



Approaches to Establishing Tolerance in Immune Mediated Diseases

Michelle F. Huffaker^{1†}, Srinath Sanda^{1†}, Sindhu Chandran^{1†}, Sharon A. Chung^{1†}, E. William St. Clair², Gerald T. Nepom³ and Dawn E. Smilek^{1*}

¹ Immune Tolerance Network, University of California San Francisco, San Francisco, CA, United States, ² Immune Tolerance Network, Duke University, Durham, NC, United States, ³ Immune Tolerance Network, Benaroya Research Institute, Seattle, WA, United States

OPEN ACCESS

Edited by:

Amy Rosenberg,
United States Food and Drug
Administration, United States

Reviewed by:

Nathan Karin,
Technion Israel Institute of
Technology, Israel
Matthias G. Von Herrath,
La Jolla Institute for Immunology (LJI),
United States

*Correspondence:

Dawn E. Smilek
dsmilek@immunetolerance.org

[†]These authors have contributed
equally to this work and share
first authorship

Specialty section:

This article was submitted to
Immunological Tolerance
and Regulation,
a section of the journal
Frontiers in Immunology

Received: 20 July 2021

Accepted: 25 August 2021

Published: 20 September 2021

Citation:

Huffaker MF, Sanda S, Chandran S,
Chung SA, St. Clair EW, Nepom GT
and Smilek DE (2021) Approaches
to Establishing Tolerance in
Immune Mediated Diseases.
Front. Immunol. 12:744804.
doi: 10.3389/fimmu.2021.744804

The development of rational approaches to restore immune tolerance requires an iterative approach that builds on past success and utilizes new mechanistic insights into immune-mediated pathologies. This article will review concepts that have evolved from the clinical trial experience of the Immune Tolerance Network, with an emphasis on lessons learned from the innovative mechanistic studies conducted for these trials and new strategies under development for induction of tolerance.

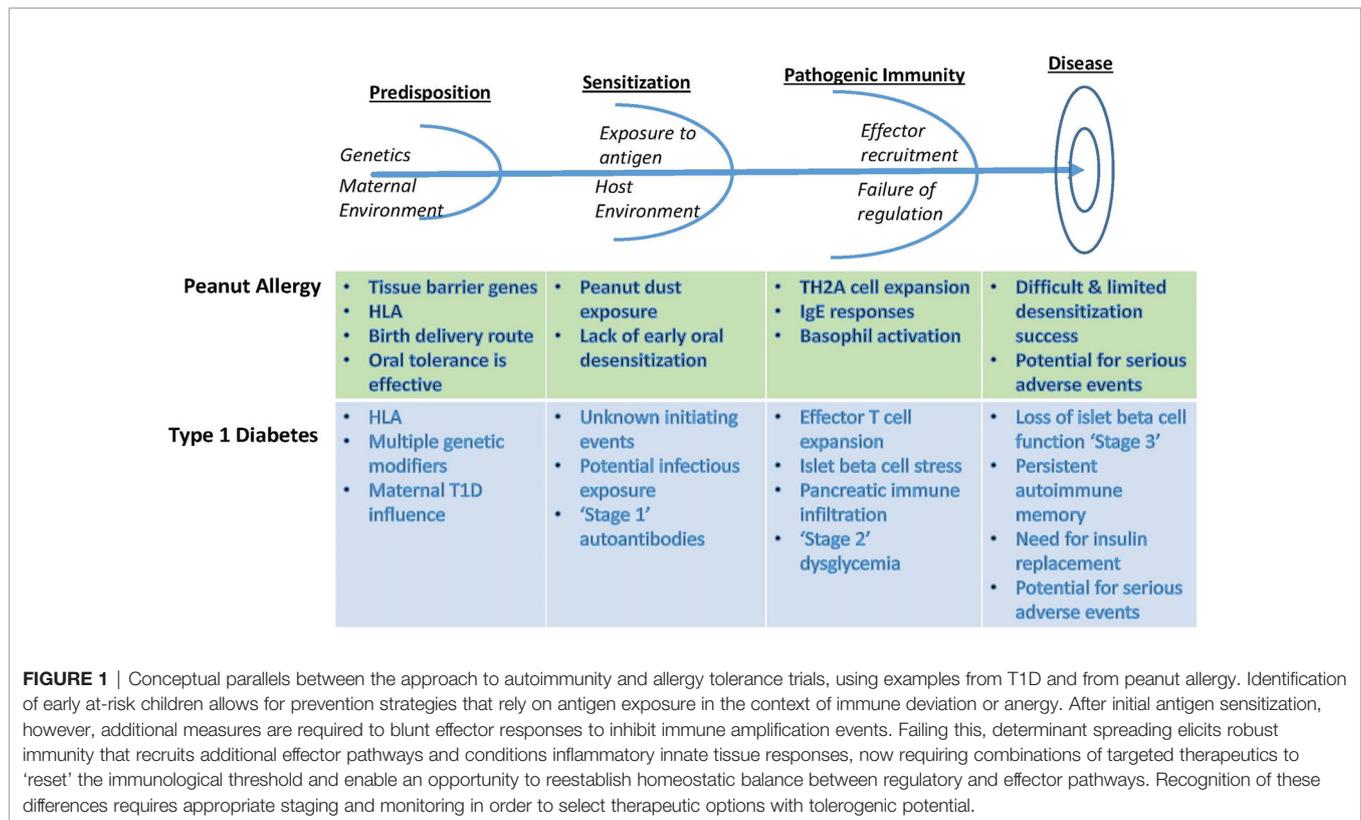
Keywords: immune tolerance, allergy, transplantation, autoimmunity, clinical trial, immunomodulation, immunosuppression

INTRODUCTION

The mission of the Immune Tolerance Network (ITN) is to advance the development of immune tolerance strategies in autoimmunity, allergy, and transplantation by conducting high-quality clinical trials with emerging therapeutic agents. Integrated mechanism-based research is a critical component of these trials that provide new insights into the success or failure of the intervention, as well as further understanding of disease pathogenesis. These results in turn provide the building blocks for further clinical trials and the design of new treatment strategies and incremental advancement towards the tolerance goal.

Central to the ITN's mission is the concept of immunologic tolerance. While specific clinical definitions of tolerance vary across the immune mediated diseases, they all center on differentiating permanent or prolonged improvements in disease that represent a significant clinical benefit over the expected natural history of disease or the standard of care. Conceptual parallels are illustrated in **Figure 1**. In the transplant setting, tolerance is defined as graft acceptance without the need for a continuous immunosuppressive regimen, or a greatly reduced regimen. In allergic disease, tolerance may be defined as prolonged unresponsiveness to antigen challenge or exposure after withdrawal of allergen immunotherapy. For autoimmune diseases, tolerance is reflected in reduced need for disease-modifying therapy or prolonged improvement of disease manifestations, such as retention of residual insulin secretion in type 1 diabetes.

However, at a molecular level, immunologic tolerance represents prolonged or permanent modulation of aberrant immune responses towards a homeostatic state. Immune self-tolerance normally occurs in T and B lymphocytes by central and peripheral mechanisms, reviewed previously (1). Central tolerance involves elimination of lymphocytes with high affinity receptors for self-components, a process that takes place in T lymphocytes and B lymphocytes during cellular maturation in the thymus and bone marrow, respectively. Self-reactive lymphocytes that are not



eliminated centrally during development exit into the periphery where they are restrained by a variety of tolerance mechanisms, included induction of anergy, cellular exhaustion, and suppression by regulatory T cells, B cells, innate immune cells, and inhibitory cytokines.

In disease conditions, immunologic tolerance can in theory be re-established through a variety of mechanisms which may expand or augment regulatory cells (e.g. Tr1, Treg, Breg) and suppress effector responses (e.g. effector cell depletion, co-stimulation blockade, anti-cytokine therapy) that in tandem act to restore immune homeostasis and disease quiescence. In this review, we provide an overview of ITN successes, challenges, and new strategies to achieve immune tolerance in the fields of allergy, autoimmune disease, and solid organ transplantation. **Table 1** summarizes the ITN trials mentioned in this review. Summaries of all ITN trials are available at www.immunetolerance.org.

