Check for updates

OPEN ACCESS

EDITED BY Aditya Kulkarni, Avammune Therapeutics, Inc, United States

REVIEWED BY Yihang Qi, Chinese Academy of Medical Sciences and Peking Union Medical College, China Claudio Costantini, University of Perugia, Italy

*CORRESPONDENCE Jibin Lu Valujb@sj-hospital.org

RECEIVED 12 June 2023 ACCEPTED 16 August 2023 PUBLISHED 28 August 2023

CITATION

Liu Y and Lu J (2023) Mechanism and clinical application of thymosin in the treatment of lung cancer. *Front. Immunol.* 14:1237978. doi: 10.3389/fimmu.2023.1237978

COPYRIGHT

© 2023 Liu and Lu. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Mechanism and clinical application of thymosin in the treatment of lung cancer

Yafeng Liu and Jibin Lu*

Department of Thoracic Surgery, Shengjing Hospital of China Medical University, Shenyang, China

Cancer is one of the leading causes of death worldwide. The burden of cancer on public health is becoming more widely acknowledged. Lung cancer has one of the highest incidence and mortality rates of all cancers. The prevalence of early screening, the emergence of targeted therapy, and the development of immunotherapy have all significantly improved the overall prognosis of lung cancer patients. The current state of affairs, however, is not encouraging, and there are issues like poor treatment outcomes for some patients and extremely poor prognoses for those with advanced lung cancer. Because of their potent immunomodulatory capabilities, thymosin drugs are frequently used in the treatment of tumors. The effectiveness of thymosin drugs in the treatment of lung cancer has been demonstrated in numerous studies, which amply demonstrates the potential and future of thymosin drugs for the treatment of lung cancer. The clinical research on thymosin peptide drugs in lung cancer and the basic research on the mechanism of thymosin drugs in anti-lung cancer are both systematically summarized and analyzed in this paper, along with future research directions.

KEYWORDS

thymosin, lung cancer, therapy, thymosin alpha 1, thymalfasin, thymopentin, immunotherapy

1 Introduction

Cancer is one of the main causes of death on a global scale (1). Cancer has been acknowledged to be an increasing public health burden (2). Lung cancer (LC) is the leading cause of cancer deaths worldwide, and it has a relatively high incidence (3). With the popularity of low-dose computed tomography as an early screening test, more and more early lung cancers, particularly early lung adenocarcinoma (LUAD), are being detected (4, 5). LC includes non-small cell lung cancer (NSCLC) and small cell lung cancer (SCLC), and NSCLC contains numerous pathological types such as lung squamous cell carcinoma (LUSC) and lung adenocarcinoma, of which LUAD is the most prevalent pathological type of LC (6). After surgical resection, early-stage LC, especially LUAD including ground-glass opacity components, has a very favorable long-term survival outcome (7, 8). In recent years, the emergence and development of targeted therapy and immunotherapy as well as the application of these methods to neoadjuvant therapy, have also greatly improved the

prognosis of LC patients (9–12). Even though the survival rate of lung cancer patients has gradually increased, the survival of patients with advanced lung cancer is not encouraging, and the 5-year survival rate of patients with advanced LC is even lower than 20% (13). Today, there are still issues such as drug resistance, negative gene mutations, and negative immunotherapy targets for LC patients or poor treatment effects and incapacity to tolerate the toxic side effects of treatment. Therefore, for LC, whether it is the discovery of innovative, safe, and effective treatment options or the investigation of auxiliary therapeutic drugs, the need is still very urgent.

The thymus gland is an organ in the body that has immunological and endocrine functions (14, 15). In the 1960s, researchers extracted hormones such as thymosin from the thymus gland, based on this discovery, the primary active substance of thymosin, thymosin fraction 5, was then isolated and further separated and purified as thymosin II and thymosin alpha1 (T α 1) (16). Thymosin is a class of physiologically active polypeptides released by the thymus gland, and the most widely used thymosin drugs in clinical practice mainly include thymosin, thymosin alpha-1, and thymopentin (17). Thymosin is a mixture of peptides extracted from the thymus glands of animals. Tα1, also known as thymalfasin, is a medication completely consistent with human $T\alpha 1$, which is a small molecule bioactive polypeptide obtained by isolation and purification of thymosin component 5, made up of 28 amino acids and a molecular weight of 3,018 Da (18, 19). The immunoreactive center of thymosin II, a polypeptide fragment composed of 5 amino acids, located in the 32-36th position of the polypeptide, is called thymopentin (20, 21). The most common dosage forms of thymosin drugs include thymosin injection, Tal injection, thymopentin injection, and thymosin enteric-coated tablets/capsules. Thymosin drugs have an immunomodulating and boosting immunity function, suited for treating individuals with a compromised immune status caused by sepsis, and other illnesses (22-24). Additionally, thymosin drugs are also frequently used in the treatment and adjuvant treatment of cancer (25-27). There have been numerous clinical studies and basic mechanism studies that have demonstrated the effectiveness of either thymosin monotherapy or combination therapy for the treatment of LC (26). This fully illustrates the potential of thymosin for the treatment of LC. However, there is currently a lack of systematic summarization of thymosin in the treatment of LC. Therefore, this study organizes and analyzes basic and clinical research in this field (Figure 1), to clarify the current situation and explore future research directions.

2 Summary of the mechanism of thymosin in the treatment of LC

Many scholars have advanced the mechanism of thymosin in LC and undertaken in-depth and extensive research. Numerous mechanisms have been identified so far, including inhibition of LC growth, proliferation, invasion, and migration, and immunomodulatory effects that regulate the activity and function of natural killer cells, T lymphocytes, and immunosuppression, to exert anti-LC effects. The mechanism and effects of thymosin in the



treatment of LC are summarized in detail in Figures 2, 3, and Table 1.

2.1 Inhibit the growth, proliferation, invasion, and migration of LC

At present, a large number of studies have confirmed that thymosin drugs in combination with other drugs or monotherapy have the effect of inhibiting the growth of LC (28-30, 33-35). Both in vitro and in vivo experiments have demonstrated that Ta1 monotherapy can significantly inhibit the growth of NSCLC (28). Chemoprevention, a method, and strategy to reduce cancer incidence and mortality, is defined as the use of natural or synthetic substances, biological agents, and other pharmacological agents in the early stages of cancer to achieve the purpose of impeding, arresting, or even reversing the development of cancer (41). Chemoprevention has a wide range of applications in the field of LC and is one of the potential therapeutic options to reduce the risk of LC (42). According to studies, $T\alpha 1$ can be used as a chemopreventive drug that can effectively prevent the occurrence of LUAD, and Tal also has a direct inhibitory effect on the growth of LUAD (29, 30). In addition to the monotherapy of thymosin drugs, many scholars have also paid attention to the effect and mechanism of thymosin drugs combined with other drugs on LC, these studies shown that $T\alpha 1$ in combination with interferon (IFN) α/β or interleukin-2 (IL-2) can inhibit LUAD growth and that T α 1 has a synergistic effect with IFN α/β or IL-2, and the combination has a better effect of inhibiting tumor growth than Tal monotherapy (33-35). The dysregulation of the JAK/STAT pathway is closely related to LC and other cancers, STAT3 plays an important role in the JAK-STAT signaling pathway, and STAT3 is primarily involved in tumor growth, differentiation, migration, and invasion regulation (43, 44). A kind of matrix metalloproteinase named MMP2 can degrade type IV and type V collagen and elastic fibers, thereby destroying the basement membrane and extracellular matrix, and promoting tumor invasion and metastasis (45). Bo et al. discovered that $T\alpha 1$ inhibited the transcription of MMP2 by inhibiting phosphorylation of STAT3, resulting in a decrease in MMP2



protein expression and inhibiting the migration and invasion of NSCLC cells with programmed cell death ligand 1 (PD-L1) highexpressing, which suggests that $T\alpha 1$ may inhibit the STAT3-MMP2 signaling pathway by stimulating PD-L1 (31). One of the main



FIGURE 3

The major mechanism of thymosin in the treatment of lung cancer By upregulating IFN- γ and CD86, as well as inhibiting CD31, thymosin inhibited the growth of LCLC/LUAD and boosted the immune response of T lymphocytes. Thymosin inhibited the proliferation, migration, and antioxidant activity of LC by inhibiting the expression of ROS, upregulating catalase, peroxide dismutase, and glutathione peroxidase. Thymosin increased CD8⁺ T cell levels, inhibited the growth of LUAD. Thymosin inhibited the invasion and migration of LC by inhibiting STAT3 and then MMP2. ROS, reactive oxygen species; IFN- γ , interferon- γ , STAT3, signal transducer and activator of transcription 3; MMP2, matrix metalloproteinase 2; LCLC, large cell lung cancer; LUAD, lung adenocarcinoma; LC, lung cancer. The figure is partly generated using Servier Medical Art provided by Servier, licensed under a Creative Commons Attribution 3.0 Unported License. components produced by oxidative stress in the body is reactive oxygen species (ROS), which has the role of driving the development of cancer and other diseases, and the increase of ROS can promote the growth, migration, and invasion of tumors (46). ROS is also one of the main causes of LC occurrence and progression (47). A study has confirmed that $T\alpha 1$ has the effect of inhibiting the proliferation and migration of LUAD cells, and $T\alpha 1$ also can inhibit the antioxidant activity of LUAD cells by reducing ROS levels and increasing the activity of antioxidant enzymes like catalase, superoxide dismutase, and glutathione peroxidase (32). Thymosin $\beta 4$ (T $\beta 4$) is also thought to be associated with LC in addition to T α 1. T β 4, a polypeptide with 43 amino acids that is distributed in various tissues of the human body and is one of the main actin regulators in the human body, makes up the largest proportion of β -thymosin (48, 49). According to Huang et al., T β 4 was overexpressed in NSCLC, and after T β 4 was silenced, the activation of Notch1 was inhibited, which inhibited the proliferation and migration, and invasion of LUAD cells (40).

