Tyring SK. Shingrix for Herpes Zoster: A Review. Skin Ther Lett (2019) 24(4):5–7.
- Johnson RW, Levin MJ. Herpes Zoster and Its Prevention by Vaccination. Interdiscip Top Gerontol Geriatr (2020) 43:131–45. doi: 10.1159/000504484
- Levin MJ, Cai GY, Manchak MD, Pizer LI. Varicella-Zoster Virus DNA in Cells Isolated From Human Trigeminal Ganglia. J Virol (2003) 77(12):6979– 87. doi: 10.1128/JVI.77.12.6979-6987.2003
- 282. Ouwendijk WJD, Depledge DP, Rajbhandari L, Lenac Rovis T, Jonjic S, Breuer J, et al. Varicella-Zoster Virus VLT-ORF63 Fusion Transcript Induces Broad Viral Gene Expression During Reactivation From Neuronal Latency. *Nat Commun* (2020) 11(1):6324. doi: 10.1038/s41467-020-20031-4
- Amanna IJ, Carlson NE, Slifka MK. Duration of Humoral Immunity to Common Viral and Vaccine Antigens. N Engl J Med (2007) 357(19):1903–15. doi: 10.1056/NEJMoa066092
- 284. Weinberg A, Zhang JH, Oxman MN, Johnson GR, Hayward AR, Caulfield MJ, et al. Varicella-Zoster Virus-Specific Immune Responses to Herpes Zoster in Elderly Participants in a Trial of a Clinically Effective Zoster Vaccine. J Infect Dis (2009) 200(7):1068–77. doi: 10.1086/605611
- 285. Steain M, Sutherland JP, Rodriguez M, Cunningham AL, Slobedman B, Abendroth A. Analysis of T Cell Responses During Active Varicella-Zoster Virus Reactivation in Human Ganglia. J Virol (2014) 88(5):2704–16. doi: 10.1128/JVI.03445-13
- Kennedy PGE, Gershon AA. Clinical Features of Varicella-Zoster Virus Infection. Viruses (2018) 10(11):609. doi: 10.3390/v10110609
- Gnann JWJr. Varicella-Zoster Virus: Atypical Presentations and Unusual Complications. J Infect Dis (2002) 186 Suppl 1:S91–8. doi: 10.1086/342963
- Arvin AM. Human Herpesviruses : Biology, Therapy, and Immunoprophylaxis Vol. 1388. Cambridge, New York: Cambridge University Press (2007). doi: 10.1017/CBO9780511545313
- Lachiewicz AM, Srinivas ML. Varicella-Zoster Virus Post-Exposure Management and Prophylaxis: A Review. *Prev Med Rep* (2019) 16:101016. doi: 10.1016/j.pmedr.2019.101016
- Levin MJ. Immune Senescence and Vaccines to Prevent Herpes Zoster in Older Persons. Curr Opin Immunol (2012) 24(4):494–500. doi: 10.1016/ j.coi.2012.06.002
- 291. Takahashi M, Otsuka T, Okuno Y, Asano Y, Yazaki T. Live Vaccine Used to Prevent the Spread of Varicella in Children in Hospital. *Lancet* (1974) 2 (7892):1288–90. doi: 10.1016/S0140-6736(74)90144-5
- 292. Shaw J, Gershon AA. Varicella Virus Vaccination in the United States. Viral Immunol (2018) 31(2):96–103. doi: 10.1089/vim.2017.0136
- 293. Woodward M, Marko A, Galea S, Eagel B, Straus W. Varicella Virus Vaccine Live: A 22-Year Review of Postmarketing Safety Data. Open Forum Infect Dis (2019) 6(8):ofz295. doi: 10.1093/ofid/ofz295
- 294. Alexander KE, Tong PL, Macartney K, Beresford R, Sheppeard V, Gupta M. Live Zoster Vaccination in an Immunocompromised Patient Leading to Death Secondary to Disseminated Varicella Zoster Virus Infection. *Vaccine* (2018) 36(27):3890–3. doi: 10.1016/j.vaccine.2018.05.078
- 295. Levin MJ, Weinberg A. Immune Responses to Zoster Vaccines. Hum Vaccin Immunother (2019) 15(4):772–7. doi: 10.1080/21645515.2018.1560918
- 296. Blom K, Yin L, Arnheim-Dahlstrom L. Effectiveness of the Herpes Zoster Vaccine Zostavax(R) in Stockholm County, Sweden. Vaccine (2019) 37 (31):4401–6. doi: 10.1016/j.vaccine.2019.06.008
- 297. Dooling KL, Guo A, Patel M, Lee GM, Moore K, Belongia EA, et al. Recommendations of the Advisory Committee on Immunization Practices for Use of Herpes Zoster Vaccines. *MMWR Morb Mortal Wkly Rep* (2018) 67(3):103–8. doi: 10.15585/mmwr.mm6703a5
- 298. Winston DJ, Mullane KM, Cornely OA, Boeckh MJ, Brown JW, Pergam SA, et al. Inactivated Varicella Zoster Vaccine in Autologous Haemopoietic Stem-Cell Transplant Recipients: An International, Multicentre, Randomised, Double-Blind, Placebo-Controlled Trial. *Lancet* (2018) 391 (10135):2116–27. doi: 10.1016/S0140-6736(18)30631-7
- 299. Mullane KM, Morrison VA, Camacho LH, Arvin A, McNeil SA, Durrand J, et al. Safety and Efficacy of Inactivated Varicella Zoster Virus Vaccine in Immunocompromised Patients With Malignancies: A Two-Arm, Randomised, Double-Blind, Phase 3 Trial. *Lancet Infect Dis* (2019) 19 (9):1001–12. doi: 10.1016/S1473-3099(19)30310-X
- 300. Arnold N, Messaoudi I. Herpes Zoster and the Search for an Effective Vaccine. Clin Exp Immunol (2017) 187(1):82–92. doi: 10.1111/cei.12809

- 301. Heineman TC, Cunningham A, Levin M. Understanding the Immunology of Shingrix, a Recombinant Glycoprotein E Adjuvanted Herpes Zoster Vaccine. *Curr Opin Immunol* (2019) 59:42–8. doi: 10.1016/j.coi.2019.02.009
