



Dermatomyositis With Anti-MDA5 Antibodies: Bioclinical Features, Pathogenesis and Emerging Therapies

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Anti-MDA5 dermatomyositis is a rare systemic autoimmune disease, historically described in Japanese patients with clinically amyopathic dermatomyositis and life-threatening rapidly progressive interstitial lung disease. Subsequently, the complete clinical spectrum of the disease was enriched by skin, articular and vascular manifestations. Depending on the predominance of these symptoms, three distinct clinical phenotypes with different prognosis are now defined. To date, the only known molecular component shared by the three entities are specific antibodies targeting MDA5, a cytosolic protein essential for antiviral host immune responses. Several biological tools have emerged to detect these antibodies, with drawbacks and limitations for each of them. However, the identification of this highly specific serological marker of the disease raises the question of its role in the pathogenesis. Although current knowledge on the pathogenic mechanisms that take place in the disease are still in their infancy, several lines of evidence support a central role of interferon-mediated vasculopathy in the development of skin and lung lesions, as well as a possible pathogenic involvement of anti-MDA5 antibodies. Here, we review the clinical and biological evidences in favor of these hypothesis, and we discuss the contribution of emerging therapies that shed some light on the pathogenesis of the disease.

Keywords: myositis, dermatomyositis, idiopathic inflammatory myopathies, MDA5, COVID-19, SARS-CoV-2, autoantibodies, autoantibody

INTRODUCTION

The idiopathic inflammatory myopathies (IIM) are a heterogeneous group of rare connective tissue diseases, characterized by inflammation of several organs and tissues other than the muscles, such as the skin and the lungs. IIM include necrotizing immune-mediated myositis, inclusion body myositis, antisynthetase syndrome and dermatomyositis (DM) (1). These four subgroups are

very heterogeneous in their clinical, prognostic and pathological features, which renders the diagnosis and the treatment challenging. However, the discovery and the inclusion of myositis specific autoantibodies (MSA) in the diagnostic algorithm of myositis allowed a better definition of subgroups of patients in terms of clinical phenotypes, prognosis and response to treatment. One of these MSA, the anti-melanoma differentiation-associated gene 5 (MDA5) antibodies (Abs), was originally identified in a specific subset of DM, named clinically amyopathic DM (CADM), associated with an increased risk for rapidly progressive interstitial lung disease (RP-ILD). The clinical presentation of anti-MDA5 DM differs substantially from the other forms of DM, with three distinct clinical phenotypes, according to the predominance of pulmonary, skin-articular or vascular symptoms (2). The pathogenesis of these three forms of anti-MDA5 DM is largely unknown, and to date, the only common molecular characteristic of these entities is the presence in the blood of the patients of Abs targeting the antigen MDA5, a highly specific biomarker of the disease, sometimes difficult to detect with usual techniques. MDA5 is a cytosolic protein, essential for antiviral host immune responses, which functions as a virus RNA sensor and induces, once activated, the production of type I interferons (IFN-I) and pro-inflammatory cytokines by the cell. The pathogenic role of anti-MDA5 Abs is currently unknown, but its involvement in the disease by targeting a critical actor of the immune system could be congruent with the concept of autoimmunity induced by infectious agents.

In this review, we outline the clinical phenotypes of the DM with anti-MDA5 Abs, the distribution and the functions of MDA5, as well as the biological tools available for the detection of anti-MDA5 Abs and their limitations. We focus on recent biological data that provide insight into the pathogenesis of the disease, and we propose a pathophysiological model centered on vascular dysfunction and dysregulated immune system. In this proposed model, we will discuss whether the Abs specific of the disease could be critical players in disease pathogenesis, and not just biomarkers.

CLINICAL SPECTRUM IN ADULTS OF THE DERMATOMYOSITIS WITH ANTI-MDA5 ANTIBODIES

Anti-MDA5 DM is a rare disease representing less than 2% of IIM in Europe (3). Among the subgroup of DM, the prevalence of anti-MDA5 DM ranges from 7 to 60%, with higher prevalence in Asian (11–60%) than in Caucasian (7–16%) (Table 1) (1, 4–19). Similar to other autoimmune diseases, anti-MDA5 DM occurs mainly in women, with a female/male ratio that ranges from 0.6 to 7.3 (F/M >1 in 14 out of 16 studies) (Table 1) (4–19).

Systemic autoimmune DM are characterized by skin manifestations accompanying or preceding muscle weakness, and, to various extents, lung lesions. The DM associated with anti-MDA5 Abs shares clinical features with DM but also has

disease-specific traits. Indeed, the patients with anti-MDA5 DM may have the hallmark cutaneous manifestations of DM, but the disease is also associated with specific skin manifestations. Moreover, the muscle disease is minimal or absent, and pulmonary interstitial lesions may be rapidly progressive which is not the case in other forms of DM.

Muscular Manifestations of Anti-MDA5 Dermatomyositis

Anti-MDA5 DM was first described by Sato et al. in 2005, in a Japanese cohort with CADM and RP-ILD (4). As defined by Sontheimer et al. (20), CADM patients display the hallmark cutaneous manifestations of DM for at least 6 months, without muscle weakness. The hypomyopathic forms of CADM are associated with elevated muscle enzymes and/or abnormalities in EMG or muscle biopsy, whereas amyopathic DM patients have the cutaneous manifestations of DM for at least 6 months, without clinical or biological signs of muscle damage. However, CADM is not sufficient to define anti-MDA5 DM, as not all CADM patients have anti-MDA5 Abs. The proportion of anti-MDA5 Abs positive patients among CADM ranges from 23% to 100%, depending on the cohorts (Table 1) (4–7, 9–14, 17, 19). Conversely, the prevalence of CADM among anti-MDA5 Abs positive patients is also very inconsistent (17 to 100%, Table 1) (4–19).

When present, muscle damage mainly affects proximal muscles (2, 12). Several histological features of muscle biopsies are shared by different forms of DM, and include perifascicular fiber atrophy, perivascular inflammation, infiltrates of T and B cells, diffuse class I major histocompatibility complex (MHC-I) expression and deposition of complement attack complex (21, 22). In contrast, muscle biopsies of patients with anti-MDA5 DM are often normal or highlight only rare anomalies. Inflammation is absent or scarce and cellular infiltrates, mostly macrophages, cluster focally in the perimysium. MHC-I expression is focal or absent. Complement membrane attack complex deposition is rarely observed. Furthermore, IFN signature in skeletal muscles of patients with anti-MDA5 DM is up-regulated, compared with healthy subjects but lower than patients with anti-MDA5 negative DM (21, 22). These observations are consistent with the mild muscle phenotype frequently associated with anti-MDA5 DM.

Mucocutaneous and Articular Manifestations of Anti-MDA5 Dermatomyositis

The hallmark cutaneous manifestations of the DM spectrum occur with similar prevalence in patients with anti-MDA5 DM. They include periorbital heliotrope (blue–purple) rash with edema, erythematous rash on the face, or the anterior chest (in a V-sign), and back and shoulders (in a shawl sign), violaceous papules or plaques located on the dorsal part of the metacarpophalangeal or interphalangeal joints, called Gottron's papules and cracked palmar fingertips ("mechanic's hands"). It is important to underline here the danger of potential misdiagnosis of patients with anti-MDA5 DM, given the absence of clinical

TABLE 1 | Ethnicity and prevalence of dermatomyositis with anti-MDA5 antibodies.

Country	DM cases* (n)	Anti-MDA5+	Detection technique	CADM among anti-MDA5+	CADM cases (n)	Anti-MDA5+ among CADM	Prevalence of women anti-MDA5+	ILD among anti-MDA5+	RP-ILD among anti-MDA5+	Ref	Publication year
Japan	42	19%	IP/IB/IIF	100%	15	53%	75%	88%	50%	(4)	2005
Japan	30	27%	IP	75%	13	46%	88%	100%	100%	(5)	2009
Japan	65	22%	IP/ELISA	57%	15	53%	79%	100%	71%	(6)	2010
Japan	37	35%	IP/IB	85%	15	73%	69%	92%	54%	(7)	2010
Japan	55	11%	IP	17%	ND	ND	50%	83%	50%	(8)	2011
Japan	376	11%	IP/IB/ELISA	77%	51	65%	79%	93%	65%	(9)	2011
Japan	79	22%	IP/ELISA	82%	21	67%	88%	94%	71%	(10)	2012
China	84	23%	ELISA	26%	8	63%	58%	ND	79%	(11)	2012
China	64	23%	ELISA	80%	32	38%	60%	100%	60%	(12)	2012
China	43	60%	ELISA	35%	9	100%	39%	100%	39%	(13)	2013
China	213	21%	ID	77%	81	42%	71%	82%	64%	(14)	2020
North America	77	13%	IP	50%	ND	ND	88%	67%	20%	(15)	2011
North America	160	7%	IP	46%	ND	ND	73%	73%	ND	(16)	2013
Spain	117	12%	ELISA/IB/IIF	57%	15	53%	64%	64%	57%	(17)	2014
Italy	34	15%	IP-WB/ELISA/IIF	100%	ND	ND	60%	60%	20%	(18)	2014
Brazil	131	16%	ELISA	24%	22	23%	71%	38%	ND	(19)	2018
France	54	9%	IIF/ID	ND	ND	ND	ND	ND	ND	(1)	2018

*Selection of cohorts with at least 30 patients (cohorts with JDM patients excluded).

DM, dermatomyositis; CADM, clinically amyopathic dermatomyositis; JDM, juvenile dermatomyositis; ILD, interstitial lung disease; RP-ILD, rapidly progressive interstitial lung disease; IP, immunoprecipitation; IB, immunoblot; IIF, indirect immunofluorescence; ELISA, enzyme-linked immunosorbent assay; WB, western blot; ID, immunodot assay; ND, not done.

evidence of myopathy and also sometimes the similarities of DM-specific manifestations, such as Gottron's papules, with other skin lesions, such as psoriatic lesions (23, 24).

Anti-MDA5 DM is also associated with a more specific cutaneous phenotype, that includes palmar papules and skin ulcerations, reported in both Caucasian (2, 15, 16, 25–28) and Asian populations (9, 10, 12, 29–31). Unlike Gottron's papules, palmar papules are often located on the palmar surface or lateral sides of the fingers, especially over metacarpophalangeal and interphalangeal joints. Many of these lesions have a central ivory coloration, and they are frequently painful. Palmar papules can be associated with hyperkeratosis, and complicated of ulcerations.

Skin ulcerations associated with anti-MDA5 DM usually manifest as deep painful ulcers localized over Gottron's papule, involving the digital pulp and nail folds, or over Gottron's sign on the knees, elbows, or both. The histology of skin ulcerations shows vasculopathy, pauci-inflammatory or characterized by medium vessel wall infiltration with mononuclear cells, and intravascular thrombus (12, 15). Major complications can occur, such as gangrene and osteomyelitis, potentially leading to digital amputation (25, 32, 33).

Less specific auricular skin lesions have also been described in DM with anti-MDA5 Abs, such as antihelix/helix violaceous macules and erythematous auricular papules (34, 35). These particular cutaneous phenotypes, seldom mentioned in the

literature, could be clinical markers of poor prognosis, although most previous studies examining auricular skin lesions in the disease have been limited to isolated case reports (36). Other less specific mucocutaneous lesions have been described in DM with anti-MDA5 Abs. They include oral ulcers, panniculitis, alopecia and flagellate erythema (15, 17, 37, 38).

Since 2010, many studies reported the increased prevalence of arthritis (42–82%) and arthralgia in patients with anti-MDA5 DM (2, 9, 15, 16). The arthritis described in the patients closely resembles that found in rheumatoid arthritis, potentially leading to misdiagnosis in the absence of other DM symptoms. They are associated with morning stiffness and are typically symmetric, affecting the small joints of the hands, but also the wrists and the ankles (16, 23, 39). Although rarely described in the literature, the conventional radiography do not show any bone erosion. However, erosions have been exceptionally reported by magnetic resonance imaging (16, 39). When explored, the rheumatoid factor was positive in some cases, but anti-cyclic citrullinated peptide Abs were not detected in any of the patients (27, 39).

Another differential diagnosis of these articular forms of anti-MDA5 DM is the psoriatic arthritis. Indeed arthritis is a common feature associated with psoriasis and some cases of anti-MDA5 DM initially manifest as psoriasiform skin lesions (23, 24, 40). In the literature, one case has been reported of a

patient with psoriasiform lesions, associated with severe symmetrical polyarthritis of large and small joints, and a mild proximal weakness in upper and lower extremities. All of these elements initially led to the misdiagnosis of psoriatic arthritis (23). However, thoracic CT scan subsequently showed a bilateral ground-glass pattern, and laboratory tests revealed blood positivity for anti-MDA5 Abs. Thus, the association of arthritis and psoriasiform lesions, associated with weak muscle or pulmonary involvement may lead to a misdiagnosis of psoriatic arthritis, if anti-MDA5 Abs are not explored.

Finally, some patients have a phenotype close to the antisynthetase syndrome (i.e. arthritis, Raynaud's syndrome, mechanic's hands, ILD), further complicating the diagnosis (16, 41).

Lung Manifestations of Anti-MDA5 Dermatomyositis

Anti-MDA5 DM is associated with poor prognosis due to a high prevalence of RP-ILD. ILD is the result of inflammation and fibrosis of the lung parenchyma and specific patterns of ground-glass attenuation are observed on high-resolution computed tomography (42). Significant disparities between association of anti-MDA5 Abs and RP-ILD have been described in the cohorts. For instance, in Japan and in East Asia populations, ILD occurs in 82 to 100% of patients with anti-MDA5 DM, and RP-ILD in 39 to 100% of patients (**Table 1**) (4–14). The incidence of RP-ILD appears less important in Caucasian populations with 38 to 73% of anti-MDA5 DM patients having ILD and 20 to 57% having RP-ILD (**Table 1**) (15–19), some studies reporting a significant association (17, 25) while others do not (15, 16, 19).

It is of importance to note that ILD can be absent at diagnosis and develop many years later, or completely absent in some patients over years of follow-up. Conversely, ILD can sometimes reveals anti-MDA5 DM. For instance, three Japanese patients with ILD and anti-MDA5 Abs, but without cutaneous nor muscular manifestation, had a fatal outcome within two months after onset (43). Similarly, two patients initially hospitalized for fever and dyspnea developed skin symptoms characteristic of DM one month after onset (44).

The mortality rate of patients who develop RP-ILD is reported to be approximately 50%, with most deaths occurring during the very early stages of the illness (6, 7, 9, 10, 13, 43, 45, 46). Some studies suggest that beyond 6 months after onset, disease progression tends to settle down, and relapse seems uncommon (10). However, recurrences have also been described in the form of cutaneous and/or severe respiratory relapse many years after onset and after months of clinical remission and new treatments are needed to contain the exacerbation (47, 48).

