



Systematically Displaying the Pathogenesis of Keratoconus *via* Multi-Level Related Gene Enrichment-Based Review

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Keratoconus (KC) is an etiologically heterogeneous corneal ectatic disorder. To systematically display the pathogenesis of keratoconus (KC), this study reviewed all the reported genes involved in KC, and performed an enrichment analysis of genes identified at the genome, transcription, and protein levels respectively. Combined analysis of multi-level results revealed their shared genes, gene ontology (GO), and pathway terms, to explore the possible pathogenesis of KC. After an initial search, 80 candidate genes, 2,933 transcriptional differential genes, and 947 differential proteins were collected. The candidate genes were significantly enriched in extracellular matrix (ECM) related terms, Wht signaling pathway and cytokine activities. The enriched GO/pathway terms of transcription and protein levels highlight the importance of ECM, cell adhesion, and inflammatory once again. Combined analysis of multi-levels identified 13 genes, 43 GOs, and 12 pathways. The pathogenic relationships among these overlapping factors maybe as follows. The gene mutations/variants caused insufficient protein dosage or abnormal function, together with environmental stimulation, leading to the related functions and pathways changes in the corneal cells. These included response to the glucocorticoid and reactive oxygen species; regulation of various signaling (P13K-AKT, MAPK and NF-kappaB), apoptosis and aging; upregulation of cytokines and collagen-related enzymes; and downregulation of collagen and other ECM-related proteins. These undoubtedly lead to a reduction of extracellular components and induction of cell apoptosis, resulting in the loosening and thinning of corneal tissue structure. This study, in addition to providing information about the genes involved, also provides an integrated insight into the gene-based etiology and pathogenesis of KC.

Keywords: keratoconus, candidate genes, multi-level combined analysis, gene enrichment, pathogenesis

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INTRODUCTION

Keratoconus (KC) is a complex multifactor degenerative disorder of the cornea, characterized by corneal ectasia, thinning, and cone-shaped protrusion, leading to reduced vision, irregular astigmatism, and corneal scarring (1-4). The worldwide prevalence of KC is approximately 1:2000 (4). KC usually manifests during puberty, and the clinical manifestation vary depending on disease severity (3). Myopia and astigmatism in one or both eyes were the main symptoms in the early stage. With disease progression, visual acuity of patients is progressive loss, and cannot be corrected with spectacles. KC at completion stage often has typical clinical sign, including Munson sign, a V-shape deformation of the lower eyelid in downward position; Fleischer ring, a hemosiderin arc or circle line around the cone base; Vogt's striae, fine vertical lines produced by Descemet's membrane compression (3, 4). In addition, the central or lower temporal part of the cornea shows obvious conical protrusion, and the central cornea becomes thinner obviously. In the completion stage, KC spontaneously or due to external forces such as eye rubbing, rupture of the posterior elastic layer of the cornea occurs, resulting in acute corneal edema and significant decline in visual acuity (3, 4). Because of the unclear pathogenesis and limited availability of medical treatments, KC has become a significant clinical problem worldwide and a leading indication for corneal transplantation (5).

Probing KC's etiology and pathogenesis and adopting effective control methods are the fundamentals of prevention and treatment of KC. KC has a clear genetic tendency. Genetic factors are involved in the development of KC (2, 6, 7). Until now, more than 70 candidate genes and regions have been screened and identified by genome-wide linkage analysis, wholeexome sequencing (WES), candidate gene sequencing, genomewide association study (GWAS), or candidate gene association study (7–90). Due to the genetic heterogeneity and population differences among KC patients, the genetic cause of most cases has not been effectively identified, and the pathogenesis underlying the genetic mutation remains unclear. This represents the current bottleneck in KC etiology research, so it is very important to find a breakthrough point to explore the key related genes, and the common pathogenesis, of KC.

Traditional genetic studies have typically focused on highquality families to map and identify new disease-causing genes or screen susceptible sites through population association analysis. However, the pathogenesis caused by mutations or susceptible sites is obscure, leading to the slow progress of pathogenesis studies of most genetic diseases, especially complex ones (91). The combined analysis of multi-levels can achieve the display of the candidate genes screened at the DNA level at the transcription level, the analysis of mutations in the genome at the RNA level for significantly differentially expressed genes, and the enrichment analysis of key genes in the pathway. Integrated genomics, transcription, and protein data can be leveraged to systematically analyze multiple consecutive events occurring in diseases, including changes in expression levels caused by gene mutations, and various forms of heterogeneity in transcriptional regulation, translation, and post-translational regulation body and feedback regulation. According to the changes in candidate factors at different levels, the candidate pathogenic factors can be thoroughly explored and the target of pathogenicity can be identified. Multiomics analysis can also be used to build a gene regulatory network in order to clarify the regulatory and causal relationships between various molecules, so as to gain a deeper understanding of the molecular mechanism and genetic basis of complex traits in genetic diseases.

In this analytical study, we examined all the reported genes involved in KC and performed an enrichment analysis of genes identified at the genome, transcription, and protein levels, respectively. In addition, by using gene set enrichment analysis, we attempted to explore the important mechanisms at different levels. Combined analysis of multi-level results revealed their shared genes, GOs, and pathways, allowing us to explore the possible pathogenesis of KC. The results of this study, in addition to providing information about the changed genes involved in the disease, provide an integrated insight into the common pathogenesis of KC.

MATERIALS AND METHODS

Literature Search to Find Relevant Genes

To find genes associated with KC, the literature was reviewed and data were collected manually. All the studies describing genome changes (including pathogenic mutations or susceptible variants) and differentially expressed encoding genes at the transcription and protein levels were scrutinized using the following keywords in the PubMed and Web of Science databases: "keratoconus" AND ("gene" OR "expression" OR "transcriptome"). We limited our search to articles published up to search date that were written in English. The search was conducted in November 2020. Then, each article was read and classified carefully. For the genome level, only those studies on patients with keratoconus were collected, excluding those on other syndromes patients with keratoconus, central corneal thickness and corneal curvature of normal person unless their results were confirmed in keratoconus samples. For the transcription and protein levels, only those human studies that directly used in situ KC corneal tissue or primary KC corneal cells were selected. Finally, we identified and recorded all the reported genes with pathogenic mutations or susceptible variants, differentially expressed genes at the transcription level, and differentially expressed or abnormally distributed genes at the protein level. This study did not search other databases, such as clinical trials and so. The relevant genes collected just included the genes published in the PubMed and Web of Science databases.

Enrichment Analysis

Enrichment analysis is a statistics-based method for classifying genes that are overrepresented in a specific set of genes. All the genes associated with KC were classified into three groups according to changed levels, and gene ontology (GO) and Kyoto Encyclopedia of Genes and Genomes (KEGG) pathway analysis were performed *via* the online Database for Annotation, Visualization, and Integrated Discovery (DAVID) software, version 6.8 (92, 93).

Combined Analysis of Multi-Levels

Combined analysis of multi-level enrichment revealed their shared GOs and pathways, allowing us to explore the possible pathogenesis of KC. To further identify the putative pathogenicity of gene mutations/susceptible variants and to detect the KC-related gene function and pathway changes, we conducted a combined analysis of the multi-level results. Online tools (http://bioinformatics.psb.ugent.be/webtools/ Venn/) were used to calculate and draw custom Venn diagrams depict the genes, GOs, and KEGG pathways shared by the multi-level results.

RESULTS

After an extensive review of resources, more than 200 studies were selected from the published articles and were reviewed in greater detail. Our study confirmed the 80 candidate genes, 2,933 differentially expressed genes at the transcription level, and 947 differentially expressed genes at the protein level.

Candidate Genes and GO/Pathway Enrichment at the Genomic Level

KC has a clear genetic tendency (2, 6, 7). In the past 20 years, scholars have extensively investigated the genetic cause of KC (7-90). Thus far, more than 40 candidate genes and regions have been located and screened via genome-wide linkage analysis, WES, or candidate gene sequencing. In addition, candidate gene association studies and GWAS have been carried out successively. Thus far, research has found that SNP (Single Nucleotide Polymorphism) loci of 30 genes are related to KC, which can increase or decrease the risk of KC. After an initial search, 80 candidate genes were collected for further analysis. As the results show, a few genes were identified by more than one type of analysis, which represents stronger evidence of their involvement in KC (Figure 1A). The top seven genes were COL5A1, MIR184, LOX, ZNF469, VSX1, COL4A3 and COL4A4 (Figure 1A). Detailed literature informations of reported KC associated genes at the genomic level were shown in Supplementary Table S1.

On the basis of the DAVID results, we tested whether the genes associated with KC clustered into certain GO terms and KEGG pathways. Figures 1B,C shows the significant GO terms and KEGG pathways with a p < 0.05. We found that the top-ranking GO terms were proteinaceous extracellular matrix (ECM) (GO: 0005578, $p = 6.00 \times 10^{-17}$), extracellular space (GO: 0005615, p $= 2.10 \times 10^{-10}$) and their related pathways in the cell component category, including extracellular region, collagen timer, basement membrane, extracellular exosome, as well as ECM (Figure 1B). The top five enriched biological processes were collagen catabolic process (GO: 0010033, $p = 5.00 \times 10^{-7}$), Wnt signaling pathway (GO: 0009611, $p = 2.60 \times 10^{-5}$), ECM organization (GO: 0033554, $p = 3.50 \times 10^{-5}$), canonical Wnt signaling pathway (GO: 0006916, $p = 4.20 \times 10^{-5}$), and negative regulation of fat cell differentiation (GO: 0010941, $p = 4.20 \times 10^{-5}$). The top molecular function terms most were involved in or related to various binding (frizzled binding, interleukin-1 receptor binding, collagen binding, transcription regulatory region DNA binding and ECM binding etc.), and activities of cytokine, receptor agonist, and metalloendopeptidase inhibitor.

The top-ranking KEGG pathways were associated with certain complex diseases, including cancer, type 1 diabetes mellitus, and graft-vs.-host disease (**Figure 1C**). These results suggested that KC might overlap with some of the pathogenesis of these diseases. The others top-ranking KEGG pathways were Wnt signaling pathway, Hippo signaling pathway, and Focal adhesion, which suggest that these pathways may play a role in the pathogenicity of KC.

Differential Genes and GO/Pathway Enrichment at the Transcription Level

Detailed literature information of differentially expressed encoding genes were shown in **Supplementary Table S2** (49, 90, 94–136). In addition to many studies that focused on particular genes or gene families, other studies investigated the transcriptome. Quantitative real-time PCR (qRT-PCR), RNA-seq and microarray were the main methods used in the differential expression studies (49, 90, 94–136). All the significantly differentially expressed encoding genes were collected regardless of whether the different corneal tissue types including corneal tissue, corneal epithelia, corneal stroma and primary stromal fibroblast.

After an initial search, a total of 2,933 reported differentially expressed encoding genes between KC corneas and normal corneas at the transcription level were collected (Figure 2A). There are 1,948 downregulated genes, 802 upregulated genes, and 183 genes with opposite results in different studies (Supplementary Table S2). About two-thirds of differentially expressed encoding genes were reported only once in single studies and need to be verified by further research. The remaining 933 genes were reported more than twice in the same or different corneal tissue types (Figure 2A). Among them, 13 genes, including GPNMB, TIMP3, CA12, CTGF, ID3, IGFBP3, JUN, PRELP, RGS5, SFRP1, SOD2, TIMP1 and VEGFA were reported in five or more studies (Figure 2A). All of these genes were reported significantly down regulated in KC corneal tissues, except for a few opposite results in individual studies. The changes in these genes indicate the decrease of growth factor (CTGF, IGFBP3 and VEGFA), transcription factor (ID3 and JUN), superoxide dismutase (SOD2), and metalloproteinase inhibitor (TIMP3 and TIMP1) in KC corneas. These results highlight the importance of these changes in the pathogenicity of KC.

