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Editorial: Tuberculosis and humoral immunity

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Editorial on the Research Topic

Tuberculosis and humoral immunity

Tuberculosis (TB), caused by *Mycobacterium tuberculosis* (Mtb), remains a formidable global health threat, afflicting nearly one-quarter of the world's population. A complete understanding of the immune components involved in natural protective defenses against Mtb is a crucial step to control this epidemic. Unfortunately, we are still lacking effective therapies and vaccines that confer sterilizing immunity, despite decades of immunology and TB research co-development. The parallel progress of the two sciences allowed historical advances in one area to have great impact on the other and vice versa. For instance, much of what constituted the foundation of cellular immunology was based on Mtb as the ideal model for studying delayed-type hypersensitivity.

As a result, observations derived from the TB field are typically generalized to other areas of immunopathology and vaccinology. One of such generalization is the assumption that T cell responses (a very destructive weapon of the immune system) are the chief protective mechanism against intracellular bacteria. Thus, TB research has emphasized the role of cell-mediated immunity (CMI), particularly CD4+ and CD8+ T cells, in combating Mtb (1–14). However, vaccine candidates designed to promote class one T helper (Th1) cell responses have failed to control this infection (15–17), suggesting that other components of the immune system are essential for protection. Indeed, emerging evidence (18–27), including our own findings (28–30), is reshaping this perspective by underscoring the critical contributions of humoral immunity in host defense against TB.

This editorial synthesizes the current understanding of humoral immunity in TB, its potential to enhance diagnostics and vaccine strategies, and the interplay between cellular and humoral responses. Our aim was to provide a collection of selected manuscripts contributing to clarify the misconception that B cells and antibodies are ineffective against intracellular pathogens like Mtb, an assumption that has been challenged by recent studies demonstrating that humoral immunity is not only relevant but also potentially protective in TB.

A provoking reflection necessary to challenge current dogmas in TB research is the idea of tissues as crucial orchestrators of the class of immune response deployed against invading microorganism (31). As such, the lungs must determine the ultimate combination of effector mechanisms more suitable to ensure bacilli clearance while preserving the local architecture of the tissue. With this in mind, what is the benefit of evoking such a destructive stereotyped response like CMI within a local microenvironment so delicate to allow the vital function of gas exchange? Perhaps other immune components at the site of

encounter with Mtb [including mucosal antibodies and B cells of bronchial-associated lymphoid tissue (BALT)] are better at controlling the infection while preserving the structure of the lungs due to their less harmful effects.

B cells, for instance, can act as antigen-presenting cells, modulate T cell responses, and produce antibodies that neutralize Mtb or mediate effector functions. Notably, B-cell aggregates in Mtb-infected lungs have been correlated with reduced bacterial burden and improved outcomes of TB disease (20, 23, 30). Antibodies have been shown to enhance macrophage uptake and intracellular killing of Mtb via Fc receptor-mediated pathways (18, 32). Furthermore, Mtb-specific IgA and IgG antibodies contribute to mucosal immunity and bacterial clearance in experimental models (33, 34).

The research highlighted in this Research Topic further emphasizes some of these ideas and provide proof-of-concept evidence. First, Flores-Gonzalez et al. demonstrated distinct alterations in B cell subsets and reduced cytokine production, such as IFN- γ and IL-10, in patients with active TB, particularly drug-resistant cases. These findings suggest that phenotypical alterations of B cells may serve as readouts of the failure of lung tissue-instructed defenses to eliminate Mtb. Also, in their review article, McIntyre et al. highlighted increasing evidence supporting the role of antibody-mediated immunity in TB, demonstrating that antibodies contribute to pathogen neutralization, promote phagocytosis, and mediate antibody-dependent cellular cytotoxicity (ADCC). These effector mechanisms of antibodies might be pivotal at the early stages of the disease when first contact with Mtb occurs. Especially during early childhood and adolescence, social factors like breastfeeding, undernutrition, hygiene, poverty, exposure to environment mycobacteria, and respiratory infection history might influence the repertoire of antibodies available in the respiratory tract. Thus, analyzing humoral immunity to TB in distinct age groups is of significant relevance, as here represented by studies on maternal and infant immunity by Hjelm et al., which suggest that antibodies against Mtb antigens, such as lipoarabinomannan (LAM), may offer protection against disease progression during early childhood. Although significant progress has been made, the diversity of TB-specific antibodies presents challenges for vaccine and diagnostic development. Advances in deep-learning models can improve the high-throughput analysis of B cell receptors, revealing maturation pathways and functional diversity, which could improve vaccine design and diagnostics (Lee et al.).

Despite these promising findings, several questions remain. First, what are the mechanisms underlying antibody-mediated

protection against Mtb. Second, what are the antibody-independent roles of B cells in TB? Finally, how can we unravel the intricate crosstalk between humoral and cellular immunity to inform the design of more effective vaccines and therapies?

In conclusion, the role of humoral immunity in TB represents a paradigm shift that challenges the traditional focus on CMI. While Th1 cells remain central to controlling Mtb infection, antibodies, and B cells play complementary roles that cannot be ignored. The integration of humoral responses into TB vaccine design, diagnostics, and therapeutics holds immense potential for improving disease management and outcomes. Future research should prioritize characterizing antibody and B cell repertoires in lung specimens of TB patients, uncovering the mechanisms by which antibodies mediate protection, identifying key antigenic targets, and leveraging these insights to develop novel interventions.

As we continue to unravel the complexities of the immune response to TB, a more comprehensive approach that incorporates both cellular and humoral immunity will be essential. Such an approach not only broadens our understanding of TB pathogenesis but also paves the way for more effective vaccines and therapies, ultimately bringing us closer to the goal of TB eradication.

Author contributions

TD: Conceptualization, Writing – original draft, Writing – review & editing. JC-P: Conceptualization, Writing – original draft, Writing – review & editing.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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References

1. Panda S, Morgan J, Cheng C, Saito M, Gilman RH, Ciobanu N, et al. Identification of differentially recognized T cell epitopes in the spectrum of tuberculosis infection. *Nat Commun.* (2024) 15:765. doi: 10.1038/s41467-024-45058-9
2. Mpande CAM, Rozot V, Mosito B, Musvosvi M, Dintwe OB, Bilek N, et al. Immune profiling of Mycobacterium tuberculosis-specific T cells in recent and remote infection. *eBioMedicine.* (2021) 64:103233. doi: 10.1016/j.ebiom.2021.103233
3. Urdahl KB, Shafiani S, Ernst JD. Initiation and regulation of T-cell responses in tuberculosis. *Mucosal Immunol.* (2011) 4:288–93. doi: 10.1038/mi.2011.10
4. Lu Y-J, Barreira-Silva P, Boyce S, Powers J, Cavallo K, Behar SM. CD4 T cell help prevents CD8 T cell exhaustion and promotes control of Mycobacterium tuberculosis infection. *Cell Rep.* (2021) 36:109696. doi: 10.1016/j.celrep.2021.109696
5. Behar SM, Carpenter SM, Booty MG, Barber DL, Jayaraman P. Orchestration of pulmonary T cell immunity during Mycobacterium tuberculosis infection: immunity interruptus. *Semin Immunol.* (2014) 26:559–77. doi: 10.1016/j.smim.2014.09.003
6. Cardona P, Cardona P-J. Regulatory T cells in mycobacterium tuberculosis infection. *Front Immunol.* (2019) 10:2139. doi: 10.3389/fimmu.2019.02139

7. Coppola M, Villar-Hernández R, van Meijgaarden KE, Latorre I, Muriel Moreno B, Garcia-Garcia E, et al. Cell-Mediated Immune Responses to *in vivo*-Expressed and Stage-Specific Mycobacterium tuberculosis Antigens in Latent and Active Tuberculosis Across Different Age Groups. *Front Immunol.* (2020) 11:103. doi: 10.3389/fimmu.2020.00103
8. Jasenosky LD, Scriba TJ, Hanekom WA, Goldfeld AE. T cells and adaptive immunity to Mycobacterium tuberculosis in humans. *Immunol Rev.* (2015) 264:74–87. doi: 10.1111/immr.12274
9. Winslow GM, Cooper A, Reiley W, Chatterjee M, Woodland DL. Early T-cell responses in tuberculosis immunity. *Immunol Rev.* (2008) 225:10. doi: 10.1111/j.1600-065X.2008.00693.x
10. Orme IM, Henao-Tamayo MI. Trying to See the Forest through the Trees: Deciphering the Nature of Memory Immunity to Mycobacterium tuberculosis. *Front Immunol.* (2018) 9:461. doi: 10.3389/fimmu.2018.00461
11. Henao-Tamayo M, Ordway DJ, Orme IM. Memory T cell subsets in tuberculosis: what should we be targeting? *Tuberc Edinb Scotl.* (2014) 94:455–61. doi: 10.1016/j.tube.2014.05.001
12. Ordway D, Palanisamy G, Henao-Tamayo M, Smith EE, Shanley C, Orme IM, et al. The cellular immune response to mycobacterium tuberculosis infection in the Guinea pig. *J Immunol.* (2007) 179:2532–41. doi: 10.4049/jimmunol.179.4.2532
13. Yao S, Huang D, Chen CY, Halliday L, Wang RC, Chen ZW. CD4+ T cells contain early extrapulmonary tuberculosis (TB) dissemination and rapid TB progression and sustain multi-effector functions of CD8+ T and CD3- lymphocytes: mechanisms of CD4+ T cell immunity. *J Immunol Baltim Md 1950.* (2014) 192:2120–32. doi: 10.4049/jimmunol.1301373
14. Flynn JL, Goldstein MM, Triebold KJ, Koller B, Bloom BR. Major histocompatibility complex class I-restricted T cells are required for resistance to Mycobacterium tuberculosis infection. *Proc Natl Acad Sci U.S.A.* (1992) 89:12013–7. doi: 10.1073/pnas.89.24.12013
15. Mittrücker H-W, Steinhoff U, Köhler A, Krause M, Lazar D, Mex P, et al. Poor correlation between BCG vaccination-induced T cell responses and protection against tuberculosis. *Proc Natl Acad Sci U.S.A.* (2007) 104:12434–9. doi: 10.1073/pnas.0703510104
16. Kagina BMN, Abel B, Scriba TJ, Hughes EJ, Keyser A, Soares A, et al. Specific T cell frequency and cytokine expression profile do not correlate with protection against tuberculosis after bacillus Calmette-Guérin vaccination of newborns. *Am J Respir Crit Care Med.* (2010) 182:1073–9. doi: 10.1164/rccm.201003-0334OC
17. Fletcher HA, Snowden MA, Landry B, Rida W, Satti I, Harris SA, et al. T-cell activation is an immune correlate of risk in BCG vaccinated infants. *Nat Commun.* (2016) 7:11290. doi: 10.1038/ncomms11290
18. Lu LL, Chung AW, Rosebrock T, Ghebremichael M, Yu WH, Grace PS, et al. A functional role for antibodies in tuberculosis. *Cell.* (2016) 167:433–443.e14. doi: 10.1016/j.cell.2016.08.072
19. Teitelbaum R, Glatman-Freedman A, Chen B, Robbins JB, Unanue E, Casadevall A, et al. A mAb recognizing a surface antigen of Mycobacterium tuberculosis enhances host survival. *Proc Natl Acad Sci U.S.A.* (1998) 95:15688–93. doi: 10.1073/pnas.95.26.15688
20. Phuah JY, Mattila JT, Lin PL, Flynn JL. Activated B cells in the granulomas of nonhuman primates infected with Mycobacterium tuberculosis. *Am J Pathol.* (2012) 181:508–14. doi: 10.1016/j.ajpath.2012.05.009
21. Li H, Javid B. Antibodies and tuberculosis: finally coming of age? *Nat Rev Immunol.* (2018) 18:591–6. doi: 10.1038/s41577-018-0028-0
22. Achkar JM, Chan J, Casadevall A. B cells and antibodies in the defense against Mycobacterium tuberculosis infection. *Immunol Rev.* (2015) 264:167–81. doi: 10.1111/immr.12276
23. Maglione PJ, Xu J, Chan J. B cells moderate inflammatory progression and enhance bacterial containment upon pulmonary challenge with Mycobacterium tuberculosis. *J Immunol Baltim Md 1950.* (2007) 178:7222–34. doi: 10.4049/jimmunol.178.11.7222
24. Kozakiewicz L, Chen Y, Xu J, Wang Y, Dunussi-Joannopoulos K, Ou Q, et al. B cells regulate neutrophilia during Mycobacterium tuberculosis infection and BCG vaccination by modulating the interleukin-17 response. *PloS Pathog.* (2013) 9:e1003472. doi: 10.1371/journal.ppat.1003472
25. Rao M, Valentini D, Poirer T, Dodoo E, Parida S, Zumla A, et al. B in TB: B cells as mediators of clinically relevant immune responses in tuberculosis. *Clin Infect Dis.* (2015) 61:S225–34. doi: 10.1093/cid/civ614
26. Linge I, Dyatlov A, Kondratieva E, Avdienko V, Apt A, Kondratieva T. B-lymphocytes forming follicle-like structures in the lung tissue of tuberculosis-infected mice: Dynamics, phenotypes and functional activity. *Tuberc Edinb Scotl.* (2017) 102:16–23. doi: 10.1016/j.tube.2016.11.005
27. Krause R, Ogongo P, Tezera L, Ahmed M, Mbano I, Chambers M, et al. B cell heterogeneity in human tuberculosis highlights compartment-specific phenotype and functional roles. *Commun Biol.* (2024) 7:1–15. doi: 10.1038/s42003-024-06282-7
28. Choreño-Parra JA, Bobba S, Rangel-Moreno J, Ahmed M, Mehra S, Rosa B, et al. Mycobacterium tuberculosis HN878 infection induces human-like B-cell follicles in mice. *J Infect Dis.* (2020) 221:1636–46. doi: 10.1093/infdis/jiz663
29. Choreño-Parra JA, Weinstein LI, Yunis EJ, Zúñiga J, Hernández-Pando R. Thinking outside the box: innate- and B cell-memory responses as novel protective mechanisms against tuberculosis. *Front Immunol.* (2020) 11:226. doi: 10.3389/fimmu.2020.00226
30. Dutt TS, Karger BR, Fox A, Youssef N, Dadhwal R, Ali MZ, et al. Mucosal exposure to non-tuberculous mycobacteria elicits B cell-mediated immunity against pulmonary tuberculosis. *Cell Rep.* (2022) 41:111783. doi: 10.1016/j.celrep.2022.111783
31. Matzinger P, Kamala T. Tissue-based class control: the other side of tolerance. *Nat Rev Immunol.* (2011) 11:221–30. doi: 10.1038/nri2940
32. Zimmermann N, Thormann V, Hu B, Köhler A-B, Imai-Matsushima A, Loch C, et al. Human isotype-dependent inhibitory antibody responses against Mycobacterium tuberculosis. *EMBO Mol Med.* (2016) 8:1325–39. doi: 10.15252/emmm.201606330
33. Ishida E, Corrigan DT, Chen T, Liu Y, Kim RS, Song L, et al. Mucosal and systemic antigen-specific antibody responses correlate with protection against active tuberculosis in nonhuman primates. *eBioMedicine.* (2024) 99:104897. doi: 10.1016/j.ebiom.2023.104897
34. Li W, Deng G, Li M, Liu X, Wang Y. Roles of Mucosal Immunity against Mycobacterium tuberculosis Infection. *Tuberc Res Treat.* (2012) 2012:791728. doi: 10.1155/2012/791728