2.2 Immunomodulatory effects

2.2.1 Natural killer cell

Natural killer (NK) cells are important immune cells in the human body and the main effector cells of innate immunity. The use of anti-tumor immunotherapy based on NK cells is being investigated (50). Some of the side effects of chemotherapy are related to its suppression of the immune system. In animal models of LC after chemotherapy, NK cell activity was restored simultaneously with the inhibition of LC growth and an improvement in the prognosis for LC which may be related to the role of NK cells in inhibiting LC growth and metastasis (33, 34, 36). The benefits of T α 1 and IFNs in animal models of LC after chemotherapy also disappeared after the consumption of NK cells, further proving that the effectiveness of T α 1 in LC treatment is likely to be obtained by modulating NK cells (35).

Drug/gene	Major mechanism and effects	References
Τα1	STAT3-MMP2 ↓, ROS↓, catalase↑, peroxide dismutase↑, glutathione peroxidase↑; Prevented lung carcinogenesis, and inhibited the growth, proliferation, invasion, migration, and the antioxidant activity of lung cancer	(28–32),
Tα1 and IFN/IL-2	Inhibited the growth of lung cancer, restored NK cell activity	(33)
T α 1 and IFN α/β and cyclophosphamide	Inhibited the growth of lung cancer, enhanced the effectiveness of cyclophosphamide and restored NK cell activity	(34)
Tα1 and IL-2 and cyclophosphamide	Inhibited the growth of lung cancer, enhanced the effectiveness of cyclophosphamide	(35)
$T\alpha 1$ and $\alpha\beta\text{-}IFN$ and cyclophosphamide	Restored NK cell activity	(36)
$T\alpha 1$ and gemcitabine	ARG1↑, TLRs/MyD88↑; Increased CD8+ T cell levels, inhibited the growth of lung adenocarcinoma, promoted the activation of myeloid- derived suppressor cells	(37)
Tα1-iRGD	CD86 [†] ; Inhibited the growth of large cell lung cancer, promoted T cell activation	(38)
Tα1-RGDR	IL-2↑, IFN-γ↑, CD31↓; Inhibited the growth of large cell lung cancer/lung adenocarcinoma, boosted the immune response of T lymphocytes	(39)
Tβ4 (silencing)	Notch1↓; Inhibited proliferation and invasion of lung adenocarcinoma	(40)

TABLE 1 The major mechanism and effects of thymosin in the treatment of lung cancer.

Tα1, thymosin alpha1; STAT3, signal transducer and activator of transcription 3; MMP2, matrix metalloproteinase 2; ROS, reactive oxygen species; IFN, interferon; IL-2, interleukin-2; ARG1, arginase 1; TLRs, toll-like receptors; MyD88, myeloid differentiation primary response gene 88; RGD, arginine-glycine-aspartic acid; Tβ4, thymosin β4.

2.2.2 T lymphocytes

T lymphocytes are the main components of lymphocytes and play a crucial role in cellular immunity (51). For instance, the success of anti-T lymphocyte inhibitory receptor therapy for PD-L1 and cytotoxic T-lymphocyte associated antigen-4 (CTLA-4) in LC also highlights the important role of CD8⁺T cells in anti-tumor immunotherapy (52, 53). In animal models with LUAD, Tal monotherapy, and $T\alpha 1$ in combination with gencitabine have both been shown to increase CD8⁺T cell levels (37). The antitumor positive effects of Tal and IFNs were counteracted when T lymphocytes are consumed, and after the application of $T\alpha 1$ and IFNs, there was a significant T lymphocyte infiltration in the necrotic area of LC (35). These observations all confirm the idea that the regulation of T lymphocytes is one of how Ta1 exerts anti-LC effects. Targeted drug ligands like the arginine-glycine-aspartic acid (RGD) peptide are a class of polypeptides that have garnered a lot of attention for their use in the transportation of anti-cancer drugs (54). RGD peptide can target drug delivery into tumors by binding to integrins, such as $\alpha v\beta 3$ and $\alpha v\beta 5$, which are highly expressed in tumor blood vessels (55). RGD peptides with high affinity with αv integrins and neuropilin-1 are known as iRGD peptides (56). It has been demonstrated in the large cell lung cancer (LCLC) model that the use of iRGD and T α 1 fusion to form modified Ta1, or Ta1-iRGD, can promote T cell activation, can restore thymus index and spleen index, and also activate T lymphocytes by increasing the expression of CD86 to exert their immune effect (38). In addition, compared to $T\alpha 1$, the modified Tal has better immunomodulatory activity and stronger LC inhibition and also exhibits lower tumor vascular density, which is associated with the improved targeting of the modified T α 1iRGD. A study also confirmed that binding $T\alpha 1$ to RGD peptides compensated for the lack of tumor targeting of T α 1 (39). RGDR can bind to highly expressed $\alpha\nu\beta$ 3 and NRP-1 domains on the tumor surface because it includes the RGD sequence. Integrin $\alpha\nu\beta$ 3 is an integrin protein closely related to tumor angiogenesis and specifically able to recognize RGD sequences (55). NRP-1 is a transmembrane glycoprotein that is widely expressed in tumor tissues and plays a key role in tumor angiogenesis (57, 58). Therefore, T α 1-RGDR formed by fusing T α 1 with RGDR can achieve a good tumor-targeting effect. T α 1-RGDR showed stronger anti-LUAD and anti-LCLC effects than the T α 1, and it can boost the immune response of T lymphocytes by increasing IL-2 and IFN- γ levels and promoting the invasion of CD4⁺T cells and CD8⁺T cells into tumor tissues to exert anti-LC effects.

2.2.3 Immunosuppression

Tal has potent anti-LC and immunomodulatory effects when used alone or in combination with other medications. However, $T\alpha 1$ and chemotherapy drugs show a stronger anti-lung cancer and immunomodulatory effect than $T\alpha 1$ alone, which may be related to the synergistic effect of $T\alpha 1$ with other medications. A study showed that neither Tal nor gemcitabine (GEM) alone could inhibit the growth of LUAD, but when combined Ta1 with GEM showed higher levels of CD8⁺T cells and the ability to significantly inhibit LUAD growth, indicating that GEM and Ta1 do have a certain synergistic effect (37). The study also discovered that $T\alpha 1$ monotherapy can upregulate Arginase 1 (ARG1) to activate myeloid-derived suppressor cells (MDSCs) in animal models of LUAD to produce inhibitory anti-tumor immune effects, which may account for the limited effect of Ta1 monotherapy. According to some reports, Ta1 interacts with toll-like receptors (TLRs) and activates MyD88 signaling in dendritic cells, and upregulation of ARG1 relies on the

TLRs/MyD88 signaling pathway (59, 60). That is, T α 1 increases the expression of ARG1 by activating TLRs/MyD88 signaling pathways, thereby activating the immunosuppressive ability of MDSCs.

3 Summary of the clinical studies of thymosin in the treatment of LC

T α 1 is frequently used in clinical practice because it acts as an immunomodulator, boosting the body's defenses and enhancing the quality of life for LC patients (61). Numerous clinical studies have demonstrated that the use of T α 1 in combination with chemotherapy, radiotherapy, concurrent chemoradiotherapy (CCRT), targeted therapy, or as postoperative adjuvant therapy can significantly increase the clinical efficacy of LC patients, lower their risk of recurrence, and lengthen their time of survival. All the clinical studies mentioned in the paper are summarized in Table 2.

3.1 Combination with chemotherapy

Chemotherapy is a common treatment for patients with advanced LC and those with early-stage, high-risk LC (69, 70). Compared to the control group of NSCLC patients receiving only ifosfamide chemotherapy, patients receiving T α 1 and low-dose interferon- α in addition to chemotherapy had higher response rates (33% vs. 10%) and longer times to progression (TTP) (p=0.0059), according to a phase II clinical study. In the chemotherapy group, CD4⁺, CD8⁺, and NK cell counts significantly declined. Combination therapy with T α 1 reduced the toxicity of chemotherapy while leaving lymphocyte counts essentially unchanged from before chemotherapy (62). The

effectiveness of thymosin in combination with chemotherapy was assessed in a meta-analysis of 10 randomized controlled trials (RCTs) in a total of 724 patients with advanced (stage III, IV) NSCLC regimens, primarily cisplatin plus vinorelbine (NP) regimens and gemcitabine plus cisplatin (GP) regimens. Thymosin plus NP was found to increase overall response rate (ORR) (odds ratio (OR) 1.86) compared with the control group using NP-only chemotherapy, and the addition of thymosin also increased tumor control rate (TCR) (OR 3.06), 1-year survival rate (OR 3.05) and improved the quality of life (QOL) (OR 3.39). Thymosin and GP-containing chemotherapy showed superior ORR (OR 1.67), TCR (OR 2.38), and QOL (OR 3.84) compared to GP-only chemotherapy. These findings imply that for patients with advanced NSCLC, thymosin plus NP/GP chemotherapy is preferable to NP/GP chemotherapy alone (63). Another metaanalysis included 27 RCTs involving a total of 1925 patients with stage IIIa-IV NSCLC, of whom 976 received combination therapy with synthetic thymic peptides (sTPs) (including Tal and thymopentin) and chemotherapy and 949 received chemotherapy alone to evaluate the clinical effects of sTPs plus chemotherapy (64). In comparison to chemotherapy, the use of sTPs significantly improved QOL [relative risk (RR) = 2.05, P<0.00001], ORR [RR = 1.28, P = 0.0001], and one-year overall survival (OS) rate [RR = 1.43, P = 0.001], as well as increased disease control rate (DCR) [RR = 1.10, P = 0.02]. The combination of sTPs and chemotherapy demonstrated that the combination therapy was clinically effective. Combination therapy with sTPs and chemotherapy increased CD3⁺T cell levels [P<0.00001], CD3⁺CD4⁺T cell levels [P<0.00001], NK cell numbers [P<0.00001], and the proportion of CD4⁺/CD8⁺T cells [P<0.00001] at the lymphocyte level, and it can lessen the immunotoxic effects of chemotherapy when combined with sTPs. Additionally, the risk of chemotherapy-related side

