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Polyphenols: immunonutrients tipping the balance of immunometabolism in chronic diseases

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Mounting evidence progressively appreciates the vital interplay between immunity and metabolism in a wide array of immunometabolic chronic disorders, both autoimmune and non-autoimmune mediated. The immune system regulates the functioning of cellular metabolism within organs like the brain, pancreas and/or adipose tissue by sensing and adapting to fluctuations in the microenvironment's nutrients, thereby reshaping metabolic pathways that greatly impact a pro- or antiinflammatory immunophenotype. While it is agreed that the immune system relies on an adequate nutritional status to function properly, we are only just starting to understand how the supply of single or combined nutrients, all of them termed immunonutrients, can steer immune cells towards a less inflamed, tolerogenic immunophenotype. Polyphenols, a class of secondary metabolites abundant in Mediterranean foods, are pharmacologically active natural products with outstanding immunomodulatory actions. Upon binding to a range of receptors highly expressed in immune cells (e.g. AhR, RAR, RLR), they act in immunometabolic pathways through a mitochondria-centered multi-modal approach. First, polyphenols activate nutrient sensing via stress-response pathways, essential for immune responses. Second, they regulate mammalian target of rapamycin (mTOR)/AMP-activated protein kinase (AMPK) balance in immune cells and are well-tolerated caloric restriction mimetics. Third, polyphenols interfere with the assembly of NLR family pyrin domain containing 3 (NLRP3) in endoplasmic reticulum-mitochondria contact sites, inhibiting its activation while improving mitochondrial biogenesis and autophagosomelysosome fusion. Finally, polyphenols impact chromatin remodeling and coordinates both epigenetic and metabolic reprogramming. This work moves beyond the well-documented antioxidant properties of polyphenols, offering new insights into the multifaceted nature of these compounds. It proposes a mechanistical appraisal on the regulatory pathways through which polyphenols modulate the immune response, thereby alleviating chronic low-grade

inflammation. Furthermore, it draws parallels between pharmacological interventions and polyphenol-based immunonutrition in their modes of immunomodulation across a wide spectrum of socioeconomically impactful immunometabolic diseases such as Multiple Sclerosis, Diabetes (type 1 and 2) or even Alzheimer's disease. Lastly, it discusses the existing challenges that thwart the translation of polyphenols-based immunonutritional interventions into long-term clinical studies. Overcoming these limitations will undoubtedly pave the way for improving precision nutrition protocols and provide personalized guidance on tailored polyphenol-based immunonutrition plans.

KEYWORDS

polyphenols, Mediterranean phytochemicals, immunometabolism, immunonutrition, pharmacological immunomodulation, senolytics, immunometabolic diseases, precision nutrition

1 Polyphenols: leadingedge immunonutrients

The immune system, a complex interactive network of many different immune cells, mediators, and cellular mechanisms, is highly dynamic in the response to changes in the tissue environment and plays a vital role in the balance between health and disease (1). It generally comprises two lines of defense: the innate (or unspecific) which comprises the physical barriers (e.g. skin, mucosal membranes, commensal microbiota) and several innate immune cells such as neutrophils, macrophages (phagocytes), innate lymphoid cells and nonspecific mediators that rapidly detect antigens, and the adaptive (or specific) immunity that involves B and T cells (2).

Strong evidence links undernutrition to immunosuppression, decreased vaccination efficacy (3, 4) and/or a greater difficulty in recovering from infections, broadly recapitulated during the COVID-19 pandemic (5, 6). On the other hand, overnutrition is closely associated with chronic low-grade inflammation and an increased risk of metabolic diseases (7). Thus, nutritional interventions tagging specific metabolic pathways in immune cells are promising to tackle the increasing prevalence of chronic diseases featuring a dysfunctional immunometabolic status (8), as well as the immunosenescence characterizing the aging process (9).

While it is agreed that immune function relies on an adequate nutritional status to function properly, we are only just starting to understand how the supply of single or combined nutrients, all of them denominated immunonutrients, can redrive the polarization of immune cells towards a tolerogenic or less inflamed immunophenotype (1, 6, 10). Many nutrients fall within the definition of immunonutrients, the most well-known being omega-3 fatty acids, glutamine, arginine, branched-chain amino acids (BCAAs; leucine, isoleucine, valine) and nucleotides (11, 12). Immunonutrition, a branch of precision nutrition, outlines the opportunity to integrate specific nutrients, or foods, in the usual diet (12) and has been drawing the attention of the scientific and medical communities due to its promising health benefits arising from immune system modulation in varied contexts, from individuals undergoing surgical procedures to critically ill patients, subjects with immune-related diseases, the elderly and, in a distinct scope, professional athletes (1, 2). In a multidisciplinary perspective, immunonutrition is defined as the modulation of immune system by nutrients and non-nutritive substances (e.g. antioxidants, prebiotics or probiotics), collectively termed immunonutrients, which are administered in doses above those normally obtained from the diet (1). These molecular compounds display a double function: they act as dietary constituents and, at the same time, may optimize immune responses by improving defense function while maintaining diet and commensal tolerance (1, 12). One may consider immunonutrition as a set of four main mutually dependent concepts: immune system, nutrition, body organ metabolism and the microbiome (1). Besides acting as a physical barrier, the microbiome interacts dynamically with both the innate and adaptive immune system of mucosa-associated lymphoid tissue (MALT) (13). Consequently, it has a chief role in MALT-dependent processes such as oral tolerance induction, cytokine secretion and overall regulation of immune responses. The possibility to reshape microbiota through immunonutrition in the form of functional foods, nutraceuticals and/or dietary supplements, is therefore an exciting approach to switch off oxidative stress and low-grade inflammation present in a plethora of immunometabolic diseases (2, 14).

A wide variety of non-nutritive phytochemicals have shown to benefit immune homeostasis, polyphenols the most-representative ones (12, 15, 16). This group of secondary plant metabolites is a promising class of phytochemicals that hold the potential to simultaneously balance the gut microbiome (14, 17) and the immune system by reprogramming immunometabolic pathways towards the repolarization of immune cells into a tolerogenic, less inflamed phenotype (6). Accordingly, much interest has been created on their potential use as prophylactic or nutritional interventions targeting immunometabolic diseases.

In this work, we aim to shed light on the immunomodulatory effects of polyphenols, leading-edge immunonutrients, on noncommunicable chronic diseases that share immunometabolic impairments, both auto-immune and non-autoimmune mediated. It provides a critical appraisal into their capacity to modulate immunometabolic reprogramming, emphasizing polyphenols' immunomodulatory roles in the maldaptation of organ-specific immune functions as well as their potential use as precise immunonutritional interventions in immunometabolic diseases.

2 Polyphenols: dietary sources, structural diversity and bioactivity

2.1 Dietary sources

The therapeutic potential of plant-based natural compounds and the phytochemicals composing them has been a significant point of interest in the last years. The most abundant and widely distributed bioactive molecules are polyphenolic compounds (PCs) (18). PCs are significantly abundant in a series of foods including olive oil, herbs, vegetables, fruits, seeds, nuts, whole-grain cereals, and wine that are frequently held accountable for the health benefits of the Mediterranean dietary pattern (19). Each of the referred food groups is enriched in specific PCs classes: phenolic acids predominate in cereals and whole-grains such as wheat, oats, rice, corn, and triticale (20); flavones and hydroxycinnamic acids in dried herbs such as oregano and peppermint (21); catechins, hydroxycinnamic acids, anthocyanins, and proanthocyanidins in red wine (22); flavonoids, phenolic acids and dihydrochalcones in fruits such as apples, mangos and pomegranates for instance (23, 24), and anthocyanins in berries, in which they are responsible for their unique pigmentation and aroma (25). In fact, Mediterranean nutritional patterns are associated with the consumption of colorful meals composed of a high variety of plant-based foods whose sensory and nutritional qualities, namely astringency, color and scent partially derive from the PCs composing them (26, 27).

2.2 Chemical structures

Polyphenolic compounds present a phenolic ring as their basic monomer (18). Due to their chemical structure, PCs present strong free radical scavenging capacity which confers them the ability to activate biological antioxidant responses (28, 29). Besides scavenging free radicals, some PCs are also capable of inhibiting the formation and/or activation of their precursors (28, 29). Depending on their chemical structure, origin and biological function, PCs can be divided in different classes, the largest ones being (1) flavonoids and (2) phenolic acids (18). Examples of more narrow classes are (3) tannins, which include pro- and antoanthocyanidins, gallotannins and ellagitannins, (4) coumarins, (5) lignans, (6) quinones, (7) stilbenes, including resveratrol and pterostilbene for instance, and (8) curcuminoids such as curcumin and ginerol analogues (30).

2.2.1 Flavonoids

In plants, flavonoids are responsible for the coloring and aroma of flowers and fruits (24) and the majority are found as glycosides (18). The general structural backbone of flavonoids is C6–C3–C6, the carbon of the C ring on which the B ring is attached to being the determinator of the subgroup the compound belongs to (18, 24). When the link between the B and the C rings is in the position 3, they are isoflavones, and when this link happens in position 4 we stand before neoflavonoids (24). Those in which the B ring is attached to the C one in the position 2 are further classified into different subgroups depending on the structure of the C ring, them being flavanones, flavanonols, flavones, flavonols, flavanols and anthocyanins (24). Figure 1 presents the chemical structure of the most common compounds belonging to flavonoid subclasses.

2.2.2 Phenolic acids

Phenolic acids are PCs that possess one carboxylic acid group, and can be divided into two major subtypes: benzoic acids, which present a skeletal structure C6-C1, and cinnamic acids, whose structure is C6-C3 (31). They are present in innumerous plantbased foods, such as fruits, vegetables, seeds, legumes, cereal and coffee, being mainly in a bound form, such as amides, esters and glycosides (31). The most abundant hydroxycinnamic acid found in food is chlorogenic acid (CGA), which is an ester formed between caffeic and quinic acids (31). On another hand, the most common hydroxybenzoic acids are gallic, vanillic, ellagic, syringic, phydroxybenzoic, and protocatechuic acids (31). These compounds might act as neuroprotective agents through radical-scavenging activity, being useful in the context of chronic diseases associated with oxidative stress (31). Figure 2 presents the chemical structure of the most common compounds belonging to phenolic acids subclasses.

2.3 Bioavailability and bioactivity

The overall bioavailability of PCs is determined mainly by their chemical structure, their absorption, distribution, metabolism, excretion (ADME), the form of administration, and food matrix (32). Pharmacokinetic studies show that PCs classes vary in terms of bioavailability and can be placed as follows: phenolic acids > isoflavones > flavonols > catechins > flavanones, proanthocyanidins > anthocyanins (32–34).

The polyphenolic content of several plants and fruits is greatly affected by exogenous factors (e.g. climatic conditions, culture types, the degree of ripeness), storage, cooking methods and processing mechanisms (33). Besides, food related factors such as the presence of specific macro and micronutrients can as well modify PCs bioavailability and bioactivity (35). Interestingly, it has been recently suggested that the association between PCs and fiber delays their absorption through the gastrointestinal (GI) tract, potentially optimizing their assimilation (36).



FIGURE 1

Chemical structure of the main subclasses of flavonoids, examples of compounds belonging to each subclass and examples of foodstuff containing them. The largest subgroup of flavonoids are flavanols, in which the hydroxyl group is positioned in the C3 of the C ring. Flavanones and flavones display a hydroxyl group in the C5 of the A ring with the difference between them residing in the double bond formed between positions 2 and 3, which is saturated in flavanones. Isoflavones differ from flavones on the position of the phenyl group, being structurally similar to estrogens. Flavanonols present the hydroxyl group linked to the C ring in the position 3, and no double bound between this and position 2. Anthocyanin hydroxyl groups of the A and C rings is what dictates their color. All subclasses can be found in fruits and vegetables. Figure created in **BioRender.com**.



Chemical structure of the subclasses of phenolic acids, examples of compounds belonging to each subclass and examples of foodstuff containing them. The most abundant hydroxycinnamic acid found in food is chlorogenic acid (CGA), which is an ester formed between caffeic and quinic acids. On another hand, the most common hydroxybenzoic acids are gallic, vanillic, ellagic, syringic, p-hydroxybenzoic, and protocatechuic acids. Figure created in **BioRender.com**.

To exert their bioactivity, PCs must be delivered to the GI and absorbed, reach circulation and, posteriorly the target tissues, being subjected to a significant degree of transformation along their journey through the GI tract. As consequence, a single PC is able to generate several different metabolites displaying different activities and properties relatively to the original compound. In general, PCs display low oral bioavailability (5-10%) (37), due to factors such as decreased solubility, the interaction with the food

matrix, difficulties in membrane-crossing, as well as their extensive hepatic and intestinal metabolism and rapid clearance (32). Still, they display a plethora of scientifically proved and extensively documented dose-dependent beneficial effects (38). Accumulating evidence on the health-promoting effects of many PCs make them a topic of interest for scientists, nutritionists, and consumers in general. Their advantageous features, including marked antioxidant, anti-inflammatory, antimicrobial and anti-adipogenic properties grant them great potential to be incorporated in functional foods, nutraceuticals and/or dietary supplements (Table 1). Notably, PCs are closely intertwined to the therapeutic potentiation of the immune system, adding an extra layer of complexity to their pleiotropic actions. Polyphenols immunomodulatory and anti-inflammatory activities correlate to the number, positions and types of substitutions as well as the degree of polymerization based on the chromane ring (53). Moreover, the high degree of hydroxylation in the B-ring of cathecins and anthocyanidins favor metabolic reprogramming and polyphenols' bioactivity (54). In the upcoming section, the impact of polyphenols in immunometabolic reprogramming of both innate and adaptive immune responses will be discussed.

3 Polyphenols and immunometabolic reprogramming: a multiorganelle approach

Immunometabolic reprogramming heavily relies on interorganelle communication and mitochondria, key organelles for cellular metabolism, act as masters regulators of multi-organelle connections and immune cell-fate determination (55, 56). During the immune response, cells shift from metabolic quiescence to an active phase, and the preferential utilization of specific metabolic pathways can dictate immune cells' differentiation towards a pro- or anti-inflammatory immunophenotype depending on their specialization for mounting protective immunity or tolerance to self or external antigens (57).

The interaction among nutrient signaling networks, adenosine triphosphate (ATP) availability, and immunological cues is crucial to meet the energy demands and functional modifications in immune cell metabolism. AMPK and its downstream target, mTOR, serve as central hubs to nutrient availability by sensing intracellular energy levels (AMP/ADP: ATP ratio). In energy-depleted states, activated AMPK typically inhibits mTOR signaling and promotes mitochondrial biogenesis via the peroxisome proliferator-activated receptor-gamma (PPAR γ) co-activator-1 alpha (PGC1 α) signaling axis (58). Consequently, cellular metabolism skews towards increased oxidative phosphorylation (OXPHOS) activity and enhanced expression of genes encoding key mitochondrial enzymes. Conversely, in states of overnutrition, mTOR upregulates protein and lipid synthesis to promote immune cell growth and proliferation (59).

Quiescent immune cells, such as naïve T cells, memory T cells (Tmem), Treg or tolerogenic DCs, alongside M2 macrophages, predominantly favor mitochondria-driven catabolic metabolism characterized by OXPHOS and fatty acid oxidation (FAO) to sustain ATP supply for long-term survival (60, 61). Autophagy, a conserved lysosomal degradation pathway that supports immune cell differentiation, is enforced by AMPK activation, thereby restraining glycolysis and maintaining cellular quiescence (7, 62). Contrariwise, activated immune cells, such as M1 macrophages and effector T cells (e.g. Th1, Th17), exit the quiescence state by metabolizing nutrients to ensure an adequate supply of macromolecules for the energy demands associated with cellular growth (55). They shift the balance towards mTOR activation and aerobic glycolysis as a rapid source of ATP, akin to the Warburg effect, to meet the high nutritional and energetic requirements of

TABLE 1 Bioactivity of polyphenolic compounds.

Bioactivity	Polyphenolic compound	Evidence	Reference
Antioxidant	Catechins	Catechins displayed the most favorable results regarding a series of antioxidant activity evaluation assays	(39)
	(-)-Epigallocatechin-3-gallate from Green Tea	Stimulate nuclear factor erythroid 2-related factor 2 (Nrf2) translocation to the nucleus	(40)
	Kaempferol, Gallic Acid, Resveratrol	Potentiate the activity of enzymes belonging to the endogenous biological antioxidant system, such as catalase, superoxide dismutase, glutathione peroxidase and glutathione-S-transferase	(41-43)
Anti- inflammatory	Hesperidin	Reduce interleukin (IL)-6, tumor necrosis factor alpha (TNF-α) and nitric oxide levels both <i>in vitro</i> and <i>in vivo</i>	(44)
	Dehydroxylated Phenolic Acids: 3,4- dihydroxyphenylpropionic acid, 3-hydroxyphenylacetic acid, 4-hydroxyhippuric acid	Attenuate lipopolysaccharide (LPS)-induced secretion of TNF- α , IL-6 and IL-1 β in human peripheral blood mononuclear cells (PBMCs)	(45)
Antimicrobial	Epigallocatechin Gallate, Tea Polyphenols, A- type Proanthocyanidins	Disturbing the cell wall of specific bacteria, their inner cytoplasmatic membrane, or reducing their motility and biofilm-forming ability	(46-48)
Anti- adipogenic	Vanillic Acid, Catechins, Resveratrol	Adipose tissue "browning" Suppress the expression of genes and transcription factors related to adipogenesis	(49–52)

short-term clonal expansion and effector function (4, 10, 60). For example, mTORC1 sustains aerobic glycolysis and upregulates hypoxia-inducible factor 1 alpha (HIF-1a) expression to support Th17 cell differentiation, counteracting Treg expansion (58). Similarly, a significant transition from OXPHOS to aerobic glycolysis occurs in bone marrow-derived DCs upon Toll-like receptor (TRL) activation, resulting in inducible nitric oxide synthase (NOS)-dependent generation of nitric oxide (NO) and blockade of mitochondrial electron transport (61). Metabolic reprograming of activated immune cells also involves glutaminolysis. Glutamine is converted into glutamate and ketoglutarate, two well-known tricarboxylic acid cycle (TCA) intermediates that support the oxidative metabolism of immune cells, particularly macrophages. A high ketoglutarate/succinate ratio promotes alternative (M2) activation and FAO engagement, while a low ratio strengthens the proinflammatory phenotype observed in classically activated (M1) macrophages (63).

Metabolic rewiring entails significant modifications in mitochondrial biogenesis and dynamics, as well as redox signaling pathways, all of which are crucial for immune function. For instance, the immunometabolism of T cells heavily relies on the continuous dynamic reshaping of mitochondria through fusion and fission events to maintain mitochondrial quality. Memory T cells undergo increased mitochondrial fusion to support OXPHOS and fatty acid oxidation (FAO) metabolism. In contrast, activated effector T cells demonstrate heightened rates of mitochondrial fission and reduced cristae, an adaptation to facilitate aerobic glycolysis (64). Membrane-bound organelles such as mitochondria, endoplasmic reticulum (ER) and lysosomes must establish inter-organelle connections through specialized cytosolic microdomains to facilitate the intersection of metabolic signaling and the utilization of products from one pathway efficiently as intermediates for another (65). ER-mitochondria junction signaling provides a regulatory platform for various overarching immune cellular functions. The mitochondria-ER network brings together signaling components to potentiate mitochondria fission and Warburg metabolism, key events for the rapid recall response of newly activated memory CD8+ T cells (66, 67). Similarly, the activation of NLRP3 spatially correlates to mitochondria-derived reactive oxygen species (mtROS) and excessive mitochondrial fission in ER-mitochondria contact sites of macrophages undergoing glycolytic reprogramming (64, 68, 69). In summary, the dynamic behavior of mitochondria and inter-organelle communication, particularly with the ER network and endolysosomal system, is crucial for enabling immune cells to seamlessly adjust to fluctuations in nutrient availability. This aptitude is vital for effectively meeting the functional demands during immune cell remodeling.

Evidence regarding the immunomodulatory effects of PCs have been significantly emerging in the last decades (8, 70, 71). A main reason relies on the fact that different immune cell populations express various kinds of polyphenols' receptors (72). Examples of immune cellular receptors targeted by PCs include the retinoic acidinducible gene like receptors (RLRs), aryl hydrocarbon receptor (AhR), 67 kDa laminin receptor (67LR), zeta chain-associated 70 kDa protein (ZAP-70), T cell receptor (TCR) $\alpha\beta$, secretory IgM- (sIgM-) B-cell receptor, Toll-like receptor (TLR) 4 (73–79) and Retinoic Acid Receptors (RARs) (80, 81). Upon binding, PCs are able to modulate immune cells metabolism and activity through a multi-modal approach encompassing nutrient-sensing mechanisms, AMPK/mTOR signaling balance, regulation of interorganellar communication and modulation of metabolismepigenetic axis (Figure 3).

3.1 Nutrient-sensing mechanisms

Nutrients not only act as building blocks but also activate nutrient sensing via stress-response pathways and growth factors, essential for immune responses (82). Under amino acid starvation conditions', immune cells activate the amino acid response (AAR)', a cytoprotective signaling pathway that transiently reduce protein synthesis while enhancing stress-induced gene expression (83). AAR pathway is a potent regulator of inflammatory T cell differentiation. Accordingly, glutamine uptake and glutaminolysis largely cooperate in Th1/Th17 inflammatory T cell response (84). Glutamine, the most abundant amino acid in human plasma, is an important substrate of various ATP generating pathways (e.g. glycolysis, OXPHOS) (85). It is transported across the plasma membrane in mammalian cells by different transporters such as the alanine serine cysteine transporter 2 (ASCT2). Gallate-type procyanidin PCB2 3,3 (PCB2DG) polyphenol, a dimer of epicatechin, interacts directly with ASCT2 glutamine transporter and antagonizes glutamine influx, mTOR/HIF-1 pathway, Th1/ Th17 cell production and inflammatory response through interferon gamma (IFNy) and interleukin-17 (IL-17) production.

Sirtuin-1 (SIRT1) serves as another crucial energy sensor. It is activated by NAD+ in nutrient-deficient states and modulates mitochondrial biogenesis by deacetylating and activating transcription factors such as PCG-1a, signal transducer and activator of transcription 3 (STAT3) or the nuclear factor E2related factor (NRF)-2. In CD4+ T cells, SIRT1 impedes the process of differentiation of T lymphocytes into Th17 cells through STAT3 deacetylation (86). Therefore, SIRT1 agonists have emerged as promising pharmacological approaches to broaden the array of current therapeutic options focused on reducing the Th17 profile. Among these possibilities, the deacetylase activator resveratrol stands out as particularly promising. In CD4+ T cells, this polyphenol has been observed to encourage Th2 and Treg polarization, immunomodulatory effects that are linked to the diverse beneficial impacts of resveratrol in various pathologies characterized by imbalanced lymphocyte subtypes ratios (87).

3.2 AMPK/mTOR signaling balance

mTOR and AMPK stand out as two additional master regulators of cellular metabolism, enabling adaptation to challenges of nutrient scarcity or excess, ultimately promoting cell survival. They are intricately linked to cell-specific adjustments in response to metabolic stress, and disruptions in these signaling



FIGURE 3

Polyphenols and immunomodulation: a mitochondria-centered multi-modal approach. Polyphenols impact immunometabolic reprogramming through four regulatory axes: first, they activate nutrient sensing via stress-response pathways and growth factors, essential for immune responses. Second, polyphenols regulate mTOR/AMPK balance and inflammatory responses in immune cells and serve as well-tolerated caloric restriction mimetics. Third, they interfere with the assembly of NLRP3 in endoplasmic reticulum-mitochondria contact sites, inhibiting its activation while improving mitochondrial biogenesis and autophagosome-lysosome fusion. Finally, polyphenols impact chromatin remodeling through modulation of histone deacetylase/acetyltransferase, thereby coordinating both epigenetic and metabolic reprogramming. Figure created in BioRender.com.

pathways are closely associated with various pathological conditions (88). When nutrients are abundant, organisms prioritize fuel utilization to support cellular growth, with mTOR signaling playing a central role in this process. Conversely, upon nutrient depletion, organisms suppress anabolic pathways and promote autophagy via AMPK signaling to adopt a state geared towards preserving the structural and functional integrity of existing cells. Importantly, SIRT1/PGC1 α can exert negative regulation on the phosphoinositide 3-kinase (PI3K)-alpha serine/threonine-protein kinase/(Akt/mTOR) pathway, likely through their influence on the cellular maintenance of autophagy (89). Senolytic drugs can simultaneously upregulate nutrient deprivation signaling (AMPK) and suppress pathways associated with nutrient surplus (mTOR),

consequently boosting autophagic flux (90). Accordingly, caloric restriction mimetics are the most extensively studied metabolic interventions and have long been associated to lifespan extension and immunosenescence improvement (91, 92).

PCs are well-tolerated caloric restriction mimetics due to their ability to activate AMPK, a cellular energy sensor, thus improving mitochondrial turnover (93, 94). Quercetin and fisetin, two wellestablished senolytic drugs, belong to the flavonoid class of polyphenols and are key modulators of immune cell function. In lipopolysaccharide (LPS)-treated macrophages, fisetin inhibited PI3K/AKT/mTOR signaling and inflammatory cytokines secretion (95). In addition, the acetyltransferase inhibitor epigallocatechin-3-gallate (EGCG) was found to downregulate mTOR-HIF1 α signaling, a metabolic checkpoint of Th17/Treg differentiation, leading to the downregulation of glycolysisassociated molecules and inhibition of Th17 differentiation (70). Likewise, carnosol and curcumin effectively inhibit mTOR activation in response to LPS stimulation in human DCs via AMPK-dependent induction of heme oxygenase-1 (HO-1), an important antioxidant enzyme that assist the maintenance of DCs in a tolerogenic state (71).

3.3 Mitochondria and ER-lysosomes interorganellar communication

The NLRP3 inflammasome, a critical junction between innate and adaptive immunity, relies on ER-mitochondria contact sites to facilitate the association of mitochondria-driven ligands, including dysfunctional mitochondria themselves, as upstream signals for NLRP3 activation. Additionally, self-derived or foreign-derived particulates can be endocytosed by lysosomes, leading to membrane damage and further release of cathepsin B, another common upstream signal for NLRP3 activation (96). In LPS-treated microglia cells, quercetin enhances the mitophagic clearance of damaged mitochondria, countering mtROS accumulation and NLRP3 inflammasome activation during the assembly stage (97). Similarly, resveratrol inhibits the acetylated α -tubulin-mediated spatial arrangement of mitochondria and their ER contacts in macrophages. Consequently, it interferes with the assembly of NLRP3 and its adaptor protein, apoptosis-associated speck-like protein containing a caspase recruitment domain (ASC), thereby inhibiting NLRP3 inflammasome activation triggered by mitochondrial damage (98).

Interestingly, it has been proposed that polyphenols can be directly endocytosed into lysosomes, regulating key signaling pathways of phagocytic cells such as macrophages and DCs (99). Accordingly, chrysin (a flavone) and hesperidin (a flavonoid) enhanced lysosomal phosphatase activity in a concentrationdependent manner in LPS-stimulated macrophages (100). Similarly, the senolytic drug fisetin facilitates the autophagosomelysosome fusion and degradation processes in LPS-treated macrophages by regulating a set of genes primarily involved in autophagosome assembly/maturation (95). Comparable effects were observed in LPS-treated DCs, where cocoa procyanidins strongly upregulated gene pathways associated with lysosomal metabolic function and nutrient metabolism, suggesting a significant impact on DC metabolic activity (99).

3.4 Modulation of metabolismepigenetic axis

Recent research has revealed that alterations in metabolic status can coordinate the function of immune cells by influencing epigenetic changes. This regulatory axis between metabolism and epigenetic enables the microenvironment to mold immune cells, and disruption of this process can contribute to the development of various diseases (101). For example, in LPS-stimulated THP-1 promonocyte cells, TLR4 stimulation triggers glucose-dependent ATP production alongside gene-specific chromatin remodeling. Sirt1 deacetylation activates PGC1 α transcriptional activity and orchestrates sequential metabolic reprograming, sensing processes dependent on NAD+, thereby reducing HIF1 α -dependent glycolysis and enhancing PGC1 α -dependent FAO (102). Notably, quercetin upregulates Sirt1/PGC1 α signaling and improves mitochondrial function and morphology (e.g. mtROS, mitochondrial membrane potential, ATP production) in LPSinduced inflammatory macrophages (103).

Chromatin remodeling involves structural changes such as DNA methylation, histone methylation, and acetylation, which greatly impact transcriptional changes of different genes. Several polyphenols have been identified as histone deacetylase (HDAC) inhibitors (EGCG, curcumin, genistein, quercetin), histone acetyltranferase (HAT) activators (genistein) or HAT inhibitors (EGCG, curcumin) (104). For instance, gallic and ellagic acids, along with fisetin, were found to decrease HAT activity in THP-1 cells, resulting in the deacetylation of the p65 subunit of NF- κ B and attenuation of pro-inflammatory cytokine release (105, 106). Moreover, EGCG enhances HDAC activity in Treg cells, leading to suppressed nuclear factor kappa-light-chain-enhancer of activated B cells (NF-kB) signaling and elevated synthesis of the anti-inflammatory IL-10 (107). Finally, treatment with dihydrocaffeic acid (DHCA) led to a decrease in DNA methylation levels in peripheral leukocytes from mice exposed to stressful conditions as well as human and mice peripheral leucocytes exposed to lipopolysaccharide (LPS) in vitro (108).

4 Polyphenols and immunomodulation

4.1 Polyphenolic modulation of innate immunity

In a simplified manner, one might consider that innate immunity includes two distinct components: the cellular system and the non-cellular system (70). As its name suggests, the cellular system is composed of a set of different cell populations, such as granulocytes, monocytes, macrophages, natural killer and dendritic cells (70). On another hand, the non-cellular one includes diverse kinds of mechanisms that range from mucous barriers to signaling pathways (70). Both components act in a synergistic manner in order to prevent pathogens' access to the organism or promote their destruction in case the referred barriers have already been broken (70).

4.1.1 Effects of polyphenols on dendritic cells

Due to their antigen-presenting activity, dendritic cells (DCs) are indispensable for initiating and regulating innate immune responses (70). PCs have been showing to influence several aspects related to DCs, including differentiation and maturation, as well as their antigen presentation and cytokine secretion functions. For instance, resveratrol was shown to regulate the

differentiation of healthy human monocytes from the blood into DCs (109). Analogously, both EGCG and quercetin exerted immunosuppressive effects in bone marrow (BM)-derived DCs, impairing their maturation and their expression of major histocompatibility complex (MHC) (110, 111). An in vitro study has shown that quercetin's inhibition of DC maturation results from downregulated steroid receptor coactivator (Src)/PI3K-Akt-NF-kB-inflammatory pathways (112). Furthermore, EGCG exposure induced apoptosis of blood monocyte-derived DCs from healthy individuals and modulated developing DCs' phenotype by downregulating MHC II molecules and the surface markers CD11c, CD80 and CD83, which are needed for the process of antigen presentation (113). Interestingly, polyphenols of different natures have been shown to possess immunosuppressive properties towards murine BM-derived DCs stimulated with LPS, including curcumin (114), apigenin (115), daidzein (116), baicalin (117), fisetin (118) and silybin (119). These PCs significantly inhibited the expression of surface markers associated with DC maturation such as CD40, MHC II molecules, as well as costimulatory receptors namely CD80 and CD86, in a dose-dependent manner. As a consequence, they impacted the induction of Th1-mediated immune responses. Additionally, the referred study employing curcumin has also reported a decreased production of IL-1 β by DCs, once more repressing their immunostimulant activity (114). Many of these effects seem to derive from the polyphenols' ability to modulate DC metabolism, namely through suppressing mitogen-activated protein kinases (MAPKs) p38, c-Jun-N-terminal kinase (JNK), extracellular regulated kinase (ERK) 1 and 2, and NF-KB activation (111, 114, 115, 119, 120). Analogously, carnosol and curcumin were found to affect AMPK activation and downstream inhibition of the mTOR pathway in lipopolysaccharide-prime DCs (121). The reduced glycolytic flux promoted by the two polyphenols also impacted mitochondria, inhibiting the LPS-induced increase of spare respiratory capacity.

4.1.2 Effects of polyphenols on monocytes and macrophages

Similarly to DCs, macrophages play an important role in antigen presentation mechanisms, as well as tissue inflammation and repair processes (70). Remarkably, the shift between M1 and M2 phenotypes has shown to be influenced by PCs. For instance, in vitro culturing of THP-1 macrophages with a cocoa extract resulted in suppressed M1-mediated inflammation and promoted polarization to M2 (122). A similar effect has been observed with resveratrol regarding tumor-associated macrophages (123). Moreover, quercetin, kaempferol, daidzein, genistein (124) and apigenin (125) have exhibited the ability to reduce proinflammatory cytokines' secretion by these cells. Quercetin has shown to prevent the secretion of IL-6, IL-1β and tumor necrosis factor alpha (TNF-α) by macrophages by suppressing LPS-induced MAPK and ERK activation (126). Plum polyphenols have also been linked to decreased pro-inflammatory cytokines, ROS and malondialdehyde production by RAW 264.7 macrophages treated with monosodium urate through different signaling pathways involving HIF-1, ErbB and Forkhead box transcription factor O (FoxO) (127). A similar effect has been reported for hesperidin which besides decreasing *ex vivo* IL-12 secretion in LPS-stimulated mouse macrophages also suppressed their migration and adhesion properties *in vitro* (128). An interesting study aiming to evaluate the impact of the flavonoids quercetin, naringenin and naringin on the metabolism of cultured human macrophages has highlighted that the flavonoid-mediated immunomodulation derived from glycolytic downregulation, as well as anti-inflammatory reprogramming of the TCA cycle and antioxidant protection (mainly quercetin), membrane modification (naringenin) and osmoregulation (naringin) (129).

PCs are also able to modulate macrophagic ROS production and iNOS activity, as has been reported for curcumin (130, 131), resveratrol (132, 133), EGCG (134, 135), and genistein (136), to name a few.

Polyphenols further seem to improve macrophages' phagocytic capacity. EGCG and curcumin, for instance, have been shown to trigger murine peritoneal macrophages and RAW 264.7 macrophages' phagocytosis *in vitro* (137, 138). The synergistic effect of these two polyphenols together with resveratrol has been demonstrated against glioblastoma and human papillomavirus (HPV)-infected cells, leading to the repolarization of tumorassociated macrophages and tumor suppression (139).

Interestingly, PCs seem to not only influence macrophages but also their precursors -monocytes – as evidenced by an increase in nitric oxide (NO) production by blood monocytes observed in healthy individuals consuming red wine (72). Moreover, blueberry supplementation has been shown to decrease monocyte expression of monocyte-to-macrophage differentiation-associated (MMD) and C-C motif chemokine receptor 2 (CCR2), reducing inflammation in metabolic syndrome patients (140). EGCG prevented monocyte adhesion to cultured endothelial cells from pig pulmonary aortas by reducing the expression of vascular cell adhesion molecule-1 (VCAM-1) and monocyte chemotactic protein-1 (MCP-1) (141).

4.1.3 Effects of polyphenols on neutrophils

As has been observed for DCs and macrophages, studies highlighting the immunomodulatory effects of PCs on this cell population have been arising, particularly regarding their ability to inhibit in vitro neutrophils' oxidative capacity, which correlates with exacerbated neutrophilic inflammation (142, 143). Accordingly, a study performed by Drábiková et al. reported that a series of polyphenols including curcumin, pinosylvin, resveratrol, pterostilbene, piceatannol and N-feruloylserotonin significantly reduced ROS production by human neutrophils in vitro (144). Furthermore, human blood cultured neutrophils' exposed to treatment with grape polyphenols exhibit improved chemokinetic accuracy and motility in association with enhanced CD16 shedding and CD66b expression (145). On another hand, a study evaluating the impact of phenolic acids in a mouse model of colitis exalted the ability of ferulic acid to alleviate the disease by suppressing the formation of neutrophil extracellular traps (146).

4.1.4 Effects of polyphenols on natural killer cells

Natural Killer (NK) cells are recognized by their robust cytotoxicity and lytic activity, as well as effector functions (147).

Contrarily to what has been described above regarding the effects of PCs on macrophages, DCs and neutrophils, which are essentially immunosuppressive, their impact on NK cells appears to have a stimulatory nature, increasing their number and activity. As an example, green tea polyphenols and quercetin are able to promote murine NK-mediated cytotoxicity (148) and lytic activity (149), respectively. Similarly, low-dose resveratrol supplementation has promoted NK cell killing capacity in different experimental contexts (150-152), potentially by activating JNK and ERK (152, 153). Nevertheless, this seems to be dose-dependent since high doses of resveratrol exerted the opposite effect. Similarly, a study performed by Oo et al. reported that luteolin, apigenin and quercetin at doses of 12.5 µg/ml and 25µg/ml significantly increased the NK-cellmediated cytotoxic activity against lung cancer cells (154). Contrastingly, genistein blocks NK cells' activity at low doses (155) but enhances their cytotoxicity at high concentrations (156). These results highlight the dose-dependent behavior displayed by the vast majority of polyphenolic compounds.

In humans, clinical studies showed that blueberry supplementation increases NK cell count in the blood of healthy subjects (157, 158).

4.2 Polyphenolic modulation of adaptive immunity

Alternatively to the innate immune system, the adaptive branch of the immune system involves a unique type of cells - lymphocytes (70). Two primary lymphocyte populations prevail (1): T lymphocytes, which are responsible for cytokines' secretion, cytotoxic destruction of unviable cells and activation of other immune cells, and (2) B lymphocytes, known by their antibodyproducing capacity (70).

4.2.1 Effects of polyphenols on T and B cells

The immunomodulatory potential of PCs goes beyond innate immunity, considerably impacting lymphocyte numbers and functionality. For instance, incorporating EGCG in the diet for one week has proven to elevate T regulatory (Treg) cells' number in mice's spleen, mesenteric and pancreatic lymph nodes (159). Furthermore, these cells were able to repress cytotoxic T cell action and proliferation as well as interferon gamma (IFNy) production (159). A study evaluating EGCG's impact on naïve CD4⁺ T cell differentiation showed that the green tea polyphenol inhibited Th1, Th9, and Th17 differentiation by downregulating the respective transcription factors T-bet, PU.1, and RORyt, while also preventing IL-6-induced suppression of Treg development. These effects were considered to result from downregulation of Signal transducer and activator of transcription p-STAT1 and p-STAT4 for Th1, and p-STAT3 for Th17 cells, as well as inhibition of IL-6induced STAT3 phosphorylation, respectively. Analogously, naringenin displayed the potential to induce Treg cells through AhR-mediated pathways (72) and baicalin has shown to inhibit Th17 cell differentiation both in vitro and in vivo via reducing RARrelated orphan receptor gamma t (RORyt) expression and upregulating Forkhead box p3 (Foxp3) expression (160). Interestingly, EGCG has also shown to induce Treg cells by repressing DNA methylation, inducing Foxp3 and IL-12 expression both in vitro and in vivo (159). These outcomes exalt a novel epigenetic mechanism underlying the polyphenol's immunomodulatory activity associated with DNA methyltransferases inhibition. Moreover, Ning et al. provided new evidence for the effectiveness of the green tea flavonoid in vitiligo treatment via Janus kinase 2 (JAK2) kinase activity inhibition, reducing the protein levels of CD11a, CXCR3, and CCR2 receptors in human T lymphocytes, suppressing their adhesion to melanocytes induced by IFN- γ (161). Importantly, EGCG's immunomodulatory properties are not limited to CD4⁺ T cells. In fact, there are several reports on the flavonoid's competence on increasing CD8⁺ T cell number and activity in tumorigenic contexts (162, 163). Genistein has exhibited a similar effect, while also enhancing CD8⁺ T cell IFNy expression both ex vivo and in vivo, leading to immune stimulation (156).

A study performed by Ramiro-Puig et al. evaluating the effects of a cocoa-enriched diet in the spleen lymphocyte function of young rats reported that a 10% cocoa intake increased lymphocyte proliferation rate, but down-regulated Th2-associated cytokine levels and decreased immunoglobulin (Ig) secretion (164). Additionally, spleen B cell proportion was raised, and Th cell percentage declined (164).

Similarly, auraptene, a citrus fruit-derived coumarin, was able to suppress the activation of murine inguinal lymph node-derived Th1 cells (165). Finally, genistein has also shown to increase the number of both helper and cytotoxic T cells as well as B lymphocytes in rat spleen (166). Likewise, curcumin administration to Min/+ mice increases mucosal CD4⁺ T and B cell numbers by modulating CD28, CTLA-4, STAT and NF-kB expression, preventing the formation of intestinal tumors (167). In addition, through inhibiting STAT4 phosphorylation curcumin has also shown to suppress human CD4⁺ T cells differentiation into the Th1 phenotype (168). Curiously, curcumin's impact appears to depend on the stimulous to which lymphocytes have been exposed, since other studies exalt its immunossupressive activity. For instance, Sharma et al. reported that both resveratrol and curcumin suppressed the activity of concavilin A-stimulated T and B cells by inhibiting their proliferation, antibody production and lymphokine secretion (169). In fact, curcumin's ability to suppress B cell proliferation has also been demonstrated in human Epstein-Barr infected cells (170). Curiously, polyphenoldriven apoptosis of leukemic B cells was shown to correlate with caspase 3 activation, reduced mitochondrial transmembrane potential as well as downregulation of antiapoptotic protein beclin 2 and iNOS expression (171).

5 Polyphenol-based immunonutrition in immunometabolic diseases

Over the past two decades, the pivotal interplay between immunity and metabolism in chronic diseases has become increasingly evident (172). The burgeoning field of Immunometabolism has progressively illuminated how the immune system orchestrates the functionality of key homeostatic systems within tissues, such as the brain, pancreas, liver and adipose tissue. This modulation occurs through the sensing and adaptation to microenvironmental nutrient fluctuations, driving flexibly reprogramming of metabolic pathways in immune cells that greatly impact their polarization towards a pro- or anti-inflammatory phenotype (172, 173). Accordingly, mounting body of evidence progressively appreciates the mobilization of the innate and adaptive immune systems not only in autoimmune diseases featured by the loss of self-tolerance but also in supposedly non-immune pathologies encompassing neurodegeneration and metabolic disorders (174). Consequently, there's a rising interest in immunonutritional approaches aimed at optimizing immune cells functions to enhance effective defense responses while preserving tolerance.

The following sections delve into the characteristics of immunological disturbances within the spectrum of both autoimmune and non-autoimmune metabolic disorders (175). Additionally, it sheds light on the immunomodulatory roles of polyphenols and draws a mechanistical parallel between their effects and the pleiotropic immunomodulatory actions of drugs currently integrated into corresponding therapeutic algorithms (Figure 4).

5.1 Autoimmune immunometabolic diseases

5.1.1 Type 1 diabetes mellitus

Type 1 diabetes mellitus (T1DM) is currently accepted as being a T cell-mediated disease (176). Nevertheless, other adaptive immune cells as well as elements from innate immunity are believed to be involved in T1DM physiopathology.

Namely, T1DM patients display an impaired complement system function (177) and monocytes from these patients display decreased chemotaxis and phagocytic activity (178). Furthermore, infiltration of macrophages (179),neutrophils, and NK cells (180) in the pancreatic Langerhans islands has been detected in NOD mice and human patients. Moreover, hyperglycemia has shown to impair macrophages' autophagic mechanisms (181, 182). Interestingly, insulin has shown to reestablish the normal phenotype in diabetogenic macrophages through Akt and ERK signaling (183), as well as to repress TLRs and CD14 transcription (184). Furthermore, Yu et al. reported insulin's ability to promote phenotype transition of macrophages from M1 to M2 through PI3K/Akt pathways, and PPAR- γ signaling during diabetic wound healing (185).

Nonetheless, cellular infiltrates found on the pancreas of diabetic subjects are also composed of adaptive immune cells, such as CD4⁺ and CD8⁺ T as well as B lymphocytes (176). Remarkably, diabetogenic CD4⁺/CD8⁺ T lymphocytes are more dependent on aerobic glycolysis and rely less on OXPHOS (186). Accordingly, glycolysis inhibition induced terminal CD4⁺ T cell exhaustion in an animal model of T1DM, delaying disease onset (187). Treg cells are also found to be dysfunctional in the pancreatic lymph nodes of T1DM patients (188) and an increase in IL-17-

producing T cells has also been detected (188, 189). Diabetogenic T cells are further characterized by mitochondrial membrane hyperpolarization and dysfunction, resulting in increased ROS levels and diminished ATP production (190).

PCs supplementation has been emerging as a potential therapeutic strategy for alleviating the immune dysfunction characterizing T1DM, in part by improving mitochondrial function. In fact, several kinds of polyphenols have shown to improve mitochondrial function, namely through the activation of the key mitochondrial biogenesis' PGC-1 α , including ursolic acid (191), resveratrol (192), quercetin (193, 194) and olive hydroxytyrosol (195).

Nevertheless, PCs' effects are not limited to mitochondrial function. For instance, a pomegranate peel extract was able to inhibit immune cell infiltration into pancreatic islets (196). Similarly, oral administration of capsaicin to several mice strains showed to attenuate the proliferation and activation of autoreactive T cells in pancreatic lymph nodes, protecting them from disease development (197). The authors considered these effects to be mediated by capsaicin-mediated enhancement of a discreet population of CD11b⁺/F4/80⁺ macrophages in the pancreatic lymph nodes, which express the anti-inflammatory factors interleukin IL -10 and programmed death-ligand 1 (PD-L1). Moreover, procyanidin B2 gallate has been revealed as a suppressor of TNF- α production by activated CD4⁺ T cells by inhibiting their glycolytic function via mTOR- HIF-1α interaction (198). Lastly, a study evaluating the impact of black seeds and garlic intake in diabetic rats demonstrated a significant increase in the blood levels of monocytes and granulocytes, while lymphocyte proliferation was suppressed (199). A similar output was verified when administering fenugreek oil to a rat model of T1DM, which blunted the diabetes-induced increase of pancreatic lymphocyte counts (200).

5.1.2 Inflammatory bowel disease

Inflammatory Bowel Disease (IBD) presents defects in peripheral and intestinal immune function (201). A deep analysis of the peripheral immune system of IBD patients has found decreased numbers of NK cells and B lymphocytes opposing to increased counts of neutrophils and memory CD8⁺ T cells in the blood (202). Besides displaying elevated phagocytosis and cytokine production (201), IBD-associated macrophages also go through the Warburg effect by HIF-1a stabilization and subsequent increased expression of glycolytic enzymes, a process that is modulated by pyruvate kinase 2 (PKM2) (203). The disease further entails gut DCs overactivation, resulting in increased levels of IL-6in the serum and intestine of IBD patients. Decreased numbers of Treg cells in mice peripheral blood and patients' intestinal mucosa, alongside the expansion of Th17 cells and increased production of IL-17 and IL-23 in the intestinal mucosa and lamina propria (LP) have also been detected (201). Interestingly, the expression of pro-inflammatory cytokines by Th17 cells was found to be epigenetically controlled by the glucose transporter GLUT3, which is upregulated in models of IBD (204). Collectively, evidence points to a relevant role of glycolysis in the immunologic dysfunction characterizing IBD.



FIGURE 4

Mechanistical parallelism between pharmacological interventions and polyphenol-based immunonutrition in their modes of immunomodulation. Current therapeutic algorithms of immunometabolic disorders include drugs with pleiotropic immunomodulatory actions shared by a plethora of polyphenol-based immunonutritional approaches. For instance, the anti-diabetic metformin suppresses the production of pro-inflammatory cytokines by immune cells through Nf-KB signaling modulation, the same effect being reported for curcumin and the flavonoid EGCG. The anti-diabetic has also displayed the ability to counteract unbalances in T cell subpopulations by activating AMPK, an effect once again shared by the polyphenol's curcumin and carnosol. Tissue plasminogen activator (tPA), a serine protease used in stroke therapy, was found to improve chemotaxis and phagocytic ability of immune cells through metabolic pathways' modulation, including ERK 1/2 and Akt signaling. An equivalent effect has been reported for curcumin and anthocyanins, due to suppression of PI3K signaling and iNOS enzymatic activity. Through these same mechanisms, curcumin also attenuates immune cell infiltration, exerting an effect similar to the NMDA receptor antagonist memantine, an anti-dementia drug widely used in Alzheimer's disease, which blocks T cells' potassium channels. Minocycline, a tetracycline antibiotic currently being studied as a therapeutic strategy for stroke, prevents DCs and microglia cells from excessive activation by modulating the JAK/STAT signaling pathway. Likewise, anthocyanins are suggested to modulate the PI3K/Akt/Nrf2/HO-1 axis in DCs and microglia cells, suppressing their overactivation. This is also achieved with the polyphenol hydroxytyrosol, which reduces MAPK activation. A similar mechanism has been observed for polyphenols belonging to different classes, such as resveratrol and kaempferol, which also regulate PPARy, inhibiting macrophage and microglia polarization towards a pro-inflammatory (M1) phenotype. These polarization-shifting properties are likewise reported for the flavonoid phloretin and are considered to be mediated by an increased AMPK expression. These effects are analogous to the ones of insulin, which is directed to T1DM patients and interferes with PI3K/Akt and PPARy signaling, and to the decarboxylase inhibitor carbidopa, which promotes M2 macrophage polarization in the context of Parkinson's Disease by suppressing glucose uptake. Under environmental stimuli, immune cell activation occurs accompanied by metabolic reprogramming. In most cases, this primarily consists of a transition from mitochondrial OXPHOS to aerobic glycolysis. Drugs purposed for Multiple Sclerosis treatment, such as dimethyl fumarate, IFNy and glatarimater acetate, among others, are known to modulate this shift, suppressing glycolysis and promoting OXPHOS in T cells. The same is observed for methotrexate – an antimetabolite used to treat IBD. Analogously, by interfering with mTOR signaling and PKM2 activity, flavonoids such as shikonin and procyanidin B2 gallate, respectively, also modulate immune cell metabolic reprogramming in the context of immunometabolic dysfunctions. Figure created in BioRender.com.

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Accordingly, therapy with methotrexate, which is used for different autoimmune conditions including IBD, appears to suppress glycolytic mechanisms in varied immune cell populations (205), counteracting the metabolic reprogramming associated with disease pathophysiology.

Furthermore, intestinal barrier function is also impaired in the context of the disease, presenting less mucus secretion by goblet cells, reduced antimicrobial peptides (AMPs)' production by Paneth cells and several mutations in genes coding tight junction proteins, resulting in their dysfunction and consequent loss of barrier integrity (201). Defects in mucosa mitochondrial function are also a feature of IBD, including reduced complex I activity, membrane potential, biogenesis, OXPHOS, TCA cycle and fatty acid metabolism alongside increased mitochondrial fragmentation due to fission (206, 207).

Furthermore, the disease is characterized by increased susceptibility to dysfunctional autophagy of macrophages, DCs, Paneth cells and GCs (208). Particularly, Autophagy Related 16 Like 1 (ATG16L1) gene deficiency in macrophages increases the risk of Chron's Disease development (209) and suppresses DCs' ability to induce Treg cells in contexts of intestinal inflammation (210).

Remarkably, a part of PCs' beneficial effects regarding IBD is related to their impact on autophagy. As an example, the flavonoid galangin has shown to alleviate DSS-induced colitis' symptoms in mice by increasing the expression of autophagy-related proteins and promoting colonic autophagosome formation (211). On another hand, resveratrol displayed autophagy-promoting properties in cultured macrophages through sirtuin modulation (212, 213), highlighting its potential to counteract the macrophagic autophagy dysfunction underlying IBD.

PCs further display relevant potential to maintain intestinal homeostasis by protecting the intestinal barrier. Several studies evaluating the impact of polyphenolic supplementation in experimental models of the disease have revealed improved gut barrier function, consequently limiting inflammatory cell infiltration. This has proved to be true for grape seed PCs which increase colonic goblet cell density and mucin 2 mRNA expression (214); anthocyanins by enhancing tight junction molecules (zonulin-1, claudin-1, occludin) and Muc 1/2 expressions (215), just to mention a few. Interestingly, resveratrol and resveratrolrelated PCs (e.g. pterostilbene) have further demonstrated to alleviate intestinal inflammation in mice with colitis by regulating the Th17/Treg balance and control the levels of plasmatic and intestinal mucosal cytokines such as transforming growth factor beta (TGF-β), IL-6, IL-10 and IL-17 (216, 217), restoring the percentage of CD4⁺ T cells in mesenteric lymph nodes (MLNs) and decrease their number in the intestinal LP, as well as reducing the percentage of macrophages in both regions (218). Similarly, curcumin appears to promote colonic Treg cell expansion while decreasing the counts of inflammatory DCs; inhibiting proinflammatory cytokines' secretion, T cell infiltration and NF-kB activation (219), as well as to suppress macrophage activation and regulate M1/M2 polarization (220). Chlorogenic acid has also shown to mitigate DSS-induced colitis in mice by inhibiting M1 macrophage polarization through suppressing PKM2-dependent glycolysis and Nod-like receptor protein 3 activation (146).

Moreover, a study performed by Wu et al. reported that the lignan arctigenin inhibits Th17 and Th1 differentiation *in vitro* by repressing STAT3 and STAT4 phosphorylation respectively through mTORC1 downregulation, ameliorating DSS-induced colitis in mice (221).

On another hand, shikonin – a polyphenol widely used in Chinese traditional medicine – has shown to suppress glucose consumption and lactate production as well as inhibit the nuclear translocation and enzymatic activity of PKM2, which is responsible for stimulating the Warburg effect in macrophages, in a DSSinduced colitis mouse model (222).

Furthermore, PCs are known for inducing short-chain fatty acids (SCFAs) production by the gut microbiota, namely butyrate (223, 224), which displays several gut health-promoting properties: it promotes colonic mucus production (225); potentiates the extrathymic conversion of CD4-positive T lymphocytes into Treg cells; is able to reduce mTOR activation and glycolysis in intestinal macrophages, while simultaneously promoting their metabolic reprogramming to OXPHOS and lipid metabolism (203) as well as downregulating their expression of pro-inflammatory cytokines (226). Remarkably, some of the aforementioned effects equally emerge from methotrexate therapy for CD, which elevates OXPHOS in T cells by activating AMPK and blocking mTORC1 (227).

In humans, consumption of mango by IBD patients significantly improved Simple Clinical Colitis Activity Index (SCCAI) score and decreased the plasma levels of pro-inflammatory cytokines related to neutrophil-induced inflammation (228).

5.1.3 Multiple sclerosis

Similarly to T1DM and IBD, Multiple Sclerosis (MS)' pathophysiology is characterized by a series of immunological alterations, the most well-known pathophysiological components of the disease (229). Although the primary events leading to the autoimmune attack characterizing MS are not yet established, a possible explanation is based on molecular mimicry consisting of the activation of autoreactive T lymphocytes through cross-reactivity by viral and/or bacterial antigens structurally similar to central nervous system (CNS) proteins, such as myelin basic protein (MBP), myelin oligodendrocyte glycoprotein (MOG) and proteolipid protein (PLP) (229). These cells migrate to the CNS fueling neuroinflammatory events that promote BBB opening allow a second wave of immune cells to access the CNS, namely CD8⁺ T cells, B lymphocytes and macrophages (230). Macrophages within the perivascular cuff of post-capillary venules of animals with EAE display altered metabolism featured by increased expression of the glycolytic enzyme lactate dehydrogenase (LDHA) as well as monocarboxylate transporter-4 (MCT-4), specialized in secreting lactate from glycolytic cells, potentially inducing macrophage infiltration in the CNS (231). Analogously to what is described for T1DM, activated CD4⁺ T cells from MS patients display an up-regulation of aerobic glycolysis and down-regulation of OXPHOS (232), as well as altered mitochondrial structure, mitochondrial DNA (mtDNA) levels and membrane potential (233). Furthermore, abnormal expression of autophagyrelated markers and genes has been found in T cells from MS patients and EAE animals (234). It is worth noting that the role of autophagy in MS pathophysiology is controversial since there is evidence of both protective and deleterious effects of autophagy

induction in immune cells in the context of the disease (234), but there seems to be a consensus regarding the fact that autophagy contributes to MS pathology in macrophages, DCs, T and B cells while having a protective role in neurons and glial cells (235).

On another hand, there is a decreased count of circulating Treg cells in MS patients (236), which also seem to play an important role in EAE development (237). In fact, the immunometabolism of T cells is extremely relevant in the pathophysiology of MS, as evidenced by the fact that several of its therapeutic strategies modulate T cell metabolic features: Dimethyl Fumarate suppresses glycolysis; IFNy decreases ATP levels, mitochondrial membrane potential and modulates OXPHOS; Teriflunomide limits T cell activation by blocking mitochondrial respiratory chain's complex III; and Glatiramer Acetate promotes OXPHOS and represses glycolysis in CD4⁺ T cells (238). Likewise, a study evaluating the effects of cinnamic acid in EAE reported that the polyphenol acted as an MCT-4 inhibitor, attenuating immune cell infiltration into the CNS, suppressing glycolysis and lactate production by macrophages and ultimately reducing disease severity (231). The EAE-associated inflammatory phenotype of macrophages has also been reduced by the dihydrochalcone phloretin, which activated Nrf2 by stimulating AMPK-dependent autophagy (239). Additionally, a large number of PCs have shown to attenuate EAE clinical severity or inhibit its development by reducing immune cell infiltration, referring to EGCG (240), curcumin (241) and hesperidin (242). The latter two also seem to impact the Th17/Treg balance in EAE animals, promoting Treg cell expansion and Th17 suppression in the spleen (241), lymph nodes (240, 242) and the CNS, accompanied by repressed proinflammatory cytokine secretion (240, 241). Analogously, EGCG as well as naringenin for instance are known to impact the Th1mediated immune response associated with EAE (240, 243). The described outcomes are potentially attributed to the impact of PCs on the expression of transcription factors associated with each of the referred T cell subsets: Foxp3 for Tregs (242, 243), RORyt for Th17 cells (241-243), and T-bet for Th1 ones (243). Inhibition of Th17 cells' differentiation by curcumin further entails down-regulating IL-6 and IL-21 as well as STAT3 phosphorylation (241). Considering that MS is a T cell-mediated disorder, inhibition of CD4⁺ T cells' activation might comprise a promising therapeutical strategy. Interestingly, curcumin has shown to induce human T cell death through increased expression of ER stress-related transcriptional factors (244). Analogously, resveratrol inhibits CD4⁺ T cells' activation and cytokine production by promoting SIRT1 expression and activity both in vitro and in vivo (245). The stilbene has further been highlighted as able to counteract the decline in brain mitochondrial function characterizing the cuprizone-induced demyelination model by enhancing cytochrome oxidase activity and elevating ATP levels (245).

5.2 Non-autoimmune immunometabolic diseases

5.2.1 Obesity and type 2 diabetes mellitus

Findings arising from pre-clinical and clinical research have been elucidating the mechanisms of immunological dysfunction

associated with obesity and type 2 diabetes mellitus (T2DM). Regarding innate immunity, metabolic dysfunctions are characterized by an altered neutrophil functionality, increased M1 macrophage and inflammatory DCs numbers, and abnormal NK phenotypes (246). In obese individuals, neutrophils display augmented chemotaxis and non-directed migration, as well as increased basal levels of superoxide, while neutrophils from diabetic subjects lose a variety of their functions, including migration capacity, phagocytosis and ROS production (246). By increasing leptin levels, obesity alters adipose tissue macrophages' (ATMs) metabolism through Janus kinase 3 (JAK3) and STAT3, and PI3K-Akt-mTOR pathways, increasing glycolytic enzymes' activity and glucose uptake as well as inducing mitochondrial dysfunctions (247). PI3K-Akt-mTOR activation in brain macrophages of diabetic rats has further been implicated in autophagy impairment (248), which originates protein aggregates and fosters damaged mitochondria due to defective mitophagy (249). Dysfunctional mitochondria accumulation leads to increased ROS production and consequent NLRP3 dependentinflammation by macrophages in both T2DM (250) and obesity (251). Mitochondrial dynamics are likewise affected in the context of both disorders, as evidenced by induced activation of the fission regulator dynamin-related protein 1 (Drp1) by a high-fat diet (252) as well as increased mitochondrial fission and decreased fusion in leukocytes from T2DM patients (253).

NK cells are also dysfunctional in contexts of obesity and T2DM, displaying increased proliferation rates and IFN γ secretion, and impaired degranulation, respectively (246). A dysfunctional mTOR function has also been observed in NK cells from obese patients (254). Furthermore, DCs activation and maturation is promoted in cases of diabetes, and obesity-associated DCs present an inflammatory phenotype triggering Th17 cells' activation (246).

Adaptive immunity is likewise affected by metabolic impairments, resulting in increased numbers of $\gamma\delta$ T, Th17 and Th22 cells and a reduction in Tregs (246). B cells display altered functionality, promoting an abnormal antibody response (246). In obesity, CD4⁺ T cells also reveal a distinct metabolic profile characterized by the activation of glycolysis and OXPHOS (255). Furthermore, mitochondria from T2DM patients' CD8⁺ T cells display higher oxidative capacity together with elevated ROS levels and fatty acid uptake as well as decreased FAO and AMPK activity (256). In fact, metformin, a widely used oral antidiabetic, has shown to facilitate T cells' shift from a glucose-dependent anabolic state to a catabolic one through mTOR signaling blockage and by restoring mitochondrial FAO (257). Furthermore, it shuts down glycolysis and promotes OXPHOS by activating pathways involving carnitine palmitoyltransferase (CPT)-1 alpha and PGC-1 α (258).

The impact of PCs in immune system dysfunctions associated with obesity and T2DM have been consistently highlighted in both *in vitro* and *in vivo* experiments. Analogously to metformin, a variety of polyphenol formulations as well as isolated compounds are described as PGC-1 α inducers in the context of T2DM and obesity, such as ginger polyphenols 6-gingerol and 6-chrysophanol (259), epicatechin-enriched cocoa (260), sudachitin (261), and EGCG (262). The latter has also shown to inhibit T2DM- associated mitochondrial deficiency and dysfunction in diabetic Goto-Kakizaki rats by suppressing enhanced autophagy in muscle cells (263, 264). Furthermore, resveratrol administration to older adult diabetics showed to improve mitochondrial biogenesis and function through SIRT1 upregulation, alleviating the oxidative damage and promoting insulin sensitivity (265).

On another hand, PCs that include capsaicin, curcumin, and anthocyanins for instance, have shown to attenuate macrophage migration (266-268), in part by suppressing MCP-1 expression (268, 269). Curcumin has also displayed relevant suppressive effects on NFkB signaling in immune cells, leading to a reduction in iNOS expression by macrophages and DCs (270), as well as neutrophils (271). Neutrophils chemotaxis is also apparently impacted by curcumin's ability to suppress PI3K activity and Akt phosphorylation (271). On another hand, evidence suggests that PCs present in the small fruit lingonberry promote macrophage polarization to an anti-inflammatory (M2) phenotype by upregulating PPARy and STAT6 phosphorylation in experimentally induced obesity (272). Quercetin has also shown to abolish NLRP3 inflammasome activation in macrophages by upregulating Akt signaling, reducing insulin resistance in mice with particulate matter-induced metabolic disorder (273). A similar effect has been reported for red raspberry polyphenols (274).

Regarding adaptive immunity, studies employing cafeteria dietinduced obesity as well as the alloxan-induced model of diabetes in rats have reported that PCs intake lowered the production of proinflammatory mediators including ILs, TNF α , IFN γ and TGF- β by MLN and splenic lymphocytes (275, 276). Contrasting to what is observed for macrophages, PCs seem to promote Treg cell recruitment, namely through elevation of Foxp3 gene expression (277). A study employing EGCG in the context of murine dietinduced obesity has reported an increased Treg/Th17 cell balance by decreasing the ratio of STAT3/STAT5 expression (278).

5.2.2 Neurological diseases

Evidence from genome-wide association studies highlight the association between immune cells-mediated inflammation and increased risk of neurodegeneration (279). Most neurodegenerative diseases involve deposition of misfolded proteins, leading to aggregate formation and consequent neuronal loss (279). The initial phases of the referred disorders are characterized by the activation of the immune system and neuroinflammation, partially mediated by a CNS resident macrophage cell population - microglia - that are activated in virtually all neurodegenerative conditions (279). Despite the pivotal role of microglia cells, infiltrations composed of astrocytes, monocytes and/or lymphocytes are also frequent in these contexts (279). Immune cell dysfunctions in the mitochondrial respiratory chain are likewise preponderant features of neurodegenerative diseases, evidence suggesting that leucocytes, neutrophils, monocytes/macrophages, and T cells display increased levels of ROS and NO accompanied by elevated mitochondrial membrane potential and decreased complex activity (238).

Considering the above, previously mentioned reports of PCs ability to limit immune cell activation and cellular infiltration point to these natural substances as promising therapeutic agents in the context of neurological disturbances.

5.2.2.1 Alzheimer's Disease

Alzheimer's Disease (AD) is the most common neurodegenerative disease worldwide (279, 280). Its main features include β -amyloid protein (AB) deposition and tau protein hyperphosphorylation, originating senile plaques and neurofibrillary tangles (NTFs), respectively (279, 280). The referred aggregates promote microglial activation, which surprisingly appears to play a dual role in disease pathophysiology (280). Initially, activated microglia seem to have a beneficial effect by phagocytizing excessive AB, but as the disease progresses, they may lose this ability and acquire a dysfunctional senescent phenotype or become neurotoxic by remaining chronically activated (280). The inflammatory mediators produced by these cells stimulate an analogous response on astrocytes, resulting in neuronal death (280). Furthermore, brain parenchyma infiltration by neutrophils and NK cells also seems to contribute to the neuroinflammatory changes reported in AD (279, 280). On another hand, the role of adaptive immune system in the disease is controversial, since there are studies supporting a neuroprotective role for the adaptive immune cells in AD animal models (281) while others exalt its requirement for disease progression (282). Noticeably, numerous lines of evidence on the impact of metabolic perturbations in microglia mediated-neuroinflammation in AD have been arising. Actually, the age-related decline in glucose metabolism in the brain is associated with cognitive dysfunctions in AD patients (283). A β deposition seems to induce mTOR phosphorylation and HIF-1 α expression by microglia, originating inflammatory cascades (283). Microglial cells adopt a neuroimmunomodulator phenotype, exhibiting ineffective glycolysis, and TCA cycle accompanied by impaired chemotaxis and phagocytic ability (283). Furthermore, AD features a great degree of mitochondrial dysfunction with concomitant cardiolipin exposure, leading to increased microglial phagocytosis and synthesis of inflammatory mediators, fostering neuroinflammation (284). Moreover, damaged mitochondria release mtDNA which can induce the NLRP3 inflammasome and the NF-kB pathway, exacerbating inflammation (284). AD-associated microglia further display a suppressed autophagic flux due to a reduced expression of key regulatory proteins such as Beclin-1 (285).

Interestingly, Chen et al. has reported that microglial cells mediate T cell infiltration in experimental models of AD as well as human brains, driving neuroinflammation (286). Accordingly, memantine – a drug approved for the treatment of advanced AD – acts on T cell metabolism by blocking potassium channels, normalizing these cells' response (287).

Likewise, PCs exhibit vast potential as therapeutic agents in neurological pathologies by acting on different strands of their etiology, among which are immune dysfunctions. Interestingly, oleuropein aglycone – a polyphenol abundantly present in extra virgin olive oil – is reported to induce autophagy in AD mouse models by modulating AMPK signaling (288, 289) as well as sirtuin activity and histone acetylation (288). Likewise, curcumin has shown to downregulate PI3K, phosphorylated Akt and mTOR protein levels, inducing autophagy in brain tissue of APP/PS1 double transgenic AD mice (290).

Resveratrol, which is currently under investigation in several clinical trials for AD, has demonstrated to possess remarkable

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immunomodulatory properties in immune cell populations highly relevant in the context of the disease. It inhibited microglia activation, proliferation and cytokine production (IL-6 and TNF- α) (291) and promoted its polarization towards an antiinflammatory phenotype in animal models (292), suppressing neuroinflammation. Furthermore, a retrospective study with AD patients demonstrated that treatment with resveratrol induced adaptive immune responses, increasing IL-4, fibroblast growth factor (FGF) 2 and macrophage-derived chemokine (MDC) secretion by macrophages (293). Still regarding microglia cells, anthocyanins were also able to mitigate oxidative stress and neurodegeneration in a mouse model of AD by modulating the PI3K/Akt/Nuclear factor erythroid 2-related factor 2 (Nrf2)/heme oxygenase 1 (HO-1) axis (294). Additionally, supplementation of cultured microglia exposed to $A\beta$ with a polyphenol abundantly found in extra virgin olive oil named hydroxytyrosol attenuated mitogen-activated protein kinases (MAPKs) activation as well as ROS generation (295). The flavonoid baicalein has also shown to inhibit microglia-induced neuroinflammation in a mouse model of AD by suppressing NLRP3 activation and the TLR4/NF-kB pathway (296). Similarly, a study performed by Kim et al. described gallic acid's profile as a histone acetyltransferase inhibitor, highlighting its ability to inhibit NF-kB acetylation and reducing cytokine production by cultured microglia (297).

5.2.2.2 Parkinson's Disease

On the frequency ranking for neurodegenerative diseases, Parkinson's Disease (PD) follows AD at second place and is characterized by the accumulation of α -synuclein in neurons, glial cells, and nerve fibers (279). The histopathological hallmarks of PD include loss of dopaminergic neurons in the *substancia nigra pars compacta* (SNpc), presence of activated microglia, astrogliosis and lymphocytic infiltration (279).

Similarly to what happens in AD, the accumulated protein aggregates promote microglia activation, which proceed to release excessive amounts of neurotoxic factors generating a selfamplifying cycle that contributes to progressive neuronal degeneration (279). PD-associated microglia also display impaired mitochondrial function associated with mutations in genes involved in mitophagy and oxidative stress such as Pink1 and Parkin, resulting in inflammasome activation that fosters dopaminergic neurodegeneration (298, 299). Interestingly, research in PD experimental models has shown that NLRP3 inflammasome activation is exacerbated in microglia cells deficient in autophagy related protein 5 (ATG5) (300), highlighting autophagy's relevance in suppressing inflammation. Nevertheless, the role of other immune cells such as astrocytes and NK cells in this inflammatory cascade remains to be enlightened (279, 280). On another hand, the expansion of dysfunctional monocytes appears to be an essential element in PD pathogenesis and might be related to the secretion of inflammatory mediators by microglia cells as well as pro-inflammatory monocytes' recruitment to the brain, fomenting neuroinflammation (279, 280). Furthermore, evidence suggest that M1 macrophages' activation is linked to disease susceptibility and progression (301). Accordingly, the decarboxylase inhibitor carbidopa that is used for PD management has been shown to favor macrophage differentiation to an M2 phenotype (302).

Contrasting to the occurring in AD, the role of the adaptive immune system in PD's pathophysiology is becoming clearer. In fact, numerous authors have found adaptive immune populations, namely Th17 cells, in PD patients' brain samples (303). In parallel to what is observed for AD, glucose hypometabolism has been implicated in disease pathophysiology, being associated with the development of dementia occurring in the brain cortex (283). Furthermore, PD is characterized by deregulation of several glycolytic enzymes and transporters such as pyruvate dehydrogenase kinase 1 (PDK1), PKM2, LDHA, GLUT1, MCT-1 and MCT-4, as well as increased mitochondrial respiratory activity and oxidative damage in neurons (283).

Recently, PD supplementation with PCs has been drawing attention. Once more, resveratrol exposes its neuroimmunomodulatory properties, being able to suppress microglia activation and decrease the levels of TNF- α , IL-1 β and IL-6 and their receptors' expression in the SNpc of mice with 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine (MPTP)-induced PD (304). Analogously, curcumin administration has shown to inhibit microglial morphological alterations in an *in vitro* model (305). Curcumin has further demonstrated protective effects against neurodegeneration in the A57T α -synuclein model of PD by downregulating mTOR/p70S6 kinase (P70S6K) signaling and recovering macro autophagy (306). Accordingly, researchers employing a nanoformulation of α -mangostin discovered that the polyphenol reprogrammes microglia metabolism from glycolysis to OXPHOS and promotes its autophagic capacity, increasing microglial A β clearance (307).

Regarding the mitochondrial dysfunction featuring the disease, morin and mangiferin displayed the ability to attenuate membrane potential loss in neurons (308), as well as quercetin which also enhanced mitophagy by upregulating Pink and Parkin gene expression (309).

On another hand, a study evaluating the impact of genistein in dopaminergic neurodegeneration reported a dose-dependent inhibition of neuronal loss in rats' glial cells (310). The same authors reported the soybean isoflavone's ability to suppress microglia cell activation as well as NO and superoxide production by these cells (310).

It is worth noting that evidence regarding the importance of the gut-brain and spleen-brain axes in PD has been emerging, suggesting an involvement of the intestinal and splenic immune systems in this disease development (311). Wang et al. evaluated the impact of chicory acid in mice with MPTP-induced PD and verified that this PC prevented dopaminergic brain lesions and glial activation, simultaneously reverting the disease-induced alterations in IL-17, IFN- γ and TGF- β levels in both the spleen and colon (311).

5.2.2.3 Stroke and stroke-induced neurodegeneration

Stroke is one of the global leading causes of disability and mortality (312, 313). It is currently acknowledged that the immune system is an active player in stroke's pathogenesis, possibly causing subsequent damage which are collectively designated as stroke-induced secondary neurodegeneration (SND), a condition that shares numerous features with AD, namely A β accumulation (314).

Similarly to AD and PD, stroke-associated neuroinflammatory events include microglial activation and consequent release of neurotoxic mediators, as well as stimulation of macrophages and DCs (315). Stroke-associated microglial cells display dysfunctional phagocytosis and chemotaxis, severely compromising neuroinflammation resolution and neurorestoration (316). Furthermore, after stroke events activated M1 microglia cells display enhanced mitochondrial fission, leading to NF-kB and MAPK activation which induces pro-inflammatory mediators' expression (317). These cells have also shown to release damaged mitochondria to neurons where they fuse with neuronal mitochondria, damaging them and promoting mitochondria-mediated neuronal death (318). Additionally, stroke-associated microglia feature increased autophagy in associated with an enhanced inflammatory response (319-321). In agreement, treatment of permanent middle cerebral artery occlusion (pMCAO) mice with an autophagy inhibitor alleviated the inflammatory response, while an autophagy inducer exerted the opposite effect (320).

NK cells and CD4⁺, CD8⁺ and $\gamma\delta$ T lymphocytes have likewise revealed to be involved in stroke's initial stages, with B cellmediated neurodegeneration becoming prominent later on in the disease course (315). Therefore, components of immunoreactivity can be found in each phase of stroke pathology. Another similarity between AD, PD and stroke is the metabolic reprogramming of microglia cells shifting from OXPHOS to glycolysis (322). In fact, stroke brains display increased concentrations of numerous glycolytic intermediates, including glucose-6-phosphate, fructose-6-phosphate, LDHA, PKM2, pyruvate, and lactate (322). Furthermore, blocking microglial hexokinase-2, the enzyme responsible for glucose phosphorylation into glucose-6-phosphate, has shown to suppress their activation and reduce the infarct area in male Sprague-Dawley rats subjected to transient middle cerebral artery occlusion (323), highlighting the role of glucose metabolism in microglia-mediated neuroinflammation characterizing stroke. Interestingly, the ability of already existing drugs to modulate macrophage and microglia metabolism in the context of stroke is being studied (324). Minocycline, which has exhibited neuroprotective activity in the context of stroke (325), has shown to promote microglia polarization from an M1 to an M2 phenotype through STAT1 and STAT6 pathways (326). Moreover, studies suggest that tissue plasminogen activator (tPA), a widely employed fibrinolytic agent in stroke therapy, is able to normalize microglial chemotaxis and phagocytosis through metabolic pathways' modulation, including Akt and ERK 1/2 signaling (327, 328).

Likewise, PCs immunomodulatory effects have been demonstrated in situations of stroke and SND as well. For instance, a study evaluating fisetin effects in a mouse model of ischemic stroke highlighted the flavanol's ability to inhibit postischemic infiltration of macrophages and DCs as well as repress the intracerebral activation of immune cells (329). In addition, fisetin shown to suppress TNF- α production by macrophages and microglia cells *in vitro* (329). Regarding PCs' impact on microglia, a study performed by Lan et al. showed that the flavanone pinocembrin was able to suppress microglia activation and consequent production of IL-6, IL-1 β and TNF- α (330), while also decreasing the expression of TLR4 and its downstream target proteins TRIF and myeloid differentiation primary response 88 (MyD88) (330). Similarly, both curcumin (331) and baicalein (332) appeared to ameliorate ischemic brain damage a by modulating microglia polarization and suppressing TLR4 and NF-kB signaling. Gallic acid has also shown to induce microglia M2 polarization in a MCAO mouse model (333).

PCs further seem to impact stroke-associated microglia mitochondria dysfunction, as evidenced by the reduced postischemia neuronal mitochondrial damage resulting from kaempferol administration to rat PC12 cells (334). This effect was considered to derive from an upregulated SIRT1 expression alongside to inhibited gene acetylation of the pro-apoptotic protein P66shc as well as Drp1 recruitment.

Remarkably, recent research has demonstrated that consumption of resveratrol after stroke events might exert neuroprotection through gut-brain-axis modulation (335). In fact, the authors determined that the polyphenolic supplementation promoted a polarization shift of Th cells from Th1 to Th2, reducing intestinal inflammation and vascular permeability, which culminated in mitigation of inflammatory brain lesions (335).

6 Conclusion and future directions

In recent years, the term "immunometabolism" has gained traction within the scientific and research communities as a descriptor of the interface between the immune system and metabolism. The disruption of such complex interactions is increasingly recognized as a common denominator of a wide range of socioeconomically impactful diseases of both autoimmune and non-autoimmune nature (172). The escalating prevalence of immunometabolic disorders and the intricate interplay between metabolic irregularities and scenarios of chronic inflammation underscore the imperative to unravel the mechanisms that dictate the programming of immune cell metabolism. In fact, cells from the immune system display unique energy requirements depending on their activation state, anabolic and catabolic mechanisms, being associated with pro- and anti-inflammatory responses, respectively (336). Therefore, modulating immune cells' metabolic pathways through their respective energetic substrates may significantly impact disease outcome. From this perspective, nutritional interventions emerge as promising tools within the realm of preventive and/or adjunct therapeutic approaches, framing the concept of immunonutrition, a branch of precision nutrition aimed to fine tune pro- or anti-inflammatory immunophenotypes through personalized protocols tailored to individual requirements, health status and metabolic variability (57, 337).

Within the context of immunometabolic disorders, polyphenols stand out due to a wealth of evidence supporting their potential health benefits. Remarkably, varied polyphenols have shown to exert immunomodulatory effects, such as curtailing immune cell hyperactivity and rebalancing pro- and anti-inflammatory T cell subsets, naming a few (70). Particularly, these bioactive compounds are reported to influence immune cell metabolic reprogramming, driving a tolerogenic phenotype and mitigating inflammation, thus showcasing significant potential as key immunonutrients. Such outcomes stem from their influence on nutrient-sensing pathways primarily involved in processes like glycolysis, mitochondrial biogenesis and dynamics as well as mitochondria-ER-lysosome inter-organelle connections, leading to epigenetic and metabolic reprogramming that yield diverse immunomodulatory effects across different cell populations as depicted in Figure 3 (338). It is worth emphasizing that a substantial portion of these regulatory pathways is similarly influenced by the pleiotropic effects of drugs included in current therapeutic algorithms for the aforementioned diseases (as illustrated in Figure 4), encouraging further exploration on how to best leverage polyphenols as immunonutrients, including optimal dosing, administration routes and potential drug-nutrients interactions requiring clarification. Likewise, precision nutrition practices must account for the metabolic and immunological changes occurring in various life stages, particularly focusing on aging and associated immunosenescence, an imperative yet unmet need.

Additional gaps surface when one considers clinical trial's experimental design and the selection of immune biomarkers in studying the efficacy of immunonutrition approaches in the scope of chronic diseases. The intricate nature of nutritional interventions, their multi-target profile, as well as defining control groups, blinding, randomization, and insufficient adherence pose substantial hurdles to study design, results interpretation, and implementation. Overcoming these limitations will undoubtedly improve the level of precision in the clinical application of polyphenols-based immunonutrition and attenuate the massive burden of immunometabolic disorders currently compose. Multi-omics models and the integration of multi-dimensional datasets comprising nutritional genomics, phenotypes and lifestyles are paramount to understand the metabolic variability between individuals and achieve personalized guidance for tailored polyphenol-based immunonutritional plans.

Author contributions

CF: Writing – review & editing, Writing – original draft, Software, Investigation, Conceptualization. PV: Writing – review & editing. HS: Writing – review & editing. JM: Writing – review & editing, Funding acquisition. MC: Writing – review & editing, Supervision. FR: Writing – review & editing, Validation, Supervision, Funding acquisition, Formal analysis. SV: Writing – review & editing, Writing – original draft, Validation, Supervision, Resources, Project administration, Investigation, Funding acquisition, Formal analysis, Conceptualization.

References

1. Fernández-García V, González-Ramos S, Martín-Sanz P, Laparra JM, Boscá L. NOD1-targeted immunonutrition approaches: on the way from disease to health. *Biomedicines.* (2021) 9(5):519. doi: 10.3390/biomedicines9050519

2. Zapatera B, Prados A, Gómez-Martínez S, Marcos A. Immunonutrition: methodology and applications. *Nutricion hospitalaria*. (2015) 31 Suppl 3:145–54. doi: 10.3305/nh.2015.31.sup3.8762

3. Kaufman DR, De Calisto J, Simmons NL, Cruz AN, Villablanca EJ, Mora JR, et al. Vitamin A deficiency impairs vaccine-elicited gastrointestinal immunity. *J Immunol* (*Baltimore Md: 1950*). (2011) 187:1877–83. doi: 10.4049/jimmunol.1101248

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Conflict of interest

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4. Noor S, Piscopo S, Gasmi A. Nutrients interaction with the immune system. Arch Razi Institute. (2021) 76:1579–88. doi: 10.22092/ari.2021.356098.1775

5. Calder PC. Nutrition, immunity and COVID-19. *BMJ nutrition Prev Health*. (2020) 3:74–92. doi: 10.1136/bmjnph-2020-000085

6. García-Montero C, Fraile-Martínez O, Gómez-Lahoz AM, Pekarek L, Castellanos AJ, Noguerales-Fraguas F, et al. Nutritional components in western diet versus mediterranean diet at the gut microbiota-immune system interplay. *Implications Health Disease Nutrients*. (2021) 13:699. doi: 10.3390/nu13020699

7. Alwarawrah Y, Kiernan K, MacIver NJ. Changes in nutritional status impact immune cell metabolism and function. *Front Immunol.* (2018) 9:1055. doi: 10.3389/fimmu.2018.01055

8. Man AWC, Zhou Y, Xia N, Li H. Involvement of gut microbiota, microbial metabolites and interaction with polyphenol in host immunometabolism. *Nutrients*. (2020) 12(10):3054. doi: 10.3390/nu12103054

9. Wang Y, Dong C, Han Y, Gu Z, Sun C. Immunosenescence, aging and successful aging. *Front Immunol.* (2022) 13:942796. doi: 10.3389/fimmu.2022.942796

10. Alba G, Dakhaoui H, Santa-Maria C, Palomares F, Cejudo-Guillen M, Geniz I, et al. Nutraceuticals as potential therapeutic modulators in immunometabolism. *Nutrients.* (2023) 15:411. doi: 10.3390/nu15020411

11. Grimble RF. Basics in clinical nutrition: Immunonutrition – Nutrients which influence immunity: Effect and mechanism of action. *e-SPEN Eur e-Journal Clin Nutr Metab.* (2009) 4:e10–e3. doi: 10.1016/j.eclnm.2008.07.015

12. Wu D, Lewis ED, Pae M, Meydani SN. Nutritional modulation of immune function: analysis of evidence, mechanisms, and clinical relevance. *Front Immunol.* (2018) 9:3160. doi: 10.3389/fimmu.2018.03160

13. Shi N, Li N, Duan X, Niu H. Interaction between the gut microbiome and mucosal immune system. *Military Med Res.* (2017) 4:14. doi: 10.1186/s40779-017-0122-9

14. Zhao L, Zhang Q, Ma W, Tian F, Shen H, Zhou M. A combination of quercetin and resveratrol reduces obesity in high-fat diet-fed rats by modulation of gut microbiota. *Food Funct.* (2017) 8:4644–56. doi: 10.1039/C7FO01383C

15. Grammatikopoulou MG, Marakis G, Gkiouras K, Athanatou D, Maraki MI, Bogdanos DP. Fly me to the immune: immunonutrition in rheumatic diseases. *Mediterr J Rheumatol.* (2023) 34:30-6. doi: 10.31138/mjr.34.1.30

16. Nieman DC. Immunonutrition support for athletes. *Nutr Rev.* (2008) 66:310–20. doi: 10.1111/j.1753-4887.2008.00038.x

17. Queipo-Ortuño MI, Boto-Ordóñez M, Murri M, Gomez-Zumaquero JM, Clemente-Postigo M, Estruch R, et al. Influence of red wine polyphenols and ethanol on the gut microbiota ecology and biochemical biomarkers. *Am J Clin Nutr.* (2012) 95:1323–34. doi: 10.3945/ajcn.111.027847

18. Abbas M, Saeed F, Anjum F, Afzaal M, Tufail T, Bashir M, et al. Natural polyphenols: an overview. Int J Food Properties. (2017) 20(8):1689–99. doi: 10.1080/10942912.2016.1220393

19. Castro-Barquero S, Lamuela-Raventós RM, Doménech M, Estruch R. Relationship between mediterranean dietary polyphenol intake and obesity. *Nutrients.* (2018) 10(10):1523. doi: 10.3390/nu10101523

20. Irakli MN, Samanidou VF, Biliaderis CG, Papadoyannis IN. Development and validation of an HPLC-method for determination of free and bound phenolic acids in cereals after solid-phase extraction. *Food Chem.* (2012) 134:1624–32. doi: 10.1016/j.foodchem.2012.03.046

21. Pérez-Jiménez J, Neveu V, Vos F, Scalbert A. Identification of the 100 richest dietary sources of polyphenols: an application of the Phenol-Explorer database. *Eur J Clin Nutr.* (2010) 64 Suppl 3:S112–20. doi: 10.1038/ejcn.2010.221

22. Koroušić Seljak B. Web-based eHealth applications with reference to food composition data. *Eur J Clin Nutr.* (2010) 64 Suppl 3:S121-7. doi: 10.1038/ejcn.2010.222

23. GutiErrez-Grijalva EP, Ambriz-Pere DL, Leyva-Lopez N, Castillo-Lopez RI, Heiedia JB. Review: dietary phenolic compounds, health benefits and bioaccessibility. *Archivos latinoamericanos nutricion*. (2016) 66:87–100.

24. Panche AN, Diwan AD, Chandra SR. Flavonoids: an overview. J Nutr sci. (2016) 5:e47. doi: 10.1017/jns.2016.41

25. Tsao R. Chemistry and biochemistry of dietary polyphenols. *Nutrients*. (2010) 2:1231-46. doi: 10.3390/nu2121231

26. Bertelli A, Biagi M, Corsini M, Baini G, Cappellucci G, Miraldi E. Polyphenols: from theory to practice. *Foods (Basel Switzerland)*. (2021) 10(11):2595. doi: 10.3390/foods10112595

27. Issaoui M, Delgado AM, Caruso G, Micali M, Barbera M, Atrous H, et al. Phenols, flavors, and the mediterranean diet. J AOAC Int. (2020) 103:915-24. doi: 10.1093/jaocint/qsz018

28. Wang X, Qi Y, Zheng H. Dietary polyphenol, gut microbiota, and health benefits. *Antioxidants.* (2022) 11:1212. doi: 10.3390/antiox11061212

29. Zhang Z, Li X, Sang S, McClements DJ, Chen L, Long J, et al. Polyphenols as plant-based nutraceuticals: health effects, encapsulation, nano-delivery, and application. *Foods (Basel Switzerland)*. (2022) 11:2189. doi: 10.3390/foods11152189

30. Gan R-Y, Chan C-L, Yang Q-Q, Li H-B, Zhang D, Ge Y-Y, et al. 9 - Bioactive compounds and beneficial functions of sprouted grains. In: Feng H, Nemzer B, DeVries JW, editors. *Sprouted Grains*. St. Paul, MN, United States of America: AACC International Press (2019). p. 191–246.

31. Kumar N, Goel N. Phenolic acids: Natural versatile molecules with promising therapeutic applications. *Biotechnol Rep (Amsterdam Netherlands)*. (2019) 24:e00370. doi: 10.1016/j.btre.2019.e00370

32. de Souza JE, Casanova LM, Costa SS. Bioavailability of phenolic compounds: a major challenge for drug development? *Rev Fitos Eletronica*. (2015) 9:55–67. doi: 10.5935/2446-4775.20150006

33. D'Archivio M, Filesi C, Varì R, Scazzocchio B, Masella R. Bioavailability of the polyphenols: status and controversies. *Int J Mol Sci.* (2010) 11:1321–42. doi: 10.3390/ ijms11041321

34. Salah N, Miller NJ, Paganga G, Tijburg L, Bolwell GP, Rice-Evans C. Polyphenolic flavanols as scavengers of aqueous phase radicals and as chainbreaking antioxidants. *Arch Biochem biophysics*. (1995) 322:339–46. doi: 10.1006/ abbi.1995.1473

35. Ortega N, Reguant J, Romero MP, Macià A, Motilva MJ. Effect of fat content on the digestibility and bioaccessibility of cocoa polyphenol by an in *vitro* digestion model. *J Agric Food Chem.* (2009) 57:5743–9. doi: 10.1021/jf900591q

36. Pérez-Jiménez J, Serrano J, Tabernero M, Arranz S, Díaz-Rubio ME, García-Diz L, et al. Bioavailability of phenolic antioxidants associated with dietary fiber: plasma antioxidant capacity after acute and long-term intake in humans. *Plant foods Hum Nutr* (*Dordrecht Netherlands*). (2009) 64:102–7. doi: 10.1007/s11130-009-0110-7

37. Chen L, Cao H, Xiao J. 2 - Polyphenols: Absorption, bioavailability, and metabolomics. In: Galanakis CM, editor. *Polyphenols: Properties, Recovery, and Applications*. Sawston, Cambridge, United Kingdom: Woodhead Publishing (2018). p. 45–67.

38. Kyselova Z. Toxicological aspects of the use of phenolic compounds in disease prevention. *Interdiscip toxicol.* (2011) 4:173–83. doi: 10.2478/v10102-011-0027-5

39. Grzesik M, Naparło K, Bartosz G, Sadowska-Bartosz I. Antioxidant properties of catechins: Comparison with other antioxidants. *Food Chem.* (2018) 241:480–92. doi: 10.1016/j.foodchem.2017.08.117

40. Mi Y, Zhang W, Tian H, Li R, Huang S, Li X, et al. EGCG evokes Nrf2 nuclear translocation and dampens PTP1B expression to ameliorate metabolic misalignment under insulin resistance condition. *Food Funct.* (2018) 9:1510–23. doi: 10.1039/C7FO01554B

 Lv Q-z, Long J-t, Gong Z-f, Nong K-y, Liang X-m, Qin T, et al. Current state of knowledge on the antioxidant effects and mechanisms of action of polyphenolic compounds. *Natural Product Commun.* (2021) 16:1934578X211027745. doi: 10.1177/ 1934578X211027745

42. Rudrapal M, Khairnar SJ, Khan J, Dukhyil AB, Ansari MA, Alomary MN, et al. Dietary polyphenols and their role in oxidative stress-induced human diseases: insights into protective effects, antioxidant potentials and mechanism(s) of action. *Front Pharmacol.* (2022) 13:806470. doi: 10.3389/fphar.2022.806470

43. Scalbert A, Johnson IT, Saltmarsh M. Polyphenols: antioxidants and beyond. *Am J Clin Nutr.* (2005) 81:215s–7s. doi: 10.1093/ajcn/81.1.215S

44. Ding HW, Huang AL, Zhang YL, Li B, Huang C, Ma TT, et al. Design, synthesis and biological evaluation of hesperetin derivatives as potent anti-inflammatory agent. *Fitoterapia*. (2017) 121:212–22. doi: 10.1016/j.fitote.2017.07.016

45. Monagas M, Khan N, Andrés-Lacueva C, Urpi-Sardá M, Vázquez-Agell M, Lamuela-Raventós RM, et al. Dihydroxylated phenolic acids derived from microbial metabolism reduce lipopolysaccharide-stimulated cytokine secretion by human peripheral blood mononuclear cells. *Br J Nutr.* (2009) 102:201–6. doi: 10.1017/S0007114508162110

46. Yi S, Wang W, Bai F, Zhu J, Li J, Li X, et al. Antimicrobial effect and membraneactive mechanism of tea polyphenols against Serratia marcescens. *World J Microbiol Biotechnol.* (2014) 30:451–60. doi: 10.1007/s11274-013-1464-4

47. Ulrey RK, Barksdale SM, Zhou W, van Hoek ML. Cranberry proanthocyanidins have anti-biofilm properties against Pseudomonas aeruginosa. *BMC complementary Altern Med.* (2014) 14:499. doi: 10.1186/1472-6882-14-499

48. Zhao WH, Hu ZQ, Hara Y, Shimamura T. Inhibition of penicillinase by epigallocatechin gallate resulting in restoration of antibacterial activity of penicillin against penicillinase-producing Staphylococcus aureus. *Antimicrobial Agents chemother*. (2002) 46:2266–8. doi: 10.1128/AAC.46.7.2266-2268.2002

49. Nirengi S, Amagasa S, Homma T, Yoneshiro T, Matsumiya S, Kurosawa Y, et al. Daily ingestion of catechin-rich beverage increases brown adipose tissue density and decreases extramyocellular lipids in healthy young women. *SpringerPlus.* (2016) 5:1363. doi: 10.1186/s40064-016-3029-0

50. Tung YC, Lin YH, Chen HJ, Chou SC, Cheng AC, Kalyanam N, et al. Piceatannol exerts anti-obesity effects in C57BL/6 mice through modulating adipogenic proteins and gut microbiota. *Molecules (Basel Switzerland).* (2016) 21(11):1419. doi: 10.3390/molecules21111419

51. Andrade JM, Paraíso AF, de Oliveira MV, Martins AM, Neto JF, Guimarães AL, et al. Resveratrol attenuates hepatic steatosis in high-fat fed mice by decreasing lipogenesis and inflammation. *Nutr (Burbank Los Angeles County Calif).* (2014) 30:915–9. doi: 10.1016/j.nut.2013.11.016

52. Han X, Guo J, You Y, Yin M, Liang J, Ren C, et al. Vanillic acid activates thermogenesis in brown and white adipose tissue. *Food Funct.* (2018) 9:4366–75. doi: 10.1039/C8FO00978C

53. Sassi A, Mokdad Bzéouich I, Mustapha N, Maatouk M, Ghedira K, Chekir-Ghedira L. Immunomodulatory potential of hesperetin and chrysin through the cellular and humoral response. *Eur J Pharmacol.* (2017) 812:91–6. doi: 10.1016/j.ejphar.2017.07.017

54. Gerdemann A, Broenhorst M, Behrens M, Humpf HU, Esselen M. Polyphenols cause structure dependent effects on the metabolic profile of human hepatocarcinogenic cells. *Mol Nutr Food Res.* (2023) 67:e2300052. doi: 10.1002/mnfr.202300052

55. Saravia J, Raynor JL, Chapman NM, Lim SA, Chi H. Signaling networks in immunometabolism. *Cell Res.* (2020) 30:328–42. doi: 10.1038/s41422-020-0301-1

56. Luby A, Alves-Guerra M-C. UCP2 as a cancer target through energy metabolism and oxidative stress control. *Int J Mol Sci.* (2022) 23:15077. doi: 10.3390/ijms232315077

57. Tan J, Ni D, Ribeiro RV, Pinget GV, Macia L. How changes in the nutritional landscape shape gut immunometabolism. *Nutrients.* (2021) 13(3):823. doi: 10.3390/nu13030823

58. Blagih J, Coulombe F, Vincent EE, Dupuy F, Galicia-Vázquez G, Yurchenko E, et al. The energy sensor AMPK regulates T cell metabolic adaptation and effector responses in vivo. *Immunity*. (2015) 42:41–54. doi: 10.1016/j.immuni.2014.12.030

59. Hoong CWS, Chua MWJ. SGLT2 inhibitors as calorie restriction mimetics: insights on longevity pathways and age-related diseases. *Endocrinology*. (2021) 162(8): bqab079. doi: 10.1210/endocr/bqab079

60. O'Neill LA, Kishton RJ, Rathmell J. A guide to immunometabolism for immunologists. *Nat Rev Immunol.* (2016) 16:553-65. doi: 10.1038/nri.2016.70

61. Adamik J, Munson PV, Hartmann FJ, Combes AJ, Pierre P, Krummel MF, et al. Distinct metabolic states guide maturation of inflammatory and tolerogenic dendritic cells. *Nat Commun.* (2022) 13:5184. doi: 10.1038/s41467-022-32849-1

62. García-Prat L, Martínez-Vicente M, Perdiguero E, Ortet L, Rodríguez-Ubreva J, Rebollo E, et al. Autophagy maintains stemness by preventing senescence. *Nature*. (2016) 529:37–42. doi: 10.1038/nature16187

 Liu PS, Wang H, Li X, Chao T, Teav T, Christen S, et al. α-ketoglutarate orchestrates macrophage activation through metabolic and epigenetic reprogramming. *Nat Immunol.* (2017) 18:985–94. doi: 10.1038/ni.3796

64. Angajala A, Lim S, Phillips JB, Kim JH, Yates C, You Z, et al. Diverse roles of mitochondria in immune responses: novel insights into immuno-metabolism. *Front Immunol.* (2018) 9:1605. doi: 10.3389/fimmu.2018.01605

65. Jain A, Zoncu R. Organelle transporters and inter-organelle communication as drivers of metabolic regulation and cellular homeostasis. *Mol Metab.* (2022) 60:101481. doi: 10.1016/j.molmet.2022.101481

66. Bantug GR, Fischer M, Grählert J, Balmer ML, Unterstab G, Develioglu L, et al. Mitochondria-endoplasmic reticulum contact sites function as immunometabolic hubs that orchestrate the rapid recall response of memory CD8(+) T cells. *Immunity*. (2018) 48:542–55.e6. doi: 10.1016/j.immuni.2018.02.012

67. Li W, Zhang L. Rewiring mitochondrial metabolism for CD8(+) T cell memory formation and effective cancer immunotherapy. *Front Immunol.* (2020) 11:1834. doi: 10.3389/fimmu.2020.01834

68. Cardoso M, Gaspar VM, Ferreira C, Silvestre R, Duarte IF, Mano JF. Macrophage-targeted shikonin-loaded nanogels for modulation of inflammasome activation. *Nanomedicine: nanotechnology biology Med.* (2022) 42:102548. doi: 10.1016/j.nano.2022.102548

 Finucane OM, Sugrue J, Rubio-Araiz A, Guillot-Sestier MV, Lynch MA. The NLRP3 inflammasome modulates glycolysis by increasing PFKFB3 in an IL-1β-dependent manner in macrophages. Sci Rep. (2019) 9:4034. doi: 10.1038/s41598-019-40619-1

70. Shakoor H, Feehan J, Apostolopoulos V, Platat C, Al Dhaheri AS, Ali HI, et al. Immunomodulatory effects of dietary polyphenols. *Nutrients*. (2021) 13:728. doi: 10.3390/nu13030728

71. Wang S, Li Z, Ma Y, Liu Y, Lin C-C, Li S, et al. Immunomodulatory effects of green tea polyphenols. *Molecules (Basel Switzerland)*. (2021) 26:3755. doi: 10.3390/molecules26123755

72. Ding S, Jiang H, Fang J. Regulation of immune function by polyphenols. J Immunol Res. (2018) 2018:1264074. doi: 10.1155/2018/1264074

73. González R, Ballester I, López-Posadas R, Suárez MD, Zarzuelo A, Martínez-Augustin O, et al. Effects of flavonoids and other polyphenols on inflammation. *Crit Rev Food Sci Nutr.* (2011) 51:331–62. doi: 10.1080/10408390903584094

74. Hassanain E, Silverberg JI, Norowitz KB, Chice S, Bluth MH, Brody N, et al. Green tea (Camelia sinensis) suppresses B cell production of IgE without inducing apoptosis. *Ann Clin Lab sci.* (2010) 40:135–43. doi: 10.1016/j.jaci.2009.12.079

75. Sanbongi C, Suzuki N, Sakane T. Polyphenols in chocolate, which have antioxidant activity, modulate immune functions in humans in vitro. *Cell Immunol.* (1997) 177:129–36. doi: 10.1006/cimm.1997.1109

76. Murray PJ, Wynn TA. Protective and pathogenic functions of macrophage subsets. *Nat Rev Immunol.* (2011) 11:723–37. doi: 10.1038/nri3073

77. Essafi-Benkhadir K, Refai A, Riahi I, Fattouch S, Karoui H, Essafi M. Quince (Cydonia oblonga Miller) peel polyphenols modulate LPS-induced inflammation in human THP-1-derived macrophages through NF- κ B, p38MAPK and Akt inhibition. *Biochem Biophys Res Commun.* (2012) 418:180–5. doi: 10.1016/j.bbrc.2012.01.003

78. Soromou LW, Zhang Z, Li R, Chen N, Guo W, Huo M, et al. Regulation of inflammatory cytokines in lipopolysaccharide-stimulated RAW 264.7 murine macrophage by 7-O-methyl-naringenin. *Molecules (Basel Switzerland)*. (2012) 17:3574–85. doi: 10.3390/molecules17033574

79. Zunino SJ, Storms DH. Resveratrol alters proliferative responses and apoptosis in human activated B lymphocytes in vitro. *J Nutr.* (2009) 139:1603–8. doi: 10.3945/jn.109.105064

80. Oh H-J, Kang Y-G, Na T-Y, Kim H-J, Park JS, Cho W-J, et al. Identification of daidzein as a ligand of retinoic acid receptor that suppresses expression of matrix metalloproteinase-9 in HaCaT cells. *Mol Cell Endocrinology*. (2013) 376:107-13. doi: 10.1016/j.mce.2013.06.015

81. Zeng W, Zhang C, Cheng H, Wu Y-L, Liu J, Chen Z, et al. Targeting to the nongenomic activity of retinoic acid receptor-gamma by acacetin in hepatocellular carcinoma. *Sci Rep.* (2017) 7:348. doi: 10.1038/s41598-017-00233-5

82. Sanchez-Garrido J, Shenoy AR. Regulation and repurposing of nutrient sensing and autophagy in innate immunity. *Autophagy.* (2021) 17:1571-91. doi: 10.1080/15548627.2020.1783119

83. Yang L, Chu Z, Liu M, Zou Q, Li J, Liu Q, et al. Amino acid metabolism in immune cells: essential regulators of the effector functions, and promising opportunities to enhance cancer immunotherapy. *J Hematol Oncol.* (2023) 16:59. doi: 10.1186/s13045-023-01453-1

84. Nakaya M, Xiao Y, Zhou X, Chang JH, Chang M, Cheng X, et al. Inflammatory T cell responses rely on amino acid transporter ASCT2 facilitation of glutamine uptake and mTORC1 kinase activation. *Immunity*. (2014) 40:692–705. doi: 10.1016/j.immuni.2014.04.007

85. Walls J, Sinclair L, Finlay D. Nutrient sensing, signal transduction and immune responses. *Semin Immunol.* (2016) 28:396–407. doi: 10.1016/j.smim.2016.09.001

86. Limagne E, Thibaudin M, Euvrard R, Berger H, Chalons P, Végan F, et al. Sirtuin-1 activation controls tumor growth by impeding th17 differentiation via STAT3 deacetylation. *Cell Rep.* (2017) 19:746–59. doi: 10.1016/j.celrep.2017.04.004

87. Delmas D, Limagne E, Ghiringhelli F, Aires V. Immune Th17 lymphocytes play a critical role in the multiple beneficial properties of resveratrol. *Food Chem toxicology: an Int J published Br Ind Biol Res Assoc.* (2020) 137:111091. doi: 10.1016/j.fct.2019.111091

88. Sadria M, Layton AT. Interactions among mTORC, AMPK and SIRT: a computational model for cell energy balance and metabolism. *Cell communication signaling: CCS.* (2021) 19:57. doi: 10.1186/s12964-021-00706-1

89. Packer M. Critical reanalysis of the mechanisms underlying the cardiorenal benefits of SGLT2 inhibitors and reaffirmation of the nutrient deprivation signaling/ autophagy hypothesis. *Circulation*. (2022) 146:1383-405. doi: 10.1161/CIRCULATIONAHA.122.061732

90. Power H, Valtchev P, Dehghani F, Schindeler A. Strategies for senolytic drug discovery. *Aging Cell.* (2023) 22:e13948. doi: 10.1111/acel.13948

91. Ji Z, Liu GH, Qu J. Mitochondrial sirtuins, metabolism, and aging. J Genet Genomics = Yi Chuan xue bao. (2022) 49:287-98. doi: 10.1016/j.jgg.2021.11.005

92. Di Giosia P, Stamerra CA, Giorgini P, Jamialahamdi T, Butler AE, Sahebkar A. The role of nutrition in inflammaging. *Ageing Res Rev.* (2022) 77:101596. doi: 10.1016/j.arr.2022.101596

93. Dong D, Cai GY, Ning YC, Wang JC, Lv Y, Hong Q, et al. Alleviation of senescence and epithelial-mesenchymal transition in aging kidney by short-term caloric restriction and caloric restriction mimetics via modulation of AMPK/mTOR signaling. *Oncotarget.* (2017) 8:16109–21. doi: 10.18632/oncotarget.v8i10

94. Davinelli S, De Stefani D, De Vivo I, Scapagnini G. Polyphenols as caloric restriction mimetics regulating mitochondrial biogenesis and mitophagy. *Trends Endocrinol metabolism: TEM.* (2020) 31:536–50. doi: 10.1016/j.tem.2020.02.011

95. Sun Y, Qin H, Zhang H, Feng X, Yang L, Hou DX, et al. Fisetin inhibits inflammation and induces autophagy by mediating PI3K/AKT/mTOR signaling in LPS-induced RAW264.7 cells. *Food Nutr Res.* (2021) 65:6355. doi: 10.29219/ fnr.v65.6355

96. Zhang Y, Yang W, Li W, Zhao Y. NLRP3 inflammasome: checkpoint connecting innate and adaptive immunity in autoimmune diseases. *Front Immunol.* (2021) 12:732933. doi: 10.3389/fimmu.2021.732933

97. Han X, Xu T, Fang Q, Zhang H, Yue L, Hu G, et al. Quercetin hinders microglial activation to alleviate neurotoxicity via the interplay between NLRP3 inflammasome and mitophagy. *Redox Biol.* (2021) 44:102010. doi: 10.1016/j.redox.2021.102010

98. Misawa T, Saitoh T, Kozaki T, Park S, Takahama M, Akira S. Resveratrol inhibits the acetylated α -tubulin-mediated assembly of the NLRP3-inflammasome. *Int Immunol.* (2015) 27:425–34. doi: 10.1093/intimm/dxv018

99. Midttun H, Ramsay A, Müller-Harvey I, Williams A. Cocoa procyanidins modulate transcriptional pathways linked to inflammation and metabolism in human dendritic cells. *Food Funct*. (2018) 9:2883–90. doi: 10.1039/C8FO00387D

100. da Rosa MM, de Amorim LC, de Oliveira Alves JV, de Silva Aguiar IF, da Silva Oliveira F, da Silva MV, et al. The promising role of natural products in Alzheimer's disease. *Brain Disord*. (2022) 7:100049. doi: 10.1016/j.dscb.2022.100049

101. Britt EC, John SV, Locasale JW, Fan J. Metabolic regulation of epigenetic remodeling in immune cells. *Curr Opin Biotechnol.* (2020) 63:111–7. doi: 10.1016/j.copbio.2019.12.008

102. Liu TF, Vachharajani VT, Yoza BK, McCall CE. NAD+-dependent sirtuin 1 and 6 proteins coordinate a switch from glucose to fatty acid oxidation during the acute inflammatory response. *J Biol Chem.* (2012) 287:25758–69. doi: 10.1074/jbc.M112.362343

103. Peng J, Yang Z, Li H, Hao B, Cui D, Shang R, et al. Quercetin reprograms immunometabolism of macrophages via the SIRT1/PGC-1 α Signaling pathway to ameliorate lipopolysaccharide-induced oxidative damage. *Int J Mol Sci.* (2023) 24 (6):5542. doi: 10.3390/ijms24065542

104. Rajendran P, Ho E, Williams DE, Dashwood RH. Dietary phytochemicals, HDAC inhibition, and DNA damage/repair defects in cancer cells. *Clin epigenetics*. (2011) 3:4. doi: 10.1186/1868-7083-3-4

105. Kiss AK, Granica S, Stolarczyk M, Melzig MF. Epigenetic modulation of mechanisms involved in inflammation: Influence of selected polyphenolic substances on histone acetylation state. *Food Chem.* (2012) 131:1015–20. doi: 10.1016/j.foodchem.2011.09.109

106. Kim HJ, Kim SH, Yun JM. Fisetin inhibits hyperglycemia-induced proinflammatory cytokine production by epigenetic mechanisms. *Evidence-Based complementary Altern medicine: eCAM.* (2012) 2012:639469. doi: 10.1155/2012/639469

107. Yun JM, Jialal I, Devaraj S. Effects of epigallocatechin gallate on regulatory T cell number and function in obese v. lean volunteers. *Br J Nutr.* (2010) 103:1771–7. doi: 10.1017/S000711451000005X

108. Blaze J, Wang J, Ho L, Mendelev N, Haghighi F, Pasinetti GM. Polyphenolic compounds alter stress-induced patterns of global DNA methylation in brain and blood. *Mol Nutr Food Res.* (2018) 62:e1700722. doi: 10.1002/mnfr.201700722

109. Švajger U, Obermajer N, Jeras M. Dendritic cells treated with resveratrol during differentiation from monocytes gain substantial tolerogenic properties upon activation. *Immunology.* (2010) 129:525–35. doi: 10.1111/j.1365-2567.2009.03205.x

110. Gupta SC, Tyagi AK, Deshmukh-Taskar P, Hinojosa M, Prasad S, Aggarwal BB. Downregulation of tumor necrosis factor and other proinflammatory biomarkers by polyphenols. *Arch Biochem biophysics*. (2014) 559:91–9. doi: 10.1016/j.abb.2014.06.006

111. Huang RY, Yu YL, Cheng WC, OuYang CN, Fu E, Chu CL. Immunosuppressive effect of quercetin on dendritic cell activation and function. J Immunol (Baltimore Md: 1950). (2010) 184:6815–21. doi: 10.4049/jimmunol.0903991

112. Lin W, Wang W, Wang D, Ling W. Quercetin protects against atherosclerosis by inhibiting dendritic cell activation. *Mol Nutr Food Res.* (2017) 61(9):1700031. doi: 10.1002/mnfr.v61.9

113. Yoneyama S, Kawai K, Tsuno NH, Okaji Y, Asakage M, Tsuchiya T, et al. Epigallocatechin gallate affects human dendritic cell differentiation and maturation. J Allergy Clin Immunol. (2008) 121:209–14. doi: 10.1016/j.jaci.2007.08.026

114. Kim GY, Kim KH, Lee SH, Yoon MS, Lee HJ, Moon DO, et al. Curcumin inhibits immunostimulatory function of dendritic cells: MAPKs and translocation of NF-kappa B as potential targets. *J Immunol (Baltimore Md: 1950)*. (2005) 174:8116–24. doi: 10.4049/jimmunol.174.12.8116

115. Yoon MS, Lee JS, Choi BM, Jeong YI, Lee CM, Park JH, et al. Apigenin inhibits immunostimulatory function of dendritic cells: Implication of immunotherapeutic adjuvant. *Mol Pharmacol.* (2006) 70:1033–44. doi: 10.1124/mol.106.024547

116. Yum MK, Jung MY, Cho D, Kim TS. Suppression of dendritic cells' maturation and functions by daidzein, a phytoestrogen. *Toxicol Appl Pharmacol.* (2011) 257:174– 81. doi: 10.1016/j.taap.2011.09.002

117. Kim ME, Kim HK, Park HY, Kim DH, Chung HY, Lee JS. Baicalin from Scutellaria baicalensis impairs Th1 polarization through inhibition of dendritic cell maturation. J Pharmacol Sci. (2013) 121:148–56. doi: 10.1254/jphs.12200FP

118. Liu SH, Lin CH, Hung SK, Chou JH, Chi CW, Fu SL. Fisetin inhibits lipopolysaccharide-induced macrophage activation and dendritic cell maturation. J Agric Food Chem. (2010) 58:10831–9. doi: 10.1021/jf1017093

119. Lee JS, Kim SG, Kim HK, Lee TH, Jeong YI, Lee CM, et al. Silibinin polarizes Th1/Th2 immune responses through the inhibition of immunostimulatory function of dendritic cells. *J Cell Physiol*. (2007) 210:385–97. doi: 10.1002/jcp.20852

120. Ahn SC, Kim GY, Kim JH, Baik SW, Han MK, Lee HJ, et al. Epigallocatechin-3gallate, constituent of green tea, suppresses the LPS-induced phenotypic and functional maturation of murine dendritic cells through inhibition of mitogen-activated protein kinases and NF-kappaB. *Biochem Biophys Res Commun.* (2004) 313:148–55. doi: 10.1016/j.bbrc.2003.11.108

121. Campbell NK, Fitzgerald HK, Fletcher JM, Dunne A. Plant-derived polyphenols modulate human dendritic cell metabolism and immune function via AMPK-dependent induction of heme oxygenase-1. *Front Immunol.* (2019) 10:345. doi: 10.3389/fimmu.2019.00345

122. Dugo L, Belluomo MG, Fanali C, Russo M, Cacciola F, Maccarrone M, et al. Effect of cocoa polyphenolic extract on macrophage polarization from proinflammatory M1 to anti-inflammatory M2 state. *Oxid Med Cell longevity*. (2017) 2017:6293740. doi: 10.1155/2017/6293740

123. Kimura Y, Sumiyoshi M. Resveratrol prevents tumor growth and metastasis by inhibiting lymphangiogenesis and M2 macrophage activation and differentiation in tumor-associated macrophages. *Nutr cancer*. (2016) 68:667–78. doi: 10.1080/01635581.2016.1158295

124. Hämäläinen M, Nieminen R, Vuorela P, Heinonen M, Moilanen E. Antiinflammatory effects of flavonoids: genistein, kaempferol, quercetin, and daidzein inhibit STAT-1 and NF-kappaB activations, whereas flavone, isorhamnetin, naringenin, and pelargonidin inhibit only NF-kappaB activation along with their inhibitory effect on iNOS expression and NO production in activated macrophages. *Med Inflamm.* (2007) 2007:45673. doi: 10.1155/2007/45673

125. Zhang X, Wang G, Gurley EC, Zhou H. Flavonoid apigenin inhibits lipopolysaccharide-induced inflammatory response through multiple mechanisms in macrophages. *PloS One.* (2014) 9:e107072. doi: 10.1371/journal.pone.0107072

126. Houde V, Grenier D, Chandad F. Protective effects of grape seed proanthocyanidins against oxidative stress induced by lipopolysaccharides of periodontopathogens. *J periodontology*. (2006) 77:1371–9. doi: 10.1902/jop.2006. 050419

127. Li Y, Deng W, Wu L, Chen S, Zheng Z, Song H. Anti-inflammatory effects of polyphenols from plum (Prunus salicina lindl) on RAW264.7 macrophages induced by monosodium urate and potential mechanisms. *Foods (Basel Switzerland)*. (2023) 12 (2):254. doi: 10.3390/foods12020254

128. Zanotti Simoes Dourado GK, de Abreu Ribeiro LC, Zeppone Carlos I, Borges César T. Orange juice and hesperidin promote differential innate immune response in macrophages ex vivo. Int J vitamin Nutr Res Internationale Z fur Vitamin- und Ernahrungsforschung J Int vitaminologie Nutr. (2013) 83:162–7. doi: 10.1024/0300-9831/a000157

129. Mendes LF, Gaspar VM, Conde TA, Mano JF, Duarte IF. Flavonoid-mediated immunomodulation of human macrophages involves key metabolites and metabolic pathways. *Sci Rep.* (2019) 9:14906. doi: 10.1038/s41598-019-51113-z

130. Brouet I, Ohshima H. Curcumin, an anti-tumour promoter and antiinflammatory agent, inhibits induction of nitric oxide synthase in activated macrophages. *Biochem Biophys Res Commun.* (1995) 206:533–40. doi: 10.1006/ bbrc.1995.1076

131. Joe B, Lokesh BR. Dietary n-3 fatty acids, curcumin and capsaicin lower the release of lysosomal enzymes and eicosanoids in rat peritoneal macrophages. *Mol Cell Biochem.* (2000) 203:153–61. doi: 10.1023/a:1007005605869

132. Tsai SH, Lin-Shiau SY, Lin JK. Suppression of nitric oxide synthase and the down-regulation of the activation of NFkappaB in macrophages by resveratrol. *Br J Pharmacol.* (1999) 126:673–80. doi: 10.1038/sj.bjp.0702357

133. Chan MM, Mattiacci JA, Hwang HS, Shah A, Fong D. Synergy between ethanol and grape polyphenols, quercetin, and resveratrol, in the inhibition of the inducible nitric oxide synthase pathway. *Biochem Pharmacol.* (2000) 60:1539–48. doi: 10.1016/S0006-2952(00)00471-8

134. Lin YL, Lin JK. (-)-Epigallocatechin-3-gallate blocks the induction of nitric oxide synthase by down-regulating lipopolysaccharide-induced activity of transcription factor nuclear factor-kappaB. *Mol Pharmacol.* (1997) 52:465–72. doi: 10.1124/mol.52.3.465

135. Lee SJ, Lee IS, Mar W. Inhibition of inducible nitric oxide synthase and cyclooxygenase-2 activity by 1,2,3,4,6-penta-O-galloyl-beta-D-glucose in murine macrophage cells. Arch pharmacal Res. (2003) 26:832–9. doi: 10.1007/BF02980029

136. Marczin N, Papapetropoulos A, Catravas JD. Tyrosine kinase inhibitors suppress endotoxin- and IL-1 beta-induced NO synthesis in aortic smooth muscle cells. *Am J Physiology-Heart Circulatory Physiol.* (1993) 265:H1014–H8. doi: 10.1152/ ajpheart.1993.265.3.H1014

137. Bisht K, Choi WH, Park SY, Chung MK, Koh WS. Curcumin enhances noninflammatory phagocytic activity of RAW264.7 cells. *Biochem Biophys Res Commun.* (2009) 379:632–6. doi: 10.1016/j.bbrc.2008.12.135

138. Huang AC, Cheng HY, Lin TS, Chen WH, Lin JH, Lin JJ, et al. Epigallocatechin gallate (EGCG), influences a murine WEHI-3 leukemia model in *vivo* through enhancing phagocytosis of macrophages and populations of T- and B-cells. *In Vivo* (*Athens Greece*). (2013) 27:627–34.

139. Mukherjee S, Hussaini R, White R, Atwi D, Fried A, Sampat S, et al. TriCurin, a synergistic formulation of curcumin, resveratrol, and epicatechin gallate, repolarizes tumor-associated macrophages and triggers an immune response to cause suppression of HPV+ tumors. *Cancer immunology immunotherapy: CII.* (2018) 67:761–74. doi: 10.1007/s00262-018-2130-3

140. Stull AJ, Cash KC, Champagne CM, Gupta AK, Boston R, Beyl RA, et al. Blueberries improve endothelial function, but not blood pressure, in adults with metabolic syndrome: a randomized, double-blind, placebo-controlled clinical trial. *Nutrients.* (2015) 7:4107–23. doi: 10.3390/nu7064107

141. Han SG, Han SS, Toborek M, Hennig B. EGCG protects endothelial cells against PCB 126-induced inflammation through inhibition of AhR and induction of Nrf2-regulated genes. *Toxicol Appl Pharmacol.* (2012) 261:181–8. doi: 10.1016/j.taap.2012.03.024

142. Saroni Arwa P, Zeraik ML, Ximenes VF, da Fonseca LM, Bolzani Vda S, Siqueira Silva DH. Redox-active biflavonoids from Garcinia brasiliensis as inhibitors of neutrophil oxidative burst and human erythrocyte membrane damage. *J ethnopharmacology*. (2015) 174:410–8. doi: 10.1016/j.jep.2015.08.041

143. Hwang TL, Leu YL, Kao SH, Tang MC, Chang HL. Viscolin, a new chalcone from Viscum coloratum, inhibits human neutrophil superoxide anion and elastase release via a cAMP-dependent pathway. *Free Radical Biol Med.* (2006) 41:1433–41. doi: 10.1016/j.freeradbiomed.2006.08.001

144. Drábiková K, Perečko T, Nosáľ R, Harmatha J, Smidrkal J, Jančinová V. Polyphenol derivatives - potential regulators of neutrophil activity. *Interdiscip toxicol.* (2012) 5:65–70. doi: 10.2478/v10102-012-0011-8

145. Petersen KS, Marnewick JL, Smith C. Grape polyphenols corrects ageingrelated detriments in neutrophil functionality via modulation of specific molecular targets. *Inflammopharmacology*. (2018) 26:1349–58. doi: 10.1007/s10787-018-0511-z

146. Han D, Wu Y, Lu D, Pang J, Hu J, Zhang X, et al. Polyphenol-rich diet mediates interplay between macrophage-neutrophil and gut microbiota to alleviate intestinal inflammation. *Cell Death disease*. (2023) 14:656. doi: 10.1038/s41419-023-06190-4

147. Kucuksezer UC, Aktas Cetin E, Esen F, Tahrali I, Akdeniz N, Gelmez MY, et al. The role of natural killer cells in autoimmune diseases. *Front Immunol.* (2021) 12:622306. doi: 10.3389/fimmu.2021.622306 148. Kim YH, Won YS, Yang X, Kumazoe M, Yamashita S, Hara A, et al. Green tea catechin metabolites exert immunoregulatory effects on CD4(+) T cell and natural killer cell activities. *J Agric Food Chem.* (2016) 64:3591–7. doi: 10.1021/acs.jafc.6b01115

149. Mariee AD, Abd-Allah GM, El-Beshbishy HA. Protective effect of dietary flavonoid quercetin against lipemic-oxidative hepatic injury in hypercholesterolemic rats. *Pharm Biol.* (2012) 50:1019–25. doi: 10.3109/13880209.2012.655424

150. Falchetti R, Fuggetta MP, Lanzilli G, Tricarico M, Ravagnan G. Effects of resveratrol on human immune cell function. *Life Sci.* (2001) 70:81–96. doi: 10.1016/S0024-3205(01)01367-4

151. Lu CC, Lai HC, Hsieh SC, Chen JK. Resveratrol ameliorates Serratia marcescens-induced acute pneumonia in rats. *J leukocyte Biol.* (2008) 83:1028–37. doi: 10.1189/jlb.0907647

152. Lu CC, Chen JK. Resveratrol enhances perforin expression and NK cell cytotoxicity through NKG2D-dependent pathways. *J Cell Physiol.* (2010) 223:343–51. doi: 10.1002/jcp.22043

153. Li C, Ge B, Nicotra M, Stern JN, Kopcow HD, Chen X, et al. JNK MAP kinase activation is required for MTOC and granule polarization in NKG2D-mediated NK cell cytotoxicity. *Proc Natl Acad Sci United States America*. (2008) 105:3017–22. doi: 10.1073/pnas.0712310105

154. Oo AM, Mohd Adnan LH, Nor NM, Simbak N, Ahmad NZ, Lwin OM. Immunomodulatory effects of flavonoids: An experimental study on natural-killer-cellmediated cytotoxicity against lung cancer and cytotoxic granule secretion profile. *Proc Singapore Healthcare*. (2021) 30:279–85. doi: 10.1177/2010105820979006

155. Jiang X, Patterson NM, Ling Y, Xie J, Helferich WG, Shapiro DJ. Low concentrations of the soy phytoestrogen genistein induce proteinase inhibitor 9 and block killing of breast cancer cells by immune cells. *Endocrinology*. (2008) 149:5366–73. doi: 10.1210/en.2008-0857

156. Guo TL, Chi RP, Hernandez DM, Auttachoat W, Zheng JF. Decreased 7,12dimethylbenz[a]anthracene-induced carcinogenesis coincides with the induction of antitumor immunities in adult female B6C3F1 mice pretreated with genistein. *Carcinogenesis*. (2007) 28:2560–6. doi: 10.1093/carcin/bgm223

157. McAnulty LS, Nieman DC, Dumke CL, Shooter LA, Henson DA, Utter AC, et al. Effect of blueberry ingestion on natural killer cell counts, oxidative stress, and inflammation prior to and after 2.5 h of running. *Appl physiology nutrition Metab* = *Physiologie appliquee Nutr metabolisme*. (2011) 36:976–84. doi: 10.1139/h11-120

158. McAnulty LS, Collier SR, Landram MJ, Whittaker DS, Isaacs SE, Klemka JM, et al. Six weeks daily ingestion of whole blueberry powder increases natural killer cell counts and reduces arterial stiffness in sedentary males and females. *Nutr Res (New York NY)*. (2014) 34:577–84. doi: 10.1016/j.nutres.2014.07.002

159. Wong CP, Nguyen LP, Noh SK, Bray TM, Bruno RS, Ho E. Induction of regulatory T cells by green tea polyphenol EGCG. *Immunol Lett.* (2011) 139:7–13. doi: 10.1016/j.imlet.2011.04.009

160. Yang J, Yang X, Chu Y, Li M. Identification of Baicalin as an immunoregulatory compound by controlling T(H)17 cell differentiation. *PloS One.* (2011) 6:e17164. doi: 10.1371/journal.pone.0017164

161. Ning W, Wang S, Dong X, Liu D, Fu L, Jin R, et al. Epigallocatechin-3-gallate (EGCG) suppresses the trafficking of lymphocytes to epidermal melanocytes via inhibition of JAK2: its implication for vitiligo treatment. *Biol Pharm bulletin.* (2015) 38:1700–6. doi: 10.1248/bpb.b15-00331

162. Kang TH, Lee JH, Song CK, Han HD, Shin BC, Pai SI, et al. Epigallocatechin-3gallate enhances CD8+ T cell-mediated antitumor immunity induced by DNA vaccination. *Cancer Res.* (2007) 67:802–11. doi: 10.1158/0008-5472.CAN-06-2638

163. Mantena SK, Roy AM, Katiyar SK. Epigallocatechin-3-gallate inhibits photocarcinogenesis through inhibition of angiogenic factors and activation of CD8+ T cells in tumors. *Photochem photobiology*. (2005) 81:1174–9. doi: 10.1562/2005-04-11-RA-487

164. Ramiro-Puig E, Pérez-Cano FJ, Ramírez-Santana C, Castellote C, Izquierdo-Pulido M, Permanyer J, et al. Spleen lymphocyte function modulated by a cocoaenriched diet. *Clin Exp Immunol.* (2007) 149:535–42. doi: 10.1111/j.1365-2249.2007.03430.x

165. Niu X, Huang Z, Zhang L, Ren X, Wang J. Auraptene has the inhibitory property on murine T lymphocyte activation. *Eur J Pharmacol.* (2015) 750:8–13. doi: 10.1016/j.ejphar.2015.01.017

166. Guo TL, White KL Jr., Brown RD, Delclos KB, Newbold RR, Weis C, et al. Genistein modulates splenic natural killer cell activity, antibody-forming cell response, and phenotypic marker expression in F(0) and F(1) generations of Sprague-Dawley rats. *Toxicol Appl Pharmacol.* (2002) 181:219–27. doi: 10.1006/taap.2002.9418

167. Churchill M, Chadburn A, Bilinski RT, Bertagnolli MM. Inhibition of intestinal tumors by curcumin is associated with changes in the intestinal immune cell profile. *J Surg Res.* (2000) 89:169–75. doi: 10.1006/jsre.2000.5826

168. Fahey AJ, Adrian Robins R, Constantinescu CS. Curcumin modulation of IFNbeta and IL-12 signalling and cytokine induction in human T cells. *J Cell Mol Med.* (2007) 11:1129–37. doi: 10.1111/j.1582-4934.2007.00089.x

169. Sharma S, Chopra K, Kulkarni SK, Agrewala JN. Resveratrol and curcumin suppress immune response through CD28/CTLA-4 and CD80 co-stimulatory pathway. *Clin Exp Immunol.* (2007) 147:155–63. doi: 10.1111/j.1365-2249.2006.03257.x

170. Ranjan D, Siquijor A, Johnston TD, Wu G, Nagabhuskahn M. The effect of curcumin on human B-cell immortalization by Epstein-Barr virus. *Am surgeon.* (1998) 64:47–51; discussion -2.

171. Billard C, Izard JC, Roman V, Kern C, Mathiot C, Mentz F, et al. Comparative antiproliferative and apoptotic effects of resveratrol, epsilon-viniferin and vine-shots derived polyphenols (vineatrols) on chronic B lymphocytic leukemia cells and normal human lymphocytes. *Leukemia lymphoma*. (2002) 43:1991–2002. doi: 10.1080/1042819021000015952

172. Makowski L, Chaib M, Rathmell JC. Immunometabolism: From basic mechanisms to translation. *Immunol Rev.* (2020) 295:5-14. doi: 10.1111/imr.12858

173. Procaccini C, de Candia P, Russo C, De Rosa G, Lepore MT, Colamatteo A, et al. Caloric restriction for the immunometabolic control of human health. *Cardiovasc Res.* (2023) 119(18):2787–800. doi: 10.1093/cvr/cvad035

174. Mathis D, Shoelson SE. Immunometabolism: an emerging frontier. Nat Rev Immunol. (2011) 11:81. doi: 10.1038/nri2922

175. Childs CE, Calder PC, Miles EA. Diet and immune function. *Nutrients*. (2019) 11:1933. doi: 10.3390/nu11081933

176. Anaya J-M, Shoenfeld Y, Rojas-Villarraga A, Levy R, Cervera R. Autoimmunity. from bench to bedside. Bogota, Colombia: El Rosario University Press (2013).

177. Vergani D, Johnston C, B-Abdullah N, Barnett AH. Low serum C4 concentrations: an inherited predisposition to insulin dependent diabetes? *Br Med J (Clinical Res ed).* (1983) 286:926–8. doi: 10.1136/bmj.286.6369.926

178. Sun L, Xi S, He G, Li Z, Gang X, Sun C, et al. Two to tango: dialogue between adaptive and innate immunity in type 1 diabetes. *J Diabetes Res.* (2020) 2020:4106518. doi: 10.1155/2020/4106518

179. Parsa R, Andresen P, Gillett A, Mia S, Zhang XM, Mayans S, et al. Adoptive transfer of immunomodulatory M2 macrophages prevents type 1 diabetes in NOD mice. *Diabetes*. (2012) 61:2881–92. doi: 10.2337/db11-1635

180. Poirot L, Benoist C, Mathis D. Natural killer cells distinguish innocuous and destructive forms of pancreatic islet autoimmunity. *Proc Natl Acad Sci United States America*. (2004) 101:8102–7. doi: 10.1073/pnas.0402065101

181. Gonzalez CD, Lee MS, Marchetti P, Pietropaolo M, Towns R, Vaccaro MI, et al. The emerging role of autophagy in the pathophysiology of diabetes mellitus. *Autophagy*. (2011) 7:2–11. doi: 10.4161/auto.7.1.13044

182. Sousa ESA, Queiroz LAD, Guimarães JPT, Pantoja KC, Barros RS, Epiphanio S, et al. The influence of high glucose conditions on macrophages and its effect on the autophagy pathway. *Front Immunol.* (2023) 14:1130662. doi: 10.3389/fimmu.2023.1130662

183. Xiu F, Stanojcic M, Diao L, Jeschke MG. Stress hyperglycemia, insulin treatment, and innate immune cells. *Int J endocrinology*. (2014) 2014:486403. doi: 10.1155/2014/486403

184. van Niekerk G, Christowitz C, Conradie D, Engelbrecht AM. Insulin as an immunomodulatory hormone. *Cytokine Growth factor Rev.* (2020) 52:34–44. doi: 10.1016/j.cytogfr.2019.11.006

185. Yu T, Gao M, Yang P, Liu D, Wang D, Song F, et al. Insulin promotes macrophage phenotype transition through PI3K/Akt and PPAR-γ signaling during diabetic wound healing. J Cell Physiol. (2019) 234:4217–31. doi: 10.1002/jcp.27185

186. Zhang M, Zhou Y, Xie Z, Luo S, Zhou Z, Huang J, et al. New developments in T cell immunometabolism and therapeutic implications for type 1 diabetes. *Front endocrinology.* (2022) 13:914136. doi: 10.3389/fendo.2022.914136

187. Martins CP, New LA, O'Connor EC, Previte DM, Cargill KR, Tse IL, et al. Glycolysis inhibition induces functional and metabolic exhaustion of CD4(+) T cells in type 1 diabetes. *Front Immunol.* (2021) 12:669456. doi: 10.3389/fimmu.2021.669456

188. Ferraro A, Socci C, Stabilini A, Valle A, Monti P, Piemonti L, et al. Expansion of Th17 cells and functional defects in T regulatory cells are key features of the pancreatic lymph nodes in patients with type 1 diabetes. *Diabetes*. (2011) 60:2903–13. doi: 10.2337/db11-0090

189. Jain R, Tartar DM, Gregg RK, Divekar RD, Bell JJ, Lee HH, et al. Innocuous IFNgamma induced by adjuvant-free antigen restores normoglycemia in NOD mice through inhibition of IL-17 production. *J Exp Med.* (2008) 205:207–18. doi: 10.1084/ jem.20071878

190. Chen J, Chernatynskaya AV, Li JW, Kimbrell MR, Cassidy RJ, Perry DJ, et al. T cells display mitochondria hyperpolarization in human type 1 diabetes. *Sci Rep.* (2017) 7:10835. doi: 10.1038/s41598-017-11056-9

191. Chen J, Wong HS, Leong PK, Leung HY, Chan WM, Ko KM. Ursolic acid induces mitochondrial biogenesis through the activation of AMPK and PGC-1 in C2C12 myotubes: a possible mechanism underlying its beneficial effect on exercise endurance. *Food Funct*. (2017) 8:2425–36. doi: 10.1039/C7FO00127D

192. Lagouge M, Argmann C, Gerhart-Hines Z, Meziane H, Lerin C, Daussin F, et al. Resveratrol improves mitochondrial function and protects against metabolic disease by activating SIRT1 and PGC-1alpha. *Cell.* (2006) 127:1109–22. doi: 10.1016/j.cell.2006.11.013

193. Davis JM, Murphy EA, Carmichael MD, Davis B. Quercetin increases brain and muscle mitochondrial biogenesis and exercise tolerance. *Am J Physiol Regulatory Integr Comp Physiol*. (2009) 296:R1071–7. doi: 10.1152/ajpregu.90925.2008

194. Nieman DC, Williams AS, Shanely RA, Jin F, McAnulty SR, Triplett NT, et al. Quercetin's influence on exercise performance and muscle mitochondrial biogenesis. *Med Sci sports exercise*. (2010) 42:338–45. doi: 10.1249/MSS.0b013e3181b18fa3

195. Zhu L, Liu Z, Feng Z, Hao J, Shen W, Li X, et al. Hydroxytyrosol protects against oxidative damage by simultaneous activation of mitochondrial biogenesis and phase II detoxifying enzyme systems in retinal pigment epithelial cells. *J Nutr Biochem.* (2010) 21:1089–98. doi: 10.1016/j.jnutbio.2009.09.006

196. Stojanović I, Šavikin K, Đedović N, Živković J, Saksida T, Momčilović M, et al. Pomegranate peel extract ameliorates autoimmunity in animal models of multiple sclerosis and type 1 diabetes. *J Funct Foods*. (2017) 35:522–30. doi: 10.1016/ j.jff.2017.06.021

197. Nevius E, Srivastava PK, Basu S. Oral ingestion of Capsaicin, the pungent component of chili pepper, enhances a discreet population of macrophages and confers protection from autoimmune diabetes. *Mucosal Immunol.* (2012) 5:76–86. doi: 10.1038/mi.2011.50

198. Endo K, Matsui R, Sugiyama M, Asami T, Inaba C, Kobayashi S, et al. Procyanidin B2 gallate regulates TNF- α production from T cells through inhibiting glycolytic activity via mTOR-HIF-1 pathway. *Biochem Pharmacol.* (2020) 177:113952. doi: 10.1016/j.bcp.2020.113952

199. Abel-Salam BK. Immunomodulatory effects of black seeds and garlic on alloxan-induced Diabetes in albino rat. *Allergologia immunopathologia*. (2012) 40:336-40. doi: 10.1016/j.aller.2011.07.002

200. Hamden K, Masmoudi H, Carreau S, Elfeki A. Immunomodulatory, beta-cell, and neuroprotective actions of fenugreek oil from alloxan-induced diabetes. *Immunopharmacol immunotoxicology*. (2010) 32:437-45. doi: 10.3109/08923970903490486

201. Lu Q, Yang MF, Liang YJ, Xu J, Xu HM, Nie YQ, et al. Immunology of inflammatory bowel disease: molecular mechanisms and therapeutics. *J Inflammation Res.* (2022) 15:1825–44. doi: 10.2147/JIR.S353038

202. Kosoy R, Kim-Schulze S, Rahman A, Friedman JR, Huang R, Peters LA, et al. Deep analysis of the peripheral immune system in IBD reveals new insight in disease subtyping and response to monotherapy or combination therapy. *Cell Mol Gastroenterol hepatology.* (2021) 12:599–632. doi: 10.1016/j.jcmgh.2021.03.012

203. Pan X, Zhu Q, Pan LL, Sun J. Macrophage immunometabolism in inflammatory bowel diseases: From pathogenesis to therapy. *Pharmacol Ther.* (2022) 238:108176. doi: 10.1016/j.pharmthera.2022.108176

204. Hochrein SM, Wu H, Eckstein M, Arrigoni L, Herman JS, Schumacher F, et al. The glucose transporter GLUT3 controls T helper 17 cell responses through glycolytic-epigenetic reprogramming. *Cell Metab.* (2022) 34:516–32.e11. doi: 10.1016/j.cmet.2022.02.015

205. Pålsson-McDermott EM, O'Neill LAJ. Targeting immunometabolism as an anti-inflammatory strategy. *Cell Res.* (2020) 30:300–14. doi: 10.1038/s41422-020-0291-z

206. Lee H, Jeon JH, Kim ES. Mitochondrial dysfunctions in T cells: focus on inflammatory bowel disease. *Front Immunol.* (2023) 14:1219422. doi: 10.3389/fimmu.2023.1219422

207. Sánchez-Quintero MJ, Rodríguez-Díaz C, Rodríguez-González FJ, Fernández-Castañer A, García-Fuentes E, López-Gómez C. Role of mitochondria in inflammatory bowel diseases: A systematic review. *Int J Mol Sci.* (2023) 24(23):17124. doi: 10.3390/ ijms242317124

208. Alula KM, Theiss AL. Autophagy in crohn's disease: converging on dysfunctional innate immunity. *Cells.* (2023) 12(13):1779. doi: 10.3390/cells12131779

209. Gao P, Liu H, Huang H, Sun Y, Jia B, Hou B, et al. The Crohn Diseaseassociated ATG16L1(T300A) polymorphism regulates inflammatory responses by modulating TLR- and NLR-mediated signaling. *Autophagy*. (2022) 18:2561–75. doi: 10.1080/15548627.2022.2039991

210. Chu H, Khosravi A, Kusumawardhani IP, Kwon AH, Vasconcelos AC, Cunha LD, et al. Gene-microbiota interactions contribute to the pathogenesis of inflammatory bowel disease. *Sci (New York NY).* (2016) 352:1116–20. doi: 10.1126/science.aad9948

211. Xuan H, Ou A, Hao S, Shi J, Jin X. Galangin protects against symptoms of dextran sodium sulfate-induced acute colitis by activating autophagy and modulating the gut microbiota. *Nutrients*. (2020) 12(2):347. doi: 10.3390/nu12020347

212. Ohtsu A, Shibutani Y, Seno K, Iwata H, Kuwayama T, Shirasuna K. Advanced glycation end products and lipopolysaccharides stimulate interleukin-6 secretion via the RAGE/TLR4-NF- κ B-ROS pathways and resveratrol attenuates these inflammatory responses in mouse macrophages. *Exp Ther Med.* (2017) 14:4363–70. doi: 10.3892/etm

213. Duan WJ, Li YF, Liu FL, Deng J, Wu YP, Yuan WL, et al. A SIRT3/AMPK/ autophagy network orchestrates the protective effects of trans-resveratrol in stressed peritoneal macrophages and RAW 264.7 macrophages. *Free Radical Biol Med.* (2016) 95:230–42. doi: 10.1016/j.freeradbiomed.2016.03.022

214. Yang G, Xue Y, Zhang H, Du M, Zhu MJ. Favourable effects of grape seed extract on intestinal epithelial differentiation and barrier function in IL10-deficient mice. *Br J Nutr.* (2015) 114:15–23. doi: 10.1017/S0007114515001415

215. Chen F, Liu Q, Xiong Y, Xu L. Current strategies and potential prospects of nanomedicine-mediated therapy in inflammatory bowel disease. *Int J nanomedicine*. (2021) 16:4225–37. doi: 10.2147/IJN.S310952

216. Yao J, Wei C, Wang JY, Zhang R, Li YX, Wang LS. Effect of resveratrol on Treg/ Th17 signaling and ulcerative colitis treatment in mice. *World J gastroenterology*. (2015) 21:6572–81. doi: 10.3748/wjg.v21.i21.6572

217. Yashiro T, Yura S, Tobita A, Toyoda Y, Kasakura K, Nishiyama C. Pterostilbene reduces colonic inflammation by suppressing dendritic cell activation and promoting regulatory T cell development. *FASEB J.* (2020) 34:14810–9. doi: 10.1096/fj.202001502R

218. Singh UP, Singh NP, Singh B, Hofseth LJ, Price RL, Nagarkatti M, et al. Resveratrol (trans-3,5,4'-trihydroxystilbene) induces silent mating type information

regulation-1 and down-regulates nuclear transcription factor-kappaB activation to abrogate dextran sulfate sodium-induced colitis. *J Pharmacol Exp Ther.* (2010) 332:829–39. doi: 10.1124/jpet.109.160838

219. Zhong Y-B, Kang Z-P, Wang M-X, Long J, Wang H-Y, Huang J-Q, et al. Curcumin ameliorated dextran sulfate sodium-induced colitis via regulating the homeostasis of DCs and Treg and improving the composition of the gut microbiota. *J Funct Foods.* (2021) 86:104716. doi: 10.1016/j.jff.2021.104716

220. Kang ZP, Wang MX, Wu TT, Liu DY, Wang HY, Long J, et al. Curcumin alleviated dextran sulfate sodium-induced colitis by regulating M1/M2 macrophage polarization and TLRs signaling pathway. *Evidence-Based complementary Altern medicine: eCAM.* (2021) 2021:3334994. doi: 10.1155/2021/3334994

221. Wu X, Dou Y, Yang Y, Bian D, Luo J, Tong B, et al. Arctigenin exerts anti-colitis efficacy through inhibiting the differentiation of Th1 and Th17 cells via an mTORC1-dependent pathway. *Biochem Pharmacol.* (2015) 96:323–36. doi: 10.1016/j.bcp.2015.06.008

222. Huang B, Wang Q, Jiang L, Lu S, Li C, Xu C, et al. Shikonin ameliorated mice colitis by inhibiting dimerization and tetramerization of PKM2 in macrophages. *Front Pharmacol.* (2022) 13:926945. doi: 10.3389/fphar.2022.926945

223. Alves-Santos AM, Sugizaki CSA, Lima GC, Naves MMV. Prebiotic effect of dietary polyphenols: A systematic review. J Funct Foods. (2020) 74:104169. doi: 10.1016/j.jff.2020.104169

224. Zhao Y, Jiang Q. Roles of the polyphenol-gut microbiota interaction in alleviating colitis and preventing colitis-associated colorectal cancer. *Adv Nutr* (*Bethesda Md*). (2021) 12:546–65. doi: 10.1093/advances/nmaa104

225. Shimotoyodome A, Meguro S, Hase T, Tokimitsu I, Sakata T. Short chain fatty acids but not lactate or succinate stimulate mucus release in the rat colon. *Comp Biochem Physiol Part A Mol Integr Physiol.* (2000) 125:525–31. doi: 10.1016/S1095-6433(00)00183-5

226. Chang PV, Hao L, Offermanns S, Medzhitov R. The microbial metabolite butyrate regulates intestinal macrophage function via histone deacetylase inhibition. *Proc Natl Acad Sci United States America*. (2014) 111:2247–52. doi: 10.1073/pnas.1322269111

227. Zaiatz Bittencourt V, Jones F, Doherty G, Ryan EJ. Targeting immune cell metabolism in the treatment of inflammatory bowel disease. *Inflammatory bowel diseases*. (2021) 27:1684–93. doi: 10.1093/ibd/izab024

228. Kim H, Venancio VP, Fang C, Dupont AW, Talcott ST, Mertens-Talcott SU. Mango (Mangifera indica L.) polyphenols reduce IL-8, GRO, and GM-SCF plasma levels and increase Lactobacillus species in a pilot study in patients with inflammatory bowel disease. *Nutr Res (New York NY)*. (2020) 75:85–94. doi: 10.1016/j.nuttres.2020.01.002

229. Maghzi A-H, Borazanci A, McGee J, Steven Alexander J, Gonzalez-Toledo E, Minagar A. 1 - Multiple Sclerosis: Pathophysiology, Clinical Features, Diagnosis, and Management. In: Minagar A, editor. *Neuroinflammation*. Elsevier, London (2011). p. 1–23.

230. Miljković Đ, Jevtić B, Stojanović I, Dimitrijević M. ILC3, a central innate immune component of the gut-brain axis in multiple sclerosis. *Front Immunol.* (2021) 12:657622. doi: 10.3389/fimmu.2021.657622

231. Kaushik DK, Bhattacharya A, Mirzaei R, Rawji KS, Ahn Y, Rho JM, et al. Enhanced glycolytic metabolism supports transmigration of brain-infiltrating macrophages in multiple sclerosis. *J Clin Invest.* (2019) 129:3277–92. doi: 10.1172/ JCI124012

232. Chen Z, Liu M, Li L, Chen L. Involvement of the Warburg effect in non-tumor diseases processes. J Cell Physiol. (2018) 233:2839–49. doi: 10.1002/jcp.25998

233. Wang P-F, Jiang F, Zeng Q-M, Yin W-F, Hu Y-Z, Li Q, et al. Mitochondrial and metabolic dysfunction of peripheral immune cells in multiple sclerosis. *J neuroinflammation*. (2024) 21:28. doi: 10.1186/s12974-024-03016-8

234. Shen D, Liu K, Wang H, Wang H. Autophagy modulation in multiple sclerosis and experimental autoimmune encephalomyelitis. *Clin Exp Immunol.* (2022) 209:140–50. doi: 10.1093/cei/uxac017

235. Misrielal C, Mauthe M, Reggiori F, Eggen BJL. Autophagy in multiple sclerosis: two sides of the same coin. *Front Cell Neurosci.* (2020) 14:603710. doi: 10.3389/fncel.2020.603710

236. Rossi B, Santos-Lima B, Terrabuio E, Zenaro E, Constantin G. Common peripheral immunity mechanisms in multiple sclerosis and alzheimer's disease. *Front Immunol.* (2021) 12:639369. doi: 10.3389/fimmu.2021.639369

237. Koutrolos M, Berer K, Kawakami N, Wekerle H, Krishnamoorthy G. Treg cells mediate recovery from EAE by controlling effector T cell proliferation and motility in the CNS. *Acta neuropathologica Commun.* (2014) 2:163. doi: 10.1186/s40478-014-0163-1

238. Runtsch MC, Ferrara G, Angiari S. Metabolic determinants of leukocyte pathogenicity in neurological diseases. *J neurochemistry*. (2021) 158:36–58. doi: 10.1111/jnc.15169

239. Dierckx T, Haidar M, Grajchen E, Wouters E, Vanherle S, Loix M, et al. Phloretin suppresses neuroinflammation by autophagy-mediated Nrf2 activation in macrophages. *J neuroinflammation*. (2021) 18:148. doi: 10.1186/s12974-021-02194-z

240. Wang J, Ren Z, Xu Y, Xiao S, Meydani SN, Wu D. Epigallocatechin-3-gallate ameliorates experimental autoimmune encephalomyelitis by altering balance among CD4+ T-cell subsets. *Am J pathology*. (2012) 180:221–34. doi: 10.1016/ j.ajpath.2011.09.007

241. Xie L, Li XK, Funeshima-Fuji N, Kimura H, Matsumoto Y, Isaka Y, et al. Amelioration of experimental autoimmune encephalomyelitis by curcumin treatment through inhibition of IL-17 production. *Int immunopharmacology*. (2009) 9:575–81. doi: 10.1016/j.intimp.2009.01.025

242. Haghmorad D, Mahmoudi MB, Salehipour Z, Jalayer Z, Momtazi Brojeni AA, Rastin M, et al. Hesperidin ameliorates immunological outcome and reduces neuroinflammation in the mouse model of multiple sclerosis. *J neuroimmunology*. (2017) 302:23–33. doi: 10.1016/j.jneuroim.2016.11.009

243. Wang J, Qi Y, Niu X, Tang H, Meydani SN, Wu D. Dietary naringenin supplementation attenuates experimental autoimmune encephalomyelitis by modulating autoimmune inflammatory responses in mice. *J Nutr Biochem.* (2018) 54:130–9. doi: 10.1016/j.jnutbio.2017.12.004

244. Zheng M, Zhang Q, Joe Y, Lee BH, Ryu DG, Kwon KB, et al. Curcumin induces apoptotic cell death of activated human CD4+ T cells via increasing endoplasmic reticulum stress and mitochondrial dysfunction. *Int immunopharmacology*. (2013) 15:517–23. doi: 10.1016/j.intimp.2013.02.002

245. Zou T, Yang Y, Xia F, Huang A, Gao X, Fang D, et al. Resveratrol Inhibits CD4+ T cell activation by enhancing the expression and activity of Sirt1. *PloS One*. (2013) 8: e75139. doi: 10.1371/journal.pone.0075139

246. Frydrych LM, Bian G, O'Lone DE, Ward PA, Delano MJ. Obesity and type 2 diabetes mellitus drive immune dysfunction, infection development, and sepsis mortality. *J leukocyte Biol.* (2018) 104:525-34. doi: 10.1002/JLB.5VMR0118-021RR

247. Monteiro L, Pereira J, Palhinha L, Moraes-Vieira PMM. Leptin in the regulation of the immunometabolism of adipose tissue-macrophages. J leukocyte Biol. (2019) 106:703-16. doi: 10.1002/JLB.MR1218-478R

248. Wang B, Zhong Y, Li Q, Cui L, Huang G. Autophagy of macrophages is regulated by PI3k/Akt/mTOR signalling in the development of diabetic encephalopathy. *Aging.* (2018) 10:2772-82. doi: 10.18632/aging.v10i10

249. Rocha M, Apostolova N, Diaz-Rua R, Muntane J, Victor VM. Mitochondria and T2D: role of autophagy, ER stress, and inflammasome. *Trends Endocrinol metabolism: TEM.* (2020) 31:725–41. doi: 10.1016/j.tem.2020.03.004

250. Nakahira K, Haspel JA, Rathinam VA, Lee SJ, Dolinay T, Lam HC, et al. Autophagy proteins regulate innate immune responses by inhibiting the release of mitochondrial DNA mediated by the NALP3 inflammasome. *Nat Immunol.* (2011) 12:222–30. doi: 10.1038/ni.1980

251. Xu L, Yan X, Zhao Y, Wang J, Liu B, Yu S, et al. Macrophage polarization mediated by mitochondrial dysfunction induces adipose tissue inflammation in obesity. *Int J Mol Sci.* (2022) 23(16):9252. doi: 10.3390/ijms23169252

252. Wang L, Hu J, Zhou H. Macrophage and adipocyte mitochondrial dysfunction in obesity-induced metabolic diseases. *World J men's Health*. (2021) 39:606–14. doi: 10.5534/wjmh.200163

253. Diaz-Morales N, Rovira-Llopis S, Bañuls C, Escribano-Lopez I, de Marañon AM, Lopez-Domenech S, et al. Are mitochondrial fusion and fission impaired in leukocytes of type 2 diabetic patients? *Antioxidants Redox Signaling*. (2016) 25:108–15. doi: 10.1089/ars.2016.6707

254. Schmidt V, Hogan AE, Fallon PG, Schwartz C. Obesity-mediated immune modulation: one step forward, (Th)2 steps back. *Front Immunol.* (2022) 13. doi: 10.3389/fimmu.2022.932893

255. Boutens L, Hooiveld GJ, Dhingra S, Cramer RA, Netea MG, Stienstra R. Unique metabolic activation of adipose tissue macrophages in obesity promotes inflammatory responses. *Diabetologia*. (2018) 61:942–53. doi: 10.1007/s00125-017-4526-6

256. Callender LA, Carroll EC, Garrod-Ketchley C, Schroth J, Bystrom J, Berryman V, et al. Altered nutrient uptake causes mitochondrial dysfunction in senescent CD8(+) EMRA T cells during type 2 diabetes. *Front aging*. (2021) 2:681428. doi: 10.3389/fragi.2021.681428

257. Verdura S, Cuyàs E, Martin-Castillo B, Menendez JA. Metformin as an archetype immuno-metabolic adjuvant for cancer immunotherapy. *Oncoimmunology.* (2019) 8:e1633235. doi: 10.1080/2162402X.2019.1633235

258. Bahrambeigi S, Shafiei-Irannejad V. Immune-mediated anti-tumor effects of metformin; targeting metabolic reprogramming of T cells as a new possible mechanism for anti-cancer effects of metformin. *Biochem Pharmacol.* (2020) 174:113787. doi: 10.1016/j.bcp.2019.113787

259. Deng X, Zhang S, Wu J, Sun X, Shen Z, Dong J, et al. Promotion of Mitochondrial Biogenesis via Activation of AMPK-PGC1 α Signaling Pathway by Ginger (Zingiber officinale Roscoe) Extract, and Its Major Active Component 6-Gingerol. J Food sci. (2019) 84:2101–11. doi: 10.1111/1750-3841.14723

260. Taub PR, Ramirez-Sanchez I, Ciaraldi TP, Perkins G, Murphy AN, Naviaux R, et al. Alterations in skeletal muscle indicators of mitochondrial structure and biogenesis in patients with type 2 diabetes and heart failure: effects of epicatechin rich cocoa. *Clin Trans sci.* (2012) 5:43–7. doi: 10.1111/j.1752-8062.2011.00357.x

261. Tsutsumi R, Yoshida T, Nii Y, Okahisa N, Iwata S, Tsukayama M, et al. Sudachitin, a polymethoxylated flavone, improves glucose and lipid metabolism by increasing mitochondrial biogenesis in skeletal muscle. *Nutr Metab.* (2014) 11:32. doi: 10.1186/1743-7075-11-32

262. Lee MS, Shin Y, Jung S, Kim Y. Effects of epigallocatechin-3-gallate on thermogenesis and mitochondrial biogenesis in brown adipose tissues of diet-induced obese mice. *Food Nutr Res.* (2017) 61:1325307. doi: 10.1080/16546628.2017.1325307

263. Yan J, Feng Z, Liu J, Shen W, Wang Y, Wertz K, et al. Enhanced autophagy plays a cardinal role in mitochondrial dysfunction in type 2 diabetic Goto-Kakizaki (GK) rats: ameliorating effects of (-)-epigallocatechin-3-gallate. *J Nutr Biochem.* (2012) 23:716–24. doi: 10.1016/j.jnutbio.2011.03.014

264. Liu J, Tang Y, Feng Z, Liu J, Liu J, Long J. (-)-Epigallocatechin-3-gallate attenuated myocardial mitochondrial dysfunction and autophagy in diabetic Goto-Kakizaki rats. *Free Radical Res.* (2014) 48:898–906. doi: 10.3109/10715762.2014.920955

265. García-Martínez BI, Ruiz-Ramos M, Pedraza-Chaverri J, Santiago-Osorio E, Mendoza-Núñez VM. Effect of resveratrol on markers of oxidative stress and sirtuin 1 in elderly adults with type 2 diabetes. *Int J Mol Sci.* (2023) 24(8):7422. doi: 10.3390/ ijms24087422

266. Kang JH, Kim CS, Han IS, Kawada T, Yu R. Capsaicin, a spicy component of hot peppers, modulates adipokine gene expression and protein release from obesemouse adipose tissues and isolated adipocytes, and suppresses the inflammatory responses of adipose tissue macrophages. *FEBS letters*. (2007) 581:4389–96. doi: 10.1016/j.febslet.2007.07.082

267. Prior RL, Wu X, Gu L, Hager T, Hager A, Wilkes S, et al. Purified berry anthocyanins but not whole berries normalize lipid parameters in mice fed an obesogenic high fat diet. *Mol Nutr Food Res.* (2009) 53:1406–18. doi: 10.1002/mnfr.200900026

268. Kang MK, Li J, Kim JL, Gong JH, Kwak SN, Park JH, et al. Purple corn anthocyanins inhibit diabetes-associated glomerular monocyte activation and macrophage infiltration. *Am J Physiol Renal Physiol.* (2012) 303:F1060-9. doi: 10.1152/ajprenal.00106.2012

269. Kang JH, Tsuyoshi G, Le Ngoc H, Kim HM, Tu TH, Noh HJ, et al. Dietary capsaicin attenuates metabolic dysregulation in genetically obese diabetic mice. *J medicinal Food*. (2011) 14:310–5. doi: 10.1089/jmf.2010.1367

270. Yekollu SK, Thomas R, O'Sullivan B. Targeting curcusomes to inflammatory dendritic cells inhibits NF- κ B and improves insulin resistance in obese mice. *Diabetes*. (2011) 60:2928–38. doi: 10.2337/db11-0275

271. Arafa MG, Ghalwash D, El-Kersh DM, Elmazar MM. Propolis-based niosomes as oromuco-adhesive films: A randomized clinical trial of a therapeutic drug delivery platform for the treatment of oral recurrent aphthous ulcers. *Sci Rep.* (2018) 8:18056. doi: 10.1038/s41598-018-37157-7

272. Ryyti R, Hämäläinen M, Leppänen T, Peltola R, Moilanen E. Phenolic Compounds Known to Be Present in Lingonberry (Vaccinium vitis-idaea L.) Enhance Macrophage Polarization towards the Anti-Inflammatory M2 Phenotype. *Biomedicines.* (2022) 10(12):3045. doi: 10.3390/biomedicines10123045

273. Jiang J, Zhang G, Yu M, Gu J, Zheng Y, Sun J, et al. Quercetin improves the adipose inflammatory response and insulin signaling to reduce "real-world" particulate matter-induced insulin resistance. *Environ Sci pollut Res Int.* (2022) 29:2146–57. doi: 10.1007/s11356-021-15829-8

274. Fan R, You M, Toney AM, Kim J, Giraud D, Xian Y, et al. Red Raspberry Polyphenols Attenuate High-Fat Diet-Driven Activation of NLRP3 Inflammasome and its Paracrine Suppression of Adipogenesis via Histone Modifications. *Mol Nutr Food Res.* (2020) 64:e1900995. doi: 10.1002/mnfr.201900995

275. Molina N, Bolin AP, Otton R. Green tea polyphenols change the profile of inflammatory cytokine release from lymphocytes of obese and lean rats and protect against oxidative damage. *Int immunopharmacology*. (2015) 28:985–96. doi: 10.1016/j.intimp.2015.08.011

276. Widyaningsih T, Siska A, Fanani R, Martati E. Traditional drink of black cincau (Mesona palustris BL)-based wedang uwuh as immunomodulator on alloxan-induced diabetic rats. *Nutr Food Sci.* (2020) 50(6):1123–33. doi: 10.1108/NFS-05-2019-0165

277. Martinez-Micaelo N, González-Abuín N, Mulero M, Pinent M, Ardévol A, Blay M. Procyanidins and docosahexaenoic acid suppress inflammation and boost immune system in cafeteria diet-fed rats. *J Funct Foods*. (2015) 15:61–71. doi: 10.1016/j.jff.2015.02.017

278. Byun JK, Yoon BY, Jhun JY, Oh HJ, Kim EK, Min JK, et al. Epigallocatechin-3gallate ameliorates both obesity and autoinflammatory arthritis aggravated by obesity by altering the balance among CD4+ T-cell subsets. *Immunol Lett.* (2014) 157:51–9. doi: 10.1016/j.imlet.2013.11.006

279. Doty KR, Guillot-Sestier MV, Town T. The role of the immune system in neurodegenerative disorders: Adaptive or maladaptive? *Brain Res.* (2015) 1617:155–73. doi: 10.1016/j.brainres.2014.09.008

280. Zang X, Chen S, Zhu J, Ma J, Zhai Y. The emerging role of central and peripheral immune systems in neurodegenerative diseases. *Front Aging Neurosci.* (2022) 14:872134. doi: 10.3389/fnagi.2022.872134

281. Marsh SE, Abud EM, Lakatos A, Karimzadeh A, Yeung ST, Davtyan H, et al. The adaptive immune system restrains Alzheimer's disease pathogenesis by modulating microglial function. *Proc Natl Acad Sci United States America*. (2016) 113:E1316–25. doi: 10.1073/pnas.1525466113

282. Kim K, Wang X, Ragonnaud E, Bodogai M, Illouz T, DeLuca M, et al. Therapeutic B-cell depletion reverses progression of Alzheimer's disease. *Nat Commun.* (2021) 12:2185. doi: 10.1038/s41467-021-22479-4

283. Mitra S, Banik A, Saurabh S, Maulik M, Khatri SN. Neuroimmunometabolism: A new pathological nexus underlying neurodegenerative disorders. *J neuroscience: Off J Soc Neurosci.* (2022) 42:1888–907. doi: 10.1523/JNEUROSCI.0998-21.2022

 Lin MM, Liu N, Qin ZH, Wang Y. Mitochondrial-derived damage-associated molecular patterns amplify neuroinflammation in neurodegenerative diseases. Acta pharmacologica Sinica. (2022) 43:2439–47. doi: 10.1038/s41401-022-00879-6 285. Houtman J, Freitag K, Gimber N, Schmoranzer J, Heppner FL, Jendrach M. Beclin1-driven autophagy modulates the inflammatory response of microglia via NLRP3. *EMBO J.* (2019) 38(4):e99430. doi: 10.15252/embj.201899430

286. Chen X, Firulyova M, Manis M, Herz J, Smirnov I, Aladyeva E, et al. Microgliamediated T cell infiltration drives neurodegeneration in tauopathy. *Nature.* (2023) 615:668–77. doi: 10.1038/s41586-023-05788-0

287. Lowinus T, Bose T, Busse S, Busse M, Reinhold D, Schraven B, et al. Immunomodulation by memantine in therapy of Alzheimer's disease is mediated through inhibition of Kv1.3 channels and T cell responsiveness. *Oncotarget.* (2016) 7:53797–807. doi: 10.18632/oncotarget.v7i33

288. Cordero JG, García-Escudero R, Avila J, Gargini R, García-Escudero V. Benefit of oleuropein aglycone for alzheimer's disease by promoting autophagy. Oxid Med Cell longevity. (2018) 2018:5010741. doi: 10.1155/2018/5010741

289. Rigacci S, Miceli C, Nediani C, Berti A, Cascella R, Pantano D, et al. Oleuropein aglycone induces autophagy via the AMPK/mTOR signalling pathway: a mechanistic insight. *Oncotarget.* (2015) 6:35344–57. doi: 10.18632/oncotarget.v6i34

290. Wang C, Zhang X, Teng Z, Zhang T, Li Y. Downregulation of PI3K/Akt/mTOR signaling pathway in curcumin-induced autophagy in APP/PS1 double transgenic mice. *Eur J Pharmacol.* (2014) 740:312–20. doi: 10.1016/j.ejphar.2014.06.051

291. Capiralla H, Vingtdeux V, Zhao H, Sankowski R, Al-Abed Y, Davies P, et al. Resveratrol mitigates lipopolysaccharide- and A β -mediated microglial inflammation by inhibiting the TLR4/NF-KB/STAT signaling cascade. *J neurochemistry*. (2012) 120:461–72. doi: 10.1111/j.1471-4159.2011.07594.x

292. Yang X, Xu S, Qian Y, Xiao Q. Resveratrol regulates microglia M1/M2 polarization via PGC-1 α in conditions of neuroinflammatory injury. *Brain behavior immunity.* (2017) 64:162–72. doi: 10.1016/j.bbi.2017.03.003

293. Moussa C, Hebron M, Huang X, Ahn J, Rissman RA, Aisen PS, et al. Resveratrol regulates neuro-inflammation and induces adaptive immunity in Alzheimer's disease. *J neuroinflammation*. (2017) 14:1. doi: 10.1186/s12974-016-0779-0

294. Ali T, Kim T, Rehman SU, Khan MS, Amin FU, Khan M, et al. Natural dietary supplementation of anthocyanins via PI3K/akt/nrf2/HO-1 pathways mitigate oxidative stress, neurodegeneration, and memory impairment in a mouse model of alzheimer's disease. *Mol neurobiology*. (2018) 55:6076–93. doi: 10.1007/s12035-017-0798-6

295. Gallardo-Fernández M, Hornedo-Ortega R, Alonso-Bellido IM, Rodríguez-Gómez JA, Troncoso AM, García-Parrilla MC, et al. Hydroxytyrosol decreases LPSand α -synuclein-induced microglial activation in vitro. *Antioxidants (Basel Switzerland)*. (2019) 9(1):36. doi: 10.3390/antiox9010036

296. Jin X, Liu MY, Zhang DF, Zhong X, Du K, Qian P, et al. Baicalin mitigates cognitive impairment and protects neurons from microglia-mediated neuroinflammation via suppressing NLRP3 inflammasomes and TLR4/NF- κ B signaling pathway. *CNS Neurosci Ther.* (2019) 25:575–90. doi: 10.1111/cns.13086

297. Kim MJ, Seong AR, Yoo JY, Jin CH, Lee YH, Kim YJ, et al. Gallic acid, a histone acetyltransferase inhibitor, suppresses β -amyloid neurotoxicity by inhibiting microglial-mediated neuroinflammation. *Mol Nutr Food Res.* (2011) 55:1798–808. doi: 10.1002/mnfr.201100262

298. Hamzeh O, Rabiei F, Shakeri M, Parsian H, Saadat P, Rostami-Mansoor S. Mitochondrial dysfunction and inflammasome activation in neurodegenerative diseases: Mechanisms and therapeutic implications. *Mitochondrion*. (2023) 73:72–83. doi: 10.1016/j.mito.2023.10.003

299. Sarkar S, Malovic E, Harishchandra DS, Ghaisas S, Panicker N, Charli A, et al. Mitochondrial impairment in microglia amplifies NLRP3 inflammasome proinflammatory signaling in cell culture and animal models of Parkinson's disease. *NPJ Parkinson's disease*. (2017) 3:30. doi: 10.1038/s41531-017-0032-2

300. Qin Y, Qiu J, Wang P, Liu J, Zhao Y, Jiang F, et al. Impaired autophagy in microglia aggravates dopaminergic neurodegeneration by regulating NLRP3 inflammasome activation in experimental models of Parkinson's disease. *Brain behavior immunity.* (2021) 91:324–38. doi: 10.1016/j.bbi.2020.10.010

301. Yan A, Zhang Y, Lin J, Song L, Wang X, Liu Z. Partial depletion of peripheral M1 macrophages reverses motor deficits in MPTP-treated mouse by suppressing neuroinflammation and dopaminergic neurodegeneration. *Front Aging Neurosci.* (2018) 10:160. doi: 10.3389/fnagi.2018.00160

302. Zhu H, Lemos H, Bhatt B, Islam B, Singh A, Gurav A, et al. Carbidopa, a drug in use for management of Parkinson disease inhibits T cell activation and autoimmunity. *PloS One.* (2017) 12:e0183484. doi: 10.1371/journal.pone.0183484

303. Sommer A, Marxreiter F, Krach F, Fadler T, Grosch J, Maroni M, et al. Th17 lymphocytes induce neuronal cell death in a human iPSC-based model of parkinson's disease. *Cell Stem Cell*. (2018) 23:123–31.e6. doi: 10.1016/j.stem.2018.06.015

304. Lofrumento DD, Nicolardi G, Cianciulli A, De Nuccio F, La Pesa V, Carofiglio V, et al. Neuroprotective effects of resveratrol in an MPTP mouse model of Parkinson'slike disease: possible role of SOCS-1 in reducing pro-inflammatory responses. *Innate immunity*. (2014) 20:249–60. doi: 10.1177/1753425913488429

305. Yang S, Zhang D, Yang Z, Hu X, Qian S, Liu J, et al. Curcumin protects dopaminergic neuron against LPS induced neurotoxicity in primary rat neuron/glia culture. *Neurochemical Res.* (2008) 33:2044–53. doi: 10.1007/s11064-008-9675-z

306. Jiang TF, Zhang YJ, Zhou HY, Wang HM, Tian LP, Liu J, et al. Curcumin ameliorates the neurodegenerative pathology in A53T α -synuclein cell model of Parkinson's disease through the downregulation of mTOR/p70S6K signaling and the

recovery of macroautophagy. J neuroimmune Pharmacol. (2013) 8:356-69. doi: 10.1007/s11481-012-9431-7

307. Wang D, Gu X, Ma X, Chen J, Zhang Q, Yu Z, et al. Nanopolyphenol rejuvenates microglial surveillance of multiple misfolded proteins through metabolic reprogramming. *Acta Pharm Sin B.* (2023) 13:834–51. doi: 10.1016/j.apsb.2022.07.014

308. Alberdi E, Sánchez-Gómez MV, Ruiz A, Cavaliere F, Ortiz-Sanz C, Quintela-López T, et al. Mangiferin and morin attenuate oxidative stress, mitochondrial dysfunction, and neurocytotoxicity, induced by amyloid beta oligomers. *Oxid Med Cell longevity.* (2018) 2018:2856063. doi: 10.1155/2018/2856063

309. Wang WW, Han R, He HJ, Li J, Chen SY, Gu Y, et al. Administration of quercetin improves mitochondria quality control and protects the neurons in 6-OHDA-lesioned Parkinson's disease models. *Aging.* (2021) 13:11738-51. doi: 10.18632/aging.v13i8

310. Wang X, Chen S, Ma G, Ye M, Lu G. Genistein protects dopaminergic neurons by inhibiting microglial activation. *Neuroreport.* (2005) 16:267–70. doi: 10.1097/00001756-200502280-00013

311. Wang N, Li R, Feng B, Cheng Y, Guo Y, Qian H. Chicoric acid prevents neuroinflammation and neurodegeneration in a mouse parkinson's disease model: immune response and transcriptome profile of the spleen and colon. *Int J Mol Sci.* (2022) 23(4):2031. doi: 10.3390/ijms23042031

312. Malone K, Amu S, Moore AC, Waeber C. The immune system and stroke: from current targets to future therapy. *Immunol Cell Biol.* (2019) 97:5–16. doi: 10.1111/ imcb.12191

313. Wu F, Liu Z, Zhou L, Ye D, Zhu Y, Huang K, et al. Systemic immune responses after ischemic stroke: From the center to the periphery. *Front Immunol.* (2022) 13:911661. doi: 10.3389/fimmu.2022.911661

314. Ong LK, Walker F, Nilsson M. Is stroke a neurodegenerative condition? A critical review of secondary neurodegeneration and amyloid-beta accumulation after stroke. *AIMS Med Science*. (2017) 4:1–16. doi: 10.3934/medsci.2017.1.1

315. Kamel H, Iadecola C. Brain-immune interactions and ischemic stroke: clinical implications. *Arch neurology*. (2012) 69:576–81. doi: 10.1001/archneurol.2011.3590

316. Chen W, Zhang Y, Zhai X, Xie L, Guo Y, Chen C, et al. Microglial phagocytosis and regulatory mechanisms after stroke. *J Cereb Blood Flow Metab*. (2022) 42:1579–96. doi: 10.1177/0271678X221098841

317. Park J, Choi H, Min JS, Park SJ, Kim JH, Park HJ, et al. Mitochondrial dynamics modulate the expression of pro-inflammatory mediators in microglial cells. *J neurochemistry*. (2013) 127:221–32. doi: 10.1111/jnc.12361

318. Liu W, Qi Z, Li W, Liang J, Zhao L, Shi Y. M1 microglia induced neuronal injury on ischemic stroke via mitochondrial crosstalk between microglia and neurons. *Oxid Med Cell longevity.* (2022) 2022:4335272. doi: 10.1155/2022/4335272

319. Mo Y, Sun YY, Liu KY. Autophagy and inflammation in ischemic stroke. Neural regeneration Res. (2020) 15:1388-96. doi: 10.4103/1673-5374.274331

320. Yang Z, Zhong L, Zhong S, Xian R, Yuan B. Hypoxia induces microglia autophagy and neural inflammation injury in focal cerebral ischemia model. *Exp Mol pathology.* (2015) 98:219–24. doi: 10.1016/j.yexmp.2015.02.003

321. Beccari S, Sierra-Torre V, Valero J, Pereira-Iglesias M, García-Zaballa M, Soria FN, et al. Microglial phagocytosis dysfunction in stroke is driven by energy depletion and induction of autophagy. *Autophagy*. (2023) 19:1952–81. doi: 10.1080/15548627.2023.2165313

322. Takeda H, Yamaguchi T, Yano H, Tanaka J. Microglial metabolic disturbances and neuroinflammation in cerebral infarction. *J Pharmacol Sci.* (2021) 145:130–9. doi: 10.1016/j.jphs.2020.11.007

323. Li Y, Lu B, Sheng L, Zhu Z, Sun H, Zhou Y, et al. Hexokinase 2-dependent hyperglycolysis driving microglial activation contributes to ischemic brain injury. *J neurochemistry*. (2018) 144:186–200. doi: 10.1111/jnc.14267

324. Fu Y, Liu Q, Anrather J, Shi FD. Immune interventions in stroke. Nat Rev Neurology. (2015) 11:524-35. doi: 10.1038/nrneurol.2015.144

325. Cankaya S, Cankaya B, Kilic U, Kilic E, Yulug B. The therapeutic role of minocycline in Parkinson's disease. *Drugs context.* (2019) 8:212553. doi: 10.7573/dic.212553

326. Lu Y, Zhou M, Li Y, Li Y, Hua Y, Fan Y. Minocycline promotes functional recovery in ischemic stroke by modulating microglia polarization through STAT1/ STAT6 pathways. *Biochem Pharmacol.* (2021) 186:114464. doi: 10.1016/ j.bcp.2021.114464

327. Li X, Syrovets T, Genze F, Pitterle K, Oberhuber A, Orend KH, et al. Plasmin triggers chemotaxis of monocyte-derived dendritic cells through an Akt2-dependent pathway and promotes a T-helper type-1 response. *Arteriosclerosis thrombosis Vasc Biol.* (2010) 30:582–90. doi: 10.1161/ATVBAHA.109.202044

328. Liberale L, Bertolotto M, Minetti S, Contini P, Verzola D, Ameri P, et al. Recombinant tissue plasminogen activator (r-tPA) induces *in-vitro* human neutrophil migration via low density lipoprotein receptor-related protein 1 (LRP-1). *Int J Mol Sci.* (2020) 21(19):7014. doi: 10.3390/ijms21197014

329. Gelderblom M, Leypoldt F, Lewerenz J, Birkenmayer G, Orozco D, Ludewig P, et al. The flavonoid fisetin attenuates postischemic immune cell infiltration, activation and infarct size after transient cerebral middle artery occlusion in mice. *J Cereb Blood Flow Metab.* (2012) 32:835–43. doi: 10.1038/jcbfm.2011.189

330. Lan X, Han X, Li Q, Li Q, Gao Y, Cheng T, et al. Pinocembrin protects hemorrhagic brain primarily by inhibiting toll-like receptor 4 and reducing M1 phenotype microglia. *Brain behavior immunity*. (2017) 61:326–39. doi: 10.1016/j.bbi.2016.12.012

331. Wang Q, Sun AY, Simonyi A, Jensen MD, Shelat PB, Rottinghaus GE, et al. Neuroprotective mechanisms of curcumin against cerebral ischemia-induced neuronal apoptosis and behavioral deficits. *J Neurosci Res.* (2005) 82:138–48. doi: 10.1002/(ISSN) 1097-4547

332. Ran Y, Qie S, Gao F, Ding Z, Yang S, Tian G, et al. Baicalein ameliorates ischemic brain damage through suppressing proinflammatory microglia polarization via inhibiting the TLR4/NF- κ B and STAT1 pathway. *Brain Res.* (2021) 1770:147626. doi: 10.1016/j.brainres.2021.147626

333. Qu Y, Wang L, Mao Y. Gallic acid attenuates cerebral ischemia/re-perfusioninduced blood-brain barrier injury by modifying polarization of microglia. *J Immunotoxicology.* (2022) 19:17–26. doi: 10.1080/1547691X.2022.2043494 334. Zhou YP, Li GC. Kaempferol protects cell damage in *in vitro* ischemia reperfusion model in rat neuronal PC12 cells. *BioMed Res Int.* (2020) 2020:2461079. doi: 10.1155/2020/2461079

335. Dou Z, Rong X, Zhao E, Zhang L, Lv Y. Neuroprotection of resveratrol against focal cerebral ischemia/reperfusion injury in mice through a mechanism targeting gutbrain axis. *Cell Mol neurobiology*. (2019) 39:883–98. doi: 10.1007/s10571-019-00687-3

336. Kominsky DJ, Campbell EL, Colgan SP. Metabolic shifts in immunity and inflammation. J Immunol (Baltimore Md: 1950). (2010) 184:4062–8. doi: 10.4049/jimmunol.0903002

337. Chen Y, Michalak M, Agellon LB. Importance of nutrients and nutrient metabolism on human health. Yale J Biol Med. (2018) 91:95–103.

338. Cuevas A, Saavedra N, Salazar LA, Abdalla DS. Modulation of immune function by polyphenols: possible contribution of epigenetic factors. *Nutrients*. (2013) 5:2314–32. doi: 10.3390/nu5072314