#### Check for updates

#### OPEN ACCESS

EDITED BY Luca Marsili, University of Cincinnati, United States

REVIEWED BY Barbara Rossi, University of Verona, Italy Chiara Marini-Bettolo, Newcastle Hospitals, United Kingdom

\*CORRESPONDENCE Julia Pereira Lemos i j.lemos@institut-myologie.org Wilson Savino wilson.savino@fiocruz.br Piera Smeriglio piera.smeriglio@inserm.fr

RECEIVED 09 April 2023 ACCEPTED 02 October 2023 PUBLISHED 18 October 2023

#### CITATION

Lemos JP, Tenório LPG, Mouly V, Butler-Browne G, Mendes-da-Cruz DA, Savino W and Smeriglio P (2023) T cell biology in neuromuscular disorders: a focus on Duchenne Muscular Dystrophy and Amyotrophic Lateral Sclerosis. *Front. Immunol.* 14:1202834. doi: 10.3389/fimmu.2023.1202834

#### COPYRIGHT

© 2023 Lemos, Tenório, Mouly, Butler-Browne, Mendes-da-Cruz, Savino and Smeriglio. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

## T cell biology in neuromuscular disorders: a focus on Duchenne Muscular Dystrophy and Amyotrophic Lateral Sclerosis

Julia Pereira Lemos<sup>1\*</sup>, Liliane Patrícia Gonçalves Tenório<sup>2,3,4</sup>, Vincent Mouly<sup>1</sup>, Gillian Butler-Browne<sup>1</sup>, Daniella Arêas Mendes-da-Cruz<sup>3,4,5,6</sup>, Wilson Savino<sup>3,4,5,6\*</sup> and Piera Smeriglio<sup>1\*</sup>

<sup>1</sup>Sorbonne Université, INSERM, Institut de Myologie, Centre de Recherche en Myologie, Paris, France, <sup>2</sup>Laboratory of Cell Biology, Institute of Biological and Health Sciences, Federal University of Alagoas, Maceio, Alagoas, Brazil, <sup>3</sup>Laboratory on Thymus Research, Oswaldo Cruz Institute, Oswaldo Cruz Foundation, Rio de Janeiro, Brazil, <sup>4</sup>National Institute of Science and Technology on Neuroimmunomodulation (INCT-NIM), Oswaldo Cruz Institute, Oswaldo Cruz Foundation, Rio de Janeiro, Brazil, <sup>5</sup>Rio de Janeiro Research Network on Neuroinflammation (RENEURIN), Oswaldo Cruz Institute, Oswaldo Cruz Foundation, Rio de Janeiro, Brazil, <sup>6</sup>INOVA-IOC Network on Neuroimmunomodulation, Oswaldo Cruz Institute, Oswaldo Cruz Foundation, Rio de Janeiro, Brazil, <sup>6</sup>INOVA-IOC Network on

Growing evidence demonstrates a continuous interaction between the immune system, the nerve and the muscle in neuromuscular disorders of different pathogenetic origins, such as Duchenne Muscular Dystrophy (DMD) and Amyotrophic Lateral Sclerosis (ALS), the focus of this review. Herein we highlight the complexity of the cellular and molecular interactions involving the immune system in neuromuscular disorders, as exemplified by DMD and ALS. We describe the distinct types of cell-mediated interactions, such as cytokine/ chemokine production as well as cell-matrix and cell-cell interactions between T lymphocytes and other immune cells, which target cells of the muscular or nervous tissues. Most of these interactions occur independently of exogenous pathogens, through ligand-receptor binding and subsequent signal transduction cascades, at distinct levels of specificity. Although this issue reveals the complexity of the system, it can also be envisioned as a window of opportunity to design therapeutic strategies (including synthetic moieties, cell and gene therapy, as well as immunotherapy) by acting upon one or more targets. In this respect, we discuss ongoing clinical trials using VLA-4 inhibition in DMD, and in ALS, with a focus on regulatory T cells, both revealing promising results.

#### KEYWORDS

Duchenne Muscular Dystrophy, Amyotrophic Lateral Sclerosis, effector T cells, regulatory T cells, immunotherapy

## 1 Introduction

# 1.1 Neuromuscular diseases: a focus on DMD and ALS

Neuromuscular disorders (NMD) encompass a vast number of pathological conditions affecting the nervous system and the skeletal muscle and their communication, frequently causing muscle wasting and weakness. Some NMD, such as Duchenne Muscular Dystrophy (DMD), have a defined genetic origin and others, like Amyotrophic Lateral Sclerosis (ALS), can have a genetic origin or arise from sporadic mutations.

DMD is a genetic disorder due to mutations in the gene encoding the dystrophin protein. It is an X-linked disease mainly affecting boys, with a relatively high incidence (1/1500 boys born alive) It is characterized by continuous cycles of skeletal muscle degeneration and regeneration ultimately resulting in muscle wasting and loss of ambulation between 8-14 years of age (1). It has been shown that intramuscular inflammatory infiltrates, which are found in DMD patients, are mainly composed by T lymphocytes (especially CD8+ T cells) and macrophages (2). The most commonly used DMD model is the C57BL/10ScSn (mdx) mouse, which carries a naturally occurring point mutation in exon 23 of the dystrophin gene, an absence of the full-length Dp427 protein isoform (3, 4) and to a lesser extent genetic variation of the mdx model (5-7). Other DMD murine models include the D2.mdx mouse, corresponding to the mdx mice bred on the DBA2/J background (8-10), and the dystrophin/utrophin doubleknockout (dko) mouse (11-14), among others.

ALS is a neurological disorder characterized by the loss of motor neurons (MN) and paralysis of the skeletal musculature (15). It affects mostly middle-aged individuals (16), being the most common MN disorder in adults (17). The disease is characterized by muscular weakness due to progressive atrophy leading to paralysis and death, 3 to 5 years after diagnosis (18), secondary to respiratory failure (19). The diagnosis of ALS is clinical, based on the patient's history and symptomatic examination, usually supported by electrophysiological investigation (20). Around 10% of the ALS cases have a familial history (fALS), caused by mutations in one or several genes, mostly inherited in an autosomal dominant pattern (21). Of these, around 20% are associated with mutations in the Cu/Zn superoxide dismutase 1 (mSOD1) gene and approximately 40% in the chromosome 9 open reading frame 72 (C9orf72) gene, in Europe and the United States, and can drastically vary, notably in Asian patients (21-23). Experimental models for both mutations have been developed, namely the mSOD1 and the C9orf72 mice (24) that despite some limitations, recapitulate some of the main pathophysiological aspects of the disease.

## 1.2 General features of T-cell development and generation of regulatory T cells

T cells develop in the thymus, from common lymphoid progenitors of bone marrow origin. These precursors migrate into

the bloodstream and enter the thymus through vessels present at the corticomedullary junction of the thymic lobules, where they receive a signal from the microenvironment that activates specific genes, leading to commitment to the T cell lineage (25).

T cells maturation occur via positive and negative selection processes occurring through the interaction of the T-cell receptor (TCR) with class I or class II major histocompatibility complex (MHC) molecules associated with self-peptides. Those complexes are expressed by the thymic epithelium (TEC) and thymic dendritic cells (DC) (25) and largely prevent the development of cells with autoimmune potential. During the double-negative (DN; CD4-CD8<sup>-</sup>) stage, cells undergo gene rearrangement of the  $\beta$  chain or the  $\gamma\delta$  chains of the T-cell receptor, determining whether the thymocytes will be  $\alpha\beta$  or  $\gamma\delta$ , although most develop into  $\alpha\beta$  (25). Cells that do not successfully rearrange die; in contrast, cells that undergo a successful rearrangement proliferate intensively. At this stage, cells express a pre-TCR that is associated with the CD3 molecular complex necessary for receptor signaling. Pre-TCR expression leads to cell proliferation, arrest of rearrangement of β-chain genes and expression of CD4 and CD8 coreceptors, determining the double-positive stage (DP; CD4<sup>+</sup>CD8<sup>+</sup>), which represents about 80% of thymocytes (25). At this stage, the rearrangement of the  $\alpha$  chain genes begins, leading to the expression of the definitive TCR $\alpha\beta$  complex, making DP cells responsive to specific antigens and then subject to positive or negative selection processes (25). Cells whose TCRs are able to bind pMHC with moderate avidity are positively selected, following the differentiation process (26). Thymocytes that express TCRs associated with MHC class I-bound self-peptides loose expression of the CD4 molecule and become TCR<sup>hi</sup>CD4<sup>-</sup>CD8<sup>+</sup> (about 5%), whereas those that express TCRs associated with MHC class II become TCR<sup>hi</sup>CD4<sup>+</sup>CD8<sup>-</sup> (about 10%). At this final stage of intrathymic T-cell development, the single-positive (SP) thymocytes are now mature cells ready to leave the thymus and exert their effector and regulatory functions in the periphery of the immune system. In fact, some clones that recognize self-antigens with high avidity differentiate into CD4+CD8-CD25+Foxp3+ regulatory T cells (Treg), through a mechanism that appears to be dependent on the avidity and duration of TCR signaling and TGF-B and cytokine-mediated survival, such as IL-2, IL-7, and IL-15 (27, 28). Treg cells are  $CD4^+$  T lymphocytes, characterized by the expression of the transcription factor FOXP3, surface expression of CTLA-4, the glucocorticoid-induced TNF receptor family related protein (GITR) and CD25 (the IL-2 receptor alpha chain).

Tregs can also differentiate from CD4<sup>+</sup> T cells in the periphery of the immune system (peripheral Tregs) or within the thymus (central Tregs); the latter maturing under the direct influence of the thymic microenvironment (29). These thymus-derived cells represent the majority of Tregs in the periphery compared to Tregs differentiated from conventional naïve T cells (30).

In the periphery, after encountering the specific antigen and being activated, naïve T CD4<sup>+</sup> cells even differentiate into types 1, 2 or 17 T-helper (Th) cells, among others, depending on the stimulus they receive. Although morphologically indistinguishable, these cells present distinct patterns of secreted cytokines and, consequently, specific responses (31).

