Check for updates

OPEN ACCESS

EDITED BY Dehong Yan, Chinese Academy of Sciences (CAS), China

REVIEWED BY

Dmitry Aleksandrovich Zinovkin, Gomel State Medical University, Belarus Zhao Jian, Chinese People's Liberation Army General Hospital, China

[†]These authors have contributed equally to this work

RECEIVED 09 September 2024 ACCEPTED 12 November 2024 PUBLISHED 02 December 2024

CITATION

Peng K, Zhang X, Li Z, Wang Y, Sun H-W, Zhao W, Pan J, Zhang X-Y, Wu X, Yu X, Wu C, Weng Y, Lin X, Liu D, Zhan M, Xu J, Zheng L, Zhang Y and Lu L (2024) Myeloid response evaluated by noninvasive CT imaging predicts post-surgical survival and immune checkpoint therapy benefits in patients with hepatocellular carcinoma. *Front. Immunol.* 15:1493735. doi: 10.3389/fimmu.2024.1493735

COPYRIGHT

© 2024 Peng, Zhang, Li, Wang, Sun, Zhao, Pan, Zhang, Wu, Yu, Wu, Weng, Lin, Liu, Zhan, Xu, Zheng, Zhang and Lu. This is an open-access article distributed under the terms of the **Creative Commons Attribution License (CC BY)**. The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Myeloid response evaluated by noninvasive CT imaging predicts post-surgical survival and immune checkpoint therapy benefits in patients with hepatocellular carcinoma

Kangqiang Peng^{1†}, Xiao Zhang^{2,3†}, Zhongliang Li^{2†}, Yongchun Wang^{1,4†}, Hong-Wei Sun², Wei Zhao^{2,5}, Jielin Pan^{2,6}, Xiao-Yang Zhang⁷, Xiaoling Wu⁸, Xiangrong Yu^{2,6}, Chong Wu⁹, Yulan Weng⁹, Xiaowen Lin², Dingjie Liu^{2,10}, Meixiao Zhan^{2,11}, Jing Xu¹, Limin Zheng ^{1,9*}, Yaojun Zhang ^{1*} and Ligong Lu ^{2,11*}

¹State Key Laboratory of Oncology in South China, Guangdong Provincial Clinical Research Center for Cancer, Sun Yat-sen University Cancer Center, Guangzhou, China, ²Guangdong Provincial Key Laboratory of Tumor Interventional Diagnosis and Treatment, Zhuhai Institute of Translational Medicine, Zhuhai People's Hospital (Zhuhai Clinical Medical College), Jinan University, Zhuhai, China, ³Medical AI Lab, Hebei Provincial Engineering Research Center for AI-Based Cancer Treatment Decision-Making, The First Hospital of Hebei Medical University, Shijiazhuang, China, ⁴Department of Pathology, Xiangya Hospital, Central South University, Changsha, China, ⁵Department of Management, School of Business, Macau University of Science and Technology, Macau, Macau SAR, China, ⁶Department of Radiology, Zhuhai People's Hospital, Jinan University, Zhuhai, China, ⁷College of Medicine and Biological Information Engineering, Northeastern University, Shenyang, China, ⁸Department of Radiology, The First Affiliated Hospital of Jinan University, Guangzhou, China, ⁹Ministry of Education (MOE) Key Laboratory of Gene Function and Regulation, School of Life Sciences, Sun Yat-sen University, Guangzhou, China, 10 The Department of Cerebrovascular Disease, Zhuhai People's Hospital, Jinan University, Zhuhai, China, ¹¹Guangzhou First People's Hospital, The Second Affiliated Hospital, School of Medicine, South China University of Technology, Guangzhou, China

Background: The potential of preoperative CT in the assessment of myeloid immune response and its application in predicting prognosis and immune-checkpoint therapy outcomes in hepatocellular carcinoma (HCC) has not been explored.

Methods: A total of 165 patients with pathological slides and multi-phase CT images were included to develop a radiomics signature for predicting the imaging-based myeloid response score (iMRS). Overall survival (OS) and recurrence-free survival (RFS) were assessed according to the iMRS risk group and validated in a surgical resection cohort (n = 98). The complementary advantage of iMRS incorporating significant clinicopathologic factors was investigated by the Cox proportional hazards analysis. Additionally, the iMRS in inferring the benefits of immune checkpoint therapy was explored in an immunotherapy cohort (n = 36).

Results: We showed that AUCs of the optimal radiomics signature for iMRS were 0.941 [95% confidence interval (CI), 0.909–0.973] and 0.833 (0.798–0.868) in the training and test cohorts, respectively. High iMRS was associated with poor

RFS and OS. The prognostic performance of the Clinical-iMRS nomogram was better than that of a single parameter (p < 0.05), with a 1-, 3-, and 5-year C-index for RFS of 0.729, 0.709, and 0.713 in the training, test, and surgical resection cohorts, respectively. A high iMRS score predicted a higher proportion of objective response (vs. progressive disease or stable disease; odds ratio, 2.311; 95% Cl, 1.144–4.672; p = 0.020; AUC, 0.718) in patients treated with anti-PD-1 and PD-L1.

Conclusions: iMRS may provide a promising method for predicting local myeloid immune responses in HCC patients, inferring postsurgical prognosis, and evaluating benefits of immune checkpoint therapy.

KEYWORDS

hepatocellular carcinoma, radiomics, myeloid cells, prognosis, immunotherapy

Background

Hepatocellular carcinoma (HCC) is one of the leading causes of cancer-related death worldwide (1). Most HCCs arise from persistent inflammation, including hepatitis B or C virus (HBV or HCV) infections and nonalcoholic fatty liver disease (2). Therefore, immune cells constitute a highly complex and interactive milieu that contributes to the development and progression of HCC (3). Increasing evidence has also shown that tumor-infiltrating immune cells are potential prognostic and predictive factors for patient survival and therapeutic outcome (4–6). Substantial efforts have been made to depict the tumor microenvironment (TME) by integrating the information of local immune cells (7–10).

Myeloid cells are a population of heterogenous innate cells in the TME, including monocytes/macrophages, dendritic cells, neutrophils, and myeloid-derived suppressor cells (11). These cells are major components and critical regulators in the tumor contexture, which play a vital role in tumor initiation, progression, and therapy response (12, 13). Therefore, myeloid-based biomarkers have attracted particular attention to predict the prognosis and clinical benefit of patients (14-16). However, it usually needs to involve several myeloid markers due to the heterogenicity and plasticity of local myeloid cells in tumor tissues (8, 17), resulting in extra and redundant pathological examination. Previously, Wu et al. used 18 myeloid-related features to fit clinical data of patients with HCC and finally constructed a myeloid-specific prognostic signature (based on CD11b and CD169) named myeloid response score (MRS) (8). MRS reflects the changes in the myeloid response balance from antitumor to protumor activities and is closely related to the immune tolerance of CD8⁺ T cells. The findings indicate that MRS is accurate and useful in predicting post-surgery HCC prognosis and sorafenib efficacy for recurrent HCC. However, the small specimens cannot comprehensively capture the biological characteristics and reflect MRS levels in the whole tumor due to tumor heterogeneity and limitations in pathology specimen acquisition. Given the highly dynamic evolution and spatial heterogeneity of TME (18), a noninvasive, economical, and comprehensive panoramic view of the MRS assessment is still in great demand to decipher the tumor immune infiltrate.

