



Gut Microbiota as Regulators of Th17/Treg Balance in Patients With Myasthenia Gravis

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Myasthenia gravis (MG) is an acquired neurological autoimmune disorder characterized by dysfunctional transmission at the neuromuscular junction, with its etiology associated with genetic and environmental factors. Anti-inflammatory regulatory T cells (Tregs) and proinflammatory T helper 17 (Th17) cells functionally antagonize each other, and the immune imbalance between them contributes to the pathogenesis of MG. Among the numerous factors influencing the balance of Th17/Treg cells, the gut microbiota have received attention from scholars. Gut microbial dysbiosis and altered microbial metabolites have been seen in patients with MG. Therefore, correcting Th17/Treg imbalances may be a novel therapeutic approach to MG by modifying the gut microbiota. In this review, we initially review the association between Treg/Th17 and the occurrence of MG and subsequently focus on recent findings on alterations of gut microbiota on Th17/ Treg balance in patients with MG, which may provide a new direction for the prevention and treatment of this disease.

Keywords: myasthenia gravis, Th17, Treg, gut microbiota, short-chain fatty acids

INTRODUCTION

Myasthenia gravis (MG) is an autoantibody-mediated neurological disorder caused by transmission defects at the neuromuscular junction (1). Pathogenic antibodies that can be detected using current technological methods mainly include antibodies against acetylcholine receptors (AChR), muscle-specific kinase (MuSK), and lipoprotein receptor-related protein 4 (LRP4) (2). The antibodies induce fatigue and weakness of the eye muscles or skeletal muscles, which are the main clinical hallmarks (3, 4). Patients with MG whose weakness is limited to the extraocular muscles are referred to as patients with oculomotor myasthenia gravis (OMG). Once fatigue spreads to the skeletal muscles, the disease can progress to generalized myasthenia gravis (GMG). Depending on whether patients are over 50 years of age at onset, MG can be divided into early-onset MG (EOMG) and late-onset MG (LOMG) (4). Myasthenia gravis has an annual worldwide incidence of 8–10 per 1 million people per year, and the total prevalence of MG has been reported to be 150–250 per 1 million people (5). Genetic factors play a crucial role in the induction of myasthenia gravis. MG has a concordance rate of 35.5% in monozygotic twins and 4-5% in heterozygous twins (6). As *reported*, some genes correlates strongly with *the onset of MG*, such as HLA-DQ5 and polymorphisms of CTLA-4 (4, 7). Based on these findings, genetic susceptibility may contribute to the development of

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MG.With a low rate of spontaneous remission, most patients require long-term immunosuppressive agents to achieve minimal manifestation status or improvement after MG intervention (2). Monoclonal antibodies targeting specific molecules have become a research hotspot in recent years; however, they are not yet widely applied in clinical practice, and finding treatments to prevent or cure this disorder is a major challenge in this field that requires an in-depth understanding of disease pathogenesis.

Myasthenia gravis is an acquired autoimmune disease with complex pathogenesis, mediated by humoral and cellular immunity and implicated by complement. The thymus is affected in most AChR-MG patients, with thymic hyperplasia being the most common (8). The formation of germinal centers in thymus is a typical pathological change of MG patients (9).Take early-onset AChR-MG for example, in response to AChR stimulation, the thymic germinal centers overexpress pro-inflammatory cytokines and thymic epithelial cells, presenting AChR subunits to autoreactive CD4+ T cells, thereby upregulating IL-4 and IL-6 and stimulating B cell proliferation to promote the production of anti-AChR antibodies (10). Meanwhile, the titers of IFN- γ and IL-17 are increased, implying the involvement of T helper (Th)1 and Th17 cells in MG pathogenesis (11). Moreover, the ability of Treg to suppress T cell responses are also greatly impaired (11). Tregs are critical for maintaining immune tolerance and immune homeostasis (12). Th17 cells play an essential role in mediating chronic inflammation in autoimmune diseases (13), and multiple studies have shown abnormalities in the function or expression levels of Treg and Th17 in animal models or patients with MG (14-20). Moreover, antibody titer tends to correlate with disease severity, indicating a possible involvement of a Treg/Th17 imbalance in MG disease pathogenesis. Rebuilding Thl7/Treg balance has a promising application for the biological target therapy of MG.

