



## Prealbumin as a Predictor of Prognosis in Patients With Coronavirus Disease 2019

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**Background:** The predictive value of prealbumin for the prognosis of coronavirus disease 2019 (COVID-19) has not been extensively investigated.

**Methods:** A total of 1,115 patients with laboratory-confirmed COVID-19 were enrolled at Tongji hospital from February to April 2020 and classified into fatal (n = 129) and recovered (n = 986) groups according to the patient's outcome. Prealbumin and other routine laboratory indicators were measured simultaneously.

**Results:** The level of prealbumin on admission was significantly lower in fatal patients than in recovered patients. For predicting the prognosis of COVID-19, the performance of prealbumin was better than most routine laboratory indicators, such as albumin, lymphocyte count, neutrophil count, hypersensitive C-reactive protein, d-dimer, lactate dehydrogenase, creatinine, and hypersensitive cardiac troponin I. When a threshold of 126 mg/L was used to discriminate between fatal and recovered patients, the sensitivity and specificity of prealbumin were, respectively, 78.29 and 90.06%. Furthermore, a model based on the combination of nine indexes showed an improved performance in predicting the death of patients with COVID-19. Using a cut-off value of 0.19, the prediction model was able to distinguish between fatal and recovered individuals with a sensitivity of 86.82% and a specificity of 90.37%.

**Conclusions:** A lower level of prealbumin on admission may indicate a worse outcome of COVID-19. Immune and nutritional status may be vital factors for predicting disease progression in the early stage of COVID-19.

Keywords: coronavirus disease 2019, severe acute respiratory syndrome coronavirus 2, prealbumin, routine laboratory tests, prognosis, immune status, fatal patients, recovered patients

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### **INTRODUCTION**

Coronavirus disease 2019 (COVID-19) as a rampant disease caused by the emerging infection of severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2), the outbreak of which has initiated an extreme health concern (1, 2). Many patients might progress to acute respiratory disease or other complications in a short period of time (3, 4). Since no effective vaccine or anti-viral treatment is currently available, this epidemic is difficult to manage and control. As of 7 May 2020, the number of patients infected with SARS-CoV-2 has exceeded 3.5 million globally, and there has been more

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than 250,000 reported cases of COVID-19-related deaths worldwide (5). Therefore, it is urgent and desirable to establish a model which can be used to predict the progression of disease and help clinicians to better choose a therapeutic strategy.

Up to now, many studies have reported that the risk factors for death in patients with COVID-19 are attributed to advanced age and co-morbidities including hypertension, myocardial injury, liver damage, and kidney failure (6-10). Some indicators such as creatinine (CR), lactate dehydrogenase (LDH), aspartate aminotransferase (AST), and hypersensitive cardiac troponin I (hs-cTnI) have been found to be helpful to assess the severity of the disease and predict the prognosis of COVID-19 (9, 11, 12). In addition, some studies discovered and elaborated the potential value of coagulation indicators represented by prothrombin time (PT) and d-dimer (DD) in predicting the prognosis and outcome of patients with COVID-19 (13-15). Moreover, some recent studies focused on the body's immune status, including inflammatory responses and the number and phenotype of lymphocytes, and found that some inflammatory indicators or cytokines including hypersensitive C-reactive protein (hsCRP), procalcitonin (PCT), interleukin-6, interleukin-2 receptor, interferon-gamma, and the number of lymphocytes also contributed to the outcome of the disease (16-19).

Additionally, some indicators, such as albumin (ALB) and prealbumin (PAB), can partially reflect the nutritional and immune status of the host (20-23). However, the potential value of them for the prognosis of COVID-19 has not been fully explored. Theoretically, in view of the decrease in the number of lymphocytes and their subsets in the early stage of the disease (24, 25) and the poor immune function potentially caused by various complications (26, 27), ALB and PAB are potential and feasible predictors for the prognosis of COVID-19. In this study, we did a comprehensive evaluation of various laboratory indicators in fatal and recovered patients with confirmed COVID-19 on admission. We also compared the predictive value of PAB and other routine laboratory makers for the prognosis of COVID-19. It is hoped that the information obtained in this study will offer a better understanding on the disease progression that occurs after SARS-CoV-2 infection, and establish a basis to optimize the current treatment.

