

TREM-1 Modulation Strategies for Sepsis

Sara Siskind^{1,2,3}, Max Brenner^{1,2,3*†} and Ping Wang^{1,2,3*†}

¹ Center for Immunology and Inflammation, The Feinstein Institutes for Medical Research, Manhasset, NY, United States, ² Department of Surgery, Zucker School of Medicine at Hofstra/Northwell, Manhasset, NY, United States, ³ Elmezzi Graduate School of Molecular Medicine, Manhasset, NY, United States

OPEN ACCESS

Edited by:

Daniela Novick, Weizmann Institute of Science, Israel

Reviewed by:

Vikrant Rai, Western University of Health Sciences, United States Alexander B. Sigalov, SignaBlok, Inc., United States

*Correspondence:

Ping Wang pwang@northwell.edu Max Brenner mbrenner@northwell.edu

[†]These authors have contributed equally to this work and share senior authorship

Specialty section:

This article was submitted to Inflammation, a section of the journal Frontiers in Immunology

Received: 29 March 2022 Accepted: 17 May 2022 Published: 15 June 2022

Citation:

Siskind S, Brenner M and Wang P (2022) TREM-1 Modulation Strategies for Sepsis. Front. Immunol. 13:907387. doi: 10.3389/fimmu.2022.907387 The triggering receptor expressed on myeloid cells-1 (TREM-1) is a pattern recognition receptor, which can be upregulated in inflammatory diseases as an amplifier of immune responses. Once activated, TREM-1 induces the production and release of proinflammatory cytokines and chemokines, in addition to increasing its own expression and circulating levels of the cleaved soluble extracellular portion of TREM-1 (sTREM-1). This amplification of the inflammatory response by TREM-1 has now been considered as a critical contributor to the dysregulated immune responses in sepsis. Studies have shown that in septic patients there is an elevated expression of TREM-1 on immune cells and increased circulating levels of sTREM-1, associated with increased mortality. As a result, a considerable effort has been made towards identifying endogenous ligands of TREM-1 and developing TREM-1 inhibitory peptides to attenuate the exacerbated inflammatory response in sepsis. TREM-1 modulation has proven a promising strategy for the development of therapeutic agents to treat sepsis. Therefore, this review encompasses the ligands investigated as activators of TREM-1 thus far and highlights the development and efficacy of novel inhibitors for the treatment of sepsis and septic shock.

Keywords: sepsis, inflammation, TREM-1, shock, DAMP

INTRODUCTION

Sepsis is a complex disorder, defined by a dysregulated host response to infection leading to organ dysfunction (1, 2). The pro-inflammatory response to invading pathogens is initiated by pattern recognitions receptors (PRRs) located on the surface and intracellularly of immune and non-immune cells, that recognize pathogen-associated molecular patterns (PAMPs), molecules released from pathogens, and damage-associated molecular patterns (DAMPs), molecules released from damaged cells (3, 4). Activation of PRRs on innate immune cells initiates microbicidal and pro-inflammatory responses to contain and eliminate the invading pathogens and activates the adaptive immune response, particularly T lymphocytes (3). Toll-like receptors (TLRs) are a well-studied family of PRRs for their role in innate immunity that amplify the inflammatory response synergistically with triggering receptor expressed on myeloid cells-1 (TREM-1), a more recently characterized PRR (5–8).

1

TREM-1 was first identified on circulating neutrophils and monocytes in response to lipopolysaccharide (LPS), a glycan mostly present on the surface of Gram-negative bacteria that strongly activates the host immune response (9-11). TREM-1 is part of a family of TREM receptors that also includes TREM-2, TREM-3, and TREM-like transcript-1 and 2 (TLT-1, TLT-2) (12). TREM-3 is only expressed in mice, and like TREM-1, is upregulated in response to LPS. TREM-2, on the other hand, is downregulated in response to LPS and, upon activation, attenuates the inflammatory response (12, 13). Activation of TREM-1 in neutrophils and monocytes triggers the release of pro-inflammatory cytokines and chemokines, in addition to upregulating the gene expression of TREM-1 and surface expression of cell activation markers (9). This amplification of the inflammatory response by TREM-1 has gained interest as a critical contributor to the dysregulated immune response in sepsis (4). Patients admitted to the ICU with septic shock have higher surface expression of TREM-1 on monocytes and elevated circulating levels of the cleaved soluble extracellular portion of TREM-1 (sTREM-1) (14, 15). Elevated levels of circulating sTREM-1 have also been associated with increased mortality in patients with septic shock (16, 17). Additionally, mice genetically deficient in TREM-1 had less severe disease in response to multiple pathogens without affecting their ability to clear the infection, resulting in similar pathogen loads (18). The inhibition of TREM-1 remains a promising target for development of therapeutic agents to treat sepsis. This review aims to give an overview of the role of TREM-1 and its endogenous ligands in sepsis, and the development of novel inhibitors of this pathway and their efficacy for the treatment of sepsis.

STRUCTURE AND SIGNALING PATHWAY

TREM-1 is a member of the immunoglobin superfamily, a group of cell surface receptors with related extracellular Ig-like domains (19, 20). In addition to the extracellular Ig domain, it consists of a transmembrane region with a conserved lysine residue, and a short cytoplasmic domain that does not contain a signaling motif (21, 22). Propagation of signaling is instead dependent on association with the immunoreceptor adaptor protein DNAX activation protein 12 (DAP12). TREM-1 contains a positively charged transmembrane lysine residue that associates with a negatively charged aspartate residue of DAP12 (21-24). Upon receptor activation, the immunoreceptor tyrosine-based activation motif (ITAM) of DAP12 is phosphorylated, signaling the recruitment and activation of spleen tyrosine kinase (Syk), a nonreceptor tyrosine kinase (20). Syk activates multiple downstream signal transduction pathways including the PI3K/Akt pathway, the Ras/ERK/MAPK pathway, NF-KB signaling, and phospholipase C phosphorylation, leading to increased intracellular calcium and proinflammatory cytokine secretion (19-22, 25).

TREM-1 activation independently triggers downstream inflammatory cascades that can synergize with TLR signaling pathways (22, 25). TLR4 activation upregulates the expression of

TREM-1, and the concomitant activation of TREM-1 and TLR4 leads to a synergistic increase in proinflammatory cytokine and chemokine release by 25-fold compared with that of TLR4 activation alone (8, 26–28). One proposed mechanism for their synergistic proinflammatory response is that TREM-1 increases the availability of TLR4 downstream signaling molecules such as MyD88, CD14, NF- κ B, and I κ B α (26). When TREM-1 s blocked, RAW cells stimulated with LPS maintain TLR4 expression but have decreased genetic expression of these signaling molecules as well as inflammatory cytokines (21, 26, 29). Understanding the structure and signaling pathway of TREM-1 remains an important focus of research to better elucidate its role in sepsis and to develop novel therapeutic targets.

TREM-1 LIGANDS

The identification of TREM-1's endogenous ligands is crucial for studying the role of TREM-1 in the pathogenesis of sepsis. Moreover, knowing TREM-1's endogenous ligands and their structure offers critical insights to develop TREM-1-targeting pharmacological strategies to reduce sepsis hyperinflammation, such as the inhibitory peptides discussed below. Although TREM-1 was characterized in 2000, it took 14 years until the first ligand was identified. Since then, multiple ligands have been implicated as activators of TREM-1, furthering our knowledge on this receptor's role in innate immunity, sepsis, and noninfectious inflammatory diseases.

Peptidoglycan Receptor Protein 1 (PGLYRP1)

PGLYRP1 (also named Tag7) was one of the first identified ligands of TREM-1 (30). PGLYRP1 is an antimicrobial protein that is secreted from polymorphonuclear leukocyte granules in response to infection (5, 30, 31). It subsequently binds to peptidoglycan and LPS, essential components of the bacterial membrane, where it induces lethal membrane depolarization and oxidative stress to the bacteria (32–34). After it was discovered that an endogenous, unknown ligand of TREM-1 exists on bacterially activated peritoneal neutrophils, PGLYRP1 was investigated as a potential ligand (30, 35). Neutrophils were stimulated with peptidoglycan and cross-linked to sTREM-1 and, using mass spectrometry, identified that the resulting complexes contained PGLYRP1 peptides (30).

PGLYRP1 as a ligand for TREM-1 was further studied using affinity chromatography, which demonstrated conclusively that immobilized soluble TREM-1 binds PGLYRP1 (36). It has also been shown that PGLYRP1 is an activating ligand of TREM-1 on monocytes and induces the development of cytotoxic lymphocyte subpopulations in peripheral blood mononuclear cells (PBMCs) (37). In the presence of LP17, an inhibitory peptide of TREM-1 discussed further below, monocytes treated with PGLYRP1 failed to induce cytotoxic lymphocyte transformation and had reduced secretion of IL-2, thus providing further evidence that PGLYRP1 is an endogenous ligand for TREM-1 (36, 37). Interestingly, soluble PGLYRP1 alone does not activate TREM-1. Only when complexed with peptidoglycan, anchored to HEK293 cell surface, or bound to a plate, was PGLYRP1 able to elicit an inflammatory response *via* TREM-1 (30). This should be kept in mind for future research on PGLYRP1/TREM-1 interaction, as activation may only be reproducible when TREM-1 is bound to a cell surface.

