



# Molecular Inferences on Scomberomorus brasiliensis, From the Western South Atlantic, Based on Two Mitochondrial Genes

Divino B. da Cunha<sup>1,2\*</sup>, Luis Fernando S. Rodrigues-Filho<sup>3</sup>, João Braúllio de Luna Sales<sup>4</sup>, Pericles Rêgo<sup>4</sup>, Cleonilde Queiroz<sup>5</sup>, Iracilda Sampaio<sup>6</sup> and Marcelo Vallinoto<sup>2</sup>

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> \*Correspondence: Divino B. da Cunha divinobruno@yahoo.com.br

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The Serra Spanish Mackerel, Scomberomorus brasiliensis is one of the principal fishery resources found in the tropical and subtropical waters of the western South Atlantic. Declining catches of S. brasiliensis off the coast of northeastern Brazil indicate that this species is being overfished. Despite the importance of this species to local fisheries, few data are available on its genetic diversity or population parameters. Given this scenario, the present study evaluated the genetic variability of S. brasiliensis off the coast of the western South Atlantic, based on mitochondrial genes. We compiled two databases, one with partial sequences of the MT-CYB gene (N = 105), and the other of the MT-ND4 gene of the Serra Spanish Mackerel, sampled from eight localities, ranging from Cumana in Venezuela to Paranaguá, in southern Brazil. The results of the analysis indicate low levels of genetic diversity in the study. Samples from Cumana, on the coast of Venezuela were the most differentiated, although the results of the AMOVA were not significant. The PhiST values, the genetic divergence, and the haplotype network all indicate the widespread sharing of haplotypes by the mackerel from all the different localities. Analyses of the historical demography, based on both markers, indicate the occurrence of past population expansion, coinciding with fluctuations in sea level that occurred 10,000 years ago. The demographic and phylogeographic analyses of the mitochondrial DNA revealed the existence of a single genetic stock of S. brasiliensis on the coast of the western South Atlantic according to. These findings are fundamental to the development of effective conservation strategies.

Keywords: demographic history, genetic diversity, western South Atlantic, mitochondrial genes, S. brasiliensis

# INTRODUCTION

The Serra Spanish mackerel, *Scomberomorus brasiliensis* (Collette et al., 1978), is a pelagic marine fish distributed in the Caribbean Sea and western South Atlantic, between southern Belize and the extreme south of Brazil, in Rio Grande do Sul (Collette and Russo, 1984). This mackerel is common near islands and open beaches and appears to be able to migrate long distances on a seasonal basis (Collette and Russo, 1984; Gold et al., 2010). The diet of *S. brasiliensis* is omnivorous, and includes a range of fish, crustaceans, and mollusks (Fonteles-Filho, 1988). Reproduction is oviparous, with pelagic eggs and larvae (Gesteira and Mesquita, 1976).

The Serra Spanish mackerel is one of the most important fishery resources in the western Atlantic. In Brazil alone, between 2009 and 2011, the annual catch of this fishery resource was approximately 9,500 t (Maia et al., 2015). In Brazil, 2,733,000 tons of *S. brasiliensis* were caught between 1970 and 2000 (FAO, 2000), with the state of Ceará contributing 40.7% of the total catch. The Brazilian bulletin of fishery and agricultural statistics reported a mean annual catch of 433,000 tons between 2009 and 2011 (MPA, 2011).

Nóbrega and Lessa (2009) found evidence of the overfishing of S. brasiliensis off the coast of the Brazilian Northeast, observing declining catches of this species and its congener, the King mackerel, Somberomorus cavalla (Gonçalves et al., 2003; Nóbrega and Lessa, 2009). In this case, the analysis of phylogeographic patterns and the genetic diversity of commercially exploited organisms is essential for the development of adequate conservation and management policies for these species (Oliveira et al., 2009, 2014; Reiss et al., 2009; Silva et al., 2015). There are a few studies of the genetic diversity of fish species exploited commercially in Brazil (Santa Brígida et al., 2007; Rodrigues et al., 2008; Silva et al., 2015) although none has contemplated S. brasiliensis. Using microsatellite markers, Gold et al. (2010) found evidence of structuring in the S. brasiliensis populations between the coast of Venezuela (Margarita Island and Cumana) and Trinidad and Tobago, which was corroborated by mitochondrial sequences. Up until now, however, there has been no molecular investigation of the Brazilian populations of S. brasiliensis, so it is not known whether these stocks are genetically distinct from those of the Caribbean region, or whether the populations of the western South Atlantic are genetically contiguous.

