Check for updates

OPEN ACCESS

EDITED BY Helen Carolyn Steel, University of Pretoria, South Africa

REVIEWED BY Meganathan Kannan, Central University of Tamil Nadu, India

*CORRESPONDENCE Saravanan Subramaniam Saravanan subramaniam@mcphs.edu

RECEIVED 26 February 2024 ACCEPTED 08 April 2024 PUBLISHED 23 April 2024

CITATION

Subramaniam S, Mohiuddin N and Jose A (2024) Does SARS-CoV-2 infect platelets? *Front. Immunol.* 15:1392000. doi: 10.3389/fimmu.2024.1392000

COPYRIGHT

© 2024 Subramaniam, Mohiuddin and Jose. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Does SARS-CoV-2 infect platelets?

Saravanan Subramaniam¹*, Naila Mohiuddin¹ and Asha Jose²

¹Department of Pharmacology and Toxicology, School of Pharmacy, Massachusetts College of Pharmacy and Health Sciences, Boston, MA, United States, ²Renal Section, Department of Medicine, Boston Medical Center, Boston University School of Medicine, Boston, MA, United States

KEYWORDS

platelets, COVID-19, inflammation, protease-activated receptors, coagulation

Introduction

COVID-19 is characterized by dysregulated thrombo-inflammatory responses and coagulation, leading to an increased risk of mortality in affected patients (1–3). Platelets play a crucial role as rapid responders to the presence of pathogens, alerting nearby immune cells and contributing to intravascular coagulation (4). In cases of Acute Respiratory Distress Syndrome (ARDS), respiratory failure and thrombocytopenia account for 70% of COVID-19-related deaths (5). The remaining fatalities result from a sepsis-like condition triggered by a cytokine storm in response to viral infection and secondary infections (6). Various studies have demonstrated that COVID-19 patients have elevated levels of inflammatory cytokines, including IL-6, TNF- α , IL-1 β , and IL-18 (7, 8).

Administration of steroids or anti-IL-6 monoclonal antibodies has been associated with increased survival (9). Initial observations of severe lung necrosis, hyperinflammation, vascular damage, thrombosis, and coagulation prompted therapeutic trials targeting classical platelet activation or the coagulation cascade (4, 10). Microvascular thrombosis of the pulmonary vasculature and other vascular beds is frequently observed in COVID-19 (11). Patients in the intensive care unit with COVID-19 exhibit an elevated risk of ischemic stroke and disseminated intravascular coagulation, as indicated by increased C-reactive protein, D-dimer, and P-selectin levels (12). Notably, antiplatelet and anticoagulant trials did not consistently demonstrate a beneficial effect, and the lack of efficacy often coincided with the timing of infection and inflammation levels (13). The pathophysiology of COVID-19 has been explored in recent reviews (reviewed in (3, 14, 15). While evidence confirms inevitable platelet activation during COVID-19, the direct infectivity of platelets remains uncertain.

Functions of platelets

Platelets, or thrombocytes, represent indispensable small blood cells crucial for hemostasis, aimed at preventing bleeding. Originating from megakaryocytes, platelets assume a pivotal role in the coagulation cascade (16). Their primary functions encompass the formation of blood clots to staunch wounds, thereby averting excessive bleeding. Platelets exhibit adherence to damaged blood vessels, releasing signaling chemicals that attract

10.3389/fimmu.2024.1392000

additional platelets, thereby instigating clot formation. Furthermore, platelets house granules containing clotting factors and growth factors imperative for tissue repair (17). Deviations in platelet function may precipitate bleeding disorders or hypercoagulability. Regular assessment of platelet levels through blood tests is standard practice. Disorders affecting platelets span thrombocytosis, characterized by elevated platelet counts, and thrombocytopenia, indicative of low platelet levels. In addition to their well-established hemostatic role, emerging research underscores the active involvement of platelets in immune responses.

