

OPEN ACCESS

EDITED BY Chunmei Ma, Nanjing Medical University, China

REVIEWED BY Chakrapani Vemulawada, The University of Iowa, United States

[†]These authors have contributed equally to this work

RECEIVED 08 July 2025 ACCEPTED 04 August 2025 PUBLISHED 21 August 2025

CITATION

Xin C, Liu P, Zhan Q and Cao W (2025) Gzmk⁺ CD8 T cells in inflammatory diseases. Front. Immunol. 16:1661755. doi: 10.3389/fimmu.2025.1661755

COPYRIGHT

© 2025 Xin, Liu, Zhan and Cao. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Gzmk⁺ CD8 T cells in inflammatory diseases

Cui Xin^{1†}, Peiyun Liu^{2†}, Qiangian Zhan^{2†} and Wengiang Cao^{2*}

¹Department of Criminal Science and Technology, Hunan Police College, Changsha, China, ²Key Laboratory of Major Chronic Diseases of Nervous System of Liaoning Province, Health Sciences Institute of China Medical University, Shenyang, China

T cells are integral to the immune response, with distinct subsets exhibiting specialized functions, a phenomenon well-characterized in helper CD4⁺ T cells. Recent advancements in single-cell RNA sequencing (scRNA-seq) have facilitated the identification of numerous novel CD8 T cell subsets, each characterized by unique functional properties. As cytotoxic T lymphocytes, the primary focus has been on the cytotoxic capabilities and antigen specificity of these subsets. A recently identified subset, Granzyme k (Gzmk)⁺ CD8 T cells, has been closely associated with inflammatory diseases, independent of their cytotoxic function. Unlike other granzymes, granzyme K predominantly induces proinflammatory responses in tissues or cells rather than mediating cytotoxicity. This review synthesizes current evidence regarding the regulation, functional roles, and underlying mechanisms of Gzmk⁺ CD8 T cells in inflammatory conditions. Elucidating these processes may reveal potential therapeutic targets for treating inflammatory diseases.

KEYWORDS

GZMK, CD8, inflammatory disease, inflammaging, aging

Introduction

Inflammatory diseases are mediated by immune cells, which can act in either an autoimmune or non-autoimmune manner. In autoimmune disorders such as rheumatoid arthritis (RA) and multiple sclerosis (MS), CD4 T cells were traditionally considered the primary effectors. However, autoreactive CD8 T cells also play a pathogenic role in several autoimmune diseases by directly damaging self-cells (1). Additionally, CD8 regulatory T cells can negatively impact autoimmune responses by killing autoreactive CD4 T cells (2–4). These functions related to autoimmune activity are associated with cytotoxicity. In non-autoimmune diseases, CD8 T cells are often found in higher concentrations in inflamed tissues, suggesting they likely play a critical role in these conditions as well (5).

Response to different stimulatory milieus, CD4 T cells differentiate into distinct subsets with specialized phenotypes and functions (6). Similarly, CD8 T cells are also categorized into subsets such as Tc1, Tc2 and Tc17, but this classification is controversial (7). Upon stimulation, naïve CD8 T cells differentiate into effector/memory T cells. Effector CD8 T cells can directly kill target cells through interactions involving Fas/Fas ligand, or by secreting perforin and granzymes. Within effector or memory CD8 T cells, several different

populations have been identified, including short-lived effector CD8 T cells, long-live memory CD8 T cells (Tm), exhausted CD8 T cells (Tex), memory precursor CD8 T cells (Tmp), central and effector memory CD8 T cells (Tcm and Tem), tissue-resident memory cells (Trm) (8) and terminally differentiated T effector memory CD45RA+ (Temra) in human (9). However, these classifications do not entirely capture the specific functions of CD8 T cells in particular diseases. Recent scRNA-seq studies have revealed that Gzmk+ CD8 T cells are enriched in the inflamed tissues of both autoimmune and non-autoimmune diseases (10–16). In this review, we have included all diseases in which the presence of Gzmk+ CD8 T cells has been identified using single-cell RNA sequencing (scRNA-seq) and flow cytometry. We will discuss the differentiation, function, and underlying mechanisms of Gzmk+ CD8 T cells in the context of inflammatory diseases.

Identification and generation of Gzmk⁺ CD8 T cells

Conventional subsets such as Tem, Tcm, and Temra do not fully reflect the heterogeneity of CD8 T cells. Advances in single-cell RNA sequencing (scRNA-seq) have enhanced our understanding of CD8 T cell diversity, leading to the identification of several new CD8 T cell subsets in specific diseases or conditions. For instance, Gzmk+ CD8 T cells have been identified in aging tissues in mice, exhibiting exhaustion-like phenotypes characterized by the coexpression of TOX, PD1, and other co-inhibitory receptors (17). Further studies have revealed that Gzmk+ CD8 T cells also increase in peripheral blood mononuclear cells (PBMCs) from older individuals (17). Aging brings significant changes to CD8 T cells, including an increase in Temra and virtual memory cells (18-20). A key question is whether Gzmk+ CD8 T cells represent a specific agerelated subset of CD8 T cells. Gzmk+ CD8 T cells are primarily found in the Tem and Tcm subsets and are less prevalent in the Temra. Notably, Gzmk+ Tem cells are characterized as CD27+ CD28⁺ CD57⁻. Transcriptional analysis reveals that Gzmk⁺ CD8 T cells differ from virtual memory cells, which are driven by IL-15 stimulation and do not express CD49d (18). In contrast, Gzmk+ CD8 T cells exhibit high CD49d expression (17). Therefore, Gzmk⁺ CD8 T cells should not be classified as typical age-related CD8 T cells. Moreover, it has been observed that CD39⁺ memory CD8 T cells are elevated in older individuals (21), and CD39 expression is also a hallmark of cell exhaustion (22-24), raising the question of whether Gzmk and CD39 co-express in the same CD8 T cell population. Although Gzmk and granzyme b (Gzmb) are both granzymes, their expression profiles differ significantly. Cytometry and scRNA-seq analyses have demonstrated that Gzmk and Gzmb express mutually within the Tem subset (17). In summary, Gzmk⁺ CD8 T cells are a distinct subset associated with aging, highlighting the complexity of CD8 T cell heterogeneity.

Gzmk⁺ CD8 T cells were initially identified as conserved ageassociated T cells (Taa). Still, they have also been found to be enriched in inflamed tissues and circulation during many inflammation-related diseases (10, 12–16, 25–28). This suggests that Gzmk+ CD8 T cells play a crucial role in these diseases, in addition to their involvement in age-related immune responses. However, the reasons for Gzmk⁺ CD8 T cells accumulating in aged or inflamed tissues remain unclear. Mogilenko et al. revealed that the old environment drives the generation of Gzmk⁺ CD8 T cells (17), indicating that shared factors between aged and inflamed tissue are responsible for their differentiation. Single-cell ATAC-seq analysis suggests that the transcriptional activity of Eomes is elevated, which may be involved in the differentiation of Gzmk⁺ CD8 T cells (17). Eomes has been shown to promote CD8 T cells exhaustion and directly regulate exhaustion-associated genes (29, 30), raising a question of whether Eomes is involved in the regulation of Gzmk. Indeed, Eomes was reported to bind to the promoter of *Gzmk* and induce its expression in CD4 T cells (31, 32). This leads to the speculation that factors within inflamed and aged tissues trigger Eomes expression, which in turn causes the generation of Gzmk+ CD8 T cells, warranting future investigation.