### **METHODS**

### **Study Design and Participants**

The current study was conducted at Tongji hospital (the largest hospital in the central region of China) in Wuhan, China. Consecutive hospitalized patients with confirmed COVID-19 were enrolled between February and April 2020. COVID-19 was diagnosed if patients met the following criteria: (1) having typical clinical symptoms, (2) having typical imaging findings, and (3) positive for SARS-CoV-2 real-time reverse transcription-polymerase chain reaction. The patients who died during hospitalization were defined as the fatal group, and those who recovered and were finally discharged after hospitalization were defined as the recovered group. All recovered patients with COVID-19 met the following criteria: having completely

resolved symptoms and signs, having significant improvement in pulmonary and extrapulmonary organ dysfunction, no longer need supportive care, and with confirmed viral clearance by repeated tests for SARS-CoV-2 before hospital discharge. This study was reviewed and approved by the ethical committee of Tongji Hospital, Tongji Medical College, Huazhong University of Science and Technology, Wuhan, China (TJ-C20200128).

## Real Time Reverse Transcription-Polymerase Chain Reaction

The clinical samples, including throat and nasal swab obtained from patients at admission or during the hospital stay, were maintained in a viral-transport medium. SARS-CoV-2 was confirmed by using TaqMan One-Step reverse transcription-polymerase chain reaction (RT-PCR) Kits from Shanghai Huirui Biotechnology Co., Ltd and Shanghai BioGerm Medical Biotechnology Co., Ltd. Briefly, RNA was extracted from clinical samples. 5  $\mu L$  of RNA was used for real-time RT-PCR, which targeted the ORF1ab and N gene. Real-time RT-PCR was performed using the following conditions: 50°C for 15 min and 95°C for 5 min, 45 cycles of amplification at 95°C for 10 s and 55°C for 45 s. The positive SARS-CoV-2 real time RT-PCR result was defined if both ORF1ab and N cycle thresholds were <35.

### **Laboratory Tests**

The measurements of white blood cell count (WBC#), neutrophil count (NEU#), lymphocyte count (LYM#), and platelet count (PLT#) were performed using XN-9000 Sysmex (Sysmex Co., Kobe, Japan). The measurements of total protein (TP), PAB, ALB, globulin (GLB), CR, AST, LDH, hsCRP, PCT, and hs-cTnI were performed using ROCHE COBAS 8000 (Mannheim, Germany). PT and DD were detected using STA-R coagulation analyzers (Diagnostic Stago, France) according to the manufacturer's instructions.

### **Statistical Analysis**

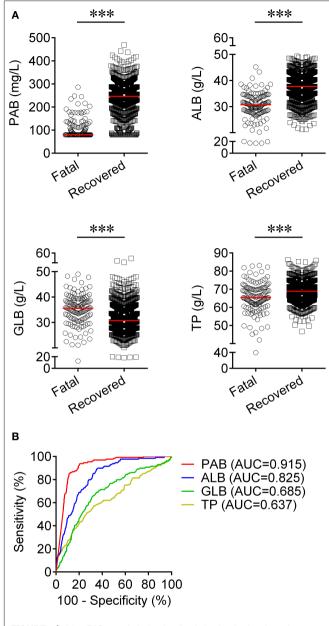
Continuous variables were expressed as mean ± standard deviation (SD) and categorical variables were reported as numbers and percentages (%). The comparison between continuous variables was performed using the Wilcoxon test or Mann-Whitney U test. The chi-square test was used for comparison of categorical data. The area under the curves (AUCs) were compared using the z statistic with the procedure of (28). A two-sided P-value below 0.05 was considered to be statistically significant. A prediction model for predicting the outcome of death was established by using a multivariate logistic regression method. All variables with statistical significance were taken as candidates for multivariable logistic regression analyses, and the regression equation (prediction model) was obtained. The regression coefficients of the prediction model were regarded as the weights for the respective variables and a score for each patient was calculated. Receiver operating characteristic (ROC) analysis was performed on these scores to assess the ability for distinguishing between fatal and recovered COVID-19 patients. AUC, sensitivity, specificity, positive predictive value (PPV), negative predictive value (NPV), positive likelihood ratio (PLR), and negative likelihood ratio (NLR), together with their 95%

