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## Review: Mechanisms and perspective treatment of radioresistance in non-small cell lung cancer

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Radiotherapy is the major treatment of non-small cell lung cancer (NSCLC). The radioresistance and toxicity are the main obstacles that leading to therapeutic failure and poor prognosis. Oncogenic mutation, cancer stem cells (CSCs), tumor hypoxia, DNA damage repair, epithelial-mesenchymal transition (EMT), and tumor microenvironment (TME) may dominate the occurrence of radioresistance at different stages of radiotherapy. Chemotherapy drugs, targeted drugs, and immune checkpoint inhibitors are combined with radiotherapy to treat NSCLC to improve the efficacy. This article reviews the potential mechanism of radioresistance and the advantages of Traditional Chinese medicine (TCM) in improving the efficacy and reducing the toxicity of radiotherapy.

#### KEYWORDS

non-small cell lung cancer, radiotherapy, radioresistance mechanisms, toxicity, Traditional Chinese Medicine

## 1 Introduction

Radiotherapy is the mainstay of treatment for patients with non-small cell lung cancer (NSCLC), and usually combined with surgery, chemotherapy, immunotherapy, and targeted therapy (1–6). The radiation commonly used in radiotherapy mainly includes photo radiation, such as X-ray and  $\gamma$ -ray and particle rays such as neutrons, electrons, protons and heavy ions (2, 7–10). However, no matter what radiation and dose segmentation methods are used, radioresistance will inevitably occur, leading to radiotherapy failure and local recurrence. Simply increasing the radiation dose does not improve survival benefits but leads to adverse reactions and poor prognosis (11).

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Radioresistance of NSCLC can be classified as inherent radioresistance and acquired radioresistance, of which acquired radioresistance plays a major role. Various mechanisms that cause radioresistance run through the whole process of radiotherapy. Whether they cause inherent radioresistance or acquired radioresistance is relative. We have simply classified them according to their leading role in different stages of radiotherapy. Acquired radioresistance is mainly related to DNA damage repair, tumor microenvironment (TME) remodeling by inflammation, immune response, tumor metabolic reprogramming, tumor microbiota and senescence cells, and epithelial-mesenchymal transition (EMT) (12–20). Inherent radioresistance is mainly associated with oncogenic mutation, cancer stem cells (CSCs), and tumor hypoxia (21–29).

Targeted drugs or small molecule drugs combined with radiotherapy are used in the treatment of NSCLC. However, there is no systematic review on the mechanism of radioresistance and radiosensitizers of NSCLC. Therefore, we will discuss the radioresistance mechanisms of NSCLC and the study of drugs that can enhance the efficacy and reduce the toxicity of radiotherapy, especially TCM. So as to provide a theoretical basis for clinical development of anti-tumor drugs in combination with radiotherapy.

# 2 Mechanisms of acquired radioresistance

Acquired radioresistance occurs mainly with radiation induction, the underlying mechanisms involve DNA damage repair and EMT in

cancer cell itself, as well as changes in TME such as inflammation, immune response, tumor metabolic reprogramming etc. (15, 16, 19, 30-32). Acquired radioresistance may enhance invasion and metastasis of surviving cancer cells after primary radiotherapy and affect the prognosis and life quality of patients.

### 2.1 DNA damage repair and radioresistance

Radiation causes direct DNA single-strand breaks (SSBs) and double-strand breaks (DSBs), as well as indirect DNA damage from oxidative stress such as ROS and reactive nitrogen species (RNS) (30) (Figure 1). SSBs is repaired by poly (ADP-ribose) polymerase (PARP) (33), as well as ATR/Chk1 pathway (34). ATR can be activated by recruitment to its partner protein ATRIP, binding to topoisomerase (DNA) II binding protein 1 (TopBP1) and claspin, phosphorylated Chk1 (34), activated Chk1 phosphorylated WEE1, leading to cell cycle arrest and DNA repair (35), resulting in radioresistance of NSCLC (36, 37). DSBs are repaired by homologous recombination (HR) and non-homologous ending joining (NHEJ) (33). In HR, the MRN complex comprising meiotic recombination 11 homolog1 (MRE11), ATP-binding cassette-ATPase (RAD50), and Nijmegen breakage syndrome protein 1 (NBS1), binds to C-terminal binding protein (CtBP)-interacting protein (CtIP) and BRCA1, was recruited by RAD17 to DSB sites, activated ATM, inducing the phosphorylation Chk2 and H2AX, as well as accumulation of RAD51, thus leading to cell cycle arrest and DNA repair (34, 35, 38, 39). Radiation delayed tumor growth and increased local control in Atm deletion tumor



#### FIGURE 1

Mechanisms of radioresistance caused by DNA damage repair in NSCLC. Radiation induced DNA damage response can activate the DNA repair pathway, of which SSBs can be repaired through ATR/Chk1 pathway and PARP1, DSBs can be repaired through NHEJ and HR. In the red box are proteins related to the repair of DSBs that may become new targets for radiosensitization. The solid line represents a clear target, and the dotted line represents a possible action through this target.

model of lung adenocarcinoma (40). In NHEJ, DNA-dependent protein kinase catalytic subunit (DNA-PKcs) is directly recruited to DNA damage sites via Ku heterodimers (Ku70/80), inducing Artemis-mediated end processing and DNA ligase 4 (LIG4) mediated ligation, leading to DNA repair (35). Besides, heat shock protein 90 (HSP90) (41), acidic nucleoplasmic DNA binding protein1 (AND-1) (42), sirtuin 3 (Sirt 3) (43), IQ motif containing GTPaseactivating protein 3 (IQGAP3) (44), and serine proteinase inhibitor clade E member 2 (SERPINE2) (45) can enhance radioresistance via activating ATM to activate the HR pathway in NSCLC. G2 and S phase-expressed 1(GTSE1) (46), plakophilin2 (PKP2) (47), ephrin type-A receptor 2 (EphA2) (48, 49), and cancer-derived IgG (cancer-IgG) (50) can participate in NHEJ pathway, leading to radioresistance in NSCLC. AAA ATPases RUVBL1/2 can activate both HR and NHEJ pathway, inducing radioresistance in NSCLC (51). In addition to Chk1 and Chk2, WEE1 can also regulate cell cycle arrest by inhibiting the phosphorylation of cyclin-dependent kinases (CDKs) (52, 53), inducing radioresistance (54). Thus, targeting DNA repair proteins or cell cycle regulator may overcome radiation-induced radioresistance in NSCLC.

It can be seen from the above that tumor cells have multiple ways to repair radiation-induced DNA damage. Drugs targeting a single

target may not have obvious effect on overcoming radioresistance, and are prone to drug resistance. Therefore, the combination of multiple targets for DNA repair will be the direction of tumor treatment in the future, but how to effectively combine the targets still needs more research.

## 2.2 Tumor microenvironment remodeling and radioresistance

Tumor microenvironment (TME) is closely related to tumor progression, and the remodeling of TME after radiotherapy can lead to radioresistance (18). Here, we focus on the effects of inflammation and immune response, tumor microbiota, tumor metabolic reprogramming, and cell senescence on TME to clarify the role of radiation-induced TME remodeling in the radioresistance of NSCLC (Figure 2).

#### 2.2.1 Inflammation and immune response

Radiation generated ROS and RNS can activate NF- $\kappa$ B, Janusassociated kinase (JAK)2/signal transducer and activator of transcription (STAT)3 pathway to increase the release of growth



and immune response can cause immunosuppression. The local lung microbiota can induce tumor growth by regulating the immune cells. Tumor metabolic reprogramming after radiation will lead to increase of acidity, hypoxia, and GSH production, unbalance of ferroptosis, and redox homeostasis maintaining, being conductive to tumor growth. Cell senescence after radiotherapy can also induce cell proliferation and tumor growth. The above factors together form TME remodeling that promotes tumor growth, metastasis and recurrence after radiotherapy. (This figure is adapted from an image created from BioRender.com).

factors and cytokines, such as VEGF, TGF-β, IL-1, IL-8, TNF-α and IL-6 etc. (15, 31, 55, 56), which are involved in inflammation and immune response (18, 57–60). TGF $\beta$  and TNF $\alpha$  in TME are very important in the formation and development of tumor (61). Our previous study found that irradiation of NSCLC cell lines A549 and H1299 can cause overexpression of TGF $\beta$  and TNF $\alpha$ , resulting in HIF1a activation and cause genomic instability of BMSCs and may promote tumor progression (62). We also found that TGF $\beta$  mainly causes short-term side effects, while TNF mainly causes long-term side effects. This may provide a guideline for clinical use of combined drugs at different stages of radiotherapy for NSCLC. In addition, radiation-induced inflammation reduces tumor-suppressing immune cells such as CD4+T cells, CD8+T cells, and natural killer (NK) cells, while tumor-promoting immune cells such as regulatory T cells (Treg), myeloid-derived suppressor cells (MDSC), and M2 tumorassociated macrophages (TAMs) increase, eventually forming an immunosuppressive environment, and high level of TGF $\beta$  (19, 63). Moreover, M2 macrophages and B cells infiltrate into TME also participate in the radioresistance of NSCLC (64, 65). Thus targeting inflammation not only enhance the tumor killing effects of radiotherapy, but also protect the normal cells and tissues from radiation-induced bystander effects.

#### 2.2.2 Tumor microbiota

It is well known that intestinal microbiome can regulate host physiological and pathological processes through metabolism, inflammation and immune response (66, 67). On the one hand, tumor microbiota can enhance anti-tumor immunity through STING signal activation, T and NK cell activation. On the other hand, it can upregulate ROS, induce anti-inflammatory environment, and inactivate T cell to promote immunosuppression and cancer progression (68). Moreover, tumor microbiota stimulates  $\gamma\delta T$  cells to produce IL-17 that triggering inflammation and cancer progression (69), and increases Th17 cells that enhancing lung cancer proliferation and angiogenesis (70). It is found that the dysbiosis of local lung microbiota in patients with advanced lung cancer can lead to Th1, Th17, γδT cells and PD-1 positive T cells increase, neutrophils recruitment, dendritic cells (DCs) activation, CD4+T cells decrease, cytokines IL-1, IL-1b, IL-6, IL-17, IL-23 upregulation, as well as upregulation of ERK/MAPK and inflammation pathways, all these above alterations promoted tumor progression (71). Although there is no research report on the correlation between tumor microbiota and radioresistance, there is no doubt that tumor microbiota will participate in tumor response to radiotherapy through inflammation and immune response and affect the final efficacy. The microbiome may serve as an indicator for cancer diagnosis or prognostic assessment and could be used as potential targets to develop new cancer therapies.

#### 2.2.3 Tumor metabolic reprogramming

Tumor metabolic reprogramming after radiotherapy, such as glucose metabolism, lipid metabolism, and redox metabolism, contributes to TME remodeling and acquired radioresistance (19). The increased expression of glucose transporter1 (GLUT1), pyruvate kinase M2 isoform (PKM2), and lactate dehydrogenase (LDHA) after irradiation promoted glycolysis (19, 72, 73), while the expression of

glycogen synthetase 1 (GYS1) was also up-regulated, leading to glycogen accumulation (74). These changes enable cancer cells to survive, proliferate and resist radiation. In addition, pentose phosphate pathway (PPP) after radiation induced more NADPH production, which promoted the production of reduced glutathione (GSH) (19), may lead to radioresistance of NSCLC.

IR can induce the expression of ACSL4, which is a lipid metabolizing enzyme in cancer cells, to increase lipid peroxidation and ferroptosis (75). Intriguingly, IR can also induce the expression of ferroptosis inhibitors SLC7A11 and GPX4, causing radioresistance (75). Perhaps, radioresistant cells have formed a balance between promoting ferroptosis and inhibiting ferroptosis, enabling cells to survive. In radioresistant lung cancer cells, the expression of lipid Droplet (LD) and ferritin heavy chain (FTH1) increased and correlated with each other, which can be targeted and synergistically inhibit tumor radioresistance (76). In addition, the activation of sPLA2-PKC $\delta$ -MAPKs-cPLA2 $\alpha$  pathway promoted tumor progression and radioresistance in NSCLC (77).

ROS levels are elevated in lung cancer radioresistant cells, which exhibit radioresistance by maintaining oxidative stress and NRF2dependent metabolic adaptation (78). The expression of TP53regulated inhibitor of apoptosis 1 (TRIAP1) in NSCLC cell lines A549 and H460 increased after irradiation, and caused upregulation of antioxidant proteins such as thioredoxin-related transmembrane protein (TMX) 1, TMX2, thioredoxin (TXN), glutaredoxin (GLRX) 2, GLRX3, peroxiredoxin (PRDX) 3, PRDX4 and PRDX6, participated in redox metabolism and enhanced the ROS scavenging, leading to radioresistance in NSCLC (79).

In conclusion, NSCLC cells that survived after radiation have altered metabolism that favors cell proliferation and tumor progression. Therefore, targeting tumor metabolism will be an effective measure to improve the radiotherapy prognosis and survival rate of NSCLC patients.

