#### Check for updates

#### **OPEN ACCESS**

EDITED BY Susan A. Boackle, University of Colorado Denver, United States

REVIEWED BY Takashi Matsushita, Kanazawa University, Japan David Fox, University of Michigan, United States

\*CORRESPONDENCE Claire F. Beesley Claire.beesley.20@ucl.ac.uk

#### SPECIALTY SECTION

This article was submitted to Autoimmune and Autoinflammatory Disorders: Autoimmune Disorders, a section of the journal Frontiers in Immunology

RECEIVED 20 July 2022 ACCEPTED 22 December 2022 PUBLISHED 16 January 2023

#### CITATION

Beesley CF, Goldman NR, Taher TE, Denton CP, Abraham DJ, Mageed RA and Ong VH (2023) Dysregulated B cell function and disease pathogenesis in systemic sclerosis. *Front. Immunol.* 13:999008. doi: 10.3389/fimmu.2022.999008

#### COPYRIGHT

© 2023 Beesley, Goldman, Taher, Denton, Abraham, Mageed and Ong. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

## Dysregulated B cell function and disease pathogenesis in systemic sclerosis

Claire F. Beesley<sup>1\*</sup>, Nina R. Goldman<sup>1</sup>, Taher E. Taher<sup>2</sup>, Christopher P. Denton<sup>1</sup>, David J. Abraham<sup>1</sup>, Rizgar A. Mageed<sup>3</sup> and Voon H. Ong<sup>1</sup>

<sup>1</sup>Centre for Rheumatology, Division of Medicine, University College London, London, United Kingdom, <sup>2</sup>Institute of Immunology and Immunotherapy, University of Birmingham, Birmingham, United Kingdom, <sup>3</sup>Centre for Translational Medicine and Therapeutics, William Harvey Research Institute, Queen Mary University of London, London, United Kingdom

Systemic sclerosis (SSc) is a complex, immune-mediated rheumatic disease characterised by excessive extracellular matrix deposition in the skin and internal organs. B cell infiltration into lesional sites such as the alveolar interstitium and small blood vessels, alongside the production of defined clinically relevant autoantibodies indicates that B cells play a fundamental role in the pathogenesis and development of SSc. This is supported by B cell and fibroblast coculture experiments revealing that B cells directly enhance collagen and extracellular matrix synthesis in fibroblasts. In addition, B cells from SSc patients produce large amounts of profibrotic cytokines such as IL-6 and TGF- $\beta$ , which interact with other immune and endothelial cells, promoting the profibrotic loop. Furthermore, total B cell counts are increased in SSc patients compared with healthy donors and specific differences can be found in the content of naïve, memory, transitional and regulatory B cell compartments. B cells from SSc patients also show differential expression of activation markers such as CD19 which may shape interactions with other immune mediators such as T follicular helper cells and dendritic cells. The key role of B cells in SSc is further supported by the therapeutic benefit of B cell depletion with rituximab in some patients. It is notable also that B cell signaling is impaired in SSc patients, and this could underpin the failure to induce tolerance in B cells as has been shown in murine models of scleroderma.

#### KEYWORDS

autoantibodies, systemic sclerosis (scleroderma), autoimmunity, B cells, fibrosis

### 1 Introduction

Systemic sclerosis (SSc) is a rare, immune-mediated rheumatic disease characterised by pathogenic microvascular damage and progressive fibrosis of the skin and internal organs. It has the highest case-specific mortality of any rheumatic disease and carries significant morbidity for the patient. Although the updated American College of Rheumatology (ACR) and European League Against Rheumatism (EULAR) classification criteria have improved sensitivity and specificity of diagnosis, understanding of disease aetiopathogenesis and development remains unclear (1–3). In addition, SSc is highly heterogenous, so it can be difficult to stratify patients and devise optimal treatment strategies (2, 4).

Skin fibrosis is the major diagnostic feature of SSc, and the extent of skin involvement is used to stratify patients into two subsets. These are limited cutaneous systemic sclerosis (lcSSc) and diffuse cutaneous systemic sclerosis (dcSSc) (5). Patients with lcSSc generally exhibit skin fibrosis which is distal to the elbows and knees, whereas diffuse involvement occurs proximally to the elbows and knees (3). Patients with dcSSc are at an increased risk of complications such as scleroderma renal crisis and interstitial lung disease (ILD), although some patients with lcSSc can also develop these complications (4, 6, 7). Importantly, there is no difference in the frequency and timing of development of significant ILD between patients for both skin subsets (8). Therefore, classification based upon skin subset alone does not offer accurate risk stratification of internal organ involvement.

Pathogenesis of systemic sclerosis is mediated by several immune and inflammatory cells. Currently, it is thought that vascular injury drives an infiltration of mast cells, T lymphocytes and macrophages into lesional tissues early on in the disease. Ultimately, this results in an unresolving pro-inflammatory and pro-fibrotic response mediated by myofibroblast differentiation and the production of cytokines such as interleukin-6 (IL-6) and transforming growth factor  $\beta$  (TGF- $\beta$ ). Evidence for B cell involvement comes from multiple studies outlining dysregulated B cell signaling and homeostasis within SSc patients, as well as evidence from mouse models and B cell modulating therapeutics. The key immunological feature of SSc is the presence of high levels of self-reactive antibodies in the blood. Patients present with various autoantibody profiles and, in clinical practice, this is one of the best indications to stratify patients and predict organ involvement (9). In addition, detection of autoantibodies can precede clinical onset of SSc, highlighting their pathological relevance (10). As such it is likely that autoreactive B cells are a driving factor in SSc, but the complete relevance and origin of these cells has not been fully determined.

### 2 B cell biology

B lymphocytes are pleiotropic cells with multiple functions including antibody and cytokine production, antigen presentation to T cells, modulation of dendritic cell function, and lymphoid organogenesis. As a result, B cells can orchestrate immune responses and influence the local environment at sites of infection and tissue injury/inflammation. With regards to the latter, dysregulated B cell responses have been implicated in a number of autoimmune diseases including systemic lupus erythematosus (SLE), Sjogren's syndrome and rheumatoid arthritis (RA) (11).

B lymphocytes develop within the bone marrow from hematopoietic stem cells where they acquire a functional B cell receptor (BCR) following rearrangement of their immunoglobulin (Ig) heavy (H) and light (L) genes in the presence of selective growth factors. Following Ig gene rearrangements, immature B cells express surface IgM and are then purged of autoreactive cells that recognise self-antigens in the bone marrow with high affinity. Most self-reactive clones are tolerised at this central checkpoint through mechanisms of receptor editing and clonal deletion. Immature B cell clones that do not recognise local self-antigens leave the bone marrow and migrate to the periphery for development as transitional cells to naïve mature B cells in a linear pathway (Figure 1) (12, 13).

### 3 B cell regulation in scleroderma

Differences in B cell homeostasis in SSc patients compared with healthy individuals have been documented in a number of studies (14–16). Typically, these studies have reported an overall increase in the number of B cells and/or differences in the distribution of B cell subsets in scleroderma patients compared with healthy individuals (14–16). However, it is pertinent to highlight that disease duration and immunosuppressive medications are likely to have impacts on lymphocyte homeostasis and subset distribution. Therefore, inclusive analyses are important to mitigate the effect of these factors on altered B cell homeostasis in SSc.

An early study by Sato et al. analysed B cell subsets in 39 Japanese SSc patients who had not received any immunosuppressive therapy. This study observed an increase in the number of circulating CD27<sup>-</sup> naïve B cells compared with healthy individuals with a reduction in CD27<sup>+</sup> memory B cells and CD27<sup>+</sup> plasmablasts (15). Additionally, memory B cells from SSc patients displayed augmented ability to undergo apoptosis and an overexpression of CD19 – possibly relating to their functional hyperactivity (15). These findings have been



autoreactive B cells becoming anergic-like T3 cells. T2 cells can mature to become either follicular (Fo) or marginal zone (MZ) B cells expressing IgM and IgD. Fo B cells can differentiate to low affinity antibody producing plasmablasts (PB) or undergo class switching and affinity maturation in germinal centres (GC) to become high affinity antibody producing plasma cells or memory cells. Figure created with **BioRender.com**.

confirmed in subsequent studies which have revealed further differences between CD27<sup>-</sup> naïve B cells in healthy individuals and patients. Following from these earlier studies, transitional CD27<sup>-</sup>CD24<sup>hi</sup>CD38<sup>hi</sup> B cells have also been implicated in scleroderma (17-19). Transitional B cells encompass a heterogenous population comprised of at least three distinct subsets (T1 - T3) with differential expression of IgM and IgD and a differential capacity to respond to antigen stimulation. T1 B cells define a population which has recently emigrated from the bone marrow and must acquire survival signals and undergo peripheral tolerance before the cells can proceed through the T2/ 3 pathway and develop to maturity (20). The T1 and T2 subsets are distinguishable by their differential capacity to survive or undergo apoptosis following BCR engagement, whilst the T3 subset display a functional status which is similar to that of anergic B cells (20). In some studies, a CD27<sup>+</sup> subset has also been identified (18, 20). This subset responds rapidly to T cell independent stimulation and is able to produce natural IgM antibodies and secrete high levels of IL-10 (18). Taher et al. studied transitional B cells in SSc patients and reported reduced numbers of T1 cells and expanded T2 cells in SSc patients compared with matched healthy individuals (18). Importantly, T1 cells from SSc patients displayed reduced capacity to undergo apoptosis and contained large numbers of B cells that were specific against the SSc-associated antigen Scl-70. Furthermore, B cell specificity for Scl-70 was demonstrated in patients who were seropositive for Scl-70 autoantibodies, highlighting the significance of this finding (18). This study also analysed the phosphorylation of the STAT-3 signalling pathway when transitional B cells were stimulated through toll-like receptor (TLR)9. STAT-3 has a fundamental role in suppressing proinflammatory signal transduction through the TLRs and is

crucial for anti-inflammatory IL-10 signalling (21). Taher and colleagues observed a significant reduction in STAT-3 phosphorylation in transitional T1 and CD27<sup>+</sup> cells from SSc patients and this is consistent with reduced IL-10 production from these cells (18). Taken together this may implicate a failed tolerance checkpoint at the developmental T1 – T2 stage in SSc patients and similar findings have been reported in a recent study by Glauzy and colleagues who concluded that central and peripheral B cell tolerance checkpoints are likely breached in SSc patients (18, 19). However, STAT-3 is also involved in the regulation of several other cytokines including IL-6, indicating that the observation of reduced IL-10 production in SSc B cells may relate to a signalling molecule upstream of STAT-3 (22, 23).

Differences in the number and function of IL-10 producing regulatory B cells, or Bregs have been noted in multiple studies of SSc (24–26). There is no definitive marker to define Bregs, therefore, current classification is based upon Breg capacity to produce high levels of anti-inflammatory IL-10 and immune-regulatory IL-35 and TGF- $\beta$  – a key cytokine also involved in fibrotic pathways (27). In SSc patients, IL-10 producing B cells are markedly reduced compared with healthy individuals and B cell capacity to produce IL-10 is reduced (16, 18, 24, 25). Diminished Breg populations have been linked with an increased risk of ILD, whilst IL-10 specific B cell expansion has been shown to correlate positively with patient responses to autologous haematopoietic stem cell transplant (16, 25, 28).

A study to examine CD27<sup>+</sup> memory B cell populations in SSc patients highlighted a reduction in the non-switched memory compartment, resulting in an imbalance between tolerogenic and activated memory B cells (29). In addition, elevated numbers of switched and activated memory B cells were associated with dcSSc and are likely to be relevant through

autoantibody and cytokine production. However a significant proportion (52.6%) of the dcSSc patients had received immunosuppressive medications suggesting that these results should be interpreted with caution (29).

