



# Biochemical Pathways of Sarcopenia and Their Modulation by Physical Exercise: A Narrative Review

Mohammad Mosaferi Ziaaldini<sup>1</sup>, Emanuele Marzetti<sup>2\*</sup>, Anna Picca<sup>2</sup> and Zsolt Murlasits<sup>3</sup>

<sup>1</sup>Sport Physiology Department, Ferdowsi University of Mashhad, Mashhad, Iran, <sup>2</sup>Department of Geriatrics, Neurosciences and Orthopedics, Catholic University of the Sacred Heart, Rome, Italy, <sup>3</sup>Sport Science Program, College of Arts and Sciences, Qatar University, Doha, Qatar

## OPEN ACCESS

### Edited by:

Gary Sinoff,  
University of Haifa, Israel

### Reviewed by:

Mario Ulises Pérez-Zepeda,  
Instituto Nacional  
de Geriatria, Mexico  
Philippe De Souto Barreto,  
Centre Hospitalier Universitaire (CHU)  
de Toulouse, France

### \*Correspondence:

Emanuele Marzetti  
emarzetti@live.com

### Specialty section:

This article was submitted  
to Geriatric Medicine,  
a section of the journal  
Frontiers in Medicine

**Received:** 28 June 2017

**Accepted:** 21 September 2017

**Published:** 04 October 2017

### Citation:

Ziaaldini MM, Marzetti E, Picca A and  
Murlasits Z (2017) Biochemical  
Pathways of Sarcopenia and Their  
Modulation by Physical Exercise: A  
Narrative Review.  
Front. Med. 4:167.  
doi: 10.3389/fmed.2017.00167

Aging is a complex process characterized by progressive multisystem derangement predisposing individuals to increased risk of developing negative health outcomes. Sarcopenia is the age-related decline of muscle mass and function/strength and represents a highly prevalent correlate of aging. Several factors have been indicated to play a role in the onset and progression of sarcopenia; however, its pathophysiology is still unclear. Physical exercise is to date one of the few strategies able to improve muscle health in old age through multiple metabolic and transcriptional adaptations. Although the benefits of different exercise modalities on the function and structure of aged myocytes is acknowledged, the cellular and molecular mechanisms underlying such effects are not yet fully identified. Here, we briefly overview the current knowledge on the biochemical pathways associated with the onset and progression of sarcopenia. We subsequently describe the effects of exercise on relevant signaling pathways involved in sarcopenia pathophysiology.

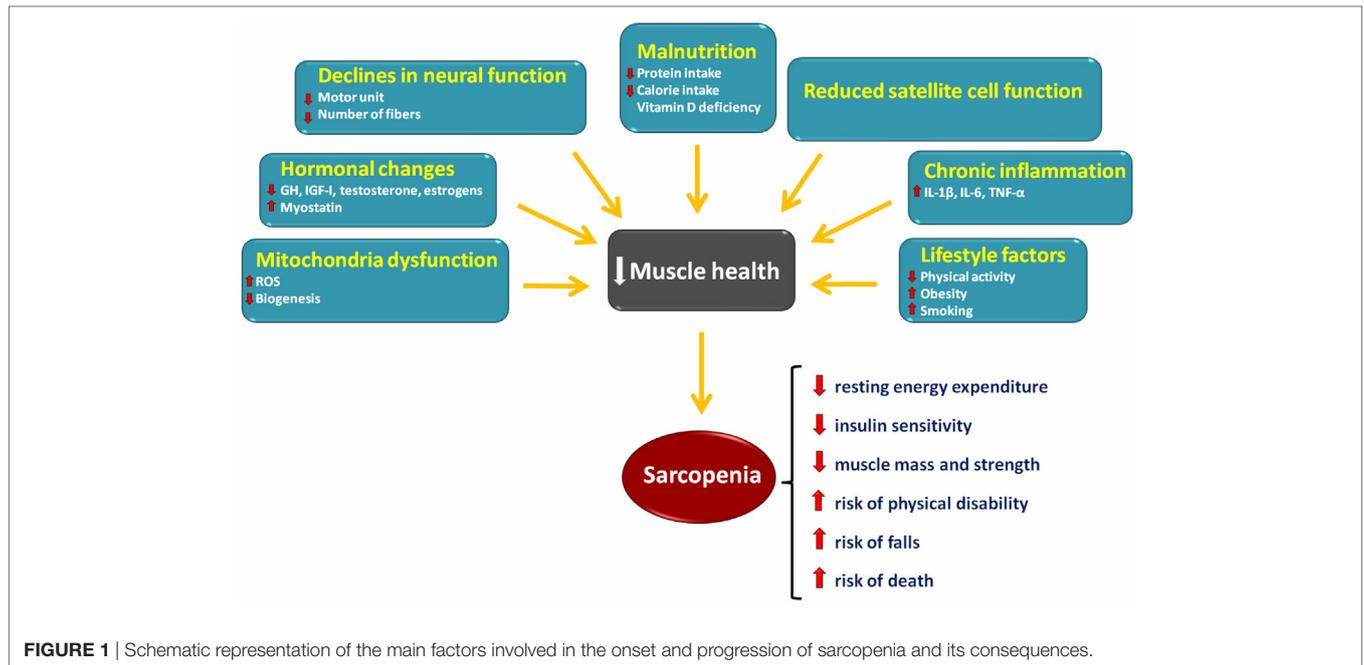
**Keywords:** muscle atrophy, physical activity, apoptosis, inflammation, mitochondria

## INTRODUCTION

Sarcopenia is the progressive loss of muscle mass and strength/function during aging and has been increasingly recognized as a relevant factor for the occurrence of negative health outcomes in late life (e.g., falls, morbidity, disability, loss of independence, and mortality) (1). Indeed, sarcopenia is endorsed as a reliable biomarker allowing for the discrimination, at a clinical level, of biological from chronological age (2).

Despite the growing interest surrounding muscle aging, several intrinsic limitations still exist impeding its appreciation as a paradigm to study the aging process. First of all, the lack of a univocal operational definition of sarcopenia beside an unbiased method for the assessment of muscle mass and function hampers the incorporation of sarcopenia in everyday clinical practice (1). In addition to this, the incomplete knowledge of the pathophysiology of this condition halts the identification of targets that could be exploited for the development of intervention strategies (3).