The GO analysis of differentially expressed genes showed that there were 497 biological process, 84 cell component, and 127 molecular factor terms with a p < 0.05. The top ten significantly enriched biological process terms were involved in or related to ECM organization, regulation of cell proliferation, cell adhesion, angiogenesis, wound healing, response to drug and inflammatory (**Figure 2B**). The top ten significant cell component terms were involved in or related to proteinaceous ECM, ECM, extracellular exosome, extracellular space, focal adhesion, cell surface, plasma membrane, extracellular region, endoplasmic reticulum lumen,



FIGURE 1 | Reported candidate genes in keratoconus at the DNA level. (A) The associated genes identified by different analysis techniques. (B) Top ten enriched terms of each GO category at the DNA level. The three colors represent biological process (blue), cell component (green), and molecular function (yellow), respectively. (C) Top ten enriched KEGG pathways at the DNA level.

and membrane raft (Figure 2B). The top ten significant enriched molecular function terms were associated with various bindings, such as protein, heparin, collagen, integrin, extracellular matrix, fibronectin, actin, calcium ion, and glycoprotein bindings. These results further highlight the importance of extracellular region and ECM related proteins in the pathogenicity of KC. In addition, cell proliferation, cell adhesion angiogenesis and response to drug and inflammatory may be involved in the process of KC, which requires further study. The KEGG pathway analysis identified 86 enriched pathways. The top five significantly enriched KEGG pathways were pathways in cancer (hsa05200, $p = 3.20 \times 10^{-12}$), focal adhesion (hsa04510, $p = 6.50 \times 10^{-9}$), PI3K-Akt signaling pathway (hsa 04210, $p = 1.20 \times 10^{-8}$), MAPK signaling pathway (hsa04010, $p = 3.7 \times 10^{-8}$), and ECM-receptor interaction (hsa04512, $p = 4.60 \times 10^{-8}$) (Figure 2C). These results suggest that these pathways may play a role in the pathogenicity of KC.

Differential Genes and GO/Pathway Enrichment at the Protein Level

After an initial search, a total of 946 reported differentially expressed proteins between KC corneas and normal corneas were collected (98–103, 109, 114, 117, 118, 120, 125, 126, 128, 130, 131, 133, 137–193). Immunohistochemistry, immunofluorescence, and western blot were used for candidate protein level detection, and proteomic analysis was performed *via* mass spectrometry. All the reported differentially expressed proteins were collected, including downregulated, upregulated, and abnormally distributed proteins. There are 427 downregulated genes, 398 upregulated genes, 14 abnormally distributed proteins, and 107 genes with opposite results in different studies (**Supplementary Table S3**). About half differentially expressed genes were reported only once. The remaining 466 genes were reported more than twice in the same or different corneal tissue



according to the number and results of related studies. (B) Top 10 enriched terms of each GO category at the transcription level. The three colors represent biological process (blue), cell component (green), and molecular function (yellow), respectively. (C) Top ten enriched KEGG pathways at the transcription level.

types (Figure 3A). Among them, 30 genes were reported in four or more times, including five genes (*HSPB1*, *VIM*, *HMGA1*, *TSTD1*, *RUVBL2*) with upregulation and two genes (*BZW1*, *LOX*) with downregulation in at least four studies. The changes in these genes indicate the decrease of transcription factor (*BZW1*), lysyl oxidase (*LOX*), and increase of small heat shock protein (*HSPB1*), vimentins (*VIM*), thiosulfate sulfurtransferase (*TSTD1*) and ATPase (*RUVBL2*) in KC corneas. These results highlight the importance of these changes in the pathogenicity of KC. The other genes with conflicting results suggest that the pathogenesis of KC is very complex, and further research is needed to clarify the role of these genes. The GO analysis of differentially expressed proteins showed that there were 344 biological process, 127 cell component, and 98 molecular factor terms with a p < 0.05. The top ten significant enriched biological process terms were involved in or related to ECM disassembly, ECM organization, collagen catabolic process, proteolysis, cell-cell adhesion, platelet degranulation, complement activation, retina homeostasis, protein folding and translational initiation (**Figure 3B**). The top ten significant cell component terms were involved in or related to extracellular exosome, ECM, extracellular space, extracellular region, cell-cell adherens junction, focal adhesion, cytosol, membrane, cytoplasm, and blood microparticle (**Figure 3B**).



according to the number and results of related studies. (B) Top 10 enriched terms of each GO category at the protein level. The three colors represent biological process (blue), cell component (green), and molecular function (yellow), respectively. (C) Top ten enriched KEGG pathways at the protein level.

The top 10 significant enriched molecular function terms were associated various bindings (cadherin, RNA, protein, identical protein, protease and enzyme bindings), endopeptidase activity, protein homodimerization activity and electron carrier activity (**Figure 3B**). These results once again highlight the importance of collagen and ECM in the pathogenicity of KC at the protein level. Except for ECM and its related GO terms, the results showed that proteolysis, cell-cell adhesion, focal adhesion, protein binding, and endopeptidase may be involved in the process of KC (**Figure 3B**). The top five significant enriched KEGG pathways were Amoebiasis, Carbon metabolism, ECM-receptor interaction, Biosynthesis of antibiotics and Biosynthesis of amino acids (**Figure 3C**). These results provided further evidence of the

important role of ECM pathways in the pathogenicity of KC at the protein level.

Combined Analysis of Multi-Levels

To further identify the putative pathogenicity of gene variants and detect the KC-related gene function and pathway changes, we conducted a combined analysis of DNA, RNA, and protein level results. First, we analyzed the genes shared between the different levels. The results showed that there were 13 overlapping genes between all three levels (**Figure 4A**, **Table 1**). They were *COL4A3*, *COL6A2*, *MMP9*, *TIMP1*, *LOX*, *TGFBI*, *TNF*, *IL1A*, *IL1RN*, *SOD1*, *CAT*, *VSX1* and *TF*. All the genes except *TGFBI* and *SOD1* were reported to have KC-susceptibility SNPs. Potential



pathogenic mutations of seven overlapping genes (LOX, IL1RN, COL4A3, VSX1, TGFBI, SOD1 and COL6A2) were identified in KC patients. In addition, LOX and IL1RN were located in the susceptible loci detected by linkage analysis. Five of the 13 genes (LOX, IL1RN, VSX1, COL4A3, and TGFBI) were identified by multiple types of analysis at the DNA level. Three of the 13 genes (LOX, TIMP1, and TNF) were verified in multiple studies with consistent expression changes at the transcription level. Five of the 13 proteins (LOX, IL1RN, COL6A2, MMP9, and TNF) were verified in multiple studies with consistent changes at the protein level. These results suggested that these overlapping genes might be key genes of KC and might play an important role in the pathogenesis of KC. As the results show (Table 1), five genes (TNF, MMP9, IL1A, CAT, and VSX1) had significant upregulation, and six genes (TIMP1, COL6A2, SOD1, TGFBI, COL4A3, and LOX) had significant downregulation in KC at both the transcription and protein levels. The coincident changes of these genes also indicated the decrease of collagen (*COL4A3* and *COL6A2*), metallopeptidase inhibitor (*TIMP1*), lysyl oxidase (*LOX*), superoxide dismutase (*SOD1*), and the increase of metallopeptidase (*MMP9*), antioxidant enzyme (*CAT*) and inflammatory cytokines (*TNF* and *IL1A*). These results highlight the importance of these changes in the pathogenicity of KC.

Combined analysis of different levels' enrichment revealed their shared significant GOs and pathways, allowing the researchers to explore the possible pathogenesis of KC. The overlapping significant GOs between all three levels, including 24 biological process, 13 cell component, and six molecular function terms, are shown in **Figure 4B**. The shared biological process terms can divided into four groups, including various responserelated GOs (response to drug, glucocorticoid, hydrogen

Genes	Analysis techniques of DNA level	Changes of RNA level (N)	Changes of protein level (N
LOX	Linkage analysis; candidate gene mutation analysis; GWAS; candidate gene association studies	down (4)	down (4)
IL1RN	Linkage analysis; candidate gene association studies	down (1)	up (2)
COL4A3	Candidate gene mutation analysis; NGS; GWAS; candidate gene association studies	down (2)/up (1)	down (1)
VSX1	Candidate gene mutation analysis; NGS; candidate gene association studies	up (1)	up (1)
TGFBI	Candidate gene mutation analysis; NGS	down (1)	down (2) /up (1)
SOD1	Candidate gene mutation analysis	down (1)	down (1) /up (2)
COL6A2	NGS	down (1) /up (1)	down (3)
CAT	Candidate gene association studies	up (1)	up (1)
IL1A	Candidate gene association studies	down (3)/ up (1)	up (1)
MMP9	Candidate gene association studies	down (1) /up (3)	up (2)
TF	Candidate gene association studies	down (1)/up (1)	down (1)/up (3)
TIMP1	Candidate gene association studies	down (5)	down (1) /up (1)
TNF	Candidate gene association studies	up (3)	up (3)

TABLE 1 | Overlapping genes between multi-levels.

GWAS, Genome-Wide Association Studies; NGS, Next-Generation Sequencing; N, the number of studies.

The genes with consistent changes in transcription and protein levels are bold.

peroxide, reactive oxygen species, and hypoxia), apoptosisrelated GOs (activation of cysteine-type endopeptidase activity involved in apoptotic process, negative regulation of apoptotic process, and extrinsic apoptotic signaling pathway via death domain receptors), ECM-related GOs (ECM organization, collagen catabolic process, and collagen fibril organization) and activation and positive regulation of various signaling (MAPK activity, NF-kappaB activity, and I-kappaB kinase/NF-kappaB signaling). For the cell components, most of the overlapping GOs were ECM related (including basement membrane, ECM, proteinaceous ECM, collagen type IV trimer, and collagen trimer), extracellular related (extracellular region, extracellular space and extracellular exosome) and pinocytosis related (caveola and cytoplasmic vesicle). For the molecular functions, the overlapping GOs were ECM related (ECM structural constituent and collagen binding), and various binding-related GOs (protein, identical protein, protease and chromatin DNA binding). The combined analysis of the KEGG pathway showed that Protein digestion and absorption, Focal adhesion, ECM-receptor interaction, PI3K-Akt signaling pathway, apoptosis, and various diseases related pathways (including cancer, Prion diseases, Malaria, Amoebiasis and Chagas disease) were significantly enriched at DNA, RNA, and protein levels (Figure 4C). Most GO and pathway enrichments shared at DNA, RNA, and protein levels were related to collagen, ECM, extracellular, various responses and apoptosis, suggesting that these GO and pathway changes might have been etiological-serving as mechanisms of KC.

DISCUSSION

KC is an etiologically heterogeneous corneal ectatic disorder, and both environmental and genetic factors play a role in

its etiopathogenesis (194). Based on results from studies that have investigated the genetic etiology, expression, and translation changes in the process of development, it is becoming increasingly clear that KC is a complex disease with a complex etiology or convergence of multiple disease pathways. However, the common pathogenesis underlying the different etiologies remains unclear. In this study, we reviewed all the studies of KCrelated genes identified at the genome, transcription, and protein levels. Through multi-level related gene enrichment-based review, we systematically explored the schematic representing factors responsible for KC at different levels. The results of this study, in addition to providing information about the genes involved in the disease, clearly provide an integrated insight into the gene-based etiology and pathogenesis of KC.

Genetic changes play an important role in the etiopathogenesis of KC (2, 6, 7, 194). Many forms of gene variation, such as inheritance gene mutation, de novo mutation, and polymorphism, have been reported to be involved in the etiology of KC (7-90). More than half of the genes were reported in one type of study or in single studies. A few genes, encoded chains of collagens (COL5A1, COL4A3, and COL4A4) (57, 59, 61, 67, 72, 76, 78, 84, 87, 88, 90), collagen cross-linking enzyme (LOX) (18, 30, 90), factor for the synthesis or organization of collagen fibers (ZNF469) (27, 32, 34, 46, 47, 62, 65), and others (MIR184 and VSX1) (6, 7, 16, 19, 24, 40, 56, 82, 85) were identified in different types of studies, such as pathogenic mutation analysis, polymorphism association analysis, and family-based linkage analysis. However, the occurrence rate of these gene mutations in the population was relatively low, and in many populations, it could not even be verified (25, 195-202), which suggested that KC is genetically heterogeneous. Among the reported KC associated genetic changes, there were 11 genes responsible for apoptosis related

process (FAS, FASLG, TNF, BIRC2, SMAD3, WNT5A, CAT, TIMP1, MMP9, FOXO1, and COL4A3), 14 reported corneal biomechanics loci (MPDZ, COL6A2, MYOF, LOX, ZNF469, SMAD3, NFIB, FNDC3B, COL5A1, WNT10A, TGFBI, SLC4A11, FOXO1 and COL4A3) (203), and five genes responsible for inflammatory processes (IL1A, IL1B, FAS, TNF and IL17B). Genetic changes in these genes might lead to the changes in related functions and pathways in the corneal cells, then lead to induction of apoptosis, inflammatory and altered biomechanics of cornea, which have been reported involved in the etiopathogenesis of KC (204-206), and lead to the occurrence of KC. The top candidate gene based enrichment, including ECM and their related pathways (76, 90), Wnt signaling pathway (60), and cytokine activities (204), have been reported involved in the etiopathogenesis of KC. The role of negative regulation of fat cell differentiation in the etiopathogenesis of KC has not been reported, but body mass index was reported associated with keratoconus before (207). So, more studies are needed on the relationship between this functional change in order to clarify its relationship and mechanism of action in KC.

