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# Targeting STING signaling for the optimal cancer immunotherapy

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Despite the transformative impact of anti-PD-1/PD-L1 therapies, challenges such as low response rates persist. The stimulator of interferon genes (STING) pathway, a crucial element of innate immunity, emerges as a strategic target to overcome these limitations. Understanding its multifaceted functions in cancer, including antigen presentation and response to DNA damage, provides valuable insights. STING agonists, categorized into cyclic dinucleotides (CDNs) and non-CDNs, exhibit promising safety and efficacy profiles. Innovative delivery systems, including antibody-drug conjugates, nanocarriers, and exosome-based therapies, address challenges associated with systemic administration and enhance targeted tumor delivery. Personalized vaccines, such as DT-Exo-STING, showcase the adaptability of STING agonists for individualized treatment. These advancements not only offer new prospects for combination therapies but also pave the way for overcoming resistance mechanisms. This review focuses on the potential of targeting STING pathway to enhance cancer immunotherapy. The integration of STING agonists into cancer immunotherapy holds promise for more effective, personalized, and successful approaches against malignancies, presenting a beacon of hope for the future of cancer treatment.

KEYWORDS

STING, PD-1, PD-L1, cancer immunotherapy, the tumor microenvironment

# 1 Background

In recent years, immunotherapy, with a particular focus on anti-PD-1/PD-L1 antibodies, has emerged as a groundbreaking paradigm in cancer treatment (1, 2). The remarkable advances in this field have revolutionized the therapeutic landscape, harnessing the immune system's potential to combat malignancies. The introduction of anti-PD-1/PD-L1 therapies has marked a significant stride forward, offering a promising avenue for more effective and targeted cancer interventions (3–11). However, despite the promise and success observed in some cases, the clinical utility of anti-PD-1/PD-L1 therapies faces substantial challenges, primarily characterized by a low response rate among patients (12–14). This limitation underscores the need for a comprehensive understanding of the factors influencing treatment outcomes. The intricacies of the tumor microenvironment (TME) play a critical role in shaping the efficacy of anti-PD-1/PD-L1 therapies (15). Within the TME, a myriad of immunosuppressive factors acts synergistically to impede the optimal function of these therapies (16–19).

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Multiple hurdles within the TME contribute to the suboptimal response rates observed with anti-PD-1/PD-L1 antibodies (20). These obstacles include defects in immune checkpoint signaling, the accumulation of immunosuppressive cells, and antigen presentation deficiency (21). The dynamic interplay of these factors underscores the complexity of the TME and its role in modulating tumor progression (22-24). Recognizing the challenges posed by the multifaceted immunosuppressive factors, there is growing interest in exploring novel strategies to enhance the efficacy of immunotherapy. One such promising avenue is the targeting of the stimulator of interferon genes (STING) pathway (25). The STING pathway, an integral component of the innate immune system, is implicated in recognizing cellular stress and infection (26-29). Recent findings suggest that manipulating the STING pathway could offer a means to overcome the limitations associated with anti-PD-1/PD-L1 therapies (30, 31).

Understanding the interplay between immunotherapy and the intricate dynamics of the TME is crucial for advancing cancer treatment strategies (32). This review aims to summarize the advances and limitations of anti-PD-1/PD-L1 immunotherapies, unravel the complexities of immunosuppressive factors within the TME, and explore the potential of targeting the STING pathway as a strategic approach to augment the efficacy of immunotherapy. By elucidating these aspects, we aim to contribute valuable insights for more effective and personalized cancer treatment modalities.

#### 2 The cGAS-STING signaling pathway

The cGAS-STING signaling pathway, a prominent cytosolic DNA-sensing mechanism, stands as a cornerstone in the innate immune system, orchestrating responses against pathogens (33–36). Beyond its primary role in pathogen recognition, this pathway intricately regulates a spectrum of cellular functions, spanning from the induction of antiviral interferon responses, cytokine production, autophagy, metabolism, senescence, metastasis to apoptosis (37–45). At the heart of this pathway lies the STING, an endoplasmic reticulum-associated transmembrane protein activated by the endogenous cyclic dinucleotide (CDN) second messenger, 2'3'-cGAMP, produced by the enzyme cGAMP synthase (cGAS) upon binding to cytosolic DNA (46). STING's activation initiates a cascade involving downstream effectors TANK-binding kinase-1 (TBK1) and IFN regulatory factor-3 (IRF3), resulting in robust innate immune responses (47).

The STING-TBK1-IRF3 signaling transduction pathway emerges as a pivotal element in the cGAS-STING cascade, translating STING activation into effective immune responses (Figure 1). Activated STING undergoes conformational changes, engaging TBK1 and IRF3 (48). TBK1, forming a homodimer, interacts with two STING molecules, inducing a conformational shift that releases the C-terminal tail of STING (49). This enables recruitment and activation of TBK1, leading to phosphorylation of serine 366 of the STING C-terminal tail. Phosphorylated IRF3 undergoes homodimerization, translocates to the nucleus, and triggers the transcription of type I interferon (IFN) and other interferon-stimulated genes (ISGs) (50, 51). The STING-TBK1-IRF3 axis acts as a critical link in antiviral defense, orchestrating IFN production to curb viral replication and spread. Importantly, the STING-TBK1-IRF3 pathway also intersects with canonical NF- $\kappa$ B signaling, involving the NF- $\kappa$ B subunit p65 (52). Although the precise mechanisms of STING-induced NF-κB activation are under investigation, it seems to be independent of ER-to-Golgi trafficking (49). This intricate crosstalk adds another layer of complexity to the diverse cellular functions regulated by the cGAS-STING pathway (53). Therapeutic activation of STING to boost immune responses may inadvertently overactivate NF-KB, leading to harmful inflammation (54). Combining STING agonists with inhibitors of negative regulators in the NF-KB pathway may enhance anti-tumor immunity while controlling inflammation (55). Modulating the crosstalk can help in designing therapies that suppress pathological immune responses without compromising host defense. Understanding individual variations in the STING and NF-KB pathways can aid in predicting patient responses and tailoring treatments. Additionally, STING is involved in extensive crosstalk with various other immune associated signaling pathways, referred as non-canonical STING pathway (40, 56-58). Developing drugs that specifically target components of the crosstalk may provide therapeutic benefits with fewer side effects.

The intricate regulation of the cGAS-STING pathway is paramount, as aberrant activation may lead to severe autoinflammatory or autoimmune diseases (59–61). Recent advances have illuminated the mechanisms of STING activation and the meticulous regulation of this pathway to prevent excessive signaling (62). Furthermore, the cGAS-STING pathway assumes a pivotal role in antimicrobial immunity, influencing responses to diverse pathogens, including viruses and bacteria, thereby shaping immune regulation, and offering potential avenues for therapeutic interventions (63).

# 3 The role of STING signaling in cancer

The STING signaling has been believed to be a central player in the dynamic landscape of cancer immunology, with a particular emphasis on its intricate involvement in cancer antigen presentation (64–66). While the pro-inflammatory role of IFN signaling has fueled interest in STING as a mediator of effective antitumor immunity, recent insights reveal the multifaceted functions of this pathway in cancer, demanding careful contextual consideration (67). A unique feature of cancer is chromosomal instability, marked by accumulated chromosome mis-segregation during mitosis (68). This results in the formation of micronuclei, rupturing during S-phase, exposing genomic double-stranded DNA (dsDNA) to the cytoplasm (69–71). The chronic activation of the

Abbreviations: TME, the tumor microenvironment; STING, stimulator of interferon genes; CDN, cyclic dinucleotide; cGAS, cGAMP synthase; TBK1, TANK-binding kinase-1; IRF3, IFN regulatory factor-3; ISG, interferon-stimulated gene; cGAMP, cyclic GMP-AMP; NK, natural killer cell; ORR, overall response rate; ADC, antibody-drug conjugate.



