Check for updates

OPEN ACCESS

EDITED BY Guendalina Froechlich, University of Naples Federico II, Italy

REVIEWED BY Shaosen Zhang, Chinese Academy of Medical Sciences and Peking Union Medical College, China Pingwei Xu, Wenzhou Medical University, China

CORRESPONDENCE Weijie Qiang i qiangwj92@163.com Weihong Ge i 6221230@sina.com

RECEIVED 12 October 2024 ACCEPTED 06 November 2024 PUBLISHED 25 November 2024

CITATION

Wang L, Wang J, Qiang W and Ge W (2024) Stanniocalcin-1 in tumor immunity: acts via macrophages. *Front. Immunol.* 15:1510182. doi: 10.3389/fimmu.2024.1510182

COPYRIGHT

© 2024 Wang, Wang, Qiang and Ge. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Stanniocalcin-1 in tumor immunity: acts via macrophages

Lele Wang^{1,2}, Jianjun Wang^{1,2}, Weijie Qiang^{3*} and Weihong Ge^{3*}

¹Department of Pharmacy, Nanjing Drum Tower Hospital, School of Basic Medicine and Clinical Pharmacy, China Pharmaceutical University, Nanjing, Jiangsu, China, ²School of Basic Medicine and Clinical Pharmacy, China Pharmaceutical University, Nanjing, Jiangsu, China, ³Department of Pharmacy, Nanjing Drum Tower Hospital, Affiliated Hospital of Medical School, Nanjing University, Nanjing, Jiangsu, China

Tumor immune escape has become a research hotspot in the field of cancer immunotherapy. Tumor-associated macrophages (TAMs) are the key component of tumor microenvironment, which play a pivotal role in tumor immune escape by regulating the immunity checkpoints, inhibiting the activity of T lymphocytes and natural killer (NK) cells, and modulating proportion of different T cells. Stanniocalcin-1(STC1) is ubiquitously expressed in human body, which is proven to involve with tumor progression and clinical prognosis. Recently, STC1 is implicated in tumor microenvironment as a phagocytosis checkpoint, as well as regulates the immunity via macrophages. In the review, we discussed the role of STC1 and TAMs in tumor immunity and their crosstalk, hoping to provide references for the research of STC1 in tumor immunotherapy.

KEYWORDS

tumor immunity, tumor-associated macrophages, Stanniocalcin-1, immunotherapy, tumor microenvironment

1 Introduction

Tumor is one of the leading causes of death worldwide. In 2022, there were an estimated 20 million cases of cancer diagnosed and 9.7 million deaths due to cancer globally, which severely threatens human life and health (1). Traditional treatments for cancer include surgery, radiotherapy, and chemotherapy. However, these options, alone or in combination, are not effective and there is a pressing need for therapeutic progress. In recent years, molecular targeted therapy and immunotherapy have gradually attracted researchers' interest in the field of oncology. Unfortunately, in clinical practice, only approximately 20%-40% of patients could benefit from immunotherapy, and the majority of patients would appear drug resistance and adverse drug reactions (2). "Tumor immune escape" is considered as the main reason for the immunotherapy failure, tumor cells evade the attack of immune system by decreasing tumor immunogenicity, regulating the infiltration of immune cells, interfering with immune checkpoints, destroying

the recognition function of immune cells, recruiting immunosuppressive cells, etc (3). Accordingly, the research and identification of underlying genes and molecules involved in tumor immune escape is of great significance to decrease drug resistance and improve the efficacy of immunotherapy.

Tumor progression is the crosstalk between cancer cells and their living environment-the tumor microenvironment (TME), which consists of immune cells, blood vessels, fibroblasts, and mesenchymal stromal cells, etc (4). Hence, TME greatly influences the efficacy of immunotherapy (5). As the key component of TME, macrophages are recruited around solid tumors via tumor cell-derived chemokines, thus are named as tumor-associated macrophages (TAMs) (6). It has been found that high TAMs infiltration in tumors is typically associated with poor prognosis and tumor progression, suggesting that TAMs correlate with tumorigenesis and development (7, 8). Reportedly, TAMs can be involved in tumor immune escape by regulating PD-1/PD-L1 axis (9, 10), inhibiting the activity of T lymphocytes and natural killer (NK) cells (11), as well as modulating the conversion of T cells into regulatory T cells (Tregs) (12). In conclusion, TAMs play a pivotal role in tumor immune surveillance and immune escape, so the discovery of underlying molecular mechanisms regulating TAMs function is of great significance for tumor immunotherapy strategies.

Stanniocalcin-1 (STC-1), a glycoprotein hormone first identified in bony fish, relatives to calcium and phosphorus metabolism (13). STC1 is ubiquitously expressed in human body, encompassing the brain, thyroid, spleen, thymus, parathyroid glands, lungs, heart, and skeletal muscle, and acts through autocrine and paracrine (14). So far, multiple experimental studies have demonstrated that STC1 affected cancer cell proliferation (15), migration (16), metastasis (17), apoptosis (18), stem cell properties (19), epithelial-mesenchymal transition (EMT) (20), etc. Moreover, a number of clinical studies have validated that STC1 is aberrantly expressed in diverse tumor tissues and associated with patients prognosis (13, 21, 22). In addition, STC1 is involved in many cancer-related signaling pathways, such as notch1-sex determining region Y-box 2 (Notch1-SOX2) (23), extracellular signal-regulated kinases1/2 (ERK1/2) (24), nuclear factor-kappaB (NF-KB) (17) and jun-N-terminal kinase (JNK) (25) signaling pathways, etc. In brief, STC1 is associated with multiple physiological and pathophysiological processes of tumorigenesis and progression.

In recent years, some studies have found that STC1 is implicated in tumor microenvironment as a phagocytosis checkpoint. In detail, STC1 participates in tumor immunity by affecting the TME, participating in the EMT, and interfering the phagocytic signals. Moreover, multiple researches have demonstrated that STC1 serves as a regulator of tumor immunity affecting differentiation, activation, inflammatory response, and antigen presentation of macrophages. In the review, we introduced biological function and expression pattern of STC1, discussed the role of STC1 and TAMs in tumor immunity, explored how STC1 regulates the function of macrophages. In addition, we proposed the future development directions and prospects for immunotherapy, hoping to provide references for the research of STC1 in tumor immunotherapy.

2 Biological function and expression pattern of STC1

2.1 Structure and molecular characteristics of STC1

The STC family consists of STC1 and STC2, and STC1 is a glycoprotein hormone that was first discovered in bony fish. In 1996, STC1 is shown to exist in mammals and there is 73% homology of STC1 amino acid sequences between human and bony fish (26). In human, the STC1 gene is consisted with four exons with a 5'UTR rich in CAG trinucleotide repeats (27) and locates on the short arm of chromosome 8p11.2-p21. Meanwhile, STC1 protein is a homologous dimer glycoprotein containing 11 conserved cysteine residues, composed of 247 amino acids, with a molecular weight of 56 kDa (28). STC1 is ubiquitously distributed in brain, thyroid, spleen, thymus, parathyroid gland, lung, heart, skeletal muscle, kidney, pancreas, small intestine, colon, placenta, ovary, testis and prostate, etc (14). The widespread expression of STC1 indicates that it acts through autocrine and paracrine. STC1 is considered as a regulator of calcium and phosphorus metabolism and associated with tumor progression and metastasis (29, 30). What's more, its localization in the thymus and spleen reveals that it may make a difference in immune and inflammatory processes (10). STC1 is secreted to extracellular environment in vivo (31), and binds to protein on the cell membrane (32), followed by internalization and targeting to the inner mitochondrial membrane (33, 34). There is no clear consensus on the receptor of STC1 at present, but STC1 can play a regulatory role in the occurrence and development of cancer through signaling pathways such as Notch1-SOX2 (23), ERK1/2 (24), NF-Kb (17) and JNK (25) signaling pathways, etc.

2.2 Expression of STC1 in tumor cells

Emerging evidences have demonstrated that STC1 is abnormally expressed in tumor tissues and is associated with tumor cell proliferation, migration, apoptosis and disease prognosis, such as non-small cell lung cancer, renal cell carcinoma, leukemia, lung adenocarcinoma, glioma, colon cancer, prostate cancer, ovarian cancer, etc.