#### 2.2.4 Cell senescence

Studies have shown that induction of premature senescence can radiosensitize NSCLC cells (80). On the contrary, the cytokines secreted by senescent cells which not killed by radiation will affect the adjacent surviving cancer cells in TME, stimulate tumor cell proliferation, invasion and metastasis, escape apoptosis, induce angiogenesis, and promote tumor phenotype (20). In addition, lung cancer cells survived from radiotherapy experienced senescence and entered a dormant state, reprogrammed their biology months or years after irradiation, reactivated stemness, and produced tumors with enhanced growth and metastasis (81).

To sum up, the TME itself is inflammatory, radiation further promotes the release of inflammatory factors, aggravates inflammatory reaction, causes immunosuppression, and hinders anti-tumor immunity. In addition, tumor microbiota can activate inflammatory pathways to promote tumor progression. Moreover, the oxidative stress reaction induced by radiation promotes the occurrence of tumor metabolism reprogramming, causes the depletion of T cells, and makes tumor cells resistant to radiation. The senescent cells in TME can promote tumor proliferation, invasion and metastasis by secreting cytokines. The interaction of the above factors leads to TME remodeling and radioresistance. In addition to these above, tumor hypoxia also play a key role in TME. It is a challenge to overcome radioresistance by targeting TME, because any single targeting action may not achieve the desired therapeutic effect.

## 2.3 Epithelial-mesenchymal transition and radioresistance

Radiotherapy induced EMT and CSCs characteristics, enhanced the invasion and metastasis of cancer cells surviving after primary radiotherapy, leading to acquired resistance of NSCLC, which in turn weakened the radiation-induced tumor killing effect (16, 17, 82). TWIST1 (83, 84) increased after radiation in NSCLC, resulting in downregulation of E-cadherin, overexpression of Snail1, Vimentin, N-cadherin, and PDGFR (85), which promotes cell proliferation, invasion and vascular regeneration, leading to tumor metastasis and radioresistance. PDGF secretion also increased after radiation and resulted in radiation-induced pneumonitis and fibrosis (86), PDGF binds to PDGFR to induce proliferation and vascular regeneration. In addition, Yes-associated protein 1 (YAP1) (87), Snail1 (88), PAK1-LIM domain kinase 1 (LIMK1)-cofilins signaling (89), and zinc-finger E-box-binding homeobox factor 1 (ZEB1) (90)were upregulated after radiotherapy, leading to EMT (Figure 3). Radiation survived NSCLC cells also show cancer stemness properties, such as highly expressed CD166, CD24, CD44, Sox-2, CD133, etc. (76, 85, 91). EMT are critical targets and biomarkers for radiotherapy, radiosensitizers targeting EMT are promising treatment for NSCLC in clinical.

# 3 Mechanisms of inherent radioresistance

Inherent radioresistance is an important factor leading to radiotherapy failure, which is mainly related to the gene mutation status of tumor cells, hypoxia status, distribution of CSCs (28, 92, 93), etc.

## 3.1 Mutation status of oncogenes and radioresistance

Mutations of oncogenes, such as epidermal growth factor receptor (EGFR) (94, 95) and kirsten rat sarcoma viral oncogene (KRAS) (96), or tumor suppressor genes such as Kelch-like ECH-associated protein 1 (KEAP1) (97, 98) and tumor protein P53 (TP53) (99, 100), can cause activation of cell proliferation and resisting cell death signals in NSCLC, leading to radioresistance (Figure 4).

EGFR is overexpressed in 40% to 80% of patients with NSCLC and is associated with poor prognosis (101, 102). Radiation can stimulate wild-type EGFR to enter nucleus and bind with DNA-PK catalytic subunit (DNA-PKcs) to promote DNA repair, thus leading to radioresistance of NSCLC (103, 104). However, the NSCLC with



#### FIGURE 3

Mechanisms of EMT contributes to radioresistance in NSCLC. The surviving cells included enriched CSCs subpopulations that highly expressed CSCs markers (CD24, CD44, CD133, and CD166). miR-30b was upregulated and targeted to FBXL14, activating the transcription factor Twist1, resulting in increased expression of Vimentin, N-cadherin and Fibronectin, and decreased E-cadherin expression. Integrin ß1 is overexpressed after radiotherapy and targets YAP1 to activate ATM/Chk2 signaling, leading to EMT. Furthermore, RAC1 is upregulated in radioresistant NSCLCs, activates the RAC1/PAK1/ LIMK1/Cofilins pathway and the PAK1/Snail pathway, causes EMT and promotes tumor metastasis and invasion. LKB1/SIK1 signaling is downregulated after radiotherapy, prompting activation of the transcription factor ZEB1, leading to EMT and tumor metastasis and invasion. (This figure is adapted from an image created from BioRender.com).



Inherent radioresistance is related to the mutation status of oncogene or tumor suppressor gene in NSCLC. Wildtype EGFR and mutant EGFR both caused cell proliferation and resisting cell death by activation of downstream pathway. In addition, wildtype EGFR can be stimulated by radiation and translocate to nucleus to activate DNA-PKcs, leading to DNA repair. Mutant KRAS can also promote the expression of DNA repair protein RAD51 by activating MYC and improving DNA repair. Mutant p53 will induce anti-apoptosis of NSCLC cells. Mutant KEAP1 cannot inhibit activation of NRF2, leading to the expression of antioxidant proteins to eliminate ROS. (This figure is adapted from an image created from BioRender.com).

mutant EGFR is not necessarily more sensitive to radiation (94), since the mutant EGFR can trigger the downstream pathways, including extracellular signal-regulated kinase (RAS/ERK), p38MAPK, c-Jun Nterminal kinase (JNK), and ERK5, and also activate PI3K/AKT/ mTOR signaling pathway, leading to cell proliferation and antiapoptosis, migration, DNA repair, cell cycle arrest, etc., which in turn increases the radioresistance of NSCLC (22, 23, 103, 105–108).

KRAS can regulate cell growth, differentiation and apoptosis, its mutations mainly occur at codons 12 and 13, and have been found approximately in 20-30% of NSCLC tumor samples (109). KRASmutant NSCLC cells are more radioresistant than KRAS wild-type cells (96, 110) or EGFR-mutant cells (21), because mutant KRAS can upregulate RAD51 expression through oncogene MYC, thereby enhancing DNA damage repair and cell survival (111). In addition, EGFR-dependent chromatin condensation, namely mitotic-like concentrated chromatin (MLCC), can protect KRAS-mutant NSCLC cells from ionizing radiation (IR)-induced DSBs and premature senescence (110), and enhance the expression of CSC marker protein CD133 *via* osteopontin/EGFR pathway, promoting tumor invasion and radioresistance (112).

KEAP1/nuclear factor E2-related factor (NRF2) mutation happens approximately 30% of NSCLC (92), and 7% of patients have co-mutations of KEAP1/NRF2 and EGFR (113). Mutant KEAP1 lost its inhibitory effect on NRF2 in cytoplasm, the activated NRF2 translocated to the nucleus and activated the expression of antioxidant proteins by binding to antioxidant-responsive elements (AREs), resulting in the scavenging of ROS and anti-apoptosis of tumor cells, leading to unlimited proliferation, metastasis and radioresistance of NSCLC (114–116).

TP53 mutation occurs in more than half of human cancers, including NSCLC (92). p53 can not only induce cell cycle arrest, senescence, and apoptosis, but also regulate tumor metabolism, promote ferroptosis, and inhibit tumor development (117). Mutant p53 can neither induce the expression of pro-apoptotic protein Bax, nor inhibit the expression of anti-apoptotic proteins (Bcl-2, Bcl-xl) (118), which makes cancer cells survive in radiotherapy (119). Compared with single mutations in TP53, NSCLC with comutation in KRAS and TP53, or co-mutation in TP53 and KEAP1 are more resistance to IR-induced cell death (100, 114).

In addition to the oncogenic mutation, high levels of inhibitor of apoptosis proteins, such as cIAP1/2, XIAP, and survivin, participate in cell death and survival by binding and inhibiting caspases, leading to radioresistance of NSCLC (120–123). In addition, microRNAs can also affect the radiosensitivity of NSCLC by regulating the cell proliferation and apoptosis. miR-99a and miR-770-5p can induce apoptosis and improve radiosensitivity (124, 125). On the contrary, miR-410 and miR-208a promote radioresistance by inducing EMT and increasing cell proliferation respectively in NSCLC (126, 127).

According to the potential mechanism of inherent radioresistance mentioned above, genome sequencing can be carried out before treatment to determine the mutation type and formulate effective treatment strategies, so as to improve the tumor-killing rate and reduce the tumor recurrence rate after primary radiotherapy of NSCLC.

### 3.2 Cancer stem cells and radioresistance

Cancer stem cells (CSCs) are subpopulations of tumor cells with the ability to self-renew and differentiate into heterogeneous tumor cells, promote tumor growth and metastasis through unlimited proliferation and migration, leading to tumor recurrence and treatment failure (128-131). Antioxidant signaling pathways (NRF2, Wnt, etc.) in CSCs are activated, and more antioxidant proteins are expressed to clear ROS, so as to maintain a low level of ROS in cells, which contributes to the quiescence and self-renewal of CSCs (130, 132-134) (Figure 5). In addition, CSCs have a strong ability to repair DNA damage (135). CD133 is one of the markers of CSCs. As early as 2006, Bao et al. reported that CD133-positive glioma cells showed higher expression of DNA damage checkpoint proteins (i.e. ATM, Rad17, Chk1 and Chk2) than CD133-negative cells (136). In 2014, Desai et al. clarified that CD133-positive NSCLC cells highly express DNA damage repair proteins RAD51 and Exo1, which can promote radioresistance, but had cell type specificity (137). Therefore, low levels of ROS and enhanced DNA damage repair make CSCs insensitive to radiotherapy, leading to tumor relapse (138). CD44 is another CSCs marker, which is related to tumor recurrence and metastasis, as well as prediction of the outcome after radiotherapy (139, 140). The overexpression of CD44 promotes the proliferation of NSCLC, and up-regulates the expression of PD-L1 to promote tumorigenesis, immunosuppression and chemotherapy resistance (141, 142). It is worth noting that due to the low sensitivity of CSCs to IR, the CSCs not killed by radiation will be enriched, which leads to acquired radioresistance. Therefore, the treatment efficiency of NSCLC can be improved by inducing CSCs to differentiate to reduce their proportion in tumor tissues, or by combining drugs targeting key proteins of CSCs antioxidant pathways or surface marker proteins with radiotherapy.

#### 3.3 Tumor hypoxia and radioresistance

As one of the cancer hallmarks, hypoxia induces angiogenesis, tumor invasion, metastasis, and treatment resistance (27, 28). Hypoxic tumor cells are very insensitive to radiation (143). In this article, we mainly introduce the research on HIFs and radioresistance in NSCLC (Figure 6). Hypoxia environment and radiation activate HIFs (144, 145), activated HIF-1 $\alpha$  and its partner subunits HIF-1 $\beta$ enter nucleus and bind to hypoxia-responsive element (HRE) to trigger gene transcription regulating VEGF and glucose metabolism etc., leading to cell survival (146–148). HIF-2 $\alpha$  mainly induces angiogenesis and tumor growth (149–151). Research showed that the upregulation of HIF-1 $\alpha$  enhanced the radioresistance of NSCLC (152). Conversely, the decrease of HIF-1 $\alpha$  in acidic environment caused radioresistance in NSCLC cells and blocking simultaneously of both HIF-1 $\alpha$  and HIF-2 $\alpha$  radiosensitized NSCLC, suggesting the role of HIF-2 $\alpha$  in radioresistance (153). In addition, miR-210 promoted the hypoxic phenotype of NSCLC cells and increased radioresistance by inducing and stabilizing HIF-1 $\alpha$  (154). By contrast, another microRNA, miR-320a targeted HIF-1 $\alpha$  to promote methylation of PTEN, thereby reducing the radioresistance of NSCLC *in vitro* and *in vivo* (155). In conclusion, targeting HIFs may enhance radiosensitivity of NSCLC in clinical, but the pH value should be considered when targeting HIF-1 $\alpha$ .

# 4 Radiosensitizing non-small cell lung cancer

As we discussed above, the mechanisms of radioresistance of NSCLC are complex and not isolated from each other. Whether surgery, chemotherapy, targeted therapy, or immunotherapy, the goal of their combination with radiotherapy is to kill tumor cells, but what cannot be avoided is limited effect, tumor recurrence and metastasis, as well as side effects.

At present, chemotherapy drugs, targeted drugs, immune checkpoint inhibitors, etc. can be used as radiosensizitizers in NSCLC, but these drugs are prone to drug resistance and toxic side effects (11, 30, 156, 157). In addition, most of the current studies about radiosensitizers used in NSCLC is still limited to experimental research, and only a few drugs have been used in clinical. We summarized the drugs or agents for NSCLC radiosensitization in Table 1, including antibody drugs and small molecule inhibitors from non-natural sources. In addition to drug toxicity, radiotherapy itself can also cause lung toxicity in patients, manifesting as different grades of pneumonia, pulmonary fibrosis (32, 183). In addition, the mechanisms of radioresistance are complex, and the efficacy of these single targeted drugs is limited. Therefore, it is an urgent need to develop drugs targeting multiple targets, with synergistic effects and low toxicity in NSCLC radiotherapy.

