Check for updates

OPEN ACCESS

EDITED BY Reinaldo B. Oria, Federal University of Ceara, Brazil

REVIEWED BY Yanhua Zeng, University of South China, China Iwona Rudkowska, Laval University, Canada

*CORRESPONDENCE Omid Asbaghi omid.asbaghi@gmail.com Mohammad Zamani

SPECIALTY SECTION This article was submitted to Nutritional Immunology, a section of the journal Frontiers in Immunology

Md_zamany@yahoo.com

RECEIVED 07 November 2022 ACCEPTED 02 February 2023 PUBLISHED 22 February 2023

CITATION

Rastgoo S, Shimi G, Shiraseb F, Karbasi A, Ashtary-Larky D, Yousefi M, Golalipour E, Asbaghi O and Zamani M (2023) The effects of conjugated linoleic acid supplementation on inflammatory cytokines and adipokines in adults: A GRADE-assessed systematic review and dose-response meta-analysis. *Front. Immunol.* 14:1092077. doi: 10.3389/fimmu.2023.1092077

COPYRIGHT

© 2023 Rastgoo, Shimi, Shiraseb, Karbasi, Ashtary-Larky, Yousefi, Golalipour, Asbaghi and Zamani. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

The effects of conjugated linoleic acid supplementation on inflammatory cytokines and adipokines in adults: A GRADEassessed systematic review and dose-response meta-analysis

Samira Rastgoo^{1,2}, Ghazaleh Shimi¹, Farideh Shiraseb³, Ashkan Karbasi⁴, Damoon Ashtary-Larky⁵, Mohsen Yousefi⁶, Elnaz Golalipour⁶, Omid Asbaghi^{2,7*} and Mohammad Zamani^{8*}

¹Department of Cellular and Molecular Nutrition, Faculty of Nutrition Science and Food Technology, National Nutrition and Food Technology Research Institute, Shahid Beheshti University of Medical Sciences, Tehran, Iran, ²Research Committee, Shahid Beheshti University of Medical Sciences, Tehran, Iran, ³Department of Community Nutrition, School of Nutritional Sciences and Dietetics, Tehran University of Medical Sciences (TUMS), Tehran, Iran, ⁴Department of Clinical Biochemistry, School of Medicine, Hamadan University of Medical Sciences, Hamadan, Iran, ⁵Nutrition and Metabolic Diseases Research Center, Ahvaz Jundishapur University of Medical Sciences, Ahvaz, Iran, ⁶Faculty of Medicine, Shahid Beheshti University of Medical Sciences, Tehran, Iran, ⁷Cancer Research Center, Shahid Beheshti University of Medical Sciences, Tehran, Iran, ⁸Department of Clinical Nutrition, School of Nutritional Sciences and Dietetics, Tehran University of Medical Sciences, Tehran, Iran

Background and aims: Many studies have investigated the effect of conjugated linoleic acid (CLA) supplementation on inflammatory cytokines and adipokines. However, the results of these studies are not consistent. Therefore, this systematic review and meta-analysis were designed to comprehensively evaluate the effect of CLA supplementation on inflammatory cytokines and adipokines.

Methods: Randomized controlled trials (RCTs) examining the effects of CLA supplementation on C-reactive protein (CRP), interleukin 6 (IL-6), tumor necrosis factor-alpha (TNF- α), adiponectin, and leptin, published up to March 2022, were identified through PubMed, SCOPUS, and ISI Web of Science databases. A random-effects model was used to calculate weighted mean differences (WMDs) with 95% confidence intervals (CI) for 42 studies that included 1,109 participants.

Results: Findings from 42 studies with 58 arms indicated that CLA supplementation significantly decreased IL-6 and TNF- α levels and also slightly increased CRP levels. However, adiponectin and leptin levels did not change after CLA supplementation. A subgroup analysis found that CLA supplementation reduced adiponectin and leptin in women.

Conclusion: Our results demonstrated that CLA supplementation increased CRP levels and decreased TNF- α and IL-6 levels. Therefore, it seems that CLA can have both proinflammatory and anti-inflammatory roles.

Systematic review registration: https://www.crd.york.ac.uk/prospero/, identifier (CRD42022331110).

KEYWORDS

inflammation, cytokines, adipokines, conjugated linoleic acid, meta-analysis

Introduction

Inflammation is a protective reaction by an organism in response to injury, irritation, or infection that eliminates harmful stimuli and initiates the healing process (1). However, uncontrolled or unresolved inflammation can lead to tissue damage as well as the development of chronic inflammatory diseases, including type 2 diabetes, cardiovascular disease, cancer, etc. (2, 3). Therefore, the control and prevention of chronic inflammation should be considered in order to prevent some chronic diseases and even improve health.

Weight loss strategies such as dietary interventions and physical activity are commonly used as nonpharmacological interventions to combat and/or prevent proinflammatory conditions (4, 5). Despite the considerable benefits of pharmacological therapies, these may exert undesirable side effects and may not be tolerated by some individuals. On the other hand, they may cause drug resistance (reduction of efficiency) and even toxicity (6). Several studies have suggested that dietary strategies, such as certain supplements, can modulate inflammation. For example, a clinical trial has reported that magnesium and vitamin E co-supplementation led to a significant decrease in C-reactive protein (CRP) and a significant increase in total antioxidant capacity levels (7-9). Another study has suggested that Lglutamine supplementation in the early period of COVID-19 infection may reduce inflammatory responses and boost the immune system (10). Therefore, it is practical to find nutraceuticals and natural compounds with anti-inflammatory effects that may serve as alternative therapies to pharmacological interventions.

Conjugated linoleic acid (CLA) is a collective term for geometric isomers of linoleic acid (C18:2, n-6). This polyunsaturated fatty acid has two double bonds separated by a methylene group. This conjugation of the double bond is generally in positions 9 and 11 or 10 and 12 and may be a cis or trans configuration. Depending on the position and geometry of the double bonds, several isomers of CLA have been identified, such as 9cis,11trans-CLA and 10trans,12cis-CLA (11, 12). Most of the beneficial properties of CLA are elicited by these two main isomers. For example, 10trans,12cis-CLA is involved in catabolic processes such as lipolysis and fat oxidation, whereas 9cis,11trans-CLA seems to be the active anabolic agent. In addition, based on the evidence, 10trans,12cis-CLA is thought to be anticarcinogenic, antiobesity, and antidiabetic, whereas 9cis,11trans-CLA is mainly anti-inflammatory (13). CLA is formed via biohydrogenation by bacterial enzymes known as catalyst, present in the intestine microbiota of ruminant mammals; hence, CLA is mainly found in their flesh and milk (14). Nonetheless, it is present in foods only in finite amounts; therefore, commercialized CLA supplements have been provided to offer potential benefits, such as reduction of body weight and total fat mass, anticancer effects including reducing tumor growth (15, 16), reducing insulin resistance, lipid disorders, and oxidative stress, improving liver function in nonalcoholic fatty liver (17), and immunomodulatory effects (18–20).

A large number of studies have investigated the effect of CLA supplementation on inflammatory cytokines and adipokines. However, the results of these studies are contradictory. For instance, some studies have reported that CLA supplementation had no significant effect on leptin levels (21, 22), whereas others found a significant decrease in serum leptin during CLA supplementation (23, 24). CLA supplementation has even been shown to significantly increase serum leptin levels (25). Also, some studies have reported that CLA supplements lead to a decrease in adiponectin levels (26), while another study has reported that CLA supplements have little or no effect on adiponectin levels (27). However, Sneddon et al. (28) have reported that CLA supplementation was associated with a significant increase in serum adiponectin levels (12%) in young obese subjects. Furthermore, while one study has found that CLA supplementation may increase CRP and tumor necrosis factor-alpha (TNF- α) and marginally decrease interleukin-6 (IL-6), indicating a proinflammatory state (29, 30), a recent review article suggests that CLA may have an antiinflammatory effect by reducing inflammatory mediators such as cytokines, particularly IL-6, TNF- α , IFN- γ , and IL-1 β (30). Therefore, due to the inconsistencies in previous studies, the present study sought to update previous meta-analyses in light of the plethora of new studies. Thus, the current meta-analysis sought to investigate the effects of CLA on inflammatory cytokines and adipokines in adults.

Method and materials

This systematic review has been conducted according to the PRISMA statement (31). The present study was registered at PROSPERO CRD42022331110.

We conducted a comprehensive literature search in scientific databases such as PubMed/Medline, Scopus, Web of Science, EMBASE, the Cochrane databases, and Google Scholar to find relevant prospective studies on the effects of CLA supplementation on inflammatory cytokines, adiponectin, and leptin published up to March 2022. There were no restrictions on the length of time or language of publications. The Participant, Intervention, Comparison/Control, Outcome (PICOS) search framework was used (32), which stands for population (both healthy and unhealthy adults), intervention (CLA supplementation), comparison (placebo/control group), outcome (changes in CRP, IL-6, TNF, adiponectin, and leptin), and research design (parallel and crossover randomized clinical trials).

The combination of MESH and non-MESH terms were used for the search, as follows: ("conjugated linoleic acid" OR "conjugated fatty acid" OR "boric acid" OR "rumenic acid" OR "CLA") AND (intervention OR "intervention study" OR "intervention studies" OR "controlled trial" OR randomized OR random OR randomly OR placebo OR "randomized controlled trial" OR "randomized clinical trial" OR RCT OR blinded OR "double blind" OR "double blinded" OR trial OR "clinical trial" OR trials OR "Pragmatic Clinical Trial" OR parallel OR "parallel study" OR "parallel trial").

Study selection

By reading the titles, abstracts, and full text of the papers as needed, two researchers independently (FS and OA) chose the appropriate articles. The effects of CLA supplementation on inflammatory cytokines and adipokines variables in adults with different health statuses in all human randomized clinical trials) RCTs(.

We incorporated studies that satisfied the following criteria: (1) RCTs (parallel or crossover); (2) used oral intake of CLA; (3) examined the effects of CLA supplementation on CRP, IL-6, TNF- α , adiponectin, and leptin; (4) had an intervention duration of at least 2 weeks (RCTs with two or more eligible arms were considered as separate studies); (5) were performed on adults (\geq 18 years old); (6) provided means and standard deviations (SDs) for all outcome variables, or any other effect sizes from which the calculation of mean and SD was possible. The searches were limited to human studies with no language restrictions. Animal studies, reviews, *in vitro* research, research on kids and teenagers, grey literature, conference abstracts, opinion pieces, books, and RCTs without a placebo or control group were excluded. Studies that used CLA in combination with vitamins or minerals were also excluded.

In the present study, we searched for studies that assessed the effects of CLA supplementation on all inflammatory cytokines. After screening and finding eligible studies, we found that most studies evaluated CRP, IL-6, and TNF. In addition, a limited number of studies have evaluated other inflammatory factors such as IL-1 and IL-8. Therefore, we included studies that evaluated the effects of CLA on CRP, IL-6, and TNF.

Data extraction

After quickly skimming the titles, abstracts, and full text to choose the most pertinent research following a separate review of each qualifying RCT, the following data were gleaned by two independent researchers (OA and FS). Name of the first author, country of origin, year of publication, type of clinical trial, participant characteristics (mean age, body mass index (BMI), sex), randomization, blinding, sample size, number of participants in the intervention and control groups, type and dosage of supplemented CLA, duration of the study, and related data were extracted for additional measurements. The CLA dosages were converted to milligrams per day, whether they were given in grams per day or another unit.

Quality assessment

To rate the quality of the studies, the Cochrane Collaboration technique was utilized (33). Two researchers (SR and GS) independently assessed the methods, and any disagreements between their assessments were resolved through discussion. Each study was evaluated for any bias, including those caused by randomized sequence generation, allocation concealment, participant and staff blindness, outcome assessor blindness, insufficient outcome data, selective reporting, and other biases. Thus, three categories were established: high risk of bias (general risk of bias > 2 high risks), low risk of bias (general risk of bias = 2 high risk).

