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# Based on the immune system: the role of the IL-2 family in pancreatic disease

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The IL-2 family, consisting of IL-2, IL-4, IL-7, IL-9, IL-15 and IL-21, is a key regulator of the immune response. As an important endocrine and digestive organ, the function of the pancreas is regulated by the immune system. Studies have shown that each cytokine of the IL-2 family influences the occurrence and development of pancreatic diseases by participating in the regulation of the immune system. In this paper, we review the structural and functional characteristics of IL-2 family members, focus on their molecular mechanisms in pancreatic diseases including acute pancreatitis, chronic pancreatitis and pancreatic cancer, and highlight the importance of the related proteins in the regulation of immune response and disease progression, which will provide valuable insights for new biomarkers in pancreatic diseases, early diagnosis of the diseases, assessment of the disease severity, and development of new therapeutic regimens. The insights of the study are summarized in the following sections.

#### KEYWORDS

pancreatitis, pancreatic cancer, IL-2 family, IL-2, immunity

# 1 Introduction

The interleukin-2 (IL-2) family consists of six cytokines, including IL-2, IL-4, IL-7, IL-9, IL-15, and IL-21, which share a  $\gamma$ -chain structural domain. These molecules are indispensable regulators in both innate and adaptive immune responses (1). IL-2 has multiple functions that promote T cell proliferation, enhance the cytotoxicity of natural killing (NK) cells, and promote the production of inflammatory factors by activating transcription factors such as STAT5 (2). In addition, other members of the IL-2 family have been shown to play key regulatory roles in the immune system (3).

Pancreatitis is an inflammatory disease that usually results from abnormal activation of trypsin due to a variety of factors, which in turn causes self-digestion of pancreatic tissues, swelling, bleeding, and even tissue necrosis (4). Among them, acute pancreatitis (AP) is one of the common gastrointestinal diseases in hospitals with high mortality rate. AP is an acute inflammatory response of the pancreas in which the patient experiences sudden onset of severe epigastric pain accompanied by abdominal pressure pain, nausea, vomiting, fever,

and increased pulse rate (5). And despite the low incidence of chronic pancreatitis (CP), it can cause permanent damage to the pancreas, which has a significant impact on the quality of life of patients (6). CP develops slowly, beginning with cellular injury followed by inflammation and fibrosis. The pathophysiologic process of chronic pancreatitis involves the alveolar cells, which are the primary site of initial injury or stress, leading to an inflammatory cascade response (7). Studies have revealed that local and systemic immune responses driven by inflammatory cells determine the course and severity of pancreatitis and are one of the important factors that induce pancreatitis (8). Several immune cells, including macrophages, granulocytes, dendritic cells (DCs), and T cells, play a crucial role in the pathologic process of pancreatitis (9).

The persistence of pancreatitis, especially CP, is considered an important risk factor for pancreatic cancer (PC) (10). The lethality of PC is so high that it ranks among the leading causes of cancer death worldwide (11). The tumor microenvironment (TME) exhibits immunosuppressive properties and is able to interfere with the effects of immunotherapy by promoting the recruitment of immunosuppressive cells and increasing the expression of immunosuppressive molecules (12). The TME of PC is a complex network of multiple cells and molecules, which includes PC cells, pancreatic stellate cells (PSCs), cancer-associated fibroblasts (CAFs), tumor-associated macrophages (TAMs), regulatory T cells (Tregs), extracellular matrix, vasculature, and lymphatic vessels, as well as a multitude of signaling molecules, such as growth factors, cytokines, and chemokines (13, 14). This microenvironment significantly affects PC progression and patient prognosis by enhancing fibrosis, promoting immune escape, and increasing treatment resistance. The IL-2 family, as an important player in the inflammatory response and a key factor in immune regulation, is tightly linked to the development of pancreatitis. Cytokines of the IL-2 family have been shown to influence the TME by modulating immune responses to affect TME, which in turn affects the growth and spread of tumor cells and participates in the development of PC.

Currently, there is no systematic review of the specific role of the IL-2 family in pancreatitis and PC. Therefore, in-depth studies in this area are of particular importance. In this paper, we will explore the structural features and functional properties of IL-2 family members and outline their roles and their molecular mechanisms in pancreatitis and PC, in order to deepen the understanding of the mechanisms of IL-2 family roles in pancreatic diseases and to lay a solid scientific foundation for the development of innovative therapeutic strategies for IL-2 family cytokines in pancreatic diseases.

# 2 IL-2 family

Interleukins (ILs) are proteins secreted by leukocytes that play crucial communicative roles in both the innate and adaptive immune systems, while also affecting the functions of nonimmune cells and tissues (15). These molecules promote the proliferation, differentiation, and activation of immune cells, thereby enhancing the body's immune response. Based on sequence similarity, secondary structure, receptor composition, and functional characteristics, scientists have classified over 60 types of ILs into seven different families (16). Among them, IL-2, IL-4, IL-7, IL-9, IL-15, and IL-21 are categorized as type I cytokines, collectively known as the IL-2 family, due to their similar threedimensional structure characterized by a four  $\alpha$ -helix bundle (17). These cytokines exert their effects by binding to specific receptors that typically consist of common  $\gamma$  chains and unique  $\alpha$  chains (such as IL-2R $\alpha$ , IL-4R $\alpha$ , IL-7R $\alpha$ , IL-9R $\alpha$ , and IL-15R $\alpha$ ) (18), (as shown in Figure 1). There is a conserved sequence on these chains, the WSXWs motif, which plays a key role in the activation of the receptors. Therefore, the IL-2 family is sometimes referred to as the cytokine receptor  $\gamma$ c family. Upon binding to their receptors, these cytokines can activate JAK1 and JAK3 kinases, leading to a cascade of phosphorylation reactions involving STAT proteins (such as STAT1, STAT3, STAT5, and STAT6) (19).

IL-2, the first member of the IL-2 family, was initially discovered in the supernatant of activated T cells and identified as a key factor for T cell growth (20). This 15 kDa cytokine is typically secreted by activated CD4+T cells and CD8+T cells (21). The IL-2 receptor complex is composed of three distinct subunits: IL-2R $\alpha$  (CD25), IL-2R $\beta$  (CD122), and IL-2R $\gamma$  (CD132). The primary function of IL-2 is to promote the proliferation and differentiation of effector T cells, memory T cells, and NK cells, thereby triggering immune responses (2). Additionally, IL-2 activates both T cells and B cells and regulates immune responses through activation-induced cell death (AICD) to maintain immune system balance. IL-2 is also crucial for the formation of Tregs and is essential for the function of Foxp3+ T cells in autoimmune diseases such as type 1 diabetes, systemic lupus erythematosus, pancreatitis, and pancreatic cancer (22).

IL-4 was initially identified as a pleiotropic type 2 cytokine, primarily secreted by CD4+T cells, Th2 cells, basophils, eosinophils, and mast cells (1). IL-4 can bind to its receptor, activating various signaling pathways, including JAK-STAT and PI3K/Akt/mTOR (23). IL-4 plays multiple roles in immune regulation: it promotes the survival and proliferation of mast cells, induces macrophage polarization towards the M2 phenotype (24), enhances B cell differentiation, facilitates immunoglobulin class switching, and increases the production of IgG and IgE (1).

Hunt and his colleagues first identified IL-7 as a cytokine that promotes the development of pro-B cells and early B cells (25). This active form is a 25 kDa single-chain glycoprotein, also known as lymphopoietin-1 (LP-1) (26). IL-7 plays a significant role in regulating the immune system by promoting the maturation of B and T cells, maintaining a reservoir of immature T cells, enhancing the proliferation of memory T cells, and increasing the cytotoxicity of NK cells (27, 28). Additionally, IL-7 is involved in the development and regulation of monocytes, macrophages, dendritic cells, and innate lymphoid cells (29).

Researchers identified a growth factor that promotes the sustained proliferation of human P40 T cells and named it IL-9 (30). This cytokine is a single-chain glycoprotein with a molecular weight of 32 to 39 kDa (30). IL-9 plays a crucial role in the immune system by promoting the proliferation of T cells, B cells, mast cells,



and epithelial cells through the activation of multiple members of the STAT family, such as STAT1, STAT3, and STAT5 (31). Additionally, IL-9 contributes to the proliferation of other cell types, including erythroid progenitor cells, fetal thymocytes, Human megakaryocytic leukemia cell lines, and bone marrow progenitor cells (32).

IL-15 is a cytokine composed of 114 amino acids with a molecular weight of 14-15 kDa, known for stimulating T lymphocyte proliferation (33). Structurally, IL-15 is quite similar to IL-2, as they share the same receptor subunits, the  $\beta$  chain and  $\gamma$  chain, and can activate similar signaling pathways (34). Despite these structural and signaling similarities, IL-15 and IL-2 have distinct functions in the body. For instance, IL-15 can inhibit AICD triggered by IL-2 (35). IL-15 supports the survival of T cells, B cells, and NK cells by inhibiting apoptosis (36). It plays a critical role in the development, homeostasis, and function of NK cells, intraepithelial lymphocytes, dendritic cells, macrophages, and mast cells (37, 38). Additionally, IL-15 is involved in the generation and reactivation of naive T cells, effector T cells, and memory T cells (39).

Discovered in 2000, IL-21 is the most recently identified member of the IL-2 family (40). It is an autocrine cytokine primarily produced by follicular Th cells and Th17 cells, with a molecular weight of 15 kDa (41). IL-21 is notable for its potent regulatory effects on numerous immune cells. It enhances T cell expression and promotes Th cell development through the activation of JAK/STAT, MAPK, and PI3K pathways (42). Additionally, IL-21 is involved in the maturation and proliferation of B cells, class switching, and the production of antigen-specific antibodies (43, 44).

# 3 Pancreatic diseases

The development of pancreatitis is strongly associated with gallstones, hypertriglyceridemia and alcohol consumption (45). Under normal physiological conditions, zymogen is activated in the brush border of the small intestine, triggering the normal activation process of other zymogens. However, if zymogen is prematurely activated in the pancreas, it may cause inflammation, necrosis, and severe injury to pancreatic tissues, ultimately leading to the development of pancreatitis (46). Pancreatitis can be categorized into acute and chronic types based on its clinical manifestations and pathophysiological features. AP is an acute inflammatory response of the pancreas. Over the past decade, AP hospitalizations have increased by about 30%, costing the U.S. \$2.6 billion annually in health care costs (47), and in the United States alone, approximately 300,000 people visit the emergency room each year for this condition (48). Patients with AP experience sudden onset of severe epigastric pain, which tends to radiate down the back

and worsens after a meal, and is accompanied by abdominal compression pain, nausea, vomiting, fever and accelerated pulse rate (5). CP, on the other hand, is a persistent fibroinflammatory state caused by recurrent episodes of varying degrees of pancreatic inflammation, which ultimately leads to irreversible damage to the pancreatic tissue and loss of function (49). Compared with AP, the diagnosis of CP is more challenging, and its diagnostic criteria need to be further studied and clarified.

Abnormal immune cell infiltration and cytokine overexpression play a key role in the development of pancreatitis. Macrophages exhibit different phenotypes at different stages of AP and CP. In AP, they differentiate into the pro-inflammatory M1 type, whereas in CP, they gradually change from the initial M1 type to the M2 type with anti-inflammatory and pro-fibrotic properties (50). In CP, M1type macrophages, as the main immune cells triggering the inflammatory response, promote the expression of proinflammatory factors such as TNF- $\alpha$ , TGF- $\alpha$ , and IL-6 (51), while M2-type macrophages secrete anti-inflammatory mediators such as TGF- $\beta$ 1 and IL-10 (52). It is well known that PSCs play an important role in pancreatic fibrosis by regulating the synthesis and degradation of extracellular matrices (e.g., fibronectin and type I/III collagen) (53). Interestingly, M2-type macrophages can interact with PSCs to stimulate the pro-pancreatic fibrosis effects of PSCs. M2-type macrophages activate PSCs by releasing the cytokines PDGF and TGF-B, and in turn activated PSCs secrete IL-4/IL-13 in an IL-4Ra-dependent manner, inducing macrophage polarization to the M2 phenotype (54).