The present study evaluated in detail the phylogeographic genetic diversity patterns, population dynamics, and of S. brasiliensis on the western coast of the South Atlantic, based on the sequences of two distinct regions of the mitochondrial genome (including previously published data) - MT-CYB and MT-ND4 and compared the diversity indices with those available the individuals from Venezuela, for Brazil, and Trinidad and Tobago.

# MATERIALS AND METHODS

## **Ethics Statement**

All Brazilian samples of *S. brasiliensis* were obtained directly from local fish markets or other retail outlets. The species is not subject to any legal restrictions in Brazil, according to the Brazilian Environment Institute (IBAMA, 2006).

## Sampling and DNA Procedures

The *S. brasiliensis* specimens were obtained from three different locations on the Brazilian coast, that is, the municipalities of Macapá (Amapá state; N = 76) and Fortaleza (Ceará state; N = 12), and the port of Paranaguá (Paraná state; N = 17) (Figure 1A). The total DNA was extracted from the samples of muscle tissue using the standard phenol chloroform method, followed by precipitation with sodium acetate (Sambrook et al., 2001).

The primers used to amplify the MT-CYB (Cytochrome b) gene were TRNA-GluF (5' -CTYTAACCAGGACTAATGGCTTG-3') and TRNA-ThrR (5'-CCTCCGACGTCCGGYTTACAAG-3'), which were designed specifically for the present study using FastPCR software (Kalendar et al., 2017). The PCR for the MT-CYB gene was run in a total volume of 10  $\mu$ l containing 1.5  $\mu$ l of the DNA (10-20 ng),1 µl of 10× buffer (Invitrogen<sup>TM</sup>; Tris-HCl, KCl, MgCl2, pH 7.8), 1.25 µl of MgCl2, 0.6 µl mix of dNTP; 0.1 µl of Taq polymerase, 0.5  $\mu$ l of each primer (10 pmol/  $\mu$ l), and 5.05 µl of sterile bidistilled water. The reactions were run in a Gene Amp PCR System 9700 thermocycler (Applied Biosystems), with a cycling protocol of 94°C for 3 min, followed by 30 cycles of 1 min at 94°C for denaturation and 1 min at 59°C for annealing, followed by 1 min extension at 72°C, with a final extension of 10 min at 72°C.

The primers used for the amplification of the MT-ND4 (Dehvdrogenase subunit 4) gene were SbrND4F-(5'-CCACACTTATGCTCGTCC-3') and SbrND4R-(5'-GCTTTGGGAAGTCATAGGT-3') (Gold et al., 2010). In this case, the PCR protocol was also that described by Gold et al. (2010). The PCR products were purified using the ExoSAP-IT kit (Amersham Pharmacia Biotech) and sequenced using Big Dye 3.1 in an ABI 3100 automatic DNA sequencer, following the manufacturer's instructions, and a GeneAmp® PCR System 9700 thermal cycler (Applied Biosystems, Foster City, CA, United States).

Two datasets were compiled for the present study. We obtained sequences of the MT-CTB fragment of 105 *S. brasiliensis* specimens. We generated 60 nucleotide sequences of the MT-ND4 fragment (**Figure 1A**), which were added to the 128 sequences published by Gold et al. (2010), which were obtained from GenBank (**Supplementary Materials 1, 2**).