Differences between the expression of genes in normal and KC corneas suggested disease pathways. The top 13 verified differentially expressed genes at RNA level indicated a decrease of growth factor, transcription factor, superoxide dismutase, and metalloproteinase inhibitor, which highlights the importance of these changes in the pathogenicity of KC. The top differentially expressed genes based enrichment were similar to the GOs at the genomic level, mainly including ECM and its related GO terms, response to inflammatory and various bindings. These results represent further confirmation of the importance of collagen (61, 67, 75, 78, 84, 87, 88, 113, 117, 125, 163-165), ECM (76, 90, 97, 158), cell adhesion (72, 97), and inflammatory (49, 101, 124, 128, 139, 204) in the pathogenicity of KC. In addition, cell proliferation, angiogenesis, and response to drug have not been reported before, which may be involved in the process of KC, and should be investigated further. For reported differentially expressed proteins, the genes with upregulation or downregulation in at least four studies highlight the importance of transcription factor (98, 106, 112, 128, 145, 160), lysyl oxidase (18, 30, 81, 125, 150, 151), small heat shock protein (156, 188, 193), vimentins (126, 156, 188, 193), thiosulfate sulfurtransferase and ATPase (188) in KC corneas. Gene-based enrichment analysis showed that the differentially expressed proteins were significantly enriched in ECM and its related GO terms, proteolysis, and various bindings. These results once again highlight the importance of these GO terms in the pathogenicity of KC at the protein level.

Different approaches have been used to investigate and define the phenotype, mechanisms, and causes of KC. Thousands of genes were identified at genomic, transcription, and protein levels. Observations of corneal changes that occur in KC often do not distinguish between primary changes and secondary inflammatory or degenerative effects. Although research has identified many differences that distinguish KC corneas from normal corneas, it has not been possible to trace these changes back to primary causes or to identify the triggers that precipitate the cascade of events that leads to the clinical picture of KC. The results at different levels were clearly similar. In order to explore the key points, combined analysis of multi-levels was performed. Integrated genomics, transcription, and protein data can be leveraged to systematically analyze multiple consecutive events occurring in diseases. According to the changes in candidate factors at different levels, the candidate pathogenic factors can be thoroughly explored and the target of pathogenicity can be identified. The consistent changes in these factors at different levels suggested that these factors play an important role in the pathogenesis of KC. The DNA, RNA, and protein changes represented the cause and process changes of KC, respectively.

Based on the results of multi-level combined analysis, we hypothesized that the pathogenic relationships among these related factors is as follows. The etiology of KC can be divided into environmental and genetic factors. The environmental factors may include endogenous and exogenous factors, such as glucocorticoid, hydrogen peroxide, and reactive oxygen species (ROS) (120). The gene mutations or variants involved in collagen (57, 61, 64, 67, 75, 76, 78, 88), metallopeptidase inhibitor (75, 81, 109, 157), lysyl oxidase (18, 30, 81, 90), metallopeptidase (64), antioxidant enzyme (16, 25, 81, 199), inflammatory cytokines (11, 45, 83, 201), and others cause insufficient protein dosage or abnormal function. These genetic changes, together with the aforementioned stimulation, lead to the changes in related functions and pathways in the corneal cells. The related functions include the response to the stimulation of hormones and reactive oxygen species (96, 106, 120, 121, 189), activation and positive regulation of various signaling (MAPK activity, NF-kappaB activity, and I-kappaB kinase/NF-kappaB signaling) (123, 128), upregulation of cytokines and collagen-related enzymes (49, 101, 122, 124, 147, 174, 187), and downregulation of collagen, collagen-crossing, and other ECM-related proteins (97, 103, 117, 163, 164), and regulation of apoptosis (36, 175, 186). These undoubtedly lead to the reduction of extracellular components and the induction of apoptosis and aging. The change in extracellular structure, decrease of extracellular composition, and apoptosis of corneal cells all lead to the loosening and thinning of corneal tissue structure, which leads to the occurrence of KC.

In addition to the different levels and combined analysis results of this paper, our hypothesis was supported by many other studies of molecular mechanisms and cell events of KC. A few hormones and substances have been reported to be associated with KC (208–214). However, the relationship between glucocorticoid and KC has not been studied before, and should be investigated further. Chronic inflammatory events were detected in the tears of KC patients (215-217). A significant increase in the apoptosis of KC cells has been reported in several studies (186, 218, 219). Moreover, a decrease of dulfated epitopes of keratan sulfate KC corneas was also reported (220). The electron microscope results of KC showed that the content of the stroma increases, whereas the fibril diameter is reduced, the mean diameter and interfibrillar spacing of collagen fibrils are reduced, and the collagen fibrils and proteoglycans number density and area fractions are significantly increased (138).

Our study has some limitations. All the results were obtained through multi-level related gene enrichment-based analysis. More studies are needed on the relationship between these

functional changes in order to clarify their relationship and mechanism of action, which could provide a new direction for the treatment of KC. For the expression studies collected in this study, there were several different corneal tissue types. Most of the studies used the corneal tissue, and a few studies used the corneal epithelia, corneal stroma or primary stromal fibroblast (see Supplementary Tables S1-S3). In this analysisbased study, because of the limited space, instead of categorizing different genes detected in different tissues, we just conducted a unified analysis in different levels. Analysis that is more detailed needs to be carried out in the future to find the role of different corneal cells in KC pathogenesis. Furthermore, the interaction between genetic factors and environmental factors in the pathogenesis of KC has not been effectively solved, and further research is needed. Epigenetic mechanisms might help explain environmental contributions to the pathogenesis of KC (221). There are few studies on the relationship between epigenetic changes and KC (60). Recently, certain epigenetic changes, such as circle RNA, have been confirmed to play an important role in other diseases having overlapped pathogenesis pathway with KC (222-225), suggesting its potential role in KC pathogenesis. Study the role of these epigenetic changes might be a new research direction of KC in future.

Conclusions

Keratoconus is an etiologically heterogeneous corneal ectatic disorder, and both environmental and genetic factors play a role in its etiopathogenesis. Based on results from studies that have investigated the genetic etiology, expression, and translation changes in the process of development, it is becoming increasingly clear that KC is a complex disease with a complex etiology or convergence of multiple disease pathways. The common pathogenesis underlying the different etiologies remains unclear. In this study, we reviewed all the studies of KCrelated genes identified at the genome, transcription, and protein levels. Through multi-level related gene enrichment-based review, we systematically explored the schematic representing factors responsible for KC at different levels. The results of this study, in addition to providing information about the genes involved in the disease, clearly provide an integrated insight into the gene-based etiology and pathogenesis of KC. Base on the results, we hypothesized that the pathogenic relationships among these related factors is as follows. The gene mutations/variants caused insufficient protein dosage or abnormal function, together with environmental stimulation, leading to the changes in the related functions and pathways in the corneal cells. These included response to the glucocorticoid and reactive oxygen species; regulation of various signaling (P13K-AKT, MAPK and NF-kappaB), apoptosis and aging; upregulation of cytokines

REFERENCES

- Rabinowitz YS. Keratoconus. Surv Ophthalmol. (1998) 42:297–319. doi: 10.1016/S0039-6257(97)00119-7
- Mas Tur V, MacGregor C, Jayaswal R, O'Brart D, Maycock N, A. review of keratoconus: Diagnosis, pathophysiology, and genetics. *Surv Ophthalmol.* (2017) 62:770–83. doi: 10.1016/j.survophthal.2017.06.009

and collagen-related enzymes; and downregulation of collagen and other ECM-related proteins. These undoubtedly lead to a reduction of extracellular components and induction of cell apoptosis, resulting in the loosening and thinning of corneal tissue structure. This hypothesis was supported by many other studies of molecular mechanisms and cell events of KC. More studies are needed on the relationship between these functional changes in order to clarify their relationship and mechanism of action, which could provide a new direction for the treatment of KC.

DATA AVAILABILITY STATEMENT

The original contributions presented in the study are included in the article/**Supplementary Material**, further inquiries can be directed to the corresponding authors.

AUTHOR CONTRIBUTIONS

X-DH and HG designed the research. W-HX, CS, YL, and Z-XZ performed the literature search. X-DH analyzed the data, participated in the discussion, and wrote and revised the paper. KW and P-FL participated in the revision of the paper.

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SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fmed. 2021.770138/full#supplementary-material

Supplementary Table S1 | Reported candidate genes in keratoconus at the DNA level.

Supplementary Table S2 | The gene lists of reported differential expressed genes between KC and normal cornea at the transcription level.

Supplementary Table S3 | The gene lists of reported differential expressed genes between KC and normal cornea at the protein level.

- Keratology Group OBoCA. Expert consensus on diagnosis and treatment of keratoconus in China (2019). Chinese Journal of Ophthalmology. (2019) 55:891-5.
- Ferrari G, Rama P. The keratoconus enigma: A review with emphasis on pathogenesis. Ocul Surf. (2020) 18:363–73. doi: 10.1016/j.jtos.2020.03.006
- 5. Rahman I, Carley F, Hillarby C, Brahma A, Tullo AB. Penetrating keratoplasty: indications, outcomes, and

