



The Ki-67 Proliferation Index-Related Nomogram to Predict the Response of First-Line Tyrosine Kinase Inhibitors or Chemotherapy in Non-small Cell Lung Cancer Patients With Epidermal Growth Factor Receptor-Mutant Status

OPEN ACCESS

Edited by:

Paula Tejera, Harvard University, United States

Reviewed by:

Pai-Chien Chou, Taipei Medical University Hospital, Taiwan Rudolf Maria Huber, Ludwig Maximilian University of Munich, Germany

*Correspondence:

Jinhong Mei mjhdoctor@126.com Weijia Wang 734162529@qq.com Chunliang Wang wang330006@126.com

[†]These authors share first authorship

Specialty section:

This article was submitted to Pulmonary Medicine, a section of the journal Frontiers in Medicine

Received: 21 June 2021 Accepted: 28 September 2021 Published: 05 November 2021

Citation:

Gu W, Hu M, Xu L, Ren Y, Mei J, Wang W and Wang C (2021) The Ki-67 Proliferation Index-Related Nomogram to Predict the Response of First-Line Tyrosine Kinase Inhibitors or Chemotherapy in Non-small Cell Lung Cancer Patients With Epidermal Growth Factor Receptor-Mutant Status. Front. Med. 8:728575. doi: 10.3389/fmed.2021.728575 Weiguo Gu^{1,2†}, Mingbin Hu^{1,2†}, Linlin Xu^{1,3†}, Yuanhui Ren¹, Jinhong Mei^{1,3*}, Weijia Wang^{1*} and Chunliang Wang^{4*}

¹ Department of Pathology, The First Affiliated Hospital of Nanchang University, Nanchang, China, ² Department of Oncology, The First Affiliated Hospital of Nanchang University, Nanchang, China, ³ Institute of Molecular Pathology, Nanchang University, Nanchang, China, ⁴ Department of Neurosurgery, The First Affiliated Hospital of Nanchang University, Nanchang, China

Background: The correlation between Ki-67 and epidermal growth factor receptor (EGFR)- or Kristen rat sarcoma viral oncogene homolog (KRAS)-mutant status in advanced or postoperative-recurrent non-small cell lung cancer (NSCLC) has fewer studies reported, and the prognostic role of Ki-67 with first-line EGFR-tyrosine kinase inhibitors (TKIs) or chemotherapy remains controversial.

Methods: A total of 295 patients were tested for EGFR-mutant status in advanced or postoperative-recurrent NSCLC and received first-line EGFR-TKIs or chemotherapy for treatment. Ki-67 expression was retrospectively analyzed by immunohistochemistry. The Kaplan-Meier method was used to calculate survival rates. The multivariate Cox proportional hazards model was used to generate a nomogram. The established nomogram was validated using the calibration plots.

Results: The expression levels of Ki-67 were divided into low (<60%, n = 186) and high (\geq 60%, n = 109) groups, based on the receiver operating characteristic curve. The expression levels of Ki-67 were found to be higher in patients with KRAS mutations when compared to KRAS wildtype, and EGFR wildtype was higher than EGFR mutations. The median overall survival (OS) of the low Ki-67 expression group was significantly longer than that of the high Ki-67 group, no matter in all NSCLC, EGFR mutations, EGFR wildtype, KRAS-mutant status, EGFR-TKIs, or chemotherapy of patients (P < 0.05). Subgroup analysis showed that the KRAS wildtype or EGFR mutations combine with low Ki-67 expression group had the longest median OS than KRAS mutations or EGFR wildtype combine with Ki-67 high expression group (P < 0.05). In the training cohort, the multivariate Cox analysis identified age, serum lactate dehydrogenase (LDH), serum Cyfra211, EGFR mutations, and Ki-67 as independent prognostic factors, and

1

a nomogram was developed based on these covariates. The calibration curve for predicting the 12-, 24-, and 30-month OS showed an optimal agreement between the predicted and actual observed outcomes.

Conclusions: The Ki-67 expression-based nomogram can well predict the efficacy of first-line therapy in NSCLC patients with EGFR- or KRAS-mutant status, high expression levels of Ki-67 correlated with a poor prognosis.

Keywords: non-small cell lung cancer, tyrosine kinase inhibitor, Ki-67, nomogram, survival

INTRODUCTION

Although advances in precision medicine have improved cancer treatment, lung cancer is still the main cause of morbidity and cancer-related mortality worldwide (1, 2). Epidermal growth factor receptor-tyrosine kinase inhibitor (EGFR-TKIs) remains the first-line therapeutic option for advanced non-small cell lung cancer (NSCLC) with EGFR mutation (3, 4). However, after prolonged oral targeted therapy, most patients develop slow or rapidly progressive diseases and drug resistance, such as resistance to osimertinib, a third-generation drug. These drugs fail to bind to original EGFR mutation sites and cause secondary mutations (5), resulting in primary drug resistance (6), secondary drug resistance (7), and activation of bypass pathway (8). It has been shown that tumour, node, metastasis (TNM) stage, tumor differentiation, serum laboratory indexes, or immunohistochemical (IHC) staining may be significant prognostic factors for NSCLC treated with EGFR-TKIs (9, 10).

Ki-67, a proliferative index (PI), is a nuclear marker associated with tumor cell proliferation, it is correlated with progression, metastasis, and prognosis in various malignancies (11). The expression of Ki-67 is an important prognostic indicator for prostate cancer (12), multiple myeloma (13), and breast cancer (14). Moreover, Ki-67 expression is an important biomarker for luminal classification of breast cancer. High expression levels of Ki-67 are associated with neoadjuvant or adjuvant chemotherapy in breast cancer while low expression levels of Ki-67 correlate with the better effect of treatment outcomes (14, 15). Warth et al. (16) investigated the expression levels of Ki-67 in 417 patients with stages I-IV NSCLC, with cutoff values of Ki-67 set at 25% (lung adenocarcinoma, LUAD) and 50% (lung squamous cell carcinoma, LCSC). They found that high expression levels of Ki-67 are associated with worse prognostic outcomes and were, therefore, considered an important prognostic biomarker for stratifying LUAD patients to be administered with adjuvant therapy. Del Gobbo et al. (17) showed that Ki-67 PI and metabolic activities in the LCSC group were higher than those in patients with LUAD with the highest intratumoral heterogeneity in proliferative activities and could also be used as prognostic factors. Zhu et al. (18) investigated the correlation between Ki-67 and EGFR mutations in 523 radically resected NSCLC patients. They found that high expression levels of Ki-67 are associated with poor survival outcomes in both EGFR mutation and wildtype cohorts.

Although Ki-67 is associated with EGFR mutation and poor survival outcomes for NSCLC patients, the relationship between

Ki-67 expression and first-line EGFR-TKIs or chemotherapy in NSCLC patients has not been elucidated. Moreover, a limited number of studies have evaluated the association between the KRAS-mutant status and Ki-67. Although the cutoff value of Ki-67 as a prognostic reference in NSCLC has been suggested by a large-scale meta-analysis and several reviews, there is still no consensus on the optimal cutoff value. In this study, we investigated the prognostic value of Ki-67 in NSCLC patients administered with first-line EGFR-TKIs or chemotherapy and established a nomogram for survival prediction.