BARRIERS TO TOLERANCE

Immunological memory is a hallmark of successful immune responses, and is essential for pathogen surveillance and extinction. It is also a formidable barrier to successful immune tolerance induction. The challenge of reversing pathogenic immune responses in individuals with autoimmune disease and allergy requires not only directed therapy against immune effector cells, but also prevention of recurrent memory responses when therapy is discontinued. This concept also plays a role in

transplantation, both through initial heterologous memory and subsequently in dealing with the robust alloimmune response. Innate immune activation, memory, and self-perpetuating inflammatory cascades must be restrained, allowing tissue repair to occur. Therefore, a major focus of immune tolerance strategies is to retain and expand regulatory immune mechanisms, thereby exploiting homeostatic pathways that are intrinsic to a healthy balance of immune effector and regulatory compartments.

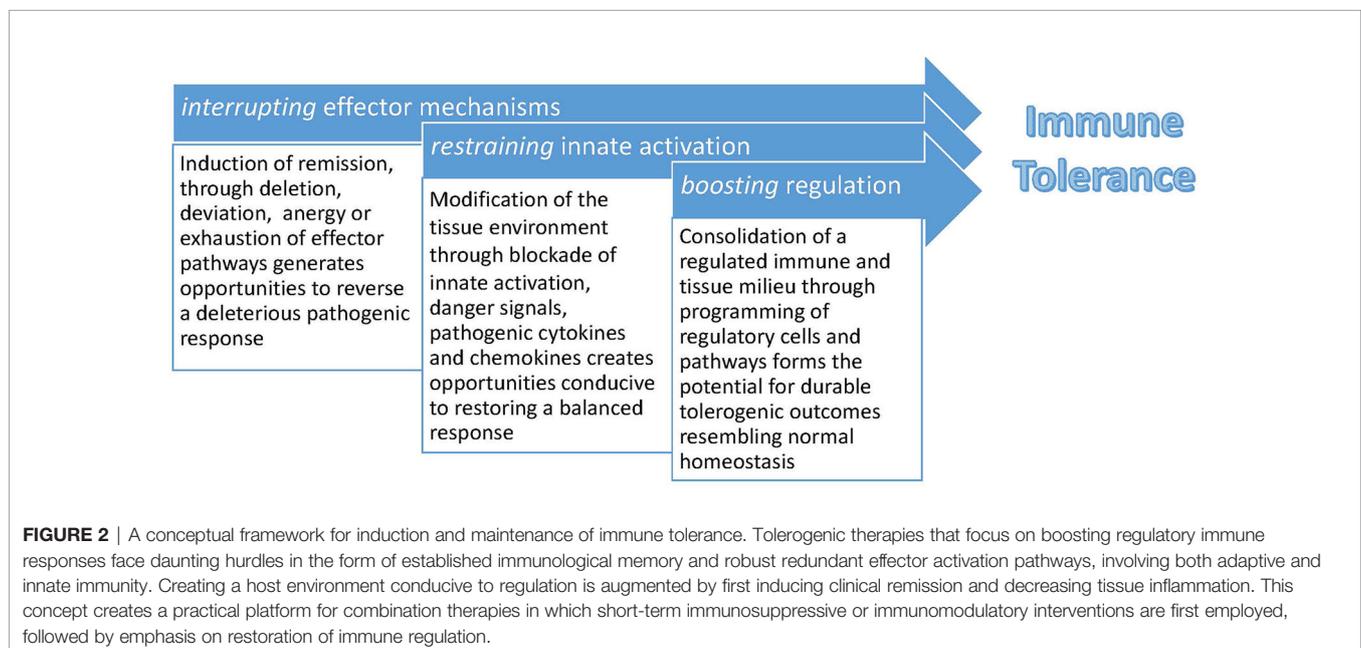
This combination of *interrupting* effector mechanisms, *restraining* innate activation, and *boosting* regulation is the central dogma of successful immune tolerance therapy, illustrated in **Figure 2**. Therapies that achieve only one of these goals without the others, as discussed in the examples below, achieve suboptimal or transient clinical benefit. And because immunological memory is very resilient, early intervention – at the time of transplant or early in the immune process prior to the onset of clinical disease in allergy or autoimmunity – can be attempted whenever at-risk individuals can be identified.

Underlying these efforts to rebalance immune effector and regulatory responses is the recognition that drastic immune suppression is accompanied by significant safety concerns, notably infectious and neoplastic risk, as well as life-long drug administration. Consequently, the targets of immune tolerance therapy need to be selective, addressing the ITN's '*interrupting/restraining/boosting*' therapeutic dogma without creating an unacceptable level of danger. Selective targets within the adaptive immune system may involve specific subsets of T or B cells, or even antigen-specific receptors and signaling pathways;

TABLE 1 | Summary of ITN Clinical Trials Included in this Review.

Disease Indication	Trial Name	Intervention	Status	References	ClinicalTrials.gov Identifier
Allergy					
At risk for peanut allergy	LEAP	Peanut	Complete	(2–7)	NCT00329784
	LEAP-On				NCT01366846
Peanut allergic	IMPACT	Peanut	Complete	(8)	NCT01867671
At risk for atopy	ACTIVATE	Vaginal microbiome seeding	Ongoing		NCT03567707
Grass allergic	GRADUATE	Timothy grass + Dupilumab	Ongoing		NCT04502966
Grass allergic	GRASS	Timothy grass	Complete	(9)	NCT01335139
Cat allergic	CATNIP	Cat allergen + Tezepelumab	Complete		NCT02237196
Transplantation					
Kidney transplant	Mixed	Immunosuppression withdrawal following donor hematopoietic stem cell transplant	Complete	(10)	NCT00063817, NCT00801632
Kidney transplant	Chimerism	Immunosuppression withdrawal following donor mesenchymal stromal cells	Ongoing		NCT03504241
Liver transplant	TEACH				
	LITTMUS	Immunosuppression withdrawal following donor specific Tregs	Ongoing		NCT03577431, NCT03654040
HLA sensitization awaiting kidney transplant	ADAPT	Carfilzomib plus belatacept	Planned		NCT05017545
	ATTAIN	Daratumumab plus belatacept	Planned		NCT04827979
Autoimmunity					
Treatment-resistant multiple sclerosis	HALT-MS	Autologous hematopoietic stem cell transplant	Complete	(11–13)	NCT00288626
	BEAT-MS	Autologous hematopoietic stem cell transplant	Ongoing		NCT04047628
ANCA associated vasculitis	RAVE	Rituximab	Complete	(14)	NCT00104299
Lupus nephritis	CALIBRATE	Rituximab + Belimumab	Complete	(15)	NCT02260934
Primary membranous nephropathy	REBOOT	Rituximab + Belimumab	Ongoing		NCT03949855
Antiphospholipid syndrome	DARE-APS	Daratumumab	Planned		Pending
New onset type 1 diabetes	AbATE	Teplizumab	Complete	(16–18)	NCT00129259
New onset type 1 diabetes	T1DAL	Alefacept	Complete	(19–21)	NCT00965458
New onset type 1 diabetes	START	Antithymocyte globulin	Complete	(22)	NCT00515099
Lupus nephritis	ACCESS	Abatacept	Complete	(23)	NCT00774852
Multiple sclerosis	ACCLAIM	Abatacept	Complete	(24, 25)	NCT01116427
Psoriasis vulgaris	PAUSE	Ustekinumab + Abatacept	Complete	(26)	NCT01999868

Summaries of all ITN trials are available at www.immunetolerance.org.



and selectivity in a broader sense may target a limited set of cytokine, chemokine, or activation pathways without global immunosuppression. Rapid development over the last two decades of novel immune therapeutic agents that meet these types of selectivity requirements has enabled the design of clinical trials with aspirations for achieving immune tolerance. Although transplantation, allergy, and autoimmune diseases have many distinct characteristics, conducting such trials within the ITN provides an opportunity to study immune mechanisms and concepts across clinical disciplines while evaluating safety, therapeutic durability, and homeostatic immune reconstitution.

ALLERGY

Desensitization strategies have been a mainstay of allergy therapy for many years, for many types of environmental allergens, including hymenoptera venom and aeroallergens such as pollens, dust mites, and animal dander. On this background of clinical experience showing that allergen specificity of an allergic response that can often be modulated by controlled antigen exposure, the ITN has designed several clinical trials intended to achieve durable, long-lasting allergen desensitization, i.e. tolerance.