Association of Anti-MDA5 Dermatomyositis With Malignancy

Several DM are associated with an increased risk for cancer, such as DM with anti-TIF1 γ Abs, and to a lesser extent, DM with anti-NXP2 Abs (3, 49). By contrast, the association of anti-MDA5 DM with cancer has been explored in large cohorts, without any

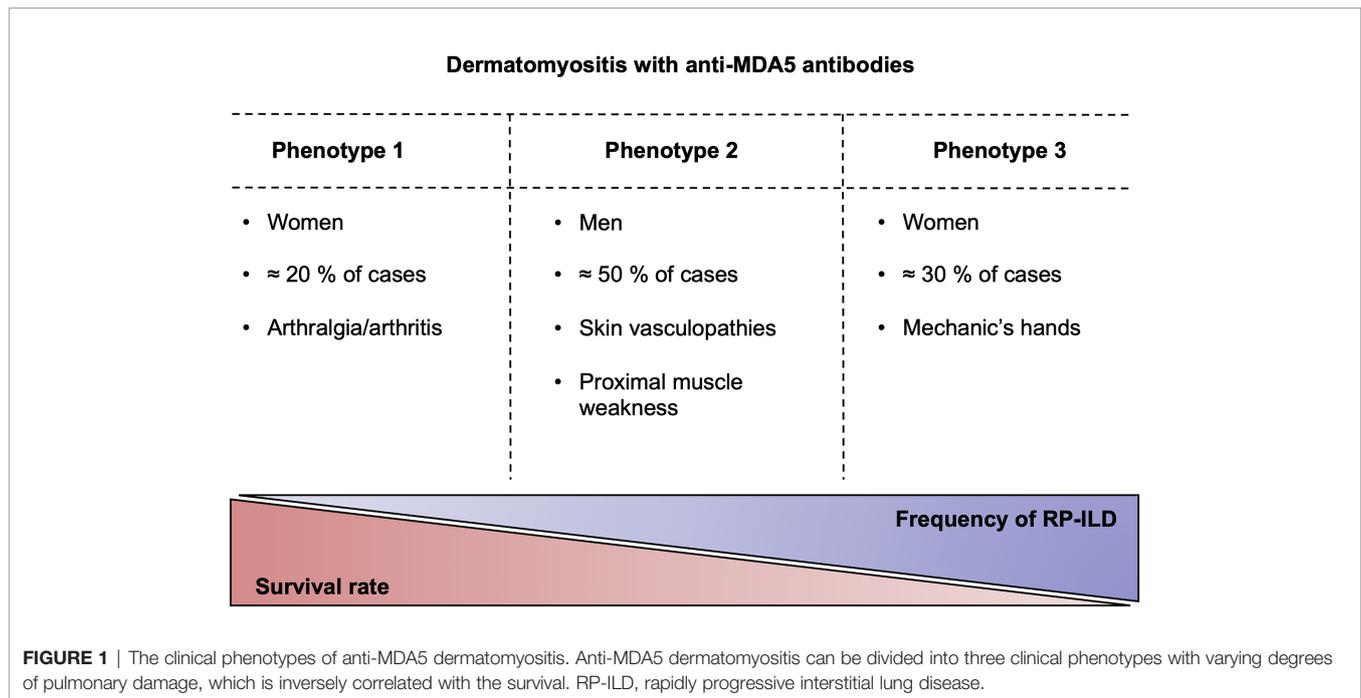
correlation reported (7, 9, 10, 12). However, few cases of cancer have been reported in patients with anti-MDA5 DM (33, 37, 50). Metastatic small cell carcinoma with liver involvement was detected 12 months after anti-MDA5 DM diagnosis in a 60 year-old French woman (50). Another study presented the case of a man diagnosed simultaneously with both anti-MDA5 DM and thyroid cancer (37). Even if no evident causal link is established, these few cases highlight that anti-MDA5 DM and cancer might not be exclusive.

Other Clinical and Biological Features of Anti-MDA5 Dermatomyositis

Among the constitutional symptoms, the fever has been described in up to 74% of anti-MDA5 DM at onset [33–74%] (7–9, 11, 13, 16). Several distinctive features in blood tests are also reported in the disease, such as elevated ferritinemia, with no significant elevation of C-reactive protein. Ferritin levels correlate with the severity of the disease and ILD (6, 7, 51, 52). Liver dysfunction is also recurrently observed in anti-MDA5 DM, as evidenced by elevated levels of alanine transaminase or gamma-glutamyl transferase, without elevated creatine kinase (6, 7, 21, 53). Liver biopsies show steatosis and hepatocyte ballooning (53). Furthermore, liver enzymes might increase as ferritinemia increases and ILD worsens (7). Decreased CD4⁺ and CD8⁺ T cell counts and a raised CD4⁺/CD8⁺ ratio are frequently described in peripheral blood of patients with anti-MDA5 DM, even before immunosuppressive treatment (11, 54). When studying the relation between pulmonary lesions and lymphopenia, an increase of CD4⁺ and CD8⁺ T cell counts is observed in parallel with pulmonary lesions improvement after treatment. Inversely, CD4⁺ and CD8⁺ T cell counts decrease and the CD4⁺/CD8⁺ ratio increases in patients with ILD refractory to treatment (54). Finally, positivity for anti-TRIM21 (Ro52) Abs (55, 56) and older age (14, 29) are indicators of poor prognosis while the female sex and articular form might be factors of good prognosis (57).

Dermatomyositis With Anti-MDA5 Antibodies: Three Distinct Clinical Phenotypes

The heterogeneity of clinical features and outcomes among patients with anti-MDA5 DM prompted to divide the clinical spectrum of anti-MDA5 DM into three distinct clinical subgroups (**Figure 1**) (2). The first one is composed mainly of women with RP-ILD associated with mechanic's hands, with the highest mortality rate (80%). A rheumatologic group (55% of cases) is also made mostly of women with arthralgia or arthritis (83%), less frequent RP-ILD (17%) and a better prognosis. The third clinical phenotype mainly encompasses men with symptoms dominated by skin vasculopathies, including a Raynaud's phenomenon (82%), skin ulcers (77%), digital necrosis (32%) and calcinosis (23%). This last subgroup is frequently associated with proximal muscles weakness (68%), with relatively few RP-ILD (23%) and it is of intermediate prognosis. This classification was already suggested by a



Spanish group in 2014 (17) and could permit an appraisal of the prognosis of patients with anti-MDA5 DM.

JUVENILE DERMATOMYOSITIS ASSOCIATED WITH ANTI-MDA5 ANTIBODIES

Juvenile DM (JDM) is associated with anti-MDA5 Abs in 6 to 12% of cases in European and North American cohorts, and constitutes the third most frequent DM-associated Abs after anti-TIF1 γ and anti-NXP2 Abs (56, 58–62). The prevalence of the anti-MDA5 Abs is higher in Japanese cohorts, with 24 to 38% of patients having anti-MDA5 Abs associated JDM (63–65). The anti-MDA5 JDM phenotype is associated with a higher frequency of constitutional symptoms (weight loss, fever and adenopathy) and milder muscle involvement compared to the other forms of JDM (58, 61, 62, 65).

Similarly to the adult form, the specific cutaneous phenotype found in anti-MDA5 DM, i.e. the palmar papules and skin ulcerations, are also observed in anti-MDA5 JDM, as well as arthritis, which is more frequent than in other subtypes of JDM (58, 59, 61, 64). These lesions distinguish anti-MDA5 JDM from the other forms of JDM. The arthritis associated with the juvenile form is similar to the one observed in adults, characterized by symmetrical pain and swelling of the small joints of the hands (58, 66). Anti-MDA5 JDM is also associated with an increased risk of ILD (58, 61, 63–65, 67), which is more frequent compared with other forms of JDM associated with MSA, except for anti-synthetase Abs-associated JDM. RP-ILD appears to be less common (<10%) in Caucasian anti-MDA5 JDM, compared to the adult form (58). Thus, a major difference between the

juvenile form and the adult form of the disease in Caucasian patients is observed in terms of outcome, since anti-MDA5 JDM has comparable outcome with the other forms of JDM, but flares are less frequent in this subset of JDM and necessitates less medication (58, 59, 61). The observations on the association between anti-MDA5 Abs JDM and RP-ILD are more contradictory in the Japanese population. Indeed, although some studies report a much higher frequency of RP-ILD in Japanese anti-MDA5 JDM patients associated with a poor prognosis, a recent multicenter study reported only 19% of RP-ILD in a cohort of 31 anti-MDA5 Abs positive patients, with a lower rate of relapse than other MSA positive patients (63, 67, 68).

MDA5 AND ANTI-MDA5 ANTIBODIES

In 2005, Sato et al. reported that Abs found in the sera of patients with CADM react with a cytoplasmic protein of 140 kDa (4). In 2009, the same team described MDA5 as the antigenic target of 140 kDa which is recognized by the Abs found in patients with CADM (69).

MDA5 was initially identified in 2002, as a type I IFN-inducible gene in human melanoma cells, and the first function described for this protein was to induce the death of cancer cells (70). Since this first description, MDA5 is now considered as a key sensor of viral infection, mediating the production by the infected cell of IFN-I and the induction of other genes that collectively establish an antiviral host response. MDA5 is encoded by the gene *IFIH1* (interferon-induced with helicase C domain 1) and is part of the Retinoic acid Inducible Gene-I-like receptor family that detect molecular patterns of

viruses that have gained access to the cytosol of the infected cell. More specifically, MDA5 senses preferentially long (> 300 bp) RNA-RNA strand pairs, which are mainly RNA viruses replicative intermediates, although some DNA viruses also produce them during their life cycle (71, 72). MDA5 senses these double-stranded RNA (dsRNA) *via* its RNA helicase domain and a C-terminal domain (CTD), and subsequently transmits a signal *via* its homotypic interacting caspase recruitment domain (CARD). Upon recognition of dsRNA, MDA5 assembles into a filament along the dsRNA axis and adopts a ring-like conformation around dsRNA, allowing MDA5 to bind with a strong affinity to its ligand. MDA5 then interacts with the mitochondrial antiviral signaling protein (MAVS), present on the outer membrane of the mitochondria, peroxisomes and other mitochondria-associated membranes. The interaction between MDA5 and MAVS through their respective CARD leads to the activation of the transcription factors interferon regulatory factor 3 and 7 (IRF3 and IRF7), as well as nuclear factor-kappa B (NF- κ B). Phosphorylated IRF3, IRF7 and NF- κ B then accumulate in the nuclei where they activate antiviral gene transcription, including IFN-I genes, leading to the production of type I IFN (IFN α and IFN β) and pro-inflammatory cytokines (73, 74).

By this way, MDA5 is able to detect and limit the replication of several RNA viruses of the picornavirus, flavivirus or coronavirus families, as well as DNA viruses of the herpesvirus family for instance (71, 75). But MDA5 is also able to sense endogenous dsRNA, such as mitochondrial dsRNA generated by bilateral transcription of mitochondrial DNA. This biological mechanism is tightly regulated, by mitochondrial degradasome enzymes such as the polynucleotide phosphorylase (PNPase), avoiding the accumulation of dsRNA and thus deleterious activation of MDA5. As a result, PNPase deficiency can cause a massive accumulation of long mitochondrial dsRNA, escaping into the cytosol and leading ultimately to an uncontrolled activation of MDA5 and an upregulation of interferon-stimulated genes (76). In line with this, patients with hypomorphic mutations in *PNPT1*, the gene that encodes for PNPase, have type I interferonopathies characterized by a constitutive activation of the IFN-I axis (77).

Tissue Distribution and Cellular Location of MDA5

A greater knowledge of the tissue distribution and cellular location of MDA5 is essential to better understand the pathophysiology of anti-MDA5 DM. MDA5 has low tissue specificity in physiological condition (78). A higher expression in the skin and lung tissues of patients with anti-MDA5 DM would be expected compared to expression in muscle tissues. Although studies on MDA5 expression in these target tissues are very rare, MDA5 expression has been shown to be enhanced in skin biopsies of patients with DM, which could offer an element of response to the severe cutaneous symptoms associated with the disease (79).

Defining the cellular location of MDA5 in a pathological context is also critical as pathogenic Abs necessitate to be

internalized when the antigenic target is cytosolic, or act by interacting with a protein expressed at the surface of a target cell. At the cellular level, MDA5 was initially described as an intracellular protein found in the cytoplasm of most cells (70). However, Berger et al. studied MDA5 expression and subcellular localization in neutrophils and showed an overexpression of MDA5 both in the cytoplasm and in secretory vesicles as well as a cell surface expression (80). Expression of MDA5 at the surface of other cell types is still being determined. The identification of transient or constitutive expression of MDA5 on the surface of target cells could constitute a critical element in the understanding of the pathogenesis of the disease, in favor of a potential pathogenic role of anti-MDA5 Abs.

Anti-MDA5 Autoantibodies: The Diagnostic Marker

The identification of the anti-MDA5 Abs in CADM was initially performed by immunoprecipitations (IP) of sera from patients incubated with ³⁵S-methionine-labeled K562 cell extracts (4). Since then, radiolabeled IP is considered as the gold standard testing method to detect anti-MDA5 Abs. Although the substitution of radiolabeled antigenic extracts by biotin-labeled recombinant MDA5 constitute a good alternative to bypass the use of radioactive materials (81), IP remains difficult to use in everyday practice, because time-consuming and expensive. Another major issue of IP assay regarding anti-MDA5 Abs identification is the comigration of the MDA5 antigen with other antigens found in DMs, such as the antigens NXP2, TIF1 γ and OJ (18, 59, 82). Great expertise is therefore required for correct identification of anti-MDA5 Abs by IP, and alternative interpretation and other assays are therefore required to confirm a serum positivity.

For all the reasons mentioned above, IP is used only in a limited number of medical laboratories, which opted now for qualitative or quantitative alternative assays. Qualitative assays include indirect immunofluorescence (IIF) staining on HEp-2 cells and immunodot assays (so-called line blot or dot blot) (83). IIF staining performed on HEp-2 cells with diluted sera from patients with anti-MDA5 DM can give rise to a characteristic cytoplasmic staining with a finely granular appearance, in rare clustered cells and in one study in all cells (**Figure 2**) (4, 84, 85). This difference could be due to the source of HEp-2 cells differentially expressing MDA5 or to the presence of other Abs. However, the scarcity of positive cells generally observed in the microscope field makes the identification of anti-MDA5 Abs by IIF very touchy, and requires a trained eye. Moreover, this IIF pattern is inconstant. In our personal experience that includes a cohort of 31 anti-MDA5 DM, this particular IIF pattern was observed with 50% of sera. Several nonspecific IIF patterns are otherwise observed and have been described, such as a granular cytoplasmic pattern in all cells (which can mask the typical cytoplasmic pattern), or a nuclear speckled pattern, associated or not with the typical cytoplasmic pattern (**Figure 2**) (4). Finally, anti-MDA5 Abs positive sera can also be negative by IIF (17, 18). IIF results should thus always be interpreted by taking in consideration the clinical context and

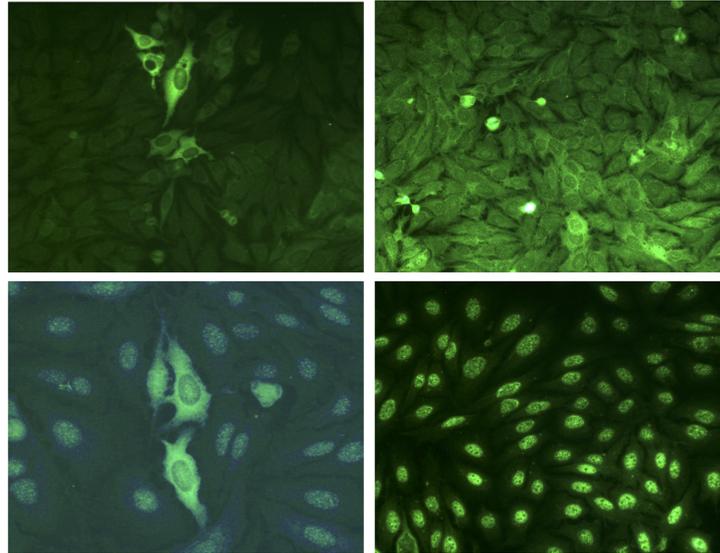


FIGURE 2 | Indirect immunofluorescence patterns of HEp-2 cells stained with anti-MDA5 positive sera. Stainings were performed on HEp-2 cells (Kallestad, Biorad) with sera from patients with anti-MDA5 Abs dermatomyositis. Top left: typical fine granular cytoplasmic staining in rare clustered cells. Top right: granular cytoplasmic pattern in all cells (which can mask the typical cytoplasmic pattern). Bottom left: nuclear speckled pattern, associated with the typical cytoplasmic pattern. Bottom right: isolated nuclear speckled pattern. Note that absence of fluorescence can also be observed with some sera.

other specific assays should be conducted to confirm a positive IIF pattern or to further explore a negative IIF, if there is a strong clinical suspicion of anti-MDA5 DM.