Statistical analysis

Stata 11.0 was used to conduct the statistical analysis (Stata Corp, College Station, TX, USA). p-values <0.05 were deemed statistically significant for all two-tailed tests. The pooled weighted mean difference (WMD) was calculated using a random-effects model to take into account any existing heterogeneity (due to the different intervention doses, duration, participant health, sample sizes, and length of intervention) developed by Der Simonian and Laird (34). We computed the mean differences in CRP, IL-6, TNF- α , adiponectin, and leptin between the CLA supplementation and control groups from the preintervention to the postintervention. Formula: SD = square root [(SD at baseline)² + (SD at end ofstudy)² (2rSD at baseline, SD at end of study)] was used to get the SD of the mean difference (35). For RTCs that only provided standard error of the mean (SEM) information, SD was determined using the following formula: Standard Error of Mean (SEM): SD =, where *n* is the number of subjects in each group (36). After visual inspection of forest plots or Cochrane's Q test, heterogeneity was evaluated using the I square (I^2) statistic $(p < 0.05 \text{ and } I^2 > 40\%)$, with $I^2 > 40\%$ considered clinically significant (37). The baseline CRP, IL-6, TNF- α , adiponectin, and leptin levels were analyzed, while the intervention duration (< 12 weeks, \geq 12 weeks) and dosage of CLA (\geq 3 and < 3 g/ day) were based on the median values of the included studies and health status (healthy: people without any disease; unhealthy: people suffering from a disease such as atherosclerosis, rheumatoid arthritis, diabetes, etc.). Other subgroup analyses were performed according to gender (man, woman), baseline BMI (normal (18.5–24.9 kg/m²), overweight (25–29.9 kg/m²), and obese (>30 kg/m²)) and isomer's type (CLA-c9t11, CLA-t10c12, both). Studies examining the effect of CLA supplementation on inflammatory cytokines and adipokines used the Begg's test and the funnel plot test to evaluate publication bias (38). We used the leave-one-out approach to do a sensitivity analysis to identify how many inferences were dependent on a single sample to examine the influence of each study on the pooled effect size (39). The possible linear association between CLA (mg/day) dose and duration on outcome variables was evaluated using meta-regression, whereas a nonlinear dose–response analysis was used to assess the effect of CLA supplementation dose and duration (40, 41).

Result

The flow of study selection

The flow chart presented in Figure 1 describes the selection process and the references retrieved from the database.

A total of 8,351 studies were identified in the first step of the literature search of electronic databases, PubMed (n = 3,582), ISI Web of Science (n = 1,991), Scopus (n = 2,310), and the Cochrane library (n = 468). After excluding duplicated (n = 2,419) and irrelevant studies based on titles and abstracts (n = 2,804), 104 potentially relevant

articles were considered for full-text review. Out of 104 studies, 62 did not have the desired data. Finally, 42 studies (11, 12, 16, 18, 19, 27, 28, 42–76) were included in the present meta-analysis, and their characteristics are illustrated in Table 1. The risk of bias assessment is summarized in Table 2.

Study characteristics

All of these studies were RCTs published between 2000 and 2020. Study design characteristics are presented in Table 1. The WMD and 95% CI of CRP (mg/dl), IL-6 (pg/ml), TNF-α (ng/ml), adiponectin (μ g/ml), and leptin (ng/ml) and their changes are presented in Figures 2A-E, respectively. The two studies were conducted in the USA (42, 67), three in Sweden (43, 45, 49), three in the Netherlands (44, 52, 65), four in Canada (12, 14, 46, 68), two in Norway (27, 47), four in the UK (28, 48, 51, 56), two in France (50, 53), two in Turkey (11, 72), one in Japan (55), one in Ireland (57), one in Korean (58), one in Finland (59), one in Denmark (61), one in China (62), three in Germany (62, 69, 70), one in Italy (64), one in Spain (16), one in Poland (75), and the others in Iran (18, 19, 60, 66, 71, 73, 74, 76). Thirteen studies included only men and seven women, and 22 included both sexes. There were 37 parallel (11, 12, 16, 18, 19, 27, 42-45, 47-63, 65-67, 69, 71-76) and five crossover (28, 46, 64, 68, 70) studies. A total of 1,192 subjects (intervention: 597/control: 595) in terms of CRP, 817 subjects (intervention: 418/control: 399) for IL-6, 779 subjects (intervention: 402/control: 377) for TNF-α, 845 subjects



TABLE 1 Characteristics of the included studies.

Studies	Country	Study design	Participant	Sex		nple ze	Trial duration	Mean	s age	Mear	is BMI		Intervention	
					IG	CG	(week)	IG	CG	IG	CG	CLA dose	lsomer's type	Control group Whigham
Medina et al. (42)	USA	Parallel, R, PC, B	Healthy	F: 17	10	7	9	20-41	20-41	23.2 ± 1.5	22.2 ± 3.9	3	22.6% 10trans,12cis; 23.6% 11cis,13trans; 17.6% 9cis,11trans; 16.6% 8trans,10cis; 7.7% 9trans,11trans and 10trans,12trans; and 11.9% other isomers	Placebo
Riserus et al. (43)	Sweden	Parallel, R, PC, DB	Obese men with the metabolic syndrome	M: 38	19	19	12	51 ± 7.1	53 ± 10.1	30.1 ± 1.8	30.2 ± 1.8	3.4	35.9% t10c12 CLA, 35.4% c9t11 CLA	Placebo
Riserus et al. (43)	Sweden	Parallel, R, PC, DB	Obese men with the metabolic syndrome	M: 38	19	19	12	55 ± 7.1	53 ± 10.1	31.2 ± 2.5	30.2 ± 1.8	3.4	CLA-t10c12	Placebo
Albers et al. (44)	Netherlands	Parallel, R, PC, DB	Healthy men	M: 47	25	22	12	52 ± 9	53 ± 10	25.3 ± 2.8	25.3 ± 2.3	1.7	50% c9,t11 CLA and 50% t10,c12	Control diet
Albers et al. (44)	Netherlands	Parallel, R, PC, DB	Healthy men	M: 46	24	22	12	52 ± 8	53 ± 10	25.7 ± 2.5	25.3 ± 2.3	1.6	80% c9,t11 CLA and 20% t10,c12	Control diet
Riserus et al. (45)	Sweden	Parallel, R, PC, DB	Obese men	M: 38	19	19	12	51 ± 7.1	53 ± 10.1	30.1 ± 1.8	30.2 ± 1.8	3.4	Both	Control diet
Riserus et al. (45)	Sweden	Parallel, R, PC, DB	Obese men	M: 38	19	19	12	55 ± 7.1	53 ± 10.1	31.2 ± 2.5	30.2 ± 1.8	3.4	CLA-t10c12	Control diet
Ramakers et al. (50)	France	Parallel, R, PC, DB	Overweight subjects	M/F (F: 13, M: 13)	14	12	13	53 ± 7	58 ± 5	29 ± 2	29 ± 2	3	CLA-c9t11	Control diet
Ramakers et al. (50)	France	Parallel, R, PC, DB	Overweight subjects	M/F (F: 12, M: 12)	12	12	13	56 ± 6	58 ± 5	29 ± 3	29 ± 2	3	CLA-t10c12	Control diet
Desroches et al. (46)	Canada	Crossover, R, PC, B	Overweight and obese	M: 17	17	17	4	36.6 ± 12.4	36.6 ± 12.4	31.2 ± 4.4	31.2 ± 4.4	4.22	Both	Control diet
Song et al. (48)	UK	Parallel, R, PC, DB	Young healthy volunteers	M/F (F: 20, M: 8)	14	14	12	31.8 ± 6.88	30.9 ± 7.14	24.3 ± 3.8	24.23 ± 3.69	3	50% c9,t11 CLA and 50% t10,c12	Control diet
Gaullier et al. (47)	Norway	Parallel, R, PC, DB	Healthy overweight humans	M/F (F: 74, M: 14)	47	41	104	48.6 ± 10.6	45.1 ± 8.8	28.3 ± 1.5	27.4 ± 1.7	3.4	50% c9,t11 CLA and 50% t10,c12	Placebo
Gaullier et al. (47)	Norway	Parallel, R, PC, DB	Healthy overweight humans	M/F (F: 69, M: 18)	46	41	104	45.1 ± 10.5	45.1 ± 8.8	28.1 ± 1.4	27.4 ± 1.7	3.4	50% c9,t11 CLA and 50% t10,c12	Placebo

(Continued)

10.3389/fimmu.2023.1092077

TABLE 1 Continued

Studies	Country	Study design	Participant	Sex		nple ze	Trial duration	Means	age	Mean	s BMI		Intervention		
					IG	CG	(week)	IG	CG	IG	CG	CLA dose	lsomer's type	Control group Whighar	
Smedman et al. (49)	Sweden	Parallel, R, PC, DB	Healthy	M/F (F: 26, M: 27)	28	25	12	43.312.8	47.8 ± 10.1	25.8 ± 3.9	24.4 ± 4.2	4.2	50% c9,t11 CLA and 50% t10,c12	Control diet	
Naumann et al. (52)	Netherlands	Parallel, R, PC, DB	Overweight subjects with LDL phenotype B	M/F: 68	34	34	13	51 ± 7	51 ± 9	28.6 ± 2.3	28 ± 2.2	3	CLA-c9t11	Control diet	
Naumann et al. (52)	Netherlands	Parallel, R, PC, DB	Overweight subjects with LDL phenotype B	M/F: 53	19	34	13	55 ± 7	51 ± 9	29.3 ± 2.4	28 ± 2.2	3	CLA-t10c12	Control diet	
Colakoglu et al. (11)	Turkey	Parallel, R, PC, SB	Healthy	F: 26	12	14	6	21.7 ± 2	20.4 ± 2.5	22.5 ± 1.7	21.6 ± 1.6	3.6	Both	Control diet-exercis	
Colakoglu et al. (11)	Turkey	Parallel, R, PC, SB	Healthy	F: 18	11	7	6	20.4 ± 1.7	21.9 ± 2	23.3 ± 1.2	20.8 ± 1.6	3.6	Both	Control diet	
Taylor et al. (51)	UK	Parallel, R, PC, DB	Healthy	M/F: 40	21	19	12	45 ± 6	47 ± 8	33 ± 3	33 ± 3	4.5	35% 9c,11t CLA, 36% t10,c12 CLA	Control diet	
Mullen et al. (57)	Ireland	Parallel, R, PC, DB	Healthy middle- aged	M: 30	15	15	8	51 ± 8.04	47.8 ± 6.36	26.2 ± 2.32	26 ± 3.4	2.2	50% c9,t11 CLA and 50% t10,c12	Placebo	
Nazare et al. (53)	France	Parallel, R, PC, DB	Healthy subjects	M/F: 44	21	23	14	29.4 ± 6.75	28.5 ± 5.7	25.2 ± 1.45	25.1 ± 1.48	3.76	50% c9,t11 CLA and 50% t10,c12	Placebo	
Iwata et al. (55)	Japan	Parallel, R, PC, DB	Overweight	M: 40	20	20	12	44.3 ± 10.2	42.5 ± 10.4	27.4 ± 2	27.8 ± 1.9	5.4	50% c9,t11 CLA and 50% t10,c12	Placebo	
Iwata et al. (55)	Japan	Parallel, R, PC, DB	Overweight	M: 40	20	20	12	40.5 ± 8.8	42.5 ± 10.4	28.1 ± 2.1	27.8 ± 1.9	10.8	50% c9,t11 CLA and 50% t10,c12	Placebo	
Steck et al. (56)	UK	Parallel, R, PC, DB	Healthy obese humans	M/F (F: 23, M: 9)	16	16	12	36.3 ± 8.9	34.9 ± 8	32.7 ± 1.8	32.7 ± 1.9	3.2	50% c9,t11 CLA and 50% t10,c12	Placebo	
Steck et al. (56)	UK	Parallel, R, PC, DB	Healthy obese humans	M/F (F: 24, M: 8)	16	16	12	34.1 ± 8.9	34.9 ± 8	32.7 ± 1.7	32.7 ± 1.9	6.4	50% c9,t11 CLA and 50% t10,c12	Placebo	