The function and regulation of Gzmk in CD8 T cells

Granzymes are primarily recognized as factors that induce cell death in target cells. For instance, Gzmb enters the target cell and cleaves caspases 3 and 7 to trigger apoptosis (33). The delivery of Gzmb relies mainly on the formation of pores in the cell membrane mediated by perforin (34). Early studies on Gzmk also focused on its cytotoxicity, demonstrating that Gzmk can induce caspase-independent apoptosis by cleaving various substrates, including SET, Ape1, HMG2, Bid, VCP and P53 (35–39). However, Gzmk⁺ CD8 T cells lack perforin (10, 14), and extracellular Gzmk is present in specific conditions (40), suggesting Gzmk has additional functions beyond its intracellular cytotoxicity. Increasing evidence indicates that Gzmk plays a proinflammatory role by cleaving extracellular substrates such as PAR1, lipopolysaccharides (LPS) and complement proteins (12, 13, 40–44).

Gzmb and perforin in CD8 T cells are synthesized and released upon T-cell receptor (TCR) activation and the formation of the immunological synapse (45, 46). Interestingly, Gzmk is constitutively synthesized and released by CD8 T cells, even in the absence of TCR stimulation. In fact, TCR activation inhibits Gzmk expression in human CD8 T cells (14, 41). Notably, coinhibitory receptors, such as PD-1, LAG3 and CTLA4, are elevated at both human and murine Gzmk+ CD8 T cells (17), which likely play a role in regulating Gzmk expression by counteracting the inhibition mediated by TCR signaling. Furthermore, aged CD8 T cells tend to acquire natural killer (NK)-related phenotypes and downregulate TCR-related molecules (47). This results in reduced TCR signaling transduction and makes aged CD8 T cells more sensitive to cytokine stimulation. IL-2/IL-12 strongly induces Gzmk expression and proliferation of Gzmk⁺ CD8 T cells (14), while IL-15 is also capable of triggering Gzmk expression in CD8 T cells (28).

There is a complex interaction between cytokines and TCR signals that determines the differentiation fate of CD8 T cells. A recent study has shown that TCR signaling inhibits the IL-15-

induced the upregulation of NK-related genes through NFATc1 (48). While IL-15 activates AP-1 to induce NKG2D expression, the binding of NFATc1 to AP-1 constrains this effect. In contrast to wild-type NFATc1, overexpression of a mutant form of NFATc1 that cannot bind to AP-1 does not abrogate IL-15-induced NKGD2 upregulation (48). Interestingly, expression of a version of NFAT1 that is incapable of interacting with AP-1 induced exhaustion of CD8 T cells characterized by upregulation of inhibitory receptors and the related transcription factors including BLIMP1, ZEB1 and TOX (49). TOX is highly expressed in Gzmk⁺ CD8 T cells, but it remains unclear whether TOX regulates the function and differentiation of Gzmk⁺ T cells through integrating cytokines and TCR signaling.

ATAC-seq and RNA-seq analysis indicated that Eomes is likely an essential transcription factor in human and murine Gzmk⁺ CD8 T cells. Tem-k (Gzmk+ Tem) and Tem-b (Gzmb+ Tem) cells were enriched for Eomes and T-bet motif, respectively (17). Moreover, Eomes plays a critical role in the effects of IL-15 and has been reported to regulate Gzmk expression directly in CD4 T cells (50). These suggest that the differentiation of Tem-k and Tem-b may rely on a balance between T-bet and Eomes-driven transcriptional regulation, similar to the differentiation of effector and exhausted CD8 T cells (51, 52). Notably, the ratio of T-bet/Eomes has been reported to be influenced by the strength of TCR signaling, with low-affinity peptides favoring a lower ratio of T-bet/Eomes (53). Further investigation is warranted to determine whether TCR or/ and cytokines signaling regulates the differentiation of Gzmk⁺ CD8 T cells through the transcriptional activity of T-bet and Eomes. Additionally, in the absence of TCR stimulation, IL-15 promotes the generation of virtual memory cells, which also dependents on Eomes (18, 54). Future research should explore how Eomes responds to environmental stimuli and controls the differentiation of virtual memory, exhausted and Gzmk+ CD8 T cells.

Gzmk⁺ CD8 T cells in autoimmune diseases

The role of CD8 T cells in autoimmune diseases has been extensively studied, which depends on their cytotoxicity towards selfcells or autoreactive CD4 cells (1-4). Recent researches have revealed that Gzmk⁺ CD8 T cells are enriched in tissues of rheumatoid arthritis (RA) and psoriasis, where they contribute to inflammation and disease progression by activating the complement system (10, 41). Notably, Gzmk has been identified in areas abundant with C3b and C5a. Experiments have shown that deficiency of Gzmk in mice leads to reduction in the severity of RA, imiguimod-induce dermatitis, and C3d deposition (41). Given that the crucial function of complements in autoimmune diseases (55), It is likely that Gzmk+ CD8 T cells are involved in autoimmune diseases via activating complement system. Beyond RA and psoriasis, Gzmk+ CD8 T cells have also been found to be enriched in at tissues of systemic lupus erythematosus (SLE) (56, 57), Sjögren's disease (28), uveitis (58) and IgG4-related disease (59). The function of complements in autoimmune diseases is complex, exhibiting pathogenic or protective roles (55). For instance, deficiencies of C1q and C4, can exacerbate the severity of SLE, whereas a lack of C3 may alleviate it (55). Gzmk is reported to cleave C2 and C4, resulting in the formation of C3 convertase, which generates C3a and triggers a pro-inflammatory reponse (12, 41). This mechanism likely contributes to the progression of RA, psoriasis, SLE and uveitis (60). In cases of Sjögren's disease and IgG4-related diseases, a higher frequency of hypocomplementemia is observed which is linked to the clinical phenotype (61, 62). Therefore, further investigation is needed to determine whether Gzmk⁺ CD8 T cells contribute to Sjögren's disease and IgG4-related diseases through the activation complements.

Apart from its roles in cleaving complements, Gzmk has also been reported to cleave protease-activated receptor 1 (PAR1), which significant in inflammation, including autoimmune dieases (63). For example, *PAR1*-/- mice exhibited less severity of arthritis induced by antigen (AIA) (64), which reflects the role of Gzmk in AIA (41). However, the deficiency of PAR1 exacerbated the collagen-induced arthritis (CIA) (65). This complexity is further highlighted by the fact that PAR1 serves different functions in various cells types involved in inflammatory bowel disease (IBD) (66), another autoimmune condition. Notably, Gzmk+ CD8 T cells have also been identified in the lamina propria and intraepithelial of patients with ileal Crohn's disease (67). Hence, the roles of Gzmk-mediated PAR1 cleavage and activation at RA and IBD warrant further investigation.