The STING signaling pathway. Activated STING undergoes conformational changes, engaging TBK1 and IRF3. TBK1, forming a homodimer, interacts with two STING molecules, inducing a conformational shift that releases the C-terminal tail of STING. This enables recruitment and activation of TBK1, leading to phosphorylation of serine 366 of the STING C-terminal tail. Phosphorylated IRF3 undergoes homodimerization, translocates to the nucleus, and triggers the transcription of type I interferon (IFN) and other interferon-stimulated genes (ISGs). The STING-TBK1-IRF3 axis acts as a critical link in antiviral defense, orchestrating IFN production to curb viral replication and spread. Importantly, the STING-TBK1-IRF3 pathway also intersects with canonical NF-κB signaling (Created with Biorender).

cGAS-STING pathway in cancers exhibiting rampant chromosome instability is further compounded by DNA damage induced by radiation therapy, chemotherapeutic agents, and mitochondrial DNA leakage due to oxidative stress (72–78). Nevertheless, tumors exhibit a remarkable ability to regulate the expression of STING pathway genes to evade its antitumor and pro-inflammatory effects. Loss of chromosome 9p, harboring the IFN-gene cluster, is common in certain cancers, allowing these tumors to signal through the NF- $\kappa$ B pathway without inducing IFN response (79). Other evasion mechanisms involve the downregulation of STING levels, observed in various cancers (80).

Moreover, tumor-derived cyclic GMP-AMP (cGAMP), a critical activator of the cGAS-STING pathway, can be derived from cancer cells and transferred into neighboring cells, directly activating STING in the TME (81). This transfer occurs through various mechanisms, including import through cell gap junctions, cGAMP importer SLC19A1, connexin, and exosomes (81–84). The delivery of cGAMP or dsDNA to non-tumor cells has been shown to exert both antitumor effects and promote tumor progression, underscoring the intricate interplay between the STING pathway and the TME

(49). For instance, the STING pathway in dendritic cells (DCs) plays a vital role by taking up tumor-derived DNA, leading to increased type I IFN expression (85). This enhances DC cross-presentation, survival, lymph node homing, and the expression of Th1 chemokines, which are crucial for immune cell trafficking (Figure 2) (86–88). STING inhibition in DCs impairs antigen presentation and reduces tumor-infiltrating lymphocytes (TIL) (74).

Furthermore, tumor-derived cGAMP also plays a pivotal role in natural killer (NK) cell activation. In mouse models, persistent cGAS activation in cancer cells, followed by paracrine cGAMP uptake by neighboring NK cells, enhances IFN-I signaling and antitumor immunity (89). Despite initial enthusiasm for the use of STING agonists in anti-cancer therapy, challenges in translating results from mouse models to humans have surfaced. The efficacy of STING agonists, designed to boost antitumor immunity, has been limited in humans, suggesting that tumors evolve mechanisms to undermine IFN-I signaling (90). These complexities highlight the need for a nuanced understanding of the cGAS-STING pathway in cancer immunotherapy, considering factors such as tumor type, dose-dependent effects, and the dynamic nature of the TME.



#### FIGURE 2

The STING signaling in dendritic cells (DCs). The STING pathway in DCs plays a vital role by taking up tumor-derived DNA, leading to increased type I IFN expression. This enhances DC cross-presentation, survival, lymph node homing, and the expression of Th1 chemokines, crucial for immune cell trafficking (Created with Biorender).

# 4 The application of STING agonist in cancer immunotherapy

The evolving field of cancer immunotherapy has witnessed remarkable progress with the emergence of STING agonists, which hold immense potential in harnessing the immune system to combat tumors (91). Here, we summarize the diverse landscape of STING agonists, categorizing them into two groups: CDNs and non-CDNs.

#### 4.1 CDNs

The early forays into CDNs introduced compounds like DMXAA and ADU-S100. Despite DMXAA's limited translational success in human trials due to its weak binding to human STING protein (92), ADU-S100 has emerged as a promising candidate (93–95). In clinical trials, ADU-S100 demonstrated safety and elicited encouraging responses, especially when combined with checkpoint inhibitors. In the phase I trial NCT02675439, ADU-S100 exhibited remarkable safety and tolerability in patients with advanced cancers (96). Despite limited clinical activity as a single agent, noteworthy treatment responses included a partial response in Merkel cell carcinoma and two unconfirmed partial responses in parotid cancer and myxofibrosarcoma (96). Besides, in the phase Ib doseescalation study NCT03172936, the safety and tolerability of

combining ADU-S100 with the PD-1 inhibitor Spartalizumab were evaluated in 106 patients with advanced solid tumors or lymphomas. Administered through weekly intratumoral injections of ADU-S100 and a fixed dose of intravenous spartalizumab (400 mg) every four weeks, the combination demonstrated favorable safety profiles, with common adverse events including pyrexia, injection site pain, and diarrhea (97). Despite minimal antitumor responses (ORR: 10.4%), the study highlighted the feasibility of this combination, even in patients with anti-PD-1 refractory disease, warranting further exploration and optimization in the ongoing quest for effective cancer immunotherapies (97).

Moreover, MK-1454, a synthetic CDN, has showcased notable antitumor activity in phase I trials, particularly when administered in combination with the anti-PD-1 agent pembrolizumab (98, 99). Besides, JNJ-4412, a novel CDN STING agonist, demonstrated efficacy in preclinical models, highlighting its potential through intratumoral injection. The results from these preclinical studies suggest that JNJ-4412 holds promise as a potent STING agonist, potentially contributing to the arsenal of therapeutic options for cancer treatment (100). Furthermore, the exploration of novel STING agonists for systemic delivery unveils a spectrum of promising candidates. SB11285, a small molecule CDN STING agonist, is currently undergoing clinical evaluation for intravenous administration (101, 102). Preclinical models have demonstrated its higher inhibition of tumor growth, hinting at its potential systemic benefits in cancer patients (101). Additionally, Sun et al. present a cancer metalloimmunotherapy prototype employing CDN STING agonists and Mn<sup>2+</sup> ions assembled into a nanoparticle (CDN-Mn<sup>2</sup> <sup>+</sup>particle, CMP). This modality administered via either intratumoral injection or systemic intravenous injection, elicited potent antitumor immune responses and demonstrated remarkable therapeutic efficacy with abridged doses of STING agonists in various preclinical models (103).

#### 4.2 Non-CDN STING agonists

The non-CDN category features innovative compounds with unique structures and mechanisms. SNX281, another small molecule agent, is being investigated in a phase I dose escalation study for advanced solid tumors (104, 105). Also, BMS-986301, exhibiting robust antitumor efficacy pre-clinically, has transitioned into clinical trials, reflecting its potential as an effective systemic STING agonist (106). The development of CRD-5500 showcases its effectiveness through both intravenous and intratumoral routes in murine models. This versatility positions CRD-5500 as a favorable candidate for future clinical development, offering flexibility in therapeutic administration (104, 107). Moreover, TTI-10001 has demonstrated safety and antitumor activity in preclinical models (108). Besides, ALG-031048 exhibited higher stability compared to ADU-S100 and demonstrated significant tumor regression in preclinical models. Its dose-dependent increase in cytokine levels and enhanced antitumor efficacy, in combination with anti-PD1 therapy position ALG-031048, make it a promising candidate for further clinical exploration (109).

Notably, some novel non-CDN compounds exhibit potential for systemic delivery, with enhanced stability, opening avenues for broader application in cancer immunotherapy. For example, identified through a screen for IFN-B secretion inducers, MSA-2 operates through a unique mechanism, binding to both human and mouse STING as a noncovalent dimer. Administered orally or subcutaneously, MSA-2 induced elevations of interferon- $\beta$  in plasma and tumors, demonstrating well-tolerated regimens and prompting tumor regressions in mice with MC38 syngeneic tumors (110). Structural analyses unveiled MSA-2's binding as a noncovalent dimer to STING in a "closed-lid" conformation, shedding light on its mechanism of action. MSA-2's ability to preferentially activate STING in tumors positions it as a promising candidate for developing human STING agonists that are amenable to systemic administration in patients (110). In the later preclinical studies, MSA-2 synergized with other immunotherapies, such as anti-TGF-β/PD-L1 bispecific antibodies, to overcome immunotherapy resistance in murine tumor models (111, 112). Also, unlike current efforts focused on modified cyclic dinucleotides for intratumoral delivery, ABZI demonstrated systemic efficacy in treating tumors (113). Developed through a linking strategy, di-ABZIs exhibit enhanced binding to STING and cellular function (113). Intravenous injection of di-ABZI STING agonist in mice with syngeneic colon tumors resulted in potent antitumor activity, leading to complete and lasting tumor regression (113). This milestone marks a significant advancement in the quest for effective immune-modulating cancer treatments (113). Additionally, JNJ-6196 also demonstrated systemic efficacy, positioning JNJ-6196 as a compelling candidate for clinical development (114).