### 4 Autoantibodies in SSc

Whilst the pathological relevance of SSc-specific autoantibodies remains incompletely understood, autoantibodies are strongly associated with the disease and are the strongest predictors of disease course and outcome. This means that autoantibodies are valuable clinical tools which are routinely used to stratify patients and predict patient prognosis (9, 30). Typically, these autoantibodies are of high specificity against nuclear antigens and are present at a high concentration (10).

The three autoantibodies which are most frequently associated with SSc are anti-centromere antibodies (ACA), anti-topisomerase I antibodies (ATA) and anti-RNA polymerase III antibodies (ARA). A patient will typically present with one dominant autoantibody specificity and is unlikely to change this autoantibody subtype. One of these three subsets of autoantibodies (ACA, ATA or ARA) are present in over 50% of those with scleroderma (9).

ACA are the most frequently observed autoantibodies in SSc patients (9). These autoantibodies are highly specific for SSc and are associated with lcSSc (31). Most commonly, ACA are specific for centromere protein B (9). Centromere protein B is a highly conserved nuclear protein which facilitates centromere formation. Patients with ACA are often thought to be at risk of developing pulmonary hypertension (PAH) but these studies are either based on enriched cohorts or have used echocardiogram-diagnosed PAH which is not sufficient for PAH diagnosis (32, 33). However, a number of studies have demonstrated that B cell depletion or B cell deficiency can be protective against vascular remodeling in rodent models of PAH (34–36).

ATA positivity is strongly associated with SSc and is present in up to 40% of individuals with the disease. ATA was initially named as anti-Scl-70 as these autoantibodies react with a 70 kDa protein on immunoblots, but it was later realised that Scl-70 was in fact a breakdown product of the larger 100 kDa topoisomerase I protein (37). ATA-positive patients can be dcSSc or lcSSc, although there is a slight predominance of dcSSc with ATA (9). ATA are a strong predictor of digital ulcer, pulmonary fibrosis and ILD development irrespective of skin involvement and ATA levels positively correlate with disease severity and activity (10). Multiple studies have shown that purified ATA from SSc sera can bind to the cell surface of fibroblasts, providing a potential mechanism by which ATA-positivity could influence disease (38, 39).

Unlike anti-RNA polymerase I and II autoantibodies that can be detected across various autoimmune rheumatic diseases, anti-RNA polymerase III antibodies (ARA) are strongly associated with SSc (9). In a recent meta-analysis, the pooled prevalence of ARA positivity in SSc patients was 11% (40). ARA positivity is a strong predictor of scleroderma renal crisis and up to 59% of those with scleroderma renal crisis are seropositive for ARA (41). ARA levels correlate with skin involvement, but they do not predict organ complications or disease outcome (9). However, patients with ARA have a higher risk of cancer (42).

In terms of disease pathogenesis, autoantibodies can amplify immune responses and initiate inflammation and fibrosis through immune complex formation. Recently, a new class of autoantibodies reactive against various cell surface receptors have been identified in those with SSc. These include antibodies with specificity for platelet-derived growth factor receptor alpha (PDGFR $\alpha$ ), angiotensin II type 1 receptor (AT1R) and endothelin-1 type A receptor (ETAR). Antibodies against PDGFR $\alpha$  have been linked with fibrosis in both *in vitro* and *in vivo* studies, whilst antibodies against AT1R and ETAR have been linked with vascular damage (43). These autoantibodies, however, are only detected in a small number of SSc patients and have low disease specificity compared with ACA, ATA and ARA positivity (44).

Additionally, autoantibodies with specificity for CD22 have been detected in a subset of patients. CD22 is an inhibitory B cell receptor that dampens BCR signalling *via* a tyrosine phosphorylation-dependent mechanism. Patients with CD22 autoantibodies exhibit significantly worse skin scores than SSc patients without these antibodies and it is hypothesised that anti-CD22 autoantibodies are likely to interfere with CD22-mediated suppression of B cell activation, resulting in further dysregulation of B cell homeostasis (45). However, these autoantibodies have been found in other autoimmune diseases and are only present in a small subset of SSc patients so are not likely to be a driving factor of disease (17, 45).

# 5 Abnormalities in B cell signalling and B cell activation

In order to mount an effective immune response, B cells must coordinate signalling through multiple innate and adaptive immune receptors. Alongside the BCR, B cells express a variety of coactivating and coinhibitory receptors that regulate B cell activation status. Remarkably, even a minor change in the expression or function of these receptors can result in a defective B cell response. These abnormalities have been reported in murine models of autoimmunity and in patients with autoimmune diseases including SSc (Figure 2).

There is considerable evidence to show that B cells in SSc patients are in a hyperactivated state induced, at least partly, by increased levels of BAFF (46). This hyperactivation state is marked by increased expression of the BCR coreceptor CD19 (approximately 20% higher) in SSc patients compared with healthy controls (15, 47). As CD19 lowers the threshold for



antigen-dependent stimulation of the BCR, enhanced CD19 expression could augment B cell signalling, resulting in an autoimmune humoral response. Indeed, various autoantibodies, including SSc-specific ATA antibodies were considerably enhanced in transgenic mice that overexpressed CD19 by 20%. This implies that a minor increase in CD19 expression in human SSc may be enough to skew B cell signalling and, thus, trigger autoantibody production (48, 49). CD19 also strengthens antigenic signals generated by the BCR and with CD40 engagement by the CD40 ligand (CD40L) expressed on activated T cells (50). Upregulation of CD19 has been observed in other autoimmune diseases and is considered to be a possible target for future therapy in SSc (17). On B cells, CD19 forms a complex with CD21, also called complement receptor 2 (CR2) which binds to cleavage products of the C3 complement component and transduces signals through CD19, thus lowering the threshold for B cell activation (48). Besides CD21, CD19 also forms a complex with CD225 and CD81, where CD225 could regulate CD19-mediated PI3K signalling (51). On the other hand, CD81 interaction with CD19 is important for CD19 expression and function.

In contrast to CD21, which delivers a costimulatory signal, the complement receptor CD35 (CR1) transduces inhibitory signals by inhibiting the induced increase of cytoplasmic  $Ca^{2+}$ levels through BCR and CD40 signalling (17, 52, 53). This suggests that CR1 is a late checkpoint to prevent autoreactive B cell maturation and reduced CR1 expression has been found in memory B cells from SSc patients, potentially augmenting their ability to respond to self-antigen (17).

Programmed cell death protein 1 (PD-1) is a cell surface molecule which is expressed on leukocytes and has been linked to the loss of B cell tolerance and development of autoimmunity. PD-1 regulates immunity by promoting self-tolerance when it is engaged with its ligands PD-L1 and PD-L2 (54, 55). PD-L1 is expressed on many cell types while PD-L2 is expressed on antigen presenting cells. The interaction between PD-1/PD-L2 on B cells and T cells is suggested to suppress tumor necrosis factor alpha (TNF- $\alpha$ ) production from antigen-specific B cells and increase IL-10 production (56, 57). This interaction can be blocked by serum soluble PD-1 (sPD-1) and PD-L2 (sPD-L2), resulting in augmented T and B cell responses. Indeed, sPD-1 and sPD-L2 levels are elevated in SSc patients and this correlates with severity of disease, as well as an increase in TNF- $\alpha$ producing B cells and a reduction in IL-10 producing B cells (56, 58).

Signalling through innate immune receptors, such as TLRs can associate with the development of autoimmunity and the loss of B cell tolerance. TLR signalling enables immune cell responses to various stimuli such as pathogen associated molecular signals (PAMPs) and danger associated molecular signals (DAMPs). Defects in TLR signalling have been implicated in several fibrotic diseases and B cell-specific TLR expression dictates developmental trajectories within these cells (59). For example, TLR4 which recognises lipopolysaccharide is thought to promote B cell survival and maturation during transitional development, whilst TLR2 arrests this process (60). In addition, DAMP induced TLR4 activation is known to be a key mediator of myofibroblast differentiation and is relevant in scleroderma as TLR4 and several associated DAMPs are significantly elevated in lesional tissues of SSc patients (61). Additionally, TLR7 and TLR9 can operate in conjunction with BCR-mediated signals and these receptors have important regulatory functions in B cell development and autoimmunity. TLR7 and TLR9 are intracellular endosomal receptors found in eosinophils, dendritic cells and B cells. They recognise bacterial and viral DNA and induce IRF7 signalling and IFN- $\alpha$ production (62). In SLE patients, high TLR7 expression driven by the TLR7 polymorphism rs3853839 C/G was associated with increased disease activity and an upregulation of IFN-responsive genes. Patients with higher TLR7 expression had greater numbers of B cells than patients with lower TLR7 expression and this was most notable in the transitional B cell subset (63). Analysis of the expression of TLR7, TLR9 and JAK2 in PBMC samples from 50 SSc patients and 13 healthy individuals revealed significant TLR7 upregulation in the SSc patients and decreased levels of TLR9 and JAK2. However, this study was carried out using total PBMC and most of the patient cohort were receiving immunosuppressive therapy (64). An in vitro study on B cells isolated from SSc patients and healthy donors reported significantly reduced IL-10 production when the SSc B cells were stimulated via TLR9 perhaps indicating defective TLR9 signalling in the B cells from SSc patients (18).

Upon BCR engagement, intracellular protein kinases such as SYK and BTK are activated. These kinases then phosphorylate CD19 and B cell adapter for PI3K (BCAP) which provide docking sites for PI3Ks leading to PI3K activation. PI3K activation leads to further downstream activation of Akt and mTOR serine/threonine kinase signalling, as well as NF- $\kappa$ B pathway activation (65–69). These pathways are critical for B cell survival, proliferation and differentiation with defective class 1A PI3K function preventing B cell development beyond the

pre-BCR stage (70). Additionally, ubiquitination is a key mechanism for regulating BCR-driven signalling where inappropriate ubiquitination has been associated with autoimmunity. A20 is a widely expressed deubiquitinating and ubiquitin-editing enzyme which restricts NF-KB signalling and protects against TNF-α induced programmed cell death. A20 has been linked with SSc in multiple genome-wide association studies (GWAS) and single nucleotide polymorphism (SNP) analyses which implicate the Tnfaip3 gene (71, 72). Tavares and colleagues studied this gene using a floxed allele of Tnfaip3 to generate mice deficient in A20 in B cells (73). The B cells from these mice were hyper-responsive and displayed enhanced NFκB signalling through CD40 induced signals. The B cells were also resistant to Fas mediated cell death, likely due to increased expression of anti-apoptotic proteins such as Bcl-x produced via the NF-KB pathway, potentially providing a mechanism by which autoreactive B cells in genetically susceptible individuals may survive tolerance checkpoints and develop to maturity in SSc (74).

Similar to other autoimmune diseases, SSc development likely results from a combination of genetic and environmental factors with genome-wide association studies (GWAS) identifying a number of polymorphisms associated with SSc. Some of these polymorphisms relate to B cell signalling, but a 2011 association study of B cell gene polymorphisms in a cohort of 900 SSc patients and 1034 heathy individuals did not find evidence of SSc-associated polymorphisms in CD19, CD20, CD22 and CD24 (68). However, there is evidence for increased SSc susceptibility resulting from genetic polymorphisms in coding domains of other B cell signalling molecules. These include polymorphisms in BANK1, BLK, PTPN22 and CSK (69, 75-77). Some of these polymorphisms are known to be associated with multiple autoimmune diseases and are associated with a patient's ethnicity and autoantibody subset. For example, the PTPN22 1858T risk allele is associated with patients who are seropositive for ATA autoantibodies and results in a memory B cell deficit with reduced responsiveness to antigen stimulation via the BCR (77). In addition, polymorphisms in the negative regulators of B and T cell activation, suppressors of cytokine signalling 2 (SOCS2) and SOCS3, were noted to be associated with SSc (78). Further research is needed to understand the functional relevance of these polymorphisms in SSc disease development.

