



Ranking 67 Florida Reefs for Survival of *Acropora cervicornis* Outplants

Raymond B. Banister and Robert van Woesik*

Institute for Global Ecology, Florida Institute of Technology, Melbourne, FL, United States

Over the past three decades, coral populations have declined across the tropical and subtropical oceans because of thermal stress, coral diseases, and pollution. Restoration programs are currently attempting to re-establish depauperate coral populations along the Florida reef tract. We took an integrated Bayesian approach to determine which Florida reefs ranked highest based on the survival of outplanted colonies of *Acropora cervicornis* from 2012 to 2018. Survival of *A. cervicornis* outplants was highly variable in the upper Florida Keys with some reefs showing the highest likelihood of survival (e.g., North Dry Rocks, Carysfort, Key Largo Dry Rocks, and Conch Reef), whereas some adjacent reefs showed the lowest likelihood of survival (e.g., an Unnamed Reef, Pickles Reef, and U47 Patch Reef). Similarly, survival was highly variable in the middle and lower Florida Keys and in the Broward-Miami subregions. Survival was high and less variable in Biscayne Bay and low and less variable in the Marquesas subregions. The reefs that ranked lowest for outplant survival were exposed to high wave energy. Partitioning out the spatial effects of reefs and subregions from the model, we detected spatial latent effects of low survival that were most evident in the middle and the upper Florida Keys, particularly between 2015 and 2017. The overall high spatial and temporal variability in survival among adjacent reefs highlights a need to outplant nursery-reared colonies strategically, in order to optimize coral-population recovery efforts in Florida.

Keywords: coral, restoration, outplants, survival, *Acropora cervicornis*, Florida reef tract

OPEN ACCESS

Edited by:

Frank S. Gilliam,
University of West Florida,
United States

Reviewed by:

Joshua Patterson,
University of Florida, United States
Iliana B. Baums,
Pennsylvania State University (PSU),
United States

*Correspondence:

Robert van Woesik
rvw@fit.edu

Specialty section:

This article was submitted to
Global Change and the Future Ocean,
a section of the journal
Frontiers in Marine Science

Received: 26 February 2021

Accepted: 15 June 2021

Published: 07 July 2021

Citation:

Banister RB and van Woesik R
(2021) Ranking 67 Florida Reefs
for Survival of *Acropora cervicornis*
Outplants. *Front. Mar. Sci.* 8:672574.
doi: 10.3389/fmars.2021.672574

INTRODUCTION

Coral populations have declined globally in the past three decades (Hoegh-Guldberg et al., 2007; Edwards and Gomez, 2007; Hughes et al., 2018; Lough et al., 2018). Two acroporid species, *Acropora cervicornis* and *Acropora palmata*, which previously dominated the Caribbean, and were major reef builders through millennia (Agassiz, 1885; Vaughan, 1919; Goldberg, 1973; Marszalek et al., 1977; Precht and Miller, 2007), have experienced some of the largest declines. Diseases and thermal anomalies have been the main causes of coral population declines in the Caribbean (Porter and Meier, 1992; Aronson and Precht, 2001; Toth et al., 2014; Precht et al., 2016; Walton et al., 2018). A major outbreak of white-band disease in the 1970s caused a 95% decline in Caribbean acroporids (Aronson and Precht, 2001; Miller et al., 2002; Gardner et al., 2003). Since then, *A. cervicornis* and *A. palmata* have been listed as “threatened” under the US Endangered Species Act in 2006 (National Marine Fisheries Service, 2006) and “critically endangered” on the World Conservation Union (IUCN) Red List in 2008 (Aronson et al., 2008). Coral restoration efforts are now attempting to restore acroporid populations along the Florida reef tract.

Reef-restoration programs first appeared in Florida and the wider Caribbean in the 1990s. *Acropora* species have been targeted in recent restoration efforts because of their: (i) threatened status (National Marine Fisheries Service, 2006), (ii) low recruitment rates (van Woesik et al., 2014), (iii) rapid growth rates (Tunncliffe, 1981), (iv) importance in reef-building framework (Jackson, 1992), and (v) reproductive strategies through fragmentation (Highsmith, 1982). These factors make acroporids ideal candidates for coral restoration programs. However, restoration through propagation and outplanting techniques alone will not be enough if the benchmarks set by the Paris Agreement are not met (Hoegh-Guldberg et al., 2014).

Restoration techniques primarily rely on either securing corals that have been naturally fragmented (Bruckner and Bruckner, 2001), rescuing and relocating colonies from habitats threatened by local stressors (Gayle et al., 2005; Seguin et al., 2010; Young et al., 2012), or outplanting nursery-based corals (Ware et al., 2020). Most recent efforts have focused on outplanting nursery-reared corals and these methods are evolving rapidly (Edwards and Gomez, 2007; Rinkevich, 2014; Forsman, 2015; Page et al., 2015) and include both land-based and marine programs (Rinkevich, 1995; Oren and Benayahu, 1997; van Treeck and Schuhmacher, 1999). Coral-restoration practitioners are also attempting to identify coral genotypes that are most tolerant to disease and thermal stress (Baums et al., 2019), essentially accelerating natural selection, with the goal of using those robust individuals to repopulate reefs (van Oppen and Gates, 2006; Baums, 2008; van Oppen et al., 2015; Pausch et al., 2018).

One of the more difficult hurdles of coral restoration, however, is developing models that can accurately predict optimal localities for coral outplanting at a variety of spatial scales (Wirt et al., 2013; van Woesik et al., 2020a). Such models aim to determine which subregions (at the 100-km scale), reefs (at the 10-km scale), or habitats (at the 0.1–1-km scale) are most favorable for *Acropora* survival along the Florida reef tract. Indeed, there is an urgent need for hierarchical models that inform restoration practitioners and managers which coral species to outplant, and where to outplant them, at a range of spatial scales along the Florida reef tract.

There are, however, some analytical barriers preventing accurate predictions, primarily because acroporids along the Florida reef tract are scarce. Most models that estimate the spatial distribution of *Acropora* species in Florida have had high specificity (i.e., skilled at predicting true negatives) but low sensitivity (i.e., not skilled at predicting true positives) (van Woesik et al., 2020b). In other words, regional models have a low accuracy of predicting where an acroporid species is likely to occur because the frequency and intensity of recent disturbances makes that niche space highly dynamic and unpredictable. Therefore, developing models that can perform with limited presence data, incorporate environmental predictors, and accurately predict optimal restoration sites are urgently needed at all spatial scales along the Florida reef tract.

At the local, habitat scale, several studies have recently made considerable progress at predicting the presence of *A. cervicornis*. For example, *A. cervicornis* colonies are most commonly found growing close to reef edges, where water-flow rates are high

(D'Antonio et al., 2016), and in habitats with moderate wave energy, between 0.5 and 1.5 kJ/m² (van Woesik et al., 2020b). In addition, *A. cervicornis* outplant survival appears highest in back-reef and fore-reef habitats (van Woesik et al., 2020a). There is, however, little information on the survival of acroporids at the reef scale, even though the reef scale has been identified as the scale of highest variability in Florida (Murdoch and Aronson, 1999). To optimize restoration efforts and improve *A. cervicornis* outplant survival we developed a spatio-temporal Integrated Nested Laplace Approximation (INLA) survival model, within a Bayesian framework, which used survival data from *A. cervicornis* outplants along the Florida reef tract from 2012 to 2018. The objectives of this study were to: (i) rank 67 reefs in terms of survival of outplanted colonies of the coral *A. cervicornis* along the Florida reef tract, and (ii) determine whether outplant survival was geographically consistent through time.