TABLE 1 Continued

Studies	Country	Study design	Participant	Sex		nple ze	Trial duration	Means	age	Mean	is BMI		Intervention	
					IG	CG	(week)	IG	CG	IG	CG	CLA dose	lsomer's type	Control group Whighar
Whigham et al. (54)	Canada	Parallel, R, PC, DB	Healthy	M/F (F: 32, M: 8)	22	18	24	34 ± 8	32 ± 7	27.6 ± 1.8	28 ± 2.2	3.2	39.2% 9cis,11 trans and 38.5% 10trans,12cis	Placebo
Gaullier et al. (27)	Norway	Parallel, R, PC, DB	Overweight and obese	M/F (F: 84, M: 21)	55	50	24	45.8 ± 10	48.7 ± 9.2	30.5 ± 10.4	30.2 ± 10.4	3.4	50% c9,t11 CLA and 50% t10,c12	Placebo
Raff et al. (61)	Denmark	Parallel, R, PC, DB	Healthy young men	M: 38	18	20	5	25.7 ± 4.2	26.1 ± 3.6	22 ± 1.9	22.5 ± 2.1	5.5	39.4% 9cis,11trans and 38.5% 10trans,12cis	Control diet
Aryaeian et al. (60)	Iran	Parallel, R, PC, DB	Rheumatoid Arthritis	M/F (F: 38, M: 6)	22	22	12	46.23 ± 13.07	47.95 ± 11.14	27.18 ± 0.99	28.48 ± 0.84	2.5	50% c9,t11 CLA and 50% t10,c12	Placebo
Kim et al. (58)	Korea	Parallel, R, PC, DB	Healthy overweight women	F: 27	15	12	12	26.33 ± 9.4	29.5 ± 10.8	25.23 ± 2.16	26.47 ± 1.8	3	50% c9,t11 CLA and 50% t10,c12	Control diet
Sneddon et al. (28)	UK	Crossover, R, PC, DB	Young lean	M: 13	13	13	12	30.5 ± 4.9	30.5 ± 4.9	23.6 ± 1.5	23.6 ± 1.5	3	50% c9,t11 CLA and 50% t10,c12	Placebo
Sneddon et al. (28)	UK	Crossover, R, PC, DB	Young obese	M: 12	12	12	12	32.4 ± 2.3	32.4 ± 2.4	32.3 ± 1.9	32.3 ± 1.9	3	50% c9,t11 CLA and 50% t10,c12	Placebo
Sneddon et al. (28)	UK	Crossover, R, PC, DB	Older lean	M: 20	20	20	12	56.3 ± 4.2	56.3 ± 4.2	23.6 ± 1.5	23.6 ± 1.5	3	50% c9,t11 CLA and 50% t10,c12	Placebo
Sneddon et al. (28)	UK	Crossover, R, PC, DB	Older obese	M: 14	14	14	12	56.9 ± 5.4	56.4 ± 5.4	32 ± 1.6	32 ± 1.6	3	50% c9,t11 CLA and 50% t10,c12	Placebo
Turpeinen et al. (59)	Finland	Parallel, R, PC	Subjects with birch pollen allergy	M/F (F: 28, M: 12)	20	20	12	20-46	20-46	NR	NR	2	65·3% 9cis,11trans-CLA and 8·5% 10trans,12cis-CLA	Placebo
Norris et al. (63)	Germany	Parallel, R, PC, DB	Postmenopausal women with type 2 diabetes mellitus	F: 55	22	33	16	59.4 ± 7.3	60.1 ± 7.3	37.1 ± 7.2	36.3 ± 6.1	6.4	41.6% c9,t11 CLA and 40.4% t10,c12	Control diet
Zhao et al. (62)	China	Parallel, R, PC, DB	Obesity-related hypertension	M/F (F: 36, M: 44)	40	40	8	62.3 ± 3.5	59.4 ± 2.4	32.3 ± 2.3	31.2 ± 1.4	4.5	50% c9,t11 CLA and 50% t10,c12	Control diet
Sluijs et al. (65)	Netherlands	Parallel, R, PC, DB	Overweight and obese adults	M/F (F: 179, M: 167)	173	173	24	58 ± 0.4	58.8 ± 0.5	28 ± 9.45	27.7 ± 12.75	4	CLA-c9t11	Placebo

10.3389/fimmu.2023.1092077

TABLE 1 Continued

Studies	Country	Study design	Participant	Sex		nple ze	Trial duration	Mean	s age	Mean	s BMI		Intervention	
					IG	CG	(week)	IG	CG	IG	CG	CLA dose	lsomer's type	Control group Whighan
Tavakkoli Darestani et al. (66)	Iran	Parallel, R, PC, DB	Postmenopausal	F: 76	38	38	12	55.1 ± 6.4	54.9 ± 6.9	27.6 ± 3.4	27 ± 3.4	3.2	50% c9,t11 CLA and 50% t10,c12	Placebo
MacRedmond et al. (12)	Canada	Parallel, R, PC, DB	Overweight mild asthmatics	M/F (F: 15, M: 13)	15	13	12	32.2 ± 8.7	29.9 ± 3.8	27.8 ± 4.5	27.3 ± 3.6	4.5	50% c9,t11 CLA and 50% t10,c12	Placebo
Sofi et al. (64)	Italy	Crossover, R, PC	Healthy middle- aged	M/F (F: 6, M: 4)	10	10	8	45.6	45.6	25 ± 4	25 ± 4	3	Both	Control diet
Joseph et al. (68)	Canada	Crossover, R, PC, DB	Overweight, hyperlipidemic	M: 27	27	27	8	18-60	18-60	31.5 ± 4	31.3 ± 4	3.5	50% c9,t11 CLA and 50% t10,c12	Placebo
Joseph et al. (68)	Canada	Crossover, R, PC, DB	Overweight, hyperlipidemic	M: 27	27	27	8	18-60	18-60	31.4 ± 4	31.3 ± 4	3.5	50% c9,t11 CLA and 50% t10,c12	Placebo
Pfeuffer et al. (69)	Germany	Parallel, R, PC, DB	Obese male subjects	M: 40	21	19	4	45-68	45-68	28.3 ± 2.3	27.8 ± 1.3	4.5	50% c9,t11 CLA and 50% t10,c12	Control diet
Brown et al. (67)	USA	Paralrell, R, PC	Health in young women	F: 18	9	9	8	20-40	20-40	19–30	19–30	1.17	Both	Control diet
Rubin et al. (70)	Germany	Crossover, R, PC, DB	Middle-aged men	M: 35	35	35	4	45-68	45-68	26 ± 2.6	26.1 ± 3	4.25	CLA-c9t11	Control diet
Rubin et al. (70)	Germany	Crossover, R, PC, DB	Middle-aged men	M: 35	35	35	4	45-68	45-68	26 ± 3.5	26.1 ± 3	4.25	CLA-t10c12	Control diet
Eftekhari et al. (73)	Iran	Parallel, R, PC	Atherosclerotic patients	M/F: 57	29	28	8	52.79 ± 14.11	55.85 ± 14.13	24.02 ± 2.76	24.66 ± 2.34	3	50% c9,t11 CLA and 50% t10,c12	Control diet
Shadman et al. (71)	Iran	Parallel, R, PC, DB	Overweight type2 diabetics	M/F (F: 21, M: 18)	19	20	8	45.1 ± 5.7	45.5 ± 4.3	27.4 ± 0.5	27.1 ± 1.8	3	50% c9,t11 CLA and 50% t10,c12	Placebo
Bulut et al. (72)	Turkey	Parallel, R, PC, DB	Young men	M: 18	9	9	4	19–31	19–31	27.5 ± 2.6	26.8 ± 1.9	3	50% c9,t11 CLA and 50% t10,c12	Placebo
Lopez-Plaza et al. (16)	Spain	Parallel, R, PC, DB	Healthy overweight people	M/F (F: 29, M: 9)	22	16	24	43 ± 8.3	44.35 ± 7.79	28.44 ± 1.08	28.56 ± 0.95	3	50% c9,t11 CLA and 50% t10,c12	Placebo

	Control group Whigham	Placebo	Placebo	Placebo	Control diet	Placebo
Intervention	lsomer's type	50% c9,t11 CLA and 50% t10,c12	50% c9,t11 CLA and 50% t10,c12	Both	Both	Both
	CLA dose	2.5	ς,	5.6	ę	3.2
s BMI	90	28.48 ± 3.94	35.36 ± 3.56	23.83 ± 2.18	35.27 ± 3.46	45.71 ± 26.11
Means BMI	ט	27.18 ± 4.63	34 ± 3.5	$\begin{array}{c} 23.13\\ \pm\\ 0.89\end{array}$	32.72 ± 4.63	25.46 ± 3.85
age	ŋ	47.95 ± 11.14	54 ± 4	18.2 ± 0.5	38.58 ± 8.24	61.55 \pm 10.81
Means age	ט	46.23 ± 13.07	54 ± 4	18.46 ± 1	36.74 ± 6.87	63.82 ± 10.58
Trial duration	(week)	12	12	7	œ	9
Sample size	SC	22	37	10	19	42
Sar	Ð	22	37	13	19	40
Sex		M/F (F: 38, M: 6)	F: 74	M: 23	M/F (F: 32, M: 6)	M/F: 82
Participant		Rheumatoid arthritis	Overweight and obese	Athletic	Nonalcoholic fatty liver disease	Chronic obstructive pulmonary disease (COPD)
Study design		Parallel, R, PC, DB	Parallel, R, PC, DB	Parallel, R, PC, DB	Parallel, R, PC, SB	Parallel, R, PC, DB
Country		Iran	Poland	Iran	Iran	Iran
Studies		Aryaeian et al. (19)	Dus- Zuchowska et al. (75)	Baghi et al. (74)	Abedi et al. (76)	Aslani et al. (18)

(intervention: 424/control: 421) for adiponectin, and 1,274 subjects (intervention: 649/control: 625) for leptin were enrolled. The duration of the intervention varied from 4 to 104 weeks. The CLA supplements were used in doses of 1.6 to 10.8 mg/day. The mean age and baseline BMI ranged from 20 to 61.55 years and 22.2 to 45.71 kg/ m², respectively. Participant's health conditions were as follows: obese men with metabolic syndrome (43), overweight subjects with LDL phenotype B (52), both older obese and young lean, older lean, young obese (28), birch pollen allergy (59), obesity-related hypertension (62), postmenopausal women with type 2 diabetes mellitus (63), overweight mild asthmatics (12), postmenopausal women (66), overweight and hyperlipidemic (68), overweight type 2 diabetes (71), healthy overweight people (16, 47, 56), healthy subjects (11, 42, 44, 48, 49, 51, 53, 54, 57, 58, 61, 64, 67, 70, 72), atherosclerotic patients (73), rheumatoid arthritis (19, 60), athletic (74), overweight/obese (27, 45, 46, 50, 55, 65, 69, 75), nonalcoholic fatty liver disease (76), and chronic obstructive pulmonary disease (COPD) (18). Iwata et al. (55), Albers et al. (44), and Steck et al. used two different CLA doses in their studies. Out of the 42 studies, CRP, IL-6, TNF- α , adiponectin, and leptin were reported in 20, 15, 16, 12, and 20, respectively. For supplementation, 47 study arms used mixed isomers (9cis,11trans-CLA and 10trans,12cis-CLA), four study arms used 9cis,11trans-CLA isomer, and five study arms used 10trans,12cis-CLA isomer. In these studies, detection methods of CRP were Behring latex-enhanced high-sensitivity assays, Eckman Synchron CX7 System, enzyme-linked immunosorbent assay, enhanced turbidimetric immunoassay, highly sensitive immunoassay with a monoclonal antibody coated with polystyrene particles, immunoturbidimetric assay, and rabbit antihuman.

Meta-analysis findings

Effect of CLA supplementation on CRP

Overall, 24 effect sizes from 20 studies for CRP were included in the analysis. The results of the analysis showed that CLA supplementation significantly increased CRP compared to placebo [(WMD = 0.26 mg/dl; 95% CI, 0.00 to 0.52; p = 0.048 ($I^2 = 56.3\%$, <0.001)] (Figure 2A). The effect of CLA on CRP was significant in studies with baseline CRP less than 3 mg/dl (WMD = 0.31 mg/dl; 95% CI, 0.05 to 0.57; p = 0.016), supplementation dose ≥ 3 (g/day) (WMD = 0.49 mg/dl; 95% CI, 0.09 to 0.90; p = 0.017), and healthy subgroup (WMD = 0.39 mg/dl; 95% CI, 0.10 to 0.68; p = 0.008). Heterogeneity disappeared in supplementation subgroups of less than 3 (g/day) ($I^2 = 36.9\%$, p = 0.104), overweight (25–29.9) ($I^2 =$ 15.9%, p = 0.288), and unhealthy subjects ($I^2 = 52.4\%$, p = 0.078). In the other subgroups, the effect of CLA supplementation on serum concentrations of CRP was not significant (Table 3).

Effect of CLA supplementation on IL-6

Pooling effect sizes from 15 studies showed that CLA supplementation significantly decreased IL-6 compared to placebo [(WMD = -0.66 mg/dl; 95% CI, -1.14 to -0.19; p = 0.006 ($I^2 = 86.3\%$; p < 0.001)] (Figure 2B). The effect of CLA on IL-6 was significant in trials lasting <12 weeks (WMD = -1.16 pg/ml; 95% CI, -1.82 to -0.50; p = 0.001), male subjects (WMD = -0.31 mg/dl; 95% CI, -0.51 to -0.12; p = 0.002), unhealthy subgroup (WMD = -0.61 mg/dl; 95% CI, -1.09 to -0.14; p = 0.010), and when a mixture of two

TABLE 1 Continued

TABLE 2 Risk of bias assessment.