Medical imaging, which allows noninvasive and comprehensive tumor evaluation, can reveal subtle relations between tumor texture and the molecular biological processes active in the TME (19). Multiphase dynamic enhancement CT scans can display lesions from multiple angles and directions, to fully reveal the blood supply and the intensification characteristics of patients with HCC. Unlike the visual interpretation of medical images, an emerging highthroughput computational method called "Radiomics" can reflect the molecular properties of tumor tissues through translating imaging data into high-dimensional quantitative data (20–22). It allows noninvasive, timely, and longitudinal evaluation of the entire tumor as well as its microenvironment to complement biopsies and tissue sections, demonstrating a great advantage in predicting vascular invasion, histological grade, prognosis, and therapeutic outcomes in HCC (23–28) and other tumors (29–32).

In view of the tumor immune microenvironment, the association and feasibility of radiomics-based biomarkers to tumor-infiltrating immune cell (33–36), the response of immunotherapy (37–41), and PD-L1/PD-L2 expression level (42–44) have also been investigated. Previously, Jiang et al. successfully developed a noninvasive radiomics-based predictor of ImmunoScore of gastric cancer from lymphoid and myeloid cells (45) and further validated its association with both disease-free survival and overall survival (OS) (35). The finding implied that it might be feasible to use radiomics to noninvasively predict MRS in HCC. Considering that only a fraction of patients benefit from the PD-1/PD-L1 monotherapy, it

Abbreviations: HCC, hepatocellular carcinoma; HR, hazard ratio; MRS, myeloid response score; OS, overall survival; RFS, recurrence-free survival; CI, confidence interval.

implies a great need to excavate a curative effect predictor for precise immunotherapy (46–48). In line with this, Yuming et al. (40) proposed a new imaging-based TME classifier, i.e., deep learning radiomics signature (DLRS), which could be used as a highly independent prognostic factor to accurately predict clinical response in patients treated with checkpoint blockade immunotherapy. The efficacy of predictive radiologic markers based on MRS in assessing the benefits of immune checkpoint therapy are unknown and need to be further explored in HCC.

In this study, we aimed to construct a predictive radiomics signature of MRS by combining triple-phase CT images and immunohistochemistry (IHC) staining from tumor biopsies for patients with HCC. A multi-phase CT (mp-CT) imaging-based predictor for MRS was validated to predict the local MRS, evaluate patient survival, and infer the benefits of immune checkpoint therapy in HCC.

Methods

Study design and patients

The retrospective study was approved by the ethics committee of Sun Yat-sen University Cancer Center, and the requirement for informed consent was waived. The overall study design is shown in Figure 1. To develop the radiomics model for MRS, patients with HCC treated at our hospital were enrolled, in line with the inclusion and exclusion criteria. The inclusion criteria were as follows: (a) pathologic confirmation of HCC with available pathology slides and (b) preoperative contrast-enhanced liver CT performed. The exclusion criteria were as follows: (a) undergoing other treatments before surgery and (b) poor-quality radiologic or pathologic images.

A total of 165 patients who performed presurgical liver CT scans and had available tumor samples for IHC staining from 2007 to 2010 were randomly split at a 7:3 ratio to cohort 1 (training cohort, n = 110) for the construction of the radiomics signature and cohort 2 (test cohort, n = 55) for performance evaluation. An additional surgical resection cohort (n = 98) including patients who were consecutively treated from 2010 to 2016 with CT scans and clinicopathological data was used for prognostic analysis based on the MRS radiomics signature. Moreover, another immunotherapy cohort (n = 36) with patients who underwent anti-PD-1/PD-L1 therapy from 2018 to 2020 was used to assess the outcome of immune checkpoint therapy in HCC. All the patients were followed up through telephone or admission notes to record recurrence and death. OS was calculated as the time from surgery to death or the last follow-up. Recurrence-free survival (RFS) was defined as the interval between the time of surgery to recurrence, the last followup for patients without recurrence, or death if no recurrence was observed.

IHC staining and definition of MRS

Paraffin-embedded tumor tissues were cut into 4- μ m sections, which were used for IHC staining. Tumor sections were sequentially deparaffinized and re-hydrated with xylene and a decreasing ethanol series. Subsequently, the slides were subjected to endogenous peroxidase activity elimination in 0.3% H₂O₂ for 10 min and heat-induced epitope retrieval in citrate buffer for 10 min. The sections were then incubated with anti-CD169 antibody (1:200, R&D Systems, Cat#AF5197) or anti-CD11b antibody (1:2000, Abcam, Cat#ab133357) overnight at 4°C. Diaminobenzidine



FIGURE 1

Study design. The training cohort and test cohort that contain data from patients with CT images and pathology slides were used to develop the radiomics signature of MRS. Two additional cohorts were used to validate the clinical and prognostic value of this radiomics signature. The surgical resection cohort comprised clinical data and the corresponding imaging data from patients with surgical resection. The immunotherapy cohort comprised advanced HCC patients who had been treated with anti-PD-1 or anti-PD-L1 therapy. MRS, myeloid response score.

(DAB) staining was performed with horseradish peroxidaseconjugated anti-rabbit/mouse Dako REALTM EnVisionTM detection systems (Dako, Cat# K5007) according to the manufacturer's instructions. Brown color indicated positive signaling. All sections were counterstained with Mayer's hematoxylin.

IHC-stained slides were scanned at ×20 magnification and evaluated by the Vectra 2.0 Automated Quantitative Pathology Imaging System and InForm Cell Analysis 2.2 (PerkinElmer) to count the numbers of CD169⁺ or CD11b⁺ cells (8). The density of cells was quantified as the number of positive cells per square millimeter area. The MRS was calculated as follows: MRS = 0.161 × CD11b – 0.106 × CD169 + 35 (0 ≤ MRS ≤ 100), to classify the HCC patients into two subgroups (MRS_{low}, 0–60.6; and MRS_{high}, 60.7–100; the cutoff was determined with the X-Tile software). IHC and CT images from two randomly selected patients in the MRS_{low} and MRS_{high} groups are shown in Figure 2A.

CT acquisition, image preprocessing, and tumor segmentation

Triple-phase CT images of the liver, namely, non-contrast phase (NP), arterial phase (AP; 20–40 s postcontrast injection), and portal venous phase (PVP; 50–70 s postcontrast injection), were obtained after intravenous injection of contrast agent (Bayer Schering Pharma AG, Berlin, Germany; Ultravist Iopromide 370 mgI/mL) at a rate of 2–3 mL/s and a dose of 1–1.5 mL/kg bodyweight. The scanning range included the whole liver (from the diaphragmatic dome to the lower edge of the liver). Details regarding CT acquisition parameters are provided in Supplementary Table S1.

All the images were retrieved from the picture archiving and communication system. The gray values were normalized to a range of 0–255 to reduce scanner- and patient-dependent intensity variability in CT imaging. The manual segmentation containing



FIGURE 2

Radiomics workflow. (A) Representative imaging and pathology data of patients from MRS-high and MRS-low groups. (B) Radiomics workflow. MRS-PM represents the mp-CT radiomics signature with the best predictive performance for MRS. Prognosis-PM represents the prognostic Clinical-iMRS nomogram by combining iMRS and the significant clinicopathological features. NP, non-contrast phase; AP, arterial phase; PVP, portal venous phase; ICC, intraclass correlation coefficient; PCC, Pearson correlation coefficient; RFE, recursive feature elimination; iMRS, imaging-based MRS; mp-CT, multi-phase CT. the entire tumor for the NP, AP, and PVP images was independently conducted by two radiologists (readers 1 and 2 with 5- and 8-year experience in abdominal diagnosis, respectively) using the 3D Slicer software (version 5.0.3). Reader 2 repeated the segmentation a month later. A total of 50 patients were randomly selected from the MRS cohort to assess intra- and intersegmentation reproducibility using dice similarity coefficient (DSC) and intraclass correlation coefficient (ICC). A feature with intraand inter-ICC \geq 0.75 was considered stable. A senior radiologist with 12 years of experience confirmed the segmentation variation. Any disagreement between the readers was discussed until a final consensus was reached.