- Levin MJ, Weinberg A. Adjuvanted Recombinant Glycoprotein E Herpes Zoster Vaccine. Clin Infect Dis (2019) 70(7):1509–15. doi: 10.1093/cid/ciz770
- 303. Short MD, Fergus C. Which Patients Should Receive the Herpes Zoster Vaccine? JAAPA (2019) 32(9):18–20. doi: 10.1097/01.JAA.0000578788.69674.2b
- 304. Chlibek R, Pauksens K, Rombo L, van Rijckevorsel G, Richardus JH, Plassmann G, et al. Long-Term Immunogenicity and Safety of an Investigational Herpes Zoster Subunit Vaccine in Older Adults. *Vaccine* (2016) 34(6):863–8. doi: 10.1016/j.vaccine.2015.09.073
- 305. Leroux-Roels G, Marchant A, Levy J, Van Damme P, Schwarz TF, Horsmans Y, et al. Impact of Adjuvants on CD4(+) T Cell and B Cell Responses to a Protein Antigen Vaccine: Results From a Phase II, Randomized, Multicenter Trial. *Clin Immunol* (2016) 169:16–27. doi: 10.1016/j.clim.2016.05.007
- 306. Weinberg A, Popmihajlov Z, Schmader KE, Johnson MJ, Caldas Y, Salazar AT, et al. Persistence of Varicella-Zoster Virus Cell-Mediated Immunity After the Administration of a Second Dose of Live Herpes Zoster Vaccine. *J Infect Dis* (2019) 219(2):335–8. doi: 10.1093/infdis/jiy514
- 307. Schmader KE, Oxman MN, Levin MJ, Johnson G, Zhang JH, Betts R, et al. Persistence of the Efficacy of Zoster Vaccine in the Shingles Prevention Study and the Short-Term Persistence Substudy. *Clin Infect Dis* (2012) 55 (10):1320–8. doi: 10.1093/cid/cis638
- 308. Sullivan NL, Eberhardt CS, Wieland A, Vora KA, Pulendran B, Ahmed R. Understanding the Immunology of the Zostavax Shingles Vaccine. *Curr Opin Immunol* (2019) 59:25–30. doi: 10.1016/j.coi.2019.02.005
- 309. Levin MJ, Bresnitz E, Popmihajlov Z, Weinberg A, Liaw KL, Willis E, et al. Studies With Herpes Zoster Vaccines in Immune Compromised Patients. *Expert Rev* Vaccines (2017) 16(12):1217–30. doi: 10.1080/14760584.2017.1395703
- 310. Tseng HF, Tartof S, Harpaz R, Luo Y, Sy LS, Hetcher RC, et al. Vaccination Against Zoster Remains Effective in Older Adults Who Later Undergo Chemotherapy. *Clin Infect Dis* (2014) 59(7):913–9. doi: 10.1093/cid/ciu498
- 311. Hoffmann M, Kleine-Weber H, Schroeder S, Kruger N, Herrler T, Erichsen S, et al. SARS-Cov-2 Cell Entry Depends on ACE2 and TMPRSS2 and Is Blocked by a Clinically Proven Protease Inhibitor. *Cell* (2020) 181(2):271–80. doi: 10.1016/j.cell.2020.02.052
- 312. Richardson S, Hirsch JS, Narasimhan M, Crawford JM, McGinn T, Davidson KW, et al. Presenting Characteristics, Comorbidities, and Outcomes Among 5700 Patients Hospitalized With Covid-19 in the New York City Area. JAMA (2020) 323(20):2052–9. doi: 10.1001/jama.2020.6775
- 313. Goyal P, Choi JJ, Pinheiro LC, Schenck EJ, Chen R, Jabri A, et al. Clinical Characteristics of Covid-19 in New York City. N Engl J Med (2020) 382 (24):2372–4. doi: 10.1056/NEJMc2010419
- 314. Wang D, Hu B, Hu C, Zhu F, Liu X, Zhang J, et al. Clinical Characteristics of 138 Hospitalized Patients With 2019 Novel Coronavirus-Infected Pneumonia in Wuhan, China. JAMA (2020) 323(11):1061–9. doi: 10.1001/jama.2020.1585
- Petersen E, Koopmans M, Go U, Hamer DH, Petrosillo N, Castelli F, et al. Comparing SARS-CoV-2 With SARS-CoV and Influenza Pandemics. *Lancet Infect Dis* (2020) 20(9):e238–44. doi: 10.1016/S1473-3099(20)30484-9
- Nichol KL, Nordin JD, Nelson DB, Mullooly JP, Hak E. Effectiveness of Influenza Vaccine in the Community-Dwelling Elderly. N Engl J Med (2007) 357(14):1373–81. doi: 10.1056/NEJMoa070844
- 317. Nassar MS, Bakhrebah MA, Meo SA, Alsuabeyl MS, Zaher WA. Middle East Respiratory Syndrome Coronavirus (Mers-CoV) Infection: Epidemiology, Pathogenesis and Clinical Characteristics. *Eur Rev Med Pharmacol Sci* (2018) 22(15):4956–61. doi: 10.26355/eurrev_201808_15635
- 318. Donnelly CA, Ghani AC, Leung GM, Hedley AJ, Fraser C, Riley S, et al. Epidemiological Determinants of Spread of Causal Agent of Severe Acute Respiratory Syndrome in Hong Kong. *Lancet* (2003) 361(9371):1761–6. doi: 10.1016/S0140-6736(03)13410-1
- 319. Meftahi GH, Jangravi Z, Sahraei H, Bahari Z. The Possible Pathophysiology Mechanism of Cytokine Storm in Elderly Adults With COVID-19 Infection: The Contribution of "Inflame-Aging". *Inflamm Res* (2020) 69(9):825–39. doi: 10.1007/s00011-020-01372-8
- 320. Rydyznski Moderbacher C, Ramirez SI, Dan JM, Grifoni A, Hastie KM, Weiskopf D, et al. Antigen-Specific Adaptive Immunity to SARS-CoV-2 in Acute Covid-19 and Associations With Age and Disease Severity. *Cell* (2020) 183(4):996–1012.e19. doi: 10.1016/j.cell.2020.09.038

- Moore JB, June CH. Cytokine Release Syndrome in Severe COVID-19. Science (2020) 368(6490):473–4. doi: 10.1126/science.abb8925