IgE-mediated peanut allergy typically develops in the first two years of life and for the majority affected, the disease persists into adulthood. Sensitization to peanut may involve cutaneous and enteral pathways that lead to production of TH2 cytokines and antigen-specific IgE. The Learning Early about Peanut Allergy (LEAP) trial demonstrated that early oral exposure to peanut prevented the development of peanut allergy in atopic infants including those who had already developed sensitization to peanut (2). This protection proved to be durable through the LEAP-On trial, which assessed peanut allergy status after one year of avoidance of peanut following completion of the oral peanut intervention (3). Nearly complete protection in the LEAP cohort of infants with family histories of peanut allergy and eczema, who were at high risk for development of peanut allergy, led to the publication of new public health guidelines for safely introducing peanut protein into the early childhood diet (4). This protection was highly allergen specific, as consumption of peanut did not prevent the development of other food allergies (5).

While it is possible that antigen specific prevention may be applicable for many food allergens, there are practical limitations to introducing multiple allergens in young infants, including the ability of young infants to comply with ingestion of multiple different foods in sufficient quantity to prevent allergic disease (27). Prevention strategies are needed that are personalized based on individual risk of developing a specific allergy. For example, subsequent whole genome sequencing from the LEAP trial revealed a novel association for peanut allergy with a single nucleotide variant in the mucosa-associated lymphoid tissue lymphoma translocation (*MALT1*) gene (6). *MALT1* encodes a paracaspase that acts in response to antigen binding to the T-cell or B-cell receptor leading to NF κ B activation (28). The association of *MALT1* with peanut allergy was found to be independent of atopic dermatitis and egg allergy, suggesting that carrier status

predisposed to a unique risk for peanut allergy specifically, and correlated with the progressive acquisition of IgE antibodies to multiple allergenic peanut protein components (6). This immunological relationship was further explored by analyzing the development of peanut specific IgE to a broad repertoire of linear epitopes after the second year of life (7). Peanut specific IgE in infants sensitized to peanut and consuming it, however, recognized conformational epitopes without expansion of peanut specific IgE reactivity to linear epitopes. These findings suggest an interaction between genetics, age of peanut exposure, and likelihood of tolerance, opening possibilities for personalized prevention and intervention strategies based on distinct phenotypic and genotypic risk factors.

Observations in another ITN study, the Oral Immunotherapy for Induction of Tolerance and Desensitization in Peanut-Allergic Children (IMPACT) trial, further emphasize this point. IMPACT was a randomized double-blind placebo controlled trial of peanut oral immunotherapy (OIT) in peanut allergic children ages 12-48 months. Successful desensitization at the end of OIT and persistent desensitization after 6 months of withholding OIT was associated with lower baseline peanut specific IgE, particularly in children below the age of 3 years (8). While tolerance was achieved in ~70% of the younger children with low initial IgE, overall the success rate was much lower, indicating an opportunity may exist early in disease during which the host immune response may be more receptive to development of tolerance. Together, LEAP and IMPACT demonstrate that the atopic march, the paradigm for the progression from atopic dermatitis to allergen-specific disease, can be halted or even reversed early in its course for at least a single allergen-specific disease.

These studies illustrate the concept that the younger immune system, not surprisingly, is more amenable to tolerization using antigen desensitization. They also suggest opportunities to attempt tolerization therapy in older, more established allergy, using agents to encourage an immune response that resembles the immature phenotype. Towards that end, two novel strategies are currently under ITN development, one using microbiome immune modulation and another using anti-cytokine agents in combination with desensitization. Microbiome modification is conceptually a way of providing an innate adjuvant to encourage tolerization, and potentially provide a complementary approach to antigen-specific preventative therapy by inhibiting the development of atopic diseases. Differences in environmental exposure and associated alterations of the microbiome (e.g., gut, nasopharyngeal, airway epithelium) have been implicated in the development of asthma and allergic disease (29). In the LEAP trial, colonization with *S. aureus* amongst young participants was associated with increased IgE production, persistence of egg allergy, and inhibition of oral tolerance to peanut (30). In the Copenhagen Prospective Studies on Asthma in Childhood 2010 mother-child cohort, the risk of asthma at 6 years [OR 2.45 (95% CI 1.32 to 4.55), $P = 0.004$] and allergic sensitization at 18 months [OR 1.68 (95% CI 1.01 to 2.79), $P = 0.046$] was higher in infants delivered by Cesarean section compared to vaginally-delivered infants. Since the increased risk of asthma was

identified in infants born by Cesarean section whose gut microbiome was less mature at 1 year of age (i.e., retained a Cesarean section microbiome signature), it is possible the infant gut microbiome may be a contributing factor to this predisposition for the development of asthma (31). In addition, in neonates who have a high risk for developing asthma, the gut microbiome produces metabolites, which in murine studies have been shown to increase pulmonary inflammation and decrease regulatory T (Treg) cell abundance in the lung (32). Taken together, these studies suggest that altering the newborn microbiome could favorably impact the risk of allergic and atopic disease. To address this hypothesis, the Vaginal Microbiome Exposure and Immune Responses in C-section Infants (ACTIVATE) trial is studying the impact of vaginal microbiome seeding on the development of allergic disease in infants delivered by Cesarean section (NCT03567707). Longitudinal samples of the gut, skin, nasal, and oral microbiomes will be collected over the child's first 3 years of life to assess for compositional factors as well as changes to the microbiome (e.g., maturation) associated with developing sensitization to food and aeroallergens.

In older individuals, with established allergic responses and mature immunological memory, a strategy to alter the allergenic immune program associated with TH2 T cell immunobiology is an alternative approach. To this end, the ITN GRADUATE trial, which is currently underway (NCT04502966), combines dupilumab (anti-IL4R) and sublingual grass desensitization to induce durable tolerance. This study builds on the previous ITN GRASS trial comparing desensitization to grass pollen between the sublingual and subcutaneous routes (9). Another cytokine strategy, using tezepelumab (anti-TSLP) to alter TH2 developmental programming, has been studied recently in the ITN CATNIP trial as a potential adjunct to desensitization in established allergic patients (NCT02237196). With recent advances in understanding TH2 diathesis, agents such as anti-OX40 (33), or other strategies targeting allergenic T effector cell subsets, known as TH2A cells (34), may provide additional therapeutic opportunities.

TRANSPLANTATION

Hematopoietic chimerism has been utilized to induce tolerance of kidney allografts in animals and humans, based on the foundational discovery that dizygotic cattle twins exhibit stable red cell chimerism and mutually accept skin grafts (35, 36). A proof of concept for chimerism as a robust mechanism of transplant tolerance was provided by recipients of allogeneic hematopoietic stem cell transplant for leukemia who did not require maintenance immunosuppression after kidney transplant from the same donor 3–11 years later (37). The ITN clinical experience, while successful at achieving transient mixed chimerism (2–3 weeks) in haplotype-matched patients, was only able to induce durable immunosuppression free graft survival in a small number of study participants (10). Similar loss of chimerism and subsequent rejection has been seen by others (38, 39), likely indicating a need for improved peripheral deletion of donor-reactive T cell clones and induction of regulatory mechanisms. Use of a

donor hematopoietic stem cell product featuring CD8+/TCR-“tolerance-promoting facilitating cells” (40), various regulatory T cell strategies (41–44), or a donor mesenchymal stem cell infusion (the ITN TEACH trial, NCT03504241) are alternatives currently being evaluated for potential for inducing transplant tolerance using therapeutic cell transfer. The ITN LITTMUS trials (NCT03577431, NCT03654040) are currently testing whether infusion of alloantigen-specific regulatory T cells, generated from liver transplant recipient cells collected within the first post-transplant year, can facilitate withdrawal of immunosuppressive anti-rejection medications.