Gut microbiota can influence various human physiological activities in many aspects, including nutrient synthesis, immune response modulation, and defense against pathogen invasion (21-23). The gut microbiome is a dynamically balanced unity of microorganisms. Although the mechanisms are still unclear, the immunomodulatory effects of gut microbiota are mostly realized through the Thl7/Treg axis (24, 25). Gut microbiota dysbiosis may lead to increased permeability of the intestinal mucosal barrier, infection, and excessive inflammatory response, as well as an imbalance of T cell subsets, which has been described in asthma (26), rheumatoid arthritis (27), inflammatory bowel disease (28), multiple sclerosis (29), and MG (30), among others. Accordingly, regulating the immune imbalance of Thl7/ Treg cells and inducing the reconstruction of immune tolerance may become a potential treatment modality for autoimmune diseases, such as MG. Studies have revealed that environmental factors, especially dietary factors, can alter the gut microbiota composition and its metabolic activity, which produce adverse implications for human health (31). For instance, alterations in the gut microbiota caused by a Western high-fat diet have recently been recognized as a major contributing factor to the

colorectal cancer epidemic (32). High-sucrose-high-fat dietary patterns were supposed to participate in the asthma pathogenesis through intestinal bacteria (33, 34). Currently, there is not related research on whether dietary factors are involved in the pathology of MG. This review paper covers three main aspects. First, we introduce the association between Treg/Th17 imbalances and MG occurrence. Second, we summarize recent findings regarding varied gut microbiota and its metabolites in patients with MG and also explore the effects of gut microbiota on Th17/ Treg in these patients. Finally, we provide new insights on MG therapy regarding the rebuilding of Th17/Treg balance by modifying the gut microbiota.

SIGNIFICANCE OF Th17/Treg IMBALANCES IN MG PROGRESSION

Th17 cells synthesize and release various inflammatory cytokines, such as interleukin (IL)-17 and IL-22, activate neutrophils, and infiltrate into lesions, thus amplifying inflammatory effect. The key transcription factor, retinoic acidrelated orphan receptor (RORyt), mediates the differentiation of Th17 cells and biosynthesis of Th17 cytokines (35-37). Evidence from previous studies has shown that Th17 cells promote inflammatory responses and mediate tissue damage. The proinflammatory effects of Th17 cells are mainly attributed to their secretion of IL-17, and the corresponding mechanisms involves the recruitment of neutrophils, enhancement of B cell function, activation of innate immune cells, and induction release of proinflammatory cytokines, such as tumor necrosis factor alpha, granulocyte-macrophage colony-stimulating factor, and IL-1β (38, 39). Various studies have found that Thl7 cells and their main secreted cytokine IL-17 is related inextricably to MG pathogenesis. Schaffert et al. observed less myasthenia symptoms and significant reductions in anti-AChR antibodies in IL-17(knock-out) mice, concluding that the process of antibody production and loss of B-cell tolerance was regulated by IL-17 and consequently confirming the involvement of Th17 cells in MG autoimmunity (14). A study in 2017 emphasized that IL-17 independent pathways drove the autoimmune response related to experimental autoimmune myasthenia gravis (EAMG) development (15). Moreover, serum IL-17 levels have shown a positive correlation to the degree of quantitative MG score and anti-AChR antibody titers, heralding a severer disease course (16). Antibody production in patients with MG is affected by Th1/Th2 cytokine balances, which is regulated by Th17 cells (40). Accordingly, Th17 cells and their released cytokines are linked to the induction of anti-AChR antibody-mediated autoimmunity at the neuromuscular junction.

Tregs could inhibit the function of other effector T cells and antigen-presenting cells, suppressing immune responses (41). Tregs release multiple inhibitory cytokines, such as transforming growth factor-beta (TGF- β) and IL-10 (42). The immunomodulatory property of Tregs is controlled by the expression of forkhead box protein 3 (FoxP3) (43). The number of CD4+CD25+FoxP3+Tregs in peripheral blood is decreased

