Check for updates

OPEN ACCESS

EDITED BY Ferdaus Mohd Altaf Hossain, Sylhet Agricultural University, Bangladesh

REVIEWED BY Wayne Robert Thomas, University of Western Australia, Australia

*CORRESPONDENCE Paul Engeroff Maul.engeroff@unibe.ch

RECEIVED 15 November 2023 ACCEPTED 11 December 2023 PUBLISHED 11 January 2024

CITATION

Storni F, Vogel M, Bachmann MF and Engeroff P (2024) IgG in the control of FccRI activation: a battle on multiple fronts. *Front. Immunol.* 14:1339171. doi: 10.3389/fimmu.2023.1339171

COPYRIGHT

© 2024 Storni, Vogel, Bachmann and Engeroff. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

IgG in the control of FcERI activation: a battle on multiple fronts

Federico Storni^{1,2}, Monique Vogel^{2,3}, Martin F. Bachmann^{2,3} and Paul Engeroff^{2,3}*

¹Department of Visceral Surgery and Medicine, Inselspital, Bern University Hospital, University of Bern, Bern, Switzerland, ²Department of BioMedical Research, University of Bern, Bern, Switzerland, ³Department of Rheumatology and Immunology, University Hospital Bern, Bern, Switzerland

The rising global incidence of IgE-mediated allergic reactions poses a significant challenge to the quality of life of affected individuals and to healthcare systems, with current treatments being limited in effectiveness, safety, and disease-modifying capabilities. IgE acts by sensitizing the high-affinity IgE receptor FccRI expressed by mast cells and basophils, tuning these cells for inflammatory degranulation in response to future allergen encounters. In recent years, IgG has emerged as an essential negative regulator of IgE-dependent allergic inflammation. Mechanistically, studies have proposed different pathways by which IgG can interfere with the activation of IgE-mediated inflammation. Here, we briefly summarize the major proposed mechanisms of action by which IgG controls the IgE-FccRI inflammatory axis and how those mechanisms are currently applied as therapeutic interventions for IgE-mediated inflammation.

KEYWORDS

allergy, IgE, mast cells, basophils, degranulation, FcyRIIB, antibody, sensitization

1 FccRI sensitization and activation

Whether IgE switching occurs from IgM or IgG remains a debated topic and has been discussed elsewhere (1, 2). Here, we focus on how IgG controls the immediate allergic reaction once IgE is produced by B cells. The activation of inflammatory mast cell and basophil degranulation by IgE is a two-step process (3, 4). First, IgE needs to sensitize mast cells and basophils by binding to the high-affinity IgE receptor FceRI. This interaction is remarkably stable and sensitization can thus persist for long periods, specifically in the more long-lived mast cells (5, 6). The second event is the encounter with an antigen that cross-links IgE bound to FceRI leading to FceRI intracellular signaling cascades and degranulation (7, 8). If improperly controlled, this reaction can have negative consequences such as in allergy, where harmless antigens (allergens) become activators of $Fc\epsilon RI$. Currently, allergic diseases are posing massive challenges to global health and thus, strategies to inhibit the IgE : $Fc\epsilon RI$ inflammatory axis have been of high interest to clinical and pharmaceutical research (9, 10). In recent years, it has become clear that IgG antibodies may be important in both types of physiological $Fc\epsilon RI$ de-granulation control, the sensitization and the activation phase.

2 IgG specific for IgE in the prevention of FcERI sensitization

We and others have shown that naturally occurring IgG anti-IgE autoantibodies are present to surprising levels in humans and mice. In both humans and mice, the antibodies can prevent basophil sensitization and FcERI activation (11-14). Interestingly, the immunogenicity of self-IgE is regulated by a single highly conserved mannose glycosylation site on IgE, which is likewise preferentially recognized by the anti-IgE autoantibodies (12, 15). The same mannose glycan is crucial for both FcERI binding and IgE sensitization (16, 17). It thus seems plausible that competition between anti-IgE autoantibodies and FcERI for this single mannose glycosylation site explains how IgE-IgG ICs fail to activate FcERI. Nevertheless, this aspect remains to be studied in further detail. We have shown another mechanism by which anti-IgE auto-antibodies act. The formation of IgG immune complexes with IgE (IgE-IgG-IC) results in preferential targeting of the lowaffinity IgE receptor CD23 over FcERI (12). CD23 is constitutively expressed in B cells but can be induced in a variety of other cells (18). We had previously shown that CD23 is essential in the clearance of IgE-allergen-IC but this is likewise the case for IgE-IgG-ICs (12, 19). Moreover, the immune response caused by IgE-IgG-ICs results in a boost of anti-IgE autoantibodies establishing a positive feedback loop that diminishes serum IgE levels further. Interestingly, studies have proposed maternal transfer of IgE-IgG-IC via the neonatal FcRn receptor (20, 21). It is important to note, that any dysregulation of those anti-IgE autoantibodies may lead to pathologies, for example they are thought to inadvertently crosslink FcERI in Chronic Spontaneous Urticaria (CSU) or atopic dermatitis (AD) thus contributing to disease (22, 23). Overall, the role of natural anti-IgE antibodies in health and disease needs to be further investigated in future studies.