- 322. de la Rica R, Borges M, Gonzalez-Freire M. Covid-19: In the Eye of the Cytokine Storm. Front Immunol (2020) 11:558898. doi: 10.3389/ fimmu.2020.558898
- 323. Takahashi T, Ellingson MK, Wong P, Israelow B, Lucas C, Klein J, et al. Sex Differences in Immune Responses That Underlie COVID-19 Disease Outcomes. *Nature* (2020) 588(7837):315–20. doi: 10.1038/s41586-020-2700-3
- 324. Tay MZ, Poh CM, Renia L, MacAry PA, Ng LFP. The Trinity of COVID-19: Immunity, Inflammation and Intervention. Nat Rev Immunol (2020) 20 (6):363–74. doi: 10.1038/s41577-020-0311-8
- 325. Pietrobon AJ, Teixeira FME, Sato MN. I Mmunosenescence and Inflammaging: Risk Factors of Severe Covid-19 in Older People. Front Immunol (2020) 11:579220. doi: 10.3389/fimmu.2020.579220
- 326. Lan J, Ge J, Yu J, Shan S, Zhou H, Fan S, et al. Structure of the SARS-CoV-2 Spike Receptor-Binding Domain Bound to the ACE2 Receptor. *Nature* (2020) 581(7807):215–20. doi: 10.1038/s41586-020-2180-5
- 327. Vaduganathan M, Vardeny O, Michel T, McMurray JJV, Pfeffer MA, Solomon SD. Renin-Angiotensin-Aldosterone System Inhibitors in Patients With Covid-19. N Engl J Med (2020) 382(17):1653–9. doi: 10.1056/NEJMsr2005760
- 328. McElhaney JE, Verschoor CP, Andrew MK, Haynes L, Kuchel GA, Pawelec G. The Immune Response to Influenza in Older Humans: Beyond Immune Senescence. *Immun Ageing* (2020) 17:10. doi: 10.1186/s12979-020-00181-1
- 329. Clegg A, Young J, Iliffe S, Rikkert MO, Rockwood K. Frailty in Elderly People. Lancet (2013) 381(9868):752–62. doi: 10.1016/S0140-6736(12)62167-9
- 330. Louie JK, Acosta M, Winter K, Jean C, Gavali S, Schechter R, et al. Factors Associated With Death or Hospitalization Due to Pandemic 2009 Influenza A(H1N1) Infection in California. *JAMA* (2009) 302(17):1896–902. doi: 10.1001/jama.2009.1583
- 331. Kwong JC, Campitelli MA, Gubbay JB, Peci A, Winter AL, Olsha R, et al. Vaccine Effectiveness Against Laboratory-Confirmed Influenza Hospitalizations Among Elderly Adults During the 2010-2011 Season. Clin Infect Dis (2013) 57(6):820–7. doi: 10.1093/cid/cit404
- 332. Dawood FS, Chung JR, Kim SS, Zimmerman RK, Nowalk MP, Jackson ML, et al. Interim Estimates of 2019-20 Seasonal Influenza Vaccine Effectiveness -United States, February 2020. MMWR Morb Mortal Wkly Rep (2020) 69 (7):177–82. doi: 10.15585/mmwr.mm6907a1
- 333. Ward BJ, Pillet S, Charland N, Trepanier S, Couillard J, Landry N. The Establishment of Surrogates and Correlates of Protection: Useful Tools for the Licensure of Effective Influenza Vaccines? *Hum Vaccin Immunother* (2018) 14(3):647–56. doi: 10.1080/21645515.2017.1413518
- 334. Carlock MA, Ingram JG, Clutter EF, Cecil NC, Ramgopal M, Zimmerman RK, et al. Impact of Age and Pre-Existing Immunity on the Induction of Human Antibody Responses Against Influenza B Viruses. *Hum Vaccin Immunother* (2019) 15(9):2030–43. doi: 10.1080/21645515.2019.1642056
- Goodwin K, Viboud C, Simonsen L. Antibody Response to Influenza Vaccination in the Elderly: A Quantitative Review. Vaccine (2006) 24 (8):1159–69. doi: 10.1016/j.vaccine.2005.08.105
- 336. Sasaki S, Sullivan M, Narvaez CF, Holmes TH, Furman D, Zheng NY, et al. Limited Efficacy of Inactivated Influenza Vaccine in Elderly Individuals is Associated With Decreased Production of Vaccine-Specific Antibodies. J Clin Invest (2011) 121(8):3109–19. doi: 10.1172/JCI57834
- 337. Ju CH, Blum LK, Kongpachith S, Lingampalli N, Mao R, Brodin P, et al. Plasmablast Antibody Repertoires in Elderly Influenza Vaccine Responders Exhibit Restricted Diversity But Increased Breadth of Binding Across Influenza Strains. *Clin Immunol* (2018) 193:70–9. doi: 10.1016/j.clim.2018.01.011
- 338. Henry C, Zheng NY, Huang M, Cabanov A, Rojas KT, Kaur K, et al. Influenza Virus Vaccination Elicits Poorly Adapted B Cell Responses in Elderly Individuals. *Cell Host Microbe* (2019) 25(3):357–66.e6. doi: 10.1016/ j.chom.2019.01.002
- 339. Dugan HL, Guthmiller JJ, Arevalo P, Huang M, Chen YQ, Neu KE, et al. Preexisting Immunity Shapes Distinct Antibody Landscapes After Influenza Virus Infection and Vaccination in Humans. *Sci Transl Med* (2020) 12(573): eabd3601. doi: 10.1126/scitranslmed.abd3601
- 340. Fonville JM, Wilks SH, James SL, Fox A, Ventresca M, Aban M, et al. Antibody Landscapes After Influenza Virus Infection or Vaccination. *Science* (2014) 346(6212):996–1000. doi: 10.1126/science.1256427

- 341. Sridhar S, Begom S, Bermingham A, Hoschler K, Adamson W, Carman W, et al. Cellular Immune Correlates of Protection Against Symptomatic Pandemic Influenza. *Nat Med* (2013) 19(10):1305–12. doi: 10.1038/nm.3350
