



OPEN ACCESS

EDITED BY

Yang Liu,
China Medical University, China

REVIEWED BY

Shaohua Qi,
Houston Methodist Research Institute,
United States
Jie Xian,
University of California, San Diego,
United States
Pablo Damian-Matsumura,
Universidad Autónoma Metropolitana, Mexico

*CORRESPONDENCE

Ming-Yu Wang
✉ wangmy@lzu.edu.cn
Cai-Juan Bai
✉ Baicj1985@hotmail.com

RECEIVED 07 June 2025

ACCEPTED 30 August 2025

PUBLISHED 12 September 2025

CITATION

Zhang C-M, Ge Z-B, Zhou H-H, Wei M-X,
Ding X-Y, Lin Z-Z, Wang M-Y and Bai C-J
(2025) Sex chromosomes/hormones and the
tumor microenvironment of non-
reproductive cancers.
Front. Immunol. 16:1642956.
doi: 10.3389/fimmu.2025.1642956

COPYRIGHT

© 2025 Zhang, Ge, Zhou, Wei, Ding, Lin, Wang
and Bai. This is an open-access article
distributed under the terms of the [Creative
Commons Attribution License \(CC BY\)](#). The
use, distribution or reproduction in other
forums is permitted, provided the original
author(s) and the copyright owner(s) are
credited and that the original publication in
this journal is cited, in accordance with
accepted academic practice. No use,
distribution or reproduction is permitted
which does not comply with these terms.

Sex chromosomes/hormones and the tumor microenvironment of non- reproductive cancers

Chun-Miao Zhang¹, Zhong-Bo Ge², Hai-Hong Zhou^{3,4},
Meng-Xiao Wei², Xin-Yuan Ding², Zhe-Zheng Lin²,
Ming-Yu Wang^{2*} and Cai-Juan Bai^{5,6*}

¹Key Laboratory of Preclinical Study for New Drugs of Gansu Province, Institute of Biochemistry and Molecular Biology, School of Basic Medical Sciences, Lanzhou University, Lanzhou, China, ²Ministry Of Education Key Laboratory of Cell Activities and Stress Adaptations, School of Life Sciences, Lanzhou University, Lanzhou, China, ³Centre for Translational Medicine, Gansu Provincial Academic Institute for Medical Research, Lanzhou, China, ⁴Centre for Translational Medicine, Sun Yat-Sen University Cancer Center Gansu Hospital, Lanzhou, China, ⁵National Health Commission Key Laboratory of Diagnosis and Therapy of Gastrointestinal Tumor, Gansu Provincial Hospital, Lanzhou, China, ⁶The Institute of Clinical Research and Translational Medicine, Gansu Provincial Hospital, Lanzhou, China

Cancer exhibits profound sexual dimorphism in incidence and therapeutic outcomes, driven by the interplay between biological sex determinants and immune regulation. Besides established environmental risk factors (e.g., male-predominant smoking/alcohol consumption), emerging evidence identifies the tumor immune microenvironment (TIME) as a pivotal mediator of sex disparities in carcinogenesis and immunotherapy response. This review synthesizes recent advances in two fundamental mechanisms: (1) Sex chromosome biology: Recent studies delineate the Ubiquitous loss of chromosome Y (LOY) of male cancers that promotes immunosuppressive TIME remodeling, while X-chromosome inactivation escape in females enhances antitumor immunity; (2) Endocrine regulation: Androgen receptor signaling induces T-cell exhaustion via PD-1 transcriptional activation in males. Estrogen-ER α boosts cancer progression via PD-L1 high expression, whereas ER β inhibits cancer progression via CD8⁺ T cell activation in females. This mechanistic synthesis provides actionable strategies for precision immuno-oncology trials targeting sex-based immunological divergence.

KEYWORDS

sexual dimorphism, tumor microenvironment, loss of Y chromosome, X-chromosome inactivation escape, sex hormone, antitumor immunity, non-reproductive cancer

1 Introduction

Cancer remains a leading cause of global mortality (1). Population-based studies reveal significant sex-based disparities in the incidence of most non-reproductive cancers (2). This dichotomy extends to mortality patterns, where male predominance persists in lung, colorectal, and gastric cancers (3, 4). Accumulating evidence indicates that cancer-related sex disparities are mediated through multifactorial mechanisms encompassing lifestyle exposures, chromosomal determinants and hormonal regulation (5–7).

Lifestyle factors (dietary patterns, smoking and alcohol consumption) contribute to sex-specific cancer disparities. For instance, a prospective cohort study demonstrates that low-fat/high-fiber dietary patterns significantly reduce the risk of colorectal cancer specifically in males, suggesting biological susceptibility to diet-modulated carcinogenesis (8). Similarly, nationwide registry data identify persistent smoking disparities as drivers of elevated lung cancer mortality in Chinese males (3). In addition, multiple population-based cohort studies have demonstrated significantly higher alcohol-associated cancer mortality (including primary liver cancer, colorectal cancer, and esophageal cancer) in males compared to females (9, 10). This disparity primarily stems from the higher prevalence of risk-lifestyle among male populations. However, following rigorous adjustment for lifestyle confounders, epidemiological analyses consistently demonstrate persistently elevated incidence and mortality ratios of multiple malignancies in males, which are potentially mediated by sex-specific chromosome determinants or hormonal regulation (2, 11–13).

Sex chromosome complement constitutes key determinants of sex-based cancer disparities (12–14) through multilayered regulatory mechanisms. Notably, A subset of X-chromosome genes can escape X-inactivation, which would protect females from complete functional loss and confer relative tolerance to carcinogenesis (14). Conversely, males exhibit X-monosomy vulnerability, so X-linked tumor suppressor loss-of-function mutations would directly drive carcinogenesis. Beyond these cell-autonomous effects, emerging evidence highlights the pivotal regulatory roles of immune microenvironment remodeling in oncogenesis and its progression (15–17). Notably, the sex chromosome harbors a large number of immune-related genes and exerts cancer-modulating effects through spatiotemporal reprogramming of tumor-immune interfaces (18). Concurrently, sex steroids, including estrogens and androgens, have profound effects on immune function which could affect autoimmunity, allergy, infectious diseases, and cancers (19).

The tumor immune microenvironment exhibits sex-specific remodeling through chromosomal dosage effects (XX vs. XaY) and steroid hormone signaling gradients. These molecular mechanisms partially explain the observed sexual dimorphism in cancer incidence and treatment outcomes. Research based on the Four Core Genotypes (FCG) model (20, 21) reveals that sex chromosomes and sex hormones often coordinate or compensate for regulation (22). Notwithstanding these complexities, this review systematizes current findings on the regulatory dynamics of sex

chromosomes and sex hormones within the tumor immune microenvironment. This review underscores the imperative to recognize sexual dimorphism in cancer pathogenesis, advocating for the integration of sex-stratified precision therapeutic frameworks to optimize clinical outcomes through personalized intervention paradigms.