PATIENTS AND METHODS

Patients and Data Collection

We retrospectively identified patients with histologically confirmed and tested for EGFR mutations in advanced or postoperative-recurrence NSCLC administered with first-line receiving EGFR-TKIs or chemotherapy treatment. This study was performed between January 2014 and March 2019. Approval was obtained by the Ethics Committee of the First Affiliated Hospital of Nanchang University, Jiangxi P.R. China, all patients provided written informed consent before received EGFR-TKIs or chemotherapy and compliance with the declaration of Helsinki. The clinic-pathological characteristics of patients included gender, smoking index, age, pathology, TNM stage, and EGFR mutation subtype (EGFR genetic tests were performed during pre-treatment) among others. The inclusion criteria were (i) all patients were diagnosed with NSCLC by histological or cytological analysis; (ii) patients with EGFR gene mutation were detected and administered with EGFR-TKIs or platinumcontaining chemotherapy as first-line therapies; and (iii) enough tissues for Ki-67 IHC staining or Ki-67 had been done in the past. The exclusion criteria were as follows: (i) patients with small-cell lung cancer combined with other malignancies; (ii) patients with unknown EGFR mutation type and incomplete clinical data; (iii) patients with Eastern Cooperative Oncology Group performance status (ECOG PS) score \geq 3; and (iv) patients who were intolerant to adverse effects or other comorbidities and those who withdrew from further treatment or lost to follow-up.

Immunohistochemistry Analysis

Expression levels of Ki-67 in paraffin-embedded tissues from 69 NSCLC patients were detected by IHC, and 226 patients were detected at their first diagnosis. NSCLC tissue sections were sliced from paraffin blocks for IHC analysis of Ki-67 antigens at our cancer pathology center. Antigen retrieval was

Ki-67 Proliferation Index Related Nomogram

performed in a water bath using an ethylenediaminetetraacetic acid (EDTA) retrieval buffer (100 ml, Zhongshan Jinqiao company, Beijing). Slides were coated with 50 µl of Ki-67 primary antibody (1:100, 7b11, Zhongshan Jinqiao company, Beijing) and incubated at 4°C refrigerators overnight. Then, they were incubated with a secondary antibody (horseradish peroxidase-labeled mouse/rabbit universal secondary antibody, Dako company, Denmark and Roche company, Switzerland) at 37°C incubators for 15 min. Chromogen diaminobenzidine (DAB Substrate System, Dako, Carpinteria, CA, USA) was used to assess immunoreactivity. Two experienced pathologists, double-blinded to the treatment given, performed histological examinations. Appropriate positive and negative controls were used. Based on the criteria of the NSCLC Working Group, Ki-67 nuclear staining (punctate or diffuse) was found to be positive by selecting the uniformly stained area of tumor cells, and the percentage of Ki-67 positive cells was determined by counting at least 500 tumor cells of immunoreactive nuclei.

Post-treatment Evaluation Criteria and Follow-Up

Patients were administered with first-line therapy for 3–4 weeks, followed by imaging examination every 2 months and disease evaluation by chest and abdominal CT, bone scan, and craniocerebral nuclear MRI. The Response Evaluation Criteria in Solid Tumors 1.1 (RECIST1.1) was established for short-term effects. Overall survival (OS) was evaluated from the starting date of first-line treatment to the date of death or last follow-up. Follow-up data of NSCLC patients were obtained between January 2014 and March 2019. The cutoff date was July 30, 2020, and the median follow-up period was 25 months.

Establishment and Validation of the Nomogram

Patients were randomized into the training cohort group (n = 206) and validation cohort group (n = 89) at a ratio of 2:1 using the R package "rms" analysis. The univariate and multivariate Cox analyses were used to identify the independent prognostic factors in the training cohort and establish a nomogram. The concordance index (C-index) was used to assess the predictive ability of the nomogram, which was internally validated by the Bootstraps method (500 repetitions) for accuracy and unbiased estimates (19, 20). Furthermore, calibration curves of the model for 12-, 24-, 30-month survival outcomes were established to compare the OS outcomes predicted by the nomogram and actual survival outcomes.

Statistical Analysis

The Ki-67 expression levels were divided into low and high groups by the receiver operating characteristic curve (ROC) (according to the treatment regimens) in all NSCLC patients. Continuous variables of clinical-pathologic factors were converted into categorical variables for analysis. Kaplan-Meier method was used to calculate survival rates while the univariate and multivariate Cox analyses were performed to determine the hazard ratio (HR) and 95% CI for the identification of independent prognostic factors. A *P*-value of <0.05 was

considered statistically significant. All statistical analyses were performed using SPSS version 22.0 software (SPSS Inc., Chicago, IL, USA), GraphPad Prism version 5.0 software (Inc., La Jolla, CA, USA), or R statistical software version 4.0.0 (http://www.Rproject.org). Cutoff points for peripheral serum LDH and tumor biomarkers were determined as previously described (21).

RESULTS

Patient Characteristics

A total of 295 NSCLC patients were included, of which 269 patients had adenocarcinoma, 17 patients had squamous cell carcinoma, two patients had adenosquamous carcinoma, five patients had unclassified NSCLC, while two patients had large cell carcinoma. A flowchart for patient selection in this study is shown in Figure 1. One hundred and fifty-five patients were men while 140 patients were women. The mean age for all patients was 57.5 years (range 22-84 years). There were a total of 188 patients with EGFR mutations (94 patients were positive for del 19, 82 patients for L858R, 25 patients for T790M, five patients for Exon18+, and 13 patients for Exon 20+), 107 patients with EGFR wild-type, 11 patients had KRAS mutations, two patients had Her2 amplifications, two patients had Met amplifications, three patients had EGFR amplifications, four patients had ALK mutations, two patients had ROS1 mutations, five patients had PIK3CA mutations, while three patients had RET mutations (Figure 2J). One hundred and sixty-nine patients who were EGFR-mutant positive were administered with EGFR-TKIs therapy (such as, gefitinib, erlotinib, icotinib, osimertinib, and afatinib) while 126 patients who were EGFR-mutant negative or had EGFR-resistance genes were administered with chemotherapy with platinum-containing regimen or radiotherapy. NSCLC staging at initial diagnosis was based on the American Joint Committee on Cancer (AJCC) staging system. There were 19 patients with unresectable IIIB-IIIC stage, 55 patients with postoperative recurrence, and 221 patients with advanced NSCLC (Table 1).

Association Between Ki-67 Expression and Clinic-Pathological Features

Ki-67 was found to be mainly expressed in the nucleus (Figures 2A-C). The Ki-67 expression levels in the EGFR-mutant group were lower than those of the EGFR-wildtype group. Moreover, Ki-67 expression levels in EGFR-TKIs group patients were lower than those of the chemotherapy group (P < 0.05, Figures 2D,E). The relationship between pathology and Ki-67 shows that squamous cell carcinoma of Ki-67 was higher than adenocarcinoma (P < 0.05, Figure 2F). Ki-67 expression levels in the KRAS-mutant group were higher than those in the KRAS-wildtype group (P < 0.05, Figure 2G). Ki-67 expression levels in the low platelet-to-lymphocyte ratio (PLR) group were lower than in the high PLR group (P < 0.05, Figure 2H), while The Ki-67 expressions in the low Cyfra211 group were lower than in the high Cyfra211 group (P < 0.05, Figure 2I). Moreover, the Ki-67 ROC curve for cutoff points for treatment regimens was established. It was found that the cutoff value of 66% had the highest sensitivity and specificity (sensitivity:



37.3%, specificity: 81.7%, Youden index: 0.19, ROC = 0.6229, P = 0.0003; **Figure 2K**). Therefore, based on Ki-67 expression levels, we divided the patients into low (Ki-67 < 60%, n = 186) and high expression groups (Ki-67 \ge 60%, n = 109).