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during the active stage of anti-AChR antibody-positive MG and becomes higher with immunotherapy, implying that decreased number of Tregs may lead to immune disorders (44). The number of Tregs seems to be influenced by disease involvement. For instance, the proportion of Tregs of generalized anti-AChR antibody positive MG without thymoma group is remarkably lower than those of ocular MG, as could be seen in the study by Hu et al. (17). However, research has demonstrated that the relative number of Tregs presented no obvious changes when patients with MG were compared to healthy controls (18). Of note, Tregs have functional plasticity. Huang et al. demonstrated that CD4+CD25-T-cells from patients with MG transformed into CD4+CD25+Tregs expressing FoxP3 when stimulated by IFN- γ in a dose-dependent fashion (19). Notably, these studies identified Tregs based on high CD25 expression, whereas the expression level of CD25 that identifies Treg was not standardized in the literature, and CD25 was also expressed in other T cells including effector T cells or T cells with pro-inflammatory properties (45). Accordingly, analyzing the number of Tregs merely according to CD25 expression may contradict results. Although there has been no agreement on whether the number of Tregs is altered yet, most findings reported functional defects in Tregs. For instance, Thiruppathi et al. demonstrated that the inhibitory function of responder T (Tresp) cells mediated by Tregs was disrupted in patients with MG, which correlated with reduced expression of FoxP3 (18). Balandina et al. found a severe functional defect in the regulatory activity of Tregs along with a decreased expression of FoxP3 (20). Therefore, the functional impairment of Tregs, both in the periphery and thymus, is more likely to be related to MG. FoxP3 has been shown to be a key player in the development and functional activity of Tregs (46). Thus, MG is associated with a decreased expression of FoxP3 on Tregs, which is corroborated by several studies (18, 20, 44, 47).

Briefly, abnormalities in the function or expression levels of Treg and Th17 cells exist in patients with MG. Therefore, an immune imbalance between Th17 and Tregs is a significant driver of MG. Based on the above-mentioned studies, improving this imbalance is a potential new avenue for treating MG. IL-6 plays pivotal roles in the regulation of the balance of Treg/Th17 cells (48). TGF- β helps induce the development of Treg and Th17 cells and solely drives the formation of Tregs from naive CD4+ T cells (49), whereas TGF- β together with IL-6 initiates Th17 lineage development (50). Anti-IL-6 monoclonal antibody has been reported to show good efficacy in patients with EAMG. Additionally, other studies have shown that rapamycin and artemisinin lessen motor symptoms in EAMG rats, with an ability of correcting Treg/Th17 imbalances (51, 52). Rapamycin is the most common inhibitor targeting mTOR, which inhibits the differentiation of Th17 cells (53) and induces the proliferation of Treg cells (54) by inhibiting the mTOR signaling pathway. Upon treatment with artemisinins, a decreased level of tumor necrosis factor- α (TNF- α) and IL-17+ cells in mononuclear cells (MNCs), and an increased level of transforming growth factor-\u03b31 (TGF-\u03b31) and Treg cells in MNCs were observed (52). These findings indicate that

Rapamycin and artemisinin may be new choices for MG treatment, although investigations are still in its infancy. However, the reasons behind imbalances in Treg/Th17 cells in patients with MG are not well established. Conventionally, numerous cytokines are responsible for the maintenance of Treg/Th17 balance, and the abnormal expression of these cytokines due to thymoma or hyperplasia is one of the causes. Furthermore, gut microbiota is another critical regulator of Th17/Treg cell balance.

DYSBIOSIS OF GUT MICROBIOTA AND MICROBIAL METABOLITES IN PATIENTS WITH MG

The gut microbiota is defined as the microbial community colonizing the gut, including dominant bacteria, archaea, protists, fungi, and viruses. More than 90% of the bacteria in the adult gut belong to the phyla of Firmicutes and Bacteroidetes, while the rest belong to the phylum other phyla, such as Proteobacteria, Actinobacteria and Verrucomicrobia (55).

Gut microbiota colonizes the host and exerts an essential impact on host immune tolerance and immunostasis (56-59). Studies on germ-free (GF) mice and antibiotic-treated models have shown that gut microbiota can regulate the differentiation and development of immune cells (60). With advancements in corresponding research, it was found that gut microbiota participate in Th17 and Treg cell balance, and the potential mechanisms were also investigated (25). Specific components of the gut microbiome are involved in the production of proinflammatory cytokines and subsequent production of Th17 cells. Similarly, commensal bacteria and their metabolites can also promote the production of Tregs to facilitate immunosuppression. Under normal conditions, microbiomemediated inflammatory factors and anti-inflammatory factors are in immune equilibrium. However, once gut dysbiosis occurs, host immune dysregulation may start, triggering chronic inflammatory disorders, such as inflammatory bowel disease (28), rheumatoid arthritis (27), multiple sclerosis (29), and MG (30). However, different diseases have different patterns of microbial dysbiosis.