MATERIALS AND METHODS

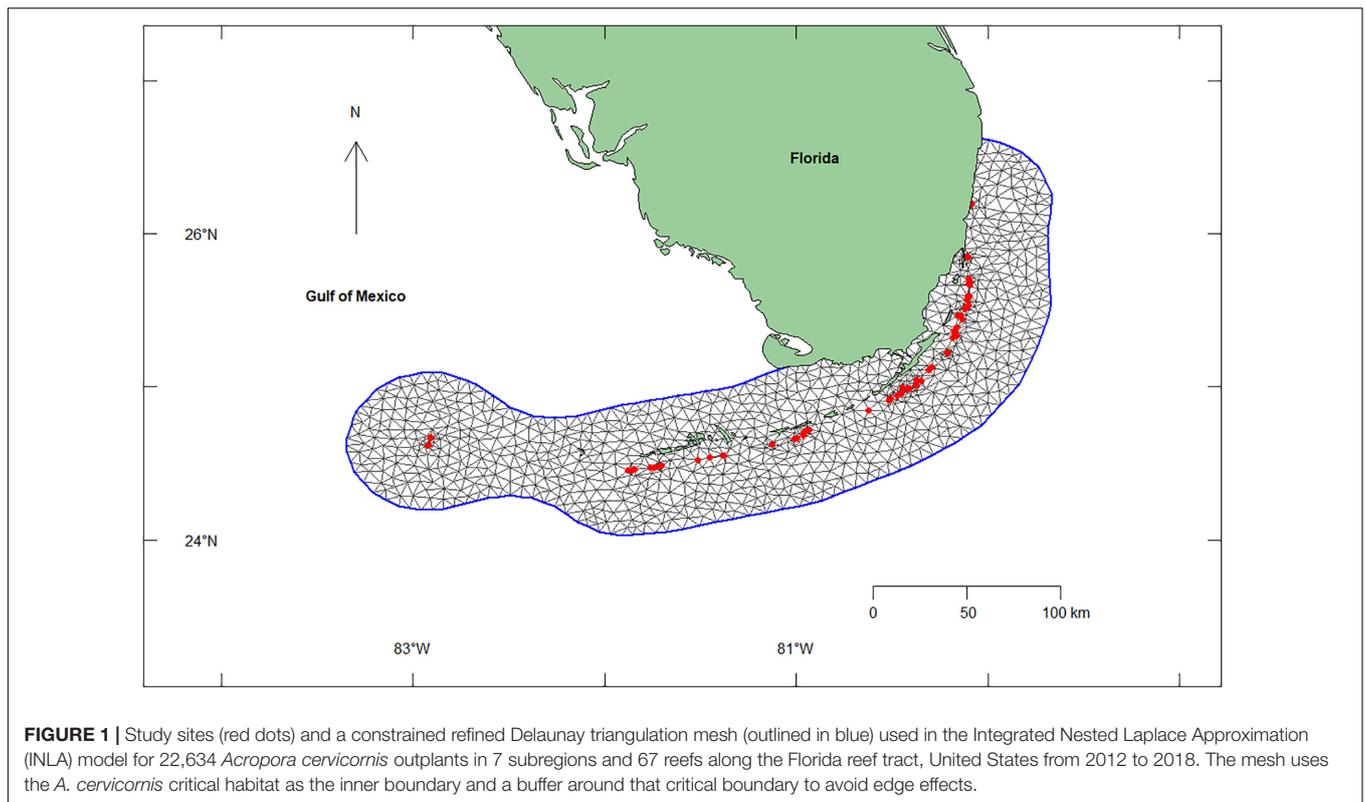
The data were a compilation of work conducted by six different coral restoration programs that examined the survival of a total of 22,634 *A. cervicornis* colonies raised in nurseries and outplanted to reef habitats along the Florida reef tract between 2012 and 2018 (van Woesik et al., 2020a). The restoration programs were conducted by: (1) The Nature Conservancy, (2) the Mote Marine Laboratory, (3) the Florida Fish and Wildlife Conservation Commission, (4) the Coral Restoration Foundation, (5) the University of Miami, and (6) Nova Southeastern University (**Supplementary Document**). All six coral restoration programs monitored the survival of *A. cervicornis* outplants after 1-month and 1-year, and several programs continued to monitor colony survival annually for 4 years (i.e., The Nature Conservancy, the Mote Marine Laboratory, the Florida Fish and Wildlife Conservation Commission, and the University of Miami). Each outplanted colony was visually assessed to determine whether it was alive or dead at each monitoring interval. An *A. cervicornis* colony was considered censored when the colony was still alive at the last monitoring interval. For analysis purposes, censored individuals were given a value of 0. By contrast, individuals that died within the timeframe of the study were given a value of 1, at the time of death. All outplanted *A. cervicornis* colonies analyzed in this study were considered as shallow (<8 m) outplants. All the data are available at: https://github.com/rvanwoesik/Acropora_survival.

Data Analysis

We used Integrated Nested Laplace Approximation (INLA) (Rue et al., 2009) within a Bayesian framework to examine spatial differences in coral survival (η) at a given site on a reef, i , expressed as:

$$\eta_i = \gamma + \sum \beta_j Z_j(s_i) + \zeta(s_i) + \varepsilon(s_i) \quad (1)$$

where γ is an intercept coefficient, β is the fixed-effect coefficient vector, Z is a matrix of covariates at the location of the data points s_i , $\zeta(s_i)$ is the spatial random effect in a spatial Gaussian



Markov Random Field (GMRF), and $\varepsilon(s_i)$ is the measurement error defined by a Gaussian white-noise process [$\sim N(0, \sigma^2_\varepsilon)$]. The GMRF combines the Gaussian field with Matérn covariance functions using stochastic partial differential equations, which in turn use a finite element representation to define the Matérn field by triangulation of the spatial domain (Lindgren et al., 2011; **Figure 1**). The GMRF computational properties have been recently enhanced by using INLA (Rue et al., 2009) for Bayesian inference, which is a computationally effective algorithm that produces fast and accurate approximations of posterior distributions (Blangiardo and Cameletti, 2015). Here we are particularly interested in the coefficients of the covariates, to determine how much of the variance is explained by the covariates reef and subregion, and the variation in the spatial latent effect explained by the variance-covariance matrix calculated using the Matérn correlation function. Model selection was based on the lowest Deviance Information Criterion (DIC) and the lowest Watanabe-Akaike Information Criterion (WAIC). To validate the model, we used spatial leave-one-out cross validation, to assess the root mean squared error (Le Rest et al., 2014). All the models were coded in R (R Core Team, 2018) and are available at: https://github.com/rvanwoesik/INLA_Florida.

In conjunction with the INLA coral survival model, we used a binary logistic regression model to examine the relationship between the probability of survival of *A. cervicornis* outplants along the Florida reef tract and wave energy. Mean wave energy was derived from site location and fetch, at a 1-km resolution (from van Woesik et al., 2020b; **Supplementary Document**) and used as the predictive covariate. For the response covariates, we

assigned 33 reefs (ranked 1–33) that had the highest likelihood of outplant survival (**Table 1**), based on the INLA results, a value of 1, and 34 reefs (ranked 34–67) that had the lowest likelihood of outplant survival (**Table 1**), based on the INLA results, a value of 0.