Platelets promote hemostasis through a series of sequential processes: adhesion, activation, and aggregation (18). Following vascular injury in classical hemostasis, the injured vessel initiates vasoconstriction to limit blood loss. Subsequently, platelets adhere to the injured vessel wall, undergo activation, and form aggregates, constituting the primary and secondary platelet plug. This plug is further stabilized by a dense fibrin mesh formed through the coagulation cascade. Platelet activation is triggered by binding to both von Willebrand factor (vWF) and collagen (19). Upon activation, platelet granules are released, mediated by GPVI, leading to an increase in platelet activators such as ADP (adenosine diphosphate; from dense δ -granules), vWF (from α granules, in addition to endothelial cell-derived vWF), and thromboxane A2 (20). These activators can, in turn, activate neighboring platelets. The binding of ADP and other potent platelet activators to their receptors induces platelet aggregation. Through an internal signaling mechanism, ADP binds to P2Y1 or P2Y12 receptors, resulting in a conformational change of the α IIb β 3 receptor on the platelet surface. In its active conformation, this receptor binds fibrinogen, facilitating platelet aggregation to form a thrombus and prevent bleeding (21).

Platelets and COVID-19

In cases of COVID-19 pneumonia, researchers have observed elevated levels of platelet-granulocyte and platelet-monocyte aggregates (22). Based on previous autopsy studies, high platelet reactivity was linked with dispersed thrombosis in multiple organs, which suggested that SARS-CoV-2-mediated platelet activation, in turn, contributed to the pathophysiology of COVID-19 (23–26). Patients hospitalized with COVID-19 have reported thrombocytopenia, and lower platelet counts have been associated with more adverse clinical outcomes (27). A meta-analysis comprising 31 studies and 7613 participants revealed a reduced platelet count in severe COVID-19 cases, correlating with a 3-fold increase in the risk of developing severe COVID-19 (28).

Increased plasma levels of proteins that could also be plateletderived, such as sCD40LG (CD154), TxB2, P-selectin, and vWF, have confirmed that platelets are being activated during COVID-19. Consistently, platelet activation is evident by increased expression of P-selectin CD40 and CD63 on the surface of platelets from patients with COVID-19. A meta-analysis of 7,613 COVID-19 patients revealed that patients with severe disease had a lower platelet count than those with non-severe disease. Additionally, the non-survivors had a much lower platelet count than the survivors. However, not all studies have found platelet counts to be a predictor of COVID-19 mortality (29). Platelet activation was obvious in individuals with severe COVID-19, as measured by surface expression of P-selectin and CD63, and it was substantially linked with D-dimer levels. Soluble P-selectin levels were considerably higher in ICU patients than in non-ICU patients (30, 31). Furthermore, increased concentrations of both types of microvesicles (EV) (CD41+EV and annexin V+ CD41+ EV) per platelet in both patients with non-severe and severe COVID-19 compared with healthy controls were observed, suggesting the contribution of platelets derived MVs to COVID-19 disease pathology (31). Patients with COVID-19 exhibit elevated levels of platelet-neutrophil aggregates and increased neutrophil extracellular traps (NETs) (32, 33). These factors can contribute to the development of immunothrombosis, leading to thromboembolic complications. Ultimately, this process results in platelet consumption and thrombocytopenia, both of which are correlated with heightened mortality (27, 34). In contrast, there are studies supporting antiviral phenotype of platelets in various viral infections, including COVID-19, regulated through IFITs/IFITM3 (35, 36).

Proposed platelet receptors interact with SARS-CoV-2

Platelets possess an array of receptors, including Cluster of differentiation(CD)40L, toll-like receptor, and the Fc receptor for IgG (FcyRIIa), enabling them to respond to stimuli (37). In other viral infections, such as Dengue haemorrhagic fever, antibody-coated virions induce substantial platelet activation through an Fc γ RIIa-dependent mechanism (38, 39). Similarly, Influenza H1N1 (40) and some Bunyaviruses like Crimea-Congo Haemorrhagic fever (41) induce platelet activation via Fc γ RIIa. However, the direct interaction mechanisms between SARS-CoV-2 and platelets or megakaryocytes remain contentious.