A meta-analysis involving 16 articles and 2,942 participants found that high expression of STC1 was significantly associated with poor prognosis, especially for digestive (HR: 1.73; 95% CI: 1.36–2.20) and nervous system tumors (HR: 2.60; 95% CI: 1.29– 2.25) (35). Chromosome 8p deficiency disorder usually appears in colorectal cancer. By analyzing gene expression on chromosome 8p in 44 patients, a total of 13 genes located in 8p21-22 2Mb region were identified, including STC1, may trigger the progression and metastasis of colorectal cancer. In ovarian cancer, Liu et al. (36) found that the expression level of STC1 protein increased. Silencing STC1 expression by siRNA showed that STC1 could promote cell proliferation, migration and colony formation. What's more, STC1 could increase the growth of xenograft tumors. Mechanically, this may be because STC1 could increase the expression of cell cycle proteins (cyclin A, cyclin B1, cyclin-dependent kinase 2, and a short cyclin E isoform) to promote G1 into S phase; and upregulate antiapoptotic proteins (B-cell lymphoma 2, B-cell lymphoma-extra large) expression. Consistent with ovarian cancer, in lung adenocarcinoma (15) and clear renal cell carcinoma (20), high

expression of STC1 had also been shown to regulate the levels of cell cycle proteins and apoptotic proteins, thereby facilitating cell proliferation. However, in breast cancer, the expression and prognostic value of STC1 are different in distinct subtypes. In triple negative breast cancer (TNBC), the expression level of STC1 is higher than that in normal tissues, and is associated with poor prognosis (17). In hormone receptor-positive breast cancer, high expression of STC1 is associated with good prognosis (28). In human epidermal growth factor receptor 2⁺ (HER2⁺) breast cancer, high STC1 expression was not associated with prognosis (37).

TABLE 1	Expression	and	function	of STC	C1 in	tumors.
---------	------------	-----	----------	--------	-------	---------

Tumor type		Detected methods	Sample sizes	Expression	Function	References
Prostate cancer		IHC	22	High expression	Promote growth and metastasis	(<mark>80</mark>)
Colorectal cancer		qRT-PCR	20	High expression	Promotes migration and invasion, associated with poor postoperative prognosis	(16)
Renal cell carcinoma		qRT-PCR, WB, IHC	48	High expression	Promotes growth, proliferation and EMT, associated with Fuhrman tumor grade and TNM stage	(20)
Laryngeal squar	nous cell carcinoma	qRT-PCR	62	High expression	Associated with advanced clinical stage	(81)
Ovarian cancer		IHC	342	High expression	Increase cell adhesion, invision, proliferation and migration, colony formation, and promote the growth of transplanted tumor	(40, 82)
Lung cancer	Non-small cell lung cancer	qRT-PCR	65	High expression	Associated with advanced clinical stage and histological subtype	(83)
	Lung adenocarcinoma	ELISA, IHC	88	High expression	Increase cell proliferation, promote G1/S transition in the cell cycle, inhibit apoptosis	(15)
Breast cancer	ER ⁺ /TAM ⁺	IHC	541	High expression	Associated with prognosis	(84)
	ER ⁻ /TAM ⁻	IHC	300	Low expression	Associated with prognosis	(84)
	TNBC	qRT-PCR	cells	High expression	Associated with RFS、OS, promote metastasis	(17)
	All subtypes	IHC	137	High expression	Promote tumor growth and drug resistance	(85)
Hepatocellular carcinoma		qRT-PCR, IHC, WB, ELISA	125	High expression	Promote metastasis, promote stemness, and positively correlate with tumor size	(19, 25)
Cervical cancer		qRT- PCR, IHC	15	Low expression	Inhibit cell proliferation, migration and invasion	(86)
Leukemia		ELISA, IHC	20	High expression	Promotes chemotherapy resistance, a marker of MRD, associated with prognosis	(87)
Gastric cancer		IHC, ELISA	83	High expression	Associated with lymph node metastasis and advanced clinical stage	(88)
Glioma		qRT- PCR, IHC	80	High expression	Associated with high pathological grade	(89)
Thyroid cancer		IHC	100	High expression	Associated with cancer differentiation	(90)
Esophageal squamous cell carcinoma		qRT-PCR	15	High expression	Associated with advanced T stage and prognosis	(22)
Bladder cancer		IHC	63	High expression	Associated with the stage and poor prognosis of bladder cancer	(39)

EMT, epithelial-mesenchymal transition; ER, Estrogen receptor; TAM⁺, tamoxifen positively; TAM⁻, tamoxifen negatively; TNBC, triple negative breast cancer; RFS, relapse-free survival; OS, overall survival; MRD, minimal residual disease.

In summary, the crosstalk between STC1 expression and prognosis is different across diverse tumor types (Table 1). Usually, STC1 is highly expressed in most tumors and is associated with poor prognosis. However, in some highly heterogeneous tumors, such as breast cancer, STC1 may be differentially expressed in different tumor subtypes and sizes, which may be affected by the copy number variations, gene expression profile of different tumor subtypes. Moreover, some studies have indicated that in gynecological malignancies, the loss of STC1 function may impact normal physiological functions, which possibly leads to the low expression of STC1 in tumor tissues. And the expression level of STC1 fluctuates throughout the entire menstrual cycle of women, the changes in calcium metabolism at different age stages in women also affect the level of STC1 (38). In the future, more clinical samples and comprehensive analysis are needed to explore the expression of STC1 in multiple tumor tissues in well-stratified disease groups. In addition, clarifying the mechanism and causes of differential expression of STC1 will help us understand the role and value of STC1 in tumor immunity.

2.3 STC1 involved in tumor immunity

It is clear that STC1 regulates tumor immunity by influencing TME, participating in EMT and interfering with phagocytosis signals. By searching TCGA database, Sun et al. (39) found that STC1 was positively correlated with the expression of immune checkpoints, such as PD-L1 (k=0.29), PD-L2 (k=0.39), OX40L (k=0.32), TIM3 (k=0.31), FOXP3 (k=0.24), CTLA4 (k=0.23). Moreover, this study found that the high expression of STC1 was correlated with the high level of immune cells in TME, encompassing TAMs, CD4⁺ T cells, CD8⁺ T cells, and total T cells, indicating that STC1 was associated with immune checkpoints and multiple immune cells infiltration. That is, STC1 exhibits key functions in tumor immunity by regulating the level of immune cells in TME. STC1 has higher expression in cancer stroma and cancer associated fibroblasts (CAFs) than normal stroma and normal fibroblasts (NFs) (40). By detecting the levels of EMT markers fibronectin, vimentin and slug, it was found that human recombinant STC1 (rhSTC1) promoted EMT thus triggering the proliferation and metastasis of ovarian cancer cells via Akt phosphorylation. While monoclonal antibody STC1 (STC1 Ab) could reverse the above effects. By constructing STC1 overexpression cell to further explore its mechanism, it was found that STC1 secreted by cancer cell promoted the transformation from NFs into CAFs, but detailed mechanisms were not be clarified (40). In this way, STC1 participate in tumor immunity. In 2021, STC1 was reported to be involved in tumor immunity as a phagocytic checkpoint. Specifically, STC1 could damage the phagocytosis of antigen presenting cells (APC) and the activation of T cells by capturing calreticulin (CRT), thereby blocking the "eat me" signal of tumor cells and mediating tumor immune escape. And it was found that CRT interacted with STC1 through protein disulfide isomerase associated 3 (PDIA3) by STC1-FLAG immunoprecipitation (IP) (41). Using an in vitro cell co-culture model including CD19 antigen receptor T-cell immunotherapy (CAR-T) cells, tumors cells and macrophages, Zhang et al. found human mesenchymal stem cell (hMSCs) could suppress the efficacy of CD19 CAR-T therapy by reducing the proportion of CD8⁺ T cells, increasing the proportion of CD4⁺ T cells and Treg cells, and promoting IDO and PD-L1 levels, while STC1 knockdown could eliminate these effects. Compared with hMSCs^{shctrl}, hMSCs^{shSTC1} significantly decreased the expression level of IDO and PD-L1 protein by more than 50%. Moreover, hMSC^{shSTC1} combined with CD19 CAR-T cells therapy showed a curative effect in a xenograft model with tumors closely disappeared at day 38. In a word, the stimulatory signals appearing in the process of CAR-T treatment enabled hMSCs to secrete STC1, and exerted immunosuppressor function (12). In summary, STC1 affects tumor immunity by regulating immune checkpoints, affecting phagocytosis signals, impacting T cell immune response, and participating in EMT.