A recent study analysed the expression of PI3K associated molecules in 21 patients with early dcSSc (79). The study identified altered mRNA expression in PI3K associated signalling molecules including TLR homolog CD180, TLR4 and C3 (79). Co-engagement of CD180 and the BCR enhanced NF- $\kappa$ B phosphorylation in dcSSc B cells, but not in healthy controls, whilst activation *via* CD180 increased the percentage of switched memory B cells in dcSSc patients compared with healthy controls (79). Additionally, in 2001, Koarada and colleagues reported that the percentage of SSc patients with CD180-negative B cells was

significantly higher than healthy controls, although not as high as those with Sjogren's syndrome or dermatomyositis (80). This may be significant as ligation of CD180 induces affinity maturation and programs immature (including T1) and mature B cell subsets to become efficient antigen presenting cells to T follicular helper cells (81, 82). Interestingly, another study confirmed these findings and reported reduced CD180 mRNA expression in B cells from dcSSc patients. In lupus CD180-negative B cells have been described as highly activated and CD180 can be internalised after stimulation, indicating that CD180-negative B cells may result from B cell activation via CD180. Of note, anti-CD180 stimulation induced natural autoantibody production and significantly increased the concentration of IL-6 in the supernatant of healthy tonsillar B cells providing a mechanism by which CD180 stimulation could increase the number of CD180-negative autoantibody producing B cells in SSc (83).

# 6 Perturbations in the B cell repertoire in SSc patients

Xiadong Shi and colleagues completed a study to analyse the B cell repertoire in SSc patients compared with healthy controls. These investigators reported differential IGHV-J gene usage in SSc patients compared with healthy controls. In addition, they noted that the average CDR3 region was significantly shorter in SSc patients compared with the healthy controls. This is important as the CDR3 region is the most variable region of the BCR and is the prime determinant of antigen specificity (84). Conversely, using immunoglobulin repertoire analyses in new emigrant/transitional B cells in SSc patients and healthy controls, Glauzy and colleagues observed a significantly higher frequency of long IgH CDR3s which are associated with selfreactivity (19). The discrepancy between these studies may reflect the different B cell populations which were sampled as the CDR3 region varies in length between subsets such as naïve and memory B cells (85). Therefore, more research is needed to determine whether the CDR3 region is involved in autoimmunity in SSc.

# 7 Profibrotic and proinflammatory B cell cytokines

B cells contribute to fibrosis *via* a number of mechanisms, including direct cell-cell contact and production of stimulating cytokines such as IL-6 (Figure 3). IL-6 is a pleotropic, proinflammatory cytokine which can promote fibrosis (86). Serum levels of IL-6 are increased in SSc patients and this has been linked to worsening disease in human and animal studies (11, 17, 86, 87). IL-6 promotes CD4<sup>+</sup> T cell differentiation into pro-inflammatory and pro-fibrotic Th1 and Th17 cells (86, 88).

Furthermore, B cell derived IL-6 drives spontaneous germinal centre formation in murine lupus, thus, providing a mechanism by which excessive IL-6 production by B cells can result in autoantibody production and autoimmunity (87). IL-6 also acts as a stimulant for B cell proliferation and it enhances plasma cell generation and antibody production, potentially inducing a pathogenic IgG autoantibody response as has been described in a murine model of lupus (89, 90).

Additionally, activated B cells produce high levels of TGF- $\beta$ which is a central player in fibrosis (91). When TGF- $\beta$  binds to its receptor on fibroblasts, it induces collagen synthesis and extracellular matrix deposition through Smad signalling (91). In addition, TGF- $\beta$  promotes fibrosis *via* the inhibition of matrix degrading proteolytic enzymes such as serine proteinases (92). Elevated levels of TGF- $\beta$  and enhanced expression of its receptors have been found in the skin of SSc patients (93).

IL-10 is a potent anti-inflammatory cytokine and is, therefore, very important in the context of autoimmune disease (94). IL-10 can suppress CD4<sup>+</sup> T cell proliferation through downregulation of CD86 in an autocrine manner. This also reduces IFN- $\gamma$  and TNF- $\alpha$  production which are functionally important in scleroderma due to their profibrotic and proinflammatory effects (88). In SSc patients, IL-10 producing B cells are markedly reduced compared with healthy controls and B cell capacity to produce IL-10 when stimulated *via* TLR9 is also reduced (18, 95).

IL-13 is another important Th2-type cytokine with several unique effector functions. IL-13 has been demonstrated to mediate tissue fibrosis in asthma, indicating that it is a key regulator of the extracellular matrix (96). In the context of B cells, IL-13 can upregulate MHC class II expression, induce Ig production, promote IgE class switching and induce B cell proliferation and differentiation (96-98). Serum levels of IL-13 are increased in SSc patients and this correlates with levels of C-reactive protein which is a biomarker of inflammation (99). In addition to IL-13, IL-33 is another important Th2 cytokine which is overexpressed in SSc. This cytokine is constitutively expressed at epithelial barrier sites and its overexpression is of interest since IL-33 can drive tissue fibrosis (100). Importantly, chronic exposure to IL-33 can also promote significant BAFF production via neutrophils and dendritic cells resulting in germinal centre formation and an IgG autoantibody response (101).

CXCL-13 is a chemokine that regulates B cell migration through secondary lymphoid tissues and is important for neogenesis of ectopic lymphoid structures in the lungs (102). It is overexpressed in SSc patients and CXCL-13 blood concentrations have been linked with worsening prognosis in patients with idiopathic pulmonary fibrosis (95, 103). It is currently thought that CXCL-13 is produced by monocyte-derived macrophages and that its gene expression is controlled by TNF- $\alpha$  and IL-10 (103, 104).

In addition to these key cytokines, high levels of BAFF have long been noted in SSc patients and are associated with disease



progression (46). BAFF is a pleiotropic cytokine (also known as BLyS) which promotes B cell proliferation and is a key regulator of peripheral tolerance. BAFF is also a fundamental survival factor and is involved in multiple cell-fate decisions during B cell development (105-107). BAFF overexpression has been linked with autoimmunity in human and animal studies and BAFF inhibition attenuated skin and lung fibrosis in a mouse model of scleroderma (108). As a homologue of BAFF, elevated levels of a proliferation-inducing ligand (APRIL) have also been reported in SSc patients and have been identified as a marker for pulmonary fibrosis, whilst high BAFF levels indicate severe skin sclerosis (109). A recent study by Glauzy and colleagues, however, did not find elevated BAFF levels in SSc patients compared with healthy controls and hence these authors concluded that BAFF is unlikely to be a driving factor in SSc pathogenesis (19).

Finally, T follicular helper (Tfh) cell production of IL-21 drives autoantibody producing plasma cell differentiation from

effector B cells and can promote fibrosis through induction of IL-6, TGF- $\beta$  and CC chemokine ligand 2 (CCL2) and B cell interaction with fibroblasts. In this respect it is notable that SSc fibroblasts have increased IL-21R expression (110).

# 8 B cell involvement in scleroderma pathogenesis: Evidence from animal models

SSc is a complex disease with high patient-patient heterogeneity. As such, it has been difficult to develop an animal model of SSc which captures all aspects of disease pathogenesis. Generally, these models are murine based and have a fibrotic skin and lung signature induced by either profibrotic agents such as bleomycin, or genetic manipulations leading to pro-fibroblast signalling (111). The tight-skin mouse (TSK) model of SSc is characterised by skin fibrosis and autoantibody production, as well as an abundance of BAFF and overexpression of CD19 (112–114). Using this model, it was shown that B cell activation is important for fibrosis and autoantibody production as CD19 deficiency decreased skin fibrosis and abrogated autoantibody production in TSK mice (113). Moreover, skin fibrosis and autoantibody production were prevented with the use of a BAFF antagonist (114).

Another mouse model of SSc involves using a subcutaneous injection of bleomycin to induce skin and lung fibrosis as well as autoantibody production. In this setting, bleomycin induces hyaluronan production which activates B cells through TLR4 (115). In the bleomycin model for lung fibrosis, it was demonstrated that CD19 signalling is crucial for B cell infiltration into the lung tissue and is associated with upregulation of the chemokine CXCR3. Loss of CD19 attenuated inflammation and reduced mortality, while CD19 overexpression increased mortality (115). Moreover, transgenic mice that overexpress CD19 spontaneously produce autoantibodies and lose immunotolerance (16, 116).

Sanges and colleagues developed a new murine model of SSc which was induced through daily intradermal injections of hypochlorous acid (HOCL) (26). The authors found significant B cell infiltration in the skin of HOCL injected mice in the later stages of disease but not in the earlier stages when compared with PBS-treated mice. In addition, splenic B cells in HOCL-treated mice produced significantly more IL-6 and CCL3 while IL-10 production was significantly reduced during the early stages of the disease, but levels of IL-10 production matched those of control mice at the later stage (26). This model showed high concordance with observations of B cell perturbations in human scleroderma patients. This is defined by B cell involvement through an early expansion of transitional B cells and late expansion of the mature naïve subset with an overall decrease in the number of Bregs, plasmablasts and memory B cells (26).

Interestingly, a study using a bleomycin-induced model of scleroderma highlighted how B cell-specific IL-6 deficient mice had attenuated skin and lung fibrosis whilst B cell-specific IL-10deficient mice had more severe fibrosis. Using this model it was also shown that IL-6 producing B effector cells or Beffs infiltrated inflamed skin and were induced to proliferate through BAFF. However, BAFF suppressed Breg generation. In addition, a BAFF antagonist attenuated skin and lung fibrosis and reduced Beffs but not Bregs. These data provide further evidence for a pathogenic role for BAFF and IL-6 produced by B cells in scleroderma models and a protective role for IL-10 (108).

Recently, a new mouse model of SSc has emerged. This model depends on topoisomerase I injection with complete Freud's adjuvant. The mice then develop skin and lung fibrosis with a defined Th2/Th17 response and increased IL-6 production. Using this model, it was shown that loss of IL-6 expression significantly improved skin and lung fibrosis (117).

# 9 B cell infiltration in the skin and lungs

Various studies have examined the cellular infiltrates in lesional tissue of SSc patients (118–122). Bosello and colleagues characterised the inflammatory cell infiltrate in scleroderma skin and found CD20<sup>+</sup> B cells in 60.7% of SSc patients. There was an increased number of CD20<sup>+</sup> B cells in patients with early disease and B cell infiltration appeared to correlate with worsening skin score. Importantly, no CD20<sup>+</sup> B cells were found in the skin of healthy individuals (120). Evidence suggests that IL-10 producing B cells migrate from the peritoneum to inflamed skin sites where they play important regulatory and protective functions. Hence, if B cell capacity to produce IL-10 is reduced in SSc patients, then the function of these regulatory skin-homing B cells are likely to be impaired (123).

B cell aggregates have been found in lung tissues of SSc patients with ILD, whilst transcriptomic data has also found evidence of lesional B cell infiltration (118, 121, 122). Furthermore, foci of B cell aggregates have been found in lung alveolar interstitium of SSc patients with ILD (122). Recent data using next-generation RNA sequencing in patients with early dcSSc demonstrated B cell signatures in 67% of patients, higher than published data in established disease (121).

# 10 Therapeutic strategies targeting B cells in SSc

The benefits of immunosuppressive therapies for scleroderma lung and skin disease provide further evidence for the role of B cell autoimmunity in SSc. However, the mixed effects of these biologic treatments also reveal the complexity of SSc pathogenesis and the strategic challenges faced when treating this disease (124).