Commercialized immunodot assays (Line blot LIA, Euroimmun, Lübeck, Germany and dot blot D-Tek, BlueDiver, Mons, Belgium) and particle-based multi-analyte technology (PMAT, Inova, Diagnostics, US) can be used for the detection of anti-MDA5 Abs. These specific qualitative assays i.e. Line Blot, Dot blot, have been validated using the IP assay as gold standard, with reported specificity of 96-99% or 98% and sensitivity of 75-93% or 76% respectively. High level of agreement was also found between IP and PMAT (86-88).

Enzyme-linked immunosorbent assays (ELISA) have also been recently developed to detect anti-MDA5 Abs and allow their quantification. An ELISA based on a recombinant MDA5 fusion protein produced in insect cells has shown an analytical sensitivity and specificity of 98% and 100% respectively, compared with IP assays (89). A major benefit of anti-MDA5 Abs ELISA is that it allows a precise quantification of the level of anti-MDA5 Abs in the serum of patients, and a follow-up of its variation during the course of the disease or after the introduction of a therapy.

Anti-MDA5 Autoantibodies: A Tool for Monitoring Disease Activity

The quantification of anti-MDA5 Abs level seems to be a way to predict disease outcome. When comparing surviving and deceased patients, anti-MDA5 Abs levels are significantly lower in surviving patients. Moreover, the outcome of patients with low titers of anti-MDA5 Abs is approximately the same as patients without anti-MDA5 Abs (90). Anti-MDA5 Abs titers also

correlate with the severity of the disease, and more particularly with the severity of ILD and cutaneous symptoms. Several Japanese studies reported higher levels of anti-MDA5 Abs in JDM patients with RP-ILD (63, 64, 67). One study reported deep necrotic ulcerations in the patients with the highest anti-MDA5 Abs levels and only superficial cutaneous symptoms in those with the lowest value (12).

Furthermore, the value of the anti-MDA5 Abs could also be useful for the evaluation of the response to treatment. In a Japanese cohort, patients with anti-MDA5 Abs levels greater than 500 units/mL (positivity threshold at 8 units/mL) were resistant to treatment by glucocorticoids/cyclophosphamide or intravenous immunoglobulins, and died (12). Inversely, patients with anti-MDA5 Abs levels lower than 500 units/mL had less severe lung lesions and cutaneous symptoms improved after treatment. Finally, monitoring the Abs levels along the course of the disease could permit to objective a remission or to detect a relapse early. Remission induces the disappearance of anti-MDA5 Abs, whereas it remains elevated in the patients who die or who later relapse (91, 92).

In conclusion, the anti-MDA5 Abs are a critical diagnostic biomarker of the disease and the level of Abs could be an important prognostic and predictive parameter to monitor in patients.

Co-Occurrence of Anti-MDA5 Autoantibodies With Other Autoantibodies

Anti-TRIM21 Abs (also known as anti-Ro52/SSA-52 Abs) are nonspecific Abs encountered in several connective diseases, and also frequently detected in the serum of patients with anti-MDA5 DM, with 27 to 62% of dual-positive patients. These patients

seem to develop RP-ILD more frequently and have a less favorable prognosis (15, 16, 55, 56).

A study reported the presence of Abs directed against a nuclear protein, the splicing factor proline/glutamine-rich (SFPQ), in the serum of 27 out of 51 patients (53%) with anti-MDA5 DM (93). SFPQ is a multifunctional nuclear protein of 110 kDa, that participates in diverse molecular functions including transcription regulation, and is also involved in the regulation of host innate immune response to viruses (94, 95). Anti-SFPQ Abs have not been detected in other form of DMs, and were identified at diagnosis in 13 patients, while the others turned positive during the disease course (93). Another study observed an apparition of anti-SFPQ Abs at recurrence (47). The clinical relevance of these Abs is unknown, and they are not researched in current practice.

PATHOGENESIS OF THE DISEASE

Owing to the rarity of anti-MDA5 DM, knowledge on the pathogenic mechanisms of the disease remain limited, but they are believed to occur as a consequence of a particular gene-environmental interaction. Although scarce, several studies have underlined an association of HLA and non-HLA alleles with the disease. Moreover, the identification of seasonal and geographical clustering at disease onset suggests that an infectious agent could be a triggering factor, an attractive hypothesis in view of the antiviral function of the MDA5 antigen. As for the pathogenic mechanisms involved in the disease, since MDA5 is an IFN-I inducible gene, IFN-I could be the starting point for most of the pathophysiological pathways.

Genetic Susceptibility

HLA allele associations have been described in Asian cohorts. The strongest disease association was found with alleles of the type II HLA alleles HLA-DRB1. Analyses of the relationship between type II HLA alleles and anti-MDA5 DM in Chinese cohorts demonstrated a higher frequency of HLA-DRB1*04:01, *12:02 and *12:01 alleles in Chinese patients with anti-MDA5 DM (96, 97). However, different risk factors are observed in particular ethnic groups, including combined frequency of HLA-DRB1*01:01 and *04:05 in Japanese patients (98), with no significant difference for the same alleles in Chinese populations (96). In addition, some alleles such as HLA-DRB1*09:01 have been associated with a worse prognosis in Chinese patients (97). To date, no association between HLA alleles and anti-MDA5 DM has been identified in Caucasian population (99).

Although HLA allele associations differ across ethnic populations, amino acid sequence variations observed in the type II HLA-DRB1 alleles might affect the structure of the antigen-binding groove of the HLA molecule and thereby influence the antigenic repertoire, increasing by this way the disease susceptibility.

One non-HLA locus, an intronic variant of WDFY4, has been recently associated with the anti-MDA5 DM in Japanese patients

(100). This variant induces a higher expression of a truncated isoform of the WDFY4 protein. Although the precise biological function of WDFY4 was unknown when this work was published, Kochi et al. demonstrated that the truncated form of WDFY4 markedly enhanced the MDA5-mediated NF- κ B activation and cell apoptosis. Interestingly, one year later, Theisen et al. uncovered the function of WDFY4, which is in fact a critical regulator of cross-presentation in the conventional CD1c⁺ dendritic cells (101). This is of particular interest in the context of anti-MDA5 DM since, as it will be discussed later, an inadequate immune response to an infectious agent could be the environmental factor triggering the disease. It is then tempting to speculate that qualitative and/or quantitative alterations of WDFY4 might induce a striking defect in cross-presentation of viral-associated antigens, which may trigger secondarily an aberrant autoimmune response (Figure 3).

Other molecular defects, unexplored so far, might also be involved in the pathogenesis of anti-MDA5 DM. For instance, given the role of mitochondrial PNPase in eliminating endogenous dsRNA, local dysregulated expression and/or

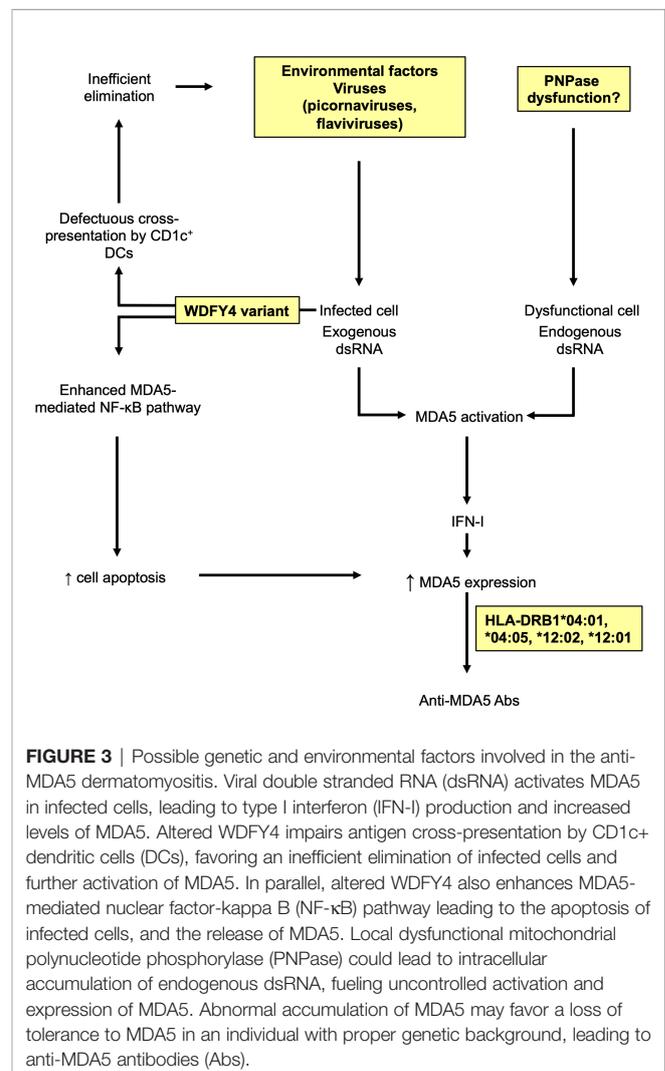


FIGURE 3 | Possible genetic and environmental factors involved in the anti-MDA5 dermatomyositis. Viral double stranded RNA (dsRNA) activates MDA5 in infected cells, leading to type I interferon (IFN-I) production and increased levels of MDA5. Altered WDFY4 impairs antigen cross-presentation by CD1c⁺ dendritic cells (DCs), favoring an inefficient elimination of infected cells and further activation of MDA5. In parallel, altered WDFY4 also enhances MDA5-mediated nuclear factor-kappa B (NF- κ B) pathway leading to the apoptosis of infected cells, and the release of MDA5. Local dysfunctional mitochondrial polynucleotide phosphorylase (PNPase) could lead to intracellular accumulation of endogenous dsRNA, fueling uncontrolled activation and expression of MDA5. Abnormal accumulation of MDA5 may favor a loss of tolerance to MDA5 in an individual with proper genetic background, leading to anti-MDA5 antibodies (Abs).

function of the enzyme could generate an accumulation of endogenous dsRNA, leading to MDA5 activation, IFN-I pathway stimulation and elevated MDA5 expression.

Environmental Factor

The hypothesis of a viral trigger of anti-MDA5 DM is suggested by epidemiological studies that highlighted a seasonal distribution of the disease. Classically, anti-MDA5 DM onset follows a seasonal repartition with an epidemic period in fall and winter, a peak in late winter and spring and a dip in summer, following respiratory virus epidemic period (102, 103). In the hypothesis of a viral infection as a possible initiator of autoimmunity, the scenario could involve an activation of MDA5 in the infected cells, leading to IFN-I production and increased levels of MDA5, followed by an excessive local apoptosis favored by a specific genetic background (intronic variant of WDFY4). The release of the MDA5 antigen into the microenvironment following cell lysis could be the cause of a loss of tolerance towards MDA5, resulting in the production of anti-MDA5 Abs. This mechanism could also be fueled by a defect in the elimination of the virus, due to an inefficient cross-presentation of viral antigens by CD1c⁺ DCs (Figure 3).

It is also interesting to note that a geographical distribution of the disease has also been reported in Japan, with an increased prevalence of anti-MDA5 DM in patients living in rural areas as opposed to urban areas as well as in patients residing near freshwater (103, 104).

Pathogenesis of the Anti-MDA5 Dermatomyositis: The Vasculopathy Hypothesis

Several pieces of evidence point toward a central role of vascular injury in the pathogenesis of anti-MDA5 DM: (i) Patients frequently show typical cutaneous features such as skin ulcers due to severe vasculopathy; (ii) Histopathology of palmar papules, as well as lung tissues highlights vasculopathy (12, 15, 105); (iii) Biomarkers of endothelial dysfunctions have been identified in the sera of patients (106, 107); (iv) Patients exhibited a strong IFN-I signature distributed in the vasculature of the skin (108, 109). It is important to emphasize that these particular features are observed in the skin and the lungs of patients with anti-MDA5 DM but that there is no or few signs of vasculopathy nor of an enhanced IFN-I signature, compared to other DM, in muscle tissue of anti-MDA5 DM patients (21, 22).

Biomarkers of endothelial dysfunctions released in the sera of anti-MDA5 DM patients include endothelin, thrombomodulin, plasminogen activator inhibitor, von Willebrand factor (vWF), soluble vascular cell adhesion molecule-1 and soluble intercellular adhesion molecule-1 (106, 107). Interestingly, serum levels of endothelin and vWF are higher in anti-MDA5 DM patients who have cutaneous ulcers and ILD and IFN-I signature correlates positively with endothelin levels (106). All together, these data suggest that blood vessels exposure to IFN-I in anti-MDA5 DM may lead to endothelial injury, ultimately

responsible for the cutaneous and pulmonary lesions associated with this disease.

Vasculopathy and Pulmonary Lesions

Anti-MDA5 DM is associated with an accumulation of activated macrophages (M2) in alveoli (110, 111). Several markers of macrophage activation are elevated in the serum of anti-MDA5 DM patients, such as a soluble form of CD206, which is highly expressed by alveolar macrophages (110, 112). Other soluble markers of activated macrophages, such as a soluble form of CD163, as well as the chitotriosidase and the ferritin, are also found in the sera of patients at significantly higher levels than in other subtypes of DM (110, 111, 113, 114). Neopterin levels, another marker of macrophage activation, are elevated in DM patients with anti-MDA5 Abs in association with RP-ILD and reduced survival. A positive correlation of neopterin levels with ferritin and markers of disease severity, and a negative correlation with pulmonary function have been demonstrated (115). Multiple chemokines can induce the recruitment of M2 macrophages. Among them, the chemokine CX3CL1 (Fractalkine) is of interest in anti-MDA5 DM since its concentration is elevated in sera of patients (116). CX3CL1 secretion can be induced by IFN-I in pulmonary vascular endothelial cells and induces the recruitment of CX3CR1⁺M2 macrophages in the lungs (117, 118). It is then tempting to speculate that high levels of IFN-I induce endothelial injury leading to the secretion of CX3CL1, responsible for the recruitment of intrapulmonary profibrotic M2 macrophages. Locally, M2 macrophages produce TGF- β to directly promote pulmonary fibrosis, and could be involved in the recruitment of profibrotic CD4⁺CXCR4⁺ T cells. In agreement with this, elevated levels of CD4⁺CXCR4⁺ T cells are observed in the peripheral blood and the bronchoalveolar lavage fluids of patients with ILD. Locally, the airway epithelial cells and the macrophages are the main sources of the stromal cell derived factor-1 (SDF-1 or CXCL12), the ligand of CXCR4 (119). Once recruited, the CD4⁺CXCR4⁺ T cells may promote pulmonary fibroblast proliferation, partly through the release of IL-21, and secondarily by the production of profibrotic agents, namely TGF- β , α -smooth muscle actin and collagen I. IL-21 is known to induce the differentiation of IL-13 producing-CD8⁺ T cells, which in turn enhance IL-21 production, creating an activation loop (120). IL-13 may fuel pulmonary fibrosis through two mechanisms: direct activation of fibroblast, and stimulation of the synthesis of TGF- β by activated macrophages (121) (Figure 4).

Vasculopathy and Cutaneous Lesions

Histology of skin biopsies from patients with anti-MDA5 DM classically show an interface dermatitis, a histopathological pattern characterized by vacuolar changes, apoptotic keratinocytes and infiltration of CD8⁺ lymphocytes at the dermoepidermal junction (122–124). The interface dermatitis is also a classical feature observed in systemic erythematosus lupus and lichen planus. Another common feature of all these diseases is the enhanced IFN-I signaling into the skin of patients.