Radiomics feature extraction

Radiomics features were calculated in accordance with the guidelines of the Image Biomarker Standardization Initiative (https://theibsi.github.io/) (49). All voxels of the tumor region were first uniformly resampled to $1 \times 1 \times 1 \text{ mm}^3$ with the cubic B-spline interpolation algorithm to improve the robustness of feature extraction. Then, a total of 962 radiomics features were extracted from each phase of CT images by using the Pyradiomics package (version 3.0.1), characterizing shape, intensity, and texture patterns of the tumor from the original and derived CT images (processed by the Laplacian of Gaussian or wavelet filter). The detailed extraction parameters and feature types are described in Supplementary Method S1 and Supplementary Table S2.

Radiomics feature selection and signature construction for MRS

Radiomics feature selection and signature construction to distinguish the MRS-high and -low group only used data from the training cohort. To eliminate the dimensions of feature magnitudes, Z-score normalization was applied. The Synthetic Minority Oversampling Technique (SMOTE) strategy was then used to remove the imbalance of the training cohort (40). In the procedure of feature selection, Pearson correlation coefficient (PCC) analysis was first conducted to obtain a feature subset with low redundancy; one of the features with PCC > 0.99 was randomly eliminated from further consideration. Recursive feature elimination (RFE) or Relief algorithm was further used for identifying the well-predictive features. Different classifiers, including random forest (RF), support vector machine (SVM), logistic regression (LR), K-nearest neighbor (KNN), and multilayer perceptron (MLP), were combined with five-fold crossvalidation to obtain the optimal radiomics signature for each CT phase. A mp-CT radiomics signature was explored by fusing the predictive score of the triple-phase CT images. Figure 2B depicts the corresponding analysis flow.

Imaging-based MRS construction for prognosis

Imaging-based MRS (iMRS) from mp-CT imaging was obtained from the predictive score of the optimal radiomics signature and then guided the risk stratification of MRS in subgroups. The cutoff value was defined by the optimal Youden's index of predictive score within the training cohort and then applied to the other validation cohorts. Patients with a score higher than the cutoff value were classified as the high-risk group, and those with a score lower than the cutoff value were classified as the low-risk group. The prognostic value of iMRS risk groups in HCC was validated by Kaplan–Meier survival curves of RFS and OS. Additionally, we explored the advantage of the iMRS in inferring with the benefits of immune checkpoint therapy.

Clinical and Clinical-iMRS score construction for prognosis

Clinicopathological features, including age, gender, hepatitis B surface antigen (HBsAg), alpha-fetoprotein (AFP), alanine aminotransferase (ALT), Child–Pugh class, tumor size, tumor number, vascular invasion, and Barcelona Clinic Liver Cancer (BCLC) stage, were enrolled. In the training cohort, significant variables associated with RFS (p < 0.05) in univariate Cox proportional hazards analysis were further adopted as covariates in a multivariate Cox proportional hazards analysis. The clinical prediction score was thus obtained by the output of the multivariate model to predict the status of RFS and OS. Considering clinical practice, we also analyzed the joint value of iMRS and the significant clinicopathological features in prognosis. A combined Clinical-iMRS nomogram was constructed by using the multivariate Cox proportional hazards analysis to calculate the Clinical-iMRS score.

Statistical analysis

The difference in continuous variables between groups was analyzed by the Mann–Whitney test. Categorical variables were compared by the chi-square test or Fisher's exact test, as appropriate. Radiomics feature extraction and signature development were performed using Python (version 3.7.6) and Pycharm (version 2020.1.5). Area under the receiver operator characteristic (ROC) curve (AUC), sensitivity, specificity, and accuracy were used to evaluate the predictive performance. We used the Delong test to compare the ROC curves of diverse radiomics signatures from the CT phases. For the survival analysis, the Kaplan–Meier curves for risk stratification were statistically tested by the log-rank test. The Harrell concordance index (C-index) was calculated for assessing the prognostic value of the scores. The calibration curve was plotted for the Clinical-iMRS nomogram. Note that a two-tailed p < 0.05 indicated statistical significance.

Results

Patient characteristics

The clinicopathological features of patients from different cohorts are summarized in Table 1. According to the MRS from IHC staining, 24.5% (27/110) and 23.6% (13/55) of patients were stratified into the MRS_{high} group in cohorts 1 and 2, respectively. Except for vascular invasion in cohort 2 (p = 0.014), there was no significant difference between the MRS_{low} and MRS_{high} groups (Table 2). The median follow-up duration was 49.0 months (range, 2–115 months) and 35.7 months (range, 2–107 months), respectively. The median OS and RFS for patients were 49.0 and 12.5 months in cohort 1, respectively. As for cohort 2, the median OS and RFS for patients were 35.7 and 11.7 months, respectively.

Intra- and intersegmentation reproducibility

The mean value and 95% confidence interval (CI) of DSC values [intra-rater DSC: 0.944 (0.936–0.953), 0.946 (0.940–0.953), and 0.946 (0.940–0.952); inter-rater DSC: 0.857 (0.838–0.876), 0.854 (0.842–0.865), and 0.858 (0.845–0.871)] for NP, AP, and PVP images were calculated, respectively. The results indicated that the difference in delineation was relatively small in this work. Furthermore, the median ICC values [intra-rater ICC: 0.967 (0.958–0.973), 0.981 (0.978–0.984), and 0.972 (0.966–0.976); inter-rater ICC: 0.854 (0.833–0.904), 0.915 (0.879–0.927), and 0.921 (0.902–0.928)] for NP, AP, and PVP images illustrated that the radiomics features also presented good reproducibility under the condition of small segmentation difference. 60.0% (577/962),

TABLE 1 Clinicopathological characteristics of patients with HCC included in four cohorts.

Characteristics	Cohort 1	Cohort 2	Surgical resection cohort	Immunotherapy cohort	<i>p</i> -value				
No. of patients	110	55	98	36					
Age (years), median (range)	52 (13-74)	48 (22–76)	54 (20-87)	52 (31–70)					
Gender									
Female	15	4	13	6	0.523				
Male	95	51	85	30					
HBsAg									
Negative	13	7	15	6	0.829				
Positive	97	48	83	30					
AFP (ng/mL), median (range)	267.85 (0-121,000)	290.00 (0-121,000)	213.10 (1-121,000)	19,902.00 (4–121,000)					
ALT (U/L), median (range)	39.00 (0-118.0)	34.00 (8.0–140.0)	34.65 (11.0–389.3)	49.2 (19.6–254.7)					
Tumor size (cm), median (range)	6.75 (1.5–17.0)	7.0 (2.0–19.0)	5.0 (1.5–30.0)	11.2 (3.0–18.5)					
Child-Pugh class									
А	107	54	96	36	0.999				
В-С	3	1	2	0					
Tumor number									
Single	74	40	80	N.A.	0.056				
Multiple	36	15	18	N.A.					
Vascular invasion									
No	96	44	89	9	< 0.0001*				
Yes	14	11	9	27					
BCLC stage	·		·						
0-A	59	33	43	0	< 0.0001*				
В-С	51	22	55	36					

HCC, hepatocellular carcinoma; HBsAg, hepatitis B surface antigen; AFP, α-fetoprotein; ALT, alanine aminotransferase; BCLC, Barcelona Clinic Liver Cancer; N.A., not applicable. *p*-values were analyzed by Fisher's exact test.

TABLE 2 Clinicopathological characteristics of different MRS risk groups.