Studies	Random sequence generation	Allocation concealment	Selective reporting	Other sources of bias	Blinding (participants and personnel)	Blinding (outcome assessment)	Incomplete outcome data	General risk of bias
Medina et al. (42)	L	Н	Н	Н	U	Н	L	Bad
RISerus et al. (43)	L	Н	Н	L	L	U	Н	Bad
Albers et al. (44)	L	L	Н	L	L	U	L	Good
Riserus et al. (45)	L	Н	Н	Н	L	U	Н	Bad
Desroches et al. (46)	L	Н	Н	Н	L	U	L	Bad
Gaullier et al. (47)	L	Н	L	L	L	U	L	Good
Song et al. (48)	L	Н	L	L	L	U	L	Good
Smedman et al. (49)	L	Н	Н	L	Н	Н	L	Bad
Ramakers et al. (50)	L	L	Н	L	L	L	L	Good
Colakoglu et al. (11)	L	Н	Н	Н	L	U	Н	Bad
Taylor et al. (51)	L	Н	Н	L	L	U	L	Fair
Naumann et al. (<mark>52</mark>)	L	Н	Н	Н	Н	Н	Н	Bad
Gaullier et al. (27)	L	L	L	Н	L	U	Н	Fair
Nazare et al. (53)	L	Н	L	L	L	U	L	Good
Iwata et al. (55)	L	Н	Н	Н	L	U	L	Bad
Watras et al. (54)	L	Н	Н	Н	L	U	L	Bad
Mullen et al. (57)	L	Н	Н	Н	L	U	L	Bad
Steck et al. (56)	L	Н	Н	L	L	U	L	Fair
Kim et al. (58)	L	Н	Н	Н	Н	Н	L	Bad
Sneddon et al. (28)	L	Н	Н	L	L	U	L	Fair
Turpeinen et al. (59)	L	Н	Н	Н	L	U	Н	Bad
Aryaeian et al. (60)	L	Н	Н	Н	L	U	Н	Bad
Raff et al. (61)	L	Н	Н	Н	L	U	Н	Bad
Zhao et al. (62)	L	Н	Н	Н	L	U	L	Bad
Norris et al. (63)	L	L	Н	L	L	U	L	Good

TABLE 2 Continued

Studies	Random sequence generation	Allocation concealment	Selective reporting	Other sources of bias	Blinding (participants and personnel)	Blinding (outcome assessment)	Incomplete outcome data	General risk of bias
MacRedmond et al. (12)	L	Н	Н	Н	Н	Н	L	Bad
Tavakkoli Darestani et al. (66)	L	L	Н	Н	L	U	L	Fair
Sofi et al. (64)	L	L	Н	L	L	U	L	Good
Sluijs et al. (65)	L	L	Н	L	L	U	L	Good
Joseph et al. (<mark>68</mark>)	L	Н	L	Н	L	U	Н	Bad
Brown et al. (67)	L	Н	Н	Н	L	U	Н	Bad
Pfeuffer et al. (69)	L	Н	Н	L	Н	Н	Н	Bad
Rubin et al. (70)	L	L	Н	Н	L	U	L	Fair
Shadman et al. (71)	L	Н	L	L	L	U	L	Good
Lopez-Plaza et al. (16)	L	L	Н	Н	L	U	L	Fair
Bulut et al. (72)	L	L	Н	L	L	U	L	Good
Eftekhari et al. (73)	L	Н	Н	Н	L	U	L	Bad
Aryaeian et al. (<mark>19</mark>)	L	Н	Н	Н	L	U	L	Bad
Baghi et al. (74)	L	Н	L	L	L	U	L	Good
Dus- Zuchowska et al. (75)	L	L	Н	Н	L	U	L	Fair
Abedi et al. (76)	L	L	L	L	Н	Н	L	Fair
Aslani et al. (18)	L	L	Н	Н	L	U	Н	Bad

U, unclear risk of bias; L, low risk of bias; H, high risk of bias.

Good < 2 high risks of bias; Fair = 2 high risks of bias; Bad > 2 high risks of bias.

isomers was used (WMD = -0.68 mg/dl; 95% CI, -1.15 to -0.21; p = 0.005). Heterogeneity disappeared in the trial duration of ≥ 12 week ($I^2 = 11.7\%$; p = 0.335) and the male subgroup ($I^2 = 15.5\%$; p = 0.335). In the other subgroups, the effect of CLA supplementation on serum concentrations of IL-6 was not significant (Table 3).

Effect of CLA supplementation on TNF- α

Results of pooled 19 random-effects size of CLA supplementation on TNF- α found a significant reduction in TNF- α compared to placebo [(WMD = -0.99 mg/dl; 95% CI, -1.76 to -0.22; p = 0.011; ($I^2 = 90.2\%$; p < 0.001)] (Figure 2C). Subgroup analysis showed a more beneficial effect of CLA supplementation in studies with an intervention dose of <3 (g/day) (WMD = −3.08 mg/dl; 95% CI, −4.91 to −1.25; p = 0.001), healthy people (WMD = −2.01 mg/dl; 95% CI, −3.31 to −0.71; p = 0.002), male subjects (WMD = −1.01 mg/ dl; 95% CI, −1.95 to −0.08; p = 0.033), subjects with normal range of BMI (18.5–24.9) (WMD = −9.29 mg/dl; 95% CI, −18.51 to −0.08; p =0.048), and when a mixture of two isomers was used (WMD = −1.41 mg/dl; 95% CI, −2.31 to −0.50; p = 0.002). Interestingly, when the 10trans12cis-CLA isomer was administered, TNF- α was significantly



Forest plot detailing weighted mean difference and 95% confidence intervals (Cls) for the effect of CLA consumption on (A) CRP (mg/L), (B) IL-6 (pg/ml), (C) TNF- α (ng/ml), (D) adiponectin (µg/ml), and (E) leptin (ng/ml). Horizontal lines represent 95% Cls. Diamonds represent pooled estimates from random-effects analysis. WMD, weighted mean difference; Cl, confidence interval; CLA, conjugated linoleic acid; CRP, C-reactive protein; IL-6, interleukin 6; TNF- α , tumor necrosis factor-alpha.

TABLE 3 Subgroup analyses of CLA supplementation on inflammatory cytokines and adipokines.

	Number of studies	WMD (95%CI)	<i>p</i> -value	Heterogeneity	
				P heterogeneity	l ²
Subgroup analyses of CLA su	upplementation on CRP (mg/dl)				
Overall effect	24	0.26 (0.00, 0.52)	0.048	<0.001	56.3%
Baseline CRP (mg/dl)					
<3	15	0.31 (0.05, 0.57)	0.016	0.003	58.0%
≥3	9	0.27 (-0.71, 1.26)	0.583	0.028	53.4%
Trial duration (week)					
≥12	14	0.44 (-0.01, 0.89)	0.056	0.008	54.3%
<12	10	0.15 (-0.18, 0.49)	0.361	0.007	60.0%
Health status					
Healthy	19	0.39 (0.10, 0.68)	0.008	0.001	57.1%
Unhealthy	5	-0.21 (-0.84, 0.41)	0.508	0.078	52.4%
Supplementation dose (g/da	y)				
≥3	13	0.49 (0.09, 0.90)	0.017	<0.001	67.3%
<3	11	0.05 (-0.29, 0.40)	0.759	0.104	36.9%
Sex					
Both sexes	16	0.34 (-0.12, 0.81)	0.149	0.009	51.3%
Male	7	0.27 (-0.10, 0.65)	0.151	0.004	68.6%
Female	1	-0.10 (-0.52, 0.32)	0.644	-	-

TABLE 3 Continued

	Number of studies	WMD (95%CI)	<i>p</i> -value	Heterogeneity	
				P heterogeneity	l ²
Baseline BMI (kg/m ²)					
Normal (18.5–24.9)	3	0.06 (-0.57, 0.71)	0.836	0.001	86.8%
Overweight (25-29.9)	12	0.25 (-0.07, 0.57)	0.131	0.288	15.9%
Obese (>30)	9	0.25 (-0.07, 0.57)	0.111	0.005	63.8%
lsomer's type		1	1		
Both	19	0.25 (-0.05, 0.56)	0.104	<0.001	64.0%
CLA-c9t11	3	0.29 (-0.35, 0.93)	0.373	0.948	0.0%
CLA-t10c12	2	0.45 (-0.08, 0.98)	0.100	0.671	0.0%
Subgroup analyses of CLA su	upplementation on IL-6 (pg/ml).	1	1		
Overall effect	19	-0.66 (-1.14, -0.19)	0.006	<0.001	86.3%
Trial duration (week)	1	1]		
≥12	10	0.32 (-0.19, 0.83)	0.224	0.335	11.7%
<12	9	-1.16 (-1.82, -0.50)	0.001	<0.001	90.4%
Health status	1				
Healthy	14	-0.55 (-1.39, 0.28)	0.192	<0.001	87.7%
Unhealthy	5	-0.61 (-1.09, -0.14)	0.010	0.017	67.0%
Supplementation dose (g/da	ау)				
≥3	9	-0.15 (-0.44, 0.14)	0.310	<0.001	71.4%
<3	10	-1.65 (-3.45, 0.14)	0.072	<0.001	79.0%
Sex					
Both sexes	13	-0.84 (-2.09, 0.40)	0.184	<0.001	89.6%
Male	6	-0.31 (-0.51, -0.12)	0.002	0.335	12.5%
Baseline BMI (kg/m ²)					
Normal (18.5–24.9)	2	-1.66 (-5.15, 1.82)	0.349	0.021	81.1%
Overweight (25-29.9)	10	-0.88 (-2.34, 0.57)	0.236	<0.001	87.4%
Obese (>30)	6	-0.15 (0.49, 0.18)	0.379	0.006	69.5%
lsomer's type	1	1]		
Both	17	-0.68 (-1.15, -0.21)	0.005	<0.001	87.2%
CLA-c9t11	1	-7.84 (-16.90, 1.22)	0.090	-	_
CLA-t10c12	1	5.77 (-0.38, 11.92)	0.066	_	_
Subgroup analyses of CLA su	upplementation on TNF- α (ng/ml))	1		
Overall effect	19	-0.99 (-1.76, -0.22)	0.011	<0.001	90.2%
Trial duration (week)	·	·	I	·	-
≥12	12	-1.21 (-2.44, 0.02)	0.055	<0.001	92.6%
<12	7	-0.73 (-1.74, 0.27)	0.155	<0.001	82.4%
Health status		· · · · · · · · · · · · · · · · · · ·			
Healthy	15	-2.01 (-3.31, -0.71)	0.002	<0.001	92.1%
Unhealthy	4	0.22 (-0.15, 0.60)	0.238	0.191	36.8%

TABLE 3 Continued

	Number of studies	WMD (95%CI)	<i>p</i> -value	Heterogeneity	
				P heterogeneity	l ²
Supplementation dose (g/da	y)				
≥3	8	0.09 (-0.53, 0.72)	0.772	<0.001	80.5%
<3	11	-3.08 (-4.91, -1.25)	0.001	<0.001	92.9%
Sex					
Both sexes	13	-1.29 (-2.62, 0.03)	0.056	<0.001	91.8%
Male	6	-1.01 (-1.95, -0.08)	0.033	<0.001	86.7%
Baseline BMI (kg/m ²)		-			
Normal (18.5–24.9)	2	-9.29 (-18.51, -0.08)	0.048	0.001	90.7%
Overweight (25–29.9)	11	-0.77 (-1.69, 0.14)	0.099	<0.001	82.1%
Obese (>30)	5	0.35 (-0.35, 1.05)	0.327	<0.001	83.5%
lsomer's type		1	L	1	
Both	17	-1.41 (-2.31, -0.50)	0.002	<0.001	90.6%
CLA-c9t11	1	-0.29 (-0.92, 0.34)	0.369	-	-
CLA-t10c12	1	1.29 (0.64, 1.93)	<0.001	-	-
Subgroup analyses of CLA su	pplementation on adiponectin (µg/ml)			
Overall effect	18	0.01 (-0.68, 0.71)	0.976	<0.001	98.09
Trial duration (week)		1	1	1	
≥12	11	-0.09 (-0.57, 0.39)	0.717	<0.001	93.29
<12	7	0.20 (-1.39, 1.79)	0.805	<0.001	93.49
Health status	I	1	1	1	
Healthy	13	0.05 (-0.38, 0.49)	0.811	<0.001	89.49
Unhealthy	5	-0.12 (-2.41, 2.17)	0.918	<0.001	98.79
Supplementation dose (g/da	у)	1	1	1	
≥3	11	-0.06 (-1.35, 1.23)	0.928	<0.001	97.39
<3	7	0.13 (-0.46, 0.73)	0.669	<0.001	94.6%
Sex					
Both sexes	6	0.41 (-1.18, 2.00)	0.614	<0.001	88.79
Male	10	-0.04 (-0.50, 0.42)	0.865	<0.001	92.19
Female	2	-1.05 (-1.58, -0.52)	<0.001	0.311	2.8%
Baseline BMI (kg/m ²)	·				
Normal (18.5–24.9)	3	0.25 (-0.50, 1.01)	0.511	<0.001	93.6%
Overweight (25-29.9)	5	-0.19 (-0.85, 0.47)	0.568	0.943	0.0%
Obese (>30)	10	-0.00 (-1.08, 1.07)	0.995	<0.001	98.5%
lsomer's type	· ·				
Both	15	0.05 (-0.72, 0.82)	0.900	<0.001	98.3%
CLA-c9t11	1	-0.00 (-1.28, 1.26)	0.989	_	-
CLA-t10c12	2	-0.25 (-1.15, 0.63)	0.572	-	90.09
Subgroup analyses of CLA su	pplementation on leptin (ng/ml)	I	I	·	