CD8<sup>+</sup> T cells, also known as cytotoxic T lymphocytes, are very important in the immune defense against intracellular pathogenic agents and also in the surveillance of tumor formation, acting through the induction of the cell death pathway (apoptosis) in the target cell (32).

## 2 T-cell immune responses in Duchenne Muscular Dystrophy

Several studies have aimed at clarifying the participation of the immune system in the pathophysiology of DMD. During the disease process, immediately after muscle damage, degenerating muscle fibers release Danger Associated Molecular Patterns (DAMP), in addition to pro-inflammatory cytokines and chemokines which attract innate immune cells. Thus, the muscle is first invaded by mast cells and neutrophils and a proinflammatory response dominates the scene with the stimulation of pro-inflammatory macrophages and proliferation of Th1 lymphocytes (33). During muscle repair, macrophages first acquire a pro-inflammatory phenotype. In this phase, proliferation of myoblast progenitors and initial differentiation occurs, followed by a switch from proinflammatory into antiinflammatory macrophages, necessary to resolve inflammation and promote myofiber growth, as shown by in vivo and ex vivo analysis using a murine model (33, 34). These events are crucial for the whole regenerative process and depend on T cells. In fact, it has been shown that genetic deletion of CD8<sup>+</sup> T lymphocytes impairs pro-inflammatory macrophage infiltration due to the absence of the chemokine CCL2 and is likely to be related to a defective muscle regeneration in a cardiotoxin-induced mouse skeletal muscle injury/regeneration model (35). These data indicate the importance of chemokines secreted by CD8<sup>+</sup> T cells upon monocyte/macrophage migration to the muscle (Figure 1A). Moreover, Rag2<sup>-</sup>Il2rb<sup>-</sup>Dmd<sup>-</sup> immunodeficient mice, which lack T, B, and NK cell activity, presented a delayed degeneration/ regeneration process when compared with the immunocompetent mdx mice (36).

In DMD it has been proposed that the lack of dystrophin has a consequence on muscle cell injury resulting in the release of intramuscular antigens that can be specifically recognized by cells of the adaptive immune response, such as B and T lymphocytes (37). Accordingly, T cells detected within the DMD muscle tissue predominantly express TCR belonging to the V $\beta$ 2 family (38). In particular, VB expansions point to T-cell clones that are selected during these immune responses, suggesting that T-lymphocytes expressing Vβ2 are involved in an antigen-driven late effector cellmediated immune response. Moreover, most DMD patients exhibit a conserved sequence of four amino acids in the complementaritydetermining region (CDR) 3 of the VB2 TCR, further suggesting specific recognition of muscle antigens (38, 39). Similarly, in the mdx mouse, T cells expressing V $\beta$ 8.1/8.2 predominated the pool of all TCR-V $\beta^+$  expressing cells. Moreover, the V $\beta$ 8.1/8.2-expressing T cells were not present in the spleen, suggesting they may expand in the dystrophic mdx muscle (40). It should be noted that DMD muscle fibers were invaded by  $CD8^+$  T cells expressed MHC class I molecules on their surfaces (41). Expression of MHC I is increased in dystrophic muscle as observed in myofibers invaded by  $CD8^+$  T cells compared to muscle from healthy individuals, which have little expression of MHC I (41, 42). Once activated, cytotoxic  $CD8^+$  T lymphocytes could then migrate and recognize specific peptides on the surface of muscle fibers triggering the release of death-related molecules including perforin, granzyme and tumor necrosis factor (TNF)- $\alpha$ , resulting in consequent tissue injury (43).

The analysis of a cohort of 75 DMD patients, has revealed that there is an increase in the percentage of circulating CD4<sup>+</sup> and CD8<sup>+</sup> T lymphocytes expressing high levels of CD49d (the alpha chain of the VLA-4 integrin) and this is positively correlated with a faster disease progression. Furthermore, T cells from patients with a more rapid disease progression exhibited higher trans-endothelial and fibronectin-driven migratory responses when compared to healthy individuals (44). The DMD patients with poorest prognosis who lost ambulation before ten years of age included in this study showed an increase in VLA-4 expression on T cells and a higher expression of the interstitial VLA-4 ligand, fibronectin, within the muscle. Furthermore, CD49d<sup>+</sup>CD4<sup>+</sup>CD3<sup>+</sup> as well as CD49d<sup>+</sup>CD8<sup>+</sup>CD3<sup>+</sup> T lymphocytes were seen intermingled with the fibronectin-containing network of the injured muscle (seen in Figure 1B). Lastly, T cells from DMD patients adhered more strongly to myotubes compared to circulating T cells from healthy subjects (44). Similar results were found in an animal model for DMD, namely the Golden Retriever Muscular Dystrophy dog. Higher numbers of circulating CD4<sup>+</sup>CD49d<sup>high</sup> T cells at early stages of the disease were found to be highly predictive for the loss of ambulation of the dogs before 6 months of age (45).

Increased expression of interferon (IFN)- $\gamma$ , transforming growth factor (TGF)- $\beta$  and chemokines, such as CCL14, CCL2, CXCL12 and CXCL14 has also been observed in the muscle of DMD patients (46). Considering that extracellular matrix (ECM) elements can interact directly with immune cells functioning as a substrate for binding soluble factors such as cytokines and chemokines, it is plausible to think that a combined action involving an increase in fibronectin expression and in chemokines/cytokines could enhance the recruitment of inflammatory cells to this area, thus maintaining and eventually enhancing inflammation and fibrosis.

As mentioned above and taking into account the studies performed in DMD patients, it is conceivable that the muscle damage initially caused by the absence of dystrophin could result in the exposure of antigens on the surface of muscle fibers. Activated T lymphocytes with high expression of CD49d (CD49d<sup>high</sup>) could migrate to muscle tissue directed by a chemotactic/haptotactic gradient. Moreover, CD49d<sup>high</sup> T cell subsets obtained from DMD patients exhibited a higher *in vitro* migration capacity across endothelia and through fibronectin, when compared to CD49d<sup>low</sup> T cell subpopulations from healthy donors (44). Once within the muscular tissue, CD8<sup>+</sup>CD49d<sup>high</sup> T lymphocytes could recognize antigens on the surface of the fibers causing their destruction.

Overall, effector T cells are important in DMD pathophysiology (43) and disease progression with a special role for the CD49d<sup>high</sup> T cell subpopulations, which in turn can be used as a prognostic



#### FIGURE 1

CD49d<sup>high</sup> T lymphocytes are involved in the Duchenne Muscular Dystrophy (DMD) pathophysiology. Effector T cells are important in DMD pathophysiology and progression, with a special role for the CD49d<sup>high</sup> T cell subpopulations. **(A)** During the post-injury phase of muscle repair, macrophages initially acquire a pro-inflammatory phenotype followed by a switch into anti-inflammatory macrophages, necessary to solve inflammation and promote myofiber growth. These events are crucial for the regenerative process and depend on T cells, notably CD8<sup>+</sup> and regulatory T cells. **(B)** Immunostaining in a quadriceps muscle biopsy obtained from a DMD patient at the onset of the disease, showing the *in situ* detection of CD8<sup>+</sup> (green) CD49d<sup>+</sup> (red) cells. Nuclei were visualized using DAPI (blue). **(C)** The scheme illustrates the notion that CD49d inhibitors can be envisioned as a therapeutic approach to decrease inflammation-mediated tissue damage, by blocking VLA-4-mediated T lymphocyte transendothelial migration; (1) as well as the intramuscular migration of the cells through a fibronectin-containing extracellular network; (2) and the CD49d<sup>high</sup> T cell adhesion to myoblasts/myotubes, (3) with consequent inhibition of muscle cell apoptosis. Figure created using the Adobe Illustrator software. Immunofluorescence micrography gently provided by Dr. Fernanda Pinto Mariz (Pediatric Institute, Federal University of Rio de Janeiro, Brazil).

10.3389/fimmu.2023.1202834

marker of disease progression (44). Moreover, CD49d inhibitors can be envisioned as a therapeutic approach to decrease inflammation-mediated tissue damage (Figure 1C), with consequent amelioration in the quality of life of DMD patients. To test this approach a phase II clinical trial using small interfering RNA (siRNA) to inhibit CD49d expression in T cells in adolescent males aged 10 to 18 years with a diagnosis of non-ambulatory DMD for at least 3 months is ongoing (http://www.anzctr.org.au/Trial/ Registration/TrialReview.aspx?ACTRN=12618000970246) (47, 48).

Within the context of T-cell biology in DMD, is also of note the presence and action of DCs, although the studies on this subject remain rather scarce. DCs are at the interface between innate and adaptive immunity, being professional antigen-presenting cells (APCs), presenting foreign antigens as peptides for T cell recognition and mediating their activation and differentiation. DCs are present in normal skeletal muscle and MHC II<sup>+</sup> cells increase after injury (49). The participation of these cells to the disease process is plausible, since they express Toll-like receptor (TLR) 7, which can bind to RNA and trigger cytokine production, enhancing or at least maintaining the cycle of inflammation/ degeneration/regeneration (50). Among the cytokines released, TGF- $\beta$  seems to be strongly induced in symptomatic DMD patients (51, 52). It is interesting to note that, at least in vitro, DCs interact with myoblasts, increasing their proliferation and migration, as well as cytokine secretion by these cells (53).

# 3 Amyotrophic Lateral Sclerosis and T cell response

In contrast to DMD, ALS is a composite disease with non-cell autonomous mechanisms contributing to neurodegeneration. For instance, activation of glial cells such as microglia and astrocytes is a typical hallmark, contributing to the disease phenotype through a local inflammatory response (54). In the healthy adult brain, resting microglia monitor the surrounding microenvironment, interact with nearby cells, and play a key role in the central nervous system (CNS) development, homeostasis and repair (55-57). Through pattern recognition receptors (PRR), microglial cells sense molecules frequently found in pathogens (the Pathogen-Associated Molecular Patterns; PAMPs) or molecules released by damaged cells, the DAMPs, leading to the production of soluble molecules and modifications in their interactions with neurons (58). The accumulation of misfolded proteins in the MNs is probably one of the first events in neurodegeneration, leading to inflammation following the release of DAMPs, such as reactive oxygen species (ROS), which, in turn, activate glial and immune cells (59).