- 342. Wilkinson TM, Li CK, Chui CS, Huang AK, Perkins M, Liebner JC, et al. Preexisting Influenza-Specific CD4+ T Cells Correlate With Disease Protection Against Influenza Challenge in Humans. *Nat Med* (2012) 18 (2):274–80. doi: 10.1038/nm.2612
- 343. Koutsakos M, Illing PT, Nguyen THO, Mifsud NA, Crawford JC, Rizzetto S, et al. Human Cd8(+) T Cell Cross-Reactivity Across Influenza a, B and C Viruses. Nat Immunol (2019) 20(5):613–25. doi: 10.1038/s41590-019-0320-6
- 344. Altenburg AF, Rimmelzwaan GF, de Vries RD. Virus-Specific T Cells as Correlate of (Cross-)Protective Immunity Against Influenza. Vaccine (2015) 33(4):500–6. doi: 10.1016/j.vaccine.2014.11.054
- 345. Nguyen THO, Sant S, Bird NL, Grant EJ, Clemens EB, Koutsakos M, et al. Perturbed Cd8(+) T Cell Immunity Across Universal Influenza Epitopes in the Elderly. *J Leukoc Biol* (2018) 103(2):321–39. doi: 10.1189/jlb.5MA0517-207R
- 346. Park HJ, Shin MS, Kim M, Bilsborrow JB, Mohanty S, Montgomery RR, et al. Transcriptomic Analysis of Human IL-7 Receptor Alpha (Low) and (High) Effector Memory CD8(+) T Cells Reveals an Age-Associated Signature Linked to Influenza Vaccine Response in Older Adults. *Aging Cell* (2019) 18(4):e12960. doi: 10.1111/acel.12960
- 347. Eaton SM, Burns EM, Kusser K, Randall TD, Haynes L. Age-Related Defects in CD4 T Cell Cognate Helper Function Lead to Reductions in Humoral Responses. J Exp Med (2004) 200(12):1613–22. doi: 10.1084/jem.20041395
- 348. Herati RS, Reuter MA, Dolfi DV, Mansfield KD, Aung H, Badwan OZ, et al. Circulating CXCR5+PD-1+ Response Predicts Influenza Vaccine Antibody Responses in Young Adults But Not Elderly Adults. *J Immunol* (2014) 193 (7):3528–37. doi: 10.4049/jimmunol.1302503
- 349. Lefebvre JS, Masters AR, Hopkins JW, Haynes L. Age-Related Impairment of Humoral Response to Influenza is Associated With Changes in Antigen Specific T Follicular Helper Cell Responses. *Sci Rep* (2016) 6:25051. doi: 10.1038/srep25051
- 350. Richards KA, Shannon I, Treanor JJ, Yang H, Nayak JL, Sant AJ. Evidence That Blunted CD4 T-Cell Responses Underlie Deficient Protective Antibody Responses to Influenza Vaccines in Repeatedly Vaccinated Human Subjects. *J Infect Dis* (2020) 222(2):273–7. doi: 10.1093/infdis/jiz433
- 351. Cowling BJ, Perera R, Valkenburg SA, Leung NHL, Iuliano AD, Tam YH, et al. Comparative Immunogenicity of Several Enhanced Influenza Vaccine Options for Older Adults: A Randomized, Controlled Trial. *Clin Infect Dis* (2020) 71(7):1704–14. doi: 10.1093/cid/ciz1034
- 352. Hart RJ. An Outbreak of Respiratory Syncytial Virus Infection in an Old People's Home. J Infect (1984) 8(3):259–61. doi: 10.1016/S0163-4453(84)94075-1
- 353. Sorvillo FJ, Huie SF, Strassburg MA, Butsumyo A, Shandera WX, Fannin SL. An Outbreak of Respiratory Syncytial Virus Pneumonia in a Nursing Home for the Elderly. J Infect (1984) 9(3):252–6. doi: 10.1016/S0163-4453(84)90530-9
- 354. Branche AR, Falsey AR. Respiratory Syncytial Virus Infection in Older Adults: An Under-Recognized Problem. *Drugs Aging* (2015) 32(4):261–9. doi: 10.1007/s40266-015-0258-9
- 355. Rossey I, Saelens X. Vaccines Against Human Respiratory Syncytial Virus in Clinical Trials, Where are We Now? *Expert Rev Vaccines* (2019) 18 (10):1053–67. doi: 10.1080/14760584.2019.1675520
- 356. Drysdale SB, Barr RS, Rollier CS, Green CA, Pollard AJ, Sande CJ. Priorities for Developing Respiratory Syncytial Virus Vaccines in Different Target Populations. *Sci Transl Med* (2020) 12(535):eaax2466. doi: 10.1126/ scitranslmed.aax2466
- 357. Swanson KA, Rainho-Tomko JN, Williams ZP, Lanza L, Peredelchuk M, Kishko M, et al. A Respiratory Syncytial Virus (RSV) F Protein Nanoparticle Vaccine Focuses Antibody Responses to a Conserved Neutralization Domain. *Sci Immunol* (2020) 5(47):eaba6466. doi: 10.1126/sciimmunol.aba6466
- 358. Capella C, Chaiwatpongsakorn S, Gorrell E, Risch ZA, Ye F, Mertz SE, et al. Prefusion F, and Disease Severity in Infants and Young Children With Acute Respiratory Syncytial Virus Infection. J Infect Dis (2017) 216(11):1398–406. doi: 10.1093/infdis/jix489
- 359. Blanco JCG, Pletneva LM, McGinnes-Cullen L, Otoa RO, Patel MC, Fernando LR, et al. Efficacy of a Respiratory Syncytial Virus Vaccine Candidate in a Maternal Immunization Model. Nat Commun (2018) 9(1):1904. doi: 10.1038/ s41467-018-04216-6

- 360. Palomo C, Mas V, Thom M, Vazquez M, Cano O, Terron MC, et al. Influence of Respiratory Syncytial Virus F Glycoprotein Conformation on Induction of Protective Immune Responses. J Virol (2016) 90(11):5485–98. doi: 10.1128/JVI.00338-16