## **MOLECULAR METHODS**

The sequences were aligned using CLUSTAL W (Thompson et al., 1994), implemented in BioEdit V.7.0.4 (Hall, 1999). The



2- Isla de Margarita; 3- San Fernando; 4- Las Cuevas; 5- Erin; 6- Macapá; 7- Fortaleza and 8- Paranaguá. (B) Neighbor-Joining and Maximum Likelihood trees for the *Scomberomorus brasiliensis* haplotypes of the MT-ND4 gene. Each haplotype is represented by a different color, yellow circles = 1- Cumana, red squares = 2- Isla de Margarita, blue = 3- San Fernando, brown = 4- Las Cuevas, green = 5- Erin, gray = 6- Macapá, pink = 7- Fortaleza and black = 8- Paranaguá.

phylogenetic relationships were analyzed in PAUP\* 4.0b10 (Swofford, 2002) using the Neighbor-Joining (NJ) and Maximum Likelihood (ML) approaches. The nodes were estimated using 1,000 bootstrap pseudo-replicates (Felsenstein, 1985) and heuristic searches. The sister species Scomberomorus cavalla was used as the outgroup (Supplementary Material 1). The ML tree generated here was also used to construct the haplotype network in the Haploviewer program (Salzburger et al., 2011). The most adequate evolutionary models (TIM1 + I for MT-ND4 and HKY for MT-CYB) for the data were selected using jModelTest 2 (Darriba et al., 2012), based on the Akaike Information Criterion (AIC). We used the ARLEQUIN 3.5 software (Excoffier and Lischer, 2010) to estimate the haplotype (h; Nei, 1987) and nucleotide diversity ( $\pi$ ; Nei, 1987) of each population and the neutrality tests. We calculated Tajima's D (Tajima, 1989) and Fu's Fs (Fu, 1997), which are the most sensitive indices for the detection of deviations from neutrality due to recent population expansion or bottlenecks. The genetic differentiation between pairs of populations was estimated using the Fst fixation index (Excoffier et al., 1992), with its significance being tested by

10,000 permutations. The genetic variability among populations was tested using a hierarchical Analysis of Molecular Variance, AMOVA (Excoffier et al., 1992) run in Arlequin v.3.5.

A Bayesian Skyline Plot (BSP) was run in BEAST2 v2.0.3 (Bouckaert et al., 2014) for 200 million generations, with samples taken at intervals of 10,000 generations, using the default parameters and priors (HKY substitution model) for both markers (MT-CYB and MT-ND4), with the strict molecular clock model, tree prior and 10% of which were discarded as burnin. The convergence and mixing of the chains were inspected visually in TRACER v1.6 (Rambaut et al., 2014). The convergence and mixing were appropriate when all of the Effective sample size (ESS) values for each of the parameters analyzed were above 200. For the MT-CYB gene with three groups and the piecewise-constant skyline model, auto-optimize operators, and log parameters every 10,000 interactions. For the MT-ND4, the plot had seven groups and a piecewise-constant skyline model, auto-optimize operators, and log parameters every 10,000 interactions. For this, two simulations were run, one based on a strict molecular clock, calibrated by the mutation rate, and the other with a relaxed, log-normal non-correlated clock. The results of the two analyses (strict and relaxed clocks) were compared using the Bayes factor, run in TRACER v1.6 (Rambaut et al., 2014). The strict model was the most adequate for our data (lnBayes Factor = 3.72) (Suchard et al., 2001), using a substitution rate of 1% per million years (Donaldson and Wilson, 1999; Craig et al., 2006), with a standard deviation of 10% for the two genes separately, MT-ND4 and MT-CYB.

#### RESULTS

We obtained 105 MT-CYB sequences with total length of 410 bps from the *S. brasiliensis* specimens representing three Brazilian populations. Overall, 19 of the nucleotides were variable, and six sites were parsimonious. A total of 19 haplotypes were detected, including 11 singletons and eight shared haplotypes, one of which was very common, being shared by 54 (51%) of the specimens and all of the Brazilian populations analyzed (**Figure 2A** and **Supplementary Material 2**).

