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## Growth hormone – releasing hormone in the context of inflammation and redox biology

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## Introduction

Inflammation and oxidative stress contribute in cancer development, severity and aggression (1–3). Many malignancies arise from tissues affected by chronic inflammation. Tumor microenvironment consists of both cancer and immune cells which secrete growth factors, cytokines and chemokines, leading to cancer spread (1). Anti-inflammatory and immunomodulatory therapeutic approaches are commonly used in oncology (3).

Reactive oxygen species (ROS) are highly active molecules, arising from physiological and pathological processes, and our body balances their excess utilizing anti-oxidative defense mechanisms. Cancers suppress antioxidative mechanisms via enzyme modulation/ mutation. Under physiological conditions ROS act as signaling molecules in cell growth, migration and differentiation. Chronic inflammation may lead to the excessive generation of ROS and reactive nitrogen species (RNS), altering immune responses, which in turn lead to oncogenic transformations (4).

The innate immune system depends on ROS, since macrophages and natural killer (NK) cells utilize those highly active molecules to maintain human tissue integrity and combat pathogens. ROS generation by mitochondria is due to activation of several proinflammatory pathways (e.g. MAPK, AMPK, PI3K/ACT) in coordination with NF- $\kappa$ B and HIF1 $\alpha$  (2).

# Growth hormone releasing hormone and its receptors

Growth hormone - releasing hormone (GHRH) is secreted by the hypothalamus and binds to the GHRH receptor (GHRH-R) of the pituitary cells to trigger the release of GH from the somatotrophs. GHRH is a 44-amino acid peptide, however its full intrinsic biological activity is retained by the NH<sub>2</sub>-terminal 29-amino acid sequence. The pituitary type GHRH receptor (pGHRH-R) is a class II G-protein-coupled receptor with seven transmembrane domains, homologous to the receptors for VIP, PACAP and calcitonin. Activation of pGHRH-R results in increased cAMP production, which acts as the second messenger in the GHRH related signal transduction. Splice variants (SVs) of the GHRH-R have been identified in various cancers. SV1 receptor possesses ligand independent activities (5, 6) and activates the mitogen-activated protein kinase (MAPK) pathway (5, 7).

#### GHRH in inflammation and tumors

The expression of GHRH has been demonstrated in prostatic, endometrial, ovarian, breast, gastroenteropatic, and lung carcinomas, glioblastomas, malignant bone tumors, human adrenal carcinomas and colorectal cancers (8). GHRH may act as an autocrine and/or paracrine growth factor in cancers (8). Knocking down of GHRH gene expression suppressed the proliferation of T47D, MDA-MB-435S, MDA-MB-468 breast cancers, LNCaP prostate cancer and NCI H838 non-SCLC (6). Moreover, GHRH can increase IL-17 secretion (9), a cytokine involved in the pathogenesis of non-alcoholic and alcoholic steatohepatitis (10). It has been reported that both conditions are associated with increased risk of hepatocellular carcinoma (HCC) development. In mice, targeting IL-17 suppressed the development of NASH-associated HCC (10). In another study, IL-17 was able to blunt the anticancer efficacy of chemotherapeutic agents in vivo (11). GHRH was able to promote TH17 cell differentiation and autoimmune inflammation (12), and MIA-690 - a GHRH antagonist - inhibited LPS-induced inflammatory and pro-oxidative markers (13).

Several splice variants of the GHRH receptor (SVs) were identified and sequenced, including SV1 (14). The major part of its cDNA sequence is identical to the corresponding sequence of pGHRH-R, with the exception of the first 334 SV1 nucleotides. The protein sequence of this transduced receptor differs from the full length receptor in the amino-terminal extracellular domain, in which a 25 amino-acid sequence replaces the first 89 amino acids of pGHRH-R (14). SV1 has been associated with strong ligand independent activities (15). Moreover, it is expressed in many cancers, including prostatic, breast, colorectal, gastric, melanomas, bone sarcomas, glioblastomas and SW13 human adrenal carcinoma cells (16). The pGHRH-R is present in human cancer tissues isolated from breast, ovarian, lung cancers, glioblastomas and lymphoma cells (16).

#### GHRH antagonists in cancers

Antagonists of growth hormone-releasing hormone inhibit the growth of various experimental cancers including prostate, breast, ovarian, colorectal, lung, renal, endometrial cancers; glioblastomas and lymphomas (16, 17). The inhibitory effect of GHRH antagonists is partially dependent on the suppression of GH secretion from the pituitary, which results in decreased IGF-I production. GHRH antagonists can also suppress tumor growth in a direct manner through blockade of autocrine GHRH action (8, 16). HeLa cells, which do not express GHRH receptors, responded to GHRH and GHRH antagonists after being transfected with the pGHRH-R or SV1 receptor (18).

