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RECEIVED 02 November 2023 ACCEPTED 04 January 2024 PUBLISHED 24 January 2024

#### CITATION

Xiong Z, Peng G, Deng J, Liu M, Ning X, Zhuang Y, Yang H and Sun H (2024) Therapeutic targets and potential delivery systems of melatonin in osteoarthritis. *Front. Immunol.* 15:1331934. doi: 10.3389/fimmu.2024.1331934

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# Therapeutic targets and potential delivery systems of melatonin in osteoarthritis

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Osteoarthritis (OA) is a highly prevalent age-related musculoskeletal disorder that typically results in chronic pain and disability. OA is a multifactorial disease, with increased oxidative stress, dysregulated inflammatory response, and impaired matrix metabolism contributing to its onset and progression. The neurohormone melatonin, primarily synthesized by the pineal gland, has emerged as a promising therapeutic agent for OA due to its potential to alleviate inflammation, oxidative stress, and chondrocyte death with minimal adverse effects. The present review provides a comprehensive summary of the current understanding regarding melatonin as a promising pharmaceutical agent for the treatment of OA, along with an exploration of various delivery systems that can be utilized for melatonin administration. These findings may provide novel therapeutic strategies and targets for inhibiting the advancement of OA.

#### KEYWORDS

osteoarthritis, melatonin, inflammation, oxidative stress, chondrocyte death, delivery systems

# **1** Introduction

Osteoarthritis (OA) is a prevalent age-related irreversible musculoskeletal disorder, recognized as a primary cause of chronic pain and disability. It is characterized by persistent synovitis, progressive degradation of articular cartilage, secondary formation of osteophytes, and remodeling of subchondral bone (1–3). The pathogenesis of OA is influenced by a myriad of risk factors, encompassing age, obesity, gender, genetic predisposition, and joint injuries (4). As a refractory condition, OA can not only give rise to localized symptoms such as pain, joint deformity, and joint dysfunction but also coexist with comorbidities including diabetes, cardiac ailments, and mental health disorders, which significantly augments the likelihood of serious adverse events (5). The global prevalence of OA stands at approximately 7% of the world's population, equating to around 500 million individuals, and the number continues to rise due to the worldwide obesity epidemic and the aging demographic (6, 7). The high incidence of adverse effects

and the rapid increase in the prevalence of OA impose a substantial financial burden on society, families, and individuals, while also posing a significant threat to public health (8, 9).

The main pathological characteristics of OA include the loss of chondrocytes, degradation of the cartilage matrix, and synovitis, ultimately leading to terminal OA (10). The treatment for OA involves halting the loss of chondrocytes, promoting the production of cartilage matrix, and reducing synovitis. A wide range of therapeutic approaches have been employed for the treatment of OA, including minimally invasive surgery, conventional surgical procedures, muscle strengthening exercises, physiotherapy interventions, sodium hyaluronate injections, corticosteroids administration, and nonsteroidal anti-inflammatory drugs (NSAIDs) (11, 12). Furthermore, several emerging therapeutic strategies have demonstrated promising initial outcomes, including the transplantation of autologous chondrocytes, and the intra-articular administration of platelet-rich plasma and mesenchymal stem cells (MSCs) (13-15). Unfortunately, current therapies for individuals with OA yield unsatisfactory outcomes due to the lack of effective interventions to impede chondrocyte loss and articular cartilage deterioration (16). The investigation of novel therapeutic targets for this intricate disease is thus imperative.

The endogenous indole hormone melatonin is primarily secreted in the pineal gland, synthesized from tryptophan through a series of derivative reactions (17). The release of melatonin into the circulation of cerebrospinal fluid and bloodstream facilitates its subsequent delivery to distant organs and tissues to regulate inflammation, provide antioxidant protection, inhibit tumor growth, and promote anti-aging effects (18-21). The findings of multiple studies have demonstrated that melatonin exerts a protective effect against the development of OA through mechanisms such as inflammation reduction, elimination of excess free radicals, and promotion of matrix synthesis (22). Consequently, the potential clinical application of melatonin characterized by minimal adverse effects, holds great promise as a viable strategy for the treatment of OA. The intra-articular injection of melatonin is an optimal choice due to the absence of lymphatic and circulatory networks in hyaline cartilage. However, due to the short half-life of melatonin, it is necessary to administer injections as frequently as twice a week (23). To minimize the frequency of intra-articular injection, several delivery systems have been employed for the sustained release of melatonin. The present review provides an overview of the therapeutic advantages and delivery systems of melatonin in the progression of OA. These findings may offer a comprehensive understanding of forthcoming studies on melatonin-based treatment for OA.

# 2 The role of oxidative stress, inflammation, and chondrocyte death in OA

# 2.1 Oxidative stress in OA

The imbalance between oxidation and antioxidants leads to oxidative stress (24). Reactive oxygen species (ROS), which are

byproducts generated during aerobic metabolism, are unstable and reactive molecules such as superoxide anion (O2), hydroxyl radical (OH-), hydrogen peroxide and (H<sub>2</sub>O<sub>2</sub>). The catalysis of ROS occurs in peroxisomes and mitochondria through the action of Nitric Oxide Synthase (NOS), Xanthine Oxidase (XO), NADPH oxidases (NOXs) (25). Under physiological conditions,  $O_2^{-1}$  is the most abundant type of ROS, with the majority being generated by mitochondria. Mitochondria, known as the "powerhouse" of eukaryotic cells, convert nutrient molecules into adenosine triphosphate (ATP) through oxidative phosphorylation (26). Although the conventional consensus posits that chondrocytes derive their energy through anaerobic glycolysis in an oxygendeprived environment, the ample oxygen supply on the surface area of articular cartilage fosters conducive conditions for aerobic respiration (27, 28). The respiratory chain, located in the inner membrane of mitochondria, is widely recognized as the primary source of ROS and generates approximately 2%-3% of O2<sup>-</sup> as a byproduct during oxidative phosphorylation (29). Additionally, mitochondria play a crucial role in regulating the synthesis of antioxidant systems such as NADH/NAD+, NADPH/NADP+, and GSH/GSSG. In pathological conditions, however, mitochondrial homeostasis is disrupted, leading to an excessive generation of O2<sup>-</sup>. Excessive production of O2<sup>-</sup> leads to mitochondrial dysfunction by reducing the membrane potential of mitochondria and causing damage to mitochondrial DNA (mtDNA), thereby amplifying the generation of  $O_2^-$ . Not only does H2O2 originate from XO during the conversion of hypoxanthine to xanthine, but it can also be generated from  $O_2^{-1}$ upon activation of superoxide dismutase (SOD). Reactive nitrogen species (RNS) encompass a group of reactive molecules derived from  $O_2^-$  and NO, which are accountable for inducing nitrosative stress that contributes to cellular damage. Endothelial NOS (eNOS), neuronal NOS (nNOS), and inducible NOS (iNOS) represent three distinct isoforms of nitric oxide synthase. The production of NO is attributed to the activity of three NOSs, namely nNOS, eNOS, and iNOS. While nNOS and eNOS generate NO at a significantly low level, iNOS induced by inflammatory cytokines such as interleukin-1 $\beta$  (IL-1 $\beta$ ), IL-17, and tumor necrosis factor  $\alpha$  (TNF $\alpha$ ) exhibits a relatively high output of NO (30, 31).

The cells possess antioxidant defense mechanisms comprising both enzymatic and non-enzymatic components to counteract the heightened production of ROS and prevent cellular dysfunction. The non-enzymatic system comprises ascorbic acid (vitamin C),  $\alpha$ tocopherol (vitamin E), and glutathione (GSH), while the enzymatic component consists of SOD, catalase (CAT), glutathione peroxidase (GPX), peroxiredoxins (PRXS), and NADPH ubiquinone oxidoreductase (NQO1) (32). The SODs, comprising three isoforms including cytosolic SOD (SOD1), mitochondrial SOD (SOD2), and extracellular SOD (SOD3), effectively eliminate ROS by converting  $O_2^-$  to  $H_2O_2$ . Subsequently, the accumulated  $H_2O_2$  is further converted to H<sub>2</sub>O through the actions of GPX, PRXS, and CATs (33-35). The presence of GSH is crucial for maintaining cellular redox potential and antioxidant defenses, as it serves as a significant reductant. GPX plays a vital role in preventing the oxidation of membrane lipids by converting H<sub>2</sub>O<sub>2</sub> to H<sub>2</sub>O through the oxidation of GSH to GSSH (35). The downregulation

of antioxidant system proteins, including SOD, CAT, and GPX, has been observed in both *in vivo* and *in vitro* studies of OA joints (36). When the production of ROS exceeds the scavenging capacity of the antioxidant system or the low activity of the antioxidant defense system, the cell is in a condition of oxidative stress which is characterized by an imbalance of oxidation and antioxidant state (37). The pathogenesis of numerous age-related disorders has been strongly associated with oxidative stress, which also serves as a pivotal contributor to the progression of OA (38–41).

The maintenance of cellular function and homeostasis necessitates a physiological level of ROS, however, excessive ROS induced by pathological processes can oxidize macromolecules such as mtDNA, genomic DNA, proteins, and lipids, thereby impairing essential cellular processes (Figure 1) (42-44). Investigations have documented that ROS-induced macromolecule compromise, including that of genomic, mtDNA, and lipids, results in synovitis worsening, extracellular matrix (ECM) degradation, and chondrocyte death, such as apoptosis and ferroptosis (45, 46). The increased level of ROS in cartilage and chondrocytes can be attributed to variations in oxygen pressure, mechanical stress, as well as the presence of inflammatory mediators such as IL-1, IL-17, and TNF- $\alpha$  (29, 47). The upregulation of ROS levels in the chondrocytes of individuals with OA have been demonstrated by numerous studies (45, 48). The most predominant ROS found in OA cartilages and chondrocytes are O2<sup>-</sup> and H2O2. Excessive generation of  $O_2^-$  can activate the transcription factor NF- $\kappa$ B, subsequently leading to elevated levels of cytokines, chemokines, and iNOS (49). Meanwhile, the cartilages and chondrocytes of individuals affected by OA also exhibit an excessive production of NO and its derivative (50). The anabolism of proteoglycans is hindered by abnormal levels of H<sub>2</sub>O<sub>2</sub> and NO, thereby impeding the production of cartilage matrix (51). In addition, studies have demonstrated that exposure of chondrocytes to pro-oxidants such as H<sub>2</sub>O<sub>2</sub>, tert-butyl hydroperoxide (TBHP), and menadione disrupts cellular redox equilibrium and induces oxidative stress, thereby leading to increased inflammation, apoptosis, and ferroptosis (52, 53). Moreover, oxidative stress accelerates telomere shortening and impairs chondrocyte replication capacity, thereby promoting chondrocyte senescence (39). A significant contributing factor to OA is the senescence of chondrocytes, which compromises the redox balance of mitochondria and leads to an increased production of ROS, which can result in the oxidation of genomic and mtDNA (54-56). Consequently, this oxidative damage can accelerate chondrocyte senescence and impede chondrocyte proliferation (57, 58). Taken collectively, these studies demonstrate that oxidative stress induced by excessive production of ROS under various adverse conditions promotes the degradation of cartilage, hinders ECM synthesis, and induces chondrocyte senescence and death. All these effects contribute to the progression of OA. Consequently, developing therapeutic interventions targeting detrimental ROS may hold promise for OA treatment.

# 2.2 Inflammation in OA

A fundamental defensive response to an infection stimulated by microorganisms or antigens is inflammation, which is mediated by



The excessive ROS caused by various pathologic processes can oxidize macromolecules including mitochondrial DNA (mtDNA), genomic, proteins, and lipids to accelerate OA.

the host immune system. The short-term, regulated inflammation contributes to tissue defense and repair, whereas the long-term, aberrant inflammation leads to tissue damage and cell death. The pathophysiology of several human diseases, such as diabetes, obesity, cancer, neurological diseases, and autoimmune diseases, are significantly influenced by chronic inflammation (59). It is well established that inflamed synovium is now recognized as a prevalent indicator of OA. To maintain the proper functioning of articular cartilage, the synovium produces synovial fluid containing hyaluronic acid and lubricin. It has been observed that patients in advanced stages of OA exhibit elevated levels of chemokines and proinflammatory cytokines in their synovial fluid (60, 61). The degeneration of cartilage and the exacerbation of synovitis are both attributed to the overexpression of prostaglandins, leukotrienes, chemokines, and cytokines in the synovium (62). Generally, chondrocytes are typically situated in an anaerobic environment, which helps maintain the articular cartilage in a state of low metabolic activity and limited turnover synthesis of ECM. However, under pathological conditions, chondrocytes overproduce chemokines and cytokines that enhance the levels of collagenases and aggrecanases, thereby disrupting the delicate balance between anabolism and catabolism in articular cartilage and leading to erosion of ECM (63). Further investigations have revealed that the elevation of cytokine levels in joints plays a pivotal

role in the pathogenesis of OA by regulating oxidative stress and

chondrocyte death (64). Consequently, targeting anti-inflammatory

strategies hold significant potential for the treatment of OA. The three most prominently expressed cytokines in patients with OA are IL-1, IL-6, and TNF- $\alpha$ , which are produced by macrophages, chondrocytes, and fibroblast-like synoviocytes, which play a significant role in the degenerative process of OA (29). Other cytokines, such as IL-17, IL-18, CXCL5, RANTES, and MCP1, have also been demonstrated to serve as key regulators in the pathogenesis of OA (65, 66). Intra-articular injections of either TNF- $\alpha$  or IL-1 into the knee joints have been demonstrated to expedite the progression of OA, with their combined effects further exacerbating this impact (29). The expression of catabolic genes such as COX-2, IL-6, iNOS, a disintegrin and metalloproteinase with thrombospondin motifs (ADAMTSs), and matrix metalloproteases (MMPs) was found upregulated in chondrocytes stimulated with IL-1 and TNF- $\alpha$ , while the expression of anabolic genes including collagen II and aggrecan was downregulated (67-69). The aberrant expression of iNOS induced by inflammatory cytokines enhances the expression of NO, thereby increasing the level of IL-1 $\beta$  and TNF- $\alpha$  to aggravate inflammation through activation of the NF- $\kappa$ B pathway (70). It is well established that MMPs and ADAMTSs are responsible for the degradation of collagen and aggrecan, respectively (71, 72). The proinflammatory cytokines TNF- $\alpha$  and IL-1 could inhibit the function of complex I, membrane potential, and lead to mtDNA damage, therefore contributing to mitochondrial dysfunction in human chondrocytes (28). It has been reported that the production of functionally compromised respiratory chain subunits was indued by mtDNA damage and mutations, thereby increasing the levels of ROS in chondrocytes (73). The impaired mitochondrial bioenergetics and increased inflammatory response ultimately contribute to chondrocyte death (73). Treatment of chondrocytes with inflammatory cytokines such as IL-1 and TNF- $\alpha$  led to a significantly elevated level of IL-6 and MMP-13 (74–76). Additionally, the administration of IL-6 through intra-articular injection in mouse knee joints promoted the destruction of articular cartilage (77). Therefore, these findings indicate that inflammatory cytokines are involved in perturbing the homeostasis of articular cartilage to involve the development of OA, and the stability of the inflammatory microenvironment is responsible for determining the function of joints.

# 2.3 Chondrocyte death in OA

Cell death plays a vital role in maintaining homeostasis and the developing of the body by eliminating senescent cells and shaping tissue during embryologic development. Additionally, cell death is an aberrant pathological phenomenon triggered by detrimental stimuli such as infections and injuries (78). The sole cell type found in articular cartilage, chondrocytes, are intricately embedded within the ECM and play a vital role in maintaining ECM homeostasis by regulating anabolic and catabolic processes, as well as repairing the damaged cartilages in OA. Therefore, the loss of chondrocytes may accelerate the remodeling of ECM, leading to abnormal structure of ECM and articular cartilage degeneration, thereby potentially hastening the progression of OA. Consequently, strategies to protect from the degeneration of articular cartilage can be developed by understanding the molecular mechanism of chondrocyte death. According to the regulation of involved processes, chondrocyte death can be categorized into nonprogrammed and programmed forms. Autophagy, pyroptosis, ferroptosis, and necroptosis are all examples of programmed cell death (PCD), while necrosis is a form of non-programmed cell death (non-PCD) that occurs due to chemical or physical stimulation under extreme conditions (79, 80).