The three *S. brasiliensis* populations presented haplotype diversity (*Hd*) of between 0.701 and 0.808, while nucleotide diversity ( $\pi$ ) ranged from 0.004 to 0.002 (**Table 1**). Only the total

population and that from Macapá presented significantly negative deviations from neutrality, whereas the other populations returned negative, but non-significant values (**Table 1**). Divergence (P) varied between 0.0 and 1.5%, with a mean population divergence of 1%. Values for intra- and inter-population divergence were all below 1%. According to the PhiST values, none of the comparisons between populations were significant (**Table 2**). This is corroborated by the AMOVA, which indicated no significant structural arrangement. The overall analysis revealed that almost 99% of the observed variance was derived from within-population differentiation (**Table 2**).

A dataset of 188 sequences of 500 bps was compiled for the MT-ND4 gene, based on the 188 *S. brasiliensis* specimens representing the South American distribution of the species (**Figure 1B** and **Supplementary Material 2**). In this case, haplotype diversity (*Hd*) ranged between 0.523 and 0.866, and nucleotide diversity ( $\pi$ ) varied from 0.002 to 0.013 (**Table 1**). A total of 48 haplotypes were identified, one of which was quite common, being shared by 88 (47%) of the specimens and found in all of the populations analyzed (**Figure 2B** and **Supplementary Material 2**). The individuals from Cumana were the most variable of all the study localities, with some haplotypes being separated by up to 12 mutations (**Figure 2B**). The *Fst* 



brasiliensis, run in the Haploviewer software, based on the HKY model. Each locality is represented by a different color, and each circle denotes a specific haplotype, the frequency of which is proportional to the scale shown. The length of the branches are also proportional to the number of mutations that separate the respective haplotypes.

<b>TABLE 1</b>   Genetic diversity and the results of the neutrality tests for the different
Scomberomorus brasiliensis localities, based on two mitochondrial genes.

Locus/Population	N	Nh	s	π	Hd	Fu's Fs	Tajima's D
MT-ND4							
Macapá-AP	34	12	14	0.005	0.70	-2.885	-0.625
Fortaleza-CE	19	05	07	0.006	0.813	0.087	-0.214
Paranaguá-PR	07	03	05	0.002	0.523	0.668	-1.486
Erin	25	14	23	0.007	0.866	-3.871	-1.285
San Fernando	23	13	22	0.007	0.784	-1.643	-1.270
Las Cuecas	25	09	13	0.005	0.813	-1.315	-0.601
Isla de Margarita	26	11	23	0.007	0.784	-1.643	-1.270
Cunama	29	11	30	0.013	0.731	0.801	-0.397
Total	188	48	57	0.007	0.771	-25.75*	-1.837*
MT-CYB							
Macapá-AP	76	14	13	0.003	0.701	<b>-9.427</b> *	-1.468
Fortaleza-CE	12	03	06	0.002	0.575	-2.092	-0.633
Paranaguá-PR	17	02	07	0.004	0.808	-1.213	-1.687
Total	105	19	19	0.003	0.706	-9.382*	-1.798*

N, number of individuals; Nh, number of haplotypes; S, number of segregating sites;  $\pi$ , nucleotide diversity (Nei, 1987); Hd, haplotype diversity (Nei, 1987). The significant (P < 0.05) values of Tajima's (1989) D and Fu's (1996) Fs are marked with an asterisk (\*). All significant values are highlighted in bold and with an asterisk.

 TABLE 2 | Results of the Analysis of Molecular Variance (AMOVA) of S. brasiliensis

 from Brazil, Venezuela, and Trinidad and Tobago.

Molecular marke	r Source of variation	% of total variance	Fixation index ( <i>Phist</i> )	Ρ
	AMOVA 1			
MT-ND4	Among groups	12.95	0.008	>0.05
	Among populations/ within groups	0.71	0.122	
MT-ND4	Within populations AMOVA 2	87.76	0.129	
	Among groups	1.67	0.023	>0.05
	Among populations/within groups	2.26	0.039	
	Within populations	96.07	0.016	
MT-CYB	Among groups Within populations	1.04 98.96	0.010	>0.05

AMOVA 1—Structure tested in the MT-ND4 group = {"Cumana" + "Total"}; AMOVA 2—Structure tested in the MT-ND4 group ={"Cumana+Isla\_Margarita" + "Las\_Cuevas+San\_Fernando+Erin" + "Macapa+Fortaleza+Paranagua"}; AMOVA—Structure tested in the MT-CYB group = {"Macapá" + "Fortaleza+Paranaguá"}.