Low levels of physical activity are among the most important factors involved in the development of sarcopenia (4, 5) (**Figure 1**). Resistance training (RT) is the exercise strategy usually recommended to counteract age-related muscle wasting (6). Recent studies have shown that aerobic exercise



training is also able to attenuate the rate of sarcopenia development (7–9). However, the precise cellular mechanisms through which resistance and aerobic training act on sarcopenia pathophysiology have not yet been fully appreciated.

Here, we briefly overview the current knowledge on the pathways involved in the onset and progression of sarcopenia. Subsequently, we describe the most notable biochemical adaptations elicited in muscle by physical exercise with the aim of pinpointing relevant pathways potentially useful for drug development.

## SIGNALING PATHWAYS INVOLVED IN SARCOPENIA

The maintenance of skeletal muscle mass depends on the balance between anabolic and catabolic pathways (10) (Figure 1). The following sections summarize the current knowledge on major signaling pathways that are involved in sarcopenia (Figure 2).

### Insulin-Like Growth Factor 1 (IGF-1)/Akt/Mammalian Target of Rapamycin (mTOR)

Muscle size is under the control of the phosphatidylinositol-3-kinases (PI3K)/Akt pathway that is modulated by IGF-1 and insulin. These hormones stimulate protein synthesis and promote muscle hypertrophy by interacting with their respective tyrosine kinase receptors to phosphorylate the insulin receptor substrate 1 (IRS-1). As a result, PI3K/Akt is activated and stimulates mTOR. The latter eventually phosphorylates the 70-kDa ribosomal S6 protein kinase (S6K) and 4E-binding protein 1 (4E-BP1) (12), ultimately promoting protein synthesis.

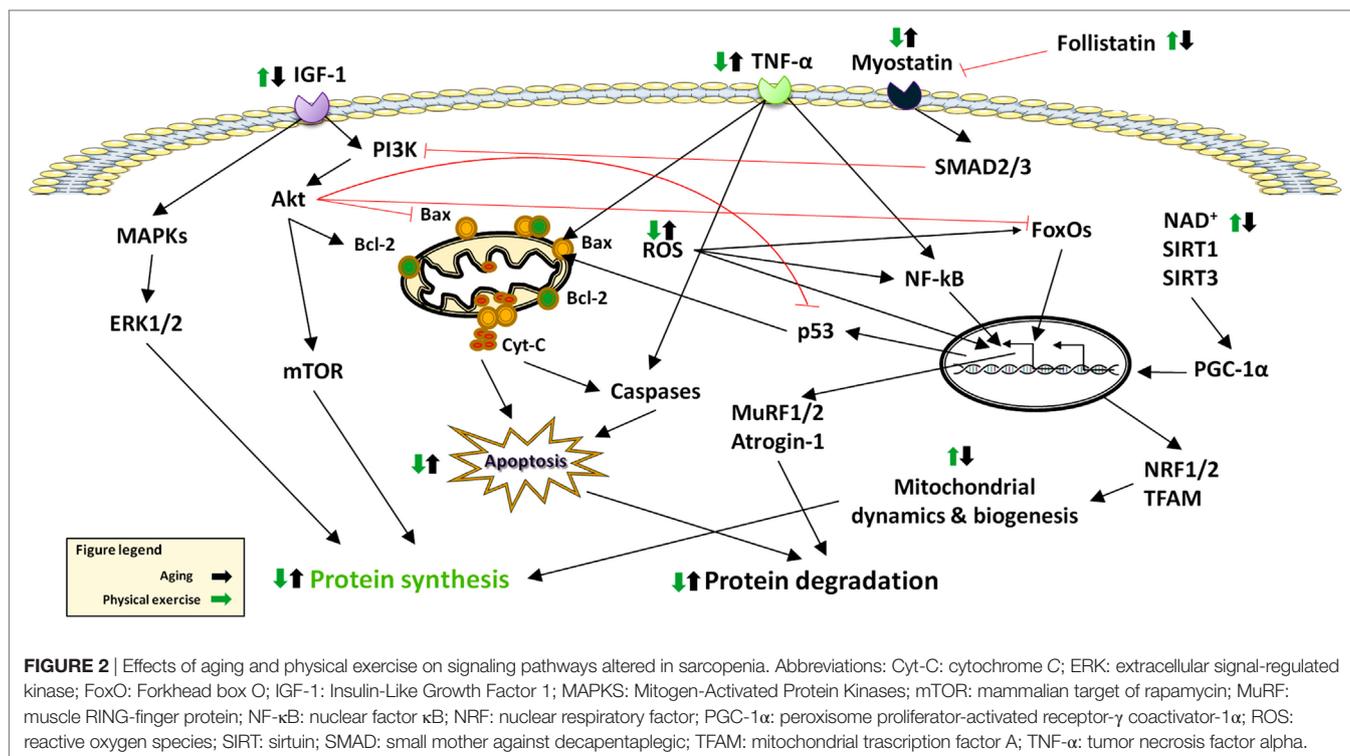
Akt/protein kinase B is a Ser/Thr kinase that has been shown to be a critical signaling component in the regulation of cell metabolism, growth, and survival (13).

### Forkhead Box O (FoxO) Transcription Factors

FoxO transcription factors consist of a large family of proteins identified by a protected DNA-binding domain referred to as FoxO (14). FoxO family members involved in skeletal muscle physiology include FoxO1, FoxO3, and FoxO4 (15). FoxOs are predominantly located in the nucleus where they regulate the expression of a number of downstream signaling proteins. However, when FoxOs are phosphorylated, mainly by Akt, they are rerouted into the cytosol and therefore become unable to transcribe genes involved in muscle atrophy (16). Recent studies have provided evidence that FoxO1 suppresses the efficiency of anabolic pathways in muscle *via* increased expression and reduced phosphorylation of the translational repressor protein 4E-BP1 and impaired signaling *via* reductions in mTOR and regulatory-associated protein of mammalian target of rapamycin (RAPTOR) levels (14). Furthermore, elevated levels of myonuclear levels of FoxO1 have been found in muscle samples from older persons compared with younger counterparts (17).