Substantial evidence indicates that MG is accompanied by dysbiosis of gut microbiota and altered metabolites. Qiu et al. (61) demonstrated for the first time the decreased microbial diversity and altered community structures in MG *via* 16S rRNA sequencing technology, offering new insights on MG-relevant host-microbes interactions. Specifically, decreased phylum Firmicutes along with increased phylum Proteobacteria and Bacteroidetes were observed in the fecal samples of patients with MG. Accordingly, the ratio of Firmicutes/Bacteroidetes (F/B ratio) in patients with MG was markedly lower, which was consistent with the decreased F/B ratio observed in systemic lupus erythematosus and inflammatory bowel disease (62, 63). In fact, the F/B ratio can reflect a pro-inflammatory environment, where the inflammatory microbiota could damage the intestinal

epithelium and subsequently trigger an immune response, leading to immunological imbalance. Regarding relative abundance at the genus level, Eubacterium and Clostridium were significantly lower in the MG group than in the control group, whereas Parasutterella and Streptococcus levels were significantly higher. Among these bacteria, Clostridium was the most depleted and comprised only approximately one-third in the healthy controls as analyzed by qPCR analyses. There is ample evidence that composition changes in the gut microbiota, particularly the abundance of Clostridium strains, have profound effects on the differentiation and development of T cells. Particularly, Clostridia can increase the expression of 2,3dioxygenase, in turn, promotes the conversion of naive CD4+ T cells to Tregs by catalyzing the tryptophan-kynurenine (Trp-Kyn) metabolic pathway (64-66). Conventionally, Tregs have a profound effect on inhibiting autoreactive B cells and the production of anti-AChR autoantibodies (67). Therefore, we can speculate that if depleted, Clostridia can be restored, the number of Tregs will increase, consequently reducing autoimmune inflammatory responses in patients with MG and providing new insights on the treatment of MG at the microbial level. Moreover, it was also observed that F. prausnitzii was significantly reduced and that streptococcus was dramatically increased in the MG group at the genus level compared with the control groups; the control group would have the ability of activating peroxisome proliferator-activated receptor (PPARy) through the suppression of immune cell function and certain signaling pathways, resulting in a tight balance of immune system responses (68, 69).

In another study (70), patients with MG were observed to harbor significantly higher proportions of Pasteurellaceae, Desulfovibrionaceae and Acidaminococcaceae compared to HCs. Moreover, patients with MG displayed a markedly lower relative abundance of the families of Bifdobacteriaceae and Verrucomicrobiaceae, as well as Leuconostocacceae, Flavobacteriaceae and Coriobacteriaceae. Results have shown an increased abundance of Bacteroidetes, among others, in patients with MG. A subsequent study in China found that the relative abundances of Bacteroidetes and Fusobacteria were increased in patients with MG, whereas those of Actinobacteria were decreased (71). A study aimed to identify the distinctive gut microbes in different subtypes of MG showed that patients with GMG harbored lower community richness and diversity and more severe gut microbial disturbances than patients with OMG. Compared with HCs, the families of Bacteroidaceae and Veillonellaceae were significantly more abundant, whereas families of Lachnospiraceae, Erysipelotrichaceae, Ruminococcaceae, Peptostreptococcaceae, Coriobacteriaceae, and Clostridiaceae_1 were scarcer in patients with MG (72). According to published literature, previous studies have suggested that Veillonellaceae and Bacteroidales are positively correlated with certain autoimmune disorders, whereas Ruminococcaceae and Lachnospiraceae, the common short chain fatty acids (SCFAs) producers (73), are negatively correlated with these diseases (74, 75). Recent research findings on the abnormal composition of gut microbiota in patients with MG are summarized in Table 1.

TABLE 1 | Altered gut microbiota compositions in patients with MG.

Altered microbiota	References
Bacteroidetes [↑] , Actinobacteria [↑] , Verrucomicrobia [↑] , Desulfovibrionaceae [↑] , Acidaminococcaceae [↑] , Pasteurellaceae [↑] , Bifdobacteriaceae [↓] , Verrucomicrobiaceae [↓] , Coriobacteriaceae [↓] , Leuconostocacceae [↓] , and Flavobacteriaceae [↓]	Moris et al. (70)
Proteobacteria [↑] , Actinobacteria [↑] , Bacteroidetes [↑] , and Firmicutes [↓] Streptococcus [↑] , Parasutterella [↑] , Escherichia [↑] , Clostridium [↓] , Eubacterium [↓] , and Lactobacillus [↓]	Qiu et al. (61)
Firmicutes†,Bacteroidetes†, and Actinobacteria↓ Bacteroidaceae†,Veillonellaceae†, and Prevotellaceae†, Lachnospiraceae↓,Ruminococcaceae↓,Erysipelotrichaceae↓, Peptostreptococcaceae↓, and Clostridiaceae↓	Zheng et al. (71)
Bacteroidaceae [↑] , and Veillonellaceae [↑] Lachnospiraceae [↓] ,Erysipelotrichaceae [↓] ,Ruminococcaceae [↓] , Peptostreptococcaceae [↓] ,Coriobacteriaceae [↓] , and Clostridiaceae ^{1↓}	Tan et al. (72)

