

# Increase in dimethylsulfide (DMS) emissions due to eutrophication of coastal waters offsets their reduction due to ocean acidification

# Nathalie Gypens<sup>1</sup>\* and Alberto V. Borges<sup>2</sup>

<sup>1</sup> Laboratoire d'Ecologie des Systèmes Aquatiques, Ecole Interfacultaire de Bioingénieurs, Université Libre de Bruxelles, Brussels, Belgium
<sup>2</sup> Unité d'Océanographie Chimique, Department of Astrophysics Geophysics and Oceanography, Institut de Physique (B5), Université de Liège, Liège, Belgium

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#### \*Correspondence:

Nathalie Gypens, Laboratoire d'Ecologie des Systèmes Aquatiques, Ecole Interfacultaire de Bioingénieurs, Université Libre de Bruxelles, CP-221, Bd du Triomphe, 1050 Brussels, Belgium e-mail: ngypens@ulb.ac.be Available information from manipulative experiments suggested that the emission of dimethylsulfide (DMS) would decrease in response to the accumulation of anthropogenic CO<sub>2</sub> in the ocean (ocean acidification). However, in coastal environments, the carbonate chemistry of surface waters was also strongly modified by eutrophication and related changes in biological activity (increased primary production and change in phytoplankton dominance) during the last 50 years. Here, we tested the hypothesis that DMS emissions in marine coastal environments also strongly responded to eutrophication in addition to ocean acidification at decadal timescales. We used the R-MIRO-BIOGAS model in the eutrophied Southern Bight of the North Sea characterized by intense blooms of *Phaeocystis* that are high producers of dimethylsulfoniopropionate (DMSP), the precursor of DMS. We showed that, for the period from 1951 to 2007, eutrophication actually led to an increase of DMS emissions much stronger than the response of DMS emissions to ocean acidification.

Keywords: dimethylsulfide (DMS), eutrophication, ocean acidification, coastal waters, Phaeocystis, modeling

## **INTRODUCTION**

Dimethylsulfide (DMS) is the largest natural source of sulfur to the atmosphere, and may influence climate regulation by affecting atmospheric chemistry and the heat balance of the atmosphere (Charlson et al., 1987) though the significance of this feedback still remains uncertain (Carslaw et al., 2010; Quinn and Bates, 2011). Dimethylsulfoniopropionate (DMSP), the precursor of DMS, is mainly produced by a limited number of marine microalgae; Haptophytes and dinoflagellates being usually characterized by higher cellular DMSP concentrations than diatoms (Keller et al., 1989; Stefels et al., 2007). The modification of microalgae dominance and production in response to climate warming (e.g., Blanchard et al., 2012), eutrophication (e.g., Mackenzie et al., 2011), or ocean acidification (e.g., Doney et al., 2009) is then expected to change DMS emissions in the future with a potential positive or a negative feedback on climate change (Bopp et al., 2003; Gabric et al., 2004; Kloster et al., 2007; Vallina et al., 2007; Cameron-Smith et al., 2011; Six et al., 2013).

Ocean acidification of surface waters corresponds to the increase of  $[CO_2]$  and of  $[H^+]$ , the decrease of pH, of  $[CO_3^{2-}]$ , and of the saturation state of calcium carbonate, all related to the input of anthropogenic CO<sub>2</sub> from the atmosphere. These changes of the carbonate chemistry can alter the rates of primary production and calcification of numerous marine organisms and communities (Doney et al., 2009). Marine ecosystems also respond to other human pressures such as the increased delivery of nutrients from rivers to coastal waters mainly related to waste water discharge and use of fertilizers. This leads to eutrophication that corresponds to a general increase of primary production but also

a change in the phytoplankton community and structure that impacts the whole food-web and ecosystem functioning (Cloern, 2001).