- Falsey AR, Walsh EE. Relationship of Serum Antibody to Risk of Respiratory Syncytial Virus Infection in Elderly Adults. J Infect Dis (1998) 177(2):463–6. doi: 10.1086/517376
- 362. Bagga B, Cehelsky JE, Vaishnaw A, Wilkinson T, Meyers R, Harrison LM, et al. Effect of Preexisting Serum and Mucosal Antibody on Experimental Respiratory Syncytial Virus (Rsv) Challenge and Infection of Adults. *J Infect Dis* (2015) 212(11):1719–25. doi: 10.1093/infdis/jiv281
- Walsh EE, Peterson DR, Falsey AR. Risk Factors for Severe Respiratory Syncytial Virus Infection in Elderly Persons. J Infect Dis (2004) 189(2):233–8. doi: 10.1086/380907
- 364. de Bree GJ, Heidema J, van Leeuwen EM, van Bleek GM, Jonkers RE, Jansen HM, et al. Respiratory Syncytial Virus-Specific CD8+ Memory T Cell Responses in Elderly Persons. J Infect Dis (2005) 191(10):1710–8. doi: 10.1086/429695
- 365. Cusi MG, Martorelli B, Di Genova G, Terrosi C, Campoccia G, Correale P. Age Related Changes in T Cell Mediated Immune Response and Effector Memory to Respiratory Syncytial Virus (RSV) in Healthy Subjects. *Immun Ageing* (2010) 7:14. doi: 10.1186/1742-4933-7-14
- 366. Cherukuri A, Patton K, Gasser RAJr., Zuo F, Woo J, Esser MT, et al. Adults 65 Years Old and Older Have Reduced Numbers of Functional Memory T Cells to Respiratory Syncytial Virus Fusion Protein. *Clin Vaccine Immunol* (2013) 20(2):239–47. doi: 10.1128/CVI.00580-12
- Hall CB, Walsh EE, Long CE, Schnabel KC. Immunity to and Frequency of Reinfection With Respiratory Syncytial Virus. *J Infect Dis* (1991) 163(4):693– 8. doi: 10.1093/infdis/163.4.693
- 368. Jozwik A, Habibi MS, Paras A, Zhu J, Guvenel A, Dhariwal J, et al. RSV-Specific Airway Resident Memory CD8+ T Cells and Differential Disease Severity After Experimental Human Infection. *Nat Commun* (2015) 6:10224. doi: 10.1038/ncomms10224
- 369. Katz PS, Siggins RW, Porretta C, Armstrong ML, Zea AH, Mercante DE, et al. Chronic Alcohol Increases CD8+ T-Cell Immunosenescence in Simian Immunodeficiency Virus-Infected Rhesus Macaques. *Alcohol* (2015) 49 (8):759–65. doi: 10.1016/j.alcohol.2015.09.003
- 370. Cho SY, Kim J, Lee JH, Sim JH, Cho DH, Bae IH, et al. Modulation of Gut Microbiota and Delayed Immunosenescence as a Result of Syringaresinol Consumption in Middle-Aged Mice. *Sci Rep* (2016) 6:39026. doi: 10.1038/ srep39026
- 371. Turner JE, Brum PC. Does Regular Exercise Counter T Cell Immunosenescence Reducing the Risk of Developing Cancer and Promoting Successful Treatment of Malignancies? Oxid Med Cell Longev (2017) 2017:4234765. doi: 10.1155/2017/ 4234765
- 372. Turner JE. Is Immunosenescence Influenced by Our Lifetime "Dose" of Exercise? *Biogerontology* (2016) 17(3):581–602. doi: 10.1007/s10522-016-9642-z
- 373. Choi IY, Lee C, Longo VD. Nutrition and Fasting Mimicking Diets in the Prevention and Treatment of Autoimmune Diseases and Immunosenescence. *Mol Cell Endocrinol* (2017) 455:4–12. doi: 10.1016/j.mce.2017.01.042
- 374. Ripa M, Chiappetta S, Tambussi G. Immunosenescence and Hurdles in the Clinical Management of Older HIV-Patients. *Virulence* (2017) 8(5):508–28. doi: 10.1080/21505594.2017.1292197
- 375. Partridge L, Deelen J, Slagboom PE. Facing Up to the Global Challenges of Ageing. *Nature* (2018) 561(7721):45–56. doi: 10.1038/s41586-018-0457-8
- 376. Scola I, Candore G, Colonna-Romano G, Crivello A, Forte GI, Paolisso G, et al. Study of the Association With -330T/G IL-2 in a Population of Centenarians From Centre and South Italy. *Biogerontology* (2005) 6(6):425–9. doi: 10.1007/ s10522-005-4909-9
- 377. Capri M, Monti D, Salvioli S, Lescai F, Pierini M, Altilia S, et al. Complexity of Anti-Immunosenescence Strategies in Humans. *Artif Organs* (2006) 30 (10):730–42. doi: 10.1111/j.1525-1594.2006.00295.x
- Laupeze B, van der Most R, Del Giudice G. Novel Technologies to Improve Vaccines for Older Adults. *Interdiscip Top Gerontol Geriatr* (2020) 43:218– 33. doi: 10.1159/000504485
- Partridge L, Fuentealba M, Kennedy BK. The Quest to Slow Ageing Through Drug Discovery. Nat Rev Drug Discovery (2020) 19(8):513–32. doi: 10.1038/ s41573-020-0067-7

- Jun JI, Lau LF. The Matricellular Protein CCN1 Induces Fibroblast Senescence and Restricts Fibrosis in Cutaneous Wound Healing. *Nat Cell Biol* (2010) 12(7):676–85. doi: 10.1038/ncb2070
- 381. Krizhanovsky V, Yon M, Dickins RA, Hearn S, Simon J, Miething C, et al. Senescence of Activated Stellate Cells Limits Liver Fibrosis. *Cell* (2008) 134 (4):657–67. doi: 10.1016/j.cell.2008.06.049
- Munoz-Espin D, Canamero M, Maraver A, Gomez-Lopez G, Contreras J, Murillo-Cuesta S, et al. Programmed Cell Senescence During Mammalian Embryonic Development. *Cell* (2013) 155(5):1104–18. doi: 10.1016/j.cell.2013.10.019