complications. *Eye* (*Lond*). (2009) 23:1288–94. doi: 10.1038/eye. 2008.305

- Karolak JA, Gajecka M. Genomic strategies to understand causes of keratoconus. *Mol Genet Genomics*. (2017) 292:251–69. doi: 10.1007/s00438-016-1283-z
- Valgaeren H, Koppen C, Van Camp G, A. new perspective on the genetics of keratoconus: why have we not been more successful? *Ophthalmic Genet*. (2018) 39:158–74. doi: 10.1080/13816810.2017.1393831
- Burdon KP, Vincent AL. Insights into keratoconus from a genetic perspective. *Clin Exp Optom.* (2013) 96:146–54. doi: 10.1111/cxo.12024
- Nielsen K, Hjortdal J, Pihlmann M, Corydon TJ. Update on the keratoconus genetics. *Acta Ophthalmol.* (2013) 91:106–13. doi: 10.1111/j.1755-3768.2012.02400.x
- Rabinowitz YS, Galvis V, Tello A, Rueda D, Garcia JD. Genetics vs chronic corneal mechanical trauma in the etiology of keratoconus. *Exp Eye Res.* (2021) 202:108328. doi: 10.1016/j.exer.2020.108328
- Kim SH, Mok JW, Kim HS, Joo CK. Association of-31T>C and-511 C>T polymorphisms in the interleukin 1 beta (IL1B) promoter in Korean keratoconus patients. *Mol Vis*. (2008) 14:2109–16.
- Guan T, Ma ZW, Ding SP. [Analyses of coding sequence point mutation and polymorphism of TGFBI gene in Chinese patients with keratoconus]. *Zhonghua Yi Xue Yi Chuan Xue Za Zhi.* (2011) 28:152–5. doi: 10.3760/cma.j.issn.1003-9406.2011.02.007
- Pathak D, Nayak B, Singh M, Sharma N, Tandon R, Sinha R, et al. Mitochondrial complex 1 gene analysis in keratoconus. *Mol Vis.* (2011) 17:1514–25.
- 14. Li X, Bykhovskaya Y, Haritunians T, Siscovick D, Aldave A, Szczotka-Flynn L, et al. A genome-wide association study identifies a potential novel gene locus for keratoconus, one of the commonest causes for corneal transplantation in developed countries. *Hum Mol Genet.* (2012) 21:421–9. doi: 10.1093/hmg/ddr460
- Czugala M, Karolak JA, Nowak DM, Polakowski P, Pitarque J, Molinari A, et al. Novel mutation and three other sequence variants segregating with phenotype at keratoconus 13q32 susceptibility locus. *Eur J Hum Genet*. (2012) 20:389–97. doi: 10.1038/ejhg.2011.203
- Saee-Rad S, Hashemi H, Miraftab M, Noori-Daloii MR, Chaleshtori MH, Raoofian R, et al. Mutation analysis of VSX1 and SOD1 in Iranian patients with keratoconus. *Mol Vis.* (2011) 17:3128–36.
- Guan T, Liu C, Ma Z, Ding S. The point mutation and polymorphism in keratoconus candidate gene TGFBI in Chinese population. *Gene.* (2012) 503:137–9. doi: 10.1016/j.gene.2012.04.061
- Bykhovskaya Y, Li X, Epifantseva I, Haritunians T, Siscovick D, Aldave A, et al. Variation in the lysyl oxidase (LOX) gene is associated with keratoconus in family-based and case-control studies. *Invest Ophthalmol Vis Sci.* (2012) 53:4152–7. doi: 10.1167/iovs.11-9268
- Wang Y, Jin T, Zhang X, Wei W, Cui Y, Geng T, et al. Common single nucleotide polymorphisms and keratoconus in the Han Chinese population. *Ophthalmic Genet.* (2013) 34:160–6. doi: 10.3109/13816810.2012.743569
- Lu Y, Vitart V, Burdon KP, Khor CC, Bykhovskaya Y, Mirshahi A, et al. Genome-wide association analyses identify multiple loci associated with central corneal thickness and keratoconus. *Nat Genet.* (2013) 45:155–63.
- Li X, Bykhovskaya Y, Tang YG, Picornell Y, Haritunians T, Aldave AJ, et al. An association between the calpastatin (CAST) gene and keratoconus. *Cornea*. (2013) 32:696–701. doi: 10.1097/ICO.0b013e3182821c1c
- 22. Mikami T, Meguro A, Teshigawara T, Takeuchi M, Uemoto R, Kawagoe T, et al. Interleukin 1 beta promoter polymorphism is associated with keratoconus in a Japanese population. *Mol Vis.* (2013) 19:845–51.
- Bae HA, Mills RA, Lindsay RG, Phillips T, Coster DJ, Mitchell P, et al. Replication and meta-analysis of candidate loci identified variation at RAB3GAP1 associated with keratoconus. *Invest Ophthalmol Vis Sci.* (2013) 54:5132–5. doi: 10.1167/iovs.13-12377
- Lechner J, Bae HA, Guduric-Fuchs J, Rice A, Govindarajan G, Siddiqui S, et al. Mutational analysis of MIR184 in sporadic keratoconus and myopia. *Invest Ophthalmol Vis Sci.* (2013) 54:5266–72. doi: 10.1167/iovs.13-12035
- Moschos MM, Kokolakis N, Gazouli M, Chatziralli IP, Droutsas D, Anagnou NP, et al. Polymorphism Analysis of VSX1 and SOD1 Genes in Greek Patients with Keratoconus. *Ophthalmic Genet.* (2015) 36:213–7. doi: 10.3109/13816810.2013.843712

- Synowiec E, Wojcik KA, Izdebska J, Binczyk E, Blasiak J, Szaflik J, et al. Polymorphisms of the homologous recombination gene RAD51 in keratoconus and Fuchs endothelial corneal dystrophy. *Dis Markers*. (2013) 35:353–62. doi: 10.1155/2013/851817
- 27. Sahebjada S, Schache M, Richardson AJ, Snibson G, MacGregor S, Daniell M, et al. Evaluating the association between keratoconus and the corneal thickness genes in an independent Australian population. *Invest Ophthalmol Vis Sci.* (2013) 54:8224–8. doi: 10.1167/iovs.13-12982
- Wojcik KA, Synowiec E, Jimenez-Garcia MP, Kaminska A, Polakowski P, Blasiak J, et al. Polymorphism of the transferrin gene in eye diseases: keratoconus and Fuchs endothelial corneal dystrophy. *Biomed Res Int.* (2013) 2013:247438. doi: 10.1155/2013/247438
- Sahebjada S, Schache M, Richardson AJ, Snibson G, Daniell M, Baird PN. Association of the hepatocyte growth factor gene with keratoconus in an Australian population. *PLoS ONE.* (2014) 9:e84067. doi: 10.1371/journal.pone.0084067
- Hasanian-Langroudi F, Saravani R, Validad MH, Bahari G, Yari D. Association of Lysyl oxidase (LOX) Polymorphisms with the risk of keratoconus in an Iranian population. *Ophthalmic Genet.* (2015) 36:309–14. doi: 10.3109/13816810.2014.881507
- Rho CR, Park JH, Jung YH, Kim MS, A. case of concomitant keratoconus and granular corneal dystrophy type II. *Cont Lens Anterior Eye.* (2014) 37:314–6. doi: 10.1016/j.clae.2014.02.001
- Lechner J, Porter LF, Rice A, Vitart V, Armstrong DJ, Schorderet DF, et al. Enrichment of pathogenic alleles in the brittle cornea gene, ZNF469, in keratoconus. *Hum Mol Genet*. (2014) 23:5527–35. doi: 10.1093/hmg/ddu253
- 33. Karolak JA, Polakowski P, Szaflik J, Szaflik JP, Gajecka M. Molecular Screening of Keratoconus Susceptibility Sequence Variants in VSX1, TGFBI, DOCK9, STK24, and IPO5 Genes in Polish Patients and Novel TGFBI Variant Identification. *Ophthalmic Genet.* (2016) 37:37–43. doi: 10.3109/13816810.2014.926375
- Vincent AL, Jordan CA, Cadzow MJ, Merriman TR, McGhee CN. Mutations in the zinc finger protein gene, ZNF469, contribute to the pathogenesis of keratoconus. *Invest Ophthalmol Vis Sci.* (2014) 55:5629–35. doi: 10.1167/iovs.14-14532
- Wojcik KA, Synowiec E, Polakowski P, Glowacki S, Izdebska J, Lloyd S, et al. Polymorphism of the flap endonuclease 1 gene in keratoconus and Fuchs endothelial corneal dystrophy. *Int J Mol Sci.* (2014) 15:14786–802. doi: 10.3390/ijms150814786
- 36. Synowiec E, Wojcik KA, Izdebska J, Blasiak J, Szaflik J, Szaflik JP. Polymorphisms of the apoptosis-related FAS and FAS ligand genes in keratoconus and Fuchs endothelial corneal dystrophy. *Tohoku J Exp Med.* (2014) 234:17–27. doi: 10.1620/tjem.234.17
- Wojcik KA, Synowiec E, Sobierajczyk K, Izdebska J, Blasiak J, Szaflik J, et al. Polymorphism of the DNA base excision repair genes in keratoconus. *Int J Mol Sci.* (2014) 15:19682–99. doi: 10.3390/ijms151119682
- Hao XD, Chen P, Chen ZL Li SX, Wang Y. Evaluating the association between keratoconus and reported genetic loci in a han Chinese population. *Ophthalmic Genet.* (2015) 36:132–6. doi: 10.3109/13816810.2015.1005317
- Synowiec E, Wojcik KA, Izdebska J, Binczyk E, Szaflik J, Blasiak J, et al. Polymorphism of the LIG3 gene in keratoconus and Fuchs endothelial corneal dystrophy. *Cell Mol Biol (Noisy-le-grand)*. (2015) 61:56–63.
- Shetty R, Nuijts RM, Nanaiah SG, Anandula VR, Ghosh A, Jayadev C, et al. Two novel missense substitutions in the VSX1 gene: clinical and genetic analysis of families with Keratoconus from India. *BMC Med Genet.* (2015) 16:33. doi: 10.1186/s12881-015-0178-x
- Cuellar-Partida G, Springelkamp H, Lucas SE, Yazar S, Hewitt AW, Iglesias AI, et al. WNT10A exonic variant increases the risk of keratoconus by decreasing corneal thickness. *Hum Mol Genet.* (2015) 24:5060–8. doi: 10.1093/hmg/ddv211
- 42. Wang Y, Wei W, Zhang C, Zhang X, Liu M, Zhu X, et al. Association of interleukin-1 gene single nucleotide polymorphisms with keratoconus in Chinese han population. *Curr Eye Res.* (2016) 41:630–5. doi: 10.3109/02713683.2015.1045083
- 43. Wojcik KA, Synowiec E, Kaminska A, Izdebska J, Polakowski P, Pawlowska E, et al. Polymorphism of the APEX nuclease 1 gene in keratoconus and Fuchs endothelial corneal dystrophy. *Cell Mol Biol Lett.* (2015) 20:48–65. doi: 10.1515/cmble-2015-0001

- 44. Karolak JA, Rydzanicz M, Ginter-Matuszewska B, Pitarque JA, Molinari A, Bejjani BA, et al. Variant c.2262A>C in DOCK9 Leads to Exon Skipping in Keratoconus Family. *Invest Ophthalmol Vis Sci.* (2015) 56:7687–90. doi: 10.1167/iovs.15-17538
- Karolak JA, Gambin T, Pitarque JA, Molinari A, Jhangiani S, Stankiewicz P. et al. Variants in SKP1, PROB1, and IL17B genes at keratoconus 5q311-q353 susceptibility locus identified by whole-exome sequencing. *Eur J Hum Genet*. (2017) 25:73–8. doi: 10.1038/ejhg.2016.130
- Yu X, Chen B, Zhang X, Shentu X. Identification of seven novel ZNF469 mutations in keratoconus patients in a Han Chinese population. *Mol Vis.* (2017) 23:296–305.
- Yildiz E, Bardak H, Gunay M, Bardak Y, Imamoglu S, Ozbas H, et al. Novel Zinc Finger Protein Gene 469 (ZNF469) Variants in Advanced Keratoconus. *Curr Eye Res.* (2017) 42:1396–400. doi: 10.1080/02713683.2017.1325910
- Rong SS, Ma STU Yu XT, Ma L, Chu WK, Chan TCY, et al. Genetic associations for keratoconus: a systematic review and meta-analysis. *Sci Rep.* (2017) 7:4620. doi: 10.1038/s41598-017-04393-2
- Arbab M, Tahir S, Niazi MK, Ishaq M, Hussain A, Siddique PM, et al. TNFalpha genetic predisposition and higher expression of inflammatory pathway components in keratoconus. *Invest Ophthalmol Vis Sci.* (2017) 58:3481–7. doi: 10.1167/iovs.16-21400
- Guan T, Wang X, Zheng LB, Wu HJ, Yao YF. Analysis of the VSX1 gene in sporadic keratoconus patients from China. *BMC Ophthalmol.* (2017) 17:173. doi: 10.1186/s12886-017-0567-3
- Hao XD, Chen P, Zhang YY Li SX, Shi WY, Gao H, De. novo mutations of TUBA3D are associated with keratoconus. *Sci Rep.* (2017) 7:13570. doi: 10.1038/s41598-017-13162-0
- Bykhovskaya Y, Fardaei M, Khaled ML, Nejabat M, Salouti R, Dastsooz H, et al. TSC1 mutations in keratoconus patients with or without tuberous sclerosis. *Invest Ophthalmol Vis Sci.* (2017) 58:6462–9. doi: 10.1167/iovs.17-22819
- Zhang J, Wu D, Li Y, Fan Y, Chen H, Xu J. Evaluating the association between calpastatin (CAST) gene and keratoconus in the Han Chinese population. *Gene.* (2018) 653:10–3. doi: 10.1016/j.gene.2018.02.016
- 54. Guan T, Wu HJ, Zhang LJ, Xu DJ, Zheng LB, Yao YF. [A novel VSX1 gene mutation identified in a sporadic keratoconus patient from China]. *Zhonghua Yan Ke Za Zhi.* (2018) 54:212–7. doi: 10.3760/cma.j.issn.0412-4081.2018.03.012
- 55. Wang YM, Ma L, Lu SY, Chan TCY, Yam JCS, Tang SM, et al. Analysis of multiple genetic loci reveals MPDZ-NF1B rs1324183 as a putative genetic marker for keratoconus. Br J Ophthalmol. (2018) 102:1736–41. doi: 10.1136/bjophthalmol-2018-312218
- da Silva DC, Gadelha BNB, Feitosa AFB, da Silva RG, Albuquerque T, Santos D, et al. Analysis of VSX1 variations in brazilian subjects with keratoconus. J Ophthalmic Vis Res. (2018) 13:266–73. doi: 10.4103/jovr.jovr_116_17
- Zhang J, Wu D, Dai Y, Xu J. Functional relevance for central cornea thickness-associated genetic variants by using integrative analyses. *BioData Min.* (2018) 11:19. doi: 10.1186/s13040-018-0179-3
- Yari D, Saravani R, Saravani S, Ebrahimian K, Galavi HR. Genetic polymorphisms of catalase and glutathione peroxidase-1 in keratoconus. *Iran J Public Health.* (2018) 47:1567–74.
- Skorodumova LO, Belodedova AV, Sharova EI, Malyugin BE. [Search for genetic markers for precise diagnostics of keratoconus]. *Biomed Khim.* (2019) 65:9–20. doi: 10.18097/PBMC20196501009
- Kabza M, Karolak JA, Rydzanicz M, Udziela M, Gasperowicz P, Ploski R, et al. Multiple Differentially Methylated Regions Specific to Keratoconus Explain Known Keratoconus Linkage Loci. *Invest Ophthalmol Vis Sci.* (2019) 60:1501–9. doi: 10.1167/iovs.18-25916
- Sargazi S, Moudi M, Heidari Nia M, Saravani R, Malek Raisi H. Association of KIF26B and COL4A4 gene polymorphisms with the risk of keratoconus in a sample of Iranian population. *Int Ophthalmol.* (2019) 39:2621–8. doi: 10.1007/s10792-019-01111-x
- Zhang W, Margines JB, Jacobs DS, Rabinowitz YS, Hanser EM, Chauhan T, et al. Corneal perforation after corneal cross-linking in keratoconus associated with potentially pathogenic ZNF469 mutations. *Cornea.* (2019) 38:1033–9. doi: 10.1097/ICO.00000000002002