# 5 Application and advantages of TCM in radiotherapy for NSCLC

As an auxiliary means of radiotherapy for NSCLC, TCM can increase the apoptosis of cancer cells, inhibit tumor metastasis, enhance the anti-tumor immunity of patients, regulate the TME homeostasis, thus improving the efficacy of radiotherapy and reducing the recurrence rate (184, 185), which reflecting the basic principles of treating tumor by reinforcing healthy qi to eliminate pathogenic factors of TCM. Some single herbs are frequently used in the treatment of NSCLC (186). Commonly used TCM for reinforcing healthy qi to eliminate pathogenic factors include Ginseng, Codonopsis pilosula, Astragalus, Angelica, and Polygonatum, etc. Astraglus has immunoregulatory, antioxidant, anti-inflammatory, and anti-cancer activities and combined with chemotherapy to increase the efficacy and reduce the toxicity in patients with advanced NSCLC (187, 188). Our previous research found that Astragalus polysaccharides can reduce the radiation induced



#### FIGURE 5

Mechanisms of radioresistance caused by CSCs in NSCLC. Hallmarks of CSCs such as self-renewal and differentiation, low levels of ROS, quiescence, DNA damage repair, invasion and metastasis ability, and immunosuppression induced by upregulated PD-L1 all contribute to radioresistance of NSCLC. (This figure is adapted from an image created from **BioRender.com**).



#### FIGURE 6

Mechanisms of radioresistance caused by hypoxia in NSCLC. Hypoxia environment of tumor and radiation can activate the expression of HIFs, which will induce the expression of growth factors (VEGF, VEGFR, EGFR etc.) and improved glucose metabolism, leading to cell survival, angiogenesis, and tumor growth of NSCLC. (This figure is adapted from an image created from BioRender.com).

### TABLE 1 Non-natural products radiosensitize NSCLC by targeting key proteins.

Mechanism of action	Targets	Radiosensitizing modulators	Experimental models/( <i>in vitro</i> or <i>in</i> <i>vivo</i> or clinical)	Conclusion	Reference
Targeting DNA repair	PARP	Olaparib+ML216	In vitro and in vivo	PARP inhibitors and BLM helicase inhibitors synergistically radiosensitized NSCLC.	(158)
	PARP	niraparib	In vitro and in vivo	Niraparib activated anti-tumor CD8+T cells in the tumor microenvironment through the STING/TBK1/IRF3 pathway in EGFR-mutant NSCLC.	(159)
	АТМ	KU55933	In vitro	ATM inhibitors radiosensitized NSCLC by inhibiting IR-induced EGFR activation and can be used in EGFR-resistant tumor cells for radiotherapy.	(160)
	DNA-PK	AZD7648	In vitro and in vivo	DNA-PK inhibitors are promising radiosensitizers for treatment of NSCLC.	(161)
	Telomerase	BIBR1532	In vitro and in vivo	Inhibiting telomerase function can radiosensitize NSCLC.	(162)
	AND-1	CH-3 compound	In vitro and in vivo	Inhibition of AND-1 will be a promising strategy for enhancing radiosensitivity in NSCLC.	(42)
	RUVBL1/2	Compound B	In vitro and in vivo	RUVBL1/2 inhibition combined with radiotherapy may be effective treatment for NSCLC.	(51)
Targeting cell cycle	BET	JQ1	In vitro and in vivo	BET inhibition may be a potential treatment for radiosensitizing NSCLC.	(163)
	CDK4/6	Palbociclib/	In vitro	Combining inhibition of CDK4/6 and radiotherapy will be promising strategy for patients with p53-wild type NSCLC.	(164)
	CDK4/6	Abemaciclib	In vitro and in vivo	Abemacilcib combined with radiotherapy radiosensitized NSCLC in preclinical models.	(165)
Targeting inflammation	ΙΚΚβ	IMD03544	In vitro and/or in vivo	Blockade of IKK-NF-κB signaling will be benefit for radiotherapy in NSCLC.	(60, 166)
Targeting immune suppression	TGF-β	LY364947	In vitro and in vivo	TGF- $\beta$ inhibition combined with radiotherapy enhanced therapeutic efficacy in NSCLC.	(167)
	STAT3	Niclosamide	In vitro and in vivo	Inhibition of STAT3 signaling will be effective treatment for NSCLC to radiotherapy.	(59)
	JAK2	TG101209	In vitro and in vivo	Targeting JAK2 may enhance radiotherapy response in KRAS- mutant NSCLC.	(168)
	PD-L1	anti-PD-L1 antibody	In vitro and in vivo	Anti-PD-L1 antibody combined with radiotherapy synergistically enhanced anti-tumor immunity in NSCLC.	(169)
	PD-1	Nivolumab	Clinical phase II (Enrollment: 65)	No published results to date	(170)
	PD-1	pembrolizumab	Clinical phase I/II (Enrollment: 104)	Pembrolizumab combined with radiotherapy may be effective in certain subgroups of NSCLC patients with low PD-L1 expression	(171)
	CTLA-4	Ipilimumab	Clinical phase I/II (Enrollment: 39)	CTLA-4 blockade and radiotherapy synergistically induced robust antitumor T cell responses	(172)
Targeting glutamine metabolism	Glutaminase	CB-839	Clinical phase I/II (Enrollment: unknown)	KEAP1/NFE2L2 mutations is associated with radioresistance and local recurrence, and glutaminase inhibition enhanced radiotherapeutic effects in NSCLC.	(97)
Targeting cell proliferation	EGFR	Erlotinib, gefitinib	Clinical phase II (Enrollment: 10)	EGFR-TKI combined with radiotherapy achieves long-term control of stage IV NSCLC with EGFR mutations.	(173)
	EGFR/ HER2/ HER3	Pan-HER	In vitro and in vivo	Pan-HER exerted anti-proliferation and tumor growth-delay effect by inhibiting multiple HER members to enhance radiation response in NSCLC.	(174)
	ERK5	XMD8-92/ERK5 knockdown	In vitro and in vivo	ERK5 contributes to lung cancer development and radioresistance and can be a novel target for radiotherapy in NSCLC.	(23)

(Continued)

### TABLE 1 Continued

Mechanism of action	Targets	Radiosensitizing modulators	Experimental models/( <i>in vitro</i> or <i>in</i> <i>vivo</i> or clinical)	Conclusion	Reference
	EGFR and Norch	Anti-EGFR/Notch CrossMab (CT16)	In vitro and in vivo	Combined blocking of EGFR and Notch signaling enhanced radiation response in NSCLC.	(175)
	EGFR and COX2	Afatinib+celecoxib	In vitro	Combined inhibition of EGFR and COX-2 radiosensitized NSCLC.	(176)
Targeting cell apoptosis	mTOR	miR-99a	In vitro and in vivo	MiR-99a enhanced radiosensitivity <i>via</i> targeting mTOR in NSCLC.	(124)
	miR-410		In vitro and in vivo	MiR-410 increased radioresistance <i>via</i> targeting PTEN/PI3K/ mTOR pathway in NSCLC.	(126)
	Bcl-XL	BXI-61, BXI-72	In vitro and in vivo	Small molecules targeting Bcl-XL conquered acquired radioresistance in NSCLC.	(177)
	Anti- apoptotic Bcl-2 family	ABT-737	In vitro and in vivo	ABT-737 radiosensitized K-ras mutant NSCLC by inhibiting the activity of Bcl-2 family proteins.	(178)
	Bak	BKA-073	In vitro and in vivo	Bak activators may be promising anticancer drugs in NSCLC treatment.	(179)
	Bcl-2/Bcl-xL	ABT-263	In vitro and in vivo	Inhibiting anti-apoptotic Bcl-2 family proteins may enhance radiosensitivity by reducing hypoxia-mediated anti-apoptosis in NSCLC.	(180)
	cIAP1/2	Birinapant	In vitro	cIAP1/2 will be promising targets for enhancing radiosensitivity of NSCLC.	(123)
Targeting angiogenesis	VEGFR2	Apatinib	In vivo	Apatinib can radiosensitize lung cancer through vascular normalization and hypoxia reduction in the xenograft model	(181)
	VEGF	Endostar	Clinical phage II (Enrollment: 67)	Continuous intravenous endurance combined with concurrent etoposide and radiation prolongs OS in patients with unresectable stage III NSCLC.	(182)

TABLE 2 Natural compound/traditional Chinese medicine used in radiotherapy of NSCLC.

Natural com- pound/Tradi- tional Chinese medicine	Type of drug	Experimental models /( <i>in</i> <i>vitro</i> or <i>in vivo</i> or clinical)	Function	Reference
Aidi injection	Compound prescription, composed of extracts from Mylabri, Ginseng Astragalus, and Acanthopanacis senticosi.	Clinical	Heat-clearing and toxin-removing, eliminating stasis and resolve masses. Aidi injection improved PSF and immune function in lung cancer patients and alleviated the myelosuppression, radiation pneumonitis, radiation esophagitis.	(197, 198)
Shengqi Fuzheng injection	Compound prescription, composed of extracts from Codonopsis and Astragalus.	Clinical	Supplement and tonify lung qi. Reduced the inflammatory response and improved immune function by regulating the expression of TNF- $\alpha$ and TGF- $\beta$ .	(199)
Compound kushen injection	Compound prescription, composed of extracts from Sophorae and Rhizoma Smilacis Glabrae.	Clinical	Clear heat and drain dampness, resolve masses and alleviate pain. Improved efficacy and reduced radiation pneumonitis, radiation esophagitis, and myelosuppression.	(200)
Kangai injection	Compound prescription, composed of extracts from Astragalus, Ginseng, and Oxymatrine.	In vitro	Increased FOXO3a-mediated apoptosis and autophagic cell death in cisplatin-resistance A549 cells.	(201)
Astragalus	Monomer herb	Meta-analysis	Reduced the toxicity and increased the efficacy of radiotherapy.	(202)
Elemene injection	Herbal extractions from Curcuma	Meta-analysis	Reduce adverse reactions, relieve symptoms.	(203)
<i>Ganoderma</i> <i>lucidum</i> polysaccharide	Herbal extractions from Ganoderm	In vitro and in vivo	Decreased the tumor immune suppression.	(204)

(Continued)

TABLE 2 Continued
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Natural com- pound/Tradi- tional Chinese medicine	Type of drug	Experimental models /(in vitro or in vivo or clinical)	Function	Reference
Kanglaite injection (KLTi)	Herbal extractions from Coix agrestis Lour	Clinical	KLTi in combination with radiotherapy improved patient survival but can induce adverse effects such as leukopenia, nausea and vomiting.	(205)
<i>Astragalus</i> polysaccharide	Herbal extractions from Astragalus.	In vitro	Reduced the radiation-induced by stander effects through regulation of MAPK/NF- $\kappa$ B signaling pathway, or TGF- $\beta$ R/MAPK/ROS pathway.	(189–191)
Micheliolide	Small molecule derived from Michelia.	In vitro	Micheliolide enhanced radiosensitivity via inducing the ubiquitination degradation of HIF-1 $\alpha$ in p53-null NSCLC.	(206)
Quercetin	Small molecule, derived from a variety of TCM such as Bupleurum, Hypericum, Astragalus, etc.	In vitro	Inhibited WEE1 signaling	(54)
Daurinol	Small molecule, derived from Haplophyllum.	In vitro	Suppressed serine/threonine family of aurora kinases A/B(AURKA/ AURKB) can radiosensitize NSCLC.	(207)
Isorhamnetin	Small molecule, derived from a variety of TCM such as Folium Ginkgo, Astragalus, etc.	In vitro	Upregulated IL-13, suppressed NF-κB signaling and increased cell apoptosis.	(208)
Cucurbitacin I	Small molecule, mainly distributed in Cucurbitaceae.	In vitro and in vivo	Inhibition of STAT3 signaling in CD133- positive NSCLC may enhance radiotherapy effect.	(209)
Phoyunnanin E	Small molecule, isolated from Dendrobium venustum.	In vitro	Induce lung cancer cell apoptosis via inhibiting survivin.	(210)
Celastrol	Small molecule, derived from Celeastrus.	In vitro	Celastrol radiosensitized NSCLC by suppressing the ATP-binding activity of Hsp90.	(211)
Curcumin	Small molecule, isolated from <i>Curcuma longa</i> L.	In vitro and in vivo	Increasing cell apoptosis <i>via</i> activating the p53-miR-192-5p/215-XIAP pathway.	(212)
Scutellarin Small molecule, isolated from Scutellaria altissima L. or S.baicalensis Georgi.		In vitro and in vivo	Promoted cell apoptosis <i>via</i> down-regulating AKT/mTOR pathway.	(213)

bystander effects (RIBE) of bone marrow mesenchymal stem cells (BMSCs) caused by X-rays and heavy ions, and reduce the toxicity of normal cells and tissues in lung cancer radiotherapy (189-191). In addition, our research also found that Guiqi Baizhu Decoction, which composed of Astragalus, Angelica, Atractylodes, Paeonia lactiflora, Tangerine peel, Rhubarb, and Licorice, can significantly reduce the radiation inflammatory reaction and immune damage, and prevent intestinal microbial imbalance and metabolic disorders caused by radiation (192). We further found that Guiqi Baizhu Decoction can regulate HIF-1a, AQP4 and Na+/K+-ATPase to reduce hypoxia and oxidative stress, thus treating radiation-induced intestinal edema (193). Moreover, in the study of gastric cancer, we used chemical informatics and cell experiments to verify that Guiqi Baizhu decoction plays a dual role of anti-tumor and immunoregulation by targeting HER2 and PD-L1 through its active components quercetin and isorhamnetin respectively, reflecting the mechanism of multipoint and synergistic effect of TCM in tumor treatment (194).

In recent decades, the advantages of natural products in antitumor have become more and more obvious, such as paclitaxel (195) and vinorelbine (196) has been used in clinical for decades. Here we listed the application of TCM compound prescription, single herb, herbal extracts, and small molecules from TCM in the preclinical and clinical research of radiotherapy toxicity reduction and efficiency enhancement for NSCLC (Table 2). The various factors that cause radioresistance are not independent, nor only appear in a certain period of time. Moreover, at different stages of radiotherapy, the mechanism that dominates radioresistance is variable and complex. However, TCM can adjust the radiation anti-tumor effect in different ways with the advantage of multi-component and multi-target, improve the efficacy of radiotherapy, reduce the toxicity of radiotherapy and improve the quality of life of patients.

## 6 Conclusions

In conclusion, the radioresistance mechanisms of NSCLC are complex. Therefore, prior to radiotherapy gene sequencing should be performed for NSCLC patients to determine the type of mutation, or abnormally expressed genes, proteins, and signaling pathways, thus to develop more effective treatment strategies. In the early stage of radiotherapy, it is suggested to combine EGFR, KRAS and other mutant targeted drugs, drugs to improve DNA damage, immune checkpoint drugs or drugs to target hypoxia. In the middle and late stage of radiotherapy, it is suggested that the combination of drugs targeting EMT or regulating TME can improve the curative effect. TCM can run through the whole radiotherapy process. However, the specific mechanism of TCM in the sensitization of NSCLC radiotherapy needs to be further explored. The combination of effective small molecules extracted from TCM and radiotherapy may provide a new application prospect for the future clinical treatment of NSCLC.

## Author contributions

TZ is responsible for literature review and article writing. LZ and JH are responsible for revising articles. ZM, YYL, YZ, and ZL are responsible for collecting and sorting materials. SZ, YC, and GZ are responsible for sorting charts. YQL provides ideas and guidance for the article. All authors contributed to the article and approved the submitted version.

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## References

1. Koh PK, Faivre-Finn C, Blackhall FH, De Ruysscher D. Targeted agents in nonsmall cell lung cancer (NSCLC): Clinical developments and rationale for the combination with thoracic radiotherapy. *Cancer Treat Rev* (2012) 38(6):626–40. doi: 10.1016/ j.ctrv.2011.11.003

2. Wirsdörfer F, de Leve S, Jendrossek V. Combining radiotherapy and immunotherapy in lung cancer: Can we expect limitations due to altered normal tissue toxicity? *Int J Mol Sci* (2019) 20(1):24. doi: 10.3390/ijms20010024

3. Sears CR, Cooney SA, Chin-Sinex H, Mendonca MS, Turchi JJ. DNA Damage response (DDR) pathway engagement in cisplatin radiosensitization of non-small cell lung cancer. DNA Repair (Amst). (2016) 40:35–46. doi: 10.1016/j.dnarep.2016.02.004

4. Provencio M, Sánchez A. Therapeutic integration of new molecule-targeted therapies with radiotherapy in lung cancer. *Transl Lung Cancer Res* (2014) 3(2):89–94. doi: 10.3978/j.issn.2218-6751.2014.03.06

5. Hirsch FR, Scagliotti GV, Mulshine JL, Kwon R, Curran WJ, Wu YL, et al. Lung cancer: current therapies and new targeted treatments. *Lancet.* (2017) 389(10066):299–311. doi: 10.1016/S0140-6736(16)30958-8

6. Trakul N, Harris JP, Quynh-Thu Le, WY H, PG M, BW L, et al. Stereotactic ablative radiotherapy for reirradiation of locally recurrent lung tumors. *J Thorac Oncol* (2012) 7 (9):1462–5. doi: 10.1097/JTO.0b013e31825f22ce

7. Iyengar P, Zhang VE, Court L, Westover K, Yan YL, Lin MH, et al. Accelerated hypofractionated image-guided vs conventional radiotherapy for patients with stage II/III non-small cell lung cancer and poor performance status: A randomized clinical trial. *JAMA Oncol* (2021) 7(10):1497–505. doi: 10.1001/jamaoncol.2021.3186

8. Senthi S, Lagerwaard FJ, Haasbeek Cornelis JA, Slotman BJ, Senan S. Patterns of disease recurrence after stereotactic ablative radiotherapy for early stage non-small-cell lung cancer: a retrospective analysis. *Lancet Oncol* (2012) 13(8):802–9. doi: 10.1016/S1470-2045(12)70242-5

9. Sato K, Shimokawa T, Imai T. Difference in acquired radioresistance induction between repeated photon and particle irradiation. *Front Oncol* (2019) 9:1213. doi: 10.3389/fonc.2019.01213

10. Catton CN, Shultz DB. Should we expand the carbon ion footprint of prostate cancer? Lancet Oncol (2019) 20(5):608–9. doi: 10.1016/S1470-2045(19)30094-4

11. Bradley JD, Hu C, Komaki RR, Masters GA, Blumenschein GR, Schild SE, et al. Long-term results of NRG oncology RTOG 0617: Standard- versus high-dose chemoradiotherapy with or without cetuximab for unresectable stage III non-small-cell lung cancer. *J Clin Oncol* (2020) 38(7):706–14. doi: 10.1200/JCO.19.01162

12. Buckley AM, Lynam-Lennon N, O'Neill H, O'Sullivan J. Targeting hallmarks of cancer to enhance radiosensitivity in gastrointestinal cancers. *Nat Rev Gastroenterol Hepatol* (2020) 17(5):298–313.

13. Lee SY, Jeong EK, Ju MK, Jeon HM, Kim MY, Kim CH, et al. Induction of metastasis, cancer stem cell phenotype, and oncogenic metabolism in cancer cells by ionizing radiation. *Mol Cance*. (2017) 16(1):10. doi: 10.1186/s12943-016-0577-4

14. Alhaddad L, Pustovalova M, Blokhina T, Chuprov-Netochin R, Osipov AN, Leonov S. IR-surviving NSCLC cells exhibit different patterns ofmolecular and cellular reactions relating to the multifraction irradiation regimen and p53-family proteins expression. *Cancers (Basel)* (2021) 13(11):2669. doi: 10.3390/cancers13112669

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## **Conflict of interest**

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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15. McKelvey KJ, Hudson AL, Back M, Eade T, Diakos CI. Radiation, inflammation and the immune response in cancer. *Mamm Genome*. (2018) 29(11-12):843-65. doi: 10.1007/s00335-018-9777-0

16. Nantajit D, Dong L, Jian JL. The network of epithelial-mesenchymal transition: potential new targets for tumor resistance. *J Cancer Res Clin* (2015) 141(10):1697–713. doi: 10.1007/s00432-014-1840-y

17. Marie-Egyptienne DT, Lohse I, Hill RP. Cancer stem cells, the epithelial to mesenchymal transition (EMT) and radioresistance: Potential role of hypoxia. *Cancer Lett* (2013) 341(1):63–72. doi: 10.1016/j.canlet.2012.11.019

18. Barker HE, Paget JT, Khan AA, Harrington KJ. The tumour microenvironment after radiotherapy: Mechanisms of resistance and recurrence. *Nat Rev Cancer*. (2015) 15 (7):409–25. doi: 10.1038/nrc3958

19. Mittal A, Nenwani M, Sarangi I, Achreja A, Lawrence TS, Nagrath D. Radiotherapy-induced metabolic hallmarks in the tumor microenvironment. *Trends Cancer*. (2022) 8(10):1–15. doi: 10.1016/j.trecan.2022.05.005

20. Hanahan D. Hallmarks of cancer: New dimensions. *Cancer Discovery* (2022) 12 (1):31–46. doi: 10.1158/2159-8290.CD-21-1059

21. Johung KL, Yao XP, Li FY, Yu JB, Gettinger SN, Goldberg S, et al. A clinical model for identifying radiosensitive tumor genotypes in non-small cell lung cancer. *Clin Cancer Res* (2013) 19(19):5523–32.

22. Sad K, Parashar P, Tripathi P, Hungyo H, Sistla R, Soni R, et al. Prochlorperazine enhances radiosensitivity of non-small cell lung carcinoma by stabilizing GDP-bound mutant KRAS conformation. *Free Radic Biol Med* (2021) 177:299–312. doi: 10.1016/j.freeradbiomed.2021.11.001

23. Jiang WW, Jin GH, Cai FF, Chen X, Cao NN, Zhang XY, et al. Extracellular signalregulated kinase 5 increases radioresistance of lung cancer cells by enhancing the DNA damage response. *Exp Mol Med* (2019) 51(2):1–20. doi: 10.1038/s12276-019-0209-3

24. Hanahan D, Weinberg RA. Hallmarks of cancer: The next generation. *Cell.* (2011) 144(5):646–74. doi: 10.1016/j.cell.2011.02.013

25. Haura EB, Cress WD, Chellappan S, Zheng Z, Bepler G. Antiapoptotic signaling pathways in non-small-cell lung cancer: Biology and therapeutic strategies. *Clin Lung Cancer.* (2004) 6(2):113–22. doi: 10.3816/CLC.2004.n.025

26. Olivares-Urbano MA, Griñán-Lisón C, Marchal JA, Núñez MI. CSC radioresistance: A therapeutic challenge to improve radiotherapy effectiveness in cancer. *Cells.* (2020) 9(7):1651. doi: 10.3390/cells9071651

27. Walsh JC, Lebedev A, Aten E, Madsen K, Marciano L, Kolb HC. The clinical importance of assessing tumor hypoxia: Relationship of tumor hypoxia to prognosis and therapeutic opportunities. *Antioxid Redox Signal* (2014) 21(10):1516–54. doi: 10.1089/ars.2013.5378

28. Salem A, Asselin MC, Reymen B, Jackson A, Lambin P, West CML, et al. Targeting hypoxia to improvenon-small cell lung cancer outcome. *J Natl Cancer Inst* (2018) 110(1): djx160. doi: 10.1093/jnci/djx160

29. Jansen J, Vieten P, Pagliari F, Hanley R, Marafioti MG, Tirinato L, et al. A novel analysis method for evaluating the interplay of oxygen and ionizing radiation at the gene level. *Front Genet* (2021) 12:597635. doi: 10.3389/fgene.2021.597635

30. Wang H, Mu XY, He H, Zhang XD. Cancer radiosensitizers. *Trends Pharmacol Sci* (2018) 39(1):24–48. doi: 10.1016/j.tips.2017.11.003

31. Havaki S, Kotsinas A, Chronopoulos E, Kletsas D, Georgakilas A, Gorgoulis VG. The role of oxidative DNA damage in radiation induced bystander effect. *Cancer Lett* (2015) 356(1):43–51. doi: 10.1016/j.canlet.2014.01.023

32. Barker HE, Paget JT, Khan AA, Harrington KJ. The tumour microenvironment after radiotherapy: Mechanisms of resistance and recurrence. *Nat Rev Cancer*. (2015) 15 (7):409–25. doi: 10.1038/nrc3958

33. Stingele J, Bellelli R, Boulton SJ. Mechanisms of DNA-protein crosslink repair. Nat Rev Mol Cell Biol (2017) 18(9):563–73. doi: 10.1038/nrm.2017.56

34. Smith J, Tho LM, Xu N, Gillespie DA. The ATM-Chk2 and ATR-Chk1 pathways in DNA damage signaling and cancer. *Adv Cancer Res* (2010) 108:73–112. doi: 10.1016/B978-0-12-380888-2.00003-0

35. Syed A, Tainer JA. The MRE11-RAD50-NBS1 complex conducts the orchestration of damage signaling and outcomes to stress in DNA replication and repair. *Annu Rev Biochem* (2018) 87:263–94. doi: 10.1146/annurev-biochem-062917-012415

36. Zou N, Xie GZ, Cui TT, Srivastava AK, Qu MH, Yang LL, et al. DDB2 increases radioresistance of NSCLC cells by enhancing DNA damage responses. *Tumor Biol* (2016) 37(10):14183–91. doi: 10.1007/s13277-016-5203-y

37. Choi SH, Yang H, Lee SH, Ki JH, Nam DH, Yoo HY. TopBP1 and claspin contribute to the radioresistance of lung cancer brain metastases. *Mol Cancer.* (2014) 13:211. doi: 10.1186/1476-4598-13-211

38. Burma S, Chen BP, Murphy M, Kurimasa A, Chen DJ. ATM Phosphorylates histone H2AX in response to DNA double-strand breaks. *J Biol Chem* (2001) 276:42462–7. doi: 10.1074/jbc.C100466200