Shen et al., using RT-PCR assay, reported the detection of CD147, GRP78, KREMEN1, cathepsin L, NRP1, and ASGR1 in megakaryoblast cell line (MEG-01), and CD147, GRP78, KREMEN1, and ASGR1 in platelets. Notably, ACE2 was not detected in MEG-01 cell line or platelets (42, 43), suggesting SARS-CoV-2 may employ receptors other than ACE2 for interaction. Structural studies have proposed CD26 as another SARS-CoV-2 receptor, although its expression on platelets is debated (24, 43). Tang et al. demonstrated that the envelope (E) protein of SARS-CoV-2 enhances platelet activation and thrombosis through a CD36/p38 MAPK/NF-KB signaling axis (44). Additionally, platelets express a multitude of immune receptors, including CD40L, Toll-like receptors (TLRs), and the Fc receptor for IgG (FcyRIIA) (37). In line with this, a few other studies indicated that the E protein can physically interact with the TLR2 transmembrane receptor, stimulating NF-KB transcription and CXCL8 production (45, 46). Additionally, the viral Spike (S) protein independently binds with CD42b and stimulates platelets (47). Carnevale and colleagues provided evidence that SARS-CoV-2 uses the S protein to activate platelets via TLR4, leading to Nox2related oxidative stress and a prothrombotic phenotype (48).

Another clinical study demonstrated that signaling through $Fc\gamma$ RIIA and the C5a-C5aR pathway mediate platelet hyperactivation in COVID-19 (49). Puhm et al. demonstrated that TF from SARS-CoV-2-infected cells activates thrombin, signaling PARs on platelets, with potential implications for COVID-19 coagulation (50). Ito and colleagues proposed platelet α IIb β 3 integrin binds to the SARS-CoV-2 S protein of the alpha strain but not wild type and omicron strains (51).

Despite the distinct biological characteristics of SARS-CoV-2 Omicron, it leads to platelet activation and desensitization, similar to observations with the Delta variant. Omicron is also found in platelets from severe patients, inducing selective autophagy. However, the mechanisms of intraplatelet processing of Omicron cargo differ from Delta, suggesting that S protein mutations modify virus-platelet interactions (52). Although several receptors on platelet surface (Figure 1), besides ACE2, proposed for viral entry in platelets, due to the limited *in vivo* experimental evidence, further studies are required to substantiate these claims.

Controversies in COVID-19 and direct infection of platelets

Initial investigations proposed the direct invasion of platelets by SARS-CoV-2, impacting their functionality (53). Autopsies

reported the presence of SARS-CoV-2 viral particles in both megakaryocytes and platelets (54). Notably, Zhang et al. reported ACE2 expression on the platelet surface (55), but this observation was not consistently replicated in other studies (24, 31, 54, 56). The conflicting findings raise questions about the interaction between SARS-CoV-2 and platelets through ACE2. RNA-seq and western blot analyses revealed no evidence of ACE2 or TMPRSS2 in CD45depleted platelets from both COVID-19 patients and healthy individuals (24). Zaid et al. conducted a similar study and also reported the absence of ACE2 on platelets from COVID-19 patients and healthy volunteers (31). A retrospective survey of plasma samples from severe and non-severe COVID-19 patients revealed increased thrombosis and elevated levels of sP-selectin, sGPVI, RANTES, and PF4 during platelet activation (43). In contrast, Zhang et al. reported significant expression of ACE2 and TMPRSS2 mRNA and protein on platelets from healthy individuals and mice. Furthermore, in vitro and in vivo experiments using humanized ACE2 transgenic mice demonstrated that SARS-CoV-2 and its S protein directly activate platelets (55). Another study by Koupenova et al. revealed that SARS-CoV-2 induces programmed cell death in platelets, leading to internalization of virions attached to microparticles, bypassing ACE2 (57). This internalization results in rapid digestion, apoptosis, necroptosis, and extracellular vesicle release, contributing to dysregulated immunity and thrombosis. However, these conflicting observations raise questions about the direct



FIGURE 1

Proposed platelet receptors interaction with SARS-CoV-2. *In vitro* studies proposed that SARS-CoV-2 virus S and E proteins interact with platelet surface receptors, such as CD42b, TLR4, Fc receptor, CD36, TLR2, and α IIb β 3 integrin. Most soluble agonists released by activated platelets, such as adenosine diphosphate (ADP), thromboxane A2 (TxA2) as well as TF/FVIIa/Xa-mediated thrombin trigger platelet activation through GPCRs. For instance, elevated levels of thrombin in the circulatory system due to the activation of coagulation cascade, also activates the platelets through PAR1/PAR4 receptors. S, spike protein; E, envelope protein.

interaction between SARS-CoV-2 and platelets and the concept of direct platelet infectivity.