RESULTS

The INLA coral survival model examining “reefs” as fixed effects showed considerable variability in outplant survival across the study region. Indeed, the top ranked reefs, where the likelihood of survival of outplants was highest, were widely distributed along the Florida reef tract (**Figure 2**). There was also considerable variability in outplant survival within each of the seven subregions (**Table 1**). Survival of *Acropora cervicornis* outplants was highly variable in the upper Florida Keys with some reefs showing high survival (e.g., North Dry Rocks, Carysfort, Key Largo Dry Rocks, and Conch Reef), whereas some adjacent or nearby reefs showed low survival (e.g., an Unnamed Reef, Pickles Reef, and U47 Patch Reef) (**Figure 2**). Similarly, survival of *A. cervicornis* was highly variable in the middle and lower Florida Keys and in the Broward-Miami subregions, with some reefs showing high survival, whereas some adjacent or nearby reefs in the same subregion showed low survival (**Table 1**). By contrast, survival of *A. cervicornis* outplants in the Biscayne subregion showed low variability, but survival was consistently high (**Table 1**). Similarly, survival of *A. cervicornis* outplants was consistent within the Marquesas and the Dry Tortugas

TABLE 1 | Posteriori mean (Mean); standard deviation (SD); 0.025, and 0.975% Quantile values; and Mode of the posteriori mean values using 67 reefs as the fixed effect in the Integrated Nested Laplace Approximation (INLA) model for 22,634 *Acropora cervicornis* outplants in 7 subregions along the Florida reef tract, United States from 2012 to 2018. The 7 subregions (from north east to south west) include: 1) Broward-Miami, 2) Biscayne, 3) Upper Keys, 4) Middle Keys, 5) Lower Keys, 6) Marquesas, and 7) Dry Tortugas. The reefs are listed from highest to lowest likelihood of survival of *A. cervicornis* outplants. Carysfort Reef, ranked # 4 and marked in bold, is included in the “Iconic Reefs” initiative.

Rank	Reef	Subregion	Mean	SD	0.025% Quantile	0.975% Quantile	Mode
1	North Dry Rocks	Upper Keys	-0.0442	7.6775	-15.1176	15.0167	-0.0442
2	Davis Ledge	Upper Keys	-0.0372	7.6774	-15.1106	15.0236	-0.0372
3	CNC2	Upper Keys	-0.0346	7.6774	-15.1080	15.0262	-0.0346
4	Carysfort	Upper Keys	-0.0320	7.6774	-15.1053	15.0288	-0.0320
5	MBNOAA1	Broward-Miami	-0.0270	11.9816	-23.5509	23.4773	-0.0270
6	Key Largo Dry Rocks	Upper Keys	-0.0264	7.6774	-15.0997	15.0344	-0.0264
7	Conch	Upper Keys	-0.0255	7.6774	-15.0988	15.0354	-0.0255
8	Coffins Patch	Middle Keys	-0.0240	10.5610	-20.7589	20.6936	-0.0240
9	MBPOM1	Broward-Miami	-0.0204	11.9816	-23.5443	23.4839	-0.0204
10	Tropical Rocks	Middle Keys	-0.0172	10.5610	-20.7521	20.7004	-0.0172
11	U	Lower Keys	-0.0171	11.2043	-22.0150	21.9625	-0.0171
12	Grecian Rocks	Upper Keys	-0.0166	7.6774	-15.0900	15.0442	-0.0166
13	M9	Middle Keys	-0.0163	10.5611	-20.7513	20.7013	-0.0163
14	DPNOAA1	Biscayne	-0.0163	5.8609	-11.5232	11.4811	-0.0163
15	Q	Lower Keys	-0.0161	11.2043	-22.0139	21.9635	-0.0161
16	Fowey	Biscayne	-0.0136	5.8609	-11.5205	11.4838	-0.0136
17	White Banks	Upper Keys	-0.0123	7.6774	-15.0857	15.0485	-0.0123
18	Western Sambo	Lower Keys	-0.0111	11.2043	-22.0090	21.9684	-0.0111
19	DP Dan	Biscayne	-0.0109	5.8609	-11.5178	11.4865	-0.0109
20	Barge	Biscayne	-0.0109	5.8609	-11.5178	11.4865	-0.0109
21	Safety Valve	Biscayne	-0.0109	5.8609	-11.5178	11.4865	-0.0109
22	KBCF2	Biscayne	-0.0109	5.8609	-11.5178	11.4865	-0.0109
23	Kpeebs	Biscayne	-0.0109	5.8609	-11.5179	11.4865	-0.0109
24	Bertha	Biscayne	-0.0108	5.8609	-11.5177	11.4867	-0.0108
25	South Inshore	Biscayne	-0.0104	5.8609	-11.5174	11.4869	-0.0104
26	Molasses	Upper Keys	-0.0098	7.6774	-15.0832	15.0509	-0.0098
27	KBPOM	Biscayne	-0.0097	5.8609	-11.5166	11.4876	-0.0097
28	Flamingo	Biscayne	-0.0097	5.8609	-11.5167	11.4877	-0.0097
29	Emerald	Biscayne	-0.0088	5.8609	-11.5157	11.4886	-0.0088
30	North Emerald	Biscayne	-0.0075	5.8609	-11.5144	11.4899	-0.0075
31	CVFD	Biscayne	-0.0070	5.8609	-11.5139	11.4904	-0.0070
32	North Midchannel	Biscayne	-0.0062	5.8609	-11.5131	11.4912	-0.0062
33	Snapper Ledge	Upper Keys	-0.0059	7.6774	-15.0793	15.0549	-0.0059
34	P	Lower Keys	-0.0047	11.2043	-22.0026	21.9748	-0.0047
35	1196	Biscayne	-0.0042	5.8609	-11.5112	11.4931	-0.0042
36	Miami Beach	Broward- Miami	-0.0027	11.9816	-23.5266	23.5016	-0.0027
37	Reef 1	Dry Tortugas	-0.0011	14.1906	-27.8621	27.8366	-0.0011
38	Reef 2	Dry Tortugas	-0.0006	14.1906	-27.8616	27.8372	-0.0006
39	Cooper	Biscayne	-0.0003	5.8609	-11.5072	11.4971	-0.0003
40	Alligator	Middle Keys	0.0005	10.5611	-20.7344	20.7181	0.0005
41	Stag Acres	Middle Keys	0.0007	10.5610	-20.7342	20.7183	0.0007
42	Reef 4	Dry Tortugas	0.0049	14.1906	-27.8562	27.8426	0.0049
43	Reef T	Marquesas	0.0051	15.8791	-31.1710	31.1552	0.0051
44	Steph's	Biscayne	0.0056	5.8609	-11.5013	11.5029	0.0056
45	Nearshore Patch	Middle Keys	0.0079	10.5610	-20.7270	20.7255	0.0079
46	Struggle Bus	Biscayne	0.0085	5.8609	-11.4985	11.5058	0.0085
47	Reef S	Marquesas	0.0097	15.8791	-31.1664	31.1598	0.0097
48	Crocker	Upper Keys	0.0097	7.6774	-15.0637	15.0705	0.0097
49	Grounding	Biscayne	0.0112	5.8609	-11.4957	11.5086	0.0112
50	CWW	Lower Keys	0.0115	11.2043	-21.9864	21.9911	0.0115
51	South Midchannel	Biscayne	0.0133	5.8609	-11.4936	11.5106	0.0133
52	Reef R	Marquesas	0.0136	15.8791	-31.1625	31.1637	0.0136

(Continued)