Autophagy is a crucial cellular process responsible for the elimination of misfolded proteins, damaged organelles, and intracellular pathogens to maintain cellular homeostasis (81-83). Autophagy can be categorized into three distinct types, including macroautophagy, microautophagy, and chaperone-mediated autophagy. Macroautophagy, commonly known as autophagy, involves the formation of bilayer membranes derived from the endoplasmic reticulum (ER) and intracellular components that encapsulate proteins and organelles, and eventually fuse with lysosomes to form autophagolysosomes (84, 85). Lysosomes contain a high concentration of hydrolytic enzymes that are capable of breaking down various substrates, including damaged macromolecules and organelles. The autophagy process consists of several consecutive phases, namely initiation, phagophore or nucleation maturation, membrane elongation, sequestering the target substrate and autophagosome formation, lysosome fusion, and substrate degradation (86-88). The autophagy process is regulated by approximately 40 autophagy-related genes, with the majority of ATG functioning in complexes to regulate autophagy through various signaling pathways (86, 89). Autophagy serves as a defense mechanism for maintaining intracellular homeostasis,

operating at a basal level under normal conditions to eliminate agingrelated damaged organelles and misfolded proteins (90). Autophagy can also be triggered by extreme conditions, such as external pressure, limited nutrient availability, hypoxia, and endoplasmic reticulum stress (ERS). The upregulation of autophagy-related proteins, including Unc-51-like kinase 1 (ULK1), LC3, and beclin-1, has been confirmed in human chondrocytes, however, the levels of these proteins decline in the aging population (91). The insufficient level of autophagy fails to effectively eliminate damaged organelles and macromolecules, leading to the disruption of chondrocyte homeostasis and ultimately resulting in OA (92). Therefore, the age-related decline in autophagy is a contributing factor to the deterioration of articular chondrocytes, thereby being associated with the occurrence and progression of OA.

Apoptosis, a tightly regulated mechanism of cell death, is indispensable for maintaining tissue homeostasis and ensuring the proper functioning of the human body. Morphological characteristics associated with cell apoptosis include DNA fragmentation, chromatin condensation, cell shrinkage, membrane blistering, and the formation of apoptotic bodies (93). Previous studies have shown that chondrocyte apoptosis is related to articular cartilage degradation (94). The intrinsic mitochondrial pathway and the extrinsic death receptor pathway are two well-established signaling pathways for apoptosis (79). External stimuli induce an increase in mitochondrial membrane permeability, facilitating the release of apoptotic factors such as cytochrome C and procaspases into the cytoplasm, thereby triggering activation of the mitochondrial pathway (95, 96). Under normal circumstances, damaged or depolarized mitochondria are selectively eliminated through autophagy to prevent cellular damage caused by dysfunctional mitochondria, which is commonly referred to as mitophagy (97, 98). The insufficient clearance of dysfunctional mitochondria through mitophagy leads to the release of apoptotic factors into the cytoplasm and subsequent initiation of apoptosis. This process is further exacerbated by the excessive production of ROS (99). The chondrocytes exhibited impaired autophagy and excessive apoptosis during the later stages of OA. Moreover, the essential antiapoptotic proteins, such as Bcl2 and Bcl-XL, can suppress autophagy by binding to the key regulators of autophagy Beclin 1, thereby inhibiting the formation of the Beclin 1 complex. The apoptosis appears to be intricately linked with autophagy. The relationship between apoptosis and autophagy in chondrocytes remains incompletely understood, necessitating further investigation for confirmation.

The different forms of cell death are classified as lytic or non-lytic based on whether the cellular contents overflow upon cell death (100). Pyroptosis, also referred to as inflammatory necrosis, is a specific form of lytic cell death primarily triggered by diverse inflammasomes. These inflammasomes, such as the NLR family pyrin domain containing 3 (NLRP3), assemble in the cytosol and activate caspase to cleave gasdermins, generating membrane toxic enzymes that contribute to the formation of cell membrane perforation (101). The influx of water into the cytosol triggers a progressive swelling of cells, ultimately leading to membrane rupture. This event results in the release of cellular debris and cytokines, which not only impair neighboring cells but also exacerbate inflammation (102–104). Pyroptosis, similar to apoptosis, is a form of caspase-dependent PCD. Pyroptosis consists primarily of two pathways, including the non-canonical pathway and the canonical

inflammasome pathway (105). The non-canonical inflammasome pathway is mediated by caspases 4, 5, and 11, whereas the canonical inflammasome pathway is mediated by caspase-1. Pyroptosis has been implicated in the pathogenesis of various diseases, including respiratory, circulatory, digestive, and urinary tract disorders since its original proposal in 2001 (106-109). The involvement of chondrocyte pyroptosis in the pathogenesis of OA has been experimentally validated (110). In addition to being commonly associated with OA, obesity, age, and basic calcium phosphate (BCP) also possess the ability to activate the NLRP3 inflammasome, thereby triggering chondrocyte pyroptosis (80). The expression of pyroptosis-related inflammasomes is upregulated in the synovial fluid of individuals affected by OA. Moreover, overexpression of inflammasomes enhances the levels of inflammatory factors such as IL-1 $\beta$  and IL-18, both contributing to chondrocyte pyroptosis and inflammatory responses (80). Additionally, the suppression of OA deterioration can be achieved by inhibiting the NLRP3 inflammasome with CY-09 (111).

Initially proposed by Stockwell's team in 2012, ferroptosis represents a distinct form of PCD (112). In contrast to autophagy, apoptosis, and pyroptosis, ferroptosis is an irondependent PCD characterized by unique morphological features including mitochondrial structural disruption and accumulation of lipid peroxides (113). The distinguishing features of ferroptosis from other PCDs primarily lie in the morphological changes observed in mitochondria, such as reduction or disappearance of mitochondrial cristae, decrease in mitochondrial volume, and rupture of the outer membrane (114). Iron-ion plays a crucial role in the process of ferroptosis, as it facilitates the generation of abundant ROS through the Fenton reaction, consequently leading to the formation of lipid peroxides (115). The accumulation of lipid peroxides ultimately contributes to an increase in membrane permeability and subsequent cell membrane rupture, resulting in cell death. Under normal circumstances, the essential antioxidant defense system known as glutathione peroxidase 4 (GPX4) effectively prevents the buildup of lipid peroxides, thereby mitigating ferroptosis (53, 116). The level of iron ion in the cartilage synovial fluid of the OA group has been found to be significantly higher in vivo, while the level of GPX4 is lower compared to that in the normal group (53). Furthermore, ferroptosis can enhance the upregulation of MMP13 and downregulation of collagen II, thereby exacerbating ECM degradation (113). A growing body of studies has demonstrated that ferroptosis plays a significant role in the pathogenesis of OA (117, 118). The occurrence of other forms of cell death, such as cuproptosis, in addition to the previously discussed chondrocyte death, is also closely associated with the onset of OA (94, 119).

# 3 Melatonin targeting oxidative stress, inflammation, and chondrocytes death in OA

# 3.1 Melatonin

The fat-soluble indole hormone melatonin (N-acetyl-5methoxytryptamine) was initially isolated by Aaron B. Lerner and colleagues in 1958 (120). The synthesis of melatonin in mammals primarily occurs in the pineal gland, although it is also secreted by non-pineal cells and tissues such as lymphocytes, platelets, megakaryocytes, retina, ovary, testis, liver, and skin. These extrapineal sources of melatonin function in an autocrine or paracrine manner (121, 122). The production of melatonin exhibits a distinct circadian rhythm, being synthesized predominantly during the night and suppressed during the day (123). Melatonin is biosynthesized through a complex enzymatic pathway originating from the essential amino acid tryptophan under the catalytic action of a series of enzymes (124). Melatonin is rapidly delivered to its targeted cells or organelles via the bloodstream or cerebrospinal fluid upon production (125). Once integrated with the target, melatonin exerts a diverse range of physiological effects through both receptor-dependent and receptor-independent pathways (126, 127). The melatonin receptors 1 (MT1) and melatonin receptor 2 (MT2) are Gprotein-coupled receptors that are localized on both the mitochondria and the cell membrane. In addition, there is a cytosolic melatonin receptor 3 (MT3) found in several species but absent in humans. Furthermore, nuclear binding receptors such as retinoid acid-related orphan receptors (RORs)/RZR also function as receptors that melatonin targets (128, 129). The production of MT1 and MT2, which respectively regulate rapid eve movement sleep and non-rapid eye movement sleep, can be synchronized by melatonin in physiological sleep to regulate circadian rhythms (130). In addition to targeting MT1 and MT2 receptors for circadian rhythm modulation, melatonin also interacts with

TABLE 1 A list of reports studying the effect of melatonin on animal.

nuclear receptors such as RORs to modulate the circadian rhythm (131). Alongside regulating circadian rhythms, the binding of melatonin to MT1 and MT2 receptors enhances the expression of silent information regulator 1 (SIRT1) while inhibiting the phosphorylation of p38 and JNK MAPKs, thereby facilitating cell survival (132). Moreover, melatonin acts as an effective scavenger of free radicals by activating antioxidant enzymes and reducing the damaged cellular macromolecules and organelles through a receptor-independent pathway (133–135). The latest research has demonstrated that melatonin exerts a mitigating effect on inflammation, oxidative stress, and chondrocyte death in order to prevent cartilage destruction and further deterioration of OA, and the effects of melatonin on animals are listed in Table 1.

# 3.2 Melatonin as an inhibitor of oxidative stress

The hydrophilic and lipophilic properties of melatonin enable it to traverse all biological barriers, exerting an antioxidative impact on the cytosol, mitochondria, and cellular membrane (146, 147). Melatonin not only directly scavenges free radicals, but also enhances the activity of antioxidant enzymes such as SOD, CAT, and GPX to effectively inhibit oxidative stress (135, 148–150) (Figure 2). The nuclear factor-erythroid 2-related factor 2 (Nrf2) functions as a crucial transcription factor for antioxidant defense. Melatonin acts as an effective antioxidant, regulating the homeostasis of the cartilage matrix through the Nrf2 signaling

Animal Model		Treatment	Effect	Reference	
Twelve-week-old male Sprague-Dawley rats	DMM	Intraarticular injection 100 µL MT (10 mg/mL) for one month	Prevents Cartilage Degrada -tion in DMM- induced OA	Zhou et al. (136)	
Seven-week-old male Sprague-Dawley rats	DMM	Intraarticular injection 100 µL MT (10 mg/mL) once a week for twelve weeks.	Recharges of chondrocyte mitochondria to protect ca- rtilage matrix homeostasis	Zhang et al. (137)	
Sprague-Dawley rats (weight 210±20 g)	ACLT	Intraarticular injection 10 mg/kg or 20 mg/kg MT (10 mg/mL) once a wee for one month.	Inhibites matrix metallopr- oteinasesa in a concentration -dependent manner	Zhao et al. (138)	
Eight-week-old male Sprague-Dawley rats	Intraarticular injection collagenase	Subcutaneous injection 10 mg/kg MT (10 mg/mL) twice daily for one month	Downregulates the levels of MMP-13 and upregulates the expression of COL2A1.	Hong et al. (139)	
Five-month-old male Sprague–Dawley rats	ACLT	Intraperitoneal injection 20 mg/kg or 60 mg/kg MT (10 mg/mL) once daily for six weeks	Abolishes proinflammatory factor expression in a conc- entration -dependent manner	Liu et al. (140)	
Nine-week-old male C57BL/6J mice	DMM	Intraarticular injection 10 µL MT (10 mg/mL) twice a week for one month.	Prevents Cartilage Degradat -ion in DMM- induced OA	Zhang et al. (141)	
Six-seven month-old female New Zealand white rabbits	ACLT	Intraarticular injection 20 mg/kg MT weekly for one month.	Anti-inflammatory effects to ameliorate OA	Lim et al. (142)	
Eight-week-old female C57BL/6 mice	ACLT	Intraarticular injection 10 µL MT (50 mM) twice a week for eight weeks	Exerts protective effect on chondrocytes against inflammatory damage	Liang et al. (143)	
Eight-week-old male Sprague-Dawley rats	DMM	Intraperitoneal injection (15mg/k- g ) or (30 mg/kg) MT (10 mg/mL) every other day for eight weeks.	Anti-Apoptosis and Autop- hagy effects in a concentra- tion-dependent manner	Chen et al. (144)	
Ten-twelve week-old male C57/BL mice	ACLT	Intraperitoneal injection (50 mg/k- g) or (150 mg/ kg) MT (10 mg/mL) once a day for eight weeks	Attenuates mouse chondroc- yte apoptosis in a concen- tration-dependent manner	Qin et al. (145)	

MT, melatonin; OA, osteoarthritis; DMM, destabilization of the medial meniscus; ACLT, anterior cruciate ligament transection.



pathway. This is evidenced by the increased expression of Nrf2 in melatonin-treated chondrocytes, which led to a reduction of intracellular ROS levels and a significant elevation in the expression of SOD1, SOD2, CAT, and HO-1 (136). The expression of Nrf2 and antioxidant enzymes could be significantly inhibited by miR-146a, which was markedly elevated in OA chondrocyte. Moreover, overexpression of miR-146a reduced the level of Nrf2, thereby diminishing the protective effects of melatonin in articular cartilage of rats (136).

Mitochondria, the primary producers of ROS, serve as the key target organelles for melatonin in inhibiting oxidative damage. An in vitro showed that melatonin treatment restored mitochondrial homeostasis in OA chondrocytes by upregulating the expression of ATP, mtDNA, and respiratory chain factors such as CoxIV2, Sdha, Nd4, and Atp5a, thereby leading to a reduction in mitochondrial ROS levels and promotes an antioxidative effect (137) (Figure 2). The antioxidative benefits of melatonin, however, are compromised in mitochondrial homeostasis when the expression of SOD2 is inhibited, suggesting that SOD2 plays an essential role as a downstream component in mediating the protective effects of melatonin. Additionally, SIRT1, a histone deacetylase enzyme involved in nicotinamide adenine dinucleotide (NAD+) metabolism, is crucial for maintaining the activities of antioxidative enzymes (151, 152). Patients with OA who exhibited lower levels of SIRT1 showed an accelerated deterioration of articular cartilage, thereby suggesting that SIRT1 plays a protective role in the development of OA (153). The administration of melatonin significantly enhanced the expression of SIRT1, thereby promoting SOD2 activity and expression through

its involvement in histone deacetylation. In contrast, the inhibition of SIRT1 significantly diminished the protective effects of melatonin, suggesting that melatonin plays a crucial role in maintaining mitochondrial function to suppress oxidative stress by modulating the level of SIRT1 in OA progression (137). The effects of melatonin on OA through SIRT1 are listed in Table 2.

# 3.3 Melatonin as an inhibitor of inflammation

The pathophysiology of OA is primarily influenced by chronic inflammation, as indicated by a growing body of research (156, 157). IL-1 $\beta$  is commonly utilized as an *in vitro* model to simulate the inflammatory process of OA. The treatment with IL-1 $\beta$  induced upregulation of MMP-3, MMP-9, MMP-13, ADAMTS-4, COX-2 and iNOS levels, while downregulation of chondrogenic marker COL2A1 in human mesenchymal stem cells (hMSCs) and chondrocytes. However, melatonin significantly mitigated the detrimental effects caused by IL-1 $\beta$  (154, 155, 158). By inhibiting the JAK2/STAT3 signaling pathway, melatonin effectively reduced the levels of MMP-3, MMP-9, and MMP-13, thereby attenuating cartilage degradation (138) (Figure 3). Moreover, Ke et al. have demonstrated that melatonin suppressed the production of IL-1β, IL-6, and COX-2 to mitigate the progression of OA in rats (159). It has been reported that TNF-a inhibited extracellular matrix synthesis by upregulating the expression of catabolic enzymes and downregulating the expression of anabolic enzymes in chondrocytes. Melatonin could effectively downregulate the levels

TABLE 2 The effects of melatonin on OA via	SIRT1.
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Reference	Signaling pathway	Effect	
Lim et al. (142)	SIRT1/NF-ĸB	Modulate the anti- inflammatory effects	
Guo et al. (154)	SIRT1/NAMPT and NFAT5	Attenuate MMP-3 and MMP- 13 production	
Zhang et al. (137)	SIRT1/SOD2	Regain the chondrocyte mitochondrial function	
		Prevent chondrocyte matrix degradation	
Qin et al. (145)	SIRT1/IRE1α- XBP1- CHOP	Eliminate chondrocyte apoptosis	

of MMP-13 and upregulate the expression of COL2A1, thereby counteracting the inhibitory effect exerted by TNF- $\alpha$  on ECM (160) (Figure 3). Interestingly, this effect was further enhanced when combined with suitable exercise (160). The study conducted by Hong et al. demonstrated that the combination of melatonin and exercise treatment effectively suppressed abnormal catabolic upregulation, thereby reducing cartilage degradation (139). Besides, melatonin could directly bind to the MT1 receptor and thus inhibit the production of proinflammatory cytokines such as TNF- $\alpha$  and IL-8 in human OA synovial fibroblasts by antagonizing the PI3K/Akt and ERK signaling pathways, subsequently leading to an upregulation of miR-185a expression (140). It was found that melatonin enhanced the expression of miR-140, thereby abolishing IL-1 $\beta$ -induced matrix degradation in chondrocytes (141). In addition, melatonin could induce the upregulation of circRNA3503 to counteract the ECM degradation induced by TNF- $\alpha$  and IL-1 $\beta$  (161).

