Check for updates

OPEN ACCESS

EDITED BY Ioanna Mylona, General Hospital of Katerini, Greece

REVIEWED BY Yanxian Chen, Shenzhen Hospital, Peking University, China Jessy Choi, Sheffield Teaching Hospitals NHS Foundation Trust, United Kingdom

*CORRESPONDENCE

Longqian Liu ⊠ b.q15651@hotmail.com Hongbin Lv ⊠ oculistlyhongbin@163.com

[†]These authors have contributed equally to this work and share first authorship

SPECIALTY SECTION

This article was submitted to Ophthalmology, a section of the journal Frontiers in Medicine

RECEIVED 17 August 2022 ACCEPTED 19 December 2022 PUBLISHED 09 January 2023

CITATION

Huang J, Nie K, Lv X, Liu Y, Yang G, Fu J, Liu L and Lv H (2023) Abnormal lens thickening in a child with Weill–Marchesani syndrome 4: A 3-year follow-up case report. *Front. Med.* 9:1021489. doi: 10.3389/fmed.2022.1021489

COPYRIGHT

© 2023 Huang, Nie, Lv, Liu, Yang, Fu, Liu and Lv. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Abnormal lens thickening in a child with Weill–Marchesani syndrome 4: A 3-year follow-up case report

Junting Huang^{1†}, Kailai Nie^{1,2†}, Xinpin Lv³, Yuting Liu³, Guiqi Yang³, Junjiang Fu⁴, Longqian Liu¹* and Hongbin Lv³*

¹Department of Ophthalmology, West China Hospital, Sichuan University, Chengdu, China, ²Research Laboratory of Ophthalmology and Vision Sciences, State Key Laboratory of Biotherapy, West China Hospital, Sichuan University, Chengdu, China, ³Department of Ophthalmology, The Affiliated Hospital of Southwest Medical University, Luzhou, China, ⁴Key Laboratory of Epigenetics and Oncology, The Research Center for Preclinical Medicine, Southwest Medical University, Luzhou, China

Background: Weill–Marchesani syndrome 4 (WMS4) is caused by *ADAMTS17* gene variant and clinical abnormalities including lenticular myopia, ectopia lentis, glaucoma, microspherophakia, brachydactyly, and short stature. Due to free of heart defects and joint stiffness compared with other WMS forms, WMS4 has an insidious onset and is often misdiagnosed as high myopia. We combined multiple imaging biometry and whole-exome sequencing to diagnose a case of WMS4 with a 3-year follow-up.

Case presentation: An 8-year-old boy presented to our ophthalmology department with progressive myopia for 1 year. He had high myopia in both eyes with normal funds, intraocular pressure, and axial length. Ocular examination revealed thicker lenses (right 4.38 mm, left 4.31 mm) with a smaller equatorial diameter (right 7.33 mm and left 7.17 mm) compared to normal children of the same age. Finger length measurement indicates brachydactyly. Whole-exome sequencing identified compound heterozygous missense variants c.2984G > A (p.Arg995Gln) and c.2254A > G (p.Ile752Val) in the *ADAMTS17* gene. During the 3 years of follow-up, the thickness of lenses increased significantly (right 4.49 mm, left 4.48 mm), but the equatorial diameter of the lenses had no significant change (right 7.32 mm, left 7.21 mm). As the equivalent lens power increased, the patient's myopia spherical refractive error rose accordingly. Although the anterior chamber angle remained open during follow-up, the intraocular pressure increased to right 20.4 mmHg and left 19.6 mmHg, Iridodonesis and short stature were present.

Conclusion: This case report highlights the abnormal thickening of the lens in WMS4 compared to the physiological thinning process during childhood.

Comprehensive clinical examinations and genetic testing may improve diagnosis, which allows early therapeutic interventions for complications and better visual outcomes for the patient.