Several experiments based on the manipulation of the seawater carbonate chemistry in micro- or mesocosms have shown that ocean acidification is expected to change the DMS(P) cycle (Vogt et al., 2008; Lee et al., 2009; Hopkins et al., 2010; Kim et al., 2010; Avgoustidi et al., 2012; Spielmeyer and Pohnert, 2012; Archer et al., 2013; Arnold et al., 2013). Six et al. (2013) used a simple relationship between DMS concentration and pH from a restricted set of experiments to predict the impact of ocean acidification on future climate through changes of the oceanic emission of DMS and its effect on atmospheric chemistry. However, in coastal environments, eutrophication is expected to counter to some extent the effect of ocean acidification on the dissolved carbonate chemistry of surface waters (Borges and Gypens, 2010). Further, DMS emissions would respond to the overall increase of primary production but also to the change of phytoplankton community structure related to eutrophication.

Here, we test the hypothesis that DMS emissions in coastal environments will also strongly respond to eutrophication in addition to ocean acidification at decadal timescales. We use as a case study the strongly eutrophied Southern Bight of the North Sea (SBNS) that is a hot spot of DMS emissions (Uher, 2006; Lana et al., 2011) because of the occurrence of intense blooms of *Phaeocystis* that are high DMSP producers. We use the R-MIRO-BIOGAS model that has been previously validated in the SBNS to which was added the parameterization of DMS change as a function of pH used in the global study of Six et al. (2013).

# **MATERIALS AND METHODS**

The R-MIRO-BIOGAS model results of the coupling between the RIVERSTRAHLER model, a biogeochemical model of the river system (Garnier et al., 2002), and the MIRO-BIOGAS model. The coupled model describes the biological transformation of C, nitrogen (N), phosphorus (P), and silicate (Si) along the rivercoastal continuum as a function of meteorological conditions and changing human activity on the watershed. C and nutrient river loads to the coastal zone are simulated by the RIVERSTRAHLER model from prescribed land use and human activities on the drainage basin (Billen et al., 2001, 2005; Passy et al., 2013). In the marine domain, MIRO-BIOGAS describes the dynamics of phytoplankton (diatoms, nanoflagellates and Phaeocystis), zooplankton and bacteria involved in the degradation of organic matter and the regeneration of inorganic nutrients in the water column and the sediment (Lancelot et al., 2005). The description of the carbonate system is based on the evolution of dissolved inorganic carbon (DIC) and total alkalinity (TA) and allows the calculation of partial pressure of CO<sub>2</sub> (pCO<sub>2</sub>), pH and air-sea CO<sub>2</sub> fluxes (Gypens et al., 2004). The DMS(P) cycle describes the DMSP and DMS dynamics, including biological transformations by phytoplankton and bacteria, and physico-chemical processes (including photodegradation and DMS air-sea exchange) (Gypens et al., 2014).

To take account in the MIRO-BIOGAS model for the taxonspecific production of DMSP, the cellular production of DMSP is proportional to phytoplankton growth using a specific DMSP:C quota for each of the 3 phytoplankton groups described in the model (diatoms, nanoflagellates and Phaeocystis). The regulation of DMSP production by abiotic factors such as temperature, light, nutrients and salinity is not accounted. Indeed, at temperate latitudes, these effects are negligible (Stefels et al., 2007) and, as a first approximation, we consider that the cellular DMSP concentration of the different phytoplankton groups described in the model is fixed and specific (Gypens et al., 2014). The evolution of DMS concentration in the model results from its production by phytoplankton and bacterial enzymatic cleavage of DMSP in DMS and losses by bacterial consumption (assimilation and/or demethylation/methiolation), photochemical oxidation and emission to the atmosphere. The empirical relationship proposed by Six et al. (2013) modifies the DMS production (DMS<sub>prod</sub>) as a function of pH change according to:

$$DMS_{prod_pH} = F * DMS_{prod_ref}$$

where DMS<sub>prod\_pH</sub> is the DMS production taking into account the effect of ocean acidification, DMS<sub>prod\_ref</sub> is the DMS production in the reference simulation (unperturbed by ocean acidification), and F is given by:

$$F = 1 + \gamma * \left( pH_{year} - pH_{1951} \right)$$

where  $\gamma$  corresponds to the value of the slope (0.58) of the linear regression of DMS concentration as function of pH in a selected number of mesocosm experiments that give a "medium" response according to Six et al. (2013), pH<sub>1951</sub> corresponds to the pH mean

daily value in 1951, and  $pH_{year}$  is the corresponding pH mean daily value computed for each year of the model simulation.

