Check for updates

OPEN ACCESS

EDITED BY Alessandro Poggi, San Martino Hospital (IRCCS), Italy

REVIEWED BY Mohit Kumar Jolly, Indian Institute of Science (IISc), India Jason T. George, Texas A and M University, United States

*CORRESPONDENCE Li-Jie He Melijie@fmmu.edu.cn Ya Zhao Zhaoya@fmmu.edu.cn

[†]These authors have contributed equally to this work

RECEIVED 20 May 2023 ACCEPTED 28 July 2023 PUBLISHED 15 August 2023

CITATION

Zhang Y-C, Zhang Y-T, Wang Y, Zhao Y and He L-J (2023) What role does PDL1 play in EMT changes in tumors and fibrosis? *Front. Immunol.* 14:1226038. doi: 10.3389/fimmu.2023.1226038

COPYRIGHT

© 2023 Zhang, Zhang, Wang, Zhao and He. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

What role does PDL1 play in EMT changes in tumors and fibrosis?

Yun-Chao Zhang^{1†}, Yu-Ting Zhang^{1†}, Yi Wang^{1†}, Ya Zhao^{2*} and Li-Jie He^{1*}

¹Department of Nephrology, Xi Jing Hospital, The Fourth Military Medical University, Xi'an, China, ²Department of Medical Microbiology and Parasitology, Fourth Military Medical University, Xi'an, China

Epithelial-mesenchymal transformation (EMT) plays a pivotal role in embryonic development, tissue fibrosis, repair, and tumor invasiveness. Emerging studies have highlighted the close association between EMT and immune checkpoint molecules, particularly programmed cell death ligand 1 (PDL1). PDL1 exerts its influence on EMT through bidirectional regulation. EMT-associated factors, such as YB1, enhance PDL1 expression by directly binding to its promoter. Conversely, PDL1 signaling triggers downstream pathways like PI3K/AKT and MAPK, promoting EMT and facilitating cancer cell migration and invasion. Targeting PDL1 holds promise as a therapeutic strategy for EMT-related diseases, including cancer and fibrosis. Indeed, PDL1 inhibitors, such as pembrolizumab and nivolumab, have shown promising results in clinical trials for various cancers. Recent research has also indicated their potential benefit in fibrosis treatment in reducing fibroblast activation and extracellular matrix deposition, thereby addressing fibrosis. In this review, we examine the multifaceted role of PDL1 in immunomodulation, growth, and fibrosis promotion. We discuss the challenges, mechanisms, and clinical observations related to PDL1, including the limitations of the PD1/PDL1 axis in treatment and PD1-independent intrinsic PDL1 signaling. Our study highlights the dynamic changes in PDL1 expression during the EMT process across various tumor types. Through interplay between PDL1 and EMT, we uncover co-directional alterations, regulatory pathways, and diverse changes resulting from PDL1 intervention in oncology. Additionally, our findings emphasize the dual role of PDL1 in promoting fibrosis and modulating immune responses across multiple diseases, with potential implications for therapeutic approaches. We particularly investigate the therapeutic potential of targeting PDL1 in type II EMT fibrosis: strike balance between fibrosis modulation and immune response regulation. This analysis provides valuable insights into the multifaceted functions of PDL1 and contributes to our understanding of its complex mechanisms and therapeutic implications.

KEYWORDS

epithelial-mesenchymal transformation, programmed death-ligand 1, immune escape, fibrosis, EMT-related disease

1 Introduction

The occurrence and development of EMT and mesenchymalepithelial transformation (MET) can be divided into three biological subtypes in current research (1): Type I, which is primarily involved in the formation and development of embryos (2); Type II, which is primarily associated with tissue repair, regeneration, and fibrosis (3, 4); and Type III, which is linked to tumor invasion and immune suppression (5).

PDL1 has a crucial role in maintaining immune homeostasis and self-tolerance by inhibiting T cell immune response via binding to PD1 on T cells (6). However, recent studies have shown that PDL1 has other functions beyond its role in regulating immunity, including its interaction with EMT in the tumor microenvironment. The expression of PDL1 can increase during EMT, which induces immunosuppression and escape, and promotes cancer metastasis (7).

EMT represents a complex and intricate cellular process that extends beyond a simple transition from an epithelial to a mesenchymal state. During this process, cells acquire one or more hybrid epithelial/mesenchymal (E/M) phenotypes. Importantly, recent evidence indicates the existence of hybrid E/M phenotype cells characterized by elevated PDL1 expression (8, 9). These cells, with their hybrid E/M phenotype, may utilize increased PDL1 expression as a strategy to evade immune surveillance during tumor progression and metastasis. Consequently, this can confer resistance to therapeutic interventions. A deeper understanding of EMT and its correlation with elevated PDL1 expression not only enhances our comprehension of tumor development and immune evasion mechanisms but also paves the way for novel therapeutic strategies and targets.

Recent studies on EMT subtype III have revealed a significant correlation between EMT and immune activation, with high expression levels of immune checkpoint molecules such as PD1, PDL1, CTLA4, OX40L, and PDL2 observed in tissues with elevated EMT levels (10). Furthermore, interfering with the PDL1 pathway can impact the development of the EMT process within the tumor microenvironment (TME) (5, 11). While the relationship between EMT and immune checkpoints, including PDL1, has been extensively studied in the context of cancer, there is still much to be learned about their interaction in other areas of biology. However, some studies have suggested that EMT status could be a combined biomarker of PDL1, and that the two may have a bidirectional regulatory relationship beyond cancer (5). Further research is needed to fully understand the molecular mechanisms and potential clinical applications of this relationship.

EMT is a widespread phenomenon in the human body, and interrupting its progression can have a profound impact on the onset and progression of related diseases, particularly fibrosis in the context of EMT-II. Hence, reversing EMT has emerged as a crucial therapeutic strategy for combating fibrosis (2). The bidirectional interplay between EMT and PDL1 has given rise to a novel concept. Drawing from their mutual regulation in oncology, experts have suggested the existence of similar mechanisms in the context of fibrosis, such as the amplification of PDL1 to facilitate pulmonary fibrosis by hindering vimentin degradation, and the upregulation of PDL1 via Golm1 in the setting of liver fibrosis (12, 13).

Aside from the aforementioned types of fibrosis, there are numerous other fibrotic conditions where EMT plays a critical role, including peritoneal fibrosis (PF) triggered by peritoneal dialysis (PD). PD is a vital therapy for end-stage renal disease and has gained widespread acceptance in clinical settings (3). Long-term PD therapy can lead to decreased peritoneal function due to PF, which is a significant complication and the primary cause of longterm ultrafiltration failure in PD patients (14). During this pathological process, the peritoneum remains in a state of chronic inflammation, where EMT and its intermediate state induced by transforming growth factor- β (TGF- β) serve as major driving forces in the development of peritoneal fibrosis and subsequent functional deterioration (4, 15, 16). At the same time, TGF- β pathway drives the generation of congenital PD1 resistance characteristics and promotes the upregulation of PDL1 (17-20). Building upon the aforementioned phenomena, this review suggests a potential correlation between EMT and PDL1 within the realm of PF.

The primary objective of this review is to provide a comprehensive overview of the physiological roles of EMT and PDL1, while emphasizing recent advancements in understanding their reciprocal regulatory mechanisms in oncology. This includes examining the influence of EMT on PDL1 expression and the effects of inhibiting the PDL1 pathway on EMT progression. Additionally, this review aims to explore the interplay between EMT and PDL1 in non-tumor settings, with a specific focus on their implications in fibrosis.

In conclusion, EMT and its association with PDL1 have emerged as critical factors in various biological processes, including tumor progression, immune evasion, and fibrosis. Understanding the intricate relationship between EMT and PDL1 not only enhances our knowledge of disease mechanisms but also opens up new avenues for therapeutic interventions. By exploring the bidirectional interplay between EMT and PDL1, this review sheds light on their significance in both cancer and fibrosis contexts, with potential implications for the development of targeted therapies. Further research is warranted to unravel the underlying molecular mechanisms and explore the clinical applications of this dynamic interaction.

2 Challenges and opportunities in the application of PDL1

The discovery and clinical application of immune checkpoint inhibitors (ICIs) targeting PD1 and PDL1 were recognized by the 2018 Nobel Prize in Medicine as a major breakthrough in cancer treatment. This monumental achievement has brought about a paradigm shift in the approach to combating cancer, with immune checkpoint modulation taking center stage (21). At the heart of this intricate network lies the PD1/PDL1 pathway, a pivotal axis that exerts critical control over the T cell response, profoundly impacting their ability to recognize and eliminate malignant cells, including cancer cells and those infected with viruses (22). PDL1, a versatile molecule, not only adorns the surface of tumor cells and immune cells but also assumes an extracellular role, contributing to the intricate web of immunoregulation (23). Among these extracellular manifestations, exosomes and shed vesicles in the extracellular supernatant harbor PDL1, serving as an additional layer of regulation that complements the membrane-bound PD1/ PDL1 axis (24).

2.1 Impact of PDL1 on patient survival in specific cancer subtypes: exploring clinical observations

Approaching the subject from a clinical standpoint, investigating the influence of PDL1 on patient survival in specific cancer subtypes provides profound insights into the mechanisms of tumor immune evasion and facilitates the development of personalized treatment strategies. Recent research has unveiled a positive correlation between heightened PDL1 activity, immune checkpoint markers, and elevated characteristics of partial EMT and glycolysis in various cancers. These findings suggest a close association between PDL1 and EMT in cancer progression, potentially synergistically fueling invasive tumor growth (8, 25).

Moreover, analyses encompassing multiple cancer types reveal that the co-enrichment of glycolysis and PDL1 is linked to shorter overall survival, compared to cases with PDL1 enrichment alone. Notably, investigations focused on colorectal cancer demonstrate a favorable prognosis associated with elevated PDL1 expression, underscoring the potential significance of PDL1 as a crucial prognostic factor and guiding personalized therapies (26).

