



PD-L1-Mediated Immunosuppression in Oral Squamous Cell Carcinoma: Relationship With Macrophage Infiltration and Epithelial to Mesenchymal Transition Markers

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To date, immune check-point inhibitors (ICIs), particularly inhibitors of programmed cell death-1 (PD-1) and PD ligand-1 (PD-L1) have become prominent in cancer treatment and also improved life expectancy of cancer patients. As key regulators of PD-1/PD-L1 axis, the recruitment of tumor-associated macrophages (TAMs) enhances aggressive and invasive properties of tumors in immunosuppressive tumor microenvironment (TME) and promotes epithelial-mesenchymal transition (EMT). The aims of the study were first to characterize the critical links among PD-L1, TME and EMT process and, further, to explore the sensitivity of different chemical agents to different PD-L1 expression groups. Bioinformatical analysis revealed that PD-L1 was highly expressed in OSCC and higher PD-L1 expression correlated with worse survival in patients. Notably, PD-L1 was positively correlated with macrophages infiltration and EMT markers gene expression. Moreover, patients in the PD-L1^{high} group were at a significant chance of benefiting from ICI treatment and they also showed higher sensitivity to the chemical drugs (olaparib, paclitaxel, docetaxel, and pazopanib). These findings implicate PD-L1 could serve as a novel target for prognostic and therapeutic approaches in OSCC patients; PD-L1mediated immune evasion might be attributable to the infiltration of macrophages, resulting EMT progress; Chemical agents in combination with PD-L1 inhibitor could be served as personalized treatment plan for OSCC patients so as to maximize patient benefit.

Keywords: PD-L1, immunosuppressive tumor microenvironment, macrophages, epithelial-mesenchymal transition, chemical drugs

INTRODUCTION

Oral squamous cell carcinoma (OSCC), as one of the 10 most frequent cancers with approximately 300,000 new cases diagnosed annually is perceived as an immunosuppressive cancer (1). Despite advances in therapeutic approaches, including surgical methods and radiotherapy, patients with locally advanced or metastatic OSCC still face the risk of a poor prognosis (2). Therefore, further efforts are still demanded to identify clinically relevant biomarkers, establish effective mechanism-based combinations, and develop effective targeted therapies for OSCC.

Cancer immunotherapy, including immune check-point inhibitors (ICIs), Oncolytic virotherapy (OVT), and chimeric antigen receptor (CAR) T cells has led to major improvements in tumor treatment over the past two decades (3). ICIs expressed on the cell surface, including programmed cell death-1 (PD-1) and PD ligand-1 (PD-L1), play crucial roles in activating negative regulatory pathways and evading immune surveillance (4). Upon activation, ICIs can dampen antitumor immune responses, leading to cancer cells escaping from host immune system (5). The interaction between these ligands can be blocked by ICIs, thereby reactivating the cytocidal immune response. Unprecedented advances in tumor control have been made using therapeutic monoclonal antibodies (mAbs) to block ICIs. Particularly, mAbs targeting PD-1/PD-L1 have brought great clinical benefits in multiple indications, either as monotherapy or in combination regimens (6-8). Pembrolizumab and nivolumab as anti-PD-1 mAbs have shown remarkable antitumor activity in the treatment of head and neck squamous cell carcinoma (HNSCC), leading to their regulatory approval (9-11). The PD-1/PD-L1 interaction inhibits antitumor activity of cytotoxic lymphocytes (CTLs) (12), which contributes to multiple Suppressive effects, such as immune escape, tumor proliferation, invasion, angiogenesis, and epithelial-mesenchymal transition (EMT) (13). At present, the links among PD-L1, TME, and EMT process in OSCC are not well understood, and the expectation that immunotherapy in combination with standard-therapy can maximize patient benefit necessitates further research on PD-L1mediated immunosuppression. The present study was aimed at characterizing the critical links among PD-L1, TME and EMT process in OSCC, and further exploring the sensitivity of ICI treatment and different chemical agents to different PD-L1 expression groups.

METHODS AND MATERIALS

Data Acquisition

All Clinical and sequencing data were obtained from the Cancer Genome Atlas (TCGA) and the Gene Expression omnibus (GEO) research network. Transcriptome data of 150 OSCC tissue samples and 30 normal oral tissue samples were extracted from TCGA. Patients' information on age, tumornode-metastasis (TNM), stage, survival time and status were organized (**Supplementary Table 1**). The GEO database was used to obtain the OSCC microarray data set GSE30784, and 167 OSCC cancer patient samples were selected as the validation set based on the sample information (14). The EMT related genes were extracted from the Molecular Signature Database (MSigDB) (15).

Bioinformatic Analysis

Differentially expressed genes (DEGs) in different groups were identified using EdgeR package and filtered by | log2 (Fold Change) | > 1 and adjusted P value < 0.05 which was adjusted using the Benjamini-Hochberg (BH) approach (16). Furthermore, the enrichment analysis including Gene ontology (GO) function and Kyoto Encyclopedia of Genes and Genomes (KEGG) pathway annotation was conducted by cluster Profiler package. The PD-L1 expression level in diverse cancer types and correlation between PD-L1 and immune infiltrates were conducted using Tumor Immune Estimation Resource (TIMER) database (17). Gene marker sets of immune cell types were obtained from Bindea et al. (18) and Newman et al. (19). Single-sample Gene Set Enrichment Analysis (ssGSEA) and relative abundance of immune cells were calculated by Gene Set Variation Analysis (GSVA) (20). The classical chemokines and markers of macrophages and EMT signaling pathway were also included (21-24). Tumor Immune Dysfunction and Exclusion (TIDE) algorithm was applied to get individual immunotherapy response (25, 26), and individual chemotherapeutic response was predicted based on Genomics of Drug Sensitivity in Cancer (GDSC) database (27, 28). Drug sensitivity prediction was perform using R package 'pRRophetic' (29).