The NF-KB pathway plays a crucial role in orchestrating the expression of multiple proinflammatory cytokines, including IL-6, TNF-a, COX-2, and iNOS (162). Melatonin inhibited the activation of NF-KB stimulated by H<sub>2</sub>O<sub>2</sub> and also blocked the phosphorylation of upstream signaling pathways including JNK, p38 MAPK, ERK, PI3K, and Akt to improve the anti-inflammatory effects in chondrocytes (142). The inhibitory effects of melatonin on NF- $\kappa$ B and its upstream signaling pathways markedly are reversed by the downregulation of the SIRT1 level, in other words, the SIRT1 pathway participates in the cytoprotective and anti-inflammatory effects of melatonin via the inhibition of NF-κB signaling pathways on H<sub>2</sub>O<sub>2</sub>-induced articular cartilage destruction (142). Zhao et al. likewise testified that melatonin downregulates IL-1β-induced phosphorylation levels of P65 and IkBa in chondrocytes via SIRT1 pathways, thus abolishing NF-KB activation to function in cytoprotective and anti-inflammatory effects (155). It has been demonstrated that the toll-like receptor (TLR) mediates inflammatory responses triggered by chemical and physical



#### FIGURE 3

Melatonin inhibits inflammation through various signaling pathways to ameliorate the cartilage destruction in OA. ECM, extracellular matrix; MMP, matrix metalloprotease; ADAMTS, a disintegrin and metalloproteinase with thrombospondin motifs; MTR1, melatonin receptor 1; SIRT1, silent information regulator 1; TLR, toll-like receptor; COL2A1, collagen type II alpha-1.

stressors, such as cytokines and mechanical damage, ultimately leading to the development of OA (163). Hence, targeting the TLR signaling pathway may potentially serve as an efficacious therapeutic strategy for OA by attenuating the inflammatory damage. It was shown that melatonin exerted its protective effect on chondrocytes against inflammatory damage by inhibiting the TLR2/4-MyD88-NF- $\kappa$ B signaling pathway (143).

The expression of nicotinamide phosphoribosyltransferase (NAMPT), the rate-limiting enzyme in NAD+ biosynthesis, is enhanced by SIRT1 (164, 165). Activation of SIRT1 also promotes the synthesis of nuclear factor of activated T cells 5 (NFAT5), thereby enhancing the expression of pro-inflammatory cytokines in articular cartilage, including IL-1β, IL-6, TNF-α, COX-2, and iNOS (166, 167). Guo et al. demonstrated that melatonin significantly alleviated the expression of MMP-3 and MMP-13 induced by IL-1 $\beta$  in chondrocytes through the inhibition of SIRT1-mediated NAMPT and NFAT5 signaling pathways (154). Moreover, melatonin enhanced the expression of COL2A1 by regulating SIRT1, thereby restoring dexamethasone-induced ECM deterioration in chondrocytes (168). Several studies have shown that the synthesis of ECM and the differentiation, migration, and adhesion of chondrocytes were all significantly influenced by TGFβ1 (169, 170). Activation of the TGF-β1/Smad2 pathway stimulated by melatonin in IL-1B-induced chondrocytes was found contributing to the synthesis of ECM (155). It was suggested that melatonin administration in chondrocytes increased the upregulation of key chondrogenic marker genes, including Sox9, aggrecan, and collagen II via the TGF- $\beta$ 1 signaling pathway (171).

# 3.4 Melatonin as a modulator of chondrocyte death

As the sole cell type in cartilage, chondrocytes function as the core factor in regulating the homeostasis in cartilage metabolism (172). Previous studies have indicated that chondrocyte apoptosis plays a significant role in the development of OA (173, 174). The initiation of apoptosis is believed to occur as an early response to the depolarization of mitochondria, which impairs the mitochondrial membrane's potential. Substantial reductions in membrane potential promote permeabilization of the outer mitochondrial membrane, facilitating the release of apoptosis-related factors that trigger apoptosis (175, 176). Treatment with melatonin could restore the reduction of mitochondrial membrane potential and decrease the levels of caspase-3 and PARP, thereby ameliorating apoptosis in chondrocytes exposed to H<sub>2</sub>O<sub>2</sub> (144) (Figure 4). A key regulator of energy homeostasis, known as 5'-AMP-activated protein kinase (AMPK), is a serine/threonine kinase composed of multiple catalytic subunits ( $\alpha$ ,  $\beta$ , and  $\gamma$ ) (177). Multiple studies indicate that AMPK activation effectively inhibits apoptosis induced by the mitochondrial pathway through sustaining redox status and maintaining mitochondrial membrane potential, thereby restoring optimal mitochondrial function (178). The mammalian forked box transcription factor Class O (Foxo) family includes Foxo3, which functions as a downstream transcriptional factor in the AMPK signaling pathway and plays a crucial role in regulating antioxidant defenses and the autophagy process (179-181). Through the activation of AMPK/Foxo3 signaling pathways, melatonin exerted



#### FIGURE 4

Treatment of melatonin inhibits chondrocyte apoptosis and autophagy. MMP, matrix metalloprotease; ADAMTS, a disintegrin and metalloproteinase with thrombospondin motifs; SIRT1, silent information regulator 1; Foxo3, forked box transcription factor Class O 3; IRE1- $\alpha$ , inositol-requiring enzyme 1- $\alpha$ ; XBP1, X-box binding protein 1; CHOP, C/EBP homologous protein.

an inhibitory effect on apoptosis and induced upregulation of autophagy in chondrocytes to attenuate the progression of OA (144) (Figure 4).

In contrast to the internal mitochondrial pathway and extrinsic death receptor pathway, the ERS-mediated apoptosis pathway is initiated by the accumulation of misfolded proteins in the ER lumen, leading to ERS. The unfolded protein response (UPR) is a defensive mechanism that alleviates ERS and restores ER homeostasis (182, 183). However, if the ERS surpasses the threshold of UPR, it can trigger cellular apoptosis (184). UPR is initiated by transmembrane proteins, namely inositol-requiring enzyme 1-alpha (IRE1-alpha), protein kinase R-like ER kinase (PERK), and activating transcription factor 6 (ATF6) (185). The three primary signaling pathways in ERS-mediated apoptosis are IRE1α-X-box binding protein 1 (XBP1)-C/EBP homologous protein (CHOP), PERK-eukaryotic initiation factor 2a (eIF2a)-CHOP, and ATF6-XBP1-CHOP (186). The signaling pathway of IRE1α-XBP1-CHOP in chondrocyte apoptosis has been extensively investigated (187). The inhibition of the IRE1 $\alpha$ -XBP1-CHOP signaling pathway is considered a promising target for delaying the progression of OA by blocking chondrocyte apoptosis (187). It was shown that melatonin enhanced the expression of SIRT1, which suppressed the IRE1a-XBP1-CHOP signaling pathway, thereby attenuating ERS-induced apoptosis in chondrocytes (145).

The utilization of BMSCs presents a promising strategy for alleviating articular cartilage degradation, given the wide availability of resources for harvesting BMSCs and their capacity to differentiate into various cell lineages including chondrocytes and osteoblasts (188, 189). The potential of regenerating damaged articular cartilage through the chondrogenesis of BMSCs is appealing, however, the inflammatory environment in cases of OA poses challenges for the survival of BMSCs. It has been demonstrated that the administration of melatonin could reduce the expression of Bax in IL-1 $\beta$ -induced BMSCs, thereby conferring protection to BMSCs against IL-1 $\beta$ -triggered apoptosis (190). Moreover, melatonin was found to inhibit the expression of proapoptotic markers such as ADAMTS4, MMP9, and MMP13, thus rescuing IL-1 $\beta$ -induced apoptosis of BMSCs and impaired chondrogenesis through the NF- $\kappa$ B signaling pathway (158).

As the leading risk factor for the development of OA, aging can induce senescence-associated phenotypes in joints, such as increased levels of cytokines, MMPs, and ROS, and reduced expression of aggrecan and collagen II (191). Due to its buffering and lubricating properties, hyaluronic acid plays a significant protective role in mitigating mechanical stresses on articular cartilage, and its synthesis can be hindered by chondrocyte senescence and death. It was reported that melatonin could effectively downregulate the expression of senescence-related proteins p16, p21, and p-p65, and thus counteract chondrocyte senescence and the subsequent downregulation of hyaluronic acid triggered by D-galactose through activation of the SIRT1 signaling pathway (192). Ferroptosis and pyroptosis, as novel forms of PCD, have been found implicated in the pathogenesis of OA (80, 118). Although there is currently no research reporting the impact of melatonin on ferroptosis and pyroptosis of chondrocytes, the antioxidative properties and anti-inflammatory actions of melatonin suggest its potential role as a significant inhibitor of ferroptosis and pyroptosis in chondrocytes.

# 4 Melatonin as a desirable painrelieving drug in OA

OA is a primary contributor to chronic pain, significantly impacting the quality of life in individuals with OA. The exacerbation of chronic pain leads to sleep disorders, including reduced sleep efficiency and shortened total sleeping duration (193). The development of drugs to enhance the management of chronic pain in OA patients is therefore of utmost urgency. Numerous studies have suggested that melatonin exhibits analgesic effects in animal models of both acute and neuropathic pain (194-197). Several clinical trials have also confirmed the analgesic effect of melatonin in chronic pain conditions such as fibromyalgia, migraine headaches, and irritable bowel syndrome (198-200). Liu et al. demonstrated that the combination of melatonin and MT2 receptor yielded analgesic effects in rats with temporomandibular OA (201). The application of auricular acupressure has been found to enhance melatonin levels, thereby providing relief for chronic pain and addressing sleep disorders in elderly individuals with OA (202). The conventional therapeutic approaches for alleviating chronic pain, such as intra-articular steroid injections and oral nonsteroidal anti-inflammatory drugs (NSAIDs), are associated with undesirable side effects. For instance, long-term oral administration of NSAIDs can lead to gastritis and peptic ulcers, while repeated intra-articular steroid injections may result in decreased bone density and infection (203, 204). Significantly, melatonin to organs such as the liver and kidneys is associated with almost no toxicity and adverse effects (205). Collectively, the antioxidative, anti-inflammatory, and analgesic properties of melatonin render it a promising pharmaceutical agent for the treatment of OA.

# 5 Novel potential delivery systems of melatonin

The closed nature of the knee joint and the absence of blood vessels in the articular cartilage pose challenges for medications to accumulate within the joint via systemic circulation, leading to reduced efficacy and potential systemic adverse effects. Intraarticular administration is considered the optimal method for treating joint disorders, as it allows direct delivery of the drug to the articular cavity, thereby overcoming the aforementioned disadvantage. The frequent intra-articular injections, however, are invasive procedures that incur additional expenses, diminish patient adherence, and increase the risk of infection (206). The development of novel drug delivery systems that minimize the frequency of injections may hold the key to overcome the limitations of conventional intra-articular injection, which lacks long-term efficacy. Due to the challenge of finding suitable delivery carriers to arrive at the chondrocytes, therapeutic or preventative options for healing damaged articular cartilage in OA remains limited (207). For this reason, numerous researchers have devoted themselves to developing melatonin sustained release delivery systems for the treatment of OA (Figure 5). Up to now, several promising melatonin sustained release delivery systems have been successfully developed (Table 3).

# 5.1 Extracellular vesicles

Extracellular vesicles (EVs), which can be classified into three subtypes based on their size, namely apoptotic bodies, ectosomes, and exosomes, are proteolipid nanoparticles secreted by diverse cell types including bacteria, archaea, and eukaryotic cells. Apoptotic bodies, ranging in diameter from 800 to 5,000 nm, are generated through cellular shedding during the process of apoptosis. Conversely, ectosomes are formed by the plasma membrane via budding mechanisms and have a size range of 50 to 1000 nm. Additionally, exosomes (40–200 nm) are secreted from intracellular multivesicular bodies that merge with the cytoplasmic membrane. Exosomes play a crucial role in facilitating intercellular communication by transporting lipids, proteins, and various nucleic acids such as mRNAs, circular RNA, and miRNA (215). Recent studies have demonstrated the significant potential of exosomes derived from MSCs as nano-carriers for delivering therapeutic genetic materials and drugs (216). Compared to



#### FIGURE 5

Melatonin sustained release delivery systems for the treatment of OA. MT, melatonin; PLEL, poly (D, L-lactide)-poly (ethylene glycol)-poly (D, L-lactide); EVs, extracellular vesicles; NVs, nanovesicles; PLGA, polylactic-co-glycolic acid; PCL, polycaprolactone; HA, hyaluronic acid; CD, cyclodextrins; MIP-NSs, molecularly imprinted nanosponges; GeIMA-DOPA, gelatin methacryloyl-dopamine; DSPC, phosphatidylcholine; PNIPAM, poly (N-isopropyl acrylamide); GG, gellan gum; LGNF, lignocellulose nanofibrils; FS, forsterite.t of OA.

TABLE 3 The promising sustained release delivery systems of melatonin.

Materials	Delivery systems	Usage	Reference
Extracellular vesicles	PLEL-EVs@MT	injection	Tao et al. ( <mark>161</mark> )
	EVs mimicking NVs@MT	injection	Kim et al. (208)
Nanopolymer	PLGA@MT	injection	Liang et al. (143)
	Cotton fabrics-PCL@MT	transdermal patch	Massella et al. ( <mark>209</mark> )
Cyclodextrin	HA-CD@MT	injection	Zhang et al., 2022 (137)
	CD-based MIP-NSs@MT	transdermal patch	Hoti et al. (210)
Liposomes	GelMA- DOPA liposomes@MT	injection	Xiao et al. (211)
	DSPC liposomes@MT	injection	Ji et al. (212)
Hydrogel	PNIPAM/ HA hydrogels@MT	injection	Atoufi et al. (213)
	GG/LGNF/ FS hydrogel@MT	injection	Kouhi et al. (214)

MT, melatonin; PLEL, poly(D, L-lactide)-poly(ethylene glycol)-poly(D, L-lactide); EVs, extracellular vesicles; NVs, nanovesicles; PLGA, polylactic-co-glycolic acid; PCL, polycaprolactone; HA, hyaluronic acid; CD, cyclodextrins; MIP-NSs, molecularly imprinted nanosponges; GelMA-DOPA, gelatin methacryloyl-dopamine; DSPC, phosphatidylcholine; PNIPAM, poly(N-isopropyl acrylamide); GG, gellan gum; LGNF, lignocellulose nanofibrils; FS, forsterite.

MSCs, exosomes are non-viable, resulting in lower costs for storage and maintenance, as a viable state is needed in maintaining cells. Additionally, exosomes possess hypoimmunogenic properties and have a nano-scale size, which significantly reduce the likelihood of rejection (217, 218). Furthermore, exosomes possess the potential to traverse the blood-brain barrier, thereby facilitating the development of therapeutic interventions targeting the central nervous system (219). The versatility of exosomes allows for facile engineering to specifically target molecules. The current focus of numerous investigations lies in elucidating the mechanisms and functions of exosomes as efficacious drug delivery systems for various disorders. The ability of EVs to penetrate cartilage and target chondrocytes renders them as promising nano-carriers for therapeutic drugs in the treatment of OA (220). The evidence has demonstrated that EVs function as nano-carriers capable of delivering drugs to chondrocytes, thereby alleviating the progression of OA (161, 221). In conclusion, it is speculated that melatonin-loaded EVs can effectively penetrate articular cartilage and selectively target chondrocytes to attenuate the degeneration of articular cartilage by inhibiting oxidative stress, inflammation, and chondrocyte death.