Extracellular Cold Inducible RNA Binding Protein (eCIRP)

eCIRP is a DAMP that has recently been identified as an endogenous ligand of TREM-1. It was originally discovered in the blood of critically ill septic and trauma-hemorrhage surgical patients. CIRP is a ubiquitously expressed nuclear protein which has been mostly studied in macrophages, lymphocytes, and neutrophils, which release it into circulation during periods of hypoxic and mild hypothermic stress, such as in sepsis (38, 39). Mice injected with recombinant murine CIRP (rmCIRP) developed a sepsis-like acute lung injury with vascular endothelial cell damage, leukocyte infiltration and increased pro-inflammatory cytokine production (40). Using surface plasmon resonance (SPR), it was determined that rmCIRP binds to recombinant murine TREM-1 (rmTREM-1) with strong affinity, with a K_D of 11.7×10^{-8} M (41). Binding was further demonstrated between rmCIRP and surface TREM-1 on murine RAW264.7 and peritoneal macrophages using fluorescence resonance energy transfer (FRET) analysis. rmCIRP-injected mice had attenuated systemic and pulmonary inflammatory response after treatment with the TREM-1 inhibitor LP17. Additionally, mice deficient in TREM-1 had reduced serum IL-6 and IL1- β after rmCIRP injection, indicating eCIRP acts an endogenous ligand of TREM-1 (41).

High Mobility Group Box 1 (HMGB1)

HMGB1 has also been investigated as an endogenous ligand to TREM-1 (42). HMGB1 was originally identified as a nuclear DNA-binding protein that functions as a cofactor in transcription regulation. It was later found that it can be released from a variety of cells and function as a DAMP, signaling through activation of TLRs and receptor for advanced glycation end products (RAGE) to induce a proinflammatory response (43-45). More recently, using a murine model of hepatocellular carcinoma, HMGB1 released from necrotic hepatocytes was found to bind TREM-1 using immunoblotting and SPR, with a binding K_D of 35.4 x 10⁻⁶ M (42, 46). HMGB1 has also been shown to induce inflammatory responses through TREM-1 activation in THP-1 human monocytic cells, a human monocyte cell line, and upregulate TREM-1 expression on macrophages (47, 48). However, recombinant HMGB1 alone was insufficient to induce TREM-1-regulated M1 polarization, suggesting it may need coactivating molecules to fully trigger TREM-1, or that recombinant HMGB1 is different from endogenous HMGB1 (33, 49).

Heat Shock Protein 70 kDa (Hsp70)

Along with HMGB1, the 70 kDa heat shock protein (Hsp70) was found to be released from necrotic cells and augment the

proinflammatory response through TREM-1 activation (42, 47). Under normal conditions, HSP70 is a molecular chaperone that participates in the maintenance of protein homeostasis through folding and remodeling processes (50). Normally expressed at low levels, Hsp70 becomes upregulated in pro-inflammatory states like bacterial endotoxemia (47). In addition to Hsp70's primary role of protein folding stabilization, there has been growing interest in its function as a pro-inflammatory mediator after it was found to upregulate TNF- α , IL-1 β , and IL-6 in human monocytes (51). Hsp70 was then investigated as a novel ligand for TREM-1 using LPS- and necrotic cell lysate-stimulated THP-1 cells. These cells, when treated with an anti-HSP70 antibody, had a reduction in expression of TNF- α , IL-6, and IL-8 that was further reduced by the addition of an inhibitory recombinant TREM-1 fusion chimera (47). This data suggests that Hsp70 is released from necrotic cells to aid pro-inflammatory responses in monocytes through TREM-1 activation of the cytokine expression cascade (47). Hsp70 as a ligand of TREM-1 was further studied using affinity chromatography and demonstrated to bind to sTREM-1 immobilized on a CNBr-Sepharose column as well as to TREM-1 on the monocyte surface (52). Interestingly, however, it was found that Hsp70 released from necrotic Kupffer cells along with HMGB1 in a murine model of hepatocellular carcinoma, does not directly bind to TREM-1, suggesting that HMGB1 may have a stronger affinity and compete for binding on TREM-1 (42).

Extracellular Actin

Actin is one of the most abundant proteins within cells, functioning in the form of filaments that polymerize to aid in cell morphology and motility (53). When cells undergo apoptosis and necrosis, such as during sepsis, actin is released and has deleterious effects once extracellular (53). After it was determined that an unknown ligand expressed on platelets activates TREM-1 in sepsis, further investigation using gel analysis of platelet total protein and rTREM-1 suggested the ligand to be actin (54, 55). Confocal microscopy was used to confirm the co-localization of TREM-1 and actin on RAW267.7 cells treated with LPS-stimulated platelets and LPS with recombinant actin. This co-localization was also demonstrated in vivo using a CLP model of polymicrobial sepsis. Actin and TREM-1 had increased expression and co-localized in the lungs of septic mice (55). Additionally, actin dose-dependently enhanced LPS-stimulated release of TNF-α from RAW267.7 cells and peritoneal macrophages (55). This response was blunted in cells treated with the TREM-1 inhibitor LP17 as well as in peritoneal macrophages isolated from TREM-1 knockout mice, strongly suggesting that actin activates inflammatory cells via TREM-1 (55).

IMMUNE RESPONSES IN TREM-1 KNOCKOUT MICE

Mice genetically deficient in TREM-1 are another valuable tool to help uncover the potential benefits of modulating TREM-1 for

the treatment of sepsis. To study this, multiple knockout models with different genetic modifications have been developed: TREM-1 knockout, endothelial cell-specific TREM-1 (endoTREM-1) knockout, TREM-1/3 double knockout, and TREM-like transcript-1 (TLT-1) knockout. To determine the effects of TREM-1 deficiency on the immune response to infection, TREM-1-/- mice were infected with Leishmania major (18). Compared to WT mice, TREM-1^{-/-} mice had reduced infiltration of neutrophils and decreased lesion size around the site of bacterial inoculation (18). TREM-1-/- mice also had reduced morbidity after infection with influenza A virus, measured by body weight and temperature, and decreased IL-6 in bronchoalveolar lavage (18). Importantly, while infectionrelated pathologies were improved, TREM-1^{-/-} mice had equivalent pathogen clearance of L. major, influenza virus, and Legionella pneumophila as WT mice, indicating these mice do not develop disseminated infection like TREM-1/3^{-/-} (18). Additionally, after LPS-induced septic shock, TREM-1-/- mice had decreased neutrophil extracellular trap (NET) release in the serum and lungs, which have pro-inflammatory functions that contribute to the progression of septic shock (56). Aortas and mesenteric arteries isolated from TREM-1^{-/-} mice were protected from in vitro NET-induced vascular dysfunction (possibly due to their inability to respond to the eCIRP present in NETs), maintaining normal contraction and relaxation (56-58). The involvement of TREM-1 in sepsis induced vascular dysfunction was further proven using endoTREM-1^{-/-} mice (57). After cecal ligation and puncture (CLP)-induced sepsis, endoTREM-1^{-/-} mice had restored vasorelaxation in addition to reduced serum VCAM-1 and IL-6, and prolonged 7-day survival (57).

In mice, the gene for TREM-3 is adjacent to TREM-1 and likely occurred from a duplication event, demonstrated by their high homogeneity (59). Additionally, the two receptors are both amplifiers of the immune response, and likely work synergistically (59-61). TREM-3 in humans, however, is a pseudogene and has no functional overlap with TREM-3 in mice (61). Therefore, it has been suggested that TREM-1/3^{-/-} mice might better reflect TREM-1 deficiency in humans than mice deficient in TREM-1 alone (61-63). Interestingly, although TREM-1^{-/-} mice were protected after exposure to infectious agents, TREM-1/3^{-/-} mice had worse outcomes after infection. TREM-1/3^{-/-} mice had increased mortality and bacterial dissemination after infection with Streptococcus pneumoniae and Klebsiella pneumoniae, despite infected primary TREM-1/ 3^{-1-} macrophages having decreased cytokine release (64, 65). The relevance of murine TREM-3 to human TREM-1 function should continue to be explored to better characterize which murine models best emulate human disease.

TLT-1 belongs to the TREM family, and its gene resides in the human TREM gene cluster, along with TREM-1, -2, -3, and TLT-2 (66). It is specific to platelets and megakaryocytes and, upon platelet activation, translocates to the cell surface where it plays a role in hemostasis/thrombosis (66). Additionally, a soluble fragment of TLT-1 is present in the circulation and is believed to function as an endogenous TREM-1 inhibitor (67, 68). TLT-1 knockout mice (treml-1^{-/-}) had increased gene and protein

expression of inflammatory cytokines in the lungs and plasma after CLP induces sepsis (68). Additionally, septic treml-1^{-/-} mice had increased mortality, indicating a protective role of TLT-1 in polymicrobial sepsis, likely by inhibiting TREM-1 (68).

TREM-1 BLOCKADE AS A THERAPEUTIC APPROACH

Studies using TREM-1 knockout mice have demonstrated that this receptor plays a critical role in the progression of sepsis. Armed with this knowledge, multiple receptor antagonists have been created, predominately based on the structure of TREM-1 and its ligands. Modulation of TREM-1 activation with many of these inhibitors has been demonstrated as an effective approach for attenuating sepsis severity (**Table 1**).

Soluble TREM-1 (sTREM-1)

The sTREM-1 molecule is the cleaved extracellular domain of TREM-1 that was found to be increased in the blood during sepsis (69, 73, 84, 85). In addition to being studied as a prognostic indicator of sepsis severity, it has also been proposed to act as a decoy receptor for TREM-1 by binding its ligands and reducing its activation and the subsequent pro-inflammatory cytokine release (73, 86–89) (**Figure 1**). To test this hypothesis, the gene for recombinant porcine sTREM-1 was transfected into *Escherichia coli* for cloning and expression (69). After purification, it was administered to LPS-stimulated primary porcine alveolar macrophages. Macrophages treated with sTREM-1 had reduced mRNA expression of type-I and type-II inflammatory cytokines, including TNF- α , IL-1 β , IL-2, IL-4, IL-8, IL-10, IL-12, IL-16, and IL-18 (69).