In addition, studies conducted in high-grade chondrosarcoma (CS), retroperitoneal liposarcoma (LS), and undifferentiated pleomorphic sarcoma (UPS) highlight the immunoreactivity of PDL1. Particularly, patients with higher PDL1 immunoreactivity (>50%) within the tumor cells may form a rational cohort for exploring potentially beneficial PDL1/PD1 targeted therapies. These findings offer valuable insights for future in-depth investigations into PDL1 as a therapeutic target within specific sarcoma subtypes (27).

2.2 Challenges in clinical treatment: application limitations of PD1/PDL1 axis

The PD1/PDL1 axis serves as the arbiter of immune homeostasis. As T cell activity heightens, PD1 expression on the

cell surface is elevated, playing a pivotal role in modulating the intensity of T cell response (28). While blocking the PD1/PDL1 axis can enhance the immune response against abnormal cells (29), excessive intervention may give rise to immune-related adverse events (irAEs) that can unpredictably affect any organ system. These irAEs encompass different aspects: firstly, there is an elevated susceptibility to lupus-like autoimmune diseases due to PD1 deficiency (30); secondly, high PDL1 expression can result in altered T cell development (31); and thirdly, extensive clinical intervention can lead to impaired organ function, such as thyroid dysfunction (hypothyroidism or thyrotoxicosis), pituitary inflammation (hypophysitis), adrenal insufficiency, pancreatic damage (diabetes), and other endocrine diseases resulting from extensive clinical intervention (32). Due to the intricacies of immune homeostasis in the human body and the unclear mechanisms of PD1/PDL1 regulation, the highly acclaimed PD1 and PDL1 inhibitors face two major challenges in clinical treatment: limitations in application and significant side effects.

2.3 PD1-independent intrinsic PDL1 signaling

Most studies have focused on PDL1's role as a ligand of PD1 and its effect on PD1-expressing T cells. However, recent research has revealed the broader significance of cancer cell-intrinsic PDL1 signaling in controlling various aspects of tumor growth, immune effects, DNA damage response, and gene expression regulation, many of which operate independently of PD1 (33). These effects include the following: firstly, PDL1 can interact with CD80, transmitting inhibitory signals to activated T cells (34); secondly, PDL1-targeted therapy can reduce mTOR activity and glycolysis in tumor cells, even in the absence of T cells (35); thirdly, in the absence of PD1, PDL1 serves as a shield for tumor cells against cytotoxicity induced by type I and II interferons and cytotoxic T lymphocytes (36); fourthly, PDL1 can regulate extracellular matrix (ECM) components and their proteolytic remodeling products, promoting immune escape and accelerated invasion of tumor cells (37); and lastly, PDL1 and TGF- β form complementary signaling pathways that promote tumor fibrosis and epithelial transformation to stroma (38).

2.4 The intricate relationship of EMT and PDL1: immune evasion and fibrotic pathologies

Based on the aforementioned studies regarding the impact of PDL1, its intrinsic effects, suggest that changes in EMT occur during the production of PDL1 (7, 39). When coupled with the examples highlighted in the introduction indicating that the relationship between EMT and PDL1 is widely prevalent in

immune escape of tumors and organ fibrosis (5, 10–13), it is reasonable to conclude that EMT is closely linked to PDL1, and the signaling pathway between EMT and PDL1 may exist in the major physiological subtypes of EMT (Figure 1).

3 Exploring the impact of EMT on PDL1 expression: dynamic changes in tumor biology

3.1 Concordant changes of PDL1 expression during the EMT process

Currently, numerous studies have demonstrated a strong correlation between the state of EMT and the expression of immune checkpoint molecules, particularly PDL1, which exhibits dynamic changes as EMT progresses (10). This view is further supported by comparing the contents of EMT-related DNA, RNA, and protein in different tumor types: firstly, in lung cancer, a positive correlation is observed between the expression of EMT markers, such as Snail and Vimentin, and the levels of PDL1 (6). Conversely, the expression of E-cadherin, a marker of epithelial characteristics, shows a negatively correlated with PDL1 expression (40); secondly, in breast cancer, mesenchymal cancer cell lines demonstrate a higher proportion of PDL1-positive cells. Activation of ZEB1 or Snail leads to an increase in PDL1 expression (41); thirdly, in patients with cholangiocarcinoma, high expression of EMT markers, such as ZEB1, N-cadherin, and Vimentin, along with low expression of E-cadherin, are associated with increased PDL1 expression (42); lastly, similar results are observed in esophageal squamous cell carcinoma, patients with elevated levels of EMT marker ZEB1 and positive PDL1 have the worst prognosis (43). These findings collectively emphasize the close relationship between EMT and PDL1 expression across various cancer types.

3.2 Unraveling the complexities: mechanisms of PDL1 expression in the EMT process

Extensive literature has consistently indicated that in the tumor microenvironment, as EMT progresses, there is a notable upregulation of PDL1 expression (5, 10, 11, 40-45). Many scholars have put forward their own views on the underlying mechanisms governing PDL1 expression during EMT progression: firstly, TGF-B, the primary inducer of EMT, boosts the expression of PDL1 via the PI3K/AKT and MEK/ERK pathways in breast cancer. It is suggested that the EMT process itself, rather than the direct influence of TGF- β , leads to the heightened expression of PDL1 (46); secondly, interference with ZEB1 at the RNA level can bring about a reduction in PDL1 mRNA and protein levels in esophageal squamous cells (43). Similar results were obtained in non-small cell lung cancer, indicating that the expression of PDL1 is regulated by the miR-200/ZEB1 channels and that there exists a miR-200 binding site on PDL1 (47). Based on this, some scholars have proposed a clear correlation between the heightened expression of PDL1 and the augmentation of ZEB1 and ZEB2, along with the reduction of miR200a and miR200c. ZEB1, which functions as an EMT activator and a transcriptional inhibitor of miR-200, can alleviate the inhibitory effect of miR-200 on PDL1 (48, 49); thirdly, PDL1 is enriched in tumor stem cells through the EMT/β -catenin/STT3/PDL1 signaling axis, in which EMT induces N-glycosyltransferase STT3 via β -catenin transcription, and then stabilizes PDL1 and upregulates (50); fourthly, EMT transcription factors, including the ZEB, Twist, and Snail family proteins, have also been shown to regulate PDL1 expression through the MAPK pathway (42, 51, 52); lastly, there are alternative viewpoints: the downregulation of E-cadherin by shRNA can reduce PDL1 expression (53). Knockout of Snail can significantly inhibit EMT but has no effect on PDL1 expression. Knockout of c-Myc suppresses PDL1 expression while not affecting EMT (54). In



Cellular changes during EMT and MET. Upon undergoing EMT, cellular adhesion is diminished, leading to the loss of the epithelial phenotype and the gradual acquisition of a mesenchymal phenotype. In this transitional state, there is an upregulation of PDL1 expression. This transformation endows cells with the capacity for distant metastasis and redifferentiation. Conversely, MET induces the reversal of these alterations. EMT, epithelial-mesenchymal transformation; MET, mesenchymal - epithelial transformation; TJ, tight junction; AJ, adhesive junctions; DS, desmosome; RJ, residual junction.

summary, the precise mechanisms governing PDL1 expression during the EMT process warrant further investigation.

4 Divergent changes in EMT following PDL1 intervention in oncology

4.1 Co-direction changes of EMT after intervention of PDL1 expression

Numerous studies have shown the therapeutic potential of PDL1 in reducing EMT and inhibiting cell invasion in tumors (11, 55-57). High expression of PDL1 plays an important role in maintaining EMT status in various types of cancer, such as breast cancer, hepatocellular carcinoma, esophageal carcinoma, and renal cell carcinoma (58, 59). Additionally, extracellular PDL1 has similar biological effects (57, 60). Corresponding to the changes observed in PDL1 during the EMT process, interventions targeting PDL1 expression also induce EMT alterations in various types of tumors. For example: firstly, in lung cancer, the upregulation of PDL1 expression by ATM through the JAK1, 2/STAT3 signaling pathway promotes EMT and tumor metastasis, while the inhibition of ATM reduces PDL1 expression via the JAK/STAT3 signaling pathway, resulting in diminished EMT and suppressed tumor metastasis in mouse models (11); secondly, in head and neck squamous cell carcinoma, the loss of CMTM6, a regulator of PDL1 expression, inhibits PDL1 expression and leads to low EMT expression. Additionally, a positive correlation between the expression of CMTM6 and EMT-related genes has been found in TCGA. The authors propose that CMTM6 modulates the EMT process in cancer through the Wnt/ β -catenin signaling pathway (55); thirdly, in esophageal cancer cells, PDL1 expression significantly promotes EMT phenotype, with the cytoplasmic domain of PDL1 plays an important role in this process (58); fourthly, in renal cell carcinoma, downregulation of PDL1 by lentivirus can inhibit the expression of EMT markers such as Vimentin, while upregulation of PDL1 can promote the expression of these EMT markers by activating SREBP-1 (59); lastly, in non-small cell lung cancer, downregulation of Circ-CPA4 suppresses the expression of PDL1 through let-7miRNA, thereby affecting the activity of T cells, and then the process of EMT is changed. Additionally, the paper proposes an intriguing phenomenon: upregulation of Circ-CPA4 enables non-small cell lung cancer cells to bind to CD8+ T cells in the way of secreting PDL1 exosomes, thereby effectively deactivating them (57). This complementary function further enhances the effect of PDL1.