## GHRH and ROS

Reactive oxygen species (ROS) and reactive nitrogen species (RNS) act as signaling molecules. They promote human tumors by contributing to oxidative stress, a common condition in cancer (2). In LNCaP prostate cancer cells GHRH antagonists exerted antioxidative properties, and in A549 lung cancer cells JV-1–36 suppressed hydrogen-peroxide induced ROS (19, 20). In bovine pulmonary artery endothelial cells, human cerebral microvascular endothelial cells, and human lung microvascular endothelial cells those peptides reduced ROS generation. 3T3 cells which do not express GHRH receptors were not affected by GHRH analog treatment (21).

It was also recently revealed that GHRH antagonists suppress IFN- $\gamma$  (22), hydrogen peroxide (23, 24) and hydrochloric acid (25) - induced inflammation. P53 is a tumor suppressor exerting antioxidative activities, which is induced by GHRH antagonists (17, 26– 29). Those data further our knowledge on the mechanisms mediating the protective effects of those peptides against human disease (30). P53 and unfolded protein response are interrelated in the intracellular niche, since UPR activation induces P53 (31). It appears that UPR – which exerts anti-inflammatory and antioxidative activities (32–37) - is involved in the effects of GHRH antagonists in endothelial cells (32). These peptides were able to induce the three UPR sensors and its downstream target, namely BiP, in normal lung cells. There is very limited information on these effects of GHRH-related analogs in cancer cells (38).

It has been demonstrated that the SV1 receptor and pGHRH-R activate mitogen activated protein kinases ERK1/2 (7), which are strongly related to the generation and metabolism of ROS. GHRH can also activate: i) JAK2/STAT3, which contributes to oxidative phenomena (39), and ii) inducible nitric oxide synthase (iNOS) in A549 lung cancer cells. GHRH antagonist treatment counteracts those events (40). This is important because iNOS is one of the three NOS isoforms. It catalyzes the oxidative deamination of L-arginine to produce cytruline and nitric oxide (NO) and it is essential for immunity and vascular function. Moreover, it has been involved in the pathogenesis of various diseases through ROS/RNS induction. Indeed, ERK1/2 activation leads to increased iNOS and NO production (41, 42).

#### Conclusions

The aforementioned studies report that GHRH induces ROS/ RNS generation. GHRH antagonists can counteract those effects eliciting anti-inflammatory responses, which contribute to their anti-cancer activities. The exact mechanisms involved in those events are not completely understood, and are currently under investigation.

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AS: Conceptualization, Writing – original draft, Writing – review & editing. NB: Writing – review & editing.

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## Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

### References

1. Coussens LM, Werb Z. Inflammation and cancer. Nature. (2002) 420:860-7. doi: 10.1038/nature01322

2. Morris G, Gevezova M, Sarafian V, Maes M. Redox regulation of the immune response. *Cell Mol Immunol.* (2022) 19:1079–101. doi: 10.1038/s41423-022-00902-0

3. Hou J, Karin M, Sun B. Targeting cancer-promoting inflammation - have antiinflammatory therapies come of age? *Nat Rev Clin Oncol.* (2021) 18:261–79. doi: 10.1038/s41571-020-00459-9

4. Yu W, Tu Y, Long Z, Liu J, Kong D, Peng J, et al. Reactive oxygen species bridge the gap between chronic inflammation and tumor development. *Oxid Med Cell Longev*. (2022) 2022:2606928. doi: 10.1155/2022/2606928

5. Barabutis N, Tsellou E, Schally AV, Kouloheri S, Kalofoutis A, Kiaris H. Stimulation of proliferation of MCF-7 breast cancer cells by a transfected splice variant of growth hormone-releasing hormone receptor. *Proc Natl Acad Sci U.S.A.* (2007) 104:5575–9. doi: 10.1073/pnas.0700407104

6. Barabutis N, Schally AV. Knocking down gene expression for growth hormonereleasing hormone inhibits proliferation of human cancer cell lines. *Br J Cancer*. (2008) 98:1790–6. doi: 10.1038/sj.bjc.6604386

7. Barabutis N, Siejka A, Schally AV, Block NL, Cai R, Varga JL. Activation of mitogen-activated protein kinases by a splice variant of GHRH receptor. *J Mol Endocrinol.* (2010) 44:127–34. doi: 10.1677/JME-09-0121

 Barabutis N, Schally AV. Growth hormone-releasing hormone: extrapituitary effects in physiology and pathology. *Cell Cycle*. (2010) 9:4110–6. doi: 10.4161/cc.9.20.13787

9. Stepien T, Lawnicka H, Komorowski J, Stepien H, Siejka A. Growth hormonereleasing hormone stimulates the secretion of interleukin 17 from human peripheral blood mononuclear cells in *vitro. Neuro Endocrinol Lett.* (2010) 31(6):852-856.

10. Ma HY, Yamamoto G, Xu J, Liu X, Karin D, Kim JY, et al. IL-17 signaling in steatotic hepatocytes and macrophages promotes hepatocellular carcinoma in alcohol-related liver disease. *J Hepatol.* (2020) 72:946–59. doi: 10.1016/j.jhep.2019.12.016

11. Bruchard M, Mignot G, Derangere V, Chalmin F, Chevriaux A, Vegran F, et al. Chemotherapy-triggered cathepsin B release in myeloid-derived suppressor cells activates the NIrp3 inflammasome and promotes tumor growth. *Nat Med.* (2013) 19:57–64. doi: 10.1038/nm.2999

12. Du L, Ho BM, Zhou L, Yip YWY, He JN, Wei Y, et al. Growth hormone releasing hormone signaling promotes Th17 cell differentiation and autoimmune inflammation. *Nat Commun.* (2023) 14:3298. doi: 10.1038/s41467-023-39023-1

13. Recinella L, Chiavaroli A, Orlando G, Ferrante C, Marconi GD, Gesmundo I, et al. Antinflammatory, antioxidant, and behavioral effects induced by administration of growth hormone-releasing hormone analogs in mice. *Sci Rep.* (2020) 10:732. doi: 10.1038/s41598-019-57292-z

 Rekasi Z, Czompoly T, Schally AV, Halmos G. Isolation and sequencing of cDNAs for splice variants of growth hormone-releasing hormone receptors from human cancers. *Proc Natl Acad Sci U.S.A.* (2000) 97:10561–6. doi: 10.1073/ pnas.180313297

15. Kiaris H, Chatzistamou I, Schally AV, Halmos G, Varga JL, Koutselini H, et al. Ligand-dependent and -independent effects of splice variant 1 of growth hormone-releasing hormone receptor. *Proc Natl Acad Sci U.S.A.* (2003) 100:9512–7. doi: 10.1073/pnas.1533185100

16. Schally AV, Varga JL, Engel JB. Antagonists of growth-hormone-releasing hormone: an emerging new therapy for cancer. *Nat Clin Pract Endocrinol Metab.* (2008) 4:33–43. doi: 10.1038/ncpendmet0677

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17. Barabutis N, Schally AV, Siejka A. P53. GHRH Inflammation cancer EBioMedicine. (2018) 37:557-62. doi: 10.1016/j.ebiom.2018.10.034

 Barabutis N, Schally AV. Antioxidant activity of growth hormone-releasing hormone antagonists in LNCaP human prostate cancer line. *Proc Natl Acad Sci U.S.A.* (2008) 105:20470–5. doi: 10.1073/pnas.0811209106

19. Kubra KT, Akhter MS, Apperley K, Barabutis N. Growth hormone-releasing hormone antagonist JV-1–36 suppresses reactive oxygen species generation in A549 lung cancer cells. *Endocrines*. (2022) 3:813–20. doi: 10.3390/endocrines3040067

20. Barabutis N, Akhter MS, Kubra KT, Jackson K. Growth hormone-releasing hormone in endothelial inflammation. *Endocrinology* (2022) 164(2):1–11. doi: 10.1210/endocr/bqae064

21. Akhter MS, Barabutis N. Suppression of reactive oxygen species in endothelial cells by an antagonist of growth hormone-releasing hormone. *J Biochem Mol Toxicol.* (2021) 35:e22879. doi: 10.1002/jbt.22879

22. Fakir S, Barabutis N. Growth hormone-releasing hormone antagonists counteract interferon-gamma - induced barrier dysfunction in bovine and human endothelial cells. *Cytokine*. (2024) 173:156416. doi: 10.1016/j.cyto.2023.156416

23. Barabutis N, Siejka A, Akhter MS. Growth hormone-releasing hormone antagonists counteract hydrogen peroxide - induced paracellular hyperpermeability in endothelial cells. *Growth Horm IGF Res.* (2023) 69–70:101534. doi: 10.1016/j.ghir.2023.101534

24. Akhter MS, Kubra KT, Barabutis N. Protective effects of GHRH antagonists against hydrogen peroxide-induced lung endothelial barrier disruption. *Endocrine*. (2023) 79:587–92. doi: 10.1007/s12020-022-03226-1