# 4.3 STING agonists with novel delivery systems

To overcome safety concerns with systemic STING agonist administration and limited accessibility with intratumoral injection, antibody-drug conjugates (ADCs) were developed. These ADCs, combining a STING agonist with tumor-targeting antibodies, demonstrated well-tolerated systemic administration and potent antitumor efficacy in mouse models. For instance, the anti-EGFR-172 ADC demonstrates the feasibility of delivering a STING agonist selectively to tumors (115). The adaptability of IMSA172 for conjugation with various antibodies and tumor-targeting agents opens avenues for exploring different ADCs, unveiling their safety and efficacy in activating STING across diverse tumor types through systemic delivery (115). Besides, Duvall et al. developed a STING agonist ADC platform to address the translational challenges of STING agonists in the clinic (116). This platform, featuring a potent non-cyclic dinucleotide STING agonist, a cleavable esterbased linker, and a hydrophilic PEG8-bisglucamine scaffold, exhibits robust and durable antitumor activity, high stability, and favorable pharmacokinetics in nonclinical species, showcasing its potential for systemic administration and localized STING activation within tumors for enhanced therapeutic efficacy and tolerability (116).

Furthermore, various nanocarriers are promising for the delivery of STING agonists. Zhou developed a nanovaccine to resolve the difficulty in delivering mRNA and nucleic acid drugs. The nanovaccine demonstrated the activation of potent antitumor immune response and long-term immune memory by transferring mRNA antigen and cGAMP (117). Optimization of key parameters, such as modifying the PBA moiety and utilizing the anionic Lipo-ORG for lymphatic delivery, resulted in suppressed tumor growth and metastasis, extended survival, and synergistic effects with PD-L1 blockade in a B16-OVA tumor model (117). Besides, some novel pH-responsive DNA nanovaccines, featuring PLA-b-PEG in the core and pH-responsive i-motif DNA on the surface, efficiently load and release CDG in immune cell endosomes, promoting potent antitumor immune responses, overcoming immunosuppression, and demonstrating superior efficacy in a murine melanoma model compared to liposomal CDG and fluoride-CDG (118). In addition, Gu et al. designed a novel antigen-inspired MnO2 nanovaccine, serving as a  $\mathrm{Mn}^{2+}$  source and functionalized with mannose for specific delivery to innate immune cells (119). This nanovaccine activated the STING pathway, enhancing radiotherapy-induced immune responses and inhibiting both local and distant tumors, while also allowing for magnetic resonance imaging to monitor in vivo distribution (119). Actually, there are many STING-activating cancer vaccines exhibiting potent antitumor activity in preclinical studies, including STINGVAX, CDN/neoantigen co-delivering nanovaccines, PC7A, and selfdegradable poly( $\beta$ -amino ester)s (120–123).

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Finally, exosome-based therapies leverage cell-derived nanovesicles to deliver STING agonists to tumors. Cheng et al. designed multifunctional hybrid exosomes to activate the cGAS-STING pathway (124). These exosomes were created by merging genetically engineered exosomes carrying CD47 from tumor cells with those from M1 macrophages, encapsulating them with a DNAtargeting agent (SN38) and a STING agonist (MnO2) (124). The hybrid exosomes exhibited excellent tumor-targeting capabilities and prolonged circulation time, inducing polarization of tumorassociated macrophages to the M1 phenotype, releasing SN38 to cause DNA damage, and stimulating cGAS/STING activation with Mn<sup>2+</sup> at the tumor site. This multifunctional approach promoted DC maturation, facilitated cytotoxic T lymphocyte infiltration, and recruited natural killer cells to the tumor region, resulting in significant antitumor and antimetastatic efficacy (124). Notably, Liu et al. explored Artemisia annua, a plant recognized for its antimalarial properties, and isolated exosome-like particles termed artemisia-derived nanovesicles (ADNVs) (125). These nanoscaled vesicles exhibited the remarkable ability to inhibit tumor growth and enhance antitumor immunity in a lung cancer mouse model. The key player identified within these vesicles was plantderived mitochondrial DNA (mtDNA), which, upon internalization by tumor-associated macrophages, activated the cGAS-STING pathway, shifting pro-tumor macrophages to an antitumor phenotype (125). Additionally, administration of ADNVs significantly improved the effectiveness of a PD-L1 inhibitor, showcasing the potential of this inter-kingdom interaction to stimulate immunostimulatory signaling and bolster antitumor immunity (125). Besides, in the work of Bao et al., DC-tumor hybrid cell-derived chimeric exosomes loaded with STING agonists (DT-Exo-STING) were engineered to address the challenge of balancing antigen-enriched delivery and optimal antigenpresentation functionality in DCs (126). These chimeric carriers, equipped with broad-spectrum antigen complexes, induce a potent T-cell response through both direct self-presentation and indirect DC-to-T immune stimulation (126). The nanovaccine-driven STING activation not only surpassed conventional CDN delivery methods in tissue-homing capacity, including penetration of the blood-brain barrier, but also ensured efficient cytosolic entry for activating STING signaling (126). This strategy not only improves antigen presentation but also transforms immunosuppressive TME into a pro-inflammatory state, resulting in a significant reduction of intracranial primary lesions (126). Moreover, the personalized DT-Exo-STING vaccines, utilizing autologous tumor tissues, enhance sensitivity to ICB and establish systemic immune memory against cancer recurrence. These findings offer a promising avenue for glioblastoma immunotherapy, with potential implications for further exploration in clinical applications (126).

However, as alluded earlier, the potent activation of the innate immune system by using STING agonists can lead to several side effects that are critical to consider in clinical settings. STING activation leads to the production of type I interferons and proinflammatory cytokines. While beneficial in fighting tumors and infections, this can result in systemic inflammation, causing symptoms like fever, chills, and fatigue. Overstimulation of the immune system may also trigger autoimmune reactions. The heightened immune activity can cause the body to attack its own tissues, potentially leading to conditions such as lupus or rheumatoid arthritis. Furthermore, STING agonists occasionally elicit Cytokine Release Syndrome (CRS), also known as a "cytokine storm," is a severe immune reaction characterized by the rapid release of large amounts of cytokines (127). This can lead to organ dysfunction and is a serious concern with immunotherapies. Hence, reaching the next milestone for oncologists is to concurrently diminish and even eliminate side effects, while enhancing immune efficiency, prolonging lifespan, and improving patients' quality of life.

## 5 Perspective and conclusion

In conclusion, the landscape of cancer immunotherapy has evolved significantly with the emergence of STING signaling as a promising target. The limitations associated with anti-PD-1/PD-L1 therapies, including low response rates attributed to the intricate dynamics of the TME, have prompted exploration into novel strategies. The STING pathway, a central player in innate immunity, presents a unique opportunity to meet these challenges and improve immunotherapy efficacy. Understanding the role of the STING pathway in cancer immunology has revealed its intricate involvement in cancer antigen presentation, response to DNA damage, and modulation of the TME. Despite the complexities and evasion mechanisms exhibited by tumors, targeting the STING pathway holds the potential to influence diverse aspects of the immune attack against cancer.

The application of STING agonists in cancer immunotherapy has seen remarkable progress, with a diverse landscape of CDNs and non-CDNs. CDNs, such as ADU-S100, MK-1454, and JNJ-4412, have demonstrated safety and efficacy in clinical trials, particularly when combined with checkpoint inhibitors (Table 1). Non-CDN STING agonists, like MSA-2 and di-ABZI, showcase unique structures and mechanisms, expanding the options for therapeutic interventions. Moreover, the development of STING agonists with novel delivery systems, including ADCs, nanocarriers, and exosome-based therapies, addresses safety concerns and enhances the potential for systemic administration. These innovative approaches demonstrate the adaptability of STING agonists for selective tumor targeting, promoting antitumor immune responses, and overcoming challenges in drug delivery. These advancements open avenues for exploring combination therapies, overcoming resistance mechanisms, and improving the overall efficacy of cancer immunotherapy. In spite of the generalized advertisement in scale of immunotherapy, presenting problems, drawbacks and frustrations still thwart their feasibility and accessibility to fide bona clinical employment. In summary, pharmacokinetic challenges, escalating adverse effects, suppressive tumor microenvironment through alternative pathways, and genetic variability individually or collectively contribute to the current unfavorable state (76, 128). More efforts are warranted to circumvent these hurdles. Furthermore, the implementation of combinative treatment strategies involving STING agonists and other antitumor agents, such as PARP inhibitors and chemotherapies other than ICBs represents a novel endeavor to overcome cancer (129, 130).