A similar method of genetic cloning was used to express and purify murine sTREM-1, which was named recombinant extracellular domain of TREM-1 (rTREM-1) (90). Mice were then infected with Streptococcus suis, a bacterium that can rapidly cause streptococcal toxic-shock-like syndrome (STSLS), followed by treatment with rTREM-1. Conversely to the in vitro study using porcine sTREM-1, mice inoculated with S. suis and treated with rTREM-1 had worse outcomes. rTREM-1 treatment was associated with increased 7-day mortality, and elevated IL1- β , TNF- α , KC, and bacterial load in both blood and peritoneal fluid (90). Interestingly, however, when rTREM-1 treatment is combined with antibiotics, mice infected with S. suis had better outcomes than with antibiotics alone (70). Mice that received combination treatment of rTREM-1 and ampicillin had a higher 7-day survival, lower serum levels of IL1- β and TNF- α , and attenuated acute lung injury compared to mice that received PBS vehicle, just ampicillin, or rTREM-1 alone (70). These data suggest that, while TREM-1 agonism may help clear some bacterial infections, it is also a key contributor to the detrimental inflammatory processes that culminate in acute lung injury. Many of the following TREM-1 inhibitory peptides developed as a treatment modality for sepsis are based on amino acid (aa) sequences found in sTREM-1, further

TABLE 1 | TREM-1 pathway inhibitory strategies.

Treatment	Sepsis Model	Cell/Animal	Effect	Ref.
sTREM-1	LPS	Macrophages	↓ mRNA of inflammatory cytokines	(69)
	S. suis intraperitoneal	Mice	\downarrow serum TNF- $\alpha,$ IL-1 $\beta,$ \downarrow acute lung injury, \uparrow survival (when given with antibiotics)	(70)
TREM-1 Fc fusion protein	LPS	Monocytes	↓ release TNF-α, IL-1β	(8)
	P. aeruginosa	Macrophages	↓ release TNF-α, IL-1β, MCP-1	(71)
	S. pyogenes	Neutrophils	\downarrow release IL-6, TNF- α	(72)
	LPS intraperitoneal	Mice	\downarrow serum TNF- α , IL-1 β , \downarrow recruitment of peritoneal macrophages and neutrophils, \uparrow survival	(8)
	CLP	Mice	↑ survival	(8)
	E. coli intraperitoneal	Mice	↑ survival	(8)
	S. pyogenes intravenous	Mice	↓ serum IL-6, TNF-α, ↑ survival	(72)
	P. aeruginosa intraperitoneal	Mice	↓ serum IL-1β, TNF-α, MCP-1, ↑ survival	(71)
LP17	LPS	Monocytes	↓ release TNF-α, IL-1β	(73)
	E. coli	Neonatal leukocytes	↓ release TNF-α, IL-6, IL-8	(74)
	LPS intraperitoneal	Mice	↑ survival	(35)
	CLP	Mice	↓ serum TNF-α, IL-1β, ↑ survival	(73)
	S. pyogenes intravenous	Mice	↑ survival	(72)
	LPS intraperitoneal		↑ hemodynamics, ↓ serum TNF-α, IL-1β	(35)
	CLP	Rats	↑ hemodynamics, ↓ TNF-α, IL-1β, IL-6, ↑ survival	(35, 75)
	P. aeruginosa intratracheal	Rats	\downarrow serum and broncoalveolar lavage TNF- α , IL-1 β , IL-6, lactic acidosis, hypoxia, \uparrow hemodynamics, \uparrow survival	(76)
GF9	LPS	Macrophages	↓ release TNF-α, IL-1β, IL-6	(77)
	LPS intraperitoneal		↓ serum TNF-α, IL-1β, IL-6, ↑ survival	(77)
SLC-TREM-1	LPS	Endothelium	↓ TREM-1 expression, ↓ MCP-1, IL-8 release	(78)
	CLP	Mice	↑ survival	(78)
M3	LPS intraperitoneal		↓ serum TNF-α, IL-6, ↑ survival	(41)
	CLP	Mice	↓ serum AST, ALT, TNF-α, IL-6, ↓ acute lung injury, ↑ survival	(41)
	Cecal Slurry	Neonatal Mice	\downarrow serum TNF-α, IL-1β, IL-6, INFγ, \downarrow cardiac and pulmonary IL-1β, IL-6, \downarrow cardiac dysfunction, \uparrow survival	(79)
N1 LR17/LR12	LPS	Mononuclear Cells/	↓ mRNA TNF-α, IFNγ, IL-1β, and IL-6	(80)
	LPS intrabronchial	Monocytes Mice	Leonum INEV II. 4. Llung domogo	(80)
	LPS	Granulocyte	\downarrow serum INFy, IL-4, \downarrow lung damage	• •
	LPS	,	\downarrow mRNA and protein TNF-α, IL-8, IL-10, \downarrow ROS production \downarrow release TNF-α	(23)
		Monocytes		(23)
	LPS intraperitoneal CLP	Mice Mice	↓ serum TNF-α, IL-6, IL-10, ↑ survival	(23)
			↓ cytokines in serum, peritoneal and bronchoalveolar fluid, liver, lung, ↓ acute lung injury, ↓ bacteria in spleen, blood, ↓ TREM-1 expression, ↑ survival	(23, 57)
	Cecal Slurry	Pigs	↑ cardiac function, \downarrow coagulopathy, \downarrow failure of lungs, liver, kidney, ↑survival	(68)
	LPS intravenous	Monkeys	↑ hemodynamics, ↓ serum cytokines	(81)
Nangibotide	Sepsis	Humans	\downarrow systemic IL-6, \downarrow SOFA scores, minimal side effects	(82, 83)

↓- decreased; ↑- increased.

demonstrating that sTREM-1 is effective as an anti-inflammatory mediator (88).

TREM-1 Fc Fusion Protein

TREM-1/Fc, a fusion protein consisting of the extracellular domain of mouse TREM-1 and the Fc portion of human IgG1, was developed as a TREM-1 decoy receptor that can also promote the clearance of TREM-1 ligands from the circulation *via* Fc-receptor mediated endocytosis (8) (**Figure 1**). Mice treated with TREM-1/Fc prior to LPS-induced endotoxemia had decreased serum TNF- α and IL-1 β and recruitment of peritoneal neutrophils and macrophages (8). Additionally, mice treated with TREM-1/Fc had improved 7-day survival in LPS,

CLP, and *Escherichia coli*-induced sepsis (8). These findings were reproduced in another study using intraperitoneal administration of *Pseudomonas aeruginosa* to induce sepsis (71). Treatment with TREM-1/Fc in these mice led to improved survival and a significant reduction in serum pro-inflammatory cytokines, including TNF- α , IL-1 β , INF- γ , and MCP-1 (71). Additionally, TREM-1/Fc provided protection in mice against sepsis after intravenous (i.v.) injection of *Streptococcus pyogenes* (72). Mice treated 2 hours before and 2 hours after inoculation of *S. pyogenes* had prolonged survival and decreased levels of IL-6 and TNF- α in the serum (72). In addition to its TREM-1 inhibitory effects, it is possible that the TREM-1/Fc fusion protein also enhances the phagocytic

5



interaction of TREM-1 with its signaling partner DAP12. (2) LR12/nangibotide, LP17, and TREM-1/Fc fusion protein act as decoy receptors and compete for binding with naturally occurring activating ligands. (3) Together with M3 and N1, LP17 also binds to TREM-1 and competitively inhibits ligand binding. M3 competitively inhibits TREM-1 binding to and activation by extracellular cold inducible RNA binding protein, while N1 inhibits TREM-1 activation by PGLYRP1 and HSP70. (4) soluble TREM-1 is generated from proteolytic cleavage of membrane bound TREM-1 by matrix metalloproteinases. Circulating soluble TREM-1 competitively binds TREM-1's ligands and prevents further activation. All inhibitors prevent the downstream signaling cascade that upregulates the translation of inflammatory cytokines and TREM-1 receptor. *Image created with BioRender*.

clearance of pathogens by bridging peptidoglycan-bound PGLYRP1 on the surface of bacteria with Fc-receptors on macrophages and neutrophils.

LP17

LP17 was one of the first peptides developed to inhibit TREM-1 (73). It has a 17-aa sequence (LQVTDSGLYRCVIYHPP) derived from on a highly conserved extracellular domain of TREM-1 in both mice and humans, and is intended to operate through a mechanism analogous to that of decoy receptors (35, 73, 76). To demonstrate its intended function, flow cytometry was used to show LP17 inhibits the binding of a fluorophore conjugatedmTREM-1/IgG1 antibody to TREM-1 on murine peritoneal exudate cells (35). There is evidence that it functions both as a direct competitive inhibitor by binding to the receptor and thus preventing its activation by the ligand, and as a decoy receptor by binding TREM-1's ligands before they are able to activate the TREM-1 receptor (4, 68) (Figure 1). Efficacy was initially demonstrated by reducing TNF- α and IL-1 β release from LP17-treated monocytes after stimulation with LPS in a dose dependent manner (73). LP17 was further tested in vivo in mice subjected to either endotoxemia or CLP-induced sepsis (73). Mice treated with LP17 had decreased levels of serum TNF-a and IL-1 β and improved survival after LPS injection and CLP (73). LP17 improved survival in endotoxemic mice when given 1 hour before or 4 hours after LPS injection (35, 73). Additionally, LP17 has been shown to improve survival in mice after i.v. inoculation with *Streptococcus pyogenes* (72).