As one of beneficial microbial metabolites, short-chain fatty acids have been reported to have anti-inflammatory properties and be involved in the regulation of inflammatory responses (76-78). The total SCFAs content of patients with MG was reported to be lower than that of the HC cohort, with propionate and butyrate being significantly decreased (61). In another study, there was no difference in short chain fatty acids profiles between MG and controls (70). Taking into consideration the limited sample size and great inter-individual variability, this finding awaits further verification. Based on a previous finding proposed by Tan et al. that different subtypes of MG have different microbiota composition patterns (72), we can speculate that fecal metabolites were different among different subtypes. Overall, all the above mentioned studies demonstrated the presence of dysregulated gut microbiota and microbial metabolites in patients with MG, thus providing a new entry point into the pathogenesis of MG and helping us understand the full picture of myasthenia gravis to propose new therapeutic strategies.

To probe whether disturbed gut microbiota is involved in the occurrence and development of MG, fecal microbiota transplantation (FMT) was conducted by Peng et al. (71).

In experimental procedures, germ-free (GF) mice were implanted with the microbiota of patients with MG or healthy volunteers; subsequently these mice were immunized in a classical murine model of MG. Thus, MMb mice, HMb mice, and CMb mice were obtained, respectively. Compared to the mice colonized with HMb, the level of interferon gamma, tumor necrosis factor alpha, and IL-10 in the serum of MMb mice significantly increased. The MMb mice were observed to exhibit substantially impaired motor ability. Furthermore, these changes were reversed by colonization in the CMb mice. Taken together, these findings indicated that gut microbiota may probably be involved in the occurrence and development of MG. Moreover, experimental data have shown that the gut flora and its metabolites in MMb mice and patients with MG have roughly the same characteristics (71), suggesting that the gut microbiota may participate in the initiation and progress of MG by affecting host metabolism. This study provided novel knowledge on the

pathogenesis of MG and a new scientific basis for the diagnosis and treatment of MG (79).

GUT MICROBIOTA SHAPES THE Th17/ Treg BALANCE

The gut microbiome contributes importantly to the induction, maturation, and maintenance of the host immune system. Gut microbiota organisms could impact the balance of Th17/Treg cells, as reported by several studies on antibiotic-treated or germ-free animals (80–83). In contrast, metabolites produced by gut microbiota, particularly SCFAs have an effect on the induction of Tregs mediated by gut microbiota (79, 84, 85). However, certain gut microbes participate in the processes of the expansion and differentiation of immune cells (86, 87).

Short-Chain Fatty Acids

SCFAs, a group of single-chain fatty acids with lengths less than 6 carbon chains, are produced by gut microbiota through the fermentation of dietary fiber, resistant starch, and other complex carbohydrates, as well as mucin (88, 89). These acids principally induce acetate, propionate, and butyrate, with an approximate molar ratio of 60:20:20 (90). The proportions of SCFAs can change depending on the diet, age, and diseases (91). There are differences in the generative process of the three most common SCFAs. Specifically, acetate is primarily produced by Bifidobacteria belonging to the Actinobacteria phylum during carbohydrate fermentation. Propionate is mainly generated by Bacteroides spp., and several Firmicutes bacteria *via* the succinate pathway (92). Butyrate is mostly produced by Clostridial clusters IV and XIVa *via* the pyruvate metabolism pathway (93).

In addition to providing an energy source for intestinal mucosal cells, SCFAs are important immune signaling molecules that influence the development of immune responses locally and distally. Multiple studies have demonstrated that SCFAs stimulate the differentiation and development of Tregs. Both acetate and propionate reduce the onset of allergic airway disease by upregulating the number of Tregs and enhancing Tregs function (94, 95). Furthermore, propionate is thought to be involved in the occurrence of allergic asthma by altering the gut microbiota and thus affecting the number and function of helper T (Th) cells (94). Extracellularly, SCFAs can serve as ligands for cell-surface G protein-coupled receptors (GPCRs) to indirectly regulate immune body function (96). Intracellularly, SCFAs regulate gene transcription by inhibiting histone deacetylases (HDACs) and enhancing histone acetyltransferase activity, thereby directly regulating immune function (97).