The Scleroderma Lung Study (SLS) 1, comparing oral cyclophosphamide for 1 year vs placebo, reported a small improvement in forced vital capacity (FVC) with the treatment alongside improvements in dyspnoea scores and skin thickening (125). Using single cell analysis, an individual's B cell profile has been associated with cyclophosphamide response. Of note, cyclophosphamide responders had increased IL-10 producing regulatory B cells and reduced IL-6 effector B cells posttreatment (126). The SLS 2 study subsequently showed equivalence between 24 months of mycophenolate mofetil (MMF) and 12-months of oral cyclophosphamide followed by placebo with the average improvement in FVC of 7-8% (127). Post-hoc analysis of both the SENSCIS trial assessing nintedanib (a tyrosine kinase inhibitor) and the RESOLVE-1 trial assessing lenabasum (a cannabinoid type 2 receptor) in SSc, further demonstrated the efficacy of MMF in SSc. MMF inhibits both B and T cell proliferation and antibody production (128-130). With cyclophosphamide and MMF both impacting B cells alongside other mechanisms of action, further research is needed to disentangle the relative importance of their effects on B cell mediated pathogenesis.

Autologous stem cell transplant (aSCT) has emerged as a treatment option for autoimmune diseases with the rationale that a subsequent new self-tolerant immune system develops posttransplant (131). Three randomised controlled trials have supported this approach in patients with severe SSc with the ASSIST and ASTIS studies performing non-myeloablative aSCT and SCOT trial using myelo-ablative aSCT (132-134). All three trials reported sustained improvements in both skin and lung disease compared with standard of care with cyclophosphamide. The SCOT trial reported a lower treatment related mortality than previous studies, however, early treatment mortality due to increased rates of infection remains a concern (133). Gernert and colleagues demonstrated that the predominant B cell population post-transplantation was naïve B cells (CD27<sup>-</sup>/IgD<sup>+</sup>) with reduced percentages of memory B cells at 1-year post aSCT. An increased regulatory B cell phenotype with increased B cell IL-10 production was also found post-aSCT (28). A further study on 22 patients pre- and post- aSCT for SSc demonstrated similar changes in B cell populations with increased naïve B cells over prolonged follow-ups and sustained decreases in unswitched, switched and double negative B cells (135).

Rituximab, a chimeric monoclonal antibody against human CD20 on B cells provides compelling and more specific evidence for the role of B cells in SSc. Initial studies including small case series and open label trials demonstrated promising results with improvements of lung and skin fibrosis with rituximab (136-144). Alongside these studies, an initial observational EUSTAR study comparing 63 patients who received rituximab in routine clinical practice to matched controls found improved skin thickening and stabilisation of ILD with rituximab treatment (145). These findings are supported by mechanistic evidence that dermal B cells are completely, or nearly completely, depleted by rituximab therapy and a significant reduction in IL-6 at 6 months post treatment occurred with IL-6 known to be predictive of decline in FVC (136, 139, 146). A downregulation of fibroblast type I collagen gene expression with rituximab has also been demonstrated and early depletion of peripheral B cells at 2 weeks after rituximab therapy was negatively correlated with % forced vital capacity improvement at week 24 (147, 148).

The subsequent larger prospective EUSTAR study including 254 patients treated with rituximab vs 9575 propensity-score matched patients showed a significant improvement in skin fibrosis but failed to show an effect of rituximab on FVC or carbon monoxide diffusion capacity (DLCO) decline (149). A recent meta-analysis of rituximab in SSc-ILD from Goswami and colleagues, included 20 studies, with 575 SSc patients receiving rituximab treatment. Rituximab improved FVC and DLCO by 4.48% and 3.47%, respectively at 6 months and 7.03% and 4.08% at 12 months. Additionally, there were concomitant reductions in

the Modified Rodnan skin score (mRSS) which is used to evaluate skin thickness, with a higher score indicating thicker skin (150). It must be noted that only 2 of the 20 studies included were randomised control trials and neither were double-blinded. Another meta-analysis published in 2020 by Tang and colleagues including 14 studies with 597 participants found rituximab resulted in stability but not improvement of FVC (151).

The DESIRES trial, a double-blind placebo-controlled trial published in 2021, randomised both diffuse and limited SSc patients with an mRSS of  $\geq 10$  to receive weekly rituximab for 4 weeks, or placebo with absolute change in mRSS at 6 months as the primary endpoint (152). An improvement in skin score with rituximab vs placebo (absolute change in mRSS at 6 months -6.30 rituximab vs 2.14 placebo, difference -8.44 p<0.0001) was found and although most patients included in this study had relatively mild associated interstitial lung disease (ILD), rituximab did improve % FVC at 24 weeks (0.09% vs -2.87%, difference 2.96%, 95% CI 0.08 - 5.84, p=0.044). This provides more definitive evidence for the role of B cell depletion. We will wait to see if these results are reinforced by the soon to be published randomised controlled RECITAL trial assessing the effect of rituximab vs cyclophosphamide in connective tissuedisease-associated ILD (153).

Changes in autoantibody levels have been inconsistently reported following rituximab therapy (154-156) with the inability of rituximab to deplete autoantibody-producing long lived plasma cells seen in SSc (157). CD19 is expressed on a broader range of B cell subsets than CD20, including earlier B cell precursor cells, plasmablasts and some plasma cells (157). Elevated plasma cell gene signatures have been found in SSc compared with healthy controls and correlated with disease activity (158). A phase-I randomised placebo-controlled trial assessing a humanised monoclonal antibody targeting CD19 (MEDI551, inebilizumab) found it effectively depleted B cells and plasma cells and appeared well tolerated in SSc (159). At baseline patients with a high plasma cell gene signature were more likely to respond to inebilizumab than those with low plasma cell gene signatures, thereby supporting a role for plasma cells in disease pathogenesis (158). Targeting CD38, a type II glycoprotein highly expressed on several B cell subsets including short and long lived plasma cells, may provide another approach to target autoantibody production; however, studies are yet to assess anti-CD38 therapy in SSc (160).

Belimumab, a treatment licensed in SLE, binds soluble BAFF/BLyS. A double blind randomised control trial of belimumab vs placebo with background MMF including 20 SSc patients failed to reach statistical significance but demonstrated greater improvement in mRSS with belimumab compared with MMF alone (161). Responders to belimumab had significant changes in gene expression involved in B cell signalling and fibrotic signalling, consistent with the mechanism of action of the drug (161).

Blocking IL-6 signalling using tocilizumab, a humanized monoclonal antibody against the human IL-6 receptor  $\alpha$  chain

has been assessed in the phase II FaSScinate trial and subsequent phase III FocuSSced trial (162, 163). Although both failed to meet their primary endpoint regarding difference in mean change in baseline mRSS at week 24 and 48, both found that tociluzumab slowed lung function decline in early active disease (FVC decline >10%: 17% placebo vs 5% tocilizumab). The interaction of IL-6 and B cells discussed earlier in this review suggests some treatment effects are likely driven by effects of the drug on B cells; however, further research is required to delineate this (89, 90).

Both the SENSCIS trial and the ongoing Scleroderma lung study III are assessing the effect of combination therapy with oral anti-fibrotics and MMF. It is therefore likely that future treatment approaches will include a combination of immune modulatory and anti-fibrotic therapy with B cells appearing to play a crucial role in the interaction of these pathways (164, 165).

### **11** Future perspectives

B cells are able to shape SSc pathogenesis through a number of mechanisms as described throughout this review. Improved understanding of these mechanisms is needed to demonstrate the overall relevance and description of these pathogenic B cells and outline their involvement in the wider context of SSc such as their interaction with other immune and endothelial cells. A potential pathway for B cell autoreactivity and pathogenic SSc involvement could be triggered by an initial environmental signal such as high levels of IL-6. This could then lead to inappropriate B cell activation and survival marked by inflammatory cytokine and autoantibody production. Autoreactive B cells are likely to interact with other immune cells throughout this process such as fibroblasts, Tfh cells during germinal centre reactions and CD4<sup>+</sup> T helper cells in antigen presentation. In the future, increased knowledge of these interactions in the context of SSc will improve overall understanding of disease pathways and could lead to new and more targeted therapeutic strategies with less side effects.

Thus, as studies continue to delineate the relevant pathogenic B cell signalling pathways, disease treatment is likely to significantly improve with better and safer biologic therapies. For example, improved understanding of the biomarkers which predict patient response to B cell modulating drugs through single-cell RNA sequencing is likely to improve patient stratification. Meanwhile, other studies may reveal B cell signalling pathways or protein targets that are responsible for maintaining the pro-fibrotic disease state.

### 12 Conclusions

SSc is a complex disease with multiple cell types and pathways likely to contribute towards pathogenesis. B cells can

interact with many of these pathways directly contributing to the inflammatory and profibrotic phenotype characteristic of SSc. It is currently thought that the initial stages of disease are marked by a proinflammatory response which later leads to a more pronounced fibrotic response. This has implications for when to treat patients and with which biologics. No matter the disease stage, B cells are key immune mediators and are, therefore, likely to be central in scleroderma pathology. As understanding of disease development, B cell involvement and patient heterogeneity continue to improve, this will result in improved patient prognosis in SSc.

### Author contributions

CB and NG wrote the draft manuscript with VO, TT, DA, CD, and RM as editors. All authors contributed to the text and approved the final submission. CB was responsible for figure design and legend assembly.

### Funding

The authors are grateful for a Versus Arthritis grant (22534) and a Medical Research Council UK/Scleroderma & Raynaud's UK co-funded grant (MR/V030108/1).

### Acknowledgments

We would like to thank Dr Fabian Flores Borja and Dr Louisa James for their continued support at Queen Mary University of London. All figures were created with BioRender.com.

### Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

### Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

### References

1. Pattanaik D, Brown M, Postlethwaite BC, Postlethwaite AE. Pathogenesis of systemic sclerosis. *Front Immunol* (2015) 6:272. doi: 10.3389/fimmu.2015.00272

2. Sakkas LI, Bogdanos DP. Systemic sclerosis: New evidence re-enforces the role of b cells. *Autoimmun Rev* (2016) 15:155–61. doi: 10.1016/j.autrev.2015.10.005

3. Van Den Hoogen F, Khanna D, Fransen J, Johnson SSR, Baron M, Tyndall A, et al. 2013 classification criteria for systemic sclerosis: an American college of rheumatology/European league against rheumatism collaborative initiative. *Ann Rheum Dis* (2013) 72:1747–55. doi: 10.1136/ANNRHEUMDIS-2013-204424

4. Denton CP, Khanna D. Systemic sclerosis. Lancet (2017) 390:1685-99. doi: 10.1016/S0140-6736(17)30933-9

5. Carwile LeRoy E, Black C, Fleischmajer R, Jablonska S, Krieg T, Medsger TA, et al. Scleroderma (systemic sclerosis): Classification, subsets and pathogenesis. *J Rheumatol* (1988) 15:202–5.