The poly (D, L-lactide)-poly (ethylene glycol)-poly (D, L-lactide) (PDLLA-PEG-PDLLA; PLEL) triblock copolymer gels, which possess reversible, injectable, and thermosensitive properties, have been widely utilized in nano-drug delivery systems (222). The use of PLEL as a carrier for EVs has been employed to significantly enhance the sustained release of drugs loaded in EVs (161). The PLEL-entrapped EVs may offer promising delivery systems for achieving sustained release of melatonin, thereby presenting a potential therapeutic approach to alleviate the progression of OA. The limited production efficiency and laborious extraction and purification procedures of EVs, however, hinder the potential utilization of EVs in clinical practice. To address the challenges associated with EVs, EVs mimicking nanovesicles (NVs) which have similar biophysical characteristics to EVs have been generated via durably extruding cells through a microfilter (223, 224). EVs mimicking NVs are promising carriers which can be engineered to load with a variety of therapeutic drugs (225). Findings have demonstrated that melatonin-loaded EVs mimicking NVs effectively alleviated atopic dermatitis induced by 2,4-Dinitrofluorobenzene through the suppression of mast cell infiltration and local inflammation. Additionally, these EVs also promoted myocardial repair in cases of myocardial infarction by enhancing mitochondrial functions and reducing oxidative stress (208, 226). Taking together, it is very likely that the use of EVs mimicking NVs is a promising approach for delivering melatonin in the treatment of OA.

## 5.2 Nanopolymers

Nanopolymers possess exceptional mechanical properties, facile assembly, high biocompatibility, remarkable stability, scalability, and chemical modifiability, thereby offering favorable conditions for the design of nano-carrier delivery systems. Nanopolymers are broadly applied for the design of sustained release and site-specific drug delivery, resulting in improved therapeutic efficacy with fewer side effects (206, 227). Polylactic-co-glycolic acid (PLGA), a type of nanopolymer material, is widely employed in the field of drug delivery (228). The melatonin-loaded nano-delivery system was formed by encapsulating melatonin in PLGA, and the surface of PLGA was then modified with collagen II targeting polypeptides to enhance the targeting of such nanoparticles (143). What's exciting was that the sustained release of melatonin for at least 14 days in the mice joint cavity was achieved by this nanoparticle, significantly reducing the frequency of injections compared to using melatonin alone (143). The PLGA nanoparticles loaded with melatonin enable precise targeting of cartilage and sustained release, thereby reducing the degradation of cartilage and the progression of OA. Consequently, intra-articular injection of these nanoparticles may represent a novel therapeutic approach for OA treatment.

Cotton fabrics functionalized by polycaprolactone (PCL) nanoparticles are designed as a transdermal patch for the release of melatonin (209). The biodegradation process of PCL could last up to one year, making it a widely used delivery system for sustained drug release (229). Melatonin-loaded PCL nanoparticles, when distributed on cotton fibers, exhibited a controlled and sustained release of melatonin (209). This transdermal delivery system significantly enhanced the skin permeation and sustained release of melatonin through a non-invasive approach. In conclusion, it is

speculated that transdermal delivery systems may hold great potential in the treatment of OA.

## 5.3 Cyclodextrins

Cyclodextrins (CDs), which are cyclic oligosaccharides derived from starch hydrolysis, possess internal hydrophobicity and external hydrophilicity (230). Due to their unique characteristics, CDs have the ability to form host-guest complexes with suitable molecules to improve their stability, bioavailability, solubility, and controlled release. The CDs-based drug delivery system has facilitated the sustained release of the medication for OA treatment (231). The CD was incorporated into the hyaluronic acid (HA) solution to construct the HA-CD drug delivery system. Subsequently, melatonin was integrated into the HA-CD-based drug delivery system. The HA-CD melatonin delivery system achieved sustained release of melatonin over an extended period, effectively repairing dysfunctional mitochondria in OA chondrocytes (137). Furthermore, CDs could undergo polymerization with the cross-linking agent citric acid, followed by the addition of melatonin as a template molecule to form CDbased molecularly imprinted nanosponges (MIP-NSs) (210). Alongside these, CD-based MIP-NSs were incorporated into cream formulations to enhance their direct applicability to the skin. These skin formulations offered an innovative transdermal delivery system that enhanced the permeation of the skin and improved the sustained release of melatonin (210). The transdermal delivery systems of CD-based MIP-NSs provided a more advanced approach for delivering melatonin into the skin compared to traditional methods, such as intra-articular injection, thereby avoiding undesirable effects.

## 5.4 Liposomes

Liposomes have been extensively employed as drug delivery systems due to their exceptional biocompatibility and proficient capacity to regulate drug release (232, 233). Due to its excellent biocompatibility and strong adhesive properties, gelatin methacryloyl-dopamine (GelMA-DOPA) is widely used in the field of bone tissue engineering (234). The liposomes loaded with melatonin were combined with a GelMA-DOPA solution to fabricate the melatonin delivery system. The GelMA-DOPA liposomes delivery system was advantageous for regulating the sustained release of melatonin (211). The GelMA-DOPA liposomes delivery system for melatonin, although its application is limited to osteoporosis therapy right now, holds significant potential for alleviating OA. In addition, phosphatidylcholine (DSPC) liposomes could be employed as highly effective lubricants to reduce friction, in addition to their role as drug delivery systems (235). It was reported that the utilization of DSPC liposomes as a carrier for glucosamine sulphate enabled the delivery of effective boundary lubrication at the outermost layer of the joint while also facilitating the controlled and sustained release of glucosamine sulphate (212). As a result, DSPC liposomes may offer a more direct and practical approach for the intra-articular administration of melatonin in OA treatment.

## 5.5 Hydrogel

Hydrogel, a type of polymeric material, is extensively investigated in tissue engineering due to its exceptional biocompatibility, predictable degradation rate, appropriate elasticity, porous structure, and resemblance to the ECM (236, 237). The thermosensitive injectable hydrogel, poly (N-isopropyl acrylamide) (PNIPAM), has gained significant attention due to its ability to be directly injected into the injured area and effectively fill irregular flaws (238). PNIPAM has minimal cell adhesiveness and bioactivity despite its significant ability to replicate the architecture of some tissues. Accordingly, the combination of in situ injectable hydrogels with cells and bioactive compounds has garnered significant attention in the field of bone/cartilage tissue regeneration (239). The lubricating polysaccharide hyaluronic acid facilitated cellular adhesion, migration, and proliferation, thereby decreasing syneresis and hydrogel shrinkage (240). Various studies have shown that surface modification of PLGA with chitosan-g-acrylic acid (PLGA-ACH) can enhance adaptability, mucoadhesive properties, and regulate drug release (241, 242). The addition of PLGA-ACH particles as crosslinkers to PNIPAM enhanced the mechanical properties of PNIPAM, resulting in a closer resemblance to natural cartilage tissue (213). Simultaneously, the PLGA core acted as a carrier for the sustained release of melatonin (213). A previous study has demonstrated the efficacy of melatonin as a delivery system for cartilage tissue engineering, wherein injectable PNIPAM/hyaluronic acid hydrogels containing PLGA-ACH nanoparticles were applicated (213).

The low immunogenicity, cost-effectiveness, and ease of handling make Gellan gum (GG), which is composed of glucose, rhamnose, and D-glucuronate residues, particularly attractive for drug delivery (243). The GG-based hydrogel is used in cartilage regeneration due to its appealing characteristics, including noncytotoxicity, biocompatibility, mild processing conditions, and structural resemblance to native glycosaminoglycans (237, 243). However, similar to other biodegradable hydrogels, it lacks the necessary mechanical strength and bioactivity required for reinforcement through nanoscale additions. Lignocellulose nanofibrils (LGNF), characterized by their high modulus, reactive surfaces, and large aspect ratio, present an ideal material for enhancing both the mechanical and biological properties of polymeric composites (244). The porous nanoparticle form of forsterite (FS), a crystalline member of the olivine family composed of magnesia and silicon, has been investigated for its potential as a sustained drug delivery system (245). Accordingly, in order to enhance the mechanical properties of GG-based hydrogel, LGNF and FS nanoparticles were incorporated, and thus an injectable delivery system based on GG/LGNF/FS hydrogel has been developed for sustained release of melatonin and articular cartilage repair (214).

# 6 Conclusions and future directions

Increasing evidence suggests that inflammation, oxidative stress, and chondrocyte death are closely linked to the severity and progression of OA, rendering them potential targets for OA treatment. The present review provides a comprehensive overview of the role of melatonin in modulating inflammation, oxidative stress, and chondrocyte death to attenuate OA progression through the regulation of various signaling pathways including SIRT1, Nrf2, NF-κB, JAK2/STAT3, TGF-β1/Smad2, AMPK/Foxo3, IRE1α-XBP1-CHOP, PI3K/Akt and ERK. Obviously, numerous signaling pathways are implicated in the potential mechanism of melatonin in the treatment of OA. The primary source of ROS leading to chondrocyte death and exacerbating inflammatory reactions, thereby aggravating articular cartilage degradation, is mitochondrial dysfunction. The melatonin-based treatment restores impaired mitochondrial functions by recovering reductions in membrane potential and enhancing the synthesis of ATP, mtDNA, and respiratory chain factors to alleviate oxidative stress and chondrocyte death. Although melatonin is considered as an effective antioxidant for maintaining mitochondria, yet the current research in this field still remains insufficient. Therefore, future studies should aim to comprehensively and profoundly investigate the interplay between melatonin and signaling pathways on mitochondria in OA chondrocytes.

The efficacy of EVs as a well-researched carrier has been demonstrated in the administration of melatonin for various disorders. However, there is a lack of direct research to substantiate the efficacy of melatonin-loaded EVs in the treatment of OA. The efficacy of melatonin-loaded EVs will be validated in future studies. In addition, various innovative bioactive materials, including nanopolymers, cyclodextrins, liposomes, and hydrogels, have been developed to ensure sustained release of melatonin and target articular cartilage. The use of novel carriers for intra-articular injection can effectively reduce the injection frequency, thereby optimizing the therapeutic efficacy and bioavailability of melatonin in the treatment of OA. Therefore, these biomaterials play an indispensable role in advancing the potential clinical efficacy of melatonin. In summary, combined melatonin with multiple bioactive agents holds great promise as a strategy for OA treatment.

# Author contributions

ZX: Writing – original draft, Data curation, Methodology, Software, Visualization. GP: Data curation, Methodology, Formal analysis, Funding acquisition, Investigation, Validation, Writing – review & editing. JD: Formal analysis, Funding acquisition, Methodology, Writing – review & editing. ML: Writing – review & editing, Investigation, Project administration, Supervision. XN: Project administration, Supervision, Writing – review & editing, Methodology, Resources. YZ: Methodology, Project administration, Supervision, Writing – review & editing, Conceptualization, Formal analysis. HY: Conceptualization, Project administration, Writing – review & editing, Funding acquisition, Investigation, Resources. HS: Conceptualization, Funding acquisition, Investigation, Project administration, Writing – review & editing, Formal analysis, Methodology.

# Funding

The author(s) declare financial support was received for the research, authorship, and/or publication of this article. The study was funded by the National Natural Science Foundation of China (82360420, 82260372), Science and Technology Fund of Guizhou Science and Technology Department (QKH-ZK [2021] 391; QKH-ZK [2023] 344), Science and Technology Fund of Guizhou Provincial Health Commission (gzwjkj2020-1-120; gzwkj2021-261), the Youth Fund cultivation program of National Natural Science Foundation of Affiliated Hospital of Guizhou Medical University (gyfynsfc-2021-12), and Graduate Scientific Research Fund project of Guizhou (YJSKYJJ [2021]157).

# Acknowledgments

The authors would like to express their gratitude to Professor Xianwen Shang in the Affiliated Hospital of Guizhou Medical University for his generous assistance and guidance in writing. All figures were authorized and created in BioRender.com.

# **Conflict of interest**

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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# References

1. Kaspiris A, Hadjimichael AC, Lianou I, Iliopoulos ID, Ntourantonis D, Melissaridou D, et al. Subchondral bone cyst development in osteoarthritis: from pathophysiology to bone microarchitecture changes and clinical implementations. *J Clin Med* (2023) 12(3):815. doi: 10.3390/jcm12030815

2. Yao Q, Wu X, Tao C, Gong W, Chen M, Qu M, et al. Osteoarthritis: pathogenic signaling pathways and therapeutic targets. *Signal Transduct Target Ther* (2023) 8 (1):56. doi: 10.1038/s41392-023-01330-w

3. Chen D. Osteoarthritis: A complicated joint disease requiring extensive studies with multiple approaches. *J Orthop Translat* (2022) 32:130. doi: 10.1016/j.jot.2022.02.009

 Prieto-Alhambra D, Judge A, Javaid MK, Cooper C, Diez-Perez A, Arden NK. Incidence and risk factors for clinically diagnosed knee, hip and hand osteoarthritis: influences of age, gender and osteoarthritis affecting other joints. *Ann Rheum Dis* (2014) 73(9):1659–64. doi: 10.1136/annrheumdis-2013-203355

5. Wang Y, Nguyen UDT, Lane NE, Lu N, Wei J, Lei G, et al. Knee osteoarthritis, potential mediators, and risk of all-cause mortality: data from the osteoarthritis initiative. *Arthritis Care Res (Hoboken)* (2021) 73(4):566–73. doi: 10.1002/acr.24151

6. Sun AR, Udduttula A, Li J, Liu Y, Ren PG, Zhang P. Cartilage tissue engineering for obesity-induced osteoarthritis: Physiology, challenges, and future prospects. *J* Orthop Translat (2021) 26:3–15. doi: 10.1016/j.jot.2020.07.004

7. Hunter DJ, March L, Chew M. Osteoarthritis in 2020 and beyond: a lancet commission. Lancet (2020) 396(10264):1711-2. doi: 10.1016/S0140-6736(20)32230-3

 Hawker GA, King LK. The burden of osteoarthritis in older adults. Clin Geriatr Med (2022) 38(2):181–92. doi: 10.1016/i.cger.2021.11.005

9. Wen C, Xiao G. Advances in osteoarthritis research in 2021 and beyond. J Orthop Translat (2022) 32:A1–A2. doi: 10.1016/j.jot.2022.02.011

10. Nelson AE. Osteoarthritis year in review 2017: clinical. Osteoarthritis Cartilage (2018) 26(3):319–25. doi: 10.1016/j.joca.2017.11.014

11. Li J, Zhang H, Han Y, Hu Y, Geng Z, Su J. Targeted and responsive biomaterials in osteoarthritis. *Theranostics* (2023) 13(3):931-54. doi: 10.7150/thno.78639

12. Eun Y, Yoo JE, Han K, Kim D, Lee KN, Lee J, et al. Female reproductive factors and risk of joint replacement arthroplasty of the knee and hip due to osteoarthritis in postmenopausal women: a nationwide cohort study of 1.13 million women. *Osteoarthritis Cartilage* (2022) 30(1):69–80. doi: 10.1016/j.joca.2021.10.012

13. Paget LDA, Reurink G, de Vos RJ, Weir A, Moen MH, Bierma-Zeinstra SMA, et al. Effect of platelet-rich plasma injections vs placebo on ankle symptoms and function in patients with ankle osteoarthritis: A randomized clinical trial. *Jama* (2021) 326(16):1595–605. doi: 10.1001/jama.2021.16602

14. Schuette HB, Kraeutler MJ, Schrock JB, McCarty EC. Primary autologous chondrocyte implantation of the knee versus autologous chondrocyte implantation after failed marrow stimulation: A systematic review. *Am J Sports Med* (2021) 49 (9):2536–41. doi: 10.1177/0363546520968284

15. Tan SHS, Kwan YT, Neo WJ, Chong JY, Kuek TYJ, See JZF, et al. Intra-articular injections of mesenchymal stem cells without adjuvant therapies for knee osteoarthritis: A systematic review and meta-analysis. *Am J Sports Med* (2021) 49(11):3113–24. doi: 10.1177/0363546520981704