PC is an extremely lethal and aggressive malignant tumor whose global incidence has more than doubled in the last 25 years (55). The cure rate for PC is extremely low due to late diagnosis and limited treatment options (56). This has led to an increase in its mortality rate from fourth to third in the ranking of cancer causes of death in 2016 (11). A sustained inflammatory response is capable of forming pro-cancer TME and accelerating the transition from CP to PC. In particular, M1-type macrophages exacerbate inflammation and tissue damage by enhancing reactive oxygen species (ROS) production and triggering genetic instability (57). It also secretes a variety of pro-inflammatory cytokines, which bind to the corresponding receptors on the cell surface and subsequently activate signaling pathways such as PI3K/AKT, RAS/MAPK, and JAK, which further induce the activation of pro-proliferative and anti-apoptotic transcription factors such as NF-κB and STAT3 (58). Eventually, CP cells exhibit proliferation, anti-apoptosis, inflammatory response, epithelial mesenchymal transition (EMT), and stem cell properties, completing its transformation to invasive PC (as shown in Figure 2). In clinical practice, the diagnosis of PC usually requires a combination of symptoms, imaging tests and serum biomarkers. Its main symptoms include abdominal pain, abnormal liver function, jaundice, new-onset diabetes mellitus, dyspepsia, nausea and vomiting, back pain, and weight loss (59). CT angiography, magnetic resonance imaging, and ultrasound endoscopy are important imaging modalities (60). Carbohydrate antigen 19-9 is used as a biomarker with high sensitivity and specificity for the



#### FIGURE 2

Macrophages in the progression of CP to PC: M2 macrophages interact with PSCs and contribute to the fibrotic process in the pancreas. At the same time, M1 macrophages release cytotoxins such as ROS, which promote genetic instability. M1 macrophages also release pro-inflammatory factors such as EGF, TNF- $\alpha$ , and IL-6, which bind to the corresponding receptors on the cell surface and activate multiple downstream oncogenic signaling pathways. Ultimately, CP cells exhibit proliferation, anti-apoptosis, inflammatory response, epithelial mesenchymal transition (EMT), and stem cell properties, completing its transformation into an invasive PC.

diagnosis of PC (61). PC cells activate immunosuppressive cells such as PSCs, myeloid-derived suppressor cells, TAMs, and Treg cells, which lead to immune escape from the tumor and exacerbate tumor progression and metastasis (10). Treatment of PC includes immunotherapy, surgery, and chemotherapy. In the treatment of PC, immunotherapy faces challenges due to factors such as fibrosis, hypoxia, and immunosuppressive properties of TME (62). However, immunotherapeutic studies for PC are still ongoing, and various aspects such as immune checkpoint blockade, vaccination, lysovirus, and overdose immunization are available (63). Therefore, continued research into the immune mechanisms of PC is crucial for future prevention and treatment strategies.

## 4 IL-2 family and pancreatic diseases

The IL-2 family plays a crucial role in immune responses by regulating immune cells such as T cells and NK cells, significantly impacting immune-mediated diseases. As our understanding of pancreatic function and its role in diseases deepens, it becomes clear that immune regulation is vital in the development of pancreatitis and PC. Therefore, investigating the potential link between the IL-2 family and these pancreatic diseases is essential for comprehending their immunological background and providing a scientific basis for the development of prevention, diagnosis, and treatment strategies.

## 4.1 IL-2

In pancreatitis, IL-2 acts as a key immunoregulatory factor, significantly promoting the proliferation of Tregs, especially the CD25+CD4+T cell subset. Tregs suppress excessive immune responses through various complex mechanisms, including the release of anti-inflammatory cytokines like TGF-B, IL-10, and IL-35; hydrolyzing ATP into immunosuppressive adenosine; blocking stimulatory signals of CD80 and CD86; and directly eliminating inflammatory cells with cytotoxic molecules (64). Early studies found that knocking out IL-2 (65), IL-2R $\beta$  (66), or CD25 (67) genes in mice leads to systemic inflammation and abnormal lymphocyte proliferation. Specifically, the absence of CD25 results in Treg apoptosis, triggering fatal autoimmune responses. Similar conclusions have been drawn from human studies, where disrupting the expression of CD25 and IL-2RB genes reduces their in vitro inhibitory capacity, causing autoimmune inflammatory diseases (68, 69). This is because, in the absence of IL-2 signaling, Treg proliferation is inhibited. Although Treg cells cannot produce IL-2, they have high-affinity CD25 on their surface, enabling them to capture IL-2 during immune responses, ensuring their survival. When IL-2 is lacking, Treg suppression is impaired, disrupting immune homeostasis and leading to autoimmune diseases like pancreatitis. Effector T cells (Teff cells), on the other hand, only have low-affinity IL-2 receptors (IL-2R $\beta$  and IL-2R $\gamma$ ) on their surface, which prevents them from binding IL-2 preferentially, thus limiting their immune activation (70). However, when T-cell receptors (TCRs) are stimulated, Teff cells can produce IL-2 (71). Concurrently, IL-2 is preferentially captured by CD25 on Treg cells, promoting Treg proliferation through IL-2R binding. In AP, IL-2 employs these mechanisms to exert its immunosuppressive effects, preventing excessive immune responses (as illustrated in Figure 3). Therefore, IL-2 not only helps maintain immune homeostasis but also potentially prevents the worsening of AP and its progression to CP.

In CP, IL-2 has a dual role. On one hand, the aforementioned anti-inflammatory mechanism of IL-2 also applies to CP, helping to reduce pancreatic fibrosis. By promoting Treg growth, IL-2 can inhibit Th2 cells and type 2 innate lymphoid cells in the type 2 immune response, thereby reducing the expression of pro-fibrotic cytokines and helping to control the progression of pancreatic fibrosis (72). On the other hand, excessive activation of IL-2 signaling may exacerbate the inflammatory response, leading to further damage to pancreatic tissue. For instance, IL-2 can increase the production of pro-inflammatory cytokines such as TNF- $\alpha$  and IL-1 $\beta$  and promote the inflammation and tissue damage (73).

In PC, the function of IL-2 is equally complex. On one hand, IL-2 can enhance the immune response against tumors by activating Treg cells. By inducing T cell expression of CD25, IL-2 enhances T cell signaling, thereby increasing the anti-tumor efficacy of cytotoxic Tlymphocytes (CTL) and chimeric antigen receptor T cells (CAR-T) used in immunotherapy (74). In pancreatic ductal adenocarcinoma (PDAC), increased levels of IL-2 correlate with improved patient survival and enhance anti-tumor immunity, particularly by increasing CD8+ T cells, B cells, and NK cells (75). Although IL-2 does not act directly on dendritic cells (DCs), it can indirectly promote the expansion and activation of DCs through activation of T-cells and NK-cells, which can lead to an improvement in anti-tumor immune response (76). In addition, IL-2 treatment improves TME by promoting tumor infiltration by DCs and enhances sustained T-cell immunity against PDAC, whereas co-culture of IL-2 with PBMC significantly enhances the anti-tumor effects of DCs (75). A separate study demonstrated that IL-2 not only facilitates the in vitro expansion of tumor-infiltrating lymphocytes (TIL) and enhances the generation of CD8-positive TIL, thereby improving their tumor recognition and attack capabilities (77). Additionally, IL-2 is used in conjunction with agonist anti-CD3 and 4-1BB antibodies to activate and strengthen the patient's immune system, particularly T cells, which in turn amplifies immune responses against PDAC (78). Sun and other scientists have designed a recombinant IL-2 immune cytokine combining tumorlocalizing antibodies and super mutant IL-2 (sumIL-2), which specifically binds to tumor cells and cytotoxic T cells (CTLs), thus effectively enhancing the anti-cancer effect (79). Moreover, preoperative treatment with sumIL-2 helps prolong patient survival compared to conventional adjuvant therapy.

On the other hand, IL-2 can promote the development of PC by affecting the TME. Tumor cells exploit the IL-2 signaling pathway to promote their proliferation and survival or alter the function of immune cells to evade immune surveillance. The IL-2-STAT5 signaling pathway is the central mechanism of this immune response, promoting T-cell proliferation and survival and regulating the immune response to prevent autoimmune diseases. The pathway activates JAK kinase and STAT5 by binding IL-2 to its receptor, and then phosphorylated STAT5 enters the nucleus to



regulate gene expression (80). In addition, IL-2 is involved in the establishment of anti-tumor immune responses and immune tolerance, plays a crucial role in T-cell development and function, and is central to the progression of a variety of immune-related diseases. In the case of PCs, for example, high expression of proinflammatory cytokine-IL-18 is strongly associated with advanced pathological staging and poor patient prognosis.IL-18 leads to CD8 + T-cell depletion by enhancing IL-2R signaling and activating JAK kinase and p-STAT5 (81). This process not only weakens the immune attack against the tumor, but also reduces the efficacy of immune surveillance, interferes with the formation of memory Tcells, enhances immunosuppression within the TME, inhibits T-cell proliferation and survival, reduces the diversity of T-cell receptors, promotes the escape of the tumor from immune attack, and may affect the efficacy of immunotherapy (82). Additionally, IL-2 can polarize TAMs to the M2 phenotype, aiding tumor cells in evading immune surveillance and promoting tumor growth and metastasis (83). Research also indicates that abnormal activation of the IL-2 signaling pathway may be associated with chemotherapy resistance in PC. To address the issue of radiotherapy (RT) resistance in PDAC, researchers have developed a novel PD-1 targeted IL-2 variant called PD1-IL2v (84). This variant selectively binds to CD122 and CD132 without binding to CD25, preventing excessive systemic spread of IL-2 and reducing potential side effects (84). PD1-IL2v specifically binds to PD-1 receptors on effector immune cells, promoting CD8+T cell proliferation and inhibiting tumor growth and metastasis (85). Additionally, PD1-

IL2v reduces Treg production while continuously generating effector T cells, further inhibiting tumor development and spread (85). When combined with RT, PD1-IL2v exhibits sustained and synergistic anti-tumor effects, significantly reducing PDAC treatment resistance. The combination of recombinant IL-2 and allicin significantly increased the proportions of CD4+, CD8+ T cells and NK cells in the peripheral blood of mice and inhibited PCs by 90.5% (86). These strategies offer new possibilities for improving cancer treatment outcomes.

In summary, IL-2 plays multiple roles in immune regulation and is commonly used in clinical treatments for inflammation and tumors. IL-2 is a key cytokine for the differentiation and activation of cytotoxic effector cells (87). To avoid severe side effects that can arise from systemic IL-2 therapy, lower doses of IL-2 are often used in clinical settings. This approach takes advantage of the high sensitivity of Tregs to IL-2, promoting immune tolerance by increasing Treg numbers (88). However, a decrease in IL-2 levels does not always indicate improvement and may signal worsening pain. In CP, for instance, patients with severe pain have lower detectable levels of IL-2 compared to those with mild or no pain (89), possibly due to IL-2's antinociceptive effects (90). Despite the limitations and potential adverse reactions associated with highdose IL-2, such as the activation of a large number of Tregs, its use is still necessary in certain cases (3). For example, high-dose IL-2 binding to CD25 can enhance IL-2R signaling in CD4+and CD8+T cells in mice, improving the efficacy of vaccines against antigens and more effectively clearing tumor cells (91). Therefore, precise

## 4.2 IL-4

In AP, various traditional Chinese medicines have been shown to alleviate symptoms by enhancing IL-4 expression. For instance, Da Cheng Qi Tang, a herbal formula containing rhubarb, magnolia bark, bitter orange, and mirabilite, boosts IL-4 production and exerts anti-inflammatory effects (92). Baicalin, a flavonoid extracted from the dried root of Scutellaria baicalensis, not only promotes IL-4 expression but also reduces pyroptosis in pancreatic alveolar cells (PACs), alleviating tissue damage and inflammation in hyperlipidemic pancreatitis (93). In models of severe acute pancreatitis (SAP) with lung injury, IL-4 promotes the shift of lung macrophages from a pro-inflammatory M1 type to an antiinflammatory M2 type, reducing inflammation associated with SAP (94). In SAP mouse models, intraperitoneal injection of IL-4 lowers levels of complement regulatory factors DAF and CD59 in the pancreas, significantly improving pancreatic necrosis (95). IL-4 displays its anti-inflammatory properties through various mechanisms: inhibiting macrophage activation and accumulation (96), preventing the production of proinflammatory cytokines TNF- $\alpha$  and IL-1 $\beta$  (97), upregulating the expression of IL-1 receptor antagonist (98), and activating macrophage 15-lipoxygenase (99). These actions may explain the molecular mechanisms by which IL-4 improves SAP and related inflammation.

In contrast to AP, IL-4 may promote disease progression in CP. Studies have shown that elevated IL-4 levels in CP mouse models lead to macrophage polarization to the M2 type, exacerbating pancreatic fibrosis (100). Thioredoxin 1, a redox balance regulator, can inactivate IL-4 by reducing its C46-C99 disulfide bond, thereby inhibiting the generation of M2 macrophages, reducing inflammation, and slowing fibrosis progression in CP (100). Notably, there may be a positive feedback mechanism involving IL-4, PSCs, and macrophages that promotes pancreatic fibrosis in CP. IL-4 released by PSCs promotes the alternative activation of macrophages (54), which in turn produce IL-4R ligands that further activate PSCs. However, IL-4 transfer to PSCs is accompanied by increased expression of the anti-inflammatory cytokine IL-10, helping to suppress inflammation and fibrosis (101). Therefore, the specific role of IL-4 in CP requires further research and exploration.

