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Single-cell and transcriptome analyses revealed CTHRC1 a potential therapeutic target mediating invasion and tumor microenvironment in TNBC: experimental validation

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Background: Investigating the pivotal role of CTHRC1 in the tumor microenvironment of triple-negative breast cancer (TNBC).

Method: The RNA transcriptomic data obtained from the Cancer Genome Atlas and single-cell sequencing data from TNBC in Gene Expression Omnibus (GEO) were acquired and subjected to analysis. A comprehensive investigation was conducted with a specific focus on characterizing CTHRC1 in TNBC and its correlation with invasive genes. Furthermore, additional analyses were performed to explore the relationship between CTHRC1, tumor immune cell infiltration, and immunotherapy in TNBC. The expression of CTHRC1 in the tumor microenvironment, cellular differentiation, and cellular communication was systematically analyzed using single-cell data from TNBC.

Result: The expression of CTHRC1 in patients with TNBC gradually increases concomitantly with the progression of tumor T-stage and N-stage. Simultaneously, there is a concurrent increase in the expression of most invasive gene sets. Furthermore, there is a significant augmentation in both infiltration abundance and activity of M2-type macrophages associated with elevated levels of CTHRC1 expression. Single-cell data reveal an upregulated expression of the invasive gene set in CTHRC1-positive cancer associated fibroblasts (CAFs), thereby modulating their interaction with M2-type macrophages. Multiple immunofluorescence analyses confirmed that CTHRC1 modulates immune cell infiltration and tumor cell invasion through the mediation of CAFs.

Conclusion: CTHRC1 was a molecule that exhibits characteristic expression in TNBC. CTHRC1 positive CAFs exert regulatory effects within the immunosuppressive microenvironment of TNBC by modulating M2-type macrophages.

KEYWORDS

TNBC, invasion, CAFs, tumor microenvironment, M2-type macrophages

Background

Breast cancer ranks as the second leading cause of cancer-related fatalities and represents the most prevalent malignancy among women (1). Breast cancer can be categorized into three primary subtypes based on the expression levels of estrogen receptor (ER), progesterone receptor (PR), and human epidermal growth factor receptor 2 (HER2): hormone receptor-positive, HER2-positive, and triple-negative breast cancer (TNBC) (2). Patients diagnosed with triple-negative breast cancer exhibit a considerably elevated 5-year recurrence rate and demonstrate the most unfavorable prognosis, compared to the other two pathological subtypes (3). Triple-negative breast cancer constitutes approximately 15 percent of breast cancer cases, and its distinct molecular expression profile renders patients unresponsive to endocrine and targeted therapies (4). Surgery and chemotherapy are still the mainstream of treatments (5).

Single-cell and bulk transcriptomic data offer significant advantages in the field of oncology: 1) Enabling the analysis of gene expression at the single-cell level, thereby uncovering intercellular heterogeneity. 2) Facilitating precise identification and classification of diverse cell types and states, which aids in the discovery of novel cellular subpopulations. 3) Revealing spatial heterogeneity within tissues or tumors. 4) Providing a high degree of data standardization, which enhances cross-study comparisons and integrations.

The extracellular matrix (ECM) protein CTHRC1 engages in intricate interactions with diverse intracellular and extracellular matrices through distinct secretory mechanisms (6). CTHRC1 exhibits robust expression in various tumor types and facilitates cancer cell proliferation, invasion, and metastasis (7, 8). Additionally, it has been documented that CTHRC1 plays a role in the regulation of macrophage recruitment (9, 10).

This study investigates the distinct expression profile of CTHRC1 in TNBC, elucidating its association with invasion-related genes. Leveraging single-cell data from triple-negative breast cancer, we comprehensively explore the role of CTHRC1 within the tumor microenvironment, with a specific focus on immunoregulation and intercellular interactions.

Abbreviations: TCGA, The Cancer Genome Atlas; CAFs, Cancer associated fibroblasts; GSEA, Gene Set Enrichment Analysis; UMAP, Unified Metamorphic Approximation and Project; GSVA, Gene set variation analysis.

Method

This study incorporates transcriptomic data obtained from 33 tumor tissues, which were log2 transformed, downloaded from The Cancer Genome Atlas (TCGA). The Gene Expression Omnibus (GEO) database encompasses transcriptomic data derived from tumor tissues as well as single-cell sequencing data. Transcriptomic profiles of TNBC patients were retrieved from GSE58812, GSE164458, and GSE157284 (11–13). Additionally, the study incorporated single-cell transcriptome data obtained from 9 patients with TNBC through the GSE176078 dataset (14). CancerSea has acquired a compilation of 14 genes associated with the biological characteristics of tumors from <http://biocc.hrbmu.edu.cn/CancerSEA/> (Supplementary Table 1).

Pan-cancer analysis

We have generated transcriptome expression profiles for 33 tumor tissues, excluding those lacking information on paracancerous tissues. By analyzing the data from both tumor and paracancerous tissues, we initially compared the alterations in CTHRC1 expression levels. Additionally, we extracted the expression patterns of the invasion gene set across various tumor tissues. Subsequently, we conducted a correlation analysis between CTHRC1 expression in different tumors and the invasion gene set. Finally, we assessed the association between CTHRC1 expression in diverse tumors and gene expression of immunosuppressant therapeutic targets.

Clinical characterization of CTHRC1 in TNBC

The distinct characteristics of CTHRC1 in TNBC and non-TNBC were comprehensively elucidated. Breast cancer samples were classified into TNBC and non-TNBC subtypes, followed by an initial analysis of the association between CTHRC1 expression and tumor T-stage. The expression of CTHRC1 was stratified into high and low groups based on the median value. We examined the differential expression of the invasive gene set in conjunction with the trend in CTHRC1 expression. Furthermore, we corroborated this trend by analyzing expression data from multiple GEO databases for patients with TNBC.

Immune cell infiltration and immunotherapeutic features

Immunotherapy is a pivotal therapeutic modality for tumor patients, and thus we investigated the correlation between CTHRC1 expression and immune cell infiltration in TNBC patients (15). The abundance and activity of immune cell infiltration in triple-negative breasts were comprehensively analyzed using advanced algorithms including CIBERSORT, ssGSEA, and ESTIMATE. TIDE has emerged as a widely adopted tool for predicting immunotherapy response in tumor patients (16, 17). Moreover, we conducted predictive analyses to assess the potential efficacy of targeting CTHRC1 in patient immunotherapy.

Single-cell data processing

The dataset GSE176078 consists of transcriptomic data obtained from single cells of nine patients diagnosed with TNBC. Initially, the data were processed individually to create Seurat objects. Rigorous quality control measures were implemented on individual samples to exclude those exhibiting (1) mitochondrial gene expression exceeding 10%, (2) bicellular contamination, and (3) erythrocyte gene expression surpassing 3%. Furthermore, cell cycle effects were accounted for and eliminated from the analysis. Subsequently, the harmony function was employed to integrate the datasets, followed by dimensionality reduction using Unified Mobility Approximation and Project (UMAP), enabling visualization of clustered cells in a two-dimensional map. Cell types were identified utilizing “SingleR” along with relevant high-quality literature references (18–21). Finally, specific cell types of interest underwent further dimensionality reduction clustering and comprehensive annotation at the subpopulation level.