Statistical Analysis

R statistical language and SPSS 22.0 software were used for statistical analyses. Expression differences among different tumor grade groups were compared using one-way analysis of variance (ANOVA) test. Overall survival (OS) distribution and survival curves were performed by R package survival. The optimal cutoff point for PD-L1 expression level was determined by the 'surv_cutpoint' function of 'survminer' R package and calculated utilizing the maximally selected rank statistics that calculated the most optimal cut-off for continuous variables using log-rank statistics. Independent prognostic factors were evaluated using univariate and multivariate Cox proportional hazards regression analyses, and relationships between variables were calculated using Pearson correlation coefficients. Differences of TIDE scores and chemotherapy responses between groups were analyzed using Wilcoxon rank sum test.

RESULTS

PD-L1 Was Highly Expressed in OSCC and Predicted Poor OS

PD-L1 was differentially expressed in different tumor types and was highly expressed in HNSCC (**Supplementary Figure 1A**). TIMER analysis revealed that PD-L1 had positive correlations with various types of immune cells in HNSCC (**Supplementary Figure 1B**). Similarly, in OSCC, PD-L1 was also highly expressed compared with control group, but it gradually decreased with the progress of

tumor stages (P < 0.05, **Figures 1A, B**). Prognostic value of PD-L1 in OSCC patients was further analyzed; Kaplan–Meier analysis of OS revealed that patients with high PD-L1 expression exhibited a shorter survival time (P < 0.05, **Figures 1C–E**); The cutoff point for PD-L1 was 3.66. Cox regression analyses revealed that PD-L1 upregulation was significantly associated with a poor OS (**Table 1**).

Functional Enrichment Analysis of DEGs

Differential expression analysis was performed to identify the DEGs between two groups, so as to further study the function of these genes (**Figure 2A**). Among these DEGs, 384 genes were upregulated in PD-L1^{high} group, and 1090 genes up-regulated in PD-L1^{low} group (**Figure 2A**).

Additionally, tumor-associated macrophages (TAMs) characteristic cytokines such as CXCL10 and CXCL11 were significantly differentially expressed (Figures 2A, B). Notably, the

GO enrichment analysis indicated 745 immune-related GO terms including lymphocyte activation, activated leukocyte adhesion, and effector cytokine production were significantly enriched in PD-L1^{high} group, while 414 GO terms including digestion, regulation of postsynaptic membrane potential, drug transport, glutamate receptor signaling pathway, regulation of membrane potential were significantly enriched in PD-L1^{low} group (**Figures 2C, D**). KEGG pathway analysis showed that 47 KEGG pathways were characteristic enriched in PD-L1^{low} group, (**Figures 2E, F**).

PD-L1 Was Correlated With Macrophage Infiltration and Macrophage-Derived Chemokines

The results of GSVA revealed that PD-L1 was positively associated with cell proliferation of 12 immune cell types in



FIGURE 1 | Comprehensive analysis of PD-L1 in OSCC. (A) PD-L1 expression in OSCC group and the control group. (B) PD-L1 expression in different clinical stages of OSCC. (C) The optimal cutoff threshold value (D) Kaplan–Meier survival analysis according to the optimal cutoff value. (E) PD-L1 expression level in different OSCC groups.

Туре	Univariate Cox Analysis			Multivariate Cox Analysis		
	P values	HR	95%CI	P values	HR	95%CI
Age	0.289998	1.205949	0.852479 - 1.705982	0.341135	1.223076	0.807963 - 1.851465
Т	0.00025	1.413316	1.174466 - 1.700741	0.031705	1.433184	1.032024 - 1.99028
Ν	0.000497	1.46845	1.18289 - 1.822947	0.063043	1.320594	0.984993 - 1.770538
Stage	0.000488	1.470667	1.184029 - 1.826695	0.834734	1.053163	0.647391 - 1.713264
Grade	0.053395	1.307115	0.996091- 1.715256	0.135115	1.294857	0.922618 - 1.81728
PD-L1	0.02162	1.502635	1.061568 - 2.126959	0.01391	1.66808	1.109532 - 2.507807

TABLE 1 | Prognostic value of PD-L1 expression in OSCC.



FIGURE 2 | Functional enrichment analysis of DEGs in PD-L1high and PD-L1low groups. (A) Volcanic map of DEGs. (B) Heat map showing top DEGs. Top 10 enriched GO terms in PD-L1 high (C) and PD-L1low group (D). Top 10 significantly enriched KEGG pathways in PD-L1 high (E) and PD-L1low group (F).

OSCC (**Supplementary Figure 2** and **Figures 3A, B**), especially with M0 macrophages (cor = 0.42, P < 0.001), M1 macrophages (cor = 0.68, P < 0.001), and M2 macrophages (cor = 0.53, P < 0.001) (**Figures 3C–E**). In addition, the results of ssGSEA in the GEO OSCC validation set showed PD-L1 was strongly positively associated with cell proliferation in several types of immune cell infiltration (**Supplementary Figures 3A–H**), especially with M1 macrophages (cor = 0.63, P < 0.001, **Supplementary Figure 3D**) and M2 macrophages (cor = 0.54, P < 0.001, **Supplementary Figure 3E**).