3 IgG specific for allergen in the inhibition of FccRI activation

Once IgE has bound to FccRI, allergen-specific IgG may still block allergic reactions by allergen-neutralization. IgG circulates the blood at very high concentrations compared to IgE and if enough specific IgG is present, is thus likely to form IgG-allergen ICs before FceRI is encountered. Numerous studies have shown a role for blocking antibodies in preventing the initiation of the allergic cascade by impeding the binding of allergens to IgE (24-26). Due to their lower ability to activate complement and Fc-dependent effects, IgG4 antibodies have been considered specifically optimal blocking antibodies in allergy (27-30). However, neutralizing IgGs are not required to suppress FcERI. Even if an IgG-allergen IC reaches the cell and is recognized by IgE, there is a second mechanism at work that suppresses the allergic response. Mast cells and basophils can express the inhibitory IgG receptor FcyRIIb, which contains an ITIM motif that activates the inositol phosphatase SHIP, a major negative immune regulator (31, 32). When FcERI is cross-linked to FcyRIIb, the activation cascade of FcERI is suppressed via SHIP (32-35). Notably, the inhibitory signal from FcyRIIb does not only act on individual allergen-IgG pairs but SHIP can spread around the cell membrane and dephosphorylate distant FcERI and activated kinases (36). While it was often speculated that IgG4 is superior in suppressing the allergic reaction via FcyRIIb, it was recently shown that IgG1 and IgG4 appear to similarly suppress FcERI activation via FcyRIIb (33, 37). Moreover, IgG affinity for the allergen seems less important for FcyRIIb engagement than for neutralizing antibodies (38). Finally, FceRI : FcyRIIb complexes can be internalized and degraded, resulting in a de-sensitizing of effector cells, coupled with a reduction of IgE (39). In actuality, both neutralization and FcyRIIb engagement most likely occur simultaneously during polyclonal responses in vivo (40).

4 The role of IgG in allergy immunotherapy

Elevated levels of IgG antibodies have been observed in individuals with natural tolerance to allergens, suggesting a possibly inherent protective role of these antibodies (41). In mice, maternal IgG-allergen complexes transferred via breast milk and the neonatal receptor FcRn were shown to protect the offspring from allergy (42). On the other hand, IgG has been extensively studied in allergen-specific immunotherapy (AIT), a diseasemodifying treatment for allergy, where increasing doses of specific allergens are administered to allergic individuals to build tolerance (43, 44). Successful AIT has been associated with elevated levels of allergen-specific IgG. Specifically, it is accepted that the induction of high IgG : IgE ratios in addition to regulatory T and B cells is a favorable and protective occurrence whereas IgE levels are initially increased with AIT before they are later reduced (30, 45-49). Moreover, high IgG levels have been associated with reduced risk of more severe, anaphylactic reactions (50). While in AIT, the IgG4 subclass is usually associated best with protection from allergy, this most likely reflects superior induction of IgG4 over IgG1 by most

AIT-protocols and does not necessarily indicate an inferiority of IgG1 to neutralize the allergen or engage FcyRIIb. Recently, it was shown that IgG1 may suppress the allergic during earlier AIT responses whereas IgG4 gains importance in later stages (37, 49). Even though some mechanistic aspects remain elusive, current findings suggest AIT, the only current disease-modifying therapy for allergic diseases involves the induction of protective IgG antibodies.