LP17 has also been shown to be efficacious in treating septic rats. Rats treated with LP17 1 hour after LPS injection had attenuated sepsis severity, measured by improved hemodynamic parameters including mean arterial pressure, aortic blood flow, mesenteric blood flow, pH, and serum lactate (35). Additionally, these rats had lower concentrations of serum TNF- α , IL-1 β , and nitrates/nitrites (35). Rats treated with LP17 after CLP also had improved hemodynamic parameters, serum TNF- α , IL-1 β , IL-6 and nitrates/nitrites, and survival at 48 hours and 7 days (35, 75). LP17 has also been shown to be an effective treatment for sepsis secondary to *Pseudomonas aeruginosa* pneumonia in rats, resulting in with improved hemodynamic status, attenuated lactic acidosis and hypoxemia, reduced serum TNF- α , IL-1 β , and IL-6, and improved 7-day survival of the septic rats (76).

In addition to the studies in adult rodents, LP17 has been considered as a potential therapeutic agent for neonatal sepsis (74). In an *in vitro* study, leukocytes isolated from umbilical cord blood from full term human neonates and treated with LP17 had

decreased production of TNF- α , IL-6, and IL-8 after exposure to *Escherichia coli* (74).

М3

M3 is another inhibitory peptide of TREM-1 that was developed to specifically inhibit activation by its novel ligand, eCIRP (41). Its 7-aa sequence (RGFFRGG) was designed based on an area of homology between PGLYRP1 and CIRP (30, 41). Using a FRET assay, M3 was shown to dramatically abrogate CIRP's binding to TREM-1 in both murine peritoneal macrophages and RAW264.7 cells (41) (**Figure 1**). To test this peptide *in vivo*, mice were subjected to LPS-endotoxemia and those treated with M3 had decreased levels of serum TNF- α and IL-6, and improved 7-day survival (41). M3 was also protective in mice with CLP-sepsis, measured by decreased levels of serum AST, ALT, TNF- α and IL-6, and attenuated severity of sepsis-associated acute lung injury (41, 91). Mice treated with intraperitoneal (i.p.) M3 at the time of CLP or 90 minutes later also showed a 10-day survival benefit (41).

Like LP17, M3 has also been shown to be effective in treating sepsis in neonates (79). Using a cecal slurry model of sepsis, neonatal mice treated with i.p. M3 had attenuated disease severity, as measured by a reduction in serum IL-6, TNF- α , IL-1 β , and IFN- γ , and improved pulmonary and cardiac inflammation (79). Additionally, M3-treated neonates had improved cardiac function, measured by cardiac output and left-ventricular end diastolic diameter. Importantly, neonatal mice treated with M3 at either the time of cecal slurry injection or 2 hours later had improved 7-day survival (79). Although M3 is a novel peptide requires further studies, it shows promise in treating sepsis in both neonates and adults.

N1

N1 is a 10-aa inhibitory peptide based on the N-terminal aa sequence (aa 77-86) of PGLYRP1 (80). Using affinity chromatography, N1 was shown to bind sTREM-1 immobilized on CNBr-activated Sepharose (80). Using immunoblotting, N1 was also shown to bind TREM-1 on the surface of monocytes (80) (Figure 1). Monocytes and lymphocytes exposed to PGLYRP1 and Hsp70, two previously mentioned ligands of TREM-1, had decreased LDH release in the presence of N1. Additionally, N1 treated cells had decreased mRNA expression of TNF- α , IFN γ , IL-1 β , and IL-6 after exposure to LPS (80). In mice with acute lung injury after bronchial instillation of LPS and α -galactosylceramide, i.v. treatment with N1 protected against the resultant cytokine storm, leading to decreased serum levels of INFy and IL-4. These mice also had reduced histologically evaluated pulmonary inflammation (80). N1 is another novel peptide that effectively inhibits TREM-1 induced inflammation, but requires additional studies looking at its efficacy in a polymicrobial model of sepsis to better evaluate it as a treatment modality.

GF9

Moving away from the strategy of preventing ligands from interacting with TREM-1, GF9 is a ligand-independent

peptide (GLLSKSLVF) derived from murine TREM-1's transmembrane region, designed using the Signaling Chain HOmoOligomerization (SCHOOL) model (92). It functions by inhibiting the interaction between TREM-1 and its signaling partner, DAP12 (77, 92) (Figure 1). Using a mouse model of endotoxemia, the efficacy of i.p. GF9 was studied, administered in its free form, and carried within high density lipoproteins (HDL). Synthetic HDL was modified to target delivery and uptake by macrophages, therefore delivering the GF9 directly to the cell of interest. Mice subjected to endotoxemia had higher survival after pretreatment with GF9 both in free form and incorporated into macrophage-targeted HDL-like particles. However, mice required a dose of 25mg/kg of free GF9 to see a survival benefit compared to vehicle, where a significantly lower dose of 5mg/kg of HDL bound GF9 was able to achieve the same survival benefit. Additionally, incorporation of GF9 into HDL prolonged the peptide's half-life, which together with its targeted macrophage delivery could contribute to its reduced effective dose (77).

SLC-TREM-1

TREM-1 sneaking ligand construct (SLC-TREM-1) is another protein that inhibits the interaction of TREM-1 with DAP12 (Figure 1), but it was specifically designed to target the endothelium (78). It is composed of 3 portions: an E-selectin targeting domain that binds to the surface of endothelial cells, Pseudomonas aeruginosa exotoxin A to facilitate translocation from the endosomal vesicular system into the cytosol, and a 7-aa sequence (LSKSLVF) derived from the transmembrane region of TREM-1, and actually contained within GF9 (77, 78, 93). Endothelial cells stimulated with LPS in vitro had decreased TREM-1 expression and activation after treatment with SLC-TREM-1 (78). Additionally, treatment with this protein intraperitoneally improved the 10-day survival in mice subjected to CLP (78). GF9 and SLC-TREM-1 demonstrate that targeting the interaction of TREM-1 with DAP12 is a viable TREM-1 inhibitory strategy with potential for drug development in sepsis.

LR17/LR12

LR17/LR12 are the most studied inhibitory peptides of TREM-1. LR-17 is a 17-aa peptide (LQEEDAGEYGCMVDGAR) based on a highly conserved sequence between TREM-1 and TREM-like transcript-1 (TLT-1), a membrane bound protein found on the surface of activated platelets (23, 68, 94) (Figure 1). After demonstrating structural similarities between TREM-1 and TLT-1 using crystallographic approaches, soluble TLT-1 (sTLT-1) was studied as a modulator of the inflammatory response and shown to decrease human neutrophil production of TNF-a, IL-6, and IL-8 after LPS stimulation (23, 94). LR17 was subsequently shown to protect mice from LPS endotoxemia (23). Mice treated with LR17 before or after LPS administration had improved 10-day survival and decreased serum proinflammatory cytokines (23). This peptide also has a protective effect in mice with CLP-sepsis (23). Mice treated with LR17 had decreased sepsis severity demonstrated by lower levels of IL-6, TNF-a, and IL-10 in serum, peritoneal and bronchoalveolar

fluid, and IL-6 and TNF-a in the liver, and lung (23). LP17 also ameliorated sepsis-induced acute lung injury, reduced bacterial load in the spleen and blood, and an improved 7-day survival (23).

It was discovered that just 12 aa accounted for the LP17's anti-inflammatory effects, and a novel peptide containing this sequence (LQEEDAGEYGCM) was created and named LR12 (4, 68). After confirming TREM-1's expression on vascular endothelial cells, the effects of LR12 on vascular reactivity during sepsis was explored (57). Using both CLP-sepsis and LPS endotoxemia, it was determined that LR12 prevents the TREM-1 upregulation induced in the murine aortas and mesenteric arteries of mice subjected to these models (57). Additionally, LR12 treatment protected mice from sepsis-induced vascular dysfunction, measured by improved vascular contractility, and attenuated IL-6, TNF- α , and IL-10 expression as well as NOS and COX signaling pathway activation in murine aortas and mesenteric arteries (57).

The effects of LR12 during sepsis were further studied in adult minipigs to better characterize its beneficial properties as a therapeutic agent (68). Polymicrobial sepsis was induced using a cecal slurry model. Pigs treated with LR12 had less cardiovascular failure, measured by MAP, vasopressor use, and cardiac index (68). Additionally, they had decreased coagulopathy and organ failure in the lungs, liver, and kidney. LR12 improved 24-hour survival in septic pigs from 40% to 100% (68). The effects of LR12 was further explored in nonhuman primates using an LPS model of endotoxemia (81). Monkeys received an i.v. bolus of LPS followed by a continuous infusion of LR12 or placebo for 8 hours. Monkeys treated with LR12 were protected from hypotension and neutropenia and had reduced plasma cytokine concentrations (81). One month after LR12 administration, no side effects were noted in the treated animals (81). In addition to i.v. infusion, a sustained release implant was tested as a novel administration route for LR12 in rats (95). An in situ poly-lactide-co-glycolide (PGLA) or poly-lactide (PLA) implant was used to deliver LR12 in a dimerized formulation to diminish peptide degradation and allow sustained release (95). In healthy rats, these in situ forming implants delivered a therapeutic concentration of LR12 for 7 days, introducing an effective method of administrating this otherwise short-lived peptide (95).