TABLE 1 | Continued

Rank	Reef	Subregion	Mean	SD	0.025% Quantile	0.975% Quantile	Mode
53	North Inshore	Biscayne	0.0139	5.8609	-11.4931	11.5112	0.0139
54	French	Upper Keys	0.0140	7.6774	-15.0594	15.0748	0.0140
55	Staghorn City	Broward-Miami	0.0161	11.9816	-23.5078	23.5204	0.0161
56	Aruba's	Broward-Miami	0.0188	11.9816	-23.5051	23.5230	0.0188
57	M8	Middle Keys	0.0194	10.5611	-20.7155	20.7370	0.0194
58	Little Conch Ledge	Upper Keys	0.0199	7.6774	-15.0535	15.0807	0.0199
59	Reef 3	Dry Tortugas	0.0199	14.1906	-27.8411	27.8577	0.0199
60	Jons	Biscayne	0.0256	5.8609	-11.4813	11.5230	0.0256
61	South Commercial	Broward-Miami	0.0259	11.9816	-23.4980	23.5301	0.0259
62	9 Foot Stake	Lower Keys	0.0326	11.2044	-21.9654	22.0122	0.0326
63	Horseshoe Low	Middle Keys	0.0347	10.5610	-20.7002	20.7523	0.0347
64	Marker 32	Lower Keys	0.0373	11.2043	-21.9606	22.0168	0.0373
65	Unnamed Reef	Upper Keys	0.0613	7.6775	-15.0122	15.1222	0.0613
66	Pickles	Upper Keys	0.0849	7.6774	-14.9884	15.1457	0.0849
67	U47 Patch	Upper Keys	0.0886	7.6774	-14.9847	15.1494	0.0886

subregions, although survival was moderate to low (Table 1). Notably, the Marquesas and the Dry Tortugas had comparably fewer outplanting sites (3 and 4, respectively) than the other subregions.

Outplant survival on reefs was significantly ($p = 0.0124$) related to mean wave energy. Reefs exposed to moderate levels of mean wave energy ($2-4 \text{ kJ m}^{-2}$) had a high probability of outplant survival. Conversely, reefs exposed to high levels of mean wave energy ($>4 \text{ kJ m}^{-2}$) had a relatively low probability of outplant survival (Figure 3).

After removing the effects of “reefs” and “subregions” from the spatio-temporal model, spatial latent effects were evident for each sampling period (Figure 4). These spatial latent effects showed consistently low survival of *A. cervicornis* outplants in the middle Florida Keys, which were consistent through time (Figure 4). There were also spatial latent effects from 2015 through to 2017 in the upper Florida Keys, suggesting lower survival of *A. cervicornis* outplants in these years. There were no obvious spatial latent effects northeast of the upper Florida Keys and west of the lower Florida Keys, although survival of *A. cervicornis* outplants was lower between 2015 and 2017 than for 2012 and 2014 in both of those subregions (Figure 2).

DISCUSSION

This study showed considerable variability in *A. cervicornis* outplant survival across the Florida reef tract from 2012 to 2018. Reefs within the upper and middle Florida Keys had high variability in survival of *A. cervicornis* outplants, supporting some of the best, but also some of the worst, reefs for outplant survival. For example, North Dry Rocks, Carysfort, Key Largo Dry Rocks, and Conch Reef, in the upper Florida Keys, and Coffins Patch and Tropical Rocks, in the middle Florida Keys were among the best reefs for outplant survival. Yet, some reefs that were immediately adjacent to or nearby these high-survival reefs were among the

worst reefs for outplant survival (Figure 2). Such results suggest that although the environmental conditions within a subregion influence outplant survival (Toth et al., 2018; van Woësik et al., 2020a), variation among reefs within a given subregion, and variation of habitats within reefs also play major roles in the probability of outplant survival.

High variability among reefs within subregions agrees with Murdoch and Aronson (1999), who examined variation in coral cover across the Florida reef tract. They suggested that coral cover on a given reef did not predict coral cover on the adjacent or nearby reefs, because reefs are disproportionately exposed to stressors and disturbances. Some of the stressors include differential exposure to the inclement waters from Florida Bay that have long been variable in temperature, salinity, nutrients, and sediment loads (Ginsburg and Shinn, 1994). Murdoch and Aronson (1999) found that most variation along the Florida reef tract occurred among reefs at the 10–20-km scale. These results from Florida contrast with a study along the Great Barrier Reef, in Australia, by Hughes et al. (1999) who showed that the highest variation in coral cover occurred among habitats at the 0.5–3-km scale. Understanding variation in coral cover and outplant survival is critical for restoration practitioners, because such variation reflects differences in key processes, such as differential recruitment and post-settlement mortality, that can influence restoration success. In addition, outplant survival does not always suggest an increase in coral cover as growth can be independent of survival. Similarly, outplant mortality does not necessarily result in a decline in coral cover as coral colonies can fragment, move, and fuse. Monitoring outplant survival is the cornerstone of monitoring restoration success, but to improve the overall understanding of restoration success, colony growth and fragmentation records should be also examined.

Coral reef restoration and monitoring is performed by various agencies throughout the Florida Keys, with each agency generally self-restricted to a particular subregion for outplanting and monitoring. Therefore, intra-regional variation in survival is

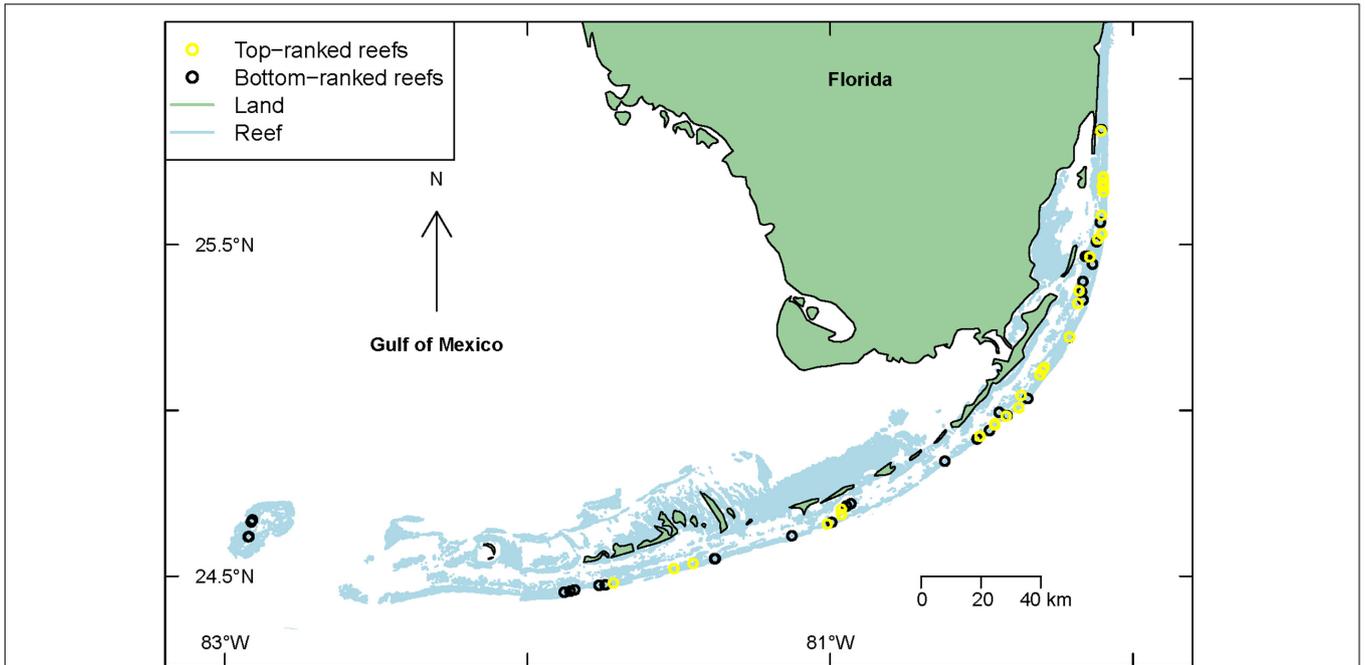


FIGURE 2 | Location of the top-ranked 33 reefs (depicted by yellow circles) with the highest likelihood of survival of *Acropora cervicornis* outplants, and the bottom-ranked 34 reefs (depicted by black circles) with the lowest likelihood of survival for *A. cervicornis* outplants in 7 subregions along the Florida reef tract, United States from 2012 to 2018—based on the *posteriori* mean values using “reef” as the fixed effect in the Integrated Nested Laplace Approximation (INLA) model.

