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Lymphatic system regulation of anti-cancer immunity and metastasis

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Cancer dissemination to lymph nodes (LN) is associated with a worse prognosis, increased incidence of distant metastases and reduced response to therapy. The LN microenvironment puts selective pressure on cancer cells, creating cells that can survive in LN as well as providing survival advantages for distant metastatic spread. Additionally, the presence of cancer cells leads to an immunosuppressive LN microenvironment, favoring the evasion of anti-cancer immune surveillance. However, recent studies have also characterized previously unrecognized roles for tumor-draining lymph nodes (TDLNs) in cancer immunotherapy response, including acting as a reservoir for pre-exhausted CD8+ T cells and stem-like CD8+ T cells. In this review, we will discuss the spread of cancer cells through the lymphatic system, the roles of TDLNs in metastasis and anti-cancer immune responses, and the therapeutic opportunities and challenges in targeting LN metastasis.

KEYWORDS

lymphatic vessels, lymph node metastasis, tumor-draining lymph node (TDLN), immunotherapy, cancer metastasis

Introduction

Cancer cell invasion of lymphatic vessels (1, 2) and the presence of metastatic cancer cells in sentinel LNs indicate that cancer has progressed to an advanced stage, which can guide treatment options and impact the overall prognosis (3–7). A higher number of affected LNs is associated with an increased risk of cancer recurrence and a worsened survival rate in many cancer types, including melanoma (8, 9), breast cancer (10, 11), head and neck cancer (12), prostate cancer (13, 14), and colon cancer (15, 16). Due to the significant importance of LN status in cancer staging, sentinel LN biopsy (SLNB) is generally recommended as standard staging practice in various solid tumors (4, 17–21). Further, in patients with clinically evident disease in lymph nodes, complete LN dissection

(CLND) is often recommended to remove potentially involved LNs. These clinical decisions assume that surgical removal of metastatic LNs could inhibit the further spreading of cancer cells. However, findings from several recent clinical trials have challenged this hypothesis (22-28). These studies indicate that CLND does not significantly improve the overall survival rates in melanoma and breast cancer patients with early-stage disease and positive SLNB. The lymphadenectomy procedures can lead to severe post-operative complications, including infections and lymphedema (29). Thus, the decision to surgically remove LNs warrants careful consideration. Clinical decision-making has become further complicated by the emerging significance of TDLNs for tumor antigen presentation (30), T cell priming (31-33), immunotherapy response (34-36), cancer vaccines (37-39) and CAR-T therapy (40-42), underscoring the need to consider the implications of LN removal on immunotherapy.

Lymphatic transport of antigen-presenting cells (APCs) and soluble tumor antigens to TDLNs is critical for the initiation of an anti-cancer immune response (43). The TDLNs are the first site where dendritic cells present tumor-antigens to CD8+ T cells to prime and activate them (44-46). The activated CD8+ T cells are subsequently recruited to the primary tumor sites. However, most intratumoral CD8+ T cells become exhausted rapidly due to the immunosuppressive tumor microenvironment (47). The circulation of TCF-1+ stem-like CD8+ T cells between the primary tumor and TDLNs is critical for systemic and intratumoral immune responses (47). Notably, the presence of stem-like CD8+ T cells in the TDLNs has been associated with durable treatment responses (31). Moreover, anti-PD-1 monoclonal antibodies not only act at the primary tumor site but also display additional activity in the periphery (48-50), particularly in the TDLNs (30, 51, 52), promoting the magnitude and quality of tumor antigen-specific CD8+ T cell responses in TDLNs (33).

To evade the important role of TDLNs in anti-cancer immunity, the progression and metastasis of cancer cells to LNs cause an immunosuppressive LN microenvironment (53–59). The elevation of Type I and Type II interferon-signaling pathways in cancer cells promotes their survival by evading NK cell killing (57) and suppressing effector T cell function by enhancing the accumulation of T regulatory (Treg) cells in the LNs (59). Moreover, the LN microenvironment accelerates epigenetic (60) and metabolic reprogramming of cancer cells (61, 62) that promote resistance to ferroptosis, enhancing survival and metastasis through the blood (62). Understanding how cancer cells escape immune surveillance in the LN, and how interactions between cancer cells and the LN microenvironment lead to immune evasion, is crucial to making clinical decisions aimed at preventing metastasis and enhancing anti-cancer immune responses.

It is likely in early-stage cancers, TDLNs play a crucial role in the generation of anti-cancer immune responses and the prevention of cancer spreading. The presence of TDLNs can create a favorable solution for neo-adjuvant and adjuvant therapy following surgical intervention. However, in late-stage cancers, the infiltration of cancer cells results in an immunosuppressive microenvironment within LNs, which will favor immune evasion and secondary metastases. Of note, patients with advanced cancers are likely to carry distal metastases, which are the primary cause of cancerrelated mortality. In such cases, the treatment strategies are likely to target systemic metastases, while the removal of metastatic LNs and targeting of non-metastatic draining LNs may potentially augment the effectiveness of immune checkpoint blockade (ICB) treatment (52, 63). In this review, we discuss the roles of TDLNs in cancer metastasis, the impact of cancer cell progression and invasion on the LN immune microenvironment and the function of TDLNs in immunotherapy, as well as the opportunities and challenges in targeting LN metastasis for improving patient care.

Cancer metastasis in the lymphatic system

Lymphangiogenesis in cancer

Tumor growth stimulates both angiogenesis (64) and lymphangiogenesis (65–70), facilitating cancer metastasis through the blood circulation and lymphatic vasculature. Tumor lymphangiogenesis is a prognostic indicator for increased risk of LN metastasis in various cancers, including melanoma (71), head and neck cancer (72-74), and breast cancer (66). Cancer cells and inflammatory cells in the tumor microenvironment secrete lymphangiogenic factors, such as VEGF-C (65, 66), VEGF-D (67), PDGF-BB (69), IGF-1 and IGF-2 (75), HGF (76, 77), FGF-2 (78), interleukin-1 (79), and COX-2 (80), promoting the proliferation and dilatation of peritumoral lymphatics, thereby favoring their metastatic spread (Figure 1). In several types of tumors, including fibrosarcoma and melanoma, lymphangiogenesis occurs mainly in the tumor margin. Inside tumors, lymphatic vessels are compressed (68, 81), possibly due to the solid stress exerted by growing cancer cells (32, 44, 45, 55, 56, 68, 82-86), but functional lymphatic vessels in the tumor periphery remain and can transport antigen and metastatic cancer cells. The presence of peritumoral lymphatic vessels accelerates LN metastasis in patients with head and neck cancer by accelerating lymphatic intravasation (72) and actively promotes cancer cell transport to LNs in preclinical models via chemoinvasion that utilizes chemokine signaling pathway such as CCR7 - CCL21 (87, 88), CXCR4 - CXCL12 (89), and CCL1-CCR8 (90) (Figure 1).

Lymphatic vessels also play a critical role in transporting antigens and antigen-presenting cells to LNs to initiate an adaptive immune response. Lymphatic endothelial cells can also archive and present antigens (91, 92). However, instead of enhancing tumor antigen transportation and immune cell trafficking to the TDLNs, tumorassociated lymphatic endothelium can impede anti-cancer T-cell response and promote cancer cell immune evasion and metastasis (93–97) (Figure 1). In the tumor microenvironment, infiltrating CD8 + T cells produce IFN- γ which drives the expression of PD-L1 on lymphatic endothelial cells (LECs) (94). The presence of PD-L1 on LECs induces T cell tolerance through PD1/PD-L1 interaction and lack of co-stimulation (98), thus restricting CD8+ T cell activation and reducing their accumulation in the tumor microenvironment (95). Additionally, PD-L1 on LECs in the LN prohibited LEC proliferation, protected them from apoptosis, and regulated LN



expansion and contraction during inflammation (99). In a murine model of B16F10 melanoma expressing both VEGF-C and chicken ovalbumin (OVA), overexpression of VEGF-C in cancer cells promotes lymphangiogenesis in both primary tumors and TDLNs, leads to CD8+ T cell immune tolerance via LEC cross-presentation (Figure 1), protects tumors against preexisting anti-cancer immunity, and promotes the local elimination of OVA-specific CD8+ T cells (93). Mice lacking dermal lymphatic vessels (K14-VEGFR3-Ig mice) with B16F10 intradermal tumors have reduced spontaneous lung metastasis and enhanced tumor infiltration of cytotoxic CD8 T cells (96). The presence of tumor-associated lymphatic vessels also promotes the exit of CD8+ T cells via the CXCL12 - CXCR4 chemotaxis, reducing the retention of tumor-specific CD8+ T cells (97). Pharmacologically blocking CXCR4 or genetically depleting CXCL12 in LECs enhances CD8+ T cell retention and promotes anticancer immunity (97).

vessels suppresses lymphatic contraction and reduces tumor-antigen transportation to the lymph nodes.

While lymphangiogenesis is associated with cancer metastasis and poor prognosis, it also improves the therapeutic benefit of immune checkpoint inhibitors. In both murine melanoma models and patients with melanoma, the presence of tumor lymphatic vessels showed a strong correlation with a local inflammatory response (96). Despite the immunosuppressive microenvironment of lymphangiogenic tumors mediated in part by LEC antigen presentation-induced T cell tolerance (94, 95, 98), they are more sensitive to systemic immunotherapy due to the elevation of CCL21 in LECs, which promotes the retention of naïve T cells and DCs in lymphangiogenic tumors via CCL21/CCR7 chemokine axis (100). Of note, in the B16F10 melanoma model, a high density of effector T cells in the tumor promoted LEC apoptosis via the IFN- γ signaling pathway (92). Subsequently, the decline of tumor lymphatic vessel density restricted lymphatic drainage (92), which might prevent the egress of tumor-specific CD8+ T cells (97). Taken together, cancer-associated lymphatic vessels can potentially drive cancer progression, but also be utilized to improve anticancer immunotherapies.

The impact of cancer cell migration on the lymphatic system

The lymphatic vascular system is a low-pressure, unidirectional circulation system that plays a critical role in tissue fluid homeostasis, tissue regeneration, lipid absorption and transportation, as well as immune surveillance (43, 101). Lymphatic vessels can be divided into initial (capillaries), precollecting, and collecting lymphatic vessels (43, 85, 102). The tissue interstitial fluid and macromolecules enter the lymphatic system through initial lymphatic vessels to form lymph and subsequently drain to the collecting lymphatic vessels and LNs. Lymph eventually transports back through the thoracic duct to reenter the blood (43).

The initial lymphatic vessels are composed of a single thin layer of LECs, forming an oakleaf-like shape with button-like junctions between them (103). The low-pressure characteristic of lymphatic

vasculature, along with the button-like junctions of the initial LECs, plays an important role in antigen transportation and dendritic cell (DC) trafficking (104). The initial LECs constitutively express CCL21 to actively recruit DCs via CCL21-CCR7 chemotaxis (105). However, the button-like junction and the presence of CCL21 in LECs also open a path for cancer cells to enter lymphatic vessels (87, 88). Additionally, the initial lymphatic vessels permit the delivery of cancer-derived cytokines (106), shed tumor-derived factors (107), metabolites (108), and exosomes (109–111) to LNs. Consequently, these processes remodel non-hematopoietic cells in the LN (112–114), creating a pre-metastatic lymphovascular niche in the LNs to facilitate cancer invasion.

In the collecting lymphatic vessels, the LECs are interconnected by tight zipper-like junctions and are surrounded by lymphatic muscle cells (103, 115). In addition, the collecting lymphatic vessels also contain intraluminal valves to prevent backflow (116), thus allowing the unidirectional propulsion of lymph. The collecting lymphatic vessels exhibit autonomous tonic and phasic contraction, driving the lymph flow. However, during cancer progression, the accumulation of inducible nitric oxide synthase positive (iNOS+) and CD11b+Gr1+ myeloid cells in the peritumoral region suppresses the contraction of tumor-draining collecting lymphatic vessels (117) (Figure 1), which might diminish tumor antigen transportation and DC trafficking to the TDLNs, reducing the immune response (118). The reduction of tumor-draining lymphatic vessel contraction could lead to the elevation of interstitial fluid pressure in primary tumor sites (119, 120), which might drive cancer cells to migrate toward functional lymphatic vessels that are mainly positioned in the tumor margin (68). Of note, in vitro experiments demonstrated that the increased pressure can trigger cancer cells to create autocrine gradients of CCR7 ligands and secrete them into the extracellular matrix (ECM), subsequently driving cancer LN metastasis via CCR7-dependent chemotaxis in the direction of flow toward functional lymphatic vessels (121). The extent to which tumor-derived factors can directly suppress lymphatic muscle cell contraction requires further investigation.

Stromal adaptations in tumor-draining LNs

The LN stromal cells provide a framework structure that maintains the niches for immune cells, acts as a backbone for immune cell migration, and forms a conduit system to filter antigens in the lymph (122–125). In addition, the LN stromal cells also provide survival signals and soluble factors that can nourish immune cells to generate strong immune responses (126, 127). The LN stromal cells can be subdivided into FRCs, LECs, blood endothelial cells (BECs), and double-negative (DN) cells (126), based on their anatomical location and gene expression. Furthermore, some blood vessels in the lymph node are morphologically and functionally specialized vessels with thick endothelial cells called high endothelial venules (HEVs) (128). HEVs express high levels of addressins, such as PNAd and MAdCAM-1, which bind L-Selectin/CD62L on lymphocytes and support the migration of naïve lymphocytes from the bloodstream into lymph nodes (128, 129). The FRCs are mainly located in the T cell and B cell zones, with high expression of podoplanin (PDPN) but diminished or absent CD31 endothelial cell marker gene expression (122). The LECs are positioned in the subcapsular sinus, the interfollicular ridges and the medullary sinus (114, 123, 125, 130), expressing both PDPN and CD31. The BECs are positive for CD31 but lack PDPN gene expression (131). Advancements in single-cell RNA sequencing have characterized previously unrecognized spatial heterogeneity and immune functions of stromal cells in the lymph nodes (114, 122, 123, 125, 130, 131), suggesting that the disintegration of the highly organized stromal cell structure will impair the anti-cancer immune responses (132). We have shown that the growth of metastatic breast cancer cells in LNs imposes solid stress on HEVs and prevents the infiltration of T cells into the metastatic lesions (56) (Figure 2). Moreover, in both 4T1 breast cancer and B16F10 melanoma models, LN LECs undergo structure and gene expression remodeling even before cancer cell invasion (113). Tumor invasion causes the expansion of the FRC network and the down-regulation of CCL21 and IL-7, leading to immunosuppressive features (112). In human lymphoma, the transcriptomic remodeling of non-hematopoietic cells has been reported, including the upregulation of CD70 in the medulla stromal cells which drives the accumulation of CD27positive malignant B cells in the medullary regions (114). Taken together, stromal cell remodeling in LNs is one of the key events that precedes and occurs during LN metastasis and lymphoma progression (Figure 2). Uncovering the heterogeneity and plasticity of stromal cells in both immune-activated and immune-suppressed lymph nodes offers potential diagnostic markers to complement lymph node biopsy in clinical settings.