The Univariate and Multivariate Survival Analyses for First-Line Therapy With Ki-67 in the Primary Cohort

Based on the EGFR-mutant status, the NSCLC patients were assigned to the EGFR-TKIs (n = 169) and chemotherapy groups (n = 126). The median OS (23.9 vs. 11.6 months, P < 0.001, Figure 3A) for the EGFR-TKIs group was longer than for those in the chemotherapy group. In addition, the EGFR mutation group had a longer median OS when compared to the EGFR-wildtype group (23.5 vs. 11.7 months, *P* < 0.001, Figure 3B). Moreover, the low Ki-67 group had a longer median OS compared to the high Ki-67 NSCLC group (21.3 vs. 12.1 months, P < 0.001, Figure 3C). Additionally, in EGFR mutation patients, subgroup analysis revealed that the low Ki-67 expression group had a high median OS when compared to the high expression group (30.2 vs. 18.3 months, *P* < 0.001, **Figure 3D**); in EGFR wildtype patients, the low Ki-67 group had a high median OS than the high expression group (14.8 vs. 9.2 months, P < 0.001, Figure 3E). In EGFR-TKIs patients, the median OS for the low Ki-67 group was longer than that of the high Ki-67 group (30.5 vs. 20.7 months, P = 0.001, Figure 3F); in chemotherapy patients, the low Ki-67 group had a longer median OS (15.5 vs. 9 months, P = 0.001, **Figure 3G**) than the high group. The KRAS-wildtype group of patients had a longer median OS (19.2 vs. 13 months, P = 0.038, **Figure 3H**) than the KRAS mutation group in all NSCLC patients. Moreover, based on KRAS-mutant status and Ki-67 expression, we found that KRAS wildtype combines with low Ki-67 expression group had the longest median OS (22.4 vs. 6 vs. 15.1 vs. 11.7 months, P < 0.001, **Figure 3I**) than the other three groups; the EGFR mutation group combines with low Ki-67 expression group also had a long median OS (30.2 vs. 14.8 vs. 18.3 vs. 9.2 months, P < 0.001, **Figure 3J**) when compared to the other three groups (**Table 2**).

Moreover, univariate analysis showed that gender, TNM stage, smoking index, treatment regimens, pathological type, EGFR mutation, Kristen rat sarcoma viral oncogene homolog (KRAS) mutation, PLR ratio, neutrophil-to-lymphocyte ratio (NLR) ratio, serum LDH, serum Cyfra211, and Ki-67 were correlated with OS and first-line therapeutic outcomes in NSCLC patients (**Table 2**). Significant factors from univariate analysis (P < 0.05) were included in the multivariate Cox analysis, which showed that the TNM stage, serum LDH, treatment regimens, and Ki-67 were independent prognostic factors for OS (P < 0.05; **Table 3**).

Association Between the Short-Term Therapeutic Effects of First-Line Therapy and Ki-67 Expressions

Disease progression was evaluated through imaging examination, every 2 months after 3–4 weeks of treatment. For all NSCLC



patients, the low Ki-67 expression groups had a higher objective remission rate (ORR, 25.1 vs. 3.1%) and disease control rate (DCR, 59.7 vs. 11.7%) than the high Ki-67 expression groups. Additionally, in the EGFR-TKIs group, subgroup analysis revealed that the low Ki-67 expression groups had a significantly higher ORR (33.7 vs. 2.4%) and DCR (73.9 vs. 12.4%) than the high Ki-67 expression group. Furthermore, in the chemotherapy group, the ORR (13.5 vs. 3.9%) and DCR (40.5 vs. 14.3%) for low Ki-67 expression subgroups were significantly longer than those of the high Ki-67 expression subgroups (**Table 4**).

The Univariate and Multivariate Survival Analyses of the Association Between First-Line Therapy and Clinic-Pathologic Features in the Training Cohort

Considering the small number of EGFR mutation cases, all NSCLC patients were analyzed by a nomogram prediction model. Advanced or postoperative-recurrent NSCLC patients in the primary cohort were randomized divided into the training cohorts (n = 206) and validation cohorts (n = 89) at a ratio of 2:1, using the "rms" R package. In addition, the clinic-pathological features and survival of patients were

analyzed in the training cohort. Univariate analysis showed that the gender, age, TNM stages, EGFR mutation, KRAS mutation, treatment regimens, PLR ratio, NLR ratio, serum LDH, serum Cyfra211, and Ki-67 were correlated with the OS following first-line therapy (P < 0.05; **Table 2**). Thereafter, all the significant factors from univariate analysis (P < 0.05) were included in the multivariate Cox analysis, which showed that age, serum LDH, serum Cyfra211, EGFR mutation, and Ki-67 were independent prognostic factors associated with first-line therapeutic in NSCLC patients (P < 0.05; **Table 3**).

Development and Internal Validation of the Nomogram

The "rms" package in R was used to generate the predictive nomogram through the multivariate Cox analysis in the training cohort. Five variables were incorporated into the nomogram, such as age, serum LDH, serum Cyfra211, EGFR mutation, and Ki-67. The nomogram showed that the EGFR-mutant status and Ki-67 had the most significant contribution to OS among NSCLC patients. Additionally, the total prognostic scores for every variable were calculated and the scores ranged from 0 to 450. Finally, the variables were used to estimate the 12-, 24-, and 30-month OS in NSCLC patients by drawing a straight line TABLE 1 | Characteristics of 295 patients received first-line therapy in NSCLC with EGFR mutation status.