39. Paull TT. Mechanisms of ATM activation. Annu Rev Biochem (2015) 84:711-38. doi: 10.1146/annurev-biochem-060614-034335

40. Torok JA, Oh P, Castle KD, Reinsvold M, Ma Y, Luo LX, et al. Deletion of atm in tumor but not endothelial cells improves radiation response in a primary mouse model of lung adenocarcinoma. *Cancer Res* (2019) 79(4):773–82. doi: 10.1158/0008-5472.CAN-17-3103

41. Koll TT, Feis SS, Wright MH, Teniola MM, Richardson MM, Robles AI, et al. HSP90 inhibitor, DMAG, synergizes with radiation of lung cancer cells by interfering with base excision and ATM-mediated DNA repair. *Mol Cancer Ther* (2008) 7(7):1985–92. doi: 10.1158/1535-7163.MCT-07-2104

42. Gou W, Yu X, Wu S, Wu H, Chang H, Chen L, et al. Targeted inhibition of acidic nucleoplasmic DNA-binding protein 1 enhances radiosensitivity of non-small cell lung cancer. *Cancer Lett* (2022) 530:100–9. doi: 10.1016/j.canlet.2022.01.020

43. Cao K, Chen Y, Zhao S, Huang Y, Liu T, Liu H, et al. Sirt3 promoted DNA damage repair and radioresistance through ATM-Chk2 in non-small cell lung cancer cells. J Cancer. (2021) 12(18):5464–72. doi: 10.7150/jca.53173

44. Zeng Y, Jie X, Wu B, Wu G, Liu L, Xu S. IQGAP3 interacts with Rad17 to recruit the Mre11-Rad50-Nbs1 complex and contributes to radioresistance in lung cancer. *Cancer Lett* (2020) 493:254–65. doi: 10.1016/j.canlet.2020.08.042

45. Zhang J, Wu Q, Zhu L, Xie S, Tu L, Yang Y, et al. SERPINE2/PN-1 regulates the DNA damage response and radioresistance by activating ATM in lung cancer. *Cancer Lett* (2022) 524:268–83. doi: 10.1016/j.canlet.2021.10.001

46. Lei X, Du L, Zhang P, Ma N, Liang Y, Han Y, et al. Knockdown GTSE1 enhances radiosensitivity in non-small-cell lung cancer through DNA damage repair pathway. *J Cell Mol Med* (2020) 24(9):5162–7. doi: 10.1111/jcmm.15165

47. Cheng C, Pei X, Li SW, Yang J, Li C, Tang J, et al. CRISPR/Cas9 library screening uncovered methylated PKP2 as a critical driver of lung cancer radioresistance by stabilizing  $\beta$ -catenin. *Oncogene*. (2021) 40(16):2842–57. doi: 10.1038/s41388-021-01692-x

48. Kaminskyy VO, Hååg P, Novak M, Végvári Á, Arapi V, Lewensohn R, et al. EPHA2 interacts with DNA-PK in cell nucleus and controls ionizing radiation responses in non-small cell lung cancer cells. *Cancers (Basel).* (2021) 13(5):1010. doi: 10.3390/ cancers13051010

49. Gong S, Li Y, Lv L, Men W. Restored microRNA-519a enhances the radiosensitivity of non-small cell lung cancer *via* suppressing EphA2. *Gene Ther* (2021) 29:1–13. doi: 10.1038/s41434-020-00213-x

50. Yang X, Wang G, You J, Gu R, Xu X, Xu C, et al. High expression of cancer-IgG is associated with poor prognosis and radioresistance *via* PI3K/AKT/DNA-PKcs pathway regulation in lung adenocarcinoma. *Front Oncol* (2021) 11:675397. doi: 10.3389/ fonc.2021.675397

51. Yenerall P, Das AK, Wang S, Kollipara RK, Li LS, Villalobos P, et al. RUVBL1/ RUVBL2 ATPase activity drives PAQosome maturation, DNA replication and radioresistance in lung cancer. *Cell Chem Biol* (2020) 27(1):105–121.e14. doi: 10.1016/ j.chembiol.2019.12.005

52. Aarts M, Sharpe R, Garcia-Murillas I, Gevensleben H, Hurd MS, Shumway SD, et al. Forced mitotic entry of s-phase cells as a therapeutic strategy induced by inhibition of WEE1. *Cancer Discovery* (2012) 2(6):524–39. doi: 10.1158/2159-8290.CD-11-0320

53. McCarthy N. Cell cycle: A WEE pointer. Nat Rev Cancer. (2012) 12(6):378-9. doi: 10.1038/nrc3286

54. Wang Q, Chen Y, Lu H, Wang H, Feng H, Xu J, et al. Quercetin radiosensitizes non-small cell lung cancer cells through the regulation of miR-16-5p/WEE1 axis. *IUBMB Life* (2020) 72(5):1012–22. doi: 10.1002/iub.2242

55. Aggarwal BB, Sung B. NF-κB in cancer: a matter of life and death. *Cancer Discovery* (2011) 1(6):469–71. doi: 10.1158/2159-8290.cd-11-0260

56. Sergei IG, Florian R, Michael K. Immunity, inflammation, and cancer. *Cell.* (2010) 140(6):883–99. doi: 10.1016/j.cell.2010.01.025

57. Bai M, Ma X, Li X, Wang X, Mei Q, Li X, et al. The accomplices of NF- $\kappa$ B lead to radioresistance. Curr Protein Pept Sci (2015) 16:279–94. doi: 10.2174/138920371604150429152328

58. Harada D, Takigawa N, Kiura K. The role of STAT3 in non-small cell lung cancer. Cancers (Basel) (2014) 6(2):708–22. doi: 10.3390/cancers6020708

59. You S, Li R, Park D, Xie M, Sica GL, Cao Y, et al. Disruption of STAT3 by niclosamide reverses radioresistance of human lung cancer. *Mol Cancer Ther* (2014) 13 (3):606–16. doi: 10.1158/1535-7163.MCT-13-0608

60. Tsolou A, Liousia M, Kalamida D, Pouliliou S, Giatromanolaki A, Koukourakis M. Inhibition of IKK-NFκB pathway sensitizes lung cancer cell lines to radiation. *Cancer Biol Med* (2017) 14(3):293–301. doi: 10.20892/j.issn.2095-3941.2017.0049

61. Zhi-Wei, L, Yi-Ming, Z, Li-Ying, Z, Ting, Z, Yang-Yang, Li, Gu-Cheng, Z, et al. Duality of interactions between TGF- $\beta$  and TNF- $\alpha$  during tumor formation. Front Immunol (2021) 12:810286(undefined). doi: 10.3389/fimmu.2021.810286

62. Yi-Ming. Z, Li-Ying. Z, Yang-Yang. Li, Heng. Z, Zhi-Ming. M, Zhi-Wei. L, et al. Radiation-induced bystander effect on the genome of bone marrow mesenchymal stem cells in lung cancer. *Antioxid Redox Signal* (2022). doi: 10.1089/ars.2022.0072. undefined (undefined), undefined. doi: 10.1089/ars.2022.0072

63. He K, Barsoumian HB, Hu Y, Sezen D, MD W, et al. Inhibition of STAT6 with antisense oligonucleotides enhances the systemic antitumor effects of radiotherapy and anti-PD1 in metastatic non-small cell lung cancer. *Cancer Immunol Res* (2023). undefined (undefined), undefined. doi: 10.1158/2326-6066.CIR-22-0547

64. Wang J, Han Q, Liu H, Luo H, Li L, Liu A, et al. Identification of radiotherapyassociated genes in lung adenocarcinoma by an integrated bioinformatics analysis approach. *Front Mol Biosci* (2021) 8:624575. doi: 10.3389/fmolb.2021.624575

65. Zhang F, Sang Y, Chen D, Wu X, Wang X, Yang W, et al. M2 macrophage-derived exosomal long non-coding RNA AGAP2-AS1 enhances radiotherapy immunity in lung cancer by reducing microRNA-296 and elevating NOTCH2. *Cell Death Dis* (2021) 12 (5):467. doi: 10.1038/s41419-021-03700-0

66. Zitvogel L, Kroemer G. Lower airway dysbiosis exacerbates lung cancer. Cancer Discovery (2021) 11(2):224–6. doi: 10.1158/2159-8290.CD-20-1641

67. Pinato DJ, Howlett S, Ottaviani D, Urus H, Patel A, Mineo T, et al. Association of prior antibiotic treatment with survival and response to immune checkpoint inhibitor therapy in patients with cancer. *JAMA Oncol* (2020) 6(2):302. doi: 10.1001/jamaoncol.2019.6921

68. Li. Y, Aitian. Li, Ying. W, Yi. Z. Intratumoral microbiota: roles in cancer initiation, development and therapeutic efficacy. *Signal Transduct Target Ther* (2023) 8(1):35. doi: 10.1038/s41392-022-01304-4

 G. Jin C, Lagoudas GK, Zhao C, Bullman S, Bhutkar A, Hu B, et al. Commensal microbiota promote lung cancer development via γδ T cells. Cell. (2019) 176(5):998–1013. doi: 10.1016/j.cell.2018.12.040

70. Xiang L, Meng X. Emerging cellular and molecular interactions between the lung microbiota and lung diseases. *Crit Rev Microbiol* (2022) 48(5):577–610. doi: 10.1080/1040841X.2021.1992345

71. Tsay JC, Wu BG, Sulaiman I, Gershner K, Schluger R, Li Y, et al. Lower airway dysbiosis affects lung cancer progression. *Cancer Discovery* (2021) 11(2):293–307. doi: 10.1158/2159-8290.CD-20-0263

72. Yang Y, Chong Y, Chen M, Dai W, Zhou X, Ji Y, et al. Targeting lactate dehydrogenase a improves radiotherapy efficacy in non-small cell lung cancer: From bedside to bench. J Transl Med (2021) 19(1):170. doi: 10.1186/s12967-021-02825-2

73. Meng MB, Wang HH, Guo WH, Wu ZQ, Zeng XL, Zaorsky NG, et al. Targeting pyruvate kinase M2 contributes to radiosensitivity of non-small cell lung cancer cells *in vitro* and *in vivo*. *Cancer Lett* (2015) 356:985–93. doi: 10.1016/j.canlet.2014.11.016

74. Tsolou A, Koparanis D, Lamprou I, Giatromanolaki A, Koukourakis MI. Increased glucose influx and glycogenesis in lung cancer cells surviving after irradiation. *Int J Radiat Biol* (2022) 1-10. doi: 10.1080/09553002.2022.2113837

75. Lei G, Zhang Y, Koppula P, Liu X, Zhang J, Lin SH, et al. The role of ferroptosis in ionizing radiation-induced cell death and tumor suppression. *Cell Res* (2020) 30(2):146–62. doi: 10.1038/s41422-019-0263-3

76. Tirinato L, Marafioti MG, Pagliari F, Jansen J, Aversa I, Hanley R, et al. Lipid droplets and ferritin heavy chain: A devilish liaison in human cancer cell radioresistance. *Elife.* (2021) 10:e72943. doi: 10.7554/eLife.72943

77. Lee S, Kim D, Kang J, Kim E, Kim W, Youn H, et al. Surfactant protein b suppresses lung cancer progression by inhibiting secretory phospholipase A2 activity and arachidonic acid production. *Cell Physiol Biochem* (2017) 42(4):1684–700. doi: 10.1159/000479418

78. Singh B, Patwardhan RS, Jayakumar S, Sharma D, Sandur SK. Oxidative stress associated metabolic adaptations regulate radioresistance in human lung cancer cells. J Photochem Photobiol B (2020) 213:112080. doi: 10.1016/j.jphotobiol.2020.112080

79. Hao CC, Luo JN, Xu CY, Zhao XY, Zhong ZB, Hu XN, et al. TRIAP1 knockdown sensitizes non-small cell lung cancer to ionizing radiation by disrupting redox homeostasis. *Thorac Cancer.* (2020) 11(4):1015–25. doi: 10.1111/1759-7714.13358

80. Luo H, Wang L, Schulte BA, Yang A, Tang S, Wang GY. Resveratrol enhances ionizing radiation-induced premature senescence in lung cancer cells. *Int J Oncol* (2013) 43(6):1999–2006. doi: 10.3892/ijo.2013.2141

81. Tsolou A, Lamprou I, Fortosi AO, Liousia M, Giatromanolaki A, Koukourakis MI. 'Stemness' and 'senescence' related escape pathways are dose dependent in lung cancer cells surviving post irradiation. *Life Sci* (2019) 232:116562. doi: 10.1016/j.lfs.2019.116562

82. Qiao L, Chen Y, Liang N, Xie J, Deng G, Chen F, et al. Targeting epithelial-tomesenchymal transition in radioresistance: crosslinked mechanisms and strategies. *Front Oncol* (2022) 12:775238. doi: 10.3389/fonc.2022.775238 83. Eckert MA, Lwin TM, Chang AT, Kim J, Danis E, Ohno-Machado L, et al. Twist1induced invadopodia formation promotes tumor metastasis. *Cancer Cell* (2011) 19 (3):372–86. doi: 10.1016/j.ccr.2011.01.036

