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RECEIVED 02 October 2023 ACCEPTED 16 November 2023 PUBLISHED 12 December 2023

CITATION

Najjary S, de Koning W, Kros JM and Mustafa DAM (2023) Unlocking molecular mechanisms and identifying druggable targets in matched-paired brain metastasis of breast and lung cancers. *Front. Immunol.* 14:1305644. doi: 10.3389/fimmu.2023.1305644

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Unlocking molecular mechanisms and identifying druggable targets in matchedpaired brain metastasis of breast and lung cancers

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Introduction: The incidence of brain metastases in cancer patients is increasing, with lung and breast cancer being the most common sources. Despite advancements in targeted therapies, the prognosis remains poor, highlighting the importance to investigate the underlying mechanisms in brain metastases. The aim of this study was to investigate the differences in the molecular mechanisms involved in brain metastasis of breast and lung cancers. In addition, we aimed to identify cancer lineage-specific druggable targets in the brain metastasis.

Methods: To that aim, a cohort of 44 FFPE tissue samples, including 22 breast cancer and 22 lung adenocarcinoma (LUAD) and their matched-paired brain metastases were collected. Targeted gene expression profiles of primary tumors were compared to their matched-paired brain metastases samples using nCounter PanCancer IO 360[™] Panel of NanoString technologies. Pathway analysis was performed using gene set analysis (GSA) and gene set enrichment analysis (GSEA). The validation was performed by using Immunohistochemistry (IHC) to confirm the expression of immune checkpoint inhibitors.

Results: Our results revealed the significant upregulation of cancer-related genes in primary tumors compared to their matched-paired brain metastases (adj. $p \le 0.05$). We found that upregulated differentially expressed genes in breast cancer brain metastasis (BM-BC) and brain metastasis from lung adenocarcinoma (BM-LUAD) were associated with the metabolic stress pathway, particularly related to the glycolysis. Additionally, we found that the upregulated genes in BM-BC and BM-LUAD played roles in immune response regulation, tumor growth, and proliferation. Importantly, we identified high expression of the immune checkpoint VTCN1 in BM-BC, and VISTA, IDO1, NT5E, and HDAC3 in BM-LUAD. Validation using immunohistochemistry further supported these findings.

Conclusion: In conclusion, the findings highlight the significance of using matched-paired samples to identify cancer lineage-specific therapies that may improve brain metastasis patients outcomes.

KEYWORDS

molecular mechanisms, brain metastasis, breast cancer, lung adenocarcinoma, gene expression

Introduction

The incidence of brain metastasis (BM) is rising. Improved therapeutic methods to control the primary tumors have led to longer survivals, but have also increased the incidence of late metastases to brain (1, 2). Despite the development of novel targeted therapies, BM remains a devastating complication of cancer. BM patients exhibit poor prognosis due to no effective long-term therapy (3, 4). BM from lung and breast cancer exhibits distinct patterns in the disease course. In lung cancer, BM typically develops within a few months after diagnosis. On the other hand, in breast cancer, metastasis often occurs in one organ before it spreads to other organs, with brain metastasis typically being a late event in the course of disease (5). The median overall survival of the patients diagnosed with BM is approximately 4-15 months in lung cancer and 6-18 months in breast cancer following treatment (2, 6). Due to the limited effectiveness of current therapies and the poor prognosis associated with BM (7-9), it is crucial to identify related genes and molecular mechanisms involved. Understanding these mechanisms can aid in the identification of new therapeutic targets and potentially improve patient survival. The signaling pathways and gene expression profiles in metastatic cancer cells play a significant role in in the formation of organ-specific metastasis. Specific gene expression patterns may dictate tendency for specific targeting specific sites, such as the brain. The unique physiological barriers in the brain, including the blood-brain-barrier (BBB), is hypothesized to limit the ability of metastatic tumor cells to infiltrate the brain (10). Site-specific metastasis may also reflect tissue-specific migration and proliferation signatures that contribute to the differential ability of tumor cells to metastasize and proliferate in specific organs (11, 12). Compared to other sites of cancer metastasis, the normal brain tissue has relatively low collagen content, lower oxygen tension, and high glucose metabolism, which may affect the survival and proliferation of disseminated tumor cells in the brain (13, 14). So far, the molecular signatures and expression patterns involved in brain metastasis remain largely unknown. Employing matched-paired samples from both primary tumors and brain metastases constitutes a distinctive and precise research approach, enabling a thorough investigation of the association between gene expression signatures and metastatic development. Despite its potential, few studies have utilized this method, underscoring the uniqueness of the presented research. Primary lung cancer and their matched-paired were used in previous studies to generate gene expression signature (15).

Similarly, 16 primary breast cancers and their matched-paired brain metastases were used to generate gene expression profiles in order to assess clinic-pathological markers, immune-related gene signatures, and identifying therapeutic targets for brain metastases (16). In addition, 21 breast cancer and their matched-paired samples were used previously to perform TrueSeq RNA-sequencing and determine clinically actionable BM target genes (17).