KEYWORDS

Weill-Marchesani syndrome, ADAMTS17 gene, lens, myopia, microspherophakia

Introduction

Weill-Marchesani syndrome (WMS) is a rare genetic disease that mainly affects the development of the eye, musculoskeletal system, and cardiovascular system. The main phenotypes are short stature, joint stiffness, thickened skin, brachydactyly, and ocular anomalies including microspherophakia, high myopia, angle-closure glaucoma, and cataract (1, 2). The autosomal dominant form of WMS is caused by variants in the FBN1 gene (WMS2) (3), whereas the autosomal recessive form is caused by variants in genes of ADAMTS10 (WMS1) (4), LTBP2 (WMS3) (5), and ADAMTS17 (WMS4) (6). The main difference between WMS4 caused by the ADAMTS17 gene variant and other WMS forms is that WMS4 is commonly free of heart defects and joint stiffness (7-9). Morales et al. (6) firstly reported ADAMTS17 variants in humans with WMS4 and defined the character of the WMS4 clinical abnormalities including lenticular myopia, ectopia lentis, glaucoma, spherophakia, brachydactyly, and short stature. This disease is also rarely reported in China (7, 10).

In this report, a combination of multiple imaging tests and whole-exome sequencing has been performed to identify a WMS4 patient with microspherophakia, short stature, and brachydactyly in a Chinese family. Abnormal thickening of the lens of the patient was observed during a 3-year follow-up.

Case report

An 8-year-old boy first visited the ophthalmic clinic with the chief complaint of a progressive decrease in binocular vision for 1 year. He had been diagnosed as high myopia in other hospitals several times with fast myopia progression. A total of 1% cyclopentolate eye drops were administered before optometry, slit lamp, biometry, and fundus examinations. Slit lamp tests revealed bilateral microspherophakia and showed that the equatorial region of the lens was fully visible after pupil dilation (**Figure 1A**). The spherical refractive error was -8.25 diopter and -7.50 diopter in the right eye and left eye respectively, but fundus examinations including ultra-widefield retinal imaging and optical coherence tomography angiography did not identify any characteristic changes in high myopia such as posterior staphyloma, myopic maculopathy, choroidal

neovascularization, and retinal detachment (11-13). IOLMaster 700 biometry showed that the axial length was 22.07 mm in the right eye and 22.01 mm left eye (Table 1), which were within mean \pm 1.96 standard deviation (SD) of the normal reference range (14-20). While the lens thickness (right 4.38 mm, left 4.31 mm) was significantly greater than normal (Table 1) (16-18, 21, 22). PENTACAM illustrated visually that the lens of the proband was thicker than normal children (control) of the same age (Figure 1B, red line). Although it was difficult for PENTACAM to measure in vivo the equatorial diameter of the lens in normal subject after adequate pupil dilatation, the patient's lens equatorial diameter (right 7.33 mm and left 7.17 mm) was seen to be smaller than that of normal control child (Figure 1B, blue line, presumed value 9.62 mm) and smaller than that of lenses at the same age measured ex vivo (23–26). The objective scatter index of OQAS II was 1.14 ± 0.20 in the right eye and 1.17 \pm 0.09 in the left eye, suggesting no significant loss of lens transparency. Ultrasound biomicroscopy demonstrated a wide angle of the anterior chamber, the iris and ciliary body showed no obvious abnormalities, the curvature of the anterior lens capsule increased and bulged forward, and the gap between the lens equatorial region of the lens and ciliary body became larger (Figure 1C). Intraocular pressure was normal (right 16.7 mmHg, left 16.0 mmHg) and corneal topography analyses excluded keratoconus (Table 1). Moreover, the patient exhibited brachydactyly (5.1 cm, -2.4 SD) (27) but did not report hand joint stiffness (Figure 1D, arrows) and was not of short stature at 8 years old (height 124 cm, -1.6 SD). No structural anomalies were found in the cardiovascular system or abdomen in ultrasound scans. Given that the patient had not yet developed symptoms such as ocular hypertension or lens ectopia, spectacles were prescribed to correct the refractive error with regular follow-up. Bestcorrected visual acuity was 20/25 in both eyes. Neither the patient's parents nor sisters had myopia, and there were no abnormalities in intraocular pressure, slit lamp and mydriatic fundus examination.

During the subsequent 3 years of follow-up, the patient presented a progressive increase in lens thickness (Figure 1E and Table 1). The equivalent lens power of the patient was calculated (28) and showed a progressive increase, which was more pronounced in the left eye. It was consistent with the change in spherical refractive error (Figure 2A).



In addition, the intraocular pressure of the patient was elevated to the upper limit of normal (right 20.4 mmHg, left 19.6 mmHg), and iridodonesis was observed (**Supplementary Video**). PENTACAM showed no significant increase in the equatorial diameter of the lens (right 7.32 mm and left 7.21 mm at 11 years of age) (23–26). At a recent follow-up height measurement, the patient (at age 10) exhibited short stature (height 131.2 cm, -2.7 standard deviation) (27).