5 Therapeutic approaches based on the anti-allergic properties of IgG

The IgG-mediated control of FcERI sensitization and activation have been increasingly translated into clinical development and application. Omalizumab, a monoclonal anti-IgE antibody which blocks FcERI sensitization has been developed a long time ago (51). Mechanistically, anti-IgE biologics act similar to natural anti-IgE autoantibodies by preventing or disrupting the FcERI-IgE interaction and by lowering serum IgE levels. We have recently shown that omalizumab function, in identical fashion to natural anti-IgE autoantibodies, depends on the single IgE mannose glycosylation site (52). A number of other IgE-targeting approaches are explored, which have been reviewed by others (53-61). The development of allergen-specific IgG for the treatment of allergy is a more recent development and for the most part, clinical trials are still ongoing (62, 63). Two major strategies are currently being employed, both aiming at harnessing the protective role of allergen-specific IgG. Neutralizing monoclonal anti-allergen IgG antibodies have demonstrated efficacy in suppressing allergy in preclinical and

clinical settings (64-72). While the results are generally positive, questions on their cost-effectiveness remain. Nevertheless, their ability to increase the safety of allergy immunotherapy and potentially boost beneficial immune responses is intriguing (63). A more cost-effective approach could be the induction of polyclonal IgG responses using immunization strategies that improve the current drawbacks of AIT including safety concerns, patient compliance, long treatment periods and varying efficacy. The challenge here is to modify the allergens in a way that reduces FcERI cross-linking but enhances the induction of allergen-specific IgG. Others have reviewed the current landscape of vaccination strategies for improving AIT (73-79). We have focused on using allergens displayed on virus-like particles (VLP), which can reduce allergen reactivity while boosting IgG responses that increase protection via FcyRIIb (80, 81).

6 Discussion: future prospects and challenges

The overall importance of IgG in the control of IgE : FcERI in basic and preclinical research is established. Ongoing and future research will define the relative importance of the here-presented individual pathways through which IgG confers protection against allergies (Figure 1). As research advances, it is expected that a more nuanced and contextual understanding of IgG in allergic reactions will be achieved. This will pave the way for innovative therapeutic strategies translating the protective potential of IgG into the clinics, thereby offering relief to millions of people suffering from allergies worldwide.



The diverse role of IgG in shutting down-the allergic reaction. (A) Naturally occurring IgG anti-IgE autoantibodies can limit the extent of FccRI sensitization and thereby prevent the allergic reaction. (B) In the case that FccRI sensitization has already occurred, neutralizing IgG anti-allergen antibodies can block allergen recognition by IgE, thus preventing FccRI activation. (C) Even in absence of neutralizing IgG antibodies, the inhibitory FcyRIIb receptor suppresses the allergic reaction when it is cross-linked to FceRI. (D) In the absence of IgG-mediated control, the allergic reaction can occur. Created with BioRender.com

Author contributions

FS: Writing – original draft. MV: Writing – review & editing. MB: Writing – review & editing. PE: Writing – original draft.

Funding

The author(s) declare financial support was received for the research, authorship, and/or publication of this article. This project was supported by funding from the following grants: SNF grant 310030_179165 to MV; SNF grant 310030_185114 to MB; SNF grant P2BEP3_188262 to PE.

References

1. Geha RS, Jabara HH, Brodeur SR. The regulation of immunoglobulin E classswitch recombination. *Nat Rev Immunol* (2003) 3:721–32. doi: 10.1038/nri1181

2. Saunders SP, Ma EGM, Aranda CJ, Curotto de Lafaille MA. Non-classical B cell memory of allergic igE responses. *Front Immunol* (2019) 10:715. doi: 10.3389/ fimmu.2019.00715

3. Bulfone-Paus S, Nilsson G, Draber P, Blank U, Levi-Schaffer F. Positive and negative signals in mast cell activation. *Trends Immunol* (2017) 38:657–67. doi: 10.1016/j.it.2017.01.008

 Gould HJ, Sutton BJ. IgE in allergy and asthma today. Nat Rev Immunol (2008) 8:205–17. doi: 10.1038/nri2273

5. Holdom MD, Davies AM, Nettleship JE, Bagby SC, Dhaliwal B, Girardi E, et al. Conformational changes in IgE contribute to its uniquely slow dissociation rate from receptor FceRI. *Nat Struct Mol Biol* (2011) 18:571–6. doi: 10.1038/nsmb.2044