**TABLE 1** | Demographic and clinical characteristics of study participants.

Variables	Fatal (n = 129)	Recovered (n = 986)	<b>P</b> *	
Sex, male, %	87 (67.44%)	476 (48.28%)	<0.001	
Age, years				
${\sf Mean} \pm {\sf SD}$	69.98±12.05	58.64±15.17	< 0.001	
<50	4 (3.10%)	251 (25.46%)	< 0.001	
50–59	16 (12.40%)	208 (21.1%)	0.021	
60–69	40 (31.01%)	300 (30.43%)	0.893	
70–79	40 (31.01%)	149 (15.11%)	< 0.001	
>79	29 (22.48%)	78 (7.91%)	< 0.001	
Symptoms on admission				
Cough	66 (51.16%)	576 (58.42%)	0.117	
Fever	80 (62.02%)	648 (65.72%)	0.406	
Shortness of breath	45 (34.88%)	126 (12.78%)	< 0.001	
Chest tightness	25 (19.38%)	198 (20.08%)	0.852	
Diarrhea	8 (6.20%)	108 (10.95%)	0.096	
Headache	2 (1.55%)	27 (2.74%)	0.425	
Nausea and vomiting	7 (5.43%)	36 (3.65%)	0.325	
Muscle ache	7 (5.43%)	54 (5.48%)	0.981	
Pharyngalgia	3 (2.33%)	45 (4.56%)	0.24	
Underlying condition or ill	ness			
Diabetes mellitus	25 (19.38%)	72 (7.30%)	< 0.001	
Hypertension	56 (43.41%)	252 (25.56%)	< 0.001	
Chronic obstructive pulmonary disease	5 (3.88%)	14 (1.42%)	0.043	
Cardiovascular disease	23 (17.83%)	95 (9.63%)	0.004	
Chronic kidney disease	6 (4.65%)	32 (3.25%)	0.408	
Chronic liver disease	3 (2.33%)	41 (4.16%)	0.315	
Hematological malignancy	2 (1.55%)	2 (0.20%)	0.016	
Solid tumor	6 (4.65%)	50 (5.07%)	0.837	
Organ transplantation	1 (0.78%)	5 (0.51%)	0.696	
Days from admission to d	eath			
${\sf Mean} \pm {\sf SD}$	$16.72 \pm 11.90$	N/A	N/A	
<3	7 (5.43%)	N/A	N/A	
3–7	20 (15.50%)	N/A	N/A	
8–14	43 (33.33%)	N/A	N/A	
15–30	41 (31.78%)	N/A	N/A	
>30	18 (13.95%)	N/A	N/A	
Days from admission to d				
Mean ± SD	N/A	21.66 ± 12.01	N/A	
<3	N/A	3 (0.30%)	N/A	
3–7	N/A	98 (9.94%)	N/A	
8–14	N/A	241 (24.44%)	N/A	
15–30	N/A	404 (40.97%)	N/A	
> 30	N/A	240 (24.34%)	N/A	

N/A, not applicable. \*Comparisons were performed between fatal and recovered groups using chi-square test or Mann-Whitney U test. Data were presented as means  $\pm$  SD or numbers (percentages).

confidence intervals (CI), were calculated. Data were analyzed by using SPSS version 25.0 (SPSS, Chicago, IL, USA), GraphPad Prism version 6 (GraphPad Software, San Diego, CA, USA), and MedCalc version 11.6 (Medcalc, Mariakerke, Belgium).



**FIGURE 1** | Using PAB on admission for discriminating fatal patients from recovered patients. **(A)** Scatter plots showing the concentrations of PAB, ALB, GLB, and TP in fatal (n=129) and recovered (n=986) patients. Horizontal lines indicate the median. \*\*\*P<0.001 (Mann-Whitney U test). **(B)** ROC analysis showing the performance of PAB, ALB, GLB, and TP in distinguishing fatal patients from recovered patients. PAB, prealbumin; ALB, albumin; GLB, globulin; TP, total protein; ROC, receiver operating characteristic curve; AUC, area under the curve.

## **RESULTS**

## Demographic and Clinical Characteristics of Study Participants

Our study enrolled 129 patients who died during hospitalization and 986 recovered patients (**Table 1**). There were 552 women (49.51%) and 563 men (50.49%) in this cohort, with ages

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**TABLE 2** | The performance of various methods for distinguishing between fatal and recovered patients.