Lipopolysaccharide (LPS) is another extracellular substrate of Gzmk (42), which strongly stimulates the innate immune response. This stimulation can exacerbate autoimmune conditions, such as SLE (68), autoimmune uveitis (69), IBD (70) and RA (71). Gzmk binds to LPS and pulls it out of its micelle conformation, enhancing the stimulation of immune cells by LPS (42). Therefore, in the context of bacterial infection or LPS leakage from damaged intestinal epithelium, Gzmk may worsen autoimmune diseases by amplifying the LPS-induced pro-inflammatory response.

Unlike Gzmk⁺ CD8 T cells, Gzmb⁺ CD8 T cells play a significant role in autoimmune diseases through their cytotoxicity (72). On one hand, auto-reactive Gzmb⁺ CD8 T cells kill target cells and cleave proteins to release autoantigens, which in turn promote the generation of autoantibodies (73, 74). On the other hand, Ly49⁺/KIR+ Gzmb⁺ CD8 T cells help prevent an excessive autoimmune response by targeting and eliminating autoreactive CD4 T cells (2–4, 75–78). This adds an additional layer of regulation by CD8 T cells to the complexity of autoimmune disease.

Gzmk⁺ CD8 T cells in airway inflammatory diseases

Clonally expanded Gzmk⁺ CD8 T cells are enriched in nasal polyps from chronic rhinosinusitis patients (12, 13), which are associated with type 2 immune cells (79). Research by Guo et al. demonstrated that Gzmk⁺ CD8 T cells interact with fibroblasts via the CXCR4-CXCL12 axis, inducing fibroblasts to release neutrophil chemoattractants. This exacerbated the airway inflammation and resistance to medical intervention in chronic rhinosinusitis with

nasal polyps (CRSwNP) (13, 80, 81), where Gzmk+ CD8 T cells likely play a crucial role. TCR analysis revealed that Gzmk⁺ CD8 T cells show a selective expansion of clones that recognize Epstein-Barr virus (EBV) (13). This indicates that Gzmk⁺ CD8 T cells are involved in airway inflammation by a bystander way, which aligns with the observation that Gzmk can be released by CD8 T cells independently of TCR stimulation (14, 41). Deletion of Gzmk in T cells resulted in reduced numbers of neutrophils, eosinophils, CD4 T cells, dendritic cells and macrophages in bronchoalveolar lavage fluid (BALF) in mice with asthma (12), This suggests that Gzmk is essential not only for the recruitment of neutrophil, but also for the recruitment of type 2 immune cells, likely due to chemokines and C3a produced locally (12). Gzmk+ CD8 T cells cannot promote eosinophil infiltration into the airway in C3-deficienct mice (12), suggesting the recruitment depends on C3 cleavage. Differently, the recruitment of neutrophils mainly depends on the interaction between Gzmk+ CD8 T cells and fibroblasts. This interaction has also been observed in the context of aging via the FN1-CD49d pathway (17). This leads to a hypothesis that fibroblasts from aged or inflamed tissues attract and retain Gzmk+ CD8 T cells, which in turn stimulate fibroblasts to produce cytokines and chemokines and generate activated C3, thereby enhancing inflammation.

In addition to directly stimulating to fibroblasts, Gzmk can cleave complements produced by fibroblasts to generate C3a, which stimulates inflammasome activation and cytokines production at fibroblasts (41, 82). The specific mechanisms by which Gzmk induces an inflammatory response in fibroblasts remain unclear, but studies involving other cell types may offer informative insights. Gzmk has been shown to trigger endothelial activation and promote the production of CCl2 and IL-6 (43). Since fibroblasts also express PAR1 and its activation induces CCL2 expression in these cells (83), it is likely that Gzmk activates fibroblasts through cleaving PAR1 as well. In summary, Gzmk⁺ CD8 T cells, whether generated locally or immigrated, drive airway inflammation by directly activating fibroblasts or by producing activated C3.

Gzmk⁺ CD8 T cells in age-related diseases

Gzmk⁺ CD8 T cells were initially recognized as conserved agerelated T cells during aging. This suggests that Gzmk⁺ CD8 T cells may increase the susceptibility of older individuals to age-related diseases, such as neurodegenerative disorders and cardiovascular disease. We summarize the current evidence regarding Gzmk⁺ CD8 T cells in the context of age-related diseases in this section.

Alzheimer's disease (AD) is an age-related neurodegenerative disorder characterized by neuroinflammation, and is considered an inflammatory disease (84). Recent studies indicate that CD8 T cells play a significant role in AD (85–89), with both pathogenic and protective effects reported. Therefore, it is essential to identify specific subsets and closely investigate their function in AD. Notably, Gzmk⁺ CD8 T cells have recently been reported to be enriched in the brains of tauopathy mice and deposited onto microgila (16). Depletion of CD8 T cells promoted the spread of

phosphorylated tau (pTau) throughout the central nervous system (CNS), suggesting that Gzmk⁺ CD8 T cells may slow the tauopathy progression (16). In a different study, infiltration of CD8 T cells mediated by microglia was found to promote the tauopathy in *APOE4* knock-in/Tau P301S mice (TE4) (89). Although the authors did not examine the expression of Gzmk or the Gzmk⁺ CD8 T cells, they identified a TOX⁺ PDCD1⁺ subset that was decreased in TE4 mice, which is likely related to Gzmk⁺ CD8 T cells (17).

The presence of the blood-brain barrier (BBB) significantly influences the profile of T cells in the brain. Gzmk+ CD8 T cells express higher CD49d, which forms very late antigen-4 (VLA-4). VLA-4 interacts with VCAM-1 at brain endothelial cells (BEC) and facilitates the infiltration of CD8 T cells into the brain, which has been reported to be upregulated at the BECs of AD mice (90). Additionally, higher levels of soluble VCAM-1 in plasma from AD patients correlate with advanced dementia (91). Therefore, Gzmk⁺ CD8 T cells are selectively infiltrating into the brains of AD patients via the VLA-4-VCAM-1 pathway. Furthermore, Gzmk can stimulate endothelial cells to upregulate VCAM-1 (43). Consistently, CCR5+ GZMK+ CD8 T cells have been observed entering the CNS parenchyma of multiple sclerosis patients via GZMK-mediated transendothelial diapedesis (92). This suggests that Gzmk⁺ CD8 T cells have a greater advantage in translocating to the brains of AD patients. The evidence about Gzmk+ CD8 T cells in AD is indirect and limited; their role in AD is uncertain. Since complements and PAR1 in brains, known substrates of Gzmk, have important roles in AD (93, 94), and the close interaction between Gzmk+ CD8 T cells and microglia, it is particularly interesting to examine the function of Gzmk+ CD8 T cells in AD using specific inhibitors or knock-out mouse models.

Sepsis results in increased mortality among the elderly (95, 96), which is linked to a dysfunctional innate immune response. Proinflammatory agents, such as LPS from the intestinal microbiome, leak from damaged intestines during aging, which is associated with inflammaging (97, 98). Given that Gzmk can enhance the LPS-induced response and trigger the production of pro-inflammatory cytokines in human monocytes and mouse macrophages respectively (42, 99), the presence of increased Gzmk⁺ CD8 T cells may make old individuals more susceptible to sepsis. Therefore, targeting Gzmk⁺ CD8 T cells presents a potential therapeutic opportunity for addressing sepsis in the elderly population.