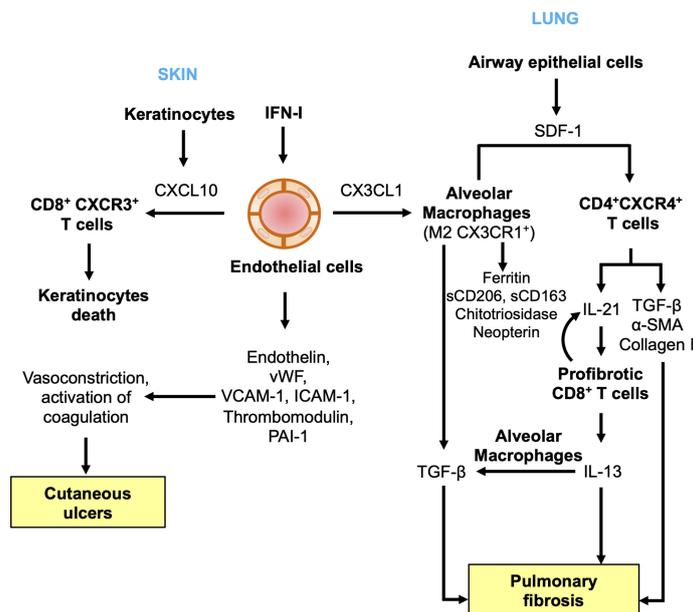


FIGURE 4 | Endothelial dysfunctions and immune alterations in anti-MDA5 dermatomyositis. In the skin, CXCL10, a keratinocyte and endothelial-derived chemokine, induces the recruitment of $CD8^+CXCR3^+$ T cells, potentially autoreactive and leading to keratinocytes death. Endothelin released by injured endothelial cells, is a strong vasoconstrictor which can induce local ischemia responsible for cutaneous ulcers. In the lung, CX3CL1 can be produced by vascular pulmonary endothelial cells following IFN-I exposure. CX3CL1 recruits $CX3CR1^+$ alternative alveolar macrophages (M2). Alveolar M2 macrophages, as well as airway epithelial cells, release stromal cell-derived factor 1 (SDF-1) which induces the accumulation of intrapulmonary $CD4^+CXCR4^+$ T cells. $CD4^+CXCR4^+$ T cells produce profibrotic agents (transformation growth factor- β (TGF- β), α -smooth muscle actin (α -SMA) and collagen I), as well as IL-21, which promotes the differentiation of profibrotic $CD8^+$ T cells. $CD8^+$ T cells secrete IL-13, which stimulates macrophages to produce profibrotic factors.

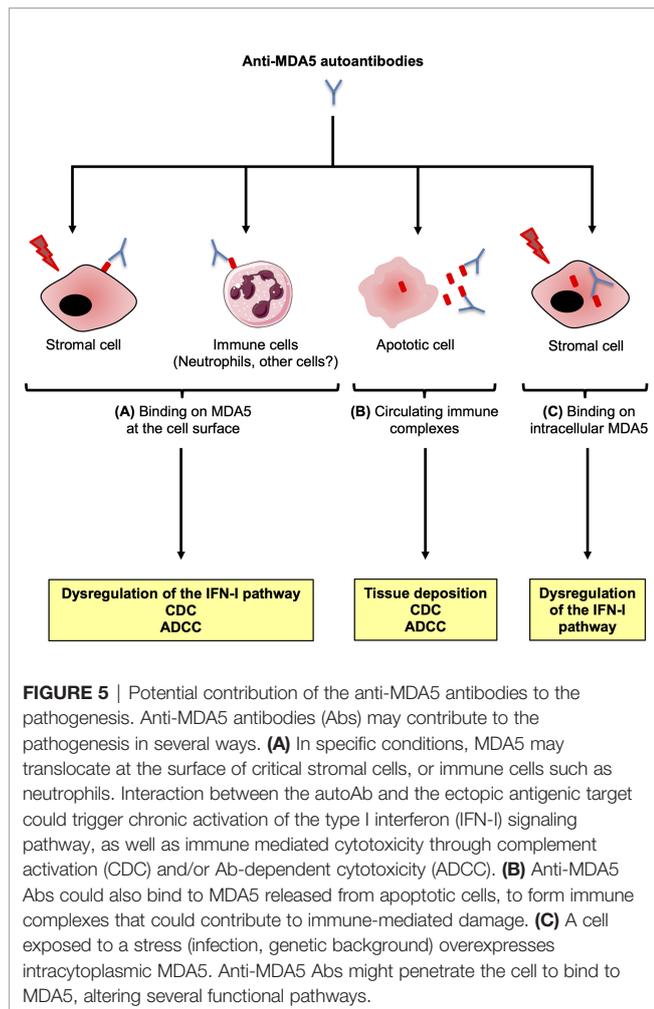
A common pathophysiological model has therefore been proposed, based on the secretion of keratinocyte-derived CXCL10 following skin exposure to IFN-I (125–128). Consistent with this observation, the expression of CXCL10 as well as the expression of IFN-I induced proteins (ISG15, MxA) are upregulated in the skin of anti-MDA5 Abs positive patients (108, 109, 126, 129). Endothelial cells stimulated by IFN-I could also constitute another source of CXCL10 (130). CXCL10 then induces the recruitment of $CXCR3^+CD8^+$ T cells, potentially autoreactive, responsible for the apoptosis of keratinocytes (125, 126) (Figure 4).

In addition, skin biopsies from anti-MDA5 DM patients also show more severe lesions, affecting the deeper layers of skin, down to the dermis. Epidermal necrosis and typical findings of vasculopathy are observed, with vascular fibrin deposition, thickening of the vascular walls and vascular thrombosis of the small and medium vessels (12, 15, 123, 131). Endothelin, released consequently to endothelial injury caused by chronic exposure to IFN-I, acts as a potent vasoconstrictor. It may in part be responsible for the skin ulcers, by inducing a local ischemia, as is the case in other diseases (132). The regression of skin ulcers in patients treated with bosentan, an endothelin-receptor inhibitor, or with a vasodilator drug, such as sildenafil, supports this hypothesis (133–135). In the same way, pro-coagulant factors (e.g. vWF) released by damaged endothelial cell could lead to blood coagulation activation, as evidenced by the vascular fibrin deposits and thromboses observed histologically (Figure 4).

Potential Contribution of Anti-MDA5 Antibodies to the Pathogenesis

Whether we consider skin or lung lesions, IFN-I seems to be the starting point for all the pathophysiological pathways described above. Indeed, IFN-I signaling is enhanced in anti-MDA5 DM, in skin and in serum, more than in other DM, suggesting the presence of a specific trigger of the IFN-I pathway in the disease (109, 129, 136). Here we assume that anti-MDA5 Abs could be the cornerstone of the dysregulation of the IFN-I pathway in anti-MDA5 DM. Several observations support this hypothesis: (i) The severity of the disease correlates with the titers of anti-MDA5 Abs (12, 63, 64, 67); (ii) The use of therapies that target the humoral immune response has shown its effectiveness in patients with anti-MDA5 DM (137–141); (iii) MDA5 expression has been shown to be enhanced in skin biopsies of patients (79).

In pathological contexts, MDA5 is overexpressed in altered tissues such as the skin of patients and ectopic expression of MDA5 at the cell surface has been reported (79, 80). In this context, anti-MDA5 Abs could then bind to MDA5-positive cells and induce an inappropriate activation of MDA5, leading to the dysregulation and the chronic activation of the IFN-I pathway in target tissues, worsening the existent lesions. Anti-MDA5 Abs binding to its antigenic target could also induce immune-mediated cytotoxicity by complement fixation or antibody-dependent cytotoxicity, further worsening the lesions (Figure 5A). Apart from binding to MDA5 expressed on cell



surface, anti-MDA5 Abs could also form immune complexes with the MDA5 proteins released from apoptotic skin and/or lung fibroblasts. These immune complexes could then deposit in organs, for example in dermal/lung vessels, inducing more vascular damage (**Figure 5B**). Finally, anti-MDA5 Abs could penetrate cells and interact with cytoplasmic MDA5, similarly to what has been described with other Abs, altering several functional pathways (**Figure 5C**) (142–144).

The isotype and the subclass of anti-MDA5 Abs might also affect their potential pathogenic function. In a Chinese cohort, anti-MDA5 IgA and IgG were the predominant isotypes. Interestingly, anti-MDA5 IgG1 were associated with higher serum ferritin levels, severe interstitial pneumonia, and a higher mortality rate. The combined positivity for anti-MDA5 IgG1 and anti-MDA5 IgG4 was predictive of poor prognosis (145). IgG1 are potent activators of the complement pathway, and as a result, anti-MDA5 IgG1 could be the main subclass involved in the pathogenesis of the disease. The identification of anti-MDA5 IgG4 is more unexpected, as IgG4 are considered non-inflammatory, owing to the unique structure of their hinge region (146). Whether the presence of anti-MDA5 IgG4 in the

most severe forms of the disease reflects a modulatory mechanism, although insufficient, remains to be defined.

Similarities Between Anti-MDA5 Dermatomyositis and COVID-19

Anti-MDA5 DM and coronavirus disease 2019 (COVID-19) share several common features, clinical and pathogenic, and exploring the pathophysiological mechanisms of COVID-19 may help to better understand the pathogenesis of anti-MDA5 DM (147).

First, as for anti-MDA5 DM, COVID-19 can be complicated by interstitial pneumonia which can lead to acute respiratory distress syndrome and death. This pulmonary damage is difficult to distinguish from the ILD associated with anti-MDA5 DM (148). Thus, it is important to carefully assess patients with RP-ILD, in particular in the case of a negative RT-PCR test for SARS-CoV-2 (severe acute respiratory syndrome coronavirus 2) as RP-ILD may be the sole symptom at anti-MDA5 CADM onset. Anti-MDA5 DM may then be easily confounded with COVID-19, delaying the diagnosis and treatment. Inversely, patients with positive test for anti-MDA5 Abs, or any other MSA, and who develop acute pneumonia should be tested for SARS-CoV-2 infection, as the infection worsens the patient's condition (149).

Second, anti-MDA5 DM and COVID-19 seem to share several pathogenic mechanisms. Both diseases are characterized by an activation of the IFN-I signaling pathway. However, although in anti-MDA5 DM, dysregulation of the IFN-I axis seems to exacerbate the disease, the role of the IFN-I pathway in COVID-19 appears more complex. MDA5 is a sensor of SARS-CoV-2 in lung cells, and its activation induces an IFN response to eliminate the virus (150–152). The IFN-I pathway plays a major role in the defense against the virus in COVID-19, as illustrated by the impaired IFN-I signaling in severe cases and the presence of anti-IFN-I Abs in the sera of more than 10% of patients with a life-threatening infection (153–156). In this sense, early activation of the IFN-I signaling pathway appears to be an essential weapon against SARS-CoV-2 infection. In contrast, a strong delayed IFN-I response could exacerbate the hyperinflammation associated with the most severe forms of COVID-19 (157, 158).

Another common pathological feature shared by the two diseases is the endothelial injury and the thrombotic manifestations associated with the more severe forms (148, 159, 160). This is highlighted by the presence of markers of endothelial and platelet activation as well as coagulation factors and fibrinolytic enzymes in the serum of patients, such as vWF, thrombomodulin, P-selectin and D-dimer (161). The important systemic inflammatory response and activation of the angiotensin converting enzyme 2 (ACE2), the major cell entry receptor for SARS-CoV-2, expressed on endothelial cells are accountable for these vascular damages (159, 162).

Finally, anti-MDA5 Abs have been identified in the serum of Chinese patients diagnosed with COVID-19 (48.2% of 274 patients), frequently in the most severe cases. Furthermore, the titers of anti-MDA5 Abs appear to correlate with the severity of the disease and were higher in the non-survival cases (163).

These preliminary results should, however, be taken with caution. Indeed, although the titers of anti-MDA5 Abs are statistically higher in the non-survivals infected SARS-CoV-2 patients *versus* the survivals, the orders of magnitude are very low (5.95 ± 5.16 U/mL *vs* 8.22 ± 6.64 U/mL, $P = 0.030$). In addition, it is well established that common viral infections, as well as SARS-CoV-2 infection, frequently trigger the induction of transient, low-titer, polyspecific autoantibodies (164, 165). In agreement with this, several myositis-specific autoantibodies have been identified as false positives in patients with COVID (166). To what extent these anti-MDA5 Abs are really specific, to what extent they are present in the infection of other RNA viruses or only during the infection by SARS-CoV-2, and what are their implications for the patient in the short and long term are all unanswered questions to be explored.

TREATMENT OF ANTI-MDA5 DERMATOMYOSITIS

Although no recommendations for the management of anti-MDA5 DM exist at this time, a combination immunosuppressive (IS) therapy is widely used. This intensive IS treatment classically consists of an association of glucocorticoids with a calcineurin inhibitor (cyclosporine A or tacrolimus) or a triple therapy with the addition of intravenous cyclophosphamide or mycophenolate mofetil. However, many cases are refractory to this treatment with a reported overall mortality rate after treatment of 40% (167). Resistance to treatment, beyond worsening clinical signs, can be assessed by monitoring the level of anti-MDA5 Abs, ferritin or IL-18 which tend to stabilize or increase in patients refractory to treatment (51, 91, 168, 169). Other studies evaluating the effectiveness of intensive IS therapy highlighted the importance of beginning the treatment early after the diagnosis to improve the prognosis (91, 170). In addition to being partially efficient, the intense IS bi- or tri-therapy is responsible for several adverse events, mostly infections and renal function alteration. Renal function alteration is mostly due to calcineurin inhibitors and lead to treatment interruption in most cases. Bacterial, viral and fungal infections are reported, the most frequent being the reactivation of cytomegalovirus (29, 46, 171, 172). *Pneumocystis jirovecii* pneumonia is also encountered (29, 173, 174). Whether the infections are the result of IS treatment or of the disease itself is not clear, although DM was associated with a higher risk of developing pneumocystosis in a French retrospective study (175).

Due to the limited effectiveness and frequent adverse events of these treatments, several alternative therapy strategies are currently evaluated to treat refractory cases. The rare studies evaluating alternative therapies, such as the use of vasodilators, inhibitors of the IFN axis, or therapies that target the humoral immune response, illustrate the importance of the dysregulated pathways discussed previously in the pathogenesis of the disease.

Data regarding the improvement of cutaneous lesions after intensive IS treatment are scarce but some case reports of

cutaneous lesions refractory to IS drugs have been published (48, 137, 176). As the vascular injury seems to have a central role in the development of skin lesions, drugs such as sildenafil, a vasodilator, or bosentan, an endothelin-receptor inhibitor, might be added to the IS treatment to treat the skin ulcerations (32, 133–135).

Considering the importance of the IFN-I signaling pathway in anti-MDA5 DM, inhibitors of Janus Kinases (JAK) appear to be a promising treatment for anti-MDA5 DM. Several studies reported the efficacy of tofacitinib and ruxolitinib in patients who experienced a relapse or were refractory to standard treatment (50, 176–179). JAK/STAT signaling pathway is activated by IFNs leading to the transcription of IFN-stimulated genes (ISGs), including MDA5. Tofacitinib and ruxolitinib inhibit this pathway, decreasing MDA5 expression and activation. Furthermore, its effectiveness on refractory forms of ILD reinforces the potential role of IL-21 in the pathophysiology of ILD as JAK-pathway is required for IL-21/IL-21R signaling (119, 180, 181). JAK inhibitors could then be used to treat or prevent severe forms of ILD. Tofacitinib seems well tolerated in most patients but one study reported cytomegalovirus reactivation in 100% of patients, varicella-zoster virus reactivation (60%) and bacterial respiratory infections (80%) (178). Whether these adverse events were caused by the disease or the therapy was not determined. The question remains whether JAK-inhibitors should be administered in patients refractory to classic IS therapy or be initiated at diagnosis to avoid worsening of the disease. A clinical trial including 50 Japanese patients with anti-MDA5 CADM-associated ILD diagnosed for less than 3 months reported a 6-month survival significantly higher (100%) in the group of patients ($n=18$) who received a glucocorticoid combined with tofacitinib than in the group ($n=32$) who received conventional immunosuppressive treatment (6-month survival of 78%) (182). JAK inhibitors could therefore have a prominent place in the first-line treatment of anti-MDA5 DM.

A pathogenic role of anti-MDA5 Abs could motivate the use of plasmapheresis, IV immunoglobulins and rituximab. Although evidence is limited to small case series, these therapies seem efficacious (137–141, 183–187), apart from one case report of a patient whose condition worsened after plasma exchange probably due to transfusion-related acute lung injury (188). However, their effectiveness in large cohorts of patients as well as data concerning long-term remission remain to be evaluated.