4.2 Shared regulatory signaling of EMT and PDL1

Currently, a wealth of evidence supports the existence of multiple signaling pathways that intricately regulate PDL1 expression. These pathways encompass a spectrum of influential factors, which can be broadly categorized into six main types: firstly, JAK/STAT pathway (61, 62); secondly, NF- κ B pathway (63, 64); thirdly, MAPK pathway (65); fourthly, PI3K/AKT/mTOR pathway (46, 66); fifthly, TGF- β /Smad pathway (60); and lastly, β -catenin/STT3 pathway (50). Remarkably, EMT marker molecules such as ZEB1 and Snail have been implicated in these six signaling cascades, further supporting the reciprocal relationship between EMT and PDL1 at the signaling pathway level (62) (Figure 2).



FIGURE 2

The main signal pathways that affect the expression of PDL1. These pathways also have downstream signals, including ZEB and Snail, which act as transcription factors for EMT and regulate the expression of PDL1, highlighting the bidirectional regulatory relationship between EMT and PDL1 at the signaling pathway level. IFN- γ , interferon- γ ; EGF, epidermal growth factor; HGF, hepatocyte growth factor; TGF- β , transforming growth factor- β ; Wht, wingless; JAK, janus kinase; STAT, signal transducer and activator of transcription; HMGB1, high mobility group protein 1; RAGE, receptor for advanced glycation end products; Ras, rasopathes; Braf, RAF family serine/threonine protein kinases; MEK, MAPK kinase; ERK, extracellular-signal regulated protein kinase; PI3K, phosphoinositide 3-kinase; AKT, protein kinase B; mTOR, mammalian target of rapamycin; STT3, N-oligosaccharide transferase complex; Smad, skin growth factor; IRF, interferon regulatory factor; ZEB, zinc finger E-box binding homeobox; PDL1, programmed death ligand 1; EMT, epithelial-mesenchymal transformation.

The transcription factors involved in promoting tissue EMT transformation contribute to the expression of PDL1. Moreover, PDL1 itself exerts an influence on the secretion of EMT transcription factors, which can be classified into the following three types (Figure 3): firstly, at the protein level, an intricate interplay between Snail and PDL1 has been observed. PDL1 has the ability to induce the phosphorylation of GSK3B through activation of the p38-MAPK or AKT pathways, ultimately promoting the expression of Snail (where GSK3β serves as a ubiquitin ligase for Snail) (67). Furthermore, the generation of the CCL2/LCN2 complex by Snail can induce cells to adopt an immunosuppressive state, leading to the upregulation of PDL1 expression (68); secondly, at the miRNA level, the impact of ZEB1 on PDL1 is notable. The miR-200 family acts as a targeted inhibitory signal for PDL1, while ZEB1 acts as both an EMT activator signal and a transcriptional suppressor for miR-200, thereby relieving the inhibitory effect of miR-200 on PDL1 expression (49). In turn, PDL1 can influence ZEB1 through the generation of CDH1 (69); thirdly, at the transcriptional level, YB1 participates in the regulation of PDL1. YB1 directly binds to the promoter sequence of PDL1, thereby inducing the production of PDL1. Excessive PDL1, in turn, triggers the release of fibrosis-promoting factors by T cells, such as TGF-B, thereby promoting the upregulation of YB1 (70).

4.3 Unique scenario: accelerated EMT transformation following downregulation of PDL1

Contrary to the anticipated alignment between EMT and PDL1 alterations discussed earlier, divergent alterations in EMT may occur after intervention in PDL1. In current therapeutic research of PDL1, a surprising revelation has emerged: high PDL1 expression does not ensure a favorable response to PD1 inhibitors, while low or even absent PDL1 expression does not necessarily signify an unresponsive state to PD1/PDL1 antibodies (71, 72). Moreover, the influence of PDL1 on EMT is highly dependent on the stage of the disease, showcasing intricate ramifications as the disease unfolds. Primarily, the reduction of PDL1 levels disrupts the binding of PD1/PDL1, effectively impeding macrophage apoptosis and mitigating the detrimental consequences of inflammation. Consequently, the progression of EMT is influenced, leading to a reduction in the magnitude of tissue fibrosis (61). Notably, the downregulation of PDL1 following primary infection plays a pivotal role in restoring protective immunity. Nevertheless, this process entails the emergence of post-infection inflammation and the consequent progression of fibrosis (73).



FIGURE 3

Bidirectional regulation of EMT transcription factors and PDL1. (A) At the protein level. PDL1 activates the p38-MAPK or AKT pathways, leading to the phosphorylation of GSK3 β and subsequent upregulation of Snail expression. Snail, in turn, promotes the generation of the CCL2/LCN2 complex, inducing an immunosuppressive state in cells and further enhancing PDL1 expression; (B) At the miRNA level, ZEB1 acts as a transcriptional repressor of miR-200, alleviating the inhibitory effect of miR-200 on PDL1 expression. Additionally, PDL1 can influence ZEB1 through the generation of CDH1; (C) At the transcriptional level, YB1 directly binds to the promoter sequence of PDL1, promoting PDL1 production. The excessive presence of PDL1 induces the release of fibrosis-promoting factors, such as TGF- β , from T cells, thereby promoting the upregulation of YB1. PDL1, programmed death ligand 1; P38-MAPK, p38 mitogen activated protein kinase; PI3K, phosphoinositide 3-kinase; AKT, protein kinase B; GSK3 β , glycogen synthease kinase 3 β ; CCL2, C-C motif chemokine ligand 2; LCN2, lipocalin 2; ZEB, zinc finger E-box binding homeobox; miRNA-200, micro RNA 200 family; YB1, Y-box binding protein 1; IL1 β , interleukin 1; IL10, interleukin 10; TGF- β , transforming growth factor- β .

5 The multifunctional role of PDL1 in EMT: from immune evasion to developmental promotion and fibrosis

Based on the strong correlation between EMT and PDL1 in the field of oncology, some scholars have carried out PDL1 studies on other subtypes of EMT, and obtained results to test the hypothesis that the mechanism linking EMT and PDL1 widely exists in the human body.

5.1 PDL1 in embryonic development of type I EMT

The physiological mechanism of EMT permeates through various facets of life, encompassing primordial cell differentiation, organ development, and even final death. Expanding upon the research conducted on PDL1 and its relationship with EMT in tumor contexts. Intriguingly, evidence has emerged to substantiate this notion: firstly, the inhibition of PDL1 has been shown to regulate the EMT process through ATF3 signaling, subsequently influencing angiogenesis and ultimately culminating in embryonic death (74); secondly, the blockade of PDL1 leads to the accumulation of T follicular regulatory cells within the uterine environment during the second trimester, provoking various immune dysfunctions and even triggering pregnancy termination (75); thirdly, PDL1 induces macrophages polarization towards the M2 phenotype during early pregnancy, facilitating the establishment of maternal-fetal tolerance (76); lastly, soluble PDL1 emerges as a pivotal factor in embryonic development during pregnancy. Trophoblast cells actively secrete soluble PDL1, thereby initiating the PD1/PDL1 pathway and orchestrating the M2 polarization of macrophages, crucial for sustaining a conducive microenvironment for embryonic growth (77).

The literature on PDL1 in embryonic development mainly focuses on its immune regulatory functions, such as its influence on M2 polarization of macrophages and pregnancy outcomes [M2 macrophages are closely related to the occurrence of EMT (78)]. Nonetheless, the precise mechanism underlying the intricate interplay between PDL1 and EMT during embryonic development remains elusive. While some evidence suggests potential roles for PDL1 beyond immune cell regulation, such as cell differentiation and tissue formation, these findings are currently limited. Further investigations are needed to unravel the intricate relationship between PDL1 and EMT during the journey of embryonic development.

5.2 PDL1 in tissue repair and fibrosis of type II EMT

Considering the well-documented impact of PDL1 on M2 macrophage polarization and the aforementioned instances of EMT and PDL1 in the field of fibrosis, scholars have extensively investigated the dynamics of PDL1 in fibrotic conditions, especially those arising from chronic inflammation. Within a chronic inflammatory microenvironment, the release of inflammatory mediators triggers an uncontrolled proliferation of fibroblasts, their conversion to an EMT phenotype, and ECM deposition, ultimately culminating in collagen overproduction and the onset of fibrosis. In this intricate cascade, the involvement of PDL1 may play a vital role (79, 80).

5.2.1 The dual role of PDL1: immunomodulation and promoting fibrosis across fibrosis diseases 5.2.1.1 The fibrotic role of PDL1 in idiopathic pulmonary fibrosis

Idiopathic pulmonary fibrosis (IPF) is a chronic, progressive, and fibrotic interstitial lung disease characterized by chronic epithelial injury and the regenerative failure of the alveolar compartment (81). Remarkably, IPF and lung cancer exhibit striking biomolecular similarities, with EMT emerging as a critical component in the pathogenesis of IPF fibrosis (82, 83).

Emerging literature highlights the potential of PDL1 inhibition in reducing fibrosis severity in pulmonary fibrosis, and the PDL1 pathway mainly plays a role in promoting fibrosis rather than its immunomodulatory role (84). The fibrotic effect of PDL1 in IPF arises from intricate cellular interactions and signaling pathways, with the following mechanisms assuming paramount significance (Figure 4): firstly, overexpression of PDL1 in lung fibroblasts exerts resistance to myoblast apoptosis and eludes macrophage phagocytosis via the inhibition of p53 pathway or activation of FAK pathway. Consequently, excessive myoblast proliferation ensues, instigating IPF development (85-87); secondly, PDL1 mediates the transition of pulmonary fibroblasts into myofibroblasts and instigates EMT induction via TGF- β, facilitated by the Smad3 and β -catenin signaling pathways, thereby promoting fibrosis (88, 89); thirdly, upregulation of PDL1 in pulmonary fibroblasts hampers autophagy-induced myofibroblast proliferation and ECM deposition, ultimately leading to pulmonary fibrosis. Overexpression of PDL1 can inhibit autophagy of fibrosis-related cells through the PI3K/AKT/ mTOR signal pathway, up regulate TGF-B and fibrosis-related factor a-SMA (90-92); fourthly, elevated PDL1 expression in pulmonary fibroblasts impedes adaptive immunity, fostering pulmonary fibrosis. Notably, immune dysregulation has emerged as a key driver of IPF, with M2 macrophages, Th17 cells, CD8+T cells, and Tregs promoting fibrosis, while Th1 and TRM cells appear to exert protective effects (93). Furthermore, current evidence suggests that upregulation of PD1/PDL1 in IPF may prompt the differentiation of CD4+T cells into Tregs and lead to depletion of macrophages, thereby enabling myofibroblasts to evade immune clearance and resist macrophage-induced phagocytosis (94, 95).