When activated, microglia exist in a variety of phenotypic states, with two extremes, as depicted in Figure 2. One corresponds to a protective phenotype, which produces anti-inflammatory cytokines, such as interleukin (IL)-4, IL-13, IL-10, as well as progranulin, and neurotrophic factors, such as insulin-like growth factor (IGF)-1, which contributes to neuroprotection by suppressing inflammation, cleaning cell debris and promoting ECM reconstruction (60–62). The other extreme is a harmful phenotype, capable of producing ROS and pro-inflammatory mediators, including the subunit of

nicotinamide-adenine-dinucleotide-phosphate (NADPH) oxidase (NOX) 2, IL-1α, IL-1β, IL-6, IL-12, IL-23, TNF-α, inducible nitric oxide synthase (iNOS), matrix metalloproteinase (MMP) 12 and IFNs, leading to neurodegeneration (61, 63). Studies using animal models of ALS have shown that microglial activation occurs before disease onset, concomitant with loss of the neuromuscular junctions (NMJ), and increases during disease progression (64-66). The NMJ, the functional unit of skeletal muscles, is composed of terminal axons of presynaptic MN, post synaptic skeletal muscle and associated glial cell types, the Schwann cells (67). Results from different ALS models have shown that MN degeneration begins in terminal axons and ascends to the cell body in a phenomenon called "axonal dieback" (68-70). Expression of the mutated protein only in skeletal muscles leads to abnormalities in the NMJ due to mitochondrial dysfunction and activation of cell death pathways (71). Transgenic mice engineered to overexpress mSOD1 in specific tissues show MN degeneration only when the mutant protein is expressed in both neurons and microglia or astrocytes (70, 72). Similarly, selective reduction of mSOD1 in microglia or astrocytes of ALS mice increases their survival (73, 74).

Astrocytes are the most abundant glial cells and a key component in the CNS environment, being involved in a variety of homeostatic functions, through the secretion of neurotrophic factors (75). They are capable of amplifying either neuroprotective or neurotoxic effects mediated by the microglia and can directly or indirectly influence the fate of MNs. Experiments using a cell transplantation approach have revealed a detrimental role for mSOD1-ALS astrocytes (76). In fact, reactive astrocytes surround both upper and lower degenerating MNs and several molecules have been linked to the astrocyte-mediated toxicity in ALS. For example, ALS astrocytes display a reduced ability to provide metabolic support to MNs and to effectively regulate the extracellular levels of neurotransmitters, such as glutamate. Excessive stimulation of glutamate receptors causes excitotoxicity, thereby increasing the influx of calcium, which can be toxic to neurons (Figure 2). Other molecules secreted by activated astrocytes that can cause neurotoxicity are pro-inflammatory cytokines and inflammatory mediators, such as NOX2, iNOS, ROS, IFN-y, prostaglandin D2 (PGD2) and TGF- $\beta$  (77–81). In fact, it has been demonstrated that astrocytes from ALS patients secrete more IL-6 and have elevated levels of NOX2, iNOS and ROS production, exacerbating the inflammatory response and MN degeneration (82, 83).

The link between microglia, astrocytes and the adaptive immune system is demonstrated by the crosstalk between these activated resident cells and the CNS-infiltrating peripheral immune components such as T cells. Accordingly, reactive microglia coupled with infiltrating T cells were found in autopsies of ALS patients (84). The CD4<sup>+</sup> T cell infiltration is likely related to an increase in the chemokine CCL2 and microglial activation (85–87). Moreover, the expression of the chemokine receptors CXCR3 and CXCR4, as well as the chemokines CCL2, and CCL5, are also increased in T cells from ALS patients, compared to healthy controls (88). Recent data highlighted the protective role of these infiltrating CD4<sup>+</sup> cells in ALS. During the dismantling of the neuromuscular junction, with axonal retraction and demyelination, several antigens are presented



#### FIGURE 2

Amyotrophic Lateral Sclerosis (ALS) different protective versus harmful phenotypic responses. Schematic representation of T-cell implication in the inflammatory events in ALS caused by toxic accumulation of mutated proteins and consequent release of damage-associated molecular patterns (DAMPs). Antigens released during the axonal degeneration are carried by antigen-presenting cells, like dendritic cells, until draining lymph nodes, where they are presented to naïve T cells. The activation of CD4<sup>+</sup> T cells generates different subtypes of T helper (Th) cells. In the CNS, quiescent microglia and astrocytes are activated and secrete pro-inflammatory factors, including oxygen-reactive species (ROS), tumor necrosis factor- $\alpha$  (TNF- $\alpha$ ), interferon gamma (IFN- $\gamma$ ). These factors attract infiltrating Th1 and Th2 CD4<sup>+</sup> cells, increasing inflammation by releasing TNF- $\alpha$ , II-6 and IFN- $\gamma$ , for instance. Likewise, the pro-inflammatory cytokines inhibit the development of the anti-inflammatory regulatory T (Treg) cells. Further, astrocytes in ALS display reduced expression of the glutamate transporter-1 (GLT-1) and fail to effectively regulate the extracellular levels of glutamate. The antigens released during axonal dismantling also activate protective responses that contribute to neuroprotection, by suppressing inflammation, cleaning cellular debris and promoting extracellular matrix rebuilding. There is activation of protective microglia, which produce anti-inflammatory cytokines, as well as neurotrophic factors and Th2 CD4<sup>+</sup> cells, which in turn confer neuroprotection by producing anti-inflammatory favoring the neuroprotective effects of activated microglia. Anti-inflammatory Treg cells are also increased in number, favoring the neuroprotective response, by inhibiting the harmful inflammatory environment, through the secretion of IL10, IL-4 and TGF- $\beta$ . Figure created using the Adobe Illustrator software.

at the site of the injury, which leads to an immune response (16). However, whether the antigenic origin is neuronal or muscular is not yet known, although it seems that the debris formed during axonal disconnection from its target musculature are the source of antigens that lead to activation of a neuroprotective immune response. Consequently, there is an induction of debris clearance and an initial repair phase culminating in exacerbation of disease progression (19).

T cells isolated from the cerebrospinal fluid of ALS patients appear to have undergone clonal expansion, also suggesting antigen-mediated activation (89). Jones et al. (2015) established that MHC class II-mediated antigen presentation is necessary for CD4<sup>+</sup> T cells to confer neuroprotection. Following axonal degeneration, MHC class II-expressing antigen presenting cells in the draining lymph nodes initially activate peripheral CD4<sup>+</sup> T cells, which are subsequently reactivated by CNS-residing MHC class  $II^+$  APCs expressing the same antigen, also supporting the idea that glial cells can present antigens to T cells (90, 91).

Th2 CD4<sup>+</sup> cells confer neuroprotection by producing antiinflammatory cytokines, such as IL-4, IL-5 and IL-13, which in turn enhance the neuroprotective effects of activated microglia (92).

However, the antigen presentation following axonal disconnection leads to the differentiation of several CD4+ T cell subsets in the draining lymph nodes and a predominant Th1 and Th17 response is observed in mSOD1 mice (93). Th1 and Th17 cells are CD4<sup>+</sup> T cell subtypes that promote neuroinflammation, by producing proinflammatory cytokines. Th1 cells produce IFN- $\gamma$  that promotes activation of a harmful microglia, increasing the release of ROS and NO. In turn, the microglia exacerbate the proliferation and function of Th1 cells (66, 92, 94). Neurotoxic effects are also attributed to IL-6 released by Th1 cells. In addition, Th1 and Th17 cells produce high amounts of TNF- $\alpha$ . Both, IL-6 and TNF- $\alpha$ , are known to inhibit the development of the anti-inflammatory IL-10-producing Treg cells (95, 96).

Whether there is a difference in the number of T cells in ALS is still controversial. There are studies showing both decreased or elevated numbers of  $CD4^+$  T cells during disease progression in the peripheral blood of ALS patients in comparison to healthy controls (97, 98). To elucidate this question, the different  $CD4^+$  T cell subpopulations should be considered, since changes in the total number may correlate with modulation in specific  $CD4^+$  T-cell subsets rather than  $CD4^+$  T cells as a whole. Indeed, it seems there is a shift from a supportive Th2 to a deleterious Th1 response as the disease progresses. In this sense, some studies showed reduced numbers of  $CD4^+$  T cells (99) and increased proportions of proinflammatory effector T cells (100, 101) in ALS patients.

Although over the course of ALS, part of the CD4<sup>+</sup> T cell response is harmful, it has been shown that total CD4<sup>+</sup> T cell deletion accelerates the disease. When mSOD1 mice were crossed with T cell receptor  $\beta$  chain (TCR $\beta$ ) deficient mice (TCR $\beta^{-/-}$ ; that lack TCR $\alpha\beta^+$ CD4<sup>+</sup> and TCR $\alpha\beta^+$ CD8<sup>+</sup> T cells), there was an increased rate of disease progression, although the time of disease onset was not modified (90). Similar results were observed when mSOD1 mice were crossed with Rag2<sup>-/-</sup> mice (lacking B and T lymphocytes) or CD4<sup>-/-</sup> mice (deficient in CD4<sup>+</sup> T lymphocytes), thus attributing a neuroprotective role to the CD4<sup>+</sup> T cells, but not to the CD4<sup>+</sup> T or B cells (102). Accordingly, the reconstruction of the CD4<sup>+</sup> T-cell populations following wild-type (WT) bone marrow transplantation to mSOD1/RAG2<sup>-/-</sup> recipient mice in a stable phase of the disease conferred neuroprotection and increased survival (102).