16. Cho Y, Jeong S, Kim H, Kang D, Lee J, Kang SB, et al. Disease-modifying therapeutic strategies in osteoarthritis: current status and future directions. *Exp Mol Med* (2021) 53(11):1689–96. doi: 10.1038/s12276-021-00710-y

17. Luchetti F, Canonico B, Bartolini D, Arcangeletti M, Ciffolilli S, Murdolo G, et al. Melatonin regulates mesenchymal stem cell differentiation: a review. *J Pineal Res* (2014) 56(4):382–97. doi: 10.1111/jpi.12133

18. Repova K, Baka T, Krajcirovicova K, Stanko P, Aziriova S, Reiter RJ, et al. Melatonin as a potential approach to anxiety treatment. *Int J Mol Sci* (2022) 23 (24):16187. doi: 10.3390/ijms232416187

19. Stauch B, Johansson LC, McCorvy JD, Patel N, Han GW, Huang XP, et al. Structural basis of ligand recognition at the human MT(1) melatonin receptor. *Nature* (2019) 569(7755):284–8. doi: 10.1038/s41586-019-1141-3

20. Hardeland R. Aging, melatonin, and the pro- and anti-inflammatory networks. Int J Mol Sci (2019) 20(5):1223. doi: 10.3390/ijms20051223

21. Galano A, Reiter RJ. Melatonin and its metabolites vs oxidative stress: From individual actions to collective protection. *J Pineal Res* (2018) 65(1):e12514. doi: 10.1111/jpi.12514

22. Zhang Y, Liu T, Yang H, He F, Zhu X. Melatonin: A novel candidate for the treatment of osteoarthritis. *Ageing Res Rev* (2022) 78:101635. doi: 10.1016/j.arr.2022.101635

23. Harpsøe NG, Andersen LP, Gögenur I, Rosenberg J. Clinical pharmacokinetics of melatonin: a systematic review. *Eur J Clin Pharmacol* (2015) 71(8):901–9. doi: 10.1007/s00228-015-1873-4

24. Rahal A, Kumar A, Singh V, Yadav B, Tiwari R, Chakraborty S, et al. Oxidative stress, prooxidants, and antioxidants: the interplay. *BioMed Res Int* (2014) 2014:761264. doi: 10.1155/2014/761264

25. Lismont C, Nordgren M, Van Veldhoven PP, Fransen M. Redox interplay between mitochondria and peroxisomes. *Front Cell Dev Biol* (2015) 3:35. doi: 10.3389/ fcell.2015.00035

26. Wallace DC. A mitochondrial bioenergetic etiology of disease. J Clin Invest (2013) 123(4):1405-12. doi: 10.1172/JCI61398

27. Lepetsos P, Papavassiliou AG. ROS/oxidative stress signaling in osteoarthritis. Biochim Biophys Acta (2016) 1862(4):576-91. doi: 10.1016/j.bbadis.2016.01.003

28. Blanco FJ, Rego I, Ruiz-Romero C. The role of mitochondria in osteoarthritis. Nat Rev Rheumatol (2011) 7(3):161–9. doi: 10.1038/nrrheum.2010.213

29. Ansari MY, Ahmad N, Haqqi TM. Oxidative stress and inflammation in osteoarthritis pathogenesis: Role of polyphenols. *BioMed Pharmacother* (2020) 129:110452. doi: 10.1016/j.biopha.2020.110452

30. Chen B, Deng Y, Tan Y, Qin J, Chen LB. Association between severity of knee osteoarthritis and serum and synovial fluid interleukin 17 concentrations. *J Int Med Res* (2014) 42(1):138–44. doi: 10.1177/0300060513501751

31. Ahmed S, Rahman A, Hasnain A, Lalonde M, Goldberg VM, Haqqi TM. Green tea polyphenol epigallocatechin-3-gallate inhibits the IL-1 beta-induced activity and expression of cyclooxygenase-2 and nitric oxide synthase-2 in human chondrocytes. *Free Radic Biol Med* (2002) 33(8):1097–105. doi: 10.1016/S0891-5849(02)01004-3

32. Lei XG, Zhu JH, Cheng WH, Bao Y, Ho YS, Reddi AR, et al. Paradoxical roles of antioxidant enzymes: basic mechanisms and health implications. *Physiol Rev* (2016) 96 (1):307–64. doi: 10.1152/physrev.00010.2014

33. Fukui M, Zhu BT. Mitochondrial superoxide dismutase SOD2, but not cytosolic SOD1, plays a critical role in protection against glutamate-induced oxidative stress and cell death in HT22 neuronal cells. *Free Radic Biol Med* (2010) 48(6):821–30. doi: 10.1016/j.freeradbiomed.2009.12.024

34. Rhee SG, Woo HA, Kil IS, Bae SH. Peroxiredoxin functions as a peroxidase and a regulator and sensor of local peroxides. *J Biol Chem* (2012) 287(7):4403–10. doi: 10.1074/jbc.R111.283432

35. Lubos E, Loscalzo J, Handy DE. Glutathione peroxidase-1 in health and disease: from molecular mechanisms to therapeutic opportunities. *Antioxid Redox Signal* (2011) 15(7):1957–97. doi: 10.1089/ars.2010.3586

36. Regan EA, Bowler RP, Crapo JD. Joint fluid antioxidants are decreased in osteoarthritic joints compared to joints with macroscopically intact cartilage and subacute injury. *Osteoarthritis Cartilage* (2008) 16(4):515–21. doi: 10.1016/j.joca.2007.09.001

37. Varesi A, Chirumbolo S, Campagnoli LIM, Pierella E, Piccini GB, Carrara A, et al. The role of antioxidants in the interplay between oxidative stress and senescence. *Antioxidants (Basel)* (2022) 11(7):1224. doi: 10.3390/antiox11071224

38. Yu H, Zhang Z, Wei F, Hou G, You Y, Wang X, et al. Hydroxytyrosol ameliorates intervertebral disc degeneration and neuropathic pain by reducing oxidative stress and inflammation. *Oxid Med Cell Longev* (2022) 2022:2240894. doi: 10.1155/2022/2240894

39. Liu L, Luo P, Yang M, Wang J, Hou W, Xu P. The role of oxidative stress in the development of knee osteoarthritis: A comprehensive research review. *Front Mol Biosci* (2022) 9:1001212. doi: 10.3389/fmolb.2022.1001212

40. Portal-Núñez S, Esbrit P, Alcaraz MJ, Largo R. Oxidative stress, autophagy, epigenetic changes and regulation by miRNAs as potential therapeutic targets in osteoarthritis. *Biochem Pharmacol* (2016) 108:1–10. doi: 10.1016/j.bcp.2015.12.012

41. Hui W, Young DA, Rowan AD, Xu X, Cawston TE, Proctor CJ. Oxidative changes and signalling pathways are pivotal in initiating age-related changes in articular cartilage. *Ann Rheum Dis* (2016) 75(2):449–58. doi: 10.1136/annrheumdis-2014-206295

42. Juan CA, Pérez de la Lastra JM, Plou FJ, Pérez-Lebeña E. The chemistry of reactive oxygen species (ROS) revisited: outlining their role in biological macromolecules (DNA, lipids and proteins) and induced pathologies. *Int J Mol Sci* (2021) 22(9):4642. doi: 10.3390/ijms22094642

43. Capozzi A, Saucier C, Bisbal C, Lambert K. Grape polyphenols in the treatment of human skeletal muscle damage due to inflammation and oxidative stress during obesity and aging: early outcomes and promises. *Molecules* (2022) 27(19):6594. doi: 10.3390/molecules27196594

44. Kinnula VL, Fattman CL, Tan RJ, Oury TD. Oxidative stress in pulmonary fibrosis: a possible role for redox modulatory therapy. *Am J Respir Crit Care Med* (2005) 172(4):417–22. doi: 10.1164/rccm.200501-017PP

45. Marchev AS, Dimitrova PA, Burns AJ, Kostov RV, Dinkova-Kostova AT, Georgiev MI. Oxidative stress and chronic inflammation in osteoarthritis: can NRF2 counteract these partners in crime? *Ann N Y Acad Sci* (2017) 1401(1):114–35. doi: 10.1111/nyas.13407

46. Liang D, Minikes AM, Jiang X. Ferroptosis at the intersection of lipid metabolism and cellular signaling. *Mol Cell* (2022) 82(12):2215-27. doi: 10.1016/j.molcel.2022.03.022

47. Koike M, Nojiri H, Ozawa Y, Watanabe K, Muramatsu Y, Kaneko H, et al. Mechanical overloading causes mitochondrial superoxide and SOD2 imbalance in chondrocytes resulting in cartilage degeneration. *Sci Rep* (2015) 5:11722. doi: 10.1038/srep11722

48. Coryell PR, Diekman BO, Loeser RF. Mechanisms and therapeutic implications of cellular senescence in osteoarthritis. *Nat Rev Rheumatol* (2021) 17(1):47–57. doi: 10.1038/s41584-020-00533-7

49. Chung HY, Cesari M, Anton S, Marzetti E, Giovannini S, Seo AY, et al. Molecular inflammation: underpinnings of aging and age-related diseases. *Ageing Res Rev* (2009) 8(1):18–30. doi: 10.1016/j.arr.2008.07.002

50. Ahmad N, Ansari MY, Bano S, Haqqi TM. Imperatorin suppresses IL-1 $\beta$ -induced iNOS expression via inhibiting ERK-MAPK/AP1 signaling in primary human OA chondrocytes. Int Immunopharmacol (2020) 85:106612. doi: 10.1016/j.intimp.2020.106612

51. Bolduc JA, Collins JA, Loeser RF. Reactive oxygen species, aging and articular cartilage homeostasis. *Free Radic Biol Med* (2019) 132:73-82. doi: 10.1016/j.freeradbiomed.2018.08.038

52. Collins JA, Wood ST, Nelson KJ, Rowe MA, Carlson CS, Chubinskaya S, et al. Oxidative stress promotes peroxiredoxin hyperoxidation and attenuates pro-survival signaling in aging chondrocytes. *J Biol Chem* (2016) 291(13):6641–54. doi: 10.1074/ jbc.M115.693523

53. Miao Y, Chen Y, Xue F, Liu K, Zhu B, Gao J, et al. Contribution of ferroptosis and GPX4's dual functions to osteoarthritis progression. *EBioMedicine* (2022) 76:103847. doi: 10.1016/j.ebiom.2022.103847

54. Xie J, Wang Y, Lu L, Liu L, Yu X, Pei F. Cellular senescence in knee osteoarthritis: molecular mechanisms and therapeutic implications. *Ageing Res Rev* (2021) 70:101413. doi: 10.1016/j.arr.2021.101413

55. Rim YA, Nam Y, Ju JH. The role of chondrocyte hypertrophy and senescence in osteoarthritis initiation and progression. *Int J Mol Sci* (2020) 21(7):2358. doi: 10.3390/ ijms21072358

56. Astrike-Davis EM, Coryell P, Loeser RF. Targeting cellular senescence as a novel treatment for osteoarthritis. *Curr Opin Pharmacol* (2022) 64:102213. doi: 10.1016/j.coph.2022.102213

57. McCulloch K, Litherland GJ, Rai TS. Cellular senescence in osteoarthritis pathology. *Aging Cell* (2017) 16(2):210-8. doi: 10.1111/acel.12562

58. Minguzzi M, Cetrullo S, D'Adamo S, Silvestri Y, Flamigni F, Borzì RM. Emerging players at the intersection of chondrocyte loss of maturational arrest, oxidative stress, senescence and low-grade inflammation in osteoarthritis. *Oxid Med Cell Longev* (2018) 2018:3075293. doi: 10.1155/2018/3075293

59. Straub RH, Schradin C. Chronic inflammatory systemic diseases: An evolutionary trade-off between acutely beneficial but chronically harmful programs. *Evol Med Public Health* (2016) 2016(1):37–51. doi: 10.1093/emph/eow001

60. Shen J, Abu-Amer Y, O'Keefe RJ, McAlinden A. Inflammation and epigenetic regulation in osteoarthritis. *Connect Tissue Res* (2017) 58(1):49-63. doi: 10.1080/03008207.2016.1208655

61. Griffin TM, Scanzello CR. Innate inflammation and synovial macrophages in osteoarthritis pathophysiology. *Clin Exp Rheumatol* (2019) 37 Suppl 120(5):57–63.

62. Scanzello CR, Goldring SR. The role of synovitis in osteoarthritis pathogenesis. Bone (2012) 51(2):249–57. doi: 10.1016/j.bone.2012.02.012

63. Khan NM, Haseeb A, Ansari MY, Devarapalli P, Haynie S, Haqqi TM. Wogonin, a plant derived small molecule, exerts potent anti-inflammatory and chondroprotective effects through the activation of ROS/ERK/Nrf2 signaling pathways in human Osteoarthritis chondrocytes. *Free Radic Biol Med* (2017) 106:288–301. doi: 10.1016/j.freeradbiomed.2017.02.041

64. Kapoor M, Martel-Pelletier J, Lajeunesse D, Pelletier JP, Fahmi H. Role of proinflammatory cytokines in the pathophysiology of osteoarthritis. *Nat Rev Rheumatol* (2011) 7(1):33-42. doi: 10.1038/nrrheum.2010.196

65. Khella CM, Horvath JM, Asgarian R, Rolauffs B, Hart ML. Anti-inflammatory therapeutic approaches to prevent or delay post-traumatic osteoarthritis (PTOA) of the knee joint with a focus on sustained delivery approaches. *Int J Mol Sci* (2021) 22 (15):8005. doi: 10.3390/ijms22158005

66. Jiang Y, Xiao Q, Hu Z, Pu B, Shu J, Yang Q, et al. Tissue levels of leukemia inhibitory factor vary by osteoarthritis grade. *Orthopedics* (2014) 37(5):e460–464. doi: 10.3928/01477447-20140430-57

67. Ansari MY, Haqqi TM. Interleukin-1 $\beta$  induced Stress Granules Sequester COX-2 mRNA and Regulates its Stability and Translation in Human OA Chondrocytes. *Sci Rep* (2016) 6:27611. doi: 10.1038/srep27611

68. Haseeb A, Chen D, Haqqi TM. Delphinidin inhibits IL-1 $\beta$ -induced activation of NF- $\kappa$ B by modulating the phosphorylation of IRAK-1(Ser376) in human articular chondrocytes. *Rheumatol (Oxford)* (2013) 52(6):998–1008. doi: 10.1093/rheumatology/kes363

69. Khan NM, Ansari MY, Haqqi TM. Sucrose, but not glucose, blocks IL1-βinduced inflammatory response in human chondrocytes by inducing autophagy *via* AKT/mTOR pathway. *J Cell Biochem* (2017) 118(3):629–39. doi: 10.1002/jcb.25750

70. Ahmad N, Ansari MY, Haqqi TM. Role of iNOS in osteoarthritis: Pathological and therapeutic aspects. J Cell Physiol (2020) 235(10):6366–76. doi: 10.1002/jcp.29607

71. Laronha H, Caldeira J. Structure and function of human matrix metalloproteinases. *Cells* (2020) 9(5):1076. doi: 10.3390/cells9051076

72. Shiomi T, Lemaître V, D'Armiento J, Okada Y. Matrix metalloproteinases, a disintegrin and metalloproteinases, and a disintegrin and metalloproteinases with thrombospondin motifs in non-neoplastic diseases. *Pathol Int* (2010) 60(7):477–96. doi: 10.1111/j.1440-1827.2010.02547.x

73. Reed KN, Wilson G, Pearsall A, Grishko VI. The role of mitochondrial reactive oxygen species in cartilage matrix destruction. *Mol Cell Biochem* (2014) 397(1-2):195–201. doi: 10.1007/s11010-014-2187-z

74. Ansari MY, Ahmad N, Haqqi TM. Butein activates autophagy through AMPK/ TSC2/ULK1/mTOR pathway to inhibit IL-6 expression in IL-1 $\beta$  Stimulated human chondrocytes. *Cell Physiol Biochem* (2018) 49(3):932–46. doi: 10.1159/000493225

 Ansari MY, Khan NM, Ahmad N, Green J, Novak K, Haqqi TM. Genetic inactivation of ZCCHC6 suppresses interleukin-6 expression and reduces the severity of experimental osteoarthritis in mice. *Arthritis Rheumatol* (2019) 71(4):583–93. doi: 10.1002/art.40751