These reports provide valuable insight into the fibrogenic role of PDL1 in IPF. It is noteworthy that the mechanism by which PDL1 operates in IPF mirrors the mutual regulation observed between PDL1 and EMT in the context of tumors. In the case of IPF, PDL1 may also impact EMT in the following ways: firstly, the PDL1 signaling pathway, including STAT3, p53, β -catenin, and PI3K/AKT/mTOR, may affect alveolar epithelial cell migration and EMT induction mediated by TGF- β (96–99); secondly, PDL1 in IPF can



FIGURE 4

The intrinsic pro-fibrotic effect of PDL1 in pulmonary fibrosis. (A) PDL1 promotes fibrosis by mediating fibroblast-to-myofibroblast transition and EMT via TGF- β /Smad3 and β -catenin signaling pathways; (B) Overexpression of PDL1 leads to myoblasts' resistance to apoptosis and escape from macrophage phagocytosis through inhibition of p53 pathway or activation of FAK pathway, ultimately leading to excessive proliferation of myoblasts and triggering IPF; (C) PDL1 can inhibit autophagy of fibrosis-related cells through the PI3K/AKT/mTOR signal pathway. PDL1, programmed death ligand 1; EMT, epithelial-mesenchymal transformation; TJ, tight junction; AJ, adhesive junctions; DS, desmosome; TGF- β , transforming growth factor- β ; Smad, skin growth factor; FAK, focal adhesion kinase; PI3K, phosphoinositide 3-kinase; AKT, protein kinase B; mTOR, mammalian target of rapamycin.

impede autophagy, a tumor suppressor process that selectively downregulates key transcription factors associated with EMT, thereby hindering metastasis in the early stages (100); lastly, PDL1 in IPF can induce Tregs and M2 macrophages. Tregs promote EMT in alveolar epithelial cells through β -catenin, while M2 macrophages foster myofibroblast activation and EMT, resulting in collagen deposition and upregulation of fibrogenic cytokines (101, 102). The above three points contribute to our comprehension of the role of PDL1 in IPF, shedding light on the potential interplay between PDL1 and EMT in this condition. Importantly, the latest research in 2023 further clarifies their correlation, as heightened PDL1 levels impede vimentin ubiquitination, consequently amplifying vimentin levels and intensifying the EMT of alveolar epithelial cells, thus exacerbating pulmonary fibrosis (12).

5.2.1.2 immunomodulatory functions of PDL1 in liver fibrosis and its potential impact on EMT

The liver, as a central immune organ in the human body, assumes a crucial role in maintaining overall immune function. Recent studies have shed light on immune dysregulation, particularly the PD1/PDL1 immune checkpoint disorder, as a significant cause of liver function damage and fibrosis (103, 104). In contrast to pulmonary fibrosis, the impact of PDL1 on liver fibrosis primarily stems from its immunomodulatory functions (Figure 5): firstly, in the context of hepatitis C, upregulated PDL1 in hepatocytes induces the generation of Tregs and follicular regulatory T cells, releasing exosomes rich in TGF- β . This inhibits

the host T cell response to persistent infection, thereby promoting liver injury and fibrosis (105); Secondly, hepatic stellate cells (HSCs), are considered to be the main effector cells of liver fibrosis, drive immunosuppressive responses in homeostasis, such as Treg induction, T cell apoptosis, or suppression of cytotoxic CD8+ T cells (106). Current evidence suggests that the activation and augmentation of HSCs are closely associated with the expression of PDL1 (107); lastly, the contribution of M2 macrophages to liver fibrosis has gained substantial recognition, with PDL1 expression believed to promote the expansion of the M2 macrophage population within the microenvironment of liver fibrosis (108).

Liver fibrosis entails a pathological process in which hepatocytes, HSCs, and bile duct cells undergo EMT and transform into myofibroblasts, leading to excessive deposition of ECM and persistent liver injury (109). The mechanism of PDL1 in hepatic fibrosis shares similarities with pulmonary fibrosis and may also affect EMT in the liver in the following ways: firstly, PDL1 can induce the production of TGF- β in liver fibrosis. Consistent with pulmonary fibrosis, EMT signaling pathways involving TGF- β, such as Smad, PI3K/AKT, STAT3 and p53 pathways, also operate in hepatic fibrosis (110-113); secondly, PDL1 activates HSCs, resulting in the secretion of various substances, including ECM components, matrix metalloproteinases, tissue inhibitors of metalloproteinases, chemokines, growth factors, and TGF- β . The latter two exert autocrine or paracrine effects on HSCs and other cells, fostering cell proliferation, migration, and EMT transformation (114); lastly, PDL1 stimulates the polarization of M2 macrophages, which in turn downregulates the expression of E-



FIGURE 5

The immune regulation of PDL1 in hepatic fibrosis, thus promoting the role of fibrosis. (A) PDL1 activates HSCs (HSCs are considered to be the main effector cells of liver fibrosis); (B) PDL1 induces the production of Tregs and releases exocrine bodies showing TGF- β ; (C) PDL1 stimulates the polarization of M2 macrophages. HSCs, hepatic stellate cells; PDL1, programmed death ligand 1; PD1, programmed cell death protein 1; TGF- β , transforming growth factor- β .

cadherin and upregulates the expression of vimentin in hepatocytes, promoting EMT (115). While these three points offer valuable insights into the role of PDL1 in hepatic fibrosis, conclusive evidence linking PDL1 and EMT in this context remains elusive, underscoring the need for further investigation.

5.2.2 The therapeutic potential of targeting PDL1 in type II EMT fibrosis: striking a balance between modulating fibrosis and regulating the immune response

Studies conducted on both individuals and mouse models of tissue fibrosis have shed light on the multifunctionality of the PDL1 axis in fibrosis, encompassing both fibrosis promotion and immunomodulation. Consequently, many preclinical investigations in mouse models have explored the potential of targeting the PD1/PDL1 axis for fibrosis treatment. Many of these studies have demonstrated that obstructing the PD1/PDL1 axis with PDL1 monoclonal antibodies can effectively reduce fibrosis severity in mice (87, 90, 95, 107, 116). However, it is important to note that not all PDL1 antagonistic therapies have yielded successful outcomes. For instance, despite human mesenchymal stem cells being utilized in IPF therapy for their immune regulatory properties, recent animal studies have revealed that administering PD1/PDL1 inhibitors can reverse their antifibrotic effect (117, 118).

The development of fibrosis stems from the intricate interplay between anti-fibrosis and pro-fibrosis mediators within a microenvironment comprised of diverse cell types (95). Research suggests that downregulating PDL1 in fibroblasts can alleviate pulmonary fibrosis through various mechanisms. However, it is important to consider that excessive activation of T cells resulting from this immune response may disrupt this delicate balance required for effective fibrosis modulation (119, 120). Thus, while targeting the PD1/PDL1 axis to reduce fibrosis holds promise as a novel therapeutic approach, extensive research is still needed to determine the optimal equilibrium between immune activation and inhibition of fibrosis.

5.2.3 Potential link between EMT and PDL1 in peritoneal fibrosis: insights from pulmonary and hepatic fibrosis

In line with pulmonary and hepatic fibrosis, peritoneal fibrosis (PF) represents a fibrotic manifestation centered around the process of EMT. Presently, it is widely acknowledged that decreased peritoneal function induced by PF is a major complication of long-term peritoneal dialysis(PD) therapy and a primary cause of long-term ultrafiltration failure of PD (14). The pathological progression of PF bears similarities to that of pulmonary and hepatic fibrosis: firstly, EMT and its intermediate state, driven by the central TGF-B signaling pathway, exert a significant driving force in the genesis and subsequent functional deterioration of PF (4, 15, 16); secondly, M2 macrophages and Tregs may also represent critical components of PF, similar to their impact on pulmonary and hepatic fibrosis (121); thirdly, as observed in the lung and liver, PD1/PDL1 expression can be detected in the abdominal cavity (122, 123); fourthly, human peritoneal mesothelial cells (HPMCs), which are the primary cells involved in PF, are also implicated in various

types of cancer and contribute to the maintenance of the tumor microenvironment. Notably, the phenotypic alterations observed in HPMCs during PF exhibit similarities to those witnessed in cancer (124). Considering these mechanisms, this review postulates a potential correlation between EMT and PDL1 in the field of PF.

6 Conclusion and prospects

Among the three subtypes of EMT, this review has focused on exploring the potential correlation between EMT and PDL1 in fibrosis, as well as the mechanism of immune escape in tumor environments. As mentioned above, EMT serves various physiological functions in the body, and the immune regulation mediated by PDL1 is a fundamental element for normal human survival. However, beyond the realm of tumors, the precise regulatory mechanisms connecting EMT and PDL1 remain elusive.

At present, the research on EMT and PDL1 primarily concentrates on tumor-related studies and membrane surface PDL1, which limits the breadth of investigation in other biological fields, such as embryonic development, wound repair, and fibrosis. An intriguing avenue to explore is the role of extracellular PDL1, which might exert physiological effects similar to PDL1 on the membrane surface. Exploring this aspect could enhance our understanding the EMT process and enable interventions in distant tissues. Furthermore, the existing mechanisms necessitate further exploration. Therefore, this review proposes several hypothetical scenarios in which the manipulation of PDL1 expression could regulate the biological effects of EMT, such as promoting the development of premature infants akin to hormone therapy, affecting the process of fibrosis caused by chronic inflammation to maintain the normal biological function of tissue, and examining the impact of extracellular PDL1 on EMT subtypes beyond tumors.