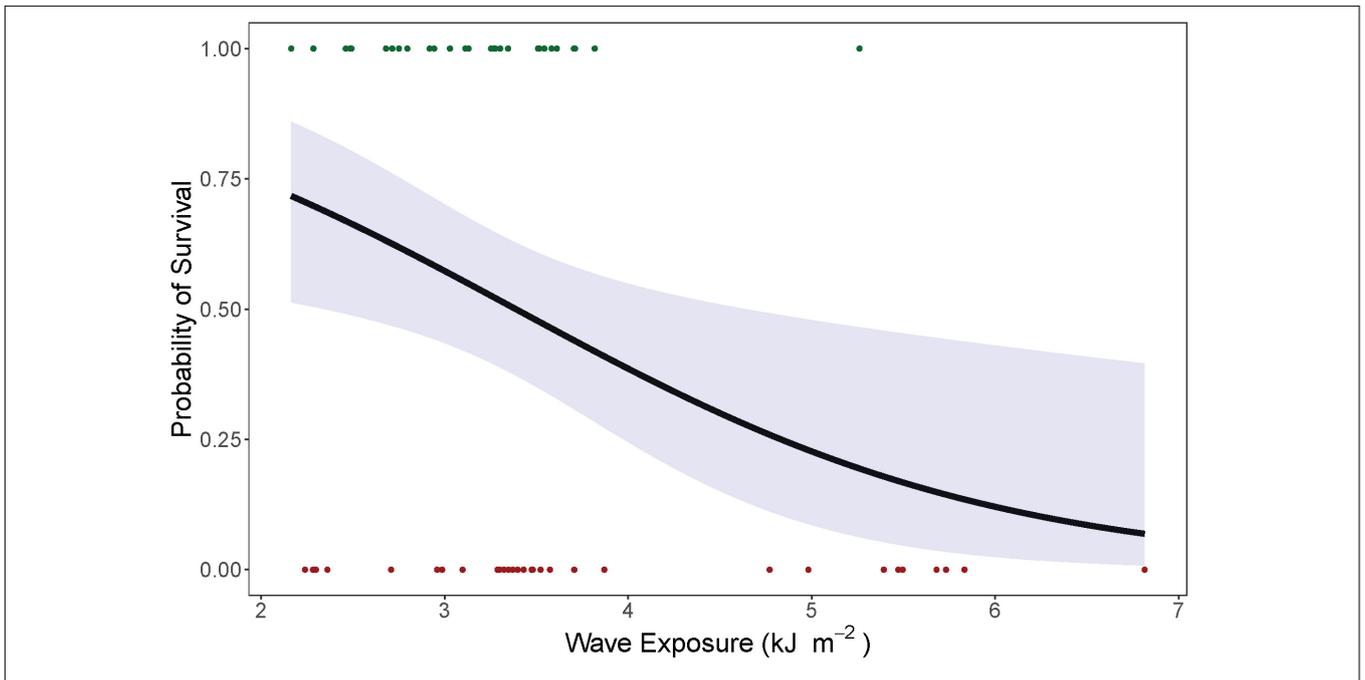
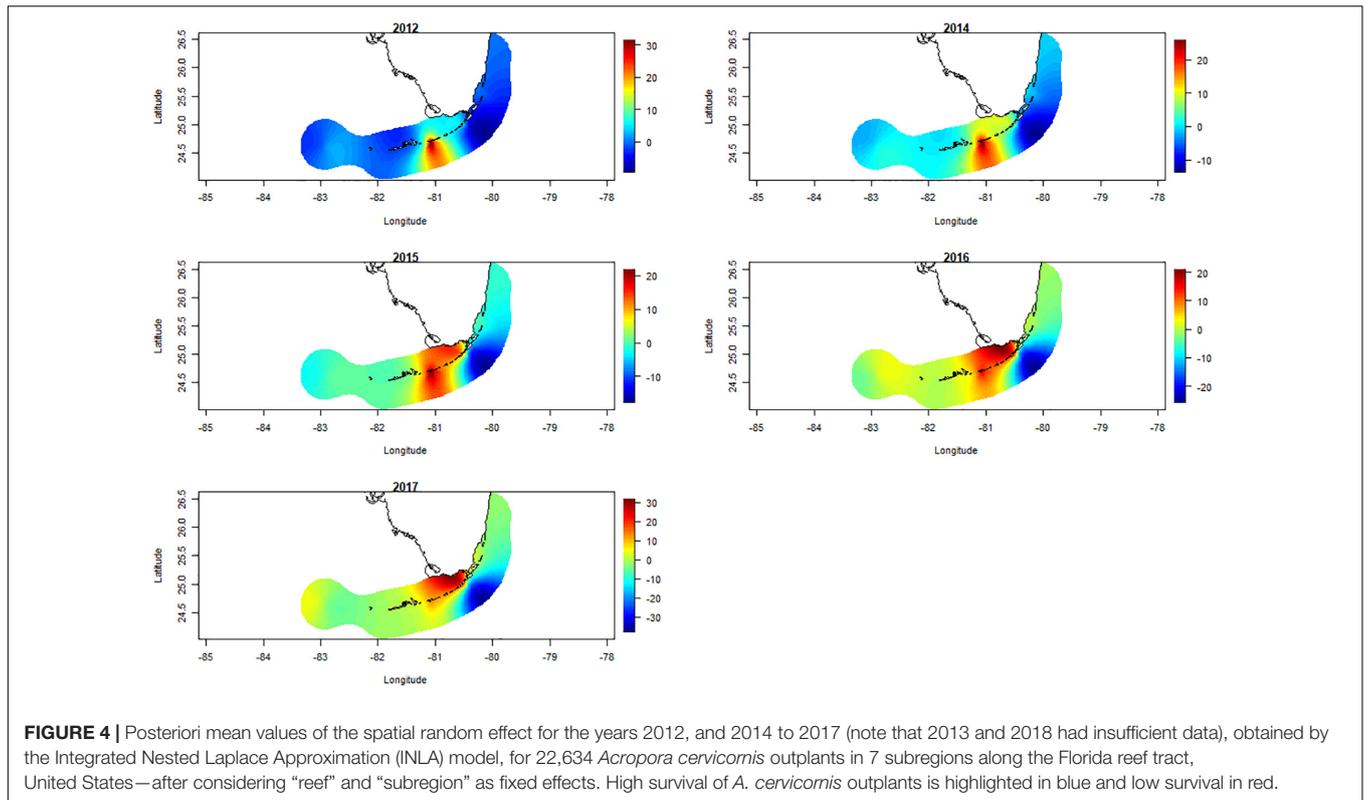


FIGURE 3 | Logistic regression (black line) showing the effect of wave energy (kJ m⁻²) on the probability of survival of *Acropora cervicornis* outplants in 7 subregions along the Florida reef tract, United States from 2012 to 2018—based on reefs with the highest likelihood of survival (green dots) and the lowest likelihood of survival (red dots). The 95% confidence intervals are shown in blue.

likely not a consequence of outplanting technique, as the same agency uses the same technique across each subregion. We did find that differences in reef exposure to water flow and

wave energy influenced outplant survival, which warrants further investigation. Based on previous findings (D’Antonio et al., 2016; van Woelik et al., 2020b) *A. cervicornis* seems to prefer reef



substrate that is exposed to moderate levels of water flow and wave energy—a finding which is supported in the present study. For example, the reefs that ranked highest for outplant survival were exposed to moderate wave energy, whereas nearby reefs that ranked lowest for outplant survival were exposed to moderate to high wave energy (Figure 3). These results point to a potential wave-energy threshold, where high wave energy is not conducive to *A. cervicornis* survival. No outplants were positioned on reefs with low wave-energy exposure, and therefore we do not know the wave-energy threshold where waters may become stagnant and thus harmful for *A. cervicornis* outplant survival.