TABLE 2	The clinical	effects of	thymosin in	the	treatment	of lung	cancer.
---------	--------------	------------	-------------	-----	-----------	---------	---------

The usage of thymosin	Drug	Conclusion	References
	Tα1 and low-dose IFN-α	Increased the response rates of chemotherapy, prolonged the time to progression	(62)
Combination with chemotherapy	Τα1	Increased ORR, TCR, 1-year survival rate and QOL	(63)
	Tα1/ thymopentin	Increased ORR, QOL, one-year OS rate, DCR, CD3 ⁺ T cell levels, CD3 ⁺ CD4 ⁺ T cell levels, NK cell numbers, and the proportion of CD4 ⁺ /CD8 ⁺ T cells, decreased the risk of neutropenia, thrombocytopenia, and gastrointestinal reactions	(64)
Combination with radiotherapy	Τα1	Improved OS, DFS, and restored T helper cell levels	(65)
Combination with concurrent chemoradiotherapy	Τα1	Decreased the incidence of grade 2 or higher RP and grade 3 to grade 4 lymphopenia, increased total lymphocyte count, and decreased CRP	(66)
Combination with targeted therapy	Tα1/ thymopentin	Improved median OS and median PFS, restored the number of CD3 ⁺ T cells and CD3 ⁺ CD4 ⁺ T/CD3 ⁺ CD8 ⁺ T subsets	(67)
Postoperative adjuvant therapy	Τα1	Increased 5-year OS rate and 5-year DFS rate	(68)

Tα1, thymosin alpha1; IFN, interferon; ORR, overall response rate; TCR, tumor control rate; QOL, quality of life; DCR, disease control rate; NK, nature killer; OS, overall survival; DFS, disease-free survival; RP, radiation pneumonitis; CRP, C-reactive protein; PFS, progression free survival.

10.3389/fimmu.2023.1237978

effects like neutropenia (RR=0.75, P=0.04), thrombocytopenia (RR=0.68, P=0.0002), and gastrointestinal reactions (RR=0.62, P<0.00001) was decreased by sTPs. To compare the two sTPs of T α 1 and thymopentin, further subgroup analysis was done. The results showed that T α 1 could significantly increase ORR [RR: 1.32, P=0.0003] and DCR [RR: 1.11, P=0.02], but that there were no statistically significant differences in OR and DCR in the thymopentin subgroup. T α 1 and thymopentin could also increase the level of peripheral blood lymphocytes. It was demonstrated that T α 1 could boost clinical effectiveness while also boosting immune function.

3.2 Combination with radiotherapy

Although radiation therapy is one of the most effective treatments for advanced lung cancer, it invariably results in side effects, such as immune response (71). An RCT study analyzed the effect of synthetic Tal on immune recovery properties in NSCLC patients after radiotherapy (65). The findings revealed that thymosin treatment following radiotherapy significantly improved OS (P=0.009) and disease-free survival (DFS) (P=0.04) compared with placebo, and thymosin restored the decrease in T helper cell levels brought on by radiotherapy (p=0.04). This proved that T α 1 can both enhance the efficacy of radiotherapy and reverse immunosuppression brought on by radiotherapy. The experimental group received two regimens of subcutaneous injection twice a week with T α 1 900 mg/m2 (body surface area) or continuous thymosin 900 mg/m2 subcutaneous injection once daily for 14 days, followed by twice-weekly subcutaneous injection. The study evaluated the difference in efficacy between different dosing regimens, with the control group receiving a placebo subcutaneous injection regimen. The results showed that the control group showed shoddy results. In the experimental group, there was no discernible difference between the two regimens, indicating that both regimens might be practicable.

3.3 Combination with concurrent chemoradiotherapy

For stage III unresectable NSCLC, consolidation immunotherapy followed by concurrent chemoradiotherapy (CCRT) is the gold standard of care (72). Radiation pneumonitis (RP) and pulmonary fibrosis are the two stages of radiation lung injury (RILI), a common side effect of radiation therapy (RT) in the chest (73, 74). One of the major barriers to the use of consolidation immunotherapy following CCRT is RP, which has a detrimental effect on survival and quality of life in patients with locally advanced NSCLC (LANSCLC). The GASTO-1043 study, a Phase II clinical trial, was explored the efficacy of T α 1 in the treatment of RP in patients with LANSCLC receiving CCRT (66). 138 patients with LANSCLC in total were enrolled in the study, 69 of whom received CCRT alone in the control group and 69 of whom received T α 1 treatment from the start of CCRT to 2 months after it ended in the experimental group. Compared to the control group, the experimental group had a lower incidence of grade 2 or higher RP (36.2% vs. 53.6%, P=0.040), lower incidence of G3 to G4 lymphopenia (19.1% vs. 62.1%, P<0.001), and significantly higher total lymphocyte count (TLC) (0.51 k/mL vs. 0.30 k/mL, P<0.001). Patients with maximum C-reactive protein (CRP) ≥ 100 mg/L (13.8% vs. 29.7%, P = 0.029) and patients with 2 to 3 weeks during CCRT CRP \geq 10 mg/L (23.5% vs. 48.5%, P = 0.003) were significantly less common in the experimental group. Patients with LANSCLC who used Ta1 during and after CCRT showed lower rates of grade 2 RP and G3 to G4 lymphopenia compared to the control group, as well as higher TLC levels and lower CRP, demonstrating that Tal can treat side effects and immune dysfunction brought on by CCRT. Additionally, patients receiving Tal had a longer median progression free survival (PFS) (14.4 months vs. 10.7 months, P=0.131) and a longer median OS (34.8 months vs. 28.7 months in the control group, P=0.062). Despite being superior to the control group in both OS and PFS, however, the Tal experimental group did not achieve a statistically significant difference, which may be related to the smaller sample size.

3.4 Combination with targeted therapy

Patients with LUAD who have an EGFR target mutation that is positive can benefit from EGFR-tyrosine kinase inhibitors (TKI) targeted therapy, which is now one of the standard treatments for LC (75). In patients with IV-stage LUAD and EGFR mutations, some researchers have compared the clinical effects of EGFR-TKI monotherapy and EGFR-TKI plus thymosin therapy (67). To balance the baseline differences, propensity score matching (PSM) was carried out in a 2:1 ratio between the control group and the experimental group, with 130 LUAD patients being treated with first-generation EGFR-TKI (gefitinib/erlotinib/icotinib) and as part of the experimental group 65 LUAD patients being treated with EGFR-TKI plus thymosin (including Tal and thymopentin). The results revealed that the median PFS (14.4 months vs. 9.2 months, HR=0.433, P<0.0001) and median OS (29.5 months vs. 19.8 months, HR=0.430, P<0.0001) in the EGFR-TKI plus thymosin group were both significantly longer than those in the EGFR-TKI group, respectively. These findings demonstrated that EGFR-TKI plus thymosin was extremely effective in lowering the risk of relapse and improving prognosis in patients with LUAD compared with EGFR-TKI alone. The safety of EGFR-TKI plus thymosin was acceptable because there was no significant difference in ORR and DCR between the two groups, nor was there any significant difference in side effects. The number of CD3⁺T cells and the number of CD3⁺CD4⁺T and CD3⁺CD8⁺T subsets after treatment were significantly reduced in the EGFR-TKI monotherapy group (P<0.05), but there was no difference between these parameters in the EGFR-TKI plus thymosin group, indicating that the addition of thymosin based on targeted therapy has some effect on restoring the body's immune activity. Further subgroup analysis revealed that in most subgroups, including age, EGFR mutation type (exon 19 deletion or exon 21 L858R mutation), presence or absence of bone metastases, central nervous system metastases, adrenal

metastases, multiple lung metastases, pleural metastases, gefitinib or erlotinib, EGFR-TKI plus thymosin demonstrated a superior PFS advantage over EGFR-TKI monotherapy. There was no statistically significant difference in OS or PFS between the EGFR-TKI plus thymosin group and the single-agent group in patients with ECOG score of 2 who had liver metastases, received icotinib, and received EGFR-TKI as second-line therapy. Male patients could benefit from combination therapy in OS, and patients with LUAD who received radiotherapy solely for bone metastases benefited from combination therapy in PFS(P=0.041), while patients with LUAD who received radiotherapy for the remaining causes did not benefit in terms of OS or PFS. A comparison of the two thymosin medications revealed no statistically significant difference in PFS or OS between T α 1 and thymopentin, suggesting that the use of both thymosin medications in combination with an EGFR-TKI may be possible.