Characteristics	Cohort 1	(<i>n</i> = 110)		Cohort 2	2 (n = 55)		
Characteristics	MRS low MRS high		<i>p</i> -value	MRS low	MRS high	<i>p</i> -value	
Age (years)				,			
≤48	33	13	0.504	20	9	0.215	
>48	50	14		22	4		
Gender			1				
Female	13	2	0.351	4	0	0.562	
Male	70	25		38	13		
HBsAg	_	1	1	1	1	1	
Negative	9	4	0.732	7	0	0.179	
Positive	74	23		35	13		
AFP (ng/mL)				·			
≤25	24	10	0.476	13	2	0.477	
>25	59	17		29	11		
ALT (U/L)	_	1	1	1	1	1	
≤40	44	14	1.000	30	8	0.511	
>40	39	13		12	5		
Tumor size (cm)		1	1	1	1	1	
≤5	33	12	0.822	18	3	0.328	
>5	50	15		24	10		
Child–Pugh class		1	1	1	1	1	
А	80	27	1.000	42	12	0.236	
B-C	3	0		0	1		
Tumor number	_	1	1	1	1		
Single	56	19	1.000	31	9	0.734	
Multiple	27	9		11	4		
Vascular invasion	·						
No	75	21	0.103	37	7	0.014*	
Yes	8	6		5	6		
BCLC stage	·						
0-A	47	12	0.374	28	5	0.106	
В-С	36	15		14	8		

 $\ensuremath{\textit{p}}\xspace$ values were analyzed by χ^2 test or Fisher's exact test, as appropriate.

HBsAg, hepatitis B surface antigen; AFP, α-fetoprotein; ALT, alanine aminotransferase; BCLC, Barcelona Clinic Liver Cancer. Data are presented as number of patients.

62.0% (596/962), and 64.3% (619/962) of the features from each phase were separately identified as good segmentation stability with ICC \geq 0.75 of both intra- and inter-rater variability. The number of features under different ICC thresholds is presented in Supplementary Table S3.

Predictive performance of the radiomics signatures for MRS

Two NP features (Relief and KNN), 8 AP features (RFE and MLP), and 11 PVP features (Relief and KNN) were identified as

well-predictive through the combination optimization of different feature selection algorithms and classifiers, respectively (Supplementary Table S4). The optimization results of the applied machine learning algorithms are shown in Supplementary Figure S1. According to the AUC and accuracy (Table 3), the optimal radiomics signature from AP images (AUC, 0.769; 95% CI, 0.718–0.820) showed better performance compared with that from NP and PVP images in cohort 2. The corresponding sensitivity, specificity, and accuracy were 69.2%, 73.8%, and 72.7%, respectively. The predictive scores from the NP, AP, and PVP phase were further fused into the mp-CT radiomics signature, which demonstrated a pleasing improvement in performance with AUCs of 0.941 (95% CI, 0.909–0.973) and 0.833 (95% CI, 0.798–0.868) in cohorts 1 and 2, respectively (Figure 3A).

Risk stratification from iMRS

Since the mp-CT radiomics signature provided the best predictive performance in both cohorts (all p < 0.05), iMRS was calculated and showed good net benefit for clinical use within the whole risk threshold range (Figure 3B). In both cohorts 1 and 2, the iMRSs of patients in the pathological MRS_{high} group were significantly higher than those in the pathological MRS_{low} group, (Figures 3C, D), which confirms the good discrimination of MRS groups based on iMRS. Moreover, the iMRSs were positively correlated with the number of tumor-infiltrating CD11b⁺ cells in both cohort 1 (r = 0.5629, p < 0.001; Figure 3E) and cohort 2 (r = 0.4271, p = 0.001; Figure 3F).

Prognostic value of iMRS

The mp-CT radiomics score was calculated and patients were classified into high- and low-risk groups by iMRS with a cutoff value of 0.19. The Kaplan–Meier survival curves showed that patients with high iMRS had shorter OS and RFS in both cohorts 1 and 2 (Figures 4A–D). At the same time, the prognostic value of iMRS was confirmed in the surgical resection cohort. High iMRS indicated

poorer OS (p = 0.009, Figure 4E) and RFS (p < 0.001, Figure 4F). The C-indexes of the iMRS were 0.627, 0.664, and 0.660 for predicting RFS in cohort 1, cohort 2, and the surgical resection cohort, respectively. Correspondingly, the C-indexes were 0.661, 0.649, and 0.648 for predicting OS.

The iMRS of patients in the complete response or partial response (CR/PR) group was significantly higher than those in the progressive disease or stable disease (PD/SD) group (p = 0.030, Figure 4G). High iMRS (odds ratio, 2.311; 95% CI, 1.144–4.672; p = 0.020) in the immunotherapy cohort predicted a higher proportion of CR/PR (vs. PD/SD; AUC, 0.718; 95% CI, 0.686–0.749) in patients who had received anti-PD-1 or anti-PD-L1 treatment. Patients with high iMRS had improved OS than those with low iMRS in the immunotherapy cohort (p = 0.030, Figure 4H).

Prognostic value of clinical score

Clinical variables including Child–Pugh class, tumor number, vascular invasion, and BCLC stage were associated with RFS in univariate analysis (p < 0.001). The multivariate Cox model revealed that Child–Pugh class, tumor number, and vascular invasion were independent negative prognostic factors for RFS in HCC patients (Supplementary Table S5). We output the predicted probability as the clinical score. The prognostic C-indexes associated with RFS were 0.676, 0.654, and 0.610 for cohort 1, cohort 2, and the surgical resection cohort, respectively. The prognostic C-indexes associated with OS were 0.701, 0.647, and 0.667, respectively.

Prognostic value of the Clinical-iMRS score

Multivariate Cox analysis of clinicopathologic characteristics and iMRS for prognosis showed that high Child–Pugh class, tumor number, vascular invasion, and high iMRS were unfavorable predictors for RFS survival of HCC patients (Table 4), resulting in the Clinical-iMRS score. A well-discriminated and calibrated nomogram (Figures 5A–D) was subsequently developed, enhancing

TABLE 3	Predictive	performance of	f the	optimal	radiomics	signatures	identified	from	triple-phase	e CT image	es.
---------	------------	----------------	-------	---------	-----------	------------	------------	------	--------------	------------	-----

Datasets	Phase	AUC (95% CI)	<i>p</i> -value	Sensitivity (%)	Specificity (%)	Accuracy (%)
Cohort 1	NP	0.898 (0.863-0.932)	0.003*	100.0	63.9	72.7
	AP	0.735 (0.653–0.817)	<0.001*	51.9	83.1	75.5
	PVP	0.756 (0.690-0.822)	<0.001*	81.5	61.4	66.4
	mp-CT	0.941 (0.909-0.973)	Ref.	88.9	83.1	84.5
Cohort 2	NP	0.680 (0.631-0.730)	<0.001*	76.9	57.1	61.8
	AP	0.769 (0.718-0.820)	0.026*	69.2	73.8	72.7
	PVP	0.734 (0.686-0.783)	<0.001*	76.9	57.1	61.8
	mp-CT	0.833 (0.798-0.868)	Ref.	76.9	71.4	72.7

p-values were analyzed by the Delong test.

AUC, area under the curve; CI, confidence interval; NP, noncontrast phase; AP, arterial phase; PVP, portal venous phase; mp-CT, multiple-phase CT.



Performance of the radiomics signature. (A) ROC curves of the optimal mp-CT radiomics signature in cohorts 1 and 2. (B) Decision curve analysis. (C, D) iMRS of patients in pMRS-high or -low groups in cohort 1 (C) and cohort 2 (D). Mann–Whitney test, ***p < 0.001. (E, F) The correlations between the number of tumor-infiltrating CD11b⁺ cells and iMRS in cohort 1 (E) and cohort 2 (F). Spearman correlation analysis. ROC, receiver operator characteristic; pMRS, pathological MRS.

predictive accuracy compared to both iMRS and clinical scores. The C-indexes for 1-, 3-, and 5-year RFS prediction were 0.729, 0.709, and 0.713 for cohort 1, cohort 2, and the surgical resection cohort (Figures 5E-G), respectively. The advantage was also observed in OS with C-indexes of 0.770, 0.697, and 0.755, respectively. The Kaplan-Meier survival curves revealed that the nomogram showed notable capacity in predicting RFS and OS among postsurgical HCC patients (Figure 6).