To validate the above results, we further analyzed correlations between PD-L1 expression and macrophage markers, which revealed that M1-related chemokines (IL12A, IL-12B, IL-23A, TNF, and IFNG) and M2-related chemokines (TGFBs, IL-10, and IL-13) showed the strongest positive correlations with PD-L1 expression (P < 0.05, **Figures 4A–D**). We also observed statistically significant correlations between M1-related chemokines and M2-related chemokines (**Figures 4B, D**).

Associations Between PD-L1 Expression, Macrophage Infiltration and EMT Biomarkers

Since the EMT process has been considered to be particularly relevant to TME, we analyzed the potential association of EMT with immune infiltration, and found significant correlations between EMT biomarkers and various immune cells infiltration (**Figure 5A**), especially with M0 macrophages (cor = 0.568, P < 0.001) and M2 macrophages (cor = 0.425, P < 0.001). We also evaluated the association between PD-L1 and EMT biomarkers, which showed significant correlations between PD-L1 and vimentin (VIM) (cor = 0.322, P < 0.001) (**Figures 5B, C**).

Sensitivity Differences to Immunotherapy/ Chemotherapy Between Groups

TIDE algorithm was employed to assess individual immunotherapy response in different PD-L1 expression groups, and higher TIDE prediction score represented a higher



FIGURE 3 | Relationship between immune cell infiltration and PD-L1 expression in OSCC samples. (A) Heat map by using the ssGSEA scores from 26 immune cell types. (B) Correlation between PD-L1 and immune infiltrates in OSCC samples. Correlation between PD-L1 and M0 (C), M1 (D) and M2 (E) macrophages infiltration in OSCC samples.

immune evasion potential (Figure 6A). The results revealed patients with higher PD-L1 expression had a higher microsatellite instability (MSI) score and T cell dysfunction score (Figures 6B, C), but a lower T cell exclusion score (Figure 6D), indicating that these patients were less likely to benefit from ICI treatment.

We also took into account the differences in the response of chemotherapy in OSCC patients, and evaluated sensitivity of two patient groups to four chemical drugs (olaparib, docetaxel, paclitaxel, and pazopanib). The IC50 value of each sample in OSCC was estimated, and significant differences were observed between groups, which revealed that PD-L1^{high} group showed higher sensitivity to all drugs (**Figures 6E–H**, *P* < 0.001).

DISCUSSION

PD-L1 overexpression has been demonstrated in many common cancers, inducing T-cell tolerance and promoting immune escape. Blockades targeted on PD-1/PD-L1 have already shown striking effectiveness in clinical applications (6–8). Immune characteristics relevant to PD-L1 in TME and EMT process during cancer

progression were depicted in our study. Firstly, PD-L1 expression was highly expressed in OSCC, but it gradually decreased with the progress of tumor stages. As an immunosuppressive cell surface molecule that promotes T cell depletion, PD-L1 upregulation may link with increased cancer aggression and poorer prognosis, as proposed in several previous studies (29, 30). In contrast to this expectation, we observed an decrease in PD-L1 expression as disease progressed; Some previous studies on the association between PD-L1 expression and improved prognosis also Supported our findings (31-34). The most likely explanation for this paradox is that PD-L1 expression can be induced by cytokines, primarily the production of interferon- γ within TME, and therefore, its expression actually reflects the contribution of endogenous anti-tumor immune response, which typically occurred in the early stages of tumor development and progression (35, 36). Moreover, our results also showed that PD-L1 was positively associated with cell proliferation in activated TAMs and EMT process. The DEGs in PD-L1^{high} group were significantly enriched in canonical signaling pathways that related to regulation of lymphocyte activation, suggesting the critical involvement of PD-L1 in regulating TAMs function. Additionally, our results suggested that patients in the PD-L1^{high}



(D) Correlation between PD-L1 and M2-derived chemokines. *P < 0.05; **P < 0.01; ***P < 0.001.

group were at a significant chance of benefiting from ICI treatment and they also showed higher sensitivity to the chemical drugs (olaparib, paclitaxel, docetaxel, and pazopanib). This study provided preliminary evidence regarding the tight correlations among PD-L1, TAMs and EMT process, further supporting the notion that clinical efficacy of both chemotherapy and immunotherapy could be greatly improved by utilizing PD-L1 inhibitors in OSCC.

A previous research on PD-L1 regulating the proliferation of macrophages revealed that the phenotype and function of macrophages could be altered by anti-PD-L1 treatment, suggesting a crucial role of PD-L1 in regulating macrophages activation and function (37). Circulating monocytes can be recruited into TME and further polarize into TAMs. Classically, in response to microenvironmental stimuli, TAMs polarize to M1like phenotype exhibiting proinflammatory and tumor-inhibiting phenotypic effects, while certain cytokines convert TAMs into an M2-like phenotype with anti-inflammatory but tumor-promoting functions (38, 39). Previous studies have shown that M2 macrophages elevated PD-L1 expression in cancer cells and at the same time facilitating immune escape (40, 41). As for M1 macrophages, the PD-L1 expressed on the surface was also reported to result in immune escape of cancer cells, resulting in bidirectional effects of both anti-tumoral and pro-tumoral activities (42): On one hand, M1-like phenotype had the unique ability to promote the activation and recruitment of various immune effectors, performing surveillance tasks (43), and their infiltration indicated good prognosis in some cancers (44). On the other hand, M1 macrophages involved in cancer phenotype maintenance and tumorigenicity regulation in vivo. For example, previous studies confirmed the important protumorigenic factor role of M1 macrophages in urethane-induced lung tumorigenesis (45); Higher aggregation level of human leukocyte antigen-DR+ (HLA-DR+) M1-like TAMs was related to poor response to



ionizing radiotherapy in rectal cancers patients (46). In the present study, positive association between PD-L1 expression and TAMs infiltration (both M0, M1 and M2) was observed in OSCC, which was in consistent with previous findings. These findings suggested that TAMs may contribute to pro-tumorigenic effects by promoting PD-L1 expression in OSCC.