Metabolic adaptations during lymphatic metastasis

Metabolic reprogramming supports the diverse energetic and proliferative demands of cancer cells but also creates unique metabolic shifts that can influence disease progression and therapeutic targets. Tumors exhibit heterogeneous metabolic functions driven by a combination of cell-intrinsic factors and extrinsic influences from the tumor microenvironment (133–135). Metabolic and lipidomic processes significantly influence cancer cell progression and metastasis (136). For example, metabolic pathway alterations in cancer cells, such as increased lactate uptake and amino acid metabolism shifts, support metastasis and cancer progression (137, 138). Moreover, cancer cells upregulate antioxidant pathways to maintain redox balance amidst oxidative stress and mitochondrial dysfunction, crucial for their survival (139-141). Metabolism also plays a critical role in altering immune cell responses, thus exacerbating cancer progression (134, 142). Cancer cells can secrete metabolites and lipids that alter the function of surrounding fibroblasts and compromise immune cell function (143). For example, lipid droplet accumulation in cancer cells has been shown to inactivate immune signaling molecules and foster a hospitable environment for cancer cell progression (144). Additionally, lipids



The remodeling of TDLN microenvironment and cancer cells. The invasion of cancer cells leads to the remodeling of stromal cells in the lymph nodes, and cancer cell growth imposes solid stress, which compresses high endothelial venules (HEVs), preventing T cell ingress. In the lymph nodes, cancer cells also undergo metabolic reprogramming, such as the elevation of oleic acid, to gain resistance against ferroptosis in the bloodstream. The unique microenvironment in the lymph nodes drives cancer cell-intrinsic interferon signaling pathways, causing the elevation of MHC class I and MHC class II in cancer cells. This leads to resistance to NK cell killing and the accumulation of Treg cells in TDLNs through anergic mechanisms.

suchas prostaglandins can modulate immune responses and promote angiogenesis, further supporting tumor growth and metastasis (145).

Although metabolomic and lipidomic techniques are now being applied to understand the role of metabolites and lipids in the lymph node microenvironment related to cancer metastasis, much remains to be explored. Innovations in these technologies have enabled profiling from low cell number samples (146, 147), allowing for the study of both cancer cell and immune cell populations from metastatic sites including TDLNs. Recent work utilizing these low cell number metabolomic and lipidomic techniques has shown that the distinct microenvironment of the lymph induces metabolic and lipidomic alterations of cancer cells which enhances the establishment of secondary metastases (62). The lymph fluid of mice has higher levels of glutathione and oleic acid, and less free iron compared to plasma. These conditions reduce cancer cell lipid oxidation and promote resistance to ferroptosis in lymph node metastases in both immunocompromised mice with humanized melanomas and immunocompetent mice with melanomas. Metabolomic analysis revealed that melanoma cells in the lymph compared to blood exhibited increased lipid metabolism; in particular, melanoma cells in lymph had higher oleic acid content in membrane phospholipids. The acyl-CoA synthetase long-chain family member 3 (ACSL3), which is critical in converting fatty acids into fatty acyl-CoA esters, enables cancer cells in the lymph to incorporate oleic acid in the cell membrane to protect them from ferroptosis (62).

Within the context of lipidomic dependencies in lymph nodes, recent studies have shown that a subpopulation of pre-metastatic oral carcinoma cells expresses high levels of the fatty acid receptor CD36 and lipid metabolism genes (148). These cells rely on dietary lipids to promote metastasis, and blocking CD36 significantly inhibited lymph node metastasis, underscoring the critical role of lipid metabolism in the metastatic process (148). Further comparative transcriptomic and metabolomic analyses of primary and LN-metastatic tumors in mice with melanoma found that LN metastases require a metabolic shift toward fatty acid oxidation (FAO) (61). Yes-associated protein (YAP) is selectively activated in metastatic lesions in LNs, which leads to increased transcription of genes in the FAO signaling pathway. Genetic deletion of YAP or pharmacological inhibition of FAO suppressed LN metastasis in mice (61). Furthermore, tumor-derived lactic acid alters the metabolic status of TDLN stroma to facilitate metastasis (108). These studies show that metastatic cancer cells in lymph nodes compared to cancer cells from the primary tumor undergo metabolomic and lipidomic alterations that confer survival advantages for metastasis. Considering that current cancer diagnoses primarily rely on features observed in cancer cells at primary sites, gaining a deeper understanding of the lipidomic and metabolic adaptations that occur during LN metastasis could offer novel solutions to prevent the spread of cancer cells. The advancements in single-cell transcriptomics and spatial metabolomics provide novel solutions to address the metabolic evolution of cancer and immune cells during LN metastasis.

LN metastasis and systemic metastasis

Different models and mechanisms of cancer metastasis have influenced clinical practice, including the role of LN metastases as predictors of distant metastasis. LN involvement has been historically considered a crucial step that preceded further metastasis, but contemporary insights suggest that distant metastases may not solely originate from LN-seeded cells. In the prevalent sequential progression model, cancer cells first spread to regional LNs and then disseminate to distant organs via the lymphatic system (149). In this model, LN metastasis serves as an important prognostic factor and triggers lymphadenectomy to prevent tumor cells from spreading further to distant organs (16, 19, 21, 150-152). However, in colorectal cancer (153-155) and breast cancer (156-158), evolutionary analysis of synchronous LN metastases and distant metastases indicates that only a small portion of patients share clonal ancestry between their LN metastases and distant metastases. In colorectal cancer, ~65% of lymphatic and distant metastases arose from independent subclones in the primary tumors, whereas ~35% of cases shared a common subclone origin (153, 155). Evolutionary analysis of matched breast cancer primary and metastatic tumors from 10 patients demonstrates the presence of both monoclonal and polyclonal origins of metastasis (156). Taken together, these data suggest that both direct hematogenous metastasis from the primary tumor and sequential progression from LNs can occur. Being able to determine which processes are occurring in an individual patient is a current unmet challenge.

Evolutionary studies of cancer in solid tumors reveal that LN metastases display higher levels of genetic diversity than distant metastases (154, 159), suggestive of their polyclonal nature (160). In colorectal cancer, LN metastases exhibit higher levels of inter-lesion and intra-lesion diversity than distant metastases (154). Additionally, colorectal cancer LN metastases develop through a wide evolutionary bottleneck, resulting in polyclonal and polyphyletic phenotypes. Distant metastases form monophyletic groups and have reduced clonality compared to LN metastases, suggesting different evolutionary mechanisms between LN metastases and distant metastases (154). A similar phenomenon was observed using a molecular barcoding system in a murine breast cancer spontaneous metastasis model, which showed LN metastases have higher levels of clonal diversity than distant metastases (159). These evolutionary studies reveal that fewer clones of cancer cells from the primary tumor site can form metastases in distant organs compared to TDLNs, which might be attributed to the different selective pressures and microenvironments in these organs, such as the varying metabolites in distant organs (161, 162). Meanwhile, cancer cells need to travel a longer distance to establish metastasis in distant organs than in locoregional lymph nodes. Since, both human (153, 155) and murine (163, 164) LN metastases can seed distant metastases, the abundance of heterogeneous cancer cell subclones in LNs could potentially give rise to more treatment-resistant cancer cells, such as the ferroptosis-resistant clones (62) or immuneresistant clones (57, 59), which spread systemically and generate

secondary metastases. Further study is needed to understand the consequences of the greater diversity of LN metastases, as well as the selective pressures in different organs.

In the early stage of cancer progression, the sentinel LNs likely retain anti-cancer immunity to prevent cancer cell intravasation and metastasis through blood circulation. This hypothesis is supported by the fact that both breast cancer (165) and melanoma (166) patients with micrometastases (<= 0.1mm in melanoma, and <=0.2mm in breast cancer) showed the same clinical outcomes as patients with a negative sentinel LN biopsy. However, in later stage cancer, TDLNs are largely immune suppressed by soluble factors and exosomes from the primary tumor and metastatic cancer cells (53-55, 59, 167-169). The suppression of TDLNs leads to the induction of tumor antigenspecific Treg cells and the reprogramming of leukocytes. Adoptive transfer of leukocytes from metastatic lymph nodes into tumornaïve recipients that were then challenged with a model of experimental metastasis showed enhanced progression of metastases in the lung, suggesting systemic immunosuppression caused by leukocytes from the metastatic lymph nodes (57). Thus, the strong predictive ability of established LN metastases may be due to the immune suppression induced by these lesions.

TDLNs in immunotherapy

TDLNs in anti-cancer immune response

In the last decade, the landscape of cancer treatment has undergone significant changes, primarily driven by the progress of immunotherapy. However, despite these advancements, a significant challenge that persists is the subset of patients who are resistant to or have limited response to immunotherapy (170), particularly in patients with low tumor mutational burden and immunogenic neoantigens (171, 172). Further, studies show that about 20 ~ 30% of patients with metastatic melanoma initially respond to immune checkpoint blockade treatment but eventually develop resistance and relapse after therapy (173, 174). The acquired resistance to ICB treatment might be due to cancer cellintrinsic and -extrinsic changes, including the loss of MHC class I on cancer cells (175, 176), genetic mutations that diminish T-cell infiltration (177, 178), or host microbiota (179-182). The underlying mechanisms responsible for the lack of response among patients are yet to be fully understood. Many researchers have focused on investigating the impact of ICB treatment on the interactions between immune cells and cancer cells in the tumor microenvironment (170, 183-185), including the tertiary lymphoid structures (186-190), as well as the T cell phenotypes in the peripheral circulation (48, 50, 191-197). Recently, TDLNs have received more attention due to their normal function as an immune organ. Several studies demonstrated that TDLNs play a critical role in anti-cancer immunity upon ICB treatment (30, 31, 33, 34, 36, 47, 51, 198, 199) (Figure 3). Traditionally, lymph node biopsy is primarily employed for cancer staging and serves as a prognostic marker in the clinic. These new studies suggest that examining the



immune microenvironment phenotypes of lymph nodes might be useful in predicting the efficacy of ICB treatment.

One key determinant of ICB treatment efficacy is the quantity and quality of infiltrating CD8+ T cells identified before or during the early stages of treatment (183). After ICB treatment, newly generated CD8+ T cells that infiltrate the tumors displayed different TCR clones compared to the pre-treatment intratumoral CD8+ T cells (50, 197, 200, 201). T cell exhaustion is a state of dysfunction that occurs during chronic infection and cancer. It is characterized by reduced effector function, the expression of inhibitory receptors, and specific transcriptional pathways that differ from those in effector or memory T cells (202). These ICB pre-treatment CD8+ T cells in the tumors exhibit reduced effector function and lack of proliferation after ICB treatment, indicating that the cells are exhausted (201). In a clinical trial (NCT02919683) of patients with head and neck squamous cell carcinomas of the oral cavity, neoadjuvant anti-PD1 or anti-PD-1/anti-CTLA4 treatment promoted the expansion of anti-cancer CD8+ T cell clones that did not exist in the tumors before the treatment (197), suggesting that these newly generated anti-cancer CD8+ T cells are derived from the TDLNs and not from the expansion of pre-existing exhausted CD8+ T cells in the tumor. Using Kaede photoconvertible mice, experiments demonstrated that CD8+ effector T cells that enter and stay in the tumor develop an exhausted phenotype within 72 hours, whereas non-effector TCF1 + stem-like CD8+ T cells are continuously trafficked between the

tumor and the TDLNs (47). In brief, the stem-like TCF1+ CD8+ T cells do not form a stable population in the tumor; they continuously egress to the TDLNs via lymphatic drainage (47, 97) (Figure 3). From the TDLNs, these stem-like TCF1+ CD8+ T cells travel through efferent lymphatics and then enter the blood circulation and circulate back to the tumor to replenish this population (47) (Figure 3). Analysis of CD8+ T cells from primary human head and neck squamous cell carcinomas, regional LNs and blood shows that the non-metastatic TDLNs are the source of pre-exhausted CD8+ T cells (Tpex), which migrate to the tumor (52). Similar findings were discovered in preclinical animal models of lung cancer (31) and melanoma (199), in which most tumor-specific CD8+ T cells in the TDLNs are functional and exhibit a gene expression signature resembling that of stem-like CD8+ T cells (31) or resident memory CD8+ T cells (199), suggesting that the antigen presentation and T cell priming in TDLNs is critical for the ICB treatment.

PD-1/PD-L1 checkpoint-blocking antibodies are thought to act primarily in the tumor microenvironment, where PD-L1 is expressed by tumor cells (174), myeloid cells (168, 203, 204), LECs (94, 95, 98), blood endothelial cells (95), B cells (205, 206), T cells (207), dendritic cells (30, 203) and NK cells (208, 209). An evaluation of the role of TDLN in PD-1/PD-L1 checkpoint blockade therapy in two mouse tumor models reveals that immune checkpoint treatment induces CD8+ T cell accumulation in the tumor-draining but not in non-draining LNs (34). Surgical removal of these TDLNs abolished PD-1/PD-L1 checkpoint blockadeinduced tumor regression and was associated with decreased immune infiltration in the primary tumors (34). Moreover, the immunomodulator FTY720-which blocks the egress of lymphocytes from secondary lymphoid organs and thymus by agnostic activation of sphingosine 1 phosphate receptors (210)abrogated checkpoint therapy, suggesting that TDLNs function as sites of T cell invigoration required for PD-1/PD-L1 checkpoint blockade therapy (34). PD-L1 can interact with other binding partners such as CD80 to form heterodimers on the same cell (211, 212). In APCs, CD80 interacts with PD-L1 in cis to disrupt PD-L1/PD-1 interaction between APCs and T cells, limiting the PD-1 coinhibitory signal, while promoting CD28/CD80 interactions and enhancing T cell activation (213). Meanwhile, the presence of PD-L1 on T cells may also interact with CD80 on APCs (214) leading to tumor-promoting tolerance through varying mechanisms (207). Notably, TDLNs are enriched for tumor-specific PD-1+ T cells, which frequently interact with PD-L1+ conventional dendritic cells (30). Targeting TDLNs with PD-L1 inhibitors induces enhanced anti-cancer T cell immunity and accelerates the infiltration of progenitor-exhausted T cells into the tumor, leading to improved tumor control in mice (30) (Figure 3). Given that a variety of cells in the lymph nodes possess the ability to express PD-L1, including LECs (94, 95, 98), FRCs (215), dendritic cells (30, 203), macrophages, B cells, T cells, and NK cells (204, 208, 209), understanding the impact of PD-1/PD-L1 immune checkpoint blockade on these cells and their interactions with T cells in the TDLNs might provide novel solutions to further enhance the efficacy of immunotherapy. In summary, TDLNs are crucial for the maintenance and trafficking of tumor antigen-specific Tpex and stem-like CD8+ T cells. These cells can be primed and activated after ICB treatment, enabling them to migrate to the tumor site and exert their effector function on cancer cells.