The aim of the present study was to investigate the mechanisms operative in the primary tumors and their matched-paired brain metastasis. The utilization of matched-paired samples provides a unique opportunity to investigate shared molecular characteristics and distinctions, advancing our understanding and potential treatments for malignancies. We compared gene expression profiles of primary tumors with their matched-paired BM. Subsequently, we thoroughly examined the upregulated genes in BM of both tumor types to identify shared and distinctive predominant molecular features in BM. We validated the identified druggable targets by using IHC in the matched-paired samples. The collective findings from our research have the potential to enhance the treatment strategies for patients with BM.

Materials and methods

Tissue samples selection and clinicopathological data

A set of 44 Formalin-fixed, paraffin-embedded (FFPE) tissue samples of primary tumors and their matched-paired brain metastases was used, including 11 pairs of lung adenocarcinoma and 11 pairs of breast cancer (Figure 1). The clinicopathological characteristics of patients are summarized in Table 1. None of the patients in either group received therapy or treatment with corticosteroids within the 6-12 months prior to brain metastasis surgery. Treatment options following surgical removal of the brain metastasis included radiotherapy, chemotherapy, stereotactic radiotherapy (SRT), and whole brain radiotherapy (WBRT). More than half of the patients with LUAD were males and the majority of patients had a smoking history. The median age at diagnosis of the brain metastases was 48 years for patients with breast cancer and 64 years for patients with LUAD. This study was approved by the Medical Ethics Committee of the Erasmus Medical Center, Rotterdam, the Netherlands, and was carried out in adherence to



the Code of Conduct of the Federation of Medical Scientific Societies in the Netherlands (MEC 02.953 & MEC-2020–0732).

RNA isolation and quality control

RNA isolation and QC were performed following our previously described protocol (18). In brief, 5 μ m H&E tissue sections from each sample were examined by a pathologist. Total RNA was isolated from 10-12 sections of 10 μ m using the RNeasy FFPE kit (Qiagen, Hilden, Germany) according to the supplier's instructions. The extracted RNA was stored in RNase/DNase-free water at -80°C. The quality and quantity of the RNA was determined using the Agilent 2100 Bioanalyzer (Santa Clara, CA, USA). RNA degradation was assessed by calculating the percentage of fragments within the range of 300-4000 nucleotides.

Targeted gene expression profiling using Nanostring[®] nCounter assay

To measure targeted gene expression profiles, the nCounter PanCancer IO360TM Panel consisting of 750 cancer-related genes and 20 housekeeping genes, was used (Nanostring Technologies, Seattle, WA, United States) (19). Briefly, 300 ng of RNA, up to a maximum of 7 µL, was hybridized with the panel probes for 17 hours at 65°C using a SimpliAmp Thermal Cycler (Applied Biosystems, Foster City, CA, USA). The removal of unannealed probes was performed through the nCounter FLEX system, following the manufacture's protocol. Gene counts were determined by scanning 490 fields-of-view (FOV). The obtained expression data were imported into the nSolver software (version 4.0), and subsequent analysis was carried out using the Advanced Analysis module (version 2.0). To normalize the raw expression data, the geNorm algorithm was applied, utilizing the 16 most stable housekeeping genes (Table S1). Genes were included for further analysis if their expression was higher than the detection limit, which was calculated as the mean of negative controls plus 2 standard deviations.

Statistical analysis

A simplified negative binomial model, log-linear models, or a mixture of negative binomial models that are embedded within the advanced analysis module, were used to identify differentially expressed genes (DEGs) between pairs of breast cancer and LUAD and their matched-paired brain metastases. To correct for multiple testing, Benjamin-Hochberg method was used and genes with a BH- P-value ≤ 0.05 were considered as DEGs. All statistical analyses were performed using R version 4.0.1. The P-values were two-sided, and statistical significance was defined as Pvalues \leq 0.05. Heatmaps were created using Log2-normalized data of significantly expressed genes (adj. $p \le 0.05$). Heatmaps of the DEGs were created using the following criteria: Log2Fold change > 1.5, adj. $p \le 0.05$. Outliers were eliminated using Tukey's rule (20). To create and visualize heatmaps, the web-based tool Morpheus by the Broad Institute (RRID: SCR_017386) was used. The scaling was performed for each and ranged from 0 to 1.

Pathways analysis

To assess the differences at the pathway level, two different methods were employed. The first was gene set analysis (GSA) was carried out using the embedded methodology within nSolver software. Pathway scores were calculated using the average expression of genes associated with each pathway. Significant changes in pathway scores between the two groups were determined using the Wilcoxon test, and a *p*-value < 0.05 was considered significant. The second method was by using Gene Set Enrichment Analysis (GSEA) that was performed using the online bioinformatics tool and database, Metascape.org. The NanoString panel was used as the background. DEGs (adj. $p \le 0.05$) in BM-BC and BM-LUAD were separately uploaded to the pathway analysis tool, which identified enriched biological pathways and conducted functional enrichment analysis. The obtained results were thoroughly examined and interpreted.