### Transforming Growth Factor Beta (TGF $\beta$ )

Muscle regeneration is primarily modulated by members of the TGF $\beta$  superfamily, which are known to suppress myogenic differentiation (18). In particular, myostatin is one of the main signaling molecules that regulate muscle growth. Myostatin is produced by skeletal myocytes and negatively regulates muscle growth (15). The effects of myostatin are mediated by the transcription factors small mother against decapentaplegic (SMAD) 2 and 3, which also interfere with IGF1–Akt signaling. Myostatin has been reported to upregulate the ubiquitin ligases atrogin1 and muscle RING-finger protein-1 (MuRF1) *via* FoxO transcription factors. Indeed, myostatin administration has been shown to block the



IGF1-PI3K-Akt pathway, thus activating FoxO1, allowing increased expression of atrogin-1. This connection between the two pathways is independent of nuclear factor κB (NF-κB) (19). In contrast, SMAD2/3 inhibition promotes muscle hypertrophy, which is partially dependent on mTOR signaling (20).

## NF-κB

NF-κB is a pleiotropic transcription factor involved in immune system modulation, inflammation, cell survival, and proliferation. NF-κB activity seems to directly regulate the expression of myogenic differentiation 1 protein (MyoD), a myogenic transcription factor, and likely other molecules, such as MuRF1, during atrophy. Reactive oxygen species and tumor necrosis factor alpha (TNF-α) both activate NF-κB (14). The binding of NF-κB to inhibitors of κB (IκB) is responsible for maintaining NF-κB in an inactive form in the cytosol. Seven isoforms of IκB exist in mammals (IκBα, IκBβ, IκBγ, IκBε, Bcl-3, p100, and p105), each possessing the ability to inhibit NF-κB. Upon certain stimuli, IκBα is phosphorylated by the IκB kinase in a step that targets IκBα for ubiquitination and subsequent proteolysis, thereby leaving NF-κB unbound. This process allows the unbound NF-κB to translocate to the nucleus where it can affect gene expression by binding NF-κB-target sequences located in the promoter region of specific genes (21).

## Mitogen-Activated Protein Kinases (MAPKs)

MAPKs are Ser/Thr kinases that transduce extracellular signals able to regulate a broad range of cellular processes. Indeed, in

eukaryotic cells, the coordination of multiple MAPK pathways control gene expression, cell division, metabolism, motility, survival, apoptosis, and differentiation (22). The MAPK protein family is composed of four distinct signaling modules in skeletal muscle: (1) extracellular signal-regulated kinase (ERK) 1/2, (2) p38 MAPK, (3) c-Jun N-terminal kinases (JNKs), and (4) ERK5 or big MAPK. MAPKs are activated by cytokines, growth factors, and cellular stressors (23) and are stimulated by phosphorylation at regulatory tyrosine and threonine residues by upstream MAPK kinases. MAPK phosphatases are instead responsible for MAPK deactivation through dephosphorylation.

## EFFECTS OF EXERCISE TRAINING ON MUSCLE PATHOPHYSIOLOGY

Sedentary lifestyle impacts muscle mass and strength as well as physical performance (24). Conversely, physical exercise, namely the body movements performed to maintain or improve components of physical fitness (25), is a powerful modulator of multiple processes involved in muscle hypertrophy and strengthening (13). Physical exercise is typically distinguished in endurance training (ET), which involves low-resistance work for protracted periods of time, and RT, characterized by more powerful movements of shorter duration (26). Both exercise regimens act on most signaling pathways involved in sarcopenia (27), including the IGF-1/Akt/mTOR axis (28–30), FoxOs (31, 32), NF-κB (33, 34), MAPKs (35, 36), mitochondrial quality control processes (37), and apoptosis (38–40). The following subsections summarize the specific effects of ET and RT on such pathways (**Figure 2**).

## Endurance Training

Aerobic exercise capacity decreases with advancing age, partly because of a decrease in the quantity and quality of muscle mitochondria (41, 42). On the other hand, ET improves maximal oxygen consumption ( $VO_{2max}$ ), mitochondrial density and activity, insulin sensitivity, and energy expenditure (43). Furthermore, ET reduces intramuscular fat accumulation and improves muscle function (44). The increase in citrate synthase (CS) activity in muscle fibers following ET supports the notion that this intervention improves mitochondrial mass (45). Numerous studies have investigated the effects of acute (13, 46–48) and chronic ET (49–63) on age-related muscle changes in both rodents and humans.

To investigate the acute effect of ET on skeletal muscle mitochondria in older persons, Bori et al. (48) studied the impact of a single bout of ET on the transcription of genes involved in mitochondrial biogenesis in sedentary versus physically active older adults. Their findings suggest that mitochondrial fission is impaired with age which could be involved in the age-associated decline in mitochondrial biogenesis-related gene expression in response to regular physical activity and exercise. These data also indicate that aging slightly affects the expression of mitochondrial biogenesis and quality control genes. Interestingly, old physically active participants showed similar levels of  $VO_{2max}$ , mitochondrial density, CS activity, and cytochrome *c* oxidase (COX) of young sedentary individuals, emphasizing the impact of regular physical activity on muscle health (48).

In contrast to acute ET, chronic ET appears to have considerably greater effects. Konopka et al. (63) examined the influence of 12 weeks of progressive ET on markers of mitochondrial content in old women. Compared with basal levels, ET significantly increased the content of the peroxisome proliferator-activated receptor- $\gamma$  coactivator-1 $\alpha$  (PGC-1 $\alpha$ ) protein along with CS,  $\beta$ -hydroxyacyl CoA dehydrogenase, succinate dehydrogenase, and COX4. In addition to this, the expression of the mitochondrial fusion factors mitofusin (Mfn) 1 and 2 and fission protein 1 (Fis1) was increased by ET (63). In keeping with these findings, 12 weeks of ET were able to stimulate mitochondrial biogenesis and improve mitochondrial networking and the efficiency of mitochondrial energy transfer in old rats (60). ET also increased the content of COX4 and dynamin-related protein 1 (Drp1), but not that of Mfn1. Finally, ATP synthase activity, an indicator of mitochondrial energy production, was increased by ET (60). Upregulation of PGC-1 $\alpha$  signaling is a major adaptation of skeletal muscle to ET (64). For instance, 12 weeks of ET were shown to increase PGC-1 $\alpha$  content by 2.3-fold in old rats (61). This adaptation was associated with significant increases in mitochondrial transcription factor A (TFAM), cytochrome *c*, and mtDNA contents. Noticeably, as a response to ET, increased levels of upstream signaling mediators modulating PGC-1 $\alpha$  activity, such as AMP-activated protein kinase (AMPK), p38MAPK, sirtuin 1 (SIRT1), and p-cAMP response element-binding protein (CREB) have been reported. These findings indicate that the age-associated decline in mitochondrial protein synthesis in skeletal muscle can be attenuated by ET (61). In this regard, 4 months of ET were found to increase the content of complexes III, IV, and V of the electron transport chain in muscles of older adults (62). Furthermore, a significant correlation was observed between TFAM and PGC-1 $\alpha$