6. Gu YS, Kong J, Cheema GS, Keen CL, Wick G, Gershwin ME. The immunobiology of systemic sclerosis. *Semin Arthritis Rheum* (2008) 38:132–60. doi: 10.1016/j.semarthrit.2007.10.010

7. Winstone TA, Assayag D, Wilcox PG, Dunne JV, Hague CJ, Leipsic J, et al. Predictors of mortality and progression in scleroderma-associated interstitial lung disease: A systematic review. *Chest* (2014) 146:422-36. doi: 10.1378/chest.13-2626

8. Nihtyanova SI, Denton CP. Pathogenesis of systemic sclerosis associated interstitial lung disease. J Scleroderma Relat Disord (2020) 5:6–16. doi: 10.1177/2397198320903867

9. Nihtyanova SI, Denton CP. Autoantibodies as predictive tools in systemic sclerosis. Nat Rev Rheumatol (2010) 6:112-6. doi: 10.1038/nrrheum.2009.238

10. Burbelo PD, Gordon SM, Waldman M, Edison JD, Little DJ, Stitt RS, et al. Autoantibodies are present before the clinical diagnosis of systemic sclerosis. *PloS One* (2019) 14:e0214202. doi: 10.1371/journal.pone.0214202

11. Bosello S, De Luca G, Tolusso B, Lama G, Angelucci C, Sica G, et al. B cells in systemic sclerosis: A possible target for therapy. *Autoimmun Rev* (2011) 10:624–30. doi: 10.1016/j.autrev.2011.04.013

12. Bemark M. Translating transitions - how to decipher peripheral human b cell development. J BioMed Res (2015) 29:264. doi: 10.7555/JBR.29.20150035

13. Martin VG, Wu YCB, Townsend CL, Lu GHC, O'Hare JS, Mozeika A, et al. Transitional b cells in early human b cell development – time to revisit the paradigm? *Front Immunol* (2016) 7:546/BIBTEX. doi: 10.3389/FIMMU. 2016.00546/BIBTEX

14. Fox DA, Lundy SK, Whitfield ML, Berrocal V, Campbell P, Rasmussen S, et al. Lymphocyte subset abnormalities in early diffuse cutaneous systemic sclerosis. *Arthritis Res Ther 2021 231* (2021) 23:1–12. doi: 10.1186/S13075–020–02383–W

15. Sato S, Fujimoto M, Hasegawa M, Takehara K. Altered blood b lymphocyte homeostasis in systemic sclerosis: Expanded naive b cells and diminished but activated memory b cells. *Arthritis Rheum* (2004) 50:1918–27. doi: 10.1002/art.20274

16. Mavropoulos A, Simopoulou T, Varna A, Liaskos C, Katsiari C, Bogdanos D, et al. Breg cells are numerically decreased and functionally impaired in patients with systemic sclerosis. *Arthritis Rheumatol* (2016) 68:494–504. doi: 10.1002/ART.39437

17. Soto L, Ferrier A, Aravena O, Fonseca E, Berendsen J, Biere A, et al. Systemic sclerosis patients present alterations in the expression of molecules involved in b-cell regulation. *Front Immunol* (2015) 6:496. doi: 10.3389/fimmu.2015.00496

18. Taher TE, Ong VH, Bystrom J, Hillion S, Simon Q, Denton CP, et al. Association of defective regulation of autoreactive interleukin-6-Producing transitional b lymphocytes with disease in patients with systemic sclerosis. *Arthritis Rheumatol* (2018) 70:450-61. doi: 10.1002/art.40390

19. Glauzy S, Olson B, May CK, Parisi D, Massad C, Hansen JE, et al. Defective early b cell tolerance checkpoints in patients with systemic sclerosis allow the production of self-antigen-specific clones. *Arthritis Rheumatol* (2022) 74(2):307–17. doi: 10.1002/ART.41927

20. Simon Q, Pers JO, Cornec D, Le Pottier L, Mageed RA, Hillion S. In-depth characterization of CD24highCD38high transitional human b cells reveals different regulatory profiles. *J Allergy Clin Immunol* (2016) 137:1577–1584.e10. doi: 10.1016/j.jaci.2015.09.014

21. Hillmer EJ, Zhang H, Li HS, Watowich SS. STAT3 signaling in immunity. Cytokine Growth Factor Rev (2016) 31:1–15. doi: 10.1016/J.CYTOGFR.2016.05.001

22. Niemand C, Nimmesgern A, Haan S, Fischer P, Schaper F, Rossaint R, et al. Activation of STAT3 by IL-6 and IL-10 in primary human macrophages is differentially modulated by suppressor of cytokine signaling 3. *J Immunol* (2003) 170:3263–72. doi: 10.4049/JIMMUNOL.170.6.3263

23. Kasmi KCEl, Holst J, Coffre M, Mielke L, Pauw A, Lhocine N, et al. General nature of the STAT3-activated anti-inflammatory response. *J Immunol* (2006) 177:7880–8. doi: 10.4049/JIMMUNOL.177.11.7880

24. Aravena O, Ferrier A, Menon M, Mauri C, Aguillón JC, Soto L, et al. TIM-1 defines a human regulatory b cell population that is altered in frequency and function in systemic sclerosis patients. *Arthritis Res Ther* (2017) 19. doi: 10.1186/s13075-016-1213-9

25. Matsushita T, Hamaguchi Y, Hasegawa M, Takehara K, Fujimoto M. Decreased levels of regulatory b cells in patients with systemic sclerosis: association with autoantibody production and disease activity. *Rheumatology* (2016) 55:263–7. doi: 10.1093/rheumatology/kev331

26. Sanges S, Jendoubi M, Kavian N, Hauspie C, Speca S, Crave J, et al. B cell homeostasis and functional properties are altered in an hypochlorous acid–induced murine model of systemic sclerosis. *Front Immunol* (2017) 8:53. doi: 10.3389/FIMMU.2017.00053

27. Rosser EC, Mauri C. Regulatory b cells: Origin, phenotype, and function. Immunity (2015) 42:607–12. doi: 10.1016/J.IMMUNI.2015.04.005

28. Gernert M, Tony HP, Schwaneck EC, Gadeholt O, Schmalzing M. Autologous hematopoietic stem cell transplantation in systemic sclerosis induces long–lasting changes in b cell homeostasis toward an anti–inflammatory b cell cytokine pattern. *Arthritis Res Ther* (2019) 21:1–10. doi: 10.1186/S13075–019–1889–8/FIGURES/4

29. Simon D, Balogh P, Bognár A, Kellermayer Z, Engelmann P, Németh P, et al. Reduced non-switched memory b cell subsets cause imbalance in b cell repertoire in systemic sclerosis. *Clin Exp Rheumatol* (2016) 34 Suppl 1:30–6.

30. Steen VD. Autoantibodies in systemic sclerosis. Semin Arthritis Rheum (2005) 35:35-42. doi: 10.1016/j.semarthrit.2005.03.005

31. Hesselstrand R, Scheja A, Shen GQ, Wiik A, A°Ckesson A. The association of antinuclear antibodies with organ involvement and survival in systemic sclerosis. *Rheumatology* (2003) 42:534–40. doi: 10.1093/RHEUMATOLOGY/KEG170

32. Bae S, Saggar R, Bolster MB, Chung L, Csuka ME, Derk C, et al. Baseline characteristics and follow-up in patients with normal haemodynamics versus borderline mean pulmonary arterial pressure in systemic sclerosis: results from the PHAROS registry. *Ann Rheum Dis* (2012) 71:1335–42. doi: 10.1136/ANNRHEUMDIS-2011-200546

33. Hudson M, Mahler M, Pope J, You D, Tatibouet S, Steele R, et al. Clinical correlates of CENP-a and CENP-b antibodies in a large cohort of patients with systemic sclerosis. *J Rheumatol* (2012) 39:787–94. doi: 10.3899/RHEUM.111133

34. Tomaszewski M, Bębnowska D, Hrynkiewicz R, Dworzyński J, Niedźwiedzka-rystwej P, Kopeć G, et al. Role of the immune system elements in pulmonary arterial hypertension. *J Clin Med* (2021) 10:3757. doi: 10.3390/JCM10163757

35. Breitling S, Hui Z, Zabini D, Hu Y, Hoffmann J, Goldenberg NM, et al. The mast cell-b cell axis in lung vascular remodeling and pulmonary hypertension. *Am J Physiol – Lung Cell Mol Physiol* (2017) 312:L710–21. doi: 10.1152/AJPLUNG.00311.2016/ASSET/IMAGES/LARGE/ZH50041772100008.JPEG

36. Mizuno S, Farkas L, Al Husseini A, Farkas D, Gomez-Arroyo J, Kraskauskas D, et al. Severe pulmonary arterial hypertension induced by SU5416 and ovalbumin immunization. *Am J Respir Cell Mol Biol* (2013) 47:679–87. doi: 10.1165/RCMB.2012-0077OC

37. Kayser C, Fritzler MJ. Autoantibodies in systemic sclerosis: Unanswered questions. *Front Immunol* (2015) 6:167. doi: 10.3389/fimmu.2015.00167

38. Senécal J–L, Hénault J, Raymond Y. The pathogenic role of autoantibodies to nuclear autoantigens in systemic sclerosis (scleroderma). *J Rheumatol* (2005) 32:1643–9. doi: 10.1177/2397198319870667

39. Hénault J, Tremblay M, Clément I, Raymond Y, Senécal J. Direct binding of anti-DNA topoisomerase I autoantibodies to the cell surface of fibroblasts in patients with systemic sclerosis. *Arthritis Rheum* (2004) 50:3265–74. doi: 10.1002/ART.20515

40. Sobanski V, Dauchet L, Lefèvre G, Lambert M, Morell–Dubois S, Sy T, et al. Prevalence of anti–RNA polymerase III antibodies in systemic sclerosis: New data from a French cohort and a systematic review and meta–analysis. *Arthritis Rheumatol* (2014) 66:407–17. doi: 10.1002/art.38219

41. Rigamonti C, Shand LM, Feudjo M, Bunn CC, Black CM, Denton CP, et al. Clinical features and prognosis of primary biliary cirrhosis associated with systemic sclerosis. *Gut* (2006) 55:388–94. doi: 10.1136/gut.2005.075002

42. Moinzadeh P, Fonseca C, Hellmich M, Shah AA, Chighizola C, Denton CP, et al. Association of anti–RNA polymerase III autoantibodies and cancer in scleroderma. *Arthritis Res Ther* (2014) 16:R53. doi: 10.1186/ar4486

43. Moroncini G, Svegliati Baroni S, Gabrielli A. Agonistic antibodies in systemic sclerosis. *Immunol Lett* (2018) 195:83–7. doi: 10.1016/J.IMLET. 2017.10.007

44. Raschi E, Privitera D, Bodio C, Lonati PA, Borghi MO, Ingegnoli F, et al. Scleroderma-specific autoantibodies embedded in immune complexes mediate endothelial damage: an early event in the pathogenesis of systemic sclerosis. *Arthritis Res Ther 2020 221* (2020) 22:1–17. doi: 10.1186/S13075-020-02360-3

45. Odaka M, Hasegawa M, Hamaguchi Y, Ishiura N, Kumada S, Matsushita T, et al. Autoantibody-mediated regulation of b cell responses by functional anti-CD22 autoantibodies in patients with systemic sclerosis. *Clin Exp Immunol* (2010) 159:176–84. doi: 10.1111/j.1365–2249.2009.04059.x

46. Matsushita T, Hasegawa M, Matsushita Y, Echigo T, Wayaku T, Horikawa M, et al. Elevated serum BAFF levels in patients with localized scleroderma in contrast to other organ–specific autoimmune diseases. *Exp Dermatol* (2007) 16:87–93. doi: 10.1111/J.1600–0625.2006.00485.X

47. Sato S, Hasegawa M, Fujimoto M, Tedder TF, Takehara K. Quantitative genetic variation in CD19 expression correlates with autoimmunity. *J Immunol* (2000) 165:6635–43. doi: 10.4049/JIMMUNOL.165.11.6635

48. Mongini PK, Vilensky MA, Highet PF, Inman JK. The affinity threshold for human b cell activation *via* the antigen receptor complex is reduced upon coligation of the antigen receptor with CD21 (CR2). *J Immunol* (1997) 159:3782–91.