3.5 Postoperative adjuvant therapy

Because it boosts human immunity and enhances patients' quality of life after LC surgery, Ta1 is frequently used in clinical practice as adjuvant therapy for postoperative LC patients. The longterm survival effects of Tal as postoperative immunomodulatory therapy were examined in a large, real-world study involving 5746 patients with NSCLC who underwent surgery and achieved R0 resection (68). The study compared 1027 operable (stage I-IIIA) NSCLC patients who received thymosin treatment to 1027 patients who did not receive thymosin treatment using PSM analysis. When compared to the control group without Tal treatment, the use of Tal in postoperative patients can lower the risk of recurrence and death in NSCLC patients. Even after multivariate analysis, Ta1 treatment still showed very significant differences in DFS (HR=0.655, P<0.0001) and OS (HR=0.548, P<0.0001). Additionally, following PSM, the T α 1 treatment group outperformed the control group in terms of 5-year DFS (77.3% vs. 64.7%, P<0.0001) and 5-year OS (83.3% vs. 72.7%, P<0.0001). Tal therapy may be an efficient postoperative adjuvant therapy in the early stage, especially in patients with stage IA NSCLC, as evidenced by the fact that the benefits of OS and DFS in patients with stage IA were more advantageous than those of all other stages in terms of staging (P<0.0001). According to pathological classification, other nonadenocarcinoma and non-squamous cell carcinoma subtypes were independently linked to worse OS (HR=2.019, P=0.0009) and DFS (HR=1.706, P=0.0038). The results of the subgroup analysis showed that patients who used T α 1 for more than 24 months had the best OS and DFS efficacy. The treatment effect was proportional to the length of treatment, with the best effect for more than 24 months, followed by treatment for 12-24 months, and the worst effects for less than 12 months. The 5-year DFS rates of the three groups were 84.7%, 81.0%, and 66.1%, respectively, and the 5-year OS rates were 92.2%, 83.7%, and 64.5%. This suggests that to reap the greatest benefits, patients with NSCLC may use T α 1 for longer than 24 months. T α 1 could show efficacy in OS and DFS in patients of any age, sex, Charlson Comorbidity Index (CCI), smoking status, and any stage of pathologic TNM, as well as patients with non-squamous cell carcinoma NSCLC who did not receive targeted therapy and BMI<28 kg/m². This was confirmed by subgroup analysis. T α 1 only demonstrated OS efficacy in postoperative NSCLC patients who received chemoradiotherapy, and it did not affect OS or DFS in patients with a BMI>28 kg/m², the pathological type of squamous cell carcinoma, targeted therapy, or combination therapy (including targeted therapy plus chemotherapy, targeted therapy plus chemoradiotherapy). T α 1 can be used as an adjuvant therapy after surgery to lower the risk of recurrence and increase the long-term survival of LC patients as well as an immunomodulatory therapy to enhance the postoperative quality of life of LC patients.

3.6 Combination with immunotherapy

Immunotherapy, an emerging therapeutic approach in recent years, has not only improved patient prognosis but also significantly altered the initial treatment regimen of LC patients. In the field of LC, immune checkpoint inhibitors (ICIs) that represent PDL1/PD-1 have the most applications and the best efficacy (76). The gold standard for immunotherapy in patients with advanced NSCLC is high PDL1 expression (PD-L1 Tumor Proportion Score 50%). Patients with NSCLC with high PDL1 expression can achieve better immunotherapy outcomes (77), this has also been confirmed by many RCTs (78, 79). Patients with high PDL1 expression in their NSCLC may respond better to immunotherapy. After long-term follow-up, a study of patients with metastatic melanoma who were given $T\alpha 1$ discovered that those who also received the CTLA-4 ICI ipilimumab had an advantage in overall survival (80), this suggests that $T\alpha 1$ and ICIs work in concert. It is expected that the combination of $T\alpha 1$ and ICIs, represented by PD-1 inhibitors, may also have a beneficial synergistic effect on the treatment of LC. The prognosis of LC patients can be significantly improved by ICIs, but their clinical use is constrained by a few drawbacks. First off, a large number of LC patients are unable to respond to ICIs treatment effectively because some LC patients belong to cold immune tumors (such as immunedesert type and immune-excluded type), which lack lymphocyte infiltration (81, 82). If cold immune tumors can be transformed into hot immune tumors, this will greatly expand the clinical application range of ICIs and increase their clinical efficacy. The degree of immune cell infiltration in the tumor microenvironment (TME) is the primary distinction between cold immune tumors and hot immune tumors. NK cells and CD3⁺, CD4⁺, and CD8⁺T lymphocytes are currently thought to be the primary immune cells in the TME, with CD8⁺T lymphocytes playing a significant role in tumor killing (83, 84). Tal, as an immunomodulator of thymic hormones, belongs to active immunity (18). As previously stated, using Ta-1 to treat LC can increase CD8⁺T cell counts, restore NK cell activity, and activate T lymphocyte immune function by increasing CD86 expression. When chemotherapy was combined with thymosin, more CD4⁺, CD8⁺, CD3⁺, and NK cells were present. Additionally, the CD4⁺/CD8⁺T cell ratios were higher than when chemotherapy was used alone. Thymosin also has the effect of increasing the helper T cell counts that have been decreased as a result of radiotherapy. Thymosin was also added to address the negative effects of EGFR-TKI treatment on the quantity

of CD3⁺T cells produced by LC patients as well as the number of CD3⁺CD4⁺T and CD3⁺CD8⁺T subsets. In addition to improving Tal targeting, Tal-RGDR promoted CD4⁺ and CD8⁺T cell infiltration of LC tissues and increased the immune activity of T cells by raising levels of IL-2 and IFN-y. These findings imply that Tal can increase CD4⁺T, CD8⁺T, CD3⁺CD4⁺T, and NK cell numbers in LC patients. Tal has the potential to convert LC patients with cold immune tumors to hot immune tumors by increasing the cellular level and cell activity of immune cells like CD8⁺T cells, enhancing the clinical effectiveness of ICIs. Similarly, Tα1 can inhibit the migration and invasion of NSCLC with high expression of PD-L1 (31) and a favorable effect on immune cells, suggesting that the combination of $T\alpha 1$ and ICIs may enhance the clinical effect of ICIs and even increase the proportion of LC patients who have an effective response to ICIs. Second, immunotherapy with ICIs is associated with adverse effects such as colitis and/or diarrhea (85). Studies have shown that Ta1 can enhance the small intestine tissue structure and barrier function of mice with cystic fibrosis (86). Additionally, a study has shown that Tal can successfully prevent ICIs-induced colitis in preclinical models by fostering the immune pathway of indoleamine 2,3dioxygenase 1-dependent tolerance (87). These studies demonstrate that $T\alpha 1$ has reduced the side effects of ICIs on the digestive system.

4 Discussion

Although the potential benefits of $T\alpha 1$ in combination with immunotherapy in the treatment of LC patients have been outlined above, some issues still need to be resolved before actual clinical use. Numerous studies have shown that Ta1 increases the number and immune activity of immune cells in LC patients, but more basic studies are required to elucidate the detailed regulatory mechanism. The degree to which T α 1 promotes TME lymphocyte infiltration in LC is another area that needs to be studied more in the future, but there is currently a dearth of research in this area. However, this hypothesis needs to be confirmed in further clinical studies. The current clinical evidence is insufficient, and before they can be cautiously applied to clinical practice, exploratory clinical trials with the support of basic research conclusions must be conducted for LC patients treated with ICIs because the addition of Ta1 may also lead to immune dysfunction and interfere with immunotherapy. Tal has demonstrated the ability to reduce immunotherapy-related adverse reactions in addition to the potential to enhance the efficacy and effectiveness of the response of LC patients to immunotherapy. However, the current basic research is momentarily insufficient, necessitating more extensive basic research and preclinical research on the precise mechanism of $T\alpha 1$ and ICIs in combination therapy for LC and immunotherapy-related adverse reactions, which is also one of the possible research directions of $T\alpha 1$ in the future. In the field of immunotherapy, NK cells have demonstrated effective antitumor immunity in addition to ICIs, whether through their immune activity, adoptive cell therapy of NK cells, or even the use of CAR-NK cell therapy (88, 89). This article summarizes how T α 1 affects NK cell activity to function as an inhibitor of LC growth and metastasis. An in-depth investigation of the precise mechanism by which T α 1 enhances NK activity in LC models following chemotherapy, as well as the precise mechanism by which T α 1 and IFNs have a synergistic effect, is a very valuable research direction. Additionally, determining whether T α 1 and CAR-NK have a synergistic effect in the treatment of LC is another important research question.