25. Barabutis N, Kubra KT, Akhter MS. Growth hormone-releasing hormone antagonists protect against hydrochloric acid-induced endothelial injury in *vitro*. *Environ Toxicol Pharmacol.* (2023) 99:104113. doi: 10.1016/j.etap.2023.104113

26. Akhter MS, Uddin MA, Barabutis N. P53 regulates the redox status of lung endothelial cells. *Inflammation*. (2020) 43:686–91. doi: 10.1007/s10753-019-01150-7

27. Akhter MS, Uddin MA, Kubra KT, Barabutis N. P53-induced reduction of lipid peroxidation supports brain microvascular endothelium integrity. *J Pharmacol Sci.* (2019) 141:83–5. doi: 10.1016/j.jphs.2019.09.008

28. Kubra KT, Uddin MA, Akhter MS, Barabutis N. P53 is subjected to lipoteichoic acid-induced phosphorylation in the lungs. *TH Open*. (2020) 4:e173–4. doi: 10.1055/s-0040-1714695

29. Uddin MA, Akhter MS, Siejka A, Catravas JD, Barabutis N. P53 supports endothelial barrier function via APE1/Ref1 suppression. *Immunobiology*. (2019) 224:532-8. doi: 10.1016/j.imbio.2019.04.008

30. Barabutis N, Fakir S. Growth hormone-releasing hormone beyond cancer. *Clin Exp Pharmacol Physiol.* (2024) 51:40–1. doi: 10.1111/1440-1681.13829

31. Akhter MS, Uddin MA, Barabutis N. Unfolded protein response regulates P53 expression in the pulmonary endothelium. *J Biochem Mol Toxicol.* (2019) 33:e22380. doi: 10.1002/jbt.22380

32. Akhter MS, Uddin MA, Schally AV, Kubra KT, Barabutis N. Involvement of the unfolded protein response in the protective effects of growth hormone releasing hormone antagonists in the lungs. *J Cell Commun Signal.* (2021) 15:125–9. doi: 10.1007/s12079-020-00593-0

33. Barabutis N. Activating transcription factor 6 in the endothelial context. *Pulm Pharmacol Ther.* (2023) 80:102216. doi: 10.1016/j.pupt.2023.102216

34. Kubra KT, Akhter MS, Saini Y, Kousoulas KG, Barabutis N. Activating transcription factor 6 protects against endothelial barrier dysfunction. *Cell Signal.* (2022) 99:110432. doi: 10.1016/j.cellsig.2022.110432

35. Kubra KT, Barabutis N, Brefeldin A. kifunensine modulate LPS-induced lung endothelial hyperpermeability in human and bovine cells. *Am J Physiol Cell Physiol.* (2021) 321:C214–20. doi: 10.1152/ajpcell.00142.2021

36. Kubra KT, Barabutis N. Ceapin-A7 potentiates lipopolysaccharide-induced endothelial injury. J Biochem Mol Toxicol. (2023) 37:e23460. doi: 10.1002/jbt.23460

37. Kubra KT, Uddin MA, Akhter MS, Barabutis N. Luminespib counteracts the Kifunensine-induced lung endothelial barrier dysfunction. *Curr Res Toxicol.* (2020) 1:111–5. doi: 10.1016/j.crtox.2020.09.003

38. Barabutis N. Growth hormone releasing hormone in the unfolded protein response context. *Endocrine*. (2020) 67:291–3. doi: 10.1007/s12020-020-02205-8

39. Siejka A, Schally AV, Barabutis N. Activation of Janus kinase/signal transducer and activator of transcription 3 pathway by growth hormone-releasing hormone. *Cell Mol Life Sci.* (2010) 67:959–64. doi: 10.1007/s00018-009-0224-y

40. Barabutis N, Siejka A, Schally AV. Growth hormone releasing hormone induces the expression of nitric oxide synthase. *J Cell Mol Med.* (2011) 15:1148–55. doi: 10.1111/jcmm.2011.15.issue-5

41. Ratajczak-Wrona W, Jablonska E, Garley M, Jablonski J, Radziwon P, Iwaniuk A. The role of MAP kinases in the induction of iNOS expression in neutrophils exposed to NDMA: the involvement transcription factors. *Adv Med Sci.* (2013) 58:265–73. doi: 10.2478/v10039-012-0074-y

42. Lowry JL, Brovkovych V, Zhang Y, Skidgel RA. Endothelial nitric-oxide synthase activation generates an inducible nitric-oxide synthase-like output of nitric oxide in inflamed endothelium. *J Biol Chem.* (2013) 288:4174–93. doi: 10.1074/jbc.M112.436022