Agents Terming	NCT numbers	Indications	Phase	Cardinal observation indications	Schedule and Outcomes
KL340399	NCT05549804/ NCT05387928	Advanced Solid Tumors	1	Tolerability and RP2D	Not referred
E7766	NCT04109092	Urinary Bladder Neoplasms	1	Tolerability and CRR	Not referred
	NCT04144140	Lymphoma, Advanced Solid Tumors	1	Tolerability and ORR	Not referred
MIW815/ ADU-\$100	NCT03937141	Head and Neck Cancer	2	ORR	Intratumoral injection (800 µg per lesion day 1 and 8 of a 21-days cycle) + Pembrolizumab; 4/8 reaching PR; 1/8 reaching SD; 3/ 8 reaching PD
	NCT02675439	Lymphoma, Advanced/Metastatic Solid Tumors	1	Tolerability and RP2D	Intratumoral injection (50 to 6,400 µg weekly, on a 3-weeks-on/1-week- off schedule); 94% of lesions reaching stable or decrease with systemic immune activity
	NCT03172936	Lymphoma, Advanced/Metastatic Solid Tumors	1	Tolerability	Intratumoral injection (50-800 µg) either weekly (3 weeks on/1 week off) or Q4W + PDR001; The regimen being tolerable; ORR=10.4%
MK-1454	NCT04220866	Head and Neck Cancer	2	ORR	Intratumoral injection (540 ug on Day 1 of every week for two 3-week then on Day 1 of each 3-week cycle for up 33 cycles + Pembrolizumab; ORR = 50%; PFS = 6.4 months
	NCT03010176	Lymphoma, Advanced/Metastatic Solid Tumors	1	Tolerability	Intratumoral injection (10-3000 ug, and 90-1500 ug Q1W*9 for 3cycles and beyond for up to 35 cycles for Arm 1, and 2 respectively), Arm 2 was combined with Pembrolizumab; Regimen being tolerable and PR = 24% (Arm 2), 0 (Arm 1); DCR = 48% (Arm 2), 20% (Arm 1)
CRD3874-SI	NCT06021626	Sarcoma, Merkel Cell Carcinoma	1	Tolerability and ORR	Not referred
TXN10128	NCT05978492	Solid Tumors	1	Tolerability	Not referred
GSK3745417	NCT03843359	Solid Tumors	1	Tolerability	Not referred
	NCT05424380	Hematologic malignancies	1	Tolerability and ORR	Not referred
IMSA101	NCT05846659/ NCT05846646	Solid Tumors	2	Progression-free rate	Not referred
SNX281	NCT04609579	Lymphoma, Advanced Solid Tumors	1	Tolerability and RP2D	Not referred
ONM-501	NCT06022029	Lymphoma, Advanced Solid Tumors	1	Tolerability	Not referred
TAK-500	NCT05070247	Solid Tumors	1	Tolerability and ORR	Not referred

#### TABLE 1 The ongoing clinical using STING agonists in cancer immunotherapy regimen.

RP2D, Recommended Phase 2 Dose; CRR, Complete Response Rate; ORR, Objective Response Rate; PR, Partial Response; SD, Stable Disease; PD, Progression Disease.

In summary, targeting the STING signaling pathway represents a promising strategy for optimizing cancer immunotherapy. As research in this field continues to unravel the complexities of the STING pathway and its interactions within the TME, the potential for innovative and effective treatment modalities grows. The integration of STING agonists into the evolving landscape of cancer immunotherapy offers hope for more personalized and successful approaches to combat malignancies.

## Author contributions

YaX: Visualization, Project administration, Methodology, Investigation, Writing – original draft, Resources. YiX: Supervision, Conceptualization, Writing – review & editing, Resources.

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### References

1. Gong J, Chehrazi-Raffle A, Reddi S, Salgia R. Development of PD-1 and PD-L1 inhibitors as a form of cancer immunotherapy: a comprehensive review of registration trials and future considerations. *J Immunother Cancer*. (2018) 6:8. doi: 10.1186/s40425-018-0316-z

2. Raval RR, Sharabi AB, Walker AJ, Drake CG, Sharma P. Tumor immunology and cancer immunotherapy: summary of the 2013 SITC primer. J Immunother Cancer. (2014) 2:14. doi: 10.1186/2051-1426-2-14

3. Cercek A, Lumish M, Sinopoli J, Weiss J, Shia J, Lamendola-Essel M, et al. PD-1 blockade in mismatch repair-deficient, locally advanced rectal cancer. *N Engl J Med.* (2022) 386:2363–76. doi: 10.1056/NEJMoa2201445

4. Topalian SL, Hodi FS, Brahmer JR, Gettinger SN, Smith DC, McDermott DF, et al. Safety, activity, and immune correlates of anti-PD-1 antibody in cancer. *N Engl J Med.* (2012) 366:2443–54. doi: 10.1056/NEJMoa1200690

5. Olson DJ, Eroglu Z, Brockstein B, Poklepovic AS, Bajaj M, Babu S, et al. Pembrolizumab plus ipilimumab following anti-PD-1/L1 failure in melanoma. J Clin Oncol. (2021) 39:2647-55. doi: 10.1200/jcc.21.00079

6. VanderWalde A, Bellasea SL, Kendra KL, Khushalani NI, Campbell KM, Scumpia PO, et al. Ipilimumab with or without nivolumab in PD-1 or PD-L1 blockade refractory metastatic melanoma: a randomized phase 2 trial. *Nat Med.* (2023) 29:2278–85. doi: 10.1038/s41591-023-02498-y

7. Ansell SM, Lesokhin AM, Borrello I, Halwani A, Scott EC, Gutierrez M, et al. PD-1 blockade with nivolumab in relapsed or refractory Hodgkin's lymphoma. *N Engl J Med.* (2015) 372:311–9. doi: 10.1056/NEJMoa1411087

8. Hamid O, Robert C, Daud A, Hodi FS, Hwu WJ, Kefford R, et al. Safety and tumor responses with lambrolizumab (anti-PD-1) in melanoma. *N Engl J Med.* (2013) 369:134–44. doi: 10.1056/NEJMoa1305133

9. Shi J, Liu J, Tu X, Li B, Tong Z, Wang T, et al. Single-cell immune signature for detecting early-stage HCC and early assessing anti-PD-1 immunotherapy efficacy. *J Immunother Cancer*. (2022) 10:e003133. doi: 10.1136/jitc-2021-003133

10. Zhang W, Tong S, Hu B, Wan T, Tang H, Zhao F, et al. Lenvatinib plus anti-PD-1 antibodies as conversion therapy for patients with unresectable intermediateadvanced hepatocellular carcinoma: a single-arm, phase II trial. *J Immunother Cancer*. (2023) 11:e007366. doi: 10.1136/jitc-2023-007366

11. Lin Z, Cai M, Zhang P, Li G, Liu T, Li X, et al. Phase II, single-arm trial of preoperative short-course radiotherapy followed by chemotherapy and camrelizumab in locally advanced rectal cancer. *J Immunother Cancer*. (2021) 9:e003554. doi: 10.1136/ jitc-2021-003554

12. Kwon M, An M, Klempner SJ, Lee H, Kim KM, Sa JK, et al. Determinants of response and intrinsic resistance to PD-1 blockade in microsatellite instability-high gastric cancer. *Cancer Discovery*. (2021) 11:2168–85. doi: 10.1158/2159-8290.Cd-21-0219

13. Ribas A, Medina T, Kirkwood JM, Zakharia Y, Gonzalez R, Davar D, et al. Overcoming PD-1 blockade resistance with cpG-A toll-like receptor 9 agonist vidutolimod in patients with metastatic melanoma. *Cancer Discovery.* (2021) 11:2998–3007. doi: 10.1158/2159-8290.Cd-21-0425

14. Kluger HM, Tawbi HA, Ascierto ML, Bowden M, Callahan MK, Cha E, et al. Defining tumor resistance to PD-1 pathway blockade: recommendations from the first meeting of the SITC Immunotherapy Resistance Taskforce. *J Immunother Cancer*. (2020) 8:e000398. doi: 10.1136/jitc-2019-000398