In transplantation, the importance of controlling immunological memory cannot be overemphasized. A significant proportion of transplant candidates have accumulated pre-formed antibodies to HLA antigens as a result of prior sensitizing events such as blood transfusion, pregnancy, or a previous transplant. These patients represent a growing challenge for the transplant community as their sensitized immune system makes it more difficult to find compatible potential donors. The achievement of “HLA desensitization” may be therefore considered a waypost on the road to B-cell tolerance. Trials of B cell depletion using rituximab and obinituzumab, although attractive in theory, were found to be unsuccessful in practice, likely reflecting the inability of these agents to eliminate long-lived plasma cells, as well as to sufficiently diminish memory B cells that are destined to develop into anti-HLA antibody-secreting cells (45, 46). Interestingly, the strategy of targeting antibody-secreting plasma cells alone, using agents highly effective against multiple myeloma, has also been found to be insufficient in producing a durable response (47). Elegant studies in allosensitized non-human primate (NHP) models have now shown that ensuing compensation by expanding germinal centers (GC) following plasma cell depletion underlies the rapid repopulation of plasma cells and consequent rebound in HLA antibody (48). Costimulation *via* the CD28 and CD40 pathways plays a critical role in GC interactions between Tfh cells and B cells (49, 50), and costimulation blockade in NHP has been shown to collapse GC and abrogate the humoral rebound seen after plasma cell depletion (51). Based on these observations, a “dual-targeting” strategy of HLA desensitization consisting of 1) plasma cell depletion (using proteasome inhibitors or anti-CD38 monoclonal antibody) and 2) suppression of the upstream humoral response by costimulation blockade is now being tested in two parallel ITN trials (ADAPT, NCT05017545, and ATTAIN, NCT04827979).

AUTOIMMUNITY

Rational strategies for tolerance induction in autoimmune diseases rely on alignment of pathogenic mechanisms and potential therapeutic targets. In some cases, selective targeting of effector T or B cell compartments may be an attractive way to induce remission, by creating a tissue environment less resistant to subsequent homeostatic regulation; in other cases, heterogeneity among patients with similar diseases may require more personalized strategies to optimize the likelihood of matching the appropriate therapy with an individual subject.

Numerous ITN trials, in diseases such as multiple sclerosis, systemic lupus erythematosus (SLE), anti-neutrophil cytoplasmic antibody (ANCA)-associated vasculitis, type 1 diabetes, and psoriasis, have documented robust changes in immune cell compartments in association with T or B cell targeting, but clinical responses are often transient. Therefore, different ITN trials have been designed to evaluate agents for selective immune depletion, immune modulation, and immune regulation, with combinations of those agents when feasible to assess synergistic mechanisms of action. A few examples that illustrate key concepts are discussed below.

Although over a dozen disease-modifying agents have been approved by the FDA for the treatment of multiple sclerosis (MS), some patients with this disease are resistant or refractory to treatment and continue to relapse and accumulate disability. In treatment-resistant relapsing MS, an immune “reset” can be accomplished with immunoablation followed by autologous hematopoietic stem cell transplant (AH SCT). In this procedure, the pathogenic immune repertoire is eliminated with interruption of autoimmune destruction of myelin in the central nervous system. Clinical trials, including the HALT-MS trial conducted by the ITN, demonstrated high efficacy in aggressive treatment-resistant relapsing MS (11, 12, 52–54). There is evidence that AH SCT alters the immune system upon reconstitution *via* thymic reactivation (55), rebalancing of regulatory and effector immune components (56–59), and T cell repertoire diversification (13). In the HALT-MS trial, dominant CD4+ T cell clones were largely replaced by a new repertoire following transplant, while CD8+ T cell clones re-emerged and better clinical outcomes post-transplant were associated with a more diverse CD8+ T cell repertoire (13). To confirm this result, the BEAT-MS multicenter trial is in progress comparing the efficacy and cost-effectiveness of AH SCT to high efficacy disease modifying agents in treatment-resistant relapsing MS (NCT04047628), with the aim of developing high quality evidence supporting consensus recommendations for utilizing AH SCT to treat resistant forms of relapsing MS (60).

Depletion of specific effector cell populations without the global changes induced by AH SCT are also effective, although with inconsistency in the durability of response. In ANCA-associated vasculitis, ANCA do not appear to cause disease by forming immune complexes or directly binding to tissues. Instead, these autoantibodies are suspected to bind neutrophils, resulting in hyperactivation of neutrophils and formation of neutrophil extracellular traps, which in turn results in vascular damage. The Rituximab for ANCA-associated Vasculitis (RAVE) trial treated patients with severe active granulomatosis with polyangiitis and microscopic polyangiitis with rituximab (anti-CD20), with the goal of depleting antibody-secreting B cells and promoting sustained disease control (14). In this study, rituximab therapy was shown to be non-inferior to cyclophosphamide, the previous standard of care for induction of disease remission. Following induction of clinical remission with rituximab therapy, the majority of patients nevertheless relapsed, indicating that immune tolerance was not achieved despite the initial clinical response. A similarly transient benefit of rituximab therapy was

seen in the TrialNet study of type 1 diabetes (61, 62); interestingly, T cell transcripts indicating activation following B cell depletion correlated with reduced preservation of residual insulin secretion, suggesting a need for targeting multiple effector arms of the immune response (63).

Despite an abundance of evidence supporting a critical role for B cells in the pathogenesis of SLE, rituximab-induced B cell depletion has failed in randomized, controlled trials to show clinical efficacy in both non-renal SLE and lupus nephritis (64, 65). One possible explanation for the early failure of rituximab therapy for SLE in clinical trials may be that levels of B-cell activating factor (BAFF) increase following B cell depletion (66). In BAFF-transgenic mice, elevated levels of BAFF rescue autoreactive B-cells and prevent them from becoming anergic (67). To address the hypothesis that inhibition of BAFF following B cell depletion might improve disease control in this setting, the ITN CALIBRATE trial was undertaken to explore in lupus nephritis the efficacy of adding the anti-BAFF agent belimumab to a B cell depletion regimen combining rituximab and cyclophosphamide. Although the percentage of autoreactive naïve B cells were decreased in the group who received belimumab, the addition of a BAFF blocker did not differentiate the clinical outcomes in lupus nephritis (15). A related rituximab and belimumab combination strategy is also being studied in primary membranous nephropathy, where anti-PLA2R autoantibody production is being assessed in the ITN REBOOT trial. In this disease, in contrast to SLE, rituximab therapy has clinical efficacy for some patients. Belimumab will be initiated 4 weeks prior to rituximab therapy based on the observation that circulating memory B cells (CD19+/CD20+/CD27+) increase in numbers after belimumab therapy (68). Thus, initiating belimumab therapy prior to the intervention with rituximab use has been hypothesized to increase the proportion of memory B cells that can be depleted by this CD20-depleting antibody.

Similar to the rationale for transplantation studies noted above, plasma cell targeting is an attractive option for some antibody-mediated autoimmune diseases. Elimination of plasma cells could be beneficial for the treatment of antibody mediated diseases such as antiphospholipid syndrome (APS), a systemic autoimmune disease characterized by thrombotic and obstetric manifestations in individuals with potentially pathogenic antiphospholipid antibodies (69). Standard treatment of APS is lifelong anticoagulation, which is not always effective. Patients with APS often have persistence of antiphospholipid antibodies and are therefore at risk for future thrombosis, shining light on the antibody-secreting cell as a plausible target for effective therapy. A candidate drug for targeting the antibody-secreting plasma cells in APS and other autoimmune conditions is daratumumab, a cytolytic monoclonal antibody developed for the treatment of multiple myeloma. Daratumumab binds to CD38 expressed on plasma cells and is cytolytic for these cells. Small case studies of patients with APS, SLE, and autoimmune hematologic conditions report that treatment with daratumumab reduces autoantibody production in concert with improved clinical manifestations of disease (70–72), providing the rationale for the ITN study of

daratumumab therapy in APS with the goal of eliminating antiphospholipid antibodies.

In parallel with these depletion studies that specifically target the pathologic humoral response, other ITN trials have focused on inducing remission and tolerance in autoimmunity through T cell depletion and modulation. The ITN trials of teplizumab (anti-CD3), alefacept (LFA3-Ig), and anti-thymocyte globulin (ATG) in type 1 diabetes (T1D) are particularly informative. Teplizumab is a monoclonal anti-CD3 antibody, investigated by the ITN for over a decade in several clinical trials. In the ITN AbATE study, treatment with teplizumab delayed the loss of residual insulin secretion in patients with type 1 diabetes in a subset of subjects, with some maintaining insulin secretory function for several years (16). Teplizumab does not deplete T cells; rather, it acts as a partial agonist to induce a series of functional and phenotypic changes. For example, whole blood transcriptomic analysis in AbATE identified *EOMES*, a transcription factor and marker of T-cell exhaustion, as a correlate of changes in insulin secretion (16–18). Moreover, CD8 effector memory T cells were found to have greater expression of *EOMES* mRNA and higher expression levels of other exhaustion markers, namely *KLRG1* and *TIGIT*, by flow cytometry. These *KLRG1+* *TIGIT+* CD8 T-cells showed impaired expression of cell cycle genes when activated, suggesting impaired proliferative capacity, and an effector T-cell exhaustion profile that correlated with improved outcomes. These studies indicate a novel mechanistic basis for tolerance, namely induction of lymphocyte exhaustion by a T cell agonist therapy. The TrialNet TN10 trial subsequently extended these findings to a population of high-risk individuals who had anti-islet cell antibodies, a marker of susceptibility to T1D, but who were not yet hyperglycemic. In this study, teplizumab induced a similar T cell exhaustion profile and resulted in an overall delay in the development of T1D by a median of 24 months (73).