Immune evasion in the LNs

Despite TDLNs being recognized as key organs in response to immunotherapy and modulating anti-cancer immunity, emerging data suggest that TDLNs are often immunosuppressed by cancer cells. In melanoma, cancer cells inhibit the activation of different DC subsets (169), including the CD5+ DC (58), and suppress immune activation in the LNs. The accumulation of Treg cells in TDLNs has been reported in several different types of solid tumors, including breast cancer (55, 59, 216), lung cancer (53), colorectal cancer (167), melanoma (57, 169), gastric cancer (217), head and neck cancer (218), and cervical cancer (168). Recently, we showed that the invasion of breast cancer cells in the LN blunts the CD4+ effector T cell proliferation and promotes the expansion of Treg cells (59). Mechanistically, we uncovered that breast cancer cells in the LN displayed high levels of interferon-induced gene signatures, including a subpopulation of cancer cells that had MHC-II gene expression and an absence of co-stimulatory molecules (Figure 2). Of note, the presence of MHC-II in cancer cells is not restricted to breast cancer. It has also been reported in other types of solid tumors (219), including colorectal cancer (220), lung cancer (221, 222), ovarian cancer (223) and melanoma (224, 225). Additionally, MHC-II is present in LECs within tumors and lymph nodes (43, 226-228), as well as the cancer-associated fibroblasts in pancreatic cancer (229). Without co-stimulatory signals on the antigenpresenting cell (in this case the cancer cell), CD4+ T cells will not differentiate into effector cells in response to antigens, which prevents an effective immune response. Subsequently, we reported that the MHC-II+ cancer cells suppress Th1 responses and promote the expansion of Tregs in the LNs. Furthermore, we found that depletion of MHC-II in 4T1 breast cancer cells significantly reduced LN metastasis and attenuated Treg cell expansion in the TDLNs. In contrast, overexpression of Ciita-a transcriptional activator of MHC-II-in 4T1 breast cancer cells promoted LN metastasis and enhanced Treg cell expansion in the TDLNs (59). While not shown, it is possible that the Treg cells in metastatic LNs subsequently enter the bloodstream to suppress systemic anti-cancer immunity and promote the establishment of distant metastases (230).

Tregs can also directly inhibit natural killer (NK) cell effector functions by the down-regulation of NKG2D receptors on the surface of NK cells caused by membrane-bound TGF-β on Tregs (231). NKG2D receptor downregulation impairs the ability of NK cells to recognize tumor cells. The depletion of mouse Treg cells enhanced NK cell proliferation and cytotoxicity in vivo (231, 232). Depletion of Treg cells from tumor-infiltrating lymphocytes also restored human NK cell-mediated tumor recognition (231). In a syngeneic melanoma mouse model featuring a serial selection of cancer cells with a preference for dissemination to the LNs, Reticker-Flynn et al. revealed that the LN microenvironment can reprogram cancer cells, leading to an elevation in interferoninduced gene signatures, which subsequently caused the upregulation of MHC class I and PD-L1 gene expression in cancer cells (57). The elevation of PD-L1 and MHC-I shielded melanoma cells from attack by (NK cells). They also found more Treg cells in LNs invaded by melanoma cells Figure 2. The tumor antigen-specific Treg cells subsequently leave the LN and systemically disseminate, thus rendering distant organs more hospitable to metastatic seeding (57). Similarly, in a murine breast cancer LN metastasis model, the expansion of Treg cells in LNs suppresses NK cell activity leading to breast cancer metastasis in LNs (216). Immune-activated T and B lymphocytes typically migrate to sites of inflammation rather than remaining in LNs for extended periods. Thus, NK cells, not T and B lymphocytes, may also participate in immune responses against targets inside LNs, opening an opportunity to target LN metastasis using NK cells.

Opportunities and challenges in targeting TDLNs

LN-targeted immunotherapy

As TDLNs are widely recognized for playing a key role in driving the immunotherapy response, it is hypothesized that the effectiveness of ICB relies predominantly on an influx of newly primed T cells into the tumor from peripheral lymphoid organs, particularly the TDLNs, rather than on reinvigorating exhausted and fully differentiated T cells that already exist in tumors (31, 33, 47, 52). Thus, leveraging the CD8+ T cell pools in the TDLNs is one of the promising approaches in immunotherapy. In the B16F10 melanoma model, directing anti-CTLA4 and anti-PD-1 treatment to the TDLN reduces the risk of immune-related adverse events, reduces distant metastases and recurrence, and enhances the anticancer T cell repertoire (233). A phase 1 clinical trial evaluated the efficacy of locally administering anti-CTLA-4 into TDLNs in patients with stage I/II melanoma (NCT04274816) (36). The results showed activation of migratory dendritic cells in sentinel LNs, an increase in effector T cells, and a decrease in Tregs in both sentinel LNs and peripheral blood (36). These findings suggest that local administration of ICB regimens is a safe and promising adjuvant treatment strategy for patients with early-stage melanoma. In preclinical breast cancer models, the CD8+ T cell viability and responses within the primary tumor were severely impaired. However, CD8+ T cell priming within LNs, which is dependent on lymphatic drainage from the primary tumor, remained intact (32). Therefore, TDLNs represent an important component of the cancer immunity cycle in melanoma and triplenegative breast cancer for which strategies may be developed to improve the effects of anti-PD-1 immunotherapy (32) (Figure 3). Notably, TDLN-derived lymphocytes with abundant tumorreactive pre-exhausted T cells (52) are relatively easy to obtain. This has been utilized in several phase I/II trial (NCT05981014, NCT06121570, NCT06121557) that investigate the safety and effectiveness of infusion of autologous lymphocytes derived from TDLNs in patients with HER2-negative breast cancer who did not respond to neoadjuvant chemotherapy.

Chemoimmunotherapy is an experimental therapeutic approach for cancer that combines traditional chemotherapy, such as paclitaxel, with immunotherapy. Mechanistically, this approach leverages chemotherapy to induce immunogenic cell death, whereby the release of danger signals and tumor antigens boost the anti-cancer efficacy of immunotherapy (234). LNtargeting nanotechnology that delivers the chemotherapeutic drug paclitaxel to direct chemoimmunotherapy to TDLNs resulted in T cell mobilization into the circulation and improved control of both primary and metastatic tumors as well as animal survival in multiple murine models of triple-negative breast cancer (35). These data show that TDLNs mediate the effects of chemoimmunotherapy in advanced murine breast cancers (35). Further, a pre-clinical study of metastatic lung cancer reported that TDLNs help control metastases after the primary tumor is removed but are not required for adjuvant immunotherapy efficacy (235). Using orthotopic murine breast cancer and melanoma that develop spontaneous LN metastases, we also showed that resection of TDLNs did not prevent ICB responses (63), which was attributed to the rerouting of antigen drainage to distant LNs. It is worth noting that these pre-clinical studies were conducted in mice, whose tumor lymphatic drainage and lymphosome are distinct from those of humans. The rate of positive SLNB after neoadjuvant ICB therapy in patients with melanoma or breast cancer will be an interesting secondary endpoint to follow.

Tumor-derived secreted factors preferentially drain to lymphatic vessels before dilution in the blood. A recent study

examined the cancer-derived secreted factors in lymph exudate in patients with metastatic melanoma after lymphadenectomy. This work demonstrated that tumor-derived extracellular vesicles were enriched in the lymph exudate (110), suggesting that exosomes may offer a novel solution for drug delivery to the LNs. However, tumorderived exosomes often cause stromal remodeling in the LNs to facilitate lymphatic metastasis by cancer cells (109). To overcome this, a recent strategy involved creating macrophage-tumor hybrid cells by introducing nuclei isolated from tumor cells into activated M1-like macrophages, resulting in the production of chimeric exosomes. These exosomes entered LNs and primed T cell activation through both the classical antigen-presenting cellinduced immunostimulatory pathways and a unique 'direct exosome interaction' mechanism (236). Of note, despite the interesting concept presented in this study, the clinical implementation of this strategy remains challenging.

LN-targeted cancer vaccination

Tumor-specific vaccines are emerging as a promising strategy to prevent tumorigenesis, as well as suppress the growth of existing tumors (39). Delivering these vaccines more efficiently to TDLNs, which are often the first sites where lymphocytes are presented with tumor-associated antigens, is under extensive exploration (38, 40, 41, 237-244). Intranodal administration of antigen plasmids (pMEL-TYR) and peptides (E-MEL and E-TYR) against Melan A Tyrosinase in patients with metastatic melanoma yielded an overall immune response rate of 50% (243). In both the B16F10 melanoma and E.G7-OVA lymphoma model, TDLN-targeting nanoparticle (NP)-conjugate vaccines induced substantially stronger local and systemic cytotoxic CD8+ T-cell responses when compared to non-TDLN-targeting vaccination, leading to enhanced tumor regression and host survival (237). Amphiphile modification improves the delivery of conjugated cancer vaccines and adjuvants to the lymph nodes, leading to 30-fold increases in T-cell priming and enhanced anti-tumor efficacy (238). A recent study demonstrated that amphiphile modification of G12D and G12R mutant KRAS (mKRAS) peptides (Amph-Peptides-2P) in combination with CpG oligonucleotide adjuvant (Amph-CpG-7909) enhanced the delivery of cancer vaccines to lymph nodes and promoted antitumor immune responses in pancreatic and colorectal cancer (38). In addition, other strategies have also been developed to enhance the lymph node-targeted vaccination, including the pullulan nanogel system (240), synthetic vaccine nanoparticles (239), LNtargeted lipid nanoparticles (242, 245), and LN-targeted immunization formulation (244, 246, 247). For example, oil immunization led to rapid transportation and retention of antigens in the interfollicular region of the LN, whereas without oil, the antigens were enriched in the medullary region (246). The PEGylated lipid NPs together with mannose facilitated the phagocytosis and antigen presentation by dendritic cells in the lymph nodes after subcutaneous administration (244). PEGylated reduced graphene oxide nanosheets (RGO-PEG, 20-30 nm in diameter) rapidly accumulated in LNs, elicited neoantigen-specific T cells and eradicated established MC-38 tumor (37). Furthermore,

in the murine melanoma model, mesoporous silica (MPS) rodbased vaccines enhanced durable expansion of lymph nodes and were associated with better and prolonged vaccine efficacy (247). Beyond LN-targeted cancer vaccines, amphiphile CAR-T ligands delivered to the lymph nodes can be presented by APCs in the native lymph node microenvironment to enhance CAR-T cell expansion (40), promote tumor antigen spreading and elicit endogenous anti-tumor T cells (41, 241).

Radiotherapy triggers abscopal effects to control tumor growth (248). In radioimmunotherapy, the synergy between radiotherapy and ICB treatment acts similar to in situ vaccination. Recent studies showed that TDLNs play a critical role in the efficacy of radioimmunotherapy (249-251), particularly in the adjuvant setting (251). In a syngeneic hepatocellular carcinoma mouse model, combining radiation and anti-PD-1 enhanced activated dendritic cells in the TDLNs, accelerated infiltration of activated cytotoxic T cells in both irradiated and non-irradiated tumors, and triggered abscopal effects in the non-irradiated tumors (248). In the B16F10 melanoma model, local tumor irradiation improved distant tumor control and was associated with the expansion of total CD8+ T cells and stem-like CD8+ T cells in the TDLNs (249). Interestingly, the delayed (adjuvant) TDLN irradiation enhanced the efficacy of radioimmunotherapy, unlike the concomitant TDLN irradiation which failed to promote the efficacy of radioimmunotherapy (251). Given that the sentinel lymph nodes or TDLNs are often surgically removed during the processes of cancer diagnosis and treatment, a better understanding of the roles of TDLNs and the tumor-draining lymphosomes in cancer vaccination and radioimmunotherapy is needed to enhance treatment efficacy.

Target lymphangiogenesis to enhance ICB treatment efficacy

VEGF-C-induced lymphangiogenesis promoted immune tolerance in murine melanoma (93, 94). Mechanistically, LECs associated with lymphangiogenesis in tumors or the draining LN promoted immune tolerance by eliminating anti-cancer CD8+ cells (95) and accelerating T cell egress (97). Thus, LECs in the local tumor microenvironment may be a target for immunomodulation. Further, data showed that pharmacologically blocking VEGFR-3 decreases Treg cell accumulation in melanoma (100). However, despite the immunosuppressive microenvironment in highly lymphangiogenic melanoma, heightened sensitivity to ICB treatment and adoptive T cell transfer therapy was measured. Mechanistically, the VEGF-C signaling pathway induces CCL21 expression in LECs, subsequently enhancing the infiltration of CCR7+ naïve T cells and DCs to the tumors (100). In glioblastoma and melanoma brain metastases, ectopic expression of VEGF-C promotes the drainage of meningeal lymphatics and enhances CD8+ T cell activation in the draining deep cervical LNs. Subsequently, activated CD8+ T cells migrate to and enter the tumor, resulting in better tumor control and long-lasting memory responses with anti-PD1 therapy (252). Taken together, these results demonstrate the multifaceted roles of VEGF-C and LECs in the tumor microenvironment. The presence of high levels of VEGF-C and tumor lymphangiogenesis is associated with immunosuppression but might also indicate better efficacy for ICB treatment. This might be due to the enhancement of T cell infiltration and increased lymphatic drainage, APC trafficking, and antigen transportation to TDLNs.

Challenges of SLNB and CLND

The sentinel LN biopsy is a standard method for the clinical staging of cancer. The decision to remove additional LNs in patients with a positive SLNB or to forgo completion lymph node dissection depends on various factors, including the type and stage of cancer, and individual patient characteristics. Emerging evidence suggests that CLND does not always provide significant clinical benefits (24, 25, 253). In both melanoma (23, 24, 253, 254) and breast cancer (22, 28, 255, 256), CLND did not show an overall survival benefit compared to SLNB alone in early-stage cancer patients. In addition, in patients with early-stage breast cancer (< 5 cm) and minimal sentinel LN involvement (one or more micrometastases < 2 mm), CLND did not show significant overall survival benefit compared to the no LN dissection group (256). In patients with clinically node-negative primary T1 to T3 breast cancer with at least one positive lymph node identified by SLNB, SLNB-only was not inferior to complete axillary LN dissection for recurrence-free survival with the addition of standard-of-care adjuvant systemic therapy and radiation (28). In patients with small primary breast tumors and negative results on ultrasonography of the axillary lymph nodes, it is safe to skip SLNB (257). Due to the limited benefits of CLND and the significantly higher incidence of lymphedema following the procedure (29), frequent follow-up observations, including the use of serial nodal ultrasound, may be offered to patients with low-risk micrometastatic melanoma (258). Currently, there is no cure and no FDA-approved drug for lymphedema making prevention an important clinical consideration. Lymphedema after cancer treatment causes swelling, discomfort, and limited mobility in the arm, hand, or leg on the affected side. It also increases the risk of infection and interferes with daily activities and quality of life. There is an urgent need to find a less aggressive strategy to target LN metastasis to prevent lymphedema and significantly improve the quality of life for cancer survivors while maintaining excellent cancer control.

Concluding remarks

It is important to note that the optimal management of LNpositive patients remains an active area of research, and personalized treatment approaches are continually evolving. Clinical trials and further studies are crucial to provide more evidence and guidance on the most effective strategies for addressing LN metastasis while maximizing anti-cancer immune responses. In some cases, sparing the LN and exploring alternative treatment strategies may be a practical option. These strategies include targeted therapies, radiation therapy, or systemic treatments such as chemotherapy, immunotherapy, and cancer vaccines. The decision should be made through a multidisciplinary approach involving input from surgeons, oncologists, and other relevant specialists, taking into account the specific characteristics of the patient's tumor type and individual factors. Ultimately, the choice between LN removal and alternative treatment strategies should be a data-driven, collaborative decision between the patient and their healthcare team. Further clinical trials exploring therapies that maximize cancer outcomes while minimizing lymphedema risk are critical to informing these decisions.