CONCLUSION

Anti-MDA5 DM is a systemic autoimmune disease that can be divided into 3 clinical subgroups, with different prognosis, linked to the incidence of RP-ILD which is influenced by the ethnic origin of the patients. Recent publications suggest a central role for IFN-I mediated vasculopathy. It might be responsible for both the pulmonary and the cutaneous lesions, through the secretion of endothelial-derived substances inducing the

recruitment and the activation of immune cells, *in fine* responsible for the lesions observed in anti-MDA5 DM. Anti-MDA5 Abs might also contribute to the pathogenesis by altering the IFN-I pathway. Of course, further studies need to be conducted to confirm these assumptions. Elucidating the precise role of anti-MDA5 Abs associated to the disease will constitute a crucial step in the understanding of the pathogenesis. An improved knowledge of the pathogenesis of the disease will also undoubtedly pave the way for the development of more effective therapeutic strategies.

REFERENCES

- Mariampillai K, Granger B, Amelin D, Guiguet M, Hachulla E, Maurier F, et al. Development of a New Classification System for Idiopathic Inflammatory Myopathies Based on Clinical Manifestations and Myositis-Specific Autoantibodies. *JAMA Neurol* (2018) 75:1528–37. doi: 10.1001/jamaneurol.2018.2598
- Allenbach Y, Uzunhan Y, Toquet S, Leroux G, Gallay L, Marquet A, et al. Different Phenotypes in Dermatomyositis Associated With Anti-MDA5 Antibody: Study of 121 Cases. *Neurology* (2020) 95:e70–8. doi: 10.1212/WNL.00000000000009727
- Betteridge Z, Tansley S, Shaddick G, Chinoy H, Cooper RG, New RP, et al. Frequency, Mutual Exclusivity and Clinical Associations of Myositis Autoantibodies in a Combined European Cohort of Idiopathic Inflammatory Myopathy Patients. *J Autoimmun* (2019) 101:48–55. doi: 10.1016/j.jaut.2019.04.001
- Sato S, Hirakata M, Kuwana M, Suwa A, Inada S, Mimori T, et al. Autoantibodies to a 140-Kd Polypeptide, CADM-140, in Japanese Patients With Clinically Amyopathic Dermatomyositis. *Arthritis Rheum* (2005) 52:1571–6. doi: 10.1002/art.21023
- Fujikawa K, Kawakami A, Kaji K, Fujimoto M, Kawashiri S, Iwamoto N, et al. Association of Distinct Clinical Subsets With Myositis-Specific Autoantibodies Towards Anti-155/140-kDa Polypeptides, Anti-140-kDa Polypeptides, and Anti-Aminoacyl tRNA Synthetases in Japanese Patients With Dermatomyositis: A Single-Centre, Cross-Sectional Study. *Scand J Rheumatol* (2009) 38:263–7. doi: 10.1080/03009740802687455
- Gono T, Kawaguchi Y, Satoh T, Kuwana M, Katsumata Y, Takagi K, et al. Clinical Manifestation and Prognostic Factor in Anti-Melanoma Differentiation-Associated Gene 5 Antibody-Associated Interstitial Lung Disease as a Complication of Dermatomyositis. *Rheumatology* (2010) 49:1713–9. doi: 10.1093/rheumatology/keq149
- Nakashima R, Imura Y, Kobayashi S, Yukawa N, Yoshifuji H, Nojima T, et al. The RIG-I-Like Receptor IFIH1/MDA5 Is a Dermatomyositis-Specific Autoantigen Identified by the Anti-CADM-140 Antibody. *Rheumatology* (2010) 49:433–40. doi: 10.1093/rheumatology/kep375
- Ikeda N, Takahashi K, Yamaguchi Y, Inasaka M, Kuwana M, Ikezawa Z. Analysis of Dermatomyositis-Specific Autoantibodies and Clinical Characteristics in Japanese Patients. *J Dermatol* (2011) 38:973–9. doi: 10.1111/j.1346-8138.2011.01262.x
- Hamaguchi Y, Kuwana M, Hoshino K, Hasegawa M, Kaji K, Matsushita T, et al. Clinical Correlations With Dermatomyositis-Specific Autoantibodies in Adult Japanese Patients With Dermatomyositis: A Multicenter Cross-Sectional Study. *Arch Dermatol* (2011) 147:391–8. doi: 10.1001/archdermatol.2011.52
- Koga T, Fujikawa K, Horai Y, Okada A, Kawashiri SY, Iwamoto N, et al. The Diagnostic Utility of Anti-Melanoma Differentiation-Associated Gene 5 Antibody Testing for Predicting the Prognosis of Japanese Patients With DM. *Rheumatology* (2012) 51:1278–84. doi: 10.1093/rheumatology/ker518
- Chen F, Wang D, Shu X, Nakashima R, Wang G. Anti-MDA5 Antibody Is Associated With A/SIP and Decreased T Cells in Peripheral Blood and Predicts Poor Prognosis of ILD in Chinese Patients With Dermatomyositis. *Rheumatol Int* (2012) 32:3909–15. doi: 10.1007/s00296-011-2323-y

AUTHOR CONTRIBUTIONS

AN and FC: writing and figures. FC and NF: concept and proof reading. All authors contributed to the article and approved the submitted version.

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- Cao H, Pan M, Kang Y, Xia Q, Li X, Zhao X, et al. Clinical Manifestations of Dermatomyositis and Clinically Amyopathic Dermatomyositis Patients With Positive Expression of Anti-Melanoma Differentiation-Associated Gene 5 Antibody. *Arthritis Care Res* (2012) 64:1602–10. doi: 10.1002/acr.21728
- Chen Z, Cao M, Plana MN, Liang J, Cai H, Kuwana M, et al. Utility of Anti-Melanoma Differentiation-Associated Gene 5 Antibody Measurement in Identifying Patients With Dermatomyositis and a High Risk for Developing Rapidly Progressive Interstitial Lung Disease: A Review of the Literature and a Meta-Analysis. *Arthritis Care Res* (2013) 65:1316–24. doi: 10.1002/acr.21985
- Li Y, Li Y, Wu J, Miao M, Gao X, Cai W, et al. Predictors of Poor Outcome of Anti-MDA5-Associated Rapidly Progressive Interstitial Lung Disease in a Chinese Cohort With Dermatomyositis. *J Immunol Res* (2020) 2020:1–8. doi: 10.1155/2020/2024869
- Fiorentino D, Chung L, Zwerner J, Rosen A, Casciola-Rosen L. The Mucocutaneous and Systemic Phenotype of Dermatomyositis Patients With Antibodies to MDA5 (CADM-140). *J Am Acad Dermatol* (2011) 65:25–34. doi: 10.1016/j.jaad.2010.09.016
- Hall JC, Casciola-Rosen L, Samedy LA, Werner J, Owoyemi K, Danoff SK, et al. Anti-Melanoma Differentiation-Associated Protein 5-Associated Dermatomyositis: Expanding the Clinical Spectrum. *Arthritis Care Res* (2013) 65:1307–15. doi: 10.1002/acr.21992
- Labrador-Horrillo M, Martinez MA, Selva-O’Callaghan A, Trallero-Araguas E, Balada E, Vilardell-Tarres M, et al. Anti-MDA5 Antibodies in a Large Mediterranean Population of Adults With Dermatomyositis. *J Immunol Res* (2014) 2014:1–8. doi: 10.1155/2014/290797
- Ceribelli A, Fredi M, Taraborelli M, Cavazzana I, Tincani A, Selmi C, et al. Prevalence and Clinical Significance of Anti-MDA5 Antibodies in European Patients With Polymyositis/Dermatomyositis. *Clin Exp Rheumatol* (2014) 32(6):891–7.
- Borges IBP, Silva MG, Shinjo SK. Prevalence and Reactivity of Anti-Melanoma Differentiation-Associated Gene 5 (Anti-MDA-5) Autoantibody in Brazilian Patients With Dermatomyositis. *Bras Dermatol* (2018) 93:517–23. doi: 10.1590/abd1806-4841.20186803
- Sontheimer RD. MDA5 Autoantibody—Another Indicator of Clinical Diversity in Dermatomyositis. *Ann Transl Med* (2017) 5(7):160. doi: 10.21037/atm.2017.03.94
- Jiang Y, Liu Y, Zhao Y, Zheng Y, Yu M, Deng J, et al. Mitochondrial Morphology and MAVS-IFN1 Signaling Pathway in Muscles of Anti-MDA5 Dermatomyositis. *Ann Clin Transl Neurol* (2021) 8:677–86. doi: 10.1002/acn3.51311
- Allenbach Y, Leroux G, Suárez-Calvet X, Preusse C, Gallardo E, Hervier B, et al. Dermatomyositis With or Without Anti-Melanoma Differentiation-Associated Gene 5 Antibodies. *Am J Pathol* (2016) 186:691–700. doi: 10.1016/j.ajpath.2015.11.010
- Cabezas-Rodríguez I, Morante-Bolado I, Brandy-García A, Queiro-Silva R, Mozo L, Ballina-García FJ. Anti-MDA5 Dermatomyositis Mimicking Psoriatic Arthritis. *Reumatol Clínica* (2018) 14:224–6. doi: 10.1016/j.reuma.2016.10.010
- Haroon M, Devlin J. Gottron’s Papule in Amyopathic Dermatomyositis Mimicking Psoriasis. *Clin Rheumatol* (2009) 28:1245–6. doi: 10.1007/s10067-009-1244-6

25. Moghadam-Kia S, Oddis CV, Sato S, Kuwana M, Aggarwal R. Antimelanoma Differentiation-Associated Gene 5 Antibody: Expanding the Clinical Spectrum in North American Patients With Dermatomyositis. *J Rheumatol* (2017) 44:319–25. doi: 10.3899/jrheum.160682
26. Hoa S, Troyanov Y, Fritzler M, Targoff I, Chartrand S, Mansour A, et al. Describing and Expanding the Clinical Phenotype of Anti-MDA5-Associated Rapidly Progressive Interstitial Lung Disease: Case Series of Nine Canadian Patients and Literature Review. *Scand J Rheumatol* (2018) 47:210–24. doi: 10.1080/03009742.2017.1334814
27. Gerfaud-Valentin M, Ahmad K, Piegay F, Fabien N, Raphanel B, Cordier J-F, et al. Pneumopathie Infiltrante Diffuse Associée À Une Dermatomyosite Amyopathique Avec Auto-Anticorps Anti-MDA5. *Rev Mal Respir* (2014) 31:849–53. doi: 10.1016/j.rmr.2014.02.010
28. Charrow A, Vleugels RA. Cutaneous Ulcerations in Anti-MDA5 Dermatomyositis. *N Engl J Med* (2019) 381:465. doi: 10.1056/NEJM1816147
29. Yamaguchi K, Yamaguchi A, Onuki Y, Itai M, Kashiwagi C, Takehara K, et al. Clinical Features of Dermatomyositis Associated With Anti-MDA5 Antibodies by Age. *Mod Rheumatol* (2020) 31:177–85. doi: 10.1080/14397595.2020.1740400
30. Narang NS, Casciola-Rosen L, Li S, Chung L, Fiorentino DF. Cutaneous Ulceration in Dermatomyositis: Association With Anti-Melanoma Differentiation-Associated Gene 5 Antibodies and Interstitial Lung Disease. *Arthritis Care Res* (2015) 67:667–72. doi: 10.1002/acr.22498
31. So H, Ip RW-K, Wong VT-L, Yip RM-L. Analysis of Anti-Melanoma Differentiation-Associated Gene 5 Antibody in Hong Kong Chinese Patients With Idiopathic Inflammatory Myopathies: Diagnostic Utility and Clinical Correlations. *Int J Rheum Dis* (2018) 21:1076–81. doi: 10.1111/1756-185X.13268
32. Xu A, Lv X, Du F, Dai M. Cutaneous Ulceration and Digital Gangrene in an Anti-MDA5-Positive Overlap Myositis. *Rheumatology* (2021), keab284. doi: 10.1093/rheumatology/keab284
33. Barailler H, Charles S, Goutorbe F, Devaux S. An Unusual Anti-Melanoma Differentiation-Associated Protein 5 Amyopathic Dermatomyositis. *JAAD Case Rep* (2019) 5:383–5. doi: 10.1016/j.jidcr.2019.02.019
34. Okiyama N, Inoue S, Saito A, Nakamura Y, Ishitsuka Y, Fujisawa Y, et al. Antihelix/helix Violaceous Macules in Japanese Patients With Anti-Melanoma Differentiation-Associated Protein 5 (MDA 5) Antibody-Associated Dermatomyositis. *Br J Dermatol* (2019) 180:1226–7. doi: 10.1111/bjd.17431
35. Motegi S, Sekiguchi A, Toki S, Kishi C, Endo Y, Yasuda M, et al. Clinical Features and Poor Prognostic Factors of Anti-Melanoma Differentiation-Associated Gene 5 Antibody-Positive Dermatomyositis With Rapid Progressive Interstitial Lung Disease. *Eur J Dermatol* (2019) 29:511–7. doi: 10.1684/ejd.2019.3634
36. Intapiboon P, Siripaitoon B. Erythematous Auricular Papules in the Fatal Cases of Anti-MDA5 Antibody-Positive Interstitial Lung Disease. *Respir Med Case Rep* (2020) 31:101299. doi: 10.1016/j.rmcr.2020.101299
37. Molina-Ruiz AM, Romero F, Carrasco L, Feltes F, Haro R, Requena L. Amyopathic Dermatomyositis Presenting as a Flagellated Skin Eruption With Positive MDA5 Antibodies and Thyroid Cancer: A Real Association? *Clin Exp Dermatol* (2015) 40:887–90. doi: 10.1111/ced.12674
38. Watanabe T, Takizawa N, Nagasaka T, Nakamura Y, Ikai H, Yamamoto M, et al. Fatal and Extensive Multiorgan Hemorrhages in Anti-Melanoma Differentiation-Associated Gene 5 Antibody-Positive Dermatomyositis: An Autopsy Case Report. *Med (Baltimore)* (2020) 99:e18600. doi: 10.1097/MD.000000000000186004
39. González-Moreno J, Raya-Cruz M, Losada-Lopez I, Cacheda AP, Oliver C, Colom B. Rapidly Progressive Interstitial Lung Disease Due to Anti-MDA5 Antibodies Without Skin Involvement: A Case Report and Literature Review. *Rheumatol Int* (2018) 38:1293–96. doi: 10.1007/s00296-018-3991-7
40. Rathore U, Haldule S, Gupta L. Psoriasiform Rashes as the First Manifestation of Anti-MDA5 Associated Myositis. *Rheumatology* (2020) 60(7):3483. doi: 10.1093/rheumatology/keaa821
41. Lega J-C, Fabien N, Reynaud Q, Durieu I, Durupt S, Dutertre M, et al. The Clinical Phenotype Associated With Myositis-Specific and Associated Autoantibodies: A Meta-Analysis Revisiting the So-Called Antisynthetase Syndrome. *Autoimmun Rev* (2014) 13:883–91. doi: 10.1016/j.autrev.2014.03.004
42. Tanizawa K, Handa T, Nakashima R, Kubo T, Hosono Y, Watanabe K, et al. HRCT Features of Interstitial Lung Disease in Dermatomyositis With Anti-CADM-140 Antibody. *Respir Med* (2011) 105:1380–7. doi: 10.1016/j.rmed.2011.05.006
43. Sakamoto N, Ishimoto H, Nakashima S, Yura H, Miyamura T, Okuno D, et al. Clinical Features of Anti-MDA5 Antibody-Positive Rapidly Progressive Interstitial Lung Disease Without Signs of Dermatomyositis. *Intern Med* (2019) 58:837–41. doi: 10.2169/internalmedicine.1516-18
44. Ortiz-Santamaria V, Babot A, Ferrer C. Anti-MDA5-Positive Dermatomyositis: An Emerging Entity With a Variable Clinical Presentation. *Scand J Rheumatol* (2017) 46:509–11. doi: 10.1080/03009742.2017.1340512
45. Li Y, Gao X, Li Y, Jia X, Zhang X, Xu Y, et al. Predictors and Mortality of Rapidly Progressive Interstitial Lung Disease in Patients With Idiopathic Inflammatory Myopathy: A Series of 474 Patients. *Front Med* (2020) 7:363. doi: 10.3389/fmed.2020.00363
46. Yamaguchi K, Yamaguchi A, Kashiwagi C, Sawada Y, Taguchi K, Umetsu K, et al. Differential Clinical Features of Patients With Clinically Amyopathic Dermatomyositis Who Have Circulating Anti-MDA5 Autoantibodies With or Without Myositis-Associated Autoantibodies. *Respir Med* (2018) 140:1–5. doi: 10.1016/j.rmed.2018.05.010
47. Endo Y, Koga T, Ishida M, Fujita Y, Tsuji S, Takatani A, et al. Recurrence of Anti-MDA5 Antibody-Positive Clinically Amyopathic Dermatomyositis After Long-Term Remission: A Case Report. *Med (Baltimore)* (2018) 97:e11024. doi: 10.1097/MD.00000000000011024
48. Ishikawa Y, Kasuya T, Fujiwara M, Kita Y. Tofacitinib for Recurrence of Antimelanoma Differentiation-Associated Gene 5 Antibody-Positive Clinically Amyopathic Dermatomyositis After Remission: A Case Report. *Med (Baltimore)* (2020) 99:e21943. doi: 10.1097/MD.00000000000021943
49. Betteridge Z, McHugh N. Myositis-Specific Autoantibodies: An Important Tool to Support Diagnosis of Myositis. *J Intern Med* (2016) 280:8–23. doi: 10.1111/joim.12451
50. Jalles C, Deroux A, Tardieu M, Lugosi M, Viel S, Benveniste O, et al. Dermatomyosite À Anticorps Anti-MDA5 Sévère Associée À Un Cancer Et Contrôlée Par Inhibiteur De JAK. *Rev Méd Interne* (2020) 41:421–4. doi: 10.1016/j.revmed.2020.02.015
51. Gono T, Sato S, Kawaguchi Y, Kuwana M, Hanaoka M, Katsumata Y, et al. Anti-MDA5 Antibody, Ferritin and IL-18 Are Useful for the Evaluation of Response to Treatment in Interstitial Lung Disease With Anti-MDA5 Antibody-Positive Dermatomyositis. *Rheumatology* (2012) 51:1563–70. doi: 10.1093/rheumatology/kes102
52. Fujiki Y, Kotani T, Isoda K, Ishida T, Shoda T, Yoshida S, et al. Evaluation of Clinical Prognostic Factors for Interstitial Pneumonia in Anti-MDA5 Antibody-Positive Dermatomyositis Patients. *Mod Rheumatol* (2018) 28:133–40. doi: 10.1080/14397595.2017.1318468
53. Nagashima T, Kamata Y, Iwamoto M, Okazaki H, Fukushima N, Minota S. Liver Dysfunction in Anti-Melanoma Differentiation-Associated Gene 5 Antibody-Positive Patients With Dermatomyositis. *Rheumatol Int* (2019) 39:901–9. doi: 10.1007/s00296-019-04255-2
54. Huang W, Ren F, Luo L, Zhou J, Huang D, Pan Z, et al. The Characteristics of Lymphocytes in Patients Positive for Anti-MDA5 Antibodies in Interstitial Lung Disease. *Rheumatology* (2020) 59:3886–91. doi: 10.1093/rheumatology/keaa266
55. Huang K, Vinik O, Shojania K, Yeung J, Shupak R, Nimmo M, et al. Clinical Spectrum and Therapeutics in Canadian Patients With Anti-Melanoma Differentiation-Associated Gene 5 (MDA5)-Positive Dermatomyositis: A Case-Based Review. *Rheumatol Int* (2019) 39:1971–81. doi: 10.1007/s00296-019-04398-2
56. Sabbagh S, Pinal-Fernandez I, Kishi T, Targoff IN, Miller FW, Rider LG, et al. Anti-Ro52 Autoantibodies Are Associated With Interstitial Lung Disease and More Severe Disease in Patients With Juvenile Myositis. *Ann Rheum Dis* (2019) 78:988–95. doi: 10.1136/annrheumdis-2018-215004
57. Toquet S, Uzunhan Y, Leroux G, Bonnotte B, Gallay L, Limal N, et al. Facteurs Pronostiques De La Dermatomyosite À Anticorps Anti-MDA5: Sexe Féminin Et Atteinte Articulaire De Bon Pronostic. *Rev Méd Interne* (2018) 39:A114. doi: 10.1016/j.revmed.2018.03.364
58. Tansley SL, Betteridge ZE, Gunawardena H, Jacques TS, Owens CM, Pilkington C, et al. Anti-MDA5 Autoantibodies in Juvenile