Nangibotide

LR12 is the first TREM-1 inhibitor to have reached the clinical stage. It is currently being studied in Phase 2 clinical trials of sepsis, COVID-19, and acute myocardial infarction under the name Inotrem (nangibotide) (16, 82, 83, 96–98). Nangibotide is an immune modulator targeting the inflammatory response amplification maintained by TREM-1 (82). In Phase 1 trials, continuous i.v. administration of nangibotide was safe and well tolerated at doses up 6 mg/kg/h for 7 hours and 45 mins following a 15-minute loading dose of 5 mg/kg, with few adverse events noted after 28 days of followup (83). In a Phase 2a multicenter, randomized, double-blind, placebo-controlled clinical trial, patients received a continuous infusion of 0.3, 1.0, or 3.0 mg/kg/h of nangibotide within 24 hours of the diagnosis of

septic shock (82). Treatment was continued until the patient was off vasopressors for 12 hours, or for a maximum of 5 days. There was no difference in adverse events, tolerability, inflammatory biomarkers and clinical efficacy between study groups (82). Treatment with nangibotide, however, was associated with a decrease in the serum levels of IL-6 (82). In a subgroup analysis, patients with high sTREM-1 treated with nangibotide had decreased SOFA scores after treatment (82, 98), suggesting that sTREM-1 could be a potential biomarker to predict which septic patients respond to nangibotide. A pivotal Phase 2b trial, looking at the efficacy, safety, and tolerability of nangibotide in patients with septic shock is currently underway (16, 97). Patients are given two doses of the peptide for 3-5 days, depending on vasopressor requirements. The primary endpoint being evaluated is total SOFA score changes from baseline to day 5 in all patients, and will include a subgroup analysis of patients with elevated sTREM-1 baseline levels (16). Larger studies are still needed to investigate the efficacy of nangibotide in attenuating sepsis severity, and to evaluate sTREM-1 as a potential biomarker of patient response to nangibotide.

SUMMARY AND PERSPECTIVES

Over the last two decades, extensive research has revealed TREM-1's critical role as a mediator of inflammation in sepsis and other diseases. Substantial work has been done to identify its endogenous ligands and develop inhibitors to be used as potential therapeutic agents for a range of diseases. There are now a variety of inhibitory molecules, predominately oligopeptides, that have shown promise as a treatment for sepsis using various preclinical sepsis models in a number of species. These peptides are capable of targeting TREM-1 activation at various points within its signaling cascade, from acting as a decoy receptor and binding up its endogenous ligands, to binding the extracellular domain and competitively inhibiting the binding of endogenous ligands, to interrupting the transmembrane association of TREM-1 with DAP12. Beyond oligopeptides, other modulators of the TREM-1 pathway have been studied in non-infectious inflammatory diseases (99). Researchers in Sweden have developed an anti-TREM-1 antibody that reduces secretion of proinflammatory cytokines from lamina propria cells isolated from patients with inflammatory bowel disease (99). Targeted monoclonal antibody treatment has been effective in treating a variety of inflammatory diseases, and TREM-1 targeted antibodies may be a beneficial approach to sepsis drug development. Future research could benefit from investigating this novel anti-TREM-1 antibody in septic preclinical models.

Although many TREM-1 inhibitory peptides have shown promise in preclinical models, there are currently no FDA approved treatments for sepsis. Strategies targeting inflammatory mediators like TNF- α and IL-1 β were not effective in clinical trials for sepsis, despite efficacy in experimental models (100). Further, drotrecogin alpha, the only treatment to receive FDA approval for the treatment of sepsis was later withdrawn from the market (101). This recombinant human activated protein C did not reduce mortality in septic patients at 28 days in a follow-up phase 3 international, randomized, controlled trial, and is no longer approved as a treatment for sepsis (101). There are likely many barriers to creating effective sepsis treatments, including poor translation between animal models and human disease, heterogeneity of the patient population selected for clinical trials, and the overall heterogeneity of the etiology of the disease (102). The physiologic response to sepsis in mice differs from humans, and mice are also more resistant to endotoxin and resilient to infection. Further, many treatments tested in murine models are given before, during, or shortly after infectious exposure. Conversely, patients often present for treatment days after infection, with sepsis developing over time. Additionally, in preclinical models, treatments are often tested on sepsis from a single, well-defined source of infection, where clinical trials often include patients who present with sepsis from a wide variety of etiologies. The host response has great variation based on the location of infectious insult and inciting organism species. Patients also have a variety of specific characteristics, like sex, age, comorbidities, and medications that can all effect response to an experimental treatment. In that regard, clinical trials of sepsis treatments would possibly benefit from selecting patients according to the source of infection and infectious agent.

LR12, a 12-aa peptide, has been studied extensively in mice, rats, pigs, and nonhuman primates as a treatment for sepsis and has been shown to be effective and safe when administered as an i.v. bolus, continuous infusion, and an *in situ* forming implant. It is currently in phase 2b clinical trials under the name nangibotide and has so far been demonstrated to be safe and

REFERENCES

- Hotchkiss RS, Moldawer LL, Opal SM, Reinhart K, Turnbull IR, Vincent JL. Sepsis and Septic Shock. *Nat Rev Dis Primers* (2016) 2:16045. doi: 10.1038/ nrdp.2016.45
- van der Poll T, van de Veerdonk FL, Scicluna BP, Netea MG. The Immunopathology of Sepsis and Potential Therapeutic Targets. *Nat Rev Immunol* (2017) 17(7):407–20. doi: 10.1038/nri.2017.36
- Amarante-Mendes GP, Adjemian S, Branco LM, Zanetti LC, Weinlich R, Bortoluci KR. Pattern Recognition Receptors and the Host Cell Death Molecular Machinery. *Front Immunol* (2018) 9:2379. doi: 10.3389/ fimmu.2018.02379
- 4. Dantas PHDS, Matos AO, da Silva Filho E, Silva-Sales M, Sales-Campos H. Triggering Receptor Expressed on Myeloid Cells-1 (TREM-1) as a Therapeutic Target in Infectious and Noninfectious Disease: A Critical Review. Int Rev Immunol (2020) 39(4):188–202. doi: 10.1080/ 08830185.2020.1762597
- Tammaro A, Derive M, Gibot S, Leemans JC, Florquin S, Dessing MC. TREM-1 and Its Potential Ligands in Non-Infectious Diseases: From Biology to Clinical Perspectives. *Pharmacol Ther* (2017) 177:81–95. doi: 10.1016/j.pharmthera.2017.02.043
- Schiechl G, Brunner SM, Kesselring R, Martin M, Ruemmele P, Mack M, et al. Inhibition of Innate Co-Receptor TREM-1 Signaling Reduces CD4(+) T Cell Activation and Prolongs Cardiac Allograft Survival. *Am J Transplant* (2013) 13(5):1168–80. doi: 10.1111/ajt.12186
- Roe K, Gibot S, Verma S. Triggering Receptor Expressed on Myeloid Cells-1 (TREM-1): A New Player in Antiviral Immunity? *Front Microbiol* (2014) 5:627. doi: 10.3389/fmicb.2014.00627

apparently effective in Phase-1 and Phase-2a studies. This peptide shows significant promise as a treatment for septic patients. It has demonstrated efficacy in seven preclinical models and in four different species, and has been administered up to 24 hours after onset of infection, addressing key issues of translatability and time of treatment in patients. Further, in the current trial, investigators are identifying patients with elevated levels of plasma sTREM-1 to evaluate whether these patients are better responder and, consequently, are more likely to benefit from treatment. By identifying a subgroup of patients who best respond to this treatment, administration can be personalized and efficacy can be better assessed. Sepsis has been called a "graveyard for pharmaceutical companies". Targeting the TREM-1 activation pathway, however, shows great promise and may finally become the first safe and effective therapeutic strategy to treat septic patients.

AUTHOR CONTRIBUTIONS

SS and MB conceived the original idea. SS prepared the figures and wrote the manuscript. PW and MB revised and edited the manuscript. PW supervised the project. All authors contributed to the article and approved the submitted version.

FUNDING

This study was partially supported by the National Institutes of Health (NIH) grants R35GM118337 and U01AI33655.

- Bouchon A, Facchetti F, Weigand MA, Colonna M. TREM-1 Amplifies Inflammation and Is a Crucial Mediator of Septic Shock. *Nature* (2001) 410 (6832):1103–7. doi: 10.1038/35074114
- Bouchon A, Dietrich J, Colonna M. Cutting Edge: Inflammatory Responses Can be Triggered by TREM-1, a Novel Receptor Expressed on Neutrophils and Monocytes. *J Immunol* (2000) 164(10):4991–5. doi: 10.4049/ jimmunol.164.10.4991
- Zamyatina A, Heine H. Lipopolysaccharide Recognition in the Crossroads of TLR4 and Caspase-4/11 Mediated Inflammatory Pathways. *Front Immunol* (2020) 11:585146. doi: 10.3389/fimmu.2020.585146
- Cohen J. TREM-1 in Sepsis. Lancet (2001) 358(9284):776-8. doi: 10.1016/ S0140-6736(01)06007-X
- Klesney-Tait J, Turnbull IR, Colonna M. The TREM Receptor Family and Signal Integration. Nat Immunol (2006) 7(12):1266–73. doi: 10.1038/ni1411
- Turnbull IR, Gilfillan S, Cella M, Aoshi T, Miller M, Piccio L, et al. Cutting Edge: TREM-2 Attenuates Macrophage Activation. J Immunol (2006) 177 (6):3520–4. doi: 10.4049/jimmunol.177.6.3520
- Gibot S. Clinical Review: Role of Triggering Receptor Expressed on Myeloid Cells-1 During Sepsis. Crit Care (2005) 9(5):485–9. doi: 10.1186/cc3732
- Gibot S, Le Renard PE, Bollaert PE, Kolopp-Sarda MN, Béné MC, Faure GC, et al. Surface Triggering Receptor Expressed on Myeloid Cells 1 Expression Patterns in Septic Shock. *Intensive Care Med* (2005) 31(4):594–7. doi: 10.1007/s00134-005-2572-x
- 16. Francois B, Lambden S, Gibot S, Derive M, Olivier A, Cuvier V, et al. Rationale and Protocol for the Efficacy, Safety and Tolerability of Nangibotide in Patients With Septic Shock (ASTONISH) Phase IIb Randomised Controlled Trial. *BMJ Open* (2021) 11(7):e042921. doi: 10.1136/bmjopen-2020-042921