The costimulatory CD28 cell surface molecule is most prominently expressed on CD4 T effector memory (Tem) cells and naïve CD8 (19). CD4 T regulatory cells (Treg) express CD28 at lower levels than any other T-cell subset. Targeting this pathway to affect the antigen-specific effector memory response preferentially over the regulatory T-cell compartment was the basis for the ITN T1DAL trial. T1DAL was a clinical trial carried out in children and adolescents with new-onset type 1 diabetes and investigated the clinical efficacy of alefacept, an LFA-3 fusion protein that blocks the costimulatory LFA-3/CD28 interaction (19–21). In this study, 30% of treated subjects retained or improved their insulin secretory function over 2 years, and another 40% showed only modest loss of islet function. Similar to treatment with teplizumab, alefacept treatment was associated with increases in the prevalence of CD8 effector memory cells expressing both *KLRG1* and *TIGIT*; as seen in AbATE, these exhausted CD8 cells correlated with beneficial clinical response. Notably, however, in T1DAL they appeared in the blood approximately 9 months after therapy, following an earlier induction of PD1 on a CD4 Tem population. Both teplizumab and alefacept treated participants demonstrated these changes in effector populations concurrent with relative preservation of regulatory T cells, providing a proof-of-concept for the sequential combination of an induction therapy followed

by consolidation of the regulatory response. This conclusion was reinforced by the findings in the ITN START trial (22) which utilized anti-thymocyte globulin in a similar study of patients with early T1D. In START, no therapeutic benefit was observed, and analysis of lymphocyte subsets indicated depletion of multiple cell lineages, notably including both regulatory and effector T cell populations. Retention or boosting of regulatory responses is therefore a major objective of current ITN therapeutic strategies, forming one of the key components in clinical trial designs.

Failure to retain regulatory T cell function may explain the disappointing outcomes in several trials using CD28 costimulatory blockade as a therapeutic tolerance strategy. Abatacept (CTLA4-Ig) is a fusion protein consisting of the extracellular domain of the CTLA4 ligand for CD80/86 coupled to a modified Fc portion of human immunoglobulin G (IgG). It acts by preventing CD80/CD86 on APCs from binding to CD28 on T cells, thereby inhibiting T cell activation and function, and serving as an effective treatment option in adult rheumatoid arthritis and juvenile idiopathic arthritis. ITN trials with abatacept in multiple sclerosis, lupus nephritis, and psoriasis, however, did not demonstrate clinical benefit for induction or maintenance of tolerance (23, 24, 26). Analysis of peripheral blood cells from the participants in the multiple sclerosis trial demonstrated that the relative proportions of activated CD4+ T follicular helper cells and regulatory T cells were both decreased in participants receiving abatacept compared with those receiving placebo (25). Similar changes following abatacept treatment, including loss of regulatory T cells, have been observed by others in a variety of other diseases (74–76). While transient immunomodulatory effects in effector cells from CD28 blockade occur, these require continued drug administration, and therefore the undesirable inhibition of regulatory T cells that also utilize CD28-dependent pathways appears to preclude effective use of abatacept as a tolerance therapy where drug discontinuation is desired.

DISCUSSION

Successful treatment interventions in transplantation, allergy, and autoimmune disease rarely allow for discontinuation of therapy and result in sustained and selective immune tolerance. However, they do occur, and in the context of clinical trials, they provide an opportunity to identify specific therapeutic targets, or combinations of targets, capable of restoring homeostatic immunity. Notably, minimization or extended drug holidays from immunosuppressive therapy also provide clinical benefit, even without achieving the ideal goal of permanent immune tolerance. Ongoing maintenance intervention may be required, for example a small dose of oral peanut (or other allergen) to maintain a non-allergic state in an individual who had previously undergone successful desensitization to that allergen.

Looking across multiple clinical trials and different clinical disciplines in ITN studies, shared patterns emerge that emphasize the need for a combination of *interrupting* effector mechanisms, *restraining* innate activation, and *boosting*

regulation. These objectives can be met through targeting of different cells and pathways in different diseases, whether B cell, plasma cell, T effector, T regulatory, cytokine or other non-lymphocytic effector mechanisms. Early intervention, and mechanism-based stratification of individuals for optimal targeting of particular immune pathways, are likely to improve success rates. Combining antigen exposure in allergic disease and transplantation with selective immunomodulation, and using initial induction strategies in autoimmunity followed by regulatory enhancement, are promising conduits for improving clinical outcomes. In the examples summarized in this article, we have learned that early intervention in the form of antigen introduction can be successful in preventing and treating peanut allergy. In autoimmune disease, immunomodulatory agents targeting T cells and inducing exhaustion pathways can delay clinical disease onset and improve the effector/regulatory T cell balance in T1D, while targeting B cells improves outcomes in ANCA-associated vasculitis. For other autoimmune diseases resistant to B cell depletion or costimulatory blockade, targeting alternative costimulatory pathways and antibody-producing plasma cells in tandem, for example, and combination strategies interrupting key hubs in the adaptive and innate immune response may be required to restore immune tolerance and tissue homeostasis. In solid organ transplantation, success has been partially achieved with an immune reset *via* hematopoietic chimerism, but other strategies including cell-based therapies and molecularly targeted agents are under investigation. Eliminating unwanted immune responses in allergy, autoimmunity, and transplantation and restoring a healthy balance of regulatory and effector immune elements is a formidable

goal. Nevertheless, improved understanding of the complexities of the immune system provide opportunities that continue to provide a foundation for future tolerance-inducing strategies and success.

AUTHOR CONTRIBUTIONS

All authors contributed to the article and approved the submitted version.

FUNDING

The work of the authors was supported by the National Institute of Allergy and Infectious Diseases of the National Institutes of Health under Award Number UM1AI109565.

ACKNOWLEDGMENTS

We gratefully acknowledge and thank our many colleagues who have pioneered and sustained ITN projects over its 20-year history, completing over 60 clinical trials and multiple mechanistic studies. We regret being unable to discuss all trials in this brief article, but encourage readers to access www.immunetolerance.org and www.ITNTrialShare.org for more complete information. We especially thank all the patients and their families who participated in ITN trials, as they are the essential generators of knowledge that will speed the quest for immune tolerance.