3 Tumor-associated macrophages participate in tumor immune escape

3.1 TAMs and tumor immune escape

TAMs contains two subtypes, the M1 type of classical activation pathway and the M2 type of alternate activation pathway (42). As shown in Figure 1, M1-type macrophages are stimulated by lipopolysaccharide (LPS), tumor necrosis factor- α (TNF- α) and interferon-gamma (IFN- γ), and secrete pro-inflammatory cytokines (IL-6, IL-12, IL-1b and TNF- α) and chemokine ligands (CCL2, CCL5, CXCL9, CXCL10 and CXCL11), which have proinflammatory and anti-tumor effects. Its common markers are CD86, CD64, CD80, CD16, CD120b, TLR2 and STAT1. While M2-type macrophages are stimulated by IL-4, IL-10 and transforming growth factor-beta (TGF-B), and secrete IL-4/10, TGF-B, VEGF and exosomes. Its common surface markers are arginase, CD36, CD206, CD266 and CD163, which have antiinflammatory and pro-tumor effects (43). What's more, cellular heterogeneity and unique subtypes of TAMs have been widely discovered recently with the development of high-dimensional flow cytometry and transcriptomic profiling (44). Generally, TAMs are mainly M1-type in the early stage of tumor development, which exert phagocytic effects on tumor cells and exhibit anti-tumor function. However, under the influence of TME, M1-type TAMs are gradually polarized into M2-type TAMs, which exhibit immunosuppressive impacts and participate in the formation of tumor immunosuppressive microenvironment (45). Compared with the tumor center, M2-type TAMs infiltration at the tumor invasive margin increased and was associated with low survival rate, indicating that TAMs distribution also had a certain spatial heterogeneity (46). In addition, the distribution of TAMs varies among different types of tumors, possibly due to differences in tumor immunosuppressive status, aggressiveness, and gene expression profiles (47). In a word, the phenotype and function of TAMs may be significantly different across tumor types and different



stages of tumor development. The time-space heterogeneity determines the specific role of TAM in TME to a large extent.

Tumor immune escape exerts key effects on unresponsive chemotherapy, immune therapy ineffective, poor prognosis, etc. The main ways of tumor cell immune escape are (1): tumor immunogenicity decreased; (2) the maturation of dendritic cells (DCs) was inhibited; (3) T lymphocytes activity was inhibited; (4) T lymphocytes migration and infiltration were inhibited; (5) the recognition function of immune cells was inhibited; (6) increased expression of immune checkpoint; (7) immunosuppressive cells were recruited (48). According to substantial evidences, TAMs play a pivotal role in tumor immune escape by regulating the recruitment and function of immune cells, promoting tumor antigen recognition disorders, regulating the secretion of immunosuppressive factors, and interfering with immune checkpoints via different mechanisms through various cytokines and pathways (Figure 2).

3.2 TAMs regulate recruitment and function of immune cells

T cells play a key role of immune response, they are mainly classified into cytotoxic T cells, helper T cells and regulatory T cells. Among them, CD8⁺ cytotoxic T cells (CTL) mainly recognize antigen peptides presented by major histocompatibility complex (MHC) class I molecules and directly kill tumor cells. Whereas

CD4⁺ T helper cells usually recognize MHC class II antigens and indirectly help clear tumor cells by secreting cytokines such as IL-2 and IFN- γ that enhance phagocytes and other T cells; Tregs prevent overimmunity, inhibit the role of cytotoxic T cells in a variety of tumors, and are generally believed to be associated with tumor immune escape (49). It has been reported that Fas ligand (FasL) on TAMs would bind to Fas receptor on immune cells, subsequently promoting the expression of caspase 3 and caspase 8, further directly regulating immune cells apoptosis, therefore, TAMs exhibit immunosuppression function (50).

In TME, TAMs recruit immunosuppressive cells and facilitate the depletion of cytotoxic T cells, thereby serving as a mediator in promoting immune escape. The infiltration of M2-type TAMs and Tregs is increased in nasopharyngeal carcinoma, and high infiltration level is associated with poor prognosis. Mechanistically, nasopharyngeal carcinoma cells induced polarization of M2-type TAMs by secreting TGF-B1 and IL-10. Subsequently, M2-type TAMs induced Tregs migration by secreting CCL22, CCL17, CXCL12, and fostered Tregs transformation by secreting TGF-B and IL-2, which led to the recruitment of Tregs and the immune escape of nasopharyngeal carcinoma cells (51). In addition, using CD4⁺ Foxp3⁺ Treg and monocytes co-culture system, it was found that the expression of CD206 and CD163 increased through IL-10, IL-13 and IL-4 secreted by Tregs (52). Exosomes are endogenous extracellular vesicles with a diameter of 30-100 nm, they are the mediators of intercellular communication (53). As reported by Yang et al. (54), exosomes derived from M2-type TAMs increased



microRNA-21 (miR-21) expression and decreased paternally expressed gene 3 (PEG3), subsequently increased the expression of Bcl-2, MMP-2 and MMP-9, reduced the proliferation and cytotoxic activity of CTL. It also enhanced Ki67 and PCNA expression, as well as the volume of tumors by downregulating the proportion of CTL, eventually inhibited the apoptosis of glioma cells, mediating immune escape. In addition, Nixon et al. (55) found that interferon regulatory factor-8 (IRF8) drives antigen presenting function of TAMs, induced the expression of PD-1 on CTL, thus leading to the depletion of CTL, weakening the immune response and forming an immunosuppressive environment. In summary, TAMs participate in tumor immune escape by regulating the recruitment and function of immune cells.

TAMs induce dysrecognition of tumor antigens. Tumor cells evade T cell recognition by removing, reducing, and transforming MHC-I molecules on their surface, resulting in tumor immune escape (48). It was found that the exosome LINC01232 antigen from M2-type TAMs promoted the transcription of NBR1 by binding to E2F2, and NBR1 binds to MHC-I protein, mediating the increased degradation of MHC-I in the autophagolysosome, eventually decreased the expression of MHC-I on the surface of tumor cells. In this way, tumor cells protected themselves from the immune attack of CTL, thus immune escape occurred (56). The surface of TAMs also express MHC-II, which presented tumor antigens to CD4⁺ T cells (57). Wu et al. (58) found that podoplanin (PDPN), a mucin-like transmembrane glycoprotein, could promote the polarization of M2-type TAMs through the secretion of exosomes. Additionally, PDPN could reduce the expression of MHC-II on the surface of macrophages through the tumor progression locus2/Erk/MHC class II transactivator (TPL2/Erk/CIITA) pathway and impair the antigen presentating ability. In turn, it affected the activity of CD4⁺ T cells and promoted the immune escape of glioblastoma. Therefore, TAMs induced tumor antigen recognition barriers by affecting MHC molecules on the surface of tumor cells, and promoted the occurrence of tumor immune escape.

3.3 TAMs regulate the secretion of immunosuppressive cytokines

Tumor immunosuppressive state would not have been possible by the recruitment of immunosuppressive cells in TME alone, but rather with the affect of immunosuppressive factors. IL-10 is a kind of immunosuppressive cytokine in TME promotes the polarization of M2-type TAMs, and in turn M2-type TAMs secrete more IL-10. A recent study have proved that microRNA let-7d targeted with IL-10 and inhibited the polarization of M2-type TAMs. That is, let-7d inhibited the crosstalk between M2-type TAMs and IL-10, thereby preventing immune escape in renal cell carcinoma (59). It is clear that IL-6 regulates macrophages, T cells and other immune cells, it has closely connection with tumor immunosuppressive microenvironment. Ma et al. (9) found that the exosome miR-1555p derived from TAMs promoted the secretion of IL-6 by down- egulating the expression of ZC3H12B in colon cancer cells both in vivo and in vitro. Subsequently, IL-6 affects the response of T cells, leading to immune escape of colon cancer and tumor progression. Similarly, in colorectal cancer, TAMs secreted the immunosuppressive factor TGF-B, promoted the expression of hypoxia-inducible factor 1α (HIF- 1α) and then increased tribbles pseudokinase 3 (TR1B3), activated the B-catenin/Wnt signaling pathway, enhanced the expression of PD-L1 on the cell membrane surface, and played a role in cell invasion and tumor immunity (10). In addition, M2-type TAMs are also involved in chemotherapy resistance through the secretion of immunosuppressive cytokines. In the lung cancer mouse model, M2-type TAMs could accumulate in well-differentiated tumor vascular sites, and promote tumor vascular reconstruction and recurrence by secreting vascular endothelial growth factor-A (VEGF-A) (60). Therefore, TAMs modulate the secretion of immunosuppressive cytokines to promote tumor immune escape.