Conclusion

The current body of evidence underscores the ambiguity surrounding the direct interaction between SARS-CoV-2 and platelets. The disparate observations of viral load within platelets from COVID-19 patients, coupled with the identification of numerous alternative receptors on the platelet surface, have contributed to the uncertainty. In vitro and in vivo experimental studies as well as proteomic and transcriptomic analyses of platelets from COVID-19 patients and mice infected with SARS-CoV-2 further highlight the unlikelihood of direct platelet infectivity for the activation of platelets by SARS-CoV-2 viral proteins. Amidst this uncertainty, critical or fatal COVID-19 cases have exhibited various pathological abnormalities, including endothelial dysfunction, cytokine storms, and the formation of NETs. These abnormalities may be exacerbated by platelet hyperactivity, resulting from the aggregation of platelets to activated endothelial cells and neutrophils.

Author contributions

SS: Conceptualization, Data curation, Methodology, Resources, Supervision, Validation, Visualization, Writing – original draft, Writing – review & editing. NM: Visualization, Writing – review & editing. AJ: Visualization, Writing – review & editing.

References

1. Conway EM, Mackman N, Warren RQ, Wolberg AS, Mosnier LO, Campbell RA, et al. Understanding COVID-19-associated coagulopathy. *Nat Rev Immunol.* (2022) 22:639–49. doi: 10.1038/s41577-022-00762-9

2. Subramaniam S, Kothari H, Bosmann M. Tissue factor in COVID-19-associated coagulopathy. *Thromb Res.* (2022) 220:35–47. doi: 10.1016/j.thromres.2022.09.025

3. Subramaniam S, Reinhardt C, Kulkarni PP, Spiezia L. Editorial: COVID-19 and thrombo-inflammatory responses. *Front Cardiovasc Med.* (2023) 10:1231909. doi: 10.3389/fcvm.2023.1231909

4. Sciaudone A, Corkrey H, Humphries F, Koupenova M. Platelets and SARS-coV-2 during COVID-19: immunity, thrombosis, and beyond. *Circ Res.* (2023) 132:1272–89. doi: 10.1161/CIRCRESAHA.122.321930

5. Hussain M, Khurram Syed S, Fatima M, Shaukat S, Saadullah M, Alqahtani AM, et al. Acute respiratory distress syndrome and COVID-19: A literature review. *J Inflammation Res.* (2021) 14:7225–42. doi: 10.2147/JIR.S334043

6. Lin HY. The severe COVID-19: A sepsis induced by viral infection? And its immunomodulatory therapy. *Chin J Traumatol.* (2020) 23:190-5. doi: 10.1016/j.cjtee.2020.06.002

7. Schultheiss C, Willscher E, Paschold L, Gottschick C, Klee B, Henkes SS, et al. The IL-1beta, IL-6, and TNF cytokine triad is associated with post-acute sequelae of COVID-19. *Cell Rep Med.* (2022) 3:100663. doi: 10.1016/j.xcrm.2022.100663

8. Faraj SS, Jalal PJ. IL1beta, IL-6, and TNF-alpha cytokines cooperate to modulate a complicated medical condition among COVID-19 patients: case-control study. *Ann Med Surg (Lond).* (2023) 85:2291–7. doi: 10.1097/MS9.0000000000000679

9. Del Valle DM, Kim-Schulze S, Huang HH, Beckmann ND, Nirenberg S, Wang B, et al. An inflammatory cytokine signature predicts COVID-19 severity and survival. *Nat Med.* (2020) 26:1636–43. doi: 10.1038/s41591-020-1051-9

10. Stark K, Massberg S. Interplay between inflammation and thrombosis in cardiovascular pathology. Nat Rev Cardiol. (2021) 18:666–82. doi: 10.1038/s41569-021-00552-1

Funding

The author(s) declare that no financial support was received for the research, authorship, and/or publication of this article.