GPR43 is found on various cells, including macrophages, neutrophils, mast cells, epithelial cells and dendritic cells (DCs), in the intestine, especially in the colon (98–100). GPR43 on colonic T-cells have an impact on the induction and enhancement of the inhibitory effect of Tregs through epigenetic modifications (101). Propionate can reportedly increase the number of colonic Tregs, an effect dependent on GPR43 (101). GPR109A is present on macrophages and dendritic cells in addition to the intestinal epithelium and acts as a receptor for both butyrate and niacin. Niacin is not present at concentrations high enough to activate the receptor under physiologic conditions (102, 103). In the colonic lumen, a high concentration of butyrate produced by the microbiota acts as an endogenous agonist of GPR109A (104). In a Niacr1-/- (the gene encoding for GPR109A) mice study, it was observed that dendritic cells and colonic macrophages from mice suppressed the differentiation and proliferation of Tregs (105). GPR41 is widely expressed in the intestine, adipose tissue, and peripheral nervous system. SCFAs may also regulate the inflammatory response to maintain organismal health through the activation of GPR41. SCFAs inhibit dendritic cell proliferation and ameliorated allergic respiratory inflammation in wild-type mice, which did not occur in GPR41 knock-out mice (106). Briefly, the GPR signaling pathway is involved in the progression of many immune-related diseases, and SCFAs can play an important role in the regulation of immune response in the whole body through the activation of the GPR signal.

HDACs are a class of enzymes that remove acetyl groups from chromatin, thereby inhibiting transcription (107) and regulating a wide range of cellular functions, such as migration, metastasis (108, 109), and survival (110, 111). SCFAs can effectively suppress histone deacetylases (HDACs). Furthermore, the suppressive effect of SCFAs against HDACs occurs in a concentration-dependent manner (112). As early as 1979, Cousens et al. revealed that high concentrations of butyrate in rat hepatoma cells inhibited histone deacetylation by directly inhibiting HDAC activity (113). In addition to butyrate, propionate inhibited HDAC6 and HDAC9 expression in Tregs and enhanced histone acetylation in mice (101). Among all SCFAs, butyrate demonstrated the strongest inhibitory effects on HDACs. FoxP3 acts as a characteristic marker molecule that maintains the anti-inflammatory effects of Tregs, and HDAC9 can degrade it by affecting the degree of deacetylation of FoxP3 (114). SCFAs can promote the differentiation of Tregs by suppressing HDACs and enhancing their suppressive function. The mechanism may be that SCFAs suppress histone H3 lysine 27 acetylation (H3K27ac) in the CNS1 and CNS3 enhancers of the FoxP3 gene, thus promoting the expression of FoxP3, a process that may require the involvement of GPR43 (79, 101). In conclusion, SCFAs can regulate the differentiation and function of Tregs and modulate body immune tolerance by inhibiting HDACs.

Commensal Gut Bacteria

Bacteroides fragilis, an anaerobic gram-negative gut microbe, activates the TLR signaling pathway to establish a host-microbe symbiotic relationship and influences the development and differentiation of T cells (115, 116). *B. fragilis* exerts its antiinflammatory effects by acting on Tregs (117). Polysaccharide A (PSA) generated by *B. fragilis*, a microbe associated with molecular patterns, has been recognized by toll-like receptor 2 (TLR2) on Tregs to induce FoxP3+ Tregs and limit Th17 response, maintaining intestinal immune tolerance (118). Moreover, *B. fragilis*-derived PSA is recognized by dendritic cells (DCs) in the intestine, which produce IL-10, consequently promoting the production of Tregs (119).

Clostridia, a gram-positive spore-forming gut microbe in the small intestine, inhibits the release of IL-17 through the production of SCFAs, thereby reducing the differentiation to Th1 and Th17 cells and further expressing FoxP3, which exerts a Tregs-like induction effect (120). Clostridia colonizes the mucus layer near intestinal epithelial cells (IECs) (81) and increases the expression of 2,3-dioxygenase and matrix metalloproteinases (MMPs) in the epithelium (81). MMPs are known to be involved in the conversion of TGF- β from the latent to the active form, which promotes the differentiation of initial CD4+ T cells to antigen-specific colonic Tregs (121). Atarashi et al. used chloroform to treat the feces of conventionally reared mice to obtain 46 strains of Clostridium and colonized them in GF mice, inducing a robust accumulation of Tregs in the colons of GF mice (81). Subsequently, Atarashi et al. colonized 17 strains of Clostridia isolated from the human intrinsic microbiota into GF mice and found that they also enhanced Treg abundance. The oral administration of the combination of 17 strains was observed to attenuate allergic diarrhea and colitis in animal models (122).