Variables	Cutoff value	AUC (95% CI)	Sensitivity (95% CI)	Specificity (95% CI)	PPV (95% CI)	NPV (95% CI)	PLR (95% CI)	NLR (95% CI)
PAB (mg/L)	126	0.915 (0.894–0.937)	78.29% (71.18%—85.41%)	90.06% (88.19%—91.93%)	50.75% (43.81%-57.70%)	96.94% (95.83%-98.06%)	7.88 (6.39–9.71)	0.24 (0.17–0.33)
ALB (g/L)	29.7	0.825† (0.792–0.859)	45.74% (37.14%-54.33%)	90.37% (88.52%-92.21%)	38.31% (30.63%-45.99%)	92.72% (91.07%-94.36%)	4.75 (3.63-6.21)	0.60 (0.51-0.70)
GLB (g/L)	39.4	0.685† (0.636–0.735)	22.48% (15.28%-29.68%)	90.26% (88.41%-92.11%)	23.20% (15.80%-30.60%)	89.90% (88.02%-91.78%)	2.31 (1.59–3.35)	0.86 (0.78-0.94)
TP (g/L)	61.4	0.637 <sup>†</sup> (0.582–0.693)	24.81% (17.35%-32.26%)	90.47% (88.63%-92.30%)	25.40% (17.80%-33.00%)	90.19% (88.34%-92.05%)	2.60 (1.82-3.72)	0.83 (0.75-0.92)
WBC# (×10 <sup>9</sup> /L)	9.83	0.751† (0.698–0.804)	45.74% (37.14%-54.33%)	90.77% (88.96%-92.58%)	39.33% (31.52%-47.15%)	92.75% (91.11%-94.38%)	4.96 (3.78-6.50)	0.60 (0.51-0.70)
LYM# (×10 <sup>9</sup> /L)	0.6	0.842 <sup>†</sup> (0.807–0.877)	46.51% (37.90%-55.12%)	90.06% (88.19%-91.93%)	37.97% (30.41%-45.54%)	92.79% (91.15%-94.43%)	4.68 (3.59-6.09)	0.59 (0.50-0.70)
NEU# (×10 <sup>9</sup> /L)	7.9	0.805† (0.757–0.853)	48.84% (40.21%-57.46%)	91.18% (89.41%-92.95%)	42.00% (34.10%-49.90%)	93.16% (91.57%-94.75%)	5.53 (4.24-7.23)	0.56 (0.47-0.66)
PLT# (×10 <sup>9</sup> /L)	145	0.722† (0.667–0.776)	44.19% (35.62%-52.76%)	90.16% (88.30%-92.02%)	37.01% (29.39%-44.64%)	92.51% (90.84%-94.17%)	4.49 (3.43-5.89)	0.62 (0.53-0.72)
PCT (ng/mL)	0.22	0.898¶ (0.873-0.923)	58.91% (50.42%-67.40%)	90.16% (88.30%-92.02%)	43.93% (36.53%-51.33%)	94.37% (92.90%-95.85%)	5.99 (4.72-7.60)	0.46 (0.37-0.56)
hsCRP (mg/L)	89	0.880§ (0.853-0.908)	58.14% (49.63%-66.65%)	90.06% (88.19%-91.93%)	43.35% (35.97%-50.74%)	94.27% (92.78%-95.75%)	5.85 (4.61-7.42)	0.46 (0.38-0.57)
PT (s)	15.1	0.839† (0.798–0.879)	58.14% (49.63%-66.65%)	91.48% (89.74%-93.22%)	47.17% (39.41%-54.93%)	94.35% (92.89%-95.81%)	6.82 (5.31-8.78)	0.46 (0.37-0.56)
DD (mg/L)	3.9	0.866‡ (0.837-0.896)	55.81% (47.24%-64.38%)	90.57% (88.74%-92.39%)	43.64% (36.07%-51.20%)	94.00% (92.49%-95.51%)	5.92 (4.62-7.58)	0.49 (0.40-0.59)
LDH (U/L)	428	0.866‡ (0.830–0.902)	59.69% (51.23%-68.15%)	91.99% (90.29%-93.68%)	49.36% (41.51%-57.20%)	94.58% (93.14%-96.01%)	7.45 (5.78–9.61)	0.44 (0.35-0.54)
AST (U/L)	51	0.753† (0.709–0.797)	34.88% (26.66%-43.11%)	90.06% (88.19%-91.93%)	31.47% (23.86%-39.08%)	91.36% (89.59%-93.12%)	3.51 (2.60-4.74)	0.72 (0.64-0.82)
CR (µmol/L)	100	0.711 <sup>†</sup> (0.659–0.764)	37.98% (29.61%-46.36%)	90.06% (88.19%-91.93%)	33.33% (25.71%-40.95%)	91.74% (90.00%-93.47%)	3.82 (2.86-5.11)	0.69 (0.60-0.79)
hs-cTnl (pg/mL)	30.1	0.864‡ (0.831-0.898)	58.91% (50.42%-67.40%)	90.97% (89.18%-92.76%)	46.06% (38.46%-53.67%)	94.42% (92.96%-95.88%)	6.53 (5.11-8.34)	0.45 (0.37-0.56)
Prediction model	0.19	0.955 (0.941-0.970)	86.82% (80.98%-92.66%)	90.37% (88.52%-92.21%)	54.11% (47.32%-60.89%)	98.13% (97.25%-99.01%)	9.01 (7.36–11.04)	0.15 (0.09-0.23)