84. Cui YH, Kang JH, Suh Y, Zhao Y, Yi JM, Bae IH, et al. Loss of FBXL14 promotes mesenchymal shift and radioresistance of non-small cell lung cancer by TWIST1 stabilization. *Signal Transduct Target Ther* (2021) 6(1):272. doi: 10.1038/s41392-021-00599-z

85. Gomez-Casal R, Bhattacharya C, Ganesh N, Bailey L, Basse P, Gibson M, et al. Non-small cell lung cancer cells survived ionizing radiation treatment display cancer stem cell and epithelial-mesenchymal transition phenotypes. *Mol Cancer*. (2013) 12(1):94. doi: 10.1186/1476-4598-12-94

86. Dadrich M, Nicolay NH, Flechsig P, Bickelhaupt S, Hoeltgen L, Roeder F, et al. Combined inhibition of TGF $\beta$  and PDGF signaling attenuates radiation-induced pulmonary fibrosis. *Oncoimmunology.* (2016) 5(5):e1123366. doi: 10.1080/2162402X. 2015.1123366

87. Li Y, Sun C, Tan Y, Zhang H, Li Y, Zou H. ITGB1 enhances the radioresistance of human non-small celllung cancer cells by modulating the DNA damage response and YAP1-induced epithelial-mesenchymal transition. *Int J Biol Sci* (2021) 17(2):635–50. doi: 10.7150/ijbs.52319

88. Kim E, Youn H, Kwon T, Son B, Kang J, Yang HJ, et al. PAK1 tyrosine phosphorylation is required to induce epithelial-mesenchymal transition and radioresistance in lung cancer cells. *Cancer Res* (2014) 74(19):5520–31. doi: 10.1158/0008-5472.CAN-14-0735

89. Tan S, Yi P, Wang H, Xia L, Han Y, Wang H, et al. RAC1 involves in the radioresistance by mediating epithelial-mesenchymal transition in lung cancer. *Front* Oncol (2020) 10:649. doi: 10.3389/fonc.2020.00649

90. Yao YH, Cui Y, Qiu XN, Zhang LZ, Zhang W, Li H, et al. Attenuated lkb1-sik1 signaling promotes epithelial-mesenchymal transition and radioresistance of non-small cell lung cancer cells. *Chin J Cancer*. (2016) 35(10):9. doi: 10.1186/s40880-016-0113-3

91. Fu W, Zhao J, Hu W, Dai L, Jiang Z, Zhong S, et al. LINC01224/ZNF91 promote stem cell-like properties and drive radioresistance in non-small cell lung cancer. *Cancer Manag Res* (2021) 13:5671–81. doi: 10.2147/CMAR.S313744

92. Cancer Genome Atlas Research Network. Comprehensive molecular profiling of lung adenocarcinoma. *Nature* (2014) 511(7511):543–50. doi: 10.1038/nature13385

93. Yang LQ, Shi PF, Zhao GC, Xu J, Peng W, Zhang JY, et al. Targeting cancer stem cell pathways for cancer therapy. *Signal Transduct Target Ther* (2020) 5(1):8. doi: 10.1038/ s41392-020-0110-5

94. Hsu F, Sit D, Pastuch A, Dingler A, Atwal P. Lung cancer epidermal growth factor receptor mutations and radiotherapy response: A multicentre clinical study. *Clin Transl Radiat Oncol* (2021) 30:15–8. doi: 10.1016/j.ctro.2021.06.006

95. Butkiewicz D, Krześniak M, Gdowicz-Kłosok A, Giglok M, Marszałek-Zeńczak M, Suwiński R. EGFR polymorphisms in gene predict clinical outcome in unresectable nonsmall cell lung cancer treated with radiotherapy and platinum-based chemoradiotherapy. *Int J Mol Sci* (2021) 22(11):5605. doi: 10.3390/ijms22115605

96. Zhu DQ, Liu Y, Yu ZJ, Zhang RH, Li AW, Gong FY, et al. The diverse analysis identifies mutated KRAS associated with radioresistance in non-small cell lung cancer. *World J Oncol* (2022) 13(2):84–95. doi: 10.14740/wjon1465

97. Binkley MS, Jeon YJ, Nesselbush M, Moding EJ, Nabet BY, Almanza D, et al. KEAP1/NFE2L2 mutations predict lung cancer radiation resistance that can be targeted by glutaminase inhibition. *Cancer Discovery* (2020) 10(12):1826–41. doi: 10.1158/2159-8290.CD-20-0282

98. Kinslow CJ, Kumar P, Cai LL, Sun RC, Chaudhary KR, Cheng SK. NRF2-pathway mutations predict radioresistance in non-small cell lung cancer. *Transl Lung Cancer Res* (2022) 11(7):1510–3. doi: 10.21037/tlcr-22-292

99. Pustovalova M, Alhaddad L, Smetanina N, Chigasova A, Blokhina T, Chuprov-Netochin R, et al. The p53-53BP1-related survival of A549 and H1299 human lung cancer cells after multifractionated radiotherapy demonstrated different response to additional acute X-ray exposure. *Int J Mol Sci* (2020) 21(9):3342. doi: 10.3390/ijms21093342

100. Gurtner K, Kryzmien Z, Koi L, Wang M, Benes CH, Hering S, et al. Radioresistance of kras/tp53-mutated lung cancer can be overcome by radiation dose escalation or egfr tyrosine kinase inhibition. *vivo. Int J Cancer.* (2020) 147(2):472–7. doi: 10.1002/ijc.32598

101. Molina JR, Yang P, Cassivi SD, Schild SE, Adjei AA. Non-small cell lung cancer: epidemiology, risk factors, treatment, and survivorship. *Mayo Clin Proc* (2008) 83(5):584–94. doi: 10.4065/83.5.584

102. Mendelsohn J, Baselga J. Status of epidermal growth factor receptor antagonists in the biology and treatment of cancer. *J Clin Oncol* (2003) 21(14):2787–99. doi: 10.1200/JCO.2003.01.504

103. Das AK, Chen BP, Story MD, Sato M, Minna JD, Chen DJ, et al. Somatic mutations in the tyrosine kinase domain of epidermal growth factor receptor (EGFR) abrogate EGFR-mediated radioprotection in non-small cell lung carcinoma. *Cancer Res* (2007) 67(11):5267–74. doi: 10.1158/0008-5472.CAN-07-0242

104. Chen GD, Kong PZ, Yang MM, Hu WL, Prise KM, Yu KN, et al. Golgi phosphoprotein 3 confers radioresistance via stabilizing EGFR in lung adenocarcinoma. *Int J Radiat Oncol Biol Phys* (2022) 112(5):1216–28. doi: 10.1016/j.ijrobp.2021.11.023

105. Liang K, Ang KK, Milas L, Hunter N, Fan Z. The epidermal growth factor receptor mediates radioresistance. *Int J Radiat Oncol Biol Phys* (2003) 57(1):246–54. doi: 10.1016/s0360-3016(03)00511-x

106. LoRusso PM. Inhibition of the PI3K/AKT/mTOR pathway in solid tumors. J Clin Oncol (2016) 34(31):3803–15. doi: 10.1200/JCO.2014.59.0018

107. Schuurbiers OC, Kaanders JH, van der Heijden H, Dekhuijzen RP, Oyen WJ, Bussink J. The PI3-K/AKT-pathway and radiation resistance mechanisms in non-small cell lung cancer. *J Thorac Oncol* (2009) 4(6):761–7. doi: 10.1097/JTO.0b013e3181a1084f

108. Raghav KP, Gonzalez-Angulo AM, Blumenschein GR. Role of HGF/MET axis in resistance of lung cancer to contemporary management. *Transl Lung Cancer Res* (2012) 1 (3):179–93. doi: 10.3978/j.issn.2218-6751.2012.09.04

109. Califano R, Landi L, Cappuzzo F. Prognostic and predictive value of K-RAS mutations in non-small cell lung cancer. *Drugs.* (2012) 72(1):28–36. doi: 10.2165/1163012-S0-00000000-00000

110. Wang M, Kern AM, Hülskötter M, Greninger P, Singh A, Pan YF, et al. EGFRmediated chromatin condensation protects KRAS-mutant cancer cells against ionizing radiation. *Cancer Res* (2014) 74(10):2825–34. doi: 10.1158/0008-5472.CAN-13-3157

111. HuJF, ZhangZG, ZhaoL, LiL, ZuoW, HanL. High expression of RAD51 promotes DNA damage repair and survival in KRAS-mutant lung cancer cells. *BMB Rep* (2019) 52 (2):151–6. doi: 10.5483/BMBRep.2019.52.2.213

112. Wang M, Han J, Marcar L, Black J, Liu Q, Li XY, et al. Radiation resistance in KRAS-mutated lung cancer is enabled by stem-like properties mediated by an osteopontin-EGFR pathway. *Cancer Res* (2017) 77(8):2018–28. doi: 10.1158/0008-5472.CAN-16-0808

113. Dempke WC, Reck M. KEAP1/NRF2 (NFE2L2) mutations in NSCLC-fuel for a superresistant phenotype? *Lung Cancer*. (2021) 159(2021):10-7. doi: 10.1016/j.lungcan.2021.07.006

114. Jeong Y, Hoang NT, Lovejoy A, Stehr H, Newman AM, Gentles AJ, et al. Role of KEAP1/NRF2 and TP53 mutations in lung squamous cell carcinoma development and radiotherapy rresponse prediction. *Cancer Discovery.* (2017) 7(1):86–101. doi: 10.1158/2159-8290.CD-16-0127

115. Pillai R, Hayashi M, Zavitsanou AM, Papagiannakopoulos T. NRF2: KEAPing tumors protected. *Cancer Discovery* (2022) 12(3):625–43. doi: 10.1158/2159-8290.CD-21-0922

116. Lee SL, Ryu H, Son AR, Seo B, Kim J, Jung SY, et al. TGF- $\beta$  and hypoxia/ reoxygenation promote radioresistance of A549 lung cancer cells through activation of Nrf2 and EGFR. Oxid Med Cell Longev (2016) 2016;6823471. doi: 10.1155/2016/6823471

117. Liu YQ, Gu W. p53 in ferroptosis regulation: The new weapon for the old guardian. Cell Death Differ (2022) 29(5):895-910. doi: 10.1038/s41418-022-00943-y

118. Bieging K, Mello S, Attardi L. Unravelling mechanisms of p53-mediated tumour suppression. *Nat Rev Cancer.* (2014) 14(5):359–70. doi: 10.1038/nrc3711

119. Moretti L, Li B, Kim KW, Chen HD, Lu B. AT-101, a pan-Bcl-2 inhibitor, leads to radiosensitization of non-small cell lung cancer. *J Thorac Oncol* (2010) 5(5):680–7. doi: 10.1097/JTO.0b013e3181d6e08e

120. Tu H, Costa M. XIAP's profile in human cancer. Biomolecules. (2020) 10 (11):1493. doi: 10.3390/biom10111493

121. Sato Y, Yoshino H, Kazama Y, Kashiwakura I. Involvement of caspase-8 in apoptosis enhancement by cotreatment with retinoic acid-inducible gene-i-like receptor agonist and ionizing radiation in human non-small cell lung cancer. *Mol Med Rep* (2018) 18(6):5286–94. doi: 10.3892/mmr.2018.9536

122. Lu B, Mu Y, Cao C, Zeng FH, Schneider S, Tan JH, et al. Survivin as a therapeutic target for radiation sensitization in lung cancer. *Cancer Res* (2004) 64(8):2840–5. doi: 10.1158/0008-5472.can-03-3547

123. Sun H, Du Y, Yao M, Wang Q, Ji K, Du L, et al. cIAP1/2 are involved in the radiosensitizing effect of birinapant on NSCLC cell line *in vitro. J Cell Mol Med* (2021) 25 (13):6125–36. doi: 10.1111/jcmm.16526

124. Yin H, Ma JQ, Chen L, Piao SQ, Zhang Y, Zhang SL, et al. MiR-99a enhances the radiation sensitivity of non-small cell lung cancer by targeting mTOR. *Cell Physiol Biochem* (2018) 46(2):471–81. doi: 10.1159/000488615

125. Lee HC, Her NG, Kang D, Jung SH, Shin J, Lee M, et al. Radiation-inducible miR-770-5p sensitizes tumors to radiation through direct targeting of PDZ-binding kinase. *Cell Death Dis* (2017) 8(3):e2693. doi: 10.1038/cddis.2017.116

126. Yuan Y, Liao H, Pu Q, Ke XX, Hu XT, Ma YF, et al. miR-410 induces both epithelial-mesenchymal transition and radioresistance through activation of the PI3K/ mTOR pathway in non-small cell lung cancer. *Signal Transduct Target Ther* (2020) 5 (1):85. doi: 10.1038/s41392-020-0182-2

127. Tang YT, Cui YY, Li ZP, Jiao ZQ, Zhang Y, He Y, et al. Radiation-induced miR-208a increases the proliferation and radioresistance by targeting p21 in human lung cancer cells. *J Exp Clin Cancer Res* (2016) 35:7. doi: 10.1186/s13046-016-0285-3