Segmented filamentous bacteria (SFB) is an anaerobic grampositive spore-forming bacterium (123). SFB is essential for the proliferation and activation of Th17 cells, a process that cannot be separated from the adhesion of SFB to intestinal epithelial cells (124). The direct contact between SFB and intestinal epithelial cells induces the production of serum amyloid A proteins (SAA) and reactive oxygen species, creating an intestinal environment conducive to Th17 cell differentiation (124). SAA induces IL-23 production in DCs; subsequently, IL-23 triggers IL-22 secretion by type 3 innate lymphoid cells. IL-22 activates Stat3, which subsequently increases SAA expression (124, 125). Moreover, various cytokines secreted by CD11c+ cells in response to SAA stimulation, including IL-1β, IL-6, and IL-23, can boost the differentiation of Th17 cells together with TGF- β and SAA, with the latter being a vector of high-density lipoprotein (HDL) and retinol; therefore, it is speculated that SAA can deliver these immunomodulatory molecules to antigenpresenting cells (APCs) and T cells to regulate the body's immune response. Thus, the SFB-mediated differentiation of Th17 cells is achieved through a complex network of interactions among multiple cells and cytokines (Figure 1).

Desulfovibrio is the major sulfate reducing bacteria (SRB) in the human gut (126). It not only inhibits the production and oxidation of butyrate, but also consumes the existing butyrate. Reduced butyrate level affects intestinal epithelial barrier function and promotes the expression of multiple inflammatory components, such as pathogen-associated molecular pattern (PAMP) or microbe-associated molecular pattern (MAMP) factors, jeopardizing the immune system of the body. Therefore, it is reasonable to presume that increased levels of desulfovibrio may contribute to the development of inflammation. When Figliuolo VR et al. examined the role of SRB in the gut immune response in GF mice, and in experimental colitis, they concluded that SRBs were capable of enhancing Th17 response in the mucosal immune system and promoting the occurrence and development of colitis (127).

POTENTIAL THERAPEUTIC POSSIBILITIES

Above all, the occurrence and development of MG is closely related to Th17/Treg cell-mediated immune imbalance. Perturbed microbial ecology is associated with the pathological process of MG, and regulating gut microbiota may improve the Th17/Treg cell imbalance. Therefore, microbiota-targeted interventions may potentially be effective in treating MG. Currently, probiotics, FMT, and other therapies targeting the gut microbiota have developed rapidly and have been widely used in clinical practice. According to relevant researches, novel therapeutic options for MG can be explored from the perspective of adjusting intestinal flora, improving pathological immune response and correcting immunity disorders.

Probiotics

According to the Food and Agriculture Organization of the United Nations and the World Health Organization, probiotics are defined as "live microorganisms which, when administered in adequate amounts, confer a health benefit to the host (128)". Probiotics are known to rebalance microbiota and modulate immune function through multiple mechanisms, including the direct inhibition of enteric pathogen activity, secretion of bacterial proteins by lowering luminal pH, induction of the defense mechanisms of the epithelium, and modification of immune regulation by reducing pro-inflammatory molecules and increasing anti-inflammatory factors (129).

Numerous animal and preclinical studies have demonstrated the potential benefits of probiotics in the therapy of diverse diseases, such as gastrointestinal (130) and neurological disorders (131). Bacterial species that are currently of research value as probiotics mainly belong to the genus Lactobacillus and Bifidobacterium. For example, Park et al. demonstrated the low levels of proinflammatory cytokines produced by Th17 cells in mouse experimental colitis models following the oral administration of Lactobacillus acidophilus. In-vitro, L. acidophilus induced Tregs and promoted the production of IL-10, whereas IL-17 secretion was inhibited in splenocytes (130). These results suggested that L. acidophilus could treat inflammatory bowel disease by regulating the Th17/Treg balance. A study has shown that the oral administration of IRT5 (a mixture of five probiotics) to experimental autoimmune encephalomyelitis mice not only enhanced Treg abundance, but reduced the number of Th17 cells in superficial lymph nodes. IRT5 dramatically suppressed experimental autoimmune encephalomyelitis when used as a pretreatment before disease onset. Moreover, probiotic treatment delayed disease progression (131). Following 8 weeks of orally administering mixed strains of bifidobacteria and lactic acid