- Dermatomyositis Identify a Distinct Clinical Phenotype: A Prospective Cohort Study. *Arthritis Res Ther* (2014) 16:R138. doi: 10.1186/ar4600
59. Tansley SL, Simou S, Shaddick G, Betteridge ZE, Almeida B, Gunawardena H, et al. Autoantibodies in Juvenile-Onset Myositis: Their Diagnostic Value and Associated Clinical Phenotype in a Large UK Cohort. *J Autoimmun* (2017) 84:55–64. doi: 10.1016/j.jaut.2017.06.007
 60. Mamyrova G, Kishi T, Targoff IN, Ehrlich A, Curiel RV, Rider LG, et al. Features Distinguishing Clinically Amyopathic Juvenile Dermatomyositis From Juvenile Dermatomyositis. *Rheumatology* (2018) 57:1956–63. doi: 10.1093/rheumatology/key190
 61. Mamyrova G, Kishi T, Shi M, Targoff IN, Huber AM, Curiel RV, et al. Anti-MDA5 Autoantibodies Associated With Juvenile Dermatomyositis Constitute a Distinct Phenotype in North America. *Rheumatology* (2020) 60(4):1839–49. doi: 10.1093/rheumatology/keaa429
 62. Deakin CT, Yasin SA, Simou S, Arnold KA, Tansley SL, Betteridge ZE, et al. Muscle Biopsy Findings in Combination With Myositis-Specific Autoantibodies Aid Prediction of Outcomes in Juvenile Dermatomyositis. *Arthritis Rheumatol* (2016) 68:2806–16. doi: 10.1002/art.39753
 63. Kobayashi I, Okura Y, Yamada M, Kawamura N, Kuwana M, Ariga T. Anti-Melanoma Differentiation-Associated Gene 5 Antibody Is a Diagnostic and Predictive Marker for Interstitial Lung Diseases Associated With Juvenile Dermatomyositis. *J Pediatr* (2011) 158:675–7. doi: 10.1016/j.jpeds.2010.11.033
 64. Ueki M, Kobayashi I, Takezaki S, Tozawa Y, Okura Y, Yamada M, et al. Myositis-Specific Autoantibodies in Japanese Patients With Juvenile Idiopathic Inflammatory Myopathies. *Mod Rheumatol* (2019) 29:351–6. doi: 10.1080/14397595.2018.1452353
 65. Iwata N, Nakaseko H, Kohagura T, Yasuoka R, Abe N, Kawabe S, et al. Clinical Subsets of Juvenile Dermatomyositis Classified by Myositis-Specific Autoantibodies: Experience at a Single Center in Japan. *Mod Rheumatol* (2019) 29:802–7. doi: 10.1080/14397595.2018.1511025
 66. Miyamae T, Hara T, Hanaya A, Tani Y, Kishi T, Yamanaka H. Effect of Adalimumab on Refractory Arthritis in Juvenile Idiopathic Inflammatory Myopathy With Anti-MDA5 Autoantibody. *Case Rep Rheumatol* (2018) 2018:1–5. doi: 10.1155/2018/2164312
 67. Kobayashi N, Takezaki S, Kobayashi I, Iwata N, Mori M, Nagai K, et al. Clinical and Laboratory Features of Fatal Rapidly Progressive Interstitial Lung Disease Associated With Juvenile Dermatomyositis. *Rheumatology* (2015) 54:784–91. doi: 10.1093/rheumatology/keu385
 68. Yamasaki Y, Kobayashi N, Akioka S, Yamazaki K, Takezaki S, Nakaseko H, et al. Clinical Impact of Myositis-Specific Autoantibodies on Long-Term Prognosis of Juvenile Idiopathic Inflammatory Myopathies: Multicentre Study. *Rheumatology* (2021) 60(10):4821–31. doi: 10.1093/rheumatology/keab108
 69. Sato S, Hoshino K, Satoh T, Fujita T, Kawakami Y, Fujita T, et al. RNA Helicase Encoded by Melanoma Differentiation-Associated Gene 5 Is a Major Autoantigen in Patients With Clinically Amyopathic Dermatomyositis: Association With Rapidly Progressive Interstitial Lung Disease. *Arthritis Rheum* (2009) 60:2193–200. doi: 10.1002/art.24621
 70. Kang DC, Gopalkrishnan RV, Wu Q, Jankowsky E, Pyle AM, Fisher PB. Mda-5: An Interferon-Inducible Putative RNA Helicase With Double-Stranded RNA-Dependent ATPase Activity and Melanoma Growth-Suppressive Properties. *Proc Natl Acad Sci* (2002) 99:637–42. doi: 10.1073/pnas.022637199
 71. Kato H, Takeuchi O, Sato S, Yoneyama M, Yamamoto M, Matsui K, et al. Differential Roles of MDA5 and RIG-I Helicases in the Recognition of RNA Viruses. *Nature* (2006) 441:101–5. doi: 10.1038/nature04734
 72. Kato H, Takeuchi O, Mikamo-Satoh E, Hirai R, Kawai T, Matsushita K, et al. Length-Dependent Recognition of Double-Stranded Ribonucleic Acids by Retinoic Acid-Inducible Gene-I and Melanoma Differentiation-Associated Gene 5. *J Exp Med* (2008) 205:1601–10. doi: 10.1084/jem.20080091
 73. Loo Y-M, Gale M. Immune Signaling by RIG-I-Like Receptors. *Immunity* (2011) 34:680–92. doi: 10.1016/j.immuni.2011.05.003
 74. Rehwinkel J, Gack MU. RIG-I-Like Receptors: Their Regulation and Roles in RNA Sensing. *Nat Rev Immunol* (2020) 20:537–51. doi: 10.1038/s41577-020-0288-3
 75. Feng Q, Hato SV, Langereis MA, Zoll J, Virgen-Slane R, Peisley A, et al. MDA5 Detects the Double-Stranded RNA Replicative Form in Picornavirus-Infected Cells. *Cell Rep* (2012) 2:1187–96. doi: 10.1016/j.celrep.2012.10.005
 76. Linder A, Hornung V. Mitochondrial dsRNA: A New DAMP for MDA5. *Dev Cell* (2018) 46:530–32. doi: 10.1016/j.devcel.2018.08.019
 77. Dhir A, Dhir S, Borowski LS, Jimenez L, Teitell M, Rötig A, et al. Mitochondrial Double-Stranded RNA Triggers Antiviral Signalling in Humans. *Nature* (2018) 560:238–42. doi: 10.1038/s41586-018-0363-0
 78. The Human Protein Atlas. *Ifih1*. Available at: <https://www.proteinatlas.org/ENSG00000115267-IFIH1/tissue> (Accessed August 12, 2021).
 79. Zahn S, Barchet W, Rehkämper C, Hornung T, Bieber T, Tüting T, et al. Enhanced Skin Expression of Melanoma Differentiation-Associated Gene 5 (MDA5) in Dermatomyositis and Related Autoimmune Diseases. *J Am Acad Dermatol* (2011) 64:988–9. doi: 10.1016/j.jaad.2010.08.004
 80. Berger M, Hsieh C-Y, Bakele M, Marcos V, Rieber N, Kormann M, et al. Neutrophils Express Distinct RNA Receptors in a Non-Canonical Way. *J Biol Chem* (2012) 287:19409–17. doi: 10.1074/jbc.M112.35357
 81. Hoshino K, Muro Y, Sugiura K, Tomita Y, Nakashima R, Mimori T. Anti-MDA5 and Anti-TIF1- γ Antibodies Have Clinical Significance for Patients With Dermatomyositis. *Rheumatology* (2010) 49:1726–33. doi: 10.1093/rheumatology/keq153
 82. Benveniste O, Stenzel W, Allenbach Y. Advances in Serological Diagnostics of Inflammatory Myopathies. *Curr Opin Neurol* (2016) 29:662–73. doi: 10.1097/WCO.0000000000000376
 83. Damoiseaux J, Vulsteke J-B, Tseng C-W, Platteel ACM, Piette Y, Showman O, et al. Autoantibodies in Idiopathic Inflammatory Myopathies: Clinical Associations and Laboratory Evaluation by Mono- and Multispecific Immunoassays. *Autoimmun Rev* (2019) 18:293–305. doi: 10.1016/j.jautrev.2018.10.004
 84. Fabien N. Anticorps Des Myopathies Autoimmunes: Actualités Des Techniques De Détection Et Intérêt Clinique. *Rev Francoph Lab* (2016) 2016:41–6. doi: 10.1016/S1773-035X(16)30243-X
 85. Pacot L, Pouchot J, De Prost N, Senant M, Tartour E, Le Pimpec-Barthes F, et al. Interstitial Lung Disease-Complicated Anti-MDA5 Antibody in Clinically Amyopathic Dermatomyositis Patients: Report of Two Cases With Distinct Clinical Features. *Front Med* (2020) 7:77. doi: 10.3389/fmed.2020.00077
 86. Tansley SL, Li D, Betteridge ZE, McHugh NJ. The Reliability of Immunoassays to Detect Autoantibodies in Patients With Myositis is Dependent on Autoantibody Specificity. *Rheumatology* (2020) 59:2109–14. doi: 10.1093/rheumatology/keaa021
 87. Cavazzana I, Richards M, Bentow C, Seaman A, Fredi M, Giudizi MG, et al. Evaluation of a Novel Particle-Based Assay for Detection of Autoantibodies in Idiopathic Inflammatory Myopathies. *J Immunol Methods* (2019) 474:112661. doi: 10.1016/j.jim.2019.112661
 88. Mahler M, Betteridge Z, Bentow C, Richards M, Seaman A, Chinoy H, et al. Comparison of Three Immunoassays for the Detection of Myositis Specific Antibodies. *Front Immunol* (2019) 10:848. doi: 10.3389/fimmu.2019.00848
 89. Sato S, Murakami A, Kuwajima A, Takehara K, Mimori T, Kawakami A, et al. Clinical Utility of an Enzyme-Linked Immunosorbent Assay for Detecting Anti-Melanoma Differentiation-Associated Gene 5 Autoantibodies. *PLoS One* (2016) 11:e0154285. doi: 10.1371/journal.pone.0154285
 90. Sakamoto S, Okamoto M, Kaieda S, Fujimoto K, Nagata S, Tominaga M, et al. Low Positive Titer of Anti-Melanoma Differentiation-Associated Gene 5 Antibody Is Not Associated With a Poor Long-Term Outcome of Interstitial Lung Disease in Patients With Dermatomyositis. *Respir Investig* (2018) 56:464–72. doi: 10.1016/j.resinv.2018.07.007
 91. Muro Y, Sugiura K, Hoshino K, Akiyama M. Disappearance of Anti-MDA-5 Autoantibodies in Clinically Amyopathic DM/interstitial Lung Disease During Disease Remission. *Rheumatology* (2012) 51:800–4. doi: 10.1093/rheumatology/ker408
 92. Matsushita T, Mizumaki K, Kano M, Yagi N, Tennichi M, Takeuchi A, et al. Antimelanoma Differentiation-Associated Protein 5 Antibody Level Is a Novel Tool for Monitoring Disease Activity in Rapidly Progressive Interstitial Lung Disease With Dermatomyositis. *Br J Dermatol* (2017) 176:395–402. doi: 10.1111/bjd.14882
 93. Hosono Y, Nakashima R, Serada S, Murakami K, Imura Y, Yoshifuji H, et al. Splicing Factor Proline/Glutamine-Rich Is a Novel Autoantigen of Dermatomyositis and Associated With Anti-Melanoma Differentiation-Associated Gene 5 Antibody. *J Autoimmun* (2017) 77:116–22. doi: 10.1016/j.jaut.2016.11.006