128. Clarke MF, Fuller M. Stem cells and cancer: Two faces of eve. *Cell.* (2006) 124 (6):1111–5. doi: 10.1016/j.cell.2006.03.011

129. Schulz A, Meyer F, Dubrovska A, Borgmann K. Cancer stem cells and radioresistance: DNA repair and beyond. *Cancers (Basel).* (2019) 11(6):862. doi: 10.3390/cancers11060862

130. Geng YY, Amante JJ, Goel HL, Zhang XZ, Walker MR, Luther DC, et al. Differentiation of cancer stem cells through nanoparticle surface engineering. ACS Nano. (2020) 14(11):15276–85. doi: 10.1021/acsnano.0c05589

131. Paul R, Dorsey JF, Fan Y. Cell plasticity, senescence, and quiescence in cancer stem cells: Biological and therapeutic implications. *Pharmacol Ther* (2022) 231:107985. doi: 10.1016/j.pharmthera.2021.107985

132. Jang YY, Sharkis SJ. A low level of reactive oxygen species selects for primitive hematopoietic stem cells that may reside in the low-oxygenic niche. *Blood.* (2007) 110 (8):3056–63. doi: 10.1182/blood-2007-05-087759

133. Lendeckel U, Wolke C. Redox-regulation in cancer stem cells. *Biomedicines* (2022) 10(10):2413. doi: 10.3390/biomedicines10102413

134. Ryoo I, Lee S, Kwak MK. Redox modulating NRF2: A potential mediator of cancer stem cell resistance. *Oxid Med Cell Longev* (2016) 2016:2428153. doi: 10.1155/2016/2428153

135. Chang L, Graham P, Hao JL, Ni J, Deng JL, Bucci J, et al. Cancer stem cells and signaling pathways in radioresistance. *Oncotarget.* (2016) 7(10):11002–17. doi: 10.18632/ oncotarget.6760

136. Bao SD, Wu QL, McLendon RE, Hao YL, Shi Q, Hjelmeland AB, et al. Glioma stem cells promote radioresistance by preferential activation of the DNA damage response. *Nature*. (2006) 444(7120):756–60. doi: 10.1038/nature05236

137. Desai A, Webb B, Gerson SL. CD133+ cells contribute to radioresistance via altered regulation of DNA repair genes in human lung cancer cells. *Radiother Oncol* (2014) 110(3):538-45. doi: 10.1016/j.radonc.2013.10.040

138. Moore N, Lyle S. Quiescent, slow-cycling stem cell populations in cancer: a review of the evidence and discussion of significance. *J Oncol* (2011) 2011:396076. doi: 10.1155/2011/396076

139. Krause M, Yaromina A, Eicheler W, Koch U, Baumann M. Cancer stem cells: targets and potential biomarkers for radiotherapy. *Clin Cancer Res* (2011) 17(23):7224–9. doi: 10.1158/1078-0432.CCR-10-2639

140. Baumann M, Krause M. CD44: a cancer stem cell-related biomarker with predictive potential for radiotherapy. *Clin Cancer Res* (2010) 16(21):5091-3. doi: 10.1158/1078-0432.CCR-10-2244

141. Hu B, Ma YY, Yang Y, Zhang LJ, Han HB, Chen JF. CD44 promotes cell proliferation in non-small cell lung cancer. Oncol Lett (2018) 15(4):5627–33. doi: 10.3892/ol.2018.8051

142. Kong T, Ahn R, Yang KN, Zhu XB, Fu Z, Morin G, et al. CD44 promotes PD-L1 expression and its tumor-intrinsic function in breast and lung cancers. *Cancer Res* (2020) 80(3):444–57. doi: 10.1158/0008-5472.CAN-19-1108

143. Bayer C, Shi K, Astner ST, Maftei CA, Vaupel P. Acute versus chronic hypoxia: why a simplified classification is simply not enough. *Int J Radiat Oncol Biol Phys* (2011) 80 (4):965–8. doi: 10.1016/j.ijrobp.2011.02.049

144. Telarovic I, Wenger RH, Pruschy M. Interfering with tumor hypoxia for radiotherapy optimization. *J Exp Clin Cancer Res* (2021) 40(1):197. doi: 10.1186/s13046-021-02000-x

145. Sun JC, He F, Yi W, Wan MH, Li R, Wei X, et al. High expression of hif- $2\alpha$  and its anti-radiotherapy effect in lung cancer stem cells. *Genet Mol Res* (2015) 14(4):18110–20. doi: 10.4238/2015.December.22.37

146. Wang Y, Roche O, Yan MS, Finak G, Evans AJ, Metcalf JL, et al. Regulation of endocytosis *via* the oxygen-sensing pathway. *Nat Med* (2009) 15(3):319–24. doi: 10.1038/ nm.1922

147. Downes NL, Laham-Karam N, Kaikkonen MU, Ylä-Herttuala S. Differential but complementary HIF1 $\alpha$  and HIF2 $\alpha$  transcriptional regulation. *Mol Ther* (2018) 26 (7):1735–45. doi: 10.1016/j.ymthe.2018.05.004

148. Gonzalez FJ, Xie C, Jiang CT. The role of hypoxia-inducible factors in metabolic diseases. *Nat Rev Endocrinol* (2018) 15(1):21–32. doi: 10.1038/s41574-018-0096-z

149. Semenza GL. Targeting HIF-1 for cancer therapy. Nat Rev Cancer. (2003) 3 (10):721-32. doi: 10.1038/nrc1187

150. Uniacke J, Holterman CE, Lachance G, Franovic A, Jacob MD, Fabian MR, et al. An oxygen-regulated switch in the protein synthesis machinery. *Nature.* (2012) 486 (7401):126–9. doi: 10.1038/nature11055

151. Choueiri TK, Kaelin WG. Targeting the HIF2-VEGF axis in renal cell carcinoma. Nat Med (2020) 26(10):1519–30. doi: 10.1038/s41591-020-1093-z

152. Moeller BJ, Cao YT, Li CY, Dewhirst MW. Radiation activates HIF-1 to regulate vascular radiosensitivity in tumors: Role of reoxygenation, free radicals, and stress granules. *Cancer Cell* (2004) 5(5):429–41. doi: 10.1016/s1535-6108(04)00115-1

153. Moreno RE, Groot AJ, Yaromina A, Hendrickx TC, Barbeau LM, Giuranno L, et al. HIF-1 $\alpha$  and HIF-2 $\alpha$  differently regulate the radiation sensitivity of NSCLC cells. *Cells.* (2019) 8(1):45. doi: 10.3390/cells8010045

154. Grosso S, Doyen J, Parks SK, Bertero T, Paye A, Cardinaud B, et al. MiR-210 promotes a hypoxic phenotype and increases radioresistance in human lung cancer cell lines. *Cell Death Dis* (2013) 4(3):e544. doi: 10.1038/cddis.2013.71

155. Xu LM, Yu H, Yuan YJ, Zhang J, Ma Y, Cao XC, et al. Overcoming of radioresistance in non-small cell lung cancer by microRNA-320a through HIF1 $\alpha$ -suppression mediated methylation of PTEN. *Front Cell Dev Biol* (2020) 8:553733. doi: 10.3389/fcell.2020.553733

156. Isla D, De Las PR, Insa A, Marsé R, Martínez-Banaclocha N, Mut P, et al. Oral vinorelbine versus etoposide with cisplatin and chemo-radiation as treatment in patients with stage III non-small cell lung cancer: A randomized phase II (RENO study). *Lung Cancer*. (2019) 135:161–8:135. doi: 10.1016/j.lungcan.2018.11.041

157. Shaverdian N, Lisberg AE, Bornazyan K, Veruttipong D, Goldman JW, Formenti SC, et al. Previous radiotherapy and the clinical activity and toxicity of pembrolizumab in the treatment of non-small-cell lung cancer: A secondary analysis of the KEYNOTE-001 phase 1 trial. *Lancet Oncol* (2017) 18(7):895–903. doi: 10.1016/S1470-2045(17)30380-7

158. Kong Y, Xu C, Sun X, Sun H, Zhao X, He N, et al. BLM helicase inhibition synergizes with PARP inhibition to improve the radiosensitivity of olaparib resistant non-small cell lung cancer cells by inhibiting homologous recombination repair. *Cancer Biol Med* (2021) 19(8):1150–71. doi: 10.20892/j.issn.2095-3941.2021.0178

159. Zhang N, Gao Y, Zeng Z, Luo Y, Xie C. PARP inhibitor niraparib as a radiosensitizer promotes antitumor immunity of radiotherapy in EGFR-mutated non-small cell lung cancer. *Clin Transl Oncol* (2021) 23(9):1827–37. doi: 10.1007/s12094-021-02591-z

160. Tang S, Li Z, Yang L, Shen L, Wang Y. A potential new role of ATM inhibitor in radiotherapy: Suppressing ionizing radiation-activated EGFR. *Int J Radiat Biol* (2020) 96 (4):461–8. doi: 10.1080/09553002.2020.1707325

161. Fok JHI, Ramos-Montoya A, Vazquez-Chantada M, Wijnhoven PWG, Follia V, James N, et al. AZD7648 is a potent and selective DNA-PK inhibitor that enhances radiation, chemotherapy and olaparib activity. *Nat Commun* (2019) 10(1):5065. doi: 10.1038/s41467-019-12836-9

162. Ding X, Cheng J, Pang Q, Wei X, Zhang X, Wang P, et al. BIBR1532, a selective telomerase inhibitor, enhances radiosensitivity of non-small cell lung cancer through increasing telomere dysfunction and ATM/CHK1 inhibition. *Int J Radiat Oncol Biol Phys* (2019) 105(4):861–74. doi: 10.1016/j.ijrobp.2019.08.009

163. Wang J, Wang Y, Mei H, Yin Z, Geng Y, Zhang T, et al. The BET bromodomain inhibitor JQ1 radiosensitizes non-small cell lung cancer cells by upregulating p21. *Cancer Lett* (2017) 391:141–51. doi: 10.1016/j.canlet.2017.01.031

164. Fernández-Aroca DM, Roche O, Sabater S, Pascual-Serra R, Ortega-Muelas M, Sánchez Pérez I, et al. P53 pathway is a major determinant in the radiosensitizing effect of palbociclib: Implication in cancer therapy. *Cancer Lett* (2019) 451:23–33. doi: 10.1016/j.canlet.2019.02.049

165. Naz S, Sowers A, Choudhuri R, Wissler M, Gamson J, Mathias A, et al. Abemaciclib, a selective CDK4/6 inhibitor enhances the radiosensitivity of non-small cell lung cancer *in vitro* and *in vivo*. *Clin Cancer Res* (2018) 24(16):3994–4005. doi: 10.1158/1078-0432.CCR-17-3575

166. Wang R, Peng S, Zhang X, Wu Z, Duan H, Yuan Y, et al. Inhibition of NF- $\kappa$ B improves sensitivity to irradiation and EGFR-TKIs and decreases irradiation-induced lung toxicity. *Int J Cancer.* (2019) 144:200–9. doi: 10.1002/ijc.31907

167. Du S, Bouquet S, Lo CH, Pellicciotta I, Bolourchi S, Parry R, et al. Attenuation of the DNA damage response by transforming growth factor- $\beta$  inhibitors enhances radiation sensitivity of non–small-cell lung cancer cells *in vitro* and *in vivo*. Int J Radiat Oncol (2015) 91(1):91–9. doi: 10.1016/j.ijrobp.2014.09.026

168. Sun Y, Moretti L, Giacalone NJ, Schleicher S, Speirs CK, Carbone DP, et al. Inhibition of JAK2 signaling by TG101209 enhances radiotherapy in lung cancer models. *J Thorac Oncol* (2011) 6(4):699–706. doi: 10.1097/JTO.0b013e31820d9d11

169. Gong X, Li X, Jiang T, Xie H, Zhu Z, Zhou F, et al. Combined radiotherapy and anti-PD-L1 antibody synergistically enhances antitumor effect in non-small cell lung cancer. J Thorac Oncol (2017) 12(7):1085–97. doi: 10.1016/j.jtho.2017.04.014

170. Bozorgmehr F, Hommertgen A, Krisam J, Lasitschka F, Kuon J, Maenz M, et al. Fostering efficacy of anti-PD-1-treatment: Nivolumab plus radiotherapy in advanced non-small cell lung cancer - study protocol of the FORCE trial. *BMC Cancer.* (2019) 19 (1):1074. doi: 10.1186/s12885-019-6205-0

171. Welsh J, Menon H, Chen D, Verma V, Heymach JV. Pembrolizumab with or without radiation therapy for metastatic non-small cell lung cancer: A randomized phase I/II trial. *J Immunother Cancer*. (2020) 8(2):e001001. doi: 10.1136/jitc-2020-001001

172. Formenti SC, Rudqvist NP, Golden E, Cooper B, Wennerberg E, Lhuillier C, et al. Radiotherapy induces responses of lung cancer to CTLA-4 blockade. *Nat Med* (2018) 24 (12):1845–51. doi: 10.1038/s41591-018-0232-2

173. Zheng LP, Wang Y, Xu Z, Yang Q, Zhu G, Liao XY, et al. Concurrent EGFR-TKI and thoracic radiotherapy as first-line treatment for stage IV non-small cell lung cancer harboring EGFR active mutations. *Oncologist.* (2019) 24(8):1031–e612. doi: 10.1634/ theoncologist.2019-0285