EMT is a novel mechanism involved in cancer metastasis, by which epithelial cells acquire both mesenchymal and epithelial phenotypes for cell migration and proliferation. The type III intermediate filament protein VIM that constitutes a key cytoskeletal element of mesenchymal cells, is a canonical marker of EMT process, and EMT process is characterized by marked VIM upregulation (47, 48). It is generally accepted that epithelial cells undergoing EMT are able to survive better under adverse environmental conditions, which enables tumor cells to evade immune destruction (49, 50). In addition, accumulating evidences have suggested strong correlations between EMT and immune evasion by activating multiple ICIs (51, 52); Cancer cells with PD-L1 upregulation displays an EMT phenotype that aids in immune escape. Macrophages and cancer cells were reported to establish a two-way cross-talk: Macrophages facilitated EMT changes in the latter while the latter skewed TAMs polarization into an M2 phenotype (53). Interestingly, recent evidences have indicated that in addition to M2 phenotype, M1-like macrophages also promoted EMT and chemoresistance (54); TAMs generally share both M1- and M2like phenotypes instead of being strictly classified into above two phenotypes (55), which does not rule out the possibility that these two phenotypes are exchangeable (56, 57).

Immuno-oncology revolutionized cancer treatment. Pembrolizumab and nivolumab as anti-PD-1 mAbs have shown remarkable anti-tumor activity in the treatment of patients with recurrent/metastatic HNSCC. Ferris et al. (10) experienced nivolumab in a population of 347 HNSCC patients to evaluate the efficacy of nivolumab comparable to that of single-agent chemotherapy (CheckMate 141): Both ORR (13.3% *vs* 5.8%) and median OS (7.5 *vs* 5.1 months) were significantly improved in the nivolumab group. The estimated 12-month OS in nivolumab group was 36% *versus* 16.6% in standard-therapy group, while there was little difference in median progression-free survival



(PFS) of these two groups. Similarly, the KEYNOTE-040 phase III study also compared the clinical efficacy of pembrolizumab versus current standard-therapy with a total of 495 patients enrolled: Both ORR (14.6% vs 10.1%) and median OS (8.4 vs 6.9 months) were significantly improved in the pembrolizumab group (11). These randomized phase III trials indicated that survival benefit could be well conferred by immunotherapy. Thus, it is of great importance to perform immune monitoring on patients, thereby identifying potential biomarkers, accurately stratifying patients and delineating responders and non-responders. Olaparib (AZD. 2281), a competitive inhibitor of poly (ADP-ribose) polymerase-1 (PARP-1), has been used in the clinical treatment of ovarian cancer with BRCA1/2 gene mutations (58, 59). An ongoing phase 1/2 study of olaparib and the PD-L1 inhibitor durvalumab in breast cancer patients with BRCA1/2 mutations demonstrated that 24 of 30 patients who were eligible for trial entry by study design achieved durable and adaptable cancer control at 12 weeks of combination therapy (60). Olaparib in combination with durvalumab exhibited promising anti-tumor efficacy and safety, which was confirmed in numerous clinical studies (61-63). In addition, paclitaxel, including nanoparticle albumin-bound paclitaxel (nab-paclitaxel), is a widely used chemotherapy drug for various cancers; Combination therapy of anti-PD-L1 mAb atezolizumab and nab-paclitaxel as a first-line treatment exhibited significantly improved PFS in patients with PD-L1-positive tumors (64, 65). In this study, we analyzed the sensitivity differences of immunotherapy and chemotherapy between different PD-L1 expression groups with the expectation of screening potential benefit populations and achieving enhanced efficacy. The results revealed that patients in the PD-L1^{high} groups were at a significant chance of benefiting from ICI treatment and they

also showed higher sensitivity to the four chemical drugs (olaparib, paclitaxel, docetaxel, and pazopanib. Further research is needed to fully confirm the promising efficacy of these agents in combination with PD-L1 inhibitor in improving personalized treatment for OSCC patients. By screening the subgroups of potential beneficiaries, immunotherapy and chemotherapy can harnessed to maximize the immunostimulatory effects of therapeutic agents.

DATA AVAILABILITY STATEMENT

The original contributions presented in the study are included in the article/**Supplementary Material**. Further inquiries can be directed to the corresponding author.

AUTHOR CONTRIBUTIONS

Conceived and designed the study: TW, CT, and RT. Data collection and analysis: TW and RT. Writing and revising the manuscript: TW, CT, XY, QJ, CF, and RT. All authors contributed to the article and approved the submitted version.