Atherosclerosis and its associated cardiovascular complications are major causes of death worldwide. Aging is a significant risk factor for atherosclerosis, which is classified as an age-related inflammatory disease (100). However, the mechanisms by which aging promotes atherosclerosis remain unclear. Research has shown that Gzmk⁺ CD8 T cells are enriched in aortas from old *Ldlr-/*mice, which develop atherosclerosis on a regular chow diet (CD). The presence of GZMK⁺ CD8 T cells has also been confirmed in human atherosclerostic plaque (15, 101). This suggests that Gzmk⁺ CD8 T cells likely play a role in the development of atherosclerosis in the context of aging. Notably, PAR1 plays a crucial role in atherosclerosis due to its pleiotropic effects on endothelial cells and macrophages (102). Vorapaxar, a selective PAR1 antagonist, has been shown to reduce atherogenesis, plaque vulnerability and

complications (102). This suggests that Gzmk from Gzmk+ CD8 T cells in older individuals may play a crucial role as an activator of PAR1 in the context of atherosclerosis. Additionally, complements also play vital roles in atherosclerosis (103), Gzmk may exacerbate the development of atherosclerosis by activating both complements and PAR1, which warrants further investigation.

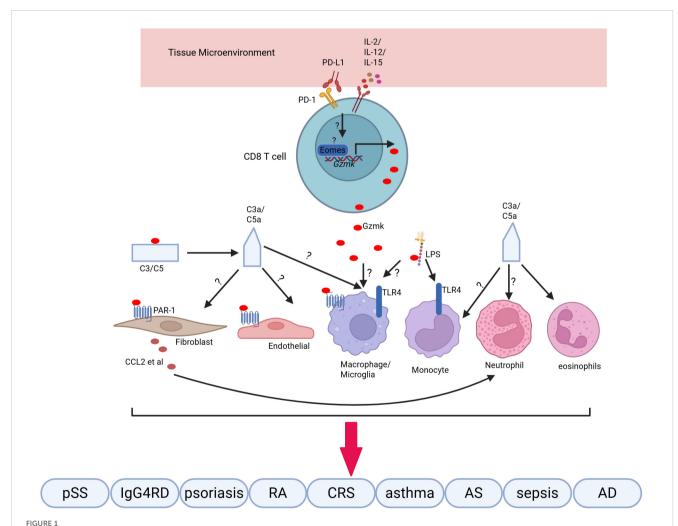
Concluding remarks

Gzmk⁺ CD8 T cells are a population of age-related T cells that accumulate in tissues as we age. This population is enriched in inflamed tissues associated with various inflammatory conditions. Gzmk plays a pro-inflammatory role by cleaving extracellular substrates, including PAR1, complement proteins, and LPS, rather than exhibiting its cytotoxic effects. This may contribute to the phenomenon of inflammaging (Figure 1). While numerous studies show a correlation between Gzmk⁺ CD8 T cells and various

inflammatory diseases, the underlying mechanisms remain limited and unclear. In addition to the known substrates, Gzmk may cleave new substrates, potentially playing unknown roles under specific conditions. Further investigations are needed to examine the factors that drive the differentiation of Gzmk⁺ CD8 T cells and the signaling pathways that regulate Gzmk expression. This research will enhance our understanding of the role of Gzmk⁺ CD8 T cells. Overall, Gzmk⁺ CD8 T cells and their substrates represent promising targets for addressing various inflammatory disorders, particularly within the context of aging.

Future direction

There is mounting evidence that Gzmk⁺ CD8 T cells may play a significant role in various inflammatory conditions by generating specific tissue-resident and/or circulating T cells. However, the origin, differentiation, and distribution profile of Gzmk+ CD8 T



Gzmk⁺ CD8 T cells synthesize and release Gzmk in response to cytokines. Extracellular Gzmk cleaves and activates PAR-1 on various cells, enhancing the inflammatory response by promoting the production of cytokines and chemokines, which recruit immune cells. Additionally, Gzmk cleaves complement proteins to generate C3a and C5a, further amplifying the inflammatory response. Finally, Gzmk binds to LPS, extracting them from their micelles and potentiating the LPS-induced pro-inflammatory response. These pro-inflammatory responses contribute to inflammation in various inflammatory diseases.

cells during aging and specific inflammatory disorders remain unclear. Future investigations should examine the common and tissue-type-specific factors responsible for generating Gzmk⁺ CD8 T cells, as well as how these factors contribute to their circulation. This research could provide potential targets for controlling Gzmk⁺ CD8 T cells. Moreover, Gzmk is the main effector of Gzmk⁺ CD8 T cells, depending on the enzymatic activity of Gzmk. Therefore, natural or small-molecule compounds that can inhibit the catalytic activity of Gzmk present a promising direction for combating inflammaging and other inflammatory conditions.

Author contributions

CX: Writing – original draft, Writing – review & editing. PL: Writing – original draft, Writing – review & editing. QZ: Writing – original draft, Writing – review & editing. WC: Conceptualization, Funding acquisition, Writing – original draft, Writing – review & editing.

Funding

The author(s) declare financial support was received for the research and/or publication of this article. This work was supported by the National Natural Science Foundation of China (32270938 to WC) and the Natural Science Foundation of Shenyang (RC220150 to WC).

References

- 1. Walter U, Santamaria P. CD8+ T cells in autoimmunity. *Curr Opin Immunol.* (2005) 17:624–31. doi: 10.1016/j.coi.2005.09.014
- 2. Nakagawa H, Wang L, Cantor H, Kim H-J. Chapter one new insights into the biology of CD8 regulatory T cells. In: Alt F, editor. *Advances in Immunology*, vol. 140. Elsevier: Academic Press (2018). p. pp 1–20.
- 3. Li J, Zaslavsky M, Su Y, Guo J, Sikora M J, van Unen V, et al. KIR⁺CD8⁺ T cells suppress pathogenic T cells and are active in autoimmune diseases and COVID-19. *Science.* (2022) 376:eabi9591. doi: 10.1126/science.abi9591
- 4. Saligrama N, Zhao F, Sikora MJ, Serratelli WS, Fernandes RA, Louis DM, et al. Opposing T cell responses in experimental autoimmune encephalomyelitis. *Nature*. (2019) 572:481-7. doi: 10.1038/s41586-019-1467-x
- 5. Giovenzana A, Codazzi V, Pandolfo M, Petrelli A. T cell trafficking in human chronic inflammatory diseases. iScience. (2024) 27:110528. doi: 10.1016/j.isci.2024.110528
- 6. Cenerenti M, Saillard M, Romero P, Jandus C. The era of cytotoxic CD4 T cells. Front Immunol. (2022) 13. doi: 10.3389/fimmu.2022.867189
- 7. Koh C-H, Lee S, Kwak M, Kim B-S, Chung Y. CD8 T-cell subsets: heterogeneity, functions, and the rapeutic potential. $\it Exp~Mol~Med.~(2023)~55:2287-99.~doi: 10.1038/s12276-023-01105-x$
- 8. Verdon DJ, Mulazzani M, Jenkins MR. Cellular and molecular mechanisms of CD8+ T cell differentiation, dysfunction and exhaustion. Int J Mol Sci. (2020) 21:7357. doi: 10.3390/ijms21197357
- 9. Goronzy JJ, Weyand CM. Mechanisms underlying T cell ageing. *Nat Rev Immunol.* (2019) 19:573–83. doi: 10.1038/s41577-019-0180-1
- 10. Jonsson AH, Zhang F, Dunlap G, Gomez-Rivas E, Watts GFM, Faust HJ, et al. Granzyme K $^+$ CD8 T cells form a core population in inflamed human tissue. *Sci Trans Med.* (2022) 14:eabo0686. doi: 10.1126/scitranslmed.abo0686
- 11. Jonsson AH. Granzyme K+ CD8 T cells in autoimmunity. Best Pract Res Clin Rheumatol. (2024) 38:101930. doi: 10.1016/j.berh.2024.101930
- 12. Lan F, Li J, Miao W, Sun F, Duan S, Song Y, et al. GZMK-expressing CD8+ T cells promote recurrent airway inflammatory diseases. *Nature*. (2025) 638:490–8. doi: 10.1038/s41586-024-08395-9