In the TME of PC, IL-4 levels are elevated, promoting cancer cell growth and acting as an immunosuppressive factor, thereby limiting the effectiveness of immunotherapy. Studies have found that plasma IL-4 levels in PDAC patients are significantly higher than in healthy individuals, and high IL-4 levels are associated with poor prognosis (102). IL-4 promotes PC progression through various mechanisms, including enhancing the activity of cathepsins B and S in TAMs (103), inducing M2-type TAM polarization (104), and activating STAT transcription factors and related signaling pathways (105), which inhibit T cell immune responses against PC. In PC cells, the

expression levels of the transcription factors STAT1, STAT3, and STAT6 differed, with the phosphorylation of STAT3 enhanced by IL-4 stimulation (105) and the nuclear translocation of STAT6 increased in response to IL-4 (106). Dual oxidase 2 (DUOX2) is a key glycoprotein that regulates cellular redox and is associated with chronic inflammation and tumor development (107). In PDAC, upregulation of IL-4 promotes nuclear translocation of STAT6 and significantly increases the expression of DUOX2 mRNA and protein, which drives the development of PDAC and enhances its invasiveness and migration (106). Studies have shown that increased expression of IRS-2 in human PC may promote tumor mitosis and growth through activation of the PI3K signaling pathway (108). Researchers such as Prokopchuk et al. found that IL-4 was able to elicit enhanced tyrosine phosphorylation of IRS-1 and IRS-2, as well as increase the activity of the MAPK and PI3K/Akt signaling pathways (105). In addition, the research team of Traub et al. found that exogenous stimulation of IL-4 could lead to enhanced phosphorylation of pro-oncogenic signals such as c-Jun, ERK-1/2, and STAT3 in Capan1 cells (109), and that the specific STAT3 phosphorylation inhibitor LLL12 was able to inhibit the survival of PC cells (110). Additionally, IL-4 weakens antigendependent T cell proliferation, increases T cell exhaustion, and reduces cytolytic capacity. Cell surface receptors, especially IL-4Ro, are highly specific targets in PC therapy and can directly kill tumor cells overexpressing these receptors while reducing damage to normal tissues. In PC, overexpression of IL-4R $\alpha$  attenuates cell growth and migration after its expression is reduced by shRNA plasmids, and impaired IL-4 signaling in PC cells enhances the inhibition of xenograft tumors (109), suggesting that IL-4R $\alpha$  is a promising therapeutic target.

Currently, CAR-T technology has proven effective in overcoming IL-4-induced immunosuppression and has become a promising option for cancer treatment. For example, the inverted cytokine receptor (ICR) of IL-4/IL-7, composed of the IL-4R extracellular domain and the IL-7R intracellular domain, reduces IL-4's inhibitory effect on CAR-T cells, promoting T cell proliferation and enhancing anti-tumor activity (111). Other types of CAR-T cells, such as 4/15NKG2D-CAR and IL-4/ IL-21 ICR, also show anti-tumor activity; the former converts IL-4 inhibitory signals downstream into IL-15 activation signals (112), while the latter activates the STAT3 pathway (113). Therefore, geneengineered therapies related to IL-4 hold promise as new treatments for PC. However, research by Seifert, M., et al. found that adding IL-4 to anti-CD40 monoclonal antibodies inhibits IgM reactivity to tumor antigens while enhancing B cell immune responses to PC antigens, aiding the immune system in recognizing and eliminating PC cells (114). This indicates that IL-4 may exert anti-tumor activity by affecting the proliferation, apoptosis, and migration of PC cells. Recombinant IL-4-PE toxins, such as IL-4-PE38QQR (115) and IL4(38-37)-PE38KDEL (116), have been shown to inhibit PC progression. Another cytotoxin, consisting of IL-4 and truncated Pseudomonas exotoxin, has specific cytotoxicity against PC cells and demonstrates synergistic antitumor effects in vitro and in various mouse models when combined with gemcitabine (117). Molecules targeting cytokine receptors may have significant toxicity in cancer therapy. In addition, a hybrid peptide (IL-4Ra-lytic) and cytotoxic cleavage peptide, which bind IL-4Ra, showed anticancer effects in PC cells expressing IL-4Ra and in xenograft mouse models (118). These findings further confirm the potential of IL-4R $\alpha$  and its related pathways in PC therapy. In addition, blocking IL-4 had a significant inhibitory effect on PC development, and in particular, the use of IL-4-neutralizing antibodies was able to inhibit the basal proliferation of COLO-357, PANC-1, and MIA PaCa-2 cell lines (105). More critically, by inhibiting IL-4 mRNA expression in the livers of patients with PC malignancy (119) and by improving the physical condition of IL-4-treated tumor-bearing mice (120), it may provide a potentially effective approach for the future treatment of PC patients with tumor-induced malignancy.

In summary, in AP, IL-4 primarily exhibits anti-inflammatory properties and promotes tissue repair. In contrast, in CP and PC, while some evidence suggests that IL-4 may inhibit disease progression, most studies support the notion that IL-4 promotes disease development.

## 4.3 IL-7

IL-7 has been shown to play a role in the progression of pancreatic diseases. In pancreatitis, IL-7 expression levels increase over time, peaking within 72 hours of the acute phase, indicating its involvement in the inflammatory response of pancreatitis (121). In CP, IL-7 can activate fibroblasts, thereby exacerbating pancreatic fibrosis (122). While the exact mechanisms of IL-7 in pancreatitis are not yet well-defined, it is known that IL-7 regulates T cell function and is involved in various immune-related diseases, including diabetes and PC (123, 124). For instance, in type 1 diabetes, IL-7 promotes T cell proliferation by binding to its receptor and activating the JAK1, JAK3, and STAT5 signaling pathways, potentially worsening the disease (123). Future research could further explore the mechanisms of IL-7's interaction with T cells in pancreatitis.

In the field of PC research, the role of IL-7 has been partially elucidated, with some studies suggesting that IL-7 could serve as a potential biomarker for diagnosing PC (125-127). Jang's study found that IL-7R and its mRNA levels in PDAC patients were significantly higher than those in patients with AP and CP (125). Heo's research indicated that IL-7R can be detected in the early stages of PDAC formation, potentially aiding doctors in providing earlier immunotherapy strategies to improve patient survival rates (126). Sahar's study showed that serum IL-7 levels in patients with autoimmune pancreatitis were significantly higher than in PDAC patients, which is important for clinical diagnosis and treatment decisions (127). IL-7 not only serves as a biomarker but also promotes PC development by affecting the apoptosis and maturation of immune cells. IL-7 binds to IL-7Rα, triggering activation of IL-7Rαrelated tyrosine kinases, JAK 1 and JAK3. Activated JAK proteins phosphorylate specific motifs on the IL-7Ra chain to form a binding site for STAT5, which is then bound and phosphorylated to form a dimer and enter the nucleus. In this process, a range of genes in the nucleus that regulate cell growth and survival are affected, other pathways such as PI3K-AKT and MEK/ERK are activated, and the anti-apoptotic proteins Bcl-XL, Bcl-2, and MCL-1 are upregulated, whereas the pro-apoptotic proteins BAX and BAD are downregulated, resulting in increased T cell survival in vivo (128). IL-7 signaling can maintain the survival of memory CD8 T cells by mediating STAT5 and STAT3 activation (129). IL-7 not only activates STAT5 and thus promotes T cell proliferation, differentiation and survival, but also regulates T cell cytotoxicity and drug resistance (130). In PDAC, IL-7 and its activation of the JAK/STAT signaling pathway may make tumor cells resistant to chemotherapeutic agents such as gemcitabine and accelerate tumor progression (131).Normally, IL-7 binding to its receptor promotes the growth and maturation of CD4+T cells by facilitating pSTAT5-NT (132). However, in an immune-deficient environment, secretory proteases like Phospholipase A2 group 1B (PLA2G1B) located on the CD4+T cell membrane may release arachidonic acid (AA), forming abnormal membrane microdomains (aMMDs) on the cell membrane (133). These aMMDs can capture IL-7Ra and yc chains, disrupting normal IL-7 receptor binding and leading to elevated plasma IL-7 levels in PDAC patients (133). High IL-7 levels inhibit STAT5 phosphorylation and nuclear translocation, reducing CD4+T cell proliferation and impairing their anti-tumor immune function, thereby promoting PDAC development (134). This may explain the pro-carcinogenic mechanism of IL-7 in the TME (as illustrated in Figure 4). Given the high expression of IL-7 in PC, researchers have developed a novel CAR-T cell therapy centered around IL-7, capable of secreting human IL-7 and CCL19. This design not only alleviates the issue of T cell infiltration in tumors but also enhances CAR-T cell survival, showing significant anti-tumor effects in PC treatment (135). Li and other investigators (136) developed a novel drug, IL-7-Fc fusion protein (NT-I7), which enhances therapeutic efficacy through enhanced proliferation, activation, and prolonged survival of CAR-T cells.NT-I7 was able to significantly boost the number of CD4+ CAR-T cells, reduce the level of depletion markers on T cells, and enhance T cell persistence and tumor penetration. The development of each of these drugs offers new strategies and hope for PC treatment.

## 4.4 IL-9

IL-9 plays a crucial role in pancreatic diseases. The activation of cathepsin B triggers the activation of trypsin, which is a key step in the onset of AP (137). Xiao et al. found that mogroside IIE, a traditional Chinese medicine component used to treat PC, can reduce cathepsin B activity (138). Their experimental results indicated that mogroside IIE can inhibit the release of IL-9 in mice with AP (138). When exogenous IL-9 was injected into the mice, the effect of mogroside IIE in reducing the activities of cathepsin B and trypsin was negated, suggesting that IL-9 is essential for activating cathepsin B in the pancreas. Further experiments showed that administering an IL-9R antibody to the model could neutralize the effects of IL-9, restoring the activity of mogroside IIE and improving AP (138). These findings support the hypothesis that the pathogenesis of AP may be related to the IL-9/ IL-9R signaling pathway. However, current research on the role of IL-9 in pancreatitis is still insufficient, and more studies are needed to further explore and validate the specific mechanisms of IL-9's action.

In PC research, the role of IL-9 remains controversial despite considerable attention. Hu et al. suggested that IL-9 may promote the proliferation and metastasis of PC cells (139). Lu's experiments



Regulation of CD4+ T cell proliferation by the IL-7 signaling pathway and its mechanism of PLA2G1B-mediated inhibition: Binding of IL-7 to its receptor triggers the phosphorylation of STAT5 (pSTAT5), a process that facilitates the translocation of pSTAT5 to the nucleus (NT), contributing to the reduction of apoptosis and the promotion of proliferation in CD4+ T cells. However, an increase in PLA2G1B leads to the release of arachidonic acid (AA), which in turn contributes to the formation of abnormal membrane microdomains (aMMDs). These aMMDs are capable of intercepting the IL-7 receptor and preventing its interaction with IL-7, which in turn inhibits pSTAT5 activation and nuclear translocation, adversely affecting the survival and proliferation of CD4+ T cells.

showed that targeting and blocking IL-9 could effectively inhibit tumor growth in a PC mouse model (140). However, Th9 cells, a major subset of CD4+T cells, have been shown to produce high levels of IL-9 and exert anti-tumor effects in certain solid tumors such as breast cancer (141) and lung cancer (142). In the TME of PC, Th9 cells are the primary source of IL-9 (143), which suggests that IL-9 should have anti-tumor functions. Chen's research indicated that Th9 cells can activate the cytolytic and secretory functions of NK cells, thus playing a role in tumor surveillance and inhibiting PDAC growth (144). Ahmed's study also found that high levels of IL-9 expression in the PDAC stroma are associated with better prognosis, though IL-9 levels in tumor epithelium and serum showed no significant correlation with patient survival rates (143). Given the complex nature of IL-9 in PC, it has become a potential target for gene-engineered T cell therapy. The IL-9 signaling pathway is not typically a significant player in T cell activation due to the scant expression of IL-9R on T cell surfaces (145). Nevertheless, researchers have ingeniously tapped into this pathway for T cell application by engineering the chimeric receptor o9R (146). This receptor is composed of the extracellular domain (ECD) of the IL-2 receptor and the intracellular domain (ICD) of the IL-9 receptor, which interacts with modified IL-2 (oIL-2), initiating JAK activation and the subsequent phosphorylation of the crucial transcription factors STAT1, STAT3, and STAT5. These phosphorylated factors then dimerize and migrate to the nucleus to modulate gene expression, influencing cell proliferation, differentiation, and survival. The o9R signaling in T cells simultaneously activates STAT1, STAT3, and STAT5, a rare event in natural IL-9 signaling, which may augment T cell stem cell memory characteristics and effector functions. T cells equipped with o9R signaling have shown enhanced antitumor efficacy in both

*in vitro* and *in vivo* models, marked by increased T cell expansion, viability, and tumor infiltration capabilities. This breakthrough is poised to mitigate T cell exhaustion and surmount the immunosuppression encountered in solid tumor therapies, including those for PC.