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SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fimmu.2021. 693881/full#supplementary-material

REFERENCES

- Seiwert TY, Zuo Z, Keck MK, Khattri A, Pedamallu CS, Stricker T, et al. Integrative and Comparative Genomic Analysis of HPV-Positive and HPV-Negative Head and Neck Squamous Cell Carcinomas. *Clin Cancer Res* (2015) 21(3):632–41. doi: 10.1158/1078-0432.CCR-13-3310
- Cohen EE, Davis DW, Karrison TG, Seiwert TY, Wong SJ, Nattam S, et al. Erlotinib and Bevacizumab in Patients With Recurrent or Metastatic Squamous-Cell Carcinoma of the Head and Neck: A Phase I/II Study. Lancet Oncol (2009) 10(3):247–57. doi: 10.1016/S1470-2045(09)70002-6
- Dougan M, Dranoff G. Immune Therapy for Cancer. Annu Rev Immunol (2009) 27:83–117. doi: 10.1146/annurev.immunol.021908.132544
- Pardoll DM. The Blockade of Immune Checkpoints in Cancer Immunotherapy. Nat Rev Cancer (2012) 12(4):252–64. doi: 10.1038/nrc3239
- Dong H, Strome SE, Salomao DR, Tamura H, Hirano F, Flies DB, et al. Tumor-Associated B7-H1 Promotes T-Cell Apoptosis: A Potential Mechanism of Immune Evasion. *Nat Med* (2002) 8(8):793-800. doi: 10.1038/nm730
- Okazaki T, Chikuma S, Iwai Y, Fagarasan S, Honjo T. A Rheostat for Immune Responses: The Unique Properties of PD-1 and Their Advantages for Clinical Application. *Nat Immunol* (2013) 14(12):1212–8. doi: 10.1038/ni.2762
- Sharma P, Allison JP. Immune Checkpoint Targeting in Cancer Therapy: Toward Combination Strategies With Curative Potential. *Cell* (2015) 161 (2):205–14. doi: 10.1016/j.cell.2015.03.030
- Sharma P, Allison JP. The Future of Immune Checkpoint Therapy. Science (2015) 348(6230):56–61. doi: 10.1126/science.aaa8172
- Seiwert TY, Burtness B, Mehra R, Weiss J, Berger R, Eder JP, et al. Safety and Clinical Activity of Pembrolizumab for Treatment of Recurrent or Metastatic Squamous Cell Carcinoma of the Head and Neck (KEYNOTE-012): An Open-Label, Multicentre, Phase 1b Trial. *Lancet Oncol* (2016) 17(7):956–65. doi: 10.1016/s1470-2045(16)30066-3
- Ferris RL, Blumenschein GJr., Fayette J, Guigay J, Colevas AD, Licitra L, et al. Nivolumab for Recurrent Squamous-Cell Carcinoma of the Head and Neck. N Engl J Med (2016) 375(19):1856–67. doi: 10.1056/NEJMoa1602252
- Cohen EEW, Soulières D, Le Tourneau C, Dinis J, Licitra L, Ahn MJ, et al. Pembrolizumab Versus Methotrexate, Docetaxel, or Cetuximab for Recurrent or Metastatic Head-and-Neck Squamous Cell Carcinoma (KEYNOTE-040): A Randomised, Open-Label, Phase 3 Study. Lancet (2019) 393(10167):156–67. doi: 10.1016/s0140-6736(18)31999-8
- Raimondi C, Carpino G, Nicolazzo C, Gradilone A, Gianni W, Gelibter A, et al. PD-L1 and Epithelial-Mesenchymal Transition in Circulating Tumor Cells From Non-Small Cell Lung Cancer Patients: A Molecular Shield to Evade Immune System? *Oncoimmunology* (2017) 6(12):e1315488. doi: 10.1080/2162402X.2017.1315488
- Zou W, Wolchok JD, Chen L. PD-L1 (B7-H1) and PD-1 Pathway Blockade for Cancer Therapy: Mechanisms, Response Biomarkers, and Combinations. *Sci Transl Med* (2016) 8(328):328rv4. doi: 10.1126/scitranslmed.aad7118
- Chen C, Méndez E, Houck J, Fan W, Lohavanichbutr P, Doody D, et al. Gene Expression Profiling Identifies Genes Predictive of Oral Squamous Cell Carcinoma. *Cancer Epidemiol Biomarkers Prev* (2008) 17(8):2152–62. doi: 10.1158/1055-9965.Epi-07-2893

Supplementary Figure 1 | PD-L1 expression level in various cancers. (A) PD-L1 expression level in various cancers was analyzed by box-plot. (B) Correlation between PD-L1 and immune infiltrates in HNSCC.

Supplementary Figure 2 | Scatter diagram of the correlation between PD-L1 expression and immune cell infiltration in OSCC.

Supplementary Figure 3 | Relationship between PD-L1 and immune infiltrates in the GEO validation set. (A) Heat map of OSCC samples by using the ssGSEA scores from 26 immune cell types. (B) PD-L1 significantly associated with Immune cells. Scatter plot showing correlation between PD-L1 and various immune cells, including aDC (C), M1 macrophages (D), M2 macrophages (E), Cytotoxic cells (F), T cells (G) and Th1 cells (H).

Supplementary Table 1 | The Clinicopathological data of patients.