Author contributions

PL: Conceptualization, Visualization, Writing – original draft, Writing – review & editing. CF: Writing – review & editing. DJ: Funding acquisition, Writing – review & editing. JU: Funding acquisition, Writing – review & editing. TP: Conceptualization, Funding acquisition, Resources, Supervision, Writing – review & editing.

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References

1. Rakha EA, Martin S, Lee AHS, Morgan D, Pharoah PDP, Hodi Z, et al. The prognostic significance of lymphovascular invasion in invasive breast carcinoma. *Cancer.* (2012) 118:3670-80. doi: 10.1002/cncr.26711

2. Houvenaeghel G, Cohen M, Classe JM, Reyal F, Mazouni C, Chopin N, et al. Lymphovascular invasion has a significant prognostic impact in patients with early breast cancer, results from a large, national, multicenter, retrospective cohort study. *ESMO Open.* (2021) 6:100316. doi: 10.1016/j.esmoop.2021.100316

3. Leong SPL. Paradigm of metastasis for melanoma and breast cancer based on the sentinel lymph node experience. *Ann Surg Oncol.* (2004) 11:192S–7S. doi: 10.1007/BF02523627

4. Morton DL, Thompson JF, Cochran AJ, Mozzillo N, Elashoff R, Essner R, et al. Sentinel-node biopsy or nodal observation in melanoma. *N Engl J Med.* (2006) 355:1307–17. doi: 10.1056/NEJMoa060992

 Lale Atahan I, Yildiz F, Ozyigit G, Sari S, Gurkaynak M, Selek U, et al. Percent positive axillary lymph node metastasis predicts survival in patients with nonmetastatic breast cancer. Acta Oncol (Madr). (2008) 47:232–8. doi: 10.1080/ 02841860701678761

 Andersson Y, Frisell J, Sylvan M, De Boniface J, Bergkvist L. Breast cancer survival in relation to the metastatic tumor burden in axillary lymph nodes. *J Clin Oncol.* (2010) 28:2868–73. doi: 10.1200/JCO.2009.24.5001

7. Leong SPL, Zuber M, Ferris RL, Kitagawa Y, Cabanas R, Levenback C, et al. Impact of nodal status and tumor burden in sentinel lymph nodes on the clinical outcomes of cancer patients. *J Surg Oncol.* (2011) 103:518–30. doi: 10.1002/jso.21815

8. Faut M, Wevers KP, van Ginkel RJ, Diercks GFH, Hoekstra HJ, Kruijff S, et al. Nodular histologic subtype and ulceration are tumor factors associated with high risk of recurrence in sentinel node-negative melanoma patients. *Ann Surg Oncol.* (2017) 24:142–9. doi: 10.1245/s10434-016-5566-8

9. Mitra D, Ologun G, Keung EZ, Goepfert RP, Amaria RN, Ross MI, et al. Nodal recurrence is a primary driver of early relapse for patients with sentinel lymph node-

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Conflict of interest

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positive melanoma in the modern therapeutic era. Ann Surg Oncol. (2021) 28:3480–9. doi: 10.1245/s10434-021-09804-3

10. JATOI I, HILSENBECK SG, CLARK GM, OSBORNE CK. Significance of axillary lymph node metastasis in primary breast cancer. J Clin Oncol. (1999) 17:2334–40. doi: 10.1200/JCO.1999.17.8.2334

11. Hernandez-Aya LF, Chavez-MacGregor M, Lei X, Meric-Bernstam F, Buchholz TA, Hsu L, et al. Nodal status and clinical outcomes in a large cohort of patients with triple-negative breast cancer. *J Clin Oncol.* (2011) 29:2628–34. doi: 10.1200/JCO.2010.32.1877

12. Ho AS, Kim S, Tighiouart M, Gudino C, Mita A, Scher KS, et al. Metastatic lymph node burden and survival in oral cavity cancer. *J Clin Oncol.* (2017) 35:3601–9. doi: 10.1200/JCO.2016.71.1176

13. Cheng L, Zincke H, Blute ML, Bergstralh EJ, Scherer B, Bostwick DG. Risk of prostate carcinoma death in patients with lymph node metastasis. *Cancer.* (2001) 91:66–73. doi: 10.1002/(ISSN)1097-0142

14. Pagliarulo V, Hawes D, Brands FH, Groshen S, Cai J, Stein JP, et al. Detection of occult lymph node metastases in locally advanced node-negative prostate cancer. *J Clin Oncol.* (2006) 24:2735–42. doi: 10.1200/JCO.2005.05.4767

15. Berger AC, Sigurdson ER, LeVoyer T, Hanlon A, Mayer RJ, Macdonald JS, et al. Colon cancer survival is associated with decreasing ratio of metastatic to examined lymph nodes. *J Clin Oncol.* (2005) 23:8706–12. doi: 10.1200/JCO.2005.02.8852

16. Chang GJ, Rodriguez-Bigas MA, Skibber JM, Moyer VA. Lymph node evaluation and survival after curative resection of colon cancer: Systematic review. *J Natl Cancer Inst.* (2007) 99:433–41. doi: 10.1093/jnci/djk092

17. Morton DL, Hoon DSB, Cochran AJ, Turner RR, Essner R, Takeuchi H, et al. Lymphatic mapping and sentinel lymphadenectomy for early-stage melanoma: therapeutic utility and implications of nodal microanatomy and molecular staging for improving the accuracy of detection of nodal micrometastases. *Ann Surg.* (2003) 238:538–50. doi: 10.1097/01.sla.0000086543.45557.cb

18. Veronesi U, Paganelli G, Viale G, Luini A, Zurrida S, Galimberti V, et al. A randomized comparison of sentinel-node biopsy with routine axillary dissection in breast cancer. *N Engl J Med.* (2003) 349:546–53. doi: 10.1056/NEJMoa012782

19. Pfister DG, Ang K-K, Brizel DM, Burtness BA, Cmelak AJ, Colevas AD, et al. Head and neck cancers. J Natl Compr Cancer Netw. (2011) 9:596–650. doi: 10.6004/jnccn.2011.0053

20. Wong SL, Balch CM, Hurley P, Agarwala SS, Akhurst TJ, Cochran A, et al. Sentinel lymph node biopsy for melanoma: American society of clinical oncology and society of surgical oncology joint clinical practice guideline. *J Clin Oncol.* (2012) 30:2912–8. doi: 10.1200/JCO.2011.40.3519

21. Parsons HM, Begun JW, Kuntz KM, Tuttle TM, McGovern PM, Virnig BA. Lymph node evaluation for colon cancer in an era of quality guidelines: Who improves? *J Oncol Pract.* (2013) 9(4):e164–71. doi: 10.1200/JOP.2012.000812

22. Giuliano AE, Hunt KK, Ballman KV, Beitsch PD, Whitworth PW, Blumencranz PW, et al. Axillary dissection vs no axillary dissection in women with invasive breast cancer and sentinel node metastasis: A randomized clinical trial. *Jama*. (2011) 305:569–75. doi: 10.1001/jama.2011.90

23. Klemen ND, Han G, Leong SP, Kashani-Sabet M, Vetto J, White R, et al. Completion lymphadenectomy for a positive sentinel node biopsy in melanoma patients is not associated with a survival benefit. *J Surg Oncol.* (2019) 119:1053–9. doi: 10.1002/jso.25444

24. Leiter U, Stadler R, Mauch C, Hohenberger W, Brockmeyer NH, Berking C, et al. Final analysis of deCOG-SLT trial: no survival benefit for complete lymph node dissection in patients with melanoma with positive sentinel node. *J Clin Oncol.* (2019) 37:3000–8. doi: 10.1200/JCO.18.02306

25. Stadler R, Leiter U, Garbe C. Lack of survival benefit in sentinel lymph nodepositive melanoma with immediate complete lymphadenectomy - a review. *J Der Dtsch Dermatologischen Gesellschaft*. (2019) 17:7–13. doi: 10.1111/ddg.13707

26. Da Cunha Cosme ML, Liuzzi Samaterra JF, Siso Cardenas SA, Chaviano Hernández JI. Lymphadenectomy after a positive sentinel node biopsy in patients with cutaneous melanoma. A systematic review. *Surg Exp Pathol.* (2021) 4:2. doi: 10.1186/s42047-020-00083-y

27. Blankenstein SA, Bonenkamp JJ, Aarts MJB, van den Berkmortel FWPJ, Blank CU, Blokx WAM, et al. Is a history of optimal staging by sentinel lymph node biopsy in the era prior to adjuvant therapy associated with improved outcome once melanoma patients have progressed to advanced disease? *Ann Surg Oncol.* (2023) 30:573–86. doi: 10.1245/s10434-022-12600-2

28. de Boniface J, Filtenborg Tvedskov T, Rydén L, Szulkin R, Reimer T, Kühn T, et al. Omitting axillary dissection in breast cancer with sentinel-node metastases. *N Engl J Med.* (2024) 390:1163–75. doi: 10.1056/NEJMoa2313487

29. Rockson SG. Lymphedema after breast cancer treatment. N Engl J Med. (2018) 379:1937-44. doi: 10.1056/NEJMcp1803290

30. Dammeijer F, van Gulijk M, Mulder EE, Lukkes M, Klaase L, van den Bosch T, et al. The PD-1/PD-L1-checkpoint restrains T cell immunity in tumor-draining lymph nodes. *Cancer Cell.* (2020) 38:685–700.e8. doi: 10.1016/j.ccell.2020.09.001

31. Connolly KA, Kuchroo M, Venkat A, Khatun A, Wang J, William I, et al. A reservoir of stem-like CD8+ T cells in the tumor-draining lymph node preserves the ongoing antitumor immune response. *Sci Immunol.* (2021) 6:1–17. doi: 10.1126/ sciimmunol.abg7836

32. O'Melia MJ, Manspeaker MP, Thomas SN. Tumor-draining lymph nodes are survival niches that support T cell priming against lymphatic transported tumor antigen and effects of immune checkpoint blockade in TNBC. *Cancer Immunol Immunother*. (2021) 70:2179–95. doi: 10.1007/s00262-020-02792-5

33. Ruggiu M, Guérin MV, Corre B, Bardou M, Alonso R, Russo E, et al. Anti-PD-1 therapy triggers Tfh cell-dependent IL-4 release to boost CD8 T cell responses in tumor-draining lymph nodes. *J Exp Med.* (2024) 221(4):e20232104. doi: 10.1084/jem.20232104

34. Fransen MF, Hall T, Ossendorp F, Fransen MF, Schoonderwoerd M, Knopf P, et al. Tumor-draining lymph nodes are pivotal in PD-1/PD-L1 checkpoint therapy. *JCI Insight*. (2018) 3:1–7. doi: 10.1172/jci.insight.124507

35. Manspeaker MP, O'Melia MJ, Thomas SN. Elicitation of stem-like CD8(+) T cell responses via lymph node-targeted chemoimmunotherapy evokes systemic tumor control. *J Immunother. Cancer.* (2022) 10(9):e005079. doi: 10.1136/jitc-2022-005079

36. van Pul KM, Notohardjo JCL, Fransen MF, Koster BD, Stam AGM, Chondronasiou D, et al. Local delivery of low-dose anti–CTLA-4 to the melanoma lymphatic basin leads to systemic Treg reduction and effector T cell activation. *Sci Immunol.* (2022) 7:eabn8097. doi: 10.1126/sciimmunol.abn8097

37. Xu C, Hong H, Lee Y, Park KS, Sun M, Wang T, et al. Efficient lymph nodetargeted delivery of personalized cancer vaccines with reactive oxygen species-inducing reduced graphene oxide nanosheets. *ACS Nano*. (2020) 14:13268–78. doi: 10.1021/ acsnano.0c05062

38. Pant S, Wainberg ZA, Weekes CD, Furqan M, Kasi PM, Devoe CE, et al. Lymphnode-targeted, mKRAS-specific amphiphile vaccine in pancreatic and colorectal cancer: the phase 1 AMPLIFY-201 trial. *Nat Med.* (2024) 30:531–42. doi: 10.1038/ s41591-023-02760-3

39. Wang J, Zhang Z, Liang R, Chen W, Li Q, Xu J, et al. Targeting lymph nodes for enhanced cancer vaccination: From nanotechnology to tissue engineering. *Mater Today Bio.* (2024) 26:101068. doi: 10.1016/j.mtbio.2024.101068

40. Ma L, Dichwalkar T, Chang JYH, Cossette B, Garafola D, Zhang AQ, et al. Enhanced CAR-T cell activity against solid tumors by vaccine boosting through the chimeric receptor. *Sci* (80-.). (2019) 365:162–8. doi: 10.1126/science.aav8692

41. Ma L, Hostetler A, Morgan DM, Maiorino L, Sulkaj I, Whittaker CA, et al. Vaccine-boosted CAR T crosstalk with host immunity to reject tumors with antigen heterogeneity. *Cell.* (2023) 186:3148–3165.e20. doi: 10.1016/j.cell.2023.06.002

42. Wang Y, Wang H. Lymph node targeting for immunotherapy. *Immuno-Oncol Technol.* (2023) 20:100395. doi: 10.1016/j.iotech.2023.100395

43. Karakousi T, Mudianto T, Lund AW. Lymphatic vessels in the age of cancer immunotherapy. *Nat Rev Cancer*. (2024) 24:363–81. doi: 10.1038/s41568-024-00681-y

44. Nouri-Shirazi M, Banchereau J, Bell D, Burkeholder S, Kraus ET, Davoust J, et al. Dendritic cells capture killed tumor cells and present their antigens to elicit tumor-specific immune responses. *J Immunol.* (2000) 165:3797–803. doi: 10.4049/jimmunol.165.7.3797

45. O'Melia MJ, Rohner NA, Manspeaker MP, Francis DM, Kissick HT, Thomas SN. Quality of CD8+ T cell immunity evoked in lymph nodes is compartmentalized by route of antigen transport and functional in tumor context. *Sci Adv.* (2020) 6(50): eabd7134. doi: 10.1126/sciadv.abd7134

46. MacNabb BW, Tumuluru S, Chen X, Godfrey J, Kasal DN, Yu J, et al. Dendritic cells can prime anti-tumor CD8(+) T cell responses through major histocompatibility complex cross-dressing. *Immunity*. (2022) 55:982–997.e8. doi: 10.1016/j.immuni.2022.04.016

47. Li Z, Tuong ZK, Dean I, Willis C, Gaspal F, Fiancette R, et al. *In vivo* labeling reveals continuous trafficking of TCF-1+ T cells between tumor and lymphoid tissue. *J Exp Med.* (2022) 219(6):e20210749. doi: 10.1084/jem.20210749

48. Spitzer MH, Carmi Y, Reticker-Flynn NE, Kwek SS, Madhireddy D, Martins MM, et al. Systemic immunity is required for effective cancer immunotherapy. *Cell.* (2017) 168(3):487–502.e15. doi: 10.1016/j.cell.2016.12.022

49. van der Leun AM, Thommen DS, Schumacher TN. CD8+ T cell states in human cancer: insights from single-cell analysis. *Nat Rev Cancer*. (2020) 20:218–32. doi: 10.1038/s41568-019-0235-4