3.4 TAMs interfere with immune checkpoints

Immune checkpoint is one of the targets of immunotherapy, including PD-1/PD-L1, CTLA-4, etc. PD-1 expresses on the surface of immune cells such as CD8⁺ T cells and CD4⁺ T cells, while PD-L1 expresses on the surface of APCs such as tumor cells and TAMs. The binding of the PD-1 and PD-L1 inhibits the cytotoxicity of immune cells and mediates immune escape. Immune checkpoint inhibitors can block these immune checkpoint ligands, so that

immune cell activity can be restored and immune escape can be suppressed. In recent years, accumulating evidences have shown that TAMs affected tumor immunity by interfering with the expression of immune checkpoints. Liu et al. (61) found that CC motif chemokine ligand 5 (CCL5) secreted by TAMs induced upregulation of PD-L1 protein in colon cancer cells through p65/ signal transducer and activator of transcription 3/COP9 signalosome 5 (p65/STAT3-CSN5) pathway, which enabled colon cancer cells to evade immune surveillance. On the other hand, inhibition of CTL mediated cytotoxicity promotes cellular immune escape and ultimately accelerated the development of colon cancer. Wang et al. (62) found that CD30L/CD30 signal increased the expression of PD-L1 on the cell membrane of TAMs. Meanwhile, PD-L1 on TAMs increased its binding to PD-1 on the surface of effector CD8⁺ T cells and CD4⁺ T cells, so it weakened the immune attack of CD8⁺ T cells and CD4⁺ T cells, and then mediated immune escape of colon cancer. In addition, in gastric cancer, TAMs-derived exosomes also mediated immune escape of gastric cancer cells by promoting PD-L1 expression in cancer cells (63). Therefore, interference with the expression of immune checkpoints is also a crucial way for TAMs to participate in tumor immune escape.

4 STC1 regulates macrophage function

4.1 STC1 regulates macrophage polarization and subpopulation differentiation

The polarization of macrophages plays a crucial role in inflammatory response and tumor immunity. Studies have shown that STC1 was secreted by CAFs and suppressed macrophage differentiation by binding to glucose-regulated protein 94 (GRP94), suggesting that STC1 exhibited an essential influence on tumor macrophages differentiation and participated in tumor immunity (64). Human acute monocytic leukemia THP-1 cells are generally used to induce M0, M1 and M2 macrophages. To explore the relationship between STC1 and TAMs differentiation, Leung et al. (65) induced THP-1 cells into M0 cells via phorbol 12myristate 13-acetate (PMA), further induced M0 into M1-type macrophages via LPS/IFN-y, and induced M0 into M2-type macrophages via IL-4/IL-3. It was found that the expression of STC1 increased in differentiated M0, M1 and M2 macrophages, among them, the level of STC1 is highest in M1-type macrophages. Additionally, using siRNA_{STC1} interference, this study investigated the effect of downregulation of STC1 gene in macrophages on the migration of hepatocellular carcinoma cells Hep3B. The results showed that the decrease of STC1 expression promoted the expression of TBC1 domain family member 3 (TBC1D3), led to the decrease of STAT phosphorylation level, and further inhibited the proliferation and migration of hepatocellular carcinoma in vitro. Different from previous studies, M1-type macrophages tend to have pro-inflammatory effects, while STC1 tends to have antiinflammatory effects. This paradoxical effect in this study may be due to the fact that STC1 negatively regulated the inflammatory response as an endogenous anti-inflammatory mediator and avoided the continuous activation of immune cells. In the myocardial infarction mouse model, Arezoo et al. (66) explored the impact of STC1 on the differentiation of monocytes/ macrophages. The release of stimuli such as pathogen-associated molecular patterns (PAMPs), damage-associated molecular patterns (DAMPs) and chemokines was elevated due to tissue injuring, these stimulants promoted the differentiation of monocytes and migration to the injured tissues. The addition of rhSTC1 decreased the expression of CD14 and the response to inflammatory stimuli, thus alleviated the continuous detrimental inflammatory response. A recent study found that STC1 promoted the immunosuppressive microenvironment by facilitating the polarization and infiltration of M2-type TAMs. By mechanism, STC1 activated Yes-associated protein (YAP) and promoted the secretion of CCL2, which induced the polarization of M2-type TAMs. Subsequently, M2-type TAMs secreted VEGF-A, which in turn increased the expression of STC1 by Akt signaling pathway. And the increase of YAP brought higher expression of PD-L1 on the melanoma cells, which decrease the interaction between macrophages and tumor cells (67). Taken together, these studies confirmed that STC1 served a role in the differentiation of macrophage subsets. It was also able to influence tumor proliferation and migration, but the role of STC1 in macrophage subpopulation differentiation needs more researches to confirm.

4.2 STC1 regulates macrophage activation and inflammatory response

Macrophages participate in the inflammatory response through the production of superoxide, inflammasome Nod-like receptor protein 3 (NLRP3), Nod-likenreceptor containing a caspase activating and recruitment domain 4 (NLRC4), IL-1B and other pathways, and studies have shown that STC1 can modulate these processes. The function of macrophages is affected by mitochondrial superoxide, and uncoupling protein 2 (UCP2) plays a major role in this process. Studies have found that STC1 can be internalized by macrophages and localized in mitochondria within 10 minutes, and STC1 induced the expression of UCP2 in macrophages, thereby weakening the influence of LPS on the generation of superoxide in macrophages. In general, STC1 exerted an impact on macrophage inflammation and immune response by regulating the generation of superoxide in macrophages (68). Mesenchymal stem cells (MSCs) are a class of stem cells with multi-differentiation potential and self-replication ability. JOO YOUN OH et al. (69) found that hMSCs derived from bone marrow reduced mitochondrial reactive oxygen species (ROS) of macrophages by secreting STC1, thus inhibited the activation of macrophage inflammasome NLRP3. What's more, STC1 secreted by adiposederived mesenchymal stem cells (ASCs) inhibited inflammasome NLRC4, alleviated the activation of caspase-1 and the secretion of IL-1B and IL-18, thus regulated the inflammation response of macrophages (70). Similarly, Yoojin Seo et al. (71) found that the

expression level of STC1 in tonsillar derived MSCs (TMSCs) was higher than that of other MSCs. Inhibition of STC1 could completely eliminate the influence of TMSCs on ROS, and further weaken the inhibition effect of TMSCs on IL-1B production in macrophages. These results have established that STC1 regulated the anti-inflammatory effect of macrophages through ROS pathway (71). To elucidate the mechanism of alveolar macrophages in acute respiratory distress syndrome, Xia et al. (72) co-cultured LPS-stimulated alveolar macrophages with human umbilical mesenchymal stem cells (HUMSC), it is found that the level of STC1, PI3K, AKT and mTOR increased, and IL-10 secreted by macrophages also elevated. Then, by knocking down STC1 gene and inhibiting mTOR with rapamycin, it was demonstrated that HUMSC secreted STC1, regulated the secretion of IL-10 by alveolar macrophages through PI3K/AKT/mTOR pathway. In conclusion, STC1 affects the inflammatory response of macrophages.

4.3 STC1 affects the phagocytosis activity and antigen presentation of macrophages

In 2004, researchers discovered that STC1 mapped to macrophages, it also reduced intracellular calcium levels, inhibited macrophages binding with chemotactic protein-1 (MCP-1) and stromal cell derived factor-1 α (SDF-1 α) on the monocytes, thereby affected the chemotaxis and response of a macrophages to antigens (13). Besides, endothelium greatly influenced the migration of macrophages from the circulation to impaired tissues, STC1 kept the tight junctions among endothelial cells by blocking TNF- α or IL-1B on the cells, ultimately inhibited the transendothelial migration of macrophages (73, 74). In other words, STC1 was capable of inducing hypofunction or defect of macrophages such as migration and response to antigens, thus leading to tumor immune escape. In 2021, STC1 was reported to be a phagocytic checkpoint. Specifically, STC1 decreased the "eat me" signal by down-regulating the expression of CRT on the membrane of cancer cells, affecting the recognition of cancer cells by antigen presenting cells such as TAMs and DCs, and further interfered with the specific recognition of T cells. That is, STC1 can drive the function of macrophages antigen presentation by influencing phagocytosis signals (75). Ferroptosis tends to occur in the injured endomentosis, large peritoneal macrophages (LPMs) will migrate to the uteri to against ferroptotic amonocytes/macrophages, which promotes anti-inflammatory response. It has been reported that MSCs facilitated the efferocytosis of LPMs, and STC1 showed an essential function in this process. By using rhSTC1 and siRNASTC1, Wang et al. (76) found MSCs-derived STC1 promoted the migration of LPMs to injured uteri, further accelerated the phagocytosis activity of LPMs. Collectively, STC1 can affect the phagocytic activity and antigen presentation of TAMs.