- Hanahan D, Weinberg RA. Hallmarks of Cancer: The Next Generation. Cell (2011) 144(5):646–74. doi: 10.1016/j.cell.2011.02.013
- Duggal P, Gillanders EM, Holmes TN, Bailey-Wilson JE. Establishing an Adjusted P-Value Threshold to Control the Family-Wide Type 1 Error in Genome Wide Association Studies. *BMC Genomics* (2008) 9:516. doi: 10.1186/1471-2164-9-516
- Li T, Fan J, Wang B, Traugh N, Chen Q, Liu JS, et al. TIMER: A Web Server for Comprehensive Analysis of Tumor-Infiltrating Immune Cells. *Cancer Res* (2017) 77(21):e108–e10. doi: 10.1158/0008-5472.Can-17-0307
- Bindea G, Mlecnik B, Tosolini M, Kirilovsky A, Waldner M, Obenauf AC, et al. Spatiotemporal Dynamics of Intratumoral Immune Cells Reveal the Immune Landscape in Human Cancer. *Immunity* (2013) 39(4):782–95. doi: 10.1016/j.immuni.2013.10.003
- Newman AM, Liu CL, Green MR, Gentles AJ, Feng W, Xu Y, et al. Robust Enumeration of Cell Subsets From Tissue Expression Profiles. *Nat Methods* (2015) 12(5):453–7. doi: 10.1038/nmeth.3337
- Hänzelmann S, Castelo R, Guinney J. GSVA: Gene Set Variation Analysis for Microarray and RNA-Seq Data. BMC Bioinf (2013) 14:7. doi: 10.1186/1471-2105-14-7
- Mantovani A, Sica A, Sozzani S, Allavena P, Vecchi A, Locati M. The Chemokine System in Diverse Forms of Macrophage Activation and Polarization. *Trends Immunol* (2004) 25(12):677–86. doi: 10.1016/ j.it.2004.09.015
- Murray PJ, Allen JE, Biswas SK, Fisher EA, Gilroy DW, Goerdt S, et al. Macrophage Activation and Polarization: Nomenclature and Experimental Guidelines. *Immunity* (2014) 41(1):14–20. doi: 10.1016/j.immuni.2014.06.008
- Li W, Graeber MB. The Molecular Profile of Microglia Under the Influence of Glioma. Neuro Oncol (2012) 14(8):958–78. doi: 10.1093/neuonc/nos116
- Karacosta LG, Anchang B, Ignatiadis N, Kimmey SC, Benson JA, Shrager JB, et al. Mapping Lung Cancer Epithelial-Mesenchymal Transition States and Trajectories With Single-Cell Resolution. *Nat Commun* (2019) 10(1):5587. doi: 10.1038/s41467-019-13441-6
- 25. Liu Z, Zhang Y, Shi C, Zhou X, Xu K, Jiao D, et al. A Novel Immune Classification Reveals Distinct Immune Escape Mechanism and Genomic Alterations: Implications for Immunotherapy in Hepatocellular Carcinoma. *J Transl Med* (2021) 19(1):5. doi: 10.1186/s12967-020-02697-y
- 26. Jiang P, Gu S, Pan D, Fu J, Sahu A, Hu X, et al. Signatures of T Cell Dysfunction and Exclusion Predict Cancer Immunotherapy Response. *Nat Med* (2018) 24(10):1550–8. doi: 10.1038/s41591-018-0136-1
- 27. Yang W, Soares J, Greninger P, Edelman EJ, Lightfoot H, Forbes S, et al. Genomics of Drug Sensitivity in Cancer (GDSC): A Resource for Therapeutic Biomarker Discovery in Cancer Cells. *Nucleic Acids Res* (2013) 41(Database issue):D955–61. doi: 10.1093/nar/gks1111
- Fu J, Li K, Zhang W, Wan C, Zhang J, Jiang P, et al. Large-Scale Public Data Reuse to Model Immunotherapy Response and Resistance. *Genome Med* (2020) 12(1):21. doi: 10.1186/s13073-020-0721-z
- Nduom EK, Wei J, Yaghi NK, Huang N, Kong LY, Gabrusiewicz K, et al. PD-L1 Expression and Prognostic Impact in Glioblastoma. *Neuro Oncol* (2016) 18 (2):195–205. doi: 10.1093/neuonc/nov172
- 30. Fanale D, Incorvaia L, Badalamenti G, De Luca I, Algeri L, Bonasera A, et al. Prognostic Role of Plasma PD-1, PD-L1, Pan-BTN3As and BTN3A1 in

Patients Affected by Metastatic Gastrointestinal Stromal Tumors: Can Immune Checkpoints Act as a Sentinel for Short-Term Survival? *Cancers* (*Basel*) (2021) 13(9):2118. doi: 10.3390/cancers13092118