50. Wu TD, Madireddi S, de Almeida PE, Banchereau R, Chen YJJ, Chitre AS, et al. Peripheral T cell expansion predicts tumour infiltration and clinical response. *Nature*. (2020) 579:274–8. doi: 10.1038/s41586-020-2056-8

51. Huang Q, Wu X, Wang Z, Chen X, Wang L, Lu Y, et al. The primordial differentiation of tumor-specific memory CD8+ T cells as bona fide responders to PD-1/PD-L1 blockade in draining lymph nodes. *Cell.* (2022) 185(22):4049–4066.e25. doi: 10.1016/j.cell.2022.09.020

52. Rahim MK, Okholm TLH, Jones KB, McCarthy EE, Liu CC, Yee JL, et al. Dynamic CD8+ T cell responses to cancer immunotherapy in human regional lymph nodes are disrupted in metastatic lymph nodes. *Cell.* (2023) 186(6):1127–1143.e18. doi: 10.1016/j.cell.2023.02.021

53. Alonso R, Flament H, Lemoine S, Sedlik C, Bottasso E, Péguillet I, et al. Induction of anergic or regulatory tumor-specific CD4+ T cells in the tumordraining lymph node. *Nat Commun.* (2018) 9:1–17. doi: 10.1038/s41467-018-04524-x

54. Van Pul KM, Vuylsteke RJCLM, Van De Ven R, Te Velde EA, Rutgers EJT, Van Den Tol PM, et al. Selectively hampered activation of lymph node-resident dendritic cells precedes profound T cell suppression and metastatic spread in the breast cancer sentinel lymph node. *J Immunother. Cancer.* (2019) 7:1–14. doi: 10.1186/s40425-019-0605-1

55. Núñez NG, Tosello Boari J, Ramos RN, Richer W, Cagnard N, Anderfuhren CD, et al. Tumor invasion in draining lymph nodes is associated with Treg accumulation in breast cancer patients. *Nat Commun.* (2020) 11:1–15. doi: 10.1038/s41467-020-17046-2

56. Jones D, Wang Z, Chen IX, Zhang S, Banerji R, Lei P-J, et al. Solid stress impairs lymphocyte infiltration into lymph-node metastases. *Nat Biomed Eng.* (2021) 5 (12):1426–36. doi: 10.1038/s41551-021-00766-1

57. Reticker-Flynn NE, Zhang W, Belk JA, Basto PA, Escalante NK, Pilarowski GOW, et al. Lymph node colonization induces tumor-immune tolerance to promote distant metastasis. *Cell.* (2022) 185(11):1924–42.e23. doi: 10.1016/j.cell.2022.04.019

58. He M, Roussak K, Ma F, Borcherding N, Garin V, White M, et al. CD5 expression by dendritic cells directs T cell immunity and sustains immunotherapy responses. *Sci (80-.).* (2023) 379(6633):eabg2752. doi: 10.1126/science.abg2752

59. Lei PJ, Pereira ER, Andersson P, Amoozgar Z, Van Wijnbergen JW, O'Melia MJ, et al. Cancer cell plasticity and MHC-II-mediated immune tolerance promote breast cancer metastasis to lymph nodes. *J Exp Med.* (2023) 220(9):e20221847. doi: 10.1084/jem.20221847

60. Leslie PL, Chao YL, Tsai YH, Ghosh SK, Porrello A, Van Swearingen AED, et al. Histone deacetylase 11 inhibition promotes breast cancer metastasis from lymph nodes. *Nat Commun.* (2019) 10(1):4192. doi: 10.1038/s41467-019-12222-5

61. Lee Ck, Jeong Sh, Jang C, Bae H, Kim YH, Park I, et al. Tumor metastasis to lymph nodes requires YAP-dependent metabolic adaptation. *Sci* (80-.). (2019) 363:644–9. doi: 10.1126/science.aav0173

62. Ubellacker JM, Tasdogan A, Ramesh V, Shen B, Mitchell EC, Martin-Sandoval MS, et al. Lymph protects metastasizing melanoma cells from ferroptosis. *Nature*. (2020) 585:113–8. doi: 10.1038/s41586-020-2623-z

63. Zhou H, Baish JW, O'Melia MJ, Darragh LB, Specht E, Czapla J, et al. Cancer immunotherapy responses persist after lymph node resection. *bioRxiv* [*Preprint*]. (2024) 2023.09.19.558262. doi: 10.1101/2023.09.19.558262

64. Carmeliet P, Jain RK. Angiogenesis in cancer and other diseases. *Nature*. (2000) 407:249–57. doi: 10.1038/35025220

65. Mandriota SJ, Jussila L, Jeltsch M, Compagni A, Baetens D, Prevo R, et al. Vascular endothelial growth factor-C-mediated lymphangiogenesis promotes tumour metastasis. *EMBO J.* (2001) 20:672–82. doi: 10.1093/emboj/20.4.672

66. Skobe M, Hawighorst T, Jackson DG, Prevo R, Janes L, Velasco P, et al. Induction of tumor lymphangiogenesis by VEGF-C promotes breast cancer metastasis. *Nat Med.* (2001) 7:192–8. doi: 10.1038/84643

67. Stacker SA, Caesar C, Baldwin ME, Thornton GE, Williams RA, Prevo R, et al. VEGF-D promotes the metastatic spread of tumor cells via the lymphatics. *Nat Med.* (2001) 7:186–91. doi: 10.1038/84635

68. Padera TP, Kadambi A, Di Tomaso E, Mouta Carreira C, Brown EB, Boucher Y, et al. Lymphatic metastasis in the absence of functional intratumor lymphatics. *Sci (80-.)*. (2002) 296:1883–6. doi: 10.1126/science.1071420

69. Cao R, Björndahl MA, Religa P, Clasper S, Garvin S, Galter D, et al. PDGF-BB induces intratumoral lymphangiogenesis and promotes lymphatic metastasis. *Cancer Cell.* (2004) 6:333–45. doi: 10.1016/j.ccr.2004.08.034

70. Stacker SA, Williams SP, Karnezis T, Shayan R, Fox SB, Achen MG. Lymphangiogenesis and lymphatic vessel remodelling in cancer. *Nat Rev Cancer.* (2014) 14:159–72. doi: 10.1038/nrc3677

71. Dadras SS, Paul T, Bertoncini J, Brown LF, Muzikansky A, Jackson DG, et al. Tumor lymphangiogenesis: A novel prognostic indicator for cutaneous melanoma metastasis and survival. *Am J Pathol.* (2003) 162:1951–60. doi: 10.1016/S0002-9440(10) 64328-3

72. Beasley NJP, Prevo R, Banerji S, Leek RD, Moore J, van Trappen P, et al. Intratumoral lymphangiogenesis and lymph node metastasis in head and neck cancer. *Cancer Res.* (2002) 62(5):1315–20.

73. Maula S-M, Luukkaa M, Grénman R, Jackson D, Jalkanen S, Ristamäki R. Intratumoral lymphatics are essential for the metastatic spread and prognosis in squamous cell carcinomas of the head and neck region. *Cancer Res.* (2003) 63 (8):1920–6.

74. Franchi A, Gallo O, Massi D, Baroni G, Santucci M. Tumor lymphangiogenesis in head and neck squamous cell carcinoma: a morphometric study with clinical correlations. *Cancer.* (2004) 101:973–8. doi: 10.1002/cncr.20454

75. Björndahl M, Cao R, Nissen LJ, Clasper S, Johnson LA, Xue Y, et al. Insulin-like growth factors 1 and 2 induce lymphangiogenesis *in vivo*. *Proc Natl Acad Sci U. S. A*. (2005) 102:15593–8. doi: 10.1073/pnas.0507865102

76. Kajiya K, Hirakawa S, Ma B, Drinnenberg I, Detmar M. Hepatocyte growth factor promotes lymphatic vessel formation and function. *EMBO J.* (2005) 24:2885–95. doi: 10.1038/sj.emboj.7600763

77. Cao R, Björndahl MA, Gallego MI, Chen S, Religa P, Hansen AJ, et al. Hepatocyte growth factor is a lymphangiogenic factor with an indirect mechanism of action. *Blood.* (2006) 107:3531-6. doi: 10.1182/blood-2005-06-2538

78. Chang LK, Garcia-Cardeña G, Farnebo F, Fannon M, Chen EJ, Butterfield C, et al. Dose-dependent response of FGF-2 for lymphangiogenesis. *Proc Natl Acad Sci U.* S. A. (2004) 101:11658–63. doi: 10.1073/pnas.0404272101

79. Watari K, Shibata T, Kawahara A, Sata KI, Nabeshima H, Shinoda A, et al. Tumor-derived interleukin-1 promotes lymphangiogenesis and lymph node metastasis through M2-type macrophages. *PloS One.* (2014) 9:1–15. doi: 10.1371/journal.pone.0099568

80. Lyons TR, Borges VF, Betts CB, Guo Q, Kapoor P, Martinson HA, et al. Cyclooxygenase-2-dependent lymphangiogenesis promotes nodal metastasis of postpartum breast cancer. J Clin Invest. (2014) 124:3901–12. doi: 10.1172/JCI73777

81. Leu AJ, Berk DA, Lymboussaki A, Alitalo K, Jain RK. Absence of functional lymphatics within a murine sarcoma: a molecular and functional evaluation. *Cancer Res.* (2000) 60(16):4324–7.

82. Nia HT, Liu H, Seano G, Datta M, Jones D, Rahbari N, et al. Solid stress and elastic energy as measures of tumour mechanopathology. *Nat Biomed Eng.* (2016) 1:0004. doi: 10.1038/s41551-016-0004

83. Nia HT, Datta M, Seano G, Huang P, Munn LL, Jain RK. Quantifying solid stress and elastic energy from excised or *in situ* tumors. *Nat Protoc.* (2018) 13:1091–105. doi: 10.1038/nprot.2018.020

84. Nia HT, Munn LL, Jain RK. Physical traits of cancer. Sci (80-.). (2020) 370(6516): eaaz0868. doi: 10.1126/science.aaz0868

85. Oliver G, Kipnis J, Randolph GJ, Harvey NL. The lymphatic vasculature in the 21st century: novel functional roles in homeostasis and disease. *Cell.* (2020) 182:270–96. doi: 10.1016/j.cell.2020.06.039

86. Padera TP, Stoll BR, Tooredman JB, Capen D, Tomaso E, Jain RK. Cancer cells compress intratumour vessels. *Nature*. (2004) 427:695. doi: 10.1038/427695a

87. Wiley HE, Gonzalez EB, Maki W, Wu MT, Hwang ST. Expression of CC chemokine receptor-7 and regional lymph node metastasis of B16 murine melanoma. *J Natl Cancer Inst.* (2001) 93:1638–43. doi: 10.1093/jnci/93.21.1638

88. Issa A, Le TX, Shoushtari AN, Shields JD, Swartz MA. Vascular endothelial growth factor-C and C-C chemokine receptor 7 in tumor cell-lymphatic cross-talk promote invasive phenotype. *Cancer Res.* (2009) 69:349–57. doi: 10.1158/0008-5472.CAN-08-1875

89. Kim M, Koh YJ, Kim KE, Koh BI, Nam DH, Alitalo K, et al. CXCR4 signaling regulates metastasis of chemoresistant melanoma cells by a lymphatic metastatic niche. *Cancer Res.* (2010) 70:10411–21. doi: 10.1158/0008-5472.CAN-10-2591

90. Das S, Sarrou E, Podgrabinska S, Cassella M, Mungamuri SK, Feirt N, et al. Tumor cell entry into the lymph node is controlled by CCL1 chemokine expressed by lymph node lymphatic sinuses. *J Exp Med.* (2013) 210:1509–28. doi: 10.1084/jem.20111627

91. Tamburini BA, Burchill MA, Kedl RM. Antigen capture and archiving by lymphatic endothelial cells following vaccination or viral infection. *Nat Commun.* (2014) 5:3989. doi: 10.1038/ncomms4989

92. Garnier L, Pick R, Montorfani J, Sun M, Brighouse D, Liaudet N, et al. IFN- γ -dependent tumor-antigen cross-presentation by lymphatic endothelial cells promotes their killing by T cells and inhibits metastasis. *Sci Adv.* (2022) 8:1–18. doi: 10.1126/sciadv.abl5162

93. Lund AW, Duraes FV, Hirosue S, Raghavan VR, Nembrini C, Thomas SN, et al. VEGF-C promotes immune tolerance in B16 melanomas and cross-presentation of tumor antigen by lymph node lymphatics. *Cell Rep.* (2012) 1:191–9. doi: 10.1016/ j.celrep.2012.01.005

94. Dieterich LC, Ikenberg K, Cetintas T, Kapaklikaya K, Hutmacher C, Detmar M. Tumor-associated lymphatic vessels upregulate PDL1 to inhibit T-cell activation. *Front Immunol.* (2017) 8:1–13. doi: 10.3389/fimmu.2017.00066

95. Lane RS, Femel J, Breazeale AP, Loo CP, Thibault G, Kaempf A, et al. IFN γ -activated dermal lymphatic vessels inhibit cytotoxic T cells in melanoma and inflamed skin. J Exp Med. (2018) 215:3057–74. doi: 10.1084/jem.20180654

96. Lund AW, Wagner M, Fankhauser M, Steinskog ES, Broggi MA, Spranger S, et al. Lymphatic vessels regulate immune microenvironments in human and murine melanoma. *J Clin Invest.* (2016) 126:3389–402. doi: 10.1172/JCI79434

97. Steele MM, Jaiswal A, Delclaux I, Dryg ID, Murugan D, Femel J, et al. T cell egress via lymphatic vessels is tuned by antigen encounter and limits tumor control. *Nat Immunol.* (2023) 24(4):664–75. doi: 10.1038/s41590-023-01491-4

98. Tewalt EF, Cohen JN, Rouhani SJ, Guidi CJ, Qiao H, Fahl SP, et al. Lymphatic endothelial cells induce tolerance via PD-L1 and lack of costimulation leading to high-level PD-1 expression on CD8 T cells. *Blood*. (2012) 120:4772–82. doi: 10.1182/blood-2012-04-427013

99. Lucas ED, Finlon JM, Burchill MA, McCarthy MK, Morrison TE, Colpitts TM, et al. Type 1 IFN and PD-L1 coordinate lymphatic endothelial cell expansion and contraction during an inflammatory immune response. *J Immunol.* (2018) 201:1735–47. doi: 10.4049/jimmunol.1800271

100. Fankhauser M, Broggi MAS, Potin L, Bordry N, Jeanbart L, Lund AW, et al. Tumor lymphangiogenesis promotes T cell infiltration and potentiates immunotherapy in melanoma. *Sci Transl Med.* (2017) 9:1–13. doi: 10.1126/scitranslmed.aal4712

101. Stritt S, Koltowska K, Mäkinen T. Homeostatic maintenance of the lymphatic vasculature. Trends Mol Med. (2021) 27:955-70. doi: 10.1016/j.molmed.2021.07.003

102. Breslin JW, Yang Y, Scallan JP, Sweat RS, Adderley SP, Murfee WL. Lymphatic vessel network structure and physiology. *Compr Physiol.* (2019) 9(1):207–99. doi: 10.1002/cphy.c180015

103. Baluk P, Fuxe J, Hashizume H, Romano T, Lashnits E, Butz S, et al. Functionally specialized junctions between endothelial cells of lymphatic vessels. *J Exp Med.* (2007) 204:2349–62. doi: 10.1084/jem.20062596