FIGURE 4

Prognostic and clinical value of the iMRS. (A–F) Overall survival and recurrence-free survival of patients relative to iMRS in cohort 1, cohort 2, and the surgical resection cohort. (G) iMRS of patients with complete CR/PR or PD/SD to anti-PD-1/PD-L1 therapy in the immunotherapy cohort. Mann– Whitney test, *p < 0.05. (H) Overall survival of patients relative to iMRS in immunotherapy cohort. CR, complete response; PR, partial response; PD, progressive disease; SD, stable disease.

Discussion

Myeloid cells are a group of innate immune cells in the TME, which play a critical role in tumor initiation, progression, and therapy response in HCC. In this study, we sought to construct a CT-based radiomics model to predict MRS, to examine the prognostic value of iMRS, and to assess the association of iMRS with the outcome of patients treated with anti-PD-1 and anti-PD-L1 in HCC.

Compared with the commonly used TNM staging, various studies have shown that the type, density, and location of immune cells have superior prognostic value (50-52). Tumor-associated myeloid cells are important regulators and prognostic factors in tumor tissues. The myeloid-specific MRS derived from

Chausatauistiaa	Univariate (Cox analysis	Multivariable Cox analysis		
Characteristics	HR (95% CI)	<i>p</i> -value	HR (95% CI)	<i>p</i> -value	
Age	1.005 (0.983-1.028)	0.678	_	-	
Gender	1 (0.507–1.974)	0.999	-	-	
HBsAg	1.503 (0.646-3.495)	0.344	_	_	
AFP group	1.133 (0.664–1.932)	0.648	_	-	
ALT group	1.017 (0.616–1.677)	0.948	_	_	
Child-Pugh class	16.040 (3.214-80.090)	<0.001*	10.375 (1.983-54.276)	0.006*	
Tumor size group	1.143 (0.685–1.909)	0.608	_	_	
Tumor number	2.607 (1.545-4.400)	<0.001*	3.183 (1.817-5.576)	<0.001*	
Vascular invasion	3.487 (1.754-6.935)	<0.001*	4.519 (2.200-9.285)	<0.001*	
BCLC stage	2.433 (1.465-4.042)	<0.001*	_	-	
iMRS	2.311 (1.144-4.672)	0.020*	3.321 (1.579-6.986)	0.002*	

TABLE 4 Univariate and multivariate Cox proportional hazards analysis of factors associated with recurrence.

HBsAg, hepatitis B surface antigen; AFP, α -fetoprotein; ALT, alanine aminotransferase; BCLC, Barcelona Clinic Liver Cancer; HR, hazard ratio; CI, confidence interval. Characteristics with a p < 0.05 are displayed in bold.

the CD169 and CD11b expression in HCC has been proven to be associated with the immune tolerance of CD8⁺ T cells, as well as the prognosis of post-surgery HCC (8). It is known that CD11b is often expressed by immunosuppressive myeloid cells in the blood and tumors, promoting tumor sensitivity to checkpoint blockade (53). This phenomenon has been confirmed in the latest study (54). As a specific marker, CD169 was useful in identifying suppressive tumor-associated macrophages in breast and endometrial cancers (55). Hence, MRS could provide a multidimensional tumor measure using various myeloid contextures. However, the evaluation of myeloid cells in tumor tissues usually requires IHC staining, which must be performed after surgery and specimen collection. The complexity and low efficiency of current methods limit the application of myeloid markers. Additionally, because of the heterogeneity and plasticity of myeloid cells in the TME, multiple antibodies are involved in the staining process, which can easily lead to statistical errors.

Our study successfully constructed a predictive model for MRS by an mp-CT radiomics signature, confirming the accuracy of iMRS and its noninvasive capacity for differentiating MRS in HCC. The derived iMRS was significantly higher in patients from the pathological MRShigh group and was positively correlated with the number of tumor-infiltrating CD11b⁺ cells. According to a previous study (56), image-derived textural diversity might possibly reflect increased immune cell infiltration, which increases tumor heterogeneity. Since the optimal mp-CT radiomics signature was constructed using multiple-phase CT images, the improved performance of the fusion model confirmed that the differences in gray levels across different phases could provide deeper supplementary information for evaluating the local myeloid response at the pre-surgery stage. In particular, the AUC of the radiomics signature from AP images was superior to that from NP and PVP images in cohort 2, which is consistent with the fact that arterial phase intensification is obvious and can present high signal characteristics (57). From the perspective of pathology, the outer layer of the tumor envelope in HCC patients presents relatively abundant compressed blood vessels and new bile ducts (58). The different blood supply of focal liver lesions is the basis of CT diagnosis and differential diagnosis. In general, the proposed method avoids the complex process of immunostaining and offers a noninvasive, efficient tool for calculating MRS.

Before the radiomics analysis of MRS, voxel resampling, graylevel discretization, and SMOTE strategy were adopted successively to reduce the data heterogeneity. SMOTE was considered the most prominent method for handling unbalanced data. As emphasized in previous studies (21), accurate segmentation of tumors remains a challenge affecting the popularization of radiomics models. Multiple segmentation by different radiologists can be used as a reliable method. Moreover, the inter- and intra-observer variability of manual segmentation and robustness of radiomics features were then investigated. The DSC and ICC values were higher than 0.85 in various segmentation scenarios, which demonstrated pleasing reproducibility in tumor delineation and made sure that our analysis was based on the accurate segmentation of tumor areas. From the perspective of features contained in the optimal signatures, we found that the features selected were all from the filtered CT images, and most of them were high-dimensional texture features. The data indicated that the processed images could amplify the differences between features, thereby providing more powerful texture information to distinguish the MRS groups.

The strength of our iMRS is that it shows good potential for predicting the survival of OS and RFS in patients with HCC as an independent prognostic biomarker. Kaplan–Meier survival curves reflected that there were significant differences between the two risk groups. Extensive studies have also reported on the similar association of tumor immunological infiltration with radiomics features and have accordingly verified the prognostic value of the noninvasive radiomics markers (33, 35, 36, 59, 60). Since the



resection cohort (G). C-index, Harrell concordance index.

predictive C-indexes of iMRS for RFS and OS in different subsets were comparable to or worse than that of clinical scores, the Clinical-iMRS score significantly improved the predictive value for RFS. Our study revealed that the radiomics signature was an important supplementation to clinical features in survival analysis for HCC patients. Previous studies have found similar results (24, 25, 30, 34, 38, 61). It is an important trend for clinical utility to provide a comprehensive evaluation system using multiple factors that reflect the different biological properties of tumors. Since the objective response rate of immunotherapy in most cancers is still relatively low (46-48), identifying reliable biomarkers from immune cells, such as the percentage of CD8⁺ cells and TLS structure, to quantify the TME associated with immunotherapy response, is still an emerging area of oncology research (33, 40, 62). In this study, a high iMRS predicted a higher proportion of objective response in patients treated with anti-PD-1 and PD-L1. Patients in the iMRS-high group had improved OS compared to those with low iMRS in the immunotherapy cohort. The results



model (high risk or low risk) in cohort 1, cohort 2, and the surgical resection cohort.

demonstrated that iMRS was significantly associated with the response of HCC patients to immune checkpoint therapy, which could identify patients most likely to benefit from anti-PD-1 and PD-L1 therapy. In mechanism, the MRS was useful to describe the immune microenvironment of HCC, and HCC patients with high MRS displayed immunosuppressive TMEs (8). On the one hand, MRS_{high} tumors are associated with CD8⁺ T-cell exhaustion, especially PD-1 expression on infiltrating T cells. On the other hand, the elevation of PD-L1 expression on M φ s and tumor cells is

significantly associated with high MRS. Therefore, we suggest that noninvasive iMRS can help promote the surveillance of the TME and the translational application of immunotherapy effectively and rationally.