<sup>†</sup>P < 0.001, compared with PAB using the z statistic; †P < 0.01, compared with PAB using the z statistic; NP > 0.05, compared with PAB using the z statistic; AUC, area under the curve; PPV, positive predictive value; NPV, negative predictive value; PLR, positive likelihood ratio; NLR, negative likelihood ratio; CI, confidence interval; PAB, prealbumin; ALB, albumin; GLB, globulin; TP, total protein; WBC#, white blood cell count; LYM#, lymphocyte count; NEU#, neutrophil count; PLT#, platelet count; PCT, procalcitonin; hsCRP, hypersensitive C-reactive protein; PT, prothrombin time; DD, d-dimer; LDH, lactate dehydrogenase; AST, aspartate aminotransferase; CR, creatinine; hs-cTnl, hypersensitive cardiac troponin I.

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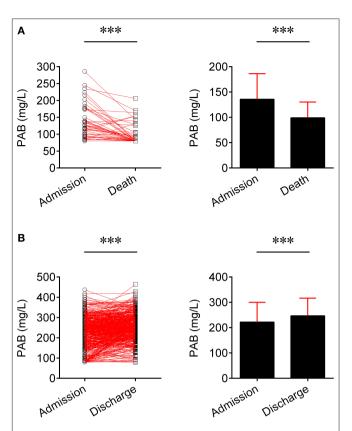
ranging from 16 to 95 years old. The mean age of fatal patients (69.98  $\pm$  12.05 years) was significantly older than recovered patients (58.64  $\pm$  15.17 years) (P < 0.001). Male sex was more predominant in fatal patients (67.44%) than in recovered patients (48.28%) (P < 0.001). Cough and fever were the most prevalent symptoms at disease onset in both fatal (51.16 and 62.02%) and recovered patients (58.42 and 65.72%). Other prevalent symptoms at the onset of illness in fatal patients included shortness of breath and chest tightness; less common symptoms included diarrhea, headache, nausea, vomiting, muscle ache, and pharyngalgia. Shortness of breath was significantly higher in fatal patients (34.88%) than in recovered patients (12.78%) (P < 0.001). Underlying diseases including diabetes mellitus, hypertension, chronic obstructive pulmonary disease, cardiovascular disease, and hematological malignancy were more frequent in fatal patients (19.38, 43.41, 3.88, 17.83, and 1.15%) than in recovered patients (7.3, 25.56, 1.42, 9.63, and 0.2%). The mean time from admission to death was 16.72  $\pm$  11.9 days in fatal patients. The mean time from admission to discharge was  $21.66 \pm 12.01$  days in recovered patients (Table 1).

## Using PAB for Predicting the Prognosis of COVID-19

We observed substantial differences in the levels of proteins including PAB, ALB, GLB, and TP between patients who died of COVID-19 and those who recovered from the disease. It was found that the concentrations of PAB, ALB, and TP on admission were markedly lower in fatal patients than in recovered patients (P < 0.001). On the contrary, the level of GLB on admission was found to be significantly higher in the fatal group than in the recovered group (P < 0.001) (Figure 1A). If using these indexes for distinguishing these two conditions, the best AUC was for PAB [0.915, (95% CI, 0.894-0.937)] (**Table 2**, **Figure 1B**). Notably, the level of PAB  $\leq$  126 mg/L produced a sensitivity of 78.29 % and a specificity of 90.06% (Table 2). In addition, ROC analysis showed that the AUC of ALB was 0.825 (95% CI, 0.792-0.859), with a sensitivity of 45.74% and a specificity of 90.37% when a cutoff value of 29.7 g/L was used to differentiate between fatal and recovered individuals (Table 2, Figure 1B).