76. Pacifici M. Osteoarthritis and chronic pain: Interleukin-6 as a common denominator and therapeutic target. *Sci Signal* (2022) 15(744):eadd3702. doi: 10.1126/scisignal.add3702

77. Ryu JH, Yang S, Shin Y, Rhee J, Chun CH, Chun JS. Interleukin-6 plays an essential role in hypoxia-inducible factor  $2\alpha$ -induced experimental osteoarthritic cartilage destruction in mice. *Arthritis Rheum* (2011) 63(9):2732–43. doi: 10.1002/art.30451

78. Tait SW, Ichim G, Green DR. Die another way-non-apoptotic mechanisms of cell death. J Cell Sci (2014) 127(Pt 10):2135-44. doi: 10.1242/jcs.093575

79. Salucci S, Falcieri E, Battistelli M. Chondrocyte death involvement in osteoarthritis. *Cell Tissue Res* (2022) 389(2):159–70. doi: 10.1007/s00441-022-03639-4

80. An S, Hu H, Li Y, Hu Y. Pyroptosis plays a role in osteoarthritis. *Aging Dis* (2020) 11(5):1146–57. doi: 10.14336/AD.2019.1127

81. Ravanan P, Srikumar IF, Talwar P. Autophagy: The spotlight for cellular stress responses. *Life Sci* (2017) 188:53–67. doi: 10.1016/j.lfs.2017.08.029

82. Thangaraj A, Sil S, Tripathi A, Chivero ET, Periyasamy P, Buch S. Targeting endoplasmic reticulum stress and autophagy as therapeutic approaches for neurological diseases. *Int Rev Cell Mol Biol* (2020) 350:285–325. doi: 10.1016/bs.ircmb.2019.11.001

83. Parzych KR, Klionsky DJ. An overview of autophagy: morphology, mechanism, and regulation. *Antioxid Redox Signal* (2014) 20(3):460–73. doi: 10.1089/ars.2013.5371

84. Klionsky DJ. Autophagy: from phenomenology to molecular understanding in less than a decade. *Nat Rev Mol Cell Biol* (2007) 8(11):931–7. doi: 10.1038/nrm2245

85. Yang Y, Klionsky DJ. Autophagy and disease: unanswered questions. *Cell Death Differ* (2020) 27(3):858-71. doi: 10.1038/s41418-019-0480-9

86. Bento CF, Renna M, Ghislat G, Puri C, Ashkenazi A, Vicinanza M, et al. Mammalian autophagy: how does it work? *Annu Rev Biochem* (2016) 85:685–713. doi: 10.1146/annurev-biochem-060815-014556

87. Dikic I, Elazar Z. Mechanism and medical implications of mammalian autophagy. *Nat Rev Mol Cell Biol* (2018) 19(6):349-64. doi: 10.1038/s41580-018-0003-4

88. Schulze-Luehrmann J, Eckart RA, Ölke M, Saftig P, Liebler-Tenorio E, Lührmann A. LAMP proteins account for the maturation delay during the establishment of the Coxiella burnetii-containing vacuole. *Cell Microbiol* (2016) 18 (2):181–94. doi: 10.1111/cmi.12494

89. Levine B, Kroemer G. Biological functions of autophagy genes: A disease perspective. Cell (2019) 176(1-2):11–42. doi: 10.1016/j.cell.2018.09.048

90. Whitehead NP. Enhanced autophagy as a potential mechanism for the improved physiological function by simvastatin in muscular dystrophy. *Autophagy* (2016) 12 (4):705–6. doi: 10.1080/15548627.2016.1144005

91. Zhang Y, Vasheghani F, Li YH, Blati M, Simeone K, Fahmi H, et al. et al: Cartilage-specific deletion of mTOR upregulates autophagy and protects mice from osteoarthritis. *Ann Rheum Dis* (2015) 74(7):1432–40. doi: 10.1136/annrheumdis-2013-204599

92. Bouderlique T, Vuppalapati KK, Newton PT, Li L, Barenius B, Chagin AS. Targeted deletion of Atg5 in chondrocytes promotes age-related osteoarthritis. *Ann Rheum Dis* (2016) 75(3):627–31. doi: 10.1136/annrheumdis-2015-207742

93. Majtnerová P, Roušar T. An overview of apoptosis assays detecting DNA fragmentation. *Mol Biol Rep* (2018) 45(5):1469–78. doi: 10.1007/s11033-018-4258-9

94. Liu S, Pan Y, Li T, Zou M, Liu W, Li Q, et al. The role of regulated programmed cell death in osteoarthritis: from pathogenesis to therapy. *Int J Mol Sci* (2023) 24 (6):5364. doi: 10.3390/ijms24065364

95. Madsen-Bouterse SA, Zhong Q, Mohammad G, Ho YS, Kowluru RA. Oxidative damage of mitochondrial DNA in diabetes and its protection by manganese superoxide dismutase. *Free Radic Res* (2010) 44(3):313–21. doi: 10.3109/10715760903494168

96. Oakes SA, Papa FR. The role of endoplasmic reticulum stress in human pathology. *Annu Rev Pathol* (2015) 10:173–94. doi: 10.1146/annurev-pathol-012513-104649

97. Yamashita SI, Kanki T. How autophagy eats large mitochondria: Autophagosome formation coupled with mitochondrial fragmentation. *Autophagy* (2017) 13(5):980–1. doi: 10.1080/15548627.2017.1291113

98. Liu D, Cai ZJ, Yang YT, Lu WH, Pan LY, Xiao WF, et al. Mitochondrial quality control in cartilage damage and osteoarthritis: new insights and potential therapeutic targets. Osteoarthritis Cartilage (2022) 30(3):395-405. doi: 10.1016/j.joca.2021.10.009

99. Lou G, Palikaras K, Lautrup S, Scheibye-Knudsen M, Tavernarakis N, Fang EF. Mitophagy and neuroprotection. *Trends Mol Med* (2020) 26(1):8–20. doi: 10.1016/j.molmed.2019.07.002

100. Jorgensen I, Miao EA. Pyroptotic cell death defends against intracellular pathogens. Immunol Rev (2015) 265(1):130–42. doi: 10.1111/imr.12287

101. Frank D, Vince JE. Pyroptosis versus necroptosis: similarities, differences, and crosstalk. *Cell Death Differ* (2019) 26(1):99–114. doi: 10.1038/s41418-018-0212-6

102. Burdette BE, Esparza AN, Zhu H, Wang S. Gasdermin D in pyroptosis. Acta Pharm Sin B (2021) 11(9):2768–82. doi: 10.1016/j.apsb.2021.02.006

103. Bedoui S, Herold MJ, Strasser A. Emerging connectivity of programmed cell death pathways and its physiological implications. *Nat Rev Mol Cell Biol* (2020) 21 (11):678–95. doi: 10.1038/s41580-020-0270-8

104. Hsu SK, Li CY, Lin IL, Syue WJ, Chen YF, Cheng KC, et al. Inflammationrelated pyroptosis, a novel programmed cell death pathway, and its crosstalk with immune therapy in cancer treatment. *Theranostics* (2021) 11(18):8813–35. doi: 10.7150/thno.62521

105. Lacey CA, Mitchell WJ, Dadelahi AS, Skyberg JA. Caspase-1 and caspase-11 mediate pyroptosis, inflammation, and control of brucella joint infection. *Infect Immun* (2018) 86(9):e00361-18. doi: 10.1128/IAI.00361-18

106. D'Anna SE, Maniscalco M, Cappello F, Carone M, Motta A, Balbi B, et al. Bacterial and viral infections and related inflammatory responses in chronic obstructive pulmonary disease. *Ann Med* (2021) 53(1):135-50. doi: 10.1080/ 07853890.2020.1831050

107. Xu YJ, Zheng L, Hu YW, Wang Q. Pyroptosis and its relationship to atherosclerosis. Clin Chim Acta (2018) 476:28–37. doi: 10.1016/j.cca.2017.11.005

108. Yu J, Li S, Qi J, Chen Z, Wu Y, Guo J, et al. Cleavage of GSDME by caspase-3 determines lobaplatin-induced pyroptosis in colon cancer cells. *Cell Death Dis* (2019) 10(3):193. doi: 10.1038/s41419-019-1441-4

109. Wu Z, Liu Q, Zhu K, Liu Y, Chen L, Guo H, et al. Cigarette smoke induces the pyroptosis of urothelial cells through ROS/NLRP3/caspase-1 signaling pathway. *Neurourol Urodyn* (2020) 39(2):613–24. doi: 10.1002/nau.24271

110. Zhang Z, Fu F, Bian Y, Zhang H, Yao S, Zhou C, et al.  $\alpha$ -chaconine facilitates chondrocyte pyroptosis and nerve ingrowth to aggravate osteoarthritis progression by activating NF- $\kappa$ B signaling. J Inflamm Res (2022) 15:5873–88. doi: 10.2147/JIR.S382675

111. Zhang Y, Lin Z, Chen D, He Y. CY-09 attenuates the progression of osteoarthritis via inhibiting NLRP3 inflammasome-mediated pyroptosis. Biochem Biophys Res Commun (2021) 553:119–25. doi: 10.1016/j.bbrc.2021.03.055

112. Dixon SJ, Lemberg KM, Lamprecht MR, Skouta R, Zaitsev EM, Gleason CE, et al. Ferroptosis: an iron-dependent form of nonapoptotic cell death. *Cell* (2012) 149 (5):1060–72. doi: 10.1016/j.cell.2012.03.042

113. Tong L, Yu H, Huang X, Shen J, Xiao G, Chen L, et al. Current understanding of osteoarthritis pathogenesis and relevant new approaches. *Bone Res* (2022) 10(1):60. doi: 10.1038/s41413-022-00226-9

114. Yan HF, Zou T, Tuo QZ, Xu S, Li H, Belaidi AA, et al. Ferroptosis: mechanisms and links with diseases. *Signal Transduct Target Ther* (2021) 6(1):49. doi: 10.1038/ s41392-020-00428-9

115. Yang WS, SriRamaratnam R, Welsch ME, Shimada K, Skouta R, Viswanathan VS, et al. Regulation of ferroptotic cancer cell death by GPX4. *Cell* (2014) 156(1-2):317–31. doi: 10.1016/j.cell.2013.12.010

116. Jiang X, Stockwell BR, Conrad M. Ferroptosis: mechanisms, biology and role in disease. *Nat Rev Mol Cell Biol* (2021) 22(4):266–82. doi: 10.1038/s41580-020-00324-8

117. Hu Y, Wang Y, Liu S, Wang H. The potential roles of ferroptosis in pathophysiology and treatment of musculoskeletal diseases-opportunities, challenges, and perspectives. J Clin Med (2023) 12(6):2125. doi: 10.3390/jcm12062125

118. Al-Hetty H, Abdulameer SJ, Alghazali MW, Sheri FS, Saleh MM, Jalil AT. The role of ferroptosis in the pathogenesis of osteoarthritis. *J Membr Biol* (2023) 256 (3):223–8. doi: 10.1007/s00232-023-00282-0

119. Sun K, Guo Z, Zhang J, Hou L, Liang S, Lu F, et al. Inhibition of TRADD ameliorates chondrocyte necroptosis and osteoarthritis by blocking RIPK1-TAK1 pathway and restoring autophagy. *Cell Death Discovery* (2023) 9(1):109. doi: 10.1038/s41420-023-01406-0

120. Arendt J. Aaron Lerner, who discovered melatonin. J Pineal Res (2007) 43 (1):106-7. doi: 10.1111/j.1600-079X.2007.00457.x

121. Acuña-Castroviejo D, Escames G, Venegas C, Díaz-Casado ME, Lima-Cabello E, López LC, et al. Extrapineal melatonin: sources, regulation, and potential functions. *Cell Mol Life Sci* (2014) 71(16):2997–3025. doi: 10.1007/s00018-014-1579-2

122. Chen X, Hao A, Li X, Du Z, Li H, Wang H, et al. Melatonin inhibits tumorigenicity of glioblastoma stem-like cells *via* the AKT-EZH2-STAT3 signaling axis. *J Pineal Res* (2016) 61(2):208–17. doi: 10.1111/jpi.12341

123. Cipolla-Neto J, Amaral FGD. Melatonin as a hormone: new physiological and clinical insights. *Endocr Rev* (2018) 39(6):990–1028. doi: 10.1210/er.2018-00084

124. Stehle JH, Saade A, Rawashdeh O, Ackermann K, Jilg A, Sebestény T, et al. A survey of molecular details in the human pineal gland in the light of phylogeny, structure, function and chronobiological diseases. *J Pineal Res* (2011) 51(1):17–43. doi: 10.1111/j.1600-079X.2011.00856.x

125. Reiter RJ, Tan DX, Kim SJ, Cruz MH. Delivery of pineal melatonin to the brain and SCN: role of canaliculi, cerebrospinal fluid, tanycytes and Virchow-Robin perivascular spaces. *Brain Struct Funct* (2014) 219(6):1873–87. doi: 10.1007/s00429-014-0719-7 126. Slominski AT, Zmijewski MA, Jetten AM. ROR $\alpha$  is not a receptor for melatonin (response to DOI 10.1002/bies.201600018). *Bioessays* (2016) 38(12):1193–4. doi: 10.1002/bies.201600204

127. Slominski AT, Kleszczyński K, Semak I, Janjetovic Z, Zmijewski MA, Kim TK, et al. Local melatoninergic system as the protector of skin integrity. *Int J Mol Sci* (2014) 15(10):17705–32. doi: 10.3390/ijms151017705

128. Hosseinzadeh A, Kamrava SK, Joghataei MT, Darabi R, Shakeri-Zadeh A, Shahriari M, et al. Apoptosis signaling pathways in osteoarthritis and possible protective role of melatonin. *J Pineal Res* (2016) 61(4):411–25. doi: 10.1111/jpi.12362

129. Lu KH, Lu PW, Lu EW, Tang CH, Su SC, Lin CW, et al. The potential remedy of melatonin on osteoarthritis. *J Pineal Res* (2021) 71(3):e12762. doi: 10.1111/jpi.12762

130. Gobbi G, Comai S. Differential function of melatonin MT(1) and MT(2) receptors in REM and NREM sleep. *Front Endocrinol (Lausanne)* (2019) 10:87. doi: 10.3389/fendo.2019.00087

131. He B, Zhao Y, Xu L, Gao L, Su Y, Lin N, et al. The nuclear melatonin receptor ROR $\alpha$  is a novel endogenous defender against myocardial ischemia/reperfusion injury. J Pineal Res (2016) 60(3):313–26. doi: 10.1111/jpi.12312

132. Espino J, Ortiz Á, Bejarano I, Lozano GM, Monllor F, García JF, et al. Melatonin protects human spermatozoa from apoptosis *via* melatonin receptor- and extracellular signal-regulated kinase-mediated pathways. *Fertil Steril* (2011) 95 (7):2290–6. doi: 10.1016/j.fertnstert.2011.03.063

133. Galano A. On the direct scavenging activity of melatonin towards hydroxyl and a series of peroxyl radicals. *Phys Chem Chem Phys* (2011) 13(15):7178–88. doi: 10.1039/ c0cp02801k

134. Galano A, Medina ME, Tan DX, Reiter RJ. Melatonin and its metabolites as copper chelating agents and their role in inhibiting oxidative stress: a physicochemical analysis. *J Pineal Res* (2015) 58(1):107–16. doi: 10.1111/jpi.12196

135. Fischer TW, Kleszczyński K, Hardkop LH, Kruse N, Zillikens D. Melatonin enhances antioxidative enzyme gene expression (CAT, GPx, SOD), prevents their UVR-induced depletion, and protects against the formation of DNA damage (8-hydroxy-2'-deoxyguanosine) in ex vivo human skin. *J Pineal Res* (2013) 54(3):303–12. doi: 10.1111/jpi.12018

136. Zhou X, Zhang Y, Hou M, Liu H, Yang H, Chen X, et al. Melatonin prevents cartilage degradation in early-stage osteoarthritis through activation of miR-146a/ NRF2/HO-1 axis. J Bone Miner Res (2022) 37(5):1056–72. doi: 10.1002/jbmr.4527