Interestingly, in a slow initial stage of ALS, neuroprotective activity has been suggested by increasing the levels of CD4<sup>+</sup>CD25<sup>hi</sup>FoxP3<sup>+</sup> Treg cells. In contrast, the number of circulating Tregs decrease significantly with the rapid progression of the disease. Therefore, it has been suggested that a decrease in Tregs, together with a decrease in FoxP3 expression levels, are associated with short-term survival in ALS patients (103). It should be noted that dysfunction in Treg activity can directly influence the activation of pro-inflammatory microglia, contributing to motor neuron excitotoxicity (104). ALS patients demonstrate an unbalanced ratio among T cell subgroups with an increased number of pro-inflammatory T cells, including Th1 and Th17, and reduced Treg/Th2 lymphocytes (105). In the same way, adoptive transfer of endogenous mSOD1 Tregs during the stable phase of ALS to mSOD1/RAG2<sup>-/-</sup> recipient mice promoted neuroprotection, postponed the disease onset and considerably increased survival (106).

Recent studies have further highlighted the immunologic alterations in ALS patients and its correlation with disease progression. Proteomic analysis of plasma and peripheral blood mononuclear cells (PBMCs) from ALS patients demonstrated the activation of molecular pathways involved in immunoregulation and cell senescence in faster progressing ALS patients (107). Indeed, Yildiz and colleagues showed that lymphocytes from ALS patients are skewed towards a senescent and late memory state when compared with those from healthy controls (108). Immunosenescence is a process reflecting immune dysfunction, by remodeling of the immune system, with reduction in the number of peripheral blood naïve cells and a relative increase in the frequency of memory cells. This leads to a decline in efficacy of the immune response during aging and is closely correlated with the development of infections, autoimmune diseases, and malignant tumors (109).

In addition, a study comparing 89 newly diagnosed ALS patients in Sweden observed that high frequency of  $CD4^+$  effector T cells in the blood and cerebrospinal fluid was associated with poor survival, whereas a high frequency of activated Treg in the blood was associated with a better survival (110).

Overall, these studies attributed the neuroprotective capacity of the CD4<sup>+</sup> T cells to CD4<sup>+</sup>CD25<sup>+</sup> and CD4<sup>+</sup>CD25<sup>+</sup>FoxP3<sup>+</sup> Treg cells and suggest that the T cell phenotypes at the time of diagnosis can be good predictors of disease prognosis, as will be described below.

# 4 Immune crosstalk in DMD and ALS: role of regulatory T cells

Migration and maintenance of Tregs in the muscle depends on at least two different mechanisms (1): The first is controlled by the ATP/P2X axis and was initially demonstrated in the mdx mouse model of DMD (111), in which blocking the ATP/P2X7 interaction with periodate-oxidized ATP resulted in an increased number of Treg within the injured muscle. In the absence of ATP/P2X7 signaling, not only do Tregs migrate more intensely to the lesion but muscle damage is reduced, indicating that ATP, released by dying cells, downregulates Treg migration, and that, in the absence of Tregs, muscle damage is more severe (111) (2). Secondly, there is an effect of the IL-33 mediated mechanism of Treg arrival and maintenance within the muscle. IL-33 is an alarmin produced by fibroadipogenic progenitors (FAPs) and skeletal muscle stem cells which can bind to the IL-1 receptor-like 1 protein (also named ST2) present on CD4<sup>+</sup> T cells (including Tregs), macrophages and FAPs. In fact, FAP-derived IL-33 is crucial for Treg accumulation within the muscle, and consequently, muscle regeneration (Figure 1A) (112).

It is well known that both T cells (CD4<sup>+</sup> or CD8<sup>+</sup>) need to engage their antigen recognition receptor through MHC interaction, so that they can interact specifically with other cells. Studies in mouse models revealed that after transferring muscle specific T cells to a normal recipient, muscle Tregs were only observed within the muscle. In addition, studies in mdx mice have shown that antigen specific Treg cells induced an accelerated rate of regeneration when compared to polyclonal Treg cells (113). Similarly, muscle injury and inflammation were mitigated by the expansion of the Treg cell population, being exacerbated in conditions of Treg cell depletion (114).

As mentioned previously, in ALS, the neuroprotective capacity of the CD4<sup>+</sup> T cells has been attributed to the CD4<sup>+</sup>CD25<sup>+</sup>FoxP3<sup>+</sup> Treg cell fraction. It was observed that Tregs in the slow phase of the disease can support a microglial anti-inflammatory phenotype and the expression of neurotrophic factors in the spinal cord through IL-4 secretion. In parallel, they suppress effector T cell proliferation via the secretion of IL-4, IL-10 and TGF- $\beta$  (115). Likewise, when mSOD1 Treg isolated from spleen and lymph nodes are co-cultured with mSOD1 adult microglia they inhibit NOX2 production and induce NO synthase expression, through IL-4 secretion (115). Thus, infiltration of CD4<sup>+</sup>CD25<sup>+</sup>FoxP3<sup>+</sup> Tregs in the CNS suppresses neuroinflammation and promotes activation of protective microglia in murine models of ALS (116), with an up-regulation of anti-inflammatory cytokines with a predominance of a neuroprotective phenotype (106).

In contrast, Tregs isolated from mSOD1 mice at a later stage of the disease fail to prolong survival of mSOD1/RAG2<sup>-/-</sup> recipient mice (106). As the disease progresses, the level of pro-inflammatory cytokines increases, while the number of Treg declines, possibly due to the loss of FoxP3 expression in Treg cells (106). For instance, increased TNF- $\alpha$  release by Th1 and Th17 cells induces Treg dysfunction by inhibiting FoxP3 phosphorylation (96). Moreover, elevated levels of IL-6 released by Th1 cells inhibit the development of FoxP3<sup>+</sup> Tregs (117), leading to the loss of the ability of Treg cells to sustain an anti-inflammatory response in the microglia, exacerbating a harmful phenotype (Figure 2).

Similar to what has been reported in mice, in ALS patients the endogenous Treg cells increase at an early slowly-progressing disease stage. During the course of the disease the development of Tregs is inhibited and their number decreases in parallel to a rapid ALS progression, likely due to the loss of FoxP3 (106, 118). Moreover, in patients, specific Treg subtypes have been correlated with the rate of ALS progression. In the blood of ALS patients with rapid disease progression, there is a reduction in the number of CD45RO<sup>+</sup> Tregs, the memory cells, which are classified as functionally active with higher levels of FoxP3 expression, when compared to the resting naïve CD45RA<sup>+</sup> Treg cells (116). Moreover, the rate and severity of ALS progression was inversely correlated with the total Treg counts, total FoxP3<sup>+</sup> Treg counts and effector CD45RO<sup>+</sup> Treg counts, while the number of resting CD45RA<sup>+</sup> Treg did not correlate with disease progression (116).

In addition to this reduction in number, Tregs from patients with rapid disease progression have an impaired ability to inhibit the proliferation of their corresponding effector T cells, when compared to those of patients with slow disease progression, or with healthy controls (103). The Treg suppression capacity was correlated with FoxP3 mRNA and protein levels, indicating that a stable expression is essential to maintain their regulatory capacity (103).

One major problem in the treatment of ALS is that to be successful, the treatment needs to be administered at the time of diagnosis. At present it is estimated that there is a delay of more than one year between disease onset and clinical diagnosis (119), and most of the patients are currently diagnosed at mid stage of progression where reversing neurodegeneration may be very difficult, demonstrating the importance of a predictive factor for the disease. For this reason, increasing the number and function of Treg cells seems like a good therapeutic strategy and some clinical trials aim to target the Treg cells, either by promoting their expansion *in vivo* or by *in vitro* functional stimulation followed by autologous transplantation, as a way of improving quality of life of ALS patients (104). One trial ongoing is MIROCALS, which is a phase II study of Low-dose IL-2 (Ld-IL-2) as a Treg Enhancer for Controlling Neuroinflammation in Newly Diagnosed ALS Patients (https://clinicaltrials.gov/study/NCT03039673?a=1).

## 5 Thymus defects underlying peripheral T cell biology abnormalities in DMD and ALS

The thymus is a primary lymphoid organ responsible for the maturation and differentiation of T lymphocytes (120, 121). During this process, key maturation events correspond to the positive and negative selection, both driven by interactions between developing thymocytes and microenvironmental cells (particularly thymic epithelial cells (TECs), involving both the MHC complex and TCR, respectively present in microenvironmental components and developing thymocytes (122). Negative selection largely prevents autoimmunity and is directly under the control of the transcription factors Aire and Fezf2 expressed by TEC in the medullary region of the thymic lobules (121). Negative selection induces thymocyte death by apoptosis; positive selection rescues developing thymocytes from death and induces their progression towards CD4<sup>+</sup> or CD8<sup>+</sup> SP thymocytes, which eventually leave the organ to colonize the peripheral lymphoid organs (123). Lastly, the intrathymic production of central Tregs and subsequent export towards the periphery of the immune system further prevents the development of autoimmunity. Accordingly, central Treg deficiency is related to autoimmune events (124).

Interestingly, the thymus may play a role in the skeletal muscle regeneration process. A study carried out in a model of induced skeletal muscle injury showed that the mean cross-sectional area of regenerating myofibers and the number of myofibers in 8-10 weeks of age thymectomized mice were impaired compared to euthymic controls (125). In addition to this defect in muscle regeneration, thymectomized mice also showed decreased levels of myoblast determination proteins, MyoD 1 and myogenin (MyoG), important myogenic factors for skeletal muscle regeneration. This study further showed that the number of Pax7-positive satellite cells in the muscle was increased in the injured control, compared to thymectomized mice, providing evidence that the thymus can regulate the number of satellite cells during this process of muscle degeneration and regeneration. Interestingly, the authors showed in vitro that thymocyte-conditioned medium promoted the proliferation of satellite cells. The CD3<sup>+</sup>CD4<sup>+</sup>CD8<sup>+</sup> thymocyte subset significantly induced proliferation in satellite cells, although CD4+CD8- and CD4-CD8+ SP thymocytes also displayed a significant effect compared to the control. In contrast, Treg cells had no significant effect in a muscle injury model of thymectomized mice (125). In view of the above data, it seems that the thymus plays a role in the skeletal muscle by its influence on regeneration.