Note that pH effects on model variables other than DMS are not included in the model such as pH effects on primary production, for which no consensual parameterization is available. Results amongst manipulative studies are indeed highly variable reflecting either inconsistencies in experimental approach (Richier et al., 2014), or taxon-specific differences of the physiology response to ocean acidification (Flynn et al., 2012).

For this application, the model is implemented in a 0D frame of three successive boxes delineated on the basis of the hydrological regime and river inputs (Figure 1 in Lancelot et al., 2005) from the English Channel to the Belgian coastal zone (BCZ), and model results are analyzed for the BCZ influenced by the inflowing of Atlantic waters from the French coastal zone and the Scheldt river. Model simulations were performed from 1951 to 2007 after a 10 year spin-up run by constraining the model by daily wind speed, sea surface temperature (SST), monthly atmospheric pCO<sub>2</sub>, daily global solar radiation (climatology for 1989–1999), 10-days RIVERSTRAHLER simulations for C and nutrient river loads (Seine and Scheldt), and river DIC and TA inputs (Seine and Scheldt) computed as a function of freshwater discharge (Gypens et al., 2009).

Validation against field data and assessment of performance of the different components of the R-MIRO-BIOGAS model (RIVERSTRAHLER, R-MIRO, MIRO-CO2, MIRO-DMS) has been given and extensively discussed in other papers. Validation of the river loadings simulated by the RIVERSTRAHLER model for the Seine and Scheldt rivers during 1951-2007 was given by Billen et al. (2001, 2005) and Passy et al. (2013). Validation of the key biogeochemical variables simulated by R-MIRO such as nutrient concentration, phytoplankton composition and biomass (forced by the Seine and Scheldt river loadings simulated by RIVERSTRAHLER) in the BCZ was given by Lancelot et al. (2007) and Passy et al. (2013) for the Phaeocystis cells in particular. Validation of the seasonal and interannual variation of the seawater carbonate chemistry (DIC, TA, pCO<sub>2</sub>) simulated by MIRO-CO2 was given by Gypens et al. (2004, 2011). The validation of DMS(P) dynamics for the year 1989 simulated by MIRO-DMS was given by Gypens et al. (2014).

In addition to the reference simulation, two other simulations described in **Table 1** were carried out to analyze model sensitivity to river inputs and atmospheric  $CO_2$  forcings. The effect of changes in SST and light was not tested as it was previously shown to have little impact on the long-term trend of simulated net primary production (NPP) and phytoplankton community composition (Lancelot et al., 2007).

## **RESULTS AND DISCUSSION** DMS EMISSIONS IN THE BCZ

From 1951 to 2007, three periods can be distinguished in terms of river inputs of N and P, quality of nutrient enrichment defined by winter-time dissolved inorganic nitrogen to phosphate (DIN:PO<sub>4</sub>) ratio, NPP, and pH (**Figures 1A,B,C**), as previously reported by Lancelot et al. (2007) and Borges and Gypens (2010). From 1951 to 1965, river nutrient inputs, NPP and pH remained stable. From 1966 to 1990, the river nutrient inputs increased and

### Table 1 | Description of the R-MIRO-BIOGAS simulations.

Name	Setup	Application
Reference-Simulation	All forcings vary	Describes inter-annual variability and decadal changes of DMS fluxes from 1951 to 2007 in the SBNS
pH-Simulation	Includes the parameterization of DMS concentration change as a function of pH given by Six et al. (2013)	Allows to analyse the effect of pH change on DMS emissions from 1951 to 2007 in the SBNS
Ocean acidification only-Simulation	All forcings are maintained at the 1951 values except for atmospheric $pCO_2$ and includes the parameterization of DMS concentration change as a function of pH given by Six et al. (2013)	Allows to analyse the effect of increasing atmospheric CO <sub>2</sub> on the inter-annual variability and decadal changes of DMS emissions from 1951 to 2007 in the SBNS