Acknowledgments

The schematic illustration was adopted and modified from "platelet receptors" by BioRender.com (2024). Retrieved from https://app.biorender.com/biorender-templates. I would like to acknowledge Mitchell Lobo for her support to refining the figure for this manuscript.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

11. Ackermann M, Verleden SE, Kuehnel M, Haverich A, Welte T, Laenger F, et al. Pulmonary vascular endothelialitis, thrombosis, and angiogenesis in covid-19. *N Engl J Med.* (2020) 383:120–8. doi: 10.1056/NEJMoa2015432

12. Szegedi I, Orban-Kalmandi R, Csiba L, Bagoly Z. Stroke as a potential complication of COVID-19-associated coagulopathy: A narrative and systematic review of the literature. *J Clin Med.* (2020) 9(10):3137. doi: 10.3390/jcm9103137

13. Anchidin OI, Rosianu SH, Nemes A, Aldica M, Blendea D, Molnar A, et al. The Effectiveness of Antiplatelet Therapy and the Factors Influencing It in Patients with Acute Coronary Syndrome before and during the COVID-19 Pandemic. *Medicina (Kaunas).* (2022) 59. doi: 10.3390/medicina59010084

14. Chen B, Tian EK, He B, Tian L, Han R, Wang S, et al. Overview of lethal human coronaviruses. *Signal Transduct Target Ther.* (2020) 5:89. doi: 10.1038/s41392-020-0190-2

15. Yuki K, Fujiogi M, Koutsogiannaki S. COVID-19 pathophysiology: A review. *Clin Immunol.* (2020) 215:108427. doi: 10.1016/j.clim.2020.108427

16. Palta S, Saroa R, Palta A. Overview of the coagulation system. *Indian J Anaesth.* (2014) 58:515–23. doi: 10.4103/0019-5049.144643

17. Lasne D, Jude B, Susen S. From normal to pathological hemostasis. Can J Anaesth. (2006) 53:S2-11. doi: 10.1007/BF03022247

18. Sang Y, Roest M, De Laat B, De Groot PG, Huskens D. Interplay between platelets and coagulation. *Blood Rev.* (2021) 46:100733. doi: 10.1016/j.blre.2020.100733

19. Bryckaert M, Rosa JP, Denis CV, Lenting PJ. Of von Willebrand factor and platelets. *Cell Mol Life Sci.* (2015) 72:307–26. doi: 10.1007/s00018-014-1743-8

20. Offermanns S. Activation of platelet function through G protein-coupled receptors. *Circ Res.* (2006) 99:1293–304. doi: 10.1161/01.RES.0000251742.71301.16

21. Stalker TJ, Newman DK, Ma P, Wannemacher KM, Brass LF. Platelet signaling. Handb Exp Pharmacol. (2012) 59-85. doi: 10.1007/978-3-642-29423-5_3

22. Hottz ED, Azevedo-Quintanilha IG, Palhinha L, Teixeira L, Barreto EA, Pao CRR, et al. Platelet activation and platelet-monocyte aggregate formation trigger tissue

factor expression in patients with severe COVID-19. *Blood.* (2020) 136:1330-41. doi: 10.1182/blood.2020007252

23. Barrett TJ, Lee AH, Xia Y, Lin LH, Black M, Cotzia P, et al. Platelet and vascular biomarkers associate with thrombosis and death in coronavirus disease. *Circ Res.* (2020) 127:945–7. doi: 10.1161/CIRCRESAHA.120.317803

24. Manne BK, Denorme F, Middleton EA, Portier I, Rowley JW, Stubben C, et al. Platelet gene expression and function in patients with COVID-19. *Blood.* (2020) 136:1317–29. doi: 10.1182/blood.2020007214

25. Rapkiewicz AV, Mai X, Carsons SE, Pittaluga S, Kleiner DE, Berger JS, et al. Megakaryocytes and platelet-fibrin thrombi characterize multi-organ thrombosis at autopsy in COVID-19: A case series. *EClinicalMedicine*. (2020) 24:100434. doi: 10.1016/j.eclinm.2020.100434

26. Ji W, Chen L, Yang W, Li K, Zhao J, Yan C, et al. Transcriptional landscape of circulating platelets from patients with COVID-19 reveals key subnetworks and regulators underlying SARS-CoV-2 infection: implications for immunothrombosis. *Cell Biosci.* (2022) 12:15. doi: 10.1186/s13578-022-00750-5

27. Yang X, Yang Q, Wang Y, Wu Y, Xu J, Yu Y, et al. Thrombocytopenia and its association with mortality in patients with COVID-19. *J Thromb Haemost.* (2020) 18:1469–72. doi: 10.1111/jth.14848

28. Jiang SQ, Huang QF, Xie WM, Lv C, Quan XQ. The association between severe COVID-19 and low platelet count: evidence from 31 observational studies involving 7613 participants. *Br J Haematol.* (2020) 190:e29–33. doi: 10.1111/bjh.16817