137. Zhang Y, Hou M, Liu Y, Liu T, Chen X, Shi Q, et al. Recharge of chondrocyte mitochondria by sustained release of melatonin protects cartilage matrix homeostasis in osteoarthritis. *J Pineal Res* (2022) 73(2):e12815. doi: 10.1111/jpi.12815

138. Zhao Z, Bi B, Cheng G, Zhao Y, Wu H, Zheng M, et al. Melatonin ameliorates osteoarthritis rat cartilage injury by inhibiting matrix metalloproteinases and JAK2/ STAT3 signaling pathway. *Inflammopharmacology* (2023) 31(1):359–68. doi: 10.1007/ s10787-022-01102-y

139. Hong Y, Kim H, Lee S, Jin Y, Choi J, Lee SR, et al. Role of melatonin combined with exercise as a switch-like regulator for circadian behavior in advanced osteoarthritic knee. *Oncotarget* (2017) 8(57):97633–47. doi: 10.18632/oncotarget.19276

140. Liu SC, Tsai CH, Wang YH, Su CM, Wu HC, Fong YC, et al. Melatonin abolished proinflammatory factor expression and antagonized osteoarthritis progression. *vivo Cell Death Dis* (2022) 13(3):215. doi: 10.1038/s41419-022-04656-5

141. Zhang Y, Lin J, Zhou X, Chen X, Chen AC, Pi B, et al. Melatonin prevents osteoarthritis-Induced cartilage degradation *via* targeting microRNA-140. *Oxid Med Cell Longev* (2019) 2019:9705929. doi: 10.1155/2019/9705929

142. Lim HD, Kim YS, Ko SH, Yoon JJ, Cho SG, Chun YH, et al. Cytoprotective and anti-inflammatory effects of melatonin in hydrogen peroxide-stimulated CHON-001 human chondrocyte cell line and rabbit model of osteoarthritis *via* the SIRT1 pathway. *J Pineal Res* (2012) 53(3):225–37. doi: 10.1111/j.1600-079X.2012.00991.x

143. Liang H, Yan Y, Sun W, Ma X, Su Z, Liu Z, et al. Preparation of melatoninloaded nanoparticles with targeting and sustained release function and their application in osteoarthritis. *Int J Mol Sci* (2023) 24(10):8740. doi: 10.3390/ijms24108740

144. Chen Z, Zhao C, Liu P, Huang H, Zhang S, Wang X. Anti-apoptosis and autophagy effects of melatonin protect rat chondrocytes against oxidative stress. *via Regul AMPK/Foxo3 Pathways Cartilage* (2021) 13(2\_suppl):1041s–53s. doi: 10.1177/19476035211038748

145. Qin K, Tang H, Ren Y, Yang D, Li Y, Huang W, et al. Melatonin promotes sirtuin 1 expression and inhibits  $IRE1\alpha$ -XBP1S-CHOP to reduce endoplasmic reticulum stress-mediated apoptosis in chondrocytes. *Front Pharmacol* (2022) 13:940629. doi: 10.3389/fphar.2022.940629

146. Majidinia M, Sadeghpour A, Mehrzadi S, Reiter RJ, Khatami N, Yousefi B. Melatonin: A pleiotropic molecule that modulates DNA damage response and repair pathways. *J Pineal Res* (2017) 63(1):12416. doi: 10.1111/jpi.12416

147. Venegas C, García JA, Escames G, Ortiz F, López A, Doerrier C, et al. Extrapineal melatonin: analysis of its subcellular distribution and daily fluctuations. J Pineal Res (2012) 52(2):217–27. doi: 10.1111/j.1600-079X.2011.00931.x

148. Tan DX, Manchester LC, Esteban-Zubero E, Zhou Z, Reiter RJ. Melatonin as a potent and inducible endogenous antioxidant: synthesis and metabolism. *Molecules* (2015) 20(10):18886–906. doi: 10.3390/molecules201018886

149. Mehrzadi S, Safa M, Kamrava SK, Darabi R, Hayat P, Motevalian M. Protective mechanisms of melatonin against hydrogen-peroxide-induced toxicity in human bone-

marrow-derived mesenchymal stem cells. Can J Physiol Pharmacol (2017) 95(7):773-86. doi: 10.1139/cjpp-2016-0409

150. Goudarzi M, Khodayar MJ, Hosseini Tabatabaei SMT, Ghaznavi H, Fatemi I, Mehrzadi S. Pretreatment with melatonin protects against cyclophosphamide-induced oxidative stress and renal damage in mice. *Fundam Clin Pharmacol* (2017) 31(6):625–35. doi: 10.1111/fcp.12303

151. Robinette TM, Nicholatos JW, Francisco AB, Brooks KE, Diao RY, Sorbi S, et al. SIRT1 accelerates the progression of activity-based anorexia. *Nat Commun* (2020) 11 (1):2814. doi: 10.1038/s41467-020-16348-9

152. Olmos Y, Sánchez-Gómez FJ, Wild B, García-Quintans N, Cabezudo S, Lamas S, et al. SirT1 regulation of antioxidant genes is dependent on the formation of a FoxO3a/PGC-1α complex. *Antioxid Redox Signal* (2013) 19(13):1507–21. doi: 10.1089/ars.2012.4713

153. Li Y, Xiao W, Wu P, Deng Z, Zeng C, Li H, et al. The expression of SIRT1 in articular cartilage of patients with knee osteoarthritis and its correlation with disease severity. *J Orthop Surg Res* (2016) 11(1):144. doi: 10.1186/s13018-016-0477-8

154. Guo JY, Li F, Wen YB, Cui HX, Guo ML, Zhang L, et al. Melatonin inhibits Sirt1-dependent NAMPT and NFAT5 signaling in chondrocytes to attenuate osteoarthritis. *Oncotarget* (2017) 8(34):55967-83. doi: 10.18632/oncotarget.18356

155. Zhao M, Song X, Chen H, Ma T, Tang J, Wang X, et al. Melatonin prevents chondrocyte matrix degradation in rats with experimentally induced osteoarthritis by inhibiting nuclear factor-κB *via* SIRT1. *Nutrients* (2022) 14(19):3966. doi: 10.3390/ nu14193966

156. Liu-Bryan R, Terkeltaub R. Emerging regulators of the inflammatory process in osteoarthritis. *Nat Rev Rheumatol* (2015) 11(1):35–44. doi: 10.1038/nrrheum.2014.162

157. Liu X, Xu Y, Chen S, Tan Z, Xiong K, Li Y, et al. Rescue of proinflammatory cytokine-inhibited chondrogenesis by the antiarthritic effect of melatonin in synovium mesenchymal stem cells *via* suppression of reactive oxygen species and matrix metalloproteinases. *Free Radic Biol Med* (2014) 68:234–46. doi: 10.1016/ j.freeradbiomed.2013.12.012

158. Gao B, Gao W, Wu Z, Zhou T, Qiu X, Wang X, et al. Melatonin rescued interleukin 1 $\beta$ -impaired chondrogenesis of human mesenchymal stem cells. *Stem Cell Res Ther* (2018) 9(1):162. doi: 10.1186/s13287-018-0892-3

159. Ke C, Li H, Yang D, Ying H, Zhu H, Wang J, et al. Melatonin attenuates the progression of osteoarthritis in rats by inhibiting inflammation and related oxidative stress on the surface of knee cartilage. *Orthop Surg* (2022) 14(9):2230–7. doi: 10.1111/ os.13408

160. Hong Y, Kim H, Lee Y, Lee S, Kim K, Jin Y, et al. Salutary effects of melatonin combined with treadmill exercise on cartilage damage. *J Pineal Res* (2014) 57(1):53–66. doi: 10.1111/jpi.12143

161. Tao SC, Huang JY, Gao Y, Li ZX, Wei ZY, Dawes H, et al. Small extracellular vesicles in combination with sleep-related circRNA3503: A targeted therapeutic agent with injectable thermosensitive hydrogel to prevent osteoarthritis. *Bioact Mater* (2021) 6(12):4455–69. doi: 10.1016/j.bioactmat.2021.04.031

162. Hoesel B, Schmid JA. The complexity of NF-κB signaling in inflammation and cancer. *Mol Cancer* (2013) 12:86. doi: 10.1186/1476-4598-12-86

163. Mullen LM, Chamberlain G, Sacre S. Pattern recognition receptors as potential therapeutic targets in inflammatory rheumatic disease. *Arthritis Res Ther* (2015) 17 (1):122. doi: 10.1186/s13075-015-0645-y

164. Choi SE, Fu T, Seok S, Kim DH, Yu E, Lee KW, et al. Elevated microRNA-34a in obesity reduces NAD+ levels and SIRT1 activity by directly targeting NAMPT. *Aging Cell* (2013) 12(6):1062–72. doi: 10.1111/acel.12135

165. Rongvaux A, Shea RJ, Mulks MH, Gigot D, Urbain J, Leo O, et al. Pre-B-cell colony-enhancing factor, whose expression is up-regulated in activated lymphocytes, is a nicotinamide phosphoribosyltransferase, a cytosolic enzyme involved in NAD biosynthesis. *Eur J Immunol* (2002) 32(11):3225–34. doi: 10.1002/1521-4141(200211) 32:11<3225::AID-IMMU3225>3.0.CO;2-L

166. Timucin AC, Bodur C, Basaga H. SIRT1 contributes to aldose reductase expression through modulating NFAT5 under osmotic stress: *In vitro* and in silico insights. *Cell Signal* (2015) 27(11):2160–72. doi: 10.1016/j.cellsig.2015.08.013

167. Johnson ZI, Shapiro IM, Risbud MV. Extracellular osmolarity regulates matrix homeostasis in the intervertebral disc and articular cartilage: evolving role of TonEBP. *Matrix Biol* (2014) 40:10–6. doi: 10.1016/j.matbio.2014.08.014

168. Yang W, Kang X, Qin N, Li F, Jin X, Ma Z, et al. Melatonin protects chondrocytes from impairment induced by glucocorticoids *via* NAD(+)-dependent SIRT1. *Steroids* (2017) 126:24–9. doi: 10.1016/j.steroids.2017.08.005

169. Li J, Huang J, Dai L, Yu D, Chen Q, Zhang X, et al. miR-146a, an IL-1 $\beta$  responsive miRNA, induces vascular endothelial growth factor and chondrocyte apoptosis by targeting Smad4. *Arthritis Res Ther* (2012) 14(2):R75. doi: 10.1186/ar3798

170. Chinzei N, Brophy RH, Duan X, Cai L, Nunley RM, Sandell LJ, et al. Molecular influence of anterior cruciate ligament tear remnants on chondrocytes: a biologic connection between injury and osteoarthritis. *Osteoarthritis Cartilage* (2018) 26 (4):588–99. doi: 10.1016/j.joca.2018.01.017

171. Pei M, He F, Wei L, Rawson A. Melatonin enhances cartilage matrix synthesis by porcine articular chondrocytes. *J Pineal Res* (2009) 46(2):181–7. doi: 10.1111/j.1600-079X.2008.00646.x

172. Thomas CM, Fuller CJ, Whittles CE, Sharif M. Chondrocyte death by apoptosis is associated with cartilage matrix degradation. *Osteoarthritis Cartilage* (2007) 15 (1):27–34. doi: 10.1016/j.joca.2006.06.012

173. Dai M, Sui B, Xue Y, Liu X, Sun J. Cartilage repair in degenerative osteoarthritis mediated by squid type II collagen *via* immunomodulating activation of M2 macrophages, inhibiting apoptosis and hypertrophy of chondrocytes. *Biomaterials* (2018) 180:91–103. doi: 10.1016/j.biomaterials.2018.07.011

174. Moon HS, Kim B, Gwak H, Suh DH, Song YS. Autophagy and protein kinase RNA-like endoplasmic reticulum kinase (PERK)/eukaryotic initiation factor 2 alpha kinase (eIF2 $\alpha$ ) pathway protect ovarian cancer cells from metformin-induced apoptosis. *Mol Carcinog* (2016) 55(4):346–56. doi: 10.1002/mc.22284

175. Vringer E, Tait SWG. Mitochondria and cell death-associated inflammation. Cell Death Differ (2023) 30(2):304–12. doi: 10.1038/s41418-022-01094-w

176. Jourdain A, Martinou JC. Mitochondrial outer-membrane permeabilization and remodelling in apoptosis. *Int J Biochem Cell Biol* (2009) 41(10):1884–9. doi: 10.1016/j.biocel.2009.05.001

177. Steinberg GR, Carling D. AMP-activated protein kinase: the current landscape for drug development. *Nat Rev Drug Discovery* (2019) 18(7):527–51. doi: 10.1038/s41573-019-0019-2

178. Fan C, Feng J, Tang C, Zhang Z, Feng Y, Duan W, et al. et al: Melatonin suppresses ER stress-dependent proapoptotic effects *via* AMPK in bone mesenchymal stem cells during mitochondrial oxidative damage. *Stem Cell Res Ther* (2020) 11(1):442. doi: 10.1186/s13287-020-01948-5

179. Lin CH, Cheng YC, Nicol CJ, Lin KH, Yen CH, Chiang MC. Activation of AMPK is neuroprotective in the oxidative stress by advanced glycosylation end products in human neural stem cells. *Exp Cell Res* (2017) 359(2):367-73. doi: 10.1016/j.yexcr.2017.08.019

180. Wang R, Zhang S, Previn R, Chen D, Jin Y, Zhou G. Role of forkhead box O transcription factors in oxidative stress-induced chondrocyte dysfunction: possible therapeutic target for osteoarthritis? *Int J Mol Sci* (2018) 19(12):3794. doi: 10.3390/ ijms19123794

181. Akasaki Y, Alvarez-Garcia O, Saito M, Caramés B, Iwamoto Y, Lotz MK. FoxO transcription factors support oxidative stress resistance in human chondrocytes. *Arthritis Rheumatol* (2014) 66(12):3349–58. doi: 10.1002/art.38868

182. Hetz C, Zhang K, Kaufman RJ. Mechanisms, regulation and functions of the unfolded protein response. *Nat Rev Mol Cell Biol* (2020) 21(8):421–38. doi: 10.1038/s41580-020-0250-z

183. Tavernier SJ, Osorio F, Vandersarren L, Vetters J, Vanlangenakker N, Van Isterdael G, et al. Regulated IRE1-dependent mRNA decay sets the threshold for dendritic cell survival. *Nat Cell Biol* (2017) 19(6):698–710. doi: 10.1038/ncb3518

184. Sepulveda D, Rojas-Rivera D, Rodríguez DA, Groenendyk J, Köhler A, Lebeaupin C, et al. Interactome screening identifies the ER luminal chaperone hsp47 as a regulator of the unfolded protein response transducer IRE1α. *Mol Cell* (2018) 69 (2):238–52.e237. doi: 10.1016/j.molcel.2017.12.028

185. Abdullah A, Ravanan P. The unknown face of IRE1 $\alpha$  - Beyond ER stress. Eur J Cell Biol (2018) 97(5):359–68. doi: 10.1016/j.ejcb.2018.05.002

186. Komoike Y, Matsuoka M. Endoplasmic reticulum stress-mediated neuronal apoptosis by acrylamide exposure. *Toxicol Appl Pharmacol* (2016) 310:68–77. doi: 10.1016/j.taap.2016.09.005

187. Wu L, Liu H, Li L, Xu D, Gao Y, Guan Y, et al. 5,7,3',4'-Tetramethoxyflavone protects chondrocytes from ER stress-induced apoptosis through regulation of the IRE1 $\alpha$  pathway. *Connect Tissue Res* (2018) 59(2):157-66. doi: 10.1080/03008207.2017.1321639

188. Charbord P. Bone marrow mesenchymal stem cells: historical overview and concepts. *Hum Gene Ther* (2010) 21(9):1045–56. doi: 10.1089/hum.2010.115

189. Raghav PK, Mann Z, Ahlawat S, Mohanty S. Mesenchymal stem cell-based nanoparticles and scaffolds in regenerative medicine. *Eur J Pharmacol* (2022) 918:174657. doi: 10.1016/j.ejphar.2021.174657

190. Liu X, Gong Y, Xiong K, Ye Y, Xiong Y, Zhuang Z, et al. Melatonin mediates protective effects on inflammatory response induced by interleukin-1 beta in human mesenchymal stem cells. *J Pineal Res* (2013) 55(1):14–25. doi: 10.1111/jpi.12045