The side effects of chemotherapy, radiotherapy, surgery, targeted therapy, and CCRT can all be significantly reduced with T α 1. The impact of T α 1 on reducing complications in LC patients who receive CCRT plus immunotherapy consolidation is something to look forward to because it makes it possible to perform consolidation immunotherapy after CCRT by reducing complications like RP associated with CCRT. The combination of $T\alpha 1$ and chemotherapy for LC has the most studies, both basic and clinical studies, and the conclusion that $T\alpha 1$ increases efficacy and reduces the toxicity of chemotherapy is firm. However, the chemotherapy regimens in these studies either include LC patients receiving NP/GP regimens or use cyclophosphamide to treat animal models, $T\alpha 1$ and other chemotherapy drugs have not yet been supplemented. The activation of MDSCs' immunosuppressive activity by $T\alpha 1$ monotherapy is one of the mechanisms that $T\alpha 1$ monotherapy can inhibit anti-tumor immunity. It is also proposed that $T\alpha 1$'s anti-tumor efficacy can be restored by blocking the MyD88 signaling pathway, raising the possibility that future research may examine ways to mitigate Ta1's adverse effects when used to treat LC. The specific mechanism of potential side effects when $T\alpha 1$ is combined with targeted therapy drugs and immunotherapy drugs, as well as the investigation of whether Tal has other side effects, are also worthwhile research areas because prior basic research has only combined chemotherapy drugs. Although baseline differences were balanced using PSM analysis in the large real-world study that showed that $T\alpha 1$ significantly improved OS and PFS in patients with LUAD undergoing surgery, this is retrospective, and prospective clinical trials are still required to provide higher-level evidence-based medical evidence. Tal as an immunomodulator can improve PFS and OS and reduce postoperative complications in patients with LC, but it is unclear what effect of $T\alpha 1$ in the short term after LC surgery. A new perioperative management strategy called "enhanced recovery after surgery" (ERAS) seeks to improve surgical outcomes and quality of life by reducing postoperative complications and hospital stays in surgical patients while also speeding up their recovery and rehabilitation (90). ERAS was initially mainly used in colorectal surgery (91), the ERAS concept however, has become increasingly popular in recent years among patients undergoing thoracic surgery, especially pneumonectomy (92). The long-term OS and PFS benefits of $T\alpha 1$ in LC patients have been confirmed, and T α 1 also has the potential to reduce complications, restore immune function, and improve quality of life in the short term after surgery, so future clinical studies are needed to determine whether Ta1 can be used postoperatively to further enhance the effect of ERAS.

We discovered that some studies had some restrictions and flaws. Meta-analyses had the highest level of evidence-based

evidence, however, the quality of meta-analyses depended on the quality of the studies included in the analyses, which could vary widely from meta-analysis to meta-analysis (93). Although both meta-analyses decided to include only RCTs, many of the trials included in the studies were of poor methodological quality, except a small number of studies that explicitly proposed the use of the envelope method as a randomization regimen, and the remaining studies did not report methods of randomization. Two metaanalyses assessing the effects of thymosin in combination with chemotherapy were reviewed above. There were differences in some variables' heterogeneity and the timing of index tests across studies. In addition, compared to typical phase III RCTs, the RCTs included in these two meta-analyses still have several flaws, most studies are small studies, most studies only involve not more than 100 patients, and there are few trials and samples that can be used to analyze survival. None of the trials reported PFS, these two metaanalyses of RCTs concluded that they did not reach the highest level of evidence because the majority of studies lacked NCT numbers, and the quality of the primary and secondary outcomes was moderate to very low according to GRADE grading. These shortcomings may be related to the earlier publication years of about half of the included studies. As the saying goes, "We can't make bricks without straw," these randomized studies have been able to demonstrate the efficacy of thymosin-combined chemotherapy in treating LC, and higher-quality meta-analyses need to be updated following the completion of well-designed large-scale RCTs.

Additionally, we discovered that Tal usage duration, dosage, and frequency varied significantly between studies. Continuous injection of Ta1 for more than 24 months had the best therapeutic effect in studies of Tal as a postoperative adjuvant therapy for LC patients, but there were significant differences in the duration of T α 1 use in other studies, such as clinical trials investigating the therapeutic effect of Ta1 on CCRT-induced side effects, which were used from the beginning of receiving CCRT to 2 months after the end of CCRT. These differences may be because different treatments often correspond to different clinical stages. Although the goal of adding $T\alpha 1$ to various treatments varies, it is necessary to standardize the time of $T\alpha 1$ usage for treatments. The majority of studies used Ta1 twice weekly, while a small subset used it on a weekly schedule. T α 1 usage frequency was also inconsistent. The dosage of T α 1 is typically 1.6 mg, and 1.6 mg per time is the current mainstream usage of $T\alpha 1$. However, the use scheme is still somewhat unclear, necessitating additional research to determine the best treatment strategy and implement unified, standardized treatment.

Conclusions from various clinical studies disagree with one another (66, 68). In the GASTO-1043 study, LC patients receiving CCRT failed to achieve a statistically significant median OS dlifference after adding T α 1 (34.8 vs. 28.7 months, P=0.062), and in a retrospective analysis examining the function of T α 1 as postoperative adjuvant therapy, LC patients receiving chemoradiotherapy could benefit from OS after T α 1 as adjuvant therapy. The difference between the conclusions of the two studies may be that patients with LC who received T α 1 as postoperative adjuvant therapy, and patients with improved treatment efficacy and could receive surgery had an earlier stage of TNM, and the GASTO-1043 study included patients with advanced LC, had a relatively small sample size of 138 patients, and had relatively insufficient postoperative follow-up time, which may have contributed to the failure to achieve a statistically significant difference in OS after the addition of T α 1 to CCRT. To determine the OS benefit of T α 1 in LC patients receiving CCRT, phase III clinical trials with longer follow-up times are required.

The combination of thymosin and targeted therapy is also controversial. According to the studies reviewed above, the combination of thymosin and EGFR-TKIs showed superior benefits for OS and DFS compared to EGFR-TKI monotherapy (67), whereas postoperative adjuvant therapy with T α 1 plus EGFR-TKIs (including targeted plus chemotherapy, targeted plus chemoradiotherapy) did not show better benefits in OS and DFS in LC patients with LC who were adjuvant with $T\alpha 1$ plus EGFR-TKIs (68). As postoperative patients with LC treated with adjuvant thymosin were stage I-IIIA and primarily stage I, whereas the other study included LUAD patients in the analysis were stage IV, the different effects of EGFR-TKIs in combination with thymosin may be related to staging. Additionally, LC patients received firstgeneration EGFR-TKI in studies of thymosin combined with targeted therapy, and variations in staging and EGFR-TKIs may be the reason why the findings of the two studies are completely dissimilar. Two studies investigating the combined use of EGFR-TKIs and thymosin included 130 and 112 patients, respectively, and both are retrospective studies. The level of evidence-based medical evidence is not sufficient, and future prospective studies with larger sample sizes are needed to determine the actual benefits of EGFR-TKIs combining thymosin to treat LC in OS and DFS.

The side effects of thymosin drugs must also be taken into consideration in addition to their beneficial effects. Severe allergic reactions have been linked to thymopolypeptides injection. Thymopolypeptides injection is a mixture of peptides extracted from the thymus of animals, and complex peptide components are thought to be the main contributor to allergic reactions. Even though the thymosin drugs used in the studies discussed in this article are STPs with a single composition, it is still important to be aware of their side effects to prevent serious adverse events. It is not necessary for all LC patients to use thymosin drugs because the immune system is constantly in a state of dynamic balance and attempting to improve immunity will not necessarily result in a healthier body. The most fundamental requirement for safe drug use is a strict understanding of the indications, so using thymosin drugs blindly is not desired.

Thymosin drugs are more widely used for $T\alpha 1$ and thymopentin, the conclusions of $T\alpha 1$ were largely consistent across studies, but thymopentin was not. When combined with chemotherapy or EGFR-TKI, thymopentin has been shown to increase the number of T lymphocytes and NK cells in LC patients. Thymopentin in combination with chemotherapy did not demonstrate superior effects to chemotherapy alone in terms of ORR and DCR in the meta-analysis examining the effects of sTPs in combination with chemotherapy (64). However, there is a clinical advantage for OS when thymopentin and EGFR-TKI are combined (67). Thymopentin combined with various treatment modalities

may have different therapeutic effects. These two studies are only retrospective, the level of evidence-based medical evidence is low, and the precise conclusions need to be proven by future high-level clinical trials. Additionally, there is a lack of basic and clinical trial research on the use of thymopentin in the treatment of LC. Therefore, before clinical promotion, there must be adequate evidence-based medical evidence and basic experimental support. In addition to $T\alpha 1$ and thymopentin, other thymosins are also closely related to LC. TB4 is highly expressed in NSCLC tissues, as was previously mentioned, and part of the mechanism by which TB4 promotes LC progression has been discovered (94). TB4 high expression is associated with a poor prognosis, and its ability to promote cancer has also been demonstrated in gastric cancer and colorectal cancer (95, 96). Then, for $T\beta 4$ as a target, one of the future research directions is the development of related drugs as well as the ongoing, in-depth study of the mechanism of $T\beta 4$ promoting LC.

5 Conclusion

Thymosin is known to inhibit LC growth, proliferation, invasion, and migration as well as have immunomodulatory effects by controlling the quantity, activity, and function of NK cells and T lymphocytes, according to a large number of basic experiments. The findings of these studies lend some support to the clinical application of thymosin in LC patients. However, the findings of current basic studies on the addition of combination treatment are all based on preclinical chemotherapy models. This means that the current basic research does not include the evaluation of the combined effects of T α 1 and radiotherapy, CCRT, targeted therapy, and immunotherapy, and it is necessary to improve these preclinical models to provide theoretical support for the combined application of T α 1 and real-world clinical treatment of LC. Although the thymosin drugs represented by

 $T\alpha 1$ have some results in the treatment of LC, they urgently require additional in-depth, creative basic research and higherlevel evidence-based medical evidence support, both in basic experiments and clinical efficacy.