GSVA enrichment analysis

GSVA enrichment analysis is widely used to assess the activity scores of gene sets in samples. We evaluated the correlation between 14 gene sets related to cell biology and CTHRC1. Firstly, we selected the expression matrix of TNBC and used the GSVA algorithm to calculate the scores of the 14 gene sets in the samples. Finally, we analyzed the correlation between CTHRC1 and their activity levels. In single-cell data, we annotated detailed cell types present in the tumor microenvironment of triple-negative breast cancer. Initially, we extracted “RNA” data from single-cell sequencing results and then assessed the activity scores of these gene sets in different cellular subsets using GSVA method.

Analysis of cell differentiation trajectories

The Monocle2 method utilizes unsupervised learning techniques on the transcriptome expression matrix of individual cells to categorize them into separate branches of the developmental path. Initially, we investigated the dynamics of CTHRC1 during cell

differentiation, and subsequently employed Plot pseudotime heatmap to visualize gene expression patterns within the feature gene set across cellular evolution.

Analysis of intercellular communication

The communication between cells is achieved through the interaction of ligands and receptors. Within the tumor microenvironment, intercellular communication plays a pivotal role in facilitating immune evasion by tumor cells. CellChat utilizes gene expression levels to infer the pathways and magnitude of intercellular communication, while also characterizing the interactions between ligands and receptors. Overexpressed genes were identified, screened for ligand-receptor pairs, and mapped onto the protein-protein interaction network. To ensure high-quality data, a filter was applied excluding communication relationships involving low-quality cells with a minimum cell count of 3.

Multiple immunofluorescence assays

Based on bioinformatics discoveries, we corroborated our findings using multiple methodologies. Multiple immunofluorescence assays were utilized to authenticate the interactions between distinct molecules and cells.

Statistical analysis

The associations between variables were assessed using Pearson or Spearman coefficients. T-tests were utilized to compare continuous variables with a normal distribution, whereas the Mann-Whitney U test was employed for variables that did not follow a normal distribution. Categorical variables were compared using either the chi-square test or Fisher exact test. All data analysis was performed using R software (version 4.1.2, available at <https://www.r-project.org/>). Statistical significance was determined based on a two-sided P-value threshold of less than 0.05 (* $P < 0.05$; ** $P < 0.01$; *** $P < 0.001$; **** $P < 0.0001$).

Result

CTHRC1 showed unique features in TNBC

Significantly elevated expression levels of CTHRC1 were observed in breast, esophageal, and liver cancers compared to corresponding paraneoplastic tissues. Notably, breast cancer exhibited particularly high levels of CTHRC1 (Figure 1A, B). In non-TNBC tissues, the difference in CTHRC1 expression was not statistically significant. However, CTHRC1 expression was markedly elevated in TNBC tissues. Additionally, as the tumor T-stage and N-stage advanced, the expression of CTHRC1 exhibited an upward trend (Figure 1C–G). CTHRC1 was closely linked to the biology of TNBC.

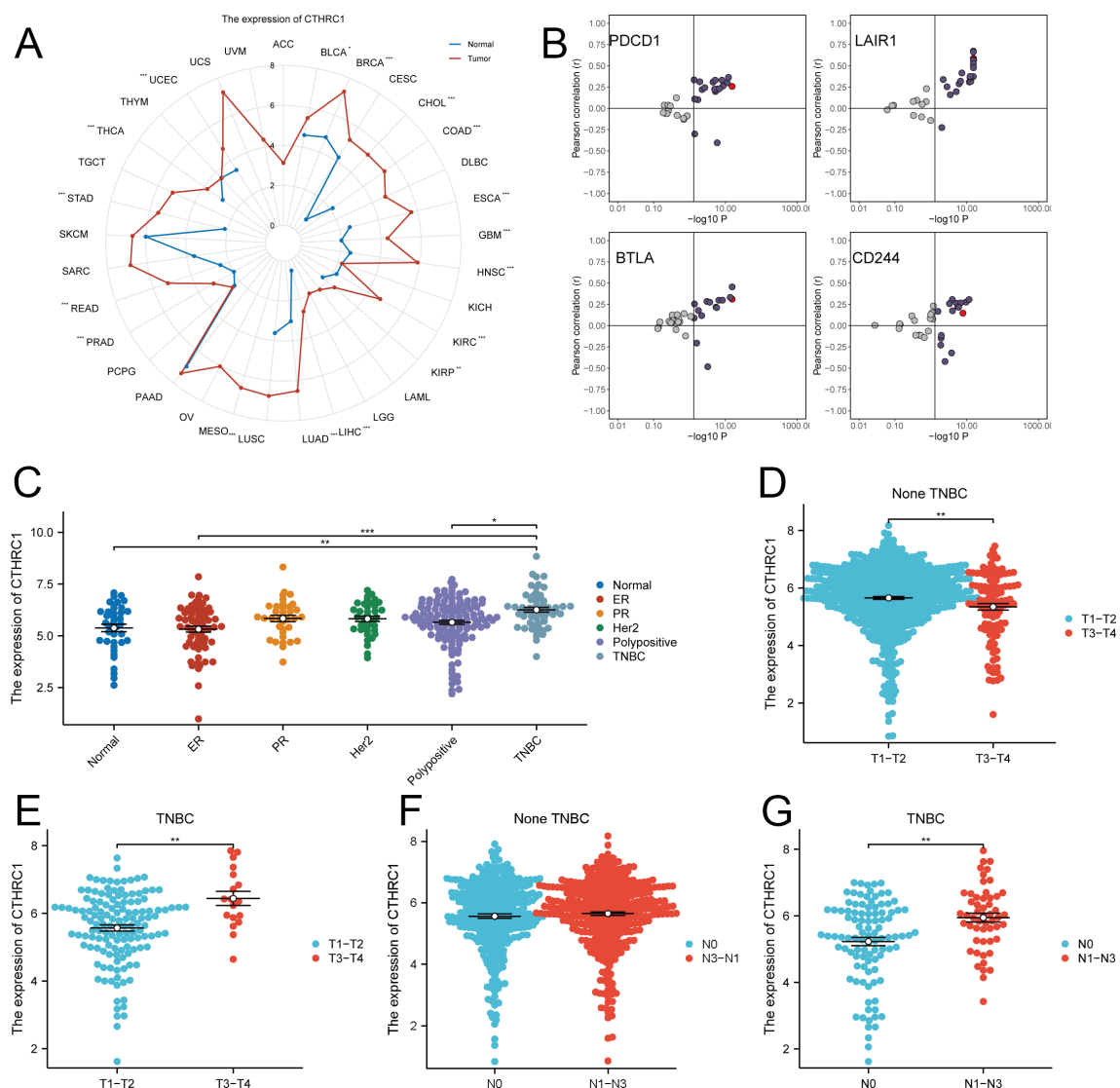


FIGURE 1

Expression analysis of CTHRC1 in multiple tumors. (A) CTHRC1 expression was evaluated in the majority of tumors. (B) Relationship between CTHRC1 and immune checkpoint inhibitory gene expression. (C) CTHRC1 expression was higher in TNBC than in other tissue types. (D-G) The expression of CTHRC1 was upregulated in triple-negative breast cancer with increasing tumor T-stage, N-stage, while no significant elevation was observed in non-triple-negative breast cancer. *P < 0.05; **P < 0.01; ***P < 0.001.

CTHRC1 was highly positively correlated with the invasion gene set

For multiple samples, we utilized the GSVA enrichment score to evaluate the invasive gene set of tumors. The results demonstrated a strong positive correlation between CTHRC1 expression and tumor invasion scores specifically in TNBC. Conversely, no significant correlation was observed between CTHRC1 expression and tumor invasion scores in non-TNBC tissues. Conversely, there was no similar pattern found in non-TNBC. After conducting an in-depth analysis, we proceeded to investigate whether the invasive gene set displayed similar characteristics in both TNBC and non-TNBC. Intriguingly, the expression of a majority of genes within the

invasive gene set exhibited an upward trend with elevated CTHRC1 gene expression specifically in the TNBC cohort (Figure 2). CTHRC1 was served as a uniquely distinctive biological molecule in TNBC relative to non-TNBC.

CTHRC1 primarily affected macrophage infiltration

In TNBC, CTHRC1 exhibited limited association with most immune cells. However, there was a significant difference in the abundance of macrophage infiltration between the CTHRC1 high-expression and low-expression groups (Figure 3A). The infiltration

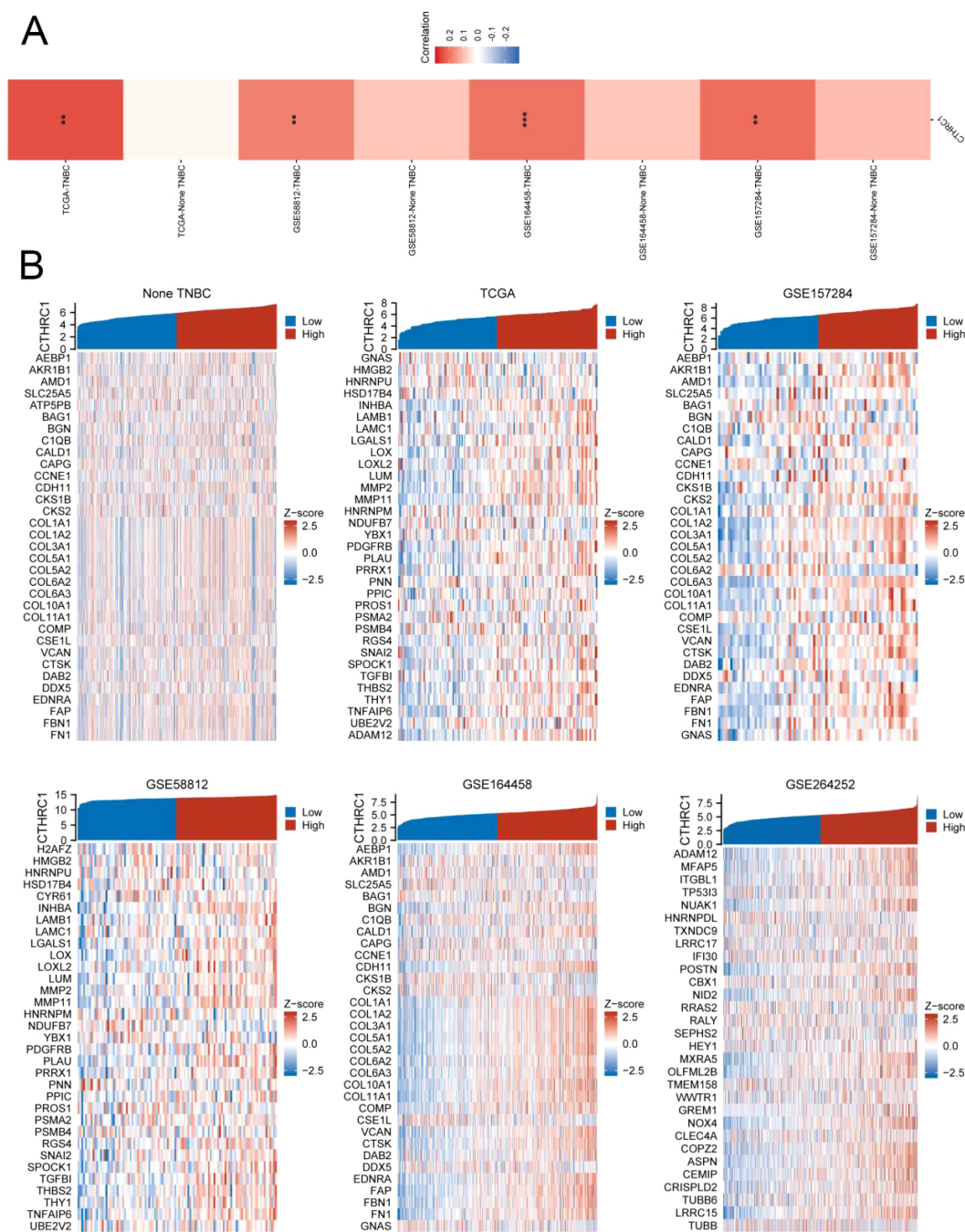
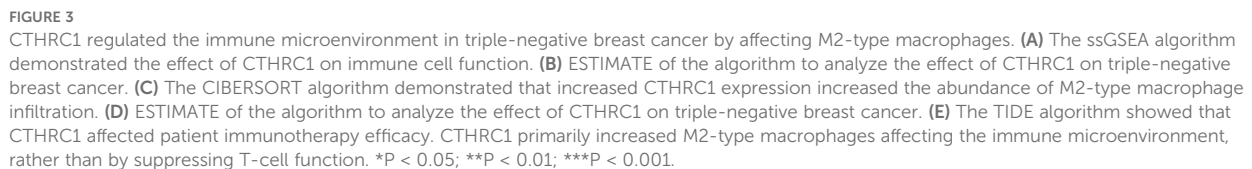


FIGURE 2

CTHRC1 showed a highly positive correlation with the tumour invasion gene set in TNBC. **(A)** CTHRC1 exhibited a significantly higher GSEA enrichment score in the tumour invasion gene set in TNBC tissues compared to non-TNBC tissues. **(B)** Invasive genes were upregulated concomitantly with increased CTHRC1 expression in triple-negative breast cancer, whereas no such correlation was observed in non-triple-negative breast cancer. ** $P < 0.01$; *** $P < 0.001$.

abundance of M2-type macrophages significantly increased with higher levels of CTHRC1 expression (Figure 3C). The ESTIMATE algorithm indicated no disparities in immune scores but revealed discrepancies in mesenchymal scores between the high and low expressing groups of CTHRC1 (Figure 3B, D). TIDE predicted that patients in the high-expression group of CTHRC1 would derive less

benefit from immunotherapy compared to those with lower expression levels. Similarly, while there were no variations observed in T-cell escape between the high and low expressing groups of CTHRC1, a difference was noted in M2-type macrophage scores (Figure 3E). CTHRC1 played a crucial role in the distinct tumour microenvironment of TNBC.



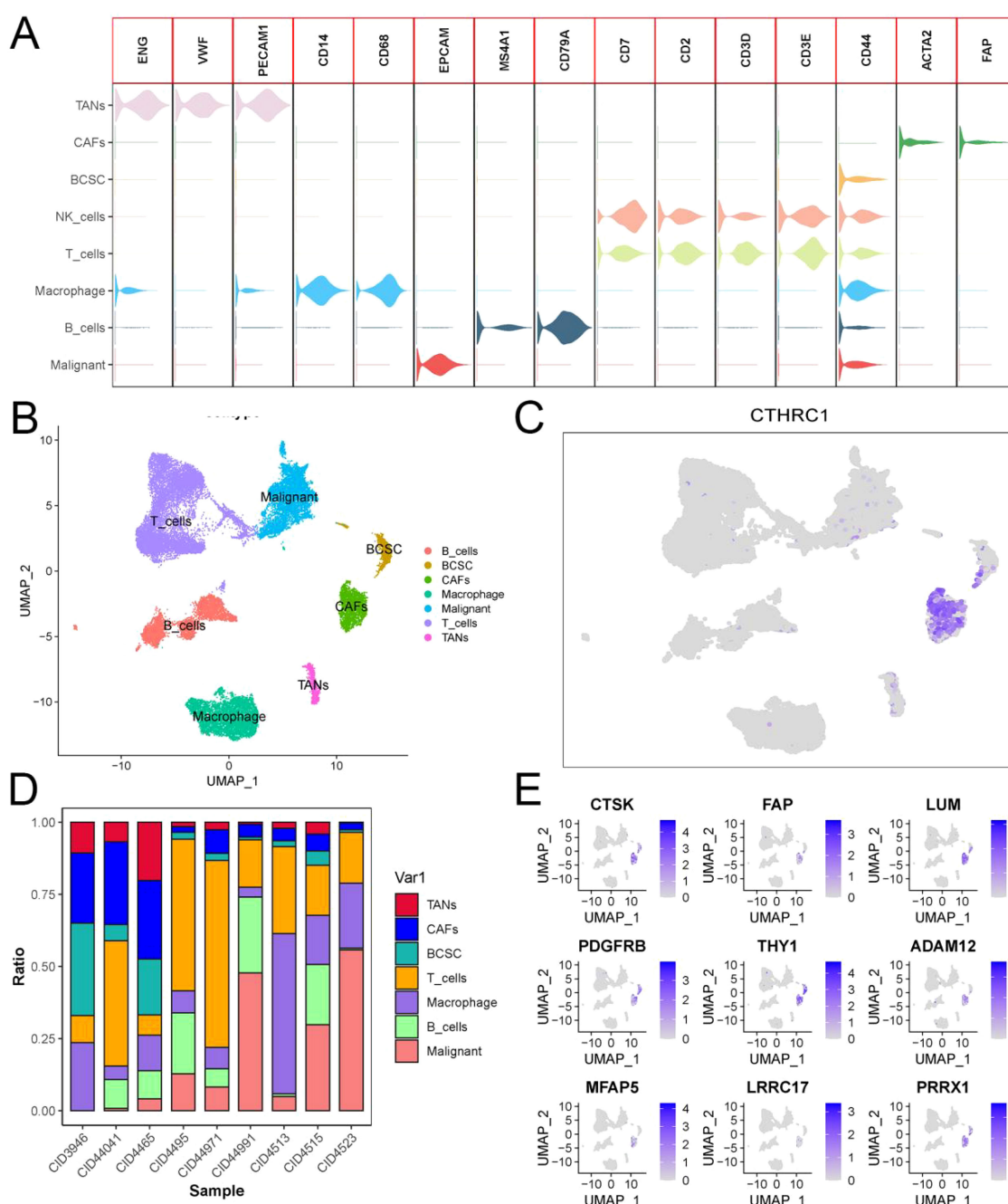


FIGURE 4

Analysis of single-cell sequencing data in triple-negative breast cancer. (A, B) Cluster cell annotation based on SingleR and previous literature. (C) CTHRC1 expression patterns in triple-negative breast cancer. (D) Plot of the proportion of cell types in single-cell sequencing samples. (E) Expression patterns of invasive genes in triple-negative breast cancer.

were classified into five distinct differentiation trajectories, wherein the expression of CTHRC1 exhibited a significant upregulation concomitant with CAFs differentiation (Figure 6D, E). This augmented expression of CTHRC1 was accompanied by a notable increase in the expression levels of most members within the invasive gene sets (Figure 6F). Alterations in CTHRC1 expression were associated with modifications in the invasive activity of TNBC.

CTHRC1 affected communication between CAFs and macrophages

Receptor-ligand interactions serve as the foundation of intercellular communication. Utilizing the 'Cellchat' function for single-cell data analysis, potential signaling between cells was speculated upon (Figures 7A, B). CTHRC1 modulates communication between

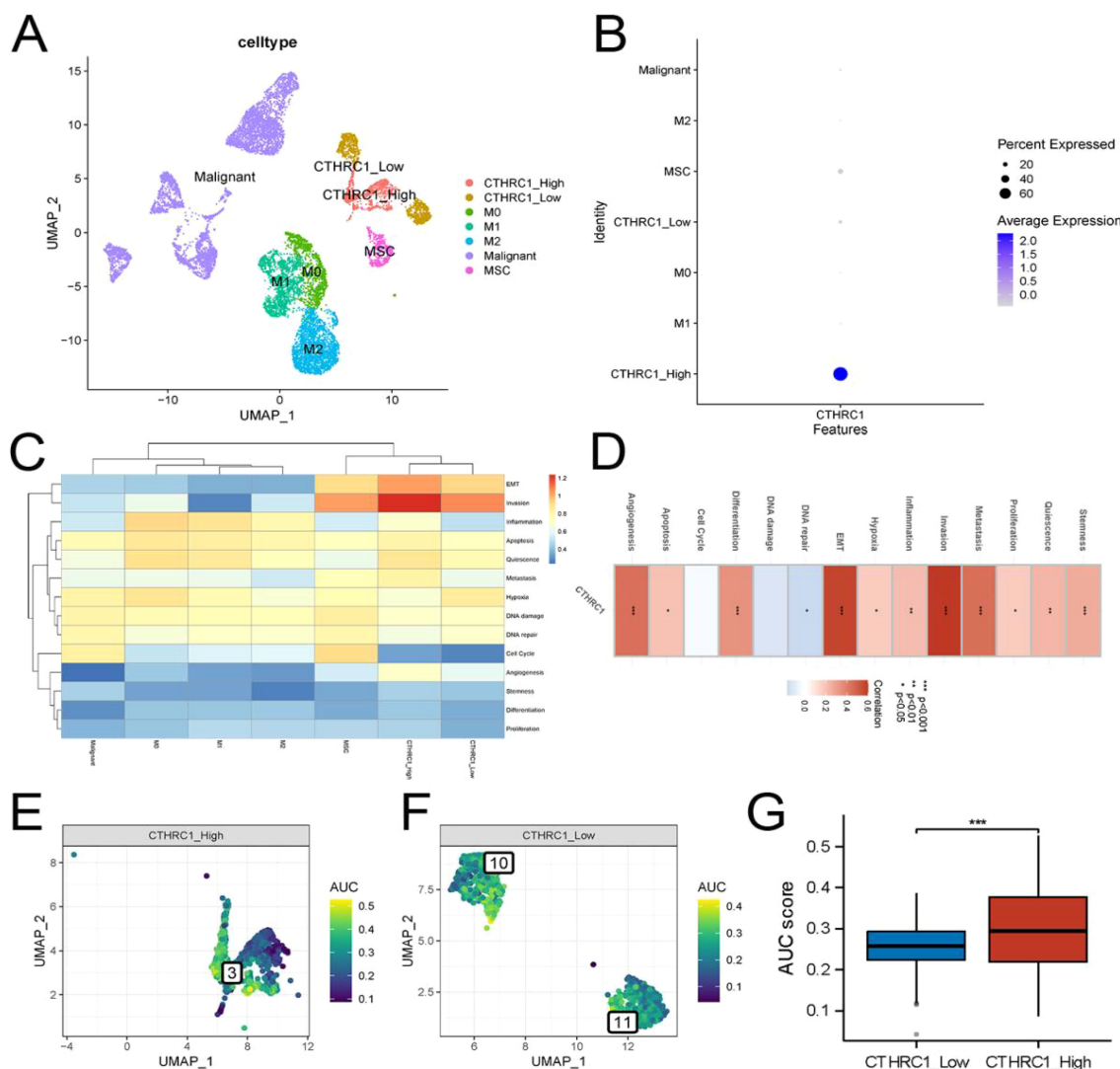


FIGURE 5

Invasive activity was strongest in CTHRC1-positive tumor-associated fibroblasts. (A, B) Re-clustering and annotation of tumor-associated fibroblasts and macrophages. (C) GSEA analysis of single-cell data demonstrated the strongest invasive activity of CTHRC1-positive fibroblasts. (D) Transcriptomic data revealed a strong positive correlation between CTHRC1 and invasive activity. (E-G) AUCCell score exhibited higher invasive activity of CTHRC1-positive cells than CTHRC1-negative cells. * $P < 0.05$; ** $P < 0.01$; *** $P < 0.001$.

tumor-associated fibroblasts and macrophages. The group with high levels of CTHRC1 demonstrated increased functionality in the complement signaling pathway and annexin signaling pathway when compared to the group with low levels of CTHRC1. Conversely, a contrasting phenomenon was observed in the PTN signaling pathway, PROS signaling pathway, ANGPTL signaling pathway, and CHEMERIN signaling pathway (Figure 7C-H). CTHRC1-positive CAFs and macrophages exhibited close communication.

CTHRC1-positive CAFs promoted infiltration of M2 macrophages

The expression levels of CTHRC1 and invasive genes were increased in TNBC patients (Figure 8B). Immunofluorescence

analysis of tumour and adjacent non-tumour tissues revealed a significant increase in M2 macrophage infiltration and elevated CTHRC1 expression levels (Figure 8A). These results validate our intriguing findings based on bioinformatics (Figure 8C). CTHRC1-positive CAFs facilitated the infiltration of M2-polarized macrophages.

Discussion

TNBC accounts for approximately 20% of breast cancer patients (22). In contrast to other pathological types, triple-negative breast cancer exhibits distinctive clinical features and tumor microenvironmental characteristics (23). Statistical data indicates that the majority of patients experience recurrence within 5 years

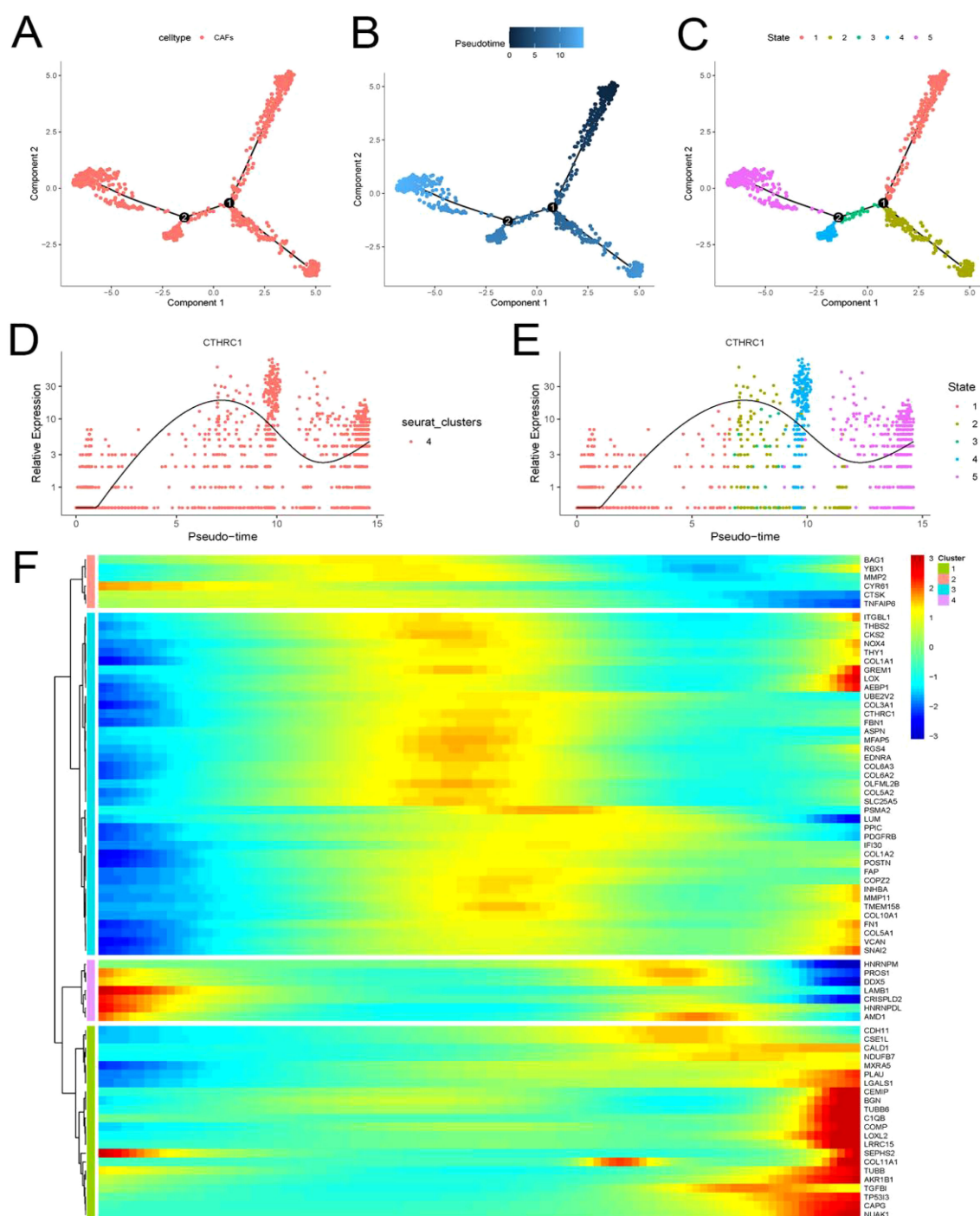


FIGURE 6

Invasive genes and CTHRC1 showed progressively higher expression with differentiation of tumor-associated fibroblasts. (A-E) CTHRC1 expression gradually increased with the differentiation of tumor-associated fibroblasts. (F) Invasive genes are accompanied by increased CTHRC1 expression in CAFs.

after diagnosis, implying that TNBC significantly worsen the burden on patients due to its high metastasis and recurrence rates (24). Considering the inter- and intra-tumor heterogeneity, identifying potential therapeutic targets based on the clinical characteristics of these tumors would greatly enhance patient prognosis (25, 26).

It is widely acknowledged that TNBC exhibits distinct invasive characteristics (27, 28). The investigation of potential targets to impede the invasion of TNBC is a highly researched topic (29, 30). SF3A2 and SNIP1 have been identified as crucial mediators implicated in the invasion of triple-negative breast cancer (31, 32). Diverse molecules and cellular components participate in regulating

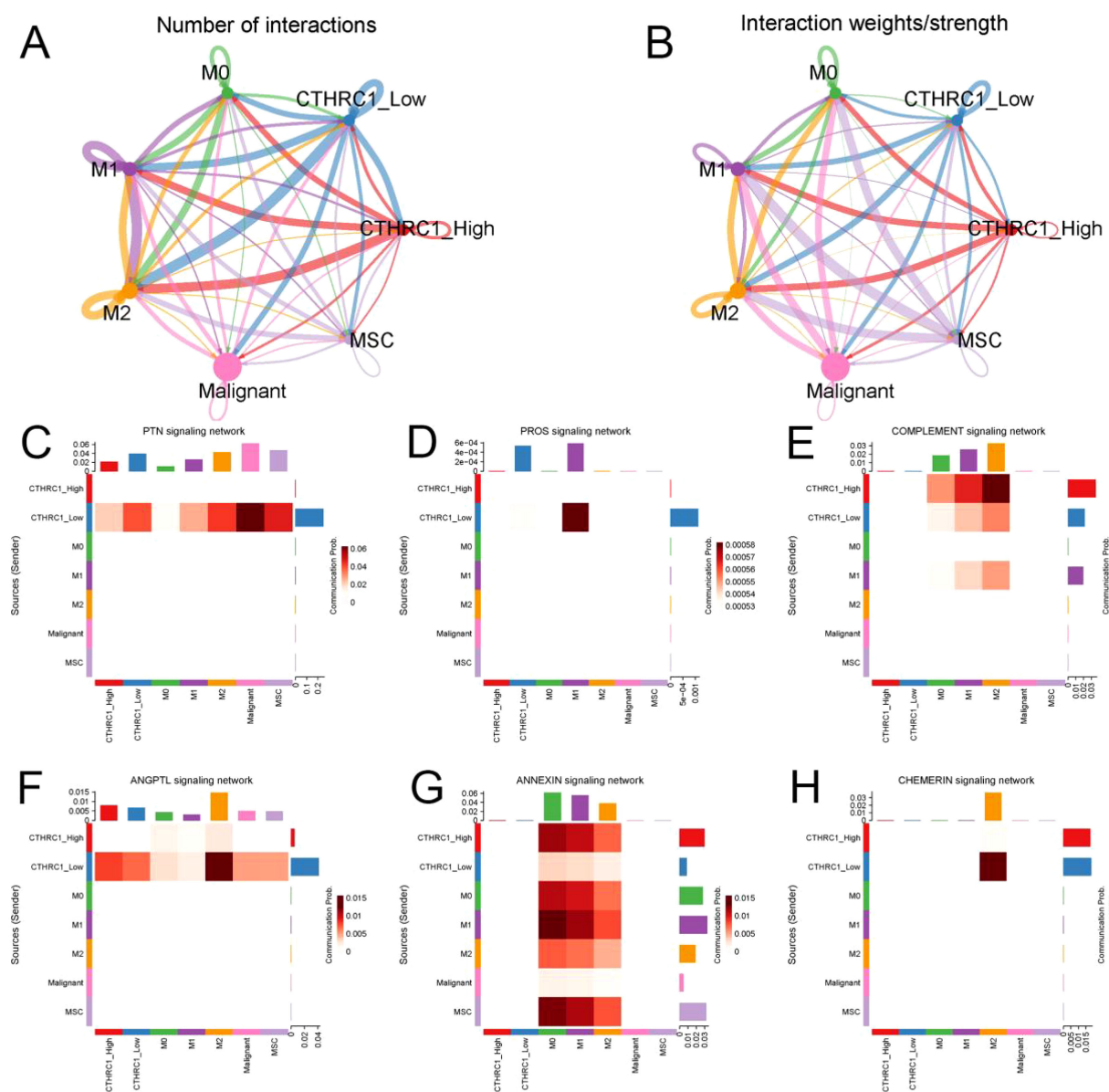


FIGURE 7

CTHRC1 expression level affected the interaction between tumor-associated fibroblasts and macrophages. (A, B) Probabilities and weights of interactions between tumor-associated fibroblasts and macrophages. (C–H) The high-expression group of CTHRC1 exhibited enhanced activity in the complement signaling pathway and annexin signaling pathway compared to the low-expression group of CTHRC1. The PTN signaling pathway, PROS signaling pathway, ANGPTL signaling pathway, and CHEMERIN signaling pathway showed high activity in CTHRC1 low group.

the microenvironment of TNBC and their malignant behaviors (33, 34). Ye et al. revealed that senescent CAFs facilitate the progression of TNBC by mediating immunosuppression (35). Mendez discovered that HMGA1 was predominantly enriched at the leading edges of primary tumor invasion and in metastatic lesions. Furthermore, inhibition of HMGA1 was found to attenuate invasion and metastasis in TNBC (36). Studies have demonstrated the involvement of CTHRC1 in gastric cancer metastasis and hepatocellular carcinoma metastasis (7, 37). Interestingly, CTHRC1-positive CAFs played a pivotal role in disease

progression. Wong et al. observed that CTHRC1-expressing CAFs suppress immune cell function to promote prostate cancer progression (38).

We observed a strong association between CTHRC1 and tumor T-stage and N-stage specifically in TNBC, while no similar correlation was found in non-TNBC. The elevated expression of CTHRC1 in TNBC was accompanied by an increase in the expression of the invasive gene set, which was not observed in non-TNBC. Therefore, it was highly likely that CTHRC1 served as a signature molecule involved in invasion specifically in TNBC.

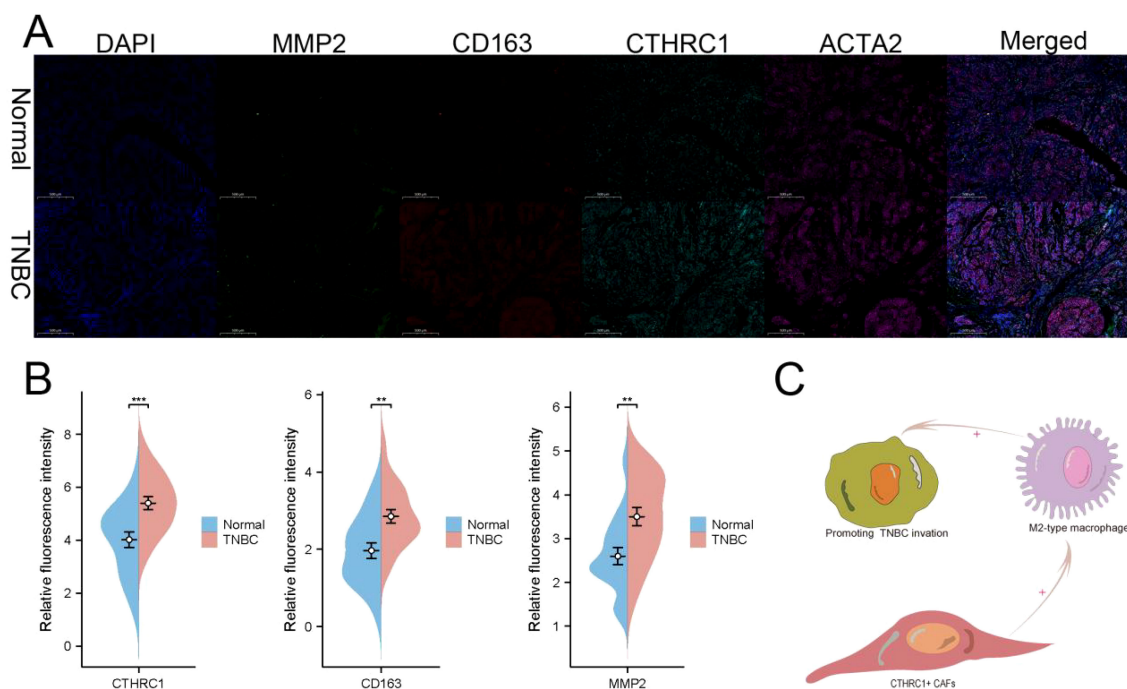


FIGURE 8

Immunofluorescence was used to validate the unique characteristics of CTHRC1 in TNBC patients. (A) Utilizing multiple immunofluorescence techniques, it was revealed that CTHRC1-positive fibroblasts exert an influence on the infiltration of M2-type macrophages. (B) CTHRC1, CD163, and MMP2 were upregulated in patients with TNBC. (C) A potential mechanism by which CTHRC1-positive CAFs regulated TNBC tumor microenvironment. ** $P < 0.01$; *** $P < 0.001$.

Single-cell data analysis revealed predominant expression of CTHRC1 in CAFs, and GSVA enrichment analysis demonstrated that these fibroblasts exhibited the highest expression levels of the invasion gene set. Further clustering identified two distinct subpopulations within tumor-associated fibroblasts: CTHRC1-positive cells and negative cells. Interestingly, the activity of the invasive gene set was significantly higher in CTHRC1-positive CAFs compared to CTHRC1-negative CAFs. Notably, a positive correlation was observed between CTHRC1 expression and the differentiated expression of CAFs. Furthermore, during differentiation, there was a gradual increase in the expression of numerous CAFs accompanied by most of the invasive genes, providing further evidence for the role of CTHRC1 in promoting TNBC progression through regulation of CAFs.

The interactions between CTHRC1 and macrophages have been previously reported. Qin et al. demonstrated that CTHRC1 upregulated infiltration of M2-type macrophages through the TGF- β signaling pathway (39). Zhang et al. discovered that CTHRC1 regulated the abundance of M2-type macrophage infiltration, thereby reducing colon cancer liver metastasis (37). Similarly, our study observed intriguing phenomena where multiple immune infiltration analysis algorithms indicated a modest association between CTHRC1 and T cells, NK cells, and dendritic cells. However, multiple algorithms have demonstrated that CTHRC1 significantly enhances the abundance of M2-type macrophage

infiltration. In addition, a comparative analysis was performed to examine the distinct interactions between macrophages and groups with high or low expression of CTHRC1. The results from single-cell intercellular communication analysis further validated that variations in CTHRC1 expression among CAFs can influence the reciprocal exchange with macrophages.

Although the concept behind this study is highly innovative and the results are remarkably intriguing, there are still certain limitations that need to be addressed. Firstly, while this study establishes a positive correlation between CTHRC1 and invasive gene set, it falls short in elucidating the underlying mechanism through which CTHRC1 regulates CAFs function. Secondly, the findings lack validation at both animal and cellular levels. Lastly, further investigation is warranted to comprehend the intricate interplay between CAFs and M2-type macrophages.

Conclusion

In conclusion, CTHRC1 is predominantly expressed in CAFs and exhibits clinical characteristics resembling TNBC by influencing invasion-related genes. Moreover, CTHRC1 modulates the immunosuppressive tumor microenvironment of TNBC through the regulation of M2-type macrophages. However, further experimental validation is necessary to corroborate the findings of this study.

Data availability statement

The original contributions presented in the study are included in the article/**Supplementary Material**. Further inquiries can be directed to the corresponding author/s.

Ethics statement

The studies involving humans were approved by The studies involving human participants are reviewed and approved by the Medical Ethics Committee of The First Affiliated Hospital of Anhui Medical University. The studies were conducted in accordance with the local legislation and institutional requirements. The participants provided their written informed consent to participate in this study.

Author contributions

HW: Software, Validation, Writing – original draft. ZL: Data curation, Software, Supervision, Writing – original draft. YX: Data curation, Writing – original draft. DY: Formal analysis, Software, Writing – original draft. HJ: Data curation, Formal analysis, Writing – original draft. ZR: Data curation, Formal analysis, Writing – original draft. XY: Writing – review & editing. JP: Writing – review & editing.

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Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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Supplementary material

The Supplementary Material for this article can be found online at: <https://www.frontiersin.org/articles/10.3389/fimmu.2025.1534981/full#supplementary-material>

References

- Giaquinto AN, Sung H, Miller KD, Kramer JL, Newman LA, Minihan A, et al. Breast cancer statistics, 2022. *CA: Cancer J Clin.* (2022) 72:524–41. doi: 10.3322/caac.21754
- Denkert C, Liedtke C, Tutt A, von Minckwitz G. Molecular alterations in triple-negative breast cancer: the road to new treatment strategies. *Lancet (London England).* (2017) 389:2430–42. doi: 10.1016/S0140-6736(16)32454-0
- Bianchini G, Balko JM, Mayer IA, Sanders ME, Gianni L. Triple-negative breast cancer: challenges and opportunities of a heterogeneous disease. *Nat Rev Clin Oncol.* (2016) 13:674–90. doi: 10.1038/nrclinonc.2016.66
- Tazzite A, Jouhadi H, Benider A, Nadifi S. BRCA mutational status is a promising predictive biomarker for platinum-based chemotherapy in triple-negative breast cancer. *Curr Drug targets.* (2020) 21:962–73. doi: 10.2174/1389450121666200203162541
- Sharma P. Biology and management of patients with triple-negative breast cancer. *oncologist.* (2016) 21:1050–62. doi: 10.1634/theoncologist.2016-0067
- Belli C, Duso BA, Ferraro E, Curigliano G. Homologous recombination deficiency in triple negative breast cancer. *Breast (Edinburgh Scotland).* (2019) 45:15–21. doi: 10.1016/j.breast.2019.02.007
- Mei D, Zhu Y, Zhang L, Wei W. The role of CTHRC1 in regulation of multiple signaling and tumor progression and metastasis. *Mediators inflammation.* (2020) 2020:9578701. doi: 10.1155/2020/9578701
- Singh CK, Fernandez S, Chhabra G, Zaemisch GR, Nihal A, Swanlund J, et al. The role of collagen triple helix repeat containing 1 (CTHRC1) in cancer development and progression. *Expert Opin Ther targets.* (2024) 28:419–35. doi: 10.1080/14728222.2024.2349686
- Li LY, Yin KM, Bai YH, Zhang ZG, Di W, Zhang S. CTHRC1 promotes M2-like macrophage recruitment and myometrial invasion in endometrial carcinoma by integrin-Akt signaling pathway. *Clin Exp metastasis.* (2019) 36:351–63. doi: 10.1007/s10585-019-09971-4
- Liu Y, Chen X, Xu Y, Yang T, Wang H, Wang Z, et al. CTHRC1 promotes colorectal cancer progression by recruiting tumor-associated macrophages via up-regulation of CCL15. *J Mol Med (Berlin Germany).* (2024) 102:81–94. doi: 10.1007/s00109-023-02399-0
- Jézéquel P, Loussouarn D, Guérin-Charbonnel C, Campion L, Vanier A, Gouraud W, et al. Gene-expression molecular subtyping of triple-negative breast cancer tumours: importance of immune response. *Breast Cancer research: BCR.* (2015) 17:43. doi: 10.1186/s13058-015-0550-y
- Ahn SG, Kim SK, Shepherd JH, Cha YJ, Bae SJ, Kim C, et al. Clinical and genomic assessment of PD-L1 SP142 expression in triple-negative breast cancer. *Breast Cancer Res Treat.* (2021) 188:165–78. doi: 10.1007/s10549-021-06193-9
- Metzger-Filho O, Collier K, Asad S, Ansell PJ, Watson M, Bae J, et al. Matched cohort study of germline BRCA mutation carriers with triple negative breast cancer in brightness. *NPJ Breast cancer.* (2021) 7:142. doi: 10.1038/s41523-021-00349-y
- Wu SZ, Al-Eryani G, Roden DL, Junankar S, Harvey K, Andersson A, et al. A single-cell and spatially resolved atlas of human breast cancers. *Nat Genet.* (2021) 53:1334–47. doi: 10.1038/s41588-021-00911-1

15. Xu S, Chen X, Fang J, Chu H, Fang S, Zeng L, et al. Comprehensive analysis of 33 human cancers reveals clinical implications and immunotherapeutic value of the solute carrier family 35 member A2. *Front Immunol.* (2023) 14:1155182. doi: 10.3389/fimmu.2023.1155182
16. Xu S, Chen X, Ying H, Chen J, Ye M, Lin Z, et al. Multi-omics identification of a signature based on Malignant cell-associated ligand-receptor genes for lung adenocarcinoma. *BMC Cancer.* (2024) 24:1138. doi: 10.1186/s12885-024-12911-5
17. Xu S, Zheng Y, Ye M, Shen T, Zhang D, Li Z, et al. Comprehensive pan-cancer analysis reveals EPHB2 is a novel predictive biomarker for prognosis and immunotherapy response. *BMC Cancer.* (2024) 24:1064. doi: 10.1186/s12885-024-12843-0
18. Bassez A, Vos H, Van Dyck L, Floris G, Arijis I, Desmedt C, et al. A single-cell map of intratumoral changes during anti-PD1 treatment of patients with breast cancer. *Nat Med.* (2021) 27:820–32. doi: 10.1038/s41591-021-01323-8
19. Shiao SL, Gouin KH 3rd, Ing N, Ho A, Basho R, Shah A, et al. Single-cell and spatial profiling identify three response trajectories to pembrolizumab and radiation therapy in triple negative breast cancer. *Cancer Cell.* (2024) 42:70–84.e8. doi: 10.1016/j.ccell.2023.12.012
20. Zhang Y, Chen H, Mo H, Hu X, Gao R, Zhao Y, et al. Single-cell analyses reveal key immune cell subsets associated with response to PD-L1 blockade in triple-negative breast cancer. *Cancer Cell.* (2021) 39:1578–93.e8. doi: 10.1016/j.ccell.2021.09.010
21. Cords L, Tietscher S, Anzeneder T, Langwieder C, Rees M, de Souza N, et al. Cancer-associated fibroblast classification in single-cell and spatial proteomics data. *Nat Commun.* (2023) 14:4294. doi: 10.1038/s41467-023-39762-1
22. Garrido-Castro AC, Lin NU, Polyak K. Insights into molecular classifications of triple-negative breast cancer: improving patient selection for treatment. *Cancer discovery.* (2019) 9:176–98. doi: 10.1158/2159-8290.CD-18-1177
23. Bai X, Ni J, Beretov J, Graham P, Li Y. Triple-negative breast cancer therapeutic resistance: Where is the Achilles' heel? *Cancer Lett.* (2021) 497:100–11. doi: 10.1016/j.canlet.2020.10.016
24. Jia H, Truica CI, Wang B, Wang Y, Ren X, Harvey HA, et al. Immunotherapy for triple-negative breast cancer: Existing challenges and exciting prospects. *Drug resistance updates: Rev commentaries antimicrobial Anticancer chemother.* (2017) 32:1–15. doi: 10.1016/j.drug.2017.07.002
25. Bose S. Triple-negative breast carcinoma: morphologic and molecular subtypes. *Adv anatomic pathol.* (2015) 22:306–13. doi: 10.1097/PAP.0000000000000084
26. Zhou L, Yu CW. Epigenetic modulations in triple-negative breast cancer: Therapeutic implications for tumor microenvironment. *Pharmacol Res.* (2024) 204:107205. doi: 10.1016/j.phrs.2024.107205
27. Borri F, Granaglia A. Pathology of triple negative breast cancer. *Semin Cancer Biol.* (2021) 72:136–45. doi: 10.1016/j.semcancer.2020.06.005
28. Ji GW, Fan Y, Sun DW, Wu MY, Wang K, Li XC, et al. Machine learning to improve prognosis prediction of early hepatocellular carcinoma after surgical resection. *J hepatocellular carcinoma.* (2021) 8:913–23. doi: 10.2147/JHC.S320172
29. Zhang Y, Wang W, Min J, Liu S, Wang Q, Wang Y, et al. ZNF451 favors triple-negative breast cancer progression by enhancing SLUG-mediated CCL5 transcriptional expression. *Cell Rep.* (2023) 42:112654. doi: 10.1016/j.celrep.2023.112654
30. Zhong W, Jian Y, Zhang C, Li Y, Yuan Z, Xiong Z, et al. SHC4 orchestrates β -catenin pathway-mediated metastasis in triple-negative breast cancer by promoting Src kinase autophosphorylation. *Cancer letters.* (2024) 582:216516. doi: 10.1016/j.canlet.2023.216516
31. Deng L, Liao L, Zhang YL, Yang SY, Hu SY, Andriani L, et al. SF3A2 promotes progression and cisplatin resistance in triple-negative breast cancer via alternative splicing of MKRN1. *Sci Adv.* (2024) 10:eadj4009. doi: 10.1126/sciadv.adj4009
32. Yu B, Su J, Shi Q, Liu Q, Ma J, Ru G, et al. KMT5A-methylated SNIP1 promotes triple-negative breast cancer metastasis by activating YAP signaling. *Nat Commun.* (2022) 13:2192. doi: 10.1038/s41467-022-29899-w
33. Karim AM, Eun Kwon J, Ali T, Jang J, Ullah I, Lee YG, et al. Triple-negative breast cancer: epidemiology, molecular mechanisms, and modern vaccine-based treatment strategies. *Biochem Pharmacol.* (2023) 212:115545. doi: 10.1016/j.bcp.2023.115545
34. Zagami P, Carey LA. Triple negative breast cancer: Pitfalls and progress. *NPJ Breast cancer.* (2022) 8:95. doi: 10.1038/s41523-022-00468-0
35. Ye J, Baer JM, Faget DV, Morikis VA, Ren Q, Melam A, et al. Senescent CAFs mediate immunosuppression and drive breast cancer progression. *Cancer discovery.* (2024) 14:1302–23. doi: 10.1158/2159-8290.CD-23-0426
36. Méndez O, Peg V, Salvans C, Pujals M, Fernández Y, Abasolo I, et al. Extracellular HMGAI promotes tumor invasion and metastasis in triple-negative breast cancer. *Clin Cancer Res.* (2018) 24:6367–82. doi: 10.1158/1078-0432.CCR-18-0517
37. Zhang XL, Hu LP, Yang Q, Qin WT, Wang X, Xu CJ, et al. CTHRC1 promotes liver metastasis by reshaping infiltrated macrophages through physical interactions with TGF- β receptors in colorectal cancer. *Oncogene.* (2021) 40:3959–73. doi: 10.1038/s41388-021-01827-0
38. Wong HY, Sheng Q, Hesterberg AB, Croessmann S, Rios BL, Giri K, et al. Single cell analysis of cribriform prostate cancer reveals cell intrinsic and tumor microenvironmental pathways of aggressive disease. *Nat Commun.* (2022) 13:6036. doi: 10.1038/s41467-022-33780-1
39. Qin S, Zheng JH, Xia ZH, Qian J, Deng CL, Yang SL. CTHRC1 promotes wound repair by increasing M2 macrophages via regulating the TGF- β and notch pathways. *Biomed pharmacother = Biomed pharmacotherapie.* (2019) 113:108594. doi: 10.1016/j.biopha.2019.01.055