94. Greco-Stewart VS, Thibault CS-L, Pelchat M. Binding of the Polypyrimidine Tract-Binding Protein-Associated Splicing Factor (PSF) to the Hepatitis Delta Virus RNA. *Virology* (2006) 356:35–44. doi: 10.1016/j.virol.2006.06.040
95. Landeras-Bueno S, Jorba N, Pérez-Cidoncha M, Ortín J. The Splicing Factor Proline-Glutamine Rich (SFPQ/PSF) Is Involved in Influenza Virus Transcription. *PLoS Pathog* (2011) 7:e1002397. doi: 10.1371/journal.ppat.1002397
96. Chen Z, Wang Y, Kuwana M, Xu X, Hu W, Feng X, et al. HLA-DRB1 Alleles as Genetic Risk Factors for the Development of Anti-MDA5 Antibodies in Patients With Dermatomyositis. *J Rheumatol* (2017) 44:1389–93. doi: 10.3899/jrheum.170165
97. Lin J, Zhang Y, Peng Q, Yang H, Shi J, Gu M, et al. Genetic Association of HLA-DRB1 Multiple Polymorphisms With Dermatomyositis in Chinese Population. *HLA* (2017) 90:354–9. doi: 10.1111/tan.13171
98. Gono T, Kawaguchi Y, Kuwana M, Sugiura T, Furuya T, Takagi K, et al. Brief Report: Association of HLA-DRB1*0101/*0405 With Susceptibility to Anti-Melanoma Differentiation-Associated Gene 5 Antibody-Positive Dermatomyositis in the Japanese Population. *Arthritis Rheum* (2012) 64:3736–40. doi: 10.1002/art.34657
99. Rothwell S, Chinoy H, Lamb JA, Miller FW, Rider LG, Wedderburn LR, et al. Focused HLA Analysis in Caucasians With Myositis Identifies Significant Associations With Autoantibody Subgroups. *Ann Rheum Dis* (2019) 78:996–1002. doi: 10.1136/annrheumdis-2019-215046
100. Kochi Y, Kamatani Y, Kondo Y, Suzuki A, Kawakami E, Hiwa R, et al. Splicing Variant of *WDFY4* Augments MDA5 Signaling and the Risk of Clinically Amyopathic Dermatomyositis. *Ann Rheum Dis* (2018) 77:602–11. doi: 10.1136/annrheumdis-2017-212149
101. Theisen DJ, Davidson JT, Briseño CG, Gargaro M, Lauron EJ, Wang Q, et al. *WDFY4* Is Required for Cross-Presentation in Response to Viral and Tumor Antigens. *Science* (2018) 362:694–9. doi: 10.1126/science.aat5030
102. Toquet S, Granger B, Uzunhan Y, Mariampillai K, Nunes H, Benveniste O, et al. The Seasonality of Dermatomyositis Associated With Anti-MDA5 Antibody: An Argument for a Respiratory Viral Trigger. *Autoimmun Rev* (2021) 20:102788. doi: 10.1016/j.autrev.2021.102788
103. Nishina N, Sato S, Masui K, Gono T, Kuwana M. Seasonal and Residential Clustering at Disease Onset of Anti-MDA5-Associated Interstitial Lung Disease. *RMD Open* (2020) 6:e001202. doi: 10.1136/rmdopen-2020-001202
104. Muro Y, Sugiura K, Hoshino K, Akiyama M, Tamakoshi K. Epidemiologic Study of Clinically Amyopathic Dermatomyositis and Anti-Melanoma Differentiation-Associated Gene 5 Antibodies in Central Japan. *Arthritis Res Ther* (2011) 13:R214. doi: 10.1186/ar3547
105. Shakshouk H, Deschaine MA, Wetter DA, Drage LA, Ernste FC, Lehman JS. Clinical and Histopathological Features of Adult Patients With Dermatomyositis and MDA5 Autoantibody Seropositivity Status, as Determined by Commercial-Based Testing: A Retrospective, Single-Institution Comparative Cohort Study. *Clin Exp Dermatol* (2021), ced.14870. doi: 10.1111/ced.14870
106. He C, Chen J, Luo X, Yan B. Evaluation of Biomarkers Related to Endothelial Dysfunction: Proof of Vasculopathy in Anti-Melanoma Differentiation-Associated Gene 5 Dermatomyositis. *Clin Exp Rheumatol* (2021) 39:151–7.
107. Funauuchi M, Shimadsu H, Tamaki C, Yamagata T, Nozaki Y, Sugiyama M, et al. Role of Endothelial Damage in the Pathogenesis of Interstitial Pneumonitis in Patients With Polymyositis and Dermatomyositis. *J Rheumatol* (2006) 33(5):903–6.
108. Ono N, Kai K, Maruyama A, Sakai M, Sadanaga Y, Koarada S, et al. The Relationship Between Type I IFN and Vasculopathy in Anti-MDA5 Antibody-Positive Dermatomyositis Patients. *Rheumatology* (2019) 58:786–91. doi: 10.1093/rheumatology/key386
109. Zhang SH, Zhao Y, Xie QB, Jiang Y, Wu YK, Yan B. Aberrant Activation of the Type I Interferon System may Contribute to the Pathogenesis of Anti-Melanoma Differentiation-Associated Gene 5 Dermatomyositis. *Br J Dermatol* (2019) 180:1090–98. doi: 10.1111/bjd.16917
110. Horiike Y, Suzuki Y, Fujisawa T, Yasui H, Karayama M, Hozumi H, et al. Successful Classification of Macrophage-Mannose Receptor CD206 in Severity of Anti-MDA5 Antibody Positive Dermatomyositis Associated ILD. *Rheumatology* (2019) 58:2143–52. doi: 10.1093/rheumatology/kez185
111. Gono T, Miyake K, Kawaguchi Y, Kaneko H, Shinozaki M, Yamanaka H. Hyperferritinaemia and Macrophage Activation in a Patient With Interstitial Lung Disease With Clinically Amyopathic DM. *Rheumatology* (2012) 51:1336–8. doi: 10.1093/rheumatology/kes012
112. Zhang L, Wang Y, Wu G, Xiong W, Gu W, Wang C-Y. Macrophages: Friend or Foe in Idiopathic Pulmonary Fibrosis? *Respir Res* (2018) 19:170. doi: 10.1186/s12931-018-0864-2
113. Fujisawa T, Hozumi H, Yasui H, Suzuki Y, Karayama M, Furuhashi K, et al. Clinical Significance of Serum Chitotriosidase Level in Anti-MDA5 Antibody-positive Dermatomyositis-Associated Interstitial Lung Disease. *J Rheumatol* (2019) 46:935–42. doi: 10.3899/jrheum.180825
114. Kawasumi H, Katsumata Y, Nishino A, Hirahara S, Kawaguchi Y, Kuwana M, et al. Association of Serum Soluble CD163 With Polymyositis and Dermatomyositis, Especially in Anti-MDA5 Antibody-positive Cases. *J Rheumatol* (2018) 45:947–55. doi: 10.3899/jrheum.170997
115. Peng QL, Zhang YM, Liang L, Liu X, Ye LF, Yang HB, et al. A High Level of Serum Neopterin Is Associated With Rapidly Progressive Interstitial Lung Disease and Reduced Survival in Dermatomyositis. *Clin Exp Immunol* (2020) 199:314–25. doi: 10.1111/cei.13404
116. Takada T, Aoki A, Asakawa K, Sakagami T, Moriyama H, Narita I, et al. Serum Cytokine Profiles of Patients With Interstitial Lung Disease Associated With Anti-CADM-140/MDA5 Antibody Positive Amyopathic Dermatomyositis. *Respir Med* (2015) 109:1174–80. doi: 10.1016/j.rmed.2015.07.004
117. Nakano M, Fujii T, Hashimoto M, Yukawa N, Yoshifuji H, Ohmura K, et al. Type I Interferon Induces CX3CL1 (Fractalkine) and CCL5 (RANTES) Production in Human Pulmonary Vascular Endothelial Cells. *Clin Exp Immunol* (2012) 170:94–100. doi: 10.1111/j.1365-2249.2012.04638.x
118. Ishida Y, Kimura A, Nosaka M, Kuninaka Y, Hemmi H, Sasaki I, et al. Essential Involvement of the CX3CL1-CX3CR1 Axis in Bleomycin-Induced Pulmonary Fibrosis via Regulation of Fibrocyte and M2 Macrophage Migration. *Sci Rep* (2017) 7:16833. doi: 10.1038/s41598-017-17007-8
119. Wang K, Zhao J, Chen Z, Li T, Tan X, Zheng Y, et al. CD4+CXCR4+ T Cells as a Novel Prognostic Biomarker in Patients With Idiopathic Inflammatory Myopathy-Associated Interstitial Lung Disease. *Rheumatol (Oxford)* (2019) 58(3):511–21. doi: 10.1093/rheumatology/key341
120. Brodeur TY, Robidoux TE, Weinstein JS, Craft J, Swain SL, Marshak-Rothstein A. IL-21 Promotes Pulmonary Fibrosis Through the Induction of Profibrotic CD8+ T Cells. *J Immunol* (2015) 195:5251–60. doi: 10.4049/jimmunol.1500777
121. Sica A, Mantovani A. Macrophage Plasticity and Polarization: *In Vivo* Veritas. *J Clin Invest* (2012) 122:787–95. doi: 10.1172/JCI59643
122. Kwan C, Milosevic S, Benham H, Scott IA. A Rare Form of Dermatomyositis Associated With Muscle Weakness and Normal Creatine Kinase Level. *BMJ Case Rep* (2020) 13:e232260. doi: 10.1136/bcr-2019-232260
123. Kushnir-Grinbaum D, Cohen-Barak E, Bieber A, Krausz J, Dodiuk-Gad R, Mader R, et al. Toxic Epidermal Necrolysis-Like Dermatomyositis Associated With Antimelanoma Differentiation Antigen 5. *JAAD Case Rep* (2019) 5:91–3. doi: 10.1016/j.jidcr.2018.10.018
124. Joshi R. Interface Dermatitis. *Indian J Dermatol Venereol Leprol* (2013) 79:349. doi: 10.4103/0378-6323.110780
125. Wenzel J, Lucas S, Zahn S, Mikus S, Metzke D, Ständer S, et al. CXCR3 <-> Ligand-Mediated Skin Inflammation in Cutaneous Lichenoid Graft-Versus-Host Disease. *J Am Acad Dermatol* (2008) 58:437–42. doi: 10.1016/j.jaad.2007.10.647
126. Wenzel J, Schmidt R, Proelss J, Zahn S, Bieber T, Tuting T. Type I Interferon-Associated Skin Recruitment of CXCR3+ Lymphocytes in Dermatomyositis. *Clin Exp Dermatol* (2006) 31:576–82. doi: 10.1111/j.1365-2230.2006.02150.x
127. Wenzel J, Wörenkämper E, Freutel S, Henze S, Haller O, Bieber T, et al. Enhanced Type I Interferon Signaling Promotes Th1-Biased Inflammation in Cutaneous Lupus Erythematosus. *J Pathol* (2005) 205:435–42. doi: 10.1002/path.1721
128. Wenzel J, Zahn S, Bieber T, Tuting T. Type I Interferon-Associated Cytotoxic Inflammation in Cutaneous Lupus Erythematosus. *Arch Dermatol Res* (2009) 301:83–6. doi: 10.1007/s00403-008-0892-8
129. Cassius C, Amode R, Delord M, Battistella M, Poirot J, How-Kit A, et al. MDA5+ Dermatomyositis Is Associated With Stronger Skin Type I Interferon Transcriptomic Signature With Upregulation of IFN- γ