Whole-exome sequencing was performed on the proband (Figure 2B; III.3, arrow) and compound heterozygous variants were successfully identified in exon 16 (c.2254A > G) and exon 21 (c.2984G > A) of *ADAMTS17* (NM_139057.4). The detailed result of whole exome sequencing was provided in the **Supplementary Tables 1**, 2. The variants of the *ADAMTS17* gene were confirmed by Sanger sequencing, and co-segregation analysis of the family was performed (Figure 2C). The result suggests that these compound heterozygous *ADAMTS17*

variants and disease were co-segregated in this family and imply their potential role in the autosomal recessive WMS.

Discussion

The human lens is almost spherical in early embryonic development and increasingly elliptical during the postnatal emmetropization process (29, 30). Measurements in school-age children between the ages of 6 and 10 years have shown that the axial dimension of the lens thins by an average of nearly 0.2 mm (22). Zadnik et al. (18) reported thickness thinning and flattening of the lens of 2,583 children aged 6–14 years. Data including 11,656 students indicated that the lens thinned within the 7–11 age range (17). A study included 596 Chinese children suggested a reduction in lens thickness and refractive lens power in children aged 6–13 years (21). Lens thinning at 7–10 years of age was also observed in a cohort of 303

| Parameters | Method | 8 years old | 9 years old | 10 years old | 11 years old |
|--------------------------------------|------------|----------------|----------------|------------------|------------------|
| Axial length (mm) | | | | | |
| Patient's (right/left) | IOLMaster | 22.07/22.01 | 22.15/22.06 | 22.39/22.32 | 22.62/22.55 |
| Lu et al. (14) (boys) | IOLMaster | 23.28 ± 0.84 | 23.61 ± 0.86 | 23.88 ± 0.99 | 23.97 ± 1.00 |
| Fledelius et al. (15) (males) | IOLMaster | 23.22 ± 0.64 | 23.70 ± 0.78 | | 23.44 ± 0.71 |
| Rozema et al. (16) | Ultrasound | 23.06 ± 0.68 | 23.15 ± 0.69 | 23.32 ± 0.72 | 23.43 ± 0.72 |
| Shih et al. (17) (boys) | Ultrasound | 23.27 ± 0.90 | 23.49 ± 0.89 | 23.79 ± 1.00 | 23.89 ± 1.00 |
| Zadnik et al. (18) (boys) | Ultrasound | 23.14 ± 0.81 | 23.40 ± 0.70 | 23.43 ± 0.80 | 23.54 ± 0.84 |
| Larsen (19) (males) | Ultrasound | 22.33 ± 0.51 | 22.43 ± 0.47 | 22.50 ± 0.47 | 22.70 ± 0.82 |
| Lens thickness (mm) | | | | | |
| Patient's (right/left) | IOLMaster | 4.38/4.31 | 4.41/4.38 | 4.46/4.47 | 4.49/4.48 |
| Lu et al. (21) (boys) | Lenstar | 3.31 ± 0.14 | 3.26 ± 0.15 | 3.31 ± 0.19 | 3.31 ± 0.15 |
| Rozema et al. (16) | Ultrasound | 3.48 ± 0.18 | 3.44 ± 0.17 | 3.45 ± 0.17 | 3.45 ± 0.19 |
| Shih et al. (17) (boys) | Ultrasound | 3.42 ± 0.27 | 3.40 ± 0.24 | 3.36 ± 0.26 | 3.32 ± 0.23 |
| Zadnik et al. (18) (boys) | Ultrasound | 3.46 ± 0.16 | 3.44 ± 0.17 | 3.43 ± 0.18 | 3.43 ± 0.17 |
| Other eye component (right/ | left) | | | | |
| Central corneal thickness (μ m) | IOLMaster | 553/548 | 564/559 | 554/552 | 547/545 |
| Anterior chamber depth (mm) | IOLMaster | 2.81/2.86 | 2.69/2.77 | 2.72/2.80 | 2.69/2.76 |
| White to white (mm) | IOLMaster | 12.0/12.0 | 12.0/11.9 | 11.9/12.0 | 11.9/12.0 |
| Mean keratometry (diopter) | IOLMaster | 44.42/44.58 | 44.12/44.35 | 44.44/44.62 | 44.14/44.41 |

