#### Check for updates

### **OPEN ACCESS**

EDITED BY Mohamad S. Hakim, Qassim University, Saudi Arabia

REVIEWED BY Maria Gazouli, National and Kapodistrian University of Athens, Greece Pavel Loskot, The Zhejiang University-University of Illinois at Urbana-Champaign Institute, United States

CORRESPONDENCE Lijun Yang ↓ lijunyang224@163.com Lingnan He ↓ 2205551@tongii.edu.cn

RECEIVED 06 December 2024 ACCEPTED 02 April 2025 PUBLISHED 24 April 2025

#### CITATION

Yang L, Li H, Tang M, He L and Yang L (2025) Circular RNAs in inflammatory bowel disease: a review of mechanisms, biomarkers and therapeutic potential. *Front. Immunol.* 16:1540768. doi: 10.3389/fimmu.2025.1540768

#### COPYRIGHT

© 2025 Yang, Li, Tang, He and Yang. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

### Circular RNAs in inflammatory bowel disease: a review of mechanisms, biomarkers and therapeutic potential

Le Yang<sup>1</sup>, Huahui Li<sup>2</sup>, Min Tang<sup>3</sup>, Lingnan He<sup>4\*</sup> and Lijun Yang<sup>3\*</sup>

<sup>1</sup>Department of Gastroenterology, Yiyang Central Hospital, Yiyang, China, <sup>2</sup>Institute of Biomedical and Health Engineering, Chinese Academy of Sciences Shenzhen Institutes of Advanced Technology, Shenzhen, China, <sup>3</sup>Department of Pharmacy, Yiyang Medical College, Yiyang, China, <sup>4</sup>Endoscopy Center, Department of Gastroenterology, Shanghai East Hospital, School of Medicine, Tongji University, Shanghai, China

Inflammatory bowel disease (IBD) is an autoimmune disease of unknown etiology characterized by recurrent chronic inflammation of the gastrointestinal tract. The incidence of IBD is increasing and has become a focus of research on digestive diseases. Despite advances in understanding its multifactorial etiology, including genetic predisposition, microbiome dysbiosis, and immune dysregulation. However, the molecular mechanisms driving IBD pathogenesis remain incompletely elucidated. Circular RNA (circRNA) is a stable single-stranded RNA with a closed-loop structure and conserved nature. circRNA possesses multiple functions, such as adsorption of microRNAs and RNA-binding proteins, and is involved in the regulation of gene splicing and transcription, as well as protein translation. However, circRNAs in IBD progression and their clinical potential as biomarkers or therapeutic targets are yet to be systematically explored. In this review, we comprehensively synthesize recent advancements in circRNA research related to IBD, integrating evidence from in vitro, in vivo, and clinical studies. We systematically analyze aberrant circRNA expression profiles in IBD tissues (e.g., intestinal mucosa, peripheral blood, and exosomes) and discuss their mechanism of action contributions to inflammation, intestinal epithelial barrier dysfunction, autophagy, intestinal fibrosis, and colitis-associated cancer (CAC). Furthermore, we evaluate methodologies for circRNA detection and therapeutic modulation, including RNA interference, viral vector delivery, and PLGA MSs delivery system strategies. This review highlights the potential of circRNA-focused strategies in the diagnosis and treatment of IBD, offering a scientific foundation for advancing precision medicine in IBD management.

#### KEYWORDS

circular RNA, inflammatory bowel disease, ulcerative colitis, Crohn's disease, biomarker, diagnosis, prognosis

### **1** Introduction

Inflammatory bowel disease (IBD), which includes ulcerative colitis (UC) and Crohn's disease (CD), is a chronic intestinal inflammatory disorder of unknown etiology. The incidence of IBD is rising globally, making it a significant health concern (1). The etiology of IBD is primarily associated with genetics, environmental factors, dysbiosis of the gut microbiome, and immune system disorders, leading to dysregulation of intestinal barriers and immune functions (2, 3). The diagnosis of IBD relies on clinical assessment, laboratory tests, imaging, endoscopy, and histological examination. However, there is currently no definitive diagnostic criterion. Identifying new therapeutic targets and diagnosis biomarkers is crucial for early detection and personalized treatment.

Non-coding RNAs, such as microRNAs(miRNA), long noncoding RNAs(lncRNA), and circRNA, do not encode proteins but play a crucial role in regulating gene expression within cellular processes (4). Early studies focused on miRNAs and lncRNAs revealed their critical roles in modulating IBD-associated pathways-such as NF-KB activation, immune homeostasis, and intestinal barrier integrity (5-7). CircRNAs are a class of covalently closed circular RNA molecules that lack a 5' cap and a 3' poly(A) tail (8). This structural feature confers exceptional stability and resistance to exonuclease-mediated degradation, enabling circRNAs to be stably expressed in the cytoplasm or stored within exosomes compared to their linear counterparts (9). They are widely expressed in mammalian tissues and cells, exhibiting tissue and cell type-specific patterns (10), and are involved in various physiological and pathological processes (11). They exhibit aberrant expression in various diseases, including (12, 13), cardiovascular diseases (14), autoimmune diseases (15), neurological disorders (16), and aging processes (17). CircRNAs can be detected in multiple body fluids (peripheral blood, saliva, urine, semen, etc.) and tissues/cells (18). The unique characteristics of circRNAs make them potential biomarkers and therapeutic targets for the diagnosis and treatment of human diseases (19, 20).

The role and molecular mechanism of circRNAs in IBD remain incompletely understood. Although circRNAs in IBD have not been extensively studied, emerging evidence has identified differential expression of specific circRNAs(e.g., circRNA\_103516,circ\_004662) between IBD patients and healthy controls, with these molecules playing pivotal roles in disease pathogenesis (21, 22). Accumulating data further suggests their potential clinical utility in IBD diagnosis, prognosis, and therapeutic interventions (7, 23). Therefore, the functional contributions and pathogenic mechanisms of circRNAs in IBD urgently require further investigation to achieve a comprehensive understanding.

Recent reviews, such as the comprehensive work by Lun et al, have systematically summarized the roles of circRNAs in IBD pathogenesis and immune regulation (24). However, emerging studies over the past years have revealed novel mechanistic insights into circRNA-mediated intestinal barrier dysfunction, autophagy modulation, intestinal fibrosis progression, and Colitisassociated cancer(CAC). Building on these findings, this review systematically synthesizes cutting-edge research advances to address critical questions regarding the roles of circRNAs in IBD. Our work provides principal contributions: Firstly, we elucidated the dynamic differential expression profile of circular RNAs (circRNAs) in inflammatory bowel disease (IBD), with validation confirming their potential as non-invasive biomarkers in plasma exosomes (e.g., circ103124). Secondly, we explored the mechanistic role of circRNAs in IBD in detail. Regarding their function as microRNA (miRNA) sponges, we discussed issues such as inflammation amplification, immune dysregulation, intestinal mucosal barrier disruption, and autophagy modulation. Concerning RNA-binding proteins (RBPs), we focused on posttranscriptional regulation, particularly in intestinal mucosal repair and autophagy regulation. Furthermore, in the field of epigenetics, we investigated the role of N6-methyladenosine(m6A)-dependent epigenetic mechanisms in IBD. Additionally, we introduced the circRNA-mediated regulatory network involved in intestinal fibrosis and colitis-associated cancer (CAC).Finally, we evaluated innovative therapeutic delivery systems, specifically the application of poly(lactic-co-glycolic acid) (PLGA) microspheres for circRNAtargeted interventions in IBD, aiming to provide a more effective and safer therapeutic strategy for patients with inflammatory bowel disease.

# 2 Aberrant expression of CircRNAs in IBD

Emerging evidence positions circRNAs as pivotal regulators in autoimmune disorders, particularly IBD. Moreover, circRNAs regulate various physiological and pathological mechanisms, and their differential expression during disease progression underscores their significance (25)(Figure 1). Elevated levels of circRNA\_103516, circAtp9b, and circ\_103756 in peripheral blood mononuclear cells (PBMCs) of IBD patients are associated with systemic inflammation (21, 26, 27). Exosomal circ\_103124 is markedly upregulated in UC plasma, where it accelerates epithelial barrier disruption (28). Yin et al. found significant changes in four circRNAs(circ\_092520, circ\_102610,circ\_004662, circ\_103124)in PBMCs of CD patients, with circRNA\_004662 in PBMCs distinguishes CD from UC, offering a non-invasive diagnostic tool (29). Importantly, recent studies highlight disease phase-dependent circRNA signature. Recent studies have further elucidated that circRNAs exhibit phase-specific expression dynamics during disease progression, with distinct profiles

Abbreviations: IBD, Inflammatory bowel disease; UC, Ulcerative Colitis; CD, Crohn's Disease; PBMCs, Peripheral Blood Mononuclear Cells; ISCs, Intestinal stem cells; NLRP3, Nucleotide-binding oligomerization domain leucine-rich repeat and pyrin domain-containing 3; LPS, Lipopolysaccharides; m6A, N6methyladenosine; METTL3, Methyltransferase-like 3; MSCs, Mesenchymal Stem Cells; TLR4, Toll-like receptor 4; MyD88, Myeloid differentiation factor 88; GSDMD, Gasdermin D; EMT, Epithelial-to-Mesenchymal Transition; CRC, Colitis-associated Cancer; SOCS1, Suppressor of cytokine signaling 1; SPTBN1, Spectrin beta non-erythrocytic 1; NCOA3, Nuclear Receptor Coactivator 3.



observed between active inflammatory phases and clinical remission.circ\_103516 and circ\_004662 are significantly elevated in IBD during active inflammation compared to remission (21, 22).CircHIPK3 is transiently upregulated in acute mucosal injury (e.g., cecal ligation and puncture models) to promote repair, but paradoxically downregulated in chronic IBD, correlating with impaired healing (30). Upregulation of circ\_102685, circ\_0062142, circSMAD4, circPRKARIB, and circ103124 in the intestinal mucosa tissues of CD patients and modulates key pathways (p53, TLR/NF- $\kappa$ B, IL-17, and JAK2/STAT3), perpetuating chronic inflammation (29, 31–34). Downregulation of circCCND1, circ\_0001021, circHECTD1, and circ\_CDKN2B-AS1 in colonic epithelial tissues of ulcerative colitis patients, while restoring their expression suppresses UC progression (35–38).

CircRNA aberrant expression correlates with disease severity, inflammatory intensity, and phase-specific progression, exhibiting mechanistic roles in mucosal homeostasis and immune dysregulation (Figure 2). These findings contribute to a deeper understanding of the molecular mechanism of IBD and provide novel directions for diagnosis and treatment.

# 3 Mechanistic roles of circRNAs in IBD pathogenesis

The pathogenesis of inflammatory bowel disease (IBD) is not yet fully understood. Many studies suggest that IBD is a multifactorial disease caused by various factors, including genetic defects, environmental abnormalities, dysbiosis of the gut microbiota, and autophagy, which can disrupt intestinal homeostasis and trigger immune-mediated inflammation in the gut (39–43). CircRNAs influence biological processes through multiple mechanisms: by acting as "sponges" for miRNAs, interacting with RNA-binding proteins (RBPs) to modulate RNA



stability, post-transcriptional gene expression regulation (44, 45). Therefore, analyzing the expression changes and functions of circRNA in IBD helps to uncover its molecular mechanisms in the disease progression, providing valuable insights into understanding IBD pathogenesis and developing new therapeutic strategies. Emerging evidence underscores the pivotal involvement of circRNAs in the pathogenesis and progression of IBD, as summarized in Table 1.

### 3.1 CircRNAs as microRNA sponge

CircRNAs harbor multiple miRNA-binding sites, functioning as molecular "sponges" to sequester miRNAs, amplifying downstream gene expression. This regulatory axis termed the competing endogenous RNA (ceRNA) mechanism, is pivotal in modulating IBD pathogenesis. CircRNAs promote the progression of IBD by sequestering miRNAs to alleviate their suppression of target mRNAs and driving pathological processes through proinflammatory signaling pathways, intestinal epithelial dysfunction, and autophagy regulation.

# 3.1.1 Pro-inflammatory signaling and intestinal immune homeostasis

CircRNAs serve as central regulators of inflammatory cytokine production and immune responses, critically influencing the pathogenesis of inflammatory diseases (46). For instance, the expression level of circ\_103516 is elevated in active CD and UC patients, its expression positively correlates with TNF- $\alpha$  and IFN- $\gamma$ levels while inversely associating with IL-10 (21). Mechanistically, it acts as a sponge for hsa-miRNA-19b-1-5p, participating in the inflammatory process of CD (47).In CD, circ\_103765 acts as a ceRNA for the miR-30 family, exerting a pro-inflammatory role (27). The imbalance between Treg and Th17 cells is a key factor in the development of CD (48). circSMAD4 functions as a ceRNA by sequestering miR-135a-5p, activating JAK2/STAT3 signaling to

PBMC, Peripheral blood mononuclear cells; SPTBN1, Spectrin beta non-erythrocytic 1.

Mechanism	CircRNA	MiRNA	Source	Up/Down	Function	Clinical value	Disease activity	Ref
	Circ_103516	MiR-19b-1-5p	РВМС	Up	Promotes intestinal inflammation	Diagnosis	Positive	(21)
	Circ_103756	MiR-30	PBMC/Cells	Up	Promotes apoptosis and intestinal inflammation	Therapeutic target	Positive	(27)
	Circ_102685	MiR-146	Tissues	Up	Promotes apoptosis and intestinal inflammation	Diagnosis/ Therapeutic	Positive	(31, 49, 50)
	CircSMAD4	MiR-135a-5p	Tissues/Cells	Up	Disruption of the intestinal epithelial barrier, pro- inflammatory, immune disorders	Therapeutic target	Positive	(33)
	CircCCND1	MiR-142-5p	Tissues/Cells	Down	Inhibits inflammation and apoptosis	Therapeutic target	Negatively	(35)
miRNA sponges	CircSnx5	miR-544	Tissues/Cells	Down	Promotes DC activation and inflammatory phenotype	Therapeutic target	Negatively	(54)
	Circ_0520	M:D 700		Up		Diagnosis	Negatively	(51)
	Circ_2243	MiR-709						
	Circ_103124	MiR-650/MiR- 1236-3P	PMBC/Cells/Tissues	Up	Promotes of macrophage M1 polarization, pro- inflammatory, pyroptosis inhibition of autophagy	Diagnosis Therapeutic target	Positive	(52, 53)
	Circ_SoD2	MiR-223/MiR-378g	Tissues/Cells	Up	Promotes intestinal inflammation and compromises the intestinal barrier	Therapeutic target	Positive	(55, 56)
	CircHIPK3	MiR-29b	Tissues/Cells	Down	Promotes the intestinal barrier	Therapeutic target		(30)
	Circ_0001021	MiR-224-5p	Tissues/Cells	Down	Promotes the intestinal barrier	Therapeutic target	Negatively	(22)
	Circ_0007919	MiR-138	Tissues/Cells	Down	Promotes the intestinal barrier	Diagnosis Therapeutic target	Negatively	(36)
	CircGMCL1	MiR-124-3p	Tissues	Down	Promotes autophagy, inhibits pyroptosis, inflammation	Therapeutic target	Negatively	(58)
	CircHECTD1	MiR-182-5p	Tissues	Down	Promotes autophagy, inhibits inflammation	Therapeutic target	Negatively	(37)
	CircPan3		Tissues		Promotes ISC self-renewal and differentiation	Therapeutic target		(61)
RNA-Binding Protein Interaction	CircBtnl1		Tissues		Inhibits ISC self-renewal and differentiation			(34)
	circPABPN1		Tissues	Up	Inhibits autophagy, Promotes inflammation	Therapeutic target	Positive	(65)
	CircPRKAR1B		Tissues/Cells	Up	Interact with SPTBN1,inhibits autophagy, promotes pyroptosis and inflammation	Therapeutic target	Positive	( <u>66</u> )
Epigenetic Modulation	CircPRKAR1B		Tissues/Cells	Up	(m6A) modification,inhibits autophagy, promotes pyroptosis and inflammation	Therapeutic target	Positive	( <del>66</del> )

Frontiers in Immunology

promote Th1/Th17 differentiation. Therapeutic inhibition of circSMAD4 restores Treg/Th17 equilibrium, attenuating intestinal inflammation (33). Research conducted by Qiao has revealed that the expression of circ\_102685 is increased in CD, and through influencing the circ\_102685/miRNA-146/NF-KB signaling axis, it regulates T regulatory cells (Treg cells) and Dendritic Cells (DCs) to alleviate intestinal inflammation (31, 49, 50). In melatonin-treated colitis models,circ\_0520 and circ\_2243 impair bone marrowderived dendritic cells(BMDCs) maturation by targeting YWHAZ/CCL19 through the PI3K/AKT pathway, limiting DC phagocytosis and migration and production of pro-inflammatory cytokines, thereby alleviating intestinal inflammation (51). A study by Yin et al. found that circ\_103124 functions as a molecular sponge for miR-650 to activate AKT2/TLR4/NF-KB signaling pathways, driving M1 macrophage polarization. Its upregulation in active CD correlates with TLR4(Toll-like receptor 4)/MyD88(Myeloid differentiation factor 88)/NLRP3/GSDMD axis activation (Gasdermin D), perpetuating pyroptosis and inflammation (52, 53). CircSnx5 acts as a sponge for miR-544 to upregulate the suppressor of cytokine signaling 1 (SOCS1), suppressing the nuclear translocation of PU.1 and DC-mediated T-cell activation. This mechanism highlights its potential in treating IBD and autoimmune disorders (54).CircRNACCND1 sponges miR-142-5p, thereby alleviating its inhibitory effect on Nuclear Receptor Coactivator 3(NCOA3) and restoring NCOA3-mediated antiinflammatory activity (35).

### 3.1.2 Intestinal epithelial barrier dysfunction

CircRNAs play a key role in preserving the intestinal barrier, interacting with tight junction proteins to exert dual functions. For example, circ\_SOD2 affects CLDN-8 or damages the intestinal barrier via the miR-378g/Snail1 pathway, promoting the progression of ulcerative colitis (55, 56).circSMAD4 functions as a ceRNA to sequester miR-135a-5p, leading to JAK2 overexpression and subsequent degradation of tight junction proteins (occludin, ZO-1). This mechanism drives epithelial barrier dysfunction in CD (33). Genome-wide analyses have revealed that circHIPK3 significantly downregulated in IBD and sepsis patients, circHIPK3 sponges miR-29b to derepress pro-repair effectors (Rac1, Cdc42, Cyclin B1), restoring the migratory and proliferative capacities of intestinal epithelial cells (30). Specific circRNAs, such as, circ\_0001021 and circ\_0007919, preserve barrier integrity by modulating Smad4,ZO-1,CLDN-2,CLDN-8,VIPR1, and EPC1 expression through miRNA-dependent mechanisms, with particular therapeutic relevance in UC (22, 36). This ceRNA mechanism plays a pivotal role in repairing mucosal damage in IBD.

### 3.1.3 Autophagy modulation

Autophagy is a critical intracellular biological process that plays an essential role in cell health and stress response. It is crucial for maintaining intestinal homeostasis, regulating immunity, and defending against pathogens. Dysregulation of autophagy is associated with various diseases, particularly when it occurs in intestinal epithelial cells, impairs mucosal immunity, and exacerbates chronic inflammation (57).circGMCL1 functioning as a molecular sponge for miR-124-3p, derepressing its target ANXA7 to enhance autophagic. This mechanism suppresses NLRP3 inflammasome activation and pyroptosis, thereby preserving barrier integrity and attenuating inflammation in CD (58). circHECTD1 sequesters miR-182-5p to stabilize HuR, an RNAbinding protein that prolongs the half-life of ATG5 and ATG9 mRNAs in Caco-2 cells. Elevated ATG5/ATG9 levels rescue impaired autophagosome formation, mitigating oxidative stress and epithelial apoptosis in UC (37). Autophagy-related genes ATG5 and ATG9 play pivotal roles in UC pathogenesis. ATG5, a critical regulator of autophagy, not only maintains cellular homeostasis but also modulates immune responses and apoptosis. Its elevated expression in UC patients suggests its involvement in disease progression. Concurrently, ATG9 facilitates autophagosome membrane formation during autophagy, further underscoring the centrality of autophagic dysregulation in UC pathogenesis (59, 60). Collectively, these findings underscore the intricate interplay between autophagy and inflammatory regulation, highlighting their dynamic balance in maintaining tissue homeostasis and disease progression.

By sponging specific miRNAs, circRNAs establish intricate competing ceRNA regulatory networks, which orchestrate multiple pathological cascades in IBD progression: inflammatory amplification, immune homeostasis disruption, barrier dysfunction, and autophagy-inflammatory crosstalk. These synergistic mechanisms collectively perpetuate the sustained chronic inflammation and progressive tissue damage characteristic of IBD.

# 3.2 CircRNAs bind to RNA-binding protein (RBP): post-transcriptional control

CircRNAs can bind to RBPs and coordinate posttranscriptional networks in the pathogenesis of IBD. This mechanism may contribute to the development and progression of IBD by influencing intestinal epithelial cell proliferation, autophagy, and inflammatory signaling pathways.

### 3.2.1 Intestinal stem cells-mediated intestinal mucosal repair

The intestinal epithelium serves as the primary barrier against luminal threats, with intestinal stem cells (ISCs) being indispensable for preserving mucosal homeostasis. Emerging evidence highlights the regulatory dominance of specific circRNAs in this biological process. For instance, circPan3 upregulation in Lgr5+ ISCs elevates IL-13R $\alpha$ 1 mRNA levels, enhancing their regenerative capacity and differentiation potential, thereby facilitating mucosal repair and epithelial barrier restoration. These findings underscore the pivotal role of the IL-13R $\alpha$ 1-mediated signaling axis in ISCdriven tissue regeneration. Conversely, genetic ablation of CircPan3 in Lgr5+ ISCs exacerbates intestinal inflammation in preclinical models, suggesting its protective function in mucosal immunity (61). In contrast, circBtnl1 demonstrates an inhibitory role in ISC dynamics. Mechanistically, it suppresses self-renewal by destabilizing Atf4 mRNA and downregulating Sox9 expression, a transcription factor essential for stem cell maintenance. This dual regulatory mechanism positions circBtnl1 as a negative modulator of ISC proliferation (34).

### 3.2.2 Regulation of intestinal epithelial cell homeostasis

HuR (Human Antigen R), an RNA-binding protein, dynamically regulates gene expression through interactions with non-coding RNAs, including miRNAs and lncRNAs (62, 63). In the context of IBD, HuR binds to the 3' untranslated region (3'UTR) of ATG16L1 mRNA, stabilizing its transcript and enhancing translational efficiency, thereby driving ATG16L1 protein synthesis. As a core component of autophagosome formation, ATG16L1 is essential for maintaining autophagic flux in intestinal epithelial cells, a critical process for epithelial homeostasis (64). Notably, emerging studies reveal HuR's functional crosstalk with circRNAs. For example, circPABPN1 suppresses HuR-mediated translational activation of ATG16L1, impairing autophagy and triggering mucosal immune dysregulation. This HuRcircPABPN1-ATG16L1 axis is implicated in perpetuating intestinal inflammation and epithelial injury in IBD, highlighting its role as a pathogenic driver (65). Therapeutic targeting of this axis may restore autophagic activity and mitigate IBD progression. Furthermore, circPRKAR1B interacts with the RNA-binding protein SPTBN1 (spectrin beta non-erythrocytic 1), disrupting autophagy while activating pyroptosis, a pro-inflammatory cell death pathway. This dual mechanism exacerbates colonic inflammation in CD, underscoring the pathological relevance of circRNA-RBP complexes in IBD pathogenesis (66).

These insights underscore the centrality of circRNA-RBP interactions in IBD pathophysiology, bridging transcriptional regulation with epithelial-immune dysfunction. Their therapeutic exploitation holds transformative potential for halting disease progression.

### 3.3 circRNAs and epigenetic modulation

CircRNAs exhibit enhanced stability due to their covalently closed structures, which confer resistance to RNase R-mediated degradation. Notably, the metabolic dynamics of circRNAs are modulated through N6-methyladenosine (m6A)-dependent epigenetic regulatory mechanisms, a dynamic RNA modification that governs diverse biological processes, including RNA decay, circRNA biogenesis, splicing, and translational regulation. Notably, m6A modifications are prevalent across circRNAs and play a critical role in IBD pathogenesis. METTL3, the core catalytic subunit of the m6A methyltransferase complex, acts as a master "writer" of this modification. Elevated METTL3 expression has been observed in colonic tissues of CD patients compared to healthy controls, suggesting its involvement in IBD-related epigenetic dysregulation (67-70). For instance, circPRKAR1B undergoes METTL3orchestrated m6A modification, which destabilizes its structure and accelerates degradation. This m6A-dependent regulation suppresses autophagic flux while activating the NLRP3 inflammasome-pyroptosis, thereby amplifying mucosal inflammation and epithelial damage in CD (66).

### 4 CircRNAs and intestinal fibrosis

Intestinal fibrosis represents a frequent complication in CD, where epithelial-mesenchymal transition (EMT) facilitates fibroblast recruitment within inflammatory foci. These fibroblasts constitute a principal reservoir of mesenchymal stromal cells(MSC) in the colonic mucosa (71).

Emerging evidence implicates circRNAs as pivotal regulators of fibrogenesis through EMT modulation. For instance, hsa\_circ\_0062142 and hsa\_circ\_0001666 critically influence CD pathogenesis by regulating EMT-mediated transcriptional reprogramming, thereby accelerating disease progression (32). Genetic depletion of circFOXP1 markedly impairs the fibrogenic differentiation potential of mesenchymal stem cells, as demonstrated in both experimental models and clinical specimens (72). Juan et al. revealed that hsa\_circRNA\_102610 functions as a competitive endogenous RNA by sequestering hsa-miR-130a-3p, which amplifies TGF- $\beta$ 1 signaling to potentiate EMT and extracellular matrix remodeling, ultimately driving intestinal fibrosis in CD (73).

Emerging evidence delineates a central role for circRNAs in driving intestinal fibrogenesis through coordinated regulation of EMT, TG- $\beta$ 1signaling hyperactivation, and MSC differentiation.

# 5 CircRNA and colitis-associated cancer(CAC)

Chronic inflammatory microenvironments serve as pivotal drivers of tumorigenesis, particularly in IBD, where persistent mucosal inflammation significantly elevates the risk of CRC (74). This review systematically integrates, for the first time, the molecular regulatory network by which circRNAs drive CAC pathogenesis. Comparative transcriptomic analyses reveal distinct circRNA expression profiles between UC, CAC, and normal colonic tissues, with specific circRNAs emerging as modulators of malignant transformation (22, 23). A study compared the expression of circRNAs in normal colon tissue versus the CAC model in mice. Key findings include: mmu\_circ\_001801, mmu\_circ\_002987, and mmu\_circ\_001155 were upregulated, while mmu\_circ\_000287, mmu\_circ\_003037, and mmu\_circ\_001226 were downregulated. Among these, mmu\_circ\_001801 and mmu\_circ\_003037 exhibit high sequence conservation with human orthologs, while mmu\_circ\_001226 and mmu\_circ\_000287 potentially regulate CAC via miRNA-mRNA networks (75). CircHIPK2 acts as a central node in Hippo pathway dysregulation. Through the circHIPK2-EIF4A3 axis, it enhances TAZ translation, promoting colonic epithelial hyperproliferation in both colitis and CRC (76). Three PRKAR2A-derived circRNAs (e.g., circPRKAR2A-1/2), sharing high homology with murine

mmu\_circ\_0001845, are markedly upregulated in CAC tissues. These circRNAs accelerate the colitis-to-CAC transition and hold promise as diagnostic biomarkers and therapeutic targets for inflammation-driven malignancies (77) (Table 2).

Emerging evidence underscores the central role of circRNAs in IBD pathogenesis, involving epithelial barrier integrity, stem cell regulation, immune cell activity, and regulation of autophagy and inflammatory response. Furthermore, circRNAs are also associated with the progression of intestinal fibrosis and Colitis-associated Cancer, offering new perspectives for the diagnosis and treatment of IBD (Figure 3).

# 6 CircRNAs as potential biomarkers for diagnosis and prognosis of IBD

While endoscopy with biopsy remains the primary and most effective diagnostic modality for IBD due to the absence of a gold standard. Epigenetic biomarkers, particularly in tissue, biofluids (serum, plasma), and exosomes are revolutionizing disease management through non-invasive, cost-efficient monitoring of diagnosis, prognosis, and treatment response.

CircRNAs emerge as superior biomarkers owing to their structural stability, evolutionary conservation across species, tissue-/phase-specific expression patterns, high abundance in bodily fluids, and can be detectable via qRT-PCR (Figure 4). Studies have pointed out that specific circRNAs may serve as biomarkers for IBD. For example, in PBMCs of CD patients, circ\_004662, circ\_092520, circ\_102610, and circ\_103124 are significantly elevated in CD patients. Notably, circ\_004662 achieves an AUC of 0.85 for distinguishing CD from UC (29). circ\_103516 levels in PBMCs correlate positively with CD activity indices (CDAI) (21). Drives UC progression via the miR-1236-3p/ MYD88 axis, serving as a dual potential diagnostic and therapeutic target.

Circ\_0001187, through interaction with the miRNA-1236-3p/ MYD88 pathway, promotes the development of UC and is a potential diagnostic and therapeutic biomarker (28).circNlgn173 is upregulated in active UC mucosa (78). Additionally, circ\_0520, circ\_2243, circ\_0062142, and circ\_0001666 have also been identified for their diagnostic potential in IBD (32, 51).

CircRNAs can also be used for prognosis assessment. For example, circ\_0085323 and circ\_0036906 predict clinical outcomes of UC (79).

# 7 Therapeutic targeting of circRNAs in inflammatory bowel disease: emerging strategies

CircRNAs are increasingly recognized as dynamic regulators of IBD pathogenesis, with their dysregulation influencing transcriptional networks and disease progression. This section delineates cutting-edge approaches for circRNA modulation, siRNA or shRNA silencing, overexpression, and innovative delivery systems.

# 7.1 RNA interference-mediated circRNA knockdown

RNA interference technology achieves the knockdown of circRNAs through small interfering RNA (siRNA) or short hairpin RNA (shRNA). For example, Silencing circ\_0085323 via siRNA suppresses TNF- $\alpha$ -induced epithelial damage by derepressing miR-495-3p/TRAF3 signaling, demonstrating therapeutic efficacy for UC (80). ShRNA-mediated inhibition of hsa\_circ\_103124 enhances autophagy, reduces NLRP3 inflammasome activation, and mitigates intestinal inflammation in CD through miR-650 sequestration, making it a potential therapeutic target (52, 53).

### 7.2 Viral vector-mediated circRNA overexpression

CircRNA expression vectors exploit endogenous back-splicing mechanisms driven by complementary intronic sequences and

TABLE 2 CircRNA and colitis-associa	ted cancer.
-------------------------------------	-------------

Disease	circRNA	miRNA	Source	Up/Down	mechanism	Ref
	mmu_circ_001801		Mice colonic tissuess	Up	involved in CAC	(75)
	mmu_circ_002987			Up		
	mmu_circ_001155			Up		
	mmu_circ_000287	Not report		Down		
AOM/DSS-induced colon carcinoma	mmu_circ_003037			Down		
	mmu_circ_001226			Down		
	circHIPK2			Up		(76)
	circPRKAR2A-1/2			Up		(77)
	mmu_circ_0001845			Up		

AOM azoxymethane,DSS dextran sulfate sodium,CAC colitis-associated cancer;



RNA-binding proteins (RBPs). Lentivirus and adenovirus vectors are commonly used for *in vivo* overexpression of circRNAs (81). For example, in UC, restoration of circRNA CCND1 via lentiviral vectors inhibits LPS-induced apoptosis in Caco-2 cells and mitigates inflammatory responses, preserving epithelial integrity (35). These findings provide experimental evidence for the development of circRNA-based targeted therapies for UC.

### 7.3 Biologics targeting circRNAinflammatory crosstalk

Biological agents, such as infliximab, a chimeric monoclonal antibody of human and mouse origin, are TNF- $\alpha$  inhibitors. They are widely used in treating autoimmune diseases like IBD, ankylosing spondylitis, and psoriatic arthritis, and are known for their high efficacy, safety, and low incidence of side effects (82). Infliximab can reverses circ\_103765 upregulation in CD mucosa, suggesting synergy between biologics and circRNA modulation (27). However, Clinical efficacy in circRNA-high IBD requires validation through multicenter trials.

### 7.4 Poly (lactic-co-glycolic acid)microspheres (PLGA MSs) delivery systems

PLGA MSs enhance low-dose drug bioavailability and enable colon-targeted circRNA delivery. PLGA MSs loaded with circGMCL1 mimics restore autophagy and inhibit pyroptosis via the miR-124-3p/ANXA7 axis, alleviating barrier dysfunction in CD (58). In animal model experiments, PLGA MSs-si-circSMAD4 treatment reduces colitis severity, evidenced by longer colon length and lower histologic inflammation scores, compared to the untreated control group (33).

### 7.5 Nanocarrier/exosome platforms

Research reports indicate that circRNA vaccines provide strong protection against SARS-CoV-2 in mice and rhesus monkeys (83). CircRNAs can be encapsulated and delivered via nanocarriers and exosomes, which may enhance the efficacy of therapeutic drugs (84, 85). However, whether circRNAs can be clinically translated for the treatment of IBD in the medical field requires further investigation.



UC and CD. CircRNA expression levels in serum can serve as an ideal biomarker for the diagnosis of IBD and provide clinicians with a basis for developing appropriate therapeutic regimens. CircRNA levels in blood and serum are also important clinical markers for IBD monitoring, treatment and prognosis.

### 8 Discussion and future perspectives

Recent advances have elucidated the multifaceted roles of circRNAs in IBD, offering novel insights into disease pathogenesis and unveiling potential therapeutic avenues. CircRNAs exhibit dysregulated expression in intestinal tissues, PBMCs, and exosomes of patients with UC and CD (28-30). These molecules orchestrate IBD progression through diverse mechanisms: acting as miRNA sponges (e.g., circ\_103516/miR-19b-1-5p axis), recruiting RNA-binding proteins (RBPs;e.g., HuR/circPABPN1/ATG16L1axis), and epigenetic modulation(e.g.,circPRKAR1B) (47, 65, 66). Such interactions drive pathological cascades, including driving inflammation, intestinal barrier dysfunction, intestinal immune homeostasis, autophagy, EMT, intestinal fibrosis, and Colitis-associated cancer. Notably, phase-specific circRNA signatures (e.g., elevated circ\_103516 in active IBD vs. remission) highlight their utility as non-invasive biomarkers for disease monitoring (21). Furthermore, circRNA-based therapeutic strategies such as PLGA-microsphere-mediated delivery and RNA interference-based silencing can attenuate colitis severity in murine models.

The mechanistic roles of circRNAs in IBD is gaining significant attention. Although its mechanisms are not yet fully understood, its key role in various diseases and potential as biomarkers have already become evident. Despite progress, critical gaps hinder clinical translation.

Current RNA sequencing (RNA-seq) struggles to differentiate circRNAs from linear isoforms due to sequence overlap, while

ribosome profiling fails to conclusively confirm their translational potential (86). Bioinformatics tools(e.g., CIRI, CIRCexplorer) exhibit reduced sensitivity in low-abundance biofluids(e.g., fecal or serum samples) (87).

While circRNAs (e.g., circAtp9b, circ\_103516, circ\_103124) correlate with IBD activity, their expression in non-IBD gastrointestinal disorders (e.g., infectious or allergic colitis) remains unvalidated. Large-scale multicenter studies are imperative to establish diagnostic thresholds and address confounding variables (e.g., microbiome composition, and medication effects). Additionally, circRNA profiles in urine, saliva, and stool, which are key non-invasive sources, are still lacking comprehensive characterization, thus limiting their diagnostic utility.

Systemic delivery of circRNA mimics or inhibitors faces challenges, including nuclease degradation, off-target effects, nonspecific tissue or cell type targeting, Mis-spliced products, and so on (83). Innovations in nanocarriers (e.g., exosome-based delivery) or engineered circRNA may enhance local bioavailability (85, 88). Immunogenicity risks of viral vectors (e.g., adenovirus) require careful evaluation. Non-viral delivery systems, such as lipid nanoparticles represent promising alternatives (84). Despite challenges in the use of circRNA drug delivery systems for the treatment of IBD, their potential in precision medicine and disease management should not be overlooked.

Breakthroughs in high-throughput sequencing and bioinformatic analytics have revolutionized our understanding of circRNAs, unveiling their previously unrecognized functionalities and establishing them as pivotal targets in contemporary biomedical research. Combining circRNA profiling with epigenomic, proteomic, and metabolomic datasets will unravel IBD complex regulatory networks. Single-cell and spatial transcriptomics can resolve cell type-specific circRNA functions within inflammatory niches. CRISPR-based tools (e.g., Cas9, Cas13d) enable targeted circRNA knockdown *in vivo*, offering a platform to validate therapeutic targets (89, 90). However, the long-term safety evaluation of circRNA-based therapeutics must prioritize the following risks:(1) Preventing unintended genomic integration of exogenous circRNAs, which may induce mutagenesis or carcinogenesis. (2) Mitigating excessive immune activation triggered by immunostimulatory circRNA motifs. (3) Minimizing prolonged retention in clearance organs (e.g., liver, kidneys) to avoid dose-dependent toxicity and organ damage.

### 9 Conclusions

This review systematically delineates the expression profiles and mechanistic roles of circRNAs in IBD pathogenesis. It underscores their dual utility as non-invasive diagnostic and prognostic biomarkers and therapeutic targets via modulation of circRNAdriven regulatory networks. Future breakthroughs in IBD therapeutics will rely on concerted interdisciplinary efforts to orchestrate circRNA-targeted strategies to transform molecular insights into clinically actionable solutions, thereby advancing personalized medicine frameworks for IBD.

### Author contributions

LY: Conceptualization, Data curation, Formal Analysis, Investigation, Writing – original draft. HHL: Formal Analysis, Writing – review & editing. MT: Formal Analysis, Writing –

### References

1. Ananthakrishnan AN, Kaplan GG, Ng SC. Changing global epidemiology of inflammatory bowel diseases: sustaining health care delivery into the 21st century. *Clin gastroenterol hepatol.* (2020) 18:1252–60. doi: 10.1016/j.cgh.2020.01.028

2. Desai D, Dhoble P. Rapidly changing epidemiology of inflammatory bowel disease: Time to gear up for the challenge before it is too late. *Indian J gastroenterol.* (2024) 43:15–7. doi: 10.1007/s12664-023-01453-6

3. Fumery M, Yzet C, Chatelain D, Yzet T, Brazier F, LeMouel JP, et al. Colonic strictures in inflammatory bowel disease: epidemiology, complications, and management. J Crohn's colitis. (2021) 15:1766–73. doi: 10.1093/ecco-jcc/jjab068

4. Yao RW, Wang Y, Chen LL. Cellular functions of long noncoding RNAs. Nat Cell Biol. (2019) 21:542–51. doi: 10.1038/s41556-019-0311-8

5. Pourmehran Y, Sadri F, Hosseini SF, Mohammadi Y, Rezaei Z. Exploring the influence of non-coding RNAs on NF-κB signaling pathway regulation in ulcerative colitis. *Biomed pharmacother* = *Biomed pharmacother*. (2024) 179:117390. doi: 10.1016/j.biopha.2024.117390

6. Hu Y, Lu Y, Fang Y, Zhang Q, Zheng Z, Zheng X, et al. Role of long non-coding RNA in inflammatory bowel disease. *Front Immunol.* (2024) 15:1406538. doi: 10.3389/fimmu.2024.1406538

7. Lin L, Zhou G, Chen P, Wang Y, Han J, Chen M, et al. Which long noncoding RNAs and circular RNAs contribute to inflammatory bowel disease? *Cell Death Dis.* (2020) 11:456. doi: 10.1038/s41419-020-2657-z

review & editing. LNH: Supervision, Validation, Writing – review & editing. LJY: Supervision, Writing – review & editing, Validation.

### Funding

The author(s) declare that no financial support was received for the research and/or publication of this article.

### Acknowledgments

Figures in this review were created with BioGDP.com. Retrieved from https://biogdp.com/.

### Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

### **Generative AI statement**

The author(s) declare that no Generative AI was used in the creation of this manuscript.

### Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

8. Liu CX, Chen LL. Circular RNAs: Characterization, cellular roles, and applications. Cell. (2022) 185:2016–34. doi: 10.1016/j.cell.2022.04.021

9. Aufiero S, Reckman YJ, Pinto YM, Creemers EE. Circular RNAs open a new chapter in cardiovascular biology. *Nat Rev Cardiol.* (2019) 16:503–14. doi: 10.1038/s41569-019-0185-2

10. Vromman M, Vandesompele J, Volders PJ. Closing the circle: current state and perspectives of circular RNA databases. *Briefings Bioinf.* (2021) 22:288–97. doi: 10.1093/bib/bbz175

11. Li J, Wang X. Functional roles of conserved lncRNAs and circRNAs in eukaryotes. *Non-coding RNA Res.* (2024) 9:1271-9. doi: 10.1016/j.ncrna.2024.06.014

12. Ma Y, Wang T, Zhang X, Wang P, Long F. The role of circular RNAs in regulating resistance to cancer immunotherapy: mechanisms and implications. *Cell Death dis.* (2024) 15:312. doi: 10.1038/s41419-024-06698-3

13. García-Rodríguez JL, Korsgaard U, Ahmadov U, Jarlstad Olesen MT, Dietrich KG, Hansen EB, et al. Spatial profiling of circular RNAs in cancer reveals high expression in muscle and stromal cells. *Cancer Res.* (2023) 83:3340–53. doi: 10.1158/0008-5472.CAN-23-0748

14. Mei X, Chen SY. Circular RNAs in cardiovascular diseases. *Pharmacol Ther.* (2022) 232:107991. doi: 10.1016/j.pharmthera.2021.107991

15. Zhao RJ, Zhang WY, Fan XX. Circular RNAs: Potential biomarkers and therapeutic targets for autoimmune diseases. *Heliyon*. (2024) 10:e23694. doi: 10.1016/j.heliyon.2023.e23694

16. Xiao F, He Z, Wang S, Li J, Fan X, Yan T, et al. Regulatory mechanism of circular RNAs in neurodegenerative diseases. *CNS Neurosci Ther.* (2024) 30:e14499. doi: 10.1111/cns.14499

17. Kim E, Kim YK, Lee SV. Emerging functions of circular RNA in aging. *Trends gene: TIG.* (2021) 37:819–29. doi: 10.1016/j.tig.2021.04.014

18. Jagtap U, Anderson ES, Slack FJ. The emerging value of circular noncoding RNA research in cancer diagnosis and treatment. *Cancer Res.* (2023) 83:809–13. doi: 10.1158/0008-5472.CAN-22-3014

19. Liu Q, Li S. Exosomal circRNAs: Novel biomarkers and therapeutic targets for urinary tumors. *Cancer lett.* (2024) 588:216759. doi: 10.1016/j.canlet.2024.216759

20. Lin Z, Ji Y, Zhou J, Li G, Wu Y, Liu W, et al. Exosomal circRNAs in cancer: Implications for therapy resistance and biomarkers. *Cancer lett.* (2023) 566:216245. doi: 10.1016/j.canlet.2023.216245

21. Ye YL, Yin J, Hu T, Zhang LP, Wu LY, Pang Z. Increased circulating circular RNA\_103516 is a novel biomarker for inflammatory bowel disease in adult patients. *World J gastroenterol.* (2019) 25:6273–88. doi: 10.3748/wjg.v25.i41.6273

22. Wang T, Chen N, Ren W, Liu F, Gao F, Ye L, et al. Integrated analysis of circRNAs and mRNAs expression profile revealed the involvement of hsa\_circ\_0007919 in the pathogenesis of ulcerative colitis. *J gastroenterol.* (2019) 54:804–18. doi: 10.1007/s00535-019-01585-7

23. Qiu C, Chen Y, Xia H, Duan J, Zhang L, Zhang Y, et al. Hsa\_circ\_0004662 accelerates the progression of ulcerative colitis via the microRNA-532/HMGB3 signalling axis. *J Cell Mol Med.* (2025) 29(6):e70430. doi: 10.1111/jcmm.70430

24. Lun J, Guo J, Yu M, Zhang H, Fang J. Circular RNAs in inflammatory bowel disease. *Front Immunol.* (2023) 14:1307985. doi: 10.3389/fimmu.2023.1307985

25. Yi Q, Feng J, Lan W, Shi H, Sun W, Sun W. CircRNA and lncRNA-encoded peptide in diseases, an update review. *Mol cancer*. (2024) 23:214. doi: 10.1186/s12943-024-02131-7

26. Li F, Fu J, Fan L, Lu S, Zhang H, Wang X, et al. Overexpression of circAtp9b in ulcerative colitis is induced by lipopolysaccharides and upregulates PTEN to promote the apoptosis of colonic epithelial cells. *Exp Ther med.* (2021) 22:1404. doi: 10.3892/etm.2021.10840

27. Ye Y, Zhang L, Hu T, Yin J, Xu L, Pang Z, et al. CircRNA\_103765 acts as a proinflammatory factor via sponging miR-30 family in Crohn's disease. *Sci reports*. (2021) 11:565. doi: 10.1038/s41598-020-80663-w

28. Ouyang W, Wu M, Wu A, Xiao H. Circular RNA\_0001187 participates in the regulation of ulcerative colitis development via upregulating myeloid differentiation factor 88. *Bioengineered*. (2022) 13:12863–75. doi: 10.1080/21655979.2022.2077572

29. Yin J, Hu T, Xu L, Li P, Li M, Ye Y, et al. Circular RNA expression profile in peripheral blood mononuclear cells from Crohn disease patients. *Medicine*. (2019) 98: e16072. doi: 10.1097/MD.00000000016072

30. Xiao L, Ma XX, Luo J, Chung HK, Kwon MS, Yu TX, et al. Circular RNA circHIPK3 promotes homeostasis of the intestinal epithelium by reducing microRNA 29b function. *Gastroenterology*. (2021) 161:1303–17.e3. doi: 10.1053/j.gastro.2021.05.060

31. Qiao YQ, Cai CW, Shen J, Zheng Q, Ran ZH. Circular RNA expression alterations in colon tissues of Crohn's disease patients. *Mol Med reports.* (2019) 19:4500-6. doi: 10.3892/mmr.2019.10070

32. Hu YA, Zhu Y, Liu G, Yao X, Yan X, Yang Y, et al. Expression profiles of circular RNAs in colon biopsies from Crohn's disease patients by microarray analysis. *J Clin Lab anal.* (2021) 35:e23788. doi: 10.1002/jcla.23788

33. Zhao J, Lin Z, Ying P, Zhao Z, Yang H, Qian J, et al. circSMAD4 promotes experimental colitis and impairs intestinal barrier functions by targeting janus kinase 2 through sponging miR-135a-5p. *J Crohn's colitis*. (2023) 17:593–613. doi: 10.1093/ecco-jcc/jjac154

34. Guo H, Zhang J, Jiang Z, Zhu X, Yang J, Mu R, et al. Noncoding RNA circBtnl1 suppresses self-renewal of intestinal stem cells via disruption of Atf4 mRNA stability. *EMBO J.* (2023) 42:e112039. doi: 10.15252/embj.2022112039

35. Xiang P, Ge T, Zhou J, Zhang Y. Protective role of circRNA CCND1 in ulcerative colitis via miR-142-5p/NCOA3 axis. *BMC gastroenterol.* (2023) 23:18. doi: 10.1186/s12876-023-02641-6

36. Li B, Li Y, Li L, Yu Y, Gu X, Liu C, et al. Hsa\_circ\_0001021 regulates intestinal epithelial barrier function via sponging miR-224-5p in ulcerative colitis. *Epigenomics*. (2021) 13:1385–401. doi: 10.2217/epi-2021-0230

37. Xu Y, Tian Y, Li F, Wang Y, Yang J, Gong H, et al. Circular RNA HECTD1 mitigates ulcerative colitis by promoting enterocyte autophagy via miR-182-5p/huR axis. *Inflamm bowel dis.* (2022) 28:273–88. doi: 10.1093/ibd/izab188

38. Rankin CR, Lokhandwala ZA, Huang R, Pekow J, Pothoulakis C, Padua D. Linear and circular CDKN2B-AS1 expression is associated with Inflammatory Bowel Disease and participates in intestinal barrier formation. *Life Sci.* (2019) 231:116571. doi: 10.1016/j.lfs.2019.116571

39. Ananthakrishnan AN, Bernstein CN, Iliopoulos D, Macpherson A, Neurath MF, Ali RAR, et al. Environmental triggers in IBD: a review of progress and evidence. *Nat Rev Gastroenterol hepatol.* (2018) 15:39–49. doi: 10.1038/nrgastro.2017.136

40. Glassner KL, Abraham BP, Quigley EMM. The microbiome and inflammatory bowel disease. J Allergy Clin Immunol. (2020) 145:16–27. doi: 10.1016/j.jaci.2019.11.003

41. Geremia A, Biancheri P, Allan P, Corazza GR, Di Sabatino A. Innate and adaptive immunity in inflammatory bowel disease. *Autoimmun Rev.* (2014) 13:3–10. doi: 10.1016/j.autrev.2013.06.004

42. Larabi A, Barnich N, Nguyen HTT. New insights into the interplay between autophagy, gut microbiota and inflammatory responses in IBD. *Autophagy*. (2020) 16:38–51. doi: 10.1080/15548627.2019.1635384

43. Holmberg FE, Seidelin JB, Yin X, Mead BE, Tong Z, Li Y, et al. Culturing human intestinal stem cells for regenerative applications in the treatment of inflammatory bowel disease. *EMBO Mol med.* (2017) 9:558–70. doi: 10.15252/emmm.201607260

44. Misir S, Wu N, Yang BB. Specific expression and functions of circular RNAs. Cell Death different. (2022) 29:481–91. doi: 10.1038/s41418-022-00948-7

45. Qu S, Yang X, Li X, Wang J, Gao Y, Shang R, et al. Circular RNA: A new star of noncoding RNAs. *Cancer lett.* (2015) 365:141–8. doi: 10.1016/j.canlet.2015.06.003

46. Ashrafizadeh M, Zarrabi A, Mostafavi E, Aref AR, Sethi G, Wang L, et al. Noncoding RNA-based regulation of inflammation. *Semin Immunol.* (2022) 59:101606. doi: 10.1016/j.smim.2022.101606

47. Cheng X, Zhang X, Su J, Zhang Y, Zhou W, Zhou J, et al. miR-19b downregulates intestinal SOCS3 to reduce intestinal inflammation in Crohn's disease. *Sci reports.* (2015) 5:10397. doi: 10.1038/srep10397

48. Zhao J, Lu Q, Liu Y, Shi Z, Hu L, Zeng Z, et al. Th17 cells in inflammatory bowel disease: cytokines, plasticity, and therapies. *J Immunol Res.* (2021) 2021:8816041. doi: 10.1155/2021/8816041

49. Nata T, Fujiya M, Ueno N, Moriichi K, Konishi H, Tanabe H, et al. MicroRNA-146b improves intestinal injury in mouse colitis by activating nuclear factor- $\kappa$ B and improving epithelial barrier function. *J Gene Med.* (2013) 15:249–60. doi: 10.1002/jgm.v15.6-7

50. Xu XM, Zhang HJ. miRNAs as new molecular insights into inflammatory bowel disease: Crucial regulators in autoimmunity and inflammation. *World J gastroenterol.* (2016) 22:2206–18. doi: 10.3748/wjg.v22.i7.2206

51. Feng S, Xu Z, Zhang Z, Mo Y, Deng Y, Li L, et al. RNA-Seq approach to investigate the effects of melatonin on bone marrow-derived dendritic cells from dextran sodium sulfate-induced colitis mice. *Toxicology*. (2022) 481:153354. doi: 10.1016/j.tox.2022.153354

52. Yin J, Hu T, Xu L, Zhang L, Zhu J, Ye Y, et al. Hsa\_circRNA\_103124 upregulation in Crohn's disease promoted macrophage M1 polarization to maintain an inflammatory microenvironment via activation of the AKT2 and TLR4/NF-κB pathways. *Int immunopharmacol.* (2023) 123:110763. doi: 10.1016/j.intimp.2023.110763

53. Yin J, Hu T, Xu LJ, Zhang LP, Ye YL, Pang Z. The mechanism by which hsa\_circRNA\_103124 highly expressed in peripheral blood of patients with active Crohn's disease regulates macrophage differentiation, pyroptosis and inflammation. *Zhonghua yi xue za zhi.* (2023) 103:3478–86. doi: 10.3760/cma.j.cn112137-20231007-00646

54. Chen Q, Mang G, Wu J, Sun P, Li T, Zhang H, et al. Circular RNA circSnx5 Controls Immunogenicity of Dendritic Cells through the miR-544/SOCS1 Axis and PU.1 Activity Regulation. *Mol Ther*. (2020) 28:2503–18. doi: 10.1016/j.ymthe.2020.07.001

55. Wang TT, Han Y, Gao FF, Ye L, Zhang YJ. Effects of circular RNA circ-SOD2 on intestinal epithelial barrier and ulcerative colitis. *Beijing da xue xue bao Yi xue ban = J Peking Univ Health Sci.* (2019) 51:805–12. doi: 10.19723/j.issn.1671-167X.2019.05.003

56. Ye G, Zhang J, Peng J, Zhou Z, Wang W, Yao S. CircSOD2: Disruption of intestinal mucosal barrier function in ulcerative colitis by regulating the miR-378g/ Snail1 axis. *J gastroenterol hepatol.* (2024) 39:1299–309. doi: 10.1111/jgh.16550

57. Levine B, Mizushima N, Virgin HW. Autophagy in immunity and inflammation. *Nature*. (2011) 469:323–35. doi: 10.1038/nature09782

58. Zhao J, Sun Y, Yang H, Qian J, Zhou Y, Gong Y, et al. PLGA-microspherescarried circGMCL1 protects against Crohn's colitis through alleviating NLRP3 inflammasome-induced pyroptosis by promoting autophagy. *Cell Death dis.* (2022) 13:782. doi: 10.1038/s41419-022-05226-5

59. Ardali R, Kazemipour N, Nazifi S, Bagheri Lankarani K, Razeghian Jahromi I, Sepehrimanesh M. Pathophysiological role of Atg5 in human ulcerative colitis. *Intest Res.* (2020) 18:421–9. doi: 10.5217/ir.2019.00120

60. Noda NN. Structural view on autophagosome formation. FEBS lett. (2024) 598:84–106. doi: 10.1002/1873-3468.14742

61. Zhu P, Zhu X, Wu J, He L, Lu T, Wang Y, et al. IL-13 secreted by ILC2s promotes the self-renewal of intestinal stem cells through circular RNA circPan3. *Nat Immunol.* (2019) 20:183–94. doi: 10.1038/s41590-018-0297-6

62. Mukherjee N, Corcoran DL, Nusbaum JD, Reid DW, Georgiev S, Hafner M, et al. Integrative regulatory mapping indicates that the RNA-binding protein HuR couples pre-mRNA processing and mRNA stability. *Mol Cell.* (2011) 43:327–39. doi: 10.1016/ j.molcel.2011.06.007

63. Noh JH, Kim KM, Abdelmohsen K, Yoon JH, Panda AC, Munk R, et al. HuR and GRSF1 modulate the nuclear export and mitochondrial localization of the lncRNA RMRP. *Genes Dev.* (2016) 30:1224–39. doi: 10.1101/gad.276022.115

64. Palanisamy K, Tsai TH, Yu TM, Sun KT, Yu SH, Lin FY, et al. RNA-binding protein, human antigen R regulates hypoxia-induced autophagy by targeting ATG7/

ATG16L1 expressions and autophagosome formation. J Cell Physiol. (2019) 234:7448– 58. doi: 10.1002/jcp.27502

65. Li XX, Xiao L, Chung HK, Ma XX, Liu X, Song JL, et al. Interaction between huR and circPABPN1 modulates autophagy in the intestinal epithelium by altering ATG16L1 translation. *Mol Cell Biol.* (2020) 40(6):e00492-19. doi: 10.1128/MCB.00492-19

66. Zhao J, Zhao Z, Ying P, Zhou Y, Xu Z, Wang H, et al. METTL3-mediated m(6) A modification of circPRKAR1B promotes Crohn's colitis by inducing pyroptosis via autophagy inhibition. *Clin Trans med.* (2023) 13:e1405. doi: 10.1002/ctm2.1405

67. Chen Y, Lin Y, Shu Y, He J, Gao W. Interaction between N(6)-methyladenosine (m(6)A) modification and noncoding RNAs in cancer. *Mol cancer*. (2020) 19:94. doi: 10.1186/s12943-020-01207-4

68. Liu J, Yue Y, Han D, Wang X, Fu Y, Zhang L, et al. A METTL3-METTL14 complex mediates mammalian nuclear RNA N6-adenosine methylation. *Nat Chem Biol.* (2014) 10:93–5. doi: 10.1038/nchembio.1432

69. Yang L, Wu G, Wu Q, Peng L, Yuan L. METTL3 overexpression aggravates LPSinduced cellular inflammation in mouse intestinal epithelial cells and DSS-induced IBD in mice. *Cell Death discov*. (2022) 8:62. doi: 10.1038/s41420-022-00849-1

70. Yang Y, Hsu PJ, Chen YS, Yang YG. Dynamic transcriptomic m(6)A decoration: writers, erasers, readers and functions in RNA metabolism. *Cell Res.* (2018) 28:616–24. doi: 10.1038/s41422-018-0040-8

71. Yoo JH, Holubar S, Rieder F. Fibrostenotic strictures in Crohn's disease. Intest Res. (2020) 18:379–401. doi: 10.5217/ir.2019.09148

72. Cherubini A, Barilani M, Rossi RL, Jalal MMK, Rusconi F, Buono G, et al. FOXP1 circular RNA sustains mesenchymal stem cell identity via microRNA inhibition. *Nucleic Acids Res.* (2019) 47:5325-40. doi: 10.1093/nar/gkz199

73. Yin J, Ye YL, Hu T, Xu LJ, Zhang LP, Ji RN, et al. Hsa\_circRNA\_102610 upregulation in Crohn's disease promotes transforming growth factor- $\beta$ 1-induced epithelial-mesenchymal transition via sponging of hsa-miR-130a-3p. *World J gastroenterol.* (2020) 26:3034–55. doi: 10.3748/wjg.v26.i22.3034

74. Shah SC, Itzkowitz SH. Colorectal cancer in inflammatory bowel disease: mechanisms and management. *Gastroenterology*. (2022) 162:715–30.e3. doi: 10.1053/j.gastro.2021.10.035

75. Yuan G, Chen T, Zhang H, Cao Q, Qiu Y, Que B, et al. Comprehensive analysis of differential circular RNA expression in a mouse model of colitis-induced colon carcinoma. *Mol carcinogen*. (2018) 57:1825–34. doi: 10.1002/mc.v57.12

76. Zeng X, Tang J, Zhang Q, Wang C, Qi J, Wei Y, et al. CircHIPK2 contributes cell growth in intestinal epithelial of colitis and colorectal cancer through promoting TAZ translation. *Adv Sci (Weinheim Baden-Wurttemberg Germany).* (2024) 11:e2401588. doi: 10.1002/advs.202401588

77. Wan D, Wang S, Xu Z, Zan X, Liu F, Han Y, et al. PRKAR2A-derived circular RNAs promote the Malignant transformation of colitis and distinguish patients with colitis-associated colorectal cancer. *Clin Trans med.* (2022) 12:e683. doi: 10.1002/ctm2.v12.2

78. Du WW, Zhou C, Yang H, Wen S, Chen Y, Chen EX, et al. Aggravated Ulcerative Colitis via circNlgn-Mediated Suppression of Nuclear Actin Polymerization. *Res (Washington DC).* (2024) 7:0441. doi: 10.34133/research.0441

79. Yuan YY, Wu H, Chen QY, Fan H, Shuai B. Construction of the underlying circRNA-miRNA-mRNA regulatory network and a new diagnostic model in ulcerative colitis by bioinformatics analysis. *World J Clin cases*. (2024) 12:1606–21. doi: 10.12998/wjcc.v12.i9.1606

80. An Q, Yang S, Tao J, Yang M, Ma Z, Gao Q, et al. CIRC\_0085323 SILENCING INHIBITS TNF-A-INDUCED NORMAL HUMAN COLONIC EPITHELIAL CELL INFLAMMATION AND APOPTOSIS THROUGH THE MIR-495-3P/TRAF3 AXIS. Shock (Augusta Ga). (2023) 60:298–305. doi: 10.1097/SHK.00000000002167

81. Meganck RM, Borchardt EK, Castellanos Rivera RM, Scalabrino MI, Wilusz JE, Marzluff WF, et al. Tissue-dependent expression and translation of circular RNAs with recombinant AAV vectors *in vivo. Mol Ther Nucleic Acids.* (2018) 13:89–98. doi: 10.1016/j.omtn.2018.08.008

82. Armstrong AW, Read C. Pathophysiology, clinical presentation, and treatment of psoriasis: A review. *Jama*. (2020) 323:1945-60. doi: 10.1001/jama.2020.4006

83. He AT, Liu J, Li F, Yang BB. Targeting circular RNAs as a therapeutic approach: current strategies and challenges. *Signal transduct target Ther.* (2021) 6:185. doi: 10.1038/s41392-021-00569-5

84. Cui X, Bao L, Wang X, Chen C. The nano-intestine interaction: understanding the location-oriented effects of engineered nanomaterials in the intestine. *Small (Weinheim an der Bergstrasse Germany).* (2020) 16:e1907665. doi: 10.1002/ smll.201907665

85. Shi X, Wang B, Feng X, Xu Y, Lu K, Sun M. circRNAs and exosomes: A mysterious frontier for human cancer. *Mol Ther Nucleic Acids*. (2020) 19:384–92. doi: 10.1016/j.omtn.2019.11.023

86. Guo JU, Agarwal V, Guo H, Bartel DP. Expanded identification and characterization of mammalian circular RNAs. *Genome Biol.* (2014) 15:409. doi: 10.1186/s13059-014-0409-z

87. Jeck WR, Sorrentino JA, Wang K, Slevin MK, Burd CE, Liu J, et al. Circular RNAs are abundant, conserved, and associated with ALU repeats. *RNA (New York NY)*. (2013) 19:141–57. doi: 10.1261/rna.035667.112

88. Wesselhoeft RA, Kowalski PS, Anderson DG. Engineering circular RNA for potent and stable translation in eukaryotic cells. *Nat commun.* (2018) 9:2629. doi: 10.1038/s41467-018-05096-6

89. Huang S, Li X, Zheng H, Si X, Li B, Wei G, et al. Loss of super-enhancer-regulated circRNA nfix induces cardiac regeneration after myocardial infarction in adult mice. *Circulation*. (2019) 139:2857–76. doi: 10.1161/CIRCULATIONAHA.118.038361

90. Toro N, Mestre MR, Martínez-Abarca F, González-Delgado A. Recruitment of reverse transcriptase-cas1 fusion proteins by type VI-A CRISPR-cas systems. *Front microbiol.* (2019) 10:2160. doi: 10.3389/fmicb.2019.02160