5 Discussion

TAMs play a critical role in tumor immunity and exert various influences on the proliferation, migration, invasion and metastasis

of cancer cells. STC1 has been widely studied as a phagocytosis checkpoint in recent years. From various researches, it has been proved that STC1 makes differences on the infiltration and function of different aspects of macrophages (Figure 3). However, in TME, how STC1 affects the functions of TAMs, such as whether STC1 reshapes the macrophage by metabolic pathway, glucose, lipid or amino acid, it still needs more experiments to explore. To be certain, current studies have confirmed that STC1 has a pro-cancer effect, so the design and development of STC1-targeted drugs are crucial for immunotherapy. There are currently no clinical trials of drugs targeting STC1, but some preclinical studies have found that targeting STC1 can slow tumor progression. For example, STC1 had been proved as a target gene for miR-146b-5p (77), and sevoflurane could upregulate the expression of miR-146b-5p and downregulate the expression of STC1 to suppress tumor growth (78); Choi et al. (79) found STC1 was the target gene of miR-606 by using the miRNA target prediction program and miR-606 mimics transfection, and it inhibited tumor growth and metastasis in vivo. In this way, the possible directions of STC1 in tumor therapy are as follows (1): Elucidate the three-dimensional structure of STC1, identify its receptor, and conduct a comprehensive study on its downstream signaling pathway; (2) Clarify the prediction value of STC1 in different tumors through multiple clinical studies, and explore the application of STC1 gene detection serve as a biomarker for tumor diagnosis, prevention and treatment; (3) Develop monoclonal antibodies against STC1 to block the action of STC1 and inhibit immune escape; (4) Develop small molecule compounds that mimic STC1 siRNA to reduce the expression level of STC1 in vivo, thereby inhibiting tumor. In addition, based on the effects of STC1 on TAMs, drugs that inhibit STC1 and TAMs, block the crosstalk between STC1 and TAMs should be an underlying direction of immunotherapy. Meanwhile, combination of STC1 inhibitor and TAMs inhibitor can be studied to achieve better immunotherapy effects. However, based on the regulation of calcium and phosphorus by STC1, safety problems targeting STC1 should be considered, and further researches are required to insure above approaches can bring clinical benefits for cancer.

In summary, tumor immunotherapy has manifested unprecedented success for cancer patients by enhancing the body immunity level to achieve the objective of killing tumor cells, whereas in clinical practice, only 20%-40% of patients can benefit from immunotherapy (2), "immune escape" phenomenon is considered to be the crucial reason for the failure of immunotherapy. Specifically, the reasons of tumor immune escape contains reducing the autoimmunogenicity, inhibiting the maturation and infiltration of immune cells, destroying the recognition function of immune cells, recruiting immunosuppressive cells, and interfering with immune checkpoints. STC1 is a glycoprotein hormone associated with metabolism of calcium and phosphorus, and a large number of studies have elucidated that STC1 is involved in the occurrence and development of tumors. It was demonstrated that STC1 could act by influencing TME, participating in EMT and interfering with phagocytosis signals. Additionally, TAMs also show its ability in tumor immune escape by promoting tumor antigen recognition disorder, regulating the recruitment and function of immune cells, regulating the secretion of immunosuppressive factors, and



interfering with immune checkpoints. In 2021, STC1 was shown to act as a phagocytic checkpoint and a TAMs function regulator, as well as participate in tumor immune escape, suggesting the interaction between STC1 and TAMs. In the future, more studies are needed to explore the relationship between STC1 and TAMs, and develop corresponding drugs to optimize the therapeutic effect of tumors. With the further studies on the crosstalk between STC1 and TAMs in the TME, may have the potential to makes STC1 to become a novel biomarker, which can improve the accuracy of early diagnosis of tumors and determine tumor metastasis. Hence, these strategies will help achieve a better anti-tumor potency of immunotherapy.

Author contributions

LW: Writing – original draft, Writing – review & editing. JW: Writing – review & editing. WQ: Funding acquisition, Methodology, Writing – review & editing. WG: Writing – review & editing.

Funding

The author(s) declare that financial support was received for the research, authorship, and/or publication of this article. This study was supported by the National Natural Science Foundation of

China (No. 82304986), Jiangsu Funding Program for Excellent Postdoctoral Talent (No. 2023ZB803) and the China Postdoctoral Science Foundation (No. 2023M741653).

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Generative AI statement

The author(s) declare that no Generative AI was used in the creation of this manuscript.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

References

1. Bray F, Laversanne M, Sung H, Ferlay J, Siegel RL, Soerjomataram I, et al. Global cancer statistics 2022: globocan estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA Cancer J Clin.* (2024) 74:229–63. doi: 10.3322/caac.21834

2. Sharma P, Hu-Lieskovan S, Wargo JA, Ribas A. Primary, adaptive, and acquired resistance to cancer immunotherapy. *Cell.* (2017) 168:707–23. doi: 10.1016/j.cell.2017.01.017

3. Evans ST, Jani Y, Jansen CS, Yildirim A, Kalemoglu E, Bilen MA. Understanding and overcoming resistance to immunotherapy in genitourinary cancers. *Cancer Biol Ther.* (2024) 25:2342599. doi: 10.1080/15384047.2024.2342599

4. Joyce JA, Pollard JW. Microenvironmental regulation of metastasis. *Nat Rev Cancer*. (2009) 9:239–52. doi: 10.1038/nrc2618

5. Basak U, Sarkar T, Mukherjee S, Chakraborty S, Dutta A, Dutta S, et al. Tumorassociated macrophages: an effective player of the tumor microenvironment. *Front Immunol.* (2023) 14:1295257. doi: 10.3389/fimmu.2023.1295257

6. Sarode P, Zheng X, Giotopoulou GA, Weigert A, Kuenne C, Günther S, et al. Reprogramming of tumor-associated macrophages by targeting B-catenin/fosl2/arid5a signaling: A potential treatment of lung cancer. *Sci Adv.* (2020) 6:eaaz6105. doi: 10.1126/sciadv.aaz6105

7. Kumar AT, Knops A, Swendseid B, Martinez-Outschoom U, Harshyne L, Philp N, et al. Prognostic significance of tumor-associated macrophage content in head and neck squamous cell carcinoma: A meta-analysis. *Front Oncol.* (2019) 9:656. doi: 10.3389/fonc.2019.00656

8. Cao J, Liu J, Xu R, Zhu X, Zhao X, Qian BZ. Prognostic role of tumour-associated macrophages and macrophage scavenger receptor 1 in prostate cancer: A systematic review and meta-analysis. *Oncotarget.* (2017) 8:83261–9. doi: 10.18632/oncotarget.18743

9. Ma YS, Wu TM, Ling CC, Yu F, Zhang J, Cao PS, et al. M2 macrophage-derived exosomal microrna-155-5p promotes the immune escape of colon cancer by downregulating zc3h12b. *Mol Ther Oncolytics*. (2021) 20:484–98. doi: 10.1016/j.omto.2021.02.005

10. Liu C, Zhang W, Wang J, Si T, Xing W. Tumor-associated macrophage-derived transforming growth factor-B Promotes colorectal cancer progression through hifl-trib3 signaling. *Cancer Sci.* (2021) 112:4198–207. doi: 10.1111/cas.15101

11. Mao X, Xu J, Wang W, Liang C, Hua J, Liu J, et al. Crosstalk between cancerassociated fibroblasts and immune cells in the tumor microenvironment: new findings and future perspectives. *Mol Cancer*. (2021) 20:131. doi: 10.1186/s12943-021-01428-1

12. Zhang R, Liu Q, Zhou S, He H, Zhao M, Ma W. Mesenchymal Stem Cell Suppresses the Efficacy of Car-T toward Killing Lymphoma Cells by Modulating the Microenvironment through Stanniocalcin-1. *NA*. (2023) 12:e82934. doi: 10.7554/eLife.82934

13. Kanellis J, Bick R, Garcia G, Truong L, Tsao CC, Etemadmoghadam D, et al. Stanniocalcin-1, an inhibitor of macrophage chemotaxis and chemokinesis. *Am J Physiol Renal Physiol*, (2004) 286:F356–62. doi: 10.1152/ajprenal.00138.2003

14. Yeung BH, Law AY, Wong CK. Evolution and roles of stanniocalcin. *Mol Cell Endocrinol.* (2012) 349:272–80. doi: 10.1016/j.mce.2011.11.007