- Brenner T, Uhle F, Fleming T, Wieland M, Schmoch T, Schmitt F, et al. Soluble TREM-1 as a Diagnostic and Prognostic Biomarker in Patients With Septic Shock: An Observational Clinical Study. *Biomarkers* (2017) 22(1):63– 9. doi: 10.1080/1354750X.2016.1204005
- Weber B, Schuster S, Zysset D, Rihs S, Dickgreber N, Schürch C, et al. TREM-1 Deficiency can Attenuate Disease Severity Without Affecting Pathogen Clearance. *PloS Pathog* (2014) 10(1):e1003900. doi: 10.1371/ journal.ppat.1003900
- Colonna M, Facchetti F. TREM-1 (Triggering Receptor Expressed on Myeloid Cells): A New Player in Acute Inflammatory Responses. J Infect Dis (2003) 187 Suppl 2:S397–401. doi: 10.1086/374754
- Arts RJ, Joosten LA, van der Meer JW, Netea MG. TREM-1: Intracellular Signaling Pathways and Interaction With Pattern Recognition Receptors. J Leukoc Biol (2013) 93(2):209–15. doi: 10.1189/jlb.0312145
- Sharif O, Knapp S. From Expression to Signaling: Roles of TREM-1 and TREM-2 in Innate Immunity and Bacterial Infection. *Immunobiology* (2008) 213(9-10):701–13. doi: 10.1016/j.imbio.2008.07.008
- Dower K, Ellis DK, Saraf K, Jelinsky SA, Lin LL. Innate Immune Responses to TREM-1 Activation: Overlap, Divergence, and Positive and Negative Cross-Talk With Bacterial Lipopolysaccharide. *J Immunol* (2008) 180 (5):3520–34. doi: 10.4049/jimmunol.180.5.3520
- Derive M, Bouazza Y, Sennoun N, Marchionni S, Quigley L, Washington V, et al. Soluble TREM-Like Transcript-1 Regulates Leukocyte Activation and Controls Microbial Sepsis. *J Immunol* (2012) 188(11):5585–92. doi: 10.4049/ jimmunol.1102674
- Ford JW, McVicar DW. TREM and TREM-Like Receptors in Inflammation and Disease. Curr Opin Immunol (2009) 21(1):38–46. doi: 10.1016/ j.coi.2009.01.009
- Fortin CF, Lesur O, Fulop T. Effects of TREM-1 Activation in Human Neutrophils: Activation of Signaling Pathways, Recruitment Into Lipid Rafts and Association With TLR4. *Int Immunol* (2007) 19(1):41–50. doi: 10.1093/ intimm/dxl119
- Klesney-Tait J, Colonna M. Uncovering the TREM-1-TLR Connection. Am J Physiol Lung Cell Mol Physiol (2007) 293(6):L1374–6. doi: 10.1152/ ajplung.00415.2007
- Bleharski JR, Kiessler V, Buonsanti C, Sieling PA, Stenger S, Colonna M, et al. A Role for Triggering Receptor Expressed on Myeloid Cells-1 in Host Defense During the Early-Induced and Adaptive Phases of the Immune Response. J Immunol (2003) 170(7):3812–8. doi: 10.4049/jimmunol. 170.7.3812
- Nathan C, Ding A. TREM-1: A New Regulator of Innate Immunity in Sepsis Syndrome. Nat Med (2001) 7(5):530–2. doi: 10.1038/87846
- Ornatowska M, Azim AC, Wang X, Christman JW, Xiao L, Joo M, et al. Functional Genomics of Silencing TREM-1 on TLR4 Signaling in Macrophages. *Am J Physiol Lung Cell Mol Physiol* (2007) 293(6):L1377– 84. doi: 10.1152/ajplung.00140.2007
- Read CB, Kuijper JL, Hjorth SA, Heipel MD, Tang X, Fleetwood AJ, et al. Cutting Edge: Identification of Neutrophil PGLYRP1 as a Ligand for TREM-1. J Immunol (2015) 194(4):1417–21. doi: 10.4049/jimmunol.1402303
- 31. Inanc N, Mumcu G, Can M, Yay M, Silbereisen A, Manoil D, et al. Elevated Serum TREM-1 Is Associated With Periodontitis and Disease Activity in Rheumatoid Arthritis. *Sci Rep* (2021) 11(1):2888. doi: 10.1038/s41598-021-82335-9
- 32. Zulfiqar F, Hozo I, Rangarajan S, Mariuzza RA, Dziarski R, Gupta D. Genetic Association of Peptidoglycan Recognition Protein Variants With Inflammatory Bowel Disease. *PloS One* (2013) 8(6):e67393. doi: 10.1371/ journal.pone.0067393
- Singh H, Rai V, Nooti SK, Agrawal DK. Novel Ligands and Modulators of Triggering Receptor Expressed on Myeloid Cells Receptor Family: 2015-2020 Updates. *Expert Opin Ther Pat* (2021) 31(6):549–61. doi: 10.1080/ 13543776.2021.1883587
- 34. Kashyap DR, Wang M, Liu LH, Boons GJ, Gupta D, Dziarski R. Peptidoglycan Recognition Proteins Kill Bacteria by Activating Protein-Sensing Two-Component Systems. *Nat Med* (2011) 17(6):676–83. doi: 10.1038/nm.2357
- 35. Gibot S, Buonsanti C, Massin F, Romano M, Kolopp-Sarda MN, Benigni F, et al. Modulation of the Triggering Receptor Expressed on the Myeloid Cell

Type 1 Pathway in Murine Septic Shock. *Infect Immun* (2006) 74(5):2823–30. doi: 10.1128/IAI.74.5.2823-2830.2006

- 36. Sharapova TN, Romanova EA, Ivanova OK, Sashchenko LP, Yashin DV. Cytokines TNFα, FN and IL-2 Are Responsible for Signal Transmission From the Innate Immunity Protein Tag7 (PGLYRP1) to Cytotoxic Effector Lymphocytes. *Cells* (2020) 9(12):2602. doi: 10.3390/cells9122602
- 37. Sharapova TN, Ivanova OK, Soshnikova NV, Romanova EA, Sashchenko LP, Yashin DV. Innate Immunity Protein Tag7 Induces 3 Distinct Populations of Cytotoxic Cells That Use Different Mechanisms to Exhibit Their Antitumor Activity on Human Leukocyte Antigen-Deficient Cancer Cells. J Innate Immun (2017) 9(6):598–608. doi: 10.1159/000479382
- Qiang X, Yang WL, Wu R, Zhou M, Jacob A, Dong W, et al. Cold-Inducible RNA-Binding Protein (CIRP) Triggers Inflammatory Responses in Hemorrhagic Shock and Sepsis. *Nat Med* (2013) 19(11):1489–95. doi: 10.1038/nm.3368
- 39. Zhang F, Brenner M, Yang WL, Wang P. A Cold-Inducible RNA-Binding Protein (CIRP)-Derived Peptide Attenuates Inflammation and Organ Injury in Septic Mice. *Sci Rep* (2018) 8(1):3052–62. doi: 10.1038/s41598-017-13139-z
- Yang WL, Sharma A, Wang Z, Li Z, Fan J, Wang P. Cold-Inducible RNA-Binding Protein Causes Endothelial Dysfunction *via* Activation of Nlrp3 Inflammasome. *Sci Rep* (2016) 6:26571. doi: 10.1038/srep26571
- Denning NL, Aziz M, Murao A, Gurien SD, Ochani M, Prince JM, et al. Extracellular CIRP as an Endogenous TREM-1 Ligand to Fuel Inflammation in Sepsis. JCI Insight (2020) 5(5):e134172. doi: 10.1172/jci.insight.134172
- 42. Wu J, Li J, Salcedo R, Mivechi NF, Trinchieri G, Horuzsko A. The Proinflammatory Myeloid Cell Receptor TREM-1 Controls Kupffer Cell Activation and Development of Hepatocellular Carcinoma. *Cancer Res* (2012) 72(16):3977–86. doi: 10.1158/0008-5472.CAN-12-0938
- Yang H, Antoine DJ, Andersson U, Tracey KJ. The Many Faces of HMGB1: Molecular Structure-Functional Activity in Inflammation, Apoptosis, and Chemotaxis. J Leukoc Biol (2013) 93(6):865–73. doi: 10.1189/jlb.1212662
- Yang H, Tracey KJ. Targeting HMGB1 in Inflammation. Biochim Biophys Acta (2010) 1799(1-2):149–56. doi: 10.1016/j.bbagrm.2009.11.019
- Weigand MA, Hörner C, Bardenheuer HJ, Bouchon A. The Systemic Inflammatory Response Syndrome. *Best Pract Res Clin Anaesthesiol* (2004) 18(3):455–75. doi: 10.1016/j.bpa.2003.12.005
- 46. Wang X, Xiang L, Li H, Chen P, Feng Y, Zhang J, et al. The Role of HMGB1 Signaling Pathway in the Development and Progression of Hepatocellular Carcinoma: A Review. Int J Mol Sci (2015) 16(9):22527–40. doi: 10.3390/ ijms160922527
- El Mezayen R, El Gazzar M, Seeds MC, McCall CE, Dreskin SC, Nicolls MR. Endogenous Signals Released From Necrotic Cells Augment Inflammatory Responses to Bacterial Endotoxin. *Immunol Lett* (2007) 111(1):36–44. doi: 10.1016/j.imlet.2007.04.011
- Zhong WJ, Duan JX, Liu T, Yang HH, Guan XX, Zhang CY, et al. Activation of NLRP3 Inflammasome Up-Regulates TREM-1 Expression in Murine Macrophages *via* HMGB1 and IL-18. *Int Immunopharmacol* (2020) 89(Pt A):107045. doi: 10.1016/j.intimp.2020.107045
- Lo TH, Tseng KY, Tsao WS, Yang CY, Hsieh SL, Chiu AW, et al. TREM-1 Regulates Macrophage Polarization in Ureteral Obstruction. *Kidney Int* (2014) 86(6):1174–86. doi: 10.1038/ki.2014.205
- Rosenzweig R, Nillegoda NB, Mayer MP, Bukau B. The Hsp70 Chaperone Network. Nat Rev Mol Cell Biol (2019) 20(11):665–80. doi: 10.1038/s41580-019-0133-3
- Asea A, Kraeft SK, Kurt-Jones EA, Stevenson MA, Chen LB, Finberg RW, et al. HSP70 Stimulates Cytokine Production Through a CD14-Dependant Pathway, Demonstrating its Dual Role as a Chaperone and Cytokine. *Nat Med* (2000) 6(4):435–42. doi: 10.1038/74697
- 52. Sharapova TN, Romanova EA, Ivanova OK, Yashin DV, Sashchenko LP. Hsp70 Interacts With the TREM-1 Receptor Expressed on Monocytes and Thereby Stimulates Generation of Cytotoxic Lymphocytes Active Against MHC-Negative Tumor Cells. *Int J Mol Sci* (2021) 22(13):6889. doi: 10.3390/ ijms22136889
- Lee WM, Galbraith RM. The Extracellular Actin-Scavenger System and Actin Toxicity. N Engl J Med (1992) 326(20):1335–41. doi: 10.1056/ NEJM199205143262006.