- Transcript. *J Invest Dermatol* (2020) 140:1276–9.e7. doi: 10.1016/j.jid.2019.10.020
130. Indraccolo S, Pfeffer U, Minuzzo S, Esposito G, Roni V, Mandruzzato S, et al. Identification of Genes Selectively Regulated by IFNs in Endothelial Cells. *J Immunol* (2007) 178:1122–35. doi: 10.4049/jimmunol.178.2.1122
 131. Pau-Charles I, Moreno PJ, Ortiz-Ibáñez K, Lucero MC, García-Herrera A, Espinosa G, et al. Anti-MDA5 Positive Clinically Amyopathic Dermatomyositis Presenting With Severe Cardiomyopathy. *J Eur Acad Dermatol Venereol* (2014) 28:1097–102. doi: 10.1111/jdv.12300
 132. Abraham D, Distler O. How Does Endothelial Cell Injury Start? The Role of Endothelin in Systemic Sclerosis. *Arthritis Res Ther* (2007) 9:S2. doi: 10.1186/ar2186
 133. Min HK, Kim H-R, Lee S-H, Park S-H, Oh J, Choi K. Refractory Digital Ulcers Treated by Botulinum Toxin and Endothelin Receptor-1 Antagonist in Anti-MDA5-Antibody-Positive Dermatomyositis. *J Clin Neurol* (2020) 16:160. doi: 10.3988/jcn.2020.16.1.160
 134. Collantes-Rodríguez C, Jiménez-Gallo D, Ossorio-García L, Villegas-Romero I, Linares-Barrios M. Image Gallery: Cutaneous Ulcers in Anti-MDA5 Dermatomyositis Successfully Treated With Sildenafil. *Br J Dermatol* (2019) 182:e1. doi: 10.1111/bjd.18376
 135. Combalia A, Giavedoni P, Tamez L, Grau-Junyent JM, Mascaró JM. Bosentan for Cutaneous Ulcers in Anti-MDA5 Dermatomyositis. *JAMA Dermatol* (2018) 154:371. doi: 10.1001/jamadermatol.2017.5462
 136. Horai Y, Koga T, Fujikawa K, Takatani A, Nishino A, Nakashima Y, et al. Serum Interferon- α Is a Useful Biomarker in Patients With Anti-Melanoma Differentiation-Associated Gene 5 (MDA5) Antibody-Positive Dermatomyositis. *Mod Rheumatol* (2015) 25:85–9. doi: 10.3109/14397595.2014.900843
 137. Ogawa Y, Kishida D, Shimojima Y, Hayashi K, Sekijima Y. Effective Administration of Rituximab in Anti-MDA5 Antibody-Positive Dermatomyositis With Rapidly Progressive Interstitial Lung Disease and Refractory Cutaneous Involvement: A Case Report and Literature Review. *Case Rep Rheumatol* (2017) 2017:5386797. doi: 10.1155/2017/5386797
 138. So H, Wong VTL, Lao VWN, Pang HT, Yip RML. Rituximab for Refractory Rapidly Progressive Interstitial Lung Disease Related to Anti-MDA5 Antibody-Positive Amyopathic Dermatomyositis. *Clin Rheumatol* (2018) 37:1983–9. doi: 10.1007/s10067-018-4122-2
 139. Ge Y, Li S, Tian X, He L, Lu X, Wang G. Anti-Melanoma Differentiation-Associated Gene 5 (MDA5) Antibody-Positive Dermatomyositis Responds to Rituximab Therapy. *Clin Rheumatol* (2021) 40(6):2311–7. doi: 10.1007/s10067-020-05530-5
 140. Endo Y, Koga T, Suzuki T, Hara K, Ishida M, Fujita Y, et al. Successful Treatment of Plasma Exchange for Rapidly Progressive Interstitial Lung Disease With Anti-MDA5 Antibody-Positive Dermatomyositis: A Case Report. *Med (Baltimore)* (2018) 97:e0436. doi: 10.1097/MD.00000000000010436
 141. Abe Y, Kusaoi M, Tada K, Yamaji K, Tamura N. Successful Treatment of Anti-MDA5 Antibody-Positive Refractory Interstitial Lung Disease With Plasma Exchange Therapy. *Rheumatology* (2020) 59:767–71. doi: 10.1093/rheumatology/kez357
 142. Vlahakos D, Foster MH, Ucci AA, Barrett KJ, Datta SK, Madaio MP. Murine Monoclonal Anti-DNA Antibodies Penetrate Cells, Bind to Nuclei, and Induce Glomerular Proliferation and Proteinuria *In Vivo*. *J Am Soc Nephrol* (1992) 2(8):1345–54. doi: 10.1681/ASN.V281345
 143. Deng SX, Hanson E, Sanz I. *In Vivo* Cell Penetration and Intracellular Transport of Anti-Sm and Anti-La Autoantibodies. *Int Immunol* (2000) 12:415–23. doi: 10.1093/intimm/12.4.415
 144. Douglas J N. Antibodies to an Intracellular Antigen Penetrate Neuronal Cells and Cause Deleterious Effects. *J Clin Cell Immunol* (2013) 04. doi: 10.4172/2155-9899.1000134
 145. Chen M, Zhao Q, Diao L, Xue K, Ruan Y, Xue F, et al. Distribution of Anti-Melanoma Differentiation Associated Gene 5 (MDA5) IgG Subclasses in MDA5+ Dermatomyositis. *Rheumatology* (2021) keab268. doi: 10.1093/rheumatology/keab268
 146. Della-Torre E, Lanzillotta M, Doglioni C. Immunology of IgG4-Related Disease: Immunology of IgG4-Related Disease. *Clin Exp Immunol* (2015) 181:191–206. doi: 10.1111/cei.12641
 147. Giannini M, Ohana M, Nespola B, Zanframundo G, Geny B, Meyer A. Similarities Between COVID-19 and Anti-MDA5 Syndrome: What can We Learn for Better Care? *Eur Respir J* (2020) 56:2001618. doi: 10.1183/13993003.01618-2020
 148. Hanley B, Naresh KN, Roufousse C, Nicholson AG, Weir J, Cooke GS, et al. Histopathological Findings and Viral Tropism in UK Patients With Severe Fatal COVID-19: A Post-Mortem Study. *Lancet Microbe* (2020) 1:e245–53. doi: 10.1016/S2666-5247(20)30115-4
 149. Quintana-Ortega C, Remesal A, Ruiz de Valbuena M, de la Serna O, Laplaza-González M, Álvarez-Rojas E, et al. Fatal Outcome of Anti-MDA5 Juvenile Dermatomyositis in a Paediatric COVID-19 Patient: A Case Report. *Mod Rheumatol Case Rep* (2021) 5:101–7. doi: 10.1080/24725625.2020.1832755
 150. Rebendenne A, Valadão ALC, Tauziet M, Maarifi G, Bonaventure B, McKellar J, et al. SARS-CoV-2 Triggers an MDA-5-Dependent Interferon Response Which Is Unable to Control Replication in Lung Epithelial Cells. *J Virol* (2021) 95(8):e02415–20. doi: 10.1128/JVI.02415-20
 151. Yin X, Riva L, Pu Y, Martin-Sancho L, Kanamune J, Yamamoto Y, et al. MDA5 Governs the Innate Immune Response to SARS-CoV-2 in Lung Epithelial Cells. *Cell Rep* (2021) 34:108628. doi: 10.1016/j.celrep.2020.108628
 152. Trouillet-Assant S, Viel S, Gaymard A, Pons S, Richard JC, Perret M, et al. Type I IFN Immunoprofiling in COVID-19 Patients. *J Allergy Clin Immunol* (2020) 146(1):206–8.e2. doi: 10.1016/j.jaci.2020.04.029
 153. Hadjadj J, Yatim N, Barnabei L, Corneau A, Boussier J, Smith N, et al. Impaired Type I Interferon Activity and Inflammatory Responses in Severe COVID-19 Patients. *Science* (2020) 369:718–24. doi: 10.1126/science.abc6027
 154. Zhang Q, Bastard P, Liu Z, Le Pen J, Moncada-Velez M, Chen J, et al. Inborn Errors of Type I IFN Immunity in Patients With Life-Threatening COVID-19. *Science* (2020) 370:eabd4570. doi: 10.1126/science.abd4570
 155. Bastard P, Rosen LB, Zhang Q, Michailidis E, Hoffmann H-H, Zhang Y, et al. Autoantibodies Against Type I IFNs in Patients With Life-Threatening COVID-19. *Science* (2020) 370:eabd4585. doi: 10.1126/science.abd4585
 156. Goncalves D, Mezidi M, Bastard P, Perret M, Saker K, Fabien N, et al. Antibodies Against Type I Interferon: Detection and Association With Severe Clinical Outcome in COVID-19 Patients. *Clin Transl Immunol* (2021) 10(8):e1327. doi: 10.1002/cti2.1327
 157. Lee JS, Shin E-C. The Type I Interferon Response in COVID-19: Implications for Treatment. *Nat Rev Immunol* (2020) 20:585–6. doi: 10.1038/s41577-020-00429-3
 158. Lee JS, Park S, Jeong HW, Ahn JY, Choi SJ, Lee H, et al. Immunophenotyping of COVID-19 and Influenza Highlights the Role of Type I Interferons in Development of Severe COVID-19. *Sci Immunol* (2020) 5:eabd1554. doi: 10.1126/sciimmunol.abd1554
 159. Nägele MP, Haubner B, Tanner FC, Ruschitzka F, Flammer AJ. Endothelial Dysfunction in COVID-19: Current Findings and Therapeutic Implications. *Atherosclerosis* (2020) 314:58–62. doi: 10.1016/j.atherosclerosis.2020.10.014
 160. Siddiqi HK, Libby P, Ridker PM. COVID-19 – A Vascular Disease. *Trends Cardiovasc Med* (2021) 31:1–5. doi: 10.1016/j.tcm.2020.10.005
 161. Goshua G, Pine AB, Meizlish ML, Chang C-H, Zhang H, Bahel P, et al. Endotheliopathy in COVID-19-Associated Coagulopathy: Evidence From a Single-Centre, Cross-Sectional Study. *Lancet Haematol* (2020) 7:e575–82. doi: 10.1016/S2352-3026(20)30216-7
 162. Evans PC, Rainger GE, Mason JC, Guzik TJ, Osto E, Stamataki Z, et al. Endothelial Dysfunction in COVID-19: A Position Paper of the ESC Working Group for Atherosclerosis and Vascular Biology, and the ESC Council of Basic Cardiovascular Science. *Cardiovasc Res* (2020) 116:2177–84. doi: 10.1093/cvr/cvaa230
 163. Liu C, Wang Q, Wang Y, Wang G, Wang L, Chen H, et al. Analysis of the Correlation Between Anti-MDA5 Antibody and the Severity of COVID-19: A Retrospective Study. *medRxiv [Preprint]* (2020) 20164780. doi: 10.1101/2020.07.29.20164780
 164. De Santis M, Isailovic N, Motta F, Ricordi C, Ceribelli A, Lanza E, et al. Environmental Triggers for Connective Tissue Disease: The Case of COVID-19 Associated With Dermatomyositis-Specific Autoantibodies. *Curr Opin Rheumatol* (2021) 33:514–21. doi: 10.1097/BOR.0000000000000844
 165. Hansen KE, Arnason J, Bridges AJ. Autoantibodies and Common Viral Illnesses. *Semin Arthritis Rheum* (1998) 27:263–71. doi: 10.1016/s0049-0172(98)80047-
 166. Gil-Vila A, Perurena-Prieto J, Nolla-Fontana C, Orozco-Galvez O, Miarons-Font M, del Castillo AG, et al. Pos1229 Anti-Mda5 and Antisynthetase

- Antibodies Screening in Severe Sars-Cov-2 Pneumonia. Be Aware of False Positive Results. *Ann Rheumatic Dis* (2021) 80:898. doi: 10.1136/annrheumdis-2021-eular.301
167. Romero-Bueno F, Diaz del Campo P, Trallero-Araguás E, Ruiz-Rodríguez JC, Castellvi I, Rodríguez-Nieto MJ, et al. Recommendations for the Treatment of Anti-Melanoma Differentiation-Associated Gene 5-Positive Dermatomyositis-Associated Rapidly Progressive Interstitial Lung Disease. *Semin Arthritis Rheum* (2020) 50:776–90. doi: 10.1016/j.semarthrit.2020.03.007
 168. Zohar DN, Seluk L, Yonath H, Shoenfeld Y, Kivity S. Anti-MDA5 Positive Dermatomyositis Associated With Rapidly Progressive Interstitial Lung Disease and Correlation Between Serum Ferritin Level and Treatment Response. *Mediterr J Rheumatol* (2019) 31:75. doi: 10.31138/mjr.31.1.75
 169. Osawa T, Morimoto K, Sasaki Y, Matsuda S, Yamana K, Yano R, et al. The Serum Ferritin Level Is Associated With the Treatment Responsivity for Rapidly Progressive Interstitial Lung Disease With Amyopathic Dermatomyositis, Irrespective of the Anti-MDA5 Antibody Level. *Intern Med* (2018) 57:387–91. doi: 10.2169/internalmedicine.8335-16
 170. Shu E, Kanoh H, Murakami A, Seishima M. Potential Inhibition of Development of Rapidly Progressive Interstitial Lung Disease by Prompt and Sufficient Immunosuppressive Treatment in Patients With Anti-Melanoma Differentiation-Associated Gene 5 Antibody-Positive Dermatomyositis. *J Dermatol* (2017) 44:e91–2. doi: 10.1111/1346-8138.13659
 171. Tsuji H, Nakashima R, Hosono Y, Imura Y, Yagita M, Yoshifuji H, et al. Multicenter Prospective Study of the Efficacy and Safety of Combined Immunosuppressive Therapy With High-Dose Glucocorticoid, Tacrolimus, and Cyclophosphamide in Interstitial Lung Diseases Accompanied by Anti-Melanoma Differentiation-Associated Gene 5-Positive Dermatomyositis. *Arthritis Rheumatol* (2020) 72:488–98. doi: 10.1002/art.41105
 172. Matsuda KM, Yoshizaki A, Kuzumi A, Fukasawa T, Ebata S, Yoshizaki-Ogawa A, et al. Combined Immunosuppressive Therapy Provides Favorable Prognosis and Increased Risk of Cytomegalovirus Reactivation in Anti-Melanoma Differentiation-Associated Gene 5 Antibody-Positive Dermatomyositis. *J Dermatol* (2020) 47:483–9. doi: 10.1111/1346-8138.15274
 173. Seiller H, Schmutz J-L, Poreaux C, Bursztejn A-C. Dermatomyosite À Anticorps Anti-MDA-5 Compliquée D'une Pneumopathie Interstitielle Et D'une Pneumocystose D'évolution Fatale. *Ann Dermatol Vénéréologie* (2020) 147:862–7. doi: 10.1016/j.annder.2020.08.046
 174. Aymonier M, Abed S, Boyé T, Barazzutti H, Fournier B, Morand J-J. Dermatomyosite Associée Aux Anticorps Anti-MDA5 Et Pneumocystose Pulmonaire: Deux Cas D'évolution Fatale. *Ann Dermatol Vénéréologie* (2017) 144:279–83. doi: 10.1016/j.annder.2016.09.677
 175. Fillatre P, Decaux O, Jouneau S, Revest M, Gacouin A, Robert-Gangneux F, et al. Incidence of Pneumocystis Jiroveci Pneumonia Among Groups at Risk in HIV-Negative Patients. *Am J Med* (2014) 127:1242. doi: 10.1016/j.amjmed.2014.07.010
 176. Wendel S, Venhoff N, Frye BC, May AM, Agarwal P, Rizzi M, et al. Successful Treatment of Extensive Calcifications and Acute Pulmonary Involvement in Dermatomyositis With the Janus-Kinase Inhibitor Tofacitinib – A Report of Two Cases. *J Autoimmun* (2019) 100:131–6. doi: 10.1016/j.jaut.2019.03.003
 177. Takatani A, Koga T, Fujita Y, Fukui S, Endo Y, Shimizu T, et al. Efficacy of Tofacitinib for Slowly Progressive Interstitial Lung Disease in a Patient With Anti-MDA5 Antibody-Positive Dermatomyositis. *Clin Immunol* (2020) 215:108451. doi: 10.1016/j.clim.2020.108451
 178. Kurasawa K, Arai S, Namiki Y, Tanaka A, Takamura Y, Owada T, et al. Tofacitinib for Refractory Interstitial Lung Diseases in Anti-Melanoma Differentiation-Associated 5 Gene Antibody-Positive Dermatomyositis. *Rheumatology* (2018) 57:2114–9. doi: 10.1093/rheumatology/key188
 179. Hosokawa Y, Oiwa H. A Case of Refractory Interstitial Lung Disease in Anti-MDA5-Positive Dermatomyositis That Improved After Switching to Tofacitinib. *J Clin Rheumatol* (2020). doi: 10.1097/RHU.0000000000001645
 180. Mehta DS, Wurster AL, Grusby MJ. Biology of IL-21 and the IL-21 Receptor. *Immunol Rev* (2004) 202:84–95. doi: 10.1111/j.0105-2896.2004.00201.x
 181. Habib T, Senadheera S, Weinberg K, Kaushansky K. The Common γ Chain (γ_c) Is a Required Signaling Component of the IL-21 Receptor and Supports IL-21-Induced Cell Proliferation via JAK3. *Biochemistry* (2002) 41:8725–31. doi: 10.1021/bi0202023
 182. Chen Z, Wang X, Ye S. Tofacitinib in Amyopathic Dermatomyositis-Associated Interstitial Lung Disease. *N Engl J Med* (2019) 381:291–3. doi: 10.1056/NEJMc1900045
 183. Scirocco C, Gubbiotti A, Sebastiani A, Sebastiani GD. Rituximab in Antimelanoma Differentiation-Associated Protein-5 Dermatomyositis With Interstitial Lung Disease. *Case Rep Rheumatol* (2020) 2020:8145790. doi: 10.1155/2020/8145790
 184. Hisanaga J, Kotani T, Fujiki Y, Yoshida S, Takeuchi T, Makino S. Successful Multi-Target Therapy Including Rituximab and Mycophenolate Mofetil in Anti-Melanoma Differentiation-Associated Gene 5 Antibody-Positive Rapidly Progressive Interstitial Lung Disease With Clinically Amyopathic Dermatomyositis. *Int J Rheum Dis* (2017) 20:2182–5. doi: 10.1111/1756-185X.13136
 185. Mao M, Xia S, Guo B, Qian W, Zheng Z, Peng X, et al. Ultra-Low Dose Rituximab as Add-on Therapy in Anti-MDA5-Positive Patients With Polymyositis/Dermatomyositis Associated ILD. *Respir Med* (2020) 172:105983. doi: 10.1016/j.rmed.2020.105983
 186. Takahashi R, Yoshida T, Morimoto K, Kondo Y, Kikuchi J, Saito S, et al. Successful Treatment of Anti-MDA5 Antibody-Positive Dermatomyositis-Associated Rapidly Progressive-Interstitial Lung Disease by Plasma Exchange: Two Case Reports. *Clin Med Insights Case Rep* (2021) 14:117954762110363. doi: 10.1177/11795476211036322
 187. Silveira MG, Selva-O'Callaghan A, Ramos-Terrades N, Arredondo-Agudelo KV, Labrador-Horrillo M, Bravo-Masgoret C. Anti-MDA5 Dermatomyositis and Progressive Interstitial Pneumonia. *QJM* (2016) 109:49–50. doi: 10.1093/qjmed/hcv068
 188. Kagawa H, Tsujino K, Yamamoto Y, Iwai A, Hara R, Matsuki T, et al. Acute Lung Injury After Plasma Exchange in a Patient With Anti-MDA5 Antibody-Positive, Rapidly Progressive, Interstitial Lung Disease: A Case Report. *Respir Med Case Rep* (2020) 29:101016. doi: 10.1016/j.rmcr.2020.101016

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