Finally, we anticipate further large-scale and carefully-planned prospective clinical trials to provide a higher level of evidence-based medical evidence support for the use of thymosin drugs in the field of LC, as well as an increasing number of in-depth basic studies to further reveal the specific mechanism of thymosin drugs for LC.

Author contributions

YL wrote the original draft, completed the visualization, JL reviewed and revised the manuscript, YL and JL both conceived and designed the study. All authors read and approved the final manuscript. All authors contributed to the article and approved the submitted version.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

References

1. Sung H, Ferlay J, Siegel RL, Laversanne M, Soerjomataram I, Jemal A, et al. Global cancer statistics 2020: globocan estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA: Cancer J Clin* (2021) 71(3):209–49. doi: 10.3322/ caac.21660

2. Kocarnik JM, Compton K, Dean FE, Fu W, Gaw BL, Harvey JD, et al. Cancer incidence, mortality, years of life lost, years lived with disability, and disability-adjusted life years for 29 cancer groups from 2010 to 2019: A systematic analysis for the global burden of disease study 2019. *JAMA Oncol* (2022) 8(3):420–44. doi: 10.1001/jamaoncol.2021.6987

3. Siegel RL, Miller KD, Fuchs HE, Jemal A. Cancer statistics, 2022. CA: Cancer J Clin (2022) 72(1):7–33. doi: 10.3322/caac.21708

4. Becker N, Motsch E, Trotter A, Heussel CP, Dienemann H, Schnabel PA, et al. Lung cancer mortality reduction by ldct screening-results from the randomized german lusi trial. *Int J Cancer* (2020) 146(6):1503–13. doi: 10.1002/ijc.32486

5. Li N, Tan F, Chen W, Dai M, Wang F, Shen S, et al. One-off low-dose ct for lung cancer screening in China: A multicentre, population-based, prospective cohort study. *Lancet Respir Med* (2022) 10(4):378–91. doi: 10.1016/S2213-2600(21)00560-9

 Zappa C, Mousa SA. Non-small cell lung cancer: current treatment and future advances. Trans Lung Cancer Res (2016) 5(3):288–300. doi: 10.21037/tlcr.2016.06.07

7. Hattori A, Suzuki K, Takamochi K, Wakabayashi M, Aokage K, Saji H, et al. Prognostic impact of a ground-glass opacity component in clinical stage ia non-small cell lung cancer. J Thorac Cardiovasc Surg (2021) 161(4):1469-80. doi: 10.1016/ j.jtcvs.2020.01.107

8. Yotsukura M, Asamura H, Motoi N, Kashima J, Yoshida Y, Nakagawa K, et al. Long-term prognosis of patients with resected adenocarcinoma in situ and minimally invasive adenocarcinoma of the lung. J Thorac Oncol Off Publ Int Assoc Study Lung Cancer (2021) 16(8):1312–20. doi: 10.1016/j.jtho.2021.04.007

9. Duma N, Santana-Davila R, Molina JR. Non-small cell lung cancer: epidemiology, screening, diagnosis, and treatment. *Mayo Clinic Proc* (2019) 94(8):1623–40. doi: 10.1016/j.mayocp.2019.01.013

10. Wang M, Herbst RS, Boshoff C. Toward personalized treatment approaches for non-small-cell lung cancer. *Nat Med* (2021) 27(8):1345–56. doi: 10.1038/s41591-021-01450-2

11. Saw SPL, Ong BH, Chua KLM, Takano A, Tan DSW. Revisiting neoadjuvant therapy in non-small-cell lung cancer. *Lancet Oncol* (2021) 22(11):e501–e16. doi: 10.1016/s1470-2045(21)00383-1

12. Ettinger DS, Wood DE, Aisner DL, Akerley W, Bauman JR, Bharat A, et al. Nonsmall cell lung cancer, version 3.2022, nccn clinical practice guidelines in oncology. J Natl Compr Cancer Network JNCCN (2022) 20(5):497–530. doi: 10.6004/ jnccn.2022.0025

13. Goldstraw P, Chansky K, Crowley J, Rami-Porta R, Asamura H, Eberhardt WE, et al. The iaslc lung cancer staging project: proposals for revision of the tnm stage groupings in the forthcoming (Eighth) edition of the tnm classification for lung cancer.

J Thorac Oncol Off Publ Int Assoc Study Lung Cancer (2016) 11(1):39–51. doi: 10.1016/ j.jtho.2015.09.009

14. Miller JF. The discovery of thymus function and of thymus-derived lymphocytes. Immunol Rev (2002) 185:7–14. doi: 10.1034/j.1600-065x.2002.18502.x

15. Zdrojewicz Z, Pachura E, Pachura P. The thymus: A forgotten, but very important organ. Adv Clin Exp Med Off Organ Wroclaw Med Univ (2016) 25 (2):369–75. doi: 10.17219/acem/58802

16. Trainin N. Thymic hormones and the immune response. *Physiol Rev* (1974) 54 (2):272–315. doi: 10.1152/physrev.1974.54.2.272

17. Wolf E, Milazzo S, Boehm K, Zwahlen M, Horneber M. Thymic peptides for treatment of cancer patients. *Cochrane Database systematic Rev* (2011) 2011(2): Cd003993. doi: 10.1002/14651858.CD003993.pub3

18. King R, Tuthill C. Immune modulation with thymosin alpha 1 treatment. Vitamins hormones (2016) 102:151-78. doi: 10.1016/bs.vh.2016.04.003

19. Dominari A, Hathaway Iii D, Pandav K, Matos W, Biswas S, Reddy G, et al. Thymosin alpha 1: A comprehensive review of the literature. *World J Virol* (2020) 9 (5):67–78. doi: 10.5501/wjv.v9.i5.67

20. Singh VK, Biswas S, Mathur KB, Haq W, Garg SK, Agarwal SS. Thymopentin and splenopentin as immunomodulators. *Curr Status. Immunologic Res* (1998) 17 (3):345–68. doi: 10.1007/bf02786456

21. Fabrizi F, Dixit V, Martin P. Meta-analysis: the adjuvant role of thymopentin on immunological response to hepatitis B virus vaccine in end-stage renal disease. *Alimentary Pharmacol Ther* (2006) 23(11):1559–66. doi: 10.1111/j.1365-2036.2006.02923.x

22. Feng Z, Shi Q, Fan Y, Wang Q, Yin W. Ulinastatin and/or thymosin A1 for severe sepsis: A systematic review and meta-analysis. *J Trauma acute Care Surg* (2016) 80(2):335–40. doi: 10.1097/ta.000000000000909

23. Liu F, Wang HM, Wang T, Zhang YM, Zhu X. The efficacy of thymosin A1 as immunomodulatory treatment for sepsis: A systematic review of randomized controlled trials. *BMC Infect Dis* (2016) 16:488. doi: 10.1186/s12879-016-1823-5

24. Matteucci C, Grelli S, Balestrieri E, Minutolo A, Argaw-Denboba A, Macchi B, et al. Thymosin alpha 1 and hiv-1: recent advances and future perspectives. *Future Microbiol* (2017) 12:141–55. doi: 10.2217/fmb-2016-0125

25. Garaci E, Pica F, Matteucci C, Gaziano R, D'Agostini C, Miele MT, et al. Historical review on thymosin A1 in oncology: preclinical and clinical experiences. *Expert Opin Biol Ther* (2015) 15 Suppl 1:S31–9. doi: 10.1517/14712598.2015.1017466

26. Costantini C, Bellet MM, Pariano M, Renga G, Stincardini C, Goldstein AL, et al. A reappraisal of thymosin alpha1 in cancer therapy. *Front Oncol* (2019) 9:873. doi: 10.3389/fonc.2019.00873

27. Nevo N, Lee Goldstein A, Bar-David S, Natanson M, Alon G, Lahat G, et al. Thymosin alpha 1 as an adjuvant to hyperthermic intraperitoneal chemotherapy in an experimental model of peritoneal metastases from colonic carcinoma. *Int Immunopharmacol* (2022) 111:109166. doi: 10.1016/j.intimp.2022.109166

28. Moody TW, Fagarasan M, Zia F, Cesnjaj M, Goldstein AL. Thymosin alpha 1 down-regulates the growth of human non-small cell lung cancer cells in vitro and in vivo. *Cancer Res* (1993) 53(21):5214–8.

29. Moody TW, Leyton J, Zia F, Tuthill C, Badamchian M, Goldstein AL. Thymosinalpha1 is chemopreventive for lung adenoma formation in a/J mice. *Cancer Lett* (2000) 155(2):121-7. doi: 10.1016/s0304-3835(00)00405-5

30. Moody TW. Thymosin alpha1 as a chemopreventive agent in lung and breast cancer. Ann New York Acad Sci (2007) 1112:297–304. doi: 10.1196/annals.1415.040

31. Bo C, Wu Q, Zhao H, Li X, Zhou Q. Thymosin A1 suppresses migration and invasion of pd-L1 high-expressing non-small-cell lung cancer cells *via* inhibition of stat3-mmp2 signaling. *OncoTargets Ther* (2018) 11:7255–70. doi: 10.2147/ott.S177943

32. Kharazmi-Khorassani J, Asoodeh A. Thymosin alpha-1; a natural peptide inhibits cellular proliferation, cell migration, the level of reactive oxygen species and promotes the activity of antioxidant enzymes in human lung epithelial adenocarcinoma cell line (A549). *Environ Toxicol* (2019) 34(8):941–9. doi: 10.1002/tox.22765

33. Garaci E, Mastino A, Favalli C. Enhanced immune response and antitumor immunity with combinations of biological response modifiers. *Bull New York Acad Med* (1989) 65(1):111–9.