- Kang TW, Yevsa T, Woller N, Hoenicke L, Wuestefeld T, Dauch D, et al. Senescence Surveillance of Pre-Malignant Hepatocytes Limits Liver Cancer Development. *Nature* (2011) 479(7374):547–51. doi: 10.1038/nature10599
- 384. Xue W, Zender L, Miething C, Dickins RA, Hernando E, Krizhanovsky V, et al. Senescence and Tumour Clearance is Triggered by p53 Restoration in Murine Liver Carcinomas. *Nature* (2007) 445(7128):656–60. doi: 10.1038/ nature05529
- 385. Ovadya Y, Landsberger T, Leins H, Vadai E, Gal H, Biran A, et al. Impaired Immune Surveillance Accelerates Accumulation of Senescent Cells and Aging. Nat Commun (2018) 9(1):5435. doi: 10.1038/s41467-018-07825-3
- 386. Childs BG, Durik M, Baker DJ, van Deursen JM. Cellular Senescence in Aging and Age-Related Disease: From Mechanisms to Therapy. *Nat Med* (2015) 21(12):1424–35. doi: 10.1038/nm.4000
- 387. Walters MS, De BP, Salit J, Buro-Auriemma LJ, Wilson T, Rogalski AM, et al. Smoking Accelerates Aging of the Small Airway Epithelium. *Respir Res* (2014) 15:94. doi: 10.1186/s12931-014-0094-1
- 388. Matthews C, Gorenne I, Scott S, Figg N, Kirkpatrick P, Ritchie A, et al. Vascular Smooth Muscle Cells Undergo Telomere-Based Senescence in Human Atherosclerosis: Effects of Telomerase and Oxidative Stress. *Circ Res* (2006) 99(2):156–64. doi: 10.1161/01.RES.0000233315.38086.bc
- 389. Cmielova J, Havelek R, Soukup T, Jiroutova A, Visek B, Suchanek J, et al. Gamma Radiation Induces Senescence in Human Adult Mesenchymal Stem Cells From Bone Marrow and Periodontal Ligaments. *Int J Radiat Biol* (2012) 88(5):393–404. doi: 10.3109/09553002.2012.666001
- 390. Kirkland JL, Tchkonia T. Senolytic Drugs: From Discovery to Translation. J Intern Med (2020) 288(5):518–36. doi: 10.1111/joim.13141
- 391. Baar MP, Brandt RMC, Putavet DA, Klein JDD, Derks KWJ, Bourgeois BRM, et al. Targeted Apoptosis of Senescent Cells Restores Tissue Homeostasis in Response to Chemotoxicity and Aging. *Cell* (2017) 169 (1):132–47.e16. doi: 10.1016/j.cell.2017.02.031
- 392. Singh R, Letai A, Sarosiek K. Regulation of Apoptosis in Health and Disease: The Balancing Act of BCL-2 Family Proteins. *Nat Rev Mol Cell Biol* (2019) 20 (3):175–93. doi: 10.1038/s41580-018-0089-8
- 393. Xu M, Pirtskhalava T, Farr JN, Weigand BM, Palmer AK, Weivoda MM, et al. Senolytics Improve Physical Function and Increase Lifespan in Old Age. *Nat Med* (2018) 24(8):1246–56. doi: 10.1038/s41591-018-0092-9
- 394. Yousefzadeh MJ, Zhu Y, McGowan SJ, Angelini L, Fuhrmann-Stroissnigg H, Xu M, et al. Fisetin is a Senotherapeutic That Extends Health and Lifespan. *EBioMedicine* (2018) 36:18–28. doi: 10.1016/j.ebiom.2018.09.015
- 395. Merkel O, Taylor N, Prutsch N, Staber PB, Moriggl R, Turner SD, et al. When the Guardian Sleeps: Reactivation of the p53 Pathway in Cancer. *Mutat Res* (2017) 773:1–13. doi: 10.1016/j.mrrev.2017.02.003
- 396. Wang AS, Dreesen O. Biomarkers of Cellular Senescence and Skin Aging. Front Genet (2018) 9:247. doi: 10.3389/fgene.2018.00247
- Mizukoshi E, Kaneko S. Telomerase-Targeted Cancer Immunotherapy. Int J Mol Sci (2019) 20(8):1823. doi: 10.3390/ijms20081823
- Fuhrmann-Stroissnigg H, Niedernhofer LJ, Robbins PD. Hsp90 Inhibitors as Senolytic Drugs to Extend Healthy Aging. *Cell Cycle* (2018) 17(9):1048–55. doi: 10.1080/15384101.2018.1475828
- 399. Kuilman T, Michaloglou C, Vredeveld LC, Douma S, van Doorn R, Desmet CJ, et al. Oncogene-Induced Senescence Relayed by an Interleukin-Dependent Inflammatory Network. *Cell* (2008) 133(6):1019–31. doi: 10.1016/j.cell.2008.03.039
- 400. Song S, Lam EW, Tchkonia T, Kirkland JL, Sun Y. Senescent Cells: Emerging Targets for Human Aging and Age-Related Diseases. *Trends Biochem Sci* (2020) 45(7):578–92. doi: 10.1016/j.tibs.2020.03.008
- 401. Prata L, Ovsyannikova IG, Tchkonia T, Kirkland JL. Senescent Cell Clearance by the Immune System: Emerging Therapeutic Opportunities. *Semin Immunol* (2018) 40:101275. doi: 10.1016/j.smim.2019.04.003

- 402. Basisty N, Kale A, Jeon OH, Kuehnemann C, Payne T, Rao C, et al. A Proteomic Atlas of Senescence-Associated Secretomes for Aging Biomarker Development. *PLoS Biol* (2020) 18(1):e3000599. doi: 10.1371/journal.pbio.3000599
- 403. Lei Q, Liu T, Gao F, Xie H, Sun L, Zhao A, et al. Microvesicles as Potential Biomarkers for the Identification of Senescence in Human Mesenchymal Stem Cells. *Theranostics* (2017) 7(10):2673–89. doi: 10.7150/thno.18915