Since suitable habitat for *A. cervicornis* outplant survival is limited throughout the Florida reef tract (van Woësik et al., 2020b), reefs showing high survival of outplants, in subregions with low overall survival, deserve special attention. Such bright spots, within subregions that are less conducive to survival, have been referred to as microrefugia and have played major roles in genetic preservation and population recovery during glacial-interglacial cycles in the past (Mosblech et al., 2011). With the continuation of climate change and ocean warming, protecting such microrefugia should be prioritized in the hope of sustaining coral populations. For example, microrefugia in the middle Florida Keys could be vital stepping-stones in maintaining connectivity among coral populations between the lower and upper Florida Keys (Frys et al., 2020). Additionally, with *A. cervicornis* and *A. palmata* listed as “threatened” and “critically endangered,” microrefuges throughout the Florida reef tract are of great importance in protecting and restoring the acroporids. Unfortunately, the genetic identity for all of the

22,634 outplants in this study were not available and prevented the assessment of individual genotypes as a confounding variable. However, we encourage the inclusion of individual genotypes as a variable in further analyses because it could reveal valuable insights into the performance of coral outplants under current and future stressors.

While the present study highlights reefs where *A. cervicornis* had the highest and lowest likelihood of survival, other coral species may have broader or narrower tolerances. Therefore, this study does not suggest abandoning coral-restoration practices in the middle Florida Keys, nor does it serve as a conduit for judging the success of other coral species. Conservation initiatives and restoration programs are only in their infancy in Florida. One such recent 2020 initiative, called “Iconic Reefs” (NOAA Fisheries, 2019), was designed to conserve and restore seven reefs along the Florida reef tract. The “Iconic Reefs” mission is an emergency restoration plan focused on providing rapid restoration efforts to seven reefs in the Florida Keys (i.e., Carysfort Reef, Horseshoe Reef, Cheeca Rocks, Sombrero Reef, Newfound Harbor, Looe Key Reef, and Eastern Dry Rocks). Carysfort Reef (north and south) is scheduled to receive 36,554 *A. cervicornis* outplants during phases 1 and 2—a notable amount of outplants, and higher than the number of outplants scheduled for the other “Iconic Reefs” (NOAA Fisheries, 2019). Carysfort Reef was ranked fourth among the highest-ranking reefs in the present study (Table 1). While we cannot comment on whether all seven reefs of the “Iconic Reefs” program will support *A. cervicornis* populations (because we did not have any outplant data on 6 of the 7 chosen reefs), the variability within our

study suggests that the seven “Iconic Reefs” will also show variable responses in outplant success, and future studies may reveal that the seven reefs vary in which coral populations they can best support.

Contemporary restoration efforts take place in a dynamic backdrop of global, regional, and local stressors (van Hooidonk et al., 2017). Coral populations in Florida have been heavily impacted by diseases and thermal-stress events (Porter and Meier, 1992; Toth et al., 2014; Precht et al., 2016; Walton et al., 2018). For example, the Stony Coral Tissue Loss Disease (SCTLD)—a particularly aggressive disease affecting more than 19 coral species throughout the Florida reef tract and wider Caribbean (Muller et al., 2020)—recently changed the composition of reefs in Florida (Muller et al., 2020). Similarly, the effects of thermal stress was evident in our study in the years 2015–2017 (Figure 4), which coincided with a global El Niño event. *A. cervicornis* outplant survival was, in general, lower between 2015 and 2017 than it was for 2012 and 2014 in the upper and lower Florida Keys. Therefore, we suggest that *A. cervicornis* restoration efforts along the Florida reef tract should: (i) continue to strive for breeding diverse yet thermally tolerant and disease resistant colonies (Baums et al., 2013; van Oppen et al., 2015), and (ii) evaluate outplant success and the dynamics of that success on a reef-by-reef basis.

The INLA approach helped us examine temporal consistency in *A. cervicornis* outplant survival, but it also helped us detect spatially latent effects that were not explicit variables in our analysis. In particular, spatially latent effects were observed in the middle Florida Keys that were independent of reef, subregion, and year of monitoring. We also noticed lower survival of *A. cervicornis* outplants in the upper Florida Keys from 2015 through to 2017 than in the other years of the study—that was most likely associated with thermal-stress related effects. In addition, the effects of Hurricane Irma may have had an effect on the survival of colonies outplanted immediately prior to its overpass in 2017 (van Woesik et al., 2020a).

In conclusion, *A. cervicornis* outplant survival was highly variable in the middle and lower Florida Keys and Broward-Miami subregions. By contrast, *A. cervicornis* outplant survival was relatively high and less variable in Biscayne Bay, and low and less variable in the Marquesas subregions. The highest variability in *A. cervicornis* outplant survival, in general, was evident among adjacent or nearby reefs, as reefs appear disproportionately exposed to stressors. Such spatial and temporal differences in survival of *A. cervicornis* outplants at a “reef” scale highlight

a need for strategic research to enhance our understanding of processes that influence growth and survival of *A. cervicornis* outplants in order to optimize population recovery along the Florida reef tract.

DATA AVAILABILITY STATEMENT

The original contributions presented in the study are included in the article/Supplementary Material, further inquiries can be directed to the corresponding author.

AUTHOR CONTRIBUTIONS

RBB and RvW analyzed the data and wrote the manuscript. RvW conceptualized the study and provided funding. Both authors contributed to the article and approved the submitted version.

FUNDING

Florida Fish and Wildlife Conservation Commission provided funds to conduct the research (award 16008).

ACKNOWLEDGMENTS

We thank extend to all the six agencies that collected the field data: The Nature Conservancy, the Mote Marine Laboratory, the Florida Fish and Wildlife Conservation Commission, the Coral Restoration Foundation, the University of Miami, and Nova Southeastern University. We would like to thank the Florida Fish and Wildlife Commission, award 16008 to RvW that partially supported this research. We would like to thank Sandra J. van Woesik for her editorial comments on the manuscript, and Lynnette Roth for her database expertise.

SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: <https://www.frontiersin.org/articles/10.3389/fmars.2021.672574/full#supplementary-material>

REFERENCES

- Agassiz, A. (1885). Explorations of the surface fauna of the Gulf Stream, under the auspices of the United States Coast Survey II. *Mem. Am. Acad. Arts Sci.* 11, 106–133.
- Aronson, R. B., Bruckner, A., Moore, J., Precht, B., and Weil, E. (2008). *Acropora cervicornis* and *A. palmata*. *The IUCN Red List of Threatened Species*. Available online at: <https://dx.doi.org/10.2305/IUCN.UK.2008.RLTS.T133006A3536699.en> (accessed May 19, 2020).
- Aronson, R. B., and Precht, W. F. (2001). White-band disease and the changing face of Caribbean coral reefs. *Hydrobiologia* 460, 25–38. doi: 10.1007/978-94-017-3284-0_2
- Baums, I. B. (2008). A restoration genetics guide for coral reef conservation. *Mol. Ecol.* 17, 2796–2811. doi: 10.1111/j.1365-294x.2008.03787.x
- Baums, I. B., Baker, A. C., Davies, S. W., Grottoli, A. G., Kenkel, C. D., Kitchen, S. A., et al. (2019). Considerations for maximizing the adaptive potential of restored coral populations in the western Atlantic. *Ecol. Appl.* 29:e01978. doi: 10.1002/eap.1978

- Baums, I. B., Devlin-Durante, M. K., Polato, N. R., Xu, D., Giri, S., Altman, N. S., et al. (2013). Genotypic variation influences reproductive success and thermal stress tolerance in the reef building coral, *Acropora palmata*. *Coral Reefs* 32, 703–717. doi: 10.1007/s00338-013-1012-6
- Blangiardo, M., and Cameletti, M. (2015). *Spatial and Spatio-temporal Bayesian Models with R – INLA*, eds M. Blangiardo and M. Cameletti (Hoboken, NJ: John Wiley & Sons), 259–302.
- Bruckner, A., and Bruckner, R. (2001). Condition of restored *Acropora palmata* fragments off Mona Island, Puerto Rico, 2 years after the Fortuna Reefer ship grounding. *Coral Reefs* 20, 235–243. doi: 10.1007/s003380100164
- D'Antonio, N. L., Gilliam, D. S., and Walker, B. K. (2016). Investigating the spatial distribution and effects of nearshore topography on *Acropora cervicornis* abundance in Southeast Florida. *PeerJ* 4:e2473. doi: 10.7717/peerj.2473
- Edwards, A., and Gomez, E. (2007). “Reef restoration concepts and guidelines: making sensible management choices in the face of uncertainty,” in *Coral Reef Targeted Research & Capacity Building for Management Programme*, ed. W. F. Precht (Newcastle upon Tyne: Newcastle University), 38.
- Forsman, A. (2015). Rethinking phenotypic plasticity and its consequences for individuals, populations and species. *Heredity* 115, 276–284. doi: 10.1038/hdy.2014.92
- Frys, C., Saint-Amand, A., Le Hénaff, M., Figueiredo, J., Kuba, A., Walker, B., et al. (2020). Fine-scale coral connectivity pathways in the Florida reef tract: implications for conservation and restoration. *Front. Mar. Sci.* 7:312. doi: 10.3389/fmars.2020.00312
- Gardner, T. A., Côte, I. M., Gill, J. A., Grant, A., and Watkinson, A. R. (2003). Long-term region-wide declines in Caribbean corals. *Science* 301, 958–960. doi: 10.1126/science.1086050
- Gayle, P. M. H., Wilson-Kelly, P., and Green, S. (2005). Transplantation of benthic species to mitigate impacts of coastal development in Jamaica. *Rev. Biol. Trop.* 53, 105–115.
- Ginsburg, R. N., and Shinn, E. A. (1994). “Preferential distribution of reefs in the Florida reef tract: the past is the key to the present,” in *Proceedings of the Colloquium on Global Aspects of Coral Reefs: Health, Hazards and History*, ed. R. N. Ginsburg (Miami, FL: University of Miami), 21–26.
- Goldberg, W. M. (1973). The ecology of the coral-octocoral communities off the southeast Florida coast: geomorphology, species composition, and zonation. *Bull. Mar. Sci.* 23, 465–488.
- Highsmith, R. C. (1982). Reproduction by fragmentation in corals. *Mar. Ecol. Prog. Ser.* 7, 207–226. doi: 10.3354/meps007207
- Hoegh-Guldberg, O., Cai, R., Poloczanska, E. S., Brewer, P. G., Sundby, S., Hilmi, K., et al. (2014). “The Ocean,” in *Climate Change 2014: Impacts, Adaptation, and Vulnerability. Part B: Regional Aspects. Contribution of Working Group II to the Fifth Assessment Report of the Intergovernmental Panel on Climate Change*, eds V. R. Barros, C. B. Field, D. J. Dokken, M. D. Mastrandrea, K. J. Mach, T. E. Bilir, et al. (Cambridge: Cambridge University Press), 1655–1731.
- Hoegh-Guldberg, O., Mumby, P. J., Hooten, A. J., Steneck, R. S., Greenfield, P., Gomezet, E., et al. (2007). Coral reefs under rapid climate change and ocean acidification. *Science* 318, 1737–1742.
- Hughes, T. P., Anderson, K. D., Connolly, S. R., Heron, S. F., Kerry, J. T., Lough, J. M., et al. (2018). Spatial and temporal patterns of mass bleaching of corals in the Anthropocene. *Science* 359, 80–83. doi:10.1126/science.aan8048
- Hughes, T. P., Baird, A. H., Dinsdale, E. A., Moltschanivskyj, N. A., Pratchett, M. S., Tanner, J. E., et al. (1999). Patterns of recruitment and abundance of corals along the Great Barrier Reef. *Nature* 397, 59–63. doi: 10.1038/16237
- Jackson, J. B. C. (1992). Pleistocene perspectives on coral reef community structure. *Am. Zool.* 32, 719–731. doi: 10.1093/icb/32.6.719
- Le Rest, K., Pinaud, D., Monestiez, P., Chadoeuf, J., and Bretagnolle, V. (2014). Spatial leave-one-out cross-validation for variable selection in the presence of spatial autocorrelation. *Glob. Ecol. Biogeogr.* 23, 811–820. doi: 10.1111/geb.12161
- Lindgren, F., Rue, H., and Lindström, J. (2011). An explicit link between Gaussian fields and Gaussian Markov random fields: the stochastic partial differential equation approach. *J. R. Stat. Soc. Ser. B Stat. Methodol.* 73, 423–498. doi: 10.1111/j.1467-9868.2011.00777.x
- Lough, J. M., Anderson, K. D., and Hughes, T. P. (2018). Increasing thermal stress for tropical coral reefs: 1871–2017. *Sci. Rep.* 8:6079.
- Marszalek, D. S., Babashoff, G. Jr., Noel, M. R., and Worley, D. R. (1977). “Reef distribution in south Florida 2,” in *Proceedings of 3rd International Coral Reef Symposium*, ed. D. L. Taylor (Miami Beach, FL: University of Miami), 223–230.
- Miller, M., Bourque, A., and Bohnsack, J. (2002). An analysis of the loss of acroporid corals at Looe Key, Florida, USA: 1983–2000. *Coral Reefs* 21, 179–182. doi: 10.1007/s00338-002-0228-7
- Mosblech, N. A. S., Bush, M. B., and van Woesik, R. (2011). On metapopulations and microrefugia: palaeoecological insights. *J. Biogeogr.* 38, 419–429. doi: 10.1111/j.1365-2699.2010.02436.x
- Muller, E. M., Sartor, C., Alcaraz, N. I., and van Woesik, R. (2020). Spatial epidemiology of the stony-coral-tissue-loss disease in Florida. *Front. Mar. Sci.* 7:163. doi: 10.3389/fmars.2020.00163
- Murdoch, T. J. T., and Aronson, R. B. (1999). Scale-dependent spatial variability of coral assemblages along the Florida Reef Tract. *Coral Reefs* 18, 341–351. doi: 10.1007/s003380050210
- National Marine Fisheries Service (2006). *Endangered and Threatened Species: Final Listing Determinations for Elkhorn Coral and Staghorn Coral*, 71(FR26852): 26852–26872. Silver Spring, MD: National Marine Fisheries Service (NMFS).
- NOAA Fisheries (2019). *Restoring Seven Iconic Reefs: A Mission to Recover the Coral Reefs of the Florida Keys. Mission: Iconic Reefs–Summary*. Available online at: https://media.fisheries.noaa.gov/dam-migration/restoring_seven_iconic_reefs_-_a_mission_to_recover_the_coral_reefs_of_the_florida_keys.pdf (accessed June 4, 2021).
- Oren, U., and Benayahu, Y. (1997). Transplantation of juvenile corals: a new approach for enhancing colonization of artificial reefs. *Mar. Biol.* 127, 499–505. doi: 10.1007/s002270050038
- Page, A. J., Cummins, C. A., Hunt, M., Wong, V. K., Reuter, S., Holden, M. T., et al. (2015). Roary: rapid large-scale prokaryote pan genome analysis. *Bioinformatics* 31, 3691–3693. doi: 10.1093/bioinformatics/btv421
- Pausch, R. E., Williams, D. E., and Miller, M. W. (2018). Impacts of fragment genotype, habitat, and size on outplanted elkhorn coral success under thermal stress. *Mar. Ecol. Prog. Ser.* 592, 109–117. doi: 10.3354/meps12488
- Porter, J. W., and Meier, O. W. (1992). Quantification of loss and change in Floridian reef coral populations. *Am. Zool.* 32, 625–640. doi: 10.1093/icb/32.6.625
- Precht, W. F., Gintert, B. E., Robbart, M. L., Fura, R., and van Woesik, R. (2016). Unprecedented disease-related coral mortality in southeastern Florida. *Sci. Rep.* 6:31374.
- Precht, W. F., and Miller, S. L. (2007). “Ecological Shifts along the Florida reef tract: the past as a key to the future,” in *Geological Approaches to Coral Reef Ecology*, ed. R. B. Aronson (New York, NY: Springer), 442.
- R Core Team (2018). *R: A Language and Environment for Statistical Computing*. Vienna: R Foundation for Statistical Computing.
- Rinkevich, B. (1995). Restoration strategies for coral reefs damaged by recreational activities: the use of sexual and asexual recruits. *Restor. Ecol.* 3, 241–251. doi: 10.1111/j.1526-100x.1995.tb00091.x
- Rinkevich, B. (2014). Rebuilding coral reefs: Does active reef restoration lead to sustainable reefs? *Curr. Opin. Environ. Sustain.* 7, 28–36. doi: 10.1016/j.cosust.2013.11.018
- Rue, H., Martino, S., and Chopin, N. (2009). Approximate Bayesian inference for latent Gaussian models by using integrated nested Laplace approximations. *J. R. Stat. Soc. Ser. B Stat. Methodol.* 71, 319–392. doi: 10.1111/j.1467-9868.2008.00700.x
- Seguín, F., Le Brun, O., Hirst, R., Al-Thary, I., and Dutrieux, E. (2010). “Large coral transplantation in Bal Haf (Yemen): an opportunity to save corals during the construction of a liquefied natural gas plant using innovative techniques,” in *Proceedings of the 11th International Coral Reef Symposium*, Fort Lauderdale, FL, 1267–1270.
- Toth, L. T., Kuffner, I. B., Stathakopoulos, A., and Shinn, E. A. (2018). A 3,000-year lag between the geological and ecological shutdown of Florida's coral reefs. *Glob. Change Biol.* 24, 5471–5483. doi: 10.1111/gcb.14389
- Toth, L. T., van Woesik, R., Smith, S. R., Murdoch, T. J. T., Ogden, J. C., Precht, W. F., et al. (2014). Do no-take reserves benefit Florida's corals? 14 years of change and stasis in the Florida Keys National Marine Sanctuary. *Coral Reefs* 33, 565–577. doi: 10.1007/s00338-014-1158-x
- Tunncliffe, V. (1981). Breakage and propagation of the stony coral *Acropora cervicornis*. *Proc. Natl. Acad. Sci. U.S.A.* 78, 2427–2431.