49. Asano N, Fujimoto M, Yazawa N, Shirasawa S, Hasegawa M, Okochi H, et al. B lymphocyte signaling established by the CD19/CD22 loop regulates autoimmunity in the tight-skin mouse. *Am J Pathol* (2004) 165:641–50. doi: 10.1016/S0002-9440(10)63328-7

50. Elgueta R, Benson MJ, De Vries VC, Wasiuk A, Guo Y, Noelle RJ. Molecular mechanism and function of CD40/CD40L engagement in the immune system. *Immunol Rev* (2009) 229:152–72. doi: 10.1111/J.1600–065X.2009.00782.X

51. Fujimoto M, Poe JC, Hasegawa M, Tedder TF. CD19 regulates intrinsic b lymphocyte signal transduction and activation through a novel mechanism of processive amplification. *Immunol Res* (2000) 22:281–98. doi: 10.1385/IR:22:2–3:281

52. Kremlitzka M, Polgar A, Fulop L, Kiss E, Poor G, Erdei A. Complement receptor type 1 (CR1, CD35) is a potent inhibitor of b-cell functions in rheumatoid arthritis patients. *Int Immunol* (2013) 25:25–33. doi: 10.1093/intimm/dxs090

53. Isaak A, Gergely P, Szekeres Z, Prechl J, Poor G, Erdei A, et al. Physiological up-regulation of inhibitory receptors fc RII and CR1 on memory b cells is lacking in SLE patients. *Int Immunol* (2008) 20:185–92. doi: 10.1093/intimm/dxm132

54. Dai S, Jia R, Zhang X, Fang Q, Huang L. The PD-1/PD-Ls pathway and autoimmune diseases. *Cell Immunol* (2014) 290:72-9. doi: 10.1016/J.CELLIMM.2014.05.006

55. Yamane H, Isozaki H, Takeyama M, Ochi N, Kudo K, Honda Y, et al. Programmed cell death protein 1 and programmed death-ligand 1 are expressed on the surface of some small-cell lung cancer lines. *Am J Cancer Res* (2015) 5:1553.

56. Fukasawa T, Yoshizaki A, Ebata S, Nakamura K, Saigusa R, Miura S, et al. Contribution of soluble forms of programmed death 1 and programmed death ligand 2 to disease severity and progression in systemic sclerosis. *Arthritis Rheumatol* (2017) 69:1879–90. doi: 10.1002/ART.40164

57. Yoshizaki A. Pathogenic roles of b lymphocytes in systemic sclerosis. Immunol Lett (2018) 195:76–82. doi: 10.1016/J.IMLET.2018.01.002

58. Yanaba K, Hayashi M, Yoshihara Y, Nakagawa H. Serum levels of soluble programmed death-1 and programmed death ligand-1 in systemic sclerosis: Association with extent of skin sclerosis. *J Dermatol* (2016) 43:954-7. doi: 10.1111/1346-8138.13339

59. Hua Z, Hou B. TLR signaling in b-cell development and activation. *Cell Mol Immunol 2013 102* (2012) 10:103–6. doi: 10.1038/cmi.2012.61

60. Cheng S, Wang H, Zhou H. The role of TLR4 on b cell activation and anti- $\beta$ 2GPI antibody production in the antiphospholipid syndrome. *J Immunol Res* (2016) 2016. doi: 10.1155/2016/1719720

61. Bhattacharyya S, Varga J. Endogenous ligands of TLR4 promote unresolving tissue fibrosis: Implications for systemic sclerosis and its targeted therapy. *Immunol Lett* (2018) 195:9–17. doi: 10.1016/J.IMLET.2017.09.011

62. Suthers AN, Sarantopoulos S. TLR7/TLR9– and b cell receptor-signaling crosstalk: Promotion of potentially dangerous b cells. *Front Immunol* (2017) 8:775/ BIBTEX. doi: 10.3389/FIMMU.2017.00775/BIBTEX

63. Wang T, Marken J, Chen J, Tran VB, Li QZ, Li M, et al. High TLR7 expression drives the expansion of CD19+CD24HiCD38hi transitional b cells and autoantibody production in SLE patients. *Front Immunol* (2019) 10:1243/BIBTEX. doi: 10.3389/FIMMU.2019.01243/BIBTEX

64. Vreća M, Zeković A, Damjanov N, Andjelković M, Ugrin M, Pavlović S, et al. Expression of TLR7, TLR9, JAK2, and STAT3 genes in peripheral blood mononuclear cells from patients with systemic sclerosis. *J Appl Genet* (2018) 59:59–66. doi: 10.1007/S13353-017-0415-4

65. Jellusova J, Rickert RC. The PI3K pathway in b cell metabolism. Crit Rev Biochem Mol Biol (2016) 51:359. doi: 10.1080/10409238.2016.1215288

66. Limon JJ, Fruman DA. Akt and mTOR in b cell activation and differentiation. *Front Immunol* (2012) 3:228/BIBTEX. doi: 10.3389/FIMMU.2012.00228/BIBTEX

67. Wang LD, Clark MR. B-cell antigen-receptor signalling in lymphocyte development. *Immunology* (2003) 110:411-20. doi: 10.1111/J.1365-2567. 2003.01756.X

68. Dawidowicz K, Dieudé P, Avouac J, Wipff J, Hachulla E, Diot E, et al. Association study of b-cell marker gene polymorphisms in European Caucasian patients with systemic sclerosis. *Clin Exp Rheumatol* (2011) 29:839–42.

69. Dieudé P, Wipff J, Guedj M, Ruiz B, Melchers I, Hachulla E, et al. BANK1 is a genetic risk factor for diffuse cutaneous systemic sclerosis and has additive effects with IRF5 and STAT4. *Arthritis Rheum* (2009) 60:3447–54. doi: 10.1002/ ART.24885

70. Abdelrasoul H, Werner M, Setz CS, Okkenhaug K, Jumaa H. PI3K induces b-cell development and regulates b cell identity. *Sci Rep* (2018) 8:1–15. doi: 10.1038/s41598-018-19460-5

71. Dieudé P, Guedj M, Wipff J, Ruiz B, Riemekasten G, Matucci-Cerinic M, et al. Association of the TNFAIP3 rs5029939 variant with systemic sclerosis in the European Caucasian population. *Ann Rheum Dis* (2010) 69:1958–64. doi: 10.1136/ARD.2009.127928

72. Koumakis E, Giraud M, Dieudé P, Cohignac V, Cuomo G, Airò P, et al. Brief report: Candidate gene study in systemic sclerosis identifies a rare and functional variant of the TNFAIP3 locus as a risk factor for polyautoimmunity. *Arthritis Rheum* (2012) 64:2746–52. doi: 10.1002/ART.34490

73. Tavares RM, Turer EE, Liu CL, Advincula R, Scapini P, Rhee L, et al. The ubiquitin modifying enayme A20 restricts b cell survival and prevents autoimmunity. *Immunity* (2010) 33:181. doi: 10.1016/J.IMMUNI.2010.07.017

74. Srinivasan L, Sasaki Y, Calado DP, Zhang B, Paik JH, DePinho RA, et al. PI3 kinase signals BCR-dependent mature b cell survival. *Cell* (2009) 139:573-86. doi: 10.1016/J.CELL.2009.08.041

75. Martin JE, Broen JC, David Carmona F, Teruel M, Simeon CP, Vonk MC, et al. Identification of CSK as a systemic sclerosis genetic risk factor through genome wide association study follow-up. *Hum Mol Genet* (2012) 21:2825–35. doi: 10.1093/HMG/DDS099

76. Gourh P, Agarwal SK, Martin E, Divecha D, Rueda B, Bunting H, et al. Association of the C8orf13–BLK region with systemic sclerosis in north–American and European populations. *J Autoimmun* (2010) 34:155–62. doi: 10.1016/J.JAUT.2009.08.014

77. Dieudé P, Guedj M, Wipff J, Avouac J, Hachulla E, Diot E, et al. The PTPN22 620W allele confers susceptibility to systemic sclerosis: findings of a large case-control study of European caucasians and a meta-analysis. *Arthritis Rheum* (2008) 58:2183–8. doi: 10.1002/ART.23601

78. Mahoney JM, Taroni J, Martyanov V, Wood TA, Greene CS, Pioli PA, et al. Systems level analysis of systemic sclerosis shows a network of immune and profibrotic pathways connected with genetic polymorphisms. *PloS Comput Biol* (2015) 11:1004005. doi: 10.1371/JOURNAL.PCBI.1004005

79. Simon D, Erdő-Bonyár S, Rapp J, Balogh P, Minier T, Nagy G, et al. Analysis of PI3K pathway associated molecules reveals dysregulated innate and adaptive functions of b cells in early diffuse cutaneous systemic sclerosis. *Int J Mol Sci* (2021) 22:1–13. doi: 10.3390/IJMS22062877

80. Koarada S, Tada Y, Kikuchi Y, Ushiyama O, Suzuki N, Ohta A, et al. CD180 (RP105) in rheumatic diseases. *Rheumatology* (2001) 40:1315–6. doi: 10.1093/ RHEUMATOLOGY/40.11.1315

81. Chaplin JW, Kasahara S, Clark EA, Ledbetter JA. Anti-CD180 (RP105) activates b cells to rapidly produce polyclonal ig *via* a T cell and MyD88-independent pathway. *J Immunol* (2011) 187:4199-209. doi: 10.4049/JIMMUNOL.1100198

82. Roe K, Shu GL, Draves KE, Giordano D, Pepper M, Clark EA. Targeting antigens to CD180 but not CD40 programs immature and mature b cell subsets to become efficient APCs. J Immunol (2019) 203:1715–29. doi: 10.4049/JIMMUNOL.1900549/-/DCSUPPLEMENTAL

83. Erdő-Bonyár S, Rapp J, Minier T, Ráth G, Najbauer J, Czirják L, et al. Tolllike receptor mediated activation of natural autoantibody producing b cell subpopulations in an autoimmune disease model. *Int J Mol Sci* (2019) 20. doi: 10.3390/IJMS20246152

84. Shi X, Shao T, Huo F, Zheng C, Li W, Jiang Z. An analysis of abnormalities in the b cell receptor repertoire in patients with systemic sclerosis using highthroughput sequencing. *PeerJ* (2020) 2020:e8370. doi: 10.7717/peerj.8370

85. Galson JD, Clutterbuck EA, Trück J, Ramasamy MN, Münz M, Fowler A, et al. BCR repertoire sequencing: different patterns of b cell activation after two meningococcal vaccines. *Immunol Cell Biol* (2015) 93:885. doi: 10.1038/ICB.2015.57

86. Fielding CA, Jones GW, McLoughlin RM, McLeod L, Hammond VJ, Uceda J, et al. Interleukin–6 signaling drives fibrosis in unresolved inflammation. *Immunity* (2014) 40:40. doi: 10.1016/J.IMMUNI.2013.10.022

87. Arkatkar T, Du SW, Jacobs HM, Dam EM, Hou B, Buckner JH, et al. B cell-derived IL-6 initiates spontaneous germinal center formation during systemic autoimmunity. *J Exp Med* (2017) 214:3207-17. doi: 10.1084/JEM.20170580

88. Zhou Y, Zhang Y, Han J, Yang M, Zhu J, Jin T. Transitional b cells involved in autoimmunity and their impact on neuroimmunological diseases. *J Transl Med* (2020) 18. doi: 10.1186/S12967-020-02289-W

89. Maeda K, Mehta H, Drevets DA, Coggeshall KM. IL-6 increases b-cell IgG production in a feed-forward proinflammatory mechanism to skew hematopoiesis and elevate myeloid production. *Blood* (2010) 115:4699. doi: 10.1182/BLOOD-2009-07-230631

90. Alarcon-Riquelme ME, Moller G, Fernandez C. Macrophage depletion decreases IgG anti-DNA in cultures from (NZB x NZW)F1 spleen cells by eliminating the main source of IL-6. *Clin Exp Immunol* (1993) 91:220–5. doi: 10.1111/j.1365-2249.1993.tb05886.x

91. Zi Z, Chapnick DA, Liu X. Dynamics of TGF– $\beta$ /Smad signaling. FEBS Lett (2012) 586:1921–8. doi: 10.1016/J.FEBSLET.2012.03.063

92. Leivonen SK, Lazaridis K, Decock J, Chantry A, Edwards DR, Kähäri VM. TGF- $\beta$ -Elicited induction of tissue inhibitor of metalloproteinases (TIMP)-3 expression in fibroblasts involves complex interplay between Smad3, p38 $\alpha$ , and ERK1/2. *PloS One* (2013) 8. doi: 10.1371/JOURNAL.PONE.0057474

93. Kubo M, Ihn H, Yamane K, Tamaki K. Upregulated expression of transforming growth factor-beta receptors in dermal fibroblasts of skin sections from patients with systemic sclerosis. *J Rheumatol* (2002) 29(12):2558–64.