REFERENCES

- Smilek DE, Ehlers MR, Nepom GT. Restoring the Balance: Immunotherapeutic Combinations for Autoimmune Disease. *Dis Model Mech* (2014) 7(5):503–13. doi: 10.1242/dmm.015099
- Du Toit G, Roberts G, Sayre PH, Bahnson HT, Radulovic S, Santos AF, et al. Randomized Trial of Peanut Consumption in Infants at Risk for Peanut Allergy. *N Engl J Med* (2015) 372(9):803–13. doi: 10.1056/NEJMoa1414850
- Du Toit G, Sayre PH, Roberts G, Sever ML, Lawson K, Bahnson HT, et al. Effect of Avoidance on Peanut Allergy After Early Peanut Consumption. *N Engl J Med* (2016) 374(15):1435–43. doi: 10.1056/NEJMoa1514209
- Togias A, Cooper SF, Acebal ML, Assa'ad A, Baker JR Jr., Beck LA, et al. Addendum Guidelines for the Prevention of Peanut Allergy in the United States: Report of the National Institute of Allergy and Infectious Diseases-Sponsored Expert Panel. *J Allergy Clin Immunol* (2017) 139(1):29–44. doi: 10.1016/j.jaci.2016.10.010
- du Toit G, Sayre PH, Roberts G, Lawson K, Sever ML, Bahnson HT, et al. Allergen Specificity of Early Peanut Consumption and Effect on Development of Allergic Disease in the Learning Early About Peanut Allergy Study Cohort. *J Allergy Clin Immunol* (2018) 141(4):1343–53. doi: 10.1016/j.jaci.2017.09.034
- Winters A, Bahnson HT, Ruczinski I, Boorgula MP, Malley C, Keramati AR, et al. The MALT1 Locus and Peanut Avoidance in the Risk for Peanut Allergy. *J Allergy Clin Immunol* (2019) 143(6):2326–9. doi: 10.1016/j.jaci.2019.02.016
- Suarez-Farinas M, Suprun M, Bahnson HT, Raghunathan R, Getts R, duToit G, et al. Evolution of Epitope-Specific IgE and IgG4 Antibodies in Children Enrolled in the LEAP Trial. *J Allergy Clin Immunol* (2021) 148(3):835–42. doi: 10.1016/j.jaci.2021.01.030
- Jones S, K E, Nadeau K, Nowak-Wegrzyn A, Wood R, Sampson H, et al. Tolerance Development Following Oral Immunotherapy (OIT) in 1-3 Year Old Children With Peanut Allergy: Findings From the Immune Tolerance Network (ITN) IMPACT TRIAL. *J Allergy Clin Immunol* (2020) 145(2):AB338. doi: 10.1016/j.jaci.2019.12.067
- Scadding GW, Calderon MA, Shamji MH, Eifan AO, Penagos M, Dumitru F, et al. Effect of 2 Years of Treatment With Sublingual Grass Pollen Immunotherapy on Nasal Response to Allergen Challenge at 3 Years Among Patients With Moderate to Severe Seasonal Allergic Rhinitis: The GRASS Randomized Clinical Trial. *JAMA* (2017) 317(6):615–25. doi: 10.1001/jama.2016.21040
- Kawai T, Sachs DH, Sprangers B, Spitzer TR, Saidman SL, Zorn E, et al. Long-Term Results in Recipients of Combined HLA-Mismatched Kidney and Bone Marrow Transplantation Without Maintenance Immunosuppression. *Am J Transplant* (2014) 14(7):1599–611. doi: 10.1111/ajt.12731
- Nash RA, Bowen JD, McSweeney PA, Pavletic SZ, Maravilla KR, Park MS, et al. High-Dose Immunosuppressive Therapy and Autologous Peripheral Blood Stem Cell Transplantation for Severe Multiple Sclerosis. *Blood* (2003) 102(7):2364–72. doi: 10.1182/blood-2002-12-3908
- Nash RA, Hutton GJ, Racke MK, Papat U, Devine SM, Steinmiller KC, et al. High-Dose Immunosuppressive Therapy and Autologous HCT for Relapsing-Remitting MS. *Neurology* (2017) 88(9):842–52. doi: 10.1212/WNL.0000000000003660
- Muraro PA, Robins H, Malhotra S, Howell M, Phippard D, Desmarais C, et al. T Cell Repertoire Following Autologous Stem Cell Transplantation for Multiple Sclerosis. *J Clin Invest* (2014) 124(3):1168–72. doi: 10.1172/JCI71691
- Stone JH, Merkel PA, Spiera R, Seo P, Langford CA, Hoffman GS, et al. Rituximab Versus Cyclophosphamide for ANCA-Associated Vasculitis. *N Engl J Med* (2010) 363(3):221–32. doi: 10.1056/NEJMoa0909905
- Atisha-Fregoso Y, Malkiel S, Harris KM, Byron M, Ding L, Kanaparthi S, et al. Phase II Randomized Trial of Rituximab Plus Cyclophosphamide Followed by Belimumab for the Treatment of Lupus Nephritis. *Arthritis Rheumatol* (2021) 73(1):121–31. doi: 10.1002/art.41466
- Herold KC, Gitelman SE, Ehlers MR, Gottlieb PA, Greenbaum CJ, Hagopian W, et al. Teplizumab (Anti-CD3 mAb) Treatment Preserves C-Peptide