15. Liu X, Hogg GD, DeNardo DG. Rethinking immune checkpoint blockade: 'Beyond the T cell'. *J Immunother Cancer*. (2021) 9:e001460. doi: 10.1136/jitc-2020-001460

16. Pu Y, Ji Q. Tumor-associated macrophages regulate PD-1/PD-L1 immunosuppression. *Front Immunol.* (2022) 13:874589. doi: 10.3389/fimmu.2022.874589

17. Fukumura D, Kloepper J, Amoozgar Z, Duda DG, Jain RK. Enhancing cancer immunotherapy using antiangiogenics: opportunities and challenges. *Nat Rev Clin Oncol.* (2018) 15:325–40. doi: 10.1038/nrclinonc.2018.29

18. Falcomatà C, Bärthel S, Schneider G, Rad R, Schmidt-Supprian M, Saur D. Context-specific determinants of the immunosuppressive tumor microenvironment in pancreatic cancer. *Cancer Discovery*. (2023) 13:278–97. doi: 10.1158/2159-8290.Cd-22-0876

19. Genova C, Dellepiane C, Carrega P, Sommariva S, Ferlazzo G, Pronzato P, et al. Therapeutic implications of tumor microenvironment in lung cancer: focus on immune checkpoint blockade. *Front Immunol.* (2021) 12:799455. doi: 10.3389/fimmu.2021.799455

20. Pitt JM, Marabelle A, Eggermont A, Soria JC, Kroemer G, Zitvogel L. Targeting the tumor microenvironment: removing obstruction to anticancer immune responses and immunotherapy. *Ann Oncol.* (2016) 27:1482–92. doi: 10.1093/annonc/mdw168

21. Vitale I, Manic G, Coussens LM, Kroemer G, Galluzzi L. Macrophages and metabolism in the tumor microenvironment. *Cell Metab.* (2019) 30:36–50. doi: 10.1016/j.cmet.2019.06.001

22. Ren X, Zhang L, Zhang Y, Li Z, Siemers N, Zhang Z. Insights gained from singlecell analysis of immune cells in the tumor microenvironment. *Annu Rev Immunol*. (2021) 39:583–609. doi: 10.1146/annurev-immunol-110519-071134

23. Gao X, Sui H, Zhao S, Gao X, Su Y, Qu P. Immunotherapy targeting myeloidderived suppressor cells (MDSCs) in tumor microenvironment. *Front Immunol.* (2020) 11:585214. doi: 10.3389/fimmu.2020.585214

24. Barnestein R, Galland L, Kalfeist L, Ghiringhelli F, Ladoire S, Limagne E. Immunosuppressive tumor microenvironment modulation by chemotherapies and targeted therapies to enhance immunotherapy effectiveness. *Oncoimmunology*. (2022) 11:2120676. doi: 10.1080/2162402x.2022.2120676

25. Samson N, Ablasser A. The cGAS-STING pathway and cancer. *Nat Cancer*. (2022) 3:1452–63. doi: 10.1038/s43018-022-00468-w

26. Liu N, Pang X, Zhang H, Ji P. The cGAS-STING pathway in bacterial infection and bacterial immunity. *Front Immunol.* (2021) 12:814709. doi: 10.3389/fimmu.2021.814709

27. Lu Q, Chen Y, Li J, Zhu F, Zheng Z. Crosstalk between cGAS-STING pathway and autophagy in cancer immunity. *Front Immunol.* (2023) 14:1139595. doi: 10.3389/fimmu.2023.1139595

28. Wan D, Jiang W, Hao J. Research advances in how the cGAS-STING pathway controls the cellular inflammatory response. *Front Immunol.* (2020) 11:615. doi: 10.3389/fimmu.2020.00615

29. Su T, Zhang Y, Valerie K, Wang XY, Lin S, Zhu G. STING activation in cancer immunotherapy. *Theranostics.* (2019) 9:7759–71. doi: 10.7150/thno.37574

30. Jiang M, Chen P, Wang L, Li W, Chen B, Liu Y, et al. cGAS-STING, an important pathway in cancer immunotherapy. *J Hematol Oncol.* (2020) 13:81. doi: 10.1186/s13045-020-00916-z

31. Tian Z, Zeng Y, Peng Y, Liu J, Wu F. Cancer immunotherapy strategies that target the cGAS-STING pathway. *Front Immunol.* (2022) 13:996663. doi: 10.3389/fimmu.2022.996663

32. Elhanani O, Ben-Uri R, Keren L. Spatial profiling technologies illuminate the tumor microenvironment. *Cancer Cell.* (2023) 41:404–20. doi: 10.1016/j.ccell.2023. 01.010

33. Domizio JD, Gulen MF, Saidoune F, Thacker VV, Yatim A, Sharma K, et al. The cGAS-STING pathway drives type I IFN immunopathology in COVID-19. *Nature*. (2022) 603:145–51. doi: 10.1038/s41586-022-04421-w

34. Alonso Paiva IM, AS R, Brito CB, Ferrero MC, Ortiz Wilczyński JM, Carrera Silva EA, et al. Role of the cGAS/STING pathway in the control of Brucella abortus infection acquired through the respiratory route. *Front Immunol.* (2023) 14:1116811. doi: 10.3389/fimmu.2023.1116811

35. Jin X, Wang W, Zhao X, Jiang W, Shao Q, Chen Z, et al. The battle between the innate immune cGAS-STING signaling pathway and human herpesvirus infection. *Front Immunol.* (2023) 14:1235590. doi: 10.3389/fimmu.2023.1235590

36. Zhang X, Bai XC, Chen ZJ. Structures and mechanisms in the cGAS-STING innate immunity pathway. *Immunity*. (2020) 53:43–53. doi: 10.1016/j.immuni.2020.05.013

37. Sun L, Wu J, Du F, Chen X, Chen ZJ. Cyclic GMP-AMP synthase is a cytosolic DNA sensor that activates the type I interferon pathway. *Science.* (2013) 339:786–91. doi: 10.1126/science.1232458

38. Zhu H, Zhang R, Yi L, Tang YD, Zheng C. UNC93B1 attenuates the cGAS-STING signaling pathway by targeting STING for autophagy-lysosome degradation. *J Med Virol.* (2022) 94:4490–501. doi: 10.1002/jmv.27860

39. Gui X, Yang H, Li T, Tan X, Shi P, Li M, et al. Autophagy induction via STING trafficking is a primordial function of the cGAS pathway. *Nature.* (2019) 567:262–6. doi: 10.1038/s41586-019-1006-9

40. Zhang D, Liu Y, Zhu Y, Zhang Q, Guan H, Liu S, et al. A non-canonical cGAS-STING-PERK pathway facilitates the translational program critical for senescence and organ fibrosis. *Nat Cell Biol.* (2022) 24:766–82. doi: 10.1038/s41556-022-00894-z

41. Xu D, Tian Y, Xia Q, Ke B. The cGAS-STING pathway: novel perspectives in liver diseases. *Front Immunol.* (2021) 12:682736. doi: 10.3389/fimmu.2021.682736

42. Gulen MF, Samson N, Keller A, Schwabenland M, Liu C, Glück S, et al. cGAS-STING drives ageing-related inflammation and neurodegeneration. *Nature.* (2023) 620:374–80. doi: 10.1038/s41586-023-06373-1

43. Victorelli S, Salmonowicz H, Chapman J, Martini H, Vizioli MG, Riley JS, et al. Apoptotic stress causes mtDNA release during senescence and drives the SASP. *Nature*. (2023) 622:627–36. doi: 10.1038/s41586-023-06621-4

44. Hu J, Sánchez-Rivera FJ, Wang Z, Johnson GN, Ho YJ, Ganesh K, et al. STING inhibits the reactivation of dormant metastasis in lung adenocarcinoma. *Nature*. (2023) 616:806–13. doi: 10.1038/s41586-023-05880-5

45. Guo Q, Chen X, Chen J, Zheng G, Xie C, Wu H, et al. STING promotes senescence, apoptosis, and extracellular matrix degradation in osteoarthritis via the NF- $\kappa$ B signaling pathway. *Cell Death Dis.* (2021) 12:13. doi: 10.1038/s41419-020-03341-9

46. Chen C, Xu P. Cellular functions of cGAS-STING signaling. Trends Cell Biol. (2023) 33:630–48. doi: 10.1016/j.tcb.2022.11.001