94. Fillatreau S, Sweenie CH, McGeachy MJ, Gray D, Anderton SM. B cells regulate autoimmunity by provision of IL-10. *Nat Immunol 2002 310* (2002) 3:944-50. doi: 10.1038/ni833

95. Forestier A, Guerrier T, Jouvray M, Giovannelli J, Lefèvre G, Sobanski V, et al. Altered b lymphocyte homeostasis and functions in systemic sclerosis. *Autoimmun Rev* (2018) 17:244–55. doi: 10.1016/J.AUTREV.2017.10.015

96. Wynn TA. IL-13 effector functions. Annu Rev Immunol (2003) 21:425–56. doi: 10.1146/ANNUREV.IMMUNOL.21.120601.141142

97. Punnonen J, de Vries JE. IL-13 induces proliferation, ig isotype switching, and ig synthesis by immature human fetal b cells. *J Immunol* (1994) 152:1094–102.

98. Cocks BG, de Waal Malefyt R, Galizzi J-P, de Vries JE, Aversa G. IL-13 induces proliferation and differentiation of human b cells activated by the CD40 ligand. *Int Immunol* (1993) 5:657–63. doi: 10.1093/INTIMM/5.6.657

99. Hasegawa M, Fujimoto M, Kikuchi K, Takehara K. Elevated serum levels of interleukin 4 (IL-4), IL-10, and IL-13 in patients with systemic sclerosis. *J Rheumatol* (1997) 24:328-32.

100. Rankin AL, Mumm JB, Murphy E, Turner S, Yu N, McClanahan TK, et al. IL–33 induces IL–13–Dependent cutaneous fibrosis. *J Immunol* (2010) 184:1526–35. doi: 10.4049/JIMMUNOL.0903306

101. Rose WA, Okragly AJ, Hu NN, Daniels MR, Martin AP, Koh YT, et al. Interleukin-33 contributes toward loss of tolerance by promoting b-Cell-Activating factor of the tumor-Necrosis-Factor family (BAFF)-dependent autoantibody production. *Front Immunol* (2018) 9:2871/BIBTEX. doi: 10.3389/ FIMMU.2018.02871/BIBTEX

102. Ebisuno Y, Tanaka T, Kanemitsu N, Kanda H, Yamaguchi K, Kaisho T, et al. Cutting edge: The b cell chemokine CXC chemokine ligand 13/B lymphocyte chemoattractari is expressed in the high endothelial venules of lymph nodes and peyer's patches and affects b cell trafficking across high endothelial venules. *J Immunol* (2003) 171:1642–6. doi: 10.4049/JIMMUNOL.171.4.1642

103. Bellamri N, Viel R, Morzadec C, Lecureur V, Joannes A, de LB, et al. TNF- $\alpha$  and IL-10 control CXCL13 expression in human macrophages. J Immunol (2020) 204:2492–502. doi: 10.4049/JIMMUNOL.1900790

104. Carlsen HS, Baekkevold ES, Morton HC, Haraldsen G, Brandtzaeg P. Monocyte-like and mature macrophages produce CXCL13 (B cell-attracting chemokine 1) in inflammatory lesions with lymphoid neogenesis. *Blood* (2004) 104:3021–7. doi: 10.1182/BLOOD-2004-02-0701

105. Kalled SL. The role of BAFF in immune function and implications for autoimmunity. *Immunol Rev* (2005) 204:43-54. doi: 10.1111/J.0105-2896.2005.00219.X

106. Nemazee D. Mechanisms of central tolerance for b cells. *Nat Rev Immunol* (2017) 17:281–94. doi: 10.1038/nri.2017.19

107. Stadanlick JE, Cancro MP. BAFF and the plasticity of peripheral b cell tolerance. *Curr Opin Immunol* (2008) 20:158. doi: 10.1016/J.COI.2008.03.015

108. Matsushita T, Kobayashi T, Mizumaki K, Kano M, Sawada T, Tennichi M, et al. BAFF inhibition attenuates fibrosis in scleroderma by modulating the regulatory and effector b cell balance. *Sci Adv* (2018) 4:9944–55. doi: 10.1126/SCIADV.AAS9944

109. Matsushita T, Hasegawa M, Yanaba K, Kodera M, Takehara K, Sato S. Elevated serum BAFF levels in patients with systemic sclerosis: Enhanced BAFF signaling in systemic sclerosis b lymphocytes. *Arthritis Rheum* (2006) 54:192–201. doi: 10.1002/art.21526

110. Worrell JC, O'Reilly S. Bi-directional communication: Conversations between fibroblasts and immune cells in systemic sclerosis. *J Autoimmun* (2020) 113. doi: 10.1016/J.JAUT.2020.102526

111. Melissaropoulos K, Daoussis D. B cells in systemic sclerosis: from pathophysiology to treatment. *Clin Rheumatol* (2021) 40:2621–31. doi: 10.1007/S10067-021-05665-Z

112. Green MC, Sweet HO, Bunker LE. Tight-skin, a new mutation of the mouse causing excessive growth of connective tissue and skeleton. *Am J Pathol* (1976) 82:493.

113. Saito E, Fujimoto M, Hasegawa M, Komura K, Hamaguchi Y, Kaburagi Y, et al. CD19–dependent b lymphocyte signaling thresholds influence skin fibrosis and autoimmunity in the tight-skin mouse. *J Clin Invest* (2002) 109:1453. doi: 10.1172/JCI15078

114. Matsushita T, Fujimoto M, Hasegawa M, Matsushita Y, Komura K, Ogawa F, et al. BAFF antagonist attenuates the development of skin fibrosis in tight-skin mice. *J Invest Dermatol* (2007) 127:2772–80. doi: 10.1038/SJ.JID.5700919

115. Komura K, Yanaba K, Horikawa M, Ogawa F, Fujimoto M, Tedder T, et al. CD19 regulates the development of bleomycin–induced pulmonary fibrosis in a mouse model. *Arthritis Rheum* (2008) 58:3574–84. doi: 10.1002/ART.23995

116. Tedder TF, Poe JC, Fujimoto M, Haas KM, Sato S. The CD19–CD21 signal transduction complex of b lymphocytes regulates the balance between health and autoimmune disease: Systemic sclerosis as a model system. *Curr Dir Autoimmun* (2005) 8:55–90. doi: 10.1159/000082087

117. Yoshizaki A, Yanaba K, Ogawa A, Asano Y, Kadono T, Sato S. Immunization with DNA topoisomerase I and freund's complete adjuvant induces skin and lung fibrosis and autoimmunity *via* interleukin-6 signaling. *Arthritis Rheum* (2011) 63:3575-85. doi: 10.1002/ART.30539

118. Whitfield ML, Finlay DR, Murray JI, Troyanskaya OG, Chi JT, Pergamenschikov A, et al. Systemic and cell type-specific gene expression patterns in scleroderma skin. *Proc Natl Acad Sci U.S.A.* (2003) 100:12319–24. doi: 10.1073/pnas.1635114100

119. Roumm AD, Whiteside TL, Medsger TA, Rodnan GP. Lymphocytes in the skin of patients with progressive systemic sclerosis. *Arthritis Rheum* (1984) 27:645–53. doi: 10.1002/art.1780270607

120. Bosello S, Angelucci C, Lama G, Alivernini S, Proietti G, Tolusso B, et al. Characterization of inflammatory cell infiltrate of scleroderma skin: B cells and skin score progression. *Arthritis Res Ther* (2018) 20:1–11. doi: 10.1186/s13075–018–1569–0

121. Skaug B, Khanna D, Swindell WR, Hinchcliff ME, Frech TM, Steen VD, et al. Global skin gene expression analysis of early diffuse cutaneous systemic sclerosis shows a prominent innate and adaptive inflammatory profile. *Ann Rheum Dis* (2020) 79:379–86. doi: 10.1136/ANNRHEUMDIS-2019-215894

122. Lafyatis R, O'Hara C, Feghali-Bostwick CA, Matteson E. B cell infiltration in systemic sclerosis-associated interstitial lung disease. *Arthritis Rheum* (2007) 56:3167–8. doi: 10.1002/art.22847

123. Geherin SA, Gómez D, Glabman RA, Ruthel G, Hamann A, Debes GF. IL-10 + innate-like b cells are part of the skin immune system and require  $\alpha$ 4 $\beta$ 1 integrin to migrate between the peritoneum and inflamed skin. *J Immunol* (2016) 196:2514–25. doi: 10.4049/JIMMUNOL.1403246/–/DCSUPPLEMENTAL

124. Khanna D, Lescoat A, Roofeh D, Bernstein EJ, Kazerooni EA, Roth MD, et al. Systemic sclerosis–associated interstitial lung disease: How to incorporate two food and drug administration–approved therapies in clinical practice. *Arthritis Rheumatol* (2022) 74:13–27. doi: 10.1002/art.41933

125. Tashkin DP, Elashoff R, Clements PJ, Goldin J, Roth MD, Furst DE, et al. Cyclophosphamide versus placebo in scleroderma lung disease. *N Engl J Med* (2006) 354:2655–66. doi: 10.1056/NEJMOA055120/SUPPL\_FILE/NEJM\_TASHKIN\_2655SA1.PDF

126. Ebata S, Yoshizaki A, Fukasawa T, Sato S. Single-cell analysis of autoreactive b cells in patients with systemic sclerosis revealed that responses to therapy are determined by effector and regulatory b cell balance. *J Immunol* (2020) 204:238. doi: 10.4049/jimmunol.204.Supp.238.16

127. Tashkin DP, Roth MD, Clements PJ, Furst DE, Khanna D, Kleerup EC, et al. Mycophenolate mofetil versus oral cyclophosphamide in scleroderma-related interstitial lung disease (SLS II): a randomised controlled, double-blind, parallel group trial. *Lancet Respir Med* (2016) 4:708–19. doi: 10.1016/S2213-2600(16) 30152-7

128. Allison AC, Eugui EM. Mycophenolate mofetil and its mechanisms of action. *Immunopharmacology* (2000) 47:85-118. doi: 10.1016/S0162-3109(00) 00188-0

129. Highland KB, Distler O, Kuwana M, Allanore Y, Assassi S, Azuma A, et al. Efficacy and safety of nintedanib in patients with systemic sclerosis-associated

interstitial lung disease treated with mycophenolate: a subgroup analysis of the SENSCIS trial. *Lancet Respir Med* (2021) 9:96–106. doi: 10.1016/S2213–2600(20) 30330–1

130. Spiera R, Furst DE, Frech TM, Kuwana M, Hummers L, Stevens W, et al. Background mycophenolate (MMF) treatment is associated with improved outcomes in a phase 3 trial of lenabasum in diffuse cutaneous systemic sclerosis (dcSSc). ACR Meet Abstr (2021).