15. Du YZ, Gu XH, Cheng SF, Li L, Liu H, Hu LP, et al. The oncogenetic role of stanniocalcin 1 in lung adenocarcinoma: A promising serum candidate biomarker for tracking lung adenocarcinoma progression. *Tumour Biol.* (2016) 37:5633–44. doi: 10.1007/s13277-015-4431-x

16. Macartney-Coxson DP, Hood KA, Shi HJ, Ward T, Wiles A, O'Connor R, et al. Metastatic susceptibility locus, an 8p hot-spot for tumour progression disrupted in colorectal liver metastases: 13 candidate genes examined at the DNA, mrna and protein level. *BMC Cancer.* (2008) 8:187. doi: 10.1186/1471-2407-8-187

17. Jeon M, Han J, Nam SJ, Lee JE, Kim S. Stc-1 expression is upregulated through an akt/nf-Kb-dependent pathway in triple-negative breast cancer cells. *Oncol Rep.* (2016) 36:1717–22. doi: 10.3892/or.2016.4972

18. Yuan X, Sun Z, Cui C. Knockdown of lncrna hottip inhibits retinoblastoma progression by modulating the mir-101-3p/stc1 axis. *Technol Cancer Res Treat.* (2021) 20:1533033821997831. doi: 10.1177/1533033821997831

19. Bai S, Zhao Y, Chen W, Peng W, Wang Y, Xiong S, et al. The stromal-tumor amplifying stc1-notch1 feedforward signal promotes the stemness of hepatocellular carcinoma. *J Transl Med.* (2023) 21:236. doi: 10.1186/s12967-023-04085-8

20. Ma X, Gu L, Li H, Gao Y, Li X, Shen D, et al. Hypoxia-induced overexpression of stanniocalcin-1 is associated with the metastasis of early stage clear cell renal cell carcinoma. *J Transl Med.* (2015) 13:56. doi: 10.1186/s12967-015-0421-4

21. Abaza HM, Elmougy MI, El Maraghy HM, Mahmoud HM. Stanniocalcin1 gene expression in patients with acute leukemia: impact on response to therapy and disease outcome. *Int J Lab Hematol.* (2016) 38:81–9. doi: 10.1111/ijlh.12445

22. Shirakawa M, Fujiwara Y, Sugita Y, Moon JH, Takiguchi S, Nakajima K, et al. Assessment of stanniocalcin-1 as a prognostic marker in human esophageal squamous cell carcinoma. *Oncol Rep.* (2012) 27:940–6. doi: 10.3892/or.2011.1607

23. Li Y, He ZC, Zhang XN, Liu Q, Chen C, Zhu Z, et al. Stanniocalcin-1 augments stem-like traits of glioblastoma cells through binding and activating notch1. *Cancer Lett.* (2018) 416:66–74. doi: 10.1016/j.canlet.2017.11.033

24. He LF, Wang TT, Gao QY, Zhao GF, Huang YH, Yu LK, et al. Stanniocalcin-1 Promotes Tumor Angiogenesis through up-Regulation of Vegf in Gastric Cancer Cells. *J BioMed Sci.* (2011) 18:39. doi: 10.1186/1423-0127-18-39

25. Chan KK, Leung CO, Wong CC, Ho DW, Chok KS, Lai CL, et al. Secretory stanniocalcin 1 promotes metastasis of hepatocellular carcinoma through activation of jnk signaling pathway. *Cancer Lett.* (2017) 403:330–8. doi: 10.1016/j.canlet.2017.06.034

26. Olsen HS, Cepeda MA, Zhang QQ, Rosen CA, Vozzolo BL, Wagner GF. Human stanniocalcin: A possible hormonal regulator of mineral metabolism. *Proc Natl Acad Sci U.S.A.* (1996) 93:1792–6. doi: 10.1073/pnas.93.5.1792

27. Zhao F, Yang G, Feng M, Cao Z, Liu Y, Qiu J, et al. Expression, function and clinical application of stanniocalcin-1 in cancer. *J Cell Mol Med.* (2020) 24:7686–96. doi: 10.1111/jcmm.15348

28. Chang AC, Jellinek DA, Reddel RR. Mammalian stanniocalcins and cancer. Endocr Relat Cancer. (2003) 10:359–73. doi: 10.1677/erc.0.0100359

29. Knösel T, Schlüns K, Stein U, Schwabe H, Schlag PM, Dietel M, et al. Chromosomal alterations during lymphatic and liver metastasis formation of colorectal cancer. *Neoplasia.* (2004) 6:23–8. doi: 10.1016/s1476-5586(04)80050-2

30. Friedman JB, Brunschwig EB, Platzer P, Wilson K, Markowitz SD. C8orf4 is a transforming growth factor B induced transcript downregulated in metastatic colon cancer. *Int J Cancer*. (2004) 111:72–5. doi: 10.1002/ijc.20235

31. Jellinek DA, Chang AC, Larsen MR, Wang X, Robinson PJ, Reddel RR. Stanniocalcin 1 and 2 are secreted as phosphoproteins from human fibrosarcoma cells. *Biochem J.* (2000) 350 Pt 2:453–61.

32. Luo CW, Kawamura K, Klein C, Hsueh AJ. Paracrine regulation of ovarian granulosa cell differentiation by stanniocalcin (Stc) 1: mediation through specific stc1 receptors. *Mol Endocrinol.* (2004) 18:2085–96. doi: 10.1210/me.2004-0066

33. Sazonova O, James KA, McCudden CR, Segal D, Talebian A, Wagner GF. Stanniocalcin-1 secretion and receptor regulation in kidney cells. *Am J Physiol Renal Physiol.* (2008) 294:F788–94. doi: 10.1152/ajprenal.00553.2007

34. McCudden CR, James KA, Hasilo C, Wagner GF. Characterization of mammalian stanniocalcin receptors. Mitochondrial targeting of ligand and receptor for regulation of cellular metabolism. *J Biol Chem.* (2002) 277:45249–58. doi: 10.1074/ jbc.M205954200

35. Li R, Liu R, Wu S, Zheng S, Ye L, Shao Y. Prognostic value of stc1 in solid tumors: A meta-analysis. *biomark Med.* (2022) 16:253–63. doi: 10.2217/bmm-2021-0835

36. Liu G, Yang G, Chang B, Mercado-Uribe I, Huang M, Zheng J, et al. Stanniocalcin 1 and ovarian tumorigenesis. *J Natl Cancer Inst.* (2010) 102:812–27. doi: 10.1093/jnci/djq127

 Zandberga E, Zayakin P, Abols A, Pūpola D, Trapencieris P, Linē A. Depletion of carbonic anhydrase ix abrogates hypoxia-induced overexpression of stanniocalcin-1 in triple negative breast cancer cells. *Cancer Biol Ther.* (2017) 18:596–605. doi: 10.1080/ 15384047.2017.1345390

38. Khatun M, Modhukur V, Piltonen TT, Tapanainen JS, Salumets A. Stanniocalcin protein expression in female reproductive organs: literature review and public cancer database analysis. *Endocrinology*. (2024) 165:bqae110. doi: 10.1210/endocr/bqae110

39. Sun J, Wei X, You J, Yue W, Ouyang J, Ling Z, et al. Stc1 is a novel biomarker associated with immune characteristics and prognosis of bladder cancer. *Int J Gen Med.* (2021) 14:5505–16. doi: 10.2147/ijgm.S329723

40. Yang Y, Yin S, Li S, Chen Y, Yang L. Stanniocalcin 1 in tumor microenvironment promotes metastasis of ovarian cancer. *Onco Targets Ther.* (2019) 12:2789–98. doi: 10.2147/ott.S196150

41. Lin H, Kryczek I, Li S, Green MD, Ali A, Hamasha R, et al. Stanniocalcin 1 is a phagocytosis checkpoint driving tumor immune resistance. *Cancer Cell*. (2021) 39:480–93.e6. doi: 10.1016/j.ccell.2020.12.023

42. Tamura R, Tanaka T, Yamamoto Y, Akasaki Y, Sasaki H. Dual role of macrophage in tumor immunity. *Immunotherapy*. (2018) 10:899–909. doi: 10.2217/ imt-2018-0006

43. Aktar T, Modak S, Majumder D, Maiti D. A detailed insight into macrophages' Role in shaping lung carcinogenesis. *Life Sci.* (2024) 352:122896. doi: 10.1016/j.lfs.2024.122896