47. Sun Z, Hornung V. cGAS-STING signaling. Curr Biol. (2022) 32:R730-r734. doi: 10.1016/j.cub.2022.05.027

48. Pan J, Fei CJ, Hu Y, Wu XY, Nie L, Chen J. Current understanding of the cGAS-STING signaling pathway: Structure, regulatory mechanisms, and related diseases. *Zool Res.* (2023) 44:183–218. doi: 10.24272/j.issn.2095-8137.2022.464

49. Chauvin SD, Stinson WA, Platt DJ, Poddar S, Miner JJ. Regulation of cGAS and STING signaling during inflammation and infection. *J Biol Chem.* (2023) 299:104866. doi: 10.1016/j.jbc.2023.104866

50. Tanaka Y, Chen ZJ. STING specifies IRF3 phosphorylation by TBK1 in the cytosolic DNA signaling pathway. *Sci Signal.* (2012) 5:ra20. doi: 10.1126/ scisignal.2002521

51. Liu S, Cai X, Wu J, Cong Q, Chen X, Li T, et al. Phosphorylation of innate immune adaptor proteins MAVS, STING, and TRIF induces IRF3 activation. *Science*. (2015) 347:aaa2630. doi: 10.1126/science.aaa2630

 Dunphy G, Flannery SM, Almine JF, Connolly DJ, Paulus C, Jønsson KL, et al. Non-canonical activation of the DNA sensing adaptor STING by ATM and IF116 mediates NF-κB signaling after nuclear DNA damage. *Mol Cell*. (2018) 71:745– 760:e745. doi: 10.1016/j.molcel.2018.07.034

53. Vashi N, Bakhoum SF. The evolution of STING signaling and its involvement in cancer. *Trends Biochem Sci.* (2021) 46:446–60. doi: 10.1016/j.tibs.2020.12.010

54. Messaoud-Nacer Y, Culerier E, Rose S, Maillet I, Rouxel N, Briault S, et al. STING agonist diABZI induces PANoptosis and DNA mediated acute respiratory distress syndrome (ARDS). *Cell Death Dis.* (2022) 13:269. doi: 10.1038/s41419-022-04664-5

55. Liu X, Hogg GD, Zuo C, Borcherding NC, Baer JM, Lander VE, et al. Contextdependent activation of STING-interferon signaling by CD11b agonists enhances antitumor immunity. *Cancer Cell.* (2023) 41:1073–1090.e1012. doi: 10.1016/ j.ccell.2023.04.018

56. Zhao H, Wu L, Yan G, Chen Y, Zhou M, Wu Y, et al. Inflammation and tumor progression: signaling pathways and targeted intervention. *Signal Transduct Target Ther.* (2021) 6:263. doi: 10.1038/s41392-021-00658-5

57. Ghosh M, Saha S, Li J, Montrose DC, Martinez LA. p53 engages the cGAS/ STING cytosolic DNA sensing pathway for tumor suppression. *Mol Cell*. (2023) 83:266–280.e266. doi: 10.1016/j.molcel.2022.12.023

58. Meibers HE, Warrick KA, VonHandorf A, Vallez CN, Kawarizadeh K, Saha I, et al. Effector memory T cells induce innate inflammation by triggering DNA damage and a non-canonical STING pathway in dendritic cells. *Cell Rep.* (2023) 42:113180. doi: 10.1016/j.celrep.2023.113180

59. Decout A, Katz JD, Venkatraman S, Ablasser A. The cGAS-STING pathway as a therapeutic target in inflammatory diseases. *Nat Rev Immunol.* (2021) 21:548–69. doi: 10.1038/s41577-021-00524-z

60. Skopelja-Gardner S, An J, Elkon KB. Role of the cGAS-STING pathway in systemic and organ-specific diseases. *Nat Rev Nephrol.* (2022) 18:558–72. doi: 10.1038/ s41581-022-00589-6

61. Kato Y, Park J, Takamatsu H, Konaka H, Aoki W, Aburaya S, et al. Apoptosisderived membrane vesicles drive the cGAS-STING pathway and enhance type I IFN production in systemic lupus erythematosus. *Ann Rheum Dis.* (2018) 77:1507–15. doi: 10.1136/annrheumdis-2018-212988

62. Konno H, Konno K, Barber GN. Cyclic dinucleotides trigger ULK1 (ATG1) phosphorylation of STING to prevent sustained innate immune signaling. *Cell.* (2013) 155:688–98. doi: 10.1016/j.cell.2013.09.049

63. Wang Y, Luo J, Alu A, Han X, Wei Y, Wei X. cGAS-STING pathway in cancer biotherapy. *Mol Cancer.* (2020) 19:136. doi: 10.1186/s12943-020-01247-w

64. Lv M, Chen M, Zhang R, Zhang W, Wang C, Zhang Y, et al. Manganese is critical for antitumor immune responses via cGAS-STING and improves the efficacy of clinical immunotherapy. *Cell Res.* (2020) 30:966–79. doi: 10.1038/s41422-020-00395-4

65. Graham PT, Nowak AK, Cornwall SMJ, Larma I, Nelson DJ. The STING agonist, DMXAA, reduces tumor vessels and enhances mesothelioma tumor antigen presentation yet blunts cytotoxic T cell function in a murine model. *Front Immunol.* (2022) 13:969678. doi: 10.3389/fimmu.2022.969678

66. Wang X, Zhang H, Wang Y, Bramasole L, Guo K, Mourtada F, et al. DNA sensing via the cGAS/STING pathway activates the immunoproteasome and adaptive T-cell immunity. *EMBO J.* (2023) 42:e110597. doi: 10.15252/embj.2022110597

67. Bakhoum SF, Ngo B, Laughney AM, Cavallo JA, Murphy CJ, Ly P, et al. Chromosomal instability drives metastasis through a cytosolic DNA response. *Nature*. (2018) 553:467–72. doi: 10.1038/nature25432

68. Drews RM, Hernando B, Tarabichi M, Haase K, Lesluyes T, Smith PS, et al. A pan-cancer compendium of chromosomal instability. *Nature.* (2022) 606:976–83. doi: 10.1038/s41586-022-04789-9

69. Hong C, Schubert M, Tijhuis AE, Requesens M, Roorda M, van den Brink A, et al. cGAS-STING drives the IL-6-dependent survival of chromosomally instable cancers. *Nature*. (2022) 607:366–73. doi: 10.1038/s41586-022-04847-2

70. Beernaert B, Parkes EE. cGAS-STING signalling in cancer: striking a balance with chromosomal instability. *Biochem Soc Trans.* (2023) 51:539–55. doi: 10.1042/bst20220838

71. Chen YH, Chen HH, Wang WJ, Chen HY, Huang WS, Kao CH, et al. TRABID inhibition activates cGAS/STING-mediated anti-tumor immunity through mitosis and autophagy dysregulation. *Nat Commun.* (2023) 14:3050. doi: 10.1038/s41467-023-38784-z

72. Yang C, Liang Y, Liu N, Sun M. Role of the cGAS-STING pathway in radiotherapy for non-small cell lung cancer. *Radiat Oncol.* (2023) 18:145. doi: 10.1186/s13014-023-02335-z

73. Yang Y, Wu M, Cao D, Yang C, Jin J, Wu L, et al. ZBP1-MLKL necroptotic signaling potentiates radiation-induced antitumor immunity via intratumoral STING pathway activation. *Sci Adv.* (2021) 7:eabf6290. doi: 10.1126/sciadv.abf6290

74. Deng L, Liang H, Xu M, Yang X, Burnette B, Arina A, et al. STING-dependent cytosolic DNA sensing promotes radiation-induced type I interferon-dependent antitumor immunity in immunogenic tumors. *Immunity*. (2014) 41:843–52. doi: 10.1016/j.immuni.2014.10.019

75. Gan Y, Li X, Han S, Liang Q, Ma X, Rong P, et al. The cGAS/STING Pathway: A novel target for cancer therapy. *Front Immunol.* (2021) 12:795401. doi: 10.3389/fimmu.2021.795401

76. Garland KM, Sheehy TL, Wilson JT. Chemical and biomolecular strategies for STING pathway activation in cancer immunotherapy. *Chem Rev.* (2022) 122:5977–6039. doi: 10.1021/acs.chemrev.1c00750