 Galli SJ, Tsai M. IgE and mast cells in allergic disease. Nat Med (2012) 18:693–704. doi: 10.1038/nm.2755

7. Kinet J-P. The high-affinity IgE receptor (FccRI): From Physiology to Pathology. Annu Rev Immunol (1999) 17:931-72. doi: 10.1146/annurev.immunol.17.1.931

8. Wilson BS, Pfeiffer JR, Oliver JM. Observing Fc epsilon RI signaling from the inside of the mast cell membrane. J Cell Biol (2000) 149:1131–42. doi: 10.1083/jcb.149.5.1131

9. Platts-Mills TAE. The allergy epidemics: 1870-2010. J Allergy Clin Immunol (2015) 136:3-13. doi: 10.1016/j.jaci.2015.03.048

 Genuneit J, Seibold AM, Apfelbacher CJ, Konstantinou GN, Koplin JJ, La Grutta S, et al. Overview of systematic reviews in allergy epidemiology. *Allergy* (2017) 72:849– 56. doi: 10.1111/all.13123

11. Engeroff P, Plattner K, Storni F, Thoms F, Frias Boligan K, Muerner L, et al. Glycan-specific IgG anti-IgE autoantibodies are protective against allergic anaphylaxis in a murine model. *J Allergy Clin Immunol* (2021) 147:1430–41. doi: 10.1016/j.jaci.2020.11.031

12. Plattner K, Gharailoo Z, Zinkhan S, Engeroff P, Bachmann MF, Vogel M. IgE glycans promote anti-IgE IgG autoantibodies that facilitate IgE serum clearance via Fc Receptors. *Front Immunol* (2022) 13:1069100. doi: 10.3389/fimmu.2022.1069100

13. Chan Y-C, Ramadani F, Santos AF, Pillai P, Ohm-Laursen L, Harper CE, et al. "Auto-anti-IgE": Naturally occurring IgG anti-IgE antibodies may inhibit allergeninduced basophil activation. *J Allergy Clin Immunol* (2014) 134:1394–1401.e4. doi: 10.1016/j.jaci.2014.06.029

14. Jensen-Jarolim E, Vogel M, de Weck AL, Stadler BM. Anti-IgE autoantibodies mistaken for specific IgG. *J Allergy Clin Immunol* (1992) 89:31–43. doi: 10.1016/S0091-6749(05)80038-7

15. Plattner K, Bachmann MF, Vogel M. On the complexity of IgE: The role of structural flexibility and glycosylation for binding its receptors. *Front Allergy* (2023) 4:1117611. doi: 10.3389/falgy.2023.1117611

16. Shade K-TC, Platzer B, Washburn N, Mani V, Bartsch YC, Conroy M, et al. A single glycan on IgE is indispensable for initiation of anaphylaxis. *J Exp Med* (2015) 212:457–67. doi: 10.1084/jem.20142182

17. Shade K-T, Conroy ME, Anthony RM. IgE glycosylation in health and disease. *Curr Top Microbiol Immunol* (2019) 423:77–93. doi: 10.1007/82_2019_151

18. Engeroff P, Vogel M. The role of CD23 in the regulation of allergic responses. *Allergy.* (2021) 76:1981–9. doi: 10.1111/all.14724

19. Engeroff P, Caviezel F, Mueller D, Thoms F, Bachmann MF, Vogel M. CD23 provides a noninflammatory pathway for IgE-allergen complexes. J Allergy Clin Immunol (2020) 145:301–311.e4. doi: 10.1016/j.jaci.2019.07.045

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

20. Bundhoo A, Paveglio S, Rafti E, Dhongade A, Blumberg RS, Matson AP. Evidence that FcRn mediates the transplacental passage of maternal IgE in the form of IgG anti-IgE / IgE immune complexes. *Clin Exp Allergy* (2015) 45(6):1085–98. doi: 10.1111/cea.12508

21. Dwyer DF, Boyce JA. Neonatal mast cells and transplacental IgE transfer: A mechanism of disease inheritance or of passive infant barrier defense? J Allergy Clin Immunol (2021) 148:76–7. doi: 10.1016/j.jaci.2021.02.046

22. Kolkhir P, Giménez-Arnau AM, Kulthanan K, Peter J, Metz M, Maurer M. Urticaria. Nat Rev Dis Prim. (2022) 8:61. doi: 10.1038/s41572-022-00389-z