(**Table 3**), AMOVA, and haplotype network all indicate the existence of a discrete genetic structure in this region.

All the results of the Analysis of Molecular Variance (AMOVA) indicated that the variation occurs primarily within populations, rather than between them, with low and non-significant PhiST values being obtained (**Table 2**). However, in the specific case of the MT-ND4 gene, when we tested Cumana and all the other regions (AMOVA 1), 12.9% of the variation was explained by the differences between this region and the other

**TABLE 3** | Matrix of the values of the *Fst* fixation index for the MT-ND4 gene sequences from eight *S. brasiliensis* localities on the coast of Venezuela, Trinidade and Tobago, and Brazil.

1	2	3	4	5	6	7
0.000						
0.781*	0.000					
0.827*	0.024	0.000				
0.786*	-0.021	-0.009	0.000			
0.780*	-0.002	-0.010	-0.015	0.000		
0.866*	0.000	0.047	0.009	0.007	0.000	
0.800*	-0.012	0.050	-0.001	0.002	0.008	
0.874*	-0.045	-0.040	-0.058	-0.050	-0.038	-0.01
	0.781* 0.827* 0.786* 0.780* 0.866* 0.800*	0.000 0.781* 0.000 0.827* 0.024 0.786* -0.021 0.780* -0.002 0.866* 0.000 0.800* -0.012	0.000         0.781*         0.000           0.827*         0.024         0.000           0.786*         -0.021         -0.009           0.780*         -0.002         -0.010           0.866*         0.000         0.047           0.860*         -0.012         0.050	0.000         0.781*         0.000           0.827*         0.024         0.000           0.786*         -0.021         -0.009         0.000           0.786*         -0.002         -0.010         -0.015           0.866*         0.000         0.047         0.009           0.800*         -0.012         0.050         -0.011	0.000         0.000         0.000           0.781*         0.000         0.000           0.827*         0.024         0.000           0.786*         -0.021         -0.009         0.000           0.780*         -0.002         -0.010         -0.015         0.000           0.866*         0.000         0.047         0.009         0.007           0.866*         -0.012         0.050         -0.011         0.002	0.000 0.781* 0.000 0.827* 0.024 0.000 0.786* −0.021 −0.009 0.000 0.780* −0.002 −0.010 −0.015 0.000 0.866* 0.000 0.047 0.009 0.007 0.000 0.800* −0.012 0.050 −0.001 0.002 0.008

\*Significant value (P < 0.005). All significant values are highlighted in bold and with an asterisk.

localities (**Table 2**). This result is supported by the *Fst* indices between localities, which were significant (**Table 3**).

In the case of the neutrality tests, both *Fu's F* and *Tajima's D* were significant only when the *S. brasiliensis* individuals were grouped within a single population for both mitochondrial genes. In accordance with *Fu's F* and *Tajima's D* for the total population, the Bayesian Skyline Plot presented evidence of demographic expansion in the *S. brasiliensis* population, dated to the Holocene in both markers. The phylogenetic analyses were run only for the MT-ND4 gene. The phylogeographic reconstructions based on the NJ and ML approaches identified two strongly supported groups of haplotypes. The first clade was formed by the four haplotypes from Cumana, which is represented by the yellow circles to highlight that there are two divergent groups (bootstrap NJ 97% and 80% ML), while the second clade contains the remaining *S. brasiliensis* haplotypes (**Figure 1B**).

## DISCUSSION

*Scomberomorus brasiliensis* is one of the most important fishery resources in the central Atlantic and northern Brazil, which may be suffering the effects of overfishing in some Brazilian regions (Nóbrega and Lessa, 2009). The present study analyzed datasets obtained from two distinct regions of the mitochondrial genome of *S. brasiliensis* specimens from the Brazilian coast, MT-CYB and MT-ND4. The MT-ND4 sequences were obtained from 188 *S. brasiliensis* specimens representing the known distribution of the species at South American.