- Haselmayer P, Grosse-Hovest L, von Landenberg P, Schild H, Radsak MP. TREM-1 Ligand Expression on Platelets Enhances Neutrophil Activation. *Blood* (2007) 110(3):1029–35. doi: 10.1182/blood-2007-01-069195
- Fu L, Han L, Xie C, Li W, Lin L, Pan S, et al. Identification of Extracellular Actin As a Ligand for Triggering Receptor Expressed on Myeloid Cells-1 Signaling. *Front Immunol* (2017) 8:917. doi: 10.3389/fimmu.2017.00917
- 56. Boufenzer A, Carrasco K, Jolly L, Brustolin B, Di-Pillo E, Derive M, et al. Potentiation of NETs Release is Novel Characteristic of TREM-1 Activation and the Pharmacological Inhibition of TREM-1 Could Prevent From the Deleterious Consequences of NETs Release in Sepsis. *Cell Mol Immunol* (2021) 18(2):452–60. doi: 10.1038/s41423-020-00591-7
- 57. Jolly L, Carrasco K, Derive M, Lemarié J, Boufenzer A, Gibot S. Targeted Endothelial Gene Deletion of Triggering Receptor Expressed on Myeloid Cells-1 Protects Mice During Septic Shock. *Cardiovasc Res* (2018) 114 (6):907–18. doi: 10.1093/cvr/cvy018
- Linders J, Madhi R, Rahman M, Mörgelin M, Regner S, Brenner M, et al. Extracellular Cold-Inducible RNA-Binding Protein Regulates Neutrophil Extracellular Trap Formation and Tissue Damage in Acute Pancreatitis. *Lab Invest* (2020) 100(12):1618–30. doi: 10.1038/s41374-020-0469-5
- Colonna M. TREMs in the Immune System and Beyond. Nat Rev Immunol (2003) 3(6):445–53. doi: 10.1038/nri1106
- 60. Chung DH, Seaman WE, Daws MR. Characterization of TREM-3, an Activating Receptor on Mouse Macrophages: Definition of a Family of Single Ig Domain Receptors on Mouse Chromosome 17. *Eur J Immunol* (2002) 32(1):59–66. doi: 10.1002/1521-4141(200201)32:1<59::AID-IMMU59>3.0.CO;2-U
- Klesney-Tait J, Keck K, Li X, Gilfillan S, Otero K, Baruah S, et al. Transepithelial Migration of Neutrophils Into the Lung Requires TREM-1. *J Clin Invest* (2013) 123(1):138–49. doi: 10.1172/JCI64181
- 62. Tammaro A, Scantlebery AML, Rampanelli E, Borrelli C, Claessen N, Butter LM, et al. TREM1/3 Deficiency Impairs Tissue Repair After Acute Kidney Injury and Mitochondrial Metabolic Flexibility in Tubular Epithelial Cells. *Front Immunol* (2019) 10:1469. doi: 10.3389/fimmu.2019.01469
- Molloy EJ. Triggering Receptor Expressed on Myeloid Cells (TREM) Family and the Application of its Antagonists. *Recent Pat Antiinfect Drug Discov* (2009) 4(1):51–6. doi: 10.2174/157489109787236292
- 64. Hommes TJ, Hoogendijk AJ, Dessing MC, Van't Veer C, Florquin S, Colonna M, et al. Triggering Receptor Expressed on Myeloid Cells-1 (TREM-1) Improves Host Defence in Pneumococcal Pneumonia. *J Pathol* (2014) 233(4):357–67. doi: 10.1002/path.4361
- Hommes TJ, Dessing MC, Veer C, Florquin S, Colonna M, de Vos AF, et al. Role of Triggering Receptor Expressed on Myeloid Cells-1/3 in Klebsiella-Derived Pneumosepsis. *Am J Respir Cell Mol Biol* (2015) 53(5):647–55. doi: 10.1165/rcmb.2014-0485OC
- Washington AV, Schubert RL, Quigley L, Disipio T, Feltz R, Cho EH, et al. A TREM Family Member, TLT-1, Is Found Exclusively in the Alpha-Granules of Megakaryocytes and Platelets. *Blood* (2004) 104(4):1042–7. doi: 10.1182/ blood-2004-01-0315
- 67. Washington AV, Gibot S, Acevedo I, Gattis J, Quigley L, Feltz R, et al. TREM-Like Transcript-1 Protects Against Inflammation-Associated Hemorrhage by Facilitating Platelet Aggregation in Mice and Humans. *J Clin Invest* (2009) 119(6):1489–501. doi: 10.1172/JCI36175
- Derive M, Boufenzer A, Bouazza Y, Groubatch F, Alauzet C, Barraud D, et al. Effects of a TREM-Like Transcript 1-Derived Peptide During Hypodynamic Septic Shock in Pigs. *Shock* (2013) 39(2):176–82. doi: 10.1097/ SHK.0b013e31827bcdfb
- Li H, Guo S, Yan L, Meng C, Hu Y, He K, et al. Expression and Purification of a Functional Porcine Soluble Triggering Receptor Expressed on Myeloid Cells 1. Anim Biotechnol (2017) 28(4):237–41. doi: 10.1080/ 10495398.2016.1267016
- Yang C, Zhao J, Lin L, Pan S, Fu L, Han L, et al. Targeting TREM-1 Signaling in the Presence of Antibiotics Is Effective Against Streptococcal Toxic-Shock-Like Syndrome (STSLS) Caused by Streptococcus Suis. Front Cell Infect Microbiol (2015) 5:79. doi: 10.3389/fcimb.2015.00079
- Wang F, Liu S, Wu S, Zhu Q, Ou G, Liu C, et al. Blocking TREM-1 Signaling Prolongs Survival of Mice With Pseudomonas Aeruginosa Induced Sepsis. *Cell Immunol* (2012) 272(2):251–8. doi: 10.1016/j.cellimm.2011.10.006

- Horst SA, Linnér A, Beineke A, Lehne S, Höltje C, Hecht A, et al. Prognostic Value and Therapeutic Potential of TREM-1 in Streptococcus Pyogenes-Induced Sepsis. J Innate Immun (2013) 5(6):581–90. doi: 10.1159/000348283
- 73. Gibot S, Kolopp-Sarda MN, Bene MC, Bollaert PE, Lozniewski A, Mory F, et al. A Soluble Form of the Triggering Receptor Expressed on Myeloid Cells-1 Modulates the Inflammatory Response in Murine Sepsis. J Exp Med (2004) 200(11):1419–26. doi: 10.1084/jem.20040708
- Qian L, Weng XW, Chen W, Sun CH, Wu J. TREM-1 as a Potential Therapeutic Target in Neonatal Sepsis. Int J Clin Exp Med (2014) 7(7):1650–8.
- Shi X, Zhang Y, Wang H, Zeng S. Effect of Triggering Receptor Expressed on Myeloid Cells 1 (TREM-1) Blockade in Rats With Cecal Ligation and Puncture (CLP)-Induced Sepsis. *Med Sci Monit* (2017) 23:5049–55. doi: 10.12659/MSM.904386
- 76. Gibot S, Alauzet C, Massin F, Sennoune N, Faure GC, Bene MC, et al. Modulation of the Triggering Receptor Expressed on Myeloid Cells-1 Pathway During Pneumonia in Rats. J Infect Dis (2006) 194(7):975–83. doi: 10.1086/506950
- 77. Sigalov AB. A Novel Ligand-Independent Peptide Inhibitor of TREM-1 Suppresses Tumor Growth in Human Lung Cancer Xenografts and Prolongs Survival of Mice With Lipopolysaccharide-Induced Septic Shock. Int Immunopharmacol (2014) 21(1):208–19. doi: 10.1016/j.intimp.2014.05.001
- Gibot S, Jolly L, Lemarié J, Carrasco K, Derive M, Boufenzer A. Triggering Receptor Expressed on Myeloid Cells-1 Inhibitor Targeted to Endothelium Decreases Cell Activation. *Front Immunol* (2019) 10:2314. doi: 10.3389/ fimmu.2019.02314
- Denning NL, Aziz M, Diao L, Prince JM, Wang P. Targeting the eCIRP/ TREM-1 Interaction With a Small Molecule Inhibitor Improves Cardiac Dysfunction in Neonatal Sepsis. *Mol Med* (2020) 26(1):121. doi: 10.1186/ s10020-020-00243-6
- 80. Sharapova TN, Romanova EA, Chernov AS, Minakov AN, Kazakov VA, Kudriaeva AA, et al. Protein PGLYRP1/Tag7 Peptides Decrease the Proinflammatory Response in Human Blood Cells and Mouse Model of Diffuse Alveolar Damage of Lung Through Blockage of the TREM-1 and TNFR1 Receptors. *Int J Mol Sci* (2021) 22(20):11213. doi: 10.3390/ijms222011213
- Derive M, Boufenzer A, Gibot S. Attenuation of Responses to Endotoxin by the Triggering Receptor Expressed on Myeloid Cells-1 Inhibitor LR12 in Nonhuman Primate. *Anesthesiology* (2014) 120(4):935–42. doi: 10.1097/ ALN.0000000000000078
- François B, Wittebole X, Ferrer R, Mira JP, Dugernier T, Gibot S, et al. Nangibotide in Patients With Septic Shock: A Phase 2a Randomized Controlled Clinical Trial. *Intensive Care Med* (2020) 46(7):1425–37. doi: 10.1007/s00134-020-06109-z
- Cuvier V, Lorch U, Witte S, Olivier A, Gibot S, Delor I, et al. A First-in-Man Safety and Pharmacokinetics Study of Nangibotide, a New Modulator of Innate Immune Response Through TREM-1 Receptor Inhibition. Br J Clin Pharmacol (2018) 84(10):2270–9. doi: 10.1111/bcp.13668
- Gómez-Piña V, Soares-Schanoski A, Rodríguez-Rojas A, Del Fresno C, García F, Vallejo-Cremades MT, et al. Metalloproteinases Shed TREM-1 Ectodomain From Lipopolysaccharide-Stimulated Human Monocytes. *J Immunol* (2007) 179(6):4065–73. doi: 10.4049/jimmunol.179.6.4065
- Palazzo SJ, Simpson T, Schnapp LM. Triggering Receptor Expressed on Myeloid Cells Type 1 as a Potential Therapeutic Target in Sepsis. *Dimens Crit Care Nurs* (2012) 31(1):1–6. doi: 10.1097/DCC.0b013e31823a5298
- Jedynak M, Siemiatkowski A, Mroczko B, Groblewska M, Milewski R, Szmitkowski M. Soluble TREM-1 Serum Level can Early Predict Mortality of Patients With Sepsis, Severe Sepsis and Septic Shock. *Arch Immunol Ther Exp* (Warsz) (2018) 66(4):299–306. doi: 10.1007/s00005-017-0499-x
- Chang W, Peng F, Meng SS, Xu JY, Yang Y. Diagnostic Value of Serum Soluble Triggering Expressed Receptor on Myeloid Cells 1 (sTREM-1) in Suspected Sepsis: A Meta-Analysis. *BMC Immunol* (2020) 21(1):2. doi: 10.1186/s12865-020-0332-x
- Gibot S, Massin F. Soluble Form of the Triggering Receptor Expressed on Myeloid Cells 1: An Anti-Inflammatory Mediator? *Intensive Care Med* (2006) 32(2):185–7. doi: 10.1007/s00134-005-0018-0
- Jolly L, Carrasco K, Salcedo-Magguilli M, Garaud JJ, Lambden S, van der Poll T, et al. sTREM-1 Is a Specific Biomarker of TREM-1 Pathway Activation. *Cell Mol Immunol* (2021) 18(8):2054–6. doi: 10.1038/s41423-021-00733-5

- 90. Yang C, Chen B, Zhao J, Lin L, Han L, Pan S, et al. TREM-1 Signaling Promotes Host Defense During the Early Stage of Infection With Highly Pathogenic Streptococcus Suis. *Infect Immun* (2015) 83(8):3293–301. doi: 10.1128/IAI.00440-15
- Tan C, Gurien SD, Royster W, Aziz M, Wang P. Extracellular CIRP Induces Inflammation in Alveolar Type II Cells via TREM-1. Front Cell Dev Biol (2020) 8:579157. doi: 10.3389/fcell.2020.579157
- Sigalov AB. SCHOOL of Nature: Ligand-Independent Immunomodulatory Peptides. Drug Discov Today (2020) 25(8):1298–306. doi: 10.1016/ j.drudis.2020.05.005
- Sigalov AB. Commentary: Triggering Receptor Expressed on Myeloid Cells-1 Inhibitor Targeted to Endothelium Decreases Cell Activation. Front Immunol (2020) 11:173. doi: 10.3389/fimmu.2020.00173
- 94. Gattis JL, Washington AV, Chisholm MM, Quigley L, Szyk A, McVicar DW, et al. The Structure of the Extracellular Domain of Triggering Receptor Expressed on Myeloid Cells Like Transcript-1 and Evidence for a Naturally Occurring Soluble Fragment. J Biol Chem (2006) 281(19):13396–403. doi: 10.1074/jbc.M600489200
- Parent M, Clarot I, Gibot S, Derive M, Maincent P, Leroy P, et al. One-Week In Vivo Sustained Release of a Peptide Formulated Into In Situ Forming Implants. Int J Pharm (2017) 521(1-2):357-60. doi: 10.1016/ j.ijpharm.2017.02.046
- 96. Advertisement Feature in Nature, Under Biopharma Dealmakers. Retrieved at: https://www.nature.com/articles/d43747-020-00839-1.
- National Library of Medicine (U.S.). Efficacy, Safety and Tolerability of Nangibotide in Patients With Septic Shock (ASTONISH) United States: NIH U.S. National Library of Medicine (2019).
- Leong K, Gaglani B, Khanna AK, McCurdy MT. Novel Diagnostics and Therapeutics in Sepsis. *Biomedicines* (2021) 9(3):311. doi: 10.3390/ biomedicines9030311

- Brynjolfsson SF, Magnusson MK, Kong PL, Jensen T, Kuijper JL, Håkansson K, et al. An Antibody Against Triggering Receptor Expressed on Myeloid Cells 1 (TREM-1) Dampens Proinflammatory Cytokine Secretion by Lamina Propria Cells From Patients With IBD. *Inflamm Bowel Dis* (2016) 22 (8):1803–11. doi: 10.1097/MIB.00000000000822
- Ulloa L, Brunner M, Ramos L, Deitch EA. Scientific and Clinical Challenges in Sepsis. Curr Pharm Des (2009) 15(16):1918–35. doi: 10.2174/138161209788453248
- 101. Ranieri VM, Thompson BT, Barie PS, Dhainaut JF, Douglas IS, Finfer S, et al. Drotrecogin Alfa (Activated) in Adults With Septic Shock. N Engl J Med (2012) 366(22):2055–64. doi: 10.1056/NEJMoa1202290
- 102. Cavaillon JM, Singer M, Skirecki T. Sepsis Therapies: Learning From 30 Years of Failure of Translational Research to Propose New Leads. *EMBO Mol Med* (2020) 12(4):e10128. doi: 10.15252/emmm.201810128

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's Note: All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Copyright © 2022 Siskind, Brenner and Wang. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

GLOSSARY

aa AKT	Amino acid Protein kinase B	
ALI	Acute lung injury	
ALT	Alanine aminotransferase	
AST	Aspartate aminotransferase	
BAL	Bronchoalveolar lavage	
BMDN	Bone marrow-derived neutrophils	
CIRP	Cold-inducible RNA-binding protein	
CLP	Cecal ligation and puncture	
DAMP	Damage-associated molecular pattern	
DAP12	DNAX activation protein 12	
eCIRP	Extracellular cold-inducible RNA-binding protein	
ERK	Extracellular signal-regulated kinase	
FRET	Fluorescence resonance energy transfer	
HDL	High density lipoprotein	
HMGB1	High mobility group box 1	
HSP70	70 kDa heat shock protein	
IFN	Interferon	
lgG	Immunoglobulin G	
lκBa	Nuclear factor of kappa light polypeptide gene enhancer in B-cells	
	inhibitor alpha	
IL	Interleukin	
IP	Intraperitoneal	
ITAM	Immunoreceptor tyrosine-based activation motif	
IV	Intravenous	
KO	Knockout	
LDH	Lactate dehydrogenase	
LPS	Lipopolysaccharides	
MAP	Mean arterial pressure	
MAPK	Mitogen-activated protein kinase	
MCP	Macrophage chemoattractant protein	
MMP	Metalloproteinases	
mRNA	Messenger ribonucleic acid	
MyD88	Myeloid differentiation primary response 88	
NET	Neutrophil extracellular trap	
NF-κB	Nuclear factor kappa-light-chain-enhancer of activated B cells	
PAMPs	Pathogen-associated molecular patterns	
PBMC	Peripheral blood mononuclear cell	
PBS	Phosphate buffered saline	
PGLA	Poly-lactide-co-glycolide	
PGLYRP1	Peptidoglycan recognition protein 1	
PGN	Peptidoglycan	
PI3K	Phosphatidylinositol 3-kinase	
PLA	Poly-lactide	
PMN	Polymorphonuclear leukocyte	
PRR	Pattern recognition receptor	
RAGE	Receptor for advanced glycation end products	
RAS	Reticular activating system	
rm	Recombinant murine	
SCHOOL	Signaling chain homooligomerization	
SLC-	Triggering receptor expressed on myeloid cells-1 sneaking ligand	
TREM-1	construct	
SPR	Surface plasmon resonance	
STREM-1	Soluble TREM-1	
STSLS	Streptococcal toxic-shock-like syndrome	
Syk	spleen tyrosine kinase Toll-like receptor	
TLR TLT-1		
TNF-α	Triggering receptor expressed on myeloid cells-1-like transcript 1 Tumor necrosis factor-α	
TREM-1		
WT	Triggering receptor expressed on myeloid cells-1 Wild-type.	
	····- ·/	