44. Yang S, Wang M, Hua Y, Li J, Zheng H, Cui M, et al. Advanced insights on tumor-associated macrophages revealed by single-cell rna sequencing: the intratumor heterogeneity, functional phenotypes, and cellular interactions. *Cancer Lett.* (2024) 584:216610. doi: 10.1016/j.canlet.2024.216610

45. Wang J, Mi S, Ding M, Li X, Yuan S. Metabolism and polarization regulation of macrophages in the tumor microenvironment. *Cancer Lett.* (2022) 543:215766. doi: 10.1016/j.canlet.2022.215766

46. Zheng X, Weigert A, Reu S, Guenther S, Mansouri S, Bassaly B, et al. Spatial density and distribution of tumor-associated macrophages predict survival in nonsmall cell lung carcinoma. *Cancer Res.* (2020) 80:4414–25. doi: 10.1158/0008-5472.Can-20-0069

47. Mantovani A, Marchesi F, Di Mitri D, Garlanda C. Macrophage diversity in cancer dissemination and metastasis. *Cell Mol Immunol.* (2024) 21:1201–14. doi: 10.1038/s41423-024-01216-z

48. Kim SK, Cho SW. The evasion mechanisms of cancer immunity and drug intervention in the tumor microenvironment. *Front Pharmacol.* (2022) 13:868695. doi: 10.3389/fphar.2022.868695

49. Farhood B, Najafi M, Mortezaee K. Cd8(+) cytotoxic T lymphocytes in cancer immunotherapy: A review. J Cell Physiol. (2019) 234:8509–21. doi: 10.1002/jcp.27782

50. Brown SB, Savill J. Phagocytosis triggers macrophage release of fas ligand and induces apoptosis of bystander leukocytes. *J Immunol.* (1999) 162:480–5. doi: 10.4049/jimmunol.162.1.480

51. Wang J, Huang H, Lu J, Bi P, Wang F, Liu X, et al. Tumor cells induced-M2 macrophage favors accumulation of treg in nasopharyngeal carcinoma. *Int J Clin Exp Pathol.* (2017) 10:8389–401.

52. Tiemessen MM, Jagger AL, Evans HG, van Herwijnen MJ, John S, Taams LS. Cd4+Cd25+Foxp3+ Regulatory T cells induce alternative activation of human monocytes/macrophages. *Proc Natl Acad Sci U.S.A.* (2007) 104:19446-51. doi: 10.1073/pnas.0706832104

53. Pegtel DM, Gould SJ. Exosomes. Annu Rev Biochem. (2019) 88:487–514. doi: 10.1146/annurev-biochem-013118-111902

54. Yang F, Wang T, Du P, Fan H, Dong X, Guo H. M2 bone marrow-derived macrophage-derived exosomes shuffle microrna-21 to accelerate immune escape of glioma by modulating peg3. *Cancer Cell Int.* (2020) 20:93. doi: 10.1186/s12935-020-1163-9

55. Nixon BG, Kuo F, Ji L, Liu M, Capistrano K, Do M, et al. Tumor-associated macrophages expressing the transcription factor irf8 promote T cell exhaustion in cancer. *Immunity*. (2022) 55:2044–58.e5. doi: 10.1016/j.immuni.2022.10.002

56. Li J, Wang K, Yang C, Zhu K, Jiang C, Wang M, et al. Tumor-associated macrophage-derived exosomal linc01232 induces the immune escape in glioma by decreasing surface mhc-I expression. *Adv Sci (Weinh).* (2023) 10:e2207067. doi: 10.1002/advs.202207067

57. Perez-Diez A, Liu X, Matzinger P. Neoantigen presentation and ifn γ Signaling on the same tumor-associated macrophage are necessary for cd4 T cell-mediated antitu mor activity in mice. *Cancer Res Commun.* (2022) 2:316–29. doi: 10.1158/2767-9764.crc-22-0052

58. Wu M, Shi Y, Liu Y, Huang H, Che J, Shi J, et al. Exosome-transmitted podoplanin promotes tumor-associated macrophage-mediated immune tolerance in glioblastoma. *CNS Neurosci Ther.* (2024) 30:e14643. doi: 10.1111/cns.14643

59. Su B, Han H, Gong Y, Li X, Ji C, Yao J, et al. Let-7d inhibits intratumoral macrophage M2 polarization and subsequent tumor angiogenesis by targeting il-13 and il-10. *Cancer Immunol Immunother*. (2021) 70:1619–34. doi: 10.1007/s00262-020-02791-6

60. Hughes R, Qian BZ, Rowan C, Muthana M, Keklikoglou I, Olson OC, et al. Perivascular M2 macrophages stimulate tumor relapse after chemotherapy. *Cancer Res.* (2015) 75:3479–91. doi: 10.1158/0008-5472.Can-14-3587

61. Liu C, Yao Z, Wang J, Zhang W, Yang Y, Zhang Y, et al. Macrophage-derived ccl5 facilitates immune escape of colorectal cancer cells via the P65/stat3-csn5-pd-L1 pathway. *Cell Death Differ*. (2020) 27:1765–81. doi: 10.1038/s41418-019-0460-0

62. Wang X, Gao Y, Zhang X, Wang X, Wang B, Meng X, et al. Cd30l/cd30 signaling regulates the formation of the tumor immune microenvironment and inhibits intestinal tumor development of colitis-associated colon cancer in mice. *Int Immunopharmacol.* (2020) 84:106531. doi: 10.1016/j.intimp.2020.106531

63. Wang Y, Shang K, Zhang N, Zhao J, Cao B. Tumor-associated macrophagederived exosomes promote the progression of gastric cancer by regulating the P38mapk signaling pathway and the immune checkpoint pd-L1. *Cancer Biother Radiopharm.* (2021). doi: 10.1089/cbr.2021.0218

64. Kamata T, So TY, Ahmed Q, Giblett S, Patel B, Luo J, et al. Fibroblast-derived stc-1 modulates tumor-associated macrophages and lung adenocarcinoma development. *Cell Rep.* (2020) 31:107802. doi: 10.1016/j.celrep.2020.107802

65. Leung CCT, Wong CKC. Characterization of stanniocalcin-1 expression in macrophage differentiation. *Transl Oncol.* (2021) 14:100881. doi: 10.1016/j.tranon.2020.100881

66. Mohammadipoor A, Lee RH, Prockop DJ, Bartosh TJ. Stanniocalcin-1 attenuates ischemic cardiac injury and response of differentiating monocytes/macrophages to inflammatory stimuli. *Transl Res.* (2016) 177:127–42. doi: 10.1016/j.trsl.2016.06.011

67. Ren Z, Xu Z, Chang X, Liu J, Xiao W. Stc1 competitively binding Bpix enhances melanoma progression via yap nuclear translocation and M2 macrophage recruitment through the yap/ccl2/vegfa/akt feedback loop. *Pharmacol Res.* (2024) 204:107218. doi: 10.1016/j.phrs.2024.107218

68. Wang Y, Huang L, Abdelrahim M, Cai Q, Truong A, Bick R, et al. Stanniocalcin-1 suppresses superoxide generation in macrophages through induction of mitochondrial ucp2. *J Leukocyte Biol.* (2009) 86:981–8. doi: 10.1189/jlb.0708454

69. Oh JY, Ko JH, Lee HJ, Yu JM, Choi H, Kim MK, et al. Mesenchymal stem/ stromal cells inhibit the nlrp3 inflammasome by decreasing mitochondrial reactive oxygen species. *Stem Cells*. (2014) 32:1553–63. doi: 10.1002/stem.1608

70. Li LL, Zhu YG, Jia XM, Liu D, Qu JM. Adipose-derived mesenchymal stem cells ameliorating pseudomonas aeruginosa-induced acute lung infection via inhibition of

nlrc4 inflammasome. Front Cell Infect Microbiol. (2020) 10:581535. doi: 10.3389/ fcimb.2020.581535

71. Seo Y, Shin TH, Ahn JS, Oh SJ, Shin YY, Yang JW, et al. Human tonsil-derived mesenchymal stromal cells maintain proliferating and ros-regulatory properties via stanniocalcin-1. *Cells.* (2020) 9:636. doi: 10.3390/cells9030636

72. Xia TT, Hu R, Shao CJ, Feng Y, Yang XL, Xie YP, et al. Stanniocalcin-1 secreted by human umbilical mesenchymal stem cells regulates interleukin-10 expression via the pi3k/akt/mtor pathway in alveolar macrophages. *Cytokine*. (2023) 162:156114. doi: 10.1016/j.cyto.2022.156114