- 31. Bang YJ, Ruiz EY, Van Cutsem E, Lee KW, Wyrwicz L, Schenker M, et al. Phase III, Randomised Trial of Avelumab Versus Physician's Choice of Chemotherapy as Third-Line Treatment of Patients With Advanced Gastric or Gastro-Oesophageal Junction Cancer: Primary Analysis of JAVELIN Gastric 300. Ann Oncol (2018) 29(10):2052–60. doi: 10.1093/annonc/mdy264
- 32. Taube JM, Anders RA, Young GD, Xu H, Sharma R, McMiller TL, et al. Colocalization of Inflammatory Response With B7-H1 Expression in Human Melanocytic Lesions Supports an Adaptive Resistance Mechanism of Immune Escape. Sci Transl Med (2012) 4(127):127ra37. doi: 10.1126/scitranslmed.3003689
- 33. Badoual C, Hans S, Merillon N, Van Ryswick C, Ravel P, Benhamouda N, et al. PD-1-Expressing Tumor-Infiltrating T Cells are a Favorable Prognostic Biomarker in HPV-Associated Head and Neck Cancer. *Cancer Res* (2013) 73(1):128–38. doi: 10.1158/0008-5472.Can-12-2606
- Roper E, Lum T, Palme CE, Ashford B, Ch'ng S, Ranson M, et al. PD-L1 Expression Predicts Longer Disease Free Survival in High Risk Head and Neck Cutaneous Squamous Cell Carcinoma. *Pathology* (2017) 49(5):499–505. doi: 10.1016/j.pathol.2017.04.004
- Davar D, Wang H, Chauvin JM, Pagliano O, Fourcade JJ, Ka M, et al. Phase Ib/II Study of Pembrolizumab and Pegylated-Interferon Alfa-2b in Advanced Melanoma. J Clin Oncol (2018) 36(35):JCO1800632. doi: 10.1200/ JCO.18.00632
- Carey CD, Gusenleitner D, Lipschitz M, Roemer MGM, Stack EC, Gjini E, et al. Topological Analysis Reveals a PD-L1-Associated Microenvironmental Niche for Reed-Sternberg Cells in Hodgkin Lymphoma. *Blood* (2017) 130 (22):2420–30. doi: 10.1182/blood-2017-03-770719
- Hartley GP, Chow L, Ammons DT, Wheat WH, Dow SW. Programmed Cell Death Ligand 1 (PD-L1) Signaling Regulates Macrophage Proliferation and Activation. *Cancer Immunol Res* (2018) 6(10):1260–73. doi: 10.1158/2326-6066.Cir-17-0537
- Cheng H, Wang Z, Fu L, Xu T. Macrophage Polarization in the Development and Progression of Ovarian Cancers: An Overview. *Front Oncol* (2019) 9:421. doi: 10.3389/fonc.2019.00421
- Guadagno E, Presta I, Maisano D, Donato A, Pirrone CK, Cardillo G, et al. Role of Macrophages in Brain Tumor Growth and Progression. *Int J Mol Sci* (2018) 19(4):1005. doi: 10.3390/ijms19041005
- Qian M, Ling W, Ruan Z. Long non-Coding RNA SNHG12 Promotes Immune Escape of Ovarian Cancer Cells Through Their Crosstalk With M2 Macrophages. Aging (Albany NY) (2020) 12(17):17122–36. doi: 10.18632/ aging.103653
- Kim SH, Go SI, Song DH, Park SW, Kim HR, Jang I, et al. Prognostic Impact of CD8 and Programmed Death-Ligand 1 Expression in Patients With Resectable Non-Small Cell Lung Cancer. Br J Cancer (2019) 120(5):547–54. doi: 10.1038/s41416-019-0398-5
- 42. Kuang DM, Zhao Q, Peng C, Xu J, Zhang JP, Wu C, et al. Activated Monocytes in Peritumoral Stroma of Hepatocellular Carcinoma Foster Immune Privilege and Disease Progression Through PD-L1. J Exp Med (2009) 206(6):1327–37. doi: 10.1084/jem.20082173
- O'Sullivan T, Saddawi-Konefka R, Vermi W, Koebel CM, Arthur C, White JM, et al. Cancer Immunoediting by the Innate Immune System in the Absence of Adaptive Immunity. J Exp Med (2012) 209(10):1869–82. doi: 10.1084/ jem.20112738
- 44. Ohri CM, Shikotra A, Green RH, Waller DA, Bradding P. Macrophages Within NSCLC Tumour Islets are Predominantly of a Cytotoxic M1 Phenotype Associated With Extended Survival. *Eur Respir J* (2009) 33 (1):118–26. doi: 10.1183/09031936.00065708
- Zaynagetdinov R, Sherrill TP, Polosukhin VV, Han W, Ausborn JA, McLoed AG, et al. A Critical Role for Macrophages in Promotion of Urethane-Induced Lung Carcinogenesis. J Immunol (2011) 187(11):5703–11. doi: 10.4049/ jimmunol.1100558
- Shaikh S, Noshirwani A, West N, Perry S, Jayne D. Can Macrophages Within the Microenvironment of Locally Invasive Rectal Cancers Predict Response to Radiotherapy? *Lancet* (2015) 385(Supl 1):S87. doi: 10.1016/s0140-6736(15) 60402-0
- Pardo-Sánchez JM, Mancheño N, Cerón J, Jordá C, Ansotegui E, Juan Ó, et al. Increased Tumor Growth Rate and Mesenchymal Properties of NSCLC-

Patient-Derived Xenograft Models During Serial Transplantation. *Cancers* (*Basel*) (2021) 13(12):2980. doi: 10.3390/cancers13122980