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Generative AI statement

The author(s) declare that no Generative AI was used in the creation of this manuscript.

Any alternative text (alt text) provided alongside figures in this article has been generated by Frontiers with the support of artificial intelligence and reasonable efforts have been made to ensure accuracy, including review by the authors wherever possible. If you identify any issues, please contact us.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

- 13. Guo C-L, Wang C-S, Wang Z-C, Liu F-F, Liu L, Yang Y, et al. Granzyme K+CD8 + T cells interact with fibroblasts to promote neutrophilic inflammation in nasal polyps. *Nat Commun.* (2024) 15:10413. doi: 10.1038/s41467-024-54685-1
- 14. Duquette D, Harmon C, Zaborowski A, Michelet X, O'Farrelly C, Winter D, et al. Human granzyme K is a feature of innate T cells in blood, tissues, and tumors, responding to cytokines rather than TCR stimulation. *J Immunol.* (2023) 211:633–47. doi: 10.4049/jimmunol.2300083
- 15. Smit V, De Mol J, Schaftenaar F, Depuydt MAC, Postel R, Smeets D, et al. Single-cell profiling reveals age-associated immune cells in atherosclerosis. *Atherosclerosis*. (2023) 379:S37. doi: 10.1016/j.atherosclerosis.2023.06.785
- 16. Mason HD, Latour YL, Boughter CT, Johnson KR, Maric D, Dorrier CE, et al. Granzyme K+ CD8 T cells slow tauopathy progression by targeting microglia. *Nat Immunol.* (2025) 26:1152–1167. doi: 10.1038/s41590-025-02198-4
- 17. Mogilenko DA, Shpynov O, Andhey PS, Arthur L, Swain A, Esaulova E, et al. Comprehensive profiling of an aging immune system reveals clonal GZMK*CD8* T cells as conserved hallmark of inflammaging. *Immunity*. (2021) 54:99–115.e112. doi: 10.1016/j.immuni.2020.11.005
- 18. White JT, Cross EW, Burchill MA, Danhorn T, McCarter MD, Rosen HR, et al. Virtual memory T cells develop and mediate bystander protective immunity in an IL-15-dependent manner. *Nat Commun.* (2016) 7:11291. doi: 10.1038/ncomms11291
- 19. Chiu B-C, Martin BE, Stolberg VR, Chensue SW. Cutting edge: central memory CD8 T cells in aged mice are virtual memory cells. *J Immunol.* (2013) 191:5793–6. doi: 10.4049/jimmunol.1302509
- 20. Quinn KM, Fox A, Harland KL, Russ BE, Li J, Nguyen THO, et al. Age-related decline in primary CD8+ T cell responses is associated with the development of senescence in virtual memory CD8+ T cells. *Cell Rep.* (2018) 23:3512–24. doi: 10.1016/j.celrep.2018.05.057
- 21. Cao W, Fang F, Gould T, Li X, Kim C, Gustafson C, et al. Ecto-NTPDase CD39 is a negative checkpoint that inhibits follicular helper cell generation. *J Clin Invest.* (2020) 130:3422-36. doi: 10.1172/JCI132417
- 22. Canale FP, Ramello MC, Núñez N, Furlan CLA, Bossio SN, Serrán MG, et al. CD39 expression defines cell exhaustion in tumor-infiltrating CD8⁺T cells. *Cancer Res.* (2018) 78:115–28. doi: 10.1158/0008-5472.CAN-16-2684