In conclusion, the intricate bidirectional regulation between EMT and PDL1 underscores the interconnectedness of these fields, as evidenced by shared responses like immune evasion in tumors and pro-fibrotic effects in fibrotic diseases. This interplay presents promising opportunities for the development of innovative therapeutic approaches to effectively manage EMT-associated diseases. However, to fully comprehend the underlying regulatory

References

 Aroeira LS, Aguilera A, Sanchez-TomeroJose'A. Epithelial to mesenchymal transition and peritoneal membrane failure in peritoneal dialysis patients:Pathologic significance and potential therapeutic interventions. J Am Soc Nephrol (2007) 18:2004– 13. doi: 10.1681/ASN.2006111292

2. Nieto Ma, Huang Ry, Jackson Ra, Thiery Jp. Emt: 2016. Cell (2016) 166(1):21-45. doi: 10.1016/j.cell.2016.06.028

3. Grassmann a, Gioberge S, Moeller S, Brown G. Esrd patients in 2004: global overview of patient numbers, treatment modalities and associated trends. *Nephrol Dial Transplant* (2005) 20:2587–93. doi: 10.1093/ndt/gfi159

4. Strippoli R, Moreno-Vicente R, Battistelli C, Cicchini C, Noce V, Amicone L, et al. Molecular mechanisms underlying peritoneal emt and fibrosis. *Stem Cells Int* (2016) 2016:3543678. doi: 10.1155/2016/3543678

mechanisms and identify potential targets for therapeutic interventions, further comprehensive research is indispensable.

Author contributions

YCZ and YTZ contributed to conception and design of the study. YCZ organized the database and wrote the first draft of the manuscript. YTZ and YW wrote sections of the manuscript. All authors contributed to manuscript revision, read, and approved the submitted version.

Funding

The study was supported by grants from the National Natural Science Foundation of China (NSFC: 81770764 and 81770669).

Acknowledgments

Thanks to the teachers of Pathogenic Microbiology Laboratory of Air Force Medical University and senior sisters for their guidance on this article.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

5. Jiang Y, Zhan H. Communication between Emt and Pd-L1 Signaling: New Insights into Tumor Immune Evasion. *Cancer Lett* (2020) 468:72–81. doi: 10.1016/ J.Canlet.2019.10.013

6. Yi M, Zhang J, Li a, Niu M, Yan Y, Jiao Y, et al. The construction, expression, and enhanced anti-tumor activity of ym101: A bispecific antibody simultaneously targeting tgf-B and pd-L1. *J Hematol Oncol* (2021) 14(1):27. doi: 10.1186/S13045-021-01045-X

7. Messeha Ss, Zarmouh No, Soliman K. Polyphenols modulating effects of pd-L1/ pd-1 checkpoint and emt-mediated pd-L1 overexpression in breast cancer. *Nutrients* (2021) 13(5):1718. doi: 10.3390/nu13051718

8. Muralidharan S, Sehgal M, Soundharya R, Mandal S, Majumdar Ss, Yeshwanth M, et al. Pd-L1 activity is associated with partial emt and metabolic reprogramming in carcinomas. *Curr Oncol* (2022) 29(11):8285–301. doi: 10.3390/Curroncol29110654

9. Dongre A, Rashidian M, Eaton En, Reinhardt F, Thiru P, Zagorulya M, et al. Direct and indirect regulators of epithelial-mesenchymal transition-mediated immunosuppression in breast carcinomas. *Cancer Discov* (2021) 11(5):1286–305. doi: 10.1158/2159-8290.Cd-20-0603

10. Mak Mp, Tong P, Diao L, Cardnell Rj, Gibbons Dl, William Wn, et al. A patientderived, pan-cancer emt signature identifies global molecular alterations and immune target enrichment following epithelial-to-mesenchymal transition. *Clin Cancer Res* (2016) 22(3):609–20. doi: 10.1158/1078-0432.Ccr-15-0876

11. Shen M, Xu Z, Xu W, Jiang K, Zhang F, Ding Q, et al. Inhibition of atm reverses emt and decreases metastatic potential of cisplatin-resistant lung cancer cells through jak/stat3/pd-L1 pathway. *J Exp Clin Cancer Res* (2019) 38(1):149. doi: 10.1186/S13046-019-1161-8

12. Li Q, Deng Ms, Wang Rt, Luo H, Luo Yy, Zhang Dd, et al. Pd-L1 upregulation promotes drug-induced pulmonary fibrosis by inhibiting vimentin degradation. *Pharmacol Res* (2023) 187:106636. doi: 10.1016/J.Phrs.2022.106636

13. Ke My, Xu T, Fang Y, Ye Yp, Li Zj, Ren Fg, et al. Liver fibrosis promotes immune escape in hepatocellular carcinoma via golm1-mediated pd-L1 upregulation. *Cancer Lett* (2021) 513:14–25. doi: 10.1016/J.Canlet.2021.05.007

14. Shi Y, Hu Y, Wang Y, Ma X, Tang L, Tao M, et al. Blockade of autophagy prevents the development and progression of peritoneal fibrosis. *Front Pharmacol* (2021) 12:724141. doi: 10.3389/Fphar.2021.724141

15. Margetts Pj, Bonniaud P, Liu L, Hoff Cm, Holmes Cj, West-Mays Ja, et al. Transient overexpression of tgf-{Beta}1 induces epithelial mesenchymal transition in the rodent peritoneum. *J Am Soc Nephrol* (2005) 16(2):425–36. doi: 10.1681/Asn.2004060436

16. Peng D, Fu M, Wang M, Wei Y, Wei X. Targeting tgf-B Signal transduction for fibrosis and cancer therapy. *Mol Cancer* (2022) 21(1):104. doi: 10.1186/S12943-022-01569-X

17. Qin W, Hu L, Zhang X, Jiang S, Li J, Zhang Z, et al. The diverse function of pd-1/ pd-L pathway beyond cancer. *Front Immunol* (2019) 10:2298. doi: 10.3389/ Fimmu.2019.02298

18. Chulkina M, Beswick Ej, Pinchuk Iv. Role of pd-L1 in gut mucosa tolerance and chronic inflammation. Int J Mol Sci (2020) 21(23):9165. doi: 10.3390/Ijms21239165

19. Bu Mt, Chandrasekhar P, Ding L, Hugo W. The roles of tgf-B and vegf pathways in the suppression of antitumor immunity in melanoma and other solid tumors. *Pharmacol Ther* (2022) 240:108211. doi: 10.1016/J.Pharmthera.2022.108211

20. Kim Bg, Malek E, Choi Sh, Ignatz-Hoover Jj, Driscoll Jj. Novel therapies emerging in oncology to target the tgf-B Pathway. *J Hematol Oncol* (2021) 14(1):55. doi: 10.1186/S13045-021-01053-X

21. Kraehenbuehl L, Weng Ch, Eghbali S, Wolchok Jd, Merghoub T. Enhancing immunotherapy in cancer by targeting emerging immunomodulatory pathways. *Nat Rev Clin Oncol* (2022) 19(1):37–50. doi: 10.1038/S41571-021-00552-7

22. Sun C, Mezzadra R, Schumacher Tn. Regulation and function of the pd-L1 checkpoint. *Immunity* (2018) 48(3):434–52. doi: 10.1016/J.Immuni.2018.03.014

23. Niu M, Liu Y, Yi M, Jiao D, Wu K. Biological characteristics and clinical significance of soluble pd-1/pd-L1 and exosomal pd-L1 in cancer. *Front Immunol* (2022) 13:827921. doi: 10.3389/Fimmu.2022.827921

24. Daassi D, Mahoney Km, Freeman Gj. The importance of exosomal pdl1 in tumour immune evasion. *Nat Rev Immunol* (2020) 20(4):209–15. doi: 10.1038/S41577-019-0264-Y

25. Miao L, Lu Y, Xu Y, Zhang G, Huang Z, Gong L, et al. Pd-L1 and C-met expression and survival in patients with small cell lung cancer. *Oncotarget* (2017) 8 (33):53978–88. doi: 10.18632/Oncotarget.9765

26. li Y, Liang L, Dai W, Cai G, Xu Y, Li X, et al. Prognostic impact of programed cell death-1 (Pd-1) and pd-ligand 1 (Pd-L1) expression in cancer cells and tumor infiltrating lymphocytes in colorectal cancer. *Mol Cancer* (2016) 15(1):55. doi: 10.1186/S12943-016-0539-X

27. Zhang Y, Chen Y, Papakonstantinou a, Tsagkozis P, Linder-Stragliotto C, Haglund F. Evaluation of pd-L1 expression in undifferentiated pleomorphic sarcomas, liposarcomas and chondrosarcomas. *Biomolecules*. (2022) 12(2):292. doi: 10.3390/Biom12020292

28. Honda T, Egen Jg, Lämmermann T, Kastenmüller W, Torabi-Parizi P, Germain Rn. Tuning of antigen sensitivity by T cell receptor-dependent negative feedback controls T cell effector function in inflamed tissues. *Immunity* (2014) 40(2):235–47. doi: 10.1016/J.Immuni.2013.11.017

29. Wolf Y, Anderson Ac, Kuchroo Vk. Tim3 comes of age as an inhibitory receptor. *Nat Rev Immunol* (2020) 20(3):173–85. doi: 10.1038/S41577-019-0224-6

30. Curran Cs, Gupta S, Sanz I, Sharon E. Pd-1 immunobiology in systemic lupus erythematosus. J Autoimmun (2019) 97:1–9. doi: 10.1016/J.Jaut.2018.10.025