Pioneer studies had previously revealed a thymic atrophy in mdx dystrophic mice, with a decrease in thymocyte numbers, densification of the TEC network and increase in ECM contents. Of note, the peak of thymocyte depletion coincided with the stage of

disease severity (126). These findings have recently been confirmed and expanded by Farini and coworkers, who showed that the disorganized thymic morphology was correlated with an abnormal intrathymic T cell development and changes in the maturation stages of DN thymocytes. These events coincide with an increase in the relative numbers of SP cells and Tregs in the thymus of mdx mice compared to controls (127). In addition, the authors showed that transplanting the involuted thymus of mdx animals into nude mice, resulted in a significant increase in the expression of the transcription factors RORyt and T-bet, as well as an increase in the numbers of CD4<sup>+</sup> and CD8<sup>+</sup> T cells infiltrating the muscle, compared to nude animals that received the thymus of healthy donors. These results indicate that the process of thymic involution may be a determining factor in DMD pathophysiology and one of the postulated mechanisms was a dysregulation in the release of autoimmune or Treg cells (33).

Despite the lack of studies that aim to characterize in detail the role of the thymus in ALS, evidence suggests that thymic function is compromised in ALS-SOD1 patients and in the mSOD1 mouse model. In 2010, Seksenyan and colleagues demonstrated a thymic defect in parallel to the MN dysfunction in mSOD1 mice and ALS patients. In this study, the authors observed a reduction in the absolute number of total thymocytes in mSOD1 mice. This reduction was reflected in all thymocyte populations, defined by the expression of CD4 and CD8 coreceptors, i.e., CD4<sup>-</sup>CD8<sup>-</sup> DN, CD4<sup>+</sup>CD8<sup>+</sup> DP, CD4<sup>+</sup>CD8<sup>-</sup> and CD4<sup>-</sup>CD8<sup>+</sup> SP cells. These changes were also observed in ALS patients at the final disease stage. Moreover, sections of the thymus of mSOD1 mice in advanced stages of the disease stained with hematoxylin and eosin, showed a loss of definition in the cortical and medullary regions (128), which may reveal an abnormal thymic architecture during ALS progression and, consequently, impaired T cell development. Indeed, the analysis of signal joint T-cell receptor rearrangement excision circle (sjTREC) in PBMCs from ALS patients, revealed decreased numbers of sjTREC compared to healthy controls regardless of the age of the patients, possibly indicating a reduction in the export of naïve T cells from the thymus of ALS patients (128).

## 6 Concluding remarks

The data discussed above provide evidence concerning the complexity of the cellular and molecular interactions involving the immune system in two very different neuromuscular disorders, DMD and ALS, which may reflect similar defects common to other neuromuscular diseases. Distinct types of interactions have been described, comprising among others cytokine/chemokine production, cell-cell and cell-matrix interactions between T lymphocytes and other cells of the immune system with target cells of the muscular or nervous tissues. Although this reveals the complexity of the system, it also can be envisioned as a window of opportunity to design novel therapeutic strategies (including synthetic moieties, cell and gene therapy, as well as immunotherapy) by acting upon one or more targets. In this respect, ongoing clinical trials on DMD patients through VLA-4 inhibition, and in ALS, focusing on regulatory T cells, have both shown promising results.

### Author contributions

JL and WS made extensive review of the literature listed and elaborated different sections of the review. JL, LT, WS, DM-d-C, and PS drafted and drew the figures. JL, WS, and PS conceived, designed and supervised the review. VM, GB-B, and DM-d-C edited the manuscript. All authors contributed to the article and approved the submitted version.

## Funding

This work was supported by Association Institut de Myologie, the Inserm-Fiocruz Associated Laboratory on Cell Therapy and Immunotherapy and the Capes-Cofecub program. WS and DAMdC are funded by CNPq, CAPES and FAPERJ (Brazil) and the MercoSur Fund for Structural Convergence (FOCEM). Participation of WS, DAMdC and LPGT is within the framework of the Brazilian National Institute of Science and Technology on Neuroimmunomodulation, the Rio de Janeiro Research Network on Neuroinflammation and the INOVA-IOC research network on Neuroimmunomodulation.

## Acknowledgments

We thank Dr. Fernanda Pinto Mariz (Pediatric Institute, Federal University of Rio de Janeiro, Brazil) for kindly providing the immunofluorescence micrography.

## Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

## Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

## References

1. Ryder S, Leadley RM, Armstrong N, Westwood M, De Kock S, Butt T, et al. The burden, epidemiology, costs and treatment for Duchenne muscular dystrophy: An evidence review. *Orphanet J Rare Dis* (2017) 12:1–21. doi: 10.1186/s13023-017-0631-3

2. Arahata K, Engel AG. Monoclonal antibody analysis of mononuclear cells in myopadues. I: quantitation of subsets according to diagnosis and sites of accumulation and demonstration and counts of muscle fibers invaded by T cells. *Ann Neurol* (1984) 16:193–208. doi: 10.1002/ana.410160206

3. Bulfield G, Siller WG, Wight PAL, Mooret KJ. X chromosome-linked muscular dystrophy (mdx) in the mouse (animal model). *Proc Nati Acad Sci USA* (1984) 81:1189–92. doi: 10.1073/pnas.81.4.1189

4. Willmann R, Possekel S, Dubach-Powell J, Meier T, Ruegg MA. Mammalian animal models for Duchenne muscular dystrophy. *Neuromuscular Disord* (2009) 19:241–9. doi: 10.1016/j.nmd.2008.11.015

5. Araki E, Nakamura K, Nakao K, Kameya S, Kobayashi O, Nonaka I, et al. Targeted disruption of exon 52 in the mouse dystrophin gene induced muscle degeneration similar to that observed in duchenne muscular dystrophy. *Biochem Biophys Res Commun* (1997) 238:492–7. doi: 10.1006/bbrc.1997.7328

6. Cox GA, Cole NM, Matsumura K, Phelps SF, Hauschka SD, Campbell KP, et al. Overexpression of dystrophin in transgenic mdx mice eliminates dystrophic symptoms without toxicity. *Nature* (1993) 364:725–9. doi: 10.1038/364725a0

7. Kudoh H, Ikeda H, Kakitani M, Ueda A, Hayasaka M, Tomizuka K, et al. A new model mouse for Duchenne muscular dystrophy produced by 2.4 Mb deletion of dystrophin gene using Cre-loxP recombination system. *Biochem Biophys Res Commun* (2005) 328:507–16. doi: 10.1016/j.bbrc.2004.12.191

8. Coley WD, Bogdanik L, Vila MC, Yu Q, van der Meulen JH, Rayavarapu S, et al. Effect of genetic background on the dystrophic phenotype in mdx mice. *Hum Mol Genet* (2016) 25:130–45. doi: 10.1093/hmg/ddv460

9. Gordish-Dressman H, Willmann R, Dalle Pazze L, Kreibich A, Van Putten M, Heydemann A, et al. "of mice and measures": A project to improve how we advance duchenne muscular dystrophy therapies to the clinic. *J Neuromuscul Dis* (2018) 5:407– 17. doi: 10.3233/JND-180324

10. van Putten M, Putker K, Overzier M, Adamzek WA, Pasteuning-Vuhman S, Plomp JJ, et al. Natural disease history of the D2-mdx mouse model for Duchenne muscular dystrophy. *FASEB J* (2019) 33:8110–24. doi: 10.1096/fj.201802488R

11. Deconinck AE, Rafael JA. Utrophin-dystrophin-deficient mice as a model for duchenne muscular dystrophy. *Cell* (1997) 90:717-27. doi: 10.1016/S0092-8674(00) 80532-2

12. Grady RM, Teng H, Nichol MC. Skeletal and cardiac myopathies in mice lacking utrophin and dystrophin: A model for duchenne muscular dystrophy. *Cell* (1997) 90:729–38. doi: 10.1016/S0092-8674(00)80533-4

13. Guo C, Willem M, Werner A, Raivich G, Emerson M, Neyses L, et al. Absence of α7 integrin in dystrophin-deficient mice causes a myopathy similar to Duchenne muscular dystrophy. *Hum Mol Genet* (2006) 15:989–98. doi: 10.1093/hmg/ddl018

14. Rooney JE, Welser JV, Dechert MA, Flintoff-Dye NL, Kaufman SJ, Burkin DJ. Severe muscular dystrophy in mice that lack dystrophin and  $\alpha$ 7 integrin. J Cell Sci (2006) 119:2185–95. doi: 10.1242/jcs.02952

15. Valko K, Ciesla L. Amyotrophic lateral sclerosis. Prog Med Chem (2019) 58:63-117. doi: 10.1016/bs.pmch.2018.12.001

16. Iyer AK, Jones KJ, Sanders VM, Walker CL. Temporospatial analysis and new players in the immunology of amyotrophic lateral sclerosis. *Int J Mol Sci* (2018) 19:1–16. doi: 10.3390/ijms19020631

17. Bensimon G, Lacomblez L, Meininger VTeh ALS/ Riluzole Study Group. A controlled trial of riluzole in amyotrophic lateral sclerosis. N Engl J Med (1994) 330:585–91. doi: 10.1056/NEJM199403033300901

18. Ravits JM, La Spada AR. ALS motor phenotype heterogeneity, focality, and spread Deconstructing motor neuron degeneration. *Neurology* (2009) 8:805–11. doi: 10.1212/WNL.0b013e3181b6bbbd

19. Iyer AK, Jones KJ. A tale of motor neurons and CD4+ T cells: Moving forward by looking back. *Neural Regener Res* (2017) 12:562–5. doi: 10.4103/1673-5374.205086

20. Brooks BR, Miller RG, Swash M, Munsat TL. El Escorial revisited: Revised criteria for the diagnosis of amyotrophic lateral sclerosis. *ALS Other Motor Neuron Disord* (2000) 1:293–9. doi: 10.1080/146608200300079536