174. Francis DM, Huang S, Armstrong EA, Werner LR, Hullett C, Li C, et al. Pan-HER inhibitor augments radiation response in human lung and head and neck cancer models. *Clin Cancer Res* (2016) 22(3):633–43. doi: 10.1158/1078-0432.CCR-15-1664

175. Hu S, Fu W, Li T, Yuan Q, Wang F, Lv G, et al. Antagonism of EGFR and notch limits resistance to EGFR inhibitors and radiation by decreasing tumor-initiating cell frequency. *Sci Transl Med* (2017) 9(380):1–15. doi: 10.1126/scitranslmed.aag0339

176. Zhang P, Song E, Jiang M, Song Y. Celecoxib and afatinib synergistic enhance radiotherapy sensitivity on human non-small cell lung cancer A549 cells. *Int J Radiat Biol* (2021) 97(2):170–8. doi: 10.1080/09553002.2021.1846817

177. Park D, Magis AT, Li R, Owonikoko TK, Sica GL, Sun SY, et al. Novel small-molecule inhibitors of bcl-XL to treat lung cancer. *Cancer Res* (2013) 73:5485–96. doi: 10.1158/0008-5472.CAN-12-2272

178. Lee JM, Kim HS, Kim A, Chang YS, Lee JG, Cho J, et al. ABT-737, a BH3 mimetic, enhances the therapeutic effects of ionizing radiation in K-ras mutant non-small cell lung cancer preclinical model. *Yonsei Med J* (2022) 63(1):16–25. doi: 10.3349/ymj.2022.63.1.16

179. Park D, Anisuzzaman AS, Magis AT, Chen G, Xie M, Zhang G, et al. Discovery of small molecule bak activator for lung cancer therapy. *Theranostics.* (2021) 11:8500–16. doi: 10.7150/thno.60349

180. Ritter V, Krautter F, Klein D, Jendrossek V, Rudner J. Bcl-2/Bcl-xL inhibitor ABT-263 overcomes hypoxia-driven radioresistence and improves radiotherapy. *Cell Death Dis* (2021) 12(7):694. doi: 10.1038/s41419-021-03971-7

181. Jiang S, Zhou Y, Zou L, Chu L, Chu X, Ni J, et al. Low- dose apatinib promotes vascular normalization and hypoxia reduction and sensitizes radiotherapy in lung cancer. *Cancer Med* (2022) 00:1–12. doi: 10.1002/cam4.5113

182. Zhai Y, Ma H, Hui Z, Zhao L, Li D, Liang J, et al. HELPER study: A phase II trial of continuous infusion of endostar combined with concurrent etoposide plus cisplatin and radiotherapy for treatment of unresectable stage III non-small-cell lung cancer. *Radiother Oncol* (2019) 131:27–34. doi: 10.1016/j.radonc.2018.10.032

183. Kong FM, Hayman JA, Griffith KA, Kalemkerian GP, Arenberg D, Lyons S, et al. Final toxicity results of a radiation-dose escalation study in patients with non-small-cell lung cancer (NSCLC): predictors for radiation pneumonitis and fibrosis. Int J Radiat Oncol Biol Phys (2006) 65(4):1075-86. doi: 10.1016/j.ijrobp.2006.01.051

184. Liu W, Yang BB, Yang L, Kaur J, Jessop C, Fadhil R, et al. Therapeutic effects of ten commonly used Chinese herbs and their bioactive compounds on cancers. *Evid Based Complement Alternat Med* (2019) 2019:6057837. doi: 10.1155/2019/6057837

185. Yang Y, Li N, Wang TM, Di L. Natural products with activity against lung cancer: A review focusing on the tumor microenvironment. *Int J Mol Sci* (2021) 22(19):10827. doi: 10.3390/ijms221910827

186. Li SG, Chen HY, Ou-Yang CS, Wang XX, Yang ZJ, Tong Y, et al. The efficacy of Chinese herbal medicine as an adjunctive therapy for advanced non-small cell lung cancer: a systematic review and meta-analysis. *PloS One* (2013) 8(2):e57604. doi: 10.1371/journal.pone.0057604

187. Balakrishnan B, Liang Q, Fenix K, Tamang B, Hauben E, Ma L, et al. Combining the anticancer and immunomodulatory effects of *Astragalus* and shiitake as an integrated therapeutic approach. *Nutrients.* (2021) 13(8):2564. doi: 10.3390/nu13082564

188. Cao A, He H, Wang Q, Li L, An Y, Zhou X. Evidence of astragalus injection combined platinum-based chemotherapy in advanced nonsmall cell lung cancer patients: A systematic review and meta-analysis. *Medicine*. (2019) 98(11):e14798. doi: 10.1097/MD.000000000014798

189. Zhang YM, Zhang LY, Zhou H, Li YY, Wei KX, Li CH, et al. Astragalus polysaccharide inhibits radiation-induced bystander effects by regulating apoptosis in bone mesenchymal stem cells (BMSCs). *Cell Cycle* (2020) 19(22):3195–207. doi: 10.1080/15384101.2020.1838793

190. Zhang L, Luo Y, Lu Z, He J, Wang L, Zhang L, et al. Astragalus polysaccharide inhibits ionizing radiation-induced bystander effects by regulating MAPK/NF-kB signaling pathway in bone mesenchymal stem cells (BMSCs). *Med Sci Monit* (2018) 24:4649–58. doi: 10.12659/MSM.909153

191. Zhang L, Yong W, Wang L, Zhang L, Yongqi L. Astragalus polysaccharide eases g1 phase-correlative bystander effects through mediation of TGF-βR/MAPK/ROS signal pathway after carbon ion irradiation in BMSCs. *Am J Chin Med* (2019) 47(3):595–612. doi: 10.1142/S0192415X19500319

192. Zhang LY, Zhou T, Zhang YM, Xu XM, Li YY, Wei KX, et al. Guiqi baizhu decoction alleviates radiation inflammation in rats by modulating the composition of the gut microbiota. *Evid Based Complement Alternat Med* (2020) 2020(undefined):9017854. doi: 10.1155/2020/9017854

193. Li YY, Zhang LY, Zhang YM, Miao ZM, Liu ZW, Zhou GC, et al. Potential molecular mechanism of guiqi baizhu decoction in radiation-induced intestinal edema by regulating HIF-1a, AQP4 and Na/K-ATPase. *Phytomedicine* (2022) 107(undefined):154445. doi: 10.1016/j.phymed.2022.154445

194. Li L, Jin XJ, Li JW, Li CH, Zhou SY, Li JJ, et al. Systematic insight into the active constituents and mechanism of guiqi baizhu for the treatment of gastric cancer. *Cancer Sci* (2021) 112(5):1772–84. doi: 10.1111/cas.14851

195. Jabbour SK, Berman AT, Decker RH, Lin Y, Feigenberg SJ, Gettinger SN, et al. Phase 1 trial of pembrolizumab administered concurrently with chemoradiotherapy for locally advanced non-small cell lung cancer: A nonrandomized controlled trial. *JAMA Oncol* (2020) 6(6):848–55. doi: 10.1001/jamaoncol.2019.6731

196. Descourt R, Vergnenegre A, Barlesi F, Lena H, Fournel P, Falchero L, et al. Oral vinorelbine and cisplatin with concurrent radiotherapy after induction chemotherapy with cisplatin and docetaxel for patients with locally advanced non-small cell lung cancer: the GFPC 05-03 study. *J Thorac Oncol* (2011) 6(2):351–7. doi: 10.1097/JTO.0b013e318200f47e

197. Xiao Z, Liang R, Wang CQ, Xu S, Li N, He Y, et al. Can aidi injection alleviate the toxicity and improve the clinical efficacy of radiotherapy in lung cancer: A meta-analysis of 16 randomized controlled trials following the PRISMA guidelines. *Med (Baltimore).* (2016) 95(35):e4517. doi: 10.1097/MD.00000000004517

198. Zhang H, Jiang H, Hu X, Jia Z. Aidi injection combined with radiation in the treatment of non-small cell lung cancer: A meta-analysis evaluation the efficacy and side effects. J Cancer Res Ther (2015) 00:C118–21. doi: 10.4103/0973-1482.163864

199. Dong XR, Wang JN, Liu L, Chen X, Chen MS, Chen J, et al. Modulation of radiation-induced tumour necrosis factor- $\alpha$  and transforming growth factor  $\beta 1$  expression in the lung tissue by shengqi fuzheng injection. Mol Med Rep (2010) 3 (4):621–7. doi: 10.3892/mmr\_00000306

200. Wang S, Lian X, Sun M, Luo L, Guo L. Efficacy of compound kushen injection plus radiotherapy on nonsmall-cell lungcancer: A systematic review and meta-analysis. J Cancer Res Ther (2016) 12(4):1298–306. doi: 10.4103/0973-1482.199538

201. Zhou H, Pan P, Zhao Q, Liu W, Sun Y, Wang J, et al. Overcoming basal autophagy, kangai injection enhances cisplatin cytotoxicity by regulating FOXO3a-dependent autophagic cell death and apoptosis in human lung adenocarcinoma A549/ DDP cells. *BioMed Res Int* (2022) 2022:6022981. doi: 10.1155/2022/6022981

202. He H, Zhou X, Wang Q, Zhao Y. Does the couse of astragalus-containing chinese herbal prescriptions and radiotherapy benefit to non-small-cell lung cancer treatment: a meta-analysis of randomized trials. *Evid Based Complement Alternat Med* (2013) 2013;426207. doi: 10.1155/2013/426207

203. Jiang X, Hidru TH, Zhang Z, Bai Y, Kong L, Li X. Evidence of elemene injection combined radiotherapy in lung cancer treatment among patients with brain metastases: A systematic review and meta-analysis. *Med (Baltimore)*. (2017) 96(21):e6963. doi: 10.1097/MD.0000000006963

204. Yu H, Yang Y, Jiang T, Zhang X, Zhao Y, Pang G, et al. Ganoderma lucidum effective radiotherapy in tumor assisted by polysaccharide-conjugated bismuth sulfide nanoparticles through radiosensitization and dendritic cell activation. ACS Appl Mater Interfaces. (2019) 11(31):27536–47. doi: 10.1021/acsami.9b07804

205. Lu T, Yu J, Gao R, Wang J, Wang H, Wang X, et al. Chinese Patent medicine kanglaite injection for non-small-cell lung cancer: An overview of systematic reviews. J Ethnopharmacol (2022) 302:115814. doi: 10.1016/j.jep.2022.115814

206. Kong P, Yu KN, Yang M, Almahi WA, Nie L, Chen G, et al. Micheliolide enhances radiosensitivities of p53-deficient non-small-cell lung cancer via promoting HIF-1 $\alpha$  degradation. Int J Mol Sci (2020) 21(9):3392. doi: 10.3390/ijms21093392

207. Woo JK, Kang JH, Shin DY, Park SH, Kang K, Nho CW, et al. Daurinol enhances the efficacy of radiotherapy in lung cancer *via* suppression of aurora kinase A/B expression. *Mol Cancer Ther* (2015) 14(7):1693–704. doi: 10.1158/1535-7163.MCT-14-0960

208. Du Y, Jia C, Liu Y, Li Y, Wang J, Sun K. Isorhamnetin enhances the radiosensitivity of A549 cells through interleukin-13 and the NF- $\kappa$ B signaling pathway. Front Pharmacol (2020) 11:610772. doi: 10.3389/fphar.2020.610772

209. Hsu HS, Huang PI, Chang YL, Tzao C, Chen YW, Shih HC, et al. CucurbitacinI inhibits tumorigenic ability and enhances radiochemosensitivity in nonsmall cell lung cancer-derived CD133-positive cells. *Cancer.* (2011) 117(13):2970–85. doi: 10.1002/cncr.25869

210. Phiboonchaiyanan PP, Petpiroon N, Sritularak B, Chanvorachote P. Phoyunnanin e induces apoptosis of non-small cell lung cancer cells via p53 activation and down-regulation of survivin. *Anticancer Res* (2018) 38(11):6281–90. doi: 10.21873/anticanres.12984

211. JiH L, KJ C, WD S, SY J, Kim M, BW L, et al. Enhancement of radiation sensitivity in lung cancer cells by celastrol is mediated by inhibition of Hsp90. *Int J Mol Med* (2011) 27(3):441–6. doi: 10.3892/ijmm.2011.601

212. Ye M, Zhang J, Zhang J, Miao Q, Yao L, Zhang J. Curcumin promotes apoptosis by activating the p53-miR-192-5p/215-XIAP pathway in non-small cell lung cancer. *Cancer Lett* (2015) 357(1):196–205. doi: 10.1016/j.canlet.2014.11.028

213. He GH, Xing DJ, Jin D, Lu Y, Guo L, Li YL, et al. Scutellarin improves the radiosensitivity of non-small cell lung cancer cells to iodine-125 seeds *via* downregulating the AKT/mTOR pathway. *Thorac Cancer.* (2021) 12(17):2352–9. doi: 10.1111/1759-7714.14077