- 23. Gupta PK, et al. CD39 expression identifies terminally exhausted CD8+ T cells. *PLoS Pathog.* (2015) 11:e1005177. doi: 10.1371/journal.ppat.1005177
- 24. Vignali PDA, DePeaux K, Watson MJ, Ye C, Ford BR, Lontos K, et al. Hypoxia drives CD39-dependent suppressor function in exhausted T cells to limit antitumor immunity. *Nat Immunol.* (2023) 24:267–79. doi: 10.1038/s41590-022-01379-9
- 25. Gao Y, Liu R, Shi J, Shan W, Zhou H, Chen Z, et al. Clonal GZMK+CD8+ T cells are identified as a hallmark of the pathogenesis of cGVHD-induced bronchiolitis obliterans syndrome after allogeneic hematopoietic stem cell transplantation. eBioMedicine. (2025) 112. doi: 10.1016/j.ebiom.2024.105535
- 26. Zhang F, Wei K, Slowikowski K, Fonseka CY, Rao DA, Kelly S, et al. Defining inflammatory cell states in rheumatoid arthritis joint synovial tissues by integrating single-cell transcriptomics and mass cytometry. *Nat Immunol.* (2019) 20:928–42. doi: 10.1038/s41590-019-0378-1
- 27. Koga R, Maehara T, Aoyagi R, Munemura R, Murakami Y, Doi A, et al. Granzyme K- and amphiregulin-expressing cytotoxic T cells and activated extrafollicular B cells are potential drivers of IgG4-related disease. *J Allergy Clin Immunol.* (2024) 153:1095–112. doi: 10.1016/j.jaci.2023.11.916
- 28. Xu T, Zhu H-X, You X, Ma J-F, Li X, Luo P-Y, et al. Single-cell profiling reveals pathogenic role and differentiation trajectory of granzyme K+CD8+ T cells in primary Sjögren's syndrome. *JCI Insight 8*. (2023). doi: 10.1172/jci.insight.167490
- 29. Li J, He Y, Hao J, Ni L, Dong C, et al. High levels of eomes promote exhaustion of anti-tumor CD8+ T cells. *Front Immunol.* (2018) 9. doi: 10.3389/fimmu.2018.02981
- 30. Chen Z, Ji Z, Ngiow SF, Manne S, Cai Z, Huang AC, et al. TCF-1-centered transcriptional network drives an effector versus exhausted CD8 T cell-fate decision. *Immunity*. (2019) 51:840–855.e845. doi: 10.1016/j.immuni.2019.09.013
- 31. Geginat J, Vasco C, Gruarin P, Bonnal R, Rossetti G, Silvestri Y, et al. Eomesodermin-expressing type 1 regulatory (EOMES+Tr1)-like T cells: Basic biology and role in immune-mediated diseases. *Eur J Immunol.* (2023) 53:2149775. doi: 10.1002/eii.202149775
- 32. Gruarin P, Maglie S, De Simone M, Häringer B, Vasco C, Ranzani V. Eomesodermin controls a unique differentiation program in human IL-10 and IFN- γ coproducing regulatory T cells. *Eur J Immunol*. (2019) 49:96–111. doi: 10.1002/eii.201847722
- 33. Darmon AJ, Nicholson DW, Bleackley RC. Activation of the apoptotic protease CPP32 by cytotoxic T-cell-derived granzyme B. *Nature*. (1995) 377:446–8. doi: 10.1038/377446a0
- 34. Thompson R, Cao X. Reassessing granzyme B: unveiling perforin-independent versatility in immune responses and therapeutic potentials. Front Immunol. (2024) 15. doi: $10.3389/\mathrm{fimmu.}2024.1392535$
- 35. Zhao T, Zhang H, Guo Y, Zhang Q, Hua G, Lu H, et al. Granzyme K cleaves the nucleosome assembly protein SET to induce single-stranded DNA nicks of target cells. *Cell Death Differentiation*. (2007) 14:489–99. doi: 10.1038/sj.cdd.4402040
- 36. Guo Y, Chen J, Zhao T, Fan Z. Granzyme K degrades the redox/DNA repair enzyme Ape1 to trigger oxidative stress of target cells leading to cytotoxicity. *Mol Immunol.* (2008) 45:2225–35. doi: 10.1016/j.molimm.2007.11.020
- 37. Zhao T, Zhang H, Guo Y, Fan Z. Granzyme K directly processes bid to release cytochrome *c* and endonuclease G leading to mitochondria-dependent cell death *. *J Biol Chem.* (2007) 282:12104–11. doi: 10.1074/jbc.M611006200
- 38. Guo Y, Chen J, Shi L, Fan Z. Valosin-containing protein cleavage by granzyme K accelerates an endoplasmic reticulum stress leading to caspase-independent cytotoxicity of target tumor cells. *J Immunol.* (2010) 185:5348–59. doi: 10.4049/jimmunol.0903792
- 39. Hua G, Wang S, Zhong C, Xue P, Fan Z. Ignition of p53 bomb sensitizes tumor cells to granzyme K-mediated cytolysis. *J Immunol.* (2009) 182:2152–9. doi: 10.4049/jimmunol.0802307
- 40. Bouwman AC, van Daalen KR, Crnko S, ten Broeke T. Bovenschen, N. Intracellular and extracellular roles of granzyme K. *Front Immunol.* (2021) 12. doi: 10.3389/fimmu.2021.677707
- 41. Donado CA, Theisen E, Zhang F, Nathan A, Fairfield ML, Rupani KV, et al. Granzyme K activates the entire complement cascade. *Nature*. (2025) 641:211–21. doi: 10.1038/s41586-025-08713-9
- 42. Wensink AC, Kemp V, Fermie J, García Laorden MI, van der Poll T, Hack CE, et al. Granzyme K synergistically potentiates LPS-induced cytokine responses in human monocytes. *Proc Natl Acad Sci.* (2014) 111:5974–9. doi: 10.1073/pnas.1317347111
- 43. Sharma M, Merkulova Y, Raithatha S, Parkinson LG, Shen Y, Cooper D, et al. Extracellular granzyme K mediates endothelial activation through the cleavage of protease-activated receptor-1. *FEBS J.* (2016) 283:1734–47. doi: 10.1111/febs.13699
- 44. Cooper DM, Pechkovsky DV, Hackett TL, Knight DA, Granville DJ. Granzyme K activates protease-activated receptor-1. *PLoS One.* (2011) 6:e21484. doi: 10.1371/journal.pone.0021484
- 45. Brownlie RJ, Zamoyska R. T cell receptor signalling networks: branched, diversified and bounded. *Nat Rev Immunol.* (2013) 13:257–69. doi: 10.1038/nri3403
- 46. Maul-Pavicic A, Chiang SCC, Rensing-Ehl A, Jessen B, Fauriat C, Wood SM, et al. ORAII-mediated calcium influx is required for human cytotoxic lymphocyte degranulation and target cell lysis. *Proc Natl Acad Sci.* (2011) 108:3324–9. doi: 10.1073/pnas.1013285108