29. Wool GD, Miller JL. The impact of COVID-19 disease on platelets and coagulation. *Pathobiology*. (2021) 88:15–27. doi: 10.1159/000512007

30. Goshua G, Pine AB, Meizlish ML, Chang CH, Zhang H, Bahel P, et al. Endotheliopathy in COVID-19-associated coagulopathy: evidence from a singlecentre, cross-sectional study. *Lancet Haematol.* (2020) 7:e575-82. doi: 10.1016/ S2352-3026(20)30216-7

31. Zaid Y, Puhm F, Allaeys I, Naya A, Oudghiri M, Khalki L, et al. Platelets can associate with SARS-cov-2 RNA and are hyperactivated in COVID-19. *Circ Res.* (2020) 127:1404–18. doi: 10.1161/CIRCRESAHA.120.317703

32. Le Joncour A, Biard L, Vautier M, Bugaut H, Mekinian A, Maalouf G, et al. Neutrophil-platelet and monocyte-platelet aggregates in COVID-19 patients. *Thromb Haemost.* (2020) 120:1733–5. doi: 10.1055/s-0040-1718732

33. Middleton EA, He XY, Denorme F, Campbell RA, Ng D, Salvatore SP, et al. Neutrophil extracellular traps contribute to immunothrombosis in COVID-19 acute respiratory distress syndrome. *Blood*. (2020) 136:1169–79. doi: 10.1182/blood.2020007008

34. Khave LJ, Zafari P, Pirsalehi A, Salari S, Baghestani A, Akbari ME, et al. Association between thrombocytopenia and platelet profile with morbidity/mortality of severe and non-severe COVID-19 patients. *Rev Assoc Med Bras (1992).* (2021) 67:1670–5. doi: 10.1590/1806-9282.20210720

35. Campbell RA, Schwertz H, Hottz ED, Rowley JW, Manne BK, Washington AV, et al. Human megakaryocytes possess intrinsic antiviral immunity through regulated induction of IFITM3. *Blood.* (2019) 133:2013–26. doi: 10.1182/blood-2018-09-873984

36. Subramaniam S, Hekman R, Jayaraman A, Kenney D, O'connell AK, Montanaro P, et al. OC 22.1 proteome and phosphoproteome analyses of SARS-coV-2 infected humanized ACE2 mice platelets revealed reprogramming of platelets from early hyperreactive to late antiviral phenotypes. *Res Pract Thromb Haemostasis.* (2023) 7. doi: 10.1016/j.rpth.2023.100353

37. Semple JW, Italiano JE Jr., Freedman J. Platelets and the immune continuum. *Nat Rev Immunol.* (2011) 11:264–74. doi: 10.1038/nri2956

38. Simmons CP, Farrar JJ, Nguyen V V, Wills B. Dengue. N Engl J Med. (2012) 366:1423-32. doi: 10.1056/NEJMra1110265

39. Screaton G, Mongkolsapaya J, Yacoub S, Roberts C. New insights into the immunopathology and control of dengue virus infection. *Nat Rev Immunol.* (2015) 15:745–59. doi: 10.1038/nri3916

40. Boilard E, Pare G, Rousseau M, Cloutier N, Dubuc I, Levesque T, et al. Influenza virus H1N1 activates platelets through FcgammaRIIA signaling and thrombin generation. *Blood.* (2014) 123:2854–63. doi: 10.1182/blood-2013-07-515536

41. Erduran E, Bahadir A, Palanci N, Gedik Y. The treatment of crimean-congo hemorrhagic fever with high-dose methylprednisolone, intravenous immunoglobulin, and fresh frozen plasma. *J Pediatr Hematol Oncol.* (2013) 35:e19–24. doi: 10.1097/MPH.0b013e3182706444

42. Rowley JW, Oler AJ, Tolley ND, Hunter BN, Low EN, Nix DA, et al. Genomewide RNA-seq analysis of human and mouse platelet transcriptomes. *Blood*. (2011) 118: e101–111. doi: 10.1182/blood-2011-03-339705

43. Shen S, Zhang J, Fang Y, Lu S, Wu J, Zheng X, et al. SARS-CoV-2 interacts with platelets and megakaryocytes via ACE2-independent mechanism. *J Hematol Oncol.* (2021) 14:72. doi: 10.1186/s13045-021-01082-6