- 404. Terlecki-Zaniewicz L, Lammermann I, Latreille J, Bobbili MR, Pils V, Schosserer M, et al. Small Extracellular Vesicles and Their miRNA Cargo are Anti-Apoptotic Members of the Senescence-Associated Secretory Phenotype. *Aging (Albany NY)* (2018) 10(5):1103–32. doi: 10.18632/aging.101452
- 405. Takasugi M, Okada R, Takahashi A, Virya Chen D, Watanabe S, Hara E. Small Extracellular Vesicles Secreted From Senescent Cells Promote Cancer Cell Proliferation Through Epha2. *Nat Commun* (2017) 8:15729. doi: 10.1038/ncomms15728
- 406. Walters HE, Deneka-Hannemann S, Cox LS. Reversal of Phenotypes of Cellular Senescence by pan-mTOR Inhibition. *Aging (Albany NY)* (2016) 8 (2):231–44. doi: 10.18632/aging.100872
- 407. Mannick JB, Morris M, Hockey HP, Roma G, Beibel M, Kulmatycki K, et al. TORC1 Inhibition Enhances Immune Function and Reduces Infections in the Elderly. *Sci Transl Med* (2018) 10(449):eaaq1564. doi: 10.1126/ scitranslmed.aaq1564
- 408. Dumas SN, Lamming DW. Next Generation Strategies for Geroprotection Via mTORC1 Inhibition. J Gerontol A Biol Sci Med Sci (2020) 75(1):14–23. doi: 10.1093/gerona/glz056
- 409. Henson SM, Lanna A, Riddell NE, Franzese O, Macaulay R, Griffiths SJ, et al. p38 Signaling Inhibits mTORC1-independent Autophagy in Senescent Human CD8(+) T Cells. J Clin Invest (2014) 124(9):4004–16. doi: 10.1172/JCI75051
- 410. Zhang S, Kaplan MH. The p38 Mitogen-Activated Protein Kinase is Required for IL-12-induced IFN-Gamma Expression. J Immunol (2000) 165(3):1374–80. doi: 10.4049/jimmunol.165.3.1374
- 411. Alimbetov D, Davis T, Brook AJ, Cox LS, Faragher RG, Nurgozhin T, et al. Suppression of the Senescence-Associated Secretory Phenotype (SASP) in Human Fibroblasts Using Small Molecule Inhibitors of P38 MAP Kinase and MK2. *Biogerontology* (2016) 17(2):305–15. doi: 10.1007/s10522-015-9610-z
- 412. Freund A, Patil CK, Campisi J. p38MAPK is a Novel DNA Damage Response-Independent Regulator of the Senescence-Associated Secretory Phenotype. *EMBO J* (2011) 30(8):1536–48. doi: 10.1038/emboj.2011.69
- 413. Lanna A, Gomes DC, Muller-Durovic B, McDonnell T, Escors D, Gilroy DW, et al. A Sestrin-Dependent Erk-Jnk-p38 MAPK Activation Complex Inhibits Immunity During Aging. *Nat Immunol* (2017) 18(3):354–63. doi: 10.1038/ni.3665
- 414. Chung HY, Kim HJ, Kim JW, Yu BP. The Inflammation Hypothesis of Aging: Molecular Modulation by Calorie Restriction. Ann N Y Acad Sci (2001) 928:327–35. doi: 10.1111/j.1749-6632.2001.tb05662.x
- 415. Chung KW, Kim DH, Park MH, Choi YJ, Kim ND, Lee J, et al. Recent Advances in Calorie Restriction Research on Aging. *Exp Gerontol* (2013) 48 (10):1049–53. doi: 10.1016/j.exger.2012.11.007
- 416. Walters HE, Cox LS. Mtorc Inhibitors as Broad-Spectrum Therapeutics for Age-Related Diseases. Int J Mol Sci (2018) 19(8):2325. doi: 10.3390/ ijms19082325
- 417. Barzilai N, Crandall JP, Kritchevsky SB, Espeland MA. Metformin as a Tool to Target Aging. *Cell Metab* (2016) 23(6):1060–5. doi: 10.1016/j.cmet.2016.05.011
- 418. Ansaldi F, Bacilieri S, Durando P, Sticchi L, Valle L, Montomoli E, et al. Cross-Protection by MF59-adjuvanted Influenza Vaccine: Neutralizing and Haemagglutination-Inhibiting Antibody Activity Against A(H3N2) Drifted Influenza Viruses. *Vaccine* (2008) 26(12):1525–9. doi: 10.1016/j.vaccine. 2008.01.019
- 419. Beran J, Reynales H, Poder A, Yu CY, Pitisuttithum P, Yuan LL, et al. Prevention of Influenza During Mismatched Seasons in Older Adults With an MF59-adjuvanted Quadrivalent Influenza Vaccine: A Randomised, Controlled, Multicentre, Phase 3 Efficacy Study. *Lancet Infect Dis* (2021) S1473-3099(20):30694-0. doi: 10.1016/S1473-3099(20)30694-0
- 420. Cioncada R, Maddaluno M, Vo HTM, Woodruff M, Tavarini S, Sammicheli C, et al. Vaccine Adjuvant MF59 Promotes the Intranodal Differentiation of Antigen-Loaded and Activated Monocyte-Derived Dendritic Cells. *PLoS One* (2017) 12(10):e0185843. doi: 10.1371/journal.pone.0185843
- O'Hagan DT, Ott GS, De Gregorio E, Seubert A. The Mechanism of Action of MF59 - an Innately Attractive Adjuvant Formulation. *Vaccine* (2012) 30 (29):4341–8. doi: 10.1016/j.vaccine.2011.09.061

- 422. Nicholson KG, Abrams KR, Batham S, Clark TW, Hoschler K, Lim WS, et al. Immunogenicity and Safety of a Two-Dose Schedule of Whole-Virion and AS03A-adjuvanted 2009 Influenza A (H1N1) Vaccines: A Randomised, Multicentre, Age-Stratified, Head-to-Head Trial. *Lancet Infect Dis* (2011) 11(2):91–101. doi: 10.1016/S1473-3099(10)70296-6
- 423. Didierlaurent AM, Laupeze B, Di Pasquale A, Hergli N, Collignon C, Garcon N. Adjuvant System AS01: Helping to Overcome the Challenges of Modern Vaccines. Expert Rev Vaccines (2017) 16(1):55–63. doi: 10.1080/14760584. 2016.1213632