- Froukh T, Hawwari A. Autosomal recessive non-syndromic keratoconus: homozygous frameshift variant in the candidate novel gene GALNT14. *Curr Mol Med.* (2019) 19:683–7. doi: 10.2174/1566524019666190730095630
- Abdullah OA, El Gazzar WB, Salem TI, Al-Kamil EA. Role of extracellular matrix remodelling gene SNPs in keratoconus. *Br J Biomed Sci.* (2020) 77:13–8. doi: 10.1080/09674845.2019.1654346
- Magalhaes OA, Kowalski TW, Wachholz GE, Schuler-Faccini L. Whole-exome sequencing in familial keratoconus: the challenges of a genetically complex disorder. *Arq Bras Oftalmol.* (2019) 82:453–9. doi: 10.5935/0004-2749.20190087
- Ilhan A, Altun S, Durukan I, Yolcu U, Erdem U. The association between genetic polymorphism of glutathione peroxidase 1 (rs1050450) and keratoconus in a Turkish population. *Arq Bras Oftalmol.* (2019) 82:501–6. doi: 10.5935/0004-2749.20190102
- Lin Q, Zheng L, Shen Z, Jie L, A. Novel Splice-Site Variation in COL5A1 Causes Keratoconus in an Indian Family. J Ophthalmol. (2019) 2019:2851380. doi: 10.1155/2019/2851380
- Cao K, Sahebjada S, Richardson AJ, Baird PN. Do age-related macular degeneration genes show association with keratoconus? *Eye Vis (Lond)*. (2019) 6:38. doi: 10.1186/s40662-019-0164-z
- 69. Khaled ML, Bykhovskaya Y, Gu C, Liu A, Drewry MD, Chen Z, et al. PPIP5K2 and PCSK1 are candidate genetic contributors to familial keratoconus. *Sci Rep.* (2019) 9:19406. doi: 10.1038/s41598-019-55866-5
- McComish BJ, Sahebjada S, Bykhovskaya Y, Willoughby CE, Richardson AJ, Tenen A, et al. Association of genetic variation with keratoconus. JAMA Ophthalmol. (2020) 138:174–81. doi: 10.1001/jamaophthalmol.2019.5293
- Abdul-Maksoud RS, Fouad RA, Elsayed TG, Ibrahem RA, Badawi AE. The impact of catalase and glutathione peroxidase-1 genetic polymorphisms on their enzyme activities among Egyptian patients with keratoconus. *J Gene Med.* (2020) 22:e3192. doi: 10.1002/jgm.3192
- 72. Karolak JA, Gambin T, Rydzanicz M, Polakowski P, Ploski R, Szaflik JP, et al. Accumulation of sequence variants in genes of Wnt signaling and focal adhesion pathways in human corneas further explains their involvement in keratoconus. *PeerJ.* (2020) 8:e8982. doi: 10.7717/peerj.8982
- Awd-Allah NA, Ismail SM, Salah El-Dine MM, Mohammed MM. Association between POLG and XRCC1 gene polymorphisms and keratoconus occurrence among Egyptian patients. Arch Soc Esp Oftalmol (Engl Ed). (2020) 95:439–46. doi: 10.1016/j.oftale.2020.03.002
- Hosoda Y, Miyake M, Meguro A, Tabara Y, Iwai S, Ueda-Arakawa N, et al. Keratoconus-susceptibility gene identification by corneal thickness genomewide association study and artificial intelligence IBM Watson. *Commun Biol.* (2020) 3:410. doi: 10.1038/s42003-020-01137-3
- 75. Yari D, Ehsanbakhsh Z, Validad MH, Langroudi FH. Association of TIMP-1 and COL4A4 gene polymorphisms with keratoconus in an Iranian population. J Ophthalmic Vis Res. (2020) 15:299–307. doi: 10.18502/jovr.v15i3.7448
- Hao XD, Chen XN, Zhang YY, Chen P, Wei C, Shi WY, et al. Multi-level consistent changes of the ECM pathway identified in a typical keratoconus twin's family by multi-omics analysis. *Orphanet J Rare Dis.* (2020) 15:227. doi: 10.1186/s13023-020-01512-7
- 77. Froukh T, Hawwari A, Al Zubi K. Whole exome sequencing highlights variants in association with Keratoconus in Jordanian families. *BMC Med Genet.* (2020) 21:177. doi: 10.1186/s12881-020-01112-z
- Abdelghany AA, Toraih EA, Abdelaziz EZ, El-Sherbeeny NA, Fawzy MS. Association of collagen gene (COL4A3) rs55703767 variant with response to riboflavin/ultraviolet a-induced collagen crosslinking in female patients with keratoconus. *Cornea.* (2021) 40:88–98. doi: 10.1097/ICO.00000000002489
- Droitcourt C, Touboul D, Ged C, Ezzedine K, Cario-Andre M, de Verneuil H, et al. A prospective study of filaggrin null mutations in keratoconus patients with or without atopic disorders. *Dermatology*. (2011) 222:336–41. doi: 10.1159/000328408
- Rathi VM, Vemuganti GK, Sangwan VS, Kannabiran C. Late occurrence of granular dystrophy in bilateral keratoconus: penetrating keratoplasty and long-term follow-up. *Indian J Ophthalmol.* (2011) 59:398–400. doi: 10.4103/0301-4738.83624

- De Bonis P, Laborante A, Pizzicoli C, Stallone R, Barbano R, Longo C, et al. Mutational screening of VSX1, SPARC, SOD1, LOX, and TIMP3 in keratoconus. *Mol Vis.* (2011) 17:2482–94.
- Hughes AE, Bradley DT, Campbell M, Lechner J, Dash DP, Simpson DA, et al. Mutation altering the miR-184 seed region causes familial keratoconus with cataract. *Am J Hum Genet*. (2011) 89:628–33. doi: 10.1016/j.ajhg.2011. 09.014
- Nowak DM, Karolak JA, Kubiak J, Gut M, Pitarque JA, Molinari A, et al. Substitution at IL1RN and deletion at SLC4A11 segregating with phenotype in familial keratoconus. *Invest Ophthalmol Vis Sci.* (2013) 54:2207–15. doi: 10.1167/iovs.13-11592
- 84. Li X, Bykhovskaya Y, Canedo AL, Haritunians T, Siscovick D, Aldave AJ, et al. Genetic association of COL5A1 variants in keratoconus patients suggests a complex connection between corneal thinning and keratoconus. *Invest Ophthalmol Vis Sci.* (2013) 54:2696–704. doi: 10.1167/iovs.13-11601
- Vincent AL, Jordan C, Sheck L, Niederer R, Patel DV, McGhee CN. Screening the visual system homeobox 1 gene in keratoconus and posterior polymorphous dystrophy cohorts identifies a novel variant. *Mol Vis.* (2013) 19:852–60.
- Dehkordi FA, Rashki A, Bagheri N, Chaleshtori MH, Memarzadeh E, Salehi A, et al. Study of VSX1 mutations in patients with keratoconus in southwest Iran using PCR-single-strand conformation polymorphism/heteroduplex analysis and sequencing method. *Acta Cytol.* (2013) 57:646–51. doi: 10.1159/000353297
- Kokolakis NS, Gazouli M, Chatziralli IP, Koutsandrea C, Gatzioufas Z, Peponis VG, et al. Polymorphism analysis of COL4A3 and COL4A4 genes in Greek patients with keratoconus. *Ophthalmic Genet.* (2014) 35:226–8. doi: 10.3109/13816810.2014.946055
- Saravani R, Hasanian-Langroudi F, Validad MH, Yari D, Bahari G, Faramarzi M, et al. Evaluation of possible relationship between COL4A4 gene polymorphisms and risk of keratoconus. *Cornea.* (2015) 34:318–22. doi: 10.1097/ICO.00000000000356
- Bardak H, Gunay M, Yildiz E, Bardak Y, Gunay B, Ozbas H, et al. Novel visual system homeobox 1 gene mutations in Turkish patients with keratoconus. *Genet Mol Res.* (2016) 15. doi: 10.4238/gmr15049024
- Xu X, Zhang X, Cui Y, Yang H, Ping X, Wu J, et al. Three novel variants identified within ECM-related genes in Chinese Han keratoconus patients. *Sci Rep.* (2020) 10:5844. doi: 10.1038/s41598-020-62572-0
- Jackson M, Marks L, May GHW, Wilson JB. The genetic basis of disease. Essays Biochem. (2018) 62:643–723. doi: 10.1042/EBC20170053
- Huang da W, Sherman BT, Lempicki RA. Systematic and integrative analysis of large gene lists using DAVID bioinformatics resources. *Nat Protoc.* (2009) 4:44–57. doi: 10.1038/nprot.2008.211
- Huang da W, Sherman BT, Lempicki RA. Bioinformatics enrichment tools: paths toward the comprehensive functional analysis of large gene lists. *Nucleic Acids Res.* (2009) 37:1–13. doi: 10.1093/nar/gkn923
- Sarker-Nag A, Hutcheon AE, Karamichos D. Mitochondrial Profile and Responses to TGF-beta Ligands in Keratoconus. *Curr Eye Res.* (2016) 41:900– 7. doi: 10.3109/02713683.2015.1078361
- 95. Atilano SR, Lee DH, Fukuhara PS, Chwa M, Nesburn AB, Udar N, et al. Corneal oxidative damage in keratoconus cells due to decreased oxidant elimination from modified expression levels of SOD enzymes, PRDX6, SCARA3, CPSF3, and FOXM1. J Ophthalmic Vis Res. (2019) 14:62–70. doi: 10.4103/jovr.jovr_80_18
- 96. Ayan B, Yuksel N, Carhan A, Gumuskaya Ocal B, Akcay E, Cagil N, et al. Evaluation estrogen, progesteron and androgen receptor expressions in corneal epithelium in keratoconus. *Cont Lens Anterior Eye.* (2019) 42:492–6. doi: 10.1016/j.clae.2018.11.015
- Bykhovskaya Y, Gromova A, Makarenkova HP, Rabinowitz YS. Abnormal regulation of extracellular matrix and adhesion molecules in corneas of patients with keratoconus. *Int J Keratoconus Ectatic Corneal Dis.* (2016) 5:63–70. doi: 10.5005/jp-journals-10025-1123
- Chiambaretta F, Nakamura H, De Graeve F, Sakai H, Marceau G, Maruyama Y, et al. Kruppel-like factor 6 (KLF6) affects the promoter activity of the alpha1-proteinase inhibitor gene. *Invest Ophthalmol Vis Sci.* (2006) 47:582– 90. doi: 10.1167/iovs.05-0551
- 99. Chiplunkar S, Chamblis K, Chwa M, Rosenberg S, Kenney MC, Brown DJ. Enhanced expression of a transmembrane phosphotyrosine phosphatase