34. Garaci E, Mastino A, Pica F, Favalli C. Combination treatment using thymosin alpha 1 and interferon after cyclophosphamide is able to cure lewis lung carcinoma in mice. *Cancer immunology immunotherapy CII* (1990) 32(3):154–60. doi: 10.1007/bf01771450

35. Mastino A, Favalli C, Grelli S, Rasi G, Pica F, Goldstein AL, et al. Combination therapy with thymosin alpha 1 potentiates the anti-tumor activity of interleukin-2 with cyclophosphamide in the treatment of the lewis lung carcinoma in mice. *Int J Cancer* (1992) 50(3):493–9. doi: 10.1002/ijc.2910500327

36. Favalli C, Jezzi T, Mastino A, Rinaldi-Garaci C, Riccardi C, Garaci E. Modulation of natural killer activity by thymosin alpha 1 and interferon. *Cancer immunology immunotherapy CII* (1985) 20(3):189–92. doi: 10.1007/bf00205574

37. Yuan C, Zheng Y, Zhang B, Shao L, Liu Y, Tian T, et al. Thymosin A1 promotes the activation of myeloid-derived suppressor cells in a lewis lung cancer model by upregulating arginase 1. *Biochem Biophys Res Commun* (2015) 464(1):249–55. doi: 10.1016/j.bbrc.2015.06.132

38. Wang F, Li B, Fu P, Li Q, Zheng H, Lao X. Immunomodulatory and enhanced antitumor activity of a modified thymosin A1 in melanoma and lung cancer. *Int J Pharmaceutics* (2018) 547(1):611–20. doi: 10.1016/j.ijpharm.2018.06.041

39. Peng R, Xu C, Zheng H, Lao X. Modified thymosin alpha 1 distributes and inhibits the growth of lung cancer in vivo. ACS omega (2020) 5(18):10374-81. doi: 10.1021/acsomega.0c00220

40. Huang D, Wang S, Wang A, Chen X, Zhang H. Thymosin beta 4 silencing suppresses proliferation and invasion of non-small cell lung cancer cells by repressing notch1 activation. *Acta Biochim Biophys Sin* (2016) 48(9):788–94. doi: 10.1093/abbs/gmw070

41. Sporn MB, Suh N. Chemoprevention: an essential approach to controlling cancer. Nat Rev Cancer (2002) 2(7):537-43. doi: 10.1038/nrc844

42. Ashraf-Uz-Zaman M, Bhalerao A, Mikelis CM, Cucullo L, German NA. Assessing the current state of lung cancer chemoprevention: A comprehensive overview. *Cancers* (2020) 12(5):1265–1292. doi: 10.3390/cancers12051265

43. Hu X, Li J, Fu M, Zhao X, Wang W. The jak/stat signaling pathway: from bench to clinic. *Signal transduction targeted Ther* (2021) 6(1):402. doi: 10.1038/s41392-021-00791-1

44. Shen M, Xu Z, Xu W, Jiang K, Zhang F, Ding Q, et al. Inhibition of atm reverses emt and decreases metastatic potential of cisplatin-resistant lung cancer cells through jak/stat3/pd-L1 pathway. *J Exp Clin Cancer Res CR* (2019) 38(1):149. doi: 10.1186/s13046-019-1161-8

45. Wu DM, Deng SH, Liu T, Han R, Zhang T, Xu Y. Tgf-B-mediated exosomal lncmmp2-2 regulates migration and invasion of lung cancer cells to the vasculature by promoting mmp2 expression. *Cancer Med* (2018) 7(10):5118–29. doi: 10.1002/ cam4.1758

46. Cheung EC, Vousden KH. The role of ros in tumour development and progression. *Nat Rev Cancer* (2022) 22(5):280–97. doi: 10.1038/s41568-021-00435-0

47. Valavanidis A, Vlachogianni T, Fiotakis K, Loridas S. Pulmonary oxidative stress, inflammation and cancer: respirable particulate matter, fibrous dusts and ozone as major causes of lung carcinogenesis through reactive oxygen species mechanisms. *Int J Environ Res Public Health* (2013) 10(9):3886–907. doi: 10.3390/ijerph10093886

48. Crockford D, Turjman N, Allan C, Angel J. Thymosin beta4: structure, function, and biological properties supporting current and future clinical applications. *Ann New York Acad Sci* (2010) 1194:179–89. doi: 10.1111/j.1749-6632.2010.05492.x

49. Sosne G, Qiu P, Goldstein AL, Wheater M. Biological activities of thymosin beta4 defined by active sites in short peptide sequences. FASEB J Off Publ Fed Am Societies Exp Biol (2010) 24(7):2144–51. doi: 10.1096/fj.09-142307

50. Maskalenko NA, Zhigarev D, Campbell KS. Harnessing natural killer cells for cancer immunotherapy: dispatching the first responders. *Nat Rev Drug Discovery* (2022) 21(8):559–77. doi: 10.1038/s41573-022-00413-7

51. Dong C. Cytokine regulation and function in T cells. Annu Rev Immunol (2021) 39:51–76. doi: 10.1146/annurev-immunol-061020-053702

52. Durgeau A, Virk Y, Corgnac S, Mami-Chouaib F. Recent advances in targeting cd8 T-cell immunity for more effective cancer immunotherapy. *Front Immunol* (2018) 9:14. doi: 10.3389/fimmu.2018.00014

53. Doroshow DB, Sanmamed MF, Hastings K, Politi K, Rimm DL, Chen L, et al. Immunotherapy in non-small cell lung cancer: facts and hopes. *Clin Cancer Res an Off J Am Assoc Cancer Res* (2019) 25(15):4592–602. doi: 10.1158/1078-0432.Ccr-18-1538

54. Sani S, Messe M, Fuchs Q, Pierrevelcin M, Laquerriere P, Entz-Werle N, et al. Biological relevance of rgd-integrin subtype-specific ligands in cancer. *Chembiochem Eur J Chem Biol* (2021) 22(7):1151–60. doi: 10.1002/cbic.202000626

55. Fu S, Xu X, Ma Y, Zhang S, Zhang S. Rgd peptide-based non-viral gene delivery vectors targeting integrin A(V)B(3) for cancer therapy. *J Drug Targeting* (2019) 27 (1):1–11. doi: 10.1080/1061186x.2018.1455841

56. Kang S, Lee S, Park S. Irgd peptide as a tumor-penetrating enhancer for tumortargeted drug delivery. *Polymers* (2020) 12(9):1906–1931. doi: 10.3390/polym12091906

57. Zhang G, Chen L, Khan AA, Li B, Gu B, Lin F, et al. Mirna-124-3p/neuropilin-1 (Nrp-1) axis plays an important role in mediating glioblastoma growth and angiogenesis. *Int J Cancer* (2018) 143(3):635–44. doi: 10.1002/ijc.31329

58. Hu C, Jiang X. Role of nrp-1 in vegf-vegfr2-independent tumorigenesis. *Targeted Oncol* (2016) 11(4):501–5. doi: 10.1007/s11523-016-0422-0

59. Bozza S, Gaziano R, Bonifazi P, Zelante T, Pitzurra L, Montagnoli C, et al. Thymosin alpha1 activates the tlr9/myd88/irf7-dependent murine cytomegalovirus sensing for induction of anti-viral responses in vivo. *Int Immunol* (2007) 19(11):1261–70. doi: 10.1093/intimm/dxm097

60. ROmani L, Bistoni F, Gaziano R, Bozza S, Montagnoli C, Perruccio K, et al. Thymosin alpha 1 activates dendritic cells for antifungal th1 resistance through toll-like receptor signaling. *Blood* (2004) 103(11):4232–9. doi: 10.1182/blood-2003-11-4036

61. Cheepsattayakorn A, Cheepsattayakorn R. Lung cancer chemotherapy, new treatment and related patents. *Recent patents anti-cancer Drug Discovery* (2014) 9 (3):372–81. doi: 10.2174/1574892809666140520120806

62. Salvati F, Rasi G, Portalone L, Antilli A, Garaci E. Combined treatment with thymosin-alpha1 and low-dose interferon-alpha after ifosfamide in non-small cell lung cancer: A phase-ii controlled trial. *Anticancer Res* (1996) 16(2):1001–4.

63. Jiang J, Wang X, Tian J, Li L, Lin Q. Thymosin plus cisplatin with vinorelbine or gemcitabine for non-small cell lung cancer: A systematic review and meta-analysis of randomized controlled trials. *Thorac Cancer* (2011) 2(4):213–20. doi: 10.1111/j.1759-7714.2011.00057.x

64. Zeng FL, Xiao Z, Wang CQ, Jiang Y, Shan JL, Hu SS, et al. Clinical efficacy and safety of synthetic thymic peptides with chemotherapy for non-small cell lung cancer in China: A systematic review and meta-analysis of 27 randomized controlled trials following the prisma guidelines. *Int Immunopharmacol* (2019) 75:105747. doi: 10.1016/j.intimp.2019.105747

65. Schulof RS, Lloyd MJ, Cleary PA, Palaszynski SR, Mai DA, Cox JW Jr., et al. A randomized trial to evaluate the immunorestorative properties of synthetic thymosinalpha 1 in patients with lung cancer. *J Biol response modifiers* (1985) 4(2):147–58.