TABLE 3 Continued

	Number of studies	WMD (95%CI)	<i>p</i> -value	Heterogeneity	
				P heterogeneity	l ²
Overall effect	28	-0.30 (-0.79, 0.17)	0.214	<0.001	95.9%
Trial duration (week)					
≥12	19	0.03 (-0.16, 0.23)	0.743	0.002	54.9%
<12	9	-0.81 (-1.71, 0.08)	0.077	0.005	63.8%
Health status					
Healthy	23	0.06 (-0.11, 0.24)	0.463	0.018	42.3%
Unhealthy	5	-1.69 (-1.80, -1.58)	<0.001	0.451	0.0%
Supplementation dose (g/da	y)				
≥3	19	-0.32 (-0.92, 0.28)	0.297	<0.001	96.8%
<3	9	-0.19 (-0.85, 0.46)	0.567	0.002	67.9%
Sex					
Both sexes	10	-0.64 (-1.47, 0.17)	0.124	<0.001	98.4%
Male	12	0.06 (-0.35, 0.48)	0.754	0.022	50.8%
Female	6	-0.95 (-1.79, -0.12)	0.025	0.696	0.0%
Baseline BMI (kg/m ²)					
Normal (18.5–24.9)	5	0.50 (0.28, 0.72)	< 0.001	0.703	0.0%
Overweight (25–29.9)	14	-0.00 (-0.14, 0.13)	0.966	0.343	10.0%
Obese (>30)	9	-0.73 (-1.59, 0.12)	0.092	<0.001	96.6%
lsomer's type					
Both	25	-0.33 (-0.85, 0.17)	0.196	<0.001	96.4%
CLA-c9t11	1	0.41 (-1.34, 2.18)	0.643	-	-
CLA-t10c12	2	-0.27 (-1.97, 1.43)	0.756	0.210	36.5%

CI, confidence interval; WMD, weighted mean differences; CLA, conjugated linoleic acid; CRP, c-reactive protein; IL-6, interleukin 6; TNF-α, tumor necrosis factor-alpha.

increased (WMD = 1.29 mg/dl; 95% CI, 0.64 to 1.93; p < 0.001). Between-study heterogeneity was out of sight in studies with unhealthy subjects ($I^2 = 36.8\%$; p = 0.191). In the other subgroups, the effect of CLA supplementation on serum concentrations of TNF- α was not significant (Table 3).

Effect of CLA supplementation on adiponectin

Combining 18 effect sizes from 12 studies showed that CLA supplementation had no significant effect on adiponectin level in the intervention group compared to placebo [(WMD = -0.01 mg/dl; 95% CI, -0.68 to 0.71; p = 0.976 ($I^2 = 98.0\%$, p < 0.001)] (Figure 2D). Subgroup analyses showed a significant decrease in adiponectin levels in the female subjects (WMD = -1.05 mg/dl; 95% CI, -1.58 to -0.52; p < 0.001). Our subgroup analyses indicated no significant betweenstudy heterogeneity in studies conducted on the female subjects ($I^2 = 2.8\%$; p = 0.311) and overweight ($I^2 = 0.0\%$; p = 0.943) (Table 3).

Effect of CLA supplementation on leptin

Combining 28 effect sizes from 20 studies showed that CLA supplementation had no significant effect on leptin level in the

intervention compared to the placebo group [(WMD = -0.30 mg/ dl; 95% CI, -0.79 to 0.17; p = 0.214 ($I^2 = 95.9\%$; p < 0.001)] (Figure 2E). Subgroup analyses showed a significant decrease in leptin levels in unhealthy subjects (WMD = -1.69 mg/dl; 95% CI, -1.80 to -1.58; p < 0.001), female subjects (WMD = -0.95 mg/ dl; 95% CI, -1.79 to -0.12; p = 0.025), and a significant increase in leptin levels in the subjects with normal BMI (18.5–24.9) (WMD = 0.50 mg/dl; 95% CI, 0.28 to 0.72; p < 0.001). Subgroup analyses indicated no significant between-study heterogeneity in studies conducted on unhealthy subjects ($I^2 = 0.0\%$; p = 0.696), overweight subjects ($I^2 = 0.0\%$; p = 0.943), and subjects with normal range of BMI ($I^2 = 0.0\%$; p = 0.703) (Table 3).

Publication bias

Although the visual inspection of funnel plots showed slight asymmetries for all of the outcome variables, no significant publication bias was detected according to Begg's test for most of the variables, such as CRP (p = 0.189), IL-6 (p = 0.441), adiponectin (p = 0.198), and leptin (p = 0.050). Publication bias was considerable just for TNF- α (p = 0.042) (Figures 3A–E).

Nonlinear dose-response analysis

The present study conducted a nonlinear dose–response regression to analyze the dose–response relationship between CLA supplementation and inflammatory cytokines and adipokines. We did not find a significant nonlinear relationship between dose (g/day) and changes in CRP [(coefficients = -0.059; $p_{nonlinearity} = 0.353$] (Figure 4A), IL-6 [(coefficients = -10.437; $p_{nonlinearity} = 0.244$)] (Figure 4B), TNF- α [(coefficients = -791.208; $p_{nonlinearity} = 0.522$)] (Figure 4C), adiponectin [(coefficients = 1.337; $p_{nonlinearity} = 0.522$)] (Figure 4D), and leptin [(coefficients = -77.607; $p_{nonlinearity} = 0.841$)] (Figure 4E). Moreover, there was a nonlinear association between the duration of intervention and CRP [(coefficients = -0.798; $p_{nonlinearity} = 0.164$)] (Figure 5A), IL-6 [(coefficients = 2.635; $p_{nonlinearity} = 0.247$)] (Figure 5B), TNF- α 9[(coefficients = -8.156; $p_{nonlinearity} = 0.622$)] (Figure 5C), adiponectin [(coefficients = -0.343; $p_{nonlinearity} = 0.752$)] (Figure 5D), and leptin [(coefficients = 0.056; $p_{nonlinearity} = 0.748$)] (Figure 5E).

Meta-regression analysis

Meta-regression analyses were performed to assess whether inflammatory cytokines and adipokines were affected by CLA supplementation doses and intervention duration. We found no significant linear association between intervention dose (g/day) and changes in CRP [(coefficients = 0.279; $p_{\text{linearity}} = 0.304$)] (Figure 6A), IL-6 [(coefficients = 0.162; $p_{\text{linearity}} = 0.384$)] (Figure 6B), TNF- α



Funnel plots for the effect of CLA consumption on (A) CRP (mg/L), (B) IL-6 (pg/ml), (C) TNF- α (ng/ml), (D) adiponectin (µg/ml), and (E) leptin (ng/ml). Horizontal lines represent 95% CIs. Diamonds represent pooled estimates from random-effects analysis. WMD, weighted mean difference; CI, confidence interval; CLA, conjugated linoleic acid; CRP, C-reactive protein; IL-6, interleukin 6; TNF- α , tumor necrosis factor-alpha.



difference; CI, confidence interval; CLA, conjugated linoleic acid; CRP, C-reactive protein; IL-6, interleukin 6; TNF- α , tumor necrosis factor-alpha.

[(coefficients = 0.051; $p_{\text{linearity}} = 0.538$)] (Figure 6C), adiponectin [(coefficients = -0.152; $p_{\text{linearity}} = 0.616$)] (Figure 6D), leptin [(coefficients = 0.138; $p_{\text{linearity}} = 0.693$)] (Figure 6E), nor duration of supplementation (week) and changes in CRP [(coefficients = 1.001; $p_{\text{linearity}} = 0.365$)] (Figure 7A), IL-6 [(coefficients = 0.327; $p_{\text{linearity}} = 0.528$)] (Figure 7B), TNF- α [(coefficients = 0.038; $p_{\text{linearity}} = 0.737$)] (Figure 7C), adiponectin [(coefficients = -0.094; $p_{\text{linearity}} = 0.944$)] (Figure 7D), and leptin [(coefficients = -0.006; $p_{\text{linearity}} = 0.994$)] (Figure 7E).

Sensitivity analysis

The sensitivity analysis demonstrated that no study had a significant impact on the overall findings. Although the overall result of studies reporting data on TNF- α was sensitive to the study by Song et al. (48) (WMD = -0.644; 95% CI, -1.353, 0.065), and IL-6

was sensitive to the study by Sofi et al. (64) (WMD = -0.284; 95% CI, -0.601, 0.033), by omitting these studies, the significance of the correlation between CLA and TNF- α and IL-6 was lost.

In the case of CRP, some studies had an impact on the overall effect size, including that of Whigham et al. (54) (WMD = 0.253; 95% CI, -0.013, 0.520), Smedman et al. (49) (WMD = 0.247; 95% CI, -0.010, 0.506), and Pfeuffer et al. (69) (WMD = 0.257; 95% CI, -0.007, 0.523); the arm "a" and "b" of the study of Ramakers et al. (50) (WMD = 0.258; 95% CI, -0.013, 0.529; WMD = 0.245; 95% CI, -0.028, 0.518), Taylor et al. (51) (WMD = 0.251; 95% CI, -0.009, 0.512), Gaullier et al. (27) (WMD = 0.190; 95% CI, -0.033, 0.434), and Mullen et al. (57) (WMD = 0.226; 95% CI, -0.036, 0.490); the arm "a" and "b" of the study of Steck et al. (56) (WMD = 0.245; 95% CI, -0.010, 0.501; WMD = 0.237; 95% CI = -0.011, 0.485), Raff et al. (61) (WMD = 0.219; 95% CI, -0.036, 0.484), and Sluijs et al. (65) (WMD = 0.262; 95% CI, -0.003, 0.529); and the arm "b" of the study of Joseph et al. (68) (WMD = 0.237; 95% CI, 0.026, 0.501) and Lopez-Plaza et al.



necrosis factor-alpha.

(16) (WMD = 0.249; 95% CI, -0.016, 0.515). It was shown that the exclusion of every individual study by the sensitivity analysis could not change the direction of the correlation but eliminated the significant effect of CLA on CRP.

GRADE assessment

Table 4 displays the Grading of Recommendations Assessment, Development, and Evaluation (GRADE) profile of CLA supplementation on inflammatory cytokine and adipokine variables together with the certainty in outcomes. For CRP, because of serious limitations in inconsistency, the quality of evidence was moderate. In the case of IL-6, because of very serious limitations in inconsistency and for TNF- α because of very serious limitations in inconsistency ad serious limitations for publication bias, the quality of evidence was low. For both outcomes, including leptin and adiponectin, because of very serious limitations in inconsistency and serious limitations in imprecision, the quality of evidence was very low.

Certainty assessment

Using the GRADE methodology, which was previously outlined, the total degree of evidence certainty across the studies was evaluated and summarized (77).

Discussion

In this review, an analysis of pooling 42 studies indicated that CLA supplementation increased CRP concentrations, decreased IL-6 and TNF- α values, and had no effect on adiponectin and leptin levels. Findings of subgroup analyses showed that CLA consumption increased CRP in subjects with normal CRP levels (less than 3 mg/dl) and those who were healthy or consumed CLA in doses of ≥ 3 g/day. Taking CLA decreased IL-6 in male individuals and unhealthy subjects if the trial duration was less than 12 weeks and when a mixture of two isomers was used. TNF- α was reduced by CLA consumption in studies with an intervention dose of <3 g/day, male



subjects, healthy persons, or those having normal baseline BMI, and when a mixture of two isomers was used. However, in subgroup analyses based on isomer type, 10trans12cis- CLA isomer significantly increased TNF- α . Supplementation with CLA reduced adiponectin in women. It also decreased leptin in women and unhealthy adults but increased in those with normal baseline BMI. Moreover, the results of the meta-regression analysis showed that there was no significant linear association between intervention doses and duration of supplementation with inflammatory cytokines and adipokines.