21. Rajabinejad M, Ranjbar S, Afshar Hezarkhani L, Salari F, Gorgin Karaji A, Rezaiemanesh A. Regulatory T cells for amyotrophic lateral sclerosis/motor neuron disease: A clinical and preclinical systematic review. *J Cell Physiol* (2020) 235:5030–40. doi: 10.1002/jcp.29401

22. Suzuki N, Nishiyama A, Warita H, Aoki M. Genetics of amyotrophic lateral sclerosis: seeking therapeutic targets in the era of gene therapy. *J Hum Genet* (2023) 68:131–52. doi: 10.1038/s10038-022-01055-8

23. Nardo G, Trolese MC, Tortarolo M, Vallarola A, Freschi M, Pasetto L, et al. New insights on the mechanisms of disease course variability in ALS from mutant SOD1 mouse models. *Brain Pathol* (2016) 26:237–47. doi: 10.1111/bpa.12351

24. Chiot A, Lobsiger CS, Boillée S. New insights on the disease contribution of neuroinflammation in amyotrophic lateral sclerosis. *Curr Opin Neurol* (2019) 32:764–70. doi: 10.1097/WCO.00000000000729

25. Carpenter AC, Bosselut R. Decision checkpoints in the thymus. *Nat Immunol* (2010) 11:666–73. doi: 10.1038/ni.1887

26. Germain RN. T-cell development and the CD4-CD8 lineage decision. *Nat Rev Immunol* (2002) 2:309–22. doi: 10.1038/nri798

27. Ohkura N, Sakaguchi S. Regulatory T cells: Roles of T cell receptor for their development and function. *Semin Immunopathol* (2010) 32:95–106. doi: 10.1007/s00281-010-0200-5

28. Josefowicz SZ, Lu LF, Rudensky AY. Regulatory T cells: Mechanisms of differentiation and function. *Annu Rev Immunol* (2012) 30:531-64. doi: 10.1146/ annurev.immunol.25.022106.141623

29. Owen DL, Mahmud SA, Sjaastad LE, Williams JB, Spanier JA, Simeonov DR, et al. Thymic regulatory T cells arise via two distinct developmental programs. *Nat Immunol* (2019) 20:195–205. doi: 10.1038/s41590-018-0289-6

30. Richards DM, Delacher M, Goldfarb Y, Kägebein D, Hofer AC, Abramson J, et al. Treg cell differentiation: from thymus to peripheral tissue. *Prog Mol Biol Transl Sci* (2015) 136:175–205. doi: 10.1016/bs.pmbts.2015.07.014

31. Swain SL, Bradley LM, Croft M, Tonkonogy S, Atkins G, Weinberg AD, et al. Helper T-cell subsets: phenotype, function and the role of lymphokines in regulating their development. *Immunol Rev* (1991) 123:115–44. doi: 10.1111/j.1600-065X.1991. tb00608.x

32. Barry M, Bleackley RC. Cytotoxic T lymphocytes: All roads lead to death. Nat Rev Immunol (2002) 2:401–9. doi: 10.1038/nri819

33. Tripodi L, Villa C, Molinaro D, Torrente Y, Farini A. The immune system in duchenne muscular dystrophy pathogenesis. *Biomedicines* (2021) 9:1–17. doi: 10.3390/biomedicines9101447

34. Arnold L, Henry A, Poron F, Baba-Amer Y, Van Rooijen N, Plonquet A, et al. Inflammatory monocytes recruited after skeletal muscle injury switch into antiinflammatory macrophages to support myogenesis. *J Exp Med* (2007) 204:1057– 69. doi: 10.1084/jem.20070075

35. Zhang J, Xiao Z, Qu C, Cui W, Wang X, Du J. CD8 T cells are involved in skeletal muscle regeneration through facilitating MCP-1 secretion and gr1high macrophage infiltration. *J Immunol* (2014) 193:5149–60. doi: 10.4049/jimmunol.1303486

36. Vallese D, Negroni E, Duguez S, Ferry A, Trollet C, Aamiri A, et al. The Rag2 -Il2rb - Dmd - Mouse: A novel dystrophic and immunodeficient model to assess innovating therapeutic strategies for muscular dystrophies. *Mol Ther* (2013) 21:1950–7. doi: 10.1038/mt.2013.186

37. Spencer MJ, Tidball JG. Do immune cells promote the pathology of dystrophinde®cient myopathies? *Neuromuscular Disord* (2001) 11:556–64. doi: 10.1016/S0960-8966(01)00198-5

38. Gussoni E, GK P, Miller RC, Panzara MA, Powell M, Blau HM, et al. Specific T cell receptor gene rearrangements at the site of muscle degeneration in duchenne muscular dystrophy. *J Immunol* (1994) 153:4798–805. doi: 10.4049/jimmunol.153. 10.4798

39. Mantegazza R, Andreetta F, Bernasconi P, Baggi F, Oksenberg JR, Simoncini O, et al. Analysis of T cell receptor repertoire of muscle-infiltrating T lymphocytes in polymyositis restricted va/f, rearrangements may indicate antigen-driven selection. *J Clin Invest* (1993) 91:2880–6. doi: 10.1172/JCI116533

40. Vetrone SA, Montecino-Rodriguez E, Kudryashova E, Kramerova I, Hoffman EP, Liu SD, et al. Osteopontin promotes fibrosis in dystrophic mouse muscle by modulating immune cell subsets and intramuscular TGF- $\beta$ . J Clin Invest (2009) 119:1583–94. doi: 10.1172/JCI37662

41. Emslie-Smith AM, Engel AG. Major histocompatibility complex class I antigen expression, Immunolocalization of interferon subtypes, and T cell-mediated cytotoxicity in myopathies. *Hum Pathol* (1989) 20:224–31. doi: 10.1016/0046-8177 (89)90128-7

42. Appleyard ST, Dubowitz V, Dunn MJ, Rose ML. Increased expression of HLA ABC class I antigens by muscle fibers in Duchenne Muscular Dystrophy, Inflammatory Myopathy, and other muscular disorders. *Lancet* (1985) 1:361–3. doi: 10.1016/S0140-6736(85)91384-4

43. Spencer MJ, Montecino-Rodriguez E, Dorshkind K, Tidball JG. Helper (CD4+) and cytotoxic (CD8+) T cells promote the pathology of dystrophin-deficient muscle. *Clin Immunol* (2001) 98:235–43. doi: 10.1006/clim.2000.4966

44. Pinto-Mariz F, Rodrigues Carvalho L, Prufer De Queiroz Campos Araujo A, De Mello W, Gonçalves Ribeiro M, Cunha MDCSA, et al. CD49d is a disease progression biomarker and a potential target for immunotherapy in Duchenne muscular dystrophy. *Skelet Muscle* (2015) 5:1–10. doi: 10.1186/s13395-015-0066-2

45. Barthélémy I, Pinto-Mariz F, Yada E, Desquilbet L, Savino W, Silva-Barbosa SD, et al. Predictive markers of clinical outcome in the GRMD dog model of Duchenne muscular dystrophy. *Dis Model Mech* (2014) 7:1253–61. doi: 10.1242/dmm.016014

46. Evans NP, Misyak SA, Robertson JL, Bassaganya-Riera J, Grange RW. Immunemediated mechanisms potentially regulate the disease time-course of duchenne muscular dystrophy and provide targets for therapeutic intervention. *PMR* (2009) 1:755–68. doi: 10.1016/j.pmrj.2009.04.010

47. Coles CA, Woodcock I, Pellicci DG, Houweling PJ. A spotlight on T lymphocytes in duchenne muscular dystrophy—Not just a muscle defect. *Biomedicines* (2022) 10:1–10. doi: 10.3390/biomedicines10030535

48. Woodcock IR, Tachas G, Desem N, Houweling PJ, Yiu EM, Kean M, et al. A Phase 2 open-label study to determine the safety and efficacy of weekly dosing of ATL1102 in patients with non-ambulatory Duchenne muscular dystrophy. *MedRxiv Preprint* (2022), 1–30. doi: 10.1101/2022.01.16.22269029

49. Pimorady-esfahani A, Grounds MD, Mcmenamin PG. Macrophages and dendritic cells in normal and regenerating murine skeletal muscle. *Muscle Nerve* (1997) 20:158–66. doi: 10.1002/(SICI)1097-4598(199702)20:2<158::AID-MUS4>3.0. CO;2-B

50. Bonomo AC, Pinto-Mariz F, Riederer I, Benjamim CF, Butler-Browne G, Mouly V, et al. Crosstalk between innate and T cell adaptive immunity with(in) the muscle. *Front Physiol* (2020) 11:573347. doi: 10.3389/fphys.2020.573347

51. Mbongue J, Nicholas D, Firek A, Langridge W. The role of dendritic cells in tissue-specific autoimmunity. *J Immunol Res* (2014) 2014:1–17. doi: 10.1155/2014/857143

52. Rosenberg AS, Puig M, Nagaraju K, Hoffman EP, Villalta SA, Rao VA, et al. Immune-mediated pathology in Duchenne muscular dystrophy. *Sci Transl Med* (2015) 7:1–26. doi: 10.1126/scitranslmed.aaa7322

53. Ladislau L, Portilho DM, Courau T, Solares-Pérez A, Negroni E, Lainé J, et al. Activated dendritic cells modulate proliferation and differentiation of human myoblasts. *Cell Death Dis* (2018) 9:1–14. doi: 10.1038/s41419-018-0426-z

54. Bowerman M, Murray LM, Scamps F, Schneider BL, Kothary R, Raoul C. Pathogenic commonalities between spinal muscular atrophy and amyotrophic lateral sclerosis: Converging roads to therapeutic development. *Eur J Med Genet* (2018) 61:685–98. doi: 10.1016/j.ejmg.2017.12.001

55. Luo X-G, Chen S-D. The changing phenotype of microglia from homeostasis to disease. *Luo Chen Trans Neurodegeneration* (2012) 1:1-13. doi: 10.1186/2047-9158-1-9

56. Cherry JD, Olschowka JA, O'Banion MK. Are "resting" microglia more "M2"? Front Immunol (2014) 5:594. doi: 10.3389/fimmu.2014.00594