Our study also has several limitations. First, the data were retrospectively collected from a single center, and the amount of data was small. It is important to note that we grouped the HCC patients over different time periods to provide a longitudinal time design to verify the generalization performance of the model. This also compensates for the shortcomings of a single center. Of course, more data from other centers will be collected in the future to further prospectively validate our findings. Second, despite our efforts to select relatively stable radiomics features through intraand inter-ICC analysis, an automated and accurate segmentation strategy is necessary and can improve the efficiency of radiomics analysis. Third, while the radiomics signature was associated with CD11b⁺ cells' infiltration, the overview of the immune infiltration pattern from single-cell RNA sequencing could better explain the biological meaning of radiomics patterns of our iMRS. Lastly, multiomics fusion holds great potential for future development, and we hope to combine other multidimensional data with iMRS to better characterize the TME of patients with HCC and assist clinical treatment decision-making (63).

Conclusions

Our study constructed an accurate and efficient predictive model for MRS, confirmed the correlation of the radiomics signature and tumor-infiltrating myeloid cells, further revealed the prognostic value of iMRS for survival, and assessed the association of iMRS with the outcome of anti-PD-1 and PD-L1 therapy in HCC. Our study provided a promising and noninvasive tool to evaluate the TME and to assist immunotherapeutic decisions in clinical trials, thus enabling a more tailored therapeutic approach with improved outcomes for HCC patients.

Data availability statement

The original contributions presented in the study are included in the article/Supplementary Material. Further inquiries can be directed to the corresponding authors.

Ethics statement

The studies involving humans were approved by the ethics committee of the Sun Yat-sen University Cancer Center. The studies were conducted in accordance with the local legislation and institutional requirements. The participants provided their written informed consent to participate in this study. Written informed consent was obtained from the individual(s) for the publication of any potentially identifiable images or data included in this article.

Author contributions

KP: Methodology, Project administration, Writing – original draft, Investigation. XZ: Methodology, Project administration, Writing – original draft. ZL: Methodology, Writing – original draft, Investigation. YCW: Methodology, Investigation, Writing original draft. H-WS: Conceptualization, Funding acquisition, Investigation, Methodology, Writing - original draft. WZ: Formal analysis, Funding acquisition, Investigation, Methodology, Writing - original draft. JP: Data curation, Formal analysis, Writing - original draft. X-YZ: Formal analysis, Methodology, Writing - original draft. XW: Methodology, Project administration, Writing - original draft. XY: Investigation, Project administration, Writing - original draft. CW: Investigation, Methodology, Writing - original draft. YLW: Investigation, Methodology, Project administration, Writing - original draft. XL: Investigation, Methodology, Writing - original draft. DL: Investigation, Project administration, Validation, Writing original draft. MZ: Data curation, Formal analysis, Funding acquisition, Methodology, Writing - original draft. JX: Conceptualization, Investigation, Writing - review & editing. LZ: Funding acquisition, Supervision, Writing - review & editing. YZ: Writing - original draft. LL: Funding acquisition, Supervision, Writing - review & editing.

Funding

The author(s) declare financial support was received for the research, authorship, and/or publication of this article. This work was supported by project grants from the National Natural Science Foundation of China (82230067, 82272103, and 82103306) and the Natural Science Foundation of Guangdong Province of China (2022B1515020010 and 2021A1515220183).

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Supplementary material

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fimmu.2024.1493735/ full#supplementary-material

References

1. Rumgay H, Arnold M, Ferlay J, Lesi O, Cabasag CJ, Vignat J, et al. Global burden of primary liver cancer in 2020 and predictions to 2040. *J Hepatol.* (2022) 77:1598–606. doi: 10.1016/j.jhep.2022.08.021

2. Llovet JM, Kelley RK, Villanueva A, Singal AG, Pikarsky E, Roayaie S, et al. Hepatocellular carcinoma. *Nat Rev Dis Primers.* (2021) 7:6. doi: 10.1038/s41572-020-00240-3

3. Donne R, Lujambio A. The liver cancer immune microenvironment: Therapeutic implications for hepatocellular carcinoma. *Hepatology*. (2023) 77:1773–96. doi: 10.1002/hep.32740

4. Gabrielson A, Wu Y, Wang H, Jiang J, Kallakury B, Gatalica Z, et al. Intratumoral CD3 and CD8 T-cell densities associated with relapse-free survival in HCC. *Cancer Immunol Res.* (2016) 4:419–30. doi: 10.1158/2326-6066.Cir-15-0110

5. Lim CJ, Lee YH, Pan L, Lai L, Chua C, Wasser M, et al. Multidimensional analyses reveal distinct immune microenvironment in hepatitis B virus-related hepatocellular carcinoma. *Gut.* (2019) 68:916–27. doi: 10.1136/gutjnl-2018-316510

6. Weng J, Liu S, Zhou Q, Xu W, Xu M, Gao D, et al. Intratumoral PPT1-positive macrophages determine immunosuppressive contexture and immunotherapy response in hepatocellular carcinoma. *J Immunother Cancer.* (2023) 11:e006655. doi: 10.1136/ jitc-2022-006655

7. Zhang Q, He Y, Luo N, Patel SJ, Han Y, Gao R, et al. Landscape and dynamics of single immune cells in hepatocellular carcinoma. *Cell.* (2019) 179:829-45.e20. doi: 10.1016/j.cell.2019.10.003

8. Wu C, Lin J, Weng Y, Zeng DN, Xu J, Luo S, et al. Myeloid signature reveals immune contexture and predicts the prognosis of hepatocellular carcinoma. *J Clin Invest.* (2020) 130:4679–93. doi: 10.1172/jci135048

9. Kurebayashi Y, Matsuda K, Ueno A, Tsujikawa H, Yamazaki K, Masugi Y, et al. Immunovascular classification of HCC reflects reciprocal interaction between immune and angiogenic tumor microenvironments. *Hepatology*. (2022) 75:1139–53. doi: 10.1002/hep.32201

10. Maestri E, Kedei N, Khatib S, Forgues M, Ylaya K, Hewitt SM, et al. Spatial proximity of tumor-immune interactions predicts patient outcome in hepatocellular carcinoma. *Hepatology.* (2024) 79:768–79. doi: 10.1097/hep.0000000000000000

11. Engblom C, Pfirschke C, Pittet MJ. The role of myeloid cells in cancer therapies. *Nat Rev Cancer.* (2016) 16:447–62. doi: 10.1038/nrc.2016.54

12. Mantovani A, Marchesi F, Jaillon S, Garlanda C, Allavena P. Tumor-associated myeloid cells: diversity and therapeutic targeting. *Cell Mol Immunol.* (2021) 18:566–78. doi: 10.1038/s41423-020-00613-4

13. Veglia F, Sanseviero E, Gabrilovich DI. Myeloid-derived suppressor cells in the era of increasing myeloid cell diversity. *Nat Rev Immunol.* (2021) 21:485–98. doi: 10.1038/s41577-020-00490-y

14. Molgora M, Esaulova E, Vermi W, Hou J, Chen Y, Luo J, et al. TREM2 modulation remodels the tumor myeloid landscape enhancing anti-PD-1 immunotherapy. *Cell.* (2020) 182:886–900.e17. doi: 10.1016/j.cell.2020.07.013