Characteristic		Ki-67 low group (<i>n</i> = 186, %)	Ki-67 high group (n = 109, %)	P value
Gender	Male	89 (47.8)	66 (60.6)	0.035
	Female	97 (52.2)	43 (39.4)	
Age (years)	≤60	99 (53.2)	57 (52.3)	0.877
	>60	87 (46.8)	52 (47.7)	
ECOG PS	0-1	128 (68.8)	80 (73.4)	0.405
	2-3	58 (31.2)	29 (26.6)	
Smoking index ^a	<400	129 (69.4)	65 (59.6)	0.089
	≥400	57 (30.6)	44 (40.4)	
Pathological type	Adenocarcinoma	175 (94.1)	94 (86.2)	0.022
	Other NSCLC	11 (5.9)	15 (13.8)	
TNM stage	Recurrent	35 (18.8)	20 (18.3)	0.87
	IIIB-C	13 (7.0)	6 (5.5)	
	IV	138 (74.2)	83 (76.1)	
Tumer diameters	≤3cm	62 (33.3)	25 (22.9)	0.059
	>3cm	124 (66.7)	84 (77.1)	
Brain metastasis	No	132 (71)	89 (81.7)	0.041
	Yes	54 (29)	20 (18.3)	
EGFR-mutant status	EGFR wildtype	61 (32.8)	46 (42.2)	0.105
	EGFR mutation ^d	125 (67.2)	63 (57.8)	
KRAS mutant	Wildtype	182 (97.8)	102 (93.6)	0.062
	Mutation	4 (2.2)	7 (6.4)	
Treatment regimens	EGFR-TKIs ^e	117 (62.9)	52 (47.7)	0.011
	Chemotherapy ^f	69 (37.1)	57 (52.3)	
PLR ratio ^b	≤184	118 (63.4)	52 (47.7)	0.008
	>184	68 (36.6)	57 (52.3)	
NLR ratio ^b	≤3.68	124 (66.7)	65 (59.6)	0.224
	>3.68	62 (33.3)	44 (30.4)	
LDH (U/L) ^c	≤250	106 (57)	55 (50.5)	0.277
	>250	80 (43)	54 (49.5)	
CEA (ng/ml) ^c	≤6.5	67 (36)	49 (45)	0.13
	>6.5	119 (64)	60 (55)	
Cyfra211 (ng/ml) ^c	≤3.3	80 (43)	33 (30.3)	0.03
	>3.3	106 (57)	76 (69.7)	

ECOG PS, Eastern Cooperative Oncology Group performance status; EGFR, Epidermal growth factor receptor; PLR, platelet-to-lymphocyte ratio; NLR, neutrophil-to-lymphocyte ratio; LDH, lactate dehydrogenase; CEA, carcinoembryonic antigen; Cyfra211, cytokeratin 19 fragment;Ki-67, nuclear proliferation antigen 67; ^anumber of cigarettes per day×smoking age, ^bthe cut-off points was used mean value, ^cthe cut-off points was used relevant assay kits. ^ddel19/L858/T790M/Exon20/Exon18, ^eGefitinib/Erlotinib/Icotinib/Osimertinib/Afatinib, ^fPemetrexed/Docetaxel/Paclitaxel/Gemcitabine+platinum or ± radiotherapy.

through the survival probability scales and obtaining total points (**Figure 4**). In the internal validation cohort, following first-line therapy, the calibration plots showed that the 12-, 24-, and 30-month OS probabilities predicted by the nomogram were close to the actually observed values in NSCLC (**Figures 5A–C**).

DISCUSSION

EGFR-TKIs remain the first-line therapeutic options for advanced NSCLC with EGFR mutations. However, most patients develop slow or rapidly progressive disease and drug resistance after oral-targeted therapy. In addition, nearly 40–50% of the drug-related resistance mechanisms are unknown (5, 6, 22). Previous studies have reported on clinical factors associated with first-line therapy in advanced EGFR positive NSCLC patients. Various nomograms have also been established to predict the survival outcomes for EGFR positive NSCLC patients, and it has been found that IHC biomarkers play an important role in predicting the survival of patients and efficacies of EGFR-TKIs (19, 20, 23–25). However, few studies have evaluated the correlation between Ki-67 expression levels and the first-line therapeutic outcomes in NSCLC patients with either EGFR- or KRAS-mutant status (**Table 5**).

The present study assessed the clinical-pathological factors and tested the expression of Ki-67 through IHC in advanced or postoperative recurrence NSCLC with EGFR mutations.



The study also assessed the relationship between Ki-67 and OS following first-line therapy. Additionally, the multivariate Cox analysis of independent prognostic factors was conducted after which a nomogram was generated to predict the value of Ki-67. To the best of our knowledge, this was the first study to incorporate the expression of Ki-67 and common clinical-pathological factors into a nomogram, to predict survival following first-line treatment in NSCLC with EGFR or KRAS mutations. This may be useful to clinicians in identifying the potential factors affecting first-line therapy.

Immunohistochemical markers are important for the diagnosis and treatment of numerous types of cancers, although the indicators are not fully understood. In addition, Ki-67 has been shown to be expressed in all mitosis in cancer cells and its overexpression increased tumor cell proliferation, invasion, and metastasis of tumors (11, 29–31). In addition, Ki-67 is an important reference index for the diagnosis and treatment of breast, lung, and prostate cancers (12, 14, 16). Notably, Peng et al. (30) assessed the relationship between the epithelial-mesenchymal transition (EMT) phenotype and the proliferative marker, Ki-67, in NSCLC. They found that Ki-67 is closely associated with EMT, and overexpressed Ki-67 is correlated with poor recurrence-free survival (RFS) and OS. The expression levels of Ki-67 have been associated with

EGFR mutations and high expression of Ki-67 correlates with poor survival in postoperative NSCLC (18, 32). However, the relationship between first-line EGFR-TKIs or chemotherapy and the EGFR mutation status in NSCLC is unknown. In addition, the cutoff value for Ki-67 as the standard for clinical diagnosis and treatment is still unclear (33).

In this study, two Ki-67 groups were established based on the ROC curve. Expression levels of Ki-67 in the EGFR-positive group were lower than those of the EGFR-wildtype group, and the EGFR-mutant patients administered with EGFR-TKIs had lower expression levels of Ki-67 when compared to the chemotherapy group. Moreover, we assessed the correlation between Ki-67 expression levels and prognostic outcomes in EGFR mutation patients. It was found that in NSCLC, EGFR-TKIs, chemotherapy, EGFR-positive, and EGFR-wildtype groups, the median OS of the low Ki-67 expression group was significantly longer than that of the high Ki-67 expression group. And our findings also revealed that the low Ki-67 expression groups also had a longer ORR and DCR than the high expression category. Furthermore, we interestingly found that EGFR-positive low Ki-67 expression subgroups had the longest median OS time when compared to EGFR-wildtype or EGFRpositive high Ki-67 expression subgroups. Therefore, it is highly likely that Ki-67 is associated with the EGFR mutation status,

TABLE 2 | Univariate analysis of primary cohort and training cohort for first-line treatment in NSCLC.