57. Liu LR, Liu JC, Bao JS, Bai QQ, Wang GQ. Interaction of microglia and astrocytes in the neurovascular unit. *Front Immunol* (2020) 11:1024. doi: 10.3389/fmmu.2020.01024

58. Amarante-Mendes GP, Adjemian S, Branco LM, Zanetti LC, Weinlich R, Bortoluci KR. Pattern recognition receptors and the host cell death molecular machinery. *Front Immunol* (2018) 9:2379. doi: 10.3389/fimmu.2018.02379

59. Drouin-Ouellet J, Cicchetti F. Inflammation and neurodegeneration: The story "retolled." *Trends Pharmacol Sci* (2012) 33:542–51. doi: 10.1016/j.tips.2012.07.002

60. Hu X, Leak RK, Shi Y, Suenaga J, Gao Y, Zheng P, et al. Microglial and macrophage polarization - New prospects for brain repair. *Nat Rev Neurol* (2015) 11:56–64. doi: 10.1038/nrneurol.2014.207

61. Du L, Zhang Y, Chen Y, Zhu J, Yang Y, Zhang HL. Role of microglia in neurological disorders and their potentials as a therapeutic target. *Mol Neurobiol* (2017) 54:7567–84. doi: 10.1007/s12035-016-0245-0

62. Tang Y, Le W. Differential roles of M1 and M2 microglia in neurodegenerative diseases. *Mol Neurobiol* (2016) 53:1181–94. doi: 10.1007/s12035-014-9070-5

63. Orihuela R, McPherson CA, Harry GJ. Microglial M1/M2 polarization and metabolic states. Br J Pharmacol (2016) 173:649–65. doi: 10.1111/bph.13139

64. Kanning KC, Kaplan A, Henderson CE. Motor neuron diversity in development and disease. *Annu Rev Neurosci* (2010) 33:409-40. doi: 10.1146/annurev.neuro. 051508.135722

65. Alexianu ME, Kozovska M, Appel SH. Immune reactivity in a mouse model of familial ALS correlates with disease progression. *Neurology* (2001) 57:1282–9. doi: 10.1212/WNL.57.7.1282

66. Zhao W, Beers DR, Appel SH. Immune-mediated mechanisms in the pathoprogression of amyotrophic lateral sclerosis. *J Neuroimmune Pharmacol* (2013) 8:888–99. doi: 10.1007/s11481-013-9489-x

67. Li L, Xiong W-C, Mei L. Neuromuscular junction formation, aging, and disorders. *Annu Rev Physiol* (2018) 80:14.1–14.30. doi: 10.1146/annurev-physiol-022516

68. Dupuis L, Loeffler JP. Neuromuscular junction destruction during amyotrophic lateral sclerosis: insights from transgenic models. *Curr Opin Pharmacol* (2009) 9:341–6. doi: 10.1016/j.coph.2009.03.007

69. Dadon-Nachum M, Melamed E, Offen D. The "dying-back" phenomenon of motor neurons in ALS. J Mol Neurosci (2011) 43:470-7. doi: 10.1007/s12031-010-9467-1

70. Moloney EB, de Winter F, Verhaagen J. ALS as a distal axonopathy: Molecular mechanisms affecting neuromuscular junction stability in the presymptomatic stages of the disease. *Front Neurosci* (2014) 8:1–18. doi: 10.3389/fnins.2014.00252

71. Wong M, Martin LJ. Skeletal muscle-restricted expression of human SOD1 causes motor neuron degeneration in transgenic mice. *Hum Mol Genet* (2010) 19:2284–302. doi: 10.1093/hmg/ddq106

72. Papadeas ST, Kraig SE, O'Banion C, Lepore AC, Maragakis NJ. Astrocytes carrying the superoxide dismutase 1 (SOD1 G93A) mutation induce wild-type motor neuron degeneration in *vivo. Proc Natl Acad Sci USA* (2011) 108:17803–8. doi: 10.1073/pnas.1103141108

73. Beers DR, Henkel JS, Xiao Q, Zhao W, Wang J, Yen AA, et al. Wild-type microglia extend survival in PU.1 knockout mice with familial amyotrophic lateral sclerosis. *Proc Nati Acad Sci USA* (2006) 103:16021–6. doi: 10.1073/pnas.0607423103

74. Yamanaka K, Chun SJ, Boillee S, Fujimori-Tonou N, Yamashita H, Gutmann DH, et al. Astrocytes as determinants of disease progression in inherited amyotrophic lateral sclerosis. *Nat Neurosci* (2008) 11:251–3. doi: 10.1038/nn2047

75. Burda JE, Sofroniew MV. Reactive gliosis and the multicellular response to CNS damage and disease. *Neuron* (2014) 81:229–48. doi: 10.1016/j.neuron.2013.12.034

76. Lepore AC, Rauck B, Dejea C, Pardo AC, Rao MS, Rothstein JD, et al. Focal transplantation-based astrocyte replacement is neuroprotective in a model of motor neuron disease. *Nat Neurosci* (2008) 11:1294–301. doi: 10.1038/nn.2210

77. Cassina P, Cassina A, Pehar M, Castellanos R, Gandelman M, De León A, et al. Mitochondrial dysfunction in SOD1G93A-bearing astrocytes promotes motor neuron degeneration: Prevention by mitochondrial-targeted antioxidants. *J Neurosci* (2008) 28:4115–22. doi: 10.1523/JNEUROSCI.5308-07.2008

78. Di Giorgio FP, Carrasco MA, Siao MC, Maniatis T, Eggan K. Non-cell autonomous effect of glia on motor neurons in an embryonic stem cell-based ALS model. *Nat Neurosci* (2007) 10:608–14. doi: 10.1038/nn1885

79. Di Giorgio FP, Boulting GL, Bobrowicz S, Eggan KC. Human embryonic stem cell-derived motor neurons are sensitive to the toxic effect of glial cells carrying an ALS-causing mutation. *Cell Stem Cell* (2008) 3:637–48. doi: 10.1016/j.stem.2008.09.017

80. Aebischer J, Cassina P, Otsmane B, Moumen A, Seilhean D, Meininger V, et al. IFN $\gamma$  triggers a LIGHT-dependent selective death of motoneurons contributing to the non-cell-autonomous effects of mutant SOD1. *Cell Death Differ* (2011) 18:754–68. doi: 10.1038/cdd.2010.143

81. Phatnani HP, Guarnieri P, Friedman BA, Carrasco MA, Muratet M, O'Keeffe S, et al. Intricate interplay between astrocytes and motor neurons in ALS. *Proc Natl Acad Sci United States America (PNAS)* (2013) 110:E756–65. doi: 10.1073/pnas.1222361110

82. Marchetto MCN, Muotri AR, Mu Y, Smith AM, Cezar GG, Gage FH. Non-cellautonomous effect of human SOD1G37R astrocytes on motor neurons derived from human embryonic stem cells. *Cell Stem Cell* (2008) 3:649–57. doi: 10.1016/ j.stem.2008.10.001

83. Chen C, Jiang ZY, Fu X, Yu D, Huang H, Tasker JG. Astrocytes amplify neuronal dendritic volume transmission stimulated by norepinephrine. *Cell Rep* (2019) 29:4349–4361.e4. doi: 10.1016/j.celrep.2019.11.092

84. Kawamata T, Akiyama H, Yamada T, Mcgeer PL. Immunologic reactions in amyotrophic lateral sclerosis brain and spinal cord tissue. *Am J Pathol* (1992) 140:691–707.

85. Henkel JS, Beers DR, Siklós L, Appel SH. The chemokine MCP-1 and the dendritic and myeloid cells it attracts are increased in the mSOD1 mouse model of ALS. *Mol Cell Neurosci* (2006) 31:427–37. doi: 10.1016/j.mcn.2005.10.016

86. Henkel JS, Beers DR, Zhao W, Appel SH. Microglia in ALS: The good, the bad, and the resting. *J Neuroimmune Pharmacol* (2009) 4:389–98. doi: 10.1007/s11481-009-9171-5

87. Henkel JS, Engelhardt JI, Siklós L, Simpson EP, Kim SH, Pan T, et al. Presence of dendritic cells, MCP-1, and activated microglia/macrophages in amyotrophic lateral sclerosis spinal cord tissue. *Ann Neurol* (2004) 55:221–35. doi: 10.1002/ana.10805

88. Perner C, Perner F, Stubendorff B, Förster M, Witte OW, Heidel FH, et al. Dysregulation of chemokine receptor expression and function in leukocytes from ALS patients. *J Neuroinflamm* (2018) 15:1–5. doi: 10.1186/s12974-018-1135-3

89. Panzara MA, Gussoni E, Begovich AB, Murray RS, Zang YQ, Appel SH, et al. T cell receptor BV gene rearrangements in the spinal cords and cerebrospinal fluid of patients with amyotrophic lateral sclerosis. *Neurobiol Dis* (1999) 6:392–405. doi: 10.1006/nbdi.1999.0252

90. Chiu IM, Chen A, Zheng Y, Kosaras B, Tsiftsoglou SA, Vartanian TK, et al. T lymphocytes potentiate endogenous neuroprotective inflammation in a mouse model of ALS. *Proc Natl Acad Sci (PNAS)* (2008) 105:17913–8. doi: 10.1073/pnas.0804610105

91. Jones KJ, Lovett-Racke AE, Walker CL, Sanders VM. CD4 + T cells and neuroprotection: relevance to motoneuron injury and disease. *J Neuroimmune Pharmacol* (2015) 10:587–94. doi: 10.1007/s11481-015-9625-x

92. Gendelman HE, Appel SH. Regulatory T cells in CNS injury: The simple, the complex and the confused. *Trends Mol Med* (2011) 17:687-8. doi: 10.1016/j.molmed.2011.05.012

93. Ni A, Yang T, Mesnard-Hoaglin NA, Gutierrez R, Stubbs EB, McGuire SO, et al. Th17 cell response in SOD1G93A mice following motor nerve injury. *Mediators Inflammation* (2016) 2016:1–7. doi: 10.1155/2016/6131234