104. Russo E, Nitschké M, Halin C. Dendritic cell interactions with lymphatic endothelium. Lymphat. Res Biol. (2013) 11:172-82. doi: 10.1089/lrb.2013.0008

105. Tal O, Lim HY, Gurevich I, Milo I, Shipony Z, Ng LG, et al. DC mobilization from the skin requires docking to immobilized CCL21 on lymphatic endothelium and intralymphatic crawling. *J Exp Med.* (2011) 208:2141–53. doi: 10.1084/jem.20102392

106. Ito M, Minamiya Y, Kawai H, Saito S, Saito H, Nakagawa T, et al. Tumorderived TGFβ-1 induces dendritic cell apoptosis in the sentinel lymph node. *J Immunol.* (2006) 176:5637–43. doi: 10.4049/jimmunol.176.9.5637

107. Bois H, Chen AL, Ventre KS, Steele MS, Karakousi T, Lund AW. Abstract 3511: Melanoma-shed, lymph-borne CSPG4 conditions the pre-metastatic lymph node niche. *Cancer Res.* (2023) 83:3511. doi: 10.1158/1538-7445.AM2023-3511

108. Riedel A, Helal M, Pedro L, Swietlik JJ, Shorthouse D, Schmitz W, et al. Tumorderived lactic acid modulates activation and metabolic status of draining lymph node stroma. *Cancer Immunol Res.* (2022) 10:482–97. doi: 10.1158/2326-6066.CIR-21-0778

109. Hood JL, San Roman S, Wickline SA. Exosomes released by melanoma cells prepare sentinel lymph nodes for tumor metastasis. *Cancer Res.* (2011) 71:3792–801. doi: 10.1158/0008-5472.CAN-10-4455

110. Broggi MAS, Maillat L, Clement CC, Bordry N, Corthésy P, Auger A, et al. Tumor-associated factors are enriched in lymphatic exudate compared to plasma in metastatic melanoma patients. *J Exp Med.* (2019) 216:1091–107. doi: 10.1084/jem.20181618

111. García-Silva S, Benito-Martín A, Nogués L, Hernández-Barranco A, Mazariegos MS, Santos V, et al. Melanoma-derived small extracellular vesicles induce lymphangiogenesis and metastasis through an NGFR-dependent mechanism. *Nat Cancer.* (2021) 2:1387–405. doi: 10.1038/s43018-021-00272-y

112. Riedel A, Shorthouse D, Haas L, Hall BA, Shields J. Tumor-induced stromal reprogramming drives lymph node transformation. *Nat Immunol.* (2016) 17:1118–27. doi: 10.1038/ni.3492

113. Commerford CD, Dieterich LC, He Y, Hell T, Montoya-Zegarra JA, Noerrelykke SF, et al. Mechanisms of tumor-induced lymphovascular niche formation in draining lymph nodes. *Cell Rep.* (2018) 25:3554–3563.e4. doi: 10.1016/j.celrep.2018.12.002

114. Abe Y, Sakata-Yanagimoto M, Fujisawa M, Miyoshi H, Suehara Y, Hattori K, et al. A single-cell atlas of non-haematopoietic cells in human lymph nodes and lymphoma reveals a landscape of stromal remodelling. *Nat Cell Biol.* (2022) 24:565–78. doi: 10.1038/s41556-022-00866-3

115. Lei P-J, Ruscic KJ, Roh K, Rajotte JJ, O'Melia MJ, Bouta EM, et al. Lymphatic muscle cells are unique cells that undergo aging induced changes. *bioRxiv* [*Preprint*]. (2023) 2023.11.18.567621. doi: 10.1101/2023.11.18.567621

116. Zawieja DC. Contractile physiology of lymphatics. Lymphat. Res Biol. (2009) 7:87–96. doi: 10.1089/lrb.2009.0007

117. Liao S, Bouta EM, Morris LM, Jones D, Jain RK, Padera TP. Inducible nitric oxide synthase and CD11b+Gr1+ Cells impair lymphatic contraction of tumordraining lymphatic vessels. *Lymphat. Res Biol.* (2019) 17:294–300. doi: 10.1089/ lrb.2018.0013

118. Liao S, Cheng G, Conner DA, Huang Y, Kucherlapati RS, Munn LL, et al. Impaired lymphatic contraction associated with immunosuppression. *Proc Natl Acad Sci U. S. A.* (2011) 108:18784–9. doi: 10.1073/pnas.1116152108

119. Hompland T, Ellingsen C, Øvrebø KM, Rofstad EK. Interstitial fluid pressure and associated lymph node metastasis revealed in tumors by dynamic contrastenhanced MRI. *Cancer Res.* (2012) 72:4899–908. doi: 10.1158/0008-5472.CAN-12-0903

120. Swartz MA, Lund AW. Lymphatic and interstitial flow in the tumour microenvironment: Linking mechanobiology with immunity. *Nat Rev Cancer*. (2012) 12:210–9. doi: 10.1038/nrc3186

121. Shields JD, Fleury ME, Yong C, Tomei AA, Randolph GJ, Swartz MA. Autologous chemotaxis as a mechanism of tumor cell homing to lymphatics via interstitial flow and autocrine CCR7 signaling. *Cancer Cell.* (2007) 11:526–38. doi: 10.1016/j.ccr.2007.04.020

122. Rodda LB, Lu E, Bennett ML, Sokol CL, Wang X, Luther SA, et al. Single-cell RNA sequencing of lymph node stromal cells reveals niche-associated heterogeneity. *Immunity.* (2018) 48:1014–1028.e6. doi: 10.1016/j.immuni.2018.04.006

123. Takeda A, Hollmén M, Dermadi D, Pan J, Brulois KF, Kaukonen R, et al. Single-cell survey of human lymphatics unveils marked endothelial cell heterogeneity and mechanisms of homing for neutrophils. *Immunity.* (2019) 51:561–572.e5. doi: 10.1016/j.immuni.2019.06.027

124. Krishnamurty AT, Turley SJ. Lymph node stromal cells: cartographers of the immune system. *Nat Immunol.* (2020) 21:369–80. doi: 10.1038/s41590-020-0635-3

125. Xiang M, Grosso RA, Takeda A, Pan J, Bekkhus T, Brulois K, et al. A single-cell transcriptional roadmap of the mouse and human lymph node lymphatic vasculature. *Front Cardiovasc Med.* (2020) 7. doi: 10.3389/fcvm.2020.00052

126. Malhotra D, Fletcher AL, Astarita J, Lukacs-Kornek V, Tayalia P, Gonzalez SF, et al. Transcriptional profiling of stroma from inflamed and resting lymph nodes defines immunological hallmarks. *Nat Immunol.* (2012) 13:499–510. doi: 10.1038/ni.2262

127. Huang HY, Rivas-Caicedo A, Renevey F, Cannelle H, Peranzoni E, Scarpellino L, et al. Identification of a new subset of lymph node stromal cells involved in regulating plasma cell homeostasis. *Proc Natl Acad Sci U. S. A.* (2018) 115:E6826–35. doi: 10.1073/pnas.1712628115

128. Ager A, May MJ. Understanding high endothelial venules: Lessons for cancer immunology. *Oncoimmunology*. (2015) 4:1–14. doi: 10.1080/2162402X.2015.1008791

129. Hussain B, Kasinath V, Ashton-Rickardt GP, Clancy T, Uchimura K, Tsokos G, et al. High endothelial venules as potential gateways for therapeutics. *Trends Immunol.* (2022) 43;728–40. doi: 10.1016/j.it.2022.07.002

130. Fujimoto N, He Y, D'Addio M, Tacconi C, Detmar M, Dieterich LC. Single-cell mapping reveals new markers and functions of lymphatic endothelial cells in lymph nodes. *PloS Biol.* (2020) 18(4):e3000704. doi: 10.1101/2020.01.09.900241

131. Veerman K, Tardiveau C, Martins F, Coudert J, Girard JP. Single-cell analysis reveals heterogeneity of high endothelial venules and different regulation of genes controlling lymphocyte entry to lymph nodes. *Cell Rep.* (2019) 26:3116–3131.e5. doi: 10.1016/j.celrep.2019.02.042

132. du Bois H, Heim TA, Lund AW. Tumor-draining lymph nodes: At the crossroads of metastasis and immunity. *Sci Immunol.* (2021) 6:1–14. doi: 10.1126/sciimmunol.abg3551

133. Kim J, DeBerardinis RJ. Mechanisms and implications of metabolic heterogeneity in cancer. *Cell Metab.* (2019) 30:434–46. doi: 10.1016/j.cmet.2019.08.013

134. Elia I, Haigis MC. Metabolites and the tumour microenvironment: from cellular mechanisms to systemic metabolism. Nat Metab. (2021) 3:21–32. doi: 10.1038/s42255-020-00317-z

135. Demicco M, Liu XZ, Leithner K, Fendt SM. Metabolic heterogeneity in cancer. *Nat Metab.* (2024) 6:18–38. doi: 10.1038/s42255-023-00963-z

136. Griffin JL, Shockcor JP. Metabolic profiles of cancer cells. Nat Rev Cancer. (2004) 4:551-61. doi: 10.1038/nrc1390

137. Elia I, Broekaert D, Christen S, Boon R, Radaelli E, Orth MF, et al. Proline metabolism supports metastasis formation and could be inhibited to selectively

target metastasizing cancer cells. Nat Commun. (2017) 8:1-11. doi: 10.1038/ ncomms15267

138. Tasdogan A, Faubert B, Ramesh V, Ubellacker JM, Shen B, Solmonson A, et al. Metabolic heterogeneity confers differences in melanoma metastatic potential. *Nature.* (2020) 577:115–20. doi: 10.1038/s41586-019-1847-2

139. Gal K, Ibrahim MX, Wiel C, Sayin VI, Akula MK, Karlsson C, et al. Antioxidants can increase melanoma metastasis in mice. *Sci Transl Med.* (2015) 7:1–7. doi: 10.1126/scitranslmed.aad3740

140. Piskounova E, Agathocleous M, Murphy MM, Hu Z, Huddlestun SE, Zhao Z, et al. Oxidative stress inhibits distant metastasis by human melanoma cells. *Nature*. (2015) 527:186–91. doi: 10.1038/nature15726

141. Harris IS, DeNicola GM. The complex interplay between antioxidants and ROS in cancer. *Trends Cell Biol.* (2020) 30:440–51. doi: 10.1016/j.tcb.2020.03.002

142. Kaymak I, Williams KS, Cantor JR, Jones RG. Immunometabolic interplay in the tumor microenvironment. *Cancer Cell.* (2021) 39:28–37. doi: 10.1016/j.ccell.2020.09.004

143. Liu Y, Cao X. Characteristics and significance of the pre-metastatic niche. Cancer Cell. (2016) 30:668-81. doi: 10.1016/j.ccell.2016.09.011

144. Jovičić EJ, Janež AP, Eichmann TO, Koren Š, Brglez V, Jordan PM, et al. Lipid droplets control mitogenic lipid mediator production in human cancer cells. *Mol Metab.* (2023) 76:1–21. doi: 10.1016/j.molmet.2023.101791

145. Garcia C, Andersen CJ, Blesso CN. The role of lipids in the regulation of immune responses. *Nutrients*. (2023) 15(18):3899. doi: 10.3390/nu15183899

146. DeVilbiss AW, Zhao Z, Martin-Sandoval MS, Ubellacker JM, Tasdogan A, Agathocleous M, et al. Metabolomic profiling of rare cell populations isolated by flow cytometry from tissues. *Elife*. (2021) 10:e61980. doi: 10.7554/eLife.61980

147. Cosgrove J, Marçais A, Hartmann FJ, Bergthaler A, Zanoni I, Corrado M, et al. A call for accessible tools to unlock single-cell immunometabolism research. *Nat Metab.* (2024) 6:779–82. doi: 10.1038/s42255-024-01031-w

148. Pascual G, Avgustinova A, Mejetta S, Martín M, Castellanos A, Attolini CSO, et al. Targeting metastasis-initiating cells through the fatty acid receptor CD36. *Nature*. (2017) 541:41–5. doi: 10.1038/nature20791

149. David Nathanson S, Leonard-Murali S, Burmeister C, Susick L, Baker P. Clinicopathological evaluation of the potential anatomic pathways of systemic metastasis from primary breast cancer suggests an orderly spread through the regional lymph nodes. *Ann Surg Oncol.* (2020) 27:4810–8. doi: 10.1245/s10434-020-08904-w

150. Carlson RW, Allred DC, Anderson BO, Burstein HJ, Carter WB, Edge SB, et al. Invasive breast cancer. *J Natl Compr Cancer Netw.* (2011) 9:136–222. doi: 10.6004/ jnccn.2011.0016

151. Gradishar WJ, Anderson BO, Blair SL, Burstein HJ, Cyr A, Elias AD, et al. Breast cancer version 3.2014. J. Natl. Compr. Cancer netw. J Natl Compr Canc Netw. (2014) 12:542–90. doi: 10.6004/jnccn.2014.0058

152. Coit DG, Thompson JA, Algazi A, Andtbacka R, Bichakjian CK, Carson WE, et al. Melanoma, version 2.2016, NCCN clinical practice guidelines in oncology. *J Natl Compr Cancer Netw.* (2016) 14:450–73. doi: 10.6004/jnccn.2016.0051

153. Naxerova K, Reiter JG, Brachtel E, Lennerz JK, Van De Wetering M, Rowan A, et al. Origins of lymphatic and distant metastases in human colorectal cancer. *Sci* (80-.). (2017) 357:55–60. doi: 10.1126/science.aai8515

154. Reiter JG, Hung WT, Lee IH, Nagpal S, Giunta P, Degner S, et al. Lymph node metastases develop through a wider evolutionary bottleneck than distant metastases. *Nat Genet.* (2020) 52:692–700. doi: 10.1038/s41588-020-0633-2

155. Zhang C, Zhang L, Xu T, Xue R, Yu L, Zhu Y, et al. Mapping the spreading routes of lymphatic metastases in human colorectal cancer. *Nat Commun.* (2020) 11:1–11. doi: 10.1038/s41467-020-15886-6

156. Brown D, Smeets D, Székely B, Larsimont D, Marcell Szász A, Adnet PY, et al. Phylogenetic analysis of metastatic progression in breast cancer using somatic mutations and copy number aberrations. *Nat Commun.* (2017) 8:14944. doi: 10.1038/ncomms15759

157. Ullah I, Karthik GM, Alkodsi A, Kjällquist U, Stålhammar G, Lövrot J, et al. Evolutionary history of metastatic breast cancer reveals minimal seeding from axillary lymph nodes. *J Clin Invest.* (2018) 128:1355–70. doi: 10.1172/JCI96149

158. Venet D, Fimereli D, Rothé F, Boeckx B, Maetens M, Majjaj S, et al. Phylogenetic reconstruction of breast cancer reveals two routes of metastatic dissemination associated with distinct clinical outcome. *EBioMedicine*. (2020) 56:102793. doi: 10.1016/j.ebiom.2020.102793

159. Wagenblast E, Soto M, Gutiérrez-Ángel S, Hartl CA, Gable AL, Maceli AR, et al. A model of breast cancer heterogeneity reveals vascular mimicry as a driver of metastasis. *Nature*. (2015) 520:358–62. doi: 10.1038/nature14403