# Change of the Level of PAB in the Same Patients

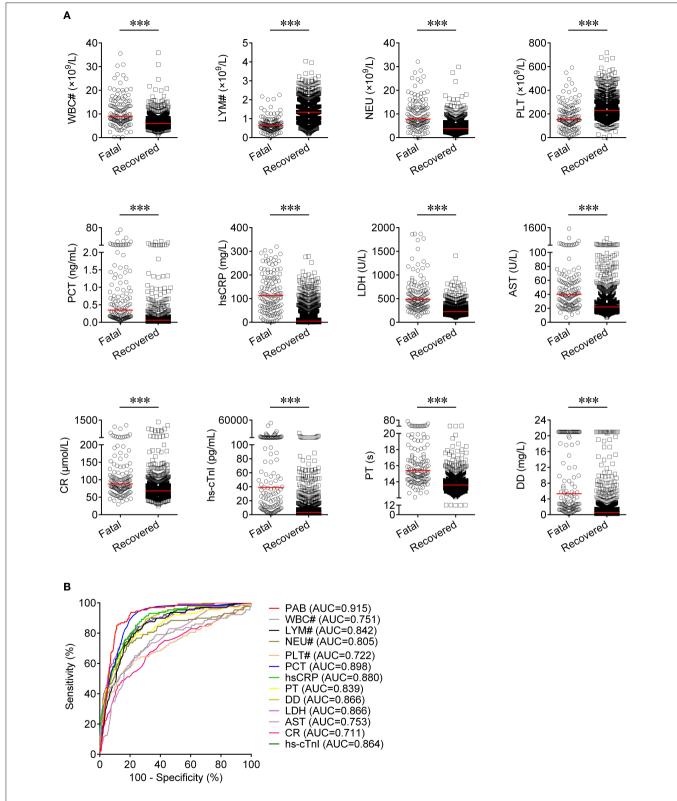
We compared the level of PAB in fatal patients between admission and death. It was found that the level of PAB was significantly decreased at the time of death compared to admission (P < 0.001) (Figure 2A). Furthermore, the concentration of PAB was further compared between admission and discharge in recovered individuals. Conversely, recovered patients showed a significantly higher level of PAB on discharge compared with on admission (P < 0.001) (Figure 2B).



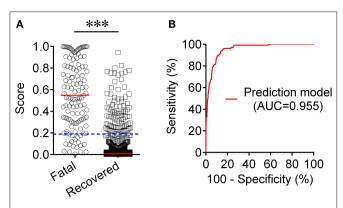
**FIGURE 2** | Change of the level of PAB in the same patients. **(A)** Line graphs showing the level of PAB for each fatal patient on admission and death (n=45). One line represents one patient. \*\*\*P<0.001 (Wilcoxon test). Bar graphs showing the level of PAB on admission and death in fatal patients (n=45). Data are shown as means  $\pm$  SD. \*\*\*P<0.001 (Wilcoxon test). **(B)** Line graphs showing the level of PAB for each recovered patient on admission and discharge (n=501). One line represents one patient. \*\*\*P<0.001 (Wilcoxon test). Bar graphs showing the level of PAB on admission and discharge in recovered patients (n=501). Data are shown as means  $\pm$  SD. \*\*\*P<0.001 (Wilcoxon test). PAB, prealbumin.

## The Comparison of Predictive Value Between PAB and Other Routine Laboratory Indicators for the Prognosis of COVID-19

Routine laboratory markers including WBC#, LYM#, NEU#, PLT#, PCT, hsCRP, PT, DD, LDH, AST, CR, and hs-cTnI were also measured in both fatal and recovered patients on admission. LYM# and PLT# were significantly lower in the fatal group than in the recovered group (P < 0.001) (**Figure 3A**). Conversely, it was found that WBC#, NEU#, PCT, hsCRP, PT, DD, LDH, AST, CR, and hs-cTnI in the fatal group was significantly higher than in the recovered group (P < 0.001) (**Figure 3A**). ROC analysis showed that the AUCs of LYM#, NEU#, PCT, hsCRP, PT, DD, LDH, and hs-cTnI were over 0.8 for distinguishing between fatal patients and recovered subjects (**Figure 3B**). Using a cut-off value of 0.22 ng/ml, the sensitivity and specificity of PCT for discriminating fatal cases from recovered individuals were 58.91 and 90.16%, respectively (**Table 2**). With a threshold of 89