- 47. Pereira BI, De Maeyer RPH, Covre LP, Nehar-Belaid D, Lanna A, Ward S, et al. Sestrins induce natural killer function in senescent-like CD8+ T cells. *Nat Immunol.* (2020) 21:684–94. doi: 10.1038/s41590-020-0643-3
- 48. Lee H, Kim S-Y, Kim S-H, Yu H, Jeong S, Kim KH, et al. TCR signaling via NFATc1 constrains IL-15-induced NK-like activation of human memory CD8 $^{+}$ T cells. bioRxiv. (2025). doi: 10.1101/2025.01.13.632884
- 49. Martinez GJ, Pereira RM, Äijö T, Kim EY, Marangoni F, Pipkin ME, et al. The transcription factor NFAT promotes exhaustion of activated CD8⁺ T cells. *Immunity*. (2015) 42:265–78. doi: 10.1016/j.immuni.2015.01.006
- 50. Intlekofer AM, Takemoto N, Wherry EJ, Longworth SA, Northrup JT, Palanivel VR, et al. Effector and memory CD8+ T cell fate coupled by T-bet and eomesodermin. *Nat Immunol.* (2005) 6:1236–44. doi: 10.1038/ni1268
- 51. Paley MA, Kroy DC, Odorizzi PM, Johnnidis JB, Dolfi DV, Barnett BE, et al. Progenitor and terminal subsets of CD8⁺ T cells cooperate to contain chronic viral infection. *Science*. (2012) 338:1220–5. doi: 10.1126/science.1229620
- 52. Kao C, Oestreich KJ, Paley MA, Crawford A, Angelosanto JM, Ali M-AA, et al. Transcription factor T-bet represses expression of the inhibitory receptor PD-1 and sustains virus-specific CD8+ T cell responses during chronic infection. *Nat Immunol.* (2011) 12:663–71. doi: 10.1038/ni.2046
- 53. Nayar R, Schutten E, Jangalwe S, Durost PA, Kenney LL, Conley JM, et al. IRF4 regulates the ratio of T-bet to comesodermin in CD8+ T cells responding to persistent LCMV infection. *PLoS One.* (2016) 10:e0144826. doi: 10.1371/journal.pone.0144826
- 54. Sosinowski T, White JT, Cross EW, Haluszczak C, Marrack P, Gapin L, et al. CD8 α ⁺ Dendritic cell trans presentation of IL-15 to naive CD8⁺ T cells produces antigen-inexperienced T cells in the periphery with memory phenotype and function. *J Immunol.* (2013) 190:1936–47. doi: 10.4049/jimmunol.1203149
- 55. Coss SL, Zhou D, Chua GT, Aziz RA, Hoffman RP, Wu YL, et al. The complement system and human autoimmune diseases. J Autoimmun. (2023) 137:102979. doi: 10.1016/j.jaut.2022.102979
- 56. Arazi A, Rao DA, Berthier CC, Davidson A, Liu Y, Hoover PJ, et al. The immune cell landscape in kidneys of patients with lupus nephritis. *Nat Immunol.* (2019) 20:902–14. doi: 10.1038/s41590-019-0398-x
- 57. Dunlap GS, Billi AC, Xing X, Ma F, Maz MP, Tsoi LC, et al. Single-cell transcriptomics reveals distinct effector profiles of infiltrating T cells in lupus skin and kidney. *JCI Insight*. (2022) 7. doi: 10.1172/jci.insight.156341
- 58. Hassman LM, Paley MA, Esaulova E, Paley GL, Ruzycki PA, Linskey N, et al. Clinicomolecular identification of conserved and individualized features of granulomatous uveitis. *Ophthalmol Sci.* (2021) 1:100010. doi: 10.1016/j.xops.2021.100010
- 59. Koga R, Maehara T, Aoyagi R, Munemura R, Murakami Y, Doi A, et al. Granzyme K- and amphiregulin-expressing cytotoxic T cells and activated extrafollicular B cells are potential drivers of IgG4-related disease. *J Allergy Clin Immunol.* (2024) 153:1095–112. doi: 10.1016/j.jaci.2023.11.916
- 60. Jha P, Bora PS, Bora NS. The role of complement system in ocular diseases including uveitis and macular degeneration. *Mol Immunol.* (2007) 44:3901–8. doi: 10.1016/j.molimm.2007.06.145
- 61. Fujita Y, Fukui S, Umeda M, Tsuji S, Iwamoto N, Nakashima Y, et al. Clinical characteristics of patients with igG4-related disease complicated by hypocomplementemia. Front Immunol. (2022) 13. doi: 10.3389/fimmu.2022.828122
- 62. Lin W, Xin Z, Wang J, Ren X, Liu Y, Yang L, et al. Hypocomplementemia in primary Sjogren's syndrome: association with serological, clinical features, and outcome. *Clin Rheumatol.* (2022) 41:2091–102. doi: 10.1007/s10067-022-06135-w
- 63. Zhuo X, Wu Y, Fu X, Liang X, Xiang Y, Li J, et al. The Yin-Yang roles of protease-activated receptors in inflammatory signalling and diseases. *FEBS J.* (2022) 289:4000–20. doi: 10.1111/febs.16406
- 64. Yang YH, Hall P, Little CB, Fosang AJ, Milenkovski G, Santos L, et al. Reduction of arthritis severity in protease-activated receptor-deficient mice. *Arthritis Rheumatism.* (2005) 52:1325–32. doi: 10.1002/art.21001
- 65. Xue M, Lin H, Liang HPH, McKelvey K, Zhao R, March L, et al. Deficiency of protease-activated receptor (PAR) 1 and PAR2 exacerbates collagen-induced arthritis in mice via differing mechanisms. *Rheumatology*. (2020) 60:2990–3003. doi: 10.1093/rheumatology/keaa701
- 66. Boucher AA, Rosenfeldt L, Mureb D, Shafer J, Sharma BK, Lane A, et al. Cell type-specific mechanisms coupling protease-activated receptor-1 to infectious colitis pathogenesis. *J Thromb Haemostasis*. (2020) 18:91–103. doi: 10.1111/jth.14641
- 67. Lutter L, Roosenboom B, Brand EC, ter Linde JJ, Oldenburg B, van Lochem EG, et al. Homeostatic function and inflammatory activation of ileal CD8+ Tissue-resident T cells is dependent on mucosal location. *Cell Mol Gastroenterol Hepatol.* (2021) 12:1567–81. doi: 10.1016/j.jcmgh.2021.06.022
- 68. CAVALLO T, GRANHOLM NA. Lipopolysaccharide from Gram-negative bacteria enhances polyclonal B cell activation and exacerbates nephritis in MRL/lpr mice. Clin Exp Immunol. (2008) 82:515–21. doi: 10.1073/pnas.1317347111
- 69. Yokochi T, Fujii Y, Nakashima I, Asai J, Kiuchi M, Kojima K, et al. A murine model of experimental autoimmune lens-induced uveitis using Klebsiella O3 lipopolysaccharide as a potent immunological adjuvant. *Int J Exp Pathol.* (1993) 74:573–82. doi: 10.1111/j.1348-0421.1999.tb02371.x

- 70. Paeng N, Morikawa A, Kato Y, Sugiyama T, Koide N, Yoshida T, et al. Experimental murine model for autoimmune enterocolitis using klebsiella pneumoniae O3 lipopolysaccharide as a potent immunological adjuvant. *Microbiol Immunol.* (1999) 43:45–52. doi: 10.1111/j.1348-0421.1999.tb02371.x
- 71. Yoshino S, Sasatomi E, Ohsawa M. Bacterial lipopolysaccharide acts as an adjuvant to induce autoimmune arthritisin mice. *Immunology*. (2000) 99:607–14. doi: 10.1046/j.1365-2567.2000.00015.x
- 72. Deng Q, Luo Y, Chang C, Wu H, Ding Y, Xiao R, et al. The emerging epigenetic role of CD8+T cells in autoimmune diseases: A systematic review. *Front Immunol.* (2019) 10. doi: 10.3389/fimmu.2019.00856
- 73. Casciola-Rosen L, Andrade F, Ulanet D, Wong WB, Rosen A. Cleavage by granzyme B is strongly predictive of autoantigen status: implications for initiation of autoimmunity. *J Exp Med.* (1999) 190:815–26. doi: 10.1084/jem.190.6.815
- 74. Blanco P, Pitard V, Viallard J-F, Taupin J-L, Pellegrin J-L, Moreau J-F, et al. Increase in activated CD8+ T lymphocytes expressing perforin and granzyme B correlates with disease activity in patients with systemic lupus erythematosus. *Arthritis Rheumatism.* (2005) 52:201–11. doi: 10.1002/art.20745
- 75. Kim H-J, Verbinnen B, Tang X, Lu L, Cantor H. Inhibition of follicular T-helper cells by CD8+ regulatory T cells is essential for self tolerance. *Nature*. (2010) 467:328–32. doi: 10.1038/nature09370
- 76. Kim H-J, Wang X, Radfar S, Sproule TJ, Roopenian DC, Cantor H, et al. CD8⁺T regulatory cells express the Ly49 Class I MHC receptor and are defective in autoimmune prone B6-Yaa mice. *Proc Natl Acad Sci.* (2011) 108:2010–5. doi: 10.1073/pnas.1018974108
- 77. Lu L, Kim H-J, Werneck MBF, Cantor H. Regulation of $\mathrm{CD8}^+$ regulatory T cells: Interruption of the NKG2A/Qa-1 interaction allows robust suppressive activity and resolution of autoimmune disease. *Proc Natl Acad Sci.* (2008) 105:19420–5. doi: 10.1073/pnas.0810383105
- 78. Lee Y-H, Ishida Y, Rifa'i M, Shi Z, Isobe K-i, Suzuki H, et al. Essential role of CD8+CD122+ Regulatory T cells in the recovery from experimental autoimmune encephalomyelitis1. *J Immunol.* (2008) 180:825–32. doi: 10.4049/jimmunol.180.2.825
- 79. Bachert C, Marple B, Hosemann W, Cavaliere C, Wen W, Zhang N, et al. Endotypes of chronic rhinosinusitis with nasal polyps: pathology and possible therapeutic implications. *J Allergy Clin Immunology: In Pract.* (2020) 8:1514–9. doi: 10.1016/j.jaip.2020.03.007
- 80. Delemarre T, Holtappels G, De Ruyck N, Zhang N, Nauwynck H, Bachert C, et al. A substantial neutrophilic inflammation as regular part of severe type 2 chronic rhinosinusitis with nasal polyps. *J Allergy Clin Immunol.* (2021) 147:179–188.e172. doi: 10.1016/j.jaci.2020.08.036
- 81. Ruan J-W, Zhao J-F, Li X-L, Liao B, Pan L, Zhu K-Z, et al. Characterizing the neutrophilic inflammation in chronic rhinosinusitis with nasal polyps. *Front Cell Dev Biol.* (2021) 9. doi: 10.3389/fcell.2021.793073
- 82. Friščić J, Böttcher M, Reinwald C, Bruns H, Wirth B, Popp S-J, et al. The complement system drives local inflammatory tissue priming by metabolic reprogramming of synovial fibroblasts. *Immunity*. (2021) 54:1002–1021.e1010. doi: 10.1016/j.immuni.2021.03.003
- 83. Deng X, Mercer PF, Scotton CJ, Gilchrist A, Chambers RC. Thrombin induces fibroblast CCL2/JE production and release via coupling of PAR1 to $G\alpha q$ and cooperation between ERK1/2 and rho kinase signaling pathways. *Mol Biol Cell.* (2008) 19:2520–33. doi: 10.1091/mbc.e07-07-0720
- 84. Bettcher BM, Tansey MG, Dorothée G, Heneka MT. Peripheral and central immune system crosstalk in Alzheimer disease a research prospectus. Nat Rev Neurol. (2021) 17:689–701. doi: 10.1038/s41582-021-00549-x
- 85. Guo L, Li X, Gould T, Wang Z-Y, Cao W. T cell aging and Alzheimer's disease. Front Immunol. (2023) 14. doi: 10.3389/fimmu.2023.1154699

- 86. Su W, Saravia J, Risch I, Rankin S, Guy C, Chapman NM, et al. CXCR6 orchestrates brain CD8+ T cell residency and limits mouse Alzheimer's disease pathology. *Nat Immunol.* (2023) 24:1735–47. doi: 10.1038/s41590-023-01604-z
- 87. Jorfi M, Park J, Hall CK, Lin C-CJ, Chen M, von Maydell D, et al. Infiltrating CD8+ T cells exacerbate Alzheimer's disease pathology in a 3D human neuroimmune axis model. *Nat Neurosci.* (2023) 26:1489–504. doi: 10.1038/s41593-023-01415-3
- 88. Gate D, Saligrama N, Leventhal O, Yang AC, Unger MS, Middeldorp J, et al. Clonally expanded CD8 T cells patrol the cerebrospinal fluid in Alzheimer's disease. *Nature*. (2020) 577:399–404. doi: 10.1038/s41586-019-1895-7
- 89. Chen X, Firulyova M, Manis M, Herz J, Smirnov I, Aladyeva E, et al. Microglia-mediated T cell infiltration drives neurodegeneration in tauopathy. *Nature*. (2023) 615:668–677. doi: 10.1002/alz.070849
- 90. Zenaro E, Pietronigro E, Bianca VD, Piacentino G, Marongiu L, Budui S, et al. Neutrophils promote Alzheimer's disease-like pathology and cognitive decline via LFA-1 integrin. *Nat Med.* (2015) 21:880–6. doi: 10.1038/nm.3913
- 91. Zenaro E, Piacentino G, Constantin G. The blood-brain barrier in Alzheimer's disease. Neurobiol Dis. (2017) 107:41-56. doi: 10.1016/i.nbd.2016.07.007
- 92. Herich S, Schneider-Hohendorf T, Rohlmann A, Khaleghi Ghadiri M, Schulte-Mecklenbeck A, Zondler L, et al. Human CCR5high effector memory cells perform CNS parenchymal immune surveillance via GZMK-mediated transendothelial diapedesis. *Brain.* (2019) 142:3411–27. doi: 10.1093/brain/awz301
- 93. Dalakas MC, Alexopoulos H, Spaeth PJ. Complement in neurological disorders and emerging complement-targeted therapeutics. *Nat Rev Neurol.* (2020) 16:601–17. doi: 10.1038/s41582-020-0400-0
- 94. Zare D, Rajizadeh MA, Maneshian M, Jonaidi H, Sheibani V, Asadi-Shekaari M, et al. Inhibition of protease-activated receptor 1 (PAR1) ameliorates cognitive performance and synaptic plasticity impairments in animal model of Alzheimer's diseases. *Psychopharmacology*. (2021) 238:1645–56. doi: 10.1007/s00213-021-05798-8
- 95. Ibarz M, Haas LEM, Ceccato A, Artigas A. The critically ill older patient with sepsis: a narrative review. *Ann Intensive Care.* (2024) 14:6. doi: 10.1186/s13613-023-01233-7
- 96. Tateda K, Matsumoto T, Miyazaki S, Yamaguchi K. Lipopolysaccharide-induced lethality and cytokine production in aged mice. *Infection Immun.* (1996) 64:769–74. doi: 10.1128/iai.64.3.769-774.1996
- 97. Steele AK, Lee EJ, Vestal B, Hecht D, Dong Z, Rapaport E, et al. Contribution of intestinal barrier damage, microbial translocation and HIV-1 infection status to an inflammaging signature. *PLoS One.* (2014) 9:e97171. doi: 10.1371/journal.pone.0097171
- 98. Walker EM, Slisarenko N, Gerrets GL, Kissinger PJ, Didier ES, Kuroda MJ, et al. Inflammaging phenotype in rhesus macaques is associated with a decline in epithelial barrier-protective functions and increased pro-inflammatory function in CD161-expressing cells. *GeroScience*. (2019) 41:739–57. doi: 10.1007/s11357-019-00099-7
- 99. Joeckel LT, Wallich R, Martin P, Sanchez-Martinez D, Weber FC, Martin SF, et al. Mouse granzyme K has pro-inflammatory potential. *Cell Death Differentiation*. (2011) 18:1112–9. doi: 10.1038/cdd.2011.5
- $100.\ Wang\ JC,$ Bennett M. Aging and atherosclerosis. Circ Res. (2012) 111:245–59. doi: 10.1161/CIRCRESAHA.111.261388
- 101. Fernandez DM, Rahman AH, Fernandez NF, Chudnovskiy A, Amir ED, Amadori L, et al. Single-cell immune landscape of human atherosclerotic plaques. *Nat Med.* (2019) 25:1576–88. doi: 10.1038/s41591-019-0590-4
- $102.\ Friebel\ J,$ et al. Pleiotropic effects of the protease-activated receptor 1 (PAR1) inhibitor, vorapaxar, on atherosclerosis and vascular inflammation. Cells. (2021) 10:3517. doi: 10.3390/cells10123517
- 103. Kiss MG, Binder CJ. The multifaceted impact of complement on atherosclerosis. Atherosclerosis. (2022) 351:29–40. doi: 10.1016/jatherosclerosis.2022.03.014