77. Zhou L, Zhang YF, Yang FH, Mao HQ, Chen Z, Zhang L. Mitochondrial DNA leakage induces odontoblast inflammation via the cGAS-STING pathway. *Cell Commun Signal.* (2021) 19:58. doi: 10.1186/s12964-021-00738-7

78. Cheng AN, Cheng LC, Kuo CL, Lo YK, Chou HY, Chen CH, et al. Mitochondrial Lon-induced mtDNA leakage contributes to PD-L1-mediated immunoescape via STING-IFN signaling and extracellular vesicles. *J Immunother Cancer*. (2020) 8: e001372. doi: 10.1136/jitc-2020-001372

79. Heyman M, Grandér D, Bröndum-Nielsen K, Liu Y, Söderhäll S, Einhorn S. Deletions of the short arm of chromosome 9, including the interferon-alpha/-beta genes, in acute lymphocytic leukemia. Studies on loss of heterozygosity, parental origin of deleted genes and prognosis. *Int J Cancer*. (1993) 54:748–53. doi: 10.1002/ijc.2910540507

80. Xia T, Konno H, Ahn J, Barber GN. Deregulation of STING signaling in colorectal carcinoma constrains DNA damage responses and correlates with tumorigenesis. *Cell Rep.* (2016) 14:282–97. doi: 10.1016/j.celrep.2015.12.029

81. Ablasser A, Schmid-Burgk JL, Hemmerling I, Horvath GL, Schmidt T, Latz E, et al. Cell intrinsic immunity spreads to bystander cells via the intercellular transfer of cGAMP. *Nature*. (2013) 503:530–4. doi: 10.1038/nature12640

82. Luteijn RD, Zaver SA, Gowen BG, Wyman SK, Garelis NE, Onia L, et al. SLC19A1 transports immunoreactive cyclic dinucleotides. *Nature*. (2019) 573:434–8. doi: 10.1038/s41586-019-1553-0

83. Chen Q, Boire A, Jin X, Valiente M, Er EE, Lopez-Soto A, et al. Carcinomaastrocyte gap junctions promote brain metastasis by cGAMP transfer. *Nature*. (2016) 533:493–8. doi: 10.1038/nature18268

84. Kitai Y, Kawasaki T, Sueyoshi T, Kobiyama K, Ishii KJ, Zou J, et al. DNAcontaining exosomes derived from cancer cells treated with topotecan activate a STING-dependent pathway and reinforce antitumor immunity. *J Immunol.* (2017) 198:1649–59. doi: 10.4049/jimmunol.1601694

85. Woo SR, Fuertes MB, Corrales L, Spranger S, Furdyna MJ. Leung MY et al: STINGdependent cytosolic DNA sensing mediates innate immune recognition of immunogenic tumors. *Immunity*. (2014) 41:830–42. doi: 10.1016/j.immuni.2014.10.017

86. Yi M, Niu M, Zhang J, Li S, Zhu S, Yan Y, et al. Combine and conquer: manganese synergizing anti-TGF-β/PD-L1 bispecific antibody YM101 to overcome immunotherapy resistance in non-inflamed cancers. *J Hematol Oncol.* (2021) 14:146. doi: 10.1186/s13045-021-01155-6

87. Zhang R, Wang C, Guan Y, Wei X, Sha M, Yi M, et al. Manganese salts function as potent adjuvants. *Cell Mol Immunol.* (2021) 18:1222–34. doi: 10.1038/s41423-021-00669-w

88. Song Y, Liu Y, Teo HY, Hanafi ZB, Mei Y, Zhu Y, et al. Manganese enhances the antitumor function of CD8(+) T cells by inducing type I interferon production. *Cell Mol Immunol.* (2021) 18:1571–4. doi: 10.1038/s41423-020-00524-4

89. Marcus A, Mao AJ, Lensink-Vasan M, Wang L, Vance RE, Raulet DH. Tumorderived cGAMP triggers a STING-mediated interferon response in non-tumor cells to activate the NK cell response. *Immunity*. (2018) 49:754–763.e754. doi: 10.1016/ j.immuni.2018.09.016

90. Bakhoum SF, Cantley LC. The multifaceted role of chromosomal instability in cancer and its microenvironment. Cell. (2018) 174:1347-60. doi: 10.1016/j.cell.2018.08.027

91. Ou L, Zhang A, Cheng Y, Chen Y. The cGAS-STING pathway: A promising immunotherapy target. Front Immunol. (2021) 12:795048. doi: 10.3389/fimmu.2021.795048

92. Shih AY, Damm-Ganamet KL, Mirzadegan T. Dynamic structural differences between human and mouse STING lead to differing sensitivity to DMXAA. *Biophys J.* (2018) 114:32–9. doi: 10.1016/j.bpj.2017.10.027

93. Papaevangelou E, Esteves AM, Dasgupta P, Galustian C. Cyto-IL-15 synergizes with the STING agonist ADU-S100 to eliminate prostate tumors and confer durable immunity in mouse models. *Front Immunol.* (2023) 14:1196829. doi: 10.3389/fimmu.2023.1196829

94. Esteves AM, Papaevangelou E, Dasgupta P, Galustian C. Combination of interleukin-15 with a STING agonist, ADU-S100 analog: A potential immunotherapy for prostate cancer. *Front Oncol.* (2021) 11:621550. doi: 10.3389/ fonc.2021.621550

95. Luo J, Pang S, Hui Z, Zhao H, Xu S, Yu W, et al. Blocking Tim-3 enhances the anti-tumor immunity of STING agonist ADU-S100 by unleashing CD4(+) T cells through regulating type 2 conventional dendritic cells. *Theranostics*. (2023) 13:4836–57. doi: 10.7150/thno.86792

96. Meric-Bernstam F, Sweis RF, Hodi FS, Messersmith WA, Andtbacka RHI, Ingham M, et al. Phase I dose-escalation trial of MIW815 (ADU-S100), an intratumoral STING agonist, in patients with advanced/metastatic solid tumors or lymphomas. *Clin Cancer Res.* (2022) 28:677–88. doi: 10.1158/1078-0432.Ccr-21-1963

97. Meric-Bernstam F, Sweis RF, Kasper S, Hamid O, Bhatia S, Dummer R, et al. Combination of the STING agonist MIW815 (ADU-S100) and PD-1 inhibitor spartalizumab in advanced/metastatic solid tumors or lymphomas: an open-label, multicenter, phase ib study. *Clin Cancer Res.* (2023) 29:110–21. doi: 10.1158/1078-0432.Ccr-22-2235

98. Chang W, Altman MD, Lesburg CA, Perera SA, Piesvaux JA, Schroeder GK, et al. Discovery of MK-1454: A potent cyclic dinucleotide stimulator of interferon genes agonist for the treatment of cancer. *J Med Chem.* (2022) 65:5675–89. doi: 10.1021/acs.jmedchem.1c02197

99. McIntosh JA, Liu Z, Andresen BM, Marzijarani NS, Moore JC, Marshall NM, et al. A kinase-cGAS cascade to synthesize a therapeutic STING activator. *Nature*. (2022) 603:439–44. doi: 10.1038/s41586-022-04422-9

100. Smith M, Chin D, Chan S, Mahady S, Campion L, Morgan C, et al. Abstract 5567: *In vivo* administration of the STING agonist, JNJ-67544412, leads to complete regression of established murine subcutaneous tumors. *Cancer Res.* (2020) 80:5567–7. doi: 10.1158/1538-7445.AM2020-5567

101. Challa S, Zhou S, Sheri A, Padmanabhan S, Delaney S, Meher G, et al. Abstract B40: Nucleotide analogs as novel STING agonists for immuno-oncology. *Cancer Immunol Res.* (2017) 5:B40–0. doi: 10.1158/2326-6074.TUMIMM16-B40

102. ABBAS A, Strauss J, Janku F, Karim R, Olszanski A, Luke J, et al. P01.01 A Phase 1a/1b dose-escalation study of intravenously administered SB 11285 alone and in combination with nivolumab in patients with advanced solid tumors. *J ImmunoTherapy Cancer*. (2020) 8:A7–8. doi: 10.1136/jitc-2020-ITOC7.14

103. Sun X, Zhang Y, Li J, Park KS, Han K, Zhou X, et al. Amplifying STING activation by cyclic dinucleotide-manganese particles for local and systemic cancer

metalloimmunotherapy. Nat Nanotechnol. (2021) 16:1260-70. doi: 10.1038/s41565-021-00962-9