31. Cha Jh, Chan Lc, Li Cw, Hsu Jl, Hung Mc. Mechanisms controlling pd-L1 expression in cancer. *Mol Cell* (2019) 76(3):359–70. doi: 10.1016/J.Molcel.2019.09.030

32. Wright Jj, Powers Ac, Johnson Db. Endocrine toxicities of immune checkpoint inhibitors. *Nat Rev Endocrinol* (2021) 17(7):389–99. doi: 10.1038/S41574-021-00484-3

33. Kornepati A, Vadlamudi Rk, Curiel Tj. Programmed death ligand 1 signals in cancer cells. *Nat Rev Cancer* (2022) 22(3):174–89. doi: 10.1038/S41568-021-00431-4

34. Cassady K, Martin Pj, Zeng D. Regulation of gvhd and gvl activity via pd-L1 interaction with pd-1 and cd80. *Front Immunol* (2018) 9:3061. doi: 10.3389/ Fimmu.2018.03061 35. Chang Ch, Qiu J, O'sullivan D, Buck Md, Noguchi T, Curtis Jd, et al. Metabolic competition in the tumor microenvironment is a driver of cancer progression. *Cell* (2015) 162(6):1229–41. doi: 10.1016/J.Cell.2015.08.016

36. Gato-Cañas M, Zuazo M, Arasanz H, Ibañez-Vea M, Lorenzo L, Fernandez-Hinojal G, et al. Pdl1 signals through conserved sequence motifs to overcome interferon-mediated cytotoxicity. *Cell Rep* (2017) 20(8):1818–29. doi: 10.1016/ J.Celrep.2017.07.075

37. Mushtaq Mu, Papadas a, Pagenkopf a, Flietner E, Morrow Z, Chaudhary Sg, et al. Tumor matrix remodeling and novel immunotherapies: The promise of matrixderived immune biomarkers. *J Immunother Cancer* (2018) 6(1):65. doi: 10.1186/ S40425-018-0376-0

38. Gulley Jl, Schlom J, Barcellos-Hoff Mh, Wang Xj, Seoane J, Audhuy F, et al. Dual inhibition of tgf-B and pd-L1: A novel approach to cancer treatment. *Mol Oncol* (2022) 16(11):2117–34. doi: 10.1002/1878-0261.13146

39. Lan Y, Moustafa M, Knoll M, Xu C, Furkel J, Lazorchak a, et al. Simultaneous targeting of tgf-B/pd-L1 synergizes with radiotherapy by reprogramming the tumor microenvironment to overcome immune evasion. *Cancer Cell* (2021) 39(10):1388–403. doi: 10.1016/J.Ccell.2021.08.008

40. Lou Y, Diao L, Cuentas Er, Denning Wl, Chen L, Fan Yh, et al. Epithelialmesenchymal transition is associated with a distinct tumor microenvironment including elevation of inflammatory signals and multiple immune checkpoints in lung adenocarcinoma. *Clin Cancer Res* (2016) 22(14):3630–42. doi: 10.1158/1078-0432.Ccr-15-1434

41. Dongre a, Rashidian M, Reinhardt F, Bagnato a, Keckesova Z, Ploegh Hl, et al. Epithelial-to-mesenchymal transition contributes to immunosuppression in breast carcinomas. *Cancer Res* (2017) 77(15):3982–9. doi: 10.1158/0008-5472.Can-16-3292

42. Ueno T, Tsuchikawa T, Hatanaka Kc, Hatanaka Y, Mitsuhashi T, Nakanishi Y, et al. Prognostic impact of programmed cell death ligand 1 (Pd-L1) expression and its association with epithelial-mesenchymal transition in extrahepatic cholangiocarcinoma. *Oncotarget* (2018) 9(28):20034-47. doi: 10.18632/ Oncotarget.25050

43. Tsutsumi S, Saeki H, Nakashima Y, Ito S, Oki E, Morita M, et al. Programmed death-ligand 1 expression at tumor invasive front is associated with epithelialmesenchymal transition and poor prognosis in esophageal squamous cell carcinoma. *Cancer Sci* (2017) 108(6):1119–27. doi: 10.1111/Cas.13237

44. Kim S, Koh J, Kim My, Kwon D, Go H, Kim Ya, et al. Pd-L1 expression is associated with epithelial-to-mesenchymal transition in adenocarcinoma of the lung. *Hum Pathol* (2016) 58:7–14. doi: 10.1016/J.Humpath.2016.07.007

45. Morales-Sanchez A, Fuentes-Panana Em. Epstein-barr virus-associated gastric cancer and potential mechanisms of oncogenesis. *Curr Cancer Drug Targets* (2017) 17 (6):534–54. doi: 10.2174/1568009616666160926124923

46. Alsuliman A, Colak D, Al-Harazi O, Fitwi H, Tulbah A, Al-Tweigeri T, et al. Bidirectional crosstalk between pd-L1 expression and epithelial to mesenchymal transition: Significance in claudin-low breast cancer cells. *Mol Cancer* (2015) 14:149. doi: 10.1186/S12943-015-0421-2

47. Akalay I, Janji B, Hasmim M, NOman Mz, André F, De Cremoux P, et al. Epithelial-to-mesenchymal transition and autophagy induction in breast carcinoma promote escape from T-cell-mediated lysis. *Cancer Res* (2013) 73(8):2418–27. doi: 10.1158/0008-5472.Can-12-2432

48. Martinez-Ciarpaglini C, Oltra S, Roselló S, Roda D, Mongort C, Carrasco F, et al. Low mir200c expression in tumor budding of invasive front predicts worse survival in patients with localized colon cancer and is related to pd-L1 overexpression. *Mod Pathol* (2019) 32(2):306–13. doi: 10.1038/S41379-018-0124-5

49. Chen L, Gibbons Dl, Goswami S, Cortez Ma, Ahn Yh, Byers La, et al. Metastasis is regulated via microrna-200/zeb1 axis control of tumour cell pd-L1 expression and intratumoral immunosuppression. *Nat Commun* (2014) 5:5241. doi: 10.1038/ Ncomms6241

50. Hsu Jm, Xia W, Hsu Yh, Chan Lc, Yu Wh, Cha Jh, et al. Stt3-dependent pd-L1 accumulation on cancer stem cells promotes immune evasion. *Nat Commun* (2018) 9 (1):1908. doi: 10.1038/S41467-018-04313-6

51. Goossens S, Vandamme N, Van Vlierberghe P, Berx G. Emt transcription factors in cancer development re-evaluated: Beyond emt and met. *Biochim Biophys Acta Rev Cancer* (2017) 1868(2):584–91. doi: 10.1016/J.Bbcan.2017.06.006

52. Taliaferro-Smith L, Oberlick E, Liu T, Mcglothen T, Alcaide T, Tobin R, et al. Fak activation is required for igf1r-mediated regulation of emt, migration, and invasion in mesenchymal triple negative breast cancer cells. *Oncotarget* (2015) 6(7):4757–72. doi: 10.18632/Oncotarget.3023

53. Suda K, Rozeboom L, Rivard Cj, Yu H, Ellison K, Melnick MAC, et al. Therapyinduced E-cadherin downregulation alters expression of programmed death ligand-1 in lung cancer cells. *Lung Cancer* (2017) 109:1–8. doi: 10.1016/J.Lungcan.2017.04.010

54. Wang Y, Hu J, Wang Y, Ye W, Zhang X, Ju H, et al. Egfr activation induced snail-dependent emt and myc-dependent pd-L1 in human salivary adenoid cystic carcinoma cells. *Cell Cycle* (2018) 17(12):1457-70. doi: 10.1080/15384101.2018.1489177

 Chen L, Yang Qc, Li Yc, Yang Ll, Liu Jf, Li H, et al. Targeting cmtm6 suppresses stem cell-like properties and enhances antitumor immunity in head and neck squamous cell carcinoma. *Cancer Immunol Res* (2020) 8(2):179–91. doi: 10.1158/ 2326-6066.Cir-19-0394 56. Wang Y, Chen L, Yu M, Fang Y, Qian K, Wang G, et al. Immune-related signature predicts the prognosis and immunotherapy benefit in bladder cancer. *Cancer Med* (2020) 9(20):7729–41. doi: 10.1002/Cam4.3400

57. Hong W, Xue M, Jiang J, Zhang Y, Gao X. Circular rna circ-cpa4/ let-7 mirna/ pd-L1 axis regulates cell growth, stemness, drug resistance and immune evasion in non-small cell lung cancer (Nsclc). *J Exp Clin Cancer Res* (2020) 39(1):149. doi: 10.1186/ S13046-020-01648-1

58. Chen L, Xiong Y, Li J, Zheng X, Zhou Q, Turner A, et al. Pd-L1 expression promotes epithelial to mesenchymal transition in human esophageal cancer. *Cell Physiol Biochem* (2017) 42(6):2267–80. doi: 10.1159/000480000

59. Wang Y, Wang H, Zhao Q, Xia Y, Hu X, Guo J. Pd-L1 induces epithelial-tomesenchymal transition via activating srebp-1c in renal cell carcinoma. *Med Oncol* (2015) 32(8):212. doi: 10.1007/S12032-015-0655-2

60. Kang Jh, Jung My, Choudhury M, Leof Eb. Transforming growth factor beta induces fibroblasts to express and release the immunomodulatory protein pd-L1 into extracellular vesicles. *FASEB J* (2020) 34(2):2213–26. doi: 10.1096/Fj.201902354r

61. Chen W, Wang J, Jia L, Liu J, Tian Y. Attenuation of the programmed cell death-1 pathway increases the M1 polarization of macrophages induced by zymosan. *Cell Death Dis* (2016) 7(2):E2115. doi: 10.1038/Cddis.2016.33