(LAR) in keratoconus cultures and corneas. *Exp Eye Res.* (1999) 68:283–93. doi: 10.1006/exer.1998.0604

- Chung ES, Lee KH, Kim M, Chang EJ, Chung TY, Kim EK, et al. Expression of neurotrophic factors and their receptors in keratoconic cornea. *Curr Eye Res.* (2013) 38:743–50. doi: 10.3109/02713683.2013.774421
- 101. Du G, Liu C, Li X, Chen W, He R, Wang X, et al. Induction of matrix metalloproteinase-1 by tumor necrosis factor-alpha is mediated by interleukin-6 in cultured fibroblasts of keratoconus. *Exp Biol Med* (*Maywood*). (2016) 241:2033–41. doi: 10.1177/1535370216650940
- 102. Engler C, Chakravarti S, Doyle J, Eberhart CG, Meng H, Stark WJ, et al. Transforming growth factor-beta signaling pathway activation in Keratoconus. *Am J Ophthalmol.* (2011) 151:752–9 e2. doi: 10.1016/j.ajo.2010.11.008
- 103. Garcia B, Garcia-Suarez O, Merayo-Lloves J, Alcalde I, Alfonso JF, Fernandez-Vega Cueto L, et al. Differential expression of proteoglycans by corneal stromal cells in keratoconus. *Invest Ophthalmol Vis Sci.* (2016) 57:2618–28. doi: 10.1167/iovs.15-16692
- 104. Garcia B, Garcia-Suarez O, Merayo-Lloves J, Ferrara G, Alcalde I, Gonzalez J, et al. Heparanase overexpresses in keratoconic cornea and tears depending on the pathologic grade. *Dis Markers.* (2017) 2017:3502386. doi: 10.1155/2017/3502386
- 105. Ha NT, Nakayasu K, Murakami A, Ishidoh K, Kanai A. Microarray analysis identified differentially expressed genes in keratocytes from keratoconus patients. *Curr Eye Res.* (2004) 28:373–9. doi: 10.1080/02713680490502201
- 106. Hao XD, Chen ZL, Qu ML, Zhao XW Li SX, Chen P. Decreased Integrity, Content, and Increased Transcript Level of Mitochondrial DNA Are Associated with Keratoconus. *PLoS ONE.* (2016) 11:e0165580. doi: 10.1371/journal.pone.0165580
- 107. Joseph R, Srivastava OP, Pfister RR. Downregulation of beta-actin gene and human antigen R in human keratoconus. *Invest Ophthalmol Vis Sci.* (2012) 53:4032–41. doi: 10.1167/iovs.11-9062
- Kanai A. [The pathogenesis and treatment of corneal disorders]. Nippon Ganka Gakkai Zasshi. (2002) 106:757–76.
- 109. Kenney MC, Chwa M, Atilano SR, Tran A, Carballo M, Saghizadeh M, et al. Increased levels of catalase and cathepsin V/L2 but decreased TIMP-1 in keratoconus corneas: evidence that oxidative stress plays a role in this disorder. *Invest Ophthalmol Vis Sci.* (2005) 46:823–32. doi: 10.1167/iovs.04-0549
- 110. Khaled ML, Bykhovskaya Y, Yablonski SER Li H, Drewry MD, Aboobakar IF, et al. Differential Expression of Coding and Long Noncoding RNAs in Keratoconus-Affected Corneas. *Invest Ophthalmol Vis Sci.* (2018) 59:2717– 28. doi: 10.1167/iovs.18-24267
- 111. Lee JE, Oum BS, Choi HY, Lee SU, Lee JS. Evaluation of differentially expressed genes identified in keratoconus. *Mol Vis.* (2009) 15:2480–7.
- 112. Mace M, Galiacy SD, Erraud A, Mejia JE, Etchevers H, Allouche M, et al. Comparative transcriptome and network biology analyses demonstrate antiproliferative and hyperapoptotic phenotypes in human keratoconus corneas. *Invest Ophthalmol Vis Sci.* (2011) 52:6181–91. doi: 10.1167/iovs.10-70981
- 113. Kabza M, Karolak JA, Rydzanicz M, Szczesniak MW, Nowak DM, Ginter-Matuszewska B, et al. Collagen synthesis disruption and downregulation of core elements of TGF-beta, Hippo, and Wnt pathways in keratoconus corneas. *Eur J Hum Genet*. (2017) 25:582–90. doi: 10.1038/ejhg.2017.4
- Mootha VV, Kanoff JM, Shankardas J, Dimitrijevich S. Marked reduction of alcohol dehydrogenase in keratoconus corneal fibroblasts. *Mol Vis.* (2009) 15:706–12.
- 115. Nielsen K, Birkenkamp-Demtroder K, Ehlers N, Orntoft TF. Identification of differentially expressed genes in keratoconus epithelium analyzed on microarrays. *Invest Ophthalmol Vis Sci.* (2003) 44:2466–76. doi: 10.1167/iovs.02-0671
- 116. Pahuja N, Kumar NR, Shroff R, Shetty R, Nuijts RM, Ghosh A, et al. Differential molecular expression of extracellular matrix and inflammatory genes at the corneal cone apex drives focal weakening in keratoconus. *Invest Ophthalmol Vis Sci.* (2016) 57:5372–82. doi: 10.1167/iovs. 16-19677
- Peters DP, Harrison DA, Brandt CR. Heterogeneity of type I collagen expression in human corneal keratoconus fibroblasts. *Ophthalmic Res.* (1993) 25:273–9. doi: 10.1159/000267325

- Priyadarsini S, McKay TB, Sarker-Nag A, Karamichos D. Keratoconus in vitro and the key players of the TGF-beta pathway. *Mol Vis.* (2015) 21:577-88.
- Rabinowitz YS, Dong L, Wistow G. Gene expression profile studies of human keratoconus cornea for NEIBank: a novel cornea-expressed gene and the absence of transcripts for aquaporin 5. *Invest Ophthalmol Vis Sci.* (2005) 46:1239–46. doi: 10.1167/iovs.04-1148
- 120. Shetty R, Sharma A, Pahuja N, Chevour P, Padmajan N, Dhamodaran K, et al. Oxidative stress induces dysregulated autophagy in corneal epithelium of keratoconus patients. *PLoS ONE*. (2017) 12:e0184628. doi: 10.1371/journal.pone.0184628
- 121. Saee-Rad S, Raoofian R, Mahbod M, Miraftab M, Mojarrad M, Asgari S, et al. Analysis of superoxide dismutase 1, dual-specificity phosphatase 1, and transforming growth factor, beta 1 genes expression in keratoconic and non-keratoconic corneas. *Mol Vis.* (2013) 19:2501–7.
- 122. Saghizadeh M, Chwa M, Aoki A, Lin B, Pirouzmanesh A, Brown DJ, et al. Altered expression of growth factors and cytokines in keratoconus, bullous keratopathy and diabetic human corneas. *Exp Eye Res.* (2001) 73:179–89. doi: 10.1006/exer.2001.1028
- 123. Sharif R, Khaled ML, McKay TB, Liu Y, Karamichos D. Transcriptional profiling of corneal stromal cells derived from patients with keratoconus. *Sci Rep.* (2019) 9:12567. doi: 10.1038/s41598-019-48983-8
- 124. Shetty R, Ghosh A, Lim RR, Subramani M, Mihir K, Reshma AR, et al. Elevated expression of matrix metalloproteinase-9 and inflammatory cytokines in keratoconus patients is inhibited by cyclosporine A. *Invest Ophthalmol Vis Sci.* (2015) 56:738–50. doi: 10.1167/iovs.14-14831
- 125. Shetty R, Sathyanarayanamoorthy A, Ramachandra RA, Arora V, Ghosh A, Srivatsa PR, et al. Attenuation of lysyl oxidase and collagen gene expression in keratoconus patient corneal epithelium corresponds to disease severity. *Mol Vis.* (2015) 21:12–25.
- 126. Shetty R, Vunnava KP, Dhamodaran K, Matalia H, Murali S, Jayadev C, et al. Characterization of Corneal Epithelial Cells in Keratoconus. *Transl Vis Sci Technol.* (2019) 8:2. doi: 10.1167/tvst.8.1.2
- 127. Shinde V, Hu N, Mahale A, Maiti G, Daoud Y, Eberhart CG, et al. RNA sequencing of corneas from two keratoconus patient groups identifies potential biomarkers and decreased NRF2-antioxidant responses. *Sci Rep.* (2020) 10:9907. doi: 10.1038/s41598-020-66735-x
- Stachon T, Latta L, Kolev K, Seitz B, Langenbucher A, Szentmary N. [Increased NF-kappaB and iNOS Expression in Keratoconus Keratocytes -Hints for an Inflammatory Component?]. *Klin Monbl Augenheilkd*. (2019) 238:1010–7. doi: 10.1055/a-1002-0100
- 129. Stachs O, Bochert A, Gerber T, Koczan D, Thiessen HJ, Guthoff RF. [The extracellular matrix structure in keratoconus]. Ophthalmologe. (2004) 101:384–9. doi: 10.1007/s00347-003-0902-3
- 130. Sutton G, Madigan M, Roufas A, McAvoy J. Secreted frizzled-related protein 1 (SFRP1) is highly upregulated in keratoconus epithelium: a novel finding highlighting a new potential focus for keratoconus research and treatment. *Clin Exp Ophthalmol.* (2010) 38:43–8. doi: 10.1111/j.1442-9071.2009.02216.x
- Wang YN, Liu XN, Wang XD, Yin Y, Chen Y, Xiao XH, et al. Expression of visual system homeobox 1 in human keratoconus. *Int J Ophthalmol.* (2019) 12:201–6. doi: 10.18240/ijo.2019.02.03
- Wentz-Hunter K, Cheng EL, Ueda J, Sugar J, Yue BY. Keratocan expression is increased in the stroma of keratoconus corneas. *Mol Med.* (2001) 7:470–7. doi: 10.1007/BF03401852
- 133. Whitelock RB, Fukuchi T, Zhou L, Twining SS, Sugar J, Feder RS, et al. Cathepsin G, acid phosphatase, and alpha 1-proteinase inhibitor messenger RNA levels in keratoconus corneas. *Invest Ophthalmol Vis Sci.* (1997) 38:529–34.
- 134. You J, Corley SM, Wen L, Hodge C, Hollhumer R, Madigan MC, et al. RNA-Seq analysis and comparison of corneal epithelium in keratoconus and myopia patients. *Sci Rep.* (2018) 8:389. doi: 10.1038/s41598-017-18480-x
- 135. Zhang LY, Zou LH. [Study on enhanced leukocyte antigen-related tyrosine phosphatase in keratoconus]. *Zhonghua Yan Ke Za Zhi.* (2005) 41:234–8.
- 136. Zhao G, Wang C, Sun W, Zhang W, Li Y, Sheng H, et al. The expression of protein betaig-h3 inducible by transforming growth factor-beta in keratoconus and normal cornea. *Zhonghua Yan Ke Za Zhi.* (2002) 38:419–21.
- 137. Akhtar S, Bron AJ, Hayes AJ, Meek KM, Caterson B. Role of keratan sulphate (sulphated poly -N-acetyllactosamine repeats) in keratoconic

cornea, histochemical, and ultrastructural analysis. *Graefes Arch Clin Exp Ophthalmol.* (2011) 249:413–20. doi: 10.1007/s00417-010-1512-9