23. Poto R, Quinti I, Marone G, Taglialatela M, de Paulis A, Casolaro V, et al. IgG autoantibodies against igE from atopic dermatitis can induce the release of cytokines and proinflammatory mediators from basophils and mast cells. *Front Immunol* (2022) 13:880412. doi: 10.3389/fimmu.2022.880412

24. Flicker S, Valenta R. Renaissance of the blocking antibody concept in type I allergy. Int Arch Allergy Immunol (2003) 132:13-24. doi: 10.1159/000073260

25. Gadermaier E, James LK, Shamji MH, Blatt K, Fauland K, Zieglmayer P, et al. Epitope specificity determines cross-protection of a SIT-induced IgG4 antibody. *Allergy.* (2016) 71:36–46. doi: 10.1111/all.12710

26. Vizzardelli C, Gindl M, Roos S, Möbs C, Nagl B, Zimmann F, et al. Blocking antibodies induced by allergen-specific immunotherapy ameliorate allergic airway disease in a human/mouse chimeric model. *Allergy.* (2018) 73:851–61. doi: 10.1111/ all.13363

27. Rispens T, Huijbers MG. The unique properties of IgG4 and its roles in health and disease. *Nat Rev Immunol* (2023) 23:763–78. doi: 10.1038/s41577-023-00871-z

28. Shamji MH, Kappen J, Abubakar-Waziri H, Zhang J, Steveling E, Watchman S, et al. Nasal allergen-neutralizing IgG4 antibodies block IgE-mediated responses: Novel biomarker of subcutaneous grass pollen immunotherapy. *J Allergy Clin Immunol* (2019) 143:1067–76. doi: 10.1016/j.jaci.2018.09.039

29. Vidarsson G, Dekkers G, Rispens T. IgG subclasses and allotypes: from structure to effector functions. *Front Immunol* (2014) 5:520. doi: 10.3389/fimmu.2014.00520

30. van der Neut Kolfschoten M, Schuurman J, Losen M, Bleeker WK, Martínez-Martínez P, Vermeulen E, et al. Anti-inflammatory activity of human IgG4 antibodies by dynamic Fab arm exchange. *Science.* (2007) 317:1554–7. doi: 10.1126/science.1144603

31. Ono M, Bolland S, Tempst P, Ravetch JV. Role of the inositol phosphatase SHIP in negative regulation of the immune system by the receptor Fc(gamma)RIIB. *Nature.* (1996) 383:263–6. doi: 10.1038/383263a0

32. Burton OT, Epp A, Fanny ME, Miller SJ, Stranks AJ, Teague JE, et al. Tissuespecific expression of the low-affinity igG receptor, fcγRIIb, on human mast cells. *Front Immunol* (2018) 9:1244. doi: 10.3389/fimmu.2018.01244

33. Bruhns P, Frémont S, Daëron M. Regulation of allergy by Fc receptors. Curr Opin Immunol (2005) 17:662–9. doi: 10.1016/j.coi.2005.09.012

34. Malbec O, Daëron M. The mast cell IgG receptors and their roles in tissue inflammation. *Immunol Rev* (2007) 217:206-21. doi: 10.1111/j.1600-065X.2007.00510.x

35. Kanagaratham C, El Ansari YS, Lewis OL, Oettgen HC. IgE and igG antibodies as regulators of mast cell and basophil functions in food allergy. *Front Immunol* (2020) 11:603050. doi: 10.3389/fimmu.2020.603050

36. Malbec O, Cassard L, Albanesi M, Jönsson F, Mancardi D, Chicanne G, et al. Trans-inhibition of activation and proliferation signals by Fc receptors in mast cells and basophils. *Sci Signal* (2016) 9(459):ra126. doi: 10.1126/scisignal.aag1401 37. Zinkhan S, Thoms F, Augusto G, Vogel M, Bachmann MF. On the role of allergen-specific IgG subclasses for blocking human basophil activation. *Front Immunol* (2022) 13:892631. doi: 10.3389/fimmu.2022.892631

38. Zha L, Leoratti FMS, He L, Mohsen MO, Cragg M, Storni F, et al. An unexpected protective role of low-affinity allergen-specific IgG through the inhibitory receptor FcγRIIb. J Allergy Clin Immunol (2018) 142:1529–1536.e6. doi: 10.1016/j.jaci.2017.09.054