94. Gao Z, Tsirka SE. Animal models of MS reveal multiple roles of microglia in disease pathogenesis. *Neurol Res Int* (2011) 2011:1–9. doi: 10.1155/2011/383087

95. Beers DR, Zhao W, Liao B, Kano O, Wang J, Huang A, et al. Neuroinflammation modulates distinct regional and temporal clinical responses in ALS mice. *Brain Behav Immun* (2011) 25:1025–35. doi: 10.1016/j.bbi.2010.12.008

96. Nie H, Zheng Y, Li R, Guo TB, He D, Fang L, et al. Phosphorylation of FOXP3 controls regulatory T cell function and is inhibited by TNF- $\alpha$  in rheumatoid arthritis. *Nat Med* (2013) 19:322–8. doi: 10.1038/nm.3085

97. Mantovani S, Garbelli S, Pasini A, Alimonti D, Perotti C, Melazzini M, et al. Immune system alterations in sporadic amyotrophic lateral sclerosis patients suggest an ongoing neuroinflammatory process. *J Neuroimmunol* (2009) 210:73–9. doi: 10.1016/ j.jneuroim.2009.02.012

98. Murdock BJ, Zhou T, Kashlan SR, Little RJ, Goutman SA, Feldman EL. Correlation of peripheral immunity with rapid Amyotrophic lateral sclerosis progression. *JAMA Neurol* (2017) 74:1446–54. doi: 10.1001/jamaneurol.2017.2255

99. Chen X, Feng W, Huang R, Guo X, Chen Y, Zheng Z, et al. Evidence for peripheral immune activation in amyotrophic lateral sclerosis. *J Neurol Sci* (2014) 347:90–5. doi: 10.1016/j.jns.2014.09.025

100. Shi N, Kawano Y, Tateishi T, Kikuchi H, Osoegawa M, Ohyagi Y, et al. Increased IL-13-producing T cells in ALS: Positive correlations with disease severity and progression rate. *J Neuroimmunol* (2007) 182:232–5. doi: 10.1016/j.jneuroim.2006.10.001

101. Saresella M, Piancone F, Tortorella P, Marventano I, Gatti A, Caputo D, et al. T helper-17 activation dominates the immunologic milieu of both amyotrophic lateral sclerosis and progressive multiple sclerosis. *Clin Immunol* (2013) 148:79–88. doi: 10.1016/j.clim.2013.04.010

102. Beers DR, Henkel JS, Zhao W, Wang J, Appel SH. CD4 T cells support glial neuroprotection, slow disease progression, and modify glial morphology in an animal model of inherited ALS. *Proc Natl Acad Sci (PNAS)* (2008) 105:15558–63. doi: 10.1073/pnas.0807419105

103. Beers DR, Zhao W, Wang J, Zhang X, Wen S, Neal D, et al. ALS patients' regulatory T lymphocytes are dysfunctional, and correlate with disease progression rate and severity. *JCI Insight* (2017) 2:1–14. doi: 10.1172/jci.insight.89530

104. Giovannelli I, Heath P, Shaw PJ, Kirby J. The involvement of regulatory T cells in amyotrophic lateral sclerosis and their therapeutic potential. *Amyotroph Lateral Scler Frontotemporal Degener* (2020) 0:1–10. doi: 10.1080/21678421.2020.1752246

105. Jin M, Günther R, Akgün K, Hermann A, Ziemssen T. Peripheral proinflammatory Th1/Th17 immune cell shift is linked to disease severity in amyotrophic lateral sclerosis. *Sci Rep* (2020) 10:1–13. doi: 10.1038/s41598-020-62756-8

106. Beers DR, Henkel JS, Zhao W, Wang J, Huang A, Wen S, et al. Endogenous regulatory T lymphocytes ameliorate amyotrophic lateral sclerosis in mice and correlate with disease progression in patients with amyotrophic lateral sclerosis. *Brain* (2011) 134:1293–314. doi: 10.1093/brain/awr074

107. Zubiri I, Lombardi V, Bremang M, Mitra V, Nardo G, Adiutori R, et al. Tissueenhanced plasma proteomic analysis for disease stratification in amyotrophic lateral sclerosis. *Mol Neurodegener* (2018) 13:1–17. doi: 10.1186/s13024-018-0292-2

108. Yildiz O, Schroth J, Tree T, Turner MR, Shaw PJ, Henson SM, et al. Senescentlike blood lymphocytes and disease progression in amyotrophic lateral sclerosis. *Neurology(R) Neuroimmunol Neuroinflamm* (2023) 10:1–13. doi: 10.1212/ NXI.000000000200042

109. Aiello A, Farzaneh F, Candore G, Caruso C, Davinelli S, Gambino CM, et al. Immunosenescence and its hallmarks: How to oppose aging strategically? A review of potential options for therapeutic intervention. *Front Immunol* (2019) 10:2247. doi: 10.3389/fmmu.2019.02247

110. Yazdani S, Seitz C, Cui C, Lovik A, Pan L, Piehl F, et al. T cell responses at diagnosis of amyotrophic lateral sclerosis predict disease progression. *Nat Commun* (2022) 13:1–13. doi: 10.1038/s41467-022-34526-9

111. Gazzerro E, Baldassari S, Assereto S, Fruscione F, Pistorio A, Panicucci C, et al. Enhancement of muscle T regulatory cells and improvement of muscular dystrophic process in mdx mice by blockade of extracellular ATP/P2X axis. *Am J Pathol* (2015) 185:3349–60. doi: 10.1016/j.ajpath.2015.08.010

112. Castiglioni A, Corna G, Rigamonti E, Basso V, Vezzoli M, Monno A, et al. FOXP3+ T cells recruited to sites of sterile skeletal muscle injury regulate the fate of satellite cells and guide effective tissue regeneration. *PloS One* (2015) 10:1–18. doi: 10.1371/journal.pone.0128094 113. Cho J, Kuswanto W, Benoist C, Mathis D. T cell receptor specificity drives accumulation of a reparative population of regulatory T cells within acutely injured skeletal muscle. *Proc Natl Acad Sci USA (PNAS)* (2019) 116:26727–33. doi: 10.1073/ pnas.1914848116/-/DCSupplemental

114. Schiaffino S, Pereira MG, Ciciliot S, Rovere-Querini P. Regulatory T cells and skeletal muscle regeneration. *FEBS J* (2017) 284:517–24. doi: 10.1111/febs.13827

115. Zhao W, Beers DR, Liao B, Henkel JS, Appel SH. Regulatory T lymphocytes from ALS mice suppress microglia and effector T lymphocytes through different cytokine-mediated mechanisms. *Neurobiol Dis* (2012) 48:418–28. doi: 10.1016/j.nbd.2012.07.008

116. Sheean RK, McKay FC, Cretney E, Bye CR, Perera ND, Tomas D, et al. Association of regulatory T-Cell Expansion with progression of amyotrophic lateral sclerosis a study of humans and a transgenic mouse model. *JAMA Neurol* (2018) 75:681–9. doi: 10.1001/jamaneurol.2018.0035

117. Bettelli E, Carrier Y, Gao W, Korn T, Strom TB, Oukka M, et al. Reciprocal developmental pathways for the generation of pathogenic effector TH17 and regulatory T cells. *Nature* (2006) 441:235–8. doi: 10.1038/nature04753

118. Henkel JS, Beers DR, Wen S, Rivera AL, Toennis KM, Appel JE, et al. Regulatory T-lymphocytes mediate amyotrophic lateral sclerosis progression and survival. *EMBO Mol Med* (2013) 5:64–79. doi: 10.1002/emmm.201201544

119. Kraemer M, Buerger M, Berlit P. Diagnostic problems and delay of diagnosis in amyotrophic lateral sclerosis. *Clin Neurol Neurosurg* (2010) 112:103–5. doi: 10.1016/j.clineuro.2009.10.014

120. Savino W, Mendes-Da-Cruz DA, Lepletier A, Dardenne M. Hormonal control of T-cell development in health and disease. *Nat Rev Endocrinol* (2016) 12:77–89. doi: 10.1038/nrendo.2015.168

121. Takaba H, Takayanagi H. The mechanisms of T cell selection in the thymus. Trends Immunol (2017) 38:805–16. doi: 10.1016/j.it.2017.07.010

122. Abramson J, Anderson G. Thymic epithelial cells. *Annu Rev Immunol* (2017) 35:85–118. doi: 10.1146/annurev-immunol-051116-052320

123. Takada K, Takahama Y. Positive-selection-inducing self-peptides displayed by cortical thymic epithelial cells. In: *Advances in Immunology*. Academic Press Inc (2015). p. 87–110. doi: 10.1016/bs.ai.2014.09.003

124. Wing JB, Tanaka A, Sakaguchi S. Human FOXP3 + Regulatory T cell heterogeneity and function in autoimmunity and cancer. *Immunity* (2019) 50:302–16. doi: 10.1016/j.immuni.2019.01.020

125. Zheng YY, Wang Y, Chen X, Wei LS, Wang H, Tao T, et al. The thymus regulates skeletal muscle regeneration by directly promoting satellite cell expansion. *J Biol Chem* (2022) 298:1–15. doi: 10.1016/j.jbc.2021.101516

126. Quirico-Santos T, Ribeiro MM, Savino W. Increased deposition of extracellular matrix components in the thymus gland of MDX mouse: correlation with the muscular lesion. *J Neuroimmunol* (1995) 59:9–18. doi: 10.1016/0165-5728(95)00010-Y

127. Farini A, Sitzia C, Villa C, Cassani B, Tripodi L, Legato M, et al. Defective dystrophic thymus determines degenerative changes in skeletal muscle. *Nat Commun* (2021) 12:1–20. doi: 10.1038/s41467-021-22305-x

128. Seksenyan A, Ron-Harel N, Azoulay D, Cahalon L, Cardon M, Rogeri P, et al. Thymic involution, a co-morbidity factor in amyotrophic lateral sclerosis. *J Cell Mol Med* (2010) 14:2470–82. doi: 10.1111/j.1582-4934.2009.00863.x