Characteristic		Primary cohort (mOS)				Training cohort (mOS)			
		Ν	months	Р	HR (95%CI)	Ν	months	Р	HR (95%CI)
Gender	Male	155	14.1		Ref	116	15		Ref
	Female	140	22.4	0.008	0.665 (0.491–0.9)	90	28.7	0.04	0.683 (0.475–0.982)
Age (years)	≤60	156	20		Ref	102	20.9		Ref
	>60	139	18.4	0.54	1.098 (0.814–1.483)	104	16.4	0.036	1.469 (1.025–2.105)
ECOG performance	0–1	208	20		Ref	144	18.7		Ref
	2–3	87	15.8	0.101	1.314 (0.948–1.823)	62	15.8	0.217	1.376 (0.942–2.011)
Smoking index	<400	194	20.9		Ref	130	20.9		Ref
-	≥400	101	14.3	0.018	1.448 (1.066–1.967)	76	15.5	0.219	1.256 (0.873-1.807)
Pathological type	Adenocarcinoma	269	20.4		Ref	193	20		Ref
0 ,,	Other NSCLC	26	11.4	<0.001	2.382 (1.466–3.87)	13	14.5	0.158	1.686 (0.816–3.481)
TNM stage	Recurrent	55	21.3	0.005	Ref	42	21.3	0.003	Ref
Ū.	IIIB-C	19	9.8	0.002	2.741 (1.439–5.221)	11	9.8	0.002	3.463 (1.563–7.673)
	IV	221	18.7	0.56	1.127 (0.753–1.687)	153	19.2	0.912	1.027 (0.643–1.639)
Brain metastasis	No	221	20.8		Ref	158	19.2		Ref
	Yes	74	16.6	0.168	1.27 (0.904–1.785)	48	16.3	0.332	1.22 (0.816–1.824)
PLR ratio	≤184	170	20.9		Ref	121	21.3		Ref
	>184	125	15.5	0.017	1.445 (1.069–1.953)	85	14	0.012	1.582 (1.104-2.266)
NLR ratio	≤3.68	189	21.3		Ref	135	21.4		Ref
	>3.68	106	14.3	0.008	1.507 (1.113–2.041)	71	13.9	0.032	1.489 (1.035–2.142)
LDH (U/L)	≤250	178	20.9	0.000	Ref	114	25	0.002	Ref
	>250	117	16.4	0.016	1.447 (1.072–1.954)	92	16.4	0.002	1.77 (1.224–2.561)
CEA (ng/ml)	≤6.5	116	18.7	0.010	Ref	81	16.5	0.002	Ref
00/((19/11))	>6.5	179	18.4	0.562	0.914 (0.673–1.24)	125	20.8	0.368	0.847 (0.59–1.216)
Cyfra211 (ng/ml)	≤3.3	112	26.4	0.002	Ref	77	32.8	0.000	Ref
Cynaz i'r (ng/nn)	>3.3	183	15	<0.001	1.832 (1.33–2.523)	129	15	0.001	1.945 (1.317–2.871)
EGFR-mutant status	EGFR wildtype	107	11.7	<0.001	Ref	76	12.9	0.001	Ref
EGITI-Indiant status	EGFR mutation	188	23.5	<0.001	0.469 (0.346–0.635)	130	29.6	<0.001	0.451 (0.314–0.649)
Treatment regimens	EGFR-TKIs	169	23.9	<0.001	Ref	118	28.7	<0.001	Ref
freathent regimens	Chemotherapy	126	11.6	<0.001	2.386 (1.764–3.228)	88	13.1	<0.001	1.969 (1.377–2.818)
Ki-67	< 60%	186	21.3	<0.001	Ref	128	30.2	<0.001	Ref
NI-07	≥ 60%	109	12.1	<0.001	2.245 (1.646–3.063)	78	12.4	<0.001	2.451 (1.695–3.545)
KRAS mutant	Wildtype	284	19.2	<0.001	Ref	199	20	<0.001	2.431 (1.095–0.043) Ref
NHAS MULANI	Mutation	204 11	19.2	0.038	2.254 (1.047–4.851)	7	13	0.02	2.68 (1.166–6.163)
		117		0.030	2.234 (1.047–4.831) Ref	1	10	0.02	2.00 (1.100-0.103)
Ki-67+EGFR-TKIs	< 60% ≥60%	52	30.5 20.7	0.001	2.147 (1.347–3.421)				
Ki 67 i Chamatharani	—			0.001	· · · · · · · · · · · · · · · · · · ·				
Ki-67+Chemotherapy	< 60%	69 57	15.5	0.000	Ref				
	≥ 60%	57	9	0.002	1.963 (1.285–2.998)				
Ki-67+EGFR mutant	< 60%	125	30.2	0.001	Ref				
	≥ 60%	63	18.3	<0.001	2.328 (1.528–3.546)				
Ki-67+EGFR wildtype	< 60%	61	14.8	0.000	Ref				
	$\geq 60\%$	46	9.2	0.003	2.025 (1.272–3.224)				
Ki-67+KRAS	KRAS ^{wildtype} + Ki-67 low	182	22.4	< 0.001	Ref				
	KRAS ^{mutation} + Ki-67 low	4	6	0.001	5.291 (1.922–14.565)				
	KRAS ^{mutation} + Ki-67 ^{high}	7	15.1	0.223	2.055 (0.645-6.55)				
	KRAS ^{wildtype} + Ki-67 ^{high}	102	11.7	< 0.001	2.352 (1.713–3.228)				
Ki-67+EGFR	EGFR ^{mutation} + Ki-67 ^{low}	125	30.2	< 0.001	Ref				
	EGFR ^{wildtype} + Ki-67 ^{low}	61	14.8	<0.001	2.115 (1.415–3.161)				
	EGFR ^{mutation} + Ki-67 ^{high}	63	18.3	<0.001	2.226 (1.47–3.371)				
	EGFR ^{wildtype} + Ki-67 ^{high}	46	9.2	< 0.001	4.641 (2.991–7.202)				

ECOG PS, Eastern Cooperative Oncology Group performance status; EGFR, Epidermal growth factor receptor; PLR, platelet-to-lymphocyte ratio; NLR, neutrophil-to-lymphocyte ratio; LDH, lactate dehydrogenase; CEA, carcinoembryonic antigen; Cyfra211, cytokeratin 19 fragment; Ki-67, nuclear proliferation antigen 67; Ref, Reference.

Variable	Group	Primary cohort (mOS)			Training cohort (mOS)			
		HR	95%CI	P value	HR	95%CI	P value	
TNM stage	Recurrent	Ref		0.025				
	IIIB-C	2.308	1.185-4.497	0.014				
	IV	1.061	0.705-1.598	0.775				
Treatment Regimens	EGFR-TKIs	Ref						
	Chemotherapy	2.343	1.722-3.188	<0.001				
Age (years)	≤60				Ref			
	>60				1.572	1.083-2.284	0.017	
LDH (U/L)	≤250	Ref			Ref			
	>250	1.807	1.321-2.471	<0.001	1.516	1.029-2.234	0.035	
Cyfra211 (ng/ml)	≤3.3				Ref			
	>3.3				1.555	1.029-2.35	0.036	
Ki-67	< 60%	Ref			Ref		< 0.001	
	$\geq 60\%$	2.277	1.66-3.123	< 0.001	2.245	1.545-3.263	< 0.001	
EGFR-mutant status	EGFR wildtype				Ref			
	EGFR mutation				0.44	0.304-0.638	< 0.001	

TABLE 3 | Multivariate analysis of independent risk factors for first-line therapy in training cohort.

ECOG PS, Eastern Cooperative Oncology Group performance status; EGFR, Epidermal growth factor receptor; LDH, lactate dehydrogenase; Cyfra211, cytokeratin 19 fragment; Ki-67, nuclear proliferation antigen 67; Ref, Reference.

TABLE 4 | The short-term efficacy comparison among EGFR-TKIs and chemotherapy with Ki-67 in NSCLC.