expression levels after 4 months of exercise intervention. However, no changes in expression levels of nuclear respiratory factor (NRF) 1 and 2 were detected in responses to ET (62).

Another mechanism through which ET positively impacts muscle aging involves the inhibition of myonuclear apoptosis (7, 14, 65). In this regard, Song et al. (51) found that 12 weeks of ET reduced the extent of apoptotic DNA fragmentation in white gastrocnemius and soleus muscles of old rats, which was attributed to downregulation of mitochondrial apoptotic signaling. Similar findings were reported by Marzetti et al. (52) in old rats following 4 weeks of treadmill running. Notably, ET prevented the age-related elevation of TNF- $\alpha$ -related apoptotic signaling in the extensor digitorum longus muscle of old rats, which was associated with improved exercise capacity and muscle strength.

A potential role for ET in increasing the circulating levels of IGF-1 has also been suggested (38). For instance, 8 weeks of ET significantly increased fasting levels of IGF-1, especially in older men relative to women (49). There was also a significant correlation between changes in  $VO_{2max}$  and IGF-1 in men (49). In addition, basal levels of growth hormone (GH), IGF-1, and IGF binding protein 1 (IGFBP-1) were found to be higher in trained middle-aged men relative to sedentary controls (46). Furthermore, acute ET increased the activity of the GH/IGF-1 axis in middle-aged men (46). In support of the anabolic effect of ET, 3 h of bicycle exercise induced a sevenfold increase in plasma levels of follistatin as opposed to a smaller effect of one-legged knee extensor exercise (47). The increase in plasma follistatin after ET may be dependent on several factors, including the intensity and duration of exercise (47). In line with this, Sakamoto et al. (13) found that Akt activity significantly increased following acute submaximal and maximal intensity ET. Increases in Akt activity were accompanied by enhanced Akt Thr308 and Ser473 phosphorylation (13). The beneficial effects of ET on anabolic pathway may depend on the frequency of training. In support to this notion, Pasini et al. (59) investigated the effects of 8 weeks of ET and training frequency [i.e., 3 (EX3) or 5 days/week (EX5)] on anabolic pathways in skeletal muscle of old rats. Aging was associated with reduced protein levels of IRS-1 and p-mTOR in control rats relative to the young control group. In response to ET, EX3 resulted in reduced insulin receptor expression and increased IRS-1 levels compared with old sedentary rats. EX5 upregulated not only IRS-1 and COX activity but also p-mTOR expression (59).

## Resistance Training

RT is effective in preventing and treating sarcopenia due to its ability to promote net muscle protein anabolism, resulting in specific metabolic and morphological muscular adaptations (27, 66, 67).

The effects of acute and chronic RT on skeletal muscle in advanced age have been thoroughly investigated (10, 26, 31, 68–70). Nevertheless, the acute effects of RT are yet to be elucidated. Fry et al. (10) measured intracellular mediators of muscle protein synthesis (MPS) following an acute bout of RT in young and old persons. At baseline and 3, 6, and 24 h after RT, muscle biopsies were taken from the vastus lateralis. No changes were observed in phosphorylation of several key signaling proteins,

**TABLE 1** | American College of Sports Medicine/American Heart Association resistance and aerobic exercise recommendations for older adults.

	Frequency	Intensity	Duration	Type
Aerobic exercise	Moderate-intensity activities, accumulate at least 30 or up to 60 (for greater benefit) min/day in bouts of at least 10 min each to total 150–300 min/week, at least 20–30 min/day or more of vigorous-intensity activities to total 75–150 min/week	On a scale of 0–10 for RPE, 5–6 for moderate-intensity, and 7–8 for vigorous intensity	Moderate-intensity activities, accumulate at least 30 min/day in bouts of at least 10 min each or at least 20 min/day of continuous activity for vigorous-intensity activities	Any modality that does not impose excessive orthopedic stress; walking is the most common type of activity. Aquatic exercise and stationary cycle exercise may be advantageous for those with limited tolerance for weight-bearing activity
Resistance exercise	At least 2 days/week	Between moderate- (5–6) and vigorous- (7–8) intensity on a scale of 0–10	It may vary, depends on number of movements, sets, and repetitions	Progressive weight training program or weight-bearing calisthenics (8–10 exercises involving major muscle groups of 8–12 repetitions each), stair climbing, and other strengthening activities that use major muscle groups

RPE, Borg rating of perceived exertion.

mTOR, S6K1, 4E-BP1, and ERK1/2 following exercise in the older group. Increased MPS factors following exercise was found only in the younger group (10). On the other hand, Raue et al. (68) investigated the mRNA expression of several myogenic modulators at rest and 4 h after a single bout of RT in young and old women. Participants performed 3 sets of 10 repetitions of bilateral knee extensions at 70% of one-repetition maximum. RT led to upregulation of MyoD (2.0-fold) and MRF4 (1.4-fold) and downregulation of myostatin (2.2-fold) (68). The same group also investigated the effect of an acute bout of RT consisting of 3 sets of 10 knee extensions at 70% of one-repetition maximum on mRNA expression of ubiquitin proteasome-related genes involved in muscle atrophy in very old women (31). Muscle biopsies were taken from the vastus lateralis before RT and 4 h after. The authors demonstrated upregulation of atrogin-1 and MuRF-1 gene expression in response to RT. These data suggest that the regulation of ubiquitin proteasome-related genes involved in muscle atrophy is altered in very old women in response to RT (31).