The MT-ND4 sequences were structured in the *S. brasiliensis* specimens from Cunama, as indicated by the formation of two well-supported haplotype groups in the NJ and ML trees (**Figure 1B**). The *Fst* indices (**Table 3**) also indicated a greater degree of differentiation of these haplotypes, in comparison with the other localities. Gold et al. (2010) found genetic structuring in the Cumana specimens, a pattern like that observed in the present study, which was supported by the phylogeographic reconstructions (**Figure 1B**). One reason for the divergence of the Cumana population may be its geographic location (**Figure 1A**), which is relatively isolated from the influence of adjacent marine environments, a situation that may limit its contact with other

populations. Even so, fidelity to spawning sites, adaptations to specific oceanographic conditions, or feeding preferences may also be relevant here (Gold et al., 2010; López et al., 2010; Zhu et al., 2014; Córdova-Alarcón et al., 2019).

It is important to note, however, that no evidence from either the population analyses or the haplotype network indicated any structuring among the other sites analyzed. According to the mitochondrial data, then, there is a single genetic stock of *S. brasiliensis* between eastern Venezuela and southern Brazil (**Figure 1B** and **Table 3**). In the MT-ND4 network, the most frequent haplotype was found in 47% of the specimens, and in all the *S. brasiliensis* populations (**Figure 2B**). As for the MT-ND4



FIGURE 3 | Bayesian Skyline plots showing changes in the effective population size over time in *Scomberomorus brasiliensis*, based on mtDNA sequences. The x axis shows time before present in years, going backward from left to right. The y axis shows the effective population size. (A) The posterior distribution of the effective population size over time based on the MT-CYB gene at three Brazilian localities, and (B) The posterior distribution of the effective population size over time based on the MT-ND4 gene from seven localities in the western South Atlantic. The dark line represents the mean, and the blue band represents the 95% highest posterior density.

gene, no population structuring was observed in the MT-CYB sequences, and the extremely low *Fst* values recorded among the three localities further reinforce the lack of population structure.

In the case of the MT-CYB, all three populations presented high levels of haplotype diversity (*Hd*: 0.701–0.808), but while low levels of nucleotide diversity were recorded in Macapá and Fortaleza, this value was relatively high in Paranaguá (**Table 1**). High levels of haplotype diversity (*Hd*: 0.523–0.866) were also recorded in the MT-ND4 gene, with low levels of nucleotide diversity ( $\pi$ : 0.002–0.013). All these values are relatively high in comparison with the sister species, *S. cavalla*, for which, Santa Brígida et al. (2007) recorded a *Hd* of 0.704.

This observed genetic homogeneity may be related to the dispersal capacity of the members of the family Scombridae (Collette and Russo, 1984; Moyle and Cech, 2004; López et al., 2010). The results of the present study are consistent with those of Batista and Fabré (2001), who concluded that the coast of the Brazilian state of Maranhão is a part of the species' migratory circuit, which may exceed 300 nautical miles. Few physical barriers to dispersal exist in the ocean, and the dispersal of pelagic eggs and larvae on ocean currents, such as the Brazilian and North Brazil currents (see Rocha-Olivares et al., 2000; Broughton et al., 2002; Córdova-Alarcón et al., 2019), may also have contributed to the genetic homogeneity observed in the present study.

A similar pattern of genetic connectivity has been observed in other fish species, such as the King mackerel, *Scomberomorus cavalla*, on the northern and northeastern coasts of Brazil (Santa Brígida et al., 2007), and the red snapper, *Lutjanus campechanus*, and Acoupa weakfish, *Cynoscion acoupa*, populations of the Brazilian coast, in which no structuring was found in mitochondrial markers (Broughton et al., 2002; Gomes et al., 2008; Rodrigues et al., 2008; Silva et al., 2015). Furthermore, Siccha-Ramírez et al. (2018) utilizing SNPs (Single Nucleotide Polymorphisms) analyze samples from two commercially important species (*S. brasiliensis* and *Thunnus albacares*), where the results suggest the absence of any genetic structure among the local populations for both species, despite the sampling points are distant approximately 3,000 km.