**FIGURE 3** | The comparison of predictive values between PAB and other route laboratory markers for the prognosis of COVID-19. **(A)** Scatter plots showing the levels of WBC#, LYM#, NEU#, PLT#, PCT, hsCRP, PT, DD, LDH, AST, CR, and hs-cTnl in fatal (n=129) and recovered (n=986) patients. Horizontal lines indicate the median. \*\*\*P < 0.001 (Mann-Whitney *U* test). **(B)** ROC analysis showing the performance of PAB, WBC#, LYM#, NEU#, PLT#, PCT, hsCRP, PT, DD, LDH, AST, CR, and hs-cTnl in distinguishing fatal patients from recovered patients. COVID-19, coronavirus disease 2019; PAB, prealbumin; WBC#, white blood cell count; LYM#, lymphocyte count; NEU#, neutrophil count; PLT#, platelet count; PCT, procalcitonin; hsCRP, hypersensitive C-reactive protein; PT, prothrombin time; DD, d-dimer; LDH, lactate dehydrogenase; AST, aspartate aminotransferase; CR, creatinine; hs-cTnl, hypersensitive cardiac troponin I; ROC, receiver operating characteristic curve: AUC, area under the curve.



**FIGURE 4** | Establishment of prediction model for prognosis of COVID-19 based on the combination of PAB and other routine laboratory indexes. **(A)** Scatter plots showing the score of prediction model in fatal patients (n=129) and survived cases (n=986). Horizontal lines indicate the median. \*\*\*P<0.001 (Mann-Whitney U test). Blue dotted lines indicate the cutoff value in distinguishing these two groups. **(B)** ROC analysis showing the performance of the prediction model in distinguishing fatal patients from recovered cases. COVID-19, coronavirus disease 2019; ROC, receiver operating characteristic curve; AUC, area under the curve.

mg/L, hsCRP was able to distinguish fatal patients from recovered patients with a sensitivity of 58.14% and a specificity of 90.06% (**Table 2**). Moreover, with a threshold of 15.1 s, PT had an AUC of 0.839 (95% CI, 0.798–0.879) with a sensitivity of 58.14% and a specificity of 91.48% (**Table 2**). The predictive utility of PAB was better than WBC#, LYM#, NEU#, PLT#, hsCRP, PT, DD, LDH, AST, CR, and hs-cTnI, and was comparable to PCT for the prognosis of patients with COVID-19 (**Table 2**).

## Establishing a Model for Predicting the Death of Patients With COVID-19

To establish a prediction model based on the combination and other routine laboratory markers on admission for distinguishing fatal patients from recovered individuals, all variables with statistical significance were used for multivariable logistic regression analysis. A prediction model was built as the following: P = 1/[1 +e-(-0.016\*PAB-0.908\*LYM#+0.067\*NEU#+0.06\*PCT+0.005\*hsCRP+0.154\*PT  $e^{-c}$ +0.003\*LDH+0.002\*CR+0.001\*hs-cTnI-3.036)] P, predictive value; e, natural logarithm (Supplementary Table 1). ROC analysis showed that the AUC of the prediction model was 0.955 (95% CI, 0.941-0.970) (Figure 4). When the cutoff value was set at 0.19, the following diagnostic parameters of the model were obtained: sensitivity, 86.82% (95% CI, 80.98–92.66%); specificity, 90.37% (95% CI, 88.52-92.21%) (Table 2). These data suggested that our established model, based on the combination of a nine-indicator biosignature, had good performance for predicting the death of patients with COVID-19.

### DISCUSSION

The rapidly increasing number of laboratory-confirmed COVID-19 cases worldwide has put a heavy burden on the medical resources in countries with large outbreaks (29–31). The World Health Organization has declared COVID-19 to be a public health emergency of international concern. The determination of the outcome of the disease is of crucial importance in regulating limited medical resources and providing better care for patients (32, 33). Meanwhile, progression at the early stage is very important to the outcome or the prognosis of the disease. Therefore, after patient admission, identifying predictors that can predict the likelihood of disease progression would help physicians to decide which group of patients can be managed safely at district hospitals and who needs early transfer to tertiary centers.