bacteria, atopic skin irritation was reduced in atopic dermatitis mouse models (132). Modulating gut microbiota by the ingestion of probiotics has been demonstrated to impact the development of mucosal and systemic immune responses, thereby altering immune profiles and immune homeostasis and positively influencing autoimmune diseases. In recent years, many international studies have demonstrated the use of probiotics to regulate MG. It was observed that the prophylactic administration of IRT5 modulated the disease course and reduced the titer of anti-AChR antibody and the level of certain pro-inflammatory factors in EAMG rats (133). A study with applications of two bifidobacteria or lactobacilli strains in a Lewis rat model also resulted in promising results (30). A subsequent study (134) reported the clinical efficacy of bifidobacterium on the course of EAMG and found that the immunomodulatory properties of viable bifidobacteria relied on the maintenance of the integrity of bacterial proteins and DNA sequences, which is denatured when exposed to high temperatures (135). Together, these studies support the effectiveness of probiotics in different immune-mediated inflammatory conditions and their possible use as a treatment in patients with MG to modulate the immune system and contribute to the restoration of the immunological tolerance to AChR. Taken together, these research studies support the validity of probiotics in different autoimmune and inflammatory disease and their use as an attractive therapeutic intervention for MG to regulate the immune microenvironment.

However, the number of studies on the application of probiotics in MG is currently insufficient, resulting in an incomplete understanding of the mechanisms explaining how probiotics improve immune disorders in myasthenia gravis. In the future, more animal and clinical studies are needed to provide detailed information and reliable evidence on the beneficial effects of probiotics.

Fecal Microbiota Transplantation

FMT refers to the administration of intestinal microbes from a healthy donor into a recipient to reconstruct the intestinal microecology of the recipient (136). If the gut microbiota plays a causal role in disease pathophysiology, altering the microbiota may influence disease course. However, in most cases, a single microorganism is not likely to be a causal pathogen or missing beneficial microbe. Therefore, an advantage of FMT over probiotics is the introduction of a complete healthy gut microbiota. The successful application of FMT in recurrent or refractory Clostridioides difficile infection has gradually became a research hotspot on biomedicine and clinical medicine.

With thorough research, the gut microbiota has been found to play a significant role in the pathophysiology of neurological disorders, and FMT is likely be a promising treatment option. In the field of neurological disease, the positive effect of FMT is supported by several animal studies or described in some case reports. Using the Parkinson's disease (PD) model, Sun et al. demonstrated that FMT reduced gut microbial dysbiosis, decreased fecal SCFAs, and alleviated physical impairment (137). Sun et al. showed that FMT treatment improved cognitive deficits in APPswe/PS1dE9 transgenic mice, which was attributed to the reduction of the brain deposition of amyloid- β (138). There are few studies on the use of FMT in patients with MG. In a study aimed at investigating whether the microbiota participated in MG pathogenesis, the impaired locomotion ability of MMb mice was restored by cocolonizing with HMb, indicating a bright future for such applications in patients with MG (71).

Currently, studies on the application of FMT have been conducted only in animals. However, the immunoreactivity observed in animal models are quite different from that in humans. Thus, a large double-blinded randomized controlled human trial should be performed.

CONCLUSIONS AND PERSPECTIVES

As the most prevalent neuromuscular junction autoimmune disease, MG is characterized by increased numbers of Th17 cells and Treg dysfunctionalities. Regulating the cytokines, receptors, and signal transduction pathways involved in the Th17/Treg immunobalance axis will provide a viable strategy for the treatment of MG. Recently, scientists have gradually realized the role of gut microbiota in affecting the balance of Th17/Treg cells. Accordingly, correcting Th17/Treg imbalances by regulating the composition of the gut microbiota would be beneficial for MG treatment.

Probiotics are a class of active microorganisms that are beneficial to the host, with a favorable effect on the microbial ecological balance of the host. Probiotics contribute to modulating immune responses and fostering immunological surveillance. Certain probiotic strains reportedly promote the production of Tregs. Therefore, probiotics may be an attractive option to treat myasthenia gravis characterized by a deficiency of Treg. It is important to be alert to the fact that certain strains of probiotics may exacerbate the disease; therefore, it is critical to fully characterize the immunological profile and dosage of probiotics to exert their beneficial effects.

FMT has an advantage over probiotics due to the introduction of a complete healthy gut microbiota. Due to the

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successful application of FMT in different areas of biomedicine, it is also currently being investigated in the domain of neurological disease with dysbiosis of gut microbiota.

Considering the low cost and high efficiency of FMT, the application of FMT for diseases with Th17/Treg cells imbalances is practical at the theoretical level. However, this therapeutic approach is unpredictable, and a single species microorganism transfer may be more effective; however, the detailed elucidation of the mechanisms involved is needed.

AUTHOR CONTRIBUTIONS

PC and XT conceived the review. PC wrote the manuscript. XT revised the manuscript. All authors contributed to the article and approved the submitted version.

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