73. Chakraborty A, Brooks H, Zhang P, Smith W, McReynolds MR, Hoying JB, et al. Stanniocalcin-1 regulates endothelial gene expression and modulates transendothelial migration of leukocytes. *Am J Physiol Renal Physiol.* (2007) 292:F895–904. doi: 10.1152/ajprenal.00219.2006

74. Chen C, Jamaluddin MS, Yan S, Sheikh-Hamad D, Yao Q. Human stanniocalcin-1 blocks tnf-alpha-induced monolayer permeability in human coronary artery endothelial cells. *Arterioscler Thromb Vasc Biol.* (2008) 28:906–12. doi: 10.1161/atvbaha.108.163667

75. Luo M, Wang X, Wu S, Yang C, Su Q, Huang L, et al. A20 promotes colorectal cancer immune evasion by upregulating stc1 expression to block "Eat-me" Signal. *Signal Transduct Target Ther.* (2023) 8:312. doi: 10.1038/s41392-023-01545-x

76. Wang J, Li J, Yin L, Wang X, Dong Y, Zhao G, et al. Mscs promote the efferocytosis of large peritoneal macrophages to eliminate ferroptotic monocytes/ macrophages in the injured endometria. *Stem Cell Res Ther.* (2024) 15:127. doi: 10.1186/s13287-024-03742-z

77. Al-Abdallah A, Jahanbani I, Mehdawi H, Ali RH, Al-Brahim N, Mojiminiyi O. The stress-activated protein kinase pathway and the expression of stanniocalcin-1 are regulated by mir-146b-5p in papillary thyroid carcinogenesis. *Cancer Biol Ther.* (2020) 21:412–23. doi: 10.1080/15384047.2020.1721250

78. Wen J, Ding Y, Zheng S, Li X, Xiao Y. Sevoflurane suppresses glioma cell proliferation, migration, and invasion both in vitro and in vivo partially via regulating kcnq1ot1/mir-146b-5p/stc1 axis. *Cancer Biother Radiopharm.* (2024) 39:105–16. doi: 10.1089/cbr.2020.3762

79. Choi S, An HJ, Yeo HJ, Sung MJ, Oh J, Lee K, et al. Microrna-606 inhibits the growth and metastasis of triple-Negative breast cancer by targeting stanniocalcin 1. *Oncol Rep.* (2024) 51:2. doi: 10.3892/or.2023.8661

80. Costa BP, Schein V, Zhao R, Santos AS, Kliemann LM, Nunes FB, et al. Stanniocalcin-1 protein expression profile and mechanisms in proliferation and cell death pathways in prostate cancer. *Mol Cell Endocrinol.* (2020) 502:110659. doi: 10.1016/j.mcc.2019.110659

81. Zhou H, Li YY, Zhang WQ, Lin D, Zhang WM, Dong WD. Expression of stanniocalcin-1 and stanniocalcin-2 in laryngeal squamous cell carcinoma and correlations with clinical and pathological parameters. *PLoS One.* (2014) 9:e95466. doi: 10.1371/journal.pone.0095466

82. Bajwa P, Kordylewicz K, Bilecz A, Lastra RR, Wroblewski K, Rinkevich Y, et al. Cancer-associated mesothelial cell-derived angptl4 and stc1 promote the early steps of ovarian cancer metastasis. *NA*. (2023) 8:e163019. doi: 10.1172/jci.insight.163019

83. Du YZ, Gu XH, Li L, Gao F. The diagnostic value of circulating stanniocalcin-1 mrna in non-small cell lung cancer. *J Surg Oncol.* (2011) 104:836–40. doi: 10.1002/jso.21948

84. Brantley KD, Kjærsgaard A, Cronin-Fenton D, Yacoub R, Nielsen AS, Lauridsen KL, et al. Stanniocalcin expression as a predictor of late breast cancer recurrence. *Cancer Epidemiol Biomarkers Prev.* (2018) 27:653–9. doi: 10.1158/1055-9965.Epi-17-0905

85. Hou J, Cheng J, Dai Z, Wei N, Chen H, Wang S, et al. Molecular and clinical significance of stanniocalcin-1 expression in breast cancer through promotion of homologous recombination-mediated DNA damage repair. *Front Cell Dev Biol.* (2021) 9:731086. doi: 10.3389/fcell.2021.731086

86. Guo F, Li Y, Wang J, Li Y, Li Y, Li G. Stanniocalcin1 (Stc1) inhibits cell proliferation and invasion of cervical cancer cells. *PLoS One.* (2013) 8:e53989. doi: 10.1371/journal.pone.0053989

87. Schelker RC, Kratzer A, Müller G, Brochhausen C, Hart C, Stempfl T, et al. Stanniocalcin 1 is overexpressed in multipotent mesenchymal stromal cells from acute myeloid leukemia patients. *Hematology*. (2021) 26:565–76. doi: 10.1080/16078454.2021.1962048

88. Fang Z, Tian Z, Luo K, Song H, Yi J. Clinical significance of stanniocalcin expression in tissue and serum of gastric cancer patients. *Chin J Cancer Res.* (2014) 26:602–10. doi: 10.3978/j.issn.1000-9604.2014.10.08

89. Su J, Guo B, Zhang T, Wang K, Li X, Liang G. Stanniocalcin-1, a new biomarker of glioma progression, is associated with prognosis of patients. *Tumour Biol.* (2015) 36:6333–9. doi: 10.1007/s13277-015-3319-0

90. Sengun S, Korkmaz H, Ciris M, Yüceer RO, Boyluboy SM, Kiran M. Diagnostic and prognostic value of stanniocalcin 1 expression in papillary thyroid cancer. *Endocrine*. (2022) 78:95–103. doi: 10.1007/s12020-022-03126-4

Glossary

	o o o o o o o			
PD-1		programmed death-1 receptor	CTL	CD8 ⁺ cytotoxic T cells
PD-L	1	programmed death-1 receptor ligand 1	MHC	major histocompatibility complex
CTLA	A-4	cytotoxic T lymphocyte-associated antigen-4	FasL	Fas ligand
IDO		indoleamine-2,3-dioxygenase	TGF-B	transforming growth factor-beta
TME		tumor microenvironment	miR-21/PEG3	microRNA-21/paternally expressed gene 3
TAM	s	tumor-associated macrophages	IRF8	interferon regulatory factor-8
NK c	ells	natural killer cells	PDPN	podoplanin
Tregs		regulatory T cells	TPL2/Erk/CIITA	tumor progression locus2/Erk/MHC class II
STC1		Stanniocalcin-1		transactivator
EMT		epithelial-mesenchymal transition	HIF-1α	hypoxia-inducible factor 1α
Notch	n1-SOX2	notch1-sex determining region Y-box 2	TR1B3	tribbles pseudokinase 3
ERK1	/2	extracellular signalregulatedkinases1/2	VEGF-A	vascular endothelial growth factor-A
NF-K	В	Nuclear Factor-kappaB	p65/STAT3-CSN5	p65/Signal transducer and activator of Transcription 3/COP9 signalosome 5
JNK		and Jun-N-terminal kinase	GRP94	glucose-regulated protein 94
TNBC	3	triple negative breast cancer	РМА	phorbol 12-myristate 13-acetate
HER2	2+	human epidermal growth factor receptor 2 ⁺	TBC1D3	TBC1 domain family member 3
CAFs		cancer associated fibroblasts	PAMPs	pathogen-associated molecular patterns
NFs		normal fibroblasts	DAMPs	damage-associated molecular patterns
rhSTO	C1	human recombinant STC1	YAP	Yes-associated protein
STC1	Ab	monoclonal antibody STC1	NLRP3	Nod-like receptor protein 3
APC		antigen presenting cells	NLRC4	Nod-liken receptor containing a caspase activating
CRT		calreticulin		and recruitment domain 4
PD1A	13	protein disulfide isomerase associated 3	UCP2	uncoupling protein 2
IP		immunoprecipitation	MSCs	mesenchymal stem cells
CAR-	T	chimeric antigen receptor T-cell immunotherapy	ROS	reactive oxygen species
hMSC	Cs	human mesenchymal stem cells	ASCs	adipose-derived mesenchymal stem cells
LPS		lipopolysaccharide	TMSCs	tonsillar derived MSCs
TNF-	α	tumor necrosis factor-α	HUMSC	human umbilical mesenchymal stem cells
IFN-γ	(interferon-gamma	MCP-1	chemotactic protein-1
CCL		CC motif chemokine ligand	SDF-1a	stromal cellderived factor-1a
CXCI	_	C-X-C motif chemokine ligand	LPMs	large peritoneal macrophages.
DCs		dendritic cells		