44. Tang Z, Xu Y, Tan Y, Shi H, Jin P, Li Y, et al. CD36 mediates SARS-CoV-2envelope-protein-induced platelet activation and thrombosis. *Nat Commun.* (2023) 14:5077. doi: 10.1038/s41467-023-40824-7

45. Zheng M, Karki R, Williams EP, Yang D, Fitzpatrick E, Vogel P, et al. TLR2 senses the SARS-CoV-2 envelope protein to produce inflammatory cytokines. *Nat Immunol.* (2021) 22:829–38. doi: 10.1038/s41590-021-00937-x

46. Planes R, Bert JB, Tairi S, Benmohamed L, Bahraoui E. SARS-coV-2 envelope (E) protein binds and activates TLR2 pathway: A novel molecular target for COVID-19 interventions. *Viruses*. (2022) 14. doi: 10.3390/v14050999

47. Li T, Yang Y, Li Y, Wang Z, Ma F, Luo R, et al. Platelets mediate inflammatory monocyte activation by SARS-CoV-2 spike protein. *J Clin Invest.* (2022) 132. doi: 10.1172/JCI150101

48. Carnevale R, Cammisotto V, Bartimoccia S, Nocella C, Castellani V, Bufano M, et al. Toll-like receptor 4-dependent platelet-related thrombosis in SARS-coV-2 infection. *Circ Res.* (2023) 132:290–305. doi: 10.1161/CIRCRESAHA.122.321541

49. Apostolidis SA, Sarkar A, Giannini HM, Goel RR, Mathew D, Suzuki A, et al. Signaling through fcgammaRIIA and the C5a-C5aR pathway mediate platelet hyperactivation in COVID-19. *Front Immunol.* (2022) 13:834988. doi: 10.3389/fimmu.2022.834988

50. Puhm F, Allaeys I, Lacasse E, Dubuc I, Galipeau Y, Zaid Y, et al. Platelet activation by SARS-CoV-2 implicates the release of active tissue factor by infected cells. *Blood Adv.* (2022) 6:3593–605. doi: 10.1182/bloodadvances.2022007444

51. Ito K, Goto K, Shirakawa R, Horiuchi H, Ogasawara K. Platelet alphaIIbbeta3 integrin binds to SARS-CoV-2 spike protein of alpha strain but not wild type and omicron strains. *Biochem Biophys Res Commun.* (2023) 657:80–5. doi: 10.1016/j.bbrc.2023.03.057

52. Garcia C, Compagnon B, Ribes A, Voisin S, Vardon-Bounes F, Payrastre B. SARS-CoV-2 Omicron variant infection affects blood platelets, a comparative analysis with Delta variant. *Front Immunol.* (2023) 14:1231576. doi: 10.3389/fimmu.2023.1231576

53. Bury L, Camilloni B, Castronari R, Piselli E, Malvestiti M, Borghi M, et al. Search for SARS-CoV-2 RNA in platelets from COVID-19 patients. *Platelets*. (2021) 32:284–7. doi: 10.1080/09537104.2020.1859104

54. Barrett TJ, Bilaloglu S, Cornwell M, Burgess HM, Virginio VW, Drenkova K, et al. Platelets contribute to disease severity in COVID-19. *J Thromb Haemost.* (2021) 19:3139–53. doi: 10.1111/jth.15534

55. Zhang S, Liu Y, Wang X, Yang L, Li H, Wang Y, et al. SARS-CoV-2 binds platelet ACE2 to enhance thrombosis in COVID-19. *J Hematol Oncol.* (2020) 13:120. doi: 10.1186/s13045-020-00954-7

56. Subramaniam S, Hekman RM, Jayaraman A, O'connell AK, Montanaro P, Blum B, et al. Platelet proteome analysis reveals an early hyperactive phenotype in SARS-CoV-2-infected humanized ACE2 mice. *bioRxiv*. (2021). doi: 10.1101/2021.08.19.457020

57. Koupenova M, Corkrey HA, Vitseva O, Tanriverdi K, Somasundaran M, Liu P, et al. SARS-coV-2 initiates programmed cell death in platelets. *Circ Res.* (2021) 129:631–46. doi: 10.1161/CIRCRESAHA.121.319117