39. Uermösi C, Zabel F, Manolova V, Bauer M, Beerli RR, Senti G, et al. IgGmediated down-regulation of IgE bound to mast cells: a potential novel mechanism of allergen-specific desensitization. *Allergy.* (2014) 69:338–47. doi: 10.1111/all.12327

40. Strait RT, Morris SC, Finkelman FD. IgG-blocking antibodies inhibit IgEmediated anaphylaxis *in vivo* through both antigen interception and FcγRIIb crosslinking. J Clin Invest. (2006) 116:833–41. doi: 10.1172/JCI25575

41. Savilahti EM, Rantanen V, Lin JS, Karinen S, Saarinen KM, Goldis M, et al. Early recovery from cow's milk allergy is associated with decreasing IgE and increasing IgG4 binding to cow's milk epitopes. *J Allergy Clin Immunol* (2010) 125:1315–1321.e9. doi: 10.1016/j.jaci.2010.03.025

42. Ohsaki A, Venturelli N, Buccigrosso TM, Osganian SK, Lee J, Blumberg RS, et al. Maternal IgG immune complexes induce food allergen–specific tolerance in offspring. *J Exp Med* (2017) 215:91–113. doi: 10.1084/jem.20171163

43. Jutel M, Agache I, Bonini S, Burks AW, Calderon M, Canonica W, et al. International consensus on allergy immunotherapy. *J Allergy Clin Immunol* (2015) 136:556–68. doi: 10.1016/j.jaci.2015.04.047

44. Durham SR, Shamji MH. Allergen immunotherapy: past, present and future. Nat Rev Immunol (2023) 23:317–28. doi: 10.1038/s41577-022-00786-1

45. Jutel M, Agache I, Bonini S, Burks AW, Calderon M, Canonica W, et al. International Consensus on Allergen Immunotherapy II: Mechanisms, standardization, and pharmacoeconomics. *J Allergy Clin Immunol* (2016) 137:358-68. doi: 10.1016/j.jaci.2015.12.1300

46. van de Veen W, Akdis M. Role of IgG₄ in IgE-mediated allergic responses. J Allergy Clin Immunol (2016) 138:1434-5. doi: 10.1016/j.jaci.2016.07.022

47. Bachmann MF, Kündig TM. Allergen specific immunotherapy: is it vaccination against toxins after all? *Allergy.* (2017) 72:13–23. doi: 10.1111/all.12890

48. James LK, Bowen H, Calvert RA, Dodev TS, Shamji MH, Beavil AJ, et al. Allergen specificity of IgG 4-expressing B cells in patients with grass pollen allergy undergoing immunotherapy. *J Allergy Clin Immunol* (2012) 130:663–670.e3. doi: 10.1016/j.jaci.2012.04.006

49. Strobl MR, Demir H, Sánchez Acosta G, Drescher A, Kitzmüller C, Möbs C, et al. The role of IgG1 and IgG4 as dominant IgE-blocking antibodies shifts during allergen immunotherapy. *J Allergy Clin Immunol* (2023) 151:1371–1378.e5. doi: 10.1016/j.jaci.2023.01.005

50. Paolucci M, Wuillemin N, Köhli A, Ballmer-Weber B, Severin Y, Waeckerle-Men Y, et al. Multivariate allergen-specific analysis and profiling of serum antibodies from patients with peanut allergy. *Clin Exp Allergy J Br Soc Allergy Clin Immunol* (2023) 53:353–8. doi: 10.1111/cea.14262

51. Lanier B, Bridges T, Kulus M, Taylor AF, Berhane I, Vidaurre CF. Omalizumab for the treatment of exacerbations in children with inadequately controlled allergic (IgE-mediated) asthma. J Allergy Clin Immunol (2009) 124(6):1210–6. doi: 10.1016/ j.jaci.2009.09.021

52. Plattner K, Augusto G, Muerner L, von Gunten S, Jörg L, Engeroff P, et al. IgE glycosylation is essential for the function of omalizumab. *Allergy* (2023) 78(9):2546–9. doi: 10.1111/all.15748

53. Gasser P, Tarchevskaya SS, Guntern P, Brigger D, Ruppli R, Zbären N, et al. The mechanistic and functional profile of the therapeutic anti-IgE antibody ligelizumab differs from omalizumab. *Nat Commun* (2020) 11(1):165. doi: 10.1038/s41467-019-13815-w