104. Amouzegar A, Chelvanambi M, Filderman JN, Storkus WJ, Luke JJ. STING agonists as cancer therapeutics. *Cancers (Basel).* (2021) 13:2695. doi: 10.3390/cancers13112695

105. Wang J, Falchook G, Nabhan S, Kulkarni M, Sandy P, Dosunmu O, et al. 495 Trial of SNX281, a systemically delivered small molecule STING agonist, in solid tumors and lymphomas. *J ImmunoTherapy Cancer*. (2021) 9:A527–7. doi: 10.1136/jitc-2021-SITC2021.495

106. Ding C, Song Z, Shen A, Chen T, Zhang A. Small molecules targeting the innate immune cGAS–STING–TBK1 signaling pathway. *Acta Pharm Sin B*. (2020) 10:2272–98. doi: 10.1016/j.apsb.2020.03.001

107. Banerjee M, Basu S, Middya S, Shrivastava R, Ghosh R, Pryde DC, et al. Abstract LB-061: CRD5500: A versatile small molecule STING agonist amenable to bioconjugation as an ADC. *Cancer Res.* (2019) 79:LB-061-LB-061. doi: 10.1158/1538-7445.AM2019-LB-061

108. Wang Z, Dove P, Rosa D, Bossen B, Helke S, Charbonneau M, et al. Abstract 3854: Preclinical characterization of a novel non-cyclic dinucleotide small molecule STING agonist with potent antitumor activity in mice. *Cancer Res.* (2019) 79:3854–4. doi: 10.1158/1538-7445.AM2019-3854

109. Jekle A, Thatikonda SK, Jaisinghani R, Ren S, Kinkade A, Stevens SK, et al. Tumor regression upon intratumoral and subcutaneous dosing of the STING agonist ALG-031048 in mouse efficacy models. *Int J Mol Sci.* (2023) 24:16274. doi: 10.3390/ ijms242216274

110. Pan BS, Perera SA, Piesvaux JA, Presland JP, Schroeder GK, Cumming JN, et al. An orally available non-nucleotide STING agonist with antitumor activity. *Science*. (2020) 369:eaba6098. doi: 10.1126/science.aba6098

111. Yi M, Niu M, Wu Y, Ge H, Jiao D, Zhu S, et al. Combination of oral STING agonist MSA-2 and anti-TGF- $\beta$ /PD-L1 bispecific antibody YM101: a novel immune cocktail therapy for non-inflamed tumors. *J Hematol Oncol.* (2022) 15:142. doi: 10.1186/s13045-022-01363-8

112. Lin Z, Wang Q, Jiang T, Wang W, Zhao JJ. Targeting tumor-associated macrophages with STING agonism improves the antitumor efficacy of osimertinib in a mouse model of EGFR-mutant lung cancer. *Front Immunol.* (2023) 14:1077203. doi: 10.3389/fimmu.2023.1077203

113. Ramanjulu JM, Pesiridis GS, Yang J, Concha N, Singhaus R, Zhang SY, et al. Design of amidobenzimidazole STING receptor agonists with systemic activity. *Nature.* (2018) 564:439–43. doi: 10.1038/s41586-018-0705-y

114. Chan SR, Bignan G, Pierson E, Mahady S, Ta H, Schepens W, et al. Abstract 5567A: JNJ-'6196: A next generation STING agonist with potent preclinical activity by the IV route. *Cancer Res.* (2020) 80:5567A–A. doi: 10.1158/1538-7445.Am2020-5567a

115. Wu YT, Fang Y, Wei Q, Shi H, Tan H, Deng Y, et al. Tumor-targeted delivery of a STING agonist improvescancer immunotherapy. *Proc Natl Acad Sci USA*. (2022) 119: e2214278119. doi: 10.1073/pnas.2214278119

116. Duvall JR, Thomas JD, Bukhalid RA, Catcott KC, Bentley KW, Collins SD, et al. Discovery and optimization of a STING agonist platform for application in antibody drug conjugates. *J Med Chem.* (2023) 66:10715–33. doi: 10.1021/acs.jmedchem.3c00907

117. Zhou L, Yi W, Zhang Z, Shan X, Zhao Z, Sun X, et al. STING agonist-boosted mRNA immunization via intelligent design of nanovaccines for enhancing cancer immunotherapy. *Natl Sci Rev.* (2023) 10:nwad214. doi: 10.1093/nsr/nwad214

118. Zhang Y, Shen T, Zhou S, Wang W, Lin S, Zhu G. pH-responsive STINGactivating DNA nanovaccines for cancer immunotherapy. *Adv Ther (Weinh)*. (2020) 3:2000083. doi: 10.1002/adtp.202000083

119. Gu Y, Lin S, Wu Y, Xu P, Zhu W, Wang Y, et al. Targeting STING activation by antigen-inspired mnO(2) nanovaccines optimizes tumor radiotherapy. *Adv Healthc Mater.* (2023) 12:e2300028. doi: 10.1002/adhm.202300028

120. Fu J, Kanne DB, Leong M, Glickman LH, McWhirter SM, Lemmens E, et al. STING agonist formulated cancer vaccines can cure established tumors resistant to PD-1 blockade. *Sci Transl Med.* (2015) 7:283ra252. doi: 10.1126/scitranslmed.aaa4306

121. Su T, Cheng F, Qi J, Zhang Y, Zhou S, Mei L, et al. Responsive multivesicular polymeric nanovaccines that codeliver STING agonists and neoantigens for combination tumor immunotherapy. *Adv Sci (Weinh).* (2022) 9:e2201895. doi: 10.1002/advs.202201895

122. Jiang X, Wang J, Zheng X, Liu Z, Zhang X, Li Y, et al. Intratumoral administration of STING-activating nanovaccine enhances T cell immunotherapy. *J Immunother Cancer*. (2022) 10:e003960. doi: 10.1136/jitc-2021-003960

123. Liu H, Hu Z, Chen H, Yan Y, Le Z, Wei C, et al. Self-degradable poly( $\beta$ -amino ester)s promote endosomal escape of antigen and agonist. *J Control Release.* (2022) 345:91–100. doi: 10.1016/j.jconrel.2022.03.006

124. Cheng L, Zhang P, Liu Y, Liu Z, Tang J, Xu L, et al. Multifunctional hybrid exosomes enhanced cancer chemo-immunotherapy by activating the STING pathway. *Biomaterials.* (2023) 301:122259. doi: 10.1016/j.biomaterials.2023.122259

125. Liu J, Xiang J, Jin C, Ye L, Wang L, Gao Y, et al. Medicinal plant-derived mtDNA via nanovesicles induces the cGAS-STING pathway to remold tumorassociated macrophages for tumor regression. *J Nanobiotechnology*. (2023) 21:78. doi: 10.1186/s12951-023-01835-0

126. Bao P, Gu HY, Ye JJ, He JL, Zhong Z, Yu AX, et al. Chimeric exosomes functionalized with STING activation for personalized glioblastoma immunotherapy. *Adv Sci (Weinh).* (2024) 11:e2306336. doi: 10.1002/advs.202306336

127. Luke JJ, Piha-Paul SA, Medina T, Verschraegen CF, Varterasian M, Brennan AM, et al. Phase I Study of SYNB1891, an Engineered E. coli Nissle Strain Expressing STING Agonist, with and without Atezolizumab in Advanced Malignancies. *Clin Cancer Res.* (2023) 29:2435–44. doi: 10.1158/1078-0432.Ccr-23-0118

128. Zhao K, Huang J, Zhao Y, Wang S, Xu J, Yin K. Targeting STING in cancer: Challenges and emerging opportunities. *Biochim Biophys Acta Rev Cancer*. (2023) 1878:188983. doi: 10.1016/j.bbcan.2023.188983

129. Wang Q, Bergholz JS, Ding L, Lin Z, Kabraji SK, Hughes ME, et al. STING agonism reprograms tumor-associated macrophages and overcomes resistance to PARP inhibition in BRCA1-deficient models of breast cancer. *Nat Commun.* (2022) 13:3022. doi: 10.1038/s41467-022-30568-1

130. Ma Z, Li Z, Mao Y, Ye J, Liu Z, Wang Y, et al. AhR diminishes the efficacy of chemotherapy via suppressing STING dependent type-I interferon in bladder cancer. *Nat Commun.* (2023) 14:5415. doi: 10.1038/s41467-023-41218-5