In contrast to acute RT, Melov et al. (69) compared the expression profile of genes related to muscle strength, in healthy young and old men and women before and after a 6-month RT program. In response to RT, a significant improvement in strength was found in both age groups. Following RT, the transcriptional signature of aging was significantly reversed toward a youthful profile for most genes. The authors concluded that mitochondrial function and muscle weakness were favorably altered at the phenotypic and transcriptome level, following 6 months of RT (69). In support to the effects of RT on age-related changes in mitochondrial function, Luo et al. (70) investigated the signaling pathways that regulate autophagy and apoptosis in the gastrocnemius muscles of 18- to 20-month-old rats in response to 9 weeks of RT. Their findings demonstrated that RT prevented the loss of muscle mass which was accompanied by reduced microtubule-associated protein 1A/1B-light chain 3 (LC3)-II/LC3-I ratio, reduced p62 protein levels, and increased levels of autophagy regulatory proteins (Atgs), including Beclin 1, Atg5/12, Atg7, and the lysosomal enzyme cathepsin L. These improvements in autophagic signaling were associated with upregulation of total and phosphorylated AMPK and FoxO3A expression. Their results also showed that RT reduced

cytochrome c release in the cytosol and caspase-3 activation suggesting an inhibition of apoptosis. Moreover, RT upregulated the expression of IGF-1 and its receptors and downregulated the phosphorylation of Akt and mTOR. As a whole, these findings suggest an anti-apoptotic effect of chronic RT most likely *via* inhibition of mitochondria-mediated apoptosis in aged skeletal muscle (70).

The exact mechanisms by which RT stimulates protein synthesis in old muscles are not yet fully understood. However, it has been speculated that in response to RT, IGF-1 and its receptors, as well as the Akt/mTOR and Akt/FoxO3a signaling pathways may be modulated (70). In fact, in response to RT, IGF-1 activates PI3K, which leads to membrane translocation and subsequent phosphorylation of Akt by phosphoinositide-dependent kinase (PDK) 1 and 2. Once activated, Akt phosphorylates mTOR and glycogen synthase kinase 3 beta (GSK3B), which play an important role in protein synthesis, transcriptional and proliferative processes related to the hypertrophic response, and control of protein degradation (16). Other mechanisms that are involved in MPS are those regulating MAPK signaling. It has been shown that in response to RT, phosphorylation of ERK1/2 by MAPK is increased, and mTOR is activated (44). mTOR activation by the ERK pathway may occur through the phosphorylation of tuberous sclerosis complex 2 (TSC2) (16).

## CONCLUSION

Sarcopenia is a major corollary of aging. Most intracellular signaling pathways involved in muscle homeostasis are affected and could therefore be exploited as targets for the development of interventions aimed at preventing, delaying, or reversing sarcopenia. To date, physical exercise, especially if in combination with appropriate nutritional supplementation (71), is considered to be the only effective intervention to manage sarcopenia and prevent its adverse outcomes (72). As for the training modality, RT is more effective in increasing muscle mass and strength, whereas ET is superior for improving maximum aerobic capacity. Based on this evidence, older people should be recommended engaging in a balanced program of both endurance and strength exercises, performed on a regular schedule (73). **Table 1** summarizes

general guidelines for resistance and aerobic exercises for older adult based on the American College of Sports Medicine in conjunction with the American Heart Association recommendations (25). Future research aimed at addressing the simultaneous effects of endurance and resistance exercise along with drug and nutritional interventions would help clarify the pathogenesis of sarcopenia and its signaling pathways.

## AUTHOR CONTRIBUTIONS

MZ, together with EM, conceived the paper, conducted, supported, and interpreted results of literature review, and drafted

the manuscript. AP assisted with the conception of the paper, contributed to the discussion, and assisted with editing the manuscript. ZM assisted with the literature review and manuscript drafting. All authors critically revised the manuscript and approved the final version.

## FUNDING

This work was supported by Fondazione Roma (NCDs call for proposals 2013), Innovative Medicine Initiative-Joint Undertaking (IMI-JU #115621), and the nonprofit research foundation “Centro Studi Achille e Linda Lorenzon.”