160. Jeong HS, Jones D, Liao S, Wattson DA, Cui CH, Duda DG, et al. Investigation of the lack of angiogenesis in the formation of lymph node metastases. *J Natl Cancer Inst.* (2015) 107:1–11. doi: 10.1093/jnci/djv155

161. Jin X, Demere Z, Nair K, Ali A, Ferraro GB, Natoli T, et al. A metastasis map of human cancer cell lines. *Nature*. (2020) 588:331–6. doi: 10.1038/s41586-020-2969-2

162. Ferraro GB, Ali A, Luengo A, Kodack DP, Deik A, Abbott KL, et al. Fatty acid synthesis is required for breast cancer brain metastasis. *Nat Cancer*. (2021) 2:414–28. doi: 10.1038/s43018-021-00183-y

163. Brown M, Assen FP, Leithner A, Abe J, Schachner H, Asfour G, et al. Lymph node blood vessels provide exit routes for metastatic tumor cell dissemination in mice. *Sci* (80-.). (2018) 359:1408–11. doi: 10.1126/science.aal3662

164. Pereira ER, Kedrin D, Seano G, Gautier O, Meijer EFJ, Jones D, et al. Lymph node metastases can invade local blood vessels, exit the node, and colonize distant organs in mice. *Sci* (80-.). (2018) 359:1403–7. doi: 10.1126/science.aal3622

165. Gobardhan PD, Elias SG, Madsen EVE, van Wely B, van den Wildenberg F, Theunissen EBM, et al. Prognostic value of lymph node micrometastases in breast cancer: a multicenter cohort study. *Ann Surg Oncol.* (2011) 18:1657–64. doi: 10.1245/s10434-010-1451-z

166. van Akkooi ACJ, de Wilt JHW, Verhoef C, Schmitz PIM, van Geel AN, Eggermont AMM, et al. Clinical relevance of melanoma micrometastases (<0.1 mm) in sentinel nodes: are these nodes to be considered negative? *Ann Oncol.* (2006) 17(10):1578–85. doi: 10.1093/annonc/mdl176

167. Deng L, Zhang H, Luan Y, Zhang J, Xing Q, Dong S, et al. Accumulation of foxp3+ T regulatory cells in draining lymph nodes correlates with disease progression and immune suppression in colorectal cancer patients. *Clin Cancer Res.* (2010) 16:4105–12. doi: 10.1158/1078-0432.CCR-10-1073

168. Heeren AM, Koster BD, Samuels S, Ferns DM, Chondronasiou D, Kenter GG, et al. High and interrelated rates of PD-L1+CD14+ antigen-presenting cells and regulatory T cells mark the microenvironment of metastatic lymph nodes from patients with cervical cancer. *Cancer Immunol Res.* (2015) 3:48–58. doi: 10.1158/2326-6066.CIR-14-0149

169. Van Den Hout MFCM, Koster BD, Sluijter BJR, Molenkamp BG, Van De Ven R, Van Den Eertwegh AJM, et al. Melanoma sequentially suppresses different DC subsets in the sentinel lymph node, affecting disease spread and recurrence. *Cancer Immunol Res.* (2017) 5:969–77. doi: 10.1158/2326-6066.CIR-17-0110

170. Morad G, Helmink BA, Sharma P, Wargo JA. Hallmarks of response, resistance, and toxicity to immune checkpoint blockade. *Cell.* (2021) 184:5309–37. doi: 10.1016/j.cell.2021.09.020

171. Yarchoan M, Hopkins A, Jaffee EM. Tumor mutational burden and response rate to PD-1 inhibition. N Engl J Med. (2017) 377:2500–1. doi: 10.1056/NEJMc1713444

172. Valero C, Lee M, Hoen D, Zehir A, Berger MF, Seshan VE, et al. Response rates to anti-PD-1 immunotherapy in microsatellite-stable solid tumors with 10 or more mutations per megabase. *JAMA Oncol.* (2021) 7:739-43. doi: 10.1001/jamaoncol.2020.7684

173. Sharma P, Hu-Lieskovan S, Wargo JA, Ribas A. Primary, adaptive, and acquired resistance to cancer immunotherapy. *Cell.* (2017) 168:707–23. doi: 10.1016/ j.cell.2017.01.017

174. Ribas A, Wolchok JD. Cancer immunotherapy using checkpoint blockade. *Science*. (2018) 359:1350-5. doi: 10.1126/science.aar4060

175. Burr ML, Sparbier CE, Chan KL, Chan Y-C, Kersbergen A, Lam EYN, et al. An evolutionarily conserved function of polycomb silences the MHC class I antigen presentation pathway and enables immune evasion in cancer. *Cancer Cell.* (2019) 36:385–401.e8. doi: 10.1016/j.ccell.2019.08.008

176. Lim SY, Shklovskaya E, Lee JH, Pedersen B, Stewart A, Ming Z, et al. The molecular and functional landscape of resistance to immune checkpoint blockade in melanoma. *Nat Commun.* (2023) 14(1):1516. doi: 10.1038/s41467-023-36979-y

177. Spranger S, Gajewski TF. Impact of oncogenic pathways on evasion of antitumour immune responses. *Nat Rev Cancer*. (2018) 18:139-47. doi: 10.1038/nrc.2017.117

178. Bergholz JS, Wang Q, Wang Q, Ramseier M, Prakadan S, Wang W, et al. PI3K β controls immune evasion in PTEN-deficient breast tumours. *Nature*. (2023) 617:139–46. doi: 10.1038/s41586-023-05940-w

179. Gopalakrishnan V, Spencer CN, Nezi L, Reuben A, Andrews MC, Karpinets TV, et al. Gut microbiome modulates response to anti-PD-1 immunotherapy in melanoma patients. *Science.* (2018) 359(6371):97–103. doi: 10.1126/science.aan4236

180. Routy B, Le Chatelier E, Derosa L, Duong CPM, Alou MT, Daillère R, et al. Gut microbiome influences efficacy of PD-1-based immunotherapy against epithelial tumors. *Science.* (2018) 359:91–7. doi: 10.1126/science.aan3706

181. Kalbasi A, Ribas A. Tumour-intrinsic resistance to immune checkpoint blockade. *Nat Rev Immunol.* (2020) 20:25–39. doi: 10.1038/s41577-019-0218-4

182. Giampazolias E, Pereira da Costa M, Lam KC, Lim KHJ, Cardoso A, Piot C, et al. Vitamin D regulates microbiome-dependent cancer immunity. *Science.* (2024) 384:428–37. doi: 10.1126/science.adh7954

183. Sade-Feldman M, Yizhak K, Bjorgaard SL, Ray JP, de Boer CG, Jenkins RW, et al. Defining T cell states associated with response to checkpoint immunotherapy in melanoma. *Cell.* (2018) 175(4):998–1013.e20. doi: 10.1016/j.cell.2018.10.038

184. Zhang Y, Chen H, Mo H, Hu X, Gao R, Zhao Y, et al. Single-cell analyses reveal key immune cell subsets associated with response to PD-L1 blockade in triple-negative breast cancer. *Cancer Cell.* (2021) 39:1578–1593.e8. doi: 10.1016/j.ccell.2021.09.010

185. Gebhardt T, Park SL, Parish IA. Stem-like exhausted and memory CD8(+) T cells in cancer. *Nat Rev Cancer.* (2023) 23:780–98. doi: 10.1038/s41568-023-00615-0

186. Hollern DP, Xu N, Thennavan A, Glodowski C, Garcia-Recio S, Mott KR, et al. B cells and T follicular helper cells mediate response to checkpoint inhibitors in high mutation burden mouse models of breast cancer. *Cell.* (2019) 179(5):1191–1206.e21. doi: 10.1016/j.cell.2019.10.028

187. Cabrita R, Lauss M, Sanna A, Donia M, Skaarup Larsen M, Mitra S, et al. Tertiary lymphoid structures improve immunotherapy and survival in melanoma. *Nature*. (2020) 577:561–5. doi: 10.1038/s41586-019-1914-8

188. Helmink BA, Reddy SM, Gao J, Zhang S, Basar R, Thakur R, et al. B cells and tertiary lymphoid structures promote immunotherapy response. *Nature.* (2020) 577 (7791):549–55. doi: 10.1038/s41586-019-1922-8

189. Vanhersecke L, Brunet M, Guégan J-P, Rey C, Bougouin A, Cousin S, et al. Mature tertiary lymphoid structures predict immune checkpoint inhibitor efficacy in solid tumors independently of PD-L1 expression. *Nat Cancer.* (2021) 2:794–802. doi: 10.1038/s43018-021-00232-6

190. Meylan M, Petitprez F, Becht E, Bougoüin A, Pupier G, Calvez A, et al. Tertiary lymphoid structures generate and propagate anti-tumor antibody-producing plasma cells in renal cell cancer. *Immunity.* (2022) 55:527–541.e5. doi: 10.1016/j.immuni.2022.02.001

191. Kamphorst AO, Pillai RN, Yang S, Nasti TH, Akondy RS, Wieland A, et al. Proliferation of PD-1+ CD8 T cells in peripheral blood after PD-1-targeted therapy in lung cancer patients. *Proc Natl Acad Sci U. S. A.* (2017) 114:4993–8. doi: 10.1073/ pnas.1705327114

192. Jansen CS, Prokhnevska N, Master VA, Sanda MG, Carlisle JW, Bilen MA, et al. An intra-tumoral niche maintains and differentiates stem-like CD8 T cells. *Nature*. (2019) 576:465–70. doi: 10.1038/s41586-019-1836-5

193. Kurtulus S, Madi A, Escobar G, Klapholz M, Nyman J, Christian E, et al. Checkpoint blockade immunotherapy induces dynamic changes in PD-1 – CD8 + Tumor-infiltrating T cells. *Immunity*. (2019) 50:181–194.e6. doi: 10.1016/jinmuni.2018.11.014

194. Li H, van der Leun AM, Yofe I, Lubling Y, Gelbard-Solodkin D, van Akkooi ACJ, et al. Dysfunctional CD8 T cells form a proliferative, dynamically regulated compartment within human melanoma. *Cell.* (2019) 176(4):775–789.e18. doi: 10.1016/j.cell.2018.11.043

195. Miller BC, Sen DR, Al Abosy R, Bi K, Virkud YV, LaFleur MW, et al. Subsets of exhausted CD8+ T cells differentially mediate tumor control and respond to checkpoint blockade. *Nat Immunol.* (2019) 20:326–36. doi: 10.1038/s41590-019-0312-6

196. Beltra JC, Manne S, Abdel-Hakeem MS, Kurachi M, Giles JR, Chen Z, et al. Developmental relationships of four exhausted CD8+ T cell subsets reveals underlying transcriptional and epigenetic landscape control mechanisms. *Immunity.* (2020) 52:825–841.e8. doi: 10.1016/j.immuni.2020.04.014

197. Luoma AM, Suo S, Wang Y, Gunasti L, Porter CBM, Nabilsi N, et al. Tissueresident memory and circulating T cells are early responders to pre-surgical cancer immunotherapy. *Cell*. (2022) 185(16):2918–2935.e29. doi: 10.1016/j.cell.2022.06.018

198. Binnewies M, Mujal AM, Pollack JL, Combes AJ, Hardison EA, Barry KC, et al. Unleashing type-2 dendritic cells to drive protective antitumor CD4 + T cell immunity. *Cell.* (2019) 177(3):556–71.e16. doi: 10.1016/j.cell.2019.02.005

199. Molodtsov AK, Khatwani N, Vella JL, Lewis KA, Zhao Y, Han J, et al. Resident memory CD8+ T cells in regional lymph nodes mediate immunity to metastatic melanoma. *Immunity*. (2021) 54:2117–2132.e7. doi: 10.1016/j.immuni.2021.08.019

200. Yost KE, Satpathy AT, Wells DK, Qi Y, Wang C, Kageyama R, et al. Clonal replacement of tumor-specific T cells following PD-1 blockade. *Nat Med.* (2019) 25:1251–9. doi: 10.1038/s41591-019-0522-3

201. Pai JA, Hellmann MD, Sauter JL, Mattar M, Rizvi H, Woo HJ, et al. Lineage tracing reveals clonal progenitors and long-term persistence of tumor-specific T cells during immune checkpoint blockade. *Cancer Cell.* (2023) 41:776–790.e7. doi: 10.1016/j.ccell.2023.03.009

202. Wherry EJ, Kurachi M. Molecular and cellular insights into T cell exhaustion. Nat Rev Immunol. (2015) 15:486–99. doi: 10.1038/nri3862

203. Lin H, Wei S, Hurt EM, Green MD, Zhao L, Vatan L, et al. Host expression of PD-L1 determines efficacy of PD-L1 pathway blockade-mediated tumor regression. *J Clin Invest.* (2018) 128:805–15. doi: 10.1172/JCI96113

204. Tang H, Liang Y, Anders RA, Taube JM, Qiu X, Mulgaonkar A, et al. PD-L1 on host cells is essential for PD-L1 blockade-mediated tumor regression. *J Clin Invest.* (2018) 128:580–8. doi: 10.1172/JCI96061

205. Chen BJ, Chapuy B, Ouyang J, Sun HH, Roemer MGM, Xu ML, et al. PD-L1 expression is characteristic of a subset of aggressive B-cell lymphomas and virus-associated Malignancies. *Clin Cancer Res.* (2013) 19:3462–73. doi: 10.1158/1078-0432.CCR-13-0855

206. Khan AR, Hams E, Floudas A, Sparwasser T, Weaver CT, Fallon PG. PD-L1hi B cells are critical regulators of humoral immunity. *Nat Commun.* (2015) 6:5997. doi: 10.1038/ncomms6997

207. Diskin B, Adam S, Cassini MF, Sanchez G, Liria M, Aykut B, et al. PD-L1 engagement on T cells promotes self-tolerance and suppression of neighboring macrophages and effector T cells in cancer. *Nat Immunol.* (2020) 21:442–54. doi: 10.1038/s41590-020-0620-x

208. Dong W, Wu X, Ma S, Wang Y, Nalin AP, Zhu Z, et al. The mechanism of antipd-l1 antibody efficacy against pd-l1-negative tumors identifies nk cells expressing pdl1 as a cytolytic effector. *Cancer Discovery*. (2019) 9:1422–37. doi: 10.1158/2159-8290.CD-18-1259

209. Sierra JM, Secchiari F, Nuñez SY, Iraolagoitia XLR, Ziblat A, Friedrich AD, et al. Tumor-experienced human NK cells express high levels of PD-L1 and inhibit CD8(+)

T cell proliferation. Front Immunol. (2021) 12:745939. doi: 10.3389/ fimmu.2021.745939

210. Chiba K. FTY720, a new class of immunomodulator, inhibits lymphocyte egress from secondary lymphoid tissues and thymus by agonistic activity at sphingosine 1-phosphate receptors. *Pharmacol Ther.* (2005) 108:308–19. doi: 10.1016/j.pharmthera.2005.05.002

211. Chaudhri A, Xiao Y, Klee AN, Wang X, Zhu B, Freeman GJ. PD-L1 Binds to B7–1 only in cis on the same cell surface. *Cancer Immunol Res.* (2018) 6:921–9. doi: 10.1158/2326-6066.CIR-17-0316