62. Nakayama Y, Mimura K, Tamaki T, Shiraishi K, Kua Lf, Koh V, et al. Phospho –Statl expression as a potential biomarker for anti–Pd–1/anti–Pd–L1 immunotherapy for breast cancer. *Int J Oncol* (2019) 54(6):2030–8. doi: 10.3892/Ijo.2019.4779

63. Li W, Tu J, Liu X, Yang W. Farnesyltransferase inhibitor fti-277 inhibits pd-L1 expression on septic spleen lymphocytes and promotes spleen lymphocyte activation. *Clin Exp Immunol* (2017) 190(1):8–18. doi: 10.1111/Cei.12995

64. Wang W, Chapman Nicole M, Zhang Bo, Li M, Fan M, Laribee RN, et al. Upregulation of pd-L1 via hmgb1-activated irf3 and nf-Kb contributes to uv radiation-induced immune suppression. *Cancer Res* (2019) 79(11):2909–2. doi: 10.1016/J.Ccr.2012.09.022

65. Zhang Y, Velez-Delgado A, Mathew E, Li D, Mendez Fm, Flannagan K, et al. Myeloid cells are required for pd-1/pd-L1 checkpoint activation and the establishment of an immunosuppressive environment in pancreatic cancer. *Gut* (2017) 66(1):124–36. doi: 10.1136/Gutjnl-2016-312078

66. Zhao R, Song Y, Wang Y, Huang Y, Li Z, Cui Y, et al. Pd-1/pd-L1 blockade rescue exhausted cd8+ T cells in gastrointestinal stromal tumours via the pi3k/akt/mtor signalling pathway. *Cell Prolif* (2019) 52(3):E12571. doi: 10.1111/Cpr.12571

67. Chen C, Li S, Xue J, Qi M, Liu X, Huang Y, et al. Pd-L1 tumor-intrinsic signaling and its therapeutic implication in triple-negative breast cancer. *JCI Insight* (2021) 6(8): E131458. doi: 10.1172/Jci.Insight.131458

68. Kudo-Saito C, Shirako H, Ohike M, Tsukamoto N, Kawakami Y. Ccl2 is critical for immunosuppression to promote cancer metastasis. *Clin Exp Metastasis* (2013) 30 (4):393–405. doi: 10.1007/S10585-012-9545-6

69. Sahoo S, Nayak Sp, Hari K, Purkait P, Mandal S, Kishore A, et al. Immunosuppressive traits of the hybrid epithelial/mesenchymal phenotype. *Front Immunol* (2021) 12:797261. doi: 10.3389/Fimmu.2021.797261

70. Tao Z, Ruan H, Sun L, Kuang D, Song Y, Wang Q, et al. Targeting the yb-1/pd-L1 axis to enhance chemotherapy and antitumor immunity. *Cancer Immunol Res* (2019) 7(7):1135–47. doi: 10.1158/2326-6066.Cir-18-0648

71. Henriksen A, Dyhl-Polk A, Chen I, Nielsen D. Checkpoint inhibitors in pancreatic cancer. *Cancer Treat Rev* (2019) 78:17–30. doi: 10.1016/J.Ctrv.2019.06.005

72. Zheng L. Pd-L1 expression in pancreatic cancer. J Natl Cancer Inst (2017) 109 (6):Djw304. doi: 10.1093/Jnci/Djw304

73. Wang Z, Wang S, Goplen Np, Li C, Cheon Is, Dai Q, et al. Pd-1hi cd8+ Resident memory T cells balance immunity and fibrotic sequelae. *Sci Immunol* (2019) 4(36): Eaaw1217. doi: 10.1126/Sciimmunol.Aaw1217

74. Al-Asmakh M, Bawadi H, Hamdan M, Gupta I, Kheraldine H, Jabeen A, et al. Dasatinib and pd-L1 inhibitors provoke toxicity and inhibit angiogenesis in the embryo. *BioMed Pharmacother* (2021) 134:111134. doi: 10.1016/J.Biopha.2020.111134

75. Zeng W, Qin S, Wang R, Zhang Y, Ma X, Tian F, et al. Pdl1 blockage increases fetal resorption and tfr cells but does not affect tfh/tfr ratio and B-cell maturation during allogeneic pregnancy. *Cell Death Dis* (2020) 11(2):119. doi: 10.1038/S41419-020-2313-7

76. Zhang Y, Ma L, Hu X, Ji J, Mor G, Liao A. The role of the pd-1/pd-L1 axis in macrophage differentiation and function during pregnancy. *Hum Reprod* (2019) 34 (1):25–36. doi: 10.1093/Humrep/Dey347

77. Zhang Yh, Aldo P, You Y, Ding J, Kaislasuo J, Petersen Jf, et al. Trophoblastsecreted soluble-pd-L1 modulates macrophage polarization and function. *J Leukoc Biol* (2020) 108(3):983–98. doi: 10.1002/Jlb.1a0420-012rr

78. Bao Z, Zeng W, Zhang D, Wang L, Deng X, Lai J, et al. Snail induces emt and lung metastasis of tumours secreting cxcl2 to promote the invasion of M2-type immunosuppressed macrophages in colorectal cancer. *Int J Biol Sci* (2022) 18 (7):2867–81. doi: 10.7150/Ijbs.66854

79. Liu Ss, Liu C, Lv Xx, Cui B, Yan J, Li Yx, et al. The chemokine ccl1 triggers an amfr-spry1 pathway that promotes differentiation of lung fibroblasts into myofibroblasts and drives pulmonary fibrosis. *Immunity* (2021) 54(9):2042–2056.E8. doi: 10.1016/J.Immuni.2021.06.008

80. Andugulapati Sb, Gourishetti K, Tirunavalli Sk, Shaikh Tb, Sistla R. Biochanin-a ameliorates pulmonary fibrosis by suppressing the tgf-B Mediated emt, myofibroblasts

differentiation and collagen deposition in in vitro and in Vivo systems. *Phytomedicine*. (2020) 78:153298. doi: 10.1016/J.Phymed.2020.153298

81. Wang J, Hu K, Cai X, Yang B, He Q, Wang J, et al. Targeting pi3k/akt signaling for treatment of idiopathic pulmonary fibrosis. *Acta Pharm Sin B* (2022) 12(1):18–32. doi: 10.1016/J.Apsb.2021.07.023

82. Tzouvelekis A, Gomatou G, Bouros E, Trigidou R, Tzilas V, Bouros D. Common pathogenic mechanisms between idiopathic pulmonary fibrosis and lung cancer. *Chest.* (2019) 156(2):383–91. doi: 10.1016/J.Chest.2019.04.114

83. Phan T, Paliogiannis P, Nasrallah Gk, Giordo R, Eid Ah, Fois Ag, et al. Emerging cellular and molecular determinants of idiopathic pulmonary fibrosis. *Cell Mol Life Sci* (2021) 78(5):2031–57. doi: 10.1007/S00018-020-03693-7

84. Jiang A, Liu N, Wang J, Zheng X, Ren M, Zhang W, et al. The role of pd-1/pd-L1 axis in idiopathic pulmonary fibrosis: Friend or Foe? *Front Immunol* (2022) 13:1022228. doi: 10.3389/Fimmu.2022.1022228

85. Fotopoulou C, Khan T, Bracinik J, Glasbey J, Abu-Rustum N, Chiva L, et al. Covidsurg gynecological cancer collaborators. Outcomes of gynecologic cancer surgery during the covid-19 pandemic: An international, multicenter, prospective covidsurg-Gynecologic oncology cancer study. *Am J Obstet Gynecol* (2022) 227(5):735.E1–735.E02. doi: 10.1016/J.Ajog.2022.06.052

86. Mohan N, Hosain S, Zhao J, Shen Y, Luo X, Jiang J, et al. Atezolizumab potentiates tcell-mediated cytotoxicity and coordinates with fak to suppress cell invasion and motility in pd-L1+ Triple negative breast cancer cells. *Oncoimmunology* (2019) 8(9):E1624128. doi: 10.1080/2162402x.2019.1624128

87. Geng Y, Liu X, Liang J, Habiel Dm, Kulur V, Coelho Al, et al. Pd-L1 on invasive fibroblasts drives fibrosis in a humanized model of idiopathic pulmonary fibrosis. *JCI Insight* (2019) 4(6):E125326. doi: 10.1172/Jci.Insight.125326

88. Flanders Kc. Smad3 as a mediator of the fibrotic response. Int J Exp Pathol (2004) 85(2):47-64. doi: 10.1111/J.0959-9673.2004.00377.X

89. Yu W, Hua Y, Qiu H, Hao J, Zou K, Li Z, et al. Pd-L1 promotes tumor growth and progression by activating wip and B-catenin signaling pathways and predicts poor prognosis in lung cancer. *Cell Death Dis* (2020) 11(7):506. doi: 10.1038/S41419-020-2701-Z

90. Lu Y, Zhong W, Liu Y, Chen W, Zhang J, Zeng Z, et al. Anti-pd-L1 antibody alleviates pulmonary fibrosis by inducing autophagy via inhibition of the pi3k/akt/mtor pathway. *Int Immunopharmacol* (2022) 104:108504. doi: 10.1016/J.Intimp.2021.108504

91. Conte E, Fruciano M, Fagone E, Gili E, Caraci F, Iemmolo M, et al. Inhibition of pi3k prevents the proliferation and differentiation of human lung fibroblasts into myofibroblasts: The role of class I P110 isoforms. *PloS One* (2011) 6(10):E24663. doi: 10.1371/Journal.Pone.0024663

92. Sun Y, Zhang Y, Chi P. Pirfenidone suppresses tgf-B1-Induced human intestinal fibroblasts activities by regulating proliferation and apoptosis via the inhibition of the smad and pi3k/akt signaling pathway. *Mol Med Rep* (2018) 18 (4):3907-13. doi: 10.3892/Mmr.2018.9423