# 4.5 IL-15

IL-15 is closely linked to the progression of pancreatitis, with its serum levels serving as a predictive marker for AP. Studies have shown that serum IL-15 levels in patients with mild AP are lower than in healthy individuals, which may be a protective mechanism by the body to limit inflammation (147). However, in SAP patients, IL-15 levels are significantly elevated and are associated with disease severity, organ dysfunction, infection risk, and poor prognosis (147). Lower serum IL-15 levels generally indicate a better prognosis, while higher levels may suggest a worse outcome. Additionally, IL-15 levels are negatively correlated with CD4+T and CD8+T cell counts, indicating that it may affect T cell proliferation (147). In a study by Saito et al., IL-15 inhibited the apoptosis of CD4+and CD8+T cells in a rat model of sepsis, improved immune suppression caused by T cell exhaustion, and enhanced the immune response to pathogens, potentially helping to prevent secondary infections (148). This suggests that the regulatory relationship between IL-15 and T cells may be influenced by the tissue environment and pathogenic factors, warranting further investigation. IL-15 exerts a range of therapeutic effects in CP through diverse mechanisms: it mitigates fibrosis by alleviating islet cell atrophy, diminishing collagen accumulation, and suppressing the expression of genes associated

with fibrosis. Additionally, it modulates immunity, safeguarding the pancreas by bolstering iNKT cell counts in the blood and stimulating their production of IFN-y. IL-15 also engages the STAT signaling pathway by binding to IL-15Rα, notably activating STAT5, which modulates the activity of NKT cells (149). Furthermore, it curbs inflammatory responses and limits the influx of inflammatory cells. Directly targeting PSCs, IL-15 curbs their activation and proliferation. It also advances the apoptosis of immune and tumor cells and governs autophagy, thereby impacting cellular survival and functionality (149). These combined effects make IL-15 a potential therapeutic tool for improving CP pathology. Another experiment confirmed these mechanisms and demonstrated that IL-15 also improved CPrelated acute lung injury (150). Thus, IL-15 presents a potential immunotherapeutic approach for CP patients, offering new treatment strategies, particularly in the context of fibrosis and malignant phenotypes.

IL-15 exerts multiple mechanisms of action in the treatment of CP: it exhibits anti-fibrotic effects, which include attenuating islet cell atrophy, decreasing collagen deposition, and inhibiting the expression of fibrosis-related genes. In addition, IL-15 enhances immune protection by elevating the number of iNKT cells in the blood and stimulating the production of IFN- $\gamma$  by these cells. It also activates STAT family signaling, particularly STAT5, by binding to IL-15R $\alpha$ , which in turn regulates NKT cell activity.IL-15 also inhibits inflammatory responses and reduces the accumulation of inflammatory cells. It acts directly on PSCs, inhibiting the activation and proliferation of these cells. In addition, IL-15 promotes apoptosis in immune cells and tumor cells and regulates the autophagic process, thereby affecting cell survival and function. These interacting mechanisms confer potential therapeutic value to IL-15 in ameliorating CP pathology.

Research on IL-15 in PC has garnered extensive attention. IL-15 is not only negatively correlated with the progression of PDAC but is also considered an important independent predictor of metastatic PDAC. In the South African population, IL-15 has been identified as a potential prognostic biomarker for metastatic disease in PDAC patients (151). IL-15 shows potential in PC treatment by promoting the generation, proliferation, and activity of anti-tumor NK cells and CD8+T cells, thereby enhancing the immune response against tumors (152). Although clinical studies have indicated that IL-15 has potential effects when combined with existing drugs, its efficacy as a monotherapy is limited due to its short half-life (153). Consequently, IL-15-based combination therapies are being widely researched. The combination of IL-15 and metformin showed excellent efficacy in PC therapy, which significantly inhibited the expression of the cell cycle-promoting Cyclin D1 and the anti-apoptotic protein Bcl-2 and promoted the upregulation of the expression of the apoptosis-promoting protein Bax in tumor cells (154). In addition, since overactivation of the Akt/mTOR/STAT3 signaling pathway inhibits autophagy, the combination therapy, by inhibiting this signaling pathway, deregulated the inhibition of autophagy and enhanced the expression of the autophagy proteins LC3 and Beclin-1. Together, these changes at the molecular level promoted autophagy and apoptosis in tumor cells, demonstrating its anti-tumor potential (154). The Van team discovered that the combined use of CD40 agonists and IL-15 can enhance immune response and anti-cancer effects (155). Their experiments showed that the combination therapy enhances the efficiency of IL-15 binding to CD8+T cells and NK cells by promoting IL-15R $\alpha$  expression through the CD40 agonist, resulting in a synergistic effect and significantly improving the long-term survival rates of PDAC mice (155). Utilizing the more stable binding characteristic of IL-15 with IL-15R $\alpha$ , four IL-15 superagonists (composed of IL-15 and truncated forms of IL-15R $\alpha$  complexes) have been designed and entered clinical development, yielding encouraging preliminary results (156). This opens new prospects for the future clinical treatment of malignant tumors such as PC.

Bioengineering based on IL-15 has opened a new avenue for the treatment of PC, providing an alternative to traditional combination therapies. For example, hIL15-ABD, a fusion of IL-15 with the albumin-binding domain (ABD), has been used to treat PDAC, enhancing anti-tumor effects and extending the drug's halflife (157). When combined with the chemotherapy drug Nabpaclitaxel, hIL15-ABD effectively reduces the number of immunosuppressive Tregs, while promoting the accumulation of CD8+T cells, NK cells, and M1-type macrophages, significantly inhibiting PDAC growth in mice (157). Nelson et al. developed an oncolvtic virus, VSV-IL-15, which enhances anti-tumor immune responses by transducing IL-15, increasing PDAC regression rates and extending patient survival (158). Additionally, a triple therapy combining NK cell immunotherapy, VSV-IL-15 virotherapy, and PD-1 blockade further strengthened immune cell activation and function, leading to more tumor regressions, with some patients achieving complete tumor eradication (158). Zhao et al. developed a dual cross-linked hydrogel system, P-aTIGIT&PDA@IL-15@gel, capable of sustained release of IL-15 and anti-TIGIT, effectively reactivating CD8+T cells and NK cells, and inhibiting residual tumor growth and distant metastasis in mouse PC models (159). Similar to previously mentioned factors, many IL-15-based genetically engineered T cells have been designed. In PDAC, cancer cells and CAFs within the TME jointly promote tumor growth and metastasis. Astrid and her team developed CD70-CAR-IL-15 NK cells, which demonstrated the ability to eliminate CAFs and improve prognosis, particularly in PDAC patients with significant desmoplasia (160). By adding IL-15 to CAR-iMacs targeting prostate stem cell antigen (PSCA), Shah et al. not only enhanced the viability of these macrophages to avoid apoptosis, but also succeeded in activating NK cells and T cells, which in turn boosted adaptive immune responses to PC (161). Teng et al. designed PSCA CAR\_s15 NK cells, which enhance IL-15 activation of NK cells, improving tumor cell killing efficiency and showing significant inhibitory effects on PSCA-positive PC cells (162). Xu and his team developed a novel CAR-NK92 cell line, incorporating the IL-15Ra-sushi/IL-15 complex and a PD-1 signal inverter, named SP (Sushi-IL15-PD1) (163). This cell line targets PD-L1-expressing tumor cells, enhancing cytotoxicity and exhibiting significant killing effects on PDAC cells (163). These studies bring innovative perspectives and approaches to PC treatment, highlighting the need for ongoing research and development in this field.

#### TABLE 1 Clinical applications of targeted IL-2 family for the treatment of pancreatic diseases.

Cytokines	Existing drug therapies	Mechanisms	Diseases	References
IL-4	Da Cheng Qi Tang	Increased IL-4	AP	(92)
IL-4	Baicalin	Decreased pancreatic alveolar cell pyroptosis	Hyperlipidemic pancreatitis	(93)
IL-4	Thioredoxin 1	Decreased IL-4, M2- type macrophages	СР	(100)
IL-15	rIL-15	Increased NK cells, IFN-γ	СР	(149)
IL-15	IL-15 and metformin	Enhanced tumor cell autophagy and apoptosis	PC	(154)
IL-15	IL-15 and CD40 agonist	Increased CD8+T and NK cells	PDAC	(155)
IL-15	hIL15-ABD and Nab-paclitaxel	Increased CD8+T, NK, and M1-type macrophages; decreased Tregs	PDAC	(157)
IL-9	Mogroside IIE	Decreased histone B and IL-9	AP	(138)
IL-21	Nab-paclitaxel and gemcitabine	Inhibition of PD-L1/PD-1 signaling pathway	PDAC	(167)

Interestingly, exercise can stimulate the release of IL-15 from skeletal muscle, prompting researchers to explore the potential of exercise-regulated IL-15 levels for treating PC. Kurz et al. found that aerobic exercise activates the IL-15/IL-15R $\alpha$  pathway, promoting CD8+T cell infiltration into PDAC cells, thereby enhancing antitumor immune responses in both mice and humans (164). Additionally, using IL-15 superagonists to activate this pathway can mimic the anti-tumor effects of exercise, including tumor growth inhibition, prolonged survival, and enhanced chemotherapy efficacy, providing a new theoretical basis for PC treatment strategies (164). Moderate exercise has also been found to increase IL-15R $\alpha$  levels in adipose tissue, reducing the risk of obesity-related PDAC (165). In non-obese mice, overexpression of IL-15 helps slow PDAC progression, while exercise itself may counteract obesity-induced PDAC progression by reducing

inflammation (165). These findings further highlight the potential value of exercise and IL-15 in PC treatment.

# 4.6 IL-21

Although research on IL-21 and pancreatitis is relatively limited, studies have begun to explore the potential connection between IL-21 and PC. It is generally believed that IL-21 exerts anti-tumor effects in PC. Early studies indicated that IL-21 inhibits tumor growth by activating NK cells (166). Recent research has found that in PDAC, the infiltration of T follicular helper (Tfh) cells promotes IL-21 secretion, which helps B cells differentiate into plasma cells, thereby enhancing the humoral immune response (167). Meanwhile, tumor cells inhibit T-cell activation by expressing PD-L1 that binds to PD-1

TABLE 2 Cellular engineering drugs based on the structure of the IL-2 family.

Cytokines	Existing drug therapies	Mechanisms	Diseases	References
IL-2	PD1-IL2v	Increased CD8+T and Teff cells; Decreased Tregs	PDAC	(85)
IL-4	IL-4/IL-7 ICR	Increased T cells	РС	(111)
IL-4	4/15NKG2D-CAR	Activated IL-15	РС	(112)
IL-4	IL-4/IL-21 ICR	Activated STAT3 pathway	РС	(113)
IL-15	VSV-IL-15	Activated IL-15	PDAC	(158)
IL-15	P-aTIGIT&PDA@IL-15@ge	Increased CD8+T and NK cells	РС	(159)
IL-15	CD70-CAR-IL-15 NK	Decreased fibroblasts	PDAC	(160)
IL-15	PSCA CAR_s15 NK	Increased NK cells	РС	(162)
IL-15	Sushi-IL15-PD1	Enhanced PD-L1 cytotoxicity	PDAC	(163)
IL-9	IL-2Rβ-ECD-IL-9R-ICD	Activated STAT pathway	РС	(146)
IL-21	VVL21	Increased CD8+T and NK cells; Enhanced macrophage M1 polarization	PC	(170)

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on T-cells, activating the PD-1/PD-L1 pathway for immune escape (168). However, the PD-L1/PD-1 pathway may inhibit the antitumor effects of IL-21. When patients receive neoadjuvant chemotherapy with Nab-paclitaxel and gemcitabine, this inhibition can be reversed, restoring the anti-tumor ability of IL-21 (167). McMichael et al. showed that IL-21 not only significantly enhanced the effector function of NK cells on cetuximab-coated PC cells and increased ADCC, an effect independent of the KRAS gene mutation status of the tumor cells; it also promoted IL-21-activated NK cells to secrete a high level of IFN-y and chemokines, enhanced T-cell chemotaxis and enhanced NK cell signal transduction by activating the ERK and STAT1 signaling pathways to enhance NK cell signaling; in addition, the combination of IL-21 and cetuximab significantly inhibited the growth of EGFR-positive PCs in a mouse model, and the effect was even better when combined with gemcitabine (169). Giulia Marelli and her team developed a novel virus, VVL21, by embedding IL-21 into a vaccinia virus strain using gene-editing technology (170). This virus enhances the accumulation of effector CD8+T cells within tumors, boosts NK cell activity, and promotes macrophage polarization to the M1 phenotype, offering an effective strategy for clinical PC treatment (170). However, some studies suggest that IL-21 may also play a role in promoting PC invasion. A study by Alica et al. found that high density of IL-21+ immune cells and expression of IL-21R and Blimp-1 in tumor tissues of PDAC patients correlated with poor patient prognosis (171). IL-21 may activate the ERK and STAT3 signaling pathways through binding to IL-21R on the surface of PC cells that which in turn promotes the upregulation of Blimp-1 expression and enhances the invasive ability of tumor cells (171). Therefore, further in-depth research is necessary to understand the complex relationship between IL-21 and PC.