54. Kuo BS, Li CH, Chen JB, Shiung Y, Chu C, Lee C, et al. IgE-neutralizing UB-221 mAb, distinct from omalizumab and ligelizumab, exhibits CD23-mediated IgE downregulation and relieves urticaria symptoms. *J Clin Invest* (2022) 132(15): e157765. doi: 10.1172/JCI157765

55. Maurer M, Giménez-Arnau AM, Sussman G, Metz M, Baker DR, Bauer A, et al. Ligelizumab for chronic spontaneous urticaria. *N Engl J Med* (2019) 381:1321–32. doi: 10.1056/NEJMoa1900408

56. Gauvreau GM, Arm JP, Boulet L-PP, Leigh R, Cockcroft DW, Davis BE, et al. Efficacy and safety of multiple doses of QGE031 (ligelizumab) versus omalizumab and placebo in inhibiting allergen-induced early asthmatic responses. *J Allergy Clin Immunol* (2016) 138:1051–9. doi: 10.1016/j.jaci.2016.02.027

57. Staubach P, Alvaro-Lozano M, Sekerel BE, Maurer M, Ben-Shoshan M, Porter M, et al. Ligelizumab in adolescents with chronic spontaneous urticaria: Results of a dedicated phase 2b randomized clinical trial supported with pharmacometric analysis. *Pediatr Allergy Immunol Off Publ. Eur Soc Pediatr Allergy Immunol* (2023) 34:e13982. doi: 10.1111/pai.13982

58. Gasser P, Eggel A. Targeting IgE in allergic disease. *Curr Opin Immunol* (2018) 54:86–92. doi: 10.1016/j.coi.2018.05.015

59. Incorvaia C, Mauro M, Makri E, Leo G, Ridolo E. Two decades with omalizumab: what we still have to learn. *Biologics* (2018) 12:135–42. doi: 10.2147/BTT.S180846

60. Bauer RN, Manohar M, Singh AM, Jay DC, Nadeau KC. The future of biologics: Applications for food allergy. *J Allergy Clin Immunol* (2015) 135:312–23. doi: 10.1016/ j.jaci.2014.12.1908

61. Jansson Å, Fuentes A, Magnusson A, Landström E, Öhman B, Kälvesten Å, et al. Efficient anti-IgE vaccination without anaphylactogenic properties. *J Allergy Clin Immunol* (2004) 113:S254. doi: 10.1016/j.jaci.2004.01.381

62. Landolina N, Levi-Schaffer F. Monoclonal antibodies: the new magic bullets for allergy: IUPHAR Review 17. Br J Pharmacol (2016) 173:793–803. doi: 10.1111/bph.13396

63. Pengo N, Wuillemin N, Bieli D, Gasser P. Anti-allergen monoclonal antibodies for the treatment of allergies. *Allergo J Int* (2023) 32:289–95. doi: 10.1007/s40629-023-00263-8

64. Paolucci M, Wuillemin N, Homère V, Bieli D, Köhli A, Ballmer-Weber B, et al. Targeting Ara h 2 with human-derived monoclonal antibodies prevents peanut-induced anaphylaxis in mice. *Allergy*. (2023) 78:1605–14. doi: 10.1111/all.15659

65. Orengo JM, Radin AR, Kamat V, Badithe A, Ben LH, Bennett BL, et al. Treating cat allergy with monoclonal IgG antibodies that bind allergen and prevent IgE engagement. *Nat Commun* (2018) 9:1421. doi: 10.1038/s41467-018-03636-8

66. Storni F, Cabral-Miranda G, Roesti E, Zha L, Engeroff P, Zeltins A, et al. A Single Monoclonal Antibody against the Peanut Allergen Ara h 2 Protects against Systemic and Local Peanut Allergy. *Int Arch Allergy Immunol* (2020) 181:334–41. doi: 10.1159/000505917

67. Kamal MA, Dingman R, Wang CQ, Lai C-H, Rajadhyaksha M, DeVeaux M, et al. REGN1908-1909 monoclonal antibodies block Fel d 1 in cat allergic subjects: Translational pharmacokinetics and pharmacodynamics. *Clin Transl Sci* (2021) 14:2440–9. doi: 10.1111/cts.13112