As migratory pelagic fish generally present low levels of geographic differentiation across their distribution in the oceans due to their way of life as pelagic larval life with external fertilization and the swimming ability of the adults, strong genetic heterogeneity has nevertheless been detected at the regional scale in *Scomberomorus commerson* (López et al., 2010; Fauvelot and Borsa, 2011). Further studies of the Brazilian *S. brasiliensis* populations, based on more variable markers, such as microsatellites and new generation sequencing, may nevertheless reveal yet undetected structuring.

The BSP reconstructions indicate that the population of *S. brasiliensis* expanded between approximately 7 and 10 million years ago. The MT-CYB data indicate that the *Ne* of the Brazilian population of *S. brasiliensis* increased around 10,000 years ago (**Figure 3A**). In the case of the MT-ND4 gene, the whole population ( $Ne = 29 \times 10^{14}$ ) expanded at approximately 7.5 million years ago (**Figure 3B**). It is important to treat the estimates obtained in the present study with caution,

however, because the divergence rates of the mitochondrial genes are highly variable in fish (Donaldson and Wilson, 1999; Waters et al., 2007).

The Holocene is characterized by an interglacial period that began approximately 10,000 years ago, when sea levels began to rise following the glaciations of the Pleistocene (Lambeck et al., 2002). The Serra Spanish mackerel is associated with areas of high productivity in which larval growth is favored (Lauth and Olson, 1996), and the increase in sea levels may have contributed to an increase in primary productivity, favoring the expansion of the *S. brasiliensis* populations into the tropical and subtropical waters of the Atlantic (Lee et al., 1995; Lambeck et al., 2002). This probable expansion of the population is supported by the negative and highly significant *Fs* and *D* values, which indicate a highly stable population with a long evolutionary history (**Table 1**).

Given the fact of S. brasiliensis is one of the most harvested marine fish species on the Brazilian coast we suggest that population of the southwestern Atlantic may correspond to a single genetic stock, and should thus be treated as a unit. The genetic variability observed in the S. brasiliensis population sampled in the present study is important for the definition of effective conservation strategies, with the primary objective of mediating the decline of the population and guaranteeing the sustainability of this fishery resource (Ward, 2000; López et al., 2010; Souza-Shibatta et al., 2018; Córdova-Alarcón et al., 2019). Given this, further research on the S. brasiliensis populations should adopt a multi-locus approach integrating microsatellites, SNPs, and RADseq (see Siccha-Ramírez et al., 2018), which would complement the findings of the present study, based on two mtDNA loci. The findings of the present study provide important insights into the genetic diversity and the potential distribution of the stocks of S. brasiliensis and provide a fundamental contribution to the development of effective, longterm management strategies for the populations of the species.

# DATA AVAILABILITY STATEMENT

The datasets generated for this study can be found in the online repositories. The names of the repository/repositories and accession number(s) can be found in the article/**Supplementary Material**.

# **ETHICS STATEMENT**

Ethical review and approval was not required for the animal study because all Brazilian samples of *S. brasiliensis* were obtained directly from local fish markets or other retail outlets. The species is not subject to any legal restrictions in Brazil, according to the Brazilian Environment Institute (IBAMA).

# **AUTHOR CONTRIBUTIONS**

DC: acquisition of samples, laboratory procedures, analysis and interpretation of the data, development of the intellectual

content, drafting and writing the manuscript, and the approval of the final version. PR: collected samples and drafted the manuscript. CQ: acquisition of samples, laboratory procedures, analysis and interpretation of data, draft of the manuscript, and the development of intellectual content. LR-F: analysis and interpretation of data, development of intellectual content, manuscript writing, and final version approval. JL: analysis and interpretation of data, draft work, development of intellectual content, writing of the manuscript, and approval of the final version. IS: interpretation of data, development of intellectual content, manuscript writing, and approval of the final version. MV: interpretation of data, development of intellectual content, manuscript writing, and final version approval. All authors contributed to the article and approved the submitted version.

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# SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fmars. 2020.558902/full#supplementary-material

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**Conflict of Interest:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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