The majority of studies included in this meta-analysis used supplements consisting of mixed amounts of CLA isomers. Therefore, since CLA isomers have shown different effects on inflammation (30, 78), it is suggested that this issue be considered in research. It is also important to consider the contents of the placebo used in studies. Some included RCTs have used vegetable oils as a placebo, such as safflower and olive oils. One study has indicated the reduced effect of safflower oil on CRP levels compared to CLA (79). Another study has elucidated the anti-inflammatory effect of olive oils through the prevention of lipooxygenase and enzymes that synthesize leukotriene (80). In other words, prescribing these vegetable oils as a placebo might misinterpret results.

Diet and physical activity can play an important role in inflammatory conditions (81). It is better to consider the effects of these two factors in the result of the present meta-analysis. Although, in some studies, the confounding variables of physical activity and diet were controlled, in some trials, these factors were not considered.

Diet is an important regulatory factor in the immune response. Malnutrition leads to suppression of the immune system, whereas overnutrition leads to immune activation. Some foods have antiinflammatory effects, and there are still controversies about others (81). For example, several studies have reported a positive association between the glycemic index/glycemic load of dietary carbohydrates and inflammatory cytokines (81). Also, a high-fat diet causes excessive body fat accumulation and impairs the immune system. On the other hand, it has been reported that trans and saturated fatty acids are significantly associated with the inflammatory state (82).



Random-effects meta-regression plots of the association between duration of intervention of CLA and weighted mean difference of (A) CRP (mg/L), (B) IL-6 (pg/ml), (C) TNF-a (ng/ml), (D) adiponectin (µg/ml), and (E) leptin (ng/ml). Horizontal lines represent 95% Cls. Diamonds represent pooled estimates from random-effects analysis. WMD, weighted mean difference; CI, confidence interval; CRP, C-reactive protein; IL-6, interleukin 6; TNF-α, tumor necrosis factor-alpha.

TABLE 4 GRADE profile of CLA supplementation for inflammatory cytokin	es and adipokines.
---	--------------------

Outcomes	Risk of bias	Inconsistency	Indirectness	Imprecision	Publication bias	Number of interventions/ control	Quality of evidence
CRP	No serious limitation	Serious limitation ^a	No serious limitation	No serious limitation	No serious limitation	1,192 (597/595)	⊕⊕⊕x̂ Moderate
IL-6	No serious limitation	Very serious limitation ^a	No serious limitation	No serious limitation	No serious limitation	817 (418/399)	⊕⊕îxî Low
TNF-α	No serious limitation	Very serious limitation ^a	No serious limitation	No serious limitation	Serious limitation ^c	779 (402/377)	⊕⊕îxî Low
Adiponectin	No serious limitation	Very serious limitation ^a	No serious limitation	Serious limitation ^b	No serious limitation	845 (424/421)	⊕îxîî Very low
Leptin	No serious limitation	Very serious limitation ^a	No serious limitation	Serious limitation ^b	No serious limitation	1,274 (649/625)	⊕îîî Very low

CLA, conjugated linoleic acid; CRP, c-reactive protein; IL-6, interleukin 6; TNF- α , tumor necrosis factor-alpha. ^aThere is a significant heterogeneity for CRP (I² = 56.3%%), IL-6 (I² = 86.3%), TNF- α (I² = 90.2%), adiponectin (I² = 98.0%), and leptin (I² = 95.9%). ^bThere is no evidence of significant effects of CLA supplementation on adiponectin and leptin. ^cThere is significant publication bias for TNF- α (p = 0.042).

Although ruminant trans fatty acids have different effects than industrial trans fatty acids, several studies show that industrial trans fatty acids promote inflammation, whereas ruminant trans fatty acids have been reported to be harmless or even beneficial to health, as well as also having anti-inflammatory properties (83).

In relation to physical activity, it has been reported that although exercise produces a short-term inflammatory response, regular exercise in the long term has an "anti-inflammatory" effect (84). One study reported that sedentary subjects had higher levels of inflammatory factors such as IL-6 and TNF- α compared to subjects with higher physical activity (85).

Other confounding factors that can affect inflammatory markers are sleep duration and sleep quality. Recently, studies have reported a relationship between longer sleep duration (>8 h) and increased levels of inflammatory markers such as CRP and IL-6 (86, 87). In addition, a study on healthy subjects reported that higher IL-6 levels were associated with lower sleep quality (88).

Here, we showed CLA consumption increased CRP concentrations, decreased IL-6 and TNF-a, and did not change adiponectin and leptin levels. There are some meta-analyses that have investigated the effect of CLA on the abovementioned markers. For example, Derakhshandeh-Rishehri et al. (2019) (89), working on 15 RCTs, revealed that long-term (24 weeks) CLA intake significantly increased CRP levels (89). A meta-analysis conducted by Haghighatdoost et al. (28) showed a significant increase in CRP (n = 8) and TNF- α levels (n = 8) along with a slight decrease in blood IL-6 (n = 9) following CLA supplementation (29). Another meta-analysis (2018) conducted on 19 RCTs demonstrated that CLA intake had a small but not significant decline in plasma leptin. However, it significantly decreased leptin in obese individuals, with trials lasting for less than 24 weeks (21). A study carried out by Mazidi et al. (89) on 14 RCTs indicated that treatment with CLA had no effect on IL-6 but significantly reduced serum TNF- α and adiponectin, besides enhancing serum CRP concentrations (90). By combining previous and current results, it can be acknowledged that except for the effect of CLA in increasing CRP levels, there is no consensus on other markers (i.e., IL-6, TNF-α, adiponectin, and leptin). Moreover, it is noteworthy to state that exercise, dietary intake, sleep duration, and sleep quality can alter the effects of CLA. Therefore, further welldesigned studies may clarify the role of CLA on the abovementioned markers.

In the present study, the gender-dependent impact of CLA has also been shown in the reduction of IL-6, TNF- α , adiponectin, and leptin. Based on evidence, CLA can induce testosterone biosynthesis (91), and subsequently, testosterone is able to decrease IL-6 expression (92). It has also been stated that circulating CLA concentrations are greater in women than in men (93). Therefore, the aforementioned evidence can justify the different effects seen between men (IL-6 and TNF- α reduction) and women (adiponectin and leptin reduction) after taking CLA.

The effect of CLA on inflammatory responses has generated inconsistent results. This may be due to isomers and tissue-specific responses of CLA. Moreover, across the included RCTs, administration doses of CLA varied from 1.6 to 10.8 mg/day, and the duration of the treatment differed from 4 to 104 weeks. Health characteristics, inflammation status, and even gut microbiome composition of the recruited individuals were also different (78). These variations in the studies can influence the results and interpretations.

For example, in relation to isomer and tissue-specific response, 9cis,11trans-CLA isomer exerts its anti-inflammatory effect by activating the peroxisome proliferator-activated receptor-y (PPAR- γ)-dependent pathway and ultimately reducing the production of proinflammatory cytokines such as TNF- α and IL-6 (30). However, the 10trans,12cis-CLA isomer has been reported to have a proinflammatory effect in adipose tissue, in contrast to its effects on innate immunity and vascular cells. In addition, in adipose tissue, activation of PPARy is contrary to the physiological effects of 10trans,12cis-CLA in vitro and in vivo (78). This isomer downregulates PPAR-y gene expression and increases TNF-a, IL-6, and CRP production in adipose tissue (78, 94). It seems that the reason for this inconsistency is cytokines secreted from adipocytes, which in turn alter PPARy expression and activity in the fat cell. Therefore, inconclusive results regarding the anti-inflammatory properties of CLA are likely due to the subtle proinflammatory effects of the 10trans,12cis-CLA isomers in specific tissues (30, 78). Overall, it appears that CLA can elicit both anti-inflammatory and proinflammatory features.

Some proposed mechanisms for the effects of CLA isomers on inflammation include the following: (1) Modulation of eicosanoid signaling. This means that CLA reduces the production of inflammatory eicosanoids derived from arachidonic acid (AA) through the inhibition of several steps in the AA cascade. (2) Modulating the transcription of some genes, such as the PPAR gene, plays an important role in the regulation of inflammation. Also, it suppresses the expression of the inducible NO synthase (iNOS) gene, which leads to a decrease in IL-6 production. (3) Inhibition of NF-kB activity, which is a transcription factor involved in cytokine gene expression, cellular adhesion, cell cycle activation, apoptosis, and carcinogenesis and is integrally related to inflammatory responses (78, 95).

This meta-analysis has some limitations that should be addressed. Most of the included studies had a relatively small sample size, which can cause an overestimation of the results. Observation of publication bias for TNF- α findings suggests overestimation of CLA efficiency on TNF- α . Moreover, TNF- α , IL-6, and CRP results were sensitive to some studies.

In conclusion, it is suggested that CLA supplementation can have both proinflammatory and anti-inflammatory roles. It can enhance CRP concentrations while reducing TNF- α and IL-6 levels. Furthermore, CLA is able to decrease adiponectin and leptin in women. It can also decrease leptin in unhealthy adults and increase it in subjects with a normal baseline BMI. Finally, in order to improve the quality of studies, future clinical trials are encouraged to carefully consider CLA-isomer-specific regulation of inflammatory markers and take notice of the contents of the placebo used in their control groups. It is also important to keep in mind the gender-dependent impact of CLA.

Data availability statement

The original contributions presented in the study are included in the article/supplementary material. Further inquiries can be directed to the corresponding authors.

Author contributions

OA and SR contributed to the conception and design of the study. DA-L, MZ, and GS contributed to data extraction. FS and AK screened articles for inclusion criteria. OA contributed to the data analysis. SR, MY, and EG contributed to manuscript drafting, OA and MZ supervised the study. All authors contributed to the article and approved the submitted version.

References

1. Asbaghi O, Ashtary-Larky D, Bagheri R, Moosavian SP, Nazarian B, Afrisham R, et al. Effects of folic acid supplementation on inflammatory markers: A grade-assessed systematic review and dose-response meta-analysis of randomized controlled trials. *Nutrients* (2021) 13(7):2327. doi: 10.3390/nu13072327

2. Ghafourian M, Ashtary-Larky D, Chinipardaz R, Eskandary N, Mehavaran M. Inflammatory biomarkers' response to two different intensities of a single bout exercise among soccer players. *Iran Red Crescent Med J* (2016) 18(2):e21498. doi: 10.5812/ ircmj.21498

3. Zouhal H, Bagheri R, Ashtary-Larky D, Wong A, Triki R, Hackney AC, et al. Effects of Ramadan intermittent fasting on inflammatory and biochemical biomarkers in males with obesity. *Phys Behavior* (2020) 225:113090. doi: 10.1016/j.physbeh.2020.113090

4. Bagheri R, Rashidlamir A, Ashtary-Larky D, Wong A, Alipour M, Motevalli MS, et al. Does green tea extract enhance the anti-inflammatory effects of exercise on fat loss? *Br J Clin Pharmacol* (2020) 86(4):753–62. doi: 10.1111/bcp.14176

5. Bagheri R, Rashidlamir A, Ashtary-Larky D, Wong A, Grubbs B, Motevalli MS, et al. Effects of green tea extract supplementation and endurance training on irisin, proinflammatory cytokines, and adiponectin concentrations in overweight middle-aged men. *Eur J Appl Physiol* (2020) 120(4):915–23. doi: 10.1007/s00421-020-04332-6

6. Kooti W, Farokhipour M, Asadzadeh Z, Ashtary-Larky D, Asadi-Samani M. The role of medicinal plants in the treatment of diabetes: A systematic review. *Electronic Physician* (2016) 8(1):1832. doi: 10.19082/1832

7. Shokrpour M, Asemi Z. The effects of magnesium and vitamin e Cosupplementation on hormonal status and biomarkers of inflammation and oxidative stress in women with polycystic ovary syndrome. *Biol Trace Elem Res* (2019) 191(1):54– 60. doi: 10.1007/s12011-018-1602-9

8. Kalmarzi RN, Naleini SN, Ashtary-Larky D, Peluso I, Jouybari L, Rafi A, et al. Antiinflammatory and immunomodulatory effects of barberry (Berberis vulgaris) and its main compounds. *Oxid Med Cell Longev* (2019) 2019:6183965. doi: 10.1155/2019/6183965

9. Dehbalaei MG, Ashtary-Larky D, Mesrkanlou HA, Talebi S, Asbaghi O. The effects of magnesium and vitamin e co-supplementation on some cardiovascular risk factors: A meta-analysis. *Clin Nutr ESPEN* (2021) 41:110–7. doi: 10.1016/j.clnesp.2020.10.021

10. Cengiz M, Uysal BB, Ikitimur H, Ozcan E, Islamoğlu MS, Aktepe E, et al. Effect of oral l-glutamine supplementation on covid-19 treatment. *Clin Nutr experimental* (2020) 33:24–31. doi: 10.1016/j.yclnex.2020.07.003

11. Colakoglu S, Colakoglu M, Taneli F, Cetinoz F, Turkmen M. Cumulative effects of conjugated linoleic acid and exercise on endurance development. *J sports Med Phys fitness.* (2006) 46:4.