# 5 Discussion

In summary, the IL-2 family plays a complex and multifaceted role in pancreatitis and PC. Unlike IL-1 family members, which act directly as inflammatory mediators, IL-2 family members primarily play a role in immune regulation and inflammatory responses. They may also influence disease progression and response to therapy by regulating the TME and cellular metabolism in the pancreas, which in turn affects disease progression and response to therapy. For example, some IL-2 family members may alleviate pancreatic damage by inhibiting inflammatory responses, while others may aid in restoring pancreatic function by promoting tissue repair. Future research needs to further elucidate the specific mechanisms by which IL-2 family members operate in pancreatic diseases. This includes understanding how they affect the survival, proliferation, and differentiation of pancreatic cells, as well as how they interact with other cell types within the pancreas. Additionally, research should focus on the expression patterns of IL-2 family members at different stages of pancreatic diseases and how they respond to existing treatment methods. As shown in Tables 1, 2.

By conducting in-depth research into the mechanisms of the IL-2 family in pancreatic diseases, scientists hope to discover new

biomarkers for early diagnosis and assessment of disease severity. This research could also provide insights for developing new therapeutic strategies, such as modulating the activity of IL-2 family members to regulate immune responses or creating targeted therapies to enhance treatment efficacy and reduce adverse effects. Current research has focused on the immune-activating effects of the family, while relatively few studies have been conducted on its inhibitors. Future studies should also take into account individual differences, such as genetic background and environmental factors, which may influence the role of IL-2 family members. Personalized medicine approaches could adjust the intervention strategies of IL-2 family members based on the specific conditions of patients, aiming for optimal therapeutic outcomes and minimal side effects. In conclusion, research on the IL-2 family in pancreatic diseases is a field filled with challenges and opportunities. As we gain a deeper understanding of the mechanisms of these cytokines, there is hope for developing more precise and effective treatments to improve the prognosis of patients with pancreatitis and PC.

# Author contributions

YZ: Conceptualization, Visualization, Writing – original draft, Writing – review & editing. ZL: Visualization, Writing – review & editing. KN: Conceptualization, Funding acquisition, Project administration, Supervision, Writing – review & editing. ZW: Funding acquisition, Project administration, Writing – review & editing. JL: Funding acquisition, Project administration, Writing – review & editing.

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# Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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# References

1. Zhou Y, Quan G, Liu Y, Shi N, Wu Y, Zhang R, et al. The application of Interleukin-2 family cytokines in tumor immunotherapy research. *Front Immunol.* (2023) 14:1090311. doi: 10.3389/fimmu.2023.1090311

2. Malek TR. The biology of interleukin-2. Annu Rev Immunol. (2008) 26:453–79. doi: 10.1146/annurev.immunol.26.021607.090357

3. Sim GC, Radvanyi L. The IL-2 cytokine family in cancer immunotherapy. *Cytokine Growth Factor Rev.* (2014) 25:377–90. doi: 10.1016/j.cytogfr.2014.07.018

4. Wang S, Ni HM, Chao X, Wang H, Bridges B, Kumer S, et al. Impaired TFEBmediated lysosomal biogenesis promotes the development of pancreatitis in mice and is associated with human pancreatitis. *Autophagy*. (2019) 15:1954–69. doi: 10.1080/ 15548627.2019.1596486

5. Lankisch PG, Apte M, Banks PA. Acute pancreatitis. Lancet. (2015) 386:85–96. doi: 10.1016/S0140-6736(14)60649-8

6. Yadav D, Lowenfels AB. The epidemiology of pancreatitis and pancreatic cancer. *Gastroenterology*. (2013) 144:1252–61. doi: 10.1053/j.gastro.2013.01.068

7. Singh VK, Yadav D, Garg PK. Diagnosis and management of chronic pancreatitis: A review. Jama. (2019) 322:2422-34. doi: 10.1001/jama.2019.19411

8. Mayerle J, Sendler M, Hegyi E, Beyer G, Lerch MM, Sahin-Tóth M. Genetics, cell biology, and pathophysiology of pancreatitis. *Gastroenterology*. (2019) 156:1951–1968.e1. doi: 10.1053/j.gastro.2018.11.081

9. Shamoon M, Deng Y, Chen YQ, Bhatia M, Sun J. Therapeutic implications of innate immune system in acute pancreatitis. *Expert Opin Ther Targets*. (2016) 20:73–87. doi: 10.1517/14728222.2015.1077227

10. Zheng L, Xue J, Jaffee EM, Habtezion A. Role of immune cells and immunebased therapies in pancreatitis and pancreatic ductal adenocarcinoma. *Gastroenterology*. (2013) 144:1230–40. doi: 10.1053/j.gastro.2012.12.042

11. Tempero MA. NCCN guidelines updates: pancreatic cancer. J Natl Compr Canc Netw. (2019) 17:603–5. doi: 10.6004/jnccn.2019.5007

12. Ullman NA, Burchard PR, Dunne RF, Linehan DC. Immunologic strategies in pancreatic cancer: making cold tumors hot. *J Clin Oncol.* (2022) 40:2789–805. doi: 10.1200/JCO.21.02616

13. De Visser KE, Joyce JA. The evolving tumor microenvironment: From cancer initiation to metastatic outgrowth. *Cancer Cell.* (2023) 41:374–403. doi: 10.1016/j.ccell.2023.02.016

14. Hartupee C, Nagalo BM, Chabu CY, Tesfay MZ, Coleman-Barnett J, West JT, et al. Pancreatic cancer tumor microenvironment is a major therapeutic barrier and target. *Front Immunol.* (2024) 15:1287459. doi: 10.3389/fimmu.2024.1287459

15. Briukhovetska D, Dörr J, Endres S, Libby P, Dinarello CA, Kobold S. Interleukins in cancer: from biology to therapy. *Nat Rev Cancer*. (2021) 21:481–99. doi: 10.1038/s41568-021-00363-z

16. Akdis M, Burgler S, Crameri R, Eiwegger T, Fujita H, Gomez E, et al. Interleukins, from 1 to 37, and interferon-γ: receptors, functions, and roles in diseases. J Allergy Clin Immunol. (2011) 127:701–21.e1-70. doi: 10.1016/j.jaci.2010.11.050

17. Bazan JF. Structural design and molecular evolution of a cytokine receptor superfamily. *Proc Natl Acad Sci U.S.A.* (1990) 87:6934–8. doi: 10.1073/pnas.87.18.6934

18. Leonard WJ, Lin JX, O'shea JJ. The  $\gamma$ (c) family of cytokines: basic biology to the rapeutic ramifications. *Immunity*. (2019) 50:832–50. doi: 10.1016/j.immuni.2019.03.028

19. Ross SH, Cantrell DA. Signaling and function of interleukin-2 in T lymphocytes. *Annu Rev Immunol.* (2018) 36:411–33. doi: 10.1146/annurev-immunol-042617-053352

20. Morgan DA, Ruscetti FW, Gallo R. Selective *in vitro* growth of T lymphocytes from normal human bone marrows. *Science.* (1976) 193:1007-8. doi: 10.1126/ science.181845

21. Liao W, Lin JX, Leonard WJ. IL-2 family cytokines: new insights into the complex roles of IL-2 as a broad regulator of T helper cell differentiation. *Curr Opin Immunol.* (2011) 23:598–604. doi: 10.1016/j.coi.2011.08.003

22. Abbas AK, Trotta E, Simeonov D,R, Marson A, Bluestone JA. Revisiting IL-2: Biology and therapeutic prospects. *Sci Immunol.* (2018) 3(25):eaat1482. doi: 10.1126/sciimmunol.aat1482

23. Shi J, Song X, Traub B, Luxenhofer M, Kornmann M. Involvement of IL-4, IL-13 and their receptors in pancreatic cancer. *Int J Mol Sci.* (2021) 22(6):2998. doi: 10.3390/ ijms22062998

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24. Naz S, Baig N, Khalil R, Ul-Haq Z. Characterization of cryptic allosteric site at IL-4 $R\alpha$ : New paradigm towards IL-4/IL-4R inhibition. *Int J Biol Macromol.* (2019) 123:239–45. doi: 10.1016/j.ijbiomac.2018.10.204

25. Hunt P, Robertson D, Weiss D, Rennick D, Lee F, Witte ON. A single bone marrow-derived stromal cell type supports the *in vitro* growth of early lymphoid and myeloid cells. *Cell*. (1987) 48:997–1007. doi: 10.1016/0092-8674(87)90708-2

26. Jiang Q, Li WQ, Aiello FB, Mazzucchelli R, Asefa B, Khaled AR, et al. Cell biology of IL-7, a key lymphotrophin. *Cytokine Growth Factor Rev.* (2005) 16:513–33. doi: 10.1016/j.cytogfr.2005.05.004

27. Bradley LM, Haynes L, Swain SL. IL-7: maintaining T-cell memory and achieving homeostasis. *Trends Immunol.* (2005) 26:172–6. doi: 10.1016/j.it.2005.01.004

28. Brenchley JM, Douek DC, Ambrozak DR, Chatterji M, Betts MR, Davis LS, et al. Expansion of activated human naïve T-cells precedes effector function. *Clin Exp Immunol.* (2002) 130:432–40. doi: 10.1046/j.1365-2249.2002.02015.x

29. Chen D, Tang TX, Deng H, Yang XP, Tang ZH. Interleukin-7 biology and its effects on immune cells: mediator of generation, differentiation, survival, and homeostasis. *Front Immunol.* (2021) 12:747324. doi: 10.3389/fimmu.2021.747324

30. Uyttenhove C, Simpson RJ, Van Snick J. Functional and structural characterization of P40, a mouse glycoprotein with T-cell growth factor activity. *Proc Natl Acad Sci U.S.A.* (1988) 85:6934–8. doi: 10.1073/pnas.85.18.6934

31. Benevides L, Costa RS, Tavares LA, Russo M, Martins GA, Da Silva LLP, et al. B lymphocyte-induced maturation protein 1 controls T(H)9 cell development, IL-9 production, and allergic inflammation. *J Allergy Clin Immunol.* (2019) 143:1119-1130.e3. doi: 10.1016/j.jaci.2018.06.046

32. Chakraborty S, Kubatzky KF, Mitra DK. An update on interleukin-9: from its cellular source and signal transduction to its role in immunopathogenesis. *Int J Mol Sci.* (2019) 20(9):2113. doi: 10.3390/ijms20092113

33. Carson WE, Giri JG, Lindemann MJ, Linett ML, Ahdieh M, Paxton R, et al. Interleukin (IL) 15 is a novel cytokine that activates human natural killer cells via components of the IL-2 receptor. *J Exp Med.* (1994) 180:1395–403. doi: 10.1084/ jem.180.4.1395

34. Waldmann TA. The biology of interleukin-2 and interleukin-15: implications for cancer therapy and vaccine design. *Nat Rev Immunol.* (2006) 6:595–601. doi: 10.1038/ nri1901

35. Waldmann TA, Miljkovic MD, Conlon KC. Interleukin-15 (dys)regulation of lymphoid homeostasis: Implications for therapy of autoimmunity and cancer. *J Exp Med.* (2020) 217(1). doi: 10.1084/jem.20191062

36. Fehniger TA, Caligiuri MA. Interleukin 15: biology and relevance to human disease. Blood. (2001) 97:14–32. doi: 10.1182/blood.V97.1.14

37. Waldmann TA, Tagaya Y. The multifaceted regulation of interleukin-15 expression and the role of this cytokine in NK cell differentiation and host response to intracellular pathogens. *Annu Rev Immunol.* (1999) 17:19–49. doi: 10.1146/ annurev.immunol.17.1.19

38. Waldmann TA, Dubois S, Miljkovic MD, Conlon KC. IL-15 in the combination immunotherapy of cancer. *Front Immunol.* (2020) 11:868. doi: 10.3389/fimmu.2020.00868

39. Kokaji AI, Hockley DL, Kane KP. IL-15 transpresentation augments CD8+ T cell activation and is required for optimal recall responses by central memory CD8+ T cells. *J Immunol.* (2008) 180:4391–401. doi: 10.4049/jimmunol.180.7.4391

40. Parrish-Novak J, Dillon SR, Nelson A, Hammond A, Sprecher C, Gross JA, et al. Interleukin 21 and its receptor are involved in NK cell expansion and regulation of lymphocyte function. *Nature*. (2000) 408:57–63. doi: 10.1038/35040504

41. Spolski R, Leonard WJ. Interleukin-21: basic biology and implications for cancer and autoimmunity. *Annu Rev Immunol.* (2008) 26:57–79. doi: 10.1146/annurev.immunol.26.021607.090316