131. Gratwohl A, Passweg J, Bocelli–Tyndall C, Fassas A, van Laar JM, Farge D, et al. Autologous hematopoietic stem cell transplantation for autoimmune diseases. *Bone Marrow Transplant 2005* 359 (2005) 35:869–79. doi: 10.1038/sj.bmt.1704892

132. Burt RK, Shah SJ, Dill K, Grant T, Gheorghiade M, Schroeder J, et al. Autologous non-myeloablative haemopoietic stem-cell transplantation compared with pulse cyclophosphamide once per month for systemic sclerosis (ASSIST): an open-label, randomised phase 2 trial. *Lancet* (2011) 378:498–506. doi: 10.1016/S0140-6736(11)60982-3

133. Sullivan KM, Goldmuntz EA, Keyes-Elstein L, McSweeney PA, Pinckney A, Welch B, et al. Myeloablative autologous stem-cell transplantation for severe scleroderma. *N Engl J Med* (2018) 378:35-47. doi: 10.1056/NEJMOA1703327

134. Van Laar JM, Farge D, Sont JK, Naraghi K, Marjanovic Z, Larghero J, et al. Autologous hematopoietic stem cell transplantation vs intravenous pulse cyclophosphamide in diffuse cutaneous systemic sclerosis: a randomized clinical trial. *JAMA* (2014) 311:2490–8. doi: 10.1001/JAMA.2014.6368

135. Lima–Júnior JR, Arruda LCM, Gonçalves MS, Dias JBE, Moraes DA, Covas DT, et al. Autologous haematopoietic stem cell transplantation restores the suppressive capacity of regulatory b cells in systemic sclerosis patients. *Rheumatol (Oxford)* (2021) 60:5538–48. doi: 10.1093/RHEUMATOLOGY/ KEAB257

136. Bosello S, De Santis M, Lama G, Spanò C, Angelucci C, Tolusso B, et al. B cell depletion in diffuse progressive systemic sclerosis: Safety, skin score modification and IL-6 modulation in an up to thirty-six months follow-up open-label trial. *Arthritis Res Ther* (2010) 12. doi: 10.1186/ar2965

137. Daoussis D, Liossis SNC, Tsamandas AC, Kalogeropoulou C, Paliogianni F, Sirinian C, et al. Effect of long-term treatment with rituximab on pulmonary function and skin fibrosis in patients with diffuse systemic sclerosis. *Clin Exp Rheumatol* (2012) 30:S17-22.

138. Moazedi-Fuerst FC, Kielhauser SM, Brickmann K, Hermann J, Lutfi A, Meilinger M, et al. Rituximab for systemic sclerosis: arrest of pulmonary disease progression in five cases. results of a lower dosage and shorter interval regimen. *Scand J Rheumatol* (2014) 43:257–8. doi: 10.3109/03009742.2013.869617

139. Smith V, Van Praet JT, Vandooren B, van der Cruyssen B, Naeyaert JM, Decuman S, et al. Rituximab in diffuse cutaneous systemic sclerosis: An open-label clinical and histopathological study. *Ann Rheum Dis* (2010) 69:193–7. doi: 10.1136/ARD.2008.095463

140. Smith V, Piette Y, Van Praet JT, Decuman S, Deschepper E, Elewaut D, et al. Two-year results of an open pilot study of a 2-treatment course with rituximab in patients with early systemic sclerosis with diffuse skin involvement. *J Rheumatol* (2013) 40:52-7. doi: 10.3899/IRHEUM.120778

141. Daoussis D, Liossis SNC, Tsamandas AC, Kalogeropoulou C, Kazantzi A, Sirinian C, et al. Experience with rituximab in scleroderma: results from a 1-year, proof-of-principle study. *Rheumatol (Oxford)* (2010) 49:271-80. doi: 10.1093/RHEUMATOLOGY/KEP093

142. Daoussis D, Melissaropoulos K, Sakellaropoulos G, Antonopoulos I, Markatseli TE, Simopoulou T, et al. A multicenter, open-label, comparative study of b-cell depletion therapy with rituximab for systemic sclerosis-associated interstitial lung disease. *Semin Arthritis Rheum* (2017) 46:625–31. doi: 10.1016/J.SEMARTHRIT.2016.10.003

143. Fraticelli P, Fischetti C, Salaffi F, Carotti M, Mattioli M, Pomponio G, et al. Combination therapy with rituximab and mycophenolate mofetil in systemic sclerosis. a single-centre case series study. *Clin Exp Rheumatol* (2018) 36:S142-5.

144. Sircar G, Goswami RP, Sircar D, Ghosh A, Ghosh P. Intravenous cyclophosphamide vs rituximab for the treatment of early diffuse scleroderma lung disease: open label, randomized, controlled trial. *Rheumatol (Oxford)* (2018) 57:2106–13. doi: 10.1093/RHEUMATOLOGY/KEY213

145. Jordan S, Distler JHW, Maurer B, Huscher D, Van Laar JM, Allanore Y, et al. Effects and safety of rituximab in systemic sclerosis: An analysis from the European scleroderma trial and research (EUSTAR) group. *Ann Rheum Dis* (2015) 74:1188–94. doi: 10.1136/annrheumdis-2013-204522

146. De Lauretis A, Sestini P, Pantelidis P, Hoyles R, Hansell DM, Goh NSL, et al. Serum interleukin 6 is predictive of early functional decline and mortality in interstitial lung disease associated with systemic sclerosis. *J Rheumatol* (2013) 40:435–46. doi: 10.3899/jrheum.120725

147. Ebata S, Yoshizaki A, Fukasawa T, Yoshizaki-Ogawa A, Asano Y, Kashiwabara K, et al. Percentage of residual b cells after 2 weeks of rituximab

treatment predicts the improvement of systemic sclerosis-associated interstitial lung disease. J Dermatol (2022) 49(1):179-83. doi: 10.1111/1346-8138.16206

148. Fraticelli P, De Vita S, Franzolini N, Svegliati S, Scott CA, Tonnini C, et al. Reduced type I collagen gene expression by skin fibroblasts of patients with systemic sclerosis after one treatment course with rituximab. *Clin Exp Rheumatol* (2015) 33:160–7.

149. Elhai M, Boubaya M, Distler O, Smith V, Matucci–Cerinic M, Alegre Sancho JJ, et al. Outcomes of patients with systemic sclerosis treated with rituximab in contemporary practice: A prospective cohort study. *Ann Rheum Dis* (2019) 78:979–87. doi: 10.1136/annrheumdis-2018-214816

150. Goswami RP, Ray A, Chatterjee M, Mukherjee A, Sircar G, Ghosh P. Rituximab in the treatment of systemic sclerosis-related interstitial lung disease: A systematic review and meta-analysis. *Rheumatol (United Kingdom)* (2021) 60:557–67. doi: 10.1093/rheumatology/keaa550

151. Tang R, Yu J, Shi Y, Zou P, Zeng Z, Tang B, et al. Safety and efficacy of rituximab in systemic sclerosis: A systematic review and meta-analysis. *Int Immunopharmacol* (2020) 83. doi: 10.1016/j.intimp.2020.106389

152. Ebata S, Yoshizaki A, Oba K, Kashiwabara K, Ueda K, Uemura Y, et al. Safety and efficacy of rituximab in systemic sclerosis (DESIRES): a double–blind, investigator–initiated, randomised, placebo–controlled trial. *Lancet Rheumatol* (2021) 3:e489–97. doi: 10.1016/S2665–9913(21)00107–7

153. Saunders P, Tsipouri V, Keir GJ, Ashby D, Flather MD, Parfrey H, et al. Rituximab versus cyclophosphamide for the treatment of connective tissue diseaseassociated interstitial lung disease (RECITAL): Study protocol for a randomised controlled trial. *Trials* (2017) 18(1):275. doi: 10.1186/s13063-017-2016-2

154. Bonroy C, Smith V, Deschepper E, De Keyser F DK. Specific antinuclear antibody level changes after b cell depletion therapy in systemic sclerosis are associated with improvement of skin thickening. *J Rheumatol* (2016) 43:681LP – 681. doi: 10.3899/jrheum.150105.C1

155. Lafyatis R, Kissin E, York M, Farina G, Viger K, Fritzler MJ, et al. B cell depletion with rituximab in patients with diffuse cutaneous systemic sclerosis. *Arthritis Rheum* (2009) 60:578-83. doi: 10.1002/art.24249

156. Sumida H, Asano Y, Tamaki Z, Aozasa N, Taniguchi T, Takahashi T, et al. Successful experience of rituximab therapy for systemic sclerosis–associated interstitial lung disease with concomitant systemic lupus erythematosus. *J Dermatol* (2014) 41:418–20. doi: 10.1111/1346–8138.12461

157. Hofmann K, Clauder AK, Manz RA. Targeting b cells and plasma cells in autoimmune diseases. *Front Immunol* (2018) 9:835/BIBTEX. doi: 10.3389/FIMMU.2018.00835/BIBTEX

158. Streicher K, Sridhar S, Kuziora M, Morehouse CA, Higgs BW, Sebastian Y, et al. Baseline plasma cell gene signature predicts improvement in systemic sclerosis skin scores following treatment with inebilizumab (MEDI-551) and correlates with disease activity in systemic lupus erythematosus and chronic obstructive pulmonary disease. *Arthritis Rheumatol (Hoboken NJ)* (2018) 70:2087-95. doi: 10.1002/ART.40656

159. Schiopu E, Chatterjee S, Hsu V, Flor A, Cimbora D, Patra K, et al. Safety and tolerability of an anti–CD19 monoclonal antibody, MEDI–551, in subjects with systemic sclerosis: A phase I, randomized, placebo–controlled, escalating single–dose study. *Arthritis Res Ther* (2016) 18:1–14. doi: 10.1186/S13075-016–1021–2/TABLES/5

160. Benfaremo D, Gabrielli A. Is there a future for anti-CD38 antibody therapy in systemic autoimmune diseases? *Cells* (2020) 9. doi: 10.3390/CELLS9010077

161. Gordon JK, Martyanov V, Franks JM, Bernstein EJ, Szymonifka J, Magro C, et al. Belimumab for the treatment of early diffuse systemic sclerosis: Results of a randomized, double-blind, placebo-controlled, pilot trial. *Arthritis Rheumatol* (2018) 70:308–16. doi: 10.1002/ART.40358/ABSTRACT

162. Khanna D, Lin CJF, Furst DE, Goldin J, Kim G, Kuwana M, et al. Tocilizumab in systemic sclerosis: a randomised, double-blind, placebocontrolled, phase 3 trial. *Lancet Respir Med* (2020) 8:963–74. doi: 10.1016/ S2213-2600(20)30318-0

163. Khanna D, Denton CP, Jahreis A, van Laar JM, Frech TM, Anderson ME, et al. Safety and efficacy of subcutaneous tocilizumab in adults with systemic sclerosis (faSScinate): a phase 2, randomised, controlled trial. *Lancet* (2016) 387:2630–40. doi: 10.1016/S0140-6736(16)00232-4/ATTACHMENT/99F33BB7-93E6-4708-94C8-06E25B85693D/MMC1.PDF

164. Wang J, Wang J, Qi X, Sun Z, Zhang T, Cui Y, et al. The efficacy and safety of pirfenidone combined with immunosuppressant therapy in connective tissue disease-associated interstitial lung disease: A 24-week prospective controlled cohort study. Front Med (Lausanne) (2022) 9:871861. doi: 10.3389/fmed.2022.871861

165. Distler O, Highland KB, Gahlemann M, Azuma A, Fischer A, Mayes MD, et al. Nintedanib for systemic sclerosis–associated interstitial lung disease. *N Engl J Med* (2019) 380:2518–28. doi: 10.1056/nejmoa1903076