15. Liao J, Zeng DN, Li JZ, Hua QM, Huang CX, Xu J, et al. Type I IFNs repolarized a CD169(+) macrophage population with anti-tumor potentials in hepatocellular carcinoma. *Mol Ther.* (2022) 30:632–43. doi: 10.1016/j.ymthe.2021.09.021

16. Nalio Ramos R, Missolo-Koussou Y, Gerber-Ferder Y, Bromley CP, Bugatti M, Nunez NG, et al. Tissue-resident FOLR2(+) macrophages associate with CD8(+) T cell infiltration in human breast cancer. *Cell.* (2022) 185:1189–207.e25. doi: 10.1016/j.cell.2022.02.021

17. Vayrynen JP, Haruki K, Vayrynen SA, Lau MC, Dias Costa A, Borowsky J, et al. Prognostic significance of myeloid immune cells and their spatial distribution in the colorectal cancer microenvironment. *J Immunother Cancer*. (2021) 9:e002297. doi: 10.1136/jitc-2020-002297

18. Lesterhuis WJ, Bosco A, Millward MJ, Small M, Nowak AK, Lake RA. Dynamic versus static biomarkers in cancer immune checkpoint blockade: unravelling complexity. *Nat Rev Drug Discov*. (2017) 16:264–72. doi: 10.1038/nrd.2016.233

19. Tomaszewski MR, Gillies RJ. The biological meaning of radiomic features. Radiology. (2021) 298:505-16. doi: 10.1148/radiol.2021202553

20. Lambin P, Rios-Velazquez E, Leijenaar R, Carvalho S, van Stiphout RG, Granton P, et al. Radiomics: extracting more information from medical images using advanced feature analysis. *Eur J Cancer*. (2012) 48:441–6. doi: 10.1016/j.ejca.2011.11.036

21. Kumar V, Gu Y, Basu S, Berglund A, Eschrich SA, Schabath MB, et al. Radiomics: the process and the challenges. *Magn Reson Imaging*. (2012) 30:1234–48. doi: 10.1016/j.mri.2012.06.010

22. Gillies RJ, Kinahan PE, Hricak H. Radiomics: images are more than pictures, they are data. *Radiology.* (2016) 278:563–77. doi: 10.1148/radiol.2015151169

23. Ji GW, Zhu FP, Xu Q, Wang K, Wu MY, Tang WW, et al. Radiomic features at contrast-enhanced CT predict recurrence in early stage hepatocellular carcinoma: A multi-institutional study. *Radiology*. (2020) 294:568–79. doi: 10.1148/radiol.2020191470

24. Jin Z, Chen L, Zhong B, Zhou H, Zhu H, Zhou H, et al. Machine-learning analysis of contrast-enhanced computed tomography radiomics predicts patients with hepatocellular carcinoma who are unsuitable for initial transarterial

chemoembolization monotherapy: A multicenter study. Transl Oncol. (2021) 14:101034. doi: 10.1016/j.tranon.2021.101034

25. Mao Y, Wang J, Zhu Y, Chen J, Mao L, Kong W, et al. Gd-EOB-DTPA-enhanced MRI radiomic features for predicting histological grade of hepatocellular carcinoma. *Hepatobiliary Surg Nutr.* (2022) 11:13–24. doi: 10.21037/hbsn-19-870

26. Feng Z, Li H, Liu Q, Duan J, Zhou W, Yu X, et al. CT radiomics to predict macrotrabecular-massive subtype and immune status in hepatocellular carcinoma. *Radiology.* (2023) 307:e221291. doi: 10.1148/radiol.221291

27. Xia TY, Zhou ZH, Meng XP, Zha JH, Yu Q, Wang W, et al. Predicting microvascular invasion in hepatocellular carcinoma using CT-based radiomics model. *Radiology*. (2023) 307:e222729. doi: 10.1148/radiol.222729

28. Summers RM. Radiomics to predict microvascular invasion in hepatocellular carcinoma: A promising biomarker for tumor recurrence. *Radiology*. (2023) 307: e230657. doi: 10.1148/radiol.230657

29. Lefebvre TL, Ueno Y, Dohan A, Chatterjee A, Vallieres M, Winter-Reinhold E, et al. Development and validation of multiparametric MRI-based radiomics models for preoperative risk stratification of endometrial cancer. *Radiology.* (2022) 305:375–86. doi: 10.1148/radiol.212873

30. Gu B, Meng M, Xu M, Feng DD, Bi L, Kim J, et al. Multi-task deep learningbased radiomic nomogram for prognostic prediction in locoregionally advanced nasopharyngeal carcinoma. *Eur J Nucl Med Mol Imaging*. (2023) 50:3996–4009. doi: 10.1007/s00259-023-06399-7

31. Dercle L, Fronheiser M, Rizvi NA, Hellmann MD, Maier S, Hayes W, et al. Baseline radiomic signature to estimate overall survival in patients with NSCLC. J Thorac Oncol.. (2023) 18:587–98. doi: 10.1016/j.jtho.2022.12.019

32. Ramtohul T, Djerroudi L, Lissavalid E, Nhy C, Redon L, Ikni L, et al. Multiparametric MRI and radiomics for the prediction of HER2-zero, -low, and -positive breast cancers. *Radiology.* (2023) 308:e222646. doi: 10.1148/radiol.222646

33. Sun R, Limkin EJ, Vakalopoulou M, Dercle L, Champiat S, Han SR, et al. A radiomics approach to assess tumour-infiltrating CD8 cells and response to anti-PD-1 or anti-PD-L1 immunotherapy: an imaging biomarker, retrospective multicohort study. *Lancet Oncol.* (2018) 19:1180–91. doi: 10.1016/s1470-2045(18)30413-3

34. Chen S, Feng S, Wei J, Liu F, Li B, Li X, et al. Pretreatment prediction of immunoscore in hepatocellular cancer: a radiomics-based clinical model based on Gd-EOB-DTPA-enhanced MRI imaging. *Eur Radiol.* (2019) 29:4177–87. doi: 10.1007/ s00330-018-5986-x

35. Jiang Y, Wang H, Wu J, Chen C, Yuan Q, Huang W, et al. Noninvasive imaging evaluation of tumor immune microenvironment to predict outcomes in gastric cancer. *Ann Oncol.* (2020) 31:760–8. doi: 10.1016/j.annonc.2020.03.295

36. Li J, Zhang L, Xing H, Geng Y, Lv S, Luo X, et al. The absence of intratumoral tertiary lymphoid structures is associated with a worse prognosis and mTOR signaling activation in hepatocellular carcinoma with liver transplantation: A multicenter retrospective study. *Adv Sci (Weinh).* (2024) 11:e2309348. doi: 10.1002/ advs.202309348

37. He B, Dong D, She Y, Zhou C, Fang M, Zhu Y, et al. Predicting response to immunotherapy in advanced non-small-cell lung cancer using tumor mutational burden radiomic biomarker. *J Immunother Cancer.* (2020) 8:e000550. doi: 10.1136/jitc-2020-000550

38. Yuan G, Song Y, Li Q, Hu X, Zang M, Dai W, et al. Development and validation of a contrast-enhanced CT-based radiomics nomogram for prediction of therapeutic efficacy of anti-PD-1 antibodies in advanced HCC patients. *Front Immunol.* (2020) 11:613946. doi: 10.3389/fimmu.2020.613946

39. Ho LM, Lam SK, Zhang J, Chiang CL, Chan AC. Association of multiphasic MR-based radiomic and dosimetric features with treatment response in unresectable hepatocellular carcinoma patients following novel sequential TACE-SBRT-immunotherapy. *Cancers (Basel).* (2023) 15:1105. doi: 10.3390/ cancers15041105