- Akhtar S, Bron AJ, Salvi SM, Hawksworth NR, Tuft SJ, Meek KM. Ultrastructural analysis of collagen fibrils and proteoglycans in keratoconus. *Acta Ophthalmol.* (2008) 86:764–72. doi: 10.1111/j.1755-3768.2007.01142.x
- Bosnar D, Dekaris I, Gabric N, Markotic A, Lazic R, Spoljaric N. Influence of interleukin-1alpha and tumor necrosis factor-alpha production on corneal graft survival. *Croat Med J.* (2006) 47:59–66.
- 140. Brookes NH, Loh IP, Clover GM, Poole CA, Sherwin T. Involvement of corneal nerves in the progression of keratoconus. *Exp Eye Res.* (2003) 77:515–24. doi: 10.1016/S0014-4835(03)00148-9
- 141. Bureau J, Pouliquen Y, Lorans G. [Fibrocyte reaction to interleukin 1 stimulation in keratoconus. Original title: Fibrocyte response to interleukin 1 stimulation in keratoconus]. *Klin Monbl Augenheilkd.* (1993) 203:269–74. doi: 10.1055/s-2008-1045679
- 142. Bystrom B, Carracedo S, Behndig A, Gullberg D, Pedrosa-Domellof F. Alpha11 integrin in the human cornea: importance in development and disease. *Invest Ophthalmol Vis Sci.* (2009) 50:5044–53. doi: 10.1167/iovs.08-3261
- 143. Caglayan M, Kocamis SI, Sarac O, Tatli Dogan H, Kosekahya P, Ayan M, et al. Investigation of Heme Oxygenase 2 Enzyme Protein Expression in Keratoconus and Normal Human Corneal Epithelium: An Immunohistochemical Study. *Curr Eye Res.* (2019) 44:25–9. doi: 10.1080/02713683.2018.1521980
- 144. Chaerkady R, Shao H, Scott SG, Pandey A, Jun AS, Chakravarti S. The keratoconus corneal proteome: loss of epithelial integrity and stromal degeneration. *J Proteomics.* (2013) 87:122–31. doi: 10.1016/j.jprot.2013.05.023
- 145. Cheng EL Li Y, Sugar J, Yue BY. Cell density regulated expression of transcription factor Sp1 in corneal stromal cultures. *Exp Eye Res.* (2001) 73:17–24. doi: 10.1006/exer.2001.1014
- 146. Cheung IM, McGhee C, Sherwin T. Deficient repair regulatory response to injury in keratoconic stromal cells. *Clin Exp Optom.* (2014) 97:234–9. doi: 10.1111/cxo.12118
- 147. Collier SA, Madigan MC, Penfold PL. Expression of membranetype 1 matrix metalloproteinase (MT1-MMP) and MMP-2 in normal and keratoconus corneas. *Curr Eye Res.* (2000) 21:662–8. doi: 10.1076/0271-3683(200008)2121-VFT662
- 148. Collier SA. Is the corneal degradation in keratoconus caused by matrix-metalloproteinases? *Clin Exp Ophthalmol.* (2001) 29:340–4. doi: 10.1046/j.1442-9071.2001.d01-17.x
- 149. Andrade FEC, Covre JL, Ramos L, Hazarbassanov RM, Santos MSD, Campos M, et al. Evaluation of galectin-1 and galectin-3 as prospective biomarkers in keratoconus. Br J Ophthalmol. (2018) 102:700–7. doi: 10.1136/bjophthalmol-2017-311495
- Dudakova L, Liskova P, Trojek T, Palos M, Kalasova S, Jirsova K. Changes in lysyl oxidase (LOX) distribution and its decreased activity in keratoconus corneas. *Exp Eye Res.* (2012) 104:74–81. doi: 10.1016/j.exer.2012. 09.005
- 151. Dudakova L, Sasaki T, Liskova P, Palos M, Jirsova K. The presence of lysyl oxidase-like enzymes in human control and keratoconic corneas. *Histol Histopathol.* (2016) 31:63–71.
- 152. Gatzioufas Z, Charalambous P, Thanos S. Reduced expression of the gap junction protein Connexin 43 in keratoconus. *Eye (Lond).* (2008) 22:294–9. doi: 10.1038/sj.eye.6702972
- Greene CA, Kuo C, Sherwin T. Aberrant Patterns of Key Epithelial Basement Membrane Components in Keratoconus. *Cornea*. (2017) 36:1549– 55. doi: 10.1097/ICO.00000000001393
- 154. Hasby EA, Saad HA. Immunohistochemical expression of Fas ligand (FasL) and neprilysin (neutral endopeptidase/CD10) in keratoconus. Int Ophthalmol. (2013) 33:125–31. doi: 10.1007/s10792-012-9651-0
- 155. Iqbal O, Fisher G, Vira S, Syed D, Sadeghi N, Freeman D, et al. Increased expression of secreted frizzled-related protein-1 and microtubuleassociated protein light chain 3 in keratoconus. *Cornea.* (2013) 32:702–7. doi: 10.1097/ICO.0b013e318282987a
- 156. Joseph R, Srivastava OP, Pfister RR. Differential epithelial and stromal protein profiles in keratoconus and normal human corneas. *Exp Eye Res.* (2011) 92:282–98. doi: 10.1016/j.exer.2011.01.008

- 157. Kenney MC, Chwa M, Alba A, Saghizadeh M, Huang ZS, Brown DJ. Localization of TIMP-1, TIMP-2, TIMP-3, gelatinase A and gelatinase B in pathological human corneas. *Curr Eye Res.* (1998) 17:238–46. doi: 10.1076/ceyr.17.3.238.5222
- Kenney MC, Nesburn AB, Burgeson RE, Butkowski RJ, Ljubimov AV. Abnormalities of the extracellular matrix in keratoconus corneas. *Cornea.* (1997) 16:345–51. doi: 10.1097/00003226-199705000-00016
- Lackner EM, Matthaei M, Meng H, Ardjomand N, Eberhart CG, Jun AS. Design and analysis of keratoconus tissue microarrays. *Cornea*. (2014) 33:49– 55. doi: 10.1097/ICO.00000000000012
- 160. Lambiase A, Merlo D, Mollinari C, Bonini P, Rinaldi AM. M DA, et al. Molecular basis for keratoconus: lack of TrkA expression and its transcriptional repression by Sp3. *Proc Natl Acad Sci U S A.* (2005) 102:16795–800. doi: 10.1073/pnas.0508516102
- 161. Li Y, Zhou L, Twining SS, Sugar J, Yue BY. Involvement of Sp1 elements in the promoter activity of the alpha1-proteinase inhibitor gene. *J Biol Chem.* (1998) 273:9959–65. doi: 10.1074/jbc.273.16.9959
- Lyon D, McKay TB, Sarkar-Nag A, Priyadarsini S, Karamichos D. Human Keratoconus Cell Contractility is Mediated by Transforming Growth Factor-Beta Isoforms. J Funct Biomater. (2015) 6:422–38. doi: 10.3390/jfb6020422
- 163. Maatta M, Heljasvaara R, Sormunen R, Pihlajaniemi T, Autio-Harmainen H, Tervo T. Differential expression of collagen types XVIII/endostatin and XV in normal, keratoconus, and scarred human corneas. *Cornea.* (2006) 25:341–9. doi: 10.1097/01.ico.0000178729.57435.96
- 164. Maatta M, Vaisanen T, Vaisanen MR, Pihlajaniemi T, Tervo T. Altered expression of type XIII collagen in keratoconus and scarred human cornea - Increased expression in scarred cornea is associated with myofibroblast transformation. *Cornea.* (2006) 25:448–53. doi: 10.1097/01.ico.0000183537.45393.1f
- Mackiewicz Z, Maatta M, Stenman M, Konttinen L, Tervo T, Konttinen YT. Collagenolytic proteinases in keratoconus. *Cornea*. (2006) 25:603–10. doi: 10.1097/01.ico.0000208820.32614.00
- 166. Malfeito M, Regueiro U, Perez-Mato M, Campos F, Sobrino T, Lema I. Innate Immunity Biomarkers for Early Detection of Keratoconus. *Ocul Immunol Inflamm.* (2019) 27:942–8. doi: 10.1080/09273948.2018.1511813
- 167. Marini M, Mencucci R, Rosa I, Favuzza E, Guasti D, Ibba-Manneschi L, et al. Telocytes in normal and keratoconic human cornea: an immunohistochemical and transmission electron microscopy study. J Cell Mol Med. (2017) 21:3602–11. doi: 10.1111/jcmm.13270
- 168. Nielsen K, Vorum H, Fagerholm P, Birkenkamp-Demtroder K, Honore B, Ehlers N, et al. Proteome profiling of corneal epithelium and identification of marker proteins for keratoconus, a pilot study. *Exp Eye Res.* (2006) 82:201–9. doi: 10.1016/j.exer.2005.06.009
- Olofsson EM, Marklund SL, Pedrosa-Domellof F, Behndig A. Interleukinlalpha downregulates extracellular-superoxide dismutase in human corneal keratoconus stromal cells. *Mol Vis.* (2007) 13:1285–90.
- 170. Priyadarsini S, Hjortdal J, Sarker-Nag A, Sejersen H, Asara JM, Karamichos D. Gross cystic disease fluid protein-15/prolactin-inducible protein as a biomarker for keratoconus disease. *PLoS ONE.* (2014) 9:e113310. doi: 10.1371/journal.pone.0113310
- 171. Regueiro U, Perez-Mato M, Hervella P, Campos F, Sobrino T, Lema I. Tolllike receptors as diagnostic targets in pellucid marginal degeneration. *Exp Eye Res.* (2020) 200:108211. doi: 10.1016/j.exer.2020.108211
- Sacchetti M, Scorcia V, Lambiase A, Bonini S. Preliminary evidence of neuropeptides involvement in keratoconus. *Acta Ophthalmol.* (2015) 93:e315–6. doi: 10.1111/aos.12483
- 173. Sawaguchi S, Yue BY, Sugar J, Gilboy JE. Lysosomal enzyme abnormalities in keratoconus. Arch Ophthalmol. (1989) 107:1507–10. doi: 10.1001/archopht.1989.01070020581044
- 174. Seppala HPS, Maatta M, Rautia M, Mackiewicz Z, Tuisku I, Tervo T, et al. EMMPRIN and MMP-1 in keratoconus. *Cornea.* (2006) 25:325–30. doi: 10.1097/01.ico.0000183534.22522.39
- Sevost'ianov EN, Giniatullin RU, Gorskova EN, Teplova SN. Keratocyte apoptosis in keratoconus. *Vestnik oftalmologii*. (2002) 118:36–8.
- 176. Sherwin T, Brookes NH, Low IP, Poole CA, Clover GM. Cellular incursion into Bowman's membrane in the peripheral cone of the keratoconic cornea. *Exp Eye Res.* (2002) 74:473–82. doi: 10.1006/exer.2001.1157