- Yao JX, Chen X, Zhu YJ, Wang H, Hu XY, Guo JM. Prognostic Value of Vimentin Is Associated With Immunosuppression in Metastatic Renal Cell Carcinoma. *Front Oncol* (2020) 10:1181. doi: 10.3389/fonc.2020.01181
- Friedl P, Alexander S. Cancer Invasion and the Microenvironment: Plasticity and Reciprocity. *Cell* (2011) 147(5):992–1009. doi: 10.1016/j.cell.2011.11.016
- Schreiber RD, Old LJ, Smyth MJ. Cancer Immunoediting: Integrating Immunity's Roles in Cancer Suppression and Promotion. *Science* (2011) 331(6024):1565–70. doi: 10.1126/science.1203486
- Chen L, Heymach JV, Qin FX, Gibbons DL. The Mutually Regulatory Loop of Epithelial-Mesenchymal Transition and Immunosuppression in Cancer Progression. Oncoimmunology (2015) 4(5):e1002731. doi: 10.1080/ 2162402x.2014.1002731
- Chouaib S, Janji B, Tittarelli A, Eggermont A, Thiery JP. Tumor Plasticity Interferes With Anti-Tumor Immunity. *Crit Rev Immunol* (2014) 34(2):91– 102. doi: 10.1615/critrevimmunol.2014010183
- Sommariva M, Gagliano N. E-Cadherin in Pancreatic Ductal Adenocarcinoma: A Multifaceted Actor During EMT. *Cells* (2020) 9 (4):1040. doi: 10.3390/cells9041040
- 54. Kuwada K, Kagawa S, Yoshida R, Sakamoto S, Ito A, Watanabe M, et al. The Epithelial-to-Mesenchymal Transition Induced by Tumor-Associated Macrophages Confers Chemoresistance in Peritoneally Disseminated Pancreatic Cancer. J Exp Clin Cancer Res (2018) 37(1):307. doi: 10.1186/ s13046-018-0981-2
- Helm O, Held-Feindt J, Grage-Griebenow E, Reiling N, Ungefroren H, Vogel I, et al. Tumor-Associated Macrophages Exhibit Pro- and Anti-Inflammatory Properties by Which They Impact on Pancreatic Tumorigenesis. *Int J Cancer* (2014) 135(4):843–61. doi: 10.1002/ijc.28736
- Pathria P, Louis TL, Varner JA. Targeting Tumor-Associated Macrophages in Cancer. Trends Immunol (2019) 40(4):310–27. doi: 10.1016/j.it.2019.02.003
- Hu W, Li X, Zhang C, Yang Y, Jiang J, Wu C. Tumor-Associated Macrophages in Cancers. *Clin Transl Oncol* (2016) 18(3):251–8. doi: 10.1007/s12094-015-1373-0
- Moore K, Colombo N, Scambia G, Kim BG, Oaknin A, Friedlander M, et al. Maintenance Olaparib in Patients With Newly Diagnosed Advanced Ovarian Cancer. N Engl J Med (2018) 379(26):2495–505. doi: 10.1056/NEJMoa1810858
- Poveda A, Floquet A, Ledermann JA, Asher R, Penson RT, Oza AM, et al. Olaparib Tablets as Maintenance Therapy in Patients With Platinum-Sensitive Relapsed Ovarian Cancer and a BRCA1/2 Mutation (SOLO2/ ENGOT-Ov21): A Final Analysis of a Double-Blind, Randomised, Placebo-Controlled, Phase 3 Trial. *Lancet Oncol* (2021) 22(5):620–31. doi: 10.1016/ S1470-2045(21)00073-5
- Domchek SM, Postel-Vinay S, Im SA, Park YH, Delord JP, Italiano A, et al. Olaparib and Durvalumab in Patients With Germline BRCA-Mutated Metastatic Breast Cancer (MEDIOLA): An Open-Label, Multicentre, Phase 1/2, Basket Study. *Lancet Oncol* (2020) 21(9):1155–64. doi: 10.1016/s1470-2045(20)30324-7
- 61. Fumet JD, Limagne E, Thibaudin M, Truntzer C, Bertaut A, Rederstorff E, et al. Precision Medicine Phase II Study Evaluating the Efficacy of a Double Immunotherapy by Durvalumab and Tremelimumab Combined With Olaparib in Patients With Solid Cancers and Carriers of Homologous Recombination Repair Genes Mutation in Response or Stable After Olaparib Treatment. BMC Cancer (2020) 20(1):748. doi: 10.1186/s12885-020-07253-x
- 62. Karzai F, VanderWeele D, Madan RA, Owens H, Cordes LM, Hankin A, et al. Activity of Durvalumab Plus Olaparib in Metastatic Castration-Resistant Prostate Cancer in Men With and Without DNA Damage Repair Mutations. *J Immunother Cancer* (2018) 6(1):141. doi: 10.1186/s40425-018-0463-2
- Lampert EJ, Zimmer A, Padget M, Cimino-Mathews A, Nair JR, Liu Y, et al. Combination of PARP Inhibitor Olaparib, and PD-L1 Inhibitor Durvalumab, in Recurrent Ovarian Cancer: A Proof-Of-Concept Phase II Study. *Clin Cancer Res* (2020) 26(16):4268–79. doi: 10.1158/1078-0432.Ccr-20-0056
- 64. Schmid P, Rugo HS, Adams S, Schneeweiss A, Barrios CH, Iwata H, et al. Atezolizumab Plus Nab-Paclitaxel as First-Line Treatment for Unresectable, Locally Advanced or Metastatic Triple-Negative Breast Cancer (IMpassion130): Updated Efficacy Results From a Randomised, Double-Blind, Placebo-Controlled, Phase 3 Trial. *Lancet Oncol* (2020) 21(1):44–59. doi: 10.1016/s1470-2045(19)30689-8

 Schmid P, Adams S, Rugo HS, Schneeweiss A, Barrios CH, Iwata H, et al. Atezolizumab and Nab-Paclitaxel in Advanced Triple-Negative Breast Cancer. N Engl J Med (2018) 379(22):2108–21. doi: 10.1056/NEJMoa1809615

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