12. MacRedmond R, Singhera G, Attridge S, Bahzad M, Fava C, Lai Y, et al. Conjugated linoleic acid improves airway hyper-reactivity in overweight mild asthmatics. *Clin Exp Allergy* (2010) 40(7):1071–8. doi: 10.1111/j.1365-2222.2010.03531.x

13. Wang T, Lee HG. Advances in research on cis-9, trans-11 conjugated linoleic acid: A major functional conjugated linoleic acid isomer. *Crit Rev Food Sci Nutr* (2015) 55 (5):720–31. doi: 10.1080/10408398.2012.674071

14. Wallace RJ, McKain N, Shingfield KJ, Devillard E. Isomers of conjugated linoleic acids are synthesized *via* different mechanisms in ruminal digesta and bacteria. *J Lipid Res* (2007) 48(10):2247–54. doi: 10.1194/jlr.M700271-JLR200

15. Yamasaki M, Nagatomo T, Matsuyama T, Ikeho Y, Kato E, Nishiyama K, et al. Conjugated linoleic acids inhibit hypoxia inducible factor- 1α stabilization under hypoxic condition in human hepatocellular carcinoma cells. *J Oleo Science*. (2012) 61(9):491–6. doi: 10.5650/jos.61.491

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

16. López-Plaza B, Bermejo LM, Weber TK, Parra P, Serra F, Hernández M, et al. Effects of milk supplementation with conjugated linoleic acid on weight control and body composition in healthy overweight people. *Nutricion Hospitalaria*. (2013) 28(6):2090–8. doi: 10.3305/nh.2013.28.6.7013

17. Ebrahimi-Mameghani M, Jamali H, Mahdavi R, Kakaei F, Abedi R, Kabir-Mamdooh B. Conjugated linoleic acid improves glycemic response, lipid profile, and oxidative stress in obese patients with non-alcoholic fatty liver disease: A randomized controlled clinical trial. *Croatian Med J* (2016) 57(4):331–42. doi: 10.3325/cmj.2016.57.331

18. Aslani MR, Matin S, Nemati A, Mesgari-Abbasi M, Ghorbani S, Ghobadi H. Effects of conjugated linoleic acid supplementation on serum levels of interleukin-6 and sirtuin 1 in COPD patients. *Avicenna J Phytomedicine* (2020) 10(3):305.

19. Aryaeian N, Djalali M, Shahram F, Djazayery A, Eshragian MR, et al. Effect of conjugated linoleic acid Effect of conjugated linoleic Acid, vitamin E, alone or combined on immunity and inflammatory parameters in adults with active rheumatoid arthritis: A randomized controlled trial Alone or combined on immunity and inflammatory parameters in adults with active rheumatoid arthritis: A randomized controlled trial Alone or combined on immunity and inflammatory parameters in adults with active rheumatoid arthritis: A randomized controlled trial. *Int J Prev Med* (2014) 5(12):1567.

20. Villacorta L, Minarrieta L, Salvatore SR, Khoo NK, Rom O, Gao Z, et al. *In situ* generation, metabolism and immunomodulatory signaling actions of nitro-conjugated linoleic acid in a murine model of inflammation. *Redox Biol* (2018) 15:522–31. doi: 10.1016/j.redox.2018.01.005

21. Mohammadi-Sartang M, Sohrabi Z, Esmaeilinezhad Z, Jalilpiran Y. Effect of conjugated linoleic acid on leptin level: A systematic review and meta-analysis of randomized controlled trials. *Hormone Metab Res* (2018) 50(02):106–16. doi: 10.1055/s-0044-100041

22. Syvertsen C, Halse J, Høivik HO, Gaullier J, Nurminiemi M, Kristiansen K, et al. The effect of 6 months supplementation with conjugated linoleic acid on insulin resistance in overweight and obese. *Int J Obes* (2007) 31(7):1148–54. doi: 10.1038/sj.ijo.0803482

23. Shahmirzadi FE, Ghavamzadeh S, Zamani T. The effect of conjugated linoleic acid supplementation on body composition, serum insulin and leptin in obese adults. *Arch Iranian Med* (2019) 22(5):255.

24. Ghobadi H, Matin S, Nemati A, Javadi H, Alipanah-Moghadam R, Saeidi-Nir M. The effect of conjugated linoleic acid supplementation on the serum leptin level, pulmonary function and quality of life in COPD patients. *J Ardabil Univ Med Sci* (2019) 19(1):53–60. doi: 10.29252/jarums.19.1.53

25. Faramarzi E, Mohammadzadeh M, Sanaie S, Andersen V, Mahdavi R. Effects of conjugated linoleic acid supplementation on serum leptin levels, oxidative stress factors and tumor marker in rectal cancer patients undergoing preoperative chemoradiotherapy. *Mediterr J Nutr Metab* (2021) 14(3):245–53. doi: 10.3233/MNM-200507

26. von Frankenberg AD, Silva FM, de Almeida JC, Piccoli V, do Nascimento FV, Sost MM, et al. Effect of dietary lipids on circulating adiponectin: A systematic review with meta-analysis of randomised controlled trials. *Br J Nutr* (2014) 112(8):1235–50. doi: 10.1017/S0007114514002013

 Gaullier J-M, Halse J, Høivik HO, Høye K, Syvertsen C, Nurminiemi M, et al. Six months supplementation with conjugated linoleic acid induces regional-specific fat mass decreases in overweight and obese. Br J Nutr (2007) 97(3):550–60. doi: 10.1017/ S0007114507381324

28. Sneddon AA, Tsofliou F, Fyfe CL, Matheson I, Jackson DM, Horgan G, et al. Effect of a conjugated linoleic acid and ω -3 fatty acid mixture on body composition and adiponectin. *Obesity* (2008) 16(5):1019–24. doi: 10.1038/oby.2008.41

29. Haghighatdoost F, Gh NM, Fatemeh B. Effect of conjugated linoleic acid on blood inflammatory markers: A systematic review and meta-analysis on randomized controlled trials. *Eur J Clin Nutr* (2018) 72(8):1071–82. doi: 10.1038/s41430-017-0048-z

30. Viladomiu M, Hontecillas R, Bassaganya-Riera J. Modulation of inflammation and immunity by dietary conjugated linoleic acid. *Eur J Pharmacol* (2016) 785:87–95. doi: 10.1016/j.ejphar.2015.03.095

31. Liberati A, Altman DG, Tetzlaff J, Mulrow C, Gøtzsche PC, Ioannidis JP, et al. The PRISMA statement for reporting systematic reviews and meta-analyses of studies that evaluate health care interventions: explanation and elaboration. *J Clin Epidemiol* (2009) 62(10):e1–e34. doi: 10.1016/j.jclinepi.2009.06.006

32. Nang C, Piano B, Lewis A, Lycett K, Woodhouse M. Using the PICOS model to design and conduct a systematic search: A speech pathology case study. (2015).

33. Higgins JGS. *Cochrane handbook for systematic reviews of interventions version 5.1.* 0 Vol. 2011. The Cochrane Collaboration (2011).

34. DerSimonian R, Laird N. Meta-analysis in clinical trials. Controlled Clin trials. (1986) 7(3):177-88. doi: 10.1016/0197-2456(86)90046-2

35. Borenstein M, Hedges LV, Higgins JP, Rothstein HR. Introduction to metaanalysis. (UK: John Wiley & Sons) (2021).

36. Hozo SP, Djulbegovic B, Hozo I. Estimating the mean and variance from the median, range, and the size of a sample. *BMC Med Res methodology*. (2005) 5(1):1–10. doi: 10.1186/1471-2288-5-13

37. Higgins JP, Thompson SG. Quantifying heterogeneity in a meta-analysis. *Stat Med* (2002) 21(11):1539–58. doi: 10.1002/sim.1186

38. Egger M, Smith GD, Schneider M, Minder C. Bias in meta-analysis detected by a simple, graphical test. *Bmj.* (1997) 315(7109):629–34. doi: 10.1136/bmj.315.7109.629

39. Duval S. The trim and fill method. publication bias in meta-analysis: Prevention, assessment and adjustments. (London: Elsevier Ltd) (2005) 127-44. doi: 10.1002/0470870168.ch8

40. Xu C, Doi SA. The robust error meta-regression method for dose-response metaanalysis. *JBI Evidence Implementation* (2018) 16(3):138-44. doi: 10.1097/ XEB.000000000000132

41. Xie Y, Gou L, Peng M, Zheng J, Chen L. Effects of soluble fiber supplementation on glycemic control in adults with type 2 diabetes mellitus: A systematic review and metaanalysis of randomized controlled trials. *Clin Nutr* (2021) 40(4):1800–10. doi: 10.1016/ j.clnu.2020.10.032

42. Medina EA, Horn WF, Keim NL, Havel PJ, Benito P, Kelley DS, et al. Conjugated linoleic acid supplementation in humans: effects on circulating leptin concentrations and appetite. *Lipids*. (2000) 35(7):783–8. doi: 10.1007/s11745-000-0586-y

43. Riserus U, Berglund L, Vessby B. Conjugated linoleic acid (CLA) reduced abdominal adipose tissue in obese middle-aged men with signs of the metabolic syndrome: A randomised controlled trial. *Int J Obes* (2001) 25(8):1129-35. doi: 10.1038/sj.ijo.0801659

44. Albers R, van der Wielen R, Brink E, Hendriks H, Dorovska-Taran V, Mohede I. Effects of cis-9, trans-11 and trans-10, cis-12 conjugated linoleic acid (CLA) isomers on immune function in healthy men. *Eur J Clin Nutr* (2003) 57(4):595–603. doi: 10.1038/ sj.ejcn.1601585

45. Risérus U, Vessby B, Ärnlöv J, Basu S. Effects of cis-9, trans-11 conjugated linoleic acid supplementation on insulin sensitivity, lipid peroxidation, and proinflammatory markers in obese men. *Am J Clin Nutr* (2004) 80(2):279–83. doi: 10.1093/ajcn/80.2.279

46. Desroches S, Chouinard PY, Galibois I, Corneau L, Delisle J, Lamarche BT, et al. Lack of effect of dietary conjugated linoleic acids naturally incorporated into butter on the lipid profile and body composition of overweight and obese men–. *Am J Clin Nutr* (2005) 82(2):309–19. doi: 10.1093/ajcn/82.2.309

47. Gaullier J-M, Halse J, Høye K, Kristiansen K, Fagertun H, Vik H, et al. Supplementation with conjugated linoleic acid for 24 months is well tolerated by and reduces body fat mass in healthy, overweight humans. *J Nutr* (2005) 135(4):778–84. doi: 10.1093/jn/135.4.778

48. Song H, Grant I, Rotondo D, Mohede I, Sattar N, Heys S, et al. Effect of CLA supplementation on immune function in young healthy volunteers. *Eur J Clin Nutr* (2005) 59(4):508–17. doi: 10.1038/sj.ejcn.1602102

49. Smedman A, Basu S, Jovinge S, Fredrikson GN, Vessby B. Conjugated linoleic acid increased c-reactive protein in human subjects. *Br J Nutr* (2005) 94(5):791–5. doi: 10.1079/BJN20041419

50. Ramakers JD, Plat J, Sébédio J-L, Mensink RP. Effects of the individual isomers cis-9, trans-11 vs. trans-10, cis-12 of conjugated linoleic acid (CLA) on inflammation parameters in moderately overweight subjects with LDL-phenotype b. *Lipids* (2005) 40 (9):909–18. doi: 10.1007/s11745-005-1451-8

51. Taylor JS, Williams SR, Rhys R, James P, Frenneaux MP. Conjugated linoleic acid impairs endothelial function. *Arteriosclerosis thrombosis Vasc Biol* (2006) 26(2):307–12. doi: 10.1161/01.ATV.0000199679.40501.ac

52. Naumann E, Carpentier YA, Saebo A, Lassel TS, Chardigny J-M, Sébédio J-L, et al. Cis-9, trans-11 and trans-10, cis-12 conjugated linoleic acid (CLA) do not affect the plasma lipoprotein profile in moderately overweight subjects with LDL phenotype b. *Atherosclerosis* (2006) 188(1):167–74. doi: 10.1016/j.atherosclerosis.2005.10.019

53. Nazare J-A, de la Perriere AB, Bonnet F, Desage M, Peyrat J, Maitrepierre C, et al. Daily intake of conjugated linoleic acid-enriched yoghurts: effects on energy metabolism and adipose tissue gene expression in healthy subjects. *Br J Nutr* (2007) 97(2):273–80. doi: 10.1017/S0007114507191911