Although previous studies have found that a series of parameters on admission are correlated with mortality risk (34), there is limited information in the existing literature regarding the relationship between nutrition indexes and disease progression in SARS-CoV-2-infected patients. Several studies have reported the value of nutritional indicators represented by PAB on determining the severity of viral infections and predicting the prognosis of patients (35, 36). In addition, there is some literature describing the roles of inflammatory indicators (hsCRP) and PCT), coagulation indicators (DD), and the number of lymphocytes in monitoring the disease progression during viral infection and prediction of disease outcome (36-38). Our study comprehensively described the differences in PAB and other routine laboratory parameters between the patients who died of COVID-19 and those who recovered from the disease. It was found that a low level of PAB on admission may indicate a poor prognosis and that the predictive value of PAB is superior to most routine laboratory indicators that reflect functional impairment or disorder of organs for prognosis of COVID-19. To our knowledge, this is the first study to report the relationship between PAB level and the outcome of COVID-19.

Host factors trigger an immune response against the pathogens during viral infection (39-41). Immune insufficiency may contribute to viral replication and cause tissue damage, resulting in a bad outcome (42, 43). The systematic overwhelming inflammation and multi-organ dysfunction are more common in deceased COVID-19 patients than in recovered patients (44). This may be caused by the poor basic immune status of patients. In accordance with recent reports, advanced age and comorbidities such as diabetes and hypertension are believed to be risk factors of death from COVID-19 (45). This suggests that immune and nutritional status may be critical in predicting disease progression at the early stage of COVID-19. In other words, poor immunity may play a role in COVID-19-related death. Thus, early vigilant monitoring along with high quality supportive care are needed for patients at high risk of death. Although the number of lymphocytes can partially reflect the host's immune function, our data indicates that the nutritional indicators represented by PAB may show this effect better. A single use of PAB could achieve a modest prediction performance for the prognosis of COVID-19. When combined with other conventional laboratory indicators, PAB could produce a better performance.

In addition, opposite trends were found in PAB level between fatal and recovered groups. The level of PAB gradually decreased with growing hospital stay in fatal patients but had an increased trend in recovered patients (Supplementary Figure 1). These data suggest that the dynamic monitoring of PAB provides a potential value for mastering the process of the disease. These findings would also alert clinicians to pay special attention not only to inflammatory indexes but also to nutritional and immune status.

Some limitations in this study should be addressed. First, since the number of participants recruited in this one-center study is limited, a further design with a large multicenter cohort will provide more conclusive and valuable data. Second, some indicators, such as PAB and DD, were over the detection limit, which would lead to bias. Finally, medical history such as malnutrition and the use of steroid drugs were not included in the regression analysis and it would affect our results.

Collectively, our study provides the evidence that PAB level on admission could be used to predict in-hospital mortality in patients with COVID-19. The information provided in our study shows potential value in enriching knowledge about this critical disease, helping clinicians to identify patients with poor prognosis at an early stage before they die from COVID-19, guiding appropriate and effective management for future patients, and consequently helping to improve patients' outcomes and decrease the fatality rate.

### **DATA AVAILABILITY STATEMENT**

All datasets presented in this study are included in the article/Supplementary Material.

### **ETHICS STATEMENT**

This study was reviewed and approved by the ethical committee of Tongji Hospital, Tongji Medical College, Huazhong University

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of Science and Technology, Wuhan, China (TJ-C20200128). The patients/participants provided their written informed consent to participate in this study.

### **AUTHOR CONTRIBUTIONS**

YL, FW, and ZS conceived the research, designed the study, interpreted data, and wrote the manuscript. YL, YX, LM, and XY contributed to the acquisition of clinical data. YL, QL, GT, and HS recruited the participants, performed experiments, and analyzed data. All authors approved the final draft of the manuscript.

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### **SUPPLEMENTARY MATERIAL**

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fmed. 2020.00374/full#supplementary-material

Supplementary Figure 1 | Dynamic monitoring of PAB in the representative of fatal and recovered patients. (A) Line diagrams showing the level of PAB in four representatives of the fatal group. (B) Line diagrams showing the level of PAB in four representatives of the recovered group. \*days from admission onset to detection. PAB, prealbumin.

Supplementary Table 1 | Logistic regression analysis.

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**Conflict of Interest:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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