93. Shenderov K, Collins Sl, Powell Jd, Horton Mr. Immune dysregulation as a driver of idiopathic pulmonary fibrosis. J Clin Invest (2021) 131(2):E143226. doi: 10.1172/Jci143226

94. Wang B, Bai W, Ma H, Li F. Regulatory effect of pd1/pd-ligand 1 (Pd-L1) on treg cells in patients with idiopathic pulmonary fibrosis. *Med Sci Monit* (2021) 27:E927577. doi: 10.12659/Msm.927577

95. Cui L, Chen Sy, Lerbs T, Lee Jw, Domizi P, Gordon S, et al. Activation of jun in fibroblasts promotes pro-fibrotic programme and modulates protective immunity. *Nat Commun* (2020) 11(1):2795. doi: 10.1038/S41467-020-16466-4

96. Su X, Tan Z, Wang G, Liu Z, Gan C, Yue L, et al. Design, synthesis and biological evaluation of novel diarylacylhydrazones derivatives for the efficient treatment of idiopathic pulmonary fibrosis. *Eur J Med Chem* (2023) 245(Pt 2):114918. doi: 10.1016/ J.Ejmech.2022.114918

97. Shaikh Sb, Prabhu A, Bhandary Yp. Curcumin targets P53-fibrinolytic system in tgf-B1 mediated alveolar epithelial mesenchymal transition in alveolar epithelial cells. Endocr Metab Immune Disord Drug Targets (2021) 21(8):1441–52. doi: 10.2174/1871 530320666200929142503

98. Borok Z. Role for alpha3 integrin in emt and pulmonary fibrosis. J Clin Invest (2009) 119(1):7–10. doi: 10.1172/Jci38084

99. Shao M, Wen Zb, Yang Hh, Zhang Cy, Xiong Jb, Guan Xx, et al. Exogenous angiotensin (1-7) directly inhibits epithelial-mesenchymal transformation induced by transforming growth factor-B1 in alveolar epithelial cells. *BioMed Pharmacother* (2019) 117:109193. doi: 10.1016/J.Biopha.2019.109193

100. Chen Ht, Liu H, Mao Mj, Tan Y, Mo Xq, Meng Xj, et al. Crosstalk between autophagy and epithelial-mesenchymal transition and its application in cancer therapy. *Mol Cancer* (2019) 18(1):101. doi: 10.1186/S12943-019-1030-2

101. Xiong S, Pan X, Xu L, Yang Z, Guo R, Gu Y, et al. Regulatory T cells promote Bcatenin-mediated epithelium-to-mesenchyme transition during radiation-induced pulmonary fibrosis. *Int J Radiat Oncol Biol Phys* (2015) 93(2):425–35. doi: 10.1016/ J.Ijrobp.2015.05.043

102. Li S, Gao S, Jiang Q, Liang Q, Luan J, Zhang R, et al. Clevudine attenuates bleomycin-induced early pulmonary fibrosis via regulating M2 macrophage polarization. *Int Immunopharmacol* (2021) 101(Pt B):108271. doi: 10.1016/J.Intimp.2021.108271

103. Aoki T, Nishida N, Kudo M. Current perspectives on the immunosuppressive niche and role of fibrosis in hepatocellular carcinoma and the development of antitumor immunity. *J Histochem Cytochem* (2022) 70(1):53–81. doi: 10.1369/00221554211056853

104. Salem Ml, El-Badawy A. Programmed death-1/programmed death-L1 signaling pathway and its blockade in hepatitis C virus immunotherapy. *World J Hepatol* (2015) 7(23):2449–58. doi: 10.4254/Wjh.V7.I23.2449

105. Park Sj, Hahn Ys. Hepatocytes infected with hepatitis C virus change immunological features in the liver microenvironment. *Clin Mol Hepatol* (2023) 29 (1):65–76. doi: 10.3350/Cmh.2022.0032

106. Weiskirchen R, Tacke F. Cellular and molecular functions of hepatic stellate cells in inflammatory responses and liver immunology. *Hepatobiliary Surg Nutr* (2014) 3(6):344–63. doi: 10.3978/J.Issn.2304-3881.2014.11.03

107. Yu Mc, Chen Ch, Liang X, Wang L, Gandhi Cr, Fung Jj, et al. Inhibition of Tcell responses by hepatic stellate cells via B7-H1-mediated T-cell apoptosis in mice. *Hepatology*. (2004) 40(6):1312–21. doi: 10.1002/Hep.20488

108. Liu X, Zhou J, Wu H, Chen S, Zhang L, Tang W, et al. Fibrotic immune microenvironment remodeling mediates superior anti-tumor efficacy of a nano-pd-L1 trap in hepatocellular carcinoma. *Mol Ther* (2023) 31(1):119–33. doi: 10.1016/J.Ymthe.2022.09.012

109. Chen Y, Fan Y, Guo Dy, Xu B, Shi Xy, Li Jt, et al. Study on the relationship between hepatic fibrosis and epithelial-mesenchymal transition in intrahepatic cells. *BioMed Pharmacother* (2020) 129:110413. doi: 10.1016/J.Biopha.2020.110413

110. Dewidar B, Meyer C, Dooley S, Meindl-Beinker An. Tgf-B in hepatic stellate cell activation and liver fibrogenesis-updated 2019. *Cells.* (2019) 8(11):1419. doi: 10.3390/Cells8111419

111. Paskeh M, Ghadyani F, Hashemi M, Abbaspour A, Zabolian A, Javanshir S, et al. Biological impact and therapeutic perspective of targeting pi3k/akt signaling in hepatocellular carcinoma: Promises and challenges. *Pharmacol Res* (2023) 187:106553. doi: 10.1016/J.Phrs.2022.106553

112. Yang Yz, Zhao Xj, Xu Hj, Wang Sc, Pan Y, Wang Sj, et al. Magnesium isoglycyrrhizinate ameliorates high fructose-induced liver fibrosis in rat by increasing mir-375-3p to suppress jak2/stat3 pathway and tgf-B1/smad signaling. *Acta Pharmacol Sin* (2019) 40(7):879–94. doi: 10.1038/S41401-018-0194-4

113. Song L, Chen Ty, Zhao Xj, Xu Q, Jiao Rq, Li Jm, et al. Pterostilbene prevents hepatocyte epithelial-mesenchymal transition in fructose-induced liver fibrosis through suppressing mir-34a/sirt1/P53 and tgf-B1/smads signalling. *Br J Pharmacol* (2019) 176 (11):1619–34. doi: 10.1111/Bph.14573

114. Zhou Wc, Zhang Qb, Qiao L. Pathogenesis of liver cirrhosis. World J Gastroenterol (2014) 20(23):7312–24. doi: 10.3748/Wjg.V20.123.7312

115. Ye Y, Xu Y, Lai Y, He W, Li Y, Wang R, et al. Long non-coding rna cox-2 prevents immune evasion and metastasis of hepatocellular carcinoma by altering M1/ M2 macrophage polarization. *J Cell Biochem* (2018) 119(3):2951–63. doi: 10.1002/ Jcb.26509

116. Celada Lj, Kropski Ja, Herazo-Maya Jd, Luo W, Creecy A, Abad at, et al. Pd-1 up-Regulation on Cd4+ T Cells Promotes Pulmonary Fibrosis through Stat3-Mediated II-17a and Tgf-B1 Production. *Sci Transl Med* (2018) 10(460):Eaar8356. doi: 10.1126/ Scitranslmed.Aar8356

117. Yang S, Liu P, Jiang Y, Wang Z, Dai H, Wang C. Therapeutic applications of mesenchymal stem cells in idiopathic pulmonary fibrosis. *Front Cell Dev Biol* (2021) 9:639657. doi: 10.3389/Fcell.2021.639657

118. Ni K, Liu M, Zheng J, Wen L, Chen Q, Xiang Z, et al. Pd-1/pd-L1 pathway mediates the alleviation of pulmonary fibrosis by human mesenchymal stem cells in humanized mice. *Am J Respir Cell Mol Biol* (2018) 58(6):684–95. doi: 10.1165/Rcmb.2017-0326oc

119. Habiel Dm, Espindola Ms, Kitson C, Azzara Av, Coelho Al, Stripp B, et al. Characterization of cd28null T cells in idiopathic pulmonary fibrosis. *Mucosal Immunol* (2019) 12(1):212-22. doi: 10.1038/S41385-018-0082-8

120. Wang W, Wang Q, Xu C, Li Z, Song Z, Zhang Y, et al. Chinese expert consensus on the multidisciplinary management of pneumonitis associated with immune checkpoint inhibitor. *Thorac Cancer* (2022) 13(23):3420–30. doi: 10.1111/1759-7714.14693

121. Chen Yt, Hsu H, Lin Cc, Pan Sy, Liu Sy, Wu Cf, et al. Inflammatory macrophages switch to ccl17-expressing phenotype and promote peritoneal fibrosis. *J Pathol* (2020) 250(1):55–66. doi: 10.1002/Path.5350

122. Chen J, Mu F, Lu T, Ma Y, Du D, Xu K. A gallbladder carcinoma patient with pseudo-progressive remission after hydrogen inhalation. *Onco Targets Ther* (2019) 12:8645–51. doi: 10.2147/Ott.S227217

123. Agaimy A, Michal M, Hadravsky L, Michal M. Follicular dendritic cell sarcoma: Clinicopathologic study of 15 cases with emphasis on novel expression of mdm2, somatostatin receptor 2a, and pd-L1. *Ann Diagn Pathol* (2016) 23:21–8. doi: 10.1016/J.Anndiagpath.2016.05.003

124. Zhang F, Guo C, Cao X, Yan Y, Zhang J, Lv S. Gastric cancer cell-derived extracellular vesicles elevate E2f7 expression and activate the mapk/erk signaling to promote peritoneal metastasis through the delivery of snhg12. *Cell Death Discov* (2022) 8(1):164. doi: 10.1038/S41420-022-00925-6