the winter-time DIN:PO<sub>4</sub> ratios remained close to phytoplankton requirements (Redfield ratio = 16:1; Redfield et al., 1963) leading to an increase of NPP and pH. From 1991 to 2007, the decrease of the total P river inputs (mainly due to removal of polyphosphates from washing powders) led to unbalanced winter-time DIN:PO<sub>4</sub> ratios (>16:1) and a P limitation of primary production (Lancelot et al., 2007) accompanied by a decline of NPP and pH. As previously described, from 1966 to 1990, eutrophication by increasing NPP countered the decline of pH due to atmospheric CO<sub>2</sub> increase (ocean acidification), while the decline of NPP between 1991 and 2007 led to a decline of pH faster than the one predicted from atmospheric CO<sub>2</sub> increase alone (Borges and Gypens, 2010). The emissions of DMS to the atmosphere (**Figure 1D**) responded strongly to the changes of NPP and in particular of *Phaeocystis* that accounts for about 80% of annual DMS emissions in the study area (Gypens et al., 2014). From 1951 to 1990,*Phaeocystis* accounted for about 50% of total annual NPP, and on average 64% from 1991 to 2007. From 1951 to 1965, the DMS emissions remained stable, but showed inter-annual variations that correspond to  $\pm 22\%$  of the overall annual mean. From 1966 to 1990, the DMS emissions strongly increased at a rate of 14.4 mgS m<sup>-2</sup> year<sup>-1</sup> decade<sup>-1</sup> ( $r^2 = 0.69$ , p < 0.0001), while from 1991 to 2007 the DMS emissions strongly decreased at a rate of -18.2 mgS m<sup>-2</sup> year<sup>-1</sup> decade<sup>-1</sup> ( $r^2 = 0.69$ , p < 0.0001). The highest DMS emission of 56.8 mgS m<sup>-2</sup> year<sup>-1</sup> simulated

in 1991 was 6.5 times higher than the one of 1951 (8.8 mgS m<sup>-2</sup> year<sup>-1</sup>). The annual DMS emission for the last year of the simulation period, 2007, (31.8 mgS m<sup>-2</sup> year<sup>-1</sup>) remained 3.6 times higher than the one of 1951. This shows that eutrophication strongly enhanced the DMS emissions to the atmosphere and also countered the effect of ocean acidification on DMS emissions. Indeed, a small (-0.11 mgS m<sup>-2</sup> year<sup>-1</sup> decade<sup>-1</sup>) but highly significant ( $r^2 = 0.98$ , p < 0.0001) decrease of DMS emissions was simulated between 1951 and 2007 when the system only reacted to the increase of atmospheric CO<sub>2</sub> (Ocean acidification only-Simulation). Since pH actually increased from 1966 to 1990 (Figure 1B), the inclusion in the model of the parameterization of DMS production as function of pH given by Six et al. (2013) led to DMS emissions (pH-Simulation) on average ~9% higher compared to the Reference-Simulation (Figure 1D).

## DMS EMISSION IN THE GLOBAL COASTAL OCEAN

The most recent estimate of global marine emission of DMS is 28.1 TgS year<sup>-1</sup> (Lana et al., 2011), although the respective part of emissions from continental shelves are not provided. We roughly evaluate DMS emissions from continental shelves assuming that DMS emissions are proportional to primary production (Bell et al., 2010; Miles et al., 2012; Kameyama et al., 2013). We used the average primary production value for coastal waters of 230 mgC m<sup>-2</sup> year<sup>-1</sup>, and for open ocean of  $100 \text{ mgC} \text{ m}^{-2}$ year<sup>-1</sup> (Wollast, 1998). Using the global surface area of  $27 \times 10^6$  $\rm km^2$  for continental shelves and of  $335 \times 10^6 \rm km^2$  for the open ocean, globally integrated primary production in coastal waters is  $6.2 \text{ PgC year}^{-1}$  vs.  $33.5 \text{ PgC year}^{-1}$  in the open ocean. This yields a global emission of DMS from continental shelves of 4.4 TgS year<sup>-1</sup> vs. 23.7 TgS year<sup>-1</sup> from the open ocean. We computed the primary production of coastal waters influenced by rivers based on the relative present day inputs of N from rivers of 43 TgN year<sup>-1</sup> (Seitzinger et al., 2010) and the input of N by mixing and upwelling at continental margins of 380 TgN year<sup>-1</sup> (Wollast, 1998) that sustains primary production in the remaining continental shelves. This implies that coastal waters influenced by rivers have a DMS emission of 0.4 TgS year<sup>-1</sup> corresponding to 2% of total oceanic DMS emissions, and 10% of DMS emissions from continental shelves. Using a value of 18 TgN year<sup>-1</sup> for pristine N river inputs (Meybeck, 1982), we can roughly evaluate that eutrophication has led to a 58% increase of DMS emissions from coastal waters influenced by rivers globally. This corresponds to the average increase globally, but regionally it is much higher, such as in the SBNS (510% from 1951 to 1990). In the SBNS, the delivery of nutrients has been declining since the early 1990's, however, it is expected that globally N river inputs will continue to increase in future (Galloway et al., 2004). By 2050, the total N inputs by rivers could range between 63 TgN year<sup>-1</sup> (Galloway et al., 2004) and 77 TgN year<sup>-1</sup> (Mackenzie et al., 2011). Using the average value of 70 TgN year<sup>-1</sup>, DMS emissions from coastal waters influenced by rivers could increase by 2050 by 63% (0.7 TgS year<sup>-1</sup>) that would correspond to about 3% of the present day total oceanic DMS emissions, for a region corresponding to only  $\sim 0.5\%$  of the total surface of the ocean.