- 424. Lal H, Cunningham AL, Godeaux O, Chlibek R, Diez-Domingo J, Hwang SJ, et al. Efficacy of an Adjuvanted Herpes Zoster Subunit Vaccine in Older Adults. N Engl J Med (2015) 372(22):2087–96. doi: 10.1056/NEJMoa1501184
- 425. Weinberger B. Adjuvant Strategies to Improve Vaccination of the Elderly Population. Curr Opin Pharmacol (2018) 41:34–41. doi: 10.1016/j.coph.2018.03.014
- 426. Holland D, Booy R, De Looze F, Eizenberg P, McDonald J, Karrasch J, et al. Intradermal Influenza Vaccine Administered Using a New Microinjection System Produces Superior Immunogenicity in Elderly Adults: A Randomized Controlled Trial. J Infect Dis (2008) 198(5):650–8. doi: 10.1086/590434
- 427. DiazGranados CA, Dunning AJ, Kimmel M, Kirby D, Treanor J, Collins A, et al. Efficacy of High-Dose Versus Standard-Dose Influenza Vaccine in Older Adults. N Engl J Med (2014) 371(7):635–45. doi: 10.1056/NEJMoa1315727
- 428. Ng TWY, Cowling BJ, Gao HZ, Thompson MG. Comparative Immunogenicity of Enhanced Seasonal Influenza Vaccines in Older Adults: A Systematic Review and Meta-Analysis. J Infect Dis (2019) 219 (10):1525–35. doi: 10.1093/infdis/jiy720
- 429. Shinde V, Fries L, Wu Y, Agrawal S, Cho I, Thomas DN, et al. Improved Titers Against Influenza Drift Variants With a Nanoparticle Vaccine. N Engl J Med (2018) 378(24):2346–8. doi: 10.1056/NEJMc1803554
- 430. Anderson EJ, Rouphael NG, Widge AT, Jackson LA, Roberts PC, Makhene M, et al. Safety and Immunogenicity of SARS-CoV-2 Mrna-1273 Vaccine in Older Adults. N Engl J Med (2020) 383(25):2427–38. doi: 10.1056/NEJMoa2028436
- 431. Andrew MK, Shinde V, Ye L, Hatchette T, Haguinet F, Dos Santos G, et al. The Importance of Frailty in the Assessment of Influenza Vaccine Effectiveness Against Influenza-Related Hospitalization in Elderly People. J Infect Dis (2017) 216(4):405–14. doi: 10.1093/infdis/jix282
- 432. Netea MG, Dominguez-Andres J, Barreiro LB, Chavakis T, Divangahi M, Fuchs E, et al. Defining Trained Immunity and its Role in Health and Disease. *Nat Rev Immunol* (2020) 20(6):375–88. doi: 10.1038/s41577-020-0285-6
- 433. Netea MG, Joosten LA, Latz E, Mills KH, Natoli G, Stunnenberg HG, et al. Trained Immunity: A Program of Innate Immune Memory in Health and Disease. *Science* (2016) 352(6284):aaf1098. doi: 10.1126/science.aaf1098
- 434. Higgins JP, Soares-Weiser K, Lopez-Lopez JA, Kakourou A, Chaplin K, Christensen H, et al. Association of BCG, DTP, and Measles Containing Vaccines With Childhood Mortality: Systematic Review. *BMJ* (2016) 355: i5170. doi: 10.1136/bmj.i5170
- 435. Aaby P, Roth A, Ravn H, Napirna BM, Rodrigues A, Lisse IM, et al. Randomized Trial of BCG Vaccination At Birth to Low-Birth-Weight Children: Beneficial Nonspecific Effects in the Neonatal Period? J Infect Dis (2011) 204(2):245–52. doi: 10.1093/infdis/jir240
- 436. Ter Horst R, Jaeger M, Smeekens SP, Oosting M, Swertz MA, Li Y, et al. Host and Environmental Factors Influencing Individual Human Cytokine Responses. *Cell* (2016) 167(4):1111–24.e13. doi: 10.1016/j.cell.2016.10.018
- 437. Wardhana EA, Sultana A, Mandang VV, Jim E. The Efficacy of Bacillus Calmette-Guerin Vaccinations for the Prevention of Acute Upper Respiratory Tract Infection in the Elderly. *Acta Med Indones* (2011) 43(3):185–90.
- 438. Leentjens J, Kox M, Stokman R, Gerretsen J, Diavatopoulos DA, van Crevel R, et al. Bcg Vaccination Enhances the Immunogenicity of Subsequent Influenza Vaccination in Healthy Volunteers: A Randomized, Placebo-Controlled Pilot Study. J Infect Dis (2015) 212(12):1930–8. doi: 10.1093/infdis/jiv332
- 439. Bulut O, Kilic G, Dominguez-Andres J, Netea MG. Overcoming Immune Dysfunction in the Elderly: Trained Immunity as a Novel Approach. Int Immunol (2020) 32(12):741–53. doi: 10.1093/intimm/dxaa052
- 440. Gorgoulis V, Adams PD, Alimonti A, Bennett DC, Bischof O, Bishop C, et al. Cellular Senescence: Defining a Path Forward. *Cell* (2019) 179(4):813–27. doi: 10.1016/j.cell.2019.10.005
- 441. Baker DJ, Wijshake T, Tchkonia T, LeBrasseur NK, Childs BG, van de Sluis B, et al. Clearance of p16Ink4a-positive Senescent Cells Delays Ageing-Associated Disorders. *Nature* (2011) 479(7372):232–6. doi: 10.1038/nature10600

442. Demaria M, Ohtani N, Youssef SA, Rodier F, Toussaint W, Mitchell JR, et al. An Essential Role for Senescent Cells in Optimal Wound Healing Through Secretion of PDGF-AA. *Dev Cell* (2014) 31(6):722–33. doi: 10.1016/j.devcel.2014.11.012

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2021 Palacios-Pedrero, Osterhaus, Becker, Elbahesh, Rimmelzwaan and Saletti. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.