42. Long D, Chen Y, Wu H, Zhao M, Lu Q. Clinical significance and immunobiology of IL-21 in autoimmunity. *J Autoimmun*. (2019) 99:1-14. doi: 10.1016/j.jaut.2019.01.013

43. Vinuesa CG, Linterman MA, Yu D, Maclennan IC. Follicular helper T cells. Annu Rev Immunol. (2016) 34:335-68. doi: 10.1146/annurev-immunol-041015-055605

44. Victora GD, Nussenzweig MC. Germinal centers. Annu Rev Immunol. (2012) 30:429–57. doi: 10.1146/annurev-immunol-020711-075032

45. Lee PJ, Papachristou GI. New insights into acute pancreatitis. Nat Rev Gastroenterol Hepatol. (2019) 16:479-96. doi: 10.1038/s41575-019-0158-2

46. Vlavcheski F, O'neill EJ, Gagacev F, Tsiani E. Effects of berberine against pancreatitis and pancreatic cancer. *Molecules.* (2022) 27(23):8630. doi: 10.3390/molecules27238630

47. Peery AF, Crockett SD, Murphy CC, Jensen ET, Kim HP, Egberg MD, et al. Burden and cost of gastrointestinal, liver, and pancreatic diseases in the United States: update 2021. *Gastroenterology*. (2022) 162:621–44. doi: 10.1053/j.gastro.2021.10.017

48. Mederos MA, Reber HA, Girgis MD. Acute pancreatitis: A review. Jama. (2021) 325:382–90. doi: 10.1001/jama.2020.20317

49. Beyer G, Habtezion A, Werner J, Lerch MM, Mayerle J. Chronic pancreatitis. *Lancet.* (2020) 396:499–512. doi: 10.1016/S0140-6736(20)31318-0

50. Gukovskaya AS, Gukovsky I, Algül H, Habtezion A. Autophagy, inflammation, and immune dysfunction in the pathogenesis of pancreatitis. *Gastroenterology*. (2017) 153:1212–26. doi: 10.1053/j.gastro.2017.08.071

51. Hu F, Lou N, Jiao J, Guo F, Xiang H, Shang D. Macrophages in pancreatitis: Mechanisms and therapeutic potential. *BioMed Pharmacother*. (2020) 131:110693. doi: 10.1016/j.biopha.2020.110693

52. Dai JJ, Jiang MJ, Wang XP, Tian L. Inflammation-related pancreatic carcinogenesis: mechanisms and clinical potentials in advances. *Pancreas*. (2017) 46:973-85. doi: 10.1097/MPA.00000000000886

53. Wynn TA, Barron L. Macrophages: master regulators of inflammation and fibrosis. Semin Liver Dis. (2010) 30:245-57. doi: 10.1055/s-0030-1255354

54. Xue J, Sharma V, Hsieh MH, Chawla A, Murali R, Pandol SJ, et al. Alternatively activated macrophages promote pancreatic fibrosis in chronic pancreatitis. *Nat Commun.* (2015) 6:7158. doi: 10.1038/ncomms8158

55. Klein AP. Pancreatic cancer epidemiology: understanding the role of lifestyle and inherited risk factors. *Nat Rev Gastroenterol Hepatol.* (2021) 18:493–502. doi: 10.1038/ s41575-021-00457-x

56. Hao L, Zeng XP, Xin L, Wang D, Pan J, Bi YW, et al. Incidence of and risk factors for pancreatic cancer in chronic pancreatitis: A cohort of 1656 patients. *Dig Liver Dis.* (2017) 49:1249–56. doi: 10.1016/j.dld.2017.07.001

57. Wu Y, Antony S, Meitzler JL, Doroshow JH. Molecular mechanisms underlying chronic inflammation-associated cancers. *Cancer Lett.* (2014) 345:164–73. doi: 10.1016/j.canlet.2013.08.014

58. Ardito CM, Grüner BM, Takeuchi KK, Lubeseder-Martellato C, Teichmann N, Mazur PK, et al. EGF receptor is required for KRAS-induced pancreatic tumorigenesis. *Cancer Cell.* (2012) 22:304–17. doi: 10.1016/j.ccr.2012.07.024

59. Schmidt-Hansen M, Berendse S, Hamilton W. Symptoms of pancreatic cancer in primary care: A systematic review. *Pancreas.* (2016) 45:814–8. doi: 10.1097/MPA.00000000000527

60. Al-Hawary MM, Francis IR, Chari ST, Fishman EK, Hough DM, Lu DS, et al. Pancreatic ductal adenocarcinoma radiology reporting template: consensus statement of the society of abdominal radiology and the american pancreatic association. *Gastroenterology*. (2014) 146:291–304.e1. doi: 10.1053/j.gastro.2013.11.004

61. Ballehaninna UK, Chamberlain RS. The clinical utility of serum CA 19-9 in the diagnosis, prognosis and management of pancreatic adenocarcinoma: An evidence based appraisal. *J Gastrointest Oncol.* (2012) 3:105–19. doi: 10.3978/j.issn.2078-6891.2011.021

62. Huber M, Brehm CU, Gress TM, Buchholz M, Alashkar Alhamwe B, Von Strandmann EP, et al. The immune microenvironment in pancreatic cancer. *Int J Mol Sci.* (2020) 21(19):7307. doi: 10.3390/ijms21197307

63. Principe DR, Korc M, Kamath SD, Munshi HG, Rana A. Trials and tribulations of pancreatic cancer immunotherapy. *Cancer Lett.* (2021) 504:1–14. doi: 10.1016/j.canlet.2021.01.031

64. Josefowicz SZ, Lu LF, Rudensky AY. Regulatory T cells: mechanisms of differentiation and function. *Annu Rev Immunol.* (2012) 30:531-64. doi: 10.1146/ annurev.immunol.25.022106.141623

65. Sadlack B, Merz H, Schorle H, Schimpl A, Feller AC, Horak I. Ulcerative colitislike disease in mice with a disrupted interleukin-2 gene. *Cell.* (1993) 75:253–61. doi: 10.1016/0092-8674(93)80067-O

66. Suzuki H, Kündig TM, Furlonger C, Wakeham A, Timms E, Matsuyama T, et al. Deregulated T cell activation and autoimmunity in mice lacking interleukin-2 receptor beta. *Science*. (1995) 268:1472–6. doi: 10.1126/science.7770771

67. Willerford DM, Chen J, Ferry JA, Davidson L, Ma A, Alt FW. Interleukin-2 receptor alpha chain regulates the size and content of the peripheral lymphoid compartment. *Immunity*. (1995) 3:521–30. doi: 10.1016/1074-7613(95)90180-9

68. Zhang Z, Gothe F, Pennamen P, James JR, Mcdonald D, Mata CP, et al. Human interleukin-2 receptor  $\beta$  mutations associated with defects in immunity and peripheral tolerance. J Exp Med. (2019) 216:1311–27. doi: 10.1084/jem.20182304

69. Sharfe N, Dadi HK, Shahar M, Roifman CM. Human immune disorder arising from mutation of the alpha chain of the interleukin-2 receptor. *Proc Natl Acad Sci U.S.A.* (1997) 94:3168–71. doi: 10.1073/pnas.94.7.3168

Xu L, Song X, Su L, Zheng Y, Li R, Sun J. New therapeutic strategies based on IL-2 to modulate Treg cells for autoimmune diseases. *Int Immunopharmacol.* (2019) 72:322–9. doi: 10.1016/j.intimp.2019.03.064

71. Boyman O, Sprent J. The role of interleukin-2 during homeostasis and activation of the immune system. *Nat Rev Immunol.* (2012) 12:180–90. doi: 10.1038/nri3156

72. Glaubitz J, Wilden A, Golchert J, Homuth G, Völker U, Bröker BM, et al. In mouse chronic pancreatitis CD25(+)FOXP3(+) regulatory T cells control pancreatic fibrosis by suppression of the type 2 immune response. *Nat Commun.* (2022) 13:4502. doi: 10.1038/s41467-022-32195-2

73. Lee HK, Nam MW, Go RE, Koo J, Kim TH, Park JE, et al. TGF- $\beta$ 2 antisense oligonucleotide enhances T-cell mediated anti-tumor activities by IL-2 via attenuation of fibrotic reaction in a humanized mouse model of pancreatic ductal adenocarcinoma. *BioMed Pharmacother*. (2023) 159:114212. doi: 10.1016/j.biopha.2022.114212

74. Luu M, Riester Z, Baldrich A, Reichardt N, Yuille S, Busetti A, et al. Microbial short-chain fatty acids modulate CD8(+) T cell responses and improve adoptive immunotherapy for cancer. *Nat Commun.* (2021) 12:4077. doi: 10.1038/s41467-021-24331-1

75. Gong T, Huang X, Wang Z, Chu Y, Wang L, Wang Q. IL-2 promotes expansion and intratumoral accumulation of tumor infiltrating dendritic cells in pancreatic cancer. *Cancer Immunol Immunother*. (2024) 73:84. doi: 10.1007/s00262-024-03669-7

76. Raeber ME, Rosalia RA, Schmid D, Karakus U, Boyman O. Interleukin-2 signals converge in a lymphoid-dendritic cell pathway that promotes anticancer immunity. *Sci Transl Med.* (2020) 12(561):eaba5464. doi: 10.1126/scitranslmed.aba5464

77. Tavera RJ, Forget MA, Kim YU, Sakellariou-Thompson D, Creasy CA, Bhatta A, et al. Utilizing T-cell activation signals 1, 2, and 3 for tumor-infiltrating lymphocytes (TIL) expansion: the advantage over the sole use of interleukin-2 in cutaneous and uveal melanoma. *J Immunother*. (2018) 41:399–405. doi: 10.1097/CJI.00000000000230

78. Amaria R, Knisely A, Vining D, Kopetz S, Overman MJ, Javle M, et al. Efficacy and safety of autologous tumor-infiltrating lymphocytes in recurrent or refractory ovarian cancer, colorectal cancer, and pancreatic ductal adenocarcinoma. *J Immunother Cancer*. (2024) 12(2):e006822. doi: 10.1136/jitc-2023-006822

79. Sun Z, Ren Z, Yang K, Liu Z, Cao S, Deng S, et al. A next-generation tumortargeting IL-2 preferentially promotes tumor-infiltrating CD8(+) T-cell response and effective tumor control. *Nat Commun.* (2019) 10:3874. doi: 10.1038/s41467-019-11782-W

80. Wang T, Wang J, Xu H, Yan H, Liu Y, Zhang N, et al. Salvianolic acid B alleviates autoimmunity in Treg-deficient mice via inhibiting IL2-STAT5 signaling. *Phytother Res.* (2024) 38:3825–36. doi: 10.1002/ptr.v38.7

81. Stromnes IM. IL18 at the crossroads between chronic inflammation and T-cell exhaustion in pancreatic cancer. *Cancer Immunol Res.* (2023) 11:400. doi: 10.1158/2326-6066.CIR-23-0145

82. St Paul M, Ohashi PS. The roles of CD8(+) T cell subsets in antitumor immunity. Trends Cell Biol. (2020) 30:695-704. doi: 10.1016/j.tcb.2020.06.003

83. Abdul-Rahman T, Ghosh S, Badar SM, Nazir A, Bamigbade GB, Aji N, et al. The paradoxical role of cytokines and chemokines at the tumor microenvironment: a comprehensive review. *Eur J Med Res.* (2024) 29:124. doi: 10.1186/s40001-024-01711-z

84. Piper M, Hoen M, Darragh LB, Knitz MW, Nguyen D, Gadwa J, et al. Simultaneous targeting of PD-1 and IL-2R $\beta\gamma$  with radiation therapy inhibits pancreatic cancer growth and metastasis. *Cancer Cell.* (2023) 41:950–969.e6. doi: 10.1016/j.ccell.2023.04.001

85. Tichet M, Wullschleger S, Chryplewicz A, Fournier N, Marcone R, Kauzlaric A, et al. Bispecific PD1-IL2v and anti-PD-L1 break tumor immunity resistance by enhancing stem-like tumor-reactive CD8(+) T cells and reprogramming macrophages. *Immunity*. (2023) 56:162–179.e6. doi: 10.1016/j.immuni.2022.12.006

86. Wang CJ, Wang C, Han J, Wang YK, Tang L, Shen DW, et al. Effect of combined treatment with recombinant interleukin-2 and allicin on pancreatic cancer. *Mol Biol Rep.* (2013) 40:6579–85. doi: 10.1007/s11033-013-2766-1

87. Baldo BA. Side effects of cytokines approved for therapy. *Drug Saf.* (2014) 37:921–43. doi: 10.1007/s40264-014-0226-z

88. Harris F, Berdugo YA, Tree T. IL-2-based approaches to Treg enhancement. *Clin Exp Immunol.* (2023) 211:149–63. doi: 10.1093/cei/uxac105