66. Liu F, Qiu B, Xi Y, Luo Y, Luo Q, Wu Y, et al. Efficacy of thymosin A1 in management of radiation pneumonitis in patients with locally advanced non-small cell lung cancer treated with concurrent chemoradiotherapy: A phase 2 clinical trial (Gasto-1043). *Int J Radiat oncology biology Phys* (2022) 114(3):433–43. doi: 10.1016/j.ijrobp.2022.07.009

67. Feng Y, Zhu G, Lang S, Hao P, Li G, Chen F, et al. The efficacy and safety of epidermal growth factor receptor tyrosine kinase inhibitor combined with thymosin in advanced non-small cell lung cancer patients harboring active epidermal growth factor receptor mutations. *Front Oncol* (2021) 11:659065. doi: 10.3389/fonc.2021.659065

68. Guo CL, Mei JD, Jia YL, Gan FY, Tang YD, Liu CW, et al. Impact of thymosin A1 as an immunomodulatory therapy on long-term survival of non-small cell lung cancer patients after R0 resection: A propensity score-matched analysis. *Chin Med J* (2021) 134(22):756. doi: 10.1097/cm9.00000000001819

69. Yang S, Zhang Z, Wang Q. Emerging therapies for small cell lung cancer. J Hematol Oncol (2019) 12(1):47. doi: 10.1186/s13045-019-0736-3

70. Griesinger F, Korol EE, Kayaniyil S, Varol N, Ebner T, Goring SM. Efficacy and safety of first-line carboplatin-versus cisplatin-based chemotherapy for non-small cell lung cancer: A meta-analysis. *Lung Cancer (Amsterdam Netherlands)* (2019) 135:196–204. doi: 10.1016/j.lungcan.2019.07.010

71. Yang WC, Hsu FM, Yang PC. Precision radiotherapy for non-small cell lung cancer. J Biomed Sci (2020) 27(1):82. doi: 10.1186/s12929-020-00676-5

72. Antonia SJ, Villegas A, Daniel D, Vicente D, Murakami S, Hui R, et al. Overall survival with durvalumab after chemoradiotherapy in stage iii nsclc. *New Engl J Med* (2018) 379(24):2342–50. doi: 10.1056/NEJMoa1809697

73. Dang J, Li G, Ma L, Diao R, Zang S, Han C, et al. Predictors of grade \geq 2 and grade \geq 3 radiation pneumonitis in patients with locally advanced non-small cell lung cancer treated with three-dimensional conformal radiotherapy. *Acta Oncol (Stockholm Sweden)* (2013) 52(6):1175–80. doi: 10.3109/0284186x.2012.747696

74. Vinod SK, Hau E. Radiotherapy treatment for lung cancer: current status and future directions. *Respirology (Carlton Vic)* (2020) 25 Suppl 2:61–71. doi: 10.1111/ resp.13870

75. Leonetti A, Sharma S, Minari R, Perego P, Giovannetti E, Tiseo M. Resistance mechanisms to osimertinib in egfr-mutated non-small cell lung cancer. *Br J Cancer* (2019) 121(9):725–37. doi: 10.1038/s41416-019-0573-8

76. Suresh K, Naidoo J, Lin CT, Danoff S. Immune checkpoint immunotherapy for non-small cell lung cancer: benefits and pulmonary toxicities. *Chest* (2018) 154 (6):1416–23. doi: 10.1016/j.chest.2018.08.1048

77. Aguilar EJ, Ricciuti B, Gainor JF, Kehl KL, Kravets S, Dahlberg S, et al. Outcomes to first-line pembrolizumab in patients with non-small-cell lung cancer and very high pd-L1 expression. *Ann Oncol Off J Eur Soc Med Oncol* (2019) 30(10):1653–9. doi: 10.1093/annonc/mdz288

78. Herbst RS, Giaccone G, de Marinis F, Reinmuth N, Vergnenegre A, Barrios CH, et al. Atezolizumab for first-line treatment of pd-L1-selected patients with nsclc. *New Engl J Med* (2020) 383(14):1328–39. doi: 10.1056/NEJMoa1917346

79. Reck M, Rodriguez-Abreu D, Robinson AG, Hui R, Csőszi T, Fülöp A, et al. Updated analysis of keynote-024: pembrolizumab versus platinum-based chemotherapy for advanced non-small-cell lung cancer with pd-L1 tumor proportion score of 50% or greater. J Clin Oncol Off J Am Soc Clin Oncol (2019) 37 (7):537–46. doi: 10.1200/jco.18.00149

80. Danielli R, Cisternino F, Giannarelli D, Calabrò L, Camerini R, Savelli V, et al. Long-term follow up of metastatic melanoma patients treated with thymosin alpha-1: investigating immune checkpoints synergy. *Expert Opin Biol Ther* (2018) 18(sup1):77– 83. doi: 10.1080/14712598.2018.1494717

81. Gerard CL, Delyon J, Wicky A, Homicsko K, Cuendet MA, Michielin O. Turning tumors from cold to inflamed to improve immunotherapy response. *Cancer Treat Rev* (2021) 101:102227. doi: 10.1016/j.ctrv.2021.102227

82. Galon J, Bruni D. Approaches to treat immune hot, altered and cold tumours with combination immunotherapies. *Nat Rev Drug Discovery* (2019) 18(3):197–218. doi: 10.1038/s41573-018-0007-y

83. Farhood B, Najafi M, Mortezaee K. Cd8(+) cytotoxic T lymphocytes in cancer immunotherapy: A review. J Cell Physiol (2019) 234(6):8509–21. doi: 10.1002/jcp.27782

84. Terrén I, Orrantia A, Vitallé J, Zenarruzabeitia O, Borrego F. Nk cell metabolism and tumor microenvironment. *Front Immunol* (2019) 10:2278. doi: 10.3389/fimmu.2019.02278

85. Thompson JA, Schneider BJ, Brahmer J, Achufusi A, Armand P, Berkenstock MK, et al. Management of immunotherapy-related toxicities, version 1.2022, nccn clinical practice guidelines in oncology. *J Natl Compr Cancer Network JNCCN* (2022) 20(4):387–405. doi: 10.6004/jnccn.2022.0020

86. ROmani L, Oikonomou V, Moretti S, Iannitti RG, D'Adamo MC, Villella VR, et al. Thymosin A1 represents a potential potent single-molecule-based therapy for cystic fibrosis. *Nat Med* (2017) 23(5):590–600. doi: 10.1038/nm.4305

87. Renga G, Bellet MM, Pariano M, Gargaro M, Stincardini C, D'Onofrio F, et al. Thymosin A1 protects from ctla-4 intestinal immunopathology. *Life Sci alliance* (2020) 3(10):202000662. doi: 10.26508/lsa.202000662

88. Wu SY, Fu T, Jiang YZ, Shao ZM. Natural killer cells in cancer biology and therapy. *Mol Cancer* (2020) 19(1):120. doi: 10.1186/s12943-020-01238-x

89. Xie G, Dong H, Liang Y, Ham JD, Rizwan R, Chen J. Car-nk cells: A promising cellular immunotherapy for cancer. *EBioMedicine* (2020) 59:102975. doi: 10.1016/j.ebiom.2020.102975

90. Rogers LJ, Bleetman D, Messenger DE, Joshi NA, Wood L, Rasburn NJ, et al. The impact of enhanced recovery after surgery (Eras) protocol compliance on morbidity from resection for primary lung cancer. *J Thorac Cardiovasc Surg* (2018) 155(4):1843–52. doi: 10.1016/j.jtcvs.2017.10.151

91. Fearon KC, Ljungqvist O, Von Meyenfeldt M, Revhaug A, Dejong CH, Lassen K, et al. Enhanced recovery after surgery: A consensus review of clinical care for patients undergoing colonic resection. *Clin Nutr (Edinburgh Scotland)* (2005) 24(3):466–77. doi: 10.1016/j.clnu.2005.02.002

92. Batchelor TJP, Rasburn NJ, Abdelnour-Berchtold E, Brunelli A, Cerfolio RJ, Gonzalez M, et al. Guidelines for Enhanced Recovery after Lung Surgery: Recommendations of the Enhanced Recovery after Surgery (Eras®) Society and the European Society of Thoracic Surgeons (Ests). Eur J cardio-thoracic Surg Off J Eur Assoc Cardio-thoracic Surg (2019) 55(1):91–115. doi: 10.1093/ejcts/ezy301

93. Muka T, Glisic M, Milic J, Verhoog S, Bohlius J, Bramer W, et al. A 24-step guide on how to design, conduct, and successfully publish a systematic review and metaanalysis in medical research. *Eur J Epidemiol* (2020) 35(1):49–60. doi: 10.1007/s10654-019-00576-5

94. Xiao Y, Chen Y, Wen J, Yan W, Zhou K, Cai W. Thymosin B4: A potential molecular target for tumor therapy. *Crit Rev eukaryotic Gene Expression* (2012) 22 (2):109–16. doi: 10.1615/critreveukargeneexpr.v22.i2.30

95. Ryu YK, Lee YS, Lee GH, Song KS, Kim YS, Moon EY. Regulation of glycogen synthase kinase-3 by thymosin beta-4 is associated with gastric cancer cell migration. *Int J Cancer* (2012) 131(9):2067–77. doi: 10.1002/ijc.27490

96. Piao Z, Hong CS, Jung MR, Choi C, Park YK. Thymosin B4 induces invasion and migration of human colorectal cancer cells through the ilk/akt/B-catenin signaling pathway. *Biochem Biophys Res Commun* (2014) 452(3):858–64. doi: 10.1016/j.bbrc.2014.09.012