- Responses in Patients With New-Onset Type 1 Diabetes in a Randomized Controlled Trial: Metabolic and Immunologic Features at Baseline Identify a Subgroup of Responders. *Diabetes* (2013) 62(11):3766–74. doi: 10.2337/db13-0345
17. Long SA, Thorpe J, DeBerg HA, Gersuk V, Eddy J, Harris KM, et al. Partial Exhaustion of CD8 T Cells and Clinical Response to Teplizumab in New-Onset Type 1 Diabetes. *Sci Immunol* (2016) 1(5):eaai7793. doi: 10.1126/sciimmunol.aai7793
 18. Long SA, Thorpe J, Herold KC, Ehlers M, Sanda S, Lim N, et al. Remodeling T Cell Compartments During Anti-CD3 Immunotherapy of Type 1 Diabetes. *Cell Immunol* (2017) 319:3–9. doi: 10.1016/j.cellimm.2017.07.007
 19. Rigby MR, DiMeglio LA, Rendell MS, Felner EI, Dostou JM, Gitelman SE, et al. Targeting of Memory T Cells With Alefacept in New-Onset Type 1 Diabetes (T1DAL Study): 12 Month Results of a Randomised, Double-Blind, Placebo-Controlled Phase 2 Trial. *Lancet Diabetes Endocrinol* (2013) 1(4):284–94. doi: 10.1016/S2213-8587(13)70111-6
 20. Diggins KE, Serti E, Muir V, Rosasco M, Lu T, Balmas E, et al. Exhausted-Like CD8+ T Cell Phenotypes Linked to C-Peptide Preservation in Alefacept-Treated T1D Subjects. *JCI Insight* (2021) 6(3):e142680. doi: 10.1172/jci.insight.142680
 21. Rigby MR, Harris KM, Pinckney A, DiMeglio LA, Rendell MS, Felner EI, et al. Alefacept Provides Sustained Clinical and Immunological Effects in New-Onset Type 1 Diabetes Patients. *J Clin Invest* (2015) 125(8):3285–96. doi: 10.1172/JCI81722
 22. Gitelman SE, Gottlieb PA, Felner EI, Willi SM, Fisher LK, Moran A, et al. Antithymocyte Globulin Therapy for Patients With Recent-Onset Type 1 Diabetes: 2 Year Results of a Randomised Trial. *Diabetologia* (2016) 59(6):1153–61. doi: 10.1007/s00125-016-3917-4
 23. The ACCESS Trial Group. Treatment of Lupus Nephritis With Abatacept. The Abatacept and Cyclophosphamide Combination Efficacy and Safety Study. *Arthritis Rheumatol* (2014) 66(11):3096–104. doi: 10.1002/art.38790
 24. Khoury SJ, Rochon J, Ding L, Byron M, Ryker K, Tosta P, et al. ACCLAIM: A Randomized Trial of Abatacept (CTLA4-Ig) for Relapsing-Remitting Multiple Sclerosis. *Mult Scler* (2017) 23(5):686–95. doi: 10.1177/1352458516662727
 25. Glatigny S, Hollbacher B, Motley SJ, Tan C, Hundhausen C, Buckner JH, et al. Abatacept Targets T Follicular Helper and Regulatory T Cells, Disrupting Molecular Pathways That Regulate Their Proliferation and Maintenance. *J Immunol* (2019) 202(5):1373–82. doi: 10.4049/jimmunol.1801425
 26. Smilek RB D, Barber K, Boh E, Feldman S, Krueger G, Helfrich Y, et al. J. Krueger. Costimulatory Blockade With Abatacept Did Not Prevent Psoriasis Relapse After Discontinuation of Ustekinumab: Results of a Clinical Trial. *Inflammatory Skin Dis Summit. Exp Dermatol* (2018) 27(Suppl 2):38–58, AB60. doi: 10.1111/exd.13795
 27. Perkin MR, Lack G. Introducing Allergenic Foods in Infants. *N Engl J Med* (2016) 375(8):e16.
 28. Thome M. CARMA1, BCL-10 and MALT1 in Lymphocyte Development and Activation. *Nat Rev Immunol* (2004) 4(5):348–59. doi: 10.1038/nri1352
 29. Fujimura KE, Lynch SV. Microbiota in Allergy and Asthma and the Emerging Relationship With the Gut Microbiome. *Cell Host Microbe* (2015) 17(5):592–602. doi: 10.1016/j.chom.2015.04.007
 30. Tsilochristou O, du Toit G, Sayre PH, Roberts G, Lawson K, Sever ML, et al. Association of Staphylococcus Aureus Colonization With Food Allergy Occurs Independently of Eczema Severity. *J Allergy Clin Immunol* (2019) 144(2):494–503. doi: 10.1016/j.jaci.2019.04.025
 31. Stokholm J, Thorsen J, Blaser MJ, Rasmussen MA, Hjelmso M, Shah S, et al. Delivery Mode and Gut Microbial Changes Correlate With an Increased Risk of Childhood Asthma. *Sci Transl Med* (2020) 12(569):eaax9929. doi: 10.1126/scitranslmed.aax9929
 32. Levan SR, Stamnes KA, Lin DL, Panzer AR, Fukui E, McCauley K, et al. Elevated Faecal 12,13-diHOME Concentration in Neonates at High Risk for Asthma Is Produced by Gut Bacteria and Impedes Immune Tolerance. *Nat Microbiol* (2019) 4(11):1851–61. doi: 10.1038/s41564-019-0498-2
 33. Burrows KE, Dumont C, Thompson CL, Catley MC, Dixon KL, Marshall D. OX40 Blockade Inhibits House Dust Mite Driven Allergic Lung Inflammation in Mice and *In Vitro* Allergic Responses in Humans. *Eur J Immunol* (2015) 45(4):1116–28. doi: 10.1002/eji.201445163
 34. Wambre E, Bajzik V, DeLong JH, O'Brien K, Nguyen QA, Speake C, et al. A Phenotypically and Functionally Distinct Human TH2 Cell Subpopulation Is Associated With Allergic Disorders. *Sci Transl Med* (2017) 9(401):eam9171. doi: 10.1126/scitranslmed.aam9171
 35. Anderson REB D, Lampkin GH, Medawar PB. The Use of Skin Grafting to Distinguish Between Monozygotic and Dizygotic Twins in Cattle. *Heredity (Edinb)* (1951) 5:379–97. doi: 10.1038/hdy.1951.38
 36. Owen RD. Immunogenetic Consequences of Vascular Anastomoses Between Bovine Twins. *Science* (1945) 102(2651):400–1. doi: 10.1126/science.102.2651.400
 37. Butcher JA, Hariharan S, Adams MB, Johnson CP, Roza AM, Cohen EP. Renal Transplantation for End-Stage Renal Disease Following Bone Marrow Transplantation: A Report of Six Cases, With and Without Immunosuppression. *Clin Transplant* (1999) 13(4):330–5. doi: 10.1034/j.1399-0012.1999.130409.x
 38. Scandling JD, Busque S, Lowsky R, Shizuru J, Shori A, Engleman E, et al. Macrochimerism and Clinical Transplant Tolerance. *Hum Immunol* (2018) 79(5):266–71. doi: 10.1016/j.humimm.2018.01.002
 39. Scandling JD, Busque S, Shizuru JA, Lowsky R, Hoppe R, Dejbakhsh-Jones S, et al. Chimerism, Graft Survival, and Withdrawal of Immunosuppressive Drugs in HLA Matched and Mismatched Patients After Living Donor Kidney and Hematopoietic Cell Transplantation. *Am J Transplant* (2015) 15(3):695–704. doi: 10.1111/ajt.13091
 40. Leventhal JR, Ildstad ST. Tolerance Induction in HLA Disparate Living Donor Kidney Transplantation by Facilitating Cell-Enriched Donor Stem Cell Infusion: The Importance of Durable Chimerism. *Hum Immunol* (2018) 79(5):272–6. doi: 10.1016/j.humimm.2018.01.007
 41. Harden PN, Game DS, Sawitzki B, van der Net JB, Hester J, Bushell A, et al. Feasibility, Long-Term Safety, and Immune Monitoring of Regulatory T Cell Therapy in Living Donor Kidney Transplant Recipients. *Am J Transplant* (2021) 21(4):1603–11. doi: 10.1111/ajt.16395
 42. Lee K, Nguyen V, Lee KM, Kang SM, Tang Q. Attenuation of Donor-Reactive T Cells Allows Effective Control of Allograft Rejection Using Regulatory T Cell Therapy. *Am J Transplant* (2014) 14(1):27–38. doi: 10.1111/ajt.12509
 43. Sawitzki B, Harden PN, Reinke P, Moreau A, Hutchinson JA, Game DS, et al. Regulatory Cell Therapy in Kidney Transplantation (The ONE Study): A Harmonised Design and Analysis of Seven Non-Randomised, Single-Arm, Phase 1/2A Trials. *Lancet* (2020) 395(10237):1627–39. doi: 10.1016/S0140-6736(20)30167-7
 44. Todo S, Yamashita K, Goto R, Zaitsumi M, Nagatsu A, Oura T, et al. A Pilot Study of Operational Tolerance With a Regulatory T-Cell-Based Cell Therapy in Living Donor Liver Transplantation. *Hepatology* (2016) 64(2):632–43. doi: 10.1002/hep.28459
 45. Jackson AM, Kraus ES, Orandi BJ, Segev DL, Montgomery RA, Zachary AA. A Closer Look at Rituximab Induction on HLA Antibody Rebound Following HLA-Incompatible Kidney Transplantation. *Kidney Int* (2015) 87(2):409–16. doi: 10.1038/ki.2014.261
 46. Redfield RR, Jordan SC, Busque S, Vincenti F, Woodle ES, Desai N, et al. Safety, Pharmacokinetics, and Pharmacodynamic Activity of Obinutuzumab, A Type 2 Anti-CD20 Monoclonal Antibody for the Desensitization of Candidates for Renal Transplant. *Am J Transplant* (2019) 19(11):3035–45. doi: 10.1111/ajt.15514
 47. Moreno Gonzales MA, Gandhi MJ, Schinstock CA, Moore NA, Smith BH, Braaten NY, et al. 32 Doses of Bortezomib for Desensitization Is Not Well Tolerated and Is Associated With Only Modest Reductions in Anti-HLA Antibody. *Transplantation* (2017) 101(6):1222–7. doi: 10.1097/TP.0000000000001330
 48. Kwun J, Burghuber C, Manook M, Iwakoshi N, Gibby A, Hong JJ, et al. Humoral Compensation After Bortezomib Treatment of Allosensitized Recipients. *J Am Soc Nephrol* (2017) 28(7):1991–6. doi: 10.1681/ASN.2016070727
 49. Klaus SJ, Pinchuk LM, Ochs HD, Law CL, Fanslow WC, Armitage RJ, et al. Costimulation Through CD28 Enhances T Cell-Dependent B Cell Activation via CD40-CD40L Interaction. *J Immunol* (1994) 152(12):5643–52.
 50. Nonoyama S, Hollenbaugh D, Aruffo A, Ledbetter JA, Ochs HD. B Cell Activation via CD40 Is Required for Specific Antibody Production by Antigen-Stimulated Human B Cells. *J Exp Med* (1993) 178(3):1097–102. doi: 10.1084/jem.178.3.1097
 51. Burghuber CK, Manook M, Ezekian B, Gibby AC, Leopardi FV, Song M, et al. Dual Targeting: Combining Costimulation Blockade and Bortezomib to Permit Kidney Transplantation in Sensitized Recipients. *Am J Transplant* (2019) 19(3):724–36. doi: 10.1111/ajt.15067