- van Hooidonk, R., Maynard, J., Tamelander, J., Gove, J., Ahmadi, G., Raymundo, L., et al. (2017). *Coral Bleaching Futures: Downscaled Projections of Bleaching Conditions for the World's Coral Reefs, Implications of Climate Policy and Management Responses*. Available online at: https://stg-wedocs.unep.org/bitstream/handle/20.500.11822/22048/Coral_Bleaching_Futures.pdf?sequence=1&isAllowed=y (accessed January 2021).
- van Oppen, M. J. H., and Gates, R. D. (2006). Conservation genetics and the resilience of reef-building corals. *Mol. Ecol.* 15, 3863–3883. doi: 10.1111/j.1365-294x.2006.03026.x
- van Oppen, M. J. H., Oliver, J. K., Putnam, H. M., and Gates, R. D. (2015). Building coral reef resilience through assisted evolution. *Proc. Natl. Acad. Sci. U.S.A.* 112, 2307–2313. doi: 10.1073/pnas.1422301112
- van Treeck, P., and Schuhmacher, H. (1999). Mass diving tourism – a new dimension calls for new management approaches. *Mar. Pollut. Bull.* 37, 499–504. doi: 10.1016/s0025-326x(99)00077-6
- van Woesik, R., Banister, R. B., Bartels, E., Gilliam, D. S., Goergen, E. A., Lusic, C., et al. (2020a). Differential survival of nursery-reared *Acropora cervicornis* outplants along the Florida reef tract. *Restor. Ecol.* 29:e13302.
- van Woesik, R., Roth, L. M., Brown, E. J., McCaffrey, K. R., and Roth, J. R. (2020b). Niche space of corals along the Florida reef tract. *PLoS One* 15:e0231104. doi: 10.1371/journal.pone.0231104
- van Woesik, R., Scott, W. J. I. V., and Aronson, R. B. (2014). Lost opportunities: coral recruitment does not translate to reef recovery in the Florida Keys. *Mar. Pollut. Bull.* 88, 110–117. doi: 10.1016/j.marpolbul.2014.09.017
- Vaughan, T. W. (1919). *Corals and the Formation of Coral Reefs*. Washington, DC: GPO.
- Walton, C. J., Hayes, N. K., and Gilliam, D. S. (2018). Impacts of a regional, multi-year, multi-species coral disease outbreak in southeast Florida. *Front. Mar. Sci.* 5:323. doi: 10.3389/fmars.2018.00323
- Ware, M., Garfield, E. N., Nedimyer, K., Levy, J., Kaufman, L., Precht, W., et al. (2020). Survivorship and growth in staghorn coral (*Acropora cervicornis*) outplanting projects in the Florida Keys National Marine Sanctuary. *PLoS One* 15:e0231817. doi: 10.1371/journal.pone.0231817
- Wirt, K. E., Hallock, P., Palandro, D., and Daly, K. L. (2013). Potential habitat of *Acropora* spp. on Florida reefs. *Appl. Geogr.* 39, 118–127. doi: 10.1016/j.apgeog.2012.12.009
- Young, C. N., Schopmeyer, S. A., and Lirman, D. (2012). A review of reef restoration and coral propagation using the threatened genus *Acropora* in the Caribbean and Western Atlantic. *Bull. Mar. Sci.* 88, 1075–1098. doi: 10.5343/bms.2011.1143

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2021 Banister and van Woesik. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.