191. Li YS, Xiao WF, Luo W. Cellular aging towards osteoarthritis. Mech Ageing Dev (2017) 162:80-4. doi: 10.1016/j.mad.2016.12.012

192. Han N, Wang Z, Li X. Melatonin alleviates d-galactose-decreased hyaluronic acid production in synovial membrane cells *via* Sirt1 signalling. *Cell Biochem Funct* (2021) 39(4):488–95. doi: 10.1002/cbf.3613

193. Jung JH, Seok H, Choi SJ, Bae J, Lee SH, Lee MH, et al. The association between osteoarthritis and sleep duration in Koreans: a nationwide cross-sectional observational study. *Clin Rheumatol* (2018) 37(6):1653–9. doi: 10.1007/s10067-018-4040-3

194. Lin TB, Hsieh MC, Lai CY, Cheng JK, Wang HH, Chau YP, et al. Melatonin relieves neuropathic allodynia through spinal MT2-enhanced PP2Ac and downstream HDAC4 shuttling-dependent epigenetic modification of hmgb1 transcription. *J Pineal Res* (2016) 60(3):263–76. doi: 10.1111/jpi.12307

195. Lin JJ, Lin Y, Zhao TZ, Zhang CK, Zhang T, Chen XL, et al. Melatonin suppresses neuropathic pain *via* MT2-dependent and -independent pathways in dorsal root ganglia neurons of mice. *Theranostics* (2017) 7(7):2015–32. doi: 10.7150/thno.19500

196. Posa L, Lopez-Canul M, Rullo L, De Gregorio D, Dominguez-Lopez S, Kaba Aboud M, et al. Nociceptive responses in melatonin MT(2) receptor knockout mice compared to MT(1) and double MT(1) /MT(2) receptor knockout mice. *J Pineal Res* (2020) 69(3):e12671. doi: 10.1111/jpi.12671

197. Xie SS, Fan WG, Liu Q, Li JZ, Zheng MM, He HW, et al. Involvement of nNOS in the antinociceptive activity of melatonin in inflammatory pain at the level of sensory neurons. *Eur Rev Med Pharmacol Sci* (2020) 24(13):7399–411. doi: 10.26355/eurrev\_202007\_21908

198. Hussain SA, Al K II, Jasim NA, Gorial FI. Adjuvant use of melatonin for treatment of fibromyalgia. *J Pineal Res* (2011) 50(3):267-71. doi: 10.1111/j.1600-079X.2010.00836.x

199. Gonçalves AL, Martini Ferreira A, Ribeiro RT, Zukerman E, Cipolla-Neto J, Peres MF. Randomised clinical trial comparing melatonin 3 mg, amitriptyline 25 mg and placebo for migraine prevention. *J Neurol Neurosurg Psychiatry* (2016) 87 (10):1127–32. doi: 10.1136/jnnp-2016-313458

200. Siah KT, Wong RK, Ho KY. Melatonin for the treatment of irritable bowel syndrome. World J Gastroenterol (2014) 20(10):2492-8. doi: 10.3748/wjg.v20.i10.2492

201. Liu W, Jiang H, Liu X, Hu S, Li H, Feng Y, et al. Melatonin abates TMJOA chronic pain by MT(2)R in trigeminal ganglion neurons. *J Dent Res* (2022) 101(1):111–9. doi: 10.1177/00220345211026551

202. Zhang Y, Zhang H. Effects of auricular acupressure on sleep and pain in elderly people who have osteoarthritis and live in nursing homes: A randomized, single-blind, placebo-controlled trial [Letter]. *Explore* (*NY*) (2023) 19(2):214–22. doi: 10.1016/ j.explore.2023.04.001

203. Shim KN, Kim JI, Kim N, Kim SG, Jo YJ, Hong SJ, et al. The efficacy and safety of irsogladine maleate in nonsteroidal anti-inflammatory drug or aspirin-induced peptic ulcer and gastritis. *Korean J Intern Med* (2019) 34(5):1008–21. doi: 10.3904/kjim.2017.370

204. Stout A, Friedly J, Standaert CJ. Systemic absorption and side effects of locally injected glucocorticoids. *Pm r* (2019) 11(4):409–19. doi: 10.1002/pmrj.12042

205. Paulino Silva KM, de Sousa FL, Alves ACB, Rocha PA, da Costa H, Ferreira WR, et al. Chondroprotective effect of melatonin and strontium ranelate in animal model of osteoarthritis. *Heliyon* (2021) 7(4):e06760. doi: 10.1016/j.heliyon.2021.e06760

206. Bruno MC, Cristiano MC, Celia C, d'Avanzo N, Mancuso A, Paolino D, et al. Injectable drug delivery systems for osteoarthritis and rheumatoid arthritis. *ACS nano* (2022) 16(12):19665–90. doi: 10.1021/acsnano.2c06393

207. Wiegant K, van Roermund PM, Intema F, Cotofana S, Eckstein F, Mastbergen SC, et al. Sustained clinical and structural benefit after joint distraction in the treatment of severe knee osteoarthritis. *Osteoarthritis Cartilage* (2013) 21(11):1660–7. doi: 10.1016/j.joca.2013.08.006

208. Kim YS, Go G, Yun CW, Yea JH, Yoon S, Han SY, et al. Topical administration of melatonin-loaded extracellular vesicle-mimetic nanovesicles improves 2,4-dinitrofluorobenzene-induced atopic dermatitis. *Biomolecules* (2021) 11(10):1450. doi: 10.3390/biom11101450

209. Massella D, Leone F, Peila R, Barresi AA, Ferri A. Functionalization of cotton fabrics with polycaprolactone nanoparticles for transdermal release of melatonin. J Funct Biomater (2017) 9(1):1. doi: 10.3390/jfb9010001

210. Hoti G, Ferrero R, Caldera F, Trotta F, Corno M, Pantaleone S, et al. A comparison between the molecularly imprinted and non-molecularly imprinted cyclodextrin-based nanosponges for the transdermal delivery of melatonin. *Polymers (Basel)* (2023) 15(6):1543. doi: 10.3390/polym15061543

211. Xiao L, Lin J, Chen R, Huang Y, Liu Y, Bai J, et al. Sustained release of melatonin from gelMA liposomes reduced osteoblast apoptosis and improved implant osseointegration in osteoporosis. *Oxid Med Cell Longev* (2020) 2020:6797154. doi: 10.1155/2020/6797154

212. Ji X, Yan Y, Sun T, Zhang Q, Wang Y, Zhang M, et al. Glucosamine sulphateloaded distearoyl phosphocholine liposomes for osteoarthritis treatment: combination of sustained drug release and improved lubrication. *Biomaterials Sci* (2019) 7(7):2716– 28. doi: 10.1039/C9BM00201D

213. Atoufi Z, Kamrava SK, Davachi SM, Hassanabadi M, Saeedi Garakani S, Alizadeh R, et al. Injectable PNIPAM/Hyaluronic acid hydrogels containing multipurpose modified particles for cartilage tissue engineering: Synthesis, characterization, drug release and cell culture study. *Int J Biol macromolecules* (2019) 139:1168–81. doi: 10.1016/j.ijbiomac.2019.08.101

214. Kouhi M, Varshosaz J, Hashemibeni B, Sarmadi A. Injectable gellan gum/ lignocellulose nanofibrils hydrogels enriched with melatonin loaded forsterite nanoparticles for cartilage tissue engineering: Fabrication, characterization and cell culture studies. *Materials Sci Eng C Materials Biol Appl* (2020) 115:111114. doi: 10.1016/j.msec.2020.111114

215. Nikfarjam S, Rezaie J, Zolbanin NM, Jafari R. Mesenchymal stem cell derivedexosomes: a modern approach in translational medicine. *J Transl Med* (2020) 18 (1):449. doi: 10.1186/s12967-020-02622-3

216. Armstrong JPK, Stevens MM. Strategic design of extracellular vesicle drug delivery systems. *Adv Drug Delivery Rev* (2018) 130:12–6. doi: 10.1016/j.addr.2018.06.017

217. Elahi FM, Farwell DG, Nolta JA, Anderson JD. Preclinical translation of exosomes derived from mesenchymal stem/stromal cells. *Stem Cells* (2020) 38(1):15–21. doi: 10.1002/stem.3061

218. Chen TS, Arslan F, Yin Y, Tan SS, Lai RC, Choo AB, et al. Enabling a robust scalable manufacturing process for therapeutic exosomes through oncogenic immortalization of human ESC-derived MSCs. *J Transl Med* (2011) 9:47. doi: 10.1186/1479-5876-9-47

219. Chen CC, Liu L, Ma F, Wong CW, Guo XE, Chacko JV, et al. Elucidation of exosome migration across the blood-Brain barrier model. *In Vitro Cell Mol Bioeng* (2016) 9(4):509–29. doi: 10.1007/s12195-016-0458-3

220. Headland SE, Jones HR, Norling LV, Kim A, Souza PR, Corsiero E, et al. Neutrophil-derived microvesicles enter cartilage and protect the joint in inflammatory arthritis. *Sci Transl Med* (2015) 7(315):315ra190. doi: 10.1126/scitranslmed.aac5608

221. Tao SC, Yuan T, Zhang YL, Yin WJ, Guo SC, Zhang CQ. Exosomes derived from miR-140-5p-overexpressing human synovial mesenchymal stem cells enhance cartilage tissue regeneration and prevent osteoarthritis of the knee in a rat model. *Theranostics* (2017) 7(1):180–95. doi: 10.7150/thno.17133

222. Tang Q, Lim T, Shen LY, Zheng G, Wei XJ, Zhang CQ, et al. Well-dispersed platelet lysate entrapped nanoparticles incorporate with injectable PDLLA-PEG-PDLLA triblock for preferable cartilage engineering application. *Biomaterials* (2021) 268:120605. doi: 10.1016/j.biomaterials.2020.120605

223. Oh K, Kim SR, Kim DK, Seo MW, Lee C, Lee HM, et al. *In vivo* differentiation of therapeutic insulin-Producing cells from bone marrow cells *via* extracellular vesicle-Mimetic nanovesicles. *ACS Nano* (2015) 9(12):11718–27. doi: 10.1021/acsnano.5b02997

224. Jo W, Jeong D, Kim J, Park J. Self-renewal of bone marrow stem cells by nanovesicles engineered from embryonic stem cells. *Adv Healthc Mater* (2016) 5 (24):3148–56. doi: 10.1002/adhm.201600810

225. Gangadaran P, Ahn BC. Extracellular vesicle- and extracellular vesicle mimetics-based drug delivery systems: new perspectives, challenges, and clinical developments. *Pharmaceutics* (2020) 12(5):442. doi: 10.3390/pharmaceutics12050442

226. Zhang Y, Yang N, Huang X, Zhu Y, Gao S, Liu Z, et al. Melatonin engineered adipose-derived biomimetic nanovesicles regulate mitochondrial functions and promote myocardial repair in myocardial infarction. *Front Cardiovasc Med* (2022) 9:789203. doi: 10.3389/fcvm.2022.789203

227. Li X, Dai B, Guo J, Zheng L, Guo Q, Peng J, et al. Nanoparticle-cartilage interaction: pathology-based intra-articular drug delivery for osteoarthritis therapy. *Nanomicro Lett* (2021) 13(1):149. doi: 10.1007/s40820-021-00670-y

228. Danhier F, Ansorena E, Silva JM, Coco R, Le Breton A, Préat V. PLGA-based nanoparticles: an overview of biomedical applications. *J Control Release* (2012) 161 (2):505–22. doi: 10.1016/j.jconrel.2012.01.043

229. Dash TK, Konkimalla VB. Poly-small je, Ukrainian-caprolactone based formulations for drug delivery and tissue engineering: A review. J Control Release (2012) 158(1):15–33. doi: 10.1016/j.jconrel.2011.09.064

230. Tian B, Hua S, Liu J. Cyclodextrin-based delivery systems for chemotherapeutic anticancer drugs: A review. *Carbohydr Polym* (2020) 232:115805. doi: 10.1016/j.carbpol.2019.115805

231. Rivera-Delgado E, Djuhadi A, Danda C, Kenyon J, Maia J, Caplan AI, et al. Injectable liquid polymers extend the delivery of corticosteroids for the treatment of osteoarthritis. *J Control Release* (2018) 284:112-21. doi: 10.1016/ j.jconrel.2018.05.037

232. Haeri A, Sadeghian S, Rabbani S, Anvari MS, Ghassemi S, Radfar F, et al. Effective attenuation of vascular restenosis following local delivery of chitosan decorated sirolimus liposomes. *Carbohydr Polym* (2017) 157:1461–9. doi: 10.1016/ j.carbpol.2016.11.021

233. Liu C, Ewert KK, Wang N, Li Y, Safinya CR, Qiao W. A multifunctional lipid that forms contrast-agent liposomes with dual-control release capabilities for precise MRI-guided drug delivery. *Biomaterials* (2019) 221:119412. doi: 10.1016/jbiomaterials.2019.119412

234. Kéri M, Forgács A, Papp V, Bányai I, Veres P, Len A, et al. Gelatin content governs hydration induced structural changes in silica-gelatin hybrid aerogels - Implications in drug delivery. *Acta Biomater* (2020) 105:131–45. doi: 10.1016/j.actbio.2020.01.016

235. Goldberg R, Klein J. Liposomes as lubricants: beyond drug delivery. *Chem Phys Lipids* (2012) 165(4):374–81. doi: 10.1016/j.chemphyslip.2011.11.007

236. Gantar A, da Silva LP, Oliveira JM, Marques AP, Correlo VM, Novak S, et al. Nanoparticulate bioactive-glass-reinforced gellan-gum hydrogels for bone-tissue engineering. *Materials Sci Eng C Materials Biol Appl* (2014) 43:27–36. doi: 10.1016/ j.msec.2014.06.045

237. Bacelar AH, Silva-Correia J, Oliveira JM, Reis RL. Recent progress in gellan gum hydrogels provided by functionalization strategies. *J materials Chem B* (2016) 4 (37):6164–74. doi: 10.1039/C6TB01488G

238. Gong C, Qi T, Wei X, Qu Y, Wu Q, Luo F, et al. Thermosensitive polymeric hydrogels as drug delivery systems. *Curr medicinal Chem* (2013) 20(1):79–94.

239. Lin X, Tsao CT, Kyomoto M, Zhang M. Injectable natural polymer hydrogels for treatment of knee osteoarthritis. *Advanced healthcare materials* (2022) 11(9): e2101479. doi: 10.1002/adhm.202101479

240. Mazumder MA, Fitzpatrick SD, Muirhead B, Sheardown H. Cell-adhesive thermogelling PNIPAAm/hyaluronic acid cell delivery hydrogels for potential application as minimally invasive retinal therapeutics. *J Biomed materials Res Part A* (2012) 100(7):1877–87. doi: 10.1002/jbm.a.34021

241. Wang Y, Li P, Kong L. Chitosan-modified PLGA nanoparticles with versatile surface for improved drug delivery. *AAPS PharmSciTech* (2013) 14(2):585–92. doi: 10.1208/s12249-013-9943-3

242. Lima IA, Khalil NM, Tominaga TT, Lechanteur A, Sarmento B, Mainardes RM. Mucoadhesive chitosan-coated PLGA nanoparticles for oral delivery of ferulic acid. *Artif cells nanomedicine Biotechnol* (2018) 46(sup2):993–1002. doi: 10.1080/ 21691401.2018.1477788

243. Bonifacio MA, Cometa S, Cochis A, Gentile P, Ferreira AM, Azzimonti B, et al. Antibacterial effectiveness meets improved mechanical properties: Manuka honey/ gellan gum composite hydrogels for cartilage repair. *Carbohydr polymers* (2018) 198:462–72. doi: 10.1016/j.carbpol.2018.06.115

244. Pereira DR, Silva-Correia J, Oliveira JM, Reis RL, Pandit A, Biggs MJ. Nanocellulose reinforced gellan-gum hydrogels as potential biological substitutes for annulus fibrosus tissue regeneration. *Nanomedicine* (2018) 14(3):897–908. doi: 10.1016/j.nano.2017.11.011

245. Hassanzadeh-Tabrizi SA, Bigham A, Rafienia M. Surfactant-assisted sol-gel synthesis of forsterite nanoparticles as a novel drug delivery system. *Materials Sci Eng C Materials Biol Appl* (2016) 58:737–41. doi: 10.1016/j.msec.2015.09.020