## REFERENCES

- Landi F, Calvani R, Cesari M, Tosato M, Martone AM, Ortolani E, et al. Sarcopenia: an overview on current definitions, diagnosis and treatment. *Curr Protein Pept Sci* (2017). doi:10.2174/1389203718666170607113459
- Marzetti E, Calvani R, Lorenzi M, Marini F, D'Angelo E, Martone AM, et al. Serum levels of C-terminal agrin fragment (CAF) are associated with sarcopenia in older hip fractured patients. *Exp Gerontol* (2014) 60:79–82. doi:10.1016/j.exger.2014.10.003
- Altun M, Grönholdt-Klein M, Wang L, Ulfhake B. Cellular degradation machineries in age-related loss of muscle mass (Sarcopenia). In: Nagata T, editor. *Senescence*. Rijeka, HR: InTech (2012). p. 271–86
- Pillard F, Laoudj-Chenivesse D, Carnac G, Mercier J, Rami J, Riviere D, et al. Physical activity and sarcopenia. *Clin Geriatr Med* (2011) 27:449–70. doi:10.1016/j.cger.2011.03.009
- Marzetti E, Calvani R, Tosato M, Cesari M, Di Bari M, Cherubini A, et al. Physical activity and exercise as countermeasures to physical frailty and sarcopenia. *Aging Clin Exp Res* (2017) 29:35–42. doi:10.1007/s40520-016-0705-4
- Law TD, Clark LA, Clark BC. Resistance exercise to prevent and manage sarcopenia and dynapenia. *Annu Rev Gerontol Geriatr* (2016) 36:205–28. doi:10.1891/0198-8794.36.205
- Ziaaldini MM, Koltai E, Csense Z, Goto S, Boldogh I, Taylor AW, et al. Exercise training increases anabolic and attenuates catabolic and apoptotic processes in aged skeletal muscle of male rats. *Exp Gerontol* (2015) 67:9–14. doi:10.1016/j.exger.2015.04.008
- Capelli C, Rittveger J, Brusaghini P, Calabria E, Tam E. Maximal aerobic power and anaerobic capacity in cycling across the age spectrum in male master athletes. *Eur J Appl Physiol* (2016) 116:1395–410. doi:10.1007/s00421-016-3396-9
- Tromm CB, Pozzi BG, Paganini CS, Marques SO, Pedrosa GS, Souza PS, et al. The role of continuous versus fractionated physical training on muscle oxidative stress parameters and calcium-handling proteins in aged rats. *Aging Clin Exp Res* (2016) 28:833–41. doi:10.1007/s40520-015-0501-6
- Fry CS, Drummond MJ, Glynn EL, Dickinson JM, Gundermann DM, Timmerman KL, et al. Aging impairs contraction-induced human skeletal muscle mTORC1 signaling and protein synthesis. *Skelet Muscle* (2011) 1:11. doi:10.1186/2044-5040-1-11
- Ziaaldini MM, Hosseini SR, Fathi M. Mitochondrial adaptations in aged skeletal muscle: effect of exercise training. *Physiol Res* (2017) 66:1–14.
- Romanick M, Thompson LV, Brown-Borg HM. Murine models of atrophy, cachexia, and sarcopenia in skeletal muscle. *Biochim Biophys Acta* (2013) 1832:1410–20. doi:10.1016/j.bbdis.2013.03.011
- Sakamoto K, Arnolds DE, Ekberg I, Thorell A, Goodyear LJ. Exercise regulates Akt and glycogen synthase kinase-3 activities in human skeletal muscle. *Biochem Biophys Res Commun* (2004) 319:419–25. doi:10.1016/j.bbrc.2004.05.020
- Meng SJ, Yu LJ. Oxidative stress, molecular inflammation and sarcopenia. *Int J Mol Sci* (2010) 11:1509–26. doi:10.3390/ijms11041509
- Schiaffino S, Dyar KA, Ciciliot S, Blaauw B, Sandri M. Mechanisms regulating skeletal muscle growth and atrophy. *FEBS J* (2013) 280:4294–314. doi:10.1111/febs.12253
- Fernandes T, Soci ÚPR, Melo SFS, Alves CR, Oliveira EM. Signaling pathways that mediate skeletal muscle hypertrophy: effects of exercise training. In: Cseri J, editor. *Skeletal Muscle – From Myogenesis to Clinical Relations*. Rijeka, HR: InTech (2012). p. 189–218.
- Giresi PG, Stevenson EJ, Theilhaber J, Koncarevic A, Parkington J, Fielding RA, et al. Identification of a molecular signature of sarcopenia. *Physiol Genomics* (2005) 21:253–63. doi:10.1152/physiolgenomics.00249.2004
- Barberi L, Scicchitano BM, De Rossi M, Bigot A, Duguez S, Wielgosik A, et al. Age-dependent alteration in muscle regeneration: the critical role of tissue niche. *Biogerontology* (2013) 14:273–92. doi:10.1007/s10522-013-9429-4
- Sandri M, Barberi L, Bijlsma AY, Blaauw B, Dyar KA, Milan G, et al. Signalling pathways regulating muscle mass in ageing skeletal muscle: the role of the IGF1-Akt-mTOR-FoxO pathway. *Biogerontology* (2013) 14:303–23. doi:10.1007/s10522-013-9432-9
- Sakuma K, Yamaguchi A. Novel intriguing strategies attenuating sarcopenia. *J Aging Res* (2012) 2012:251217. doi:10.1155/2012/251217
- LaGuire TC. The effects of aging on muscle loss and tissue-specific levels of NF-κB and SIRT6 proteins in rats. *Adv Aging Res* (2013) 02:1–9. doi:10.4236/aar.2013.21001
- Cargnello M, Roux PP. Activation and function of the MAPKs and their substrates, the MAPK-activated protein kinases. *Microbiol Mol Biol Rev* (2011) 75:50–83. doi:10.1128/MMBR.00031-10
- Kramer HF, Goodyear LJ. Exercise, MAPK, and NF-kappaB signaling in skeletal muscle. *J Appl Physiol* (1985) (2007) 103:388–95. doi:10.1152/jappphysiol.00085.2007
- Landi F, Marzetti E, Martone AM, Bernabei R, Onder G. Exercise as a remedy for sarcopenia. *Curr Opin Clin Nutr Metab Care* (2014) 17:25–31. doi:10.1097/MCO.000000000000018
- American College of Sports Medicine, Chodzko-Zajko WJ, Proctor DN, Fiaatarone Singh MA, Minson CT, Nigg CR, et al. American College of Sports Medicine position stand. Exercise and physical activity for older adults. *Med Sci Sports Exerc* (2009) 41:1510–30. doi:10.1249/MSS.0b013e3181a0c95c
- Ruas JL, White JP, Rao RR, Kleiner S, Brannan KT, Harrison BC, et al. A PGC-1alpha isoform induced by resistance training regulates skeletal muscle hypertrophy. *Cell* (2012) 151:1319–31. doi:10.1016/j.cell.2012.10.050
- Koopman R, van Loon LJ. Aging, exercise, and muscle protein metabolism. *J Appl Physiol* (1985) (2009) 106:2040–8. doi:10.1152/jappphysiol.91551.2008
- Arnarson A, Gudny Geirsdottir O, Ramel A, Jonsson PV, Thorsdottir I. Insulin-like growth factor-1 and resistance exercise in community dwelling old adults. *J Nutr Health Aging* (2015) 19:856–60. doi:10.1007/s12603-015-0547-3
- Tsai CL, Wang CH, Pan CY, Chen FC. The effects of long-term resistance exercise on the relationship between neurocognitive performance and GH, IGF-1, and homocysteine levels in the elderly. *Front Behav Neurosci* (2015) 9:23. doi:10.3389/fnbeh.2015.00023
- Ahtiainen JP, Hulmi JJ, Lehti M, Kraemer WJ, Nyman K, Selanne H, et al. Effects of resistance training on expression of IGF-I splice variants in younger and older men. *Eur J Sport Sci* (2016) 16:1055–63. doi:10.1080/17461391.2016.1185164
- Raue U, Slivka D, Jemiolo B, Hollon C, Trappe S. Proteolytic gene expression differs at rest and after resistance exercise between young and old