# **CONCLUSIONS**

We acknowledge that the change of marine DMS emissions in response to ocean acidification involves numerous and complex changes in DMS(P) sources and sinks. In particular, some studies point to an increase of DMSP production in parallel to either an increase of primary production, as both are usually correlated (e.g., Bell et al., 2010), or a change in phytoplankton dominance as a response to ocean acidification, although generally associated to a decrease of DMS emissions (Archer et al., 2013) or dissolved DMSP concentration (Lee et al., 2009). This indicates major modifications of DMS(P) cycling in particular by bacteria and zooplankton. However, the majority of manipulative experiments (micro- and mesocosms) converge to show a decrease of DMS emissions as a consequence of ocean acidification (Hopkins et al., 2010; Avgoustidi et al., 2012; Spielmeyer and Pohnert, 2012; Archer et al., 2013; Arnold et al., 2013), although some studies show the opposite response (Kim et al., 2010; Hopkins and Archer, 2014) for undetermined reasons. DMSP production by phytoplankton can also be modified by nutrient availability, not taken into account in this application. It has been suggested that Fe- and N-limitation could increase DMSP cell quota and/or DMS emission (Sunda et al., 2002, 2007; Harada et al., 2009), but primary production in the studied zone is P-limited in general and Si-limited for diatoms (Lancelot et al., 2005, 2007). There are no or little changes in DMSP cell quota under Si- or P-limitation of primary production (Bucciarelli and Sunda, 2003).

Coastal environments are hotspots of DMS emissions and are subject to eutrophication. We showed that this major anthropogenic disturbance of marine ecosystems can lead to a stronger response of DMS emissions that actually counters the effect of ocean acidification in our particular case study in the SBNS. The response of DMS emissions to eutrophication at global scale cannot be currently modeled because the resolution of general ocean circulation models or earth system models is insufficient to represent coastal areas influenced by rivers. At present, these responses can only be investigated with regional models, hence, necessarily at local scales. We nevertheless attempted a first-order estimate of the global change of DMS emissions in response to eutrophication, and we evaluated an increase of about 63% of DMS emissions from coastal waters from present to 2050 as a result of increased NPP.

The parameterization of DMS concentration as function of pH used by Six et al. (2013) covers the range of relative changes of DMS concentrations observed so far in manipulative experiments. Hence, it is unlikely that a better representation of DMS(P) responses to ocean acidification would change the overall conclusion of the present work regarding to the relative effect of ocean acidification and eutrophication on DMS emissions in eutrophied coastal environments such as the SBNS.

# **AUTHOR CONTRIBUTIONS**

Both authors contributed equally to this work Nathalie Gypens and Alberto V. Borges jointly conceived the study, designed the model simulations, and prepared the manuscript. Nathalie Gypens run the model and Alberto V. Borges conducted the global estimates of DMS emissions.

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