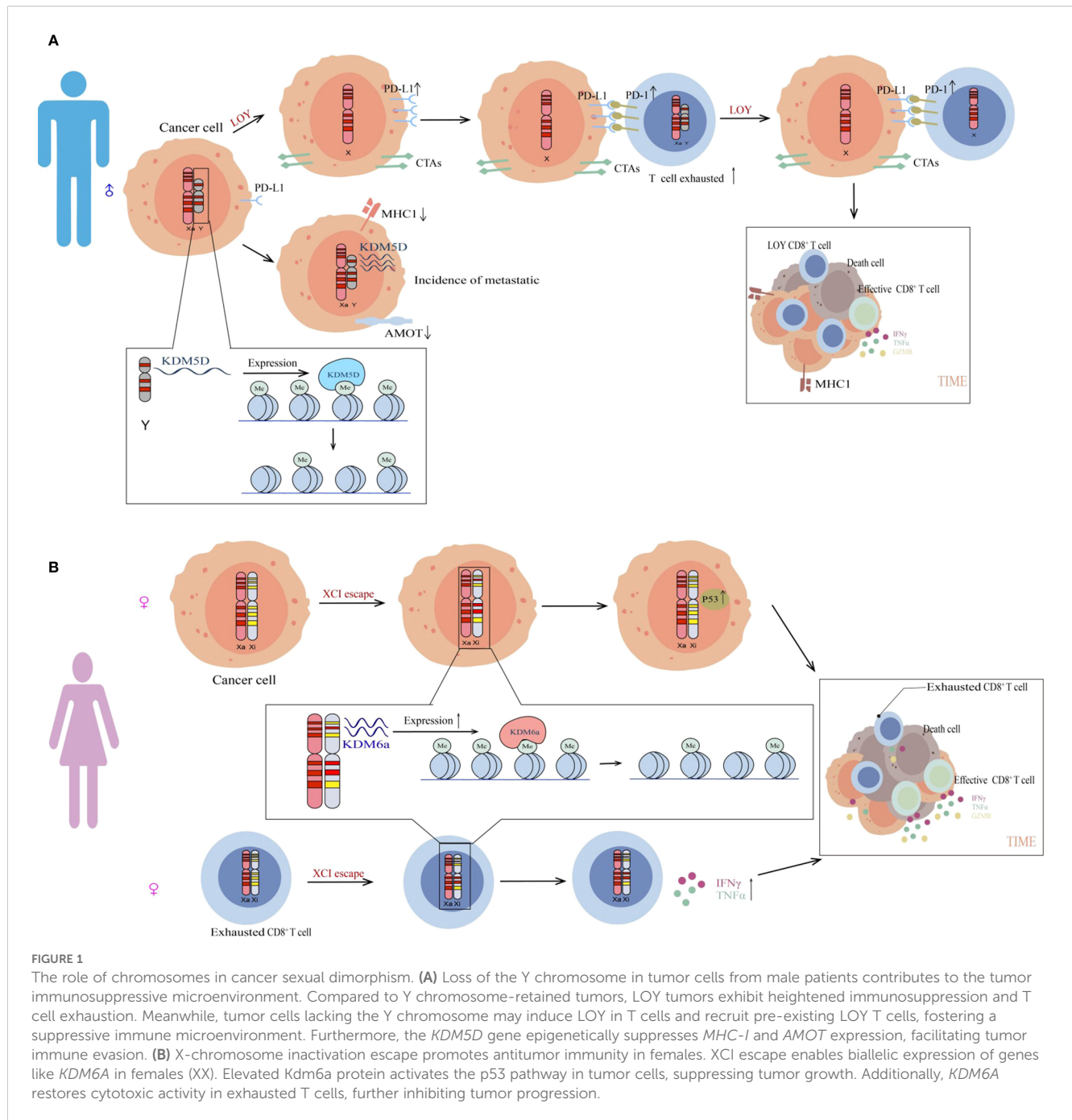
2 Sex chromosome-mediated sexual dimorphism in cancer

The X and Y chromosomes constitute distinct tumor microenvironments, with Y-chromosome loss events and X-chromosome inactivation (XCI) escape mechanisms.

The emergence of sexual dimorphism in cancer incidence and outcomes is mechanistically driven by sex-biased reconfiguration of the tumor microenvironment, particularly involving dysregulation of T cell exhaustion and cancer/testis antigens (CTAs). Deciphering the interplay between sex chromosome dynamics (LOY, XCI escape) and immunotherapy responsiveness will advance the development of sex-specific biomarkers and targeted therapeutic strategies.

2.1 Y chromosome

LOY represents a common somatic alteration in male cancer patients, frequently correlating with poor prognosis in elderly males (23–26). Emerging research reveals substantial overlap between LOY-associated genomic variants and known cancer susceptibility loci, somatic drivers of tumor progression, and genes targeted by approved or investigational anticancer therapies (27). The analysis of The Cancer Genome Atlas Program (TCGA) database cohort suggests that LOY is present in early-stage cancers and serves as a poor prognostic indicator for various tumors in men (28). In a pivotal study investigating the association of LOY with adverse outcomes in bladder cancer, TCGA data analysis demonstrated that patients with reduced expression of Y chromosome-encoded genes (e.g., *KDM5D* and *UTY* also known as *KDM6C*) exhibited shorter survival, with LOY detectable even in early-stage malignancies (29). LOY drives tumor immunosuppression by altering secretory factors or surface molecules in cancer cells, upregulating T cell exhaustion markers (e.g., *TOX*, *TIM-3*, *LAG-3*), and recruiting M2-polarized macrophages to form an immunosuppressive microenvironment (Figure 1A). The latest published results reconfirmed these conclusions through the “Y chromosome transcription signature” (YchrS) in clinical samples (30). Beyond the LOY observed in neoplastic cells, significant prevalence was detected in tumor-infiltrating immune cells, particularly within T-cell populations. Further analysis revealed a direct correlation between immune cell LOY and T-cell-mediated immunosuppression (30). Notably, Y-deficient tumor models displayed enhanced responsiveness to PD-1 inhibitors, with post-treatment reactivation of CD8⁺ T cells from an exhausted to an activated state. Clinical data corroborate that LOY-positive patients achieve significantly improved survival following anti-PD-1 therapy (28, 29).



KDM5D, encoded by the Y chromosome, is a histone-modifying enzyme (31). Obviously, LOY will inevitably lead to abnormal function of the *KDM5D* gene. Jiexi Li et al. recently reported that Y-chromosomal *KDM5D* drives male bias in *KRAS*-mutant colorectal cancer (31). *KRAS* mutation downregulates *KDM5D* expression through *STAT4* inactivation, which enhances tumor invasiveness and metastatic potential via dysfunctional CD8⁺ T cells. Murine models showed that *KDM5D* deletion reduced tumor aggressiveness and augmented CD8⁺ T cell-mediated cytotoxicity (31). Mechanistically, *KDM5D* epigenetically suppresses *AMOT*, a gene critical for maintaining intercellular tight junctions, thereby promoting metastasis. Notably, *KDM5D*

also diminishes MHC class I antigen presentation and CD8⁺ T cell-mediated cytotoxicity, enabling immune evasion (Figure 1A) (31).

Studies in hematologic malignancies further implicate LOY in male hematopoietic cells as a critical driver of leukemogenesis and disease progression (32, 33). Intriguingly, male patients receiving female-derived hematopoietic stem cell transplants exhibit elevated relapse risk, potentially attributable to weakened graft-versus-leukemia effects due to sex-mismatched H-Y antigen expression encoded by the Y chromosome (34).

Recent findings by Jonas Fischer et al. demonstrate that LOY in lung adenocarcinoma remodels tumor immunogenicity via dysregulation of CTAs, facilitating immune evasion and significantly

impacting survival outcomes in pembrolizumab-treated cohorts (35). Collectively, these discoveries suggest LOY quantification may serve as a biomarker for personalized immunotherapy selection, while CTA-targeted immunotherapies could synergize with existing regimens to enhance therapeutic efficacy.

2.2 X chromosome

XCI is an epigenetic mechanism that silences one X chromosome in female cells to balance X-linked gene expression between XX and XY individuals (36, 37). However, approximately 15–25% of X-chromosomal genes escape XCI (termed “escape genes”) (38), many of which exhibit higher expression levels in females than males, particularly those involved in immunity and tumor suppression, such as Toll-like receptor 7 (*TLR7*) and *KDM6A* (39, 40). Biallelic expression of these escape genes enhances antitumor functionality in female immune cells, contributing to lower incidence and mortality rates in females for cancers like bladder cancer.

For the past few years, researchers have increasingly focused on the *KDM6A* gene. *KDM6A*, an X-chromosome inactivation escape gene, exhibits significantly higher expression in female cells compared to males (41). Although both *KDM6A* and *KDM5D* belong to the lysine demethylase superfamily, they exhibit substrate specificity for distinct histone lysine residues—*KDM6A* catalyzing H3K27me3 demethylation and *KDM5D* targeting H3K4me3 (42). Previous studies have shown that *KDM6A* contributes to sex disparities in bladder cancer (BCa) with 3–5 times more protective effects in females (39). This study demonstrates that female bladder epithelial cells with elevated *KDM6A* expression activate p53 downstream targets (e.g., *Cdkn1a*, *Perp*) through removal of the transcriptional repressive H3K27me3, thereby inducing cell cycle arrest and apoptosis (Figure 1B) (39). Notably, even catalytically inactive *KDM6A* mutants partially suppress tumor cell proliferation. *KDM6A* knockout in mice significantly increased bladder cancer risk in females, while males remained unaffected due to compensation by the Y-chromosomal homolog *Uty* (homologous gene of *KDM6A* with redundant function) (39). The study demonstrates that *KDM6A* mutations or low expression correlate significantly with poor prognosis in female patients but not males. Similar sex-biased expression patterns are observed in other malignancies like clear cell renal carcinoma, suggesting tissue-specific tumor suppressor functions of *KDM6A* (39).

Similarly, a study investigating sex disparities in glioblastoma (GBM) reveals higher incidence and mortality rates in male dependent on *KDM6A* expression in CD8⁺ T cells (43). In detail, immunocompetent male mice exhibited reduced CD8⁺ T cell infiltration and enhanced exhaustion in tumor microenvironments. Male CD8⁺ T cells displayed elevated inhibitory receptor expression (*PD-1*, *CTLA4*, *LAG3*) and reduced cytokine production, whereas female tumors showed greater infiltration of effector T cells (Tef) (43). These findings suggest male T cells are more prone to exhaustion, while female T cells maintain effector functionality

(Figure 1B). Anti-PD-1 therapy significantly prolonged survival in male mice but showed weaker efficacy in females. Meanwhile, under anti-PD-1 therapy, male tumors exhibited enhanced CD8⁺ T cell proliferative capacity and reduced exhausted T cell subsets, with minimal changes observed in females. This implies anti-PD-1 therapy primarily activates male T cell effector functions. Further analysis revealed that lower *KDM6A* levels in male T cells promote exhaustion, whereas elevated *KDM6A* expression in female T cells helps sustain effector functionality. Low expression of *KDM6A* leads to accelerated tumor growth in males with potentially greater therapeutic benefit from anti-PD-1 treatment, while maintaining superior functional capacity to constrain tumor progression in females (43).

3 Sex hormone-mediated immunomodulation

Sex steroids (androgens/estrogens) coordinate with sex chromosomes to establish a sex-dimorphic immune microenvironment and evoke sex-based disparities in cancer incidence and therapeutic outcomes (44).

3.1 Androgens

Androgen (e.g., testosterone, dihydrotestosterone) primarily exerts its effects via the androgen receptor (AR), a ligand-dependent transcription factor regulating target gene expression (45). AR is functionally active across immune cell populations, including T cells, B cells, macrophages, and neutrophils (46–48). Androgens promote T cell exhaustion in tumor-infiltrating CD8⁺ T cells by suppressing effector molecule production via AR signaling, exacerbating male-biased progression in malignancies such as bladder, colorectal, hepatocellular, and cutaneous cancers (45, 48–52).

Multiple studies highlight synergistic antitumor effects when combining androgen deprivation therapy (ADT) with PD-1/PD-L1 blockade (45, 48). The androgen receptor (AR) signaling pathway can directly promote the differentiation of CD8⁺ T cells into a terminally exhausted state. Mechanistically, as a transcription factor, AR specifically binds to androgen response elements (AREs) within the promoter region of the *Tcf7* gene, thereby activating its transcription (49). This leads to the upregulation of TCF-1 (encoded by *Tcf7* gene) protein expression, which drives the progression of CD8⁺ T cell terminal exhaustion (Figure 2A) (49). Xiaomin Zhang et al. recently demonstrated less infiltration of CD8⁺ T cells and increased expression of exhaustion markers (such as *PD-1*/*CD39*/*TIM3*/*TIGIT*) under ADT treatment in male mouse tumors. And castration delays tumor growth and restores T cell activity (Figure 2A) (53). Their work revealed that androgen signaling suppressed antitumor T cell activity through upregulating USP18, which inhibits TAK1 phosphorylation and subsequent NF-κB activation in T cells (53). In addition, Liang Chi et al. identified elevated dendritic cell (DC) subsets (cDC1, LC,

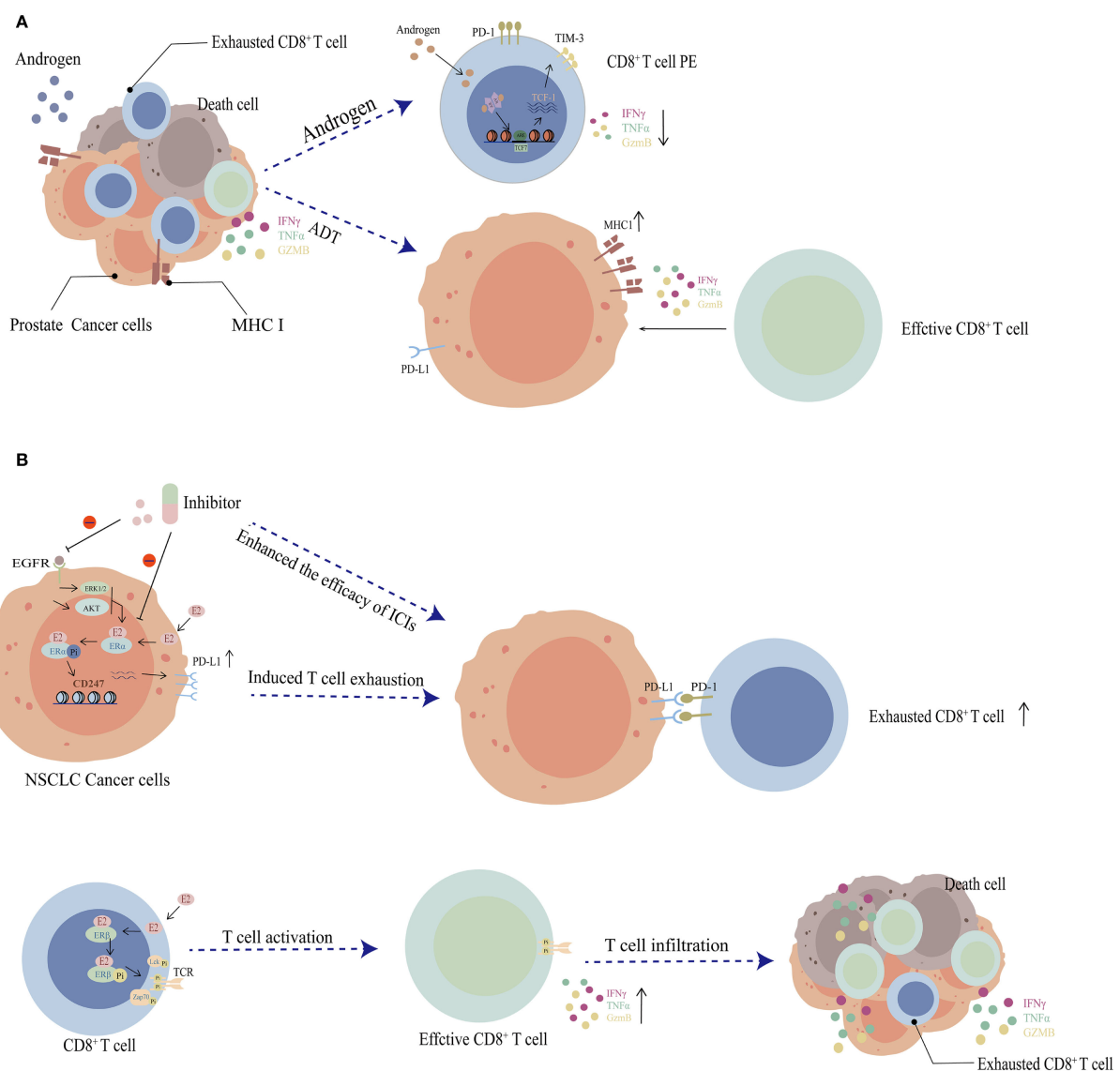


FIGURE 2

Roles of sex hormones in cancer sexual dimorphism. **(A)** Androgen receptor (AR) modulates the differentiation of tumor-infiltrating CD8⁺ T cells and impairs effector functions. AR binds to androgen response elements (AREs) located at the *TCF7* promoter, inducing its upregulation in CD8⁺ T cells. Elevated TCF1 expression promotes T cell exhaustion, thereby impairing antitumor immunity. Conversely, after androgen deprivation therapy (ADT), lower AR activity facilitates CD8⁺ T cell progression toward functional effector (Tef), enhancing antitumor immunity. These exhausted CD8⁺ T cells exhibit suppressed expression of IFN γ , TNF α and GZMB. **(B)** Estrogen activates anti-tumor immunity by binding to its receptors. E2 binding to ER α can upregulate the *CD274* expression (encoding PD-L1), driving CD8⁺ T cell exhaustion. Conversely, ER β enhances the TCR signaling pathway (such as Zap70/Lck phosphorylation) in CD8⁺ T cells, promoting T cell activation and anti-tumor cytokine secretion (e.g., IFN γ , granzyme B, and TNF α).

cDC2) in female skin compared to males. These DCs underpin antigen presentation and adaptive immune priming and are maintained by the skin group 2 innate lymphoid cells (ILC2s). Androgens negatively regulate skin ILC2s, creating DC disparities that result in weaker adaptive antitumor immunity in males (54). Consistently, AR knockout or surgical castration will enhance antitumor T cell activity and augment PD-1 blockade efficacy in males (54).

Emerging evidence reveals that androgen receptor (AR) facilitates immune evasion in prostate cancer through transcriptional suppression of MHC class I molecules (55). Mechanistically, AR

directly binds AREs within promoter regions of MHC I-associated genes (e.g., *HLA-A*, *B2M*, *TAP1/2*), inhibiting their transcriptional activity and consequently diminishing tumor antigen presentation capacity (55). AR-suppressed tumor cells exhibit heightened vulnerability to antigen-specific CD8⁺ T cell cytotoxicity in an MHC I-dependent manner (55). Notably, AR-knockdown TrampC1 tumors (TrampC1 AR-KD) demonstrated significantly restricted growth in murine models, accompanied by enhanced intra-tumoral CD8⁺ T cell infiltration and effector function (e.g., IFN γ production). However, this therapeutic effect attenuated over time, correlating with compensatory glucocorticoid receptor (GR) activation (55).

3.2 Estrogens

Estrogen, including estrone (E1), 17 β -estradiol (E2), and estriol (E3), mainly binds estrogen receptors (ER α , ER β , GPER1) to exert immunomodulation (56). Canonical estrogen receptor (ER α , ER β), functioning as the intracellular receptor for estrogen, undergoes nuclear translocation upon ligand binding and specifically binds to conserved estrogen response elements within target gene promoters, thereby regulating transcriptional activation through recruitment of coactivators and chromatin remodeling complexes. Beyond canonical ER signaling, estrogens also engage membrane-bound G protein-coupled estrogen receptor 1 (GPER1) to trigger rapid response via cyclic AMP pathways (57, 58).

ER α and ER β are expressed in various immune cells (59), but studies have shown that ER α and ER β play distinct roles in immune cells (60, 61). The estrogen-ER α signal activates the JAK2/STAT3 pathway, driving the differentiation of bone marrow myeloid precursors into Myeloid-Derived Suppressor Cells, enhancing their immunosuppressive function, thereby inhibiting the anti-tumor T-cell response and accelerating cancer progression (62). Similarly, in female melanoma, the estrogen-ER α signal drives macrophages to polarize towards the M2 phenotype, inhibits the function of CD8⁺ T cells, promotes melanoma progression and induces immune checkpoint blockade (ICB) resistance, while the antagonist of ER (Fulvestrant) can reverse the immunosuppressive microenvironment and restore T cell function (63).

Coincidentally, in non-small cell lung cancer (NSCLC) the downstream kinases of EGFR, such as Akt, ERK1/2, will phosphorylate ER α that binds to estradiol at the Ser118 site, thereby enhancing its transcriptional activity and upregulating the expression of PD-L1 (Figure 2B) (64). Pharmacological intervention with the estrogen synthesis inhibitor letrozole effectively suppresses PD-L1 expression and activates CD8⁺ T/NK cells, mimicking the therapeutic effects of PD-1/PD-L1 blockade agents. Furthermore, combinatorial administration of letrozole with PD-1/PD-L1 blockade agents demonstrates synergistic efficacy, offering a promising strategy for optimizing immunotherapy outcomes, especially in 17- β -estradiol/ER α high female NSCLC patients (64).

The estrogen-ER β pathway plays an opposite role in anti-tumor immunity. For instance, in triple-negative breast cancer and melanoma models, ER β enhances the TCR signaling pathway (such as *Zap70/Lck* phosphorylation) in CD8⁺ T cells through the tyrosine phosphorylation switch, promoting T cell activation and anti-tumor cytokine secretion in a non-genomic manner (65). The ER β selective agonist (S-equol) can activate this phosphorylation switch and significantly enhance the efficacy of anti-PD-1 immunotherapy, providing a new strategy to overcome ICB resistance (65). In a recently published study on gender differences in colon cancer, researchers knocked out ER β in the intestines of female mice, which led to decreased T cell activation and infiltration in the tumor model, increased pro-inflammatory signals (*IL-6*, *CCL2/4*), and increased infiltration of M2-type macrophages. Additionally, TCGA cohort analysis indicated that

patients with high ER β expression had a higher survival rate (66). These results suggest that targeting and activating the estrogen-ER β pathway can enhance the anti-tumor immune response in females.

Unlike androgens broadly suppress antitumor immunity, estrogens exhibit bidirectional (pro-/anti-inflammatory) effects depending on receptor subtypes and cellular contexts. Targeting the sex hormone-immune axis may yield sex-specific therapeutic strategies, necessitating a deeper exploration of hormone signaling dynamics and tumor microenvironment interactions (44).

4 Conclusions and perspectives

Males demonstrate significantly higher incidence rates and poorer prognosis across non-reproductive malignancies, with multifactorial determinants spanning sex chromosomes, sex hormones and sex-specific immune modulation (Table 1). Males experience LOY (27–34) and high androgen expression (53, 55), leading to an immunosuppressive microenvironment (characterized by T cell exhaustion, M2 macrophage infiltration, and downregulation of MHC-I expression) that promotes progression of multiple non-reproductive cancers. Conversely, females benefit from the biallelic expression of X-chromosome escape genes (e.g., *KDM6A*) and the bidirectional immunomodulatory effects of estrogen (39, 43). ER α signaling promotes immunosuppression (e.g., M2 macrophage polarization) (62–64), while ER β enhances CD8⁺ T cell function (65, 66). These collectively enhance female immune surveillance capabilities, foster an anti-tumor microenvironment, and reduce cancer incidence and mortality. In general, sex chromosomes and sex hormones coordinately reshape sex-specific tumor microenvironment, and further foster sexual dimorphism in incidence and therapeutic outcomes in non-reproductive cancers.

Studies have shown that sex differences not only lead to differences in cancer incidence rates but are also a key factor in the response to immune checkpoint inhibition therapy (13, 55). Despite the higher incidence and mortality of solid tumors in males, clinical trial data indicate superior responses to ICB in males. Meta-analyses of randomized controlled trials (RCTs) consistently report lower mortality risk in males post-ICB, though statistical significance varies across studies (76–78). In melanoma and NSCLC cohorts, males demonstrate improved overall survival (OS) and progression-free survival (PFS) following anti-PD1, anti-PDL1, or anti-CTLA4 therapy (67, 69). The observed discrepancy primarily stems from the elevated prevalence of terminally exhausted CD8⁺ T cell subsets in male patients compared to females, with these exhausted T cell populations demonstrating heightened responsiveness to ICB treatment (49, 52).

Notably, paradoxical epidemiological patterns reveal female predominance in specific non-reproductive cancer types, particularly in thyroid cancer (2, 71), Xp11 translocation renal cell carcinoma (tRCC) (75) and melanoma in pre-menopausal

TABLE 1 Summary of Major Risk Factors for Non-Reproductive Cancers*.

Non-reproductive cancer types	Sex bias		Related risk factors	Refs.
	Incidence rate escalation	Mortality rate escalation		
Lung cancer	Males	Males	Sex chromosomes: LOY	(27, 35, 67)
			Sex hormones: Estrogen	(63)
Colorectal Cancer	Males	Males	Sex chromosomes: <i>KDM5D</i> expression within Y	(31)
			Sex hormones: Androgen and Estrogen	(50, 51, 65)
Bladder Cancer	Males	Males	Sex chromosomes: LOY and XCI	(29, 39, 42)
			Sex hormones: Androgen	(49, 68)
Glioblastoma	Males	Males	Sex chromosomes: XCI	(39)
Skin cancer	Males	—	Sex hormones: Androgen	(47, 53)
Leukemia	Males	—	Sex chromosomes: LOY	(31)
Thyroid Cancer	Females	—	Overdiagnosis	(69, 70)
Xp11 translocation renal cell carcinoma	Females	—	Sex chromosomes: X chromosome translocation	(71)
Alveolar soft part sarcoma/FOXR2-activated central nervous system neuroblastoma	Females	—	Sex chromosomes: X chromosome translocation	(72–74)
Melanoma	Females (Premenopausal)	—	Sex hormones: Estrogen	(62, 64, 75)

*The risk factors were strictly confined to the predefined biological variables: sex chromosomes and hormones.

women (70). A high prevalence of thyroid cancer in females has been reported mainly attributed to healthcare utilization and overdiagnosis (72). The tRCC exhibits a higher incidence in females attributed to the vulnerability of the TFE3 gene translocation in the X chromosome (75). Similar mechanisms involving X-chromosome alterations are implicated in the female predominance of alveolar soft part sarcoma (ASPS) and FOXR2-activated central nervous system neuroblastoma (68, 73, 74). Melanoma predominance in pre-menopausal women is often attributed to the high estrogen levels upregulating $Er\alpha$ and gastrin-releasing peptide receptor (GRPR) signal (70). Therefore, female-predominant malignancies (e.g., thyroid carcinoma, tRCC) represent distinct epidemiological exceptions, while male-biased cancer incidence remains the predominant global pattern in non-reproductive cancer types.

Sex-based disparities in oncogenesis extend beyond the sex chromosome and sex hormone-mediated microenvironmental remodeling discussed herein. Many other factors could influence Sex-based disparities. Emerging data suggest that sex-associated variations in the gut microbiome directly influence innate immune responses between the sexes (79). This result demonstrates that predominant male bladder cancer patients exhibit senescence-associated neutrophil (RLSN) through defective gut microbiota-derived *Alistipes shahii* compared to females (79). In addition, pharmacokinetic sex differences, attributed to lower body weight, higher adiposity, and differential tissue perfusion in females, result in elevated drug exposure and prolonged elimination in females. For instance, ICB agents exhibit sex-dimorphic clearance. Males show faster clearance of anti-CTLA4 (tremelimumab) and anti-PD1

(nivolumab), while females metabolize anti-PDL1 (durvalumab) more rapidly (8, 80). Moreover, Age represents a significant factor in the study of sex differences in cancer. Evidence indicates that childhood tumor incidence also exhibits similar sex disparities (81). Beyond sex biology, social gender also has a multi-dimensional and throughout impact on cancer (82). A recent review article reveals that gender-sex interactions (GSI) could affect cancer biology and clinical treatment such as the timing of diagnoses, clinical trial enrolment, and the completeness of efficacy and toxicity data (82). In summary, understanding the diverse factors and mechanisms underlying sex disparities in cancer will enable optimal treatment in future clinical trials. This knowledge is crucial for developing sex-specific biomarkers (e.g., LOY, *KDM6A* and estrogen) and combination strategies targeting immune pathways.

Ultimately, research into the role of sex differences in cancer immunology holds direct translational significance. Future clinical trials should therefore be designed to maximize therapeutic efficacy and develop targeted strategies. Additionally, further investigation into whether sex-related factors can serve as biomarkers for cancer diagnosis and risk stratification will significantly enhance precision diagnostics, patient stratification, and treatment optimization.

Author contributions

C-MZ: Validation, Writing – review & editing, Writing – original draft, Visualization. Z-BG: Writing – review & editing, Visualization, Validation. H-HZ: Writing – review & editing, Validation, Funding acquisition, Visualization. M-XW:

Visualization, Validation, Writing – review & editing. X-YD: Validation, Visualization, Writing – review & editing. Z-ZL: Visualization, Validation, Writing – review & editing. M-YW: Writing – review & editing, Supervision, Validation, Funding acquisition, Visualization, Conceptualization. C-JB: Validation, Visualization, Writing – review & editing, Supervision, Conceptualization, Funding acquisition.

Funding

The author(s) declare financial support was received for the research and/or publication of this article. This work was supported by grants from the National Natural Science Foundation of China (grant number: 32170558), the Natural Science Foundation of Gansu Province (grant numbers: 25JRRA828 and 23JRRA1168), Health and Wellness Industry Research Program of Gansu Province Hospital (grant number: GSWSKY2024-13), The National Natural Science Foundation Cultivation Project of Gansu Provincial Hospital (grant number: 24GSSYA-4), The Gansu Provincial Major Scientific Research Program for Young Talents in Health Industry Innovation (grant number: GSWSQNPY2025-17), and the Gansu Youth S&T Tackling Key Problems Project (Project-Unveiling and Leader-Appointing Mechanism, grant number: GQK2024025).

References

1. Siegel RL, Kratzer TB, Giaquinto AN, Sung H, Jemal A. Cancer statistics, 2025. *CA Cancer J Clin.* (2025) 75:10–45. doi: 10.3322/caac.21871
2. Siegel RL, Giaquinto AN, Jemal A. Cancer statistics, 2024. *CA Cancer J Clin.* (2024) 74:12–49. doi: 10.3322/caac.21820
3. Qi J, Li M, Wang L, Hu Y, Liu W, Long Z. National and subnational trends in cancer burden in China, 2005–20: an analysis of national mortality surveillance data. *Lancet Public Health.* (2023) 8:e943–55. doi: 10.1016/S2468-2667(23)00211-6
4. Chen DS, Mellman I. Oncology meets immunology: the cancer-immunity cycle. *Immunity.* (2013) 39:1–10. doi: 10.1016/j.immuni.2013.07.012
5. Siegel RL, Wagle NS, Cercak A, Smith RA, Jemal A. Colorectal cancer statistics, 2023. *CA Cancer J Clin.* (2023) 73:233–54. doi: 10.3322/caac.21772
6. Nencioni A, Caffa I, Cortellino S, Longo VD. Fasting and cancer: molecular mechanisms and clinical application. *Nat Rev Cancer.* (2018) 18:707–19. doi: 10.1038/s41568-018-0061-0
7. Islami F, Goding Sauer A, Miller KD, Siegel RL, Fedewa SA, Jacobs EJ, et al. Proportion and number of cancer cases and deaths attributable to potentially modifiable risk factors in the United States. *CA Cancer J Clin.* (2018) 68:31–54. doi: 10.3322/caac.21440
8. Penley MJ, Byrd DA, Bostick RM. Associations of evolutionary-concordance diet and lifestyle pattern scores with incident, sporadic colorectal adenoma in a pooled case-control study. *Nutr Cancer.* (2022) 74:2075–87. doi: 10.1080/01635581.2021.2002919
9. Chen X, Zhu D, Li C, Lin Y, Lv L, Ai F, et al. Modifiable factors affects cancer-specific survival: findings from a large population-based prospective cohort study. *J Transl Med.* (2025) 23:486. doi: 10.1186/s12967-025-06372-y
10. Danpanichkul P, Pang Y, Mahendru T, Tothananrungraj P, Diaz LA, Arab JP, et al. Sex disparities in alcohol-associated liver disease and subtype differences in alcohol-attributable cancers in the United States. *Korean J Hepatol.* (2025) 0:1058–70. doi: 10.3350/cmh.2025.0594
11. Forsyth KS, Jiwrakja N, Lovell CD, Toothacre NE, Anguera MC. The connection between sex and immune responses. *Nat Rev Immunol.* (2024) 24:487–502. doi: 10.1038/s41577-024-00996-9
12. Dunn SE, Perry WA, Klein SL. Mechanisms and consequences of sex differences in immune responses. *Nat Rev Nephrol.* (2024) 20:37–55. doi: 10.1038/s41581-023-00787-w
13. Klein SL, Flanagan KL. Sex differences in immune responses. *Nat Rev Immunol.* (2016) 16:626–38. doi: 10.1038/nri.2016.90
14. Dunford A, Weinstock DM, Savova V, Schumacher SE, Cleary JP, Yoda A, et al. Tumor-suppressor genes that escape from X-inactivation contribute to cancer sex bias. *Nat Genet.* (2017) 49:10–6. doi: 10.1038/ng.3726
15. Chen ACY, Jaiswal S, Martinez D, Yerinde C, Ji K, Miranda V, et al. The aged tumor microenvironment limits T cell control of cancer. *Nat Immunol.* (2024) 25:1033–45. doi: 10.1038/s41590-024-01828-7
16. de Visser KE, Joyce JA. The evolving tumor microenvironment: From cancer initiation to metastatic outgrowth. *Cancer Cell.* (2023) 41:374–403. doi: 10.1016/j.ccell.2023.02.016
17. Carbone A, Gloghini A, Carlo Stella C. Tumor microenvironment contribution to checkpoint blockade therapy. Lessons learned from Hodgkin lymphoma. *Blood.* (2023) 141:2187–93. doi: 10.1182/blood.2022016590
18. Bianchi I, Lleo A, Gershwin ME, Invernizzi P. The X chromosome and immune associated genes. *J Autoimmun.* (2012) 38:187–92. doi: 10.1016/j.jaut.2011.11.012
19. Hoffmann JP, Liu JA, Seddu K, Klein SL. Sex hormone signaling and regulation of immune function. *Immunity.* (2023) 56:2472–91. doi: 10.1016/j.immuni.2023.10.008
20. Wiese CB, Soliman B, Reue K. The Four Core Genotypes mouse model: evaluating the impact of a recently discovered translocation. *Biol Sex Differ.* (2024) 15:90. doi: 10.1186/s13293-024-00665-5
21. Arnold AP. Four Core Genotypes and XY* mouse models: Update on impact on SABV research. *Neurosci Biobehav Rev.* (2020) 119:1–8. doi: 10.1016/j.neubiorev.2020.09.021
22. Grimm SL, Dong X, Zhang Y, Carisey AF, Arnold AP, Moorthy B, et al. Effect of sex chromosomes versus hormones in neonatal lung injury. *JCI Insight.* (2021) 6:e146863. doi: 10.1172/jci.insight.146863
23. Islami F, Marlow EC, Thomson B, McCullough ML, Rungay H, Gapstur SM, et al. Proportion and number of cancer cases and deaths attributable to potentially modifiable risk factors in the United States, 2019. *CA Cancer J Clin.* (2024) 74:405–32. doi: 10.3322/caac.21858
24. Büschek F, Fraune C, Garmestani S, Simon R, Kluth M, Hube-Magg C, et al. Y-chromosome loss is frequent in male renal tumors. *Ann Transl Med.* (2021) 9:209. doi: 10.21037/atm-20-3061
25. Kido T, Lau Y-FC. Roles of the Y chromosome genes in human cancers. *Asian J andrology.* (2015) 17:373–80. doi: 10.4103/1008-682X.150842

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Generative AI statement

The author(s) declare that no Generative AI was used in the creation of this manuscript.

Any alternative text (alt text) provided alongside figures in this article has been generated by Frontiers with the support of artificial intelligence and reasonable efforts have been made to ensure accuracy, including review by the authors wherever possible. If you identify any issues, please contact us.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

26. Brunelli M, Eble JN, Zhang S, Martignoni G, Cheng L. Gains of chromosomes 7, 17, 12, 16, and 20 and loss of Y occur early in the evolution of papillary renal cell neoplasia: a fluorescent *in situ* hybridization study. *Mod Pathol*. (2003) 16:1053–9. doi: 10.1097/01.MP.0000090924.90762.94
27. Thompson DJ, Genovese G, Halvardson J, Ulirsch JC, Wright DJ, Terao C, et al. Genetic predisposition to mosaic Y chromosome loss in blood. *Nature*. (2019) 575:652–7. doi: 10.1038/s41586-019-1765-3
28. Qi M, Pang J, Mitsiades I, Lane AA, Rheinbay E. Loss of chromosome Y in primary tumors. *Cell*. (2023) 186:3125–3136.e11. doi: 10.1016/j.cell.2023.06.006
29. Abdel-Hafiz HA, Schafer JM, Chen X, Xiao T, Gauntner TD, Li Z, et al. Y chromosome loss in cancer drives growth by evasion of adaptive immunity. *Nature*. (2023) 619:624–31. doi: 10.1038/s41586-023-06234-x
30. Chen X, Shen Y, Choi S, Abdel-Hafiz HA, Basu M, Hoelzen L, et al. Concurrent loss of the Y chromosome in cancer and T cells impacts outcome. *Nature*. (2025) 642:1041–50. doi: 10.1038/s41586-025-09071-2
31. Li J, Lan Z, Liao W, Horner JW, Xu X, Liu J, et al. Histone demethylase KDM5D upregulation drives sex differences in colon cancer. *Nature*. (2023) 619:632–9. doi: 10.1038/s41586-023-06254-7
32. Ouseph MM, Hasserjian RP, Dal Cin P, Lovitch SB, Steensma DP, Nardi V, et al. Genomic alterations in patients with somatic loss of the Y chromosome as the sole cytogenetic finding in bone marrow cells. *Haematologica*. (2021) 106:555–64. doi: 10.3324/haematol.2019.240689
33. Ljungström V, Mattsson J, Halvardson J, Pandzic T, Davies H, Rychlicka-Buniowska E, et al. Loss of Y and clonal hematopoiesis in blood-tissue sides of the same coin? *Leukemia*. (2022) 36:889–91. doi: 10.1038/s41375-021-01456-2
34. Tamaki M, Kameda K, Kimura S-I, Harada N, Uchida N, Doki N, et al. Deletion of Y chromosome before allogeneic hematopoietic stem cell transplantation in male recipients with female donors. *Blood Adv*. (2022) 6:1895–903. doi: 10.1182/bloodadvances.2021006456
35. Fischer J, Shutta KH, Chen C, Fanfani V, Saha E, Mandros P, et al. Selective loss of Y chromosomes in lung adenocarcinoma modulates the tumor immune environment through cancer/testis antigens. *bioRxiv*. (2024). doi: 10.1101/2024.09.19.613876
36. Boumil RM, Lee JT. Forty years of decoding the silence in X-chromosome inactivation. *Hum Mol Genet*. (2001) 10:2225–32. doi: 10.1093/hmg/10.20.2225
37. Heard E, Chaumeil J, Masui O, Okamoto I. Mammalian X-chromosome inactivation: an epigenetics paradigm. *Cold Spring Harb Symp Quant Biol*. (2004) 69:89–102. doi: 10.1101/sqb.2004.69.89
38. Tukiainen T, Villani AC, Yen A, Rivas MA, Marshall JL, Satija R, et al. Landscape of X chromosome inactivation across human tissues. *Nature*. (2017) 550:244–8. doi: 10.1038/nature24265
39. Kaneko S, Li X. X chromosome protects against bladder cancer in females via a KDM6A-dependent epigenetic mechanism. *Sci Adv*. (2018) 4:eaar5598. doi: 10.1126/sciadv.aar5598
40. Souyris M, Mejia JE, Chaumeil J, Guéry JC. Female predisposition to TLR7-driven autoimmunity: gene dosage and the escape from X chromosome inactivation. *Semin Immunopathol*. (2019) 41:153–64. doi: 10.1007/s00281-018-0712-y
41. Cabrera Zapata LE, Cisternas CD, Sosa C, Garcia-Segura LM, Arevalo MA, Cambiasso MJ. X-linked histone H3K27 demethylase Kdm6a regulates sexually dimorphic differentiation of hypothalamic neurons. *Cell Mol Life Sci*. (2021) 78:7043–60. doi: 10.1007/s00018-021-03945-0
42. Tricarico R, Nicolas E, Hall MJ, Golemis EA. X- and Y-linked chromatin-modifying genes as regulators of sex-specific cancer incidence and prognosis. *Clin Cancer Res*. (2020) 26:5567–78. doi: 10.1158/1078-0432.CCR-20-1741
43. Lee J, Nicosia M, Hong ES, Silver DJ, Li C, Bayik D, et al. Sex-biased T-cell exhaustion drives differential immune responses in glioblastoma. *Cancer Discov*. (2023) 13:2090–105. doi: 10.1158/2159-8290.CD-22-0869
44. Conforti F, Pala L, Di Mitri D, Catania C, Cocorocchio E, Laszlo D, et al. Sex hormones, the anticancer immune response, and therapeutic opportunities. *Cancer Cell*. (2025) 43:343–60. doi: 10.1016/j.ccell.2025.02.013
45. Tan MH, Li J, Xu HE, Melcher K, Yong EL. Androgen receptor: structure, role in prostate cancer and drug discovery. *Acta Pharmacol Sin*. (2015) 36:3–23. doi: 10.1038/aps.2014.18
46. Viselli SM, Reese KR, Fan J, Kovacs WJ, Olsen NJ. Androgens alter B cell development in normal male mice. *Cell Immunol*. (1997) 182:99–104. doi: 10.1006/cimm.1997.1227
47. Chuang KH, Altuwajri S, Li G, Lai JJ, Chu CY, Lai KP, et al. Neutropenia with impaired host defense against microbial infection in mice lacking androgen receptor. *J Exp Med*. (2009) 206:1181–99. doi: 10.1084/jem.20082521
48. Guan X, Polesso F, Wang C, Sehrawat A, Hawkins RM, Murray SE, et al. Androgen receptor activity in T cells limits checkpoint blockade efficacy. *Nature*. (2022) 606:791–6. doi: 10.1038/s41586-022-04522-6
49. Kwon H, Schafer JM, Song NJ, Kaneko S, Li A, Xiao T, et al. Androgen conspires with the CD8(+) T cell exhaustion program and contributes to sex bias in cancer. *Sci Immunol*. (2022) 7:eabq2630. doi: 10.1126/sciimmunol.abq2630
50. Bell HN, Huber AK, Singhal R, Korimerla N, Rebernick RJ, Kumar R, et al. Microenvironmental ammonia enhances T cell exhaustion in colorectal cancer. *Cell Metab*. (2023) 35:134–149 e6. doi: 10.1016/j.cmet.2022.11.013
51. Acha-Sagredo A, Andrei P, Clayton K, Taggart E, Antoniotti C, Woodman CA, et al. A constitutive interferon-high immunophenotype defines response to immunotherapy in colorectal cancer. *Cancer Cell*. (2025) 43:292–307. doi: 10.1016/j.ccell.2024.12.008
52. Yang C, Jin J, Yang Y, Sun H, Wu L, Shen M, et al. Androgen receptor-mediated CD8(+) T cell stemness programs drive sex differences in antitumor immunity. *Immunity*. (2022) 55:1268–1283.e9. doi: 10.1016/j.immuni.2022.05.012
53. Zhang X, Cheng L, Gao C, Chen J, Liao S, Zheng Y, et al. Androgen signaling contributes to sex differences in cancer by inhibiting NF- κ B activation in T cells and suppressing antitumor immunity. *Cancer Res*. (2023) 83:906–21. doi: 10.1158/0008-5472.CAN-22-2405
54. Chi L, Liu C, Gribonika I, Gschwend J, Corral D, Han SJ, et al. Sexual dimorphism in skin immunity is mediated by an androgen-ILC2-dendritic cell axis. *Science*. (2024) 384:eadk6200. doi: 10.1126/science.adk6200
55. Chesner LN, Polesso F, Graff JN, Hawley JE, Smith AK, Lundberg A, et al. Androgen receptor inhibition increases MHC class I expression and improves immune response in prostate cancer. *Cancer Discov*. (2025) 15:481–94. doi: 10.1158/2159-8290.CD-24-0559
56. Chakraborty B, Byemerwa J, Krebs T, Lim F, Chang CY, McDonnell DP. Estrogen receptor signaling in the immune system. *Endocr Rev*. (2023) 44:117–41. doi: 10.1210/edrv/bnac017
57. Filardo EJ, Thomas P. Minireview: G protein-coupled estrogen receptor-1, GPER-1: its mechanism of action and role in female reproductive cancer, renal and vascular physiology. *Endocrinology*. (2012) 153:2953–62. doi: 10.1210/en.2012-1061
58. Molina L, Figueroa CD, Bhoola KD, Ehrenfeld P. GPER-1/GPR30 a novel estrogen receptor sited in the cell membrane: therapeutic coupling to breast cancer. *Expert Opin Ther Targets*. (2017) 21:755–66. doi: 10.1080/14728222.2017.1350264
59. Pierdominici M, Maselli A, Colasanti T, Giammarioli AM, Delunardo F, Vacirca D, et al. Estrogen receptor profiles in human peripheral blood lymphocytes. *Immunol Lett*. (2010) 132:79–85. doi: 10.1016/j.imlet.2010.06.003
60. Gustafsson JA. What pharmacologists can learn from recent advances in estrogen signalling. *Trends Pharmacol Sci*. (2003) 24:479–85. doi: 10.1016/S0165-6147(03)00229-3
61. Heldring N, Pike A, Andersson S, Matthews J, Cheng G, Hartman J, et al. Estrogen receptors: how do they signal and what are their targets. *Physiol Rev*. (2007) 87:905–31. doi: 10.1152/physrev.00026.2006
62. Svoronos N, Perales-Puchalt A, Allegrezza MJ, Rutkowski MR, Payne KK, Tesone AJ, et al. Tumor cell-independent estrogen signaling drives disease progression through mobilization of myeloid-derived suppressor cells. *Cancer Discov*. (2017) 7:72–85. doi: 10.1158/2159-8290.CD-16-0502
63. Chakraborty B, Byemerwa J, Shepherd J, Haines CN, Baldi R, Gong W, et al. Inhibition of estrogen signaling in myeloid cells increases tumor immunity in melanoma. *J Clin Invest*. (2021) 131:e151347. doi: 10.1172/JCI151347
64. Anobile DP, Salaroglio IC, Tabbò F, La Vecchia S, Akman M, Napoli F, et al. Autocrine 17- β -estradiol/estrogen receptor- α Loop determines the response to immune checkpoint inhibitors in non-small cell lung cancer. *Clin Cancer Res*. (2023) 29:3958–73. doi: 10.1158/1078-0432.CCR-22-3949
65. Yuan B, Clark CA, Wu B, Yang J, Drerup JM, Li T, et al. Estrogen receptor beta signaling in CD8(+) T cells boosts T cell receptor activation and antitumor immunity through a phosphotyrosine switch. *J Immunother Cancer*. (2021) 9:e001932. doi: 10.1136/jitc-2020-001932
66. Birgersson M, Holm M, Gallardo-Dodd CJ, Chen B, Stepanauskaitė L, Hases L, et al. Intestinal estrogen receptor beta modulates the murine colon tumor immune microenvironment. *Cancer Lett*. (2025) 622:217661. doi: 10.1016/j.canlet.2025.217661
67. Kartolo A, Sattar J, Sahai V, Baetz T, Lakoff JM. Predictors of immunotherapy-induced immune-related adverse events. *Curr Oncol*. (2018) 25:e403–10. doi: 10.3747/co.25.4047
68. Sturm D, Orr Brent A, Toprak Umur H, Hovestadt V, Jones David TW, Capper D, et al. New brain tumor entities emerge from molecular classification of CNS-PNETs. *Cell*. (2016) 164:1060–72. doi: 10.1016/j.cell.2016.01.015
69. Cortellini A, Friedlaender A, Banna GL, Porzio G, Bersanelli M, Cappuzzo F, et al. Immune-related adverse events of pembrolizumab in a large real-world cohort of patients with NSCLC with a PD-L1 expression \geq 50% and their relationship with clinical outcomes. *Clin Lung Cancer*. (2020) 21:498–508.e2. doi: 10.1016/j.clcc.2020.06.010
70. Raymond JH, Aktary Z, Pouteaux M, Petit V, Luciani F, Wehbe M, et al. Targeting GRPR for sex hormone-dependent cancer after loss of E-cadherin. *Nature*. (2025) 643:801–9. doi: 10.1038/s41586-025-09111-x
71. Huang K, Wang N, Huang X, Qian S, Cai Y, Wu F, et al. Global, regional, and national burden of thyroid cancer in young people aged 10–24 years from 1990 to 2021: an analysis based on the Global Burden of Disease Study 2021. *BMC Public Health*. (2025) 25:1221. doi: 10.1186/s12889-025-22322-1
72. Li M, Dal Maso L, Pizzato M, Vaccarella S. Evolving epidemiological patterns of thyroid cancer and estimates of overdiagnosis in 2013–17 in 63 countries worldwide: a population-based study. *Lancet Diabetes Endocrinol*. (2024) 12:824–36. doi: 10.1016/S2213-8587(24)00223-7
73. Korshunov A, Okonechnikov K, Schmitt-Hoffner F, Ryzhova M, Sahm F, Stichel D, et al. Molecular analysis of pediatric CNS-PNET revealed nosologic heterogeneity

and potent diagnostic markers for CNS neuroblastoma with FOXR2-activation. *Acta Neuropathologica Commun.* (2021) 9:20. doi: 10.1186/s40478-021-01118-5

74. Tauziède-Espariat A, Figarella-Branger D, Métails A, Uro-Coste E, Maurage C-A, Lhermitte B, et al. CNS neuroblastoma, FOXR2-activated and its mimics: a relevant panel approach for work-up and accurate diagnosis of this rare neoplasm. *Acta Neuropathologica Commun.* (2023) 11:43. doi: 10.1186/s40478-023-01536-7

75. Achom M, Sadagopan A, Bao C, McBride F, Li J, Konda P, et al. A genetic basis for sex differences in Xp11 translocation renal cell carcinoma. *Cell.* (2024) 187:5735–5752.e25. doi: 10.1016/j.cell.2024.07.038

76. Botticelli A, Onesti CE, Zizzari I, Cerbelli B, Sciatella P, Occhipinti M, et al. The sexist behaviour of immune checkpoint inhibitors in cancer therapy? *Oncotarget.* (2017) 8:99336–46. doi: 10.18632/oncotarget.22242

77. Conforti F, Pala L, Bagnardi V, De Pas T, Martinetti M, Viale G, et al. Cancer immunotherapy efficacy and patients' sex: a systematic review and meta-analysis. *Lancet Oncol.* (2018) 19:737–46. doi: 10.1016/S1470-2045(18)30261-4

78. Grassadonia A, Sperduti I, Vici P, Iezzi L, Brocco D, Gamucci T, et al. Effect of gender on the outcome of patients receiving immune checkpoint inhibitors for advanced cancer: A systematic review and meta-analysis of phase III randomized clinical trials. *J Clin Med.* (2018) 7:e542. doi: 10.20944/preprints201808.0307.v2

79. Zhu Q, Zhang G, Cao M, Huang H, He D, Zang Z, et al. Microbiota-shaped neutrophil senescence regulates sexual dimorphism in bladder cancer. *Nat Immunol.* (2025) 26:722–36. doi: 10.1038/s41590-025-02126-6

80. Desnoyer A, Broutin S, Delahousse J, Maritaz C, Blondel L, Mir O, et al. Pharmacokinetic/pharmacodynamic relationship of therapeutic monoclonal antibodies used in oncology: Part 2, immune checkpoint inhibitor antibodies. *Eur J Cancer.* (2020) 128:119–28. doi: 10.1016/j.ejca.2020.01.003

81. Rubin JB. The spectrum of sex differences in cancer. *Trends Cancer.* (2022) 8:303–15. doi: 10.1016/j.trecan.2022.01.013

82. Rubin JB. Gender and sex interactions are intrinsic components of cancer phenotypes. *Nat Rev Cancer.* (2025) 25:634–48. doi: 10.1038/s41568-025-00829-4