68. de Blay FJ, Gherasim A, Domis N, Meier P, Shawki F, Wang CQ, et al. REGN1908/1909 prevented cat allergen-induced early asthmatic responses in an environmental exposure unit. *J Allergy Clin Immunol* (2022) 150:1437–46. doi: 10.1016/j.jaci.2022.06.025

69. Atanasio A, Franklin MC, Kamat V, Hernandez AR, Badithe A, Ben L-H, et al. Targeting immunodominant Bet v 1 epitopes with monoclonal antibodies prevents the birch allergic response. *J Allergy Clin Immunol* (2022) 149:200–11. doi: 10.1016/j.jaci.2021.05.038

70. Gevaert P, De Craemer J, De Ruyck N, Rottey S, de Hoon J, Hellings PW, et al. Novel antibody cocktail targeting Bet v 1 rapidly and sustainably treats birch allergy symptoms in a phase 1 study. *J Allergy Clin Immunol* (2022) 149:189–99. doi: 10.1016/ j.jaci.2021.05.039

71. Shamji MH, Singh I, Layhadi JA, Ito C, Karamani A, Kouser L, et al. Passive prophylactic administration with a single dose of anti-fel d 1 monoclonal antibodies REGN1908-1909 in cat allergen-induced allergic rhinitis: A randomized, double-blind, placebo-controlled clinical trial. *Am J Respir Crit Care Med* (2021) 204:23–33. doi: 10.1164/rccm.202011-4107OC

72. LaHood NA, Min J, Keswani T, Richardson CM, Amoako K, Zhou J, et al. Immunotherapy-induced neutralizing antibodies disrupt allergen binding and sustain allergen tolerance in peanut allergy. *J Clin Invest* (2023) 133(2):e164501. doi: 10.1172/JCI164501

73. Bachmann MF, Mohsen MO, Kramer MF, Heath MD. Vaccination against allergy: A paradigm shift? *Trends Mol Med* (2020) 26(4):357-68. doi: 10.1016/j.molmed.2020.01.007

74. Satitsuksanoa P, Głobińska A, Jansen K, van de Veen W, Akdis M. Modified allergens for immunotherapy. *Curr Allergy Asthma Rep* (2018) 18(2):9. doi: 10.1007/s11882-018-0766-x

75. Dorofeeva Y, Shilovskiy I, Tulaeva I, Focke-Tejkl M, Flicker S, Kudlay D, et al. Past, present, and future of allergen immunotherapy vaccines. *Allergy.* (2021) 76:131–49. doi: 10.1111/all.14300

76. Valenta R, Campana R, Focke-Tejkl M, Niederberger V. Vaccine development for allergen-specific immunotherapy based on recombinant allergens and synthetic allergen peptides: Lessons from the past and novel mechanisms of action for the future. *J Allergy Clin Immunol* (2016) 137:351–7. doi: 10.1016/j.jaci.2015.12.1299

 Niederberger V, Marth K, Eckl-Dorna J, Focke-Tejkl M, Weber M, Hemmer W, et al. Skin test evaluation of a novel peptide carrier-based vaccine, BM32, in grass pollen-allergic patients. J Allergy Clin Immunol (2015) 136:1101–3.e8. doi: 10.1016/ j.jaci.2015.03.034

78. Banerjee S, Weber M, Blatt K, Swoboda I, Focke-Tejkl M, Valent P, et al. Conversion of Der p 23, a new major house dust mite allergen, into a hypoallergenic vaccine. *J Immunol* (2014) 192:4867–75. doi: 10.4049/jimmunol.1400064

79. Mösges R, Zeyen C, Raskopf E, Acikel C, Sahin H, Allekotte S, et al. A randomized, double-blind, placebo-controlled trial with mannan-conjugated birch pollen allergoids. *Allergy* (2023). doi: 10.1111/all.15910

80. Engeroff P, Caviezel F, Storni F, Thoms F, Vogel M, Bachmann MF. Allergens displayed on virus-like particles are highly immunogenic but fail to activate human mast cells. *Allergy.* (2018) 73:341–9. doi: 10.1111/all.13268

81. Storni F, Zeltins A, Balke I, Heath MD, Kramer MF, Skinner MA, et al. Vaccine against peanut allergy based on engineered virus-like particles displaying single major peanut allergens. *J Allergy Clin Immunol* (2020) 145:1240–1253.e3. doi: 10.1016/j.jaci.2019.12.007