TABLE 1 Ocular biometric measurements of the patient and comparison with reference values reported in the literature.

emmetropic Singaporean children (16). However, the lens of the proband was thicker at 8 years of age than that of children of the same age and thickened by about 0.1 mm between 8 and 11 years old (**Figure 1E** and **Table 1**). Furthermore, lens thickness was generally thinner in myopes than emmetropes within 7–14 years of age, which may be a loss of equivalent lens power to compensate for the rapid axial growth of myopes (16, 17, 21, 31). In contrast, this WMS4 patient presented an abnormal lens thickening and a notable increase in equivalent lens power during the axial growth, resulting in progressive lens-induced high myopia.

In addition, the equatorial diameter of the patient's lens did not increase significantly and was consistently lower than that of normal peers (24–26). Measurements of intact, clear, postmortem lenses have shown that the central thickness gradually increases after 20 years of age, while the equatorial diameter of the lens increases with life, with a particularly rapid increase in equatorial diameter before 20 years of age (24– 26). As the equatorial diameter increases at nearly the same rate as the thickness, the relative shape of the lens is almost constant during adulthood (32). Thus, the lens thickness of children with WMS4 may not be thicker than that of adults (33), especially compared to elderly cataract patients (34). However, the lens thickness is still significantly thicker than that of normal children of the same age and the equatorial diameter is consistently smaller. Caution is needed when diagnosing microspherophakia in children of this age to avoid misdiagnosis.

The ADAMTS17 gene locating on human chromosome 15q26.3, encodes a protease, which belongs to a superfamily of the ADAMTS, a disintegrin-like and metalloproteinase domain with thrombospondin-type 1 motifs and may participate in the regulation of assembly of microfibrils by modulating the fibrillin isoform (35). According to the gnomAD database, c.2984G > A (rs1289240183, dbSNP) is rare with a minor allele frequency of 0.00001. In contrast, c.2254A > G has not been reported by the 1,000 Genomes Project, the ExAC, gnomAD, or the NHLBI ESP database. These variants resulted in substitutions including isoleucine (Ile) to valine (Val) at amino acid 752 (p.Ile752Val, c.2254A > G), and arginine (Arg) to glutamine (Gln) at amino acid 995 (p.Arg995Gln, c.2984G > A) in the ADAMTS17 protein (Supplementary Table 1). Two missense variants in ADAMTS17 have been reported to be associated with WMS: p.Thr343Ala and p.Cys1023Tyr. Among them, the p.Thr343Ala variant resides in a zinc-dependent metalloprotease domain, which results in reduced secretion of ADAMTS17 into the extracellular matrix (9). Evans et al. (8) first reported the p.Cys1023Tyr variant and revealed that a change in this amino residue results in significantly decreased secretion of ADAMTS17. Notably, the missense variants found in this case, p.Arg995Gln, and the nearby p.Cys1023Tyr are located in the same thrombospondin type 1 repeats domain. In addition,



FIGURE 2

(A) The spherical refractive error of the proband increased (more negative) from 8 to 11 years of age and was more pronounced in the left eye, with the same trend as the calculated equivalent lens power (28); solid line: Myopia refractive error; dashed line: equivalent lens power. (B) Pedigree with WMS4; the proband III.3 is marked with a solid square and an arrow; unaffected individuals were labeled as blank symbols. Individuals with known genotypes were labeled, and carriers were labeled with the variants (M1:c.2984G > A, M2:c.2254A > G). (C) Sanger sequencing validated the compound heterozygous *ADAMTS17* variants in the WMS family. Red arrows indicate the variant sites.

| WMS subtype (gene) | Ocular | Cardiovascular system | Musculoskeletal system | Pattern of inheritance |
|---------------------------|---|--------------------------|--|------------------------|
| WMS1 (ADAMTS10) (4) | Microspherophakia, glaucoma, ectopia lentis, cataract | Cardiac anomalies | Short stature, joint stiffness, brachycephaly, brachydactyly, thick skin | Autosomal recessive |
| WMS2 (FBN1) (3) | Microspherophakia glaucoma ectopia lentis cataract | Cardiac anomalies | Short stature, joint stiffness, brachycephaly, brachydactyly, thick skin | Autosomal dominant |
| WMS3 (<i>LTBP2</i>) (5) | Ectopia lentis, increased intraocular pressure, microspherophakia | Cardiac anomalies | