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## Overcoming resistant cancerous tumors through combined photodynamic and immunotherapy (photoimmunotherapy)

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Cancer is a major health problem as it causes significant mortality globally. In the last decades, conventional and recent therapeutic approaches have been used in oncology for cancer treatment. Despite this, the complete eradication of cancer is challenging, as the existing therapeutic strategies for cancer are typically faced with limitations. This is linked to cancer resistance to treatment, which arises because of the versatile nature of cancerous cells. Novel anticancer therapeutic procedures based on immune system activation, such as photodynamic therapy (PDT) and immunotherapy (IOT), are promising in treating resistant tumors. PDT is a minimally invasive treatment that induces cellular reactive oxygen species (ROS) production for direct elimination of cancerous cells, but can also trigger anticancer effects by activating the immune system of the host. IOT also has significant anticancer efficacy and has emerged as an advanced anticancer treatment that mainly enhances and stimulates the innate immune system of the body to identify and destroy cancerous cells. IOT can also instigate a longlasting anticancer response by harnessing the body's immune system. PDT and IOT, when used alone, cannot tackle the issue of cancer resistance. This review elucidates the principles, benefits, and setbacks of PDT and IOT, along with the unique attributes that render them suitable for cancer combination therapy. It underscores the advancement of cancer PDT when utilized in combination with IOT (photoimmunotherapy), while also encapsulating the preclinical evidence regarding the efficacy of photoimmunotherapy, and its combination with nanotechnology (Nano-photoimmunotherapy). The key findings indicate that photoimmunotherapy preclinical methods hold great promise in cancer treatment, as they can directly destroy cancer cells through PDT while also stimulating an increased anticancer immunity through co-delivery of IOT agents. Target-specific moieties can be used in nanotechnology-based anticancer photoimmunotherapy techniques to get past resistance and other therapeutic obstacles. However, clinical utilization of photoimmunotherapy procedures is greatly required to warrant the full efficacy.

#### KEYWORDS

cancer, resistant, photodynamic therapy, immunotherapy, photoimmunotherapy, nano-photoimmunotherapy

#### 1 Introduction

Cancer is a serious global health concern, as it is a great cause of mortality globally. It is accountable for about one in six deaths globally and with about 10 million deaths annually. Cancer is a transversal disease that affects both developed and emerging countries across different ethnicities and cultures (1, 2). Even though, an excessive burden is shouldered by the low and middle-income nations as a result of restricted access to early diagnosis and treatment, and increased exposure to risk factors including, unhealthy diets, poor physical activity, tobacco use, and some infections such as hepatitis B and HPV (2). Various treatment strategies are developed for cancer intervention, including surgery, radiotherapy, and chemotherapy (conventional therapy). Other recent strategies used in cancer treatment include targeted therapy, immunotherapy, stem cell therapy, ablation therapy, sonodynamic therapy, chemodynamic therapy, ferroptosis-based therapy, and radionics (3, 4). These modern cancer therapeutic procedures have significantly improved cancer treatment over the past 30 years (5); however, cancer treatment failure is still widespread due to the ineffectiveness of these therapies (6). This is linked to cancer resistance to monotherapies (5). The phenomenon of cancer resistance to treatment commonly arises in clinical practice, thus resulting in poor patient survival. Also, cancerous cells with resistant attributes frequently show crossresistance to different anticancer treatments or drugs that may not be structurally relevant, and this has been attributed to the appellation multidrug resistance (MDR). The MDR phenomenon is a major impeding obstacle to treatment success, with consequential impacts like cancer recurrence and cancer-related death (7).

Cancer treatment resistance can be grouped into acquired and intrinsic resistance depending on the time the resistance arises. Intrinsic cancer resistance is recognized as the primary resistance that originates from endogenous factors existing in tumor cells before any treatment applications. These factors give cancerous cells survival benefits and the potential to adapt to stress from primary therapy (8, 9). On the other hand, cancer-acquired drug resistance is typically mediated by adaptive changes that antagonize cancerous cells' susceptibility to an administered treatment, thus reducing the treatment's efficacy (7, 9).

The reduced response of cancerous cells to treatment is also linked to different mechanisms, most often involving the co-action of genetic and non-genetic factors. Tumor cell genetic factors are identified as major contributors to treatment resistance. These genetic factors include oncogene amplification in bypass or compensatory pathways, acquired drug target mutations, genetic diversity, and changes in epigenetics, which can also affect DNA repair, tumor cell plasticity, intratumor heterogeneity, and tumor cell susceptibility to pathways leading to cell death, hence resulting in multifactor-mediated resistance (7, 10). Yet, cancer drug resistance has been identified in which there is no genetic mutation in patients with various types of cancers (10, 11). The genotype alterations can be independent of the phenotype changes when the resistance is mediated via the metabolic inactivation of cancerous drugs, drug compartmentation, fewer intracellular transporters of drug concentrations, and reversible transcriptional or posttranslational controls on adaptive pathways induced by the drugs (7).

However, combination cancer treatments are stated to increase the likelihood and strength of treatment responses while lowering the probability of treatment resistance being developed in the patient (12). The cornerstone of cancer combination therapy is to target pathways that perpetuate or cause cancer precisely. Combination therapy often works in an additive or synergistic manner, thus requiring a reduced dose of each separate drug. Combination therapy can offer a toxic effect on cancerous cells while preventing damage to healthy cells. This can be materialized if one of the utilized drugs is cytotoxically antagonistic to a different drug within normal cells, thus shielding healthy cells from cytotoxic damage (13). An effective cancer combination therapy might overcome the shortcomings of conventional mono-therapeutic treatment, such as the non-selective targeting of active proliferating cells, which eventually results in the death of both malignant and healthy cells. Monotherapy, like chemotherapy, damages both healthy and cancerous proliferating cells, causing several hazardous effects. It can significantly weaken the immune system of patients by attacking cells in the bone marrow and amplifying the patient's susceptibility to diseases (13, 14).

Nonetheless, in the battle against tumors that are resistant to conventional therapy, novel anticancer therapy based on immune system activation is encouraged. Such therapy includes photodynamic therapy (PDT) and immunotherapy (IOT) (15). An anticancer combination therapy that explores PDT and IOT (photoimmunotherapy) may overcome the issues of cancer resistance since this type of combination therapy is documented

Abbreviations: ATP, adenosine triphosphate; ACT, Adoptive cell therapies; AbPCs, antibody-photosensitizer conjugates; Ags, antigens; BMS-202, (N-{2-[({2-Methoxy-6-[(2-Methyl[1,1'-Biphenyl]-3-Yl)methoxy]pyridin-3-Yl}methyl) amino]ethyl}acetamide); Ce6, chlorin e6; CRT, calreticulin; CAFs, cancerassociated fibroblasts; CARs, chimeric antigen receptors; CTLA-4, cytotoxic Tlymphocyte-associated antigen 4; CuAAC, copper-catalyzed azide-alkyne cycloaddition; DAMPs, damage-associated molecular patterns; DCs, dendritic cells; EpCAM, epithelial cell adhesion molecule; EphA2, ephrin type-A receptor 2; ER, endoplasmic reticulum; EPR, enhanced permeability and retention; FAP, fibroblast-activation protein; GM-CSF, granulocyte macrophage colonystimulating factor; HMGB1, high mobility group box 1; HSPs, heat-shock proteins; ICD, immunogenic cell death; ICIs, immune checkpoint inhibitors; IDO, indoleamine 2,3-dioxygenase; IFNs, interferons; IND, indoximod; IOT, immunotherapy; irAEs, immune-related adverse events; IRDye700, near-IR fluorescent dye; LAG-3, lymphocyte activation gene 3; mAbs, monoclonal antibodies; MDR, multidrug resistance; mTNBC, metastatic triple-negative breast cancer; NK, natural killer; NPs, nanoparticles; OVs, Oncolytic virus; PEG, polyethylene glycol; PD-1, programmed cell death protein 1, PD-L1 programmed death-ligand; PDT, photodynamic therapy; PRRs, pattern recognition receptors; PS, photosensitizer; ROS, reactive oxygen species; TAMs, tumor-associated macrophages; TILs, Tumor-infiltrating lymphocytes; TME, tumor microenvironment; TNFα, tumor necrosis factor alpha; VEGF, vascular endothelial growth Factor.

to prevent tumor metastasis, activate the immune system's memory cells, and stop the recurrence of tumors (16, 17). Besides, photoimmunotherapy in combination with advanced nanotechnology (nano-photoimmunotherapy) is noted to offer a better therapeutic efficacy against resistant cancer (18). In this review, we describe PDT and IOT, highlighting their significance in anticancer immune stimulation. The preclinical evidence on photoimmunotherapy and nano-photoimmunotherapy in combating cancer resistance is also discussed.

#### 2 Photodynamic therapy

PDT is a photochemical therapeutic procedure that utilizes laser irradiation at a defined wavelength to stimulate the transfer of photoelectrons to nearby oxygen molecules, thereby generating singlet oxygen, which is lethal to cancerous cells (19). This therapeutic procedure is less invasive, suitable for treating squamous cell carcinoma, and offers good spatiotemporal selectivity (20). Apart from being directly lethal to cancerous cells, PDT can stimulate inflammatory reactions that encourage the creation of tumor-associated antigens from the remnants of cancerous cells, leading to immunogenic cell death (21).

#### 2.1 Principle of photodynamic therapy

Light irradiation of photosensitizer (PS) stimulates photon absorption and the excitation of the PS to the singlet state  $(S_1)$ , where there is a shift of electrons to an orbital with higher energy (Figure 1). At this state (a short-lived state and typically not stable), the

PS may go back to the ground state  $(S_0)$  through the conversion of its energy to fluorescence or heat. This feature is so handy for applications in monitoring and diagnostic procedures (22). Alternatively, intersystem crossing may happen, resulting in the PS being excited to a triplet state (T1). The PS in the T1 can convey energy via phosphorescence or collide with different molecules, creating reactive chemical species through two kinds of molecular reactions. T<sub>1</sub> PS can also react with different types of solvents or organic substrates to transfer a single proton or electron, forming cation species or radical anions, respectively. Mostly, reactions of the PS with electron donor substrate occur, forming PS that eventually reacts with oxygen, generating radical superoxide anion. This is classified as a type I reaction. Yet, a type II reaction could take place if the PS at T<sub>1</sub> directly reacts with oxygen in its ground state (<sup>3</sup>O<sub>2</sub>) by transferring energy to generate singlet oxygen (<sup>1</sup>O<sub>2</sub>), noted as reactive oxygen species (ROS) (22, 23). The molecular products of PDT, such as superoxide anions and singlet oxygen, will promote cytotoxicity since both products can even react directly with biomolecules like nucleic acids, proteins, and lipids, leading to their degradation (24). The superoxide anions that are created from type I reactions are nondamaging, especially in biological systems. Despite that, these superoxide anions can participate in reactions that generate hydrogen peroxide. A Fenton reaction may also occur where the superoxide anion instead reacts with hydrogen peroxide, forming highly reactive hydroxyl radicals. These radicals are ultimately capable of reducing hydrogen atoms in biomolecules or adding to the side of biomolecules with double bonds. For example, reactions of fatty acids with hydroxyl radicals can generate a hydroxylated product, which is also a radical, thus originating a reaction chain involved in lipid peroxidation and consequently causing damage to the cell (22, 25). Moreover, ROS can directly damage lipids, proteins, and DNA in



Schematic representation of photodynamic therapy's principle. The absorption of light by the photosensitizer (PS) excites it from the ground state  $(S_0)$  to a higher energy orbital  $(S_1 \text{ or } S_2)$  excited state. The PS returns to the original  $S_0$  or excites to the triplet state (T1). Type I reaction (Type I energy transfer) occurs where the PS at T1 reacts with oxygen, leading to ROS production. Type II reaction (Type II energy transfer) may also occur when the PS at T1 reacts directly with oxygen in its ground state ( ${}^{3}O_2$ ) by transferring energy to generate singlet oxygen ( ${}^{1}O_2$ ). The ROS generated induces damage to cancerous cells and death.

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cancerous cells, causing alterations in ion transportation and cellular metabolism and an imbalance in homeostasis. The mitogen-activated protein kinase is stimulated in response to induced oxidative stress. Also, numerous cytokines and mediators involved in cell death processes such as autophagy, necrosis, and/or apoptosis are released. These processes or pathways could occur simultaneously and are not mutually exclusive with the same cell population. Different important parameters could contribute to the occurrence of a particular type of cell death pathway. These parameters include drug dose, PS intracellular distribution, light dose (illumination total time and fluence rate), the cell type being investigated, and the available amount of oxygen (15). Cell death following PDT activation is typically induced via autophagy, necrosis, and/or apoptosis (26-28). However, some newly identified cell death processes are linked with PDT outcomes, including paraptosis (typified with elevated cytoplasmic vacuolization that does not involve nuclear fragmentation or caspase activation) (27, 29), pyroptosis (inflammation-induced cell death) (30), ferroptosis (iron-dependent lipid peroxidation-driven cell death) (31), and necroptosis (controlled cell death mimicking attributes of necrosis and apoptosis) (32). These cell death pathways or programs can each result in the release of damage-associated molecular patterns (DAMPs) or alarmins. The pattern recognition receptors (PRRs) on immune cells are able to recognize DAMPs. Binding of DAMPs to PRRs activates the immune cell and thus immunogenic cell death (ICD) (26).

# 2.2 Photodynamic therapy photosensitizers, localization, and dose to induce ICD

Through the years, photosensitizers (PSs) have been modified to address the setbacks of their predecessors and to enhance the therapeutic efficacy of PDT. This has led to PSs being grouped or divided into various generations, including first, second, third, and even fourth generations of PDT PSs. The first-generation PSs are often natural compounds and hematoporphyrin primary derivatives like porfimer sodium (Photofrin) (33, 34). These PSs are faced with limitations such as poor tissue penetration, being prone to photobleaching, exhibiting poor solubility due to their hydrophobic nature, and long cutaneous photosensitivity (33, 35). The second-generation PSs are designed to solve these limitations. Second-generation PSs can provide decreased skin photosensitivity, reduced tissue accumulation periods, and enhanced light absorption at wavelengths from 650 to 800 nm (36, 37). On the other hand, the third-generation PSs are developed for specific tumor targeting, improving the selectivity and efficacy in intracellular delivery and multimodal therapeutic applications. They can be obtained via the conjugation of second-generation PSs with the targeted molecules or encapsulated in nanomaterials (33, 36). Specifically, the incorporation or conjugation of PSs with nanomaterials has paved the way for their utilization as nanomedicine. The fourth-generation PSs are designed by making use of porous delivery systems, such as metal-organic frameworks and mesoporous silica (34). They are generally considered to provide advanced targeting modes of action or combine many therapeutic modalities (36).

PSs can accumulate to initiate their damaging effects on various cell compartments based on the chemical attributes of the PS. The PS subcellular localization involves various organelles, including mitochondria, endoplasmic reticulum (ER), lysosomes, Golgi apparatus, nucleus, and plasma membrane (38-40). PS localization is vital in determining if the cell death following PDT will be triggered via an immune response. An important prerequisite for ICD is the formation of ROS in the ER. This induces oxidative stress, after which one of the major DAMPs [calreticulin (CRT)] is exposed, causing the activation of the host immune system to fight cancer (26, 41). This is indicative that a successful cancer eradication strategy via PDT-induced ICD will logically require the targeting of the PS within the ER. Studies have established that direct accumulation of hypericin in the ER leads to elevated ROS production and the development of a robust immune response following PDT (41, 42). But not all PSs accumulate or build up in the ER. In order to accumulate in the ER, the PS must possess amphiphilic and hydrophobic characteristics. The PS's charge also influences its ability to accumulate in the ER. Hydrophilic PSs are often found in the lysosomes/endosomes before being dispersed in the cytoplasm. When the PS is directly collected in the ER, the effectiveness of PDT and its immunogenic effects are both increased (41).

However, using PS localized in other cell compartments for PDT might also display immunogenic properties. For instance, PDT immunogenicity was established following the localization of PS in the lysosomes. The immunogenicity was exhibited in mouse fibrosarcoma (MCA205 murine prophylactic tumor vaccination mode) via dendritic cell (DC) maturation, release of DAMPs, and an effective reduction in tumor growth (43). However, a PS can be localized in multiple cellular compartments simultaneously (40, 44). Interestingly, two PSs used at the same time target and harm two cellular compartments simultaneously. For instance, studies confirm the targeting of the mitochondria and the lysosome at the same time using benzoporphyrin derivative (BPD, verteporfin) and N-aspartyl chlorin E6 (NPe6) or photofrin, respectively. This PDT technique sequentially induced photodamage starting from the lysosomal, then to the mitochondrial, leading to higher tumor eradication than using a single PS (45-47). Despite that, it was not clarified whether this method could activate an ICD. Still, the administration of two PSs sequentially to target distinct subcellular compartments might be an encouraging strategy to induce ICD, as more intriguing discoveries may be anticipated.

The administered dose of the PS also plays an instrumental role in the overall treatment outcome. High doses of PS can result in aggregation-induced quenching which causes a reduction in the PS optical properties. The systemic administration of such a high dose of PS could cause abnormal accumulation and distribution, resulting in a phototoxic effect (48). Also, high PS dosage raises the risk of adverse effects such as non-scarring skin lesions, erythema, pain, and the death of healthy cells around the area exposed to light (49, 50). It is therefore crucial for an ideal PS to be chosen for PDT, and this PS should trigger an ICD with the least amount of damage to healthy cells. The PS can also infiltrate healthy cells, so elevated PS doses can promote severe dark toxicity to healthy cells. This can harm different cell types, especially cells in the brain, since morphofunctional abnormalities such as those in the neuron-glial network can cause a drastic malfunction in the central nervous system and exacerbate the patient's situation (51, 52). Recent PDT techniques indicate that elevated doses of PS could be circumvented by employing advanced nanostructure delivery systems. The delivery system can help deposit the PS in the targeted cellular compartment in the tumor tissue, leading to the stimulation of ICD while limiting exposure to healthy cells.

#### 2.3 PDT damage, inflammation, and immune response against cancer

Tumor damage induced by PDT commonly involves the following: 1) ROS stimulating the direct killing of tumor cells by apoptosis, autophagy, and necrosis. 2) The vascular system targeted by PDT PSs forms thrombi, leading to tumor tissue hypoxic infarction. 3) An inflammatory response can occur, resulting in an antitumor immune response, and is triggered by inflammatory substances that are released by the tumor cells that undergo apoptosis or necrosis (6, 53). It is important to note that the PDT processes, including the cell cycle arrest, autophagy, and apoptosis, may all happen at the same time following a single session of PDT treatment. Sasnauskiene et al. findings reveal that there is a dosedependent correlation between the amount of cellular damage via oxidative stress. Elevated cell cycle arrest and autophagy, but without apoptosis were confirmed when the cellular toxic dose was augmented to 50% (54). Nonetheless, when the toxic dose was above 70%, the cells exhibited substantial cell cycle arrest, autophagy, and apoptosis. PDT-induced damage to the blood vessels principally depends on tumor tissue attributes of having large vascular gaps and poor integrity, thus promoting the aggregation of PS (55). The enrichment of the vascular endothelial tumor cells with PS following PDT photoactivation causes numerous physiological reactions, such as vasoconstriction and platelet aggregation. This results in ischemia, tumor vascular blockage, and hypoxia (6, 56). Moreover, different types of white blood cells, such as dendritic cells (DCs), macrophages, and neutrophils, are recruited thanks to the direct ablation action of PDT against tumor cells, which also liberate inflammatory mediators. The white blood cells subsequently trigger further tumor suppression via the stimulation of the immune cascade (6, 57).

PDT can also stimulate the interaction between the immune system of the adaptive and the innate arm (58, 59). Tumor microenvironment (TME) changes may occur by inducing the expression of mediators (acute-phase response mediators) and pro-inflammatory in the area irradiated. This could lead to a cascade process that induces systemic inflammation and adaptive and innate immunity. As a restricted treatment, PDT indirectly promotes the initiation of an acute inflammatory response while also directly damaging the tumor. PDT-induced oxidative stress in tumor cells can boost the release of inflammatory cytokines and inflammatory transcription factors, as well as improve the distribution of heat shock proteins (HSPs) (58). The invasion of the tumor site by leukocytes leads to their production of cytokines and pro-inflammatory factors. The mechanism of PDT can give rise to a robust inflammatory response, promoting neutrophils to quickly move to the treatment site, resulting in enhanced immunity and tumor response rate. In addition to impacting the proliferation/survival of tumor T cells and directing the production of PDT antitumor immunity, neutrophils also directly interact with photodamaged cells and later on eliminate the tumor cells that are photodamaged (60). Findings from in vivo studies indicate that PDT neutrophils bind and cluster on the microvascular wall. This provides supporting information that correlates antitumor response with neutrophil activity (61). Nonetheless, complement system activation has emerged as an antitumor mediator, and it also raises secondary inflammatory mediators, including histamine, coagulation factors, thromboxane, leukotrienes, and cytokines (such as IL-1 $\beta$ , IL-6, IL-10, G-CSF, and TNF- $\alpha$ ). Complement activation produces transmembrane channels, damages the plasma membrane integrity, and results in lysis and cell death. Besides the complement cascade system, natural killer cells, phagocytes [neutrophils, macrophages, and DCs], and cellular elements are all components of the innate immune system response. The activation of the complement and innate immune system, as well as the cytokine activity, all function together to activate the innate immune system in responding to PDT (6, 62, 63).

## 2.4 Photodynamic therapy in immunogenic cell death and DAMPs

Cancer PDT immune response leads to ICD of cancerous cells by inducing the liberation of tumor-related antigens from cancerous cells' remnants. It can also further excite the activation, infiltration, and proliferation of antigen-specific T lymphocytes (64). ICD is also characterized by a specific response mechanism that causes cellular and organellar stress, ultimately leading to cell death and the exposure, passive secretion, or active release of several DAMPs as presented in Figure 2.

DAMPs are spatiotemporally presented during ICD and are recognized by a unique type PRR found on antigen-presenting cells, starting a cascade of reactions that can trigger adaptive and innate immunologic responses (65, 66). DAMPs released by dying cells during ICD include the cytoplasmic protein annexin A1 (ANX1), the non-histone chromatin-binding protein high-mobility group box 1 (HMGB1), endoplasmic reticulum (ER) chaperones [like heat-shock proteins (HSPs) and calreticulin (CRT)], as well as interferons (IFNs) (*de novo* synthesized type I IFNs) and adenosine triphosphate (ATP) (67–69). The recognition of DAMPs by the PRRs expressed in immunogenic adaptive and innate cells leads to effector cells' chemoattraction, stimulation, maturation, and/or homing. These processes work together to suppress the tumor (65).

Studies indicated that oxidative stress induced by PDT treatment can trigger the production of DAMPs and tumor antigens, resulting in antitumor immunity (64). The stimulation



of DAMPs following PDT-treated cancerous cells is widely documented. Nonetheless, the DAMPs pattern may differ depending on the cancerous cell type and the treatment regimen (38). Moreover, the production of DAMPs following cell death has become a crucial component of the network of intercellular communication, as it influences both inflammatory processes and immunological responses (70). The PDT-stimulated DAMP samples after PDT were noted to have DAMPs such as high mobility group box 1 (HMGB1), HSPs (HSP60, HSP70, HSP90), CRT, and ATP (64). Table 1 presents the various types of DAMP that could be involved in ICD. Also, HSPs such as HSP34, HSP27, and HSP72/73 have been induced by PDT (38). The DAMPs are upregulated and translocated to the membrane, where they might be recognized and eliminated by the cells of the innate immunity as a danger sign, thus stimulating an innate immune response (62). It is reported that following PDT treatment, immunocompetent mice's tumor response was improved by CRT (DAMP molecule) as opposed to immunodeficient mice (78).

A typical characteristic of dying PDT-treated cancerous cells is the plasma membrane surface exposure of the calcium-binding protein CRT. Normally, CRTs are located in the lumen. Yet, when CRTs are externally exposed, they can be recognized by lipoprotein receptor-related protein 1 (LPR1, CD91) of low density to provide an 'eat me' type of signal to antigen-presenting cells. ER stress is linked with CRT exposure, which results from misfolded proteins piling up, thus causing an unfolded protein response (38). However, the mechanisms for CRT induced by PDT can vary based on PS type. For instance, treatment with Rose Bengal acetate led to CRT exposure and the co-translocation of the ER protein (79, 80), while the co-translocation of the ER protein was not noted in hypericinbased mediated PDT (41, 41). In addition, the eukaryotic initiation factor  $2\alpha$  phosphorylation is widely considered to be needed for CTR exposure and is vital for UPR induction (81). Yet in a hypericin-based PDT, this phosphorylation did not occur (41, 41). Also, macrophage activation can cause the release of CRT, where it may bind to viable cell surfaces and, in doing so, promote their clearance (82, 83). PDT-damaged cells found bound by CRT, were noted to activate migration, macrophages, and phagocytosis. Also, the host immune cells, including neutrophils, macrophages, and DCs, can be activated by DAMPs. DAMP expression induced thanks to PDT treatment can be a powerful immune response responsible for tumor ICD (6).

DAMPs are necessary for tumor-associated antigens presentation to antigen-presenting cells, which helps to stimulate an immune response against cancerous cells. Dead/dying PDT-treated cells are reported to release PDT-related DAMPs such as ATP and HMGB1 (43, 84, 85). Photosens and photodithazine have recently been reported to induce ICD which, was associated with the emission of HMGB1 and ATP (43). HMGB1 can trigger a response to innate immunity by interacting with 2 and 4 toll-like receptors and potentially with recognized receptors on antigen-presenting cells. ATP, on the other hand, encourages antigen-presenting cell recruitment by attaching to purinergic receptors, and this is read by antigen-presenting cells as a signal to 'find me.' ATP may be released actively from the cell, which is controlled by a particular signaling pathway, or released passively due to the loss of the integrity of the plasma membrane (86). Similar to the induced CRT mechanism, the exact mechanism stimulating ATP release following PDT appears to present unique attributes. For

DAMPs	Description and function	Responder cells	Reference
Heat Shock Proteins (HSPs) like HSP60, HSP70, HSP90, gp96, GRP94, GRP78	HSPs, or stress proteins, are often found in organelles or intracellular areas. They are produced by the cells in reaction to a variety of stress stimuli, like ultraviolet light. However, they are displayed on the surface of damaged or dying cells and are a key component in immunomodulatory processes. For instance, it has been discovered that surface-exposed HSP90 and HSP70 impact antigen processing/presentation and phagocytosis. Immunogenicity through dying cells is noted to be defined by HSP90. Though HSPs can be actively released through the non-classical secretory route, HSPs are more frequently secreted passively by dying cells. Vital tumor antigens can be carried by secreted HSPs, facilitating the appropriate uptake and processing of antigens by antigen- presenting cells. Moreover, they can activate immune cells to secrete different types of pro- inflammatory cytokines. Recent evidence indicates that extracellular HSP90b can hinder the stimulation of latent TGF-b1.	Monocytes, neutrophils	(59, 71, 72)
High mobility group box- 1 (HMGB1)	They are nuclear chromatin-binding proteins that can act as a DAMP molecule at the exterior of cells and as a nuclear protein when inside the nucleus. It is noted to have strong cytokine-like characteristics, and when HMGB1 is produced by dying cells, it activates immune cells to release a variety of pro-inflammatory cytokines.	Monocytes, neutrophils	(59, 71, 72)
Calreticulin (CRT)	They are multifunctional proteins that are usually present in different intracellular organelles/ regions (especially in the ER) (2, 13). Under stress conditions (commonly stress in the ER), their levels are increased extracellularly (exo-CRT) on the plasma membrane. They function as a danger "eat me" signal on the plasma membrane as they enhance the immunogenic ability of dying cells.		(59, 71–73)
Phosphatidylserine (PtdSer)	Phosphatidylserine can move from the cell's inner to the outer leaflet (compartment) if the cell is injured or dying to function as an "eat me signal" to mediate anti-inflammatory responses and effective phagocytosis, thanks to its interaction with multiple receptors on immune cells. They can also interact with opsonins (such as growth arrest-specific gene 6 (Gas6), $\beta$ 2-glycoprotein ( $\beta$ 2GP1), milk fat globule EGF/factor VIIC (MFG-E8), and Annexin-V).	Macrophage, dendritic cells	(71, 72, 74, 75)
Adenosine triphosphate (ATP)	They are often intracellular high-energy molecules, yet they can be secreted under specific stress conditions by apoptotic and necrotic cells. It is also possible for ATP generated extracellularly to help in immune cell chemoattraction.	Dendritic cells, microglia	(71, 72, 75)
Covalent/Cross-linked dimer of ribosomal protein S19 (dRP S19)	Description and function: They are small ribosomal subunit constituents and can be secreted by necrotic cells. They can attract different immune cells by acting as a chemotactic factor.	Monocytes, neutrophils	(59, 71, 72)
Calgranulin family members S100S (S100A8, S100A9, S100A12)	They are calcium-binding proteins expressed by different cell types. When calgranulins function as "find me signals" by attracting different immune cells. They can activate the pro- inflammatory cytokines, thanks to their interaction with the receptor (TLR4/RAGE) on immune cells.	Monocytes, neutrophils	(59, 71, 72)
Uric acid or Monosodium urate	The intracellular uric acid stockpiles in dying or ischemic cells are released. Also, more uric acid is produced after cell death as a result of the enzymatic degradation of nucleic acids.	Dendritic cells, neutrophils, CD4+ and CD8+ T cells	(59, 72, 76)
Spliceosome-associated protein 130 (SAP130)	SAP130 is a histone deacetylase complex subunit and can be produced by dying cells following apoptosis and regulated necrosis.	Macrophages	(59, 72, 77)

TABLE 1 Various types of DAMP that can be produced following cancer PDT treatment.

instance, hypericin-mediated PDT stimulated ATP secretion in a way that is autophagy-independent, which is contradictory to ICD stimulated by chemotherapy (87, 88).

## 2.5 Photodynamic therapy advantages and disadvantages

PDT has several advantages over conventional cancer therapeutic techniques. PDT has no long-term negative effect when correctly administered, yet first-generation PSs can induce modest, temporary photosensitivity in some areas, such as the eyes and skin (89, 90). When comparing PDT to surgical procedures, its adverse effects are frequently not severe, and the duration is not prolonged as in radiotherapy or chemotherapy. Also, PDT is typically administered as an outpatient procedure. Tumor mortality is substantially caused by the destruction of the tumor's associated vasculature (91, 92). PDT's dual selectivity allows it to be directly applied and accurately target a precise tissue (90). PDT can be administered repeatedly in the same treatment area if necessary, as opposed to being dispersed like in radiotherapy. Scarring after the PDT healing process is generally minimal. Moreover, PDT treatment typically costs less than the majority of cancer treatments (90, 93). PDT can also alter the microenvironments of the stroma and blood vessels, increasing their vulnerability to further treatments like IOT and chemotherapy (15).

Cancer tumors of the digestive tract, lung, lung lining (malignant pleural mesothelioma), head and neck, bladder, skin (basocellular carcinoma), and cervix have been treated using PDT (94). However, some disadvantages are linked with PDT. Tumor

cells are documented to be resistant to PDT (95, 96). Also, resistance to PDT photosensitizers may develop via similar mechanisms as those with conventional drugs, such as decreased uptake, elevated inactivation of the drug, drug efflux, and alterations in intracellular trafficking. A combination treatment strategy with at least two distinct treatment plans is one potential tactic for conquering tumor resistance (94). Also, PDT is ineffective for disseminated metastases because current PDT methods do not deliver whole-body laser light treatment, and the PDT effect often occurs at the treated location (97). This affirms that PDT yields limited therapeutic results on distal and metastatic tumors (23, 98). The effect of PDT relies on tumor tissue oxygenation, and this is commonly impeded by the thick tumor masses or necrotic tissue (90). The consumption of oxygen during PDT exacerbates tumor hypoxia, causing a vicious circle. Hypoxia in tumor cells can enhance the growth of immunosuppressive cells such as M2-type macrophages, hindering antitumor immunity and ICD, subsequently leading to tumor progression and relapses (99, 100).

Treating deeply infiltrated or deep-seated tumors is challenging using PDT due to visible light's (short light wavelength of about 400-700 nm) poor tissue penetration (101). Furthermore, PDT alone is not enough to trigger a substantial immune response since tumor cells produce immunosuppressive cytokines or other types of tumor-enhancing substances by a non-immunogenic mechanism. This causes an immunosuppressive TME that promotes immune suppression, preventing the fight against cancer (102). The efficacy of PDT is also reduced due to the TME-compressed tumor extracellular matrix, which impedes the infiltration of oxygen and chemical therapeutic species (23, 98). This is, therefore, suggestive of the importance of combination therapy, where PDT can be used with other forms of cancer therapy to surmount its therapeutic pitfalls. A PDT-optimized procedure in combination with IOT could produce an excellent synergistic impact against resistance in cancer treatment.

#### 3 Cancer immunotherapy

Cancer IOT is a biological therapeutic modality that works by enhancing the immune system's defenses to combat cancerous cells. The thymus, spleen, bone marrow, and lymph nodes are among the organs that constitute the immune system. Immune cells such as dendritic cells, B and T lymphocytes, monocytes, natural killer (NK) cells, and granulocytes, and signal proteins that highly include cytokines (INFa, TNFa, IL-11, IL-6, and IL-2) and chemokines (CXCL10 and CXCL9) help safeguard the host organism against cancer (15, 103). Active and passive IOT form the main categories of IOT, principally achieved by artificial stimulation of the adaptive and innate immune systems. Active IOT makes use of monoclonal antibodies [particularly the immune checkpoint inhibitors (ICIs)] or immunostimulation changes that occur during the release of cytokines. This category also includes IOT that directly modulates the immune response, like antigen-independent or antigendependent (for example, anticancer vaccines). Contrarily, passive IOT instead makes use of arginase inhibitors and small-molecule indoleamine 2,3-dioxygenase (IDO), along with genetically modified immune cells, including NK cells and T cells (adoptive therapy) (15, 104).

The immune system plays a vital role in combating cancer, and it may be broadly classified into two categories: the innate and adaptive immune systems. The system for the immune system is mostly made up of mucous membranes, epithelial barriers, neutrophils, macrophages, granulocytes, NK cells, and DCs (105). The system for the adaptive immune system mainly consists of humoral immunity, which principally consists of B cells, and cellular immunity, which is chiefly piloted by T cells (105, 106). The innate and adaptive immune systems should simultaneously function in the TME to effectively stimulate anticancer immunity. The main processes involved in stimulating an anticancer immunological effect can include (a) immature DCs recognizing cancerous cells or phagocytosing cancer-derived antigens (Ags), leading to their maturation. Also, through phagocytosis, the cancerous cells can be engulfed directly by macrophages (107); (b) fully developed DCs go through the lymphatic vessels to the lymph nodes, where they stimulate NK and T cells (108-110); (c) NK and T cells that are activated can migrate through the blood vessels to TME (111, 112); (d) cancer cell lysis is mediated by activated NK and T cells or macrophages. Nonetheless, the exhaustion of NK and T cells instead encourages tumor immune escape (113, 114).

#### 3.1 Current-day' cancer immunotherapy

Cancer IOT has greatly evolved and now has various therapeutic approaches that are designed to mimic the natural antitumor immunity of the body in order to combat cancer and extend the patient's life (115). The different therapeutic approaches are outlined below include IOT vaccines, adoptive cell therapies, immune checkpoint inhibitors IOT, monoclonal antibodies IOT, and oncolytic virus therapy.

IOT vaccines: IOT vaccines for cancer treatment are developed to increase immune cells' potential against cancer (116). They are divided into protein- and peptide-based vaccines (117), vector- and bacterial-based vaccines (118, 119), nucleic acid (RNA, DNA, selfamplifying RNAs (saRNA), and mRNA)-based vaccines (120–122), and cellular (DC and whole cell)-based vaccines (123).

Adoptive cell therapies (ACTs): In ACT, the NK and T cells or other cells from the patient are grown and expanded through engineering or without engineering, then infused into the patient to fight cancer (124, 125). The most common type of ACT is those derived from T-cells. The ACT based on T-cells can be accomplished by at least three unique T-cell approaches. Tumor-infiltratinglymphocytes (TILs)-based ACT is the first approach, where endogenous TILs obtained from the patient tumors are grown *ex vivo* and injected into the patient (125, 126). ACT, based on the fabricated T-cell receptor (TCR), is the second approach. This type of ACT helps recognize particular tumor antigens; however, it is restricted to Ags expressed by the major histocompatibility complex (126). ACT based on chimeric antigen receptors (CARs) is the third approach. Chimeric receptors comprising a domain for recognizing extracellular antigens, a signaling domain in the cytoplasm, and a transmembrane domain are engineered with T cells obtained from the patient, producing CAR T-cells that lock onto and eradicate the specific types of cancer (127, 128).

Immune checkpoint inhibitors (ICIs): Immunotherapy ICIs derived medication can boost tumor cell immune-mediated clearance procedure and obstruct the co-inhibitory signal pathways to reactivate immunity against the tumor (129, 130). For example, some types of cancerous cells excessively secrete on their surface the programmed death-ligand (PD-L1) in order to avoid immune surveillance. This causes the activation of the "cytotoxicity brake" to continuously working, leading to T cell exhaustion and positive PD-L1 cancer cell survival (131). Also, PD-L1 and programmed cell death protein 1 (PD-1) (132), T-cell immunoglobulin mucin-3 (TIM-3) (133), lymphocyte activation gene 3 (LAG-3) (134, 135), and cytotoxic T-lymphocyte-associated antigen 4 (CTLA-4) (132) are frequently used immune checkpoint proteins.

Monoclonal antibodies (mAbs) IOT: mAbs are large-molecularweight glycoproteins that are fabricated by B cells. They simulate induction of a durable antitumor response while also targeting cancerous cells directly (136). Three kinds of mAbs are commonly utilized in cancer treatment, which include the bispecific mAbs consisting of two distinct proteins attached together either covalently or non-covalently (137), antibody-drug conjugates, which are designed by conjugating mAbs with radioactive particles or chemotherapy drugs (138, 139), and naked mAbs (unconjugated mAbs) (140). However, there are several kinds of mAbs, which are linked to the nature of their heavy chain structures, and IgGs are currently the type most frequently utilized in antibody therapy (141, 142). Besides, the antibody (Ab) scaffold is a crucial component of tumor IOT. Findings from a recent study demonstrated that a VL one-domain antibody scaffold (rabbit-derived Ab scaffold) combined effectively with the drug 7-ethyl-10-hydroxycamptothecin (SN-38) to substantially hinder canine non-Hodgkin lymphoma (cNHL) cell in vivo and in vitro proliferation. This furnishes important theoretical justification for the application of Ab scaffolds in cancer IOT (143).

Oncolytic virus (OVs) therapy: OVs are one of the latest developments in cancer IOT (144, 145). OVs are a group of viruses that can be produced experimentally or exist naturally (146). OVs has the therapeutic ability to selectively replicate and spread in cancerous cells, destroying cancerous cells while avoiding damage to normal cells (147). In addition, OVs can be utilized for in situ vaccination (148), while immunological modulatory transgenes can be transferred to it and might even be utilized in conjunction with different therapies, such as chemotherapy and cell therapy (104, 149, 150). OVs are not limited to their application as oncolytic drugs; they can also serve as efficient carriers of anti-cancer genes and perform several functions at once, including gene therapy and virotherapy (151). The first OV medication authorized in 2015 by the U.S. FDA is T-vec (Talimogene laherparepvec, Imlygic) and is used for the treatment of recurrent melanoma in patients having topical nonresectable skin, lymph node, and subcutaneous lesions (152). Yet, the efficiency of OVs often depends on combination therapies, while stand-alone oncolytic virus treatments are subject to variation based on different factors, such as the immunological condition of the patient, the type of oncolytic viruses, and the kind of tumor. Moreover, current OVs exhibit poor infiltration and can be swiftly eliminated by antiviral responses (153).

For an effective cancer IOT, it might be essential to devise alternative methods to enhance immunogenicity and boost antitumor efficacy. Nonetheless, combinational therapies such as immuno-photodynamic therapy have emerged as a pivotal treatment strategy to address these shortcomings of mono-immunotherapy (154). Different research has demonstrated that PDT can stimulate an immunological response against tumors through an ICD mechanism. This type of treatment strategy is anticipated to compensate for the setbacks of each of the mono-therapeutic modalities and induce a healthy immune system to combat cancer. It may also offer stronger IOT immunogenicity as well as stop the proliferation of tumor cells that are still present in the body following PDT (102, 115).

## 3.2 Advantages and disadvantages of immunotherapy

Compared with conventional cancer therapy, radiotherapy, chemotherapy, and surgical intervention, IOT significantly increases patient survival by using various strategies, targets, and directives to combat cancer. IOT can target tumor tissues specifically while minimizing damage to normal tissues (155, 156). Two important causes of cancer death, metastasis and recurrence, could originate from cancerous cells developing defenses by evading immune surveillance (104, 157). About 90% of cancer-related fatalities worldwide are caused by metastatic malignancies, for which IOT has become a groundbreaking treatment option (18). IOT is unequivocally achieved using a variety of techniques to boost pre-existing immunity, reconstruct immune suppression within the TME to combat the targeted cancerous cells, and effectively trigger the innate and adaptive immunity. In improving surveillance and clearance function to stop cancerous cells from metastasis and recurrence, IOT not only amplifies the immune response in the initial tumor during therapy, but the therapy also excites systemic and long-lasting protective benefits (104, 158). Moreover, immune checkpoint blockade for IOT, such as PD-1 or PD-L1 inhibitors, has recently shown encouraging clinical results following treatment of patients with different types of cancer (159, 160). IOT methods such as cancer vaccines (161, 162), CAR T-cell therapy (163), and cytokine therapy (164) have evolved and have been proven to extend the progression-free survival of cancer patients and animal models under preclinical investigation. Also, to date, different advanced-stage cancers have been successfully treated using IOT, although different problems are encountered with its development (104, 165).

Solid tumor patients show a low response rate to IOT, which limits the therapy's efficacy (166). Besides, first-generation immunotherapy-based cancer vaccines have had poor outcomes in clinical trials (167, 168). Immune-related side effects also hinder the effectiveness of treating cancer through IOT. Autoimmunity and immunological toxicity are becoming more widely recognized as significant clinical problems (104, 165). Also, conventional methods for IOT are unable to convert non-immunogenic (cold) cancer to

immunogenic (hot) cancer (165, 169). "Hot" tumors are defined by an active immune response that frequently exhibits high levels of immune cell infiltration and responds well to immunotherapy, especially immune checkpoint blockade-based treatments. The "hot" tumors often exhibit an immune-inflamed property, with important infiltration of immune cells, especially CD8+ T cells (170). The TME of "hot" tumors is immunosupportive, which increases the effectiveness of immunotherapy, hence leading to better immune checkpoint blockade therapy in patients with "hot" tumors (171, 172). Conversely, "cold" tumors are non-immunogenic, immunosuppressed, and have inadequate T cell infiltration. The "cold" tumors can effectively camouflage themselves so as not to be recognized by the immune system, lowering the immune response efficiency and hence hindering antitumor therapy (102). Also, "cold" tumors are commonly described as an immune desert since they significantly lack immune cells (170). Therapeutically, cold tumors often respond poorly to most IOT immune checkpoint blockade treatments. However, some tumors may have both "hot" and "cold" tumor traits, making treatment options more difficult and requiring additional novel biomarkers and therapeutic combinations. Still, the "hot" and "cold" tumor categorization helps in understanding the therapeutic limitations and the responsiveness of cancer treatment (170, 173).

The limitations of current IOT techniques are linked to several factors that contribute to cancer's overall resistance. Cancerous cells are developing resistance to IOT, leading to primary, adaptive, and acquired resistance, which greatly impedes cancer IOT (104, 165). For instance, failure can arise due to an immunosuppressive TME (consisting of stromal and cancer cells), which expedites tumor immune escape. Immunosuppressive molecules are secreted by stromal cells such as cancer-associated fibroblasts (CAFs) or tumor-associated macrophages (TAMs), which prevent cytotoxic T lymphocyte infiltration and stimulation, hence decreasing their potential to kill tumor cells. Also, the stromal surrounding components containing dense extracellular matrix and aberrant tumor vessels hinder effector T-lymphocyte infiltration into the tumor and enhance hypoxia. This consequently enhances immune suppression by distorting the production of cytokines, recruiting myeloid cells to suppress the immune system, and hampering cytotoxic T-lymphocytes' killing action against cancerous cells. The aberrant tumor vasculature also controls solid tumors' immune escape and restricts the distribution of immunotherapeutic molecules into the tumor (174). The efficacy of IOT is also limited since it does not work for all patients, and this is linked with low clinical response rates and solid tumors' insufficient immunogenicity (175). Similarly, our body's physiological and pathological barriers may obstruct the uptake of immunotherapeutic drugs or natives, making the bioavailability of the drugs considerably more challenging. In addition, certain types of cancerous tumors poorly react to IOT due to the absence of an immunogenic TME (176, 177). Furthermore, present-day clinical IOT is faced with the challenge of over-activated autoimmunity and inadequate immune response. For instance, IOT cancer vaccines are not used for all forms of cancer, as they can only induce the activation of the immune system in specific forms of cancer (178); immune suppression mediated by the tumor tissue can render ACTs to malfunction (179); ICIs may result in unfavorable organ damage perpetuated through the immune system (180); and mAb treatments may cause an overactive immune response such as cytokine-release syndrome (181, 182).

The alluded problems, therefore, hamper the advancement and wider execution of cancer IOT, and if unresolved, could lead to the reiteration of mistakes similar to those observed in some conventional cancer therapy (104, 165). However, combination cancer treatment strategies are strongly encouraged (183). The rationale behind this combination treatment seems to be based on the fact that molecularly targeted treatment can significantly impact the antitumor immunity to evoke a potential synergy if utilized with IOT. The combinational therapy approach may increase the efficacy and comprehensiveness of treatment by simultaneously targeting several tumor pathways. As a result, novel IOT approaches needed to circumvent tumor immune evasion are desperately needed.

## 4 Combination of PDT with IOT (photoimmunotherapy)

Combined treatment of PDT and IOT (photoimmunotherapy), can trigger both systemic and local immune responses in animal studies (preclinical studies), resulting in longer-lasting immune activity, more tumor cell death, and slower tumor growth (Figure 3). Among the notable advantages of this combination treatment are enhanced antigen presentation, reduced Tregs, increased T cell stimulation, and strengthened resistance to recurrence, regardless of the type of photosensitizer that is used. It is reported that treatment combining PDT and CpG oligodeoxynucleotide can reduce metastases, potentially amplify the activation of CD8+ T cells, and prolong survival (184, 185). Similarly, a two-stage treatment intended to target the tumor directly combined a low PDT immunogenic dose followed by an elevated dose was documented. Prolongs survival and slow growth of metastases tumor was noted, albeit the effects vary depending on the tumor cell line (58, 69). PDT-based immunotherapy is generally a versatile therapeutic method and is suitable for a wide range of patients since the treatment does not depend on a precise tolerance profile or genetic predisposition (69, 186). To achieve specific immunological stimulation in cancer treatment through photoimmunotherapy, different promising approaches involving monoclonal antibodies, immune inhibitors, immune adjuvants, immune checkpoint blockade, and tumor vaccines are used in preclinical settings to overcome tumor resistance and improve treatment outcomes.

## 4.1 Preclinical methods and evidence of photoimmunotherapy

#### 4.1.1 Photoimmunotherapy antibodyphotosensitizer conjugate

To overcome anticancer treatment resistance and improve treatment efficacy, antibody-photosensitizer conjugates (AbPCs) have emerged as a viable strategy of combining PDT and targeted



activate an immunological antitumor effect through the induction of an immunogenic cell death. IOT methods (monoclonal antibodies, immune inhibitors, immune adjuvants, immune checkpoint blockade, and tumor vaccines) in combination with PDT can elicit an immune response through the stimulation of NK cells and T cells. Overall, treatment by photoimmunotherapy can result in long-lasting tumor immunity, inhibition of the original and metastasized tumor, and eradication of tumor cells.

immunotherapy. The AbPCs are designed in such a way that the antibodies target the tumor tissue precisely, and the photosensitizer is also deposited in the tumor, which, when activated by light, induces ROS production that kills tumor cells. It is important to note that AbPCs both directly destroy cancerous cells and promote an immunological response against cancer (187, 188). This indicates that PDT's selectivity and efficacy can be increased by conjugating PS with mAbs in the right way to activate an immunological response. Furthermore, the poor distribution of PS, because of the hydrophobic nature of most PSs, can be surmounted by incorporating PSs with specific antibodies. This helps to facilitate the internalization of the PS alongside the antibody (15, 188, 189). For instance, in a colorectal cancer model, a conjugate composed of chlorin e6 and cetuximab (cetuximab-maleimide-poly(ethylene oxide)-poly(propylene oxide)-poly(ethylene oxide)-chlorine e6 conjugate, CMPXC) in PDT markedly elevated the cytotoxic Tcell and dendritic cell population (187). Likewise, PDT employing AbPCs can trigger the release of danger signals and antigens, strengthening anticancer immunological responses (16). Also, developed biomimetic photosensitizers [aggregation-induced emission (AIE) photosensitizers] with hitchhiking and antigenpresenting capacity (DC@AIEdots) stimulated T-cell proliferation and activation in vivo, stimulating the immune system (190). The

conjugation of mAbs with aluminum phthalocyanine was validated. The low concentration of the free photosensitizer in the TME did not produce a significant PDT effect; however, conjugating the antibodies with the photosensitizer substantially elevates the selectivity and cytotoxicity effect of PDT while preserving the targeting molecule's stability, integrity, and immunological reactivity (15, 191).

Likewise, by specifically binding to tumor-associated antigens, an antibody conjugated to PS enables effective delivery of the therapeutic agent to cancer cells, resulting in more efficient treatment (192, 193). For instance, the conjugation of antibody against epidermal growth factor receptors (EGFR) and the near-infrared (NIR) phthalocyanine dye IR700 has demonstrated encouraging outcomes in causing selective cell death following exposure to NIR light (194). Interestingly, photoimmunotherapy can provide a special benefit when it comes to drug resistance mechanisms, especially those involving drug efflux pumps. It is demonstrated that conventional PSs like hypericin interact with breast cancer resistance protein and multidrug resistance-associated protein-1, which are known to contribute to drug resistance (195). Photoimmunotherapy treatment can potentially bypass the drug efflux mechanism, guaranteeing adequate intracellular accumulation of the PS (194, 196). For instance, photoimmunotherapy is reported to successfully

Antibody	Photosensitizer	Conjugation method	Anticancer target	Ref.
Trastuzumab (α-HER2)	IRDye700	Pye700 Formation of amide bonds with lysine residues through activated ester		(203)
Cetuximab (α-EGFR)	Benzoporphyin monoacid ring A	Formation of amide bonds with lysine residues through activated ester	Chinese hamster ovary cell line expressing EGFR (CHO-EGFR)	(204)
α-CD104	2,5-dioxopyrrolidin-1-yl 4 (4Z,10Z,14E,15Z,19Z)- 10,15,20-tri(pyridin-4-yl)-1H,21H-porphyrin-5- yl)benzoate	Formation of amide bonds with lysine residues through activated ester	Human bladder transitional cell carcinoma (UM-UC-3)	(205)
Trastuzumab (α-HER2)	(8S)-5-(carboxymethyl)-7-(3-carboxypropyl)-18-ethyl- 2,8,12,17-tetramethyl-13-vinyl 7H,8Hporphyrin- 3-carboxylic acid	Formation of amide bonds with lysine residues through activated ester	Epithelial human breast cancer (MDA-MB-231), human breast cancer	(206)
HuHMFG1	Pyropheophorbide a	Formation of amide bonds with lysine residues through activated ester	Human colorectal adenocarcinoma (HT-29), human oesophageal adenocarcinoma (OE19)	(207)
LAG-3	5-(4-isothiocyanatophenyl)-10,15,20-tris-(4-N- methylpyridiniumyl) porphyrin trichloride	Isothiocyanate to amines on lysine residue	Human colorectal cancer (Caco-2)	(208)
α-ЕрСАМ	4-(15-(4-isothiocyanatophenyl)porphyrin-5-yl)-1- methylpyridin-1-ium chloride	Isothiocyanate to amines on lysine residue	Human colorectal cancer (LoVo), Lung large cell carcinoma (CORL23)	(209)
35A7	5-(4-isothiocyanatophenyl)-10,15,20-tris-(4-N- methylpyridiniumyl) porphyrin trichloride	Isothiocyanate to amines on lysine residue	SKOv3-CEA-1B9 tumor	(210)
35A7 FSP 77	5,5′,5″-(20-(4-isothiocyanatophenyl)porphyrin-5,10,15-triyl)tris (benzene-1,3-diol)	Isothiocyanate to amines on lysine residue	Colorectal adenocarcinoma (LS174T), Ovarian adenocarcinoma (SKOv3)	(210)
SIP (LI9)	4,4',4''-(20-(4-((1-(4-((2,5-dioxopyrrolidin-1-yl)methyl) cyclohexyl)-1-oxo-5,8,11,14,17,20,23-heptaoxa-2- azapentacosan-25-yl)carbamoyl)phenyl)porphyrin-5,10,15- triyl)tris(1-methylpyridin-1-ium)	Reduction via disulfide bridge and reconnection of free thiol using maleimide	LM fibroblasts, immortalized human embryonic kidney cells (HEK293T), Chinese hamster ovary cells (CHO- S), HEK293T	(211)
SIP (LI9)	4,4',4''-(20-(4-(2,5-dioxo-2,5-dihydro-1H-pyrrol-1-yl) phenyl)porphyrin-5,10,15-triyl)tris(1-methylpyridin- 1-ium)	Reduction via disulfide bridge and reconnection of free thiol using maleimide	Vaccinia-infected mouse fibroblasts (LM fibroblasts), CHO- S, (HEK293T)	(211)
Trastuzumab (Fab)	Porphyrin	Reduction via disulfide bridge and reconnection of free thiols using maleimide propargylmaleimide proceede by CuAAC ("click" reaction)		(212)
Trastuzumab (IgG)	Porphyrin	Reduction via disulfide bridge and reconnection of free thiols using maleimide propargylmaleimide proceeded by "click" reaction.	MDA-MB-468	(213)

TABLE 2	Coniugation	methods	involved in	antibodv-	photosensitizer	coniugation	and their	anticancer	target	

lower the survival of these resistant cells, improving the effectiveness of treatment as a whole by targeting cancer stem-like cells noted to display drug efflux mechanisms (192, 193).

Antibody and PS conjugation can be formed through different conjugation techniques, including genetic and chemical techniques that exploit specific functional groups like N-chlorosuccinimide (NCS), triazoles, and thiol. Frequently used chemical methods use reactive groups found on the photosensitizer and antibody, such as carbodiimide, isothiocyanate, or NHS (succinimidyl) ester, to produce conjugates (15, 188, 189). In genetic techniques, the antibody and the photosensitizer are fused directly using a recombinant protein, producing a single molecule (197–199). Using chemical techniques in bioconjugation enables the attachment of the PS to a specific area on the antibody. The bioconjugation approach for PDT and related therapeutic applications could be divided into different categories, including isothiocyanate conjugation, amide conjugation, tetrapyrrole-nanoparticle-antibody conjugates, maleimide conjugation, and copper-catalyzed azide– alkyne cycloaddition (CuAAC) reactions ("click" reaction) (15, 200–202). Table 2 presents some of the conjugation methods involved in antibody-photosensitizer conjugation for targeting specific tumor.

Isothiocyanate-derived conjugates are formed by joining the antibody's amino group to the PS using the isothiocyanate active group (210). The LAG-3 antibody was fused with 5-(4-isothiocyanatophenyl)-10,15,20-tris-(4-N-methylpyridiniumyl) porphyrin trichloride through the isothiocyanate active group. The obtained conjugate demonstrated excellent PDT activity in the mouse

model by significantly slowing the growth of the tumor. Also, the conjugate toxicity was noted to be tolerable in the animals tested (15).

Amide bioconjugations are a frequently used approach that assists in joining two molecules together through amide bond formation and are commonly executed by fusing activated esters with amines. For instance, the conjugation of the HuHMFG1 antibody with pyropheophorbide was reported. The conjugate was observed *in vivo* and *in vitro* in preclinical investigations to kill esophageal cancerous cells effectively. The conjugated compound thus shows promise in esophageal adenocarcinoma treatment and can provide a targeted and efficient therapeutic option for esophageal cancer (207).

Maleimide conjugation is the process of joining the antibody's thiol group to the PS using the maleimide active functional group. To boost adaptability, maleimide functional groups could be integrated into bigger molecular weight linkers. For instance, the capacity of three types of porphyrins with a maleimide group linked to them to specifically mark cysteine residues on SIP-L19 antibodies [small immunoprotein (SIP)-antiangiogenic antibody (L19)] was studied. The antibody and photosensitizer complex was formed by the porphyrin's aryl ring attaching directly to the antibodies or by utilizing the succinimidyl-4-(N-maleimidomethyl) Linker cyclohexane-1-carboxylic (SMCC). The SMCC was joined to the photosensitizer through polyethylene glycol (PEG) or an aliphatic chain. The obtained conjugate preserved the photosensitizer's capacity to kill cells when activated by light and the antibody's capacity to bind to its target (211). Another photoimmunotherapy photosensitizer was developed for the localized targeting and treatment of prostate cancer (PC) and PC stem-like cells (PCSC). The photosensitizer recombinant cysteine-modified anti-EpCAM and anti-CD44 antibodies conjugated with silicon phthalocyanine dye (WB692-CB2 dye) through a maleimide linker. The developed conjugate, after red light irradiation, exhibited target-specific binding and elevated cytotoxicity on PCSC and PC. The conjugate could serve in PC-efficient treatment while protecting the prostate gland and with reduced adverse effects. It could also be employed in radical prostatectomy to destroy residual cancerous cells or metastasized tumors in the lymph node areas in scenarios where surgery is infeasible (193).

In addition, the "click" conjugation technique is noted for being a very selective and effective chemical reaction and is gaining popularity in the synthetic chemistry sector. The CuAAC reaction ("click" reaction) is now documented to be commonly used in bioconjugation because of its selectivity, excellent yields, and biocompatibility (214). PDT photosensitizer-antibody bioconjugates developed using the "click" reaction method demonstrate high phototoxicity due to their enhanced optical ability in PDT and can selectively target and destroy cancerous cells (212, 213, 215). A combination of hydrophilic PEG and hydrophobic zinc phthalocyanine PS (C11Pc) stabilized water-soluble gold nanoparticles, which were then functionalized with jacalin (lectin or monoclonal antibodies specific to HER). The complex obtained in combination with PDT induced an enhanced production of singlet oxygen and phototoxicity in SK-BR-3 (breast adenocarcinoma cells) and HT-29 (colorectal adenocarcinoma cells) (216).

#### 4.1.2 Photoimmunotherapy immune inhibitors/ immune adjuvants

Immunological adjuvants have been shown to be effective when used in conjunction with PDT. Imiquimod (a TLR7 agonist) is among the most promising adjuvants and has been approved by the FDA for the treatment of different skin diseases (217). Imiquimod causes DCs to mature and liberate pro-inflammatory cytokines by interacting with TLR7 on the endosomes and DCs (218). Imiguimod was used in a cream in combination with 5-aminolevulinic acidmediated PDT to successfully treat skin squamous cell carcinoma (219). Also, it has been demonstrated that the artificial dipeptide called pidotimod can strengthen the immune response and guard against infection in humans and mice. However, the exact mode of action of pidotimod's protective ability is unclear. According to a study exploring zebrafish models, PDT improved immune cell recruitment and enhanced the production of pro-inflammatory cytokines following the tail wound assay, yet protection from certain pathogen infections was not provided (220). PDT can also induce immunosuppression and inflammation because of contact hypersensitivity. Nonetheless, immune inhibitors are necessary to soften the immunosuppression signal of the tumor and allow for a fully induced PDT immune response (221, 222).

Anti-angiogenic peptides in combination with photofrinmediated PDT preceded T cell activation and VEGF (vascular endothelial growth Factor) inhibition, resulting in improved PDT efficacy. An enhanced PDT impact and related immune response were equally observed when PDT was conducted in combination with granulocyte macrophage colony-stimulating factor (GM-CSF), IL-7, and IL-3. The GM-CSF and IL-7 were delivered into the tumor tissue by retroviral vectors, then PDT was conducted, followed by their administration. The T lymphocyte's activity was stimulated by IL-7, and the macrophage maturation process was augmented thanks to GM-CSF activity. Recombinant cytokines, including IFN- $\gamma$ , IL-1, IL-6, IL-8, and IL-18, were also utilized in conjunction with PDT. The efficacy of PDT combined with the cytokine-based therapy strongly relies on the timing and mode of their administration, correlated with the PDT procedure (223).

Also, PDT can be utilized in combination with nonspecific types of IOT, such as cytokine-based therapy. PDT in combination with an administered tumor necrosis factor alpha (TNFa) proves to boost the therapy's efficacy. A TNFα antivascular inducer, 5,6dimethylxanthenone-4-acetic acid (DMXAA), in combination with Photofrin-mediated PDT, occasioned a reduction in tumor volume and the resumption of a prolonged growth period in mice having the radiation-induced fibrosarcoma-1. This combinational treatment strategy led to tumor tissue necrosis with a reduction in blood flow and vascular density. The utilization of GM-CSF led to similar results by initiating the suppression of the growth of the tumor, prolonging the CT26 survival period, and bearing the LLC tumor in mice. An enlargement of anticancer immunity and complete tumor cell destruction was observed in 1/3 of the mice treated (58). Besides, protoporphyrin IX (PpIX)-mediated PDT treatment was enhanced with lipopolysaccharide (LPS). The method of combined treatment caused the decreased production of IL-6 and an elevation of IL-10 and TNF $\alpha$  levels (224).

#### 4.1.3 Immunotherapy/PDT vaccines

A promising type of combination therapeutic strategy is the utilization of PDT-treated cancerous cells to immunize DCs. Thus, PDT-induced anticancer vaccines are based on the anticancer immune response that is activated by PDT. In TME, PDT can activate an immune response that could be exploited to generate an anticancer vaccine (225). Cells of the immune system implicated in this type of response often include T-cells and DCs, among others. Studies based on PDT-generated vaccines revealed that T-cell stimulation produced long-lasting anti-cancer immunity (15). Korbelik, a renowned scientist, used PDT to create a vaccination for squamous cell carcinoma (SCCVII). He demonstrated that this vaccine inhibited the growth of tumors (226). The utilization of Par-ICG-Lipo filled with indocyanine green (ICG) for endoplasmic reticulum (ER)-targeted PDT induced ICD and demonstrated in vivo to improve the in situ tumor cells' immunogenicity. This ERtargeted PDT, in conjunction with dendritic cells, may result in an effective clinical method for treating cancer by modifying cancerous cells into a vaccine for treating cancer (227). In another study, a dendritic cell vaccine containing glioma cells was stimulated by photosens-mediated PDT, resulting in ICD and the discovery of a four-gene signature linked to glioma patients' general survival. This strategy may have the capacity to enhance the treatment of glioma by triggering Th17 immunity and helping in the prediction of patient outcomes (228).

Moreover, regulatory T-cells (Tregs), which are involved in hindering immunological activity against cancerous cells, can strongly affect the immunological response induced by PDTgenerated vaccines. The depletion of these cell types can augment the efficiency of PDT-generated vaccines (229). The tumor wholecell-derived vaccine can be optimized through interconnections with phagocytic receptors. The interconnection between phagocytic receptors and cancerous cells can enhance the effectiveness of PDTgenerated vaccines. Scavenger receptors and phagocytic receptors (such as mannose receptors) are present on macrophages and dendritic cells. These receptors have the potential to recognize and attach to tumor-associated antigens, resulting in their retention and presentation to T-cells (230).

Also, tumor cell-derived vaccines can be optimized by using a different adjuvant like N-dihydrogalactochitosan (glycated chitosan) (231) or maneuvering cell death, especially that induced by necrosis (232). Dendritic cells can be activated by using adjuvants, consequently causing the adjuvant activation and thus robust production of an anti-cancer immune response for tumorassociated antigens expressing cancerous cells. Still, it is established that the activation of an immune response triggered by PDT-induced cell lysates on P815 and EMT6 cancerous cells does not need adjuvant co-administration (58). Similarly, research findings indicate that PDT-induced cell lysates activate DC maturation and the expression of IL-12 (233). A combination of PDT with immune checkpoint blockade or with immuno-regulatory activity dampening can also maximize the efficiency of PDT vaccines (15, 69). Other authors created a delivery nano drug system having doxorubicin hydrochloride and chlorin e6 (Ce6) as PS loaded on an amphipathic 4T1 breast cancer membrane that was coated by calcium carbonate. The concurrent action of PDT and PDT resulted in the liberation of tumor-associated antigens and ICD. It is anticipated that ROS produced by this technique will create PDT-DC vaccination by recruiting DCs through imitating inflammatory mechanisms (234).

## 4.1.4 Photoimmunotherapy immune checkpoint blockade therapy

Immune checkpoint blockades (commonly monoclonal antibodies) are often used in IOT to target protein interactions that typically dampen the immune system (221, 235). The immune checkpoints exhibit undesirable immunomodulatory effects (236). Treatments based on immune checkpoint blockade are guided by helpful biomarkers such as tumor mutation burden. Future approaches could use tumor mutation burden and other biomarkers to better stratify individuals for IOT, which could help overcome the problem of high tumor burdens and poor treatment results, leading to cancer patients receiving an effective treatment (237–239). Current treatment strategies utilize target-specific antibodies to stop different immune checkpoints (235). The importance of PDT in such a treatment process is to promote tumor sensitivity and immunogenicity, thereby inducing ICD (221).

Medically used photosensitizers are being examined in conjunction with various immunotherapies targeting CTLA-4, VEGF, OX40 [tumor necrosis factor receptor superfamily member 4 (TNFRSF4)], EphA2 (ephrin type-A receptor 2), and immune checkpoint blockades, such as PD-1/PD-L1. The administration of antibodies that target PD-1 and its ligand, in conjunction with a vascular-targeted PDT (V-PDT) that utilizes Tookad<sup>®</sup>Soluble as a photosensitizer, modulated the immune system reaction, causing a high number of CD8+ T cells to infiltrate the TME. It also resulted in a decrease in the number and size of metastases and stimulated a general immune response (216). V-PDT in conjunction with OX40 and PD-1 targeted therapy also led to an elevated immune response (240, 241). Treatment using a conjugate composed of anti-EphA2 antibody and IRDye700 (near-IR fluorescent dye) promoted an increased ICD, conversely to treatment utilizing the photosensitizer alone (196). Immunotherapy that targets PD-L1 and CTLA-4 receptors in combination with PDT contributed to raising the survival rate of treated mice by activating various immune responses through inflammatory induction, phagocytosis, or improved leukocyte infiltration (242). An anti-VEGF therapy in combination with PDT improved antitumor response; yet, 24 hours after treatment, the build-up of the PDT photosensitizer [5,10,15,20-tetrakis(3hydroxyphenyl) chlorin (mTHPC)] was reduced (243). Ripasudil, in conjunction with Ce6-embedded nanophotosensitizermediated PDT (FIC-PDT), triggered an ICD and incited the priming of tumor-specific cytotoxic T lymphocytes through the sensitization of antigen-presenting cells. This led to the activation of the PD-1/PD-L1 immune checkpoint blockade response, causing a strong antitumor impact in the melanoma intraocular model (244). PDT has been proposed as a possible complementary method to immune checkpoint inhibitors, including PD-1/PD-L1, CTLA-4, and CD47-targeted therapy. The principle of this treatment strategy is PDT's potential to boost the immune system and strengthen the response to cancer induced by immune checkpoint inhibitors. PDT in conjunction with immune checkpoint inhibitors has produced good results in different studies, indicating the possibility of increasing the effectiveness of cancer IOT (221, 245). Immuno checkpoint blocking can be used to rewire the immune system to target the residual malignant cells after PDT has been used to target the original tumor in a multimodal treatment paradigm that combines IOT and PDT (241).

Moreover, PDT in conjunction with different therapies that focus on regulating macrophage activity or adjusting autophagy activity through chloroquine inhibition is being studied. PDT's ability to activate a proapoptotic effect has increased due to its inhibitory effects on autophagic activity (246). The therapeutic outcome of PDT was enhanced by the macrophageactivating factor, achieving a 100% therapeutic efficacy. However, no alterations in tumor growth were perceived when the activating factor [D3-binding protein-derived macrophage-activating factor (DBPMAF)] was administered without PDT. The DBPMAF also lessened the immunosuppressive effect brought on by PDT (247).

Furthermore, transient hypoxia during PDT can change the phenotype of immune cells and tumors by upregulating PD-L1, which is reliant on hypoxia-inducible factor 1-alpha signaling. The possible changes linked with immune checkpoint homeostasis at post-PDT may work in concert with PDT's stimulation of immune cell infiltration to support PD-1/PD-L1 blockage as a beneficial supplementary tactic (241, 248, 249).

#### 4.1.5 Challenges of photoimmunotherapy

Some patients might find the combined cancer treatment strategy, such as photoimmunotherapy, to be expensive and inaccessible, which can hinder the widespread utilization of the treatment (18, 250). A major obstacle to photoimmunotherapy platform development is the absence of clinical trial statistics to help forecast the photoimmunotherapy execution in humans. This could hinder photoimmunotherapy interventions from completely replacing traditional cancer treatment, yet photoimmunotherapy is anticipated to be more suitable and widely used as an adjunct therapy. Also, it is crucial to determine which patients stand to gain from this treatment approach (18).

Moreover, different perspectives, such as cancer resistance, immune-related adverse events (irAEs), toxicity, treatment parameter optimization, accessibility, and cost, can be examined to validate the setbacks and risks linked with PDT and IOT combined treatments. PDT uses photosensitizers, which can cause localized toxicity and negative consequences if they do not target precisely the required treatment area and can harm healthy tissues (18, 94, 251). Consequently, merging PDT and IOT may cause extreme toxicity, especially if the therapies are not adequately coordinated. However, irAEs that can include autoimmune responses may arise, where the body's immune system targets healthy cells, which can be brought on by IOT. The irAEs may need to be managed carefully due to their severity (mild to severe effects). Also, treating the immune-related conditions could be complex, hence necessitating more interventions and medications, which puts an enormous burden on patients (18). Likewise, some IOT agents could induce an immunosuppressive TME that hampers the effects (dampening effect) of PDT-induced ICD (252). Additionally, PDT can worsen irAEs since inflammation brought on by PDT might cause adverse effects in both the treated area and nearby normal tissues, hindering treatment outcomes (253, 254). PDT-induced inflammation may cause or exacerbate adverse drug events such as myocardial damage and muscle weakness in individuals receiving immunotherapy, such as with tislelizumab (253). Also, PDT-induced inflammatory response may intensify the immunological activation brought on by PD-1 blockade, resulting in severe irAEs (254). Another major problem is figuring out the ideal parameters for exploring photoimmunotherapy, like dosage, treatment sequencing, and timing. Standardizing the treatment methods is challenging since every patient reacts differently. Cancer that is resistant to both PDT and IOT, and combining the two treatments, might still not be enough to get beyond the resistance mechanism (18). Nonetheless, more advanced and effective anticancer photoimmunotherapy methods using nanotechnology may use target-specific moieties for overcoming resistance and other treatment challenges.

### 5 Preclinical evidence of nanophotoimmunotherapy combating cancer resistance

Nanotechnology can improve photoimmunotherapy's accuracy and effectiveness, providing answers to some of the treatment challenges. Nanomaterials with unique therapeutic attributes can exhibit high photoactivity, low toxicity, and multifunctional characteristics, and alongside a suitable wavelength of light stimulation, can deeply penetrate tumor tissues (18). Multiple treatment strategies can be successfully integrated into one platform thanks to the multi-functionality of nanoparticles (255, 256). Photoimmunotherapy assisted by nanotechnology methods is noted to present synergistic effects, improving the antitumor immunological responses (257). This indicates that major cancer treatment resistance phenomena that are linked with the TME, metastases, enhanced permeability and retention (EPR), and nonspecific targeting may be addressed by incorporating formulated specific forms of nanoparticles in photommunotherapy treatment. This can help to overcome irAEs, toxicity, and treatment resistance.

## 5.1 Nano-photoimmunotherapy in specific tumor-targeting

Tumor-targeting through nano-photoimmunotherapy can minimize systemic toxicity, safeguard normal healthy cells, and strengthen photoimmunotherapy. Besides, the targeted delivery of

immune modulators and photosensitizers into the tumor tissue through nanoparticles can augment the treatment effectiveness (258, 259). For instance, Deng et al. produced a reduction-sensitive nanocarriers (Ds-sP NPs) [PEG-s-s-1,2-distearoyl-sn-glycero-3phosphoethanolamine-N-(amino (polyethylene glycol)-2000] that were successfully loaded with an endoplasmic reticulum ER-targeting PS (TCPP-TER) [4,4',4",4'"-(porphyrin-5,10,15,20-tetrayl)tetrakis(N-(2-((4methylphenyl)sulfonamido) ethyl)benzamide]. The obtained Ds-sP/TCPP-TER NPs complex exhibited a selective ER accumulation, resulting in local production of ROS, following treatment with the near-infrared laser. This induced an increased level of oxidative stress in the tumor cell ER, activating the DAMPs and hence producing an amplified ICD (260). Similarly, PDT combined with epidermal growth factor receptor blockade was investigated, where a nanobody-IRDye700 conjugate composite was administered to a mouse orthotopic tumor model. The results prove that the nanobody-PS conjugate composite accumulated in the tumor tissue, causing necrosis with a non-toxic impact on normal tissues (261). In addition, Hanaoka et al. developed a photoimmunotherapy photosensitizer nano-conjugate composite (drug), IR700-YP7, to target Glypican-3 (GPC3). Glypican-3 (GPC3) is a surface biomarker expressed on HCC cells and is a therapeutically attractive target since its expression is predominantly high in hepatocellular carcinoma (HCC) and not in healthy cells. Also, HCC is a deadly malignancy worldwide, and only a small percentage of patients with HCC can benefit from curative surgery (262, 263). The IR700-YP7 (IRDye700DX conjugated with anti-GPC3 antibody) photoimmunotherapy drug with nab-paclitaxel was used to treat A431/G1 tumors in mice. When the treated tumor tissue was exposed to near-infrared light treatment, the IR700-YP7 rapidly caused A431/G1 cell death. Photoimmunotherapy helped in decreasing the tumor growth in comparison with the untreated tumor and also facilitated the improved delivery of nab paclitaxel, hence boosting the treatment therapeutically (262). This is supported by the fact that AbPCs do not affect non-expressing cells, but they are only effective as a therapeutic drug when attached to the targeted cell membrane. Also, early after photoimmunotherapy, tumor vessels are not damaged and are permeable, promoting a dramatic increase in blood flow. This makes it easy for nanosized drugs of high concentration to be delivered to the specific tumor during treatment, with little uptake in the non-tumor targeted areas (264). In another study, HCC was targeted through its cell surface biomarker called EpCAM (epithelial cell adhesion molecule) (265). EpCAM plays a vital role in cell proliferation, adhesion, stemness, and migration. This makes EpCAM a possible IOT target in cancer treatment and is also useful as a prognostic marker and in diagnosis (266, 267). EpCAM has been reported to be a stem cell marker and greatly promotes the survival and metastasis of cancerous tumors, including HCC (267, 268). An anti-EpCAM-conjugated nano-micelle (anti-EpCAM-UPGs-MX) was developed by Han et al. An excellent EpCAM targeting signal was observed in HCC-bearing mice after they were treated with the conjugate. This was confirmed through a vibrant green fluorescence signal from the treated mice. The anti-EpCAM-conjugated nanomicelle also exhibited both passive and active targeting potential, leading to its elevated aggregation rate in the tumor even 48 hours

after treatment. Conversely, the untargeted micelles exhibited passive targeting and no active targeting (265).

In clinical practice, targeting tumors through the EPR effect does not always result in fruitful outcomes, as the EPR effect would rely on the tumor type and location, the macromolecular antitumor drug's physical-chemical attributes, and the tumor blood perfusion state. However, nanoparticles can promote the EPR effect, making it possible to deliver the antitumor agent precisely. This can lead to increased accumulation of the antitumor agent at the tumor site and enhance blood supply to the tumor (269, 270). A study by Sano et al. developed a novel antibody conjugate consisting of an antibody and photosensitizer [RDye 700DX (IR700)] to enhance photoimmunotherapy by the EPR effect. The antibody conjugate could deliver nanoparticles (with sizes 10-200 nm) effectively into the tumor site. Highly selective cell killing occurred rapidly following 690 nm of phototreatment. The antibody bound maximally to the cells in the perivascular tumor space and caused the fast killing of tumor cells. When these cells are killed fast, vascular permeability rises, permitting rapid leakage of nanoparticles into the tumor. As a result, the photo-immunotherapy-treated tumor accumulated the nanoparticles up to 24 times above that of the control tumor, a phenomenon referred to as "super-enhanced permeability and retention." Also, the photo-immunotherapy combined with liposome-daunorubicin treatment resulted in improved treatment and prolonged survival of the tumor-bearing mice (264). In addition, a nano-redox-activatable liposome (RAL) was developed and encapsulated with an indoleamine 2,3-dioxygenase (IDO) inhibitor (IDO@RAL). The RAL demonstrated an EPR effect by its increased tumor accumulation and prolonged blood circulation in mice with 4T1 tumors. If endocytosis occurred following treatment of the tumor, the nanovesicle may cause an exponential increase in PDT activity (>100-fold) and fluorescence signal due to the high glutathione threshold in the tumor intracellular space. Consequently, phototoxicity to healthy cells will be reduced, and tumor growth inhibition achieved thanks to the nano-activatable design. Interestingly, the RAL-mediated PDT led to cytotoxic T cells' intratumoral infiltration by triggering the tumor cells in ICD. The treatment, in combination with the IDO inhibitor, led to an augmented systemic antitumor immunologic effect (271).

# 5.2 Nano-photoimmunotherapy in overcoming tumor microenvironment setbacks

Solid tumors commonly display certain TME resistance characteristics, such as extreme hypoxia, low pH, and elevated levels of glutathione (GSH) content, compared to healthy tissues. The TME can be modulated using nanoparticles to make it less immunosuppressive and more favorable for treatment that could trigger an immune response activation. Also, smart nanotechnology TME-sensitive components or chemical linkers can be designed to overcome the TME setbacks. TME-sensitive nanoparticle components, and also advanced nanosized metal–organic frameworks can efficiently escape the aggregation-caused quenching. This allows for photosensitizers to be released quickly in the tumor, leading to great elevations in ROS production (99, 258). The elevated ROS can strengthen the immunological response by boosting the effect of ICD. In addition, TME-sensitive nanoparticles can program the pharmacokinetics and location of both immunomodulators and photosensitizers smartly. This increases the tumor-targeting ability, resulting in optimized photoimmunotherapy without producing any acute side effects (99). For instance, a study by Zhen et al. developed a nano-photoimmunotherapy (nanoparticle-based photoimmunotherapy) to modulate the TEM for effective immunity against cancer. In the study, carcinoma-associated fibroblasts (CAFs) were combated by targeting a fibroblast-activation protein (FAP), noted to be highly expressed on CAFs' surface (272). CAFs, as a major TME component, in both the original and metastatic tumors, can strongly affect the behavior of cancerous cells and exhibit multifunctions in tumor development, metastasis, angiogenesis, cancer stemness, metabolism, immunosuppression, and tumorigenesis (273-275). FAP is regarded as a universal antigen for tumor targeting, and its expression by multipotent bone marrow stem cells has been reported by different studies (272, 276, 277). A nano-photoimmunotherapy drug developed using ferritin (a solid nanoparticle-protein cage, serving as a photosensitizer carrier) was conjugated with FAP-scFv (FAP-specific single-chain variable fragment). The photo-irradiation treatment facilitated the nano-photoimmunotherapy drug to eradicate the CAFs in tumors, yet with little injury to the normal tissues because of the treatment location. Importantly, the nano-photoimmunotherapy resulted in strong suppression of the tumor in immunocompetent mice. Additional investigation revealed that the nanophotoimmunotherapy promoted a decrease in the secretion of C-X-C motif chemokine ligand 12 (CXCL12) and the deposition in the tumor extracellular matrix, which are all controlled in untreated tumors by CAFs and also regulate T cell exclusion, which stops T cells from direct contact with cancerous cells. CAFs' selective killing through nano-photoimmunotherapy leads to substantial T cell infiltration, accompanied by effective suppression of the tumor (272). Another study serendipitously discovered that pH-responsive nanovesicles (pRNVs) (which are self-assembled from block copolymer polyethylene glycol-b-cationic polypeptide) are capable of more than just acting as nanocarriers, as the pRNVs also trigger ICD through exposing calreticulin on the surface of preapoptotic cells. The pRNVs composite was developed by encapsulating with indoximod (IND) (an indoleamine 2,3-dioxygenase inhibitor) and the photosensitizer [2-(1-hexyloxyethyl)-2-devinyl pyropheophorbide-a (HPPH)], forming a pRNVs/HPPH/IND composite. A low dose of pRNVs/HPPH/IND stimulated significant anticancer impact, and the photo-irradiation led to an abscopal effect on the B16F10 melanoma model. The major outcome of the treatment includes the generation of singlet oxygen through HPPH-mediated PDT, increased recruitment of DC and immunological response following ICD stimulation by PDT and pRNVs, as well as the modulation of TME by IND, which was upregulated by P-S6K phosphorylation, leading to the inhibition of Tregs and the enlargement of CD8+ T cells. The study thus presents an "allin-one" nanocarrier that uses multifunctional materials to enhance the effectiveness of cancer immunotherapy (278).

Tumor hypoxia remains a significant barrier to immunotherapy, PDT, and other cancer treatment methods, resulting in poor clinical prognosis (279, 280). Hypoxia is essential for developing an immunosuppressive TEM, since it controls programmed death ligand 1 (PD-L1) expression and immunosuppressive TAMs infiltration (281). However, nanocarriers can deliver therapeutic agents to reduce hypoxia, which is often common in TMEs, improving the efficacy of cancer therapy. To overcome hypoxia and enhance tumor treatment, oxygen-generating or oxygen-carrying strategies mediated by nanoparticles can promote oxygen elevation in the tumor tissue (279, 280). A core-shell nanoformulation (AuNC@ MnO<sub>2</sub>, AM) was created in another study, consisting of a hollow gold nanocage (AuNCs) formulation covered with a manganese dioxide coat (282) for treating metastatic triple-negative breast cancer (mTNBC) through PDT-induced oxygen-boosted immunogenicity. mTNBC is a very aggressive form of cancer that is typified by producing elevated fatality and poor prognosis, even with systemic chemotherapy and radiotherapy interventions. The AuNC@MnO2, AM nanomaterial acts like a TME-responsive oxygen producer. The PDT-induced oxygen-boosted immunogenicity was stimulated thanks to the generation of ROS after the NIR irradiation treatment. In the presence of excessive H<sub>2</sub>O<sub>2</sub> and in an acidic microenvironment, the manganese dioxide (MnO<sub>2</sub>) undergoes a reaction: MnO<sub>2</sub> + H<sub>2</sub>O<sub>2</sub> +  $2H^+ \rightarrow Mn^{2+} + 2H_2O + O_2\uparrow$  in the tumor tissue, producing a lot of oxygen to enhance the build-up of ROS in the tumor tissue, while also increasing the effectiveness of PDT. This procedure elicits an ICD and the release of DAMPs, thereby inducing DC maturation and the stimulation of effector cells. This evokes a strong systemic immunological response against mTNBC. Also, the produced Mn<sup>2+</sup> and oxygen are useful for multimodal imaging since they can generate fluorescence (FL)/photoacoustic (PA)/magnetic resonance, hence providing the possibility for integrating the diagnosis and treatment of tumors (282).

Another study described an MnO2-containing albumin nanoformulation for enhancing IOT through immunosuppressive TME modulation and tumor hypoxia alleviation. The MnO2containing albumin nano-formulation facilitated the collaborative delivery of paclitaxel dimer, NLG919, and IR780 to augment photoimmunotherapy. An increase in oxygen supply was catalyzed by MnO2, promoting an effective paclitaxel-mediated therapy and PDT, which collectively enhanced the development of specific cytotoxic T cells and ICD. Interestingly, the increased oxygen supply relieves the tumor tissue from hypoxia, hence modifying the immunosuppressive TME by suppressing the PD-L1 expression and the M2-type TAMs infiltration in the tumor tissue. This enhanced the effectiveness and infiltration of cytotoxic T cells when combined with immune checkpoint blockade through NLG919, leading to the complete eradication of the primary tumor and almost completely halting tumor cell metastasis and relapse. This study validates a strategic therapeutic method for breast cancer by strengthening IOT through hypoxia relief by immunosuppression modulation and ICD induction (281).

#### 5.3 Nano-photoimmunotherapy in addressing metastasis and recurrence

Cancer metastasis and recurrence remain the inevitable problems despite the utilization of robust treatment methods. Also, addressing the seeding and colonization of metastatic tumor cells remains difficult. The current IOT method seems to be more suitable for solving these problems. However, the clinical utilization of IOT is hindered by poor tumor antigen presentation, tumor tissue heterogeneity, and inadequate targeting (283, 284). Combination photo-immunotherapy procedures that use nanoparticles are encouraged, as they could enhance responsiveness in patients responding to cancer treatments (258). A study by Xu et al., 2017 engineered upconversion nanoparticles (UCNPs) that were concurrently filled with imiquimod (R837) (toll-like-receptor-7 agonist) and chlorin e6 (Ce6), producing a UCNP-Ce6-R837 complex that was used for preventing the recurrence of colorectal cancer by targeting both the primary and metastasized tumor. The UCNP-Ce6-R837 treatment, followed by near-infrared (NIR) irradiation, would lead to depth penetration of the tissue, leading to effective tumor destruction by PDT. This promotes the generation of tumor-associated antigens, which, when combined with the adjuvant (R837) containing nanoparticle (UCNP-Ce6-R837), can stimulate a robust anticancer immunological response. Interestingly, UCNP-Ce6-R837 plus PDT treatment merged with checkpoint blockade (CTLA-4) not only exhibited outstanding effectiveness in eradicating the tumor when subjected to NIR irradiation but also caused a potent anticancer immunity, preventing the development of the distant tumor that remained following PDT treatment. Moreover, the long-term immunological memory capacity of such an IOT method against cancer shields the treated mice against cancer cell reactivation (285).

Besides, a nano-chimeric peptide composite (nano-PpIX-1MT) was developed to target lung cancer tumors and metastasized tumors by photoimmunotherapy. The nano-PpIX-1MT integrates an immune checkpoint inhibitor [1-methyl-tryptophan (1MT)] with the photosensitizer [protoporphyrin IX (PpIX)] using a caspaseresponsive peptide sequence [Asp-Glu-Val-Asp (DEVD)]. The nano-PpIX-1MT infiltrated the tumor site by an enhanced penetration and retention effect, followed by photo-irradiation treatment at 630 nm. This resulted in ROS production, which caused the cancerous cells to undergo apoptosis. This facilitated caspase-3 up-regulation and a strong immune response against tumor antigen production. The cleavage of caspase-3 was followed by 1MT release, subsequently enhancing the immune system and aiding in the efficient activation of CD8+ T cells. The nano-PpIX-1MT was therefore able to induce a cascaded synergic photoimmunotherapy effect by inhibiting the original tumor and metastasized lung cancer (283).

Furthermore, Ce6/BMS-202 NPs [Ce6/BMS-202/Bristol-Myers Squibb nanoparticles (NPs)] were prepared for synergistic PDT and IOT on 4T1 tumors. The NPs present beneficial properties, such as exhibiting a 100% drug loading ability and nontoxic and hydrophilic properties. BMS-202 (N-{2-[({2-Methoxy-6-[(2-Methyl[1,1'-Biphenyl]-3-Yl)methoxy]pyridin-3-Yl}methyl)amino] ethyl}acetamide) is a nonpeptidic small molecule capable of strongly inhibiting PD-1/PD-L1 interaction. Treating 4T1 tumorbearing mice with BMS-202 NPs resulted in a significant decrease in 4T1 tumor growth, which was identical to the antitumor effect induced by anti-PD-L1 monoclonal antibody ( $\alpha$ -PD-L1) treatment. Using Ce6 NPs in tandem with  $\alpha$ -PD-L1 or BMS-202 NPs constantly leads to more anticancer and antimetastatic efficacy. This was accompanied by increased maturation of dendritic cells and improvement in antigen-specific T cells infiltrating the tumor tissue, hence resulting in over 90% of the original and distant tumors being inhibited. Also, the BMS-202 NPs can combat lung cancer metastasis and stop the recurrence of the tumor by providing immune-memory protection. BMS-202 NPs could possibly be used to replace monoclonal antibodies for cancer IOT applications, as antibodies present various therapeutic limitations, such as poor immunogenicity, ineffective tumor tissue penetration, and being very costly (286).

Similarly, the study by Guo et al. utilized nano-core-shell magnetic composites (MNCs) to develop an oxygen-independent photosensitizer for the treatment of triple-negative breast cancer models. The MNCs-mediated PDT treatment led to persistent production of free radicals by promoting the polarization of macrophages into pro-inflammatory M1 phenotype, electron-hole dissociation efficacy, and stimulating a systemic immunological response against the tumor (287) However, concurrent adaptive immune resistance was observed with the MNCs-mediated PDT treatment, which was typified by increased expression of PD-L1 on tumor tissue, macrophages, and DCs. The MNCs mediated PDT treatment in combination with checkpoint blockade significantly suppressed the original and metastasized tumors through three intervention processes 'trident modality,' which includes immunosuppressive TEM modification with inhibition of PD-L1 blockade and immunosuppressive cells; increased tumor-infiltratinglymphocyte (TIL) rates; and steady generation of free radicals in both the hypoxic and normoxic states to directly eradicate the tumor. Likewise, the possible mechanisms responsible for metastasis inhibition were investigated using the lung tissue transcriptome expression profiling. The outcome indicated that the 'trident modality' modified several genes that are linked with cancer-related signal pathways and immune activation. This "trident modality" could be used widely in clinical settings and serve as a potential therapeutic strategy for managing cancer that is resistant to treatment (287). Moreover, mesoporous hexagonal core-shell zinc porphyrin-silica nanomaterial (MPSNs) with the ability to serve as a superior photosensitizer in photo-immunotherapy and also as a drug carrier to achieve a synergistic effect were loaded with R837 (imiquimod, a tolllike receptor-7 agonist) (MPSNs@R837) to stimulate photothermal therapy (PTT) and PDT ICD. This strategy led to strong immunological responses specific to 4T1 tumors in mice, by promoting dendritic cell maturation following the pH-responsive release of R837 and subsequently causing little toxicity and a strong suppression of both primary and metastatic tumors when in conjunction with the programmed death ligand-1 (PD-L1) checkpoint blockade. This treatment strategy thus demonstrates that the utilization of checkpoint blockade alongside PTT and PDT treatment can inhibit cancer metastasis (288).

#### 6 Conclusion and perspective

Cancer PDT and IOT are promising therapeutic strategies. However, these treatment strategies are still ineffective in eradicating cancer due to their inability to surmount therapeutic defeats linked with cancer resistance. The involvement of PDT in activating anticancer immunological responses is highlighted in different studies. PDT is a well-liked treatment that has become a great way to enhance immunotherapies for a more effective cancer treatment. Various methods of incorporating PDT with IOT explore immunologic adjuvants, developed DC vaccines, immune checkpoint blockades, and antibody-photosensitizer conjugates and have led to excellent synergistic effects in preclinical studies. Still, the development of photoimmunotherapy seems to be in its early stages, since it seems to lack clinical evidence to fully validate its anticancer therapeutic potential in clinical settings. Major treatment setbacks, such as toxicity, treatment accessibility, and cost, also hamper photoimmunotherapy. Nanotechnology can help address some of the therapeutic issues by increasing the precision and efficacy of photoimmunotherapy. The amalgamation of tailored nanoparticles with photoimmunotherapy (nano-photoimmunotherapy) molecules or methods is confirmed in preclinical studies to offer cancertargeted therapy, combat cancer metastasis and recurrence, and prevent resistance from the TME. This helps to reduce treatment systemic toxicity and overall cancer resistance while also promoting a long-lasting immunological anticancer response. Unfortunately, these promising outcomes are obtained from preclinical research conducted in a variety of in vitro and in vivo studies.

Photoimmunotherapy or nano-photoimmunotherapy clinical applications are strongly needed in order to fully warrant the efficacy of these combination therapies in eradicating resistant cancer in patients. To fully comprehend these anticancer therapeutic methods' potential in clinical settings and further improve them, more research is necessary. Therefore, further clinical research or multiple clinical trial studies are solicited to determine the broader efficacy of photoimmunotherapy or nanophotoimmunotherapy procedures across diverse patient tumor types. This could lead to clinical advanced photoimmunotherapy molecules for different cancers being introduced to the market. This could help in overcoming cancer resistance in cancer patients.

#### References

1. Bueschbell B, Caniceiro AB, Suzano PMS, Machuqueiro M, Rosário-Ferreira N, Moreira IS. Network biology and artificial intelligence drive the understanding of the multidrug resistance phenotype in cancer. *Drug Resist Updat.* (2022) 60:100811. doi: 10.1016/j.drup.2022.100811

2. World Health Organization. *Cancer* (2025). Available online at: https://www.who. int/news-room/fact-sheets/detail/cancer (Accessed Feb 18, 2025).

3. Debela DT, Muzazu SG, Heraro KD, Ndalama MT, Mesele BW, Haile DC, et al. New approaches and procedures for cancer treatment: Current perspectives. *SAGE Open Med.* (2021) 9:20503121211034366. doi: 10.1177/20503121211034366

4. Liu B, Zhou H, Tan L, Siu KTH, Guan XY. Exploring treatment options in cancer: tumor treatment strategies. *Sig Transduct Target Ther*. (2024) 9:1–44. doi: 10.1038/ s41392-024-01856-7

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#### Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest. This is a provisional file, not the final typeset article

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5. Wicki A, Mandalà M, Massi D, Taverna D, Tang H, Hemmings BA, et al. Acquired resistance to clinical cancer therapy: A twist in physiological signaling. *Physiol Rev.* (2016) 96:805–29. doi: 10.1152/physrev.00024.2015

6. Hua J, Wu P, Gan L, Zhang Z, He J, Zhong L, et al. Current strategies for tumor photodynamic therapy combined with immunotherapy. *Front Oncol.* (2021) 11:738323. doi: 10.3389/fonc.2021.738323

7. Lei Z, Tian Q, Teng Q, Wurpel JND, Zeng L, Pan Y, et al. Understanding and targeting resistance mechanisms in cancer. *MedComm (2020).* (2023) 4:e265. doi: 10.1002/mco2.265

8. Saleh R, Elkord E. Acquired resistance to cancer immunotherapy: Role of tumormediated immunosuppression. *Semin Cancer Biol.* (2020) 65:13–27. doi: 10.1016/ j.semcancer.2019.07.017 9. Wang X, Zhang H, Chen X. Drug resistance and combating drug resistance in cancer. *Cancer Drug Resist.* (2019) 2:141–60. doi: 10.20517/cdr.2019.10

10. Salgia R, Kulkarni P. The genetic/non-genetic duality of drug 'resistance' in cancer. Trends Cancer. (2018) 4:110-8. doi: 10.1016/j.trecan.2018.01.001

11. Marine JC, Dawson SJ, Dawson MA. Non-genetic mechanisms of therapeutic resistance in cancer. *Nat Rev Cancer.* (2020) 20:743–56. doi: 10.1038/s41568-020-00302-4

12. Palmer AC, Sorger PK. Combination cancer therapy can confer benefit via patient-topatient variability without drug additivity or synergy. *Cell.* (2017) 171:1678–91. doi: 10.1016/ j.cell.2017.11.009

13. Mokhtari RB, Homayouni TS, Baluch N, Morgatskaya E, Kumar S, Das B, et al. Combination therapy in combating cancer. *Oncotarget.* (2017) 8:38022. doi: 10.18632/ oncotarget.16723

14. Li X, Peng X, Zoulikha M, Boafo GF, Magar KT, Ju Y, et al. Multifunctional nanoparticle-mediated combining therapy for human diseases. *Sig Transduct Target Ther.* (2024) 9:1–33. doi: 10.1038/s41392-023-01668-1

15. Warszyńska M, Repetowski P, Dąbrowski JM. Photodynamic therapy combined with immunotherapy: Recent advances and future research directions. *Coordination Chem Rev.* (2023) 495:215350. doi: 10.1016/j.ccr.2023.215350

16. Kleinovink JW, van Driel PB, Snoeks TJ, Prokopi N, Fransen MF, Cruz LJ, et al. Combination of photodynamic therapy and specific immunotherapy efficiently eradicates established tumors. *Clin Cancer Res.* (2016) 22:1459–68. doi: 10.1158/1078-0432.CCR-15-0515

17. Rajendrakumar SK, Uthaman S, Cho CS, Park IK. Nanoparticle-based phototriggered cancer immunotherapy and its domino effect in the tumor microenvironment. *Biomacromolecules*. (2018) 19:1869–87. doi: 10.1021/acs.biomac.8b00460

18. Farahavar G, Abolmaali SS, Biabanikhankahdani R, Tamaddon AM. Synergistic action of combining photodynamic therapy with immunotherapy for eradicating solid tumors in animal models: A systematic review. *Crit Rev Oncology/Hematology.* (2025) 209:104691. doi: 10.1016/j.critrevonc.2025.104691

19. Kah G, Chandran R, Abrahamse H. Curcumin a natural phenol and its therapeutic role in cancer and photodynamic therapy: A review. *Pharmaceutics*. (2023) 15:639. doi: 10.3390/pharmaceutics15020639

20. Xiang H, Xue F, Yi T, Tham HP, Liu JG, Zhao Y. Cu2-xS Nanocrystals crosslinked with chlorin e6-functionalized polyethylenimine for synergistic photodynamic and photothermal therapy of cancer. ACS Appl Mater Interfaces. (2018) 10:16344–51. doi: 10.1021/acsami.8b04779

21. Wang H, Han X, Dong Z, Xu J, Wang J, Liu Z. Hyaluronidase with ph-responsive dextran modification as an adjuvant nanomedicine for enhanced photodynamicimmunotherapy of cancer. *Advanced Funct Materials*. (2019) 29:1902440. doi: 10.1002/adfm.201902440

22. van Straten D, Mashayekhi V, de Bruijn HS, Oliveira S, Robinson DJ. Oncologic photodynamic therapy: basic principles, current clinical status and future directions. *Cancers (Basel).* (2017) 9:19. doi: 10.3390/cancers9020019

23. Merlin JPJ, Crous A, Abrahamse H. Combining photodynamic therapy and targeted drug delivery systems: enhancing mitochondrial toxicity for improved cancer outcomes. *Int J Mol Sci.* (2024) 25:10796. doi: 10.3390/ijms251910796

24. Bacellar IOL, Tsubone TM, Pavani C, Baptista MS. Photodynamic efficiency: from molecular photochemistry to cell death. *Int J Mol Sci.* (2015) 16:20523–59. doi: 10.3390/ijms160920523

25. Jomova K, Raptova R, Alomar SY, Alwasel SH, Nepovimova E, Kuca K, et al. Reactive oxygen species, toxicity, oxidative stress, and antioxidants: chronic diseases and aging. *Arch Toxicol.* (2023) 97:2499–574. doi: 10.1007/s00204-023-03562-9

26. Falk-Mahapatra R, Gollnick SO. Photodynamic therapy and immunity: an update. *Photochem Photobiol.* (2020) 96:550–9. doi: 10.1111/php.13253

27. Kessel D. Apoptosis, paraptosis and autophagy: death and survival pathways associated with photodynamic therapy. *Photochem Photobiol.* (201) 95:119–25. doi: 10.1111/php.12952

28. Mishchenko T, Balalaeva I, Gorokhova A, Vedunova M, Krysko DV. Which cell death modality wins the contest for photodynamic therapy of cancer? *Cell Death Dis.* (2022) 13:1–16. doi: 10.1038/s41419-022-04851-4

29. Chang LC, Chiang SK, Chen SE, Hung MC. Exploring paraptosis as a therapeutic approach in cancer treatment. *J Biomed Science*. (2024) 31:101. doi: 10.1186/s12929-024-01089-4

30. Zeng S, Chen C, Zhang L, Liu X, Qian M, Cui H, et al. Activation of pyroptosis by specific organelle-targeting photodynamic therapy to amplify immunogenic cell death for anti-tumor immunotherapy. *Bioact Mater.* (2023) 25:580–93. doi: 10.1016/j.bioactmat.2022.07.016

31. Zhu T, Shi L, Yu C, Dong Y, Qiu F, Shen L, et al. Ferroptosis promotes photodynamic therapy: supramolecular photosensitizer-inducer nanodrug for enhanced cancer treatment. *Theranostics.* (2019) 9:3293–307. doi: 10.7150/thno.32867

32. Dhuriya YK, Sharma D. Necroptosis: a regulated inflammatory mode of cell death. J Neuroinflammation. (2018) 15:199. doi: 10.1186/s12974-018-1235-0

33. Baskaran R, Lee J, Yang SG. Clinical development of photodynamic agents and therapeutic applications. *Biomaterials Res.* (2018) 22:25. doi: 10.1186/s40824-018-0140-z

34. Escudero A, Carrillo-Carrión C, Castillejos MC, Romero-Ben E, Rosales-Barrios C, Khiar N. Photodynamic therapy: Photosensitizers and nanostructures. *Mater Chem Front.* (2021) 5:3788–812. doi: 10.1039/D0QM00922A

35. Plekhova N, Shevchenko O, Korshunova O, Stepanyugina A, Tananaev I, Apanasevich V. Development of novel tetrapyrrole structure photosensitizers for cancer photodynamic therapy. *Bioengineering (Basel).* (2022) 9:82. doi: 10.3390/bioengineering9020082

36. Aebisher D, Serafin I, Batóg-Szczęch K, Dynarowicz K, Chodurek E, Kawczyk-Krupka A, et al. Photodynamic therapy in the treatment of cancer—The selection of synthetic photosensitizers. *Pharm (Basel).* (2024) 17:932. doi: 10.3390/ph17070932

37. Jia X, Jia L. Nanoparticles improve biological functions of phthalocyanine photosensitizers used for photodynamic therapy. *Curr Drug Metab.* (2012) 13:1119–22. doi: 10.2174/138920012802850074

38. Alzeibak R, Mishchenko TA, Shilyagina NY, Balalaeva IV, Vedunova MV, Krysko DV. Targeting immunogenic cancer cell death by photodynamic therapy: past, present and future. *J Immunother Cancer*. (2021) 9:e001926. doi: 10.1136/jitc-2020-001926

39. Attar GS, Kumar M, Bhalla V. Targeting sub-cellular organelles for boosting precision photodynamic therapy. *Chem Commun.* (2024) 60:11610–24. doi: 10.1039/D4CC02702G

40. Kah G, Chandran R, Abrahamse H. Green silver nanoparticles curcumin conjugate induced photodynamic therapy of lung cancer and lung cancer stem cells. *RSC Adv.* (2025) 15:5020–41. doi: 10.1039/D4RA06035K

41. Garg AD, Krysko DV, Verfaillie T, Kaczmarek A, Ferreira GB, Marysael T, et al. A novel pathway combining calreticulin exposure and ATP secretion in immunogenic cancer cell death. *EMBO J.* (2012) 31:1062–79. doi: 10.1038/emboj.2011.497

42. Adkins I, Fucikova J, Garg AD, Agostinis P, Špíšek R. Physical modalities inducing immunogenic tumor cell death for cancer immunotherapy. *Oncoimmunology*. (2014) 3:e968434. doi: 10.4161/21624011.2014.968434

43. Turubanova VD, Balalaeva IV, Mishchenko TA, Catanzaro E, Alzeibak R, Peskova NN, et al. Immunogenic cell death induced by a new photodynamic therapy based on photosens and photodithazine. *J ImmunoTherapy Cancer.* (2019) 7:350. doi: 10.1186/s40425-019-0826-3

44. Gomes-da-Silva LC, Zhao L, Bezu L, Zhou H, Sauvat A, Liu P, et al. Photodynamic therapy with redaporfin targets the endoplasmic reticulum and Golgi apparatus. *EMBO J.* (2018) 37:e98354. doi: 10.15252/embj.201798354

45. Cincotta L, Szeto D, Lampros E, Hasan T, Cincotta AH. Benzophenothiazine and benzoporphyrin derivative combination phototherapy effectively eradicates large murine sarcomas. *Photochem Photobiol.* (1996) 63:229–37. doi: 10.1111/j.1751-1097.1996.tb03019.x

46. Kessel D, Evans CL. Promotion of proapoptotic signals by lysosomal photodamage: mechanistic aspects and influence of autophagy. *Photochem Photobiol.* (2016) 92:620–3. doi: 10.1111/php.12592

47. Kessel D, Reiners JJ. Promotion of proapoptotic signals by lysosomal photodamage. *Photochem Photobiol.* (2015) 91:931-6. doi: 10.1111/php.12456

48. Meng Z, Xue H, Wang T, Chen B, Dong X, Yang L, et al. Aggregation-induced emission photosensitizer-based photodynamic therapy in cancer: from chemical to clinical. *J Nanobiotechnology*. (2022) 20:344. doi: 10.1186/s12951-022-01553-z

49. Borgia F, Giuffrida R, Caradonna E, Vaccaro M, Guarneri F, Cannavò SP. Early and late onset side effects of photodynamic therapy. *Biomedicines.* (2018) 6:12. doi: 10.3390/biomedicines6010012

50. Ibbotson SH. Adverse effects of topical photodynamic therapy. *Photodermatology Photoimmunology Photomedicine*. (2011) 27:116–30. doi: 10.1111/j.1600-0781.2010.00560.x

51. Mishchenko TA, Turubanova VD, Mitroshina EV, Alzeibak R, Peskova NN, Lermontova SA, et al. Effect of novel porphyrazine photosensitizers on normal and tumor brain cells. *J Biophotonics*. (2020) 13:e201960077. doi: 10.1002/jbio.201960077

52. Mitroshina EV, Mishchenko TA, Turubanova VD, Alzeibak R, Balalaeva IV, Vedunova MV, et al. Effect of photosensitizers photosens, photodithazine and hypericin on glioma cells and primary neuronal cultures: A comparative analysis. *Sovrem Tehnol Med.* (2019) 11:52. doi: 10.17691/stm2019.11.4.06

53. Papa V, Furci F, Minciullo PL, Casciaro M, Allegra A, Gangemi S. Photodynamic therapy in cancer: Insights into cellular and molecular pathways. *Curr Issues Mol Biol.* (2025) 47:69. doi: 10.3390/cimb47020069

54. Sasnauskiene A, Kadziauskas J, Vezelyte N, Jonusiene V, Kirveliene V. Apoptosis, autophagy and cell cycle arrest following photodamage to mitochondrial interior. *Apoptosis.* (2009) 14:276–86. doi: 10.1007/s10495-008-0292-8

55. Zhang H, long Y, Gang YZ, Qiang LZ, Jun JH, Jun QQ. New Strategies for the treatment of solid tumors with CAR-T Cells. *Int J Biol Sci.* (2016) 12:718–29. doi: 10.7150/ijbs.14405

56. Krammer B. Vascular effects of photodynamic therapy. Anticancer Res. (2001) 21:4271-7.

57. Nath S, Obaid G, Hasan T. The course of immune stimulation by photodynamic therapy: bridging fundamentals of photochemically induced immunogenic cell death to the enrichment of T-cell repertoire. *Photochem Photobiology.* (2019) 95:1288–305. doi: 10.1111/php.13173

58. Castano AP, Mroz P, Hamblin MR. Photodynamic therapy and anti-tumour immunity. Nat Rev Cancer. (2006) 6:535-45. doi: 10.1038/nrc1894

59. Mroz P, Hashmi JT, Huang YY, Lange N, Hamblin MR. Stimulation of antitumor immunity by photodynamic therapy. *Expert Rev Clin Immunol.* (2011) 7:75–91. doi: 10.1586/eci.10.81

60. Kousis PC, Henderson BW, Maier PG, Gollnick SO. Photodynamic therapy enhancement of antitumor immunity is regulated by neutrophils. *Cancer Res.* (2007) 67:10501–10. doi: 10.1158/0008-5472.CAN-07-1778

61. de Vree WJ, Essers MC, de Bruijn HS, Star WM, Koster JF, Sluiter W. Evidence for an important role of neutrophils in the efficacy of photodynamic therapy in *vivo*. *Cancer Res.* (1996) 56:2908–11.

62. Tan L, Shen X, He Z, Lu Y. The role of photodynamic therapy in triggering cell death and facilitating antitumor immunology. *Front Oncol.* (2022) 12:863107/full. doi: 10.3389/fonc.2022.863107/full

63. Wachowska M, Muchowicz A, Demkow U. Immunological aspects of antitumor photodynamic therapy outcome. *Cent Eur J Immunol.* (2015) 40:481–5. doi: 10.5114/ ceji.2015.56974

64. Garg AD, Nowis D, Golab J, Vandenabeele P, Krysko DV, Agostinis P. Immunogenic cell death, DAMPs and anticancer therapeutics: An emerging amalgamation. *Biochim Biophys Acta (BBA) - Rev Cancer.* (2010) 1805:53-71. doi: 10.1016/j.bbcan.2009.08.003

65. Gong T, Liu L, Jiang W, Zhou R. DAMP-sensing receptors in sterile inflammation and inflammatory diseases. *Nat Rev Immunol.* (2020) 20:95–112. doi: 10.1038/s41577-019-0215-7

66. Yatim N, Cullen S, Albert ML. Dying cells actively regulate adaptive immune responses. *Nat Rev Immunol.* (2017) 17:262–75. doi: 10.1038/nri.2017.9

67. Di Virgilio F, Sarti AC, Falzoni S, De Marchi E, Adinolfi E. Extracellular ATP and P2 purinergic signalling in the tumour microenvironment. *Nat Rev Cancer*. (2018) 18:601–18. doi: 10.1038/s41568-018-0037-0

68. Rosenzweig R, Nillegoda NB, Mayer MP, Bukau B. The Hsp70 chaperone network. Nat Rev Mol Cell Biol. (2019) 20:665–80. doi: 10.1038/s41580-019-0133-3

69. Thiruppathi J, Vijayan V, Park IK, Lee SE, Rhee JH. Enhancing cancer immunotherapy with photodynamic therapy and nanoparticle: making tumor microenvironment hotter to make immunotherapeutic work better. *Front Immunol.* (2024) 15:1375767. doi: 10.3389/fimmu.2024.1375767

70. Chen R, Zou J, Zhong X, Liu J, Kang R, Tang D. *The DAMP theory: Concepts, evidence, and implications* (2024). Available online at: https://www.authorea.com/ users/868604/articles/1250265-the-damp-theory-concepts-evidence-and-implications. (Accessed April 22, 2025).

71. Garg AD, Krysko DV, Vandenabeele P, Agostinis P. DAMPs and PDT-mediated photo-oxidative stress: exploring the unknown. *Photochem Photobiol Sci.* (2011) 10:670–80. doi: 10.1039/c0pp00294a

72. Reginato E, Wolf P, Hamblin MR. Immune response after photodynamic therapy increases anti-cancer and anti-bacterial effects. *World J Immunol.* (2014) 4:1-11. doi: 10.5411/wji.v4.i1.1

73. Li W, Yang J, Luo L, Jiang M, Qin B, Yin H, et al. Targeting photodynamic and photothermal therapy to the endoplasmic reticulum enhances immunogenic cancer cell death. *Nat Commun.* (2019) 10:3349. doi: 10.1038/s41467-019-11269-8

74. Nagata S, Suzuki J, Segawa K, Fujii T. Exposure of phosphatidylserine on the cell surface. Cell Death Differ. (2016) 23:952-61. doi: 10.1038/cdd.2016.7

75. Panzarini E, Inguscio V, Dini L. Immunogenic cell death: Can it be exploited in photodynamic therapy for cancer? *BioMed Res Int*. (2013) 2013:482160. doi: 10.1371/journal.pone.0105778

76. Hernandez C, Huebener P, Schwabe RF. Damage-associated molecular patterns in cancer: A double-edged sword. *Oncogene.* (2016) 35:5931–41. doi: 10.1038/ onc.2016.104

77. Li L, Ren J, Guo M, An Z, Duan W, Lv J, et al. SAP130 mediates crosstalk between hepatocyte ferroptosis and M1 macrophage polarization in PFOS-induced hepatotoxicity. *Sci Total Environment.* (2024) 951:175612. doi: 10.1016/j.scitotenv.2024.175612

78. Korbelik M, Banáth J, Saw KM, Zhang W, Čiplys E. Calreticulin as cancer treatment adjuvant: combination with photodynamic therapy-generated vaccines. *Front Oncol.* (2015) 5:15. doi: 10.3389/fonc.2015.00015

79. Galluzzi L, Buqué A, Kepp O, Zitvogel L, Kroemer G. Immunosuppressive cell death in cancer. *Nat Rev Immunol.* (2017) 17:402–2. doi: 10.1038/nri.2017.48

80. Panzarini E, Inguscio V, Fimia GM, Dini L. Rose Bengal acetate photodynamic therapy (RBAc-PDT) induces exposure and release of damage-associated molecular patterns (DAMPs) in human HeLa cells. *PloS One.* (2014) 9:e105778. doi: 10.1371/journal.pone.0105778

81. Bezu L, Sauvat A, Humeau J, Leduc M, Kepp O, Kroemer G. eIF2 $\alpha$  phosphorylation: A hallmark of immunogenic cell death. *Oncoimmunology*. (2018) 7: e1431089. doi: 10.1080/2162402X.2018.1431089

82. Feng M, Marjon KD, Zhu F, Weissman-Tsukamoto R, Levett A, Sullivan K, et al. Programmed cell removal by calreticulin in tissue homeostasis and cancer. *Nat Commun.* (2018) 9:3194. doi: 10.1038/s41467-018-05211-7

83. Krysko DV, Ravichandran KS, Vandenabeele P. Macrophages regulate the clearance of living cells by calreticulin. *Nat Commun.* (2018) 9:4644. doi: 10.1038/ s41467-018-06807-9

84. Garg AD, Vandenberk L, Koks C, Verschuere T, Boon L, Van Gool SW, et al. Dendritic cell vaccines based on immunogenic cell death elicit danger signals and T cell-driven rejection of high-grade glioma. *Sci Transl Med.* (2016) 8:328ra27. doi: 10.1126/scitranslmed.aae0105

85. Wang X, Ji J, Zhang H, Fan Z, Zhang L, Shi L, et al. Stimulation of dendritic cells by DAMPs in ALA-PDT treated SCC tumor cells. *Oncotarget.* (2015) 6:44688–702. doi: 10.18632/oncotarget.5975

86. Krysko DV, Garg AD, Kaczmarek A, Krysko O, Agostinis P, Vandenabeele P. Immunogenic cell death and DAMPs in cancer therapy. *Nat Rev Cancer.* (2012) 12:860–75. doi: 10.1038/nrc3380

87. Garg AD, Dudek, Aleksandra M, Ferreira GB, Verfaillie T, Vandenabeele P, Krysko DV, et al. ROS-induced autophagy in cancer cells assists in evasion from determinants of immunogenic cell death. *Autophagy*. (2013) 9:1292–307. doi: 10.4161/auto.25399

88. Michaud M, Martins I, Sukkurwala AQ, Adjemian S, Ma Y, Pellegatti P, et al. Autophagy-dependent anticancer immune responses induced by chemotherapeutic agents in mice. *Science*. (2011) 334:1573–7. doi: 10.1126/science.1208347

89. Abrahamse H, Hamblin MR. New photosensitizers for photodynamic therapy. *Biochem J.* (2016) 473:347–64. doi: 10.1042/BJ20150942

90. Correia JH, Rodrigues JA, Pimenta S, Dong T, Yang Z. Photodynamic therapy review: principles, photosensitizers, applications, and future directions. *Pharmaceutics*. (2021) 13:1332. doi: 10.3390/pharmaceutics13091332

91. Nyst HJ, Tan IB, Stewart FA, Balm AJM. Is photodynamic therapy a good alternative to surgery and radiotherapy in the treatment of head and neck cancer? *Photodiagnosis Photodyn Ther.* (2009) 6:3–11. doi: 10.1016/j.pdpdt.2009.03.002

92. Prajapati S, Hinchliffe T, Roy V, Shah N, Jones CN, Obaid G. Biomimetic nanotechnology: A natural path forward for tumor-selective and tumor-specific NIR activable photonanomedicines. *Pharmaceutics.* (2021) 13:786. doi: 10.3390/pharmaceutics13060786

93. Aebisher D, Rogóż K, Myśliwiec A, Dynarowicz K, Wiench R, Cieślar G, et al. The use of photodynamic therapy in medical practice. *Front Oncol.* (2024) 14:1373263. doi: 10.3389/fonc.2024.1373263

94. Al-Jamal AN, Al-Hussainy AF, Mohammed BA, Abbas HH, Kadhim IM, Ward ZH, et al. Photodynamic Therapy (PDT) in drug delivery: Nano-innovations enhancing treatment outcomes. *Health Sci Review.* (2025) 14:100218. doi: 10.1016/j.hsr.2025.100218

95. Aniogo EC, George BP, Abrahamse H. Characterization of resistant MCF-7 breast cancer cells developed by repeated cycles of photodynamic therapy. *Front Pharmacol.* (2022) 13:964141. doi: 10.3389/fphar.2022.964141

96. Dąbrowska A, Mastalerz J, Wilczyński B, Osiecka B, Choromańska A. Determinants of photodynamic therapy resistance in cancer cells. *Int J Mol Sci.* (2024) 25:12069. doi: 10.3390/ijms252212069

97. Agostinis P, Berg K, Cengel KA, Foster TH, Girotti AW, Gollnick SO, et al. Photodynamic therapy of cancer: an update. *CA Cancer J Clin.* (2011) 61:250–81. doi: 10.3322/caac.20114

98. Yang C, Fu Y, Huang C, Hu D, Zhou K, Hao Y, et al. Chlorin e6 and CRISPR-Cas9 dual-loading system with deep penetration for a synergistic tumoral photodynamic-immunotherapy. *Biomaterials.* (2020) 255:120194. doi: 10.1016/j.biomaterials.2020.120194

99. Ji B, Wei M, Yang B. Recent advances in nanomedicines for photodynamic therapy (PDT)-driven cancer immunotherapy. *Theranostics.* (2022) 1):434–58. doi: 10.7150/thno.67300

100. Nkune NW, Abrahamse H. Anti-hypoxia nanoplatforms for enhanced photosensitizer uptake and photodynamic Therapy Effects in Cancer Cells. *Int J Mol Sci.* (2023) 24:2656. doi: 10.3390/ijms24032656

101. Sun B, Bte Rahmat JN, Zhang Y. Advanced techniques for performing photodynamic therapy in deep-seated tissues. *Biomaterials*. (2022) 291:121875. doi: 10.1186/s40364-024-00625-6

102. Zhang M, Zhao Y, Ma H, Sun Y, Cao J. How to improve photodynamic therapy-induced antitumor immunity for cancer treatment? *Theranostics.* (2022) 12:4629–55. doi: 10.7150/thno.72465

103. Topalian SL, Weiner GJ, Pardoll DM. Cancer immunotherapy comes of age. J Clin Oncol. (2011) 29:4828–36. doi: 10.1200/JCO.2011.38.0899

104. Sui C, Wu H, Li X, Wang Y, Wei J, Yu J, et al. Cancer immunotherapy and its facilitation by nanomedicine. *biomark Res.* (2024) 12:77. doi: 10.1186/s40364-024-00625-6

105. Tomar N, De RK. A brief outline of the immune system. *Methods Mol Biol.* (2014) 1184:3–12. doi: 10.1007/978-1-4939-1115-8\_1

106. Iwasaki A, Medzhitov R. Regulation of adaptive immunity by the innate immune system. *Science*. (2010) 327:291–5. doi: 10.1126/science.1183021

107. Strioga M, Schijns V, Powell DJ, Pasukoniene V, Dobrovolskiene N, Michalek J. Dendritic cells and their role in tumor immunosurveillance. *Innate Immun.* (2013) 19:98–111. doi: 10.1177/1753425912449549

108. Bousso P. T-cell activation by dendritic cells in the lymph node: lessons from the movies. *Nat Rev Immunol.* (2008) 8:675–84. doi: 10.1038/nri2379

109. Ferlazzo G, Münz C. NK cell compartments and their activation by dendritic cells. J Immunol. (2004) 172:1333–9. doi: 10.4049/jimmunol.172.3.1333

110. Ghasemi M, Abbasi L, Ghanbari Naeini L, Kokabian P, Nameh Goshay Fard N, Givtaj N. Dendritic cells and natural killer cells: The road to a successful oncolytic virotherapy. *Front Immunol.* (2023) 13:950079/full. doi: 10.3389/fimmu.2022.950079/full

111. Ager A, Watson HA, Wehenkel SC, Mohammed RN. Homing to solid cancers: A vascular checkpoint in adoptive cell therapy using CAR T-cells. *Biochem Soc Trans.* (2016) 44:377–85. doi: 10.1042/BST20150254

112. Jiang H, Jiang J. Balancing act: the complex role of NK cells in immune regulation. Front Immunol. (2023) 14:1275028/full. doi: 10.3389/fimmu.2023.1275028/full

113. Dolina JS, Van Braeckel-Budimir N, Thomas GD, Salek-Ardakani S. CD8+ T cell exhaustion in cancer. *Front Immunol.* (2021) 12:715234. doi: 10.3389/fmmu.2021.715234

114. Pesce S, Greppi M, Tabellini G, Rampinelli F, Parolini S, Olive D, et al. Identification of a subset of human natural killer cells expressing high levels of programmed death 1: A phenotypic and functional characterization. *J Allergy Clin Immunol.* (2017) 139:335–46. doi: 10.1016/j.jaci.2016.04.025

115. Lu Y, Sun W, Du J, Fan J, Peng X. Immuno-photodynamic therapy (IPDT): Organic photosensitizers and their application in cancer ablation. *JACS Au.* (2023) 3:682–99. doi: 10.1021/jacsau.2c00591

116. Liu J, Fu M, Wang M, Wan D, Wei Y, Wei X. Cancer vaccines as promising immuno-therapeutics: platforms and current progress. *J Hematol Oncol*. (2022) 15:28. doi: 10.1186/s13045-022-01247-x

117. Malonis RJ, Lai JR, Vergnolle O. Peptide-based vaccines: Current progress and future challenges. *Chem Rev.* (2020) 120:3210–29. doi: 10.1021/acs.chemrev.9b00472

118. Morse MA, Hobeika AC, Osada T, Berglund P, Hubby B, Negri S, et al. An alphavirus vector overcomes the presence of neutralizing antibodies and elevated numbers of Tregs to induce immune responses in humans with advanced cancer. *J Clin Invest.* (2010) 120:3234–41. doi: 10.1172/JCI42672

119. Toussaint B, Chauchet X, Wang Y, Polack B, Le Gouëllec A. Live-attenuated bacteria as a cancer vaccine vector. *Expert Rev Vaccines*. (2013) 12:1139–54. doi: 10.1586/14760584.2013.836914

120. Guasp P, Reiche C, Sethna Z, Balachandran VP. RNA vaccines for cancer: Principles to practice. *Cancer Cell*. (2024) 42:1163–84. doi: 10.1016/j.ccell.2024.05.005

121. Liao HC, Liu SJ. Advances in nucleic acid-based cancer vaccines. J Biomed Science. (2025) 32:10. doi: 10.1186/s12929-024-01102-w

122. Yang R, Cui J. Advances and applications of RNA vaccines in tumor treatment. *Mol Cancer.* (2024) 23:226.

123. Le DT, Pardoll DM, Jaffee EM. Cellular vaccine approaches. Cancer J. (2010) 16:304–10. doi: 10.1097/PPO.0b013e3181eb33d7

124. Bear AS, Fraietta JA, Narayan VK, O'Hara M, Haas NB. Adoptive cellular therapy for solid tumors. *Am Soc Clin Oncol Educ Book*. (2021) 41:57–65. doi: 10.1200/ EDBK\_321115

125. Rohaan MW, Wilgenhof S, Haanen JBAG. Adoptive cellular therapies: the current landscape. Virchows Arch. (2019) 474:449-61. doi: 10.1007/s00428-018-2484-0

126. Lai J, Mardiana S, House IG, Sek K, Henderson MA, Giuffrida L, et al. Adoptive cellular therapy with T cells expressing the dendritic cell growth factor Flt3L drives epitope spreading and antitumor immunity. *Nat Immunol.* (2020) 21:914–26. doi: 10.1038/s41590-020-0676-7

127. Newick K, O'Brien S, Moon E, Albelda SM. CAR T cell therapy for solid tumors. Annu Rev Med. (2017) 68:139–52. doi: 10.1146/annurev-med-062315-120245

128. Uslu U, June CH. Beyond the blood: expanding CAR T cell therapy to solid tumors. *Nat Biotechnol.* (2025) 43:506–15. doi: 10.1038/s41587-024-02446-2

129. Pardoll DM. The blockade of immune checkpoints in cancer immunotherapy. *Nat Rev Cancer.* (2012) 12:252–64. doi: 10.1038/nrc3239

130. Sharon E, Streicher H, Goncalves P, Chen HX. Immune checkpoint inhibitors in clinical trials. *Chin J Cancer*. (2014) 33:434–44. doi: 10.5732/cjc.014.10122

131. Guan J, Lim KS, Mekhail T, Chang CC. Programmed death ligand-1 (PD-L1) Expression in the programmed death receptor-1 (PD-1)/PD-L1 Blockade: A key player against various cancers. *Arch Pathol Lab Med.* (2017) 141:851–61. doi: 10.5858/ arpa.2016-0361-RA

132. Salama AKS, Hodi FS. Cytotoxic T-lymphocyte-associated antigen-4. Clin Cancer Res. (2011) 17:4622-8. doi: 10.1158/1078-0432.CCR-10-2232

133. He Y, Cao J, Zhao C, Li X, Zhou C, Hirsch FR. TIM-3, a promising target for cancer immunotherapy. *Onco Targets Ther.* (2018) 11:7005–9. doi: 10.2147/OTT.S170385

134. Lythgoe MP, Liu DSK, Annels NE, Krell J, Frampton AE. Gene of the month: lymphocyte-activation gene 3 (LAG-3). *J Clin Pathol.* (2021) 74:543–7. doi: 10.1136/ jclinpath-2021-207517

135. Yu X, Huang X, Chen X, Liu J, Wu C, Pu Q, et al. Characterization of a novel anti-human lymphocyte activation gene 3 (LAG-3) antibody for cancer immunotherapy. *MAbs.* (2019) 11:1139–48. doi: 10.1080/19420862.2019.1629239

136. Buss NAPS, Henderson SJ, McFarlane M, Shenton JM, de Haan L. Monoclonal antibody therapeutics: history and future. *Curr Opin Pharmacol.* (2012) 12:615–22. doi: 10.1016/j.coph.2012.08.001

137. Kontermann RE, Brinkmann U. Bispecific antibodies. Drug Discov Today. (2015) 20:838-47. doi: 10.1016/j.drudis.2015.02.008

138. Kimiz-Gebologlu I, Gulce-Iz S, Biray-Avci C. Monoclonal antibodies in cancer immunotherapy. *Mol Biol Rep.* (2018) 45:2935–40. doi: 10.1007/s11033-018-4427-x

139. Ponziani S, Di Vittorio G, Pitari G, Cimini AM, Ardini M, Gentile R, et al. Antibody-drug conjugates: The new frontier of chemotherapy. *Int J Mol Sci.* (2020) 21:5510. doi: 10.3390/ijms21155510

140. Bayer V. An overview of monoclonal antibodies. *Semin Oncol Nurs.* (2019) 35:150927. doi: 10.1016/j.soncn.2019.08.006

141. Malik B, Ghatol A. Understanding how monoclonal antibodies work. In: *StatPearls*. StatPearls Publishing, Treasure Island (FL (2025). Available online at: http://www.ncbi.nlm.nih.gov/books/NBK572118/.

142. Reichert JM, Rosensweig CJ, Faden LB, Dewitz MC. Monoclonal antibody successes in the clinic. Nat Biotechnol. (2005) 23:1073–8. doi: 10.1038/nbt0905-1073

143. André AS, Dias JNR, Aguiar S, Nogueira S, Bule P, Carvalho JI, et al. Rabbit derived VL single-domains as promising scaffolds to generate antibody-drug conjugates. *Sci Rep.* (2023) 13:4837. doi: 10.1038/s41598-023-31568-x

144. Rahman MM, McFadden G. Oncolytic viruses: Newest frontier for cancer immunotherapy. *Cancers (Basel).* (2021) 13:5452. doi: 10.3390/cancers13215452

145. Zhou X, Hu S, Wang X. Recent advances in oncolytic virus combined immunotherapy in tumor treatment. *Genes Dis.* (. 2025) 12:101599. doi: 10.1016/j.gendis.2025.101599

146. Chen Y, Tao M, Wu X, Tang Z, Zhu Y, Gong K, et al. Current status and research progress of oncolytic virus. *Pharm Sci Advances*. (2024) 2:100037. doi: 10.1016/j.pscia.2024.100037

147. Tian Y, Xie D, Yang L. Engineering strategies to enhance oncolytic viruses in cancer immunotherapy. *Sig Transduct Target Ther.* (2022) 7:1–21. doi: 10.1038/s41392-022-00951-x

148. Bartlett DL, Liu Z, Sathaiah M, Ravindranathan R, Guo Z, He Y, et al. Oncolytic viruses as therapeutic cancer vaccines. *Mol Cancer*. (2013), 12:103. doi: 10.1186/1476-4598-12-103

149. Su Y, Su C, Qin L. Current landscape and perspective of oncolytic viruses and their combination therapies. *Transl Oncol.* (2022) 25:101530. doi: 10.1016/j.tranon.2022.101530

150. Zhang Q, Liu F. Advances and potential pitfalls of oncolytic viruses expressing immunomodulatory transgene therapy for Malignant gliomas. *Cell Death Dis.* (2020) 11:1–11. doi: 10.1038/s41419-019-2182-0

151. Zheng M, Huang J, Tong A, Yang H. Oncolytic viruses for cancer therapy: Barriers and recent advances. *Mol Ther Oncolytics*. (2019) 15:234–47. doi: 10.1016/j.omto.2019.10.007

152. Greig SL. Talimogene Laherparepvec: First global approval. Drugs. (2016) 76:147–54. doi: 10.1007/s40265-015-0522-7

153. Aurelian L. Oncolytic virotherapy: the questions and the promise. *Oncolytic Virother.* (2013) 2:19–29. doi: 10.2147/OV.S39609

154. Duan X, Chan C, Guo N, Han W, Weichselbaum RR, Lin W. Photodynamic therapy mediated by nontoxic core-shell nanoparticles synergizes with immune checkpoint blockade to elicit antitumor immunity and antimetastatic effect on breast cancer. *J Am Chem Soc.* (2016) 138:16686–95. doi: 10.1021/jacs.6b09538

155. Koury J, Lucero M, Cato C, Chang L, Geiger J, Henry D, et al. Immunotherapies: Exploiting the immune system for cancer treatment. *J Immunol Res.* (2018) 2018:9585614. doi: 10.1155/2018/9585614

156. Tan S, Li D, Zhu X. Cancer immunotherapy: Pros, cons and beyond. *BioMed Pharmacother*. (2020) 124:109821. doi: 10.1016/j.biopha.2020.109821

157. Zhang P, Meng J, Li Y, Yang C, Hou Y, Tang W, et al. Nanotechnologyenhanced immunotherapy for metastatic cancer. *Innovation (Camb)*. (2021) 2:100174. doi: 10.1016/j.xinn.2021.100174

158. Wang T, Wang D, Yu H, Feng B, Zhou F, Zhang H, et al. A cancer vaccinemediated postoperative immunotherapy for recurrent and metastatic tumors. *Nat Commun.* (2018) 9:1532. doi: 10.1038/s41467-018-03915-4

159. Darvin P, Toor SM, Sasidharan Nair V, Elkord E. Immune checkpoint inhibitors: recent progress and potential biomarkers. *Exp Mol Med.* (2018) 50:1–11. doi: 10.1038/s12276-018-0191-1

160. Qi Y, Zhang L, Wang Z, Kong X, Zhai J, Fang Y, et al. Efficacy and safety of anti-PD-1/PD-L1 monotherapy for metastatic breast cancer: clinical evidence. *Front Pharmacol.* (2021) 12:653521/full. doi: 10.3389/fphar.2021.653521/full

161. Türeci Ö, Löwer M, Schrörs B, Lang M, Tadmor A, Sahin U. Challenges towards the realization of individualized cancer vaccines. *Nat BioMed Eng.* (2018) 2:566–9. doi: 10.1038/s41551-018-0266-2

162. Ye T, Li F, Ma G, Wei W. Enhancing therapeutic performance of personalized cancer vaccine via delivery vectors. *Advanced Drug Delivery Rev.* (2021) 177:113927. doi: 10.1016/j.addr.2021.113927

163. Depil S, Duchateau P, Grupp SA, Mufti G, Poirot L. Off-the-shelf' allogeneic CAR T cells: development and challenges. *Nat Rev Drug Discov.* (2020) 19:185–99. doi: 10.1038/s41573-019-0051-2

164. Berraondo P, Sanmamed MF, Ochoa MC, Etxeberria I, Aznar MA, Pérez-Gracia JL, et al. Cytokines in clinical cancer immunotherapy. *Br J Cancer*. (2019) 120:6– 15. doi: 10.1038/s41416-018-0328-y

165. Sharma P, Hu-Lieskovan S, Wargo JA, Ribas A. Primary, adaptive, and acquired resistance to cancer immunotherapy. Cell. (2017) 168:707–23. doi: 10.1016/j.cell.2017.01.017

166. Waldman AD, Fritz JM, Lenardo MJ. A guide to cancer immunotherapy: From T cell basic science to clinical practice. *Nat Rev Immunol.* (2020) 20:651–68. doi: 10.1038/s41577-020-0306-5

167. Fan T, Zhang M, Yang J, Zhu Z, Cao W, Dong C. Therapeutic cancer vaccines: Advancements, challenges and prospects. *Sig Transduct Target Ther.* (2023) 8:1–23. doi: 10.1038/s41392-023-01674-3

168. Hollingsworth RE, Jansen K. Turning the corner on therapeutic cancer vaccines. NPJ Vaccines. (2019) 4:1-10. doi: 10.1038/s41541-019-0103-y

169. Wu X, Yang H, Chen X, Gao J, Duan Y, Wei D, et al. Nano-herb medicine and PDT induced synergistic immunotherapy for colon cancer treatment. *Biomaterials*. (2021) 269:120654. doi: 10.1016/j.biomaterials.2021.120654

170. Wu B, Zhang B, Li B, Wu H, Jiang M. Cold and hot tumors: from molecular mechanisms to targeted therapy. *Signal Transduct Target Ther*. (2024) 9:274. doi: 10.1038/s41392-024-01979-x

171. Wang L, Geng H, Liu Y, Liu L, Chen Y, Wu F, et al. Hot and cold tumors: Immunological features and the therapeutic strategies. *MedComm (2020).* (2023) 4: e343. doi: 10.1002/mco2.343

172. Ollauri-Ibáñez C, Ayuso-Íñigo B, Pericacho M. Hot and cold tumors: Is endoglin (CD105) a potential target for vessel normalization? *Cancers (Basel).* (2021) 13:1552. doi: 10.3390/cancers13071552

173. De Guillebon E, Dardenne A, Saldmann A, Séguier S, Tran T, Paolini L, et al. Beyond the concept of cold and hot tumors for the development of novel predictive biomarkers and the rational design of immunotherapy combination. *Int J Cancer.* (2020) 147:1509–18. doi: 10.1002/ijc.32889

174. Chen Y. Nanotechnology for next-generation cancer immunotherapy: State of the art and future perspectives. *J Controlled Release.* (2023) 356:14–25. doi: 10.1016/j.jconrel.2023.02.016

175. Chin AL, Jiang S, Jang E, Niu L, Li L, Jia X, et al. Implanta ble optical fibers for immunotherapeutics delivery and tumor impedance measurement. *Nat Commun.* (2021) 12:5138. doi: 10.1038/s41467-021-25391-z

176. Bonaventura P, Shekarian T, Alcazer V, Valladeau-Guilemond J, Valsesia-Wittmann S, Amigorena S, et al. Cold tumors: A therapeutic challenge for immunotherapy. *Front Immunol.* (2019) 10:168. doi: 10.3389/fimmu.2019.00168

177. Huang J, Yang B, Peng Y, Huang J, Wong SHD, Bian L, et al. Nanomedicineboosting tumor immunogenicity for enhanced immunotherapy. *Advanced Funct Materials.* (2021) 31:2011171. doi: 10.1002/adfm.202011171

178. Ebrahimi N, Akbari M, Ghanaatian M, Roozbahani Moghaddam P, Adelian S, Borjian Boroujeni M, et al. Development of neoantigens: from identification in cancer cells to application in cancer vaccines. *Expert Rev Vaccines*. (2022) 21:941–55. doi: 10.1080/14760584.2021.1951246

179. Innamarato P, Pilon-Thomas S. Reactive Myelopoiesis and the onset of myeloid-mediated immune suppression: Implications for adoptive cell therapy. *Cell Immunol.* (2021) 361:104277. doi: 10.1016/j.cellimm.2020.104277

180. Gunturu KS, Pham TT, Shambhu S, Fisch MJ, Barron JJ, Debono D. Immune checkpoint inhibitors: Immune-related adverse events, healthcare utilization, and costs among commercial and medicare advantage patients. *Support Care Cancer*. (2022) 30:4019–26. doi: 10.1007/s00520-022-06826-9

181. Bugelski PJ, Achuthanandam R, Capocasale RJ, Treacy G, Bouman-Thio E. Monoclonal antibody-induced cytokine-release syndrome. *Expert Rev Clin Immunol.* (2009) 5:499–521. doi: 10.1586/eci.09.31

182. Shimabukuro-Vornhagen A, Gödel P, Subklewe M, Stemmler HJ, Schlößer HA, Schlaak M, et al. Cytokine release syndrome. *J Immunother Cancer.* (2018) 6:56. doi: 10.1186/s40425-018-0343-9

183. Joshi S, Durden DL. Combinatorial approach to improve cancer immunotherapy: Rational drug design strategy to simultaneously hit multiple targets to kill tumor cells and to activate the immune system. *J Oncol.* (2019) 2019:5245034. doi: 10.1155/2019/5245034

184. Anzengruber F, Avci P, de Freitas LF. Hamblin MR. T-cell mediated anti-tumor Immun after Photodyn therapy: Why does it not always work how Can we improve it? Photochem Photobiol Sci. (2015) 14:1492–509. doi: 10.1039/c4pp00455h

185. Xia Y, Gupta GK, Castano AP, Mroz P, Avci P, Hamblin MR. CpG oligodeoxynucleotide as immune adjuvant enhances photodynamic therapy response in murine metastatic breast cancer. *J Biophotonics*. (2014) 0):897–905. doi: 10.1002/jbio.201300072

186. Lintern N, Smith AM, Jayne DG, Khaled YS. Photodynamic stromal depletion in pancreatic ductal adenocarcinoma. *Cancers (Basel).* (2023) 15:4135. doi: 10.3390/ cancers15164135

187. Ahn M, Lee T, Kim KS, Lee S, Na K. Synergistic approach of antibodyphotosensitizer conjugate independent of kras-mutation and its downstream blockade pathway in colorectal cancer. *Adv Healthc Mater.* (2023) 12:e2302374. doi: 10.1002/ adhm.202302374

188. Pye H, Stamati I, Yahioglu G, Butt MA, Deonarain M. Antibody-directed phototherapy (ADP). *Antibodies*. (2013) 2:270–305. doi: 10.3390/antib2020270

189. Sandland J, Boyle R. Photosensitizer antibody-drug conjugates: Past, present, and future. *Bioconjugate Chem.* (2019) 30:975–93. doi: 10.1021/acs.bioconjchem.9b00055

190. Xu X, Deng G, Sun Z, Luo Y, Liu J, Yu X, et al. A Biomimetic aggregationinduced emission photosensitizer with antigen-presenting and hitchhiking function for lipid droplet targeted photodynamic immunotherapy. *Adv Mater*. (2021) 33:e2102322. doi: 10.1002/adma.202102322

191. Vrouenraets MB, Visser GW, Stigter M, Oppelaar H, Snow GB, van Dongen GA. Targeting of aluminum (III) phthalocyanine tetrasulfonate by use of internalizing monoclonal antibodies: improved efficacy in photodynamic therapy. *Cancer Res.* (2001) 61:1970–5.

192. Shirasu N, Shibaguchi H, Yamada H, Kuroki M, Yasunaga S. Highly versatile cancer photoimmunotherapy using photosensitizer-conjugated avidin and biotinconjugated targeting antibodies. *Cancer Cell Int.* (2019) 19:299. doi: 10.1186/s12935-019-1034-4

193. Wolf I, Schultze-Seemann S, Gratzke C, Wolf P. Targeting CD44 and EpCAM with Antibody dye conjugates for the photoimmunotherapy of prostate cancer. *Antibodies.* (2025) 14:5. doi: 10.3390/antib14010005

194. Mitsunaga M, Ogawa M, Kosaka N, Rosenblum LT, Choyke PL, Kobayashi H. Cancer cell-selective *in vivo* near-infrared photoimmunotherapy targeting specific membrane molecules. *Nat Med.* (2011)17:1685–91. doi: 10.1038/nm.2554

195. Jendzelovský R, Mikes J, Koval' J, Soucek K, Procházková J, Kello M, et al. Drug efflux transporters, MRP1 and BCRP, affect the outcome of hypericin-mediated photodynamic therapy in HT-29 adenocarcinoma cells. *Photochem Photobiol Sci.* (2009) 8:1716–23. doi: 10.1039/b9pp00086k

196. Hsu MA, Okamura SM, De Magalhaes Filho CD, Bergeron DM, Rodriguez A, West M, et al. Cancer-targeted photoimmunotherapy induces antitumor immunity and can be augmented by anti-PD-1 therapy for durable anticancer responses in an immunologically active murine tumor model. *Cancer Immunol Immunother*. (2023) 72:151-68. doi: 10.1007/s00262-022-03239-9

197. Mironova KE, Proshkina GM, Ryabova AV, Stremovskiy OA, Lukyanov SA, Petrov RV, et al. Genetically encoded immunophotosensitizer 4D5scFv-miniSOG is a highly selective agent for targeted photokilling of tumor cells *in vitro*. *Theranostics*. (2013) 3:831–40. doi: 10.7150/thno.6715

198. Serebrovskaya EO, Edelweiss EF, Stremovskiy OA, Lukyanov KA, Chudakov DM, Deyev SM. Targeting cancer cells by using an antireceptor antibody-photosensitizer fusion protein. *Proc Natl Acad Sci.* (2009) 106:9221–5. doi: 10.1073/pnas.0904140106

199. van Dongen G a. MS, Visser GWM, Vrouenraets MB. Photosensitizer-antibody conjugates for detection and therapy of cancer. *Adv Drug Delivery Rev.* (2004) 56:31–52. doi: 10.1016/j.addr.2003.09.003

200. Kang X, Guo X, Niu X, An W, Li S, Liu Z, et al. Photothermal therapeutic application of gold nanorods-porphyrin-trastuzumab complexes in HER2-positive breast cancer. *Sci Rep.* (2017) 7:42069. doi: 10.1038/srep42069

201. Rosàs E, Santomá P, Duran-Frigola M, Hernandez B, Llinàs MC, Ruiz-González R, et al. Modifications of microvascular EC surface modulate phototoxicity of a porphycene anti-ICAM-1 immunoconjugate; Therapeutic implications. *Langmuir.* (2013) 29:9734–43. doi: 10.1021/la401067d

202. Sadiki A, Kercher EM, Lu H, Lang RT, Spring BQ, Zhou ZS. Site-specific bioconjugation and convergent click chemistry enhances antibody-chromophore conjugate binding efficiency. *Photochem Photobiol.* (2020) 96:596–603. doi: 10.1111/php.13231

203. Ito K, Mitsunaga M, Arihiro S, Saruta M, Matsuoka M, Kobayashi H, et al. Molecular targeted photoimmunotherapy for HER2-positive human gastric cancer in combination with chemotherapy results in improved treatment outcomes through different cytotoxic mechanisms. *BMC Cancer.* (2016) 16:37. doi: 10.1186/s12885-016-2072-0

204. Abu-Yousif AO, Moor ACE, Zheng X, Savellano MD, Yu W, Selbo PK, et al. Epidermal growth factor receptor-targeted photosensitizer selectively inhibits EGFR signaling and induces targeted phototoxicity in ovarian cancer cells. *Cancer Lett.* (2012) 321:120–7. doi: 10.1016/j.canlet.2012.01.014

205. Pereira PMR, Carvalho JJ, Silva S, Cavaleiro JAS, Schneider RJ, Fernandes R, et al. Porphyrin conjugated with serum albumins and monoclonal antibodies boosts efficiency in targeted destruction of human bladder cancer cells. *Org Biomol Chem.* (2014) 12:1804–11. doi: 10.1039/c3ob42082e

206. Kim KS, Kim J, Kim DH, Hwang HS, Na K. Multifunctional trastuzumabchlorin e6 conjugate for the treatment of HER2-positive human breast cancer. *Biomater Sci.* (2018) 6:1217–26. doi: 10.1039/C7BM01084B

207. Adil Butt M, Pye H, Haidry RJ, Oukrif D, Khan SUR, Puccio I, et al. Upregulation of mucin glycoprotein MUC1 in the progression to esophageal adenocarcinoma and therapeutic potential with a targeted photoactive antibody-drug conjugate. *Oncotarget.* (2017) 8:25080–96. doi: 10.18632/oncotarget.15340

208. Staneloudi C, Smith KA, Hudson R, Malatesti N, Savoie H, Boyle RW, et al. Development and characterization of novel photosensitizer: scFv conjugates for use in photodynamic therapy of cancer. *Immunology.* (2007) 120:512–7. doi: 10.1111/j.1365-2567.2006.02522.x

209. Malatesti N, Smith K, Savoie H, Greenman J, Boyle RW. Synthesis and *in vitro* investigation of cationic 5,15-diphenyl porphyrin-monoclonal antibody conjugates as targeted photodynamic sensitizers. *Int J Oncol.* (2006) 28:1561–9. doi: 10.3892/ijo.28.6.1561

210. Hudson R, Carcenac M, Smith K, Madden L, Clarke OJ, Pèlegrin A, et al. The development and characterization of porphyrin isothiocyanate-monoclonal antibody conjugates for photoimmunotherapy. *Br J Cancer.* (2005) 92:1442–9. doi: 10.1038/ sj.bjc.6602517

211. Alonso CMA, Palumbo A, Bullous AJ, Pretto F, Neri D, Boyle RW. Site-specific and stoichiometric conjugation of cationic porphyrins to antiangiogenic monoclonal antibodies. *Bioconjug Chem.* (2010) 21:302–13. doi: 10.1021/bc9003537

212. Bryden F, Maruani A, Savoie H, Chudasama V, Smith MEB, Caddick S, et al. Regioselective and stoichiometrically controlled conjugation of photodynamic sensitizers to a HER2 targeting antibody fragment. *Bioconjug Chem.* (2014) 25:611–7. doi: 10.1021/bc5000324

213. Maruani A, Savoie H, Bryden F, Caddick S, Boyle R, Chudasama V. Siteselective multi-porphyrin attachment enables the formation of a next-generation antibody-based photodynamic therapeutic. *Chem Commun.* (2015) 51:15304–7. doi: 10.1039/C5CC06985H

214. García-Álvarez J, Gimeno J. "Click" copper catalyzed azide-alkyne cycloaddition (CuAAC) in aqueous medium, in: *dvances in organometallic chemistry and catalysis* (2013). John Wiley & Sons, Ltd.

215. Bryden F, Maruani A, Rodrigues JMM, Cheng MHY, Savoie H, Beeby A, et al. Assembly of high-potency photosensitizer–antibody conjugates through application of dendron multiplier technology. *Bioconjugate Chem.* (2018) 29:176–81. doi: 10.1021/acs.bioconjchem.7b00678

216. Obaid G, Chambrier I, Cook MJ, Russell DA. Cancer targeting with biomolecules: a comparative study of photodynamic therapy efficacy using antibody or lectin conjugated phthalocyanine-PEG gold nanoparticles. *Photochem Photobiol Sci.* (2015) 14:737–47. doi: 10.1039/c4pp00312h

217. Patinote C, Karroum NB, Moarbess G, Cirnat N, Kassab I, Bonnet PA, et al. Agonist and antagonist ligands of toll-like receptors 7 and 8: Ingenious tools for therapeutic purposes. *Eur J Med Chem.* (2020) 193:112238. doi: 10.1016/j.ejmech.2020.112238

218. Kim H, Griffith TS, Panyam J. Poly(d,l-lactide-co-glycolide) nanoparticles as delivery platforms for TLR7/8 agonist-based cancer vaccine. *J Pharmacol Exp Ther.* (2019) 370:715–24. doi: 10.1124/jpet.118.254953

219. Bhatta AK, Wang P, Keyal U, Zhao Z, Ji J, Zhu L, et al. Therapeutic effect of imiquimod enhanced ALA-PDT on cutaneous squamous cell carcinoma. *Photodiagnosis Photodyn Ther.* (2018) 23:273–80. doi: 10.1016/j.pdptt.2018.07.010

220. Ding L, Luo K, Feng CG, Oehlers SH. Pidotimod increases inflammation in wounded zebrafish embryos. *Fish Shellfish Immunol.* (2022) 120:429–33. doi: 10.1016/j.fsi.2021.12.013

221. Dudzik T, Domański I, Makuch S. The impact of photodynamic therapy on immune system in cancer – an update. *Front Immunol.* (2024) 15:1335920. doi: 10.3389/fimmu.2024.1335920

222. Mroz P, Hamblin MR. The immunosuppressive side of PDT. Photochem Photobiol Sci. (2011) 10:751-8. doi: 10.1039/c0pp00345j

223. Korbelik M. Immunotherapy regimens for combination with photodynamic therapy aimed at eradication of solid cancers (2000). Available online at: https://ui. adsabs.harvard.edu/abs/2000SPIE.3914.16K (Accessed April 14, 2025).

224. Tiveron RDR, Costa DA, Leite MDI, Vaz CBS, Sousa M, Carlos SMCF, et al. Evaluation of cell damage and modulation of cytokines TNF-α, IL-6, and IL-10 in macrophages exposed to PpIX-mediated photodynamic therapy. *Braz J Biol.* (2019), 80:497–505. doi: 10.1590/1519-6984.193748

225. Korbelik M. Cancer vaccines generated by photodynamic therapy. *Photochem Photobiol Sci.* (2011) 10:664–9. doi: 10.1039/c0pp00343c

226. Korbelik M, Banáth J, Zhang W, Gallagher P, Hode T, Lam SSK, et al. Ndihydrogalactochitosan as immune and direct antitumor agent amplifying the effects of photodynamic therapy and photodynamic therapy-generated vaccines. *Int Immunopharmacol.* (2019) 75:105764. doi: 10.1016/j.intimp.2019.105764

227. Liu X, Liu Y, Li X, Huang J, Guo X, Zhang J, et al. ER-targeting PDT converts tumors into in *situ* therapeutic tumor vaccines. *ACS Nano*. (2022) 16:9240–53. doi: 10.1021/acsnano.2c01669

228. Vedunova M, Turubanova V, Vershinina O, Savyuk M, Efimova I, Mishchenko T, et al. DC vaccines loaded with glioma cells killed by photodynamic therapy induce Th17 anti-tumor immunity and provide a four-gene signature for glioma prognosis. *Cell Death Dis.* (2022) 13:1062. doi: 10.1038/s41419-022-05514-0

229. Korbelik M, Banáth J, Saw KM. Immunoregulatory cell depletion improves the efficacy of photodynamic therapy-generated cancer vaccines. *Int J Mol Sci.* (2015) 16:27005–14. doi: 10.3390/ijms161126008

230. Korbelik M. Optimization of whole tumor cell vaccines by interaction with phagocytic receptors. Vaccines (Basel). (2021) 9:904. doi: 10.3390/vaccines9080904

231. Korbelik M, Banáth J, Zhang W, Hode T, Lam SSK, Gallagher P, et al. Ndihydrogalactochitosan-supported tumor control by photothermal therapy and photothermal therapy-generated vaccine. *J Photochem Photobiol B: Biol.* (2020) 204:111780. doi: 10.1016/j.jphotobiol.2020.111780

232. Korbelik M, Stott B, Sun J. Photodynamic therapy-generated vaccines: relevance of tumour cell death expression. *Br J Cancer.* (2007) 97:1381–7. doi: 10.1038/sj.bjc.6604059

233. Gollnick SO, Vaughan L, Henderson BW. Generation of effective antitumor vaccines using photodynamic therapy. *Cancer Res.* (2002) 62:1604–8.

234. Ni J, Song J, Wang B, Hua H, Zhu H, Guo X, et al. Dendritic cell vaccine for the effective immunotherapy of breast cancer. *BioMed Pharmacother*. (2020) 126:110046. doi: 10.1016/j.biopha.2020.110046

235. Sharma P, Goswami S, Raychaudhuri D, Siddiqui BA, Singh P, Nagarajan A, et al. Immune checkpoint therapy—Current perspectives and future directions. *Cell.* (2023) 186:1652–69. doi: 10.1016/j.cell.2023.03.006

236. Aversa I, Malanga D, Fiume G, Palmieri C. Molecular T-cell repertoire analysis as source of prognostic and predictive biomarkers for checkpoint blockade immunotherapy. *Int J Mol Sci.* (2020) 21:2378. doi: 10.3390/ijms21072378

237. Chan TA, Yarchoan M, Jaffee E, Swanton C, Quezada SA, Stenzinger A, et al. Development of tumor mutation burden as an immunotherapy biomarker: utility for the oncology clinic. *Ann Oncol.* (2019) 30:44–56. doi: 10.1093/annonc/mdy495

238. Dall'Olio FG, Marabelle A, Caramella C, Garcia C, Aldea M, Chaput N, et al. Tumour burden and efficacy of immune-checkpoint inhibitors. *Nat Rev Clin Oncol.* (2022) 19:75–90. doi: 10.1038/s41571-021-00564-3

239. Katsuya H, Sano H, Sano H, Mihashi T, Nakashima C, Kai K, et al. Case report: Efficacy of immune checkpoint inhibitors for high tumour mutational burden Malignant phyllodes tumours of the breast as revealed by comprehensive genomic profiling. *Front Immunol.* (2025) 16:1549452. doi: 10.3389/fimmu.2025.1549452

240. Alvim RG, Georgala P, Nogueira L, Somma AJ, Nagar K, Thomas J, et al. Combined OX40 agonist and PD-1 Inhibitor immunotherapy improves the efficacy of vascular targeted photodynamic therapy in a urothelial tumor model. *Molecules*. (2021) 26:3744. doi: 10.3390/molecules26123744

241. Cramer GM, Moon EK, Cengel KA, Busch TM. Photodynamic therapy and immune checkpoint blockade†. *Photochem Photobiol.* (2020) 96:954–61. doi: 10.1111/php.13300

242. Hao Y, Chung CK, Gu Z, Schomann T, Dong X, Veld RVH, et al. Combinatorial therapeutic approaches of photodynamic therapy and immune checkpoint blockade for colon cancer treatment. *Mol Biomed.* (2022) 3:26. doi: 10.1186/s43556-022-00086-z

243. Peng CL, Lin HC, Chiang WL, Shih YH, Chiang PF, Luo TY, et al. Antiangiogenic treatment (Bevacizumab) improves the responsiveness of photodynamic therapy in colorectal cancer. *Photodiagnosis Photodyn Ther.* (2018) 23::111–8. doi: 10.1016/j.pdpdt.2018.06.008

244. Kim S, Kim SA, Nam GH, Hong Y, Kim GB, Choi Y, et al. *In situ* immunogenic clearance induced by a combination of photodynamic therapy and rho-kinase inhibition sensitizes immune checkpoint blockade response to elicit systemic antitumor immunity against intraocular melanoma and its metastasis. *J Immunother Cancer.* (2021) 9:e001481. doi: 10.1136/jitc-2020-001481

245. Aebisher D, Przygórzewska A, Bartusik-Aebisher D. The latest look at PDT and immune checkpoints. *Curr Issues Mol Biol.* (2024) 46:7239–57. doi: 10.3390/cimb46070430

246. Ouyang G, Xiong L, Liu Z, Lam B, Bui B, Ma L, et al. Inhibition of autophagy potentiates the apoptosis-inducing effects of photodynamic therapy on human colon cancer cells. *Photodiagnosis Photodyn Ther.* (2018) 21:396–403. doi: 10.1016/j.pdpdt.2018.01.010

247. Korbelik M, Naraparaju VR, Yamamoto N. Macrophage-directed immunotherapy as adjuvant to photodynamic therapy of cancer. *Br J Cancer*. (1997) 75:202–7. doi: 10.1038/bjc.1997.34

248. Corzo CA, Condamine T, Lu L, Cotter MJ, Youn JI, Cheng P, et al. HIF-1 $\alpha$  regulates function and differentiation of myeloid-derived suppressor cells in the tumor microenvironment. *J Exp Med.* (2010) 207:2439–53. doi: 10.1084/jem.20100587

249. Noman MZ, Desantis G, Janji B, Hasmim M, Karray S, Dessen P, et al. PD-L1 is a novel direct target of HIF-1 $\alpha$ , and its blockade under hypoxia enhanced MDSC-mediated T cell activation. *J Exp Med.* (2014) 211:781–90. doi: 10.1084/jem.20131916

250. Monaco H, Yokomizo S, Choi HS, Kashiwagi S. Quickly evolving near-infrared photoimmunotherapy provides multifaceted approach to modern cancer treatment. *VIEW*. (2022) 3:20200110. doi: 10.1002/VIW.20200110

251. Naing A, Hajjar J, Gulley JL, Atkins MB, Ciliberto G, Meric-Bernstam F, et al. Strategies for improving the management of immune-related adverse events. *J Immunother Cancer.* (2020) 8:e001754. doi: 10.1136/jitc-2020-001754

252. Li Z, Lai X, Fu S, Ren L, Cai H, Zhang H, et al. Immunogenic cell death activates the tumor immune microenvironment to boost the immunotherapy efficiency. *Advanced Science*. (2022) 9:2201734. doi: 10.1002/advs.202201734

253. Li L, Bian L, Kou N, Yuan Y, Zou H. Special immune-related adverse events and subsequent photodynamic therapy in tislelizumab treatment for esophageal cancer: a case report. *Front Immunol.* (2024) 15:1497259. doi: 10.3389/fimmu.2024.1497259

254. Ashoori MD, Suzuki K, Tokumaru Y, Ikuta N, Tajima M, Honjo T, et al. Inactivation of the PD-1-dependent immunoregulation in mice exacerbates contact hypersensitivity resembling immune-related adverse events. *Front Immunol.* (2020) 11:618711. doi: 10.3389/fimmu.2020.618711

255. Hong EJ, Choi DG, Shim MS. Targeted and effective photodynamic therapy for cancer using functionalized nanomaterials. *Acta Pharm Sin B.* (2016) 6:297–307. doi: 10.1016/j.apsb.2016.01.007

256. Li G, Wang C, Jin B, Sun T, Sun K, Wang S, et al. Advances in smart nanotechnology-supported photodynamic therapy for cancer. *Cell Death Discov.* (2024) 10:466. doi: 10.1038/s41420-024-02236-4

257. Jia J, Wu X, Long G, Yu J, He W, Zhang H, et al. Revolutionizing cancer treatment: nanotechnology-enabled photodynamic therapy and immunotherapy with advanced photosensitizers. *Front Immunol.* (2023) 14:1219785. doi: 10.3389/fimmu.2023.1219785

258. Li H, Fu Q, Muluh TA, Shinge SAU, Fu S, Wu J. The application of nanotechnology in immunotherapy based combinations for cancer treatment. *Recent Pat Anticancer Drug Discov.* (2023) 18:53–65. doi: 10.2174/1574892817666220308090954

259. Zhu J, Lee H, Huang R, Zhou J, Zhang J, Yang X, et al. Harnessing nanotechnology for cancer treatment. *Front Bioeng Biotechnol.* (2025) 12:1514890. doi: 10.3389/fbioe.2024.1514890

260. Deng H, Zhou Z, Yang W, Lin LS, Wang S, Niu G, et al. Endoplasmic reticulum targeting to amplify immunogenic cell death for cancer immunotherapy. *Nano Lett.* (2020) 20:1928–33. doi: 10.1021/acs.nanolett.9b05210

261. van Driel PBAA, Boonstra MC, Slooter MD, Heukers R, Stammes MA, Snoeks TJA, et al. EGFR targeted nanobody-photosensitizer conjugates for photodynamic therapy in a pre-clinical model of head and neck cancer. *J Control Release.* (2016) 229:93–105. doi: 10.1016/j.jconrel.2016.03.014

262. Hanaoka H, Nakajima T, Sato K, Watanabe R, Phung Y, Gao W, et al. Photoimmunotherapy of hepatocellular carcinoma-targeting Glypican-3 combined with nanosized albumin-bound paclitaxel. *Nanomedicine (Lond)*. (2015) 10:1139–47. doi: 10.2217/nnm.14.194

263. Zheng X, Liu X, Lei Y, Wang G, Liu M. Glypican-3: A novel and promising target for the treatment of hepatocellular carcinoma. *Front Oncol.* (2022) 12:824208/ full. doi: 10.3389/fonc.2022.824208/full

264. Sano K, Nakajima T, Choyke PL, Kobayashi H. Markedly enhanced permeability and retention effects induced by photo-immunotherapy of tumors. *ACS Nano*. (2013) 7:717–24. doi: 10.1021/nn305011p

265. Han Y, An Y, Jia G, Wang X, He C, Ding Y, et al. Theranostic micelles based on upconversion nanoparticles for dual-modality imaging and photodynamic therapy in hepatocellular carcinoma. *Nanoscale*. (2018) 10:6511–23. doi: 10.1039/C7NR09717D

266. Gires O, Pan M, Schinke H, Canis M, Baeuerle PA. Expression and function of epithelial cell adhesion molecule EpCAM: where are we after 40 years? *Cancer Metastasis Rev.* (2020) 39:969–87. doi: 10.1007/s10555-020-09898-3

267. Xiao D, Xiong M, Wang X, Lyu M, Sun H, Cui Y, et al. Regulation of the function and expression of epcam. *Biomedicines.* (2024) 12:1129. doi: 10.3390/biomedicines12051129

268. Liu P, Zhang Q, Liu F. Biological roles and clinical applications of EpCAM in HCC. *Discov Oncol.* (2025) 16:319. doi: 10.1007/s12672-025-02095-0

269. Vagena IA, Malapani C, Gatou MA, Lagopati N, Pavlatou EA. Enhancement of EPR Effect for passive tumor targeting: Current status and future perspectives. *Appl Sci.* (2025) 15:3189. doi: 10.3390/app15063189

270. Wu J. The enhanced permeability and retention (EPR) effect: The significance of the concept and methods to enhance its application. *J Pers Med.* (2021) 11:771. doi: 10.3390/jpm11080771

271. Liu D, Chen B, Mo Y, Wang Z, Qi T, Zhang Q, et al. redox-activated porphyrinbased liposome remote-loaded with indoleamine 2,3-dioxygenase (IDO) Inhibitor for synergistic photoimmunotherapy through induction of immunogenic cell death and blockage of IDO pathway. *Nano Lett.* (2019) 19:6964–76. doi: 10.1021/ acs.nanolett.9b02306

272. Zhen Z, Tang W, Wang M, Zhou S, Wang H, Wu Z, et al. Protein nanocage mediated fibroblast-activation protein targeted photoimmunotherapy to enhance cytotoxic T cell infiltration and tumor control. *Nano Lett.* (2017) 17:862–9. doi: 10.1021/acs.nanolett.6b04150

273. Guo T, Xu J. Cancer-associated fibroblasts: A versatile mediator in tumor progression, metastasis, and targeted therapy. *Cancer Metastasis Rev.* (2024) 43:1095–116. doi: 10.1007/s10555-024-10186-7

274. Jia H, Chen X, Zhang L, Chen M. Cancer associated fibroblasts in cancer development and therapy. *J Hematol Oncol.* (2025) 18:36. doi: 10.1186/s13045-025-01688-0

275. Yang D, Liu J, Qian H, Zhuang Q. Cancer-associated fibroblasts: From basic science to anticancer therapy. *Exp Mol Med.* (2023) 55:1322–32. doi: 10.1038/s12276-023-01013-0

276. Sakemura R, Hefazi M, Siegler EL, Cox MJ, Larson DP, Hansen MJ, et al. Targeting cancer-associated fibroblasts in the bone marrow prevents resistance to CART-cell therapy in multiple myeloma. *Blood.* (2022) 139:3708–21. doi: 10.1182/ blood.2021012811

277. Tran E, Chinnasamy D, Yu Z, Morgan RA, Lee CCR, Restifo NP, et al. Immune targeting of fibroblast activation protein triggers recognition of multipotent bone marrow stromal cells and cachexia. *J Exp Med.* (2013) 210:1125–35. doi: 10.1084/jem.20130110

278. Weijing Y, Zhang F, Deng H, Lin L, Wang S, Kang F, et al. Smart nanovesiclemediated immunogenic cell death through tumor microenvironment modulation for effective photodynamic immunotherapy. ACS Nano. (2020) 14:620–31. doi: 10.1021/acsnano.9b07212

279. Ortmann BM. Hypoxia-inducible factor in cancer: from pathway regulation to therapeutic opportunity. *bmjonc*. (2024) 3:e000154. doi: 10.1136/bmjonc-2023-000154

280. Wang H, Li J, Wang Y, Gong X, Xu X, Wang J, et al. Nanoparticles-mediated reoxygenation strategy relieves tumor hypoxia for enhanced cancer therapy. *J Controlled Release*. (2020) 319:25–45. doi: 10.1016/j.jconrel.2019.12.028

281. He M, Zhang M, Xu T, Xue S, Li D, Zhao Y, et al. Enhancing photodynamic immunotherapy by reprograming the immunosuppressive tumor microenvironment with hypoxia relief. *J Controlled Release*. (2024) 368:233–50.

282. Liang R, Liu L, He H, Chen Z, Han Z, Luo Z, et al. Oxygen-boosted immunogenic photodynamic therapy with gold nanocages@manganese dioxide to inhibit tumor growth and metastases. *Biomaterials*. (2018) 177::149-60. doi: 10.1016/j.biomaterials.2018.05.051

283. Song W, Kuang J, Li CX, Zhang M, Zheng D, Zeng X, et al. Enhanced immunotherapy based on photodynamic therapy for both primary and lung metastasis tumor eradication. *ACS Nano*. (2018) 12:1978-89. doi: 10.1021/acsnano.7b09112

284. Zaoqu I, Chen J, Ren Y, Liu S, Ba Y, Zuo A, et al. Multi-stage mechanisms of tumor metastasis and therapeutic strategies. *Sig Transduct Target Ther*. (2024) 9:1–32. doi: 10.1038/s41392-024-01955-5

285. Xu J, Xu L, Wang C, Yang R, Zhuang Q, Han X, et al. Near-infrared-triggered photodynamic therapy with multitasking upconversion nanoparticles in combination with checkpoint blockade for immunotherapy of colorectal cancer. *ACS Nano.* (2017) 11:4463–74. doi: 10.1021/acsnano.7b00715

286. Zhang R, Zhu Z, Lv H, Li F, Sun S, Li J, et al. Immune checkpoint blockade mediated by a small-molecule nanoinhibitor targeting the PD-1/PD-L1 pathway synergizes with photodynamic therapy to elicit antitumor immunity and antimetastatic effects on breast cancer. *Small.* (2019) 15:e1903881. doi: 10.1002/smll.201903881

287. Guo JC, An Q, Guo M, Xiao Y, Li B, Gao F, et al. Oxygen-independent free radical generation mediated by core-shell magnetic nanocomposites synergizes with immune checkpoint blockade for effective primary and metastatic tumor treatment. *Nano Today*. (2021) 36:101024. doi: 10.1016/j.nantod.2020.101024

288. Yue J, Mei Q, Wang P, Miao P, Dong WF, Li L. Light-triggered multifunctional nanoplatform for efficient cancer photo-immunotherapy. *J Nanobiotechnology*. (2022) 20:181. doi: 10.1186/s12951-022-01388-8