54. Whigham LD, Watras AC, Schoeller DA. Efficacy of conjugated linoleic acid for reducing fat mass: A meta-analysis in humans. *Am J Clin Nutr* (2007) 85(5):1203–11. doi: 10.1093/ajcn/85.5.1203

55. Iwata T, Kamegai T, Yamauchi-Sato Y, Ogawa A, Kasai M, Aoyama T, et al. Safety of dietary conjugated linoleic acid (CLA) in a 12-weeks trial in healthy overweight Japanese male volunteers. *J Oleo Science* (2007) 56(10):517–25. doi: 10.5650/jos.56.517

56. Steck SE, Chalecki AM, Miller P, Conway J, Austin GL, Hardin JW, et al. Conjugated linoleic acid supplementation for twelve weeks increases lean body mass in obese humans. J Nutr (2007) 137(5):1188–93. doi: 10.1093/jn/137.5.1188

57. Mullen A, Moloney F, Nugent AP, Doyle L, Cashman KD, Roche HM. Conjugated linoleic acid supplementation reduces peripheral blood mononuclear cell interleukin-2 production in healthy middle-aged males. *J Nutr Biochem* (2007) 18(10):658–66. doi: 10.1016/j.jnutbio.2006.12.008

 Kim J-H, Kim O-H, Ha Y-L, Kim J-O. Supplementation of conjugated linoleic acid with γ-oryzanol for 12 weeks effectively reduces body fat in healthy overweight Korean women. *Prev Nutr Food Science* (2008) 13(3):146–56. doi: 10.3746/jfn.2008.13.3.146

59. Turpeinen AM, Ylönen N, von Willebrand E, Basu S, Aro A. Immunological and metabolic effects of cis-9, trans-11-conjugated linoleic acid in subjects with birch pollen allergy. *Br J Nutr* (2008) 100(1):112–9. doi: 10.1017/S0007114507886326

60. Aryaeian N, Shahram F, Djalali M, Eshragian MR, Djazayeri A, Sarrafnejad A, et al. Effect of conjugated linoleic acid, vitamin e and their combination on lipid profiles and blood pressure of Iranian adults with active rheumatoid arthritis. *Vasc Health Risk Management* (2008) 4(6):1423. doi: 10.2147/VHRM.S3822

61. Raff M, Tholstrup T, Basu S, Nonboe P, Sørensen MT, Straarup EM. A diet rich in conjugated linoleic acid and butter increases lipid peroxidation but does not affect atherosclerotic, inflammatory, or diabetic risk markers in healthy young men. J Nutr (2008) 138(3):509–14. doi: 10.1093/jn/138.3.509

62. Zhao W-S, Zhai J-J, Wang Y-H, Xie P-S, Yin X-J, Li L-X, et al. Conjugated linoleic acid supplementation enhances antihypertensive effect of ramipril in Chinese patients with obesity-related hypertension. *Am J Hypertension.* (2009) 22(6):680–6. doi: 10.1038/ ajh.2009.56

63. Norris LE, Collene AL, Asp ML, Hsu JC, Liu L-F, Richardson JR, et al. Comparison of dietary conjugated linoleic acid with safflower oil on body composition in obese postmenopausal women with type 2 diabetes mellitus. *Am J Clin Nutr* (2009) 90(3):468–76. doi: 10.3945/ajcn.2008.27371

64. Sofi F, Buccioni A, Cesari F, Gori AM, Minieri S, Mannini L, et al. Effects of a dairy product (pecorino cheese) naturally rich in cis-9, trans-11 conjugated linoleic acid on lipid, inflammatory and haemorheological variables: A dietary intervention study. *Nutrition Metab Cardiovasc Diseases* (2010) 20(2):117–24. doi: 10.1016/j.numecd.2009.03.004

65. Sluijs I, Plantinga Y, De Roos B, Mennen LI, Bots ML. Dietary supplementation with cis-9, trans-11 conjugated linoleic acid and aortic stiffness in overweight and obese adults. *Am J Clin Nutr* (2010) 91(1):175–83. doi: 10.3945/ajcn.2009.28192

66. Tavakkoli Darestani A, Hosseinpanah F, Tahbaz F, Amiri Z, Tavakkoli Darestani R, Hedayati M. Effects of conjugated linoleic acid supplementation on body composition and leptin concentration in post-menopausal women. *Iranian J Endocrinol Metab* (2010) 12(1):48–59.

67. Brown AW, Trenkle AH, Beitz DC. Diets high in conjugated linoleic acid from pasture-fed cattle did not alter markers of health in young women. *Nutr Res* (2011) 31 (1):33–41. doi: 10.1016/j.nutres.2010.12.003

68. Joseph SV, Jacques H, Plourde M, Mitchell PL, McLeod RS, Jones PJ. Conjugated linoleic acid supplementation for 8 weeks does not affect body composition, lipid profile, or safety biomarkers in overweight, hyperlipidemic men. *J Nutr* (2011) 141(7):1286–91. doi: 10.3945/jn.110.135087

69. Pfeuffer M, Fielitz K, Laue C, Winkler P, Rubin D, Helwig U, et al. CLA does not impair endothelial function and decreases body weight as compared with safflower oil in overweight and obese male subjects. *J Am Coll Nutr* (2011) 30(1):19–28. doi: 10.1080/07315724.2011.10719940

70. Rubin D, Herrmann J, Much D, Pfeuffer M, Laue C, Winkler P, et al. Influence of different CLA isomers on insulin resistance and adipocytokines in pre-diabetic, middle-aged men with PPAR γ 2 Pro12Ala polymorphism. *Genes Nutr* (2012) 7(4):499–509. doi: 10.1007/s12263-012-0289-3

71. Shadman Z, Taleban FA, Saadat N, Hedayati M. Effect of conjugated linoleic acid and vitamin e on glycemic control, body composition, and inflammatory markers in overweight type2 diabetics. J Diabetes Metab Disord (2013) 12(1):1–9. doi: 10.1186/2251-6581-12-42

72. Bulut S, Bodur E, Colak R, Turnagol H. Effects of conjugated linoleic acid supplementation and exercise on post-heparin lipoprotein lipase, butyrylcholinesterase, blood lipid profile and glucose metabolism in young men. *Chemico-biological Interactions* (2013) 203(1):323–9. doi: 10.1016/j.cbi.2012.09.022

73. Eftekhari MH, Aliasghari F, Babaei-Beigi MA, Hasanzadeh J. Effect of conjugated linoleic acid and omega-3 fatty acid supplementation on inflammatory and oxidative stress markers in atherosclerotic patients. *ARYA atherosclerosis*. (2013) 9(6):311.

74. Baghi AN, Mazani M, Nemati A, Amani M, Alamolhoda S, Mogadam RA. Antiinflammatory effects of conjugated linoleic acid on young athletic males. *J Pak Med Assoc* (2016) 66(3):280–4.

75. Dus-Zuchowska M, Madry E, Krzyzanowska P, Bogdanski P, Walkowiak J. Twelve-week-conjugated linoleic acid supplementation has no effects on the selected markers of atherosclerosis in obese and overweight women. *Food Nutr Res* (2016) 60 (1):32776. doi: 10.3402/fnr.v60.32776

76. Abedi R, Aref-Hosseini S-R, Khoshbaten M, Ebrahimi-Mameghani M, Laleh HJ, Jalalypour F, et al. The effect of conjugated linoleic acid (CLA) on inflammatory factors in non-alcoholic fatty liver disease (NAFLD): A randomized controlled clinical trial. *Prog Nutr* (2018) 20:173–81. doi: 10.23751/pn.v20i2-S.5593

77. Guyatt GH, Oxman AD, Vist GE, Kunz R, Falck-Ytter Y, Alonso-Coello P, et al. GRADE: An emerging consensus on rating quality of evidence and strength of recommendations. *Bmj* (2008) 336(7650):924–6. doi: 10.1136/bmj.39489.470347.AD

78. Belda BJ, Lee Y, Heuvel JPV. Conjugated linoleic acids and inflammation: isomer-and tissue-specific responses. *Clin Lipidology* (2010) 5(5):699-717. doi: 10.2217/clp.10.54

79. Asp ML, Collene AL, Norris LE, Cole RM, Stout MB, Tang S-Y, et al. Timedependent effects of safflower oil to improve glycemia, inflammation and blood lipids in obese, post-menopausal women with type 2 diabetes: A randomized, double-masked, crossover study. *Clin Nutr* (2011) 30(4):443–9. doi: 10.1016/j.clnu.2011.01.001

80. Escrich E, Moral R, Grau L, Costa I, Solanas M. Molecular mechanisms of the effects of olive oil and other dietary lipids on cancer. *Mol Nutr Food Res* (2007) 51 (10):1279–92. doi: 10.1002/mnfr.200700213

81. Lee H, Lee IS, Choue R. Obesity, inflammation and diet. *Pediatr gastroenterology* Hepatol Nutr (2013) 16(3):143–52. doi: 10.5223/pghn.2013.16.3.143

82. Fritsche KL. The science of fatty acids and inflammation. Adv Nutr (2015) 6 (3):293S-301S. doi: 10.3945/an.114.006940

83. Oteng A-B, Kersten S. Mechanisms of action of trans fatty acids. *Adv Nutr* (2020) 11(3):697–708. doi: 10.1093/advances/nmz125

84. Kasapis C, Thompson PD. The effects of physical activity on serum c-reactive protein and inflammatory markers: A systematic review. J Am Coll Cardiol (2005) 45 (10):1563–9. doi: 10.1016/j.jacc.2004.12.077

85. Cho SMJ, Lee H, Shim J-S, Jeon JY, Kim HC. Association between physical activity and inflammatory markers in community-dwelling, middle-aged adults. *Appl Physiology Nutrition Metab* (2021) 46(7):828–36. doi: 10.1139/apnm-2020-1069

86. Patel SR, Zhu X, Storfer-Isser A, Mehra R, Jenny NS, Tracy R, et al. Sleep duration and biomarkers of inflammation. *Sleep* (2009) 32(2):200–4. doi: 10.1093/sleep/32.2.200

87. Dowd JB, Goldman N, Weinstein M. Sleep duration, sleep quality, and biomarkers of inflammation in a Taiwanese population. *Ann Epidemiol* (2011) 21(11):799–806. doi: 10.1016/j.annepidem.2011.07.004

88. Hong S, Mills PJ, Loredo JS, Adler KA, Dimsdale JE. The association between interleukin-6, sleep, and demographic characteristics. *Brain Behavior Immunity* (2005) 19 (2):165–72. doi: 10.1016/j.bbi.2004.07.008

89. Derakhshandeh-Rishehri S-M, Rahbar AR, Ostovar A. Effects of conjugated linoleic acid intake in the form of dietary supplement or enriched food on c-reactive protein and lipoprotein (a) levels in humans: A literature review and meta-analysis. *Iranian J Med Sci* (2019) 44(5):359.

90. Mazidi M, Karimi E, Rezaie P, Ferns GA. Effects of conjugated linoleic acid supplementation on serum c-reactive protein: A systematic review and meta-analysis of randomized controlled trials. *Cardiovasc Ther* (2017) 35(6):e12275. doi: 10.1111/1755-5922.12275

91. Barone R, Macaluso F, Catanese P, Marino Gammazza A, Rizzuto L, Marozzi P, et al. Endurance exercise and conjugated linoleic acid (CLA) supplementation up-regulate CYP17A1 and stimulate testosterone biosynthesis. *PLoS One* (2013) 8(11):e79686. doi: 10.1371/journal.pone.0079686

92. Bellido T, Jilka R, Boyce B, Girasole G, Broxmeyer H, Dalrymple S, et al. Regulation of interleukin-6, osteoclastogenesis, and bone mass by androgens. The role of the androgen receptor. *J Clin Invest* (1995) 95(6):2886–95. doi: 10.1172/JCI117995

93. Abdelmagid SA, Clarke SE, Wong J, Roke K, Nielsen D, Badawi A, et al. Plasma concentration of cis 9trans 11 CLA in males and females is influenced by SCD1 genetic variations and hormonal contraceptives: A cross-sectional study. *Nutr Metab* (2013) 10 (1):50. doi: 10.1186/1743-7075-10-50

94. Poirier H, Shapiro JS, Kim RJ, Lazar MA. Nutritional supplementation with trans-10, cis-12-conjugated linoleic acid induces inflammation of white adipose tissue. *Diabetes* (2006) 55(6):1634-41. doi: 10.2337/db06-0036

95. Iwakiri Y, Sampson D, Allen K. Suppression of cyclooxygenase-2 and inducible nitric oxide synthase expression by conjugated linoleic acid in murine macrophages. *Prostaglandins Leukotrienes Essential Fatty Acids* (2002) 67(6):435–43. doi: 10.1054/ plef.2002.0454