89. Saloman JL, Tang G, Stello KM, Hall KE, Wang X, Alkaade S, et al. Serum biomarkers for chronic pancreatitis pain patterns. *Pancreatology.* (2021) 21:1411–8. doi: 10.1016/j.pan.2021.09.016

90. Lin E, Calvano SE, Lowry SF. Inflammatory cytokines and cell response in surgery. *Surgery*. (2000) 127:117–26. doi: 10.1067/msy.2000.101584

91. Hernandez R, Laporte KM, Hsiung S, Santos Savio A, Malek TR. High-dose IL-2/CD25 fusion protein amplifies vaccine-induced CD4(+) and CD8(+) neoantigenspecific T cells to promote antitumor immunity. *J Immunother Cancer*. (2021) 9(9): e002865. doi: 10.1136/jitc-2021-002865

92. Zhang YM, Ren HY, Zhao XL, Li J, Li JY, Wu FS, et al. Pharmacokinetics and pharmacodynamics of Da-Cheng-Qi decoction in the liver of rats with severe acute pancreatitis. *World J Gastroenterol.* (2017) 23:1367–74. doi: 10.3748/wjg.v23.i8.1367

93. Wang X, Cai H, Chen Z, Zhang Y, Wu M, Xu X, et al. Baicalein alleviates pyroptosis and inflammation in hyperlipidemic pancreatitis by inhibiting NLRP3/ Caspase-1 pathway through the miR-192-5p/TXNIP axis. *Int Immunopharmacol.* (2021) 101:108315. doi: 10.1016/j.intimp.2021.108315

94. Sun K, He SB, Qu JG, Dang SC, Chen JX, Gong AH, et al. IRF5 regulates lung macrophages M2 polarization during severe acute pancreatitis in *vitro*. *World J Gastroenterol*. (2016) 22:9368–77. doi: 10.3748/wjg.v22.i42.9368

95. Zhang C, Ge CL, Guo RX, He SG. Effect of IL-4 on altered expression of complement activation regulators in rat pancreatic cells during severe acute pancreatitis. *World J Gastroenterol.* (2005) 11:6770-4. doi: 10.3748/wjg.v11.i43.6770

96. Cook HT, Singh SJ, Wembridge DE, Smith J, Tam FW, Pusey CD. Interleukin-4 ameliorates crescentic glomerulonephritis in Wistar Kyoto rats. *Kidney Int.* (1999) 55:1319–26. doi: 10.1046/j.1523-1755.1999.00354.x

97. Hart PH, Vitti GF, Burgess DR, Whitty GA, Piccoli DS, Hamilton JA. Potential antiinflammatory effects of interleukin 4: suppression of human monocyte tumor necrosis factor alpha, interleukin 1, and prostaglandin E2. *Proc Natl Acad Sci U.S.A.* (1989) 86:3803–7. doi: 10.1073/pnas.86.10.3803

98. Vannier E, Miller LC, Dinarello CA. Coordinated antiinflammatory effects of interleukin 4: interleukin 4 suppresses interleukin 1 production but up-regulates gene expression and synthesis of interleukin 1 receptor antagonist. *Proc Natl Acad Sci U.S.A.* (1992) 89:4076–80. doi: 10.1073/pnas.89.9.4076

99. Katoh T, Lakkis FG, Makita N, Badr KF. Co-regulated expression of glomerular 12/15-lipoxygenase and interleukin-4 mRNAs in rat nephrotoxic nephritis. *Kidney Int.* (1994) 46:341–9. doi: 10.1038/ki.1994.280

100. Plugis NM, Weng N, Zhao Q, Palanski BA, Maecker HT, Habtezion A, et al. Interleukin 4 is inactivated via selective disulfide-bond reduction by extracellular thioredoxin. *Proc Natl Acad Sci U.S.A.* (2018) 115:8781-6. doi: 10.1073/ pnas.1805288115

101. Brock P, Sparmann G, Ritter T, Jaster R, Liebe S, Emmrich J. Adenovirusmediated gene transfer of interleukin-4 into pancreatic stellate cells promotes interleukin-10 expression. *J Cell Mol Med.* (2006) 10:884–95. doi: 10.1111/j.1582-4934.2006.tb00532.x

102. Piro G, Simionato F, Carbone C, Frizziero M, Malleo G, Zanini S, et al. A circulating T(H)2 cytokines profile predicts survival in patients with resectable pancreatic adenocarcinoma. *Oncoimmunology.* (2017) 6:e1322242. doi: 10.1080/2162402X.2017.1322242

103. Gocheva V, Wang HW, Gadea BB, Shree T, Hunter KE, Garfall AL, et al. IL-4 induces cathepsin protease activity in tumor-associated macrophages to promote cancer growth and invasion. *Genes Dev.* (2010) 24:241–55. doi: 10.1101/gad.1874010

104. Liu CY, Xu JY, Shi XY, Huang W, Ruan TY, Xie P, et al. M2-polarized tumorassociated macrophages promoted epithelial-mesenchymal transition in pancreatic cancer cells, partially through TLR4/IL-10 signaling pathway. *Lab Invest.* (2013) 93:844–54. doi: 10.1038/labinvest.2013.69

105. Prokopchuk O, Liu Y, Henne-Bruns D, Kornmann M. Interleukin-4 enhances proliferation of human pancreatic cancer cells: evidence for autocrine and paracrine actions. *Br J Cancer*. (2005) 92:921–8. doi: 10.1038/sj.bjc.6602416

106. Wu Y, Konaté MM, Lu J, Makhlouf H, Chuaqui R, Antony S, et al. IL-4 and IL-17A cooperatively promote hydrogen peroxide production, oxidative DNA damage, and upregulation of dual oxidase 2 in human colon and pancreatic cancer cells. *J Immunol.* (2019) 203:2532–44. doi: 10.4049/jimmunol.1800469

107. Meitzler JL, Konaté MM, Doroshow JH. Hydrogen peroxide-producing NADPH oxidases and the promotion of migratory phenotypes in cancer. *Arch Biochem Biophys.* (2019) 675:108076. doi: 10.1016/j.abb.2019.108076

108. Kornmann M, Maruyama H, Bergmann U, Tangvoranuntakul P, Beger HG, White MF, et al. Enhanced expression of the insulin receptor substrate-2 docking protein in human pancreatic cancer. *Cancer Res.* (1998) 58:4250–4.

109. Traub B, Sun L, Ma Y, Xu P, Lemke J, Paschke S, et al. Endogenously expressed IL-4R $\alpha$  Promotes the Malignant phenotype of human pancreatic cancer *in vitro* and *in vivo*. Int J Mol Sci. (2017) 18(4):716. doi: 10.3390/ijms18040716

110. Lin L, Hutzen B, Li PK, Ball S, Zuo M, Deangelis S, et al. A novel small molecule, LLL12, inhibits STAT3 phosphorylation and activities and exhibits potent growth-suppressive activity in human cancer cells. *Neoplasia*. (2010) 12:39–50. doi: 10.1593/neo.91196

111. Mohammed S, Sukumaran S, Bajgain P, Watanabe N, Heslop HE, Rooney CM, et al. Improving chimeric antigen receptor-modified T cell function by reversing the immunosuppressive tumor microenvironment of pancreatic cancer. *Mol Ther*. (2017) 25:249–58. doi: 10.1016/j.ymthe.2016.10.016

112. Zhou Y, Farooq MA, Ajmal I, He C, Gao Y, Guo D, et al. Co-expression of IL-4/ IL-15-based inverted cytokine receptor in CAR-T cells overcomes IL-4 signaling in immunosuppressive pancreatic tumor microenvironment. *BioMed Pharmacother*. (2023) 168:115740. doi: 10.1016/j.biopha.2023.115740

113. Wang Y, Jiang H, Luo H, Sun Y, Shi B, Sun R, et al. An IL-4/21 inverted cytokine receptor improving CAR-T cell potency in immunosuppressive solid-tumor microenvironment. *Front Immunol.* (2019) 10. doi: 10.3389/fimmu.2019.01691

114. Seifert M, Seifert G, Wolff-Vorbeck G, Langenmair E, Hopt UT, Wittel UA. Inducing a humoral immune response to pancreatic cancer antigen. *Cell Immunol.* (2016) 310:150–5. doi: 10.1016/j.cellimm.2016.09.002

115. Kornmann M, Kleeff J, Debinski W, Korc M. Pancreatic cancer cells express interleukin-13 and -4 receptors, and their growth is inhibited by Pseudomonas exotoxin coupled to interleukin-13 and -4. *Anticancer Res.* (1999) 19:125–31.

116. Kawakami K, Kawakami M, Husain SR, Puri RK. Targeting interleukin-4 receptors for effective pancreatic cancer therapy. *Cancer Res.* (2002) 62:3575–80.

117. Shimamura T, Royal RE, Kioi M, Nakajima A, Husain SR, Puri RK. Interleukin-4 cytotoxin therapy synergizes with gemcitabine in a mouse model of pancreatic ductal adenocarcinoma. Cancer Res. (2007) 67:9903-12. doi: 10.1158/0008-5472.CAN-06-4558

118. Yang L, Horibe T, Kohno M, Haramoto M, Ohara K, Puri RK, et al. Targeting interleukin-4 receptor  $\alpha$  with hybrid peptide for effective cancer therapy. *Mol Cancer Ther.* (2012) 11:235–43. doi: 10.1158/1535-7163.MCT-11-0363

119. Prokopchuk O, Steinacker JM, Nitsche U, Otto S, Bachmann J, Schubert EC, et al. IL-4 mRNA is downregulated in the liver of pancreatic cancer patients suffering from cachexia. *Nutr Cancer*. (2017) 69:84–91. doi: 10.1080/01635581.2017.1247885

120. Costamagna D, Duelen R, Penna F, Neumann D, Costelli P, Sampaolesi M. Interleukin-4 administration improves muscle function, adult myogenesis, and lifespan of colon carcinoma-bearing mice. *J Cachexia Sarcopenia Muscle*. (2020) 11:783–801. doi: 10.1002/jcsm.12539

121. Van Der Sijde F, Dik WA, Mustafa DAM, Vietsch EE, Besselink MG, Debets R, et al. Serum cytokine levels are associated with tumor progression during FOLFIRINOX chemotherapy and overall survival in pancreatic cancer patients. *Front Immunol.* (2022) 13:898498. doi: 10.3389/fimmu.2022.898498

122. Robinson SM, Rasch S, Beer S, Valantiene I, Mickevicius A, Schlaipfer E, et al. Systemic inflammation contributes to impairment of quality of life in chronic pancreatitis. *Sci Rep.* (2019) 9:7318. doi: 10.1038/s41598-019-43846-8

123. Vignali D, Monti P. Targeting homeostatic T cell proliferation to control betacell autoimmunity. *Curr Diabetes Rep.* (2016) 16:40. doi: 10.1007/s11892-016-0731-9

124. Pothlichet J, Meola A, Bugault F, Jeammet L, Savitt AG, Ghebrehiwet B, et al. Microbial protein binding to gC1qR drives PLA2G1B-induced CD4 T-cell anergy. *Front Immunol.* (2022) 13:824746. doi: 10.3389/fimmu.2022.824746

125. Jang SI, Lee HK, Chang EJ, Kim S, Kim SY, Hong IY, et al. Improved predictability of pancreatic ductal adenocarcinoma diagnosis using a blood immune cell biomarker panel developed from bulk mRNA sequencing and single-cell RNA-sequencing. *Cancer Immunol Immunother*. (2023) 72:2757–68. doi: 10.1007/s00262-023-03458-8

126. Heo SH, Jang SI, Kim SY, Choi B, Lee DK, Lee HK, et al. Characterization of circulating IL-7R positive cell populations for early detection of pancreatic ductal adenocarcinoma. *J Clin Med.* (2021) 10(18):4157. doi: 10.3390/jcm10184157

127. Ghassem-Zadeh S, Gaida MM, Szanyi S, Acha-Orbea H, Frossard JL, Hinz U, et al. Distinct pathophysiological cytokine profiles for discrimination between autoimmune pancreatitis, chronic pancreatitis, and pancreatic ductal adenocarcinoma. *J Transl Med.* (2017) 15:126. doi: 10.1186/s12967-017-1227-3

128. Meyer LK, Huang BJ, Delgado-Martin C, Roy RP, Hechmer A, Wandler AM, et al. Glucocorticoids paradoxically facilitate steroid resistance in T cell acute lymphoblastic leukemias and thymocytes. *J Clin Invest.* (2020) 130:863–76. doi: 10.1172/JCI130189

129. Zhang Y, Li B, Bai Q, Wang P, Wei G, Li Z, et al. The lncRNA Snhg1-Vps13D vesicle trafficking system promotes memory CD8 T cell establishment via regulating the dual effects of IL-7 signaling. *Signal Transduct Target Ther*. (2021) 6:126. doi: 10.1038/ s41392-021-00492-9