- women. *J Gerontol A Biol Sci Med Sci* (2007) 62:1407–12. doi:10.1093/gerona/62.12.1407
32. Vechetti-Junior IJ, Bertaglia RS, Fernandez GJ, de Paula TG, de Souza RW, Moraes LN, et al. Aerobic exercise recovers disuse-induced atrophy through the stimulus of the LRP130/PGC-1 $\alpha$  complex in aged rats. *J Gerontol A Biol Sci Med Sci* (2016) 71:601–9. doi:10.1093/gerona/glv064
  33. Rivas DA, Morris EP, Haran PH, Pasha EP, Morais Mda S, Dolnikowski GG, et al. Increased ceramide content and NF $\kappa$ B signaling may contribute to the attenuation of anabolic signaling after resistance exercise in aged males. *J Appl Physiol* (1985) (2012) 113:1727–36. doi:10.1152/jappphysiol.00412.2012
  34. Masuki S, Nishida K, Hashimoto S, Morikawa M, Takasugi S, Nagata M, et al. Effects of milk product intake on thigh muscle strength and NF $\kappa$ B gene methylation during home-based interval walking training in older women: a randomized, controlled pilot study. *PLoS One* (2017) 12:e0176757. doi:10.1371/journal.pone.0176757
  35. Shen H, Tong L, Balazs R, Cotman CW. Physical activity elicits sustained activation of the cyclic AMP response element-binding protein and mitogen-activated protein kinase in the rat hippocampus. *Neuroscience* (2001) 107:219–29. doi:10.1016/s0306-4522(01)00315-3
  36. Williamson D, Gallagher P, Harber M, Hollon C, Trappe S. Mitogen-activated protein kinase (MAPK) pathway activation: effects of age and acute exercise on human skeletal muscle. *J Physiol* (2003) 547:977–87. doi:10.1113/jphysiol.2002.036673
  37. Kim Y, Triolo M, Hood DA. Impact of aging and exercise on mitochondrial quality control in skeletal muscle. *Oxid Med Cell Longev* (2017) 2017:3165396. doi:10.1155/2017/3165396
  38. Li M, Li C, Parkhouse WS. Age-related differences in the des IGF-I-mediated activation of Akt-1 and p70 S6K in mouse skeletal muscle. *Mech Ageing Dev* (2003) 124:771–8. doi:10.1016/s0047-6374(03)00124-6
  39. Fan J, Yang X, Li J, Shu Z, Dai J, Liu X, et al. Spermidine coupled with exercise rescues skeletal muscle atrophy from D-gal-induced aging rats through enhanced autophagy and reduced apoptosis via AMPK-FOXO3a signal pathway. *Oncotarget* (2017) 8:17475–90. doi:10.18632/oncotarget.15728
  40. Mejias-Pena Y, Estebanez B, Rodriguez-Miguel P, Fernandez-Gonzalo R, Almar M, de Paz JA, et al. Impact of resistance training on the autophagy-inflammation-apoptosis crosstalk in elderly subjects. *Aging (Albany NY)* (2017) 9:408–18. doi:10.18632/aging.101167
  41. Forbes SC, Little JP, Candow DG. Exercise and nutritional interventions for improving aging muscle health. *Endocrine* (2012) 42:29–38. doi:10.1007/s12020-012-9676-1
  42. Johnson ML, Robinson MM, Nair KS. Skeletal muscle aging and the mitochondrion. *Trends Endocrinol Metab* (2013) 24:247–56. doi:10.1016/j.tem.2012.12.003
  43. Volpi E, Nazemi R, Fujita S. Muscle tissue changes with aging. *Curr Opin Clin Nutr Metab Care* (2004) 7:405–10. doi:10.1097/01.mco.0000134362.76653.b2
  44. Rom O, Kaisari S, Aizenbud D, Reznick AZ. Lifestyle and sarcopenia—etiology, prevention, and treatment. *Rambam Maimonides Med J* (2012) 3:e0024. doi:10.5041/RMMJ.10091
  45. Burton LA, Sumukadas D. Optimal management of sarcopenia. *Clin Interv Aging* (2010) 5:217–28. doi:10.2147/CIA.S11473
  46. Manetta J, Brun JF, Maimoun L, Callis A, Prefaut C, Mercier J. Effect of training on the GH/IGF-I axis during exercise in middle-aged men: relationship to glucose homeostasis. *Am J Physiol Endocrinol Metab* (2002) 283:E929–36. doi:10.1152/ajpendo.00539.2001
  47. Hansen J, Brandt C, Nielsen AR, Hojman P, Whitham M, Febbraio MA, et al. Exercise induces a marked increase in plasma follistatin: evidence that follistatin is a contraction-induced hepatokine. *Endocrinology* (2011) 152:164–71. doi:10.1210/en.2010-0868
  48. Bori Z, Zhao Z, Koltai E, Fatouros IG, Jamurtas AZ, Douroudos II, et al. The effects of aging, physical training, and a single bout of exercise on mitochondrial protein expression in human skeletal muscle. *Exp Gerontol* (2012) 47:417–24. doi:10.1016/j.exger.2012.03.004
  49. Poehlman ET, Rosen CJ, Copeland KC. The influence of endurance training on insulin-like growth factor-1 in older individuals. *Metabolism* (1994) 43:1401–5. doi:10.1016/0026-0495(94)90035-3
  50. Kwak HB, Song W, Lawler JM. Exercise training attenuates age-induced elevation in Bax/Bcl-2 ratio, apoptosis, and remodeling in the rat heart. *FASEB J* (2006) 20:791–3. doi:10.1096/fj.05-5116fje
  51. Song W, Kwak HB, Lawler JM. Exercise training attenuates age-induced changes in apoptotic signaling in rat skeletal muscle. *Antioxid Redox Signal* (2006) 8:517–28. doi:10.1089/ars.2006.8.517
  52. Marzetti E, Groban L, Wohlgemuth SE, Lees HA, Lin M, Jobe H, et al. Effects of short-term GH supplementation and treadmill exercise training on physical performance and skeletal muscle apoptosis in old rats. *Am J Physiol Regul Integr Comp Physiol* (2008) 294:R558–67. doi:10.1152/ajpregu.00620.2007