RECIST	First-line	EGFR-TKIs	First-line ch	emotherapy
	Ki-67 < 60%	Ki-67 ≥ 60%	Ki-67 < 60%	Ki-67 ≥ 60%
CR	2 (1.2%)	0 (0%)	0 (0%)	1 (0.8%)
PR	55 (32.5%)	4 (2.4%)	17 (13.5%)	4 (3.2%)
SD	68 (40.2%)	17 (10.1%)	32 (25.4%)	9 (7.1%)
PD	11 (6.5%)	12 (7.1%)	25 (19.8%)	36 (28.6%)
ORR	57 (33.7%)	4 (2.4%)	17 (13.5%)	5 (3.9%)
DCR	125 (73.9%)	21 (12.4%)	51 (40.5%)	14 (14.3%)

CR, Complete Response; PR, Partial Response; SD, Stable Disease; PD, Progressive Disease; DCR (disease control rate) = (CR + PR + SD)/total cases * 100%, ORR (objective remission rate) = (CR + PR)/total cases * 100%. The short-term efficacy observed at 16 weeks.

and first-line treatment regardless of whether the patients are administered with EGFR-TKIs or chemotherapy. Moreover, high expression of Ki-67 was correlated with poor survival, DCR, and ORR, and this may be important in the prognosis of NSCLC patients with either wildtype or mutant EGFR.

The prevalence of LUADs with KRAS mutations in Asia is approximately 10–15% (34), and Johnson et al. (35) showed that the KRAS mutations are associated with poor survival than KRAS wildtype in advanced LUADs. Moreover, the relationship between Ki-67 and KRAS-mutant status in NSCLC, Woo et al. (36) showed that the expression of Ki-67 and the KRAS mutation status is significantly correlated with stage I LUAD. Individuals with higher expressions of Ki-67 and KRAS mutations are at a higher risk of postoperative recurrence than those with low expressions of Ki-67 and without KRAS mutations. Assessment of the correlation between Ki-67 and first-line therapeutic outcomes in advanced or postoperative-recurrent NSCLC patients with KRAS-mutant status, we interestingly found that the Ki-67 expression levels in the KRAS mutation group were higher than KRAS wildtype, and the OS of the KRAS-mutant group was shorter than in the KRAS-wildtype group. Subgroup analysis showed that the KRAS wildtype low Ki-67 expression subgroup had the longest survival time when compared to KRAS mutation low or high Ki-67 expression subgroups. Therefore, KRAS status and Ki-67 expression levels are potential prognostic markers for the prediction of the firstline treatment in advanced or postoperative-recurrent NSCLC.

Nomograms are universally used for predicting cancer risk, and IHC and clinic-pathological data can be used in the assessment of risk factors for predicting tumor survival outcomes. For instance, using the clinicopathological factors available before surgery, Guo et al. (19) built a nomogram for predicting axillary pathologic complete response (pCR) in hormone receptor-positive breast cancer. They found that the nomogram could accurately predict axillary pCR in patients with HR-positive disease. In addition, Keam et al. (37) used clinical



FIGURE 4 | Nomogram predicting the 12-, 24- or 30-month median survival outcomes for NSCLC patients with or without EGFR mutations and treated with first-line therapy. The prediction was done by adding the total points for all the indicators for each variable. For instance, the factor of Ki-67 was vertically downward to the scale labeled "Total Points" axis, and each indicator score was added in total points. Finally, those three factors get the total scores on the "Total Points" axis to the bottom scale and are used to predict the 12-, 24-, and 30-month overall survival probability. EGFR, epidermal growth factor receptor; NSCLC, non-small cell lung cancer.



features to establish a nomogram for predicting PFS outcomes in NSCLC patients following first-line or other forms of EGFR-TKIs treatment. Their findings showed that performance status, line of chemotherapy, response to EGFR-TKI, and bone metastasis can effectively predict the 6-, 12- and 18-month PFS for NSCLC patients. Most of the studies have focused on the association between clinical factors in NSCLC patients with EGFR mutations and the survival time (20, 21, 26) while few studies have assessed the correlation between Ki-67 and first-line therapeutic outcomes in NSCLC patients with EGFR-mutant status. In this study, 188 EGFR mutant and 107 wildtypes of NSCLC patients who had received either first-line EGFR-TKI or chemotherapy were enrolled. The patients were randomized into the training and validation cohorts at a ratio of 2:1, using the "rms" package in R. The nomogram showed that the EGFR mutation status and Ki-67 had the most significant contribution to OS in NSCLC. The calibration plots also showed that the predicted 12-, 24-, and 30-month OS probabilities were similar to the observed values. Overall, the study showed that the established nomogram, which was based on Ki-67 expression, could effectively predict the efficacy of first-line therapeutic options in NSCLC patients with either wildtype or mutant EGFR.

TABLE 5 | Representative studies on risk factors of Ki-67 with EGFR in NSCLC.

Author, journal, and year of publication	Cancers	Treatment regimens	Risk factors	Advantages	Limitations
Ni et al. (26)	postoperative recurrence in completely resected EGFR-mutant NSCLC	adjuvant chemotherapy or radiotherapy without EGFR-TKIs	Tumor size, Ki-67, CK20, and N stage	IHC of Ki-67and CK20 as a new independent risk factor influencing the prognosis of postoperative recurrence NSCLC, was identified and validated. A nomogram was developed and validated	The Ki-67 relationship between EGFR-TKIs and KRAS gene was not detected, and all the patients were postoperative recurrence NSCLC.
Zhu et al. (18)	surgically resected NSCLC patients	Unknown	Former or current smokers, pathological stage and mutant p53-or TTF-1 positive status	High Ki-67 expression was the independent predictor for the worst survival in NSCLC patients with wild-type EGFR	The EGFR-TKIs or chemoradiotherapy with Ki-67 was not analysis, and Ki-67 was not an independent predictor in overall NSCLC group. Not establishment a nomogram model to analyzed the Ki-67 prediction survival of NSCLC
Lin et al. (27)	NSCLC	Unknown	Tumor grade, the slope of the spectral CT	Tumor grade and the slope of the DESCT may be useful for predicting Ki-67 expression and the presence of EGFR mutation in NSCLC	The study only analyzed the DESCT prediction of Ki-67 expression and EGFR genes status, the correlation Ki-67 with EGFR gene were not detceted. The number of case was small and nomograms were not developed and validated
Niemiec et al. (28)	Squamous cell lung cancer (LCSC) patients	Unknown	EGFR labeling index, Ki-67 labeling index	The IHC of EGFR, Ki-67 were independent negative prognostic parameters influencing survivals of LCSC patients	The EGFR gene test was not detected and only by IHC, and the number of case was small. The patients were included only LCSC and were not developed and validated a nomograms

DESCT, dual-energy spectral computed tomography; NSCLC, advanced non-small cell lung cancer; LCSC, lung squamous cell carcinoma; EGFR-TKIs, Epidermal growth factor receptor tyrosine kinase inhibitor; IHC, immunohistochemistry.

There are some limitations to this study. First, this was a retrospectively study, and the relationship between Ki-67 expression levels and various adverse reactions was not assessed. In addition, we enrolled 295 NSCLC patients administered with TKIs or chemotherapy, instead of TKIs only, which led to the weakening of the predictive effect of Ki-67 for EGFR-TKIs outcomes. We also included 26 non-adenocarcinoma patients in this study, which limited statistical comparisons with adenocarcinoma patients. Moreover, the Ki-67 ROC curve of the cutoff point was determined using treatment regimens and the levels of sensitivity levels of 37.3% meant that many patients were false negative, which reduced our interest in this prognostic marker in NSCLC patients. Finally, this study was a singlecenter, small sample size, retrospective study, which may have led to produce selective bias. Therefore, a larger sample size, multi-centered clinical study assessing the association between Ki-67 expression levels and first-line EGFR-TKIs therapeutic outcomes should be performed in NSCLC patients with EGFR mutations. Moreover, the key clinical factors for identifying high-risk patients should be verified in large sample multicentered studies.