TABLE 1 Clinical characteristics of patients.

| Characteristics | No. | % | |
|--|---------------|----------|--|
| Total samples | 44 | 100 | |
| Cancer type | | | |
| Primary breast cancer | 11 | 25 | |
| BM-BC | 11 | 25 | |
| Primary lung adenocarcinoma | 11 | 25 | |
| BM-LUAD | 11 | 25 | |
| Lung adenocarcinoma | | | |
| Median age at diagnosis with primary tumor, years (range) | 57 (37-69) | | |
| Median age at diagnosis with brain metastasis tumor, years (range) | 64 (46-74) | | |
| Sex | | | |
| Male | 6 | 54.5 | |
| Female | 5 | 45.5 | |
| Histology of primary tumor and their matched-paired brain metastases | | | |
| Adenocarcinoma | 22 | 100 | |
| Smoking status | | | |
| Never smoking | 1 | 9.1 | |
| Former smoker | 5 | 45.5 | |
| Smoking | 4 | 36.4 | |
| Unknown | 1 | 9.1 | |
| Treatment for primary tumor | | 1 | |
| Surgery | 3 | 27.3 | |
| Surgery & Chemotherapy | 1 | 9.1 | |
| surgery & Radiotherapy | 2 | 18.2 | |
| Surgery & Chemotherapy & Radiotherapy | 2 | 18.2 | |
| surgery & Radiochemotherapy | 3 | 27.3 | |
| Treatment after surgery of BM | | 1 | |
| No treatment | 1 | 9.1 | |
| Radiotherapy | 6 | 54.4 | |
| SRT | 1 | 9.1 | |
| WBRT | 2 | 18.2 | |
| Other | 1 | 9.1 | |
| Breast cancer | | | |
| Median age at diagnosis with primary tumor, years (range) | 47 (40-74) | | |
| Median age at diagnosis with brain metastasis tumor, years (range) | 48 (44-74) | | |
| Sex | | | |
| Female | 11 | 100 | |
| | (Co | ntinued) | |

TABLE 1 Continued

| Characteristics | No. | % | |
|--|-----|------|--|
| Histology of primary tumor | | | |
| ER/PR+ | 2 | 18.2 | |
| ER/HER2+ | 1 | 9.1 | |
| HER2+ | 1 | 9.1 | |
| TNBC | 5 | 45.5 | |
| Unknown | 2 | 18.2 | |
| Histology of matched-paired brain metastases | | | |
| ER+ | 3 | 16.7 | |
| PR+ | 2 | 11.1 | |
| ER/PR+ | 1 | 5.6 | |
| HER2+ | 2 | 11.1 | |
| TNBC | 2 | 11.1 | |
| Unknown | 1 | 5.6 | |
| Treatment for primary tumor | | | |
| Surgery | 2 | 18.2 | |
| Surgery & Chemotherapy | 3 | 27.3 | |
| Surgery & Chemotherapy & Radiotherapy | 6 | 54.5 | |
| Treatment after surgery of BM | | | |
| Radiotherapy | 5 | 45.5 | |
| Chemotherapy | 2 | 18.2 | |
| Radiotherapy & Chemotherapy | 2 | 18.2 | |
| Other | 2 | 18.2 | |

ER, estrogen receptor; PR, progesterone receptor; HER2, human epidermal growth factor 2; TNBC, triple-negative breast cancer; SRT, Stereotactic radiotherapy; WBRT, Whole brain radiotherapy.

Immunohistochemistry

Immunohistochemistry (IHC) using the Alkaline phosphatase was performed to investigate the expression of immune checkpoint inhibitors IDO1, VISTA, NT5E, VTCN1 and drug target HDAC3 in all the available samples. Tissue sections of 4µm from each sample were prepared. Primary antibodies were applied and incubated overnight at 4°C, followed by the application of alkaline phosphatase-conjugated secondary antibodies for detection. The details of the antibodies that were used are summarized in (Table S2). The stained slides were evaluated and interpreted by a pathologist.

Results

Expression of cancer-related genes is higher in the primary tumors than in the matched-paired BM for both LUAD and BC

Comparing the primary breast cancer samples to their matched-paired brain metastases showed that 281 genes were

upregulated in primary breast cancers (adj. $p \le 0.05$; Table S3) and 43 were upregulated in BM-BC (adj. $p \le 0.05$; Table S4) (Figures 2A, C). As to the lung adenocarcinomas 226 genes were upregulated in the primary tumors (adj. $p \le 0.05$; Table S5) while 20 genes were upregulated in BM-LUAD (adj. $p \le 0.05$; Table S6) (Figures 2B, D). Pathway analysis revealed that the upregulated genes in the primary tumors were associated mainly with cell growth and proliferation (Figures 2E, F).

Different tumor types upregulate distinct genes in their brain metastasis

In order to investigate whether different types of tumors upregulated the same genes after developing brain metastasis, the upregulated genes in BM of both tumor types were compared (Figure 3A). The DEGs in BM-BC (n=43) and BM-LUAD (n=20) revealed 12 shared upregulated genes (adj. $p \le 0.05$) (Figure 3B). Of the other genes 31 were specifically upregulated in BM-BC (adj. $p \le 0.05$) and 8 genes in BM-LUAD (adj. $p \le 0.05$) (Figures 3D, C). Pathway analysis revealed that 6 genes of the commonly upregulated DEGs in BM-BC and BM-LUAD were associated with the metabolic stress pathway (Figures 3D, E).