  53. Lenk K, Schur R, Linke A, Erbs S, Matsumoto Y, Adams V, et al. Impact of exercise training on myostatin expression in the myocardium and skeletal muscle in a chronic heart failure model. *Eur J Heart Fail* (2009) 11:342–8. doi:10.1093/eurjhf/hfp020
  54. Konopka AR, Douglass MD, Kaminsky LA, Jemiolo B, Trappe TA, Trappe S, et al. Molecular adaptations to aerobic exercise training in skeletal muscle of older women. *J Gerontol A Biol Sci Med Sci* (2010) 65:1201–7. doi:10.1093/gerona/glq109
  55. Leiter JR, Peeler J, Anderson JE. Exercise-induced muscle growth is muscle-specific and age-dependent. *Muscle Nerve* (2011) 43:828–38. doi:10.1002/mus.21965
  56. Wroblewski AP, Amati F, Smiley MA, Goodpaster B, Wright V. Chronic exercise preserves lean muscle mass in masters athletes. *Phys Sportsmed* (2011) 39:172–8. doi:10.3810/psm.2011.09.1933
  57. Gielen S, Sandri M, Kozarez I, Kratzsch J, Teupser D, Thiery J, et al. Exercise training attenuates MuRF-1 expression in the skeletal muscle of patients with chronic heart failure independent of age: the randomized Leipzig exercise intervention in chronic heart failure and aging catabolism study. *Circulation* (2012) 125:2716–27. doi:10.1161/CIRCULATIONAHA.111.047381
  58. Harber MP, Konopka AR, Udem MK, Hinkley JM, Minchev K, Kaminsky LA, et al. Aerobic exercise training induces skeletal muscle hypertrophy and age-dependent adaptations in myofiber function in young and older men. *J Appl Physiol* (1985) (2012) 113:1495–504. doi:10.1152/jappphysiol.00786.2012
  59. Pasini E, Le Douairon Lahaye S, Flati V, Assanelli D, Corsetti G, Specia S, et al. Effects of treadmill exercise and training frequency on anabolic signaling pathways in the skeletal muscle of aged rats. *Exp Gerontol* (2012) 47:23–8. doi:10.1016/j.exger.2011.10.003
  60. Bo H, Jiang N, Ji LL, Zhang Y. Mitochondrial redox metabolism in aging: effect of exercise interventions. *J Sport Health Sci* (2013) 2:67–74. doi:10.1016/j.jshs.2013.03.006
  61. Kang C, Chung E, Diffeo G, Ji LL. Exercise training attenuates aging-associated mitochondrial dysfunction in rat skeletal muscle: role of PGC-1 $\alpha$ . *Exp Gerontol* (2013) 48:1343–50. doi:10.1016/j.exger.2013.08.004
  62. Broskey NT, Greggio C, Boss A, Boutant M, Dwyer A, Schlueter L, et al. Skeletal muscle mitochondria in the elderly: effects of physical fitness and exercise training. *J Clin Endocrinol Metab* (2014) 99:1852–61. doi:10.1210/jc.2013-3983
  63. Konopka AR, Suer MK, Wolff CA, Harber MP. Markers of human skeletal muscle mitochondrial biogenesis and quality control: effects of age and aerobic exercise training. *J Gerontol A Biol Sci Med Sci* (2014) 69:371–8. doi:10.1093/gerona/glt107
  64. Sakuma K, Yamaguchi A. Sarcopenia and age-related endocrine function. *Int J Endocrinol* (2012) 2012:127362. doi:10.1155/2012/127362
  65. Arthur ST, Cooley ID. The effect of physiological stimuli on sarcopenia; impact of Notch and Wnt signaling on impaired aged skeletal muscle repair. *Int J Biol Sci* (2012) 8:731–60. doi:10.7150/ijbs.4262
  66. Gumucio JR, Mendias CL. Atrogin-1, MuRF-1, and sarcopenia. *Endocrine* (2013) 43:12–21. doi:10.1007/s12020-012-9751-7
  67. Phillips BE, Williams JP, Gustafsson T, Bouchard C, Rankinen T, Knudsen S, et al. Molecular networks of human muscle adaptation to exercise and age. *PLoS Genet* (2013) 9:e1003389. doi:10.1371/journal.pgen.1003389
  68. Raue U, Slivka D, Jemiolo B, Hollon C, Trappe S. Myogenic gene expression at rest and after a bout of resistance exercise in young (18–30 yr) and old (80–89 yr) women. *J Appl Physiol* (1985) (2006) 101:53–9. doi:10.1152/jappphysiol.01616.2005
  69. Melov S, Tarnopolsky MA, Beckman K, Felkey K, Hubbard A. Resistance exercise reverses aging in human skeletal muscle. *PLoS One* (2007) 2:e465. doi:10.1371/journal.pone.0000465
  70. Luo L, Lu AM, Wang Y, Hong A, Chen Y, Hu J, et al. Chronic resistance training activates autophagy and reduces apoptosis of muscle cells by modulating IGF-1 and its receptors, Akt/mTOR and Akt/FOXO3a signaling in aged rats. *Exp Gerontol* (2013) 48:427–36. doi:10.1016/j.exger.2013.02.009

71. Landi F, Calvani R, Tosato M, Martone AM, Ortolani E, Saveria G, et al. Protein intake and muscle health in old age: from biological plausibility to clinical evidence. *Nutrients* (2016) 8:E295. doi:10.3390/nu8050295
72. Morley JE, Argiles JM, Evans WJ, Bhasin S, Cella D, Deutz NE, et al. Nutritional recommendations for the management of sarcopenia. *J Am Med Dir Assoc* (2010) 11:391–6. doi:10.1016/j.jamda.2010.04.014
73. Iolascon G, Di Pietro G, Gimigliano F, Mauro GL, Moretti A, Giamattei MT, et al. Physical exercise and sarcopenia in older people: position paper of the Italian Society of Orthopaedics and Medicine (OrtoMed). *Clin Cases Miner Bone Metab* (2014) 11:215–21. doi:10.11138/ccmbm/2014.11.3.215

**Conflict of Interest Statement:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2017 Ziaaldini, Marzetti, Picca and Murlasits. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) or licensor are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.