40. Jiang Y, Zhou K, Sun Z, Wang H, Xie J, Zhang T, et al. Non-invasive tumor microenvironment evaluation and treatment response prediction in gastric cancer using deep learning radiomics. *Cell Rep Med.* (2023) 4:101146. doi: 10.1016/j.xcrm.2023.101146

41. Yu G, Zhang Z, Eresen A, Hou Q, Garcia EE, Yu Z, et al. MRI radiomics to monitor therapeutic outcome of sorafenib plus IHA transcatheter NK cell combination therapy in hepatocellular carcinoma. *J Transl Med.* (2024) 22:76. doi: 10.1186/s12967-024-04873-w

42. Zhang J, Wu Z, Zhang X, Liu S, Zhao J, Yuan F, et al. Machine learning: an approach to preoperatively predict PD-1/PD-L1 expression and outcome in intrahepatic cholangiocarcinoma using MRI biomarkers. *ESMO Open.* (2020) 5: e000910. doi: 10.1136/esmoopen-2020-000910

43. Gao L, Jiang W, Yue Q, Ye R, Li Y, Hong J, et al. Radiomic model to predict the expression of PD-1 and overall survival of patients with ovarian cancer. *Int Immunopharmacol.* (2022) 113:109335. doi: 10.1016/j.intimp.2022.109335

44. Tao YY, Shi Y, Gong XQ, Li L, Li ZM, Yang L, et al. Radiomic analysis based on magnetic resonance imaging for predicting PD-L2 expression in hepatocellular carcinoma. *Cancers (Basel).* (2023) 15:365. doi: 10.3390/cancers15020365

45. Jiang Y, Zhang Q, Hu Y, Li T, Yu J, Zhao L, et al. Immunoscore signature: A prognostic and predictive tool in gastric cancer. *Ann Surg.* (2018) 267:504–13. doi: 10.1097/sla.00000000002116

46. El-Khoueiry AB, Sangro B, Yau T, Crocenzi TS, Kudo M, Hsu C, et al. Nivolumab in patients with advanced hepatocellular carcinoma (CheckMate 040): an open-label, non-comparative, phase 1/2 dose escalation and expansion trial. *Lancet.* (2017) 389:2492–502. doi: 10.1016/s0140-6736(17)31046-2

47. Zhu AX, Finn RS, Edeline J, Cattan S, Ogasawara S, Palmer D, et al. Pembrolizumab in patients with advanced hepatocellular carcinoma previously treated with sorafenib (KEYNOTE-224): a non-randomised, open-label phase 2 trial. *Lancet Oncol.* (2018) 19:940–52. doi: 10.1016/s1470-2045(18)30351-6

48. Li Q, Han J, Yang Y, Chen Y. PD-1/PD-L1 checkpoint inhibitors in advanced hepatocellular carcinoma immunotherapy. *Front Immunol.* (2022) 13:1070961. doi: 10.3389/fimmu.2022.1070961

49. Zwanenburg A, Vallieres M, Abdalah MA, Aerts H, Andrearczyk V, Apte A, et al. The image biomarker standardization initiative: standardized quantitative radiomics for high-throughput image-based phenotyping. *Radiology*. (2020) 295:328–38. doi: 10.1148/radiol.2020191145

50. Galon J, Costes A, Sanchez-Cabo F, Kirilovsky A, Mlecnik B, Lagorce-Pages C, et al. Type, density, and location of immune cells within human colorectal tumors predict clinical outcome. *Science*. (2006) 313:1960–4. doi: 10.1126/science.1129139

51. Galon J, Mlecnik B, Bindea G, Angell HK, Berger A, Lagorce C, et al. Towards the introduction of the 'Immunoscore' in the classification of Malignant tumours. *J Pathol.* (2014) 232:199–209. doi: 10.1002/path.4287

52. Pages F, Mlecnik B, Marliot F, Bindea G, Ou FS, Bifulco C, et al. International validation of the consensus Immunoscore for the classification of colon cancer: a prognostic and accuracy study. *Lancet.* (2018) 391:2128–39. doi: 10.1016/S0140-6736(18)30789-X

53. Bronte V, Brandau S, Chen SH, Colombo MP, Frey AB, Greten TF, et al. Recommendations for myeloid-derived suppressor cell nomenclature and characterization standards. *Nat Commun.* (2016) 7:12150. doi: 10.1038/ncomms12150

54. Panni RZ, Herndon JM, Zuo C, Hegde S, Hogg GD, Knolhoff BL, et al. Agonism of CD11b reprograms innate immunity to sensitize pancreatic cancer to immunotherapies. *Sci Transl Med.* (2019) 11:eaau9240. doi: 10.1126/scitranslmed.aau9240

55. Cassetta L, Fragkogianni S, Sims AH, Swierczak A, Forrester LM, Zhang H, et al. Human tumor-associated macrophage and monocyte transcriptional landscapes reveal cancer-specific reprogramming, biomarkers, and therapeutic targets. *Cancer Cell.* (2019) 35:588–602.e10. doi: 10.1016/j.ccell.2019.02.009

56. Narang S, Kim D, Aithala S, Heimberger AB, Ahmed S, Rao D, et al. Tumor image-derived texture features are associated with CD3 T-cell infiltration status in glioblastoma. *Oncotarget*. (2017) 8:101244–54. doi: 10.18632/oncotarget.20643

57. Cannella R, Dioguardi Burgio M, Beaufrere A, Trapani L, Paradis V, Hobeika C, et al. Imaging features of histological subtypes of hepatocellular carcinoma: Implication for LI-RADS. *JHEP Rep.* (2021) 3:100380. doi: 10.1016/j.jhepr.2021.100380

58. Hu YX, Shen JX, Han J, Mao SY, Mao RS, Li Q, et al. Diagnosis of nonhepatocellular carcinoma Malignancies in patients with risks for hepatocellular carcinoma: CEUS LI-RADS versus CT/MRI LI-RADS. *Front Oncol.* (2021) 11:641195. doi: 10.3389/fonc.2021.641195

59. Sun R, Sundahl N, Hecht M, Putz F, Lancia A, Rouyar A, et al. Radiomics to predict outcomes and abscopal response of patients with cancer treated with immunotherapy combined with radiotherapy using a validated signature of CD8 cells. *J Immunother Cancer.* (2020) 8:e001429. doi: 10.1136/jitc-2020-001429

60. Sun R, Lerousseau M, Briend-Diop J, Routier E, Roy S, Henry T, et al. Radiomics to evaluate interlesion heterogeneity and to predict lesion response and patient outcomes using a validated signature of CD8 cells in advanced melanoma patients treated with anti-PD1 immunotherapy. *J Immunother Cancer.* (2022) 10:e004867. doi: 10.1136/jitc-2022-004867

61. Wong CW, Filippov A, Bonjoc KJC, Brown C, Badie B, Chaudhry A. Explainable prediction of survival using clinical, molecular, and radiomic profiles in recurrent high-grade glioma patients treated with CAR T-cell therapy. *J Clin Oncol.* (2021) 39:104. doi: 10.1200/JCO.2021.39.15_suppl.104

62. Petrizzo A, Buonaguro L. Application of the Immunoscore as prognostic tool for hepatocellular carcinoma. *J Immunother Cancer*. (2016) 4:71. doi: 10.1186/s40425-016-0182-5

63. Kang W, Qiu X, Luo Y, Luo J, Liu Y, Xi J, et al. Application of radiomics-based multiomics combinations in the tumor microenvironment and cancer prognosis. *J Transl Med.* (2023) 21:598. doi: 10.1186/s12967-023-04437-4