130. Wang C, Kong L, Kim S, Lee S, Oh S, Jo S, et al. The role of IL-7 and IL-7R in cancer pathophysiology and immunotherapy. *Int J Mol Sci.* (2022) 23(18):10412. doi: 10.3390/ijms231810412

131. Fu D, Yan J, Zhang Z, Liu Y, Ma X, Ding J, et al. Nuclear PLD1 combined with NPM1 induces gemcitabine resistance through tumorigenic IL7R in pancreatic adenocarcinoma. *Cancer Biol Med.* (2023) 20:599–626. doi: 10.20892/j.issn.2095-3941.2023.0039

132. Landires I, Bugault F, Lambotte O, De Truchis P, Slama L, Danckaert A, et al. HIV infection perturbs interleukin-7 signaling at the step of STAT5 nuclear relocalization. *Aids.* (2011) 25:1843–53. doi: 10.1097/QAD.0b013e32834a3678

133. Pothlichet J, Rose T, Bugault F, Jeammet L, Meola A, Haouz A, et al. PLA2G1B is involved in CD4 anergy and CD4 lymphopenia in HIV-infected patients. *J Clin Invest.* (2020) 130:2872–87. doi: 10.1172/JCI131842

134. Schmaler M, Broggi MA, Lagarde N, Stöcklin BF, King CG, Finke D, et al. IL-7R signaling in regulatory T cells maintains peripheral and allograft tolerance in mice. *Proc Natl Acad Sci U.S.A.* (2015) 112(43):13330–5. doi: 10.1073/pnas.1510045112

135. Pang N, Shi J, Qin L, Chen A, Tang Y, Yang H, et al. IL-7 and CCL19-secreting CAR-T cell therapy for tumors with positive glypican-3 or mesothelin. *J Hematol Oncol.* (2021) 14:118. doi: 10.1186/s13045-021-01128-9

136. Li D, Liang T, Hutchins LE, Wolfarth AA, Ferrando-Martinez S, Lee BH, et al. rhIL-7-hyFc, a long-acting interleukin-7, improves efficacy of CAR-T cell therapy in solid tumors. *J Immunother Cancer.* (2024) 12(7):1–15. doi: 10.1136/jitc-2024-008989

137. Xiao J, Feng X, Huang XY, Huang Z, Huang Y, Li C, et al. Spautin-1 ameliorates acute pancreatitis via inhibiting impaired autophagy and alleviating calcium overload. *Mol Med.* (2016) 22:643–52. doi: 10.2119/molmed.2016.00034

138. Xiao J, Huang K, Lin H, Xia Z, Zhang J, Li D, et al. Mogroside II(E) inhibits digestive enzymes via suppression of interleukin 9/interleukin 9 receptor signalling in acute pancreatitis. *Front Pharmacol.* (2020) 11:859. doi: 10.3389/fphar.2020.00859

139. Hu B, Qiu-Lan H, Lei RE, Shi C, Jiang HX, Qin SY. Interleukin-9 Promotes Pancreatic Cancer Cells Proliferation and Migration via the miR-200a/Beta-Catenin Axis. *BioMed Res Int.* (2017) 2017:2831056. doi: 10.1155/2017/2831056

140. Lu D, Qin Q, Lei R, Hu B, Qin S. Targeted blockade of interleukin 9 inhibits tumor growth in murine model of pancreatic cancer. *Adv Clin Exp Med.* (2019) 28:1285–92. doi: 10.17219/acem/104543

141. You FP, Zhang J, Cui T, Zhu R, Lv CQ, Tang HT, et al. Th9 cells promote antitumor immunity via IL-9 and IL-21 and demonstrate atypical cytokine expression in breast cancer. *Int Immunopharmacol.* (2017) 52:163–7. doi: 10.1016/j.intimp.2017.08.031

142. Ye ZJ, Zhou Q, Yin W, Yuan ML, Yang WB, Xiong XZ, et al. Differentiation and immune regulation of IL-9-producing CD4+ T cells in Malignant pleural effusion. *Am J Respir Crit Care Med.* (2012) 186:1168–79. doi: 10.1164/rccm.201207-1307OC

143. Ahmed A, Klotz R, Köhler S, Giese N, Hackert T, Springfeld C, et al. Immune features of the peritumoral stroma in pancreatic ductal adenocarcinoma. *Front Immunol.* (2022) 13:947407. doi: 10.3389/fimmu.2022.947407

144. Chen T, Guo J, Cai Z, Li B, Sun L, Shen Y, et al. Th9 cell differentiation and its dual effects in tumor development. *Front Immunol.* (2020) 11:1026. doi: 10.3389/ fmmu.2020.01026

145. Li H, Nourbakhsh B, Cullimore M, Zhang GX, Rostami A. IL-9 is important for T-cell activation and differentiation in autoimmune inflammation of the central nervous system. *Eur J Immunol.* (2011) 41:2197–206. doi: 10.1002/eji.201041125

146. Kalbasi A, Siurala M, Su LL, Tariveranmoshabad M, Picton LK, Ravikumar P, et al. Potentiating adoptive cell therapy using synthetic IL-9 receptors. *Nature*. (2022) 607:360–5. doi: 10.1038/s41586-022-04801-2

147. Ueda T, Takeyama Y, Yasuda T, Shinzeki M, Sawa H, Nakajima T, et al. Serum interleukin-15 level is a useful predictor of the complications and mortality in severe acute pancreatitis. *Surgery*. (2007) 142:319–26. doi: 10.1016/j.surg.2007.05.002

148. Saito M, Inoue S, Yamashita K, Kakeji Y, Fukumoto T, Kotani J. IL-15 improves aging-induced persistent T cell exhaustion in mouse models of repeated sepsis. *Shock*. (2020) 53:228–35. doi: 10.1097/SHK.00000000001352

149. Manohar M, Kandikattu HK, Verma AK, Mishra A. IL-15 regulates fibrosis and inflammation in a mouse model of chronic pancreatitis. *Am J Physiology-Gastrointestinal Liver Physiol.* (2018) 315:G954–65. doi: 10.1152/ajpgi.00139.2018

150. Kandikattu HK, Manohar M, Venkateshaiah SU, Yadavalli C, Mishra A. Chronic inflammation promotes epithelial-mesenchymal transition-mediated Malignant phenotypes and lung injury in experimentally-induced pancreatitis. *Life Sci.* (2021) 278:119640. doi: 10.1016/j.lfs.2021.119640

151. Kruger D, Lahoud N, Yako YY, Devar J, Smith M. Pancreatic ductal adenocarcinoma: Prognostic indicators of advanced disease. *PloS One.* (2022) 17: e0262439. doi: 10.1371/journal.pone.0262439

152. Waldmann TA. Cytokines in cancer immunotherapy. Cold Spring Harbor Perspect Biol. (2018) 10. doi: 10.1101/cshperspect.a028472

153. Waldmann TA, Miljkovic MD, Conlon KC. Regulation of Lymphoid Homeostasis: Implications for Therapy of Autoimmunity and Cancer. J Exp Med. (2020) 217(1). doi: 10.1084/jem.20191062

154. Shi I, Wen J, Zhang W, Meng FD, Wan Y, Wang L, et al. Combination therapy with interleukin-15 and metformin as a synergistic treatment for pancreatic cancer. *Eur Rev Med Pharmacol Sci.* (2021) 25(16):5075–89. doi: 10.26355/eurrev\_202108\_26519

155. Van Audenaerde JR, Marcq E, Von Scheidt B, Davey AS, Oliver AJ, De Waele J, et al. Novel combination immunotherapy for pancreatic cancer: potent anti-tumor effects with CD40 agonist and interleukin-15 treatment. *Clin Transl Immunol.* (2020) 9: e1165. doi: 10.1002/cti2.1165

156. Knudson KM, Hodge JW, Schlom J, Gameiro SR. Rationale for IL-15 superagonists in cancer immunotherapy. *Expert Opin Biol Ther.* (2020) 20:705–9. doi: 10.1080/14712598.2020.1738379

157. Hsu FT, Tsai CL, Chiang IT, Lan KH, Yueh PF, Liang WY, et al. Synergistic effect of Abraxane that combines human IL15 fused with an albumin-binding domain

on murine models of pancreatic ductal adenocarcinoma. J Cell Mol Med. (2022) 26:1955–68. doi: 10.1111/jcmm.17220

158. Nelson A, Gebremeskel S, Lichty BD, Johnston B. Natural killer T cell immunotherapy combined with IL-15-expressing oncolytic virotherapy and PD-1 blockade mediates pancreatic tumor regression. *J Immunother Cancer*. (2022) 10(3): e003923. doi: 10.1136/jitc-2021-003923

159. Zhao R, Xiao Q, Wu Y, Zhang W, Liu J, Zeng Y, et al. Dual-crosslinking immunostimulatory hydrogel synchronously suppresses pancreatic fistula and pancreatic cancer relapse post-resection. *Biomaterials.* (2024) 305:122453. doi: 10.1016/j.biomaterials.2023.122453

160. Van Den Eynde A, Gehrcken L, Verhezen T, Lau HW, Hermans C, Lambrechts H, et al. IL-15-secreting CAR natural killer cells directed toward the pan-cancer target CD70 eliminate both cancer cells and cancer-associated fibroblasts. *J Hematol Oncol.* (2024) 17:8. doi: 10.1186/s13045-024-01525-w

161. Shah Z, Tian L, Li Z, Jin L, Zhang J, Li Z, et al. Human anti-PSCA CAR macrophages possess potent antitumor activity against pancreatic cancer. *Cell Stem Cell.* (2024) 31:803–817.e6. doi: 10.1016/j.stem.2024.03.018

162. Teng KY, Mansour AG, Zhu Z, Li Z, Tian L, Ma S, et al. Off-the-shelf prostate stem cell antigen-directed chimeric antigen receptor natural killer cell therapy to treat pancreatic cancer. *Gastroenterology*. (2022) 162:1319–33. doi: 10.1053/j.gastro.2021.12.281

163. Xu DL, He YQ, Xiao B, Si Y, Shi J, Liu XA, et al. A novel sushi-IL15-PD1 CAR-NK92 cell line with enhanced and PD-L1 targeted cytotoxicity against pancreatic cancer cells. *Front Oncol.* (2022) 12:726985. doi: 10.3389/fonc.2022.726985

164. Kurz E, Hirsch CA, Dalton T, Shadaloey SA, Khodadadi-Jamayran A, Miller G, et al. Exercise-induced engagement of the IL-15/IL-15R $\alpha$  axis promotes anti-tumor immunity in pancreatic cancer. *Cancer Cell.* (2022) 40:720-+. doi: 10.1016/j.ccell.2022.05.006

165. Pita-Grisanti V, Vélez-Bonet E, Chasser K, Hurst Z, Liette A, Vulic G, et al. Physical activity decreases inflammation and delays development of obesity-associated pancreatic ductal adenocarcinoma. *Cancer Res.* (2024) 84(18):3058–71. doi: 10.1158/0008-5472.c.7451282.v1

166. Ugai S, Shimozato O, Yu L, Wang YQ, Kawamura K, Yamamoto H, et al. Transduction of the IL-21 and IL-23 genes in human pancreatic carcinoma cells produces natural killer cell-dependent and -independent antitumor effects. *Cancer Gene Ther.* (2003) 10:771–8. doi: 10.1038/sj.cgt.7700630

167. Lin X, Ye L, Wang X, Liao Z, Dong J, Yang Y, et al. Follicular helper T cells remodel the immune microenvironment of pancreatic cancer via secreting CXCL13 and IL-21. *Cancers (Basel).* (2021) 13(15):3678. doi: 10.3390/cancers13153678

168. Lin X, Kang K, Chen P, Zeng Z, Li G, Xiong W, et al. Regulatory mechanisms of PD-1/PD-L1 in cancers. *Mol Cancer.* (2024) 23:108. doi: 10.1186/s12943-024-02023-w

169. Mcmichael EL, Jaime-Ramirez AC, Guenterberg KD, Luedke E, Atwal LS, Campbell AR, et al. IL-21 enhances natural killer cell response to cetuximab-coated pancreatic tumor cells. *Clin Cancer Res.* (2017) 23:489–502. doi: 10.1158/1078-0432.CCR-16-0004

170. Marelli G, Chard Dunmall LS, Yuan M, Di Gioia C, Miao J, Cheng Z, et al. A systemically deliverable Vaccinia virus with increased capacity for intertumoral and intratumoral spread effectively treats pancreatic cancer. *J Immunother Cancer*. (2021) 9 (1):e001624. doi: 10.1136/jitc-2020-001624

171. Linnebacher A, Mayer P, Marnet N, Bergmann F, Herpel E, Revia S, et al. Interleukin 21 receptor/ligand interaction is linked to disease progression in pancreatic cancer. *Cells.* (2019) 8(19):1104. doi: 10.3390/cells8091104