212. Zhao Y, Lee CK, Lin CH, Gassen RB, Xu X, Huang Z, et al. PD-L1:CD80 cisheterodimer triggers the co-stimulatory receptor CD28 while repressing the inhibitory PD-1 and CTLA-4 pathways. *Immunity*. (2019) 51:1059–1073.e9. doi: 10.1016/ j.immuni.2019.11.003

213. Sugiura D, Maruhashi T, Okazaki IM, Shimizu K, Maeda TK, Takemoto T, et al. Restriction of PD-1 function by cis-PD-L1/CD80 interactions is required for optimal T cell responses. *Sci (80-.).* (2019) 364:558–66. doi: 10.1126/science.aav7062

214. Mueller SN, Vanguri VK, Ha SJ, West EE, Keir ME, Glickman JN, et al. PD-L1 has distinct functions in hematopoietic and nonhematopoietic cells in regulating T cell responses during chronic infection in mice. *J Clin Invest.* (2010) 120:2508–15. doi: 10.1172/JCI40040

215. Mueller SN, Matloubian M, Clemens DM, Sharpe AH, Freeman GJ, Gangappa S, et al. Viral targeting of fibroblastic reticular cells contributes to immunosuppression and persistence during chronic infection. *Proc Natl Acad Sci U. S. A.* (2007) 104:15430–5. doi: 10.1073/pnas.0702579104

216. Kos K, Aslam MA, van de Ven R, Wellenstein MD, Pieters W, van Weverwijk A, et al. Tumor-educated Tregs drive organ-specific metastasis in breast cancer by impairing NK cells in the lymph node niche. *Cell Rep.* (2022) 38:110447. doi: 10.1016/j.celrep.2022.110447

217. Kawaida H, Kono K, Takahashi A, Sugai H, Mimura K, Miyagawa N, et al. Distribution of CD4+CD25high regulatory T-cells in tumor-draining lymph nodes in patients with gastric cancer. *J Surg Res.* (2005) 124:151–7. doi: 10.1016/j.jss.2004.10.004

218. Piersiala K, da Silva PFN, Lagebro V, Kolev A, Starkhammar M, Elliot A, et al. Tumour-draining lymph nodes in head and neck cancer are characterized by accumulation of CTLA-4 and PD-1 expressing Treg cells. *Transl Oncol.* (2022) 23:101469. doi: 10.1016/j.tranon.2022.101469

219. Axelrod ML, Cook RS, Johnson DB, Balko JM. Biological consequences of MHC-II expression by tumor cells in cancer. *Clin Cancer Res.* (2019) 25:2392–402. doi: 10.1158/1078-0432.CCR-18-3200

220. Beyaz S, Chung C, Mou H, Bauer-Rowe KE, Xifaras ME, Ergin I, et al. Dietary suppression of MHC class II expression in intestinal epithelial cells enhances intestinal tumorigenesis. *Cell Stem Cell*. (2021) 28(11):1922–35.e5. doi: 10.1016/j.stem.2021.08.007

221. Johnson AM, Bullock BL, Neuwelt AJ, Poczobutt JM, Kaspar RE, Li HY, et al. Cancer cell-intrinsic expression of MHC class II regulates the immune microenvironment and response to anti-PD-1 therapy in lung adenocarcinoma. *J Immunol.* (2020) 204(8):2295–307. doi: 10.1101/2020.01.09.900597

222. Neuwelt AJ, Kimball AK, Johnson AM, Arnold BW, Bullock BL, Kaspar RE, et al. Cancer cell-intrinsic expression of MHC II in lung cancer cell lines is actively restricted by MEK/ERK signaling and epigenetic mechanisms. *J Immunother. Cancer.* (2020) 8:1–12. doi: 10.1136/jitc-2019-000441

223. Izar B, Tirosh I, Stover EH, Wakiro I, Cuoco MS, Alter I, et al. A single-cell landscape of high-grade serous ovarian cancer. *Nat Med.* (2020) 26:1271–9. doi: 10.1038/s41591-020-0926-0

224. Johnson DB, Nixon MJ, Wang Y, Wang DY, Castellanos E, Estrada MV, et al. Tumor-specific MHC-II expression drives a unique pattern of resistance to immunotherapy via LAG-3/FCRL6 engagement. *JCI Insight.* (2018) 3(24):e120360. doi: 10.1172/jci.insight.120360

225. Rodig SJ, Gusenleitner D, Jackson DG, Gjini E, Giobbie-Hurder A, Jin C, et al. MHC proteins confer differential sensitivity to CTLA-4 and PD-1 blockade in untreated metastatic melanoma. *Sci Transl Med.* (2018) 10:1–14. doi: 10.1126/ scitranslmed.aar3342

226. Rouhani SJ, Eccles JD, Riccardi P, Peske JD, Tewalt EF, Cohen JN, et al. Roles of lymphatic endothelial cells expressing peripheral tissue antigens in CD4 T-cell tolerance induction. *Nat Commun.* (2015) 6:6771. doi: 10.1038/ncomms7771

227. Gkountidi AO, Garnier L, Dubrot J, Angelillo J, Harlé G, Brighouse D, et al. MHC class II antigen presentation by lymphatic endothelial cells in tumors promotes intratumoral regulatory T cell-suppressive functions. *Cancer Immunol Res.* (2021) 9 (7):748–64. doi: 10.1158/2326-6066.CIR-20-0784

228. Li CY, Park HJ, Shin J, Baik JE, Mehrara BJ, Kataru RP. Tumor-associated lymphatics upregulate MHC-II to suppress tumor-infiltrating lymphocytes. *Int J Mol Sci.* (2022) 23(21):13470. doi: 10.3390/ijms232113470

229. Elyada E, Bolisetty M, Laise P, Flynn WF, Courtois ET, Burkhart RA, et al. Cross-species single-cell analysis of pancreatic ductal adenocarcinoma reveals antigenpresenting cancer-associated fibroblasts. *Cancer Discovery*. (2019) 9:1102 LP – 1123. doi: 10.1158/2159-8290.CD-19-0094

230. Janssen LME, Ramsay EE, Logsdon CD, Overwijk WW. The immune system in cancer metastasis: Friend or foe? J. Immunother. *Cancer*. (2017) 5:1–14. doi: 10.1186/ s40425-017-0283-9

231. Ghiringhelli F, Ménard C, Terme M, Flament C, Taieb J, Chaput N, et al. CD4 +CD25+ regulatory T cells inhibit natural killer cell functions in a transforming growth factor-beta-dependent manner. *J Exp Med.* (2005) 202:1075–85. doi: 10.1084/ jem.20051511

232. Smyth MJ, Teng MWL, Swann J, Kyparissoudis K, Godfrey DI, Hayakawa Y. CD4+CD25+ T regulatory cells suppress NK cell-mediated immunotherapy of cancer. *J Immunol.* (2006) 176:1582–7. doi: 10.4049/jimmunol.176.3.1582

233. Francis DM, Manspeaker MP, Schudel A, Sestito LF, O'Melia MJ, Kissick HT, et al. Blockade of immune checkpoints in lymph nodes through locoregional delivery augments cancer immunotherapy. *Sci Transl Med.* (2020) 12:1–12. doi: 10.1126/ scitranslmed.aay3575

234. Sordo-Bahamonde C, Lorenzo-Herrero S, Gonzalez-Rodriguez AP, Martínez-Pérez A, Rodrigo JP, García-Pedrero JM, et al. Chemo-immunotherapy: A new trend in cancer treatment. *Cancers (Basel).* (2023) 15:1–15. doi: 10.3390/cancers15112912

235. Fear VS, Forbes CA, Neeve SA, Fisher SA, Chee J, Waithman J, et al. Tumour draining lymph node-generated CD8 T cells play a role in controlling lung metastases after a primary tumour is removed but not when adjuvant immunotherapy is used *Cancer Immunol Immunother*. (2021) 70:3249–58. doi: 10.1007/s00262-021-02934-3

236. Wang S, Li F, Ye T, Wang J, Lyu C, Qing S, et al. Macrophage-tumor chimeric exosomes accumulate in lymph node and tumor to activate the immune response and the tumor microenvironment. *Sci Transl Med.* (2021) 13(615):eabb6981. doi: 10.1126/scitranslmed.abb6981

237. Jeanbart L, Ballester M, de Titta A, Corthésy P, Romero P, Hubbell JA, et al. Enhancing efficacy of anticancer vaccines by targeted delivery to tumor-draining lymph nodes. *Cancer Immunol Res.* (2014) 2:436–47. doi: 10.1158/2326-6066.CIR-14-0019-T

238. Liu H, Moynihan KD, Zheng Y, Szeto GL, Li AV, Huang B, et al. Structurebased programming of lymph-node targeting in molecular vaccines. *Nature*. (2014) 507:519–22. doi: 10.1038/nature12978

239. Kim S-Y, Noh Y-W, Kang TH, Kim J-E, Kim S, Um SH, et al. Synthetic vaccine nanoparticles target to lymph node triggering enhanced innate and adaptive antitumor immunity. *Biomaterials*. (2017) 130:56–66. doi: 10.1016/j.biomaterials.2017.03.034

240. Ishihara M, Nishida Y, Kitano S, Kawai A, Muraoka D, Momose F, et al. A phase 1 trial of NY-ESO-1-specific TCR-engineered T-cell therapy combined with a lymph node-targeting nanoparticulate peptide vaccine for the treatment of advanced soft tissue sarcoma. *Int J Cancer*. (2023) 152:2554–66. doi: 10.1002/ijc.34453

241. Drakes DJ, Abbas AM, Shields J, Steinbuck MP, Jakubowski A, Seenappa LM, et al. Lymph node-targeted vaccine boosting of TCR T-cell therapy enhances antitumor function and eradicates solid tumors. *Cancer Immunol Res.* (2024) 12:214-31. doi: 10.1158/2326-6066.CIR-22-0978

242. Liu M, Feng Y, Lu Y, Huang R, Zhang Y, Zhao Y, et al. Lymph-targeted highdensity lipoprotein-mimetic nanovaccine for multi-antigenic personalized cancer immunotherapy. *Sci Adv.* (2024) 10:1–12. doi: 10.1126/sciadv.adk2444

243. Ribas A, Weber JS, Chmielowski B, Comin-Anduix B, Lu D, Douek M, et al. Intra-lymph node prime-boost vaccination against Melan A and tyrosinase for the treatment of metastatic melanoma: Results of a phase 1 clinical trial. *Clin Cancer Res.* (2011) 17:2987–96. doi: 10.1158/1078-0432.CCR-10-3272

244. Wang Y, Zhang L, Xu Z, Miao L, Huang L. mRNA vaccine with antigen-specific checkpoint blockade induces an enhanced immune response against established melanoma. *Mol Ther.* (2018) 26:420–34. doi: 10.1016/j.ymthe.2017.11.009

245. Chen J, Ye Z, Huang C, Qiu M, Song D, Li Y, et al. Lipid nanoparticle-mediated lymph node-targeting delivery of mRNA cancer vaccine elicits robust CD8+ T cell response. *Proc Natl Acad Sci U. S. A.* (2022) 119(34):e2207841119. doi: 10.1073/pnas.2207841119

246. Lian J, Ozga AJ, Sokol CL, Luster AD. Targeting lymph node niches enhances type 1 immune responses to immunization. *Cell Rep.* (2020) 31:107679. doi: 10.1016/j.celrep.2020.107679

247. Najibi AJ, Lane RS, Sobral MC, Bovone G, Kang S, Freedman BR, et al. Durable lymph-node expansion is associated with the efficacy of therapeutic vaccination. *Nat Biomed Eng.* (2024) 6. doi: 10.1038/s41551-024-01209-3

248. Yoo GS, Ahn WG, Kim SY, Kang W, Choi C, Park HC. Radiation-induced abscopal effect and its enhancement by programmed cell death 1 blockade in the hepatocellular carcinoma: A murine model study. *Clin Mol Hepatol.* (2021) 27:144–56. doi: 10.3350/cmh.2020.0095

249. Buchwald ZS, Nasti TH, Lee J, Eberhardt CS, Wieland A, Im SJ, et al. Tumordraining lymph node is important for a robust abscopal effect stimulated by radiotherapy. *J Immunother. Cancer.* (2020) 8(2):e000867. doi: 10.1136/jitc-2020-000867

250. Liu Z, Yu Z, Chen D, Verma V, Yuan C, Wang M, et al. Pivotal roles of tumordraining lymph nodes in the abscopal effects from combined immunotherapy and radiotherapy. *Cancer Commun.* (2022) 42:971–86. doi: 10.1002/cac2.12348

251. Telarovic I, Yong CSM, Kurz L, Vetrugno I, Reichl S, Fernandez AS, et al. Delayed tumor-draining lymph node irradiation preserves the efficacy of combined radiotherapy and immune checkpoint blockade in models of metastatic disease. *Nat Commun.* (2024) 15:1–23. doi: 10.1038/s41467-024-49873-y

252. Song E, Mao T, Dong H, Boisserand LSB, Antila S, Bosenberg M, et al. VEGF-C-driven lymphatic drainage enables immunosurveillance of brain tumours. *Nature*. (2020) 577:689-94. doi: 10.1038/s41586-019-1912-x

253. Morton DL, Thompson JF, Cochran AJ, Mozzillo N, Nieweg OE, Roses DF, et al. Final trial report of sentinel-node biopsy versus nodal observation in melanoma. *N Engl J Med.* (2014) 370:599–609. doi: 10.1056/NEJMoa1310460

254. Faries MB, Thompson JF, Cochran AJ, Andtbacka RH, Mozzillo N, Zager JS, et al. Completion dissection or observation for sentinel-node metastasis in melanoma. *N Engl J Med.* (2017) 376(23):2211–22. doi: 10.1056/NEJMoa1613210

255. Veronesi U, Marubini E, Mariani L, Valagussa P, Zucali R. The dissection of internal mammary nodes does not improve the survival of breast cancer patients. 30-year results of a randomised trial. *Eur J Cancer*. (1999) 35:1320–5. doi: 10.1016/S0959-8049(99)00133-1

256. Galimberti V, Cole BF, Zurrida S, Viale G, Luini A, Veronesi P, et al. Axillary dissection versus no axillary dissection in patients with sentinel-node micrometastases

(IBCSG 23-01): a phase 3 randomised controlled trial. *Lancet Oncol.* (2013) 14(4):297–305. doi: 10.1016/S1470-2045(13)70035-4

257. Gentilini OD, Botteri E, Sangalli C, Galimberti V, Porpiglia M, Agresti R, et al. Sentinel lymph node biopsy vs no axillary surgery in patients with small breast cancer and negative results on ultrasonography of axillary lymph nodes: the SOUND randomized clinical trial. *JAMA Oncol.* (2023) 9(11):1557–64. doi: 10.1001/ jamaoncol.2023.3759

258. Wong SL, Faries MB, Kennedy EB, Agarwala SS, Akhurst TJ, Ariyan C, et al. Sentinel lymph node biopsy and management of regional lymph nodes in Melanoma: American society of clinical oncology and society of surgical oncology clinical practice guideline update. *J Clin Oncol.* (2018) 36:399–413. doi: 10.1200/JCO.2017.75.7724