- 177. Smith VA, Hoh HB, Littleton M, Easty DL. Over-expression of a gelatinase a activity in. keratoconus. *Eye.* (1995) 9:429–33. doi: 10.1038/eye.1995.100
- 178. Smith VA, Matthews FJ, Majid MA, Cook SD. Keratoconus: Matrix metalloproteinase-2 activation and TIMP modulation. *Biochim Biophys Acta Mol Basis Dis.* (2006) 1762:431–9. doi: 10.1016/j.bbadis.2006. 01.010
- Srivastava OP, Chandrasekaran D, Pfister RR. Molecular changes in selected epithelial proteins in human keratoconus corneas compared to normal corneas. *Molecular Vision*. (2006) 12:1615–25.
- 180. Tai TYT, Damani MR, Vo R, Rayner SA, Glasgow BJ, Hofbauer JD, et al. Keratoconus associated with corneal stromal amyloid deposition containing TGFBIP. *Cornea.* (2009) 28:589–93. doi: 10.1097/ICO.0b013e31818 c9003
- 181. Takacs L, Csutak A, Balazs E, Modis L, Berta A. Expression of beta ig-h3 is lower than normal in keratoconus corneas but increases with scarring. *Cornea*. (1999) 18:599–605. doi: 10.1097/00003226-199909000-00014
- Thanos S, Oellers P. zu Horste MM, Prokosch V, Schlatt S, Seitz B, et al. Role of thyroxine in the development of keratoconus. *Cornea*. (2016) 35:1338–46. doi: 10.1097/ICO.00000000000988
- 183. Toti P, Tosi GM, Traversi C, Schurfeld K, Cardone C, Caporossi A. CD-34 stromal expression pattern in normal and altered human corneas. *Ophthalmology*. (2002) 109:1167–71. doi: 10.1016/s0161-6420(02)01042-4
- Tuori A, Virtanen I, Aine E, Uusitalo H. The expression of tenascin and fibronectin in keratoconus, scarred and normal human cornea. *Graefes Arch Clin Exp Ophthalmol.* (1997) 235:222–9. doi: 10.1007/BF00941763
- 185. Tuori AJ, Virtanen I, Aine E, Kalluri R, Miner JH, Uusitalo HM. The immunohistochemical composition of corneal basement membrane in keratoconus. *Curr Eye Res.* (1997) 16:792–801. doi: 10.1076/ceyr.16.8.792.8989
- 186. Wang X, Zou L, Jin T, Pan Z. Apoptosis in keratoconus and its relevance to the expression of Fas-L protein. *Ophthalmic Research*. (2008) 26:53–6.
- 187. Wolf M, Clay SM, Oldenburg CE, Rose-Nussbaumer J, Hwang DG, Chan MF. Overexpression of MMPs in corneas requiring penetrating and deep anterior lamellar keratoplasty. *Invest Ophthalmol Vis Sci.* (2019) 60:1734–47. doi: 10.1167/iovs.18-25961
- 188. Yam GH-F, Fuest M, Zhou L, Liu Y-C, Deng L, Chan AS-Y, et al. Differential epithelial and stromal protein profiles in cone and noncone regions of keratoconus corneas. *Scientific Reports.* (2019) 9. doi: 10.1038/s41598-019-39182-6
- Yin H, Luo C, Tian Y, Deng Y. Altered expression of sex hormone receptors in keratoconus corneas. *Biomedical Research-India*. (2017) 28:5089–92.
- 190. You J, Wen L, Roufas A, Madigan MC, Sutton G. Expression of SFRP Family Proteins in Human Keratoconus Corneas. *Plos ONE.* (2013) 8. doi: 10.1371/journal.pone.0066770
- 191. You J, Wen L, Roufas A, Hodge C, Sutton G, Madigan MC. Expression of HGF and c-met proteins in human keratoconus corneas. J Ophthalmol. (2015). doi: 10.1155/2015/852986
- 192. Zhou LL, Sawaguchi S, Twining SS, Sugar J, Feder RS, Yue B. Expression of degradative enzymes and protease inhibitors in corneas with keratoconus. *Investigative Ophthalmology & Visual Science*. (1998) 39:1117–24.
- 193. Zhou LL, Yue B, Twining SS, Sugar J, Feder RS. Expression of wound healing and stress-related proteins in keratoconus corneas. *Curr Eye Res.* (1996) 15:1124–31. doi: 10.3109/02713689608995144
- 194. Roy S, Yadav S, Dasgupta T, Chawla S, Tandon R, Ghosh S. Interplay between hereditary and environmental factors to establish an in vitro disease model of keratoconus. *Drug Discov Today.* (2019) 24:403–16. doi: 10.1016/j.drudis.2018.10.017
- 195. Farzadfard A, Nassiri N, Moghadam TN, Paylakhi SH, Elahi E. Screening for MIR184 Mutations in Iranian Patients with Keratoconus. J Ophthalmic Vis Res. (2016) 11:3–7. doi: 10.4103/2008-322X.180715
- 196. Lucas SEM, Zhou T, Blackburn NB, Mills RA, Ellis J, Leo P, et al. Rare, potentially pathogenic variants in ZNF469 are not enriched in keratoconus in a large australian cohort of european descent. *Invest Ophthalmol Vis Sci.* (2017) 58:6248–56. doi: 10.1167/iovs.17-22417
- 197. Kalantan H, Kondkar AA, Sultan T, Azad TA, Alsabaani NA, AlQahtani MA, et al. Polymorphism rs13334190 in zinc finger protein 469 (ZNF469) is not a risk factor for keratoconus in a Saudi cohort. *BMC Res Notes.* (2017) 10:652. doi: 10.1186/s13104-017-2996-8

- Jeoung JW, Kim MK, Park SS, Kim SY, Ko HS, Wee WR, et al. VSX1 gene and keratoconus: genetic analysis in Korean patients. *Cornea.* (2012) 31:746–50. doi: 10.1097/ICO.0b013e3181e16dd0
- 199. Nejabat M, Naghash P, Dastsooz H, Mohammadi S, Alipour M, Fardaei M. VSX1 and SOD1 mutation screening in patients with keratoconus in the South of Iran. J Ophthalmic Vis Res. (2017) 12:135–40. doi: 10.4103/jovr.jovr_97_16
- 200. Verma A, Das M, Srinivasan M, Prajna NV, Sundaresan P. Investigation of VSX1 sequence variants in South Indian patients with sporadic cases of keratoconus. *BMC Res Notes*. (2013) 6:103. doi: 10.1186/1756-0500-6-103
- 201. Palamar M, Onay H, Ozdemir TR, Arslan E, Egrilmez S, Ozkinay F, et al. Relationship between IL1beta-511C>T and ILRN VNTR polymorphisms and keratoconus. *Cornea.* (2014) 33:145–7. doi: 10.1097/ICO.00000000000027
- 202. Davidson AE, Borasio E, Liskova P, Khan AO, Hassan H, Cheetham ME, et al. Brittle cornea syndrome ZNF469 mutation carrier phenotype and segregation analysis of rare ZNF469 variants in familial keratoconus. *Invest Ophthalmol Vis Sci.* (2015) 56:578–86. doi: 10.1167/iovs.14-15792
- 203. Simcoe MJ, Khawaja AP, Hysi PG, Hammond CJ, Eye UKB, Vision C. Genome-wide association study of corneal biomechanical properties identifies over 200 loci providing insight into the genetic etiology of ocular diseases. *Hum Mol Genet*. (2020) 29:3154–64. doi: 10.1093/hmg/ddaa155
- 204. Loh IP, Sherwin T. Is Keratoconus an Inflammatory Disease? The Implication of Inflammatory Pathways. Ocul Immunol Inflamm. (2020):1– 10. doi: 10.1080/09273948.2020.1780271
- 205. Kim WJ, Rabinowitz YS, Meisler DM, Wilson SE. Keratocyte apoptosis associated with keratoconus. *Exp Eye Res.* (1999) 69:475–81. doi: 10.1006/exer.1999.0719
- Wolffsohn JS, Safeen S, Shah S, Laiquzzaman M. Changes of corneal biomechanics with keratoconus. *Cornea.* (2012) 31:849–54. doi: 10.1097/ICO.0b013e318243e42d
- 207. Eliasi E, Bez M, Megreli J, Avramovich E, Fischer N, Barak A, et al. The Association Between Keratoconus and Body Mass Index: A Population-Based Cross-Sectional Study Among Half a Million Adolescents. Am J Ophthalmol. (2021) 224:200–6. doi: 10.1016/j.ajo.2020.11.021
- Gatzioufas Z, Thanos S. Acute keratoconus induced by hypothyroxinemia during pregnancy. J Endocrinol Invest. (2008) 31:262–6. doi: 10.1007/BF03345600
- Bilgihan K, Hondur A, Sul S, Ozturk S. Pregnancy-induced Progression of Keratoconus. Cornea. (2011) 30:991–4. doi: 10.1097/ICO.0b013e3182068adc
- Dutta D, Shivaprasad K, Ghosh S, Mukhopadhyay S, Chowdhury S. Adrenal myelolipoma with keratoconus: A novel clinical association. *Indian J Endocrinol Metab.* (2012) 16:S364–6. doi: 10.4103/2230-8210.104094
- McKay TB, Hjortdal J, Sejersen H, Karamichos D. Differential Effects of Hormones on Cellular Metabolism in Keratoconus In Vitro. *Scientific Reports*. (2017) 7. doi: 10.1038/srep42896
- 212. Sharif R, Bak-Nielsen S, Sejersen H, Ding K, Hjortdal J, Karamichos D. Prolactin-induced protein is a novel biomarker for Keratoconus. *Exp Eye Res.* (2019) 179:55–63. doi: 10.1016/j.exer.2018.10.015
- 213. Dawczynski J, Franke S, Blum M, Kasper M, Stein G, Strobel J. Advanced glycation end-products in corneas of patients with keratoconus. *Graefes Arch Clin Exp Ophthalmol.* (2002) 240:296–301. doi: 10.1007/s00417-002-0445-3
- Arnal E, Peris-Martinez C, Luis Menezo J, Johnsen-Soriano S, Javier Romero F. Oxidative stress in keratoconus? *Invest Ophthalmol Vis Sci.* (2011) 52:8592–7. doi: 10.1167/iovs.11-7732

- Lema I, Duran JA. Inflammatory molecules in the tears of patients with keratoconus. *Ophthalmology*. (2005) 112:654–9. doi: 10.1016/j.ophtha.2004.11.050
- 216. Sorkhabi R, Ghorbanihaghjo A, Taheri N, Ahoor MH. Tear film inflammatory mediators in patients with keratoconus. *Int Ophthalmol.* (2015) 35:467–72. doi: 10.1007/s10792-014-9971-3
- Ionescu C, Corbu CG, Tanase C, Jonescu-Cuypers C, Nicula C, Dascalescu D, et al. Inflammatory biomarkers profile as microenvironmental expression in keratoconus. *Dis Markers*. (2016). doi: 10.1155/2016/1243819
- Uzunoglu E, Lortlar N, Erdogan D, Erdamar H, Akyol ES, Akata F. Immunohistochemical and ultrastructural presentation of apoptosis and aqueous humor's nitric oxide levels in keratoconus. *Nobel Medicus*. (2013) 9:5–9.
- 219. Kaldawy RM, Wagner J, Ching S, Seigel GM. Evidence of apoptotic cell death in keratoconus. *Cornea.* (2002) 21:206–9. doi: 10.1097/00003226-200203000-00017
- 220. Rodrigues M, Nirankari V, Rajagopalan S, Jones K, Funderburgh J. Clinical and histopathologic changes in the host cornea after epikeratoplasty for keratoconus. Am J Ophthalmol. (1992) 114:161–70. doi: 10.1016/S0002-9394(14)73980-7
- 221. McMonnies CW. Epigenetic mechanisms might help explain environmental contributions to the pathogenesis of keratoconus. *Eye Contact Lens.* (2014) 40:371–5. doi: 10.1097/ICL.000000000000078
- 222. Hu W, Han Q, Zhao L, Wang L. Circular RNA circRNA_15698 aggravates the extracellular matrix of diabetic nephropathy mesangial cells *via* miR-185/TGF-beta1. *J Cell Physiol.* (2019) 234:1469–76. doi: 10.1002/jcp.26959
- 223. Wen ZJ, Xin H, Wang YC, Liu HW, Gao YY, Zhang YF. Emerging roles of circRNAs in the pathological process of myocardial infarction. *Mol Ther Nucleic Acids*. (2021) 26:828–48. doi: 10.1016/j.omtn.2021.10.002
- 224. Zhang Y, Jia DD, Zhang YF, Cheng MD, Zhu WX Li PF, et al. The emerging function and clinical significance of circRNAs in Thyroid Cancer and Autoimmune Thyroid Diseases. *Int J Biol Sci.* (2021) 17:1731–41. doi: 10.7150/ijbs.55381
- 225. Zhang Q, Qiao X, Xia W. CircSERPINE2 weakens IL-1beta-caused apoptosis and extracellular matrix degradation of chondrocytes by regulating miR-495/TGFBR2 axis. *Biosci Rep.* (2020) 40. doi: 10.1042/BSR20201601

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