In conclusion, as a proliferation index, the expression levels of Ki-67 are correlated with KRAS- or EGFR mutations and the efficacy of first-line chemotherapy or EGFR-TKIs therapy in advanced or postoperative-recurrent NSCLC. Elevated Ki-67 expression levels are associated with a poor prognosis. The established nomogram based on Ki-67 expression can well predict the efficacy of first-line therapeutic outcomes in NSCLC patients. This is the first study to incorporate Ki-67 expression and built common clinic-pathological factors in the prediction of the survival outcomes after first-line treatment in NSCLC patients with EGFR mutation status and have demonstrated its clinical accuracy, which may help clinical doctors identify potential factors associated with clinical outcomes of first-line therapies or initiate early-intervention treatment regimens for high-risk patient

DATA AVAILABILITY STATEMENT

The original contributions presented in the study are included in the article/supplementary material, further inquiries can be directed to the corresponding author/s.

ETHICS STATEMENT

This study was approved by the Ethics Institution Committee of First Affiliated Hospital of Nanchang University, Jiangxi P.R. China. All patients provided written informed consent before receiving EGFR-TKIs or chemotherapy and compliance with the declaration of Helsinki was met. All the patients' data are kept confidential.

AUTHOR CONTRIBUTIONS

WG and MH were involved in collecting the data and followup of patients. WG, YR, LX, and MH were responsible for the conception and design of the study, assisted with the statistical analysis, and wrote the manuscript. CW, WW, and JM contributed to help data analysis and corrected the manuscript. All authors approved the manuscript prior to submission.

FUNDING

This study was supported by the National Natural Science Foundation of China (Grant No. 82060517 to LX, 81760448 to CW), the Science and technology project of Jiangxi Health

REFERENCES

- Bray F, Ferlay J, Soerjomataram I, Siegel RL, Torre LA, Jemal A. Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA Cancer J Clin.* (2018) 68:394–424. doi: 10.3322/caac.21492
- Siegel RL, Miller KD, Fuchs HE, Jemal A. Cancer statistics, 2021. CA Cancer J Clin. (2021) 71:7–33. doi: 10.3322/caac.21654
- Thongprasert S, Duffield E, Saijo N, Wu YL, Yang JC, Chu DT, et al. Health-related quality-of-life in a randomized phase III first-line study of gefitinib versus carboplatin/paclitaxel in clinically selected patients from Asia with advanced NSCLC (IPASS). J Thorac Oncol. (2011) 6:1872–80. doi: 10.1097/JTO.0b013e31822adaf7
- Rosell R, Carcereny E, Gervais R, Vergnenegre A, Massuti B, Felip E, et al. Erlotinib versus standard chemotherapy as first-line treatment for European patients with advanced EGFR mutation-positive non-small-cell lung cancer (EURTAC): a multicentre, open-label, randomised phase 3 trial. *Lancet Oncol.* (2012) 13:239–46. doi: 10.1016/S1470-2045(11)70393-X
- Sun JM, Ahn MJ, Choi YL, Ahn JS, Park K. Clinical implications of T790M mutation in patients with acquired resistance to EGFR tyrosine kinase inhibitors. *Lung Cancer*. (2013) 82:294–8. doi: 10.1016/j.lungcan.2013.08.023
- Thress KS, Paweletz CP, Felip E, Cho BC, Stetson D, Dougherty B, et al. Acquired EGFR C797S mutation mediates resistance to AZD9291 in nonsmall cell lung cancer harboring EGFR T790M. *Nat Med.* (2015) 21:560–2. doi: 10.1038/nm.3854
- Zhou B, Tang C, Li J. k-RAS mutation and resistance to epidermal growth factor receptor-tyrosine kinase inhibitor treatment in patients with nonsmall cell lung cancer. J Cancer Res Ther. (2017) 13:699–701. doi: 10.4103/jcrt.JCRT_468_17
- Kim SM, Kwon OJ, Hong YK, Kim JH, Solca F, Ha SJ, et al. Activation of IL-6R/JAK1/STAT3 signaling induces *de novo* resistance to irreversible EGFR inhibitors in non-small cell lung cancer with T790M resistance mutation. *Mol Cancer Ther.* (2012) 11:2254–64. doi: 10.1158/1535-7163.MCT-12-0311
- Guan R, Lin A, Luo P, Zhang J. Association between Circulating Inflammatory Proteins and Clinical Prognosis in Chinese Patients with EGFR Mutation-Positive Non-Small Cell Lung Cancer. J Thorac Oncol. (2019) 14:e222–4. doi: 10.1016/j.jtho.2019.05.027
- Castañón E, Rolfo C, Viñal D, López I, Fusco JP, Santisteban M, et al. Impact of epidermal growth factor receptor (EGFR) activating mutations and their targeted treatment in the prognosis of stage IV non-small cell lung cancer (NSCLC) patients harboring liver metastasis. *J Transl Med.* (2015) 13:257. doi: 10.1186/s12967-015-0622-x
- Tramm T, Kyndi M, Sørensen FB, Overgaard J, Alsner J. Influence of intra-tumoral heterogeneity on the evaluation of BCL2, E-cadherin, EGFR, EMMPRIN, and Ki-67 expression in tissue microarrays from breast cancer. *Acta Oncol.* (2018) 57:102–6. doi: 10.1080/0284186X.2017.1404128
- 12. Pascale M, Aversa C, Barbazza R, Marongiu B, Siracusano S, Stoffel F, et al. The proliferation marker Ki-67, but not neuroendocrine expression, is an independent factor in the prediction of prognosis of primary prostate cancer patients. *Radiol Oncol.* (2016) 50:313–20. doi: 10.1515/raon-2016-0033
- 13. Li Q, Ma J, Li H, Xu W, Cao Z, Liu S, et al. Correlation between uptake of 18F-FDG during PET/CT and Ki-67 expression in patients newly diagnosed

Committee (Grant No. 20203121 to WW), and Science and Technology plan of Jiangxi Administration of Traditional Chinese Medicine (Grant No. 2020B0365). The funding sources had no role in the data collection, analysis, or interpretation.