52. Atkins HL, Bowman M, Allan D, Anstee G, Arnold DL, Bar-Or A, et al. Immunoablation and Autologous Haematopoietic Stem-Cell Transplantation for Aggressive Multiple Sclerosis: A Multicentre Single-Group Phase 2 Trial. *Lancet* (2016) 388(10044):576–85. doi: 10.1016/S0140-6736(16)30169-6
53. Burt RK, Balabanov R, Burman J, Sharrack B, Snowden JA, Oliveira MC, et al. Effect of Nonmyeloablative Hematopoietic Stem Cell Transplantation vs Continued Disease-Modifying Therapy on Disease Progression in Patients With Relapsing-Remitting Multiple Sclerosis: A Randomized Clinical Trial. *JAMA* (2019) 321(2):165–74. doi: 10.1001/jama.2018.18743
54. Moore JJ, Massey JC, Ford CD, Khoo ML, Zaunders JJ, Hendrawan K, et al. Prospective Phase II Clinical Trial of Autologous Haematopoietic Stem Cell Transplant for Treatment Refractory Multiple Sclerosis. *J Neurol Neurosurg Psychiatry* (2019) 90(5):514–21. doi: 10.1136/jnnp-2018-319446
55. Muraro PA, Douek DC, Packer A, Chung K, Guenaga FJ, Cassiani-Ingoni R, et al. Thymic Output Generates a New and Diverse TCR Repertoire After Autologous Stem Cell Transplantation in Multiple Sclerosis Patients. *J Exp Med* (2005) 201(5):805–16. doi: 10.1084/jem.20041679
56. Abrahamsson SV, Angelini DF, Dubinsky AN, Morel E, Oh U, Jones JL, et al. Non-Myeloablative Autologous Haematopoietic Stem Cell Transplantation Expands Regulatory Cells and Depletes IL-17 Producing Mucosal-Associated Invariant T Cells in Multiple Sclerosis. *Brain* (2013) 136(Pt 9):2888–903. doi: 10.1093/brain/awt182
57. Arruda LCM, de Azevedo JTC, de Oliveira GLV, Scortegagna GT, Rodrigues ES, Palma PVB, et al. Immunological Correlates of Favorable Long-Term Clinical Outcome in Multiple Sclerosis Patients After Autologous Hematopoietic Stem Cell Transplantation. *Clin Immunol* (2016) 169:47–57. doi: 10.1016/j.clim.2016.06.005
58. Burman J, Fransson M, Totterman TH, Fagius J, Mangsbo SM, Loskog AS. T-Cell Responses After Haematopoietic Stem Cell Transplantation for Aggressive Relapsing-Remitting Multiple Sclerosis. *Immunology* (2013) 140(2):211–9. doi: 10.1111/imm.12129
59. Darlington PJ, Touil T, Doucet JS, Gaucher D, Zeidan J, Gauchat D, et al. Diminished Th17 (Not Th1) Responses Underlie Multiple Sclerosis Disease Abrogation After Hematopoietic Stem Cell Transplantation. *Ann Neurol* (2013) 73(3):341–54. doi: 10.1002/ana.23784
60. Cohen JA, Baldassari LE, Atkins HL, Bowen JD, Bredeson C, Carpenter PA, et al. Autologous Hematopoietic Cell Transplantation for Treatment-Refractory Relapsing Multiple Sclerosis: Position Statement From the American Society for Blood and Marrow Transplantation. *Biol Blood Marrow Transplant* (2019) 25(5):845–54. doi: 10.1016/j.bbmt.2019.02.014
61. Pescovitz MD, Greenbaum CJ, Bundy B, Becker DJ, Gitelman SE, Goland R, et al. B-Lymphocyte Depletion With Rituximab and Beta-Cell Function: Two-Year Results. *Diabetes Care* (2014) 37(2):453–9. doi: 10.2337/dc13-0626
62. Pescovitz MD, Greenbaum CJ, Krause-Steinrauf H, Becker DJ, Gitelman SE, Goland R, et al. Rituximab, B-Lymphocyte Depletion, and Preservation of Beta-Cell Function. *N Engl J Med* (2009) 361(22):2143–52. doi: 10.1056/NEJMoa0904452
63. Linsley PS, Greenbaum CJ, Rosasco M, Presnell S, Herold KC, Dufort MJ. Elevated T Cell Levels in Peripheral Blood Predict Poor Clinical Response Following Rituximab Treatment in New-Onset Type 1 Diabetes. *Genes Immun* (2019) 20(4):293–307. doi: 10.1038/s41435-018-0032-1
64. Merrill JT, Neuwelt CM, Wallace DJ, Shanahan JC, Latinis KM, Oates JC, et al. Efficacy and Safety of Rituximab in Moderately-to-Severely Active Systemic Lupus Erythematosus: The Randomized, Double-Blind, Phase II/III Systemic Lupus Erythematosus Evaluation of Rituximab Trial. *Arthritis Rheum* (2010) 62(1):222–33. doi: 10.1002/art.27233
65. Rovin BH, Furie R, Latinis K, Looney RJ, Fervenza FC, Sanchez-Guerrero J, et al. Efficacy and Safety of Rituximab in Patients With Active Proliferative Lupus Nephritis: The Lupus Nephritis Assessment With Rituximab Study. *Arthritis Rheum* (2012) 64(4):1215–26. doi: 10.1002/art.34359
66. Ehrenstein MR, Wing C. The Baffling Effects of Rituximab in Lupus: Danger Ahead? *Nat Rev Rheumatol* (2016) 12(6):367–72. doi: 10.1038/nrrheum.2016.18
67. Kawabata D, Venkatesh J, Ramanujam M, Davidson A, Grimaldi CM, Diamond B. Enhanced Selection of High Affinity DNA-Reactive B Cells Following Cyclophosphamide Treatment in Mice. *PLoS One* (2010) 5(1):e8418. doi: 10.1371/journal.pone.0008418
68. Stohl W, Hiepe F, Latinis KM, Thomas M, Scheinberg MA, Clarke A, et al. Belimumab Reduces Autoantibodies, Normalizes Low Complement Levels, and Reduces Select B Cell Populations in Patients With Systemic Lupus Erythematosus. *Arthritis Rheum* (2012) 64(7):2328–37. doi: 10.1002/art.34400
69. Garcia D, Erkan D. Diagnosis and Management of the Antiphospholipid Syndrome. *N Engl J Med* (2018) 378:2010–21. doi: 10.1056/NEJMra1705454
70. Ostendorf L, Burns M, Durek P, Heinz GA, Heinrich F, Garantziotis P, et al. Targeting CD38 With Daratumumab in Refractory Systemic Lupus Erythematosus. *N Engl J Med* (2020) 383(12):1149–55. doi: 10.1056/NEJMoa2023325
71. Pleguezuelo DE, Diaz-Simon R, Cabrera-Marante O, Lalueza A, Paz-Artal E, Lumbreras C, et al. Case Report: Resetting the Humoral Immune Response by Targeting Plasma Cells With Daratumumab in Anti-Phospholipid Syndrome. *Front Immunol* (2021) 12:667515. doi: 10.3389/fimmu.2021.667515
72. Zaninoni A, Giannotta JA, Galli A, Artuso R, Bianchi P, Malcovati L, et al. The Immunomodulatory Effect and Clinical Efficacy of Daratumumab in a Patient With Cold Agglutinin Disease. *Front Immunol* (2021) 12:649441. doi: 10.3389/fimmu.2021.649441
73. Herold KC, Bundy BN, Long SA, Bluestone JA, DiMeglio LA, Dufort MJ, et al. An Anti-CD3 Antibody, Teplizumab, in Relatives at Risk for Type 1 Diabetes. *N Engl J Med* (2019) 381(7):603–13. doi: 10.1056/NEJMoa1902226
74. Maggi L, Cimaz R, Capone M, Santarlasci V, Rossi MC, Mazzoni A, et al. Immunosuppressive Activity of Abatacept on Circulating T Helper Lymphocytes From Juvenile Idiopathic Arthritis Patients. *Int Arch Allergy Immunol* (2016) 171(1):45–53. doi: 10.1159/000450948
75. Orban T, Beam CA, Xu P, Moore K, Jiang Q, Deng J, et al. Reduction in CD4 Central Memory T-Cell Subset in Costimulation Modulator Abatacept-Treated Patients With Recent-Onset Type 1 Diabetes Is Associated With Slower C-Peptide Decline. *Diabetes* (2014) 63(10):3449–57. doi: 10.2337/db14-0047
76. Verstappen GM, Meiners PM, Corneth OBJ, Visser A, Arends S, Abdulahad WH, et al. Attenuation of Follicular Helper T Cell-Dependent B Cell Hyperactivity by Abatacept Treatment in Primary Sjogren's Syndrome. *Arthritis Rheumatol* (2017) 69(9):1850–61. doi: 10.1002/art.40165

Author Disclaimer: The content is solely the responsibility of the authors and does not necessarily represent the official views of the National Institutes of Health.

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's Note: All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Copyright © 2021 Huffaker, Sanda, Chandran, Chung, St. Clair, Nepom and Smilek. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.