Common upregulation of the metabolic stress pathway in brain metastases of BC and LUAD

To further investigate the involvement of the upregulated DEGs in brain metastasis, 43 DEGs in BM-BC and 20 DEGs in BM-LUAD were uploaded separately to the GSA and GSEA tools. The results showed that the metabolic stress pathway was enriched in BM with 13 genes in BM-BC and 7 in BM-LUAD. A total of 9 genes of the metabolic stress pathway were found to be involved in the glycolysis pathway (Figure 4A). Interestingly, the expression level of all DEGs associated with glycolysis was found to be higher in BM-BC compared to that in BM-LUAD (Figure 4B).

Common upregulation of immune response modulation and tumor proliferation in brain metastasis of BC and LUAD

Further exploration of the upregulated DEGs in the brain metastases revealed their involvement in modulating the immune response. Thirty DEGs that were upregulated in BM-BC (excluding genes associated with metabolic stress) and 13 DEGs that were upregulated in BM-LUAD (excluding genes associated with metabolic stress) were included for GSA. A total of 11 genes were found to have function related to regulating (stimulation or inhibition) immune system in brain metastasis (Figures 5A, B). Importantly, an FDA-approved druggable target, HDAC3, was found to be upregulated in BM-LUAD as compared to primary LUAD (Figure 5B). These results were validated by IHC

(Figure 5C). In addition, 8 genes were associated with tumor proliferation (Figure 6A). Remarkably, different types of tumors used distinct genes to regulate their proliferation (Figures 6A, B).

Various potential druggable targets were expressed in the brain metastasis of different cancer types

A total of 12 immune checkpoint molecules were identified to be upregulated in primary tumors compared to their matchedpaired brain metastases. However, only four immune checkpoint molecules exhibited significant differences of expression between the brain metastases from BC and LUAD (Figure 7A). The VTCN1 (B7-H4) was upregulated in BM-BC, whereas NT5E (CD73), VSIR and IDO1 were upregulated in BM-LUAD (Figure 7A). The expressional differences of these four immune checkpoints were validated at the protein level. Immunohistochemistry was performed in 2/11 breast cancer and 5/11 LUAD paired samples. Due to the amount of samples that was used for RNA extraction, the tumor compartment was lost in 9 breast cancer and 6 LUAD paired samples. Consistent with the gene expression results, higher expression of the immune checkpoint protein VTCN1 was observed in BM-BC compared to primary tumors and BM-LUAD (Figure 7B). Additionally, higher expression of the immune checkpoint proteins NT5E (CD73), VISTA (VSIR) and IDO1 was observed in primary tumors, with a predominant higher expression in BM-LUAD compared to BM-BC (Figure 7B).

Discussion

Studying brain metastasis is important for the development of effective treatment options and ultimately improving patient outcomes (21). In the present study, we compared the primary tumors of breast and LUAD to their matched-paired brain metastases. We identified that cancer-related genes were upregulated in the primary tumors compared to their matchedpaired brain metastases. In the brain metastases we found common upregulated genes suggestive of the presence of shared molecular mechanisms. In addition, we identified distinct differences of gene expression in BM-BC and BM-LUAD, signifying the utilization of unique mechanisms by different tumor types in brain metastasis. This finding highlights the heterogeneity that exists among brain metastases, revealing a complex interplay between common and unique molecular pathways in this context. Moreover, we found that the upregulated differentially expressed genes in BM-BC and BM-LUAD were associated with the metabolic stress pathway in brain metastasis, with a particular involvement of the glycolysis pathway. Interestingly, the majority of these genes exhibited higher upregulation in BM-BC compared to BM-LUAD, indicating higher glycolysis and glucose uptake in BM-BC. Limited studies have also provided evidence supporting the role of metabolic pathway in cancer metastasis. Dupuy et al. (22) showed that breast cancer cells with high metastatic potential undergo metabolic alterations leading to increased glycolysis and decreased mitochondrial



LUAD (adj. $p \le 0.05$). (C) Heatmap of normalized differentially expressed genes between primary breast and BM-BC (absolute fold change ≥ 1.5 ; adj. $p \le 0.05$). (D) Heatmap of normalized differentially expressed genes between primary LUAD and BM-LUAD (absolute fold change ≥ 1.5 ; adj. $p \le 0.05$). The heatmap is scaled based on each gene. (D) Heatmap of normalized differentially expressed genes between primary LUAD and BM-LUAD (absolute fold change ≥ 1.5 ; adj. $p \le 0.05$). The heatmap is scaled based on each gene. (E) GSEA of upregulated differentially expressed genes in primary LUAD (adj. $p \le 0.05$). (F) GSEA of upregulated differentially expressed genes in primary LUAD (adj. $p \le 0.05$).