ACKNOWLEDGMENTS

We appreciate the effort of the physicians for patients and thank all the patients involved for allowing us to analyze their clinical data.

with multiple myeloma having extramedullary involvement. *Technol Cancer Res Treat.* (2019) 18:1533033819849067. doi: 10.1177/1533033819849067

- Guarneri V, Dieci MV, Bisagni G, Frassoldati A, Bianchi GV, De Salvo GL, et al. De-escalated therapy for HR+/HER2+ breast cancer patients with Ki-67 response after 2-week letrozole: results of the PerELISA neoadjuvant study. *Ann Oncol.* (2019) 30:921–6. doi: 10.1093/annonc/mdz055
- 15. Watanabe J. Reply to letters to the editor: Discordance in estrogen receptor and change of Ki-67 between primary site and metastatic site of recurrent breast cancer. *Breast Cancer Res Treat.* (2020) 182:513–4. doi: 10.1007/s10549-020-05695-2
- Warth A, Cortis J, Soltermann A, Meister M, Budczies J, Stenzinger A, et al. Tumour cell proliferation (Ki-67) in non-small cell lung cancer: a critical reappraisal of its prognostic role. *Br J Cancer*. (2014) 111:1222–9. doi: 10.1038/bjc.2014.402
- Del Gobbo A, Pellegrinelli A, Gaudioso G, Castellani M, Zito Marino F, Franco R, et al. Analysis of NSCLC tumour heterogeneity, proliferative and 18F-FDG PET indices reveals Ki-67 prognostic role in adenocarcinomas. *Histopathology*. (2016) 68:746–51. doi: 10.1111/his.12808
- Zhu WY, Hu XF, Fang KX, Kong QQ, Cui R, Li HF, et al. Prognostic value of mutant p53, Ki-67, and TTF-1 and their correlation with EGFR mutation in patients with non-small cell lung cancer. *Histol Histopathol.* (2019) 34:1269–78. doi: 10.14670/HH-18-124
- Guo R, Su Y, Si J, Xue J, Yang B, Zhang Q, et al. A nomogram for predicting axillary pathologic complete response in hormone receptor-positive breast cancer with cytologically proven axillary lymph node metastases. *Cancer*. (2020) 126:3819–29. doi: 10.1002/cncr.32830
- Deng J, Ren Z, Wen J, Wang B, Hou X, Xue Z, et al. Construction of a nomogram predicting the overall survival of patients with distantly metastatic non-small-cell lung cancer. *Cancer Manag Res.* (2018) 10:6143–56. doi: 10.2147/CMAR.S183878
- Yang Q, Zhang P, Wu R, Lu K, Zhou H. Identifying the best marker combination in CEA, CA125, CY211, NSE, and SCC for lung cancer screening by combining roc curve and logistic regression analyses: is it feasible? *Dis Markers*. (2018) 2018:2082840. doi: 10.1155/2018/2082840
- Xu J, Wang J, Zhang S. Mechanisms of resistance to irreversible epidermal growth factor receptor tyrosine kinase inhibitors and therapeutic strategies in non-small cell lung cancer. *Oncotarget.* (2017) 8:90557–78. doi: 10.18632/oncotarget.21164
- Girard N, Sima CS, Jackman DM, Sequist LV, Chen H, Yang JC, et al. Nomogram to predict the presence of EGFR activating mutation in lung adenocarcinoma. *Eur Respir J.* (2012) 39:366–72. doi: 10.1183/09031936.00010111
- 24. Seymour L, Le Teuff G, Brambilla E, Shepherd FA, Soria JC, Kratzke R, et al. LACE-bio: validation of predictive and/or prognostic immunohistochemistry/histochemistry-based biomarkers in resected non-small-cell lung cancer. *Clin Lung Cancer.* (2019) 20:66–73. doi: 10.1016/j.cllc.2018.10.001
- Deftereos G, Sandoval A, Furtado LV, Bronner M, Matynia AP. Successful lung cancer EGFR sequencing from DNA extracted from TTF-1 immunohistochemistry slides: a new means to extend insufficient tissue. *Hum Pathol.* (2020) 97:52–9. doi: 10.1016/j.humpath.2019. 12.009

- Ni J, Guo T, Li Y, Yang X, Li Y, Zou L, et al. Patterns and risks of postoperative recurrence in completely resected EGFR-mutant non-small cell lung cancer: prognostic significance of routine immunohistochemical markers. *Transl Lung Cancer Res.* (2019) 8:967–78. doi: 10.21037/tlcr.2019.12.02
- Lin L, Cheng J, Tang D, Zhang Y, Zhang F, Xu J, et al. The associations among quantitative spectral CT parameters, Ki-67 expression levels and EGFR mutation status in NSCLC. *Sci Rep.* (2020) 10:3436. doi: 10.1038/s41598-020-60445-0
- Niemiec J, Kolodziejski L, Dyczek S, EGFR LI. and Ki-67 LI are independent prognostic parameters influencing survivals of surgically treated squamous cell lung cancer patients. *Neoplasma*. (2005) 52:231–7.
- Xie S, Liu Y, Qiao X, Hua RX, Wang K, Shan XF, et al. What is the prognostic significance of Ki-67 positivity in oral squamous cell carcinoma? *J Cancer*. (2016) 7:758–67. doi: 10.7150/jca.14214
- Peng H, Tan X, Wang Y, Dai L, Liang G, Guo J, et al. Clinical significance of Ki-67 and circulating tumor cells with an epithelial-mesenchymal transition phenotype in non-small cell lung cancer. *Am J Transl Res.* (2020) 12:2916–28. doi: 10.2147/CMAR.S251271
- Mitchell KG, Parra ER, Nelson DB, Zhang J, Wistuba II, Fujimoto J, et al. Tumor cellular proliferation is associated with enhanced immune checkpoint expression in stage I non-small cell lung cancer. J Thorac Cardiovasc Surg. (2019) 158:911–9. doi: 10.1016/j.jtcvs.2019.04.084
- Jakobsen JN, Sørensen JB. Clinical impact of ki-67 labeling index in non-small cell lung cancer. Lung Cancer. (2013) 79:1–7. doi: 10.1016/j.lungcan.2012.10.008
- 33. Wei DM, Chen WJ, Meng RM, Zhao N, Zhang XY, Liao DY, et al. Augmented expression of Ki-67 is correlated with clinicopathological characteristics and prognosis for lung cancer patients: an up-dated systematic review and metaanalysis with 108 studies and 14,732 patients. *Respir Res.* (2018) 19:150. doi: 10.1186/s12931-018-0843-7
- 34. Yoshizawa A, Sumiyoshi S, Sonobe M, Kobayashi M, Fujimoto M, Kawakami F, et al. Validation of the IASLC/ATS/ERS lung adenocarcinoma classification for prognosis and association with EGFR and KRAS gene

mutations: analysis of 440 Japanese patients. J Thorac Oncol. (2013) 8:52-61. doi: 10.1097/JTO.0b013e3182769aa8

- Johnson ML, Sima CS, Chaft J, Paik PK, Pao W, Kris MG, et al. Association of KRAS and EGFR mutations with survival in patients with advanced lung adenocarcinomas. *Cancer.* (2013) 119:356–62. doi: 10.1002/cncr. 27730
- Woo T, Okudela K, Yazawa T, Wada N, Ogawa N, Ishiwa N, et al. Prognostic value of KRAS mutations and Ki-67 expression in stage I lung adenocarcinomas. *Lung Cancer.* (2009) 65:355–62. doi: 10.1016/j.lungcan.2008.11.020
- 37. Keam B, Kim DW, Park JH, Lee JO, Kim TM, Lee SH, et al. Nomogram predicting clinical outcomes in non-small cell lung cancer patients treated with epidermal growth factor receptor tyrosine kinase inhibitors. *Cancer Res Treat.* (2014) 46:323–30. doi: 10.4143/crt.2013.120

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's Note: All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Copyright © 2021 Gu, Hu, Xu, Ren, Mei, Wang and Wang. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.