metabolism. Parida et al. (23) showed that variations in metabolic pathways among brain-tropic cells influence their ability to adapt to the brain microenvironment and support tumor growth, highlighting the importance of understanding metabolic factors in brain metastasis. Tchou et al. (24) demonstrated that triple negative breast cancers (TNBCs) with high glycolytic rates exhibit an elevated proliferation index, indicating the association between glycolysis and tumor cell proliferation in this breast cancer subtype. These findings demonstrate the potential significance of metabolic pathway in promoting the survival and proliferation of metastatic cells within the brain microenvironment. However, it is imperative to validate these observations through functional



studies, including experiments on glycolysis-related genes' role in tumor metastasis, such as transwell or wound healing assays, in future research. Given our discovery of elevated glycolysis in BM-BC and the established literature on the impact of metabolic pathways on cancer progression, we hypnotize that the increased glycolysis in BM-BC may be influenced by dietary factors, particularly carbohydrate intake in daily food consumption. A comprehensive investigation into dietary habits and sugar intake patterns could offer valuable insights into the regulation of glycolysis in the context of BM-BC. Furthermore, we found that the upregulated genes in BM-BC and BM-LUAD were involved in immune response regulation, tumor growth, and proliferation. Limited studies have utilized matched-paired samples to investigate immune related signatures. A study by Iwamoto et al. (16) involved gene expression analyses on 16 matched-paired samples between primary breast cancers and brain metastases. The results aimed to confirm previously reported genes associated with brain metastases and epithelial-mesenchymal transition (EMT) and identify novel therapeutic targets among FDA-approved agents or those investigated in clinical trials for



in brain metastasis (adj. $p \le 0.05$). The word "pri" in the figure refers to primary tumor.

distinct cancers. Their results revealed that immune-related signatures exhibited significantly lower gene expression in brain metastases than in primary breast cancers. Similar to our approach, Tsakonas et al. (2) employed immune-gene expression profiling of 13 primary lung samples and their matched-paired BM. However, detailed information on the specific subtype of the matched-paired samples is not available. Their results revealed significant downregulated of 12 immune-related genes in BM compared to primary tumors. Importantly, we found an FDA-approved druggable target (HDAC3, Figures 5B, C) that was differentially expressed in LUAD and was upregulated in BM-LUAD compared to primary LUAD. This finding suggests the potential for targeted therapy in BM-LUAD. In our previous study we discovered higher infiltration of immune cells in BM-LUAD as compared to BM-BC (19). Interestingly, despite the higher immune response regulation in BM-BC, we did not find significant immune infiltration between the primary tumors and their matched-paired brain metastases. This could be attributed to factors such as the poor blood-brain



barrier (BBB) permeability, differences in genetic characteristics, or insufficient stimulation to facilitate additional infiltration. However, we discovered four immune checkpoint molecules that exhibited differential expression between BM-BC and BM-LUAD, suggesting their potential for developing lineage-specific therapeutic strategies for brain metastasis. Importantly, in line with our previous study, we found higher expression of VISTA and IDO1 in primary LUAD and BM-LUAD as compared to breast cancer, with a predominant higher expression in primary tumors. VISTA is an immune regulatory receptor expressed mainly by myeloid cells and is associated with poor overall survival in various cancers (25, 26). We previously found that VISTA is expressed on tumor cells,



0.05, **adj. $p \le 0.01$, ***adj. $p \le 0.001$).

T lymphocytes, and to a lesser extent on microglia (19). The results highlight potential for targeted therapy strategies directed against VISTA in the treatment of brain metastasis originating from LUAD.

IDO1 is an immune response regulator involved in the suppression of the anti-tumor immune response and immune evasion. IDO1 is highly expressed in various malignant tumors including lung cancer, and its overexpression is linked to unfavorable clinical outcomes (27-30). Although we found higher expression of IDO1 in BM-LUAD, its level was relatively low as compared to VISTA. Nevertheless, the predominant expression of IDO1 in BM-LUAD should be taken into account regarding the development of combination therapies targeting IDO1. Similarly, the higher expression of IDO1 in 13 primary lung cancer and their matched-paired samples we discovered previously (2), confirming the results of our study. We also found higher expression of NT5E (CD73) in BM-LUAD than in BM-BC. CD73 is a transmembrane glycoprotein and exerts diverse functions in the tumor microenvironment (31). CD73 catalyzes the hydrolysis of adenosine, which contributes to cancer progression, neovascularization, tumor cell proliferation, and immune evasion (32). Moreover, CD73 has been found to be involved in invasion and metastasis across different cancers, such as ovarian and cervical

carcinomas (32). Dual blockade of CD73-TGF β has led to promote a complex inflammatory tumor microenvironment, characterized by decreased levels of myeloid-derived suppressor cells (MDSCs) and M2-macrophages, as well as increased levels of activated dendritic cells, cytotoxic T cells, and B cells (33). The prominent expression of CD73 in BM-LUAD, not in BM-BC, prompts this molecule as another target for the treatment of BM-LUAD. VTCN1, also known as B7-H4, is an immune regulatory molecule belonging to the B7 family (34). VTCN1 is mainly expressed by antigenpresenting cells (APCs) and induces immune suppression by inhibiting the expansion of neutrophil progenitors and T cells (35-37). Overexpression of VTCN1 in tumors correlates with their clinicopathological features, and promotes tumor proliferation, metastasis and immune evasion (38-43). We found higher expression of VTCN1 in BM-BCthan in their primary tumors, and also in BM-LUAD. The significant expression of VTCN1 in BM-BC should be considered as another potential target for inclusion in the development of targeted immunotherapy, especially in targeting BM-BC. In line with the previous article (44), we hypothesize that the immune system may play a role in the development of brain metastasis in breast and lung cancer. This particular immune system involvement



FIGURE 7

Immunohistochemistry staining for immune checkpoint inhibitors in the BM-BC and BM-LUAD samples. (A) Box plots of the differentially expressed immune checkpoint molecules in BM-BC and BM-LUAD (adj. $p \le 0.05$). (B) IHC staining for the expression of therapeutic targets VISTA, IDO1, NT5E (CD73), and VTCN1 (B7-H4) in BM-BC and BM-LUAD samples. The red color in each IHC image shows the expression of associated target on tissue of BC and LUAD. The border colors of each image are as follow, light pink: primary BC, dark pink: BM-BC, light blue: primary LUAD, dark blue: BM-LUAD.

requires additional validation, particularly in the case of lung adenocarcinoma.

There are some limitations in our study. The cohort size we analyzed was relatively small, however, it is important to highlight the exceptional uniqueness of the samples we had the opportunity to analyze. Utilizing matched-paired primary tumor and brain metastasis samples is an important and unique approach in cancer research, allowing for direct comparisons of molecular changes between the primary tumor and its corresponding brain metastasis. This not only unravels the specific adaptations that enable tumor cells to invade the brain but also sheds light on the mechanisms governing the selection and survival of metastatic cells in this unique microenvironment. Moreover, a control group consisting of tumors that did not metastasize to the brain was lacking for direct comparisons, however, our study provides valuable insights into various intriguing aspects of brain metastasis. In conclusion, the comparison of molecular mechanism in primary breast and LUAD with their matched-paired brain metastases, provide valuable insights into the molecular landscape of brain metastasis. The findings of this study highlight the differential gene expression patterns, the significance of metabolic pathways specially the involvement of glycolysis pathway, and the impact of immune response modulation in brain metastasis. In addition, the identification of the FDA-approved druggable target (HDAC3) and the four immune checkpoint molecules in BM suggest that targeted therapies should be tailored for specific tumor lineages in order to effectively manage brain metastasis.

Data availability statement

The data presented in the study are deposited in the https:// www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE248830 repository, accession number GSE248830.

Ethics statement

This study was approved by the Medical Ethics Committee of the Erasmus Medical Center, Rotterdam, the Netherlands and was carried out in adherence to the Code of Conduct of the Federation of Medical Scientific Societies in the Netherlands (MEC 02.953 & MEC-2020–0732). The studies were conducted in accordance with the local legislation and institutional requirements. The participants provided their written informed consent to participate in this study.

Author contributions

SN: Data curation, Formal analysis, Investigation, Methodology, Resources, Software, Validation, Visualization, Writing – original draft. WK: Formal analysis, Writing – review & editing. JK: Conceptualization, Investigation, Project administration, Resources, Supervision, Writing – review &

References

1. Noh M-G, Kim SS, Kim YJ, Jung T-Y, Jung S, Rhee J-H, et al. Evolution of the tumor microenvironment toward immune-suppressive seclusion during brain metastasis of breast cancer: implications for targeted therapy. *Cancers* (2021) 13 (19):4895. doi: 10.3390/cancers13194895

2. Tsakonas G, Lewensohn R, Botling J, Ortiz-Villalon C, Micke P, Friesland S, et al. An immune gene expression signature distinguishes central nervous system metastases from primary tumours in non-small-cell lung cancer. *Eur J Cancer*. (2020) 132:24–34. doi: 10.1016/j.ejca.2020.03.014

3. Hamed M, Schäfer N, Bode C, Borger V, Potthoff A-L, Eichhorn L, et al. Preoperative metastatic brain tumor-associated intracerebral hemorrhage is associated with dismal prognosis. *Front Oncol* (2021) 11:3612. doi: 10.3389/fonc.2021.699860

4. Jablonska PA, Bosch-Barrera J, Serrano D, Valiente M, Calvo A, Aristu J. Challenges and novel opportunities of radiation therapy for brain metastases in non-small cell lung cancer. *Cancers* (2021) 13(9):2141. doi: 10.3390/cancers13092141

editing. DM: Conceptualization, Investigation, Project administration, Resources, Supervision, Writing – review & editing.

Funding

The author(s) declare that no financial support was received for the research, authorship, and/or publication of this article.

Acknowledgments

The authors thank the technical assistance provided by the Erasmus MC Pathology Research and Trial Service (PARTS) with the immunohistochemistry.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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Supplementary material

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fimmu.2023.1305644/full#supplementary-material

5. Bos PD, Zhang XHF, Nadal C, Shu W, Gomis RR, Nguyen DX, et al. Genes that mediate breast cancer metastasis to the brain. *Nature* (2009) 459(7249):1005–9. doi: 10.1038/nature08021

6. Lee JY, Park K, Lee E, Ahn T, Jung HH, Lim SH, et al. Gene expression profiling of breast cancer brain metastasis. *Sci Rep* (2016) 6(1):28623. doi: 10.1038/srep28623

7. Fares J, Kanojia D, Cordero A, Rashidi A, Miska J, Schwartz CW, et al. Current state of clinical trials in breast cancer brain metastases. *Neuro-oncol practice.* (2019) 6 (5):392–401. doi: 10.1093/nop/npz003

8. Fares J, Kanojia D, Rashidi A, Ahmed AU, Balyasnikova IV, Lesniak MS. Diagnostic clinical trials in breast cancer brain metastases: barriers and innovations. *Clin Breast cancer*. (2019) 19(6):383–91. doi: 10.1016/j.clbc.2019.05.018

9. Fares J, Fares MY, Fares Y. Immune checkpoint inhibitors: Advances and impact in neuro-oncology. *Surg Neurol Int* (2019) 10. doi: 10.4103/sni.sni_366_18

10. Fares J, Kanojia D, Rashidi A, Ulasov I, Lesniak MS. Genes that mediate metastasis across the blood-brain barrier. *Trends cancer.* (2020) 6(8):660–76. doi: 10.1016/j.trecan.2020.04.007

11. Ebright RY, Zachariah MA, Micalizzi DS, Wittner BS, Niederhoffer KL, Nieman LT, et al. HIF1A signaling selectively supports proliferation of breast cancer in the brain. *Nat Commun* (2020) 11(1):6311. doi: 10.1038/s41467-020-20144-w

12. Cameron MD, Schmidt EE, Kerkvliet N, Nadkarni KV, Morris VL, Groom AC, et al. Temporal progression of metastasis in lung: cell survival, dormancy, and location dependence of metastatic inefficiency. *Cancer Res* (2000) 60(9):2541–6.

13. Carreau A, Hafny-Rahbi BE, Matejuk A, Grillon C, Kieda C. Why is the partial oxygen pressure of human tissues a crucial parameter? Small molecules and hypoxia. *J Cell Mol Med* (2011) 15(6):1239–53. doi: 10.1111/j.1582-4934.2011.01258.x

14. Ruoslahti E. Brain extracellular matrix. *Glycobiology* (1996) 6(5):489-92. doi: 10.1093/glycob/6.5.489

15. Koh YW, Han J-H, Haam S, Lee HW. An immune-related gene expression signature predicts brain metastasis in lung adenocarcinoma patients after surgery: gene expression profile and immunohistochemical analyses. *Trans Lung Cancer Res* (2021) 10(2):802. doi: 10.21037/tlcr-20-1056

16. Iwamoto T, Niikura N, Ogiya R, Yasojima H, K-i W, Kanbayashi C, et al. Distinct gene expression profiles between primary breast cancers and brain metastases from pairmatched samples. *Sci Rep* (2019) 9(1):13343. doi: 10.1038/s41598-019-50099-y

17. Varešlija D, Priedigkeit N, Fagan A, Purcell S, Cosgrove N, O'Halloran PJ, et al. Transcriptome characterization of matched primary breast and brain metastatic tumors to detect novel actionable targets. *JNCI: J Natl Cancer Institute.* (2019) 111 (4):388–98. doi: 10.1093/jnci/djy110

18. Wismans LV, Lopuhaä B, de Koning W, Moeniralam H, van Oosterhout M, Ambarus C, et al. Increase of mast cells in COVID-19 pneumonia may contribute to pulmonary fibrosis and thrombosis. *Histopathology* (2022) 82:407–19. doi: 10.1111/his.14838

19. Najjary S, Kros JM, de Koning W, Vadgama D, Lila K, Wolf J, et al. Tumor lineage-specific immune response in brain metastatic disease: opportunities for targeted immunotherapy regimen? *Acta neuropathologica Commun* (2023) 11(1):64. doi: 10.1186/s40478-023-01542-9

20. Tukey JW. *Exploratory Data Analysis*. Mosteller F, editor. Boston, MA, USA: Addison-Wesley Publishing Company (1977).

21. Katano A, Yamashita H. Brain metastasis: Recent treatment modalities and future-perspectives. Oncol Letters. (2022) 23(6):1-6. doi: 10.3892/ol.2022.13311

22. Dupuy F, Tabariès S, Andrzejewski S, Dong Z, Blagih J, Annis MG, et al. PDK1dependent metabolic reprogramming dictates metastatic potential in breast cancer. *Cell Metab* (2015) 22(4):577–89. doi: 10.1016/j.cmet.2015.08.007

23. Parida PK, Marquez-Palencia M, Nair V, Kaushik AK, Kim K, Sudderth J, et al. Metabolic diversity within breast cancer brain-tropic cells determines metastatic fitness. *Cell Metab* (2022) 34(1):90–105.e7. doi: 10.1016/j.cmet.2021.12.001

24. Tchou J, Sonnad SS, Bergey MR, Basu S, Tomaszewski J, Alavi A, et al. Degree of tumor FDG uptake correlates with proliferation index in triple negative breast cancer. *Mol Imaging Biol* (2010) 12:657–62. doi: 10.1007/s11307-009-0294-0

25. Xie S, Huang J, Qiao Q, Zang W, Hong S, Tan H, et al. Expression of the inhibitory B7 family molecule VISTA in human colorectal carcinoma tumors. *Cancer Immunol Immunother* (2018) 67:1685–94. doi: 10.1007/s00262-018-2227-8

26. ElTanbouly MA, Schaafsma E, Noelle RJ, Lines JL. VISTA: Coming of age as a multi-lineage immune checkpoint. *Clin Exp Immunol* (2020) 200(2):120–30. doi: 10.1111/cei.13415

27. Hennequart M, Pilotte L, Cane S, Hoffmann D, Stroobant V, Plaen ED, et al. Constitutive IDO1 expression in human tumors is driven by cyclooxygenase-2 and mediates intrinsic immune resistance. *Cancer Immunol Res* (2017) 5(8):695–709. doi: 10.1158/2326-6066.CIR-16-0400

28. Meng Y, Wang W, Chen M, Chen K, Xia X, Zhou S, et al. GBP1 facilitates indoleamine 2, 3-dioxygenase extracellular secretion to promote the Malignant

progression of lung cancer. Front Immunol (2021) 11:622467. doi: 10.3389/fimmu.2020.622467

29. Kozuma Y, Takada K, Toyokawa G, Kohashi K, Shimokawa M, Hirai F, et al. Indoleamine 2, 3-dioxygenase 1 and programmed cell death-ligand 1 co-expression correlates with aggressive features in lung adenocarcinoma. *Eur J Cancer.* (2018) 101:20–9. doi: 10.1016/j.ejca.2018.06.020

30. Kocher F, Amann A, Zimmer K, Geisler S, Fuchs D, Pichler R, et al. High indoleamine-2, 3-dioxygenase 1 (IDO) activity is linked to primary resistance to immunotherapy in non-small cell lung cancer (NSCLC). *Trans Lung Cancer Res* (2021) 10(1):304. doi: 10.21037/tlcr-20-380

31. Resta R, Yamashita Y, Thompson LF. Ecto-enzyme and signaling functions of lymphocyte CD 7 3. *Immunol Rev* (1998) 161(1):95–109. doi: 10.1111/j.1600-065X.1998.tb01574.x

32. Takedachi M, Oohara H, Smith BJ, Iyama M, Kobashi M, Maeda K, et al. CD73generated adenosine promotes osteoblast differentiation. *J Cell Physiol* (2012) 227 (6):2622–31. doi: 10.1002/jcp.23001

33. Xing Y, Ren Z-q, Jin R, Liu L, Pei J-p, Yu K. Therapeutic efficacy and mechanism of CD73-TGF β dual-blockade in a mouse model of triple-negative breast cancer. Acta Pharmacologica Sinica. (2022) 43(9):2410–8. doi: 10.1038/s41401-021-00840-z

34. Gao A, Zhang L, Chen X, Chen Y, Xu Z, Liu Y, Xu W. Effect of VTCN1 on progression and metastasis of ovarian carcinoma *in vitro* and *vivo*. *Biomed Pharmacother* (2015) 73:129–34. doi: 10.1016/j.biopha.2015.05.016

35. Zang X, Pn L, Kim J, Murphy K, Waitz R, Allison JP. B7x: a widely expressed B7 family member that inhibits T cell activation. *Proc Natl Acad Sci* (2003) 100(18):10388–92. doi: 10.1073/pnas.1434299100

36. Sica GL, Choi I-H, Zhu G, Tamada K, Wang S-D, Tamura H, et al. B7-H4, a molecule of the B7 family, negatively regulates T cell immunity. *Immunity* (2003) 18 (6):849–61. doi: 10.1016/S1074-7613(03)00152-3

37. He C, Qiao H, Jiang H, Sun X. The inhibitory role of b7-h4 in antitumor immunity: association with cancer progression and survival. *J Immunol Res* (2011) 2011. doi: 10.1155/2011/695834

38. Krambeck AE, Thompson RH, Dong H, Lohse CM, Park ES, Kuntz SM, et al. B7-H4 expression in renal cell carcinoma and tumor vasculature: associations with cancer progression and survival. *Proc Natl Acad Sci* (2006) 103(27):10391–6. doi: 10.1073/pnas.0600937103

39. Zang X, Thompson RH, Al-Ahmadie HA, Serio AM, Reuter VE, Eastham JA, et al. B7-H3 and B7x are highly expressed in human prostate cancer and associated with disease spread and poor outcome. *Proc Natl Acad Sci* (2007) 104(49):19458–63. doi: 10.1073/pnas.0709802104

40. Assal A, Kaner J, Pendurti G, Zang X. Emerging targets in cancer immunotherapy: beyond CTLA-4 and PD-1. *Immunotherapy* (2015) 7(11):1169-86. doi: 10.2217/imt.15.78

41. John P, Wei Y, Liu W, Du M, Guan F, Zang X. The B7x immune checkpoint pathway: from discovery to clinical trial. *Trends Pharmacol Sci* (2019) 40(11):883–96. doi: 10.1016/j.tips.2019.09.008

42. Podojil JR, Glaser AP, Baker D, Courtois ET, Fantini D, Yu Y, et al. Antibody targeting of B7-H4 enhances the immune response in urothelial carcinoma. *Oncoimmunology* (2020) 9(1):1744897. doi: 10.1080/2162402X.2020.1744897

43. Yan X, Feng J, Hong B, Qian Y. The expression of PD-L1 and B7-H4 in Thymic epithelial tumor and its relationship with tumor immune-infiltrating cells. *Front Oncol* (2021) 11:662010. doi: 10.3389/fonc.2021.662010

44. Mustafa DA, Pedrosa RM, Smid M, van der Weiden M, de Weerd V, Nigg AL, et al. T lymphocytes facilitate brain metastasis of breast cancer by inducing Guanylate-Binding Protein 1 expression. *Acta neuropathologica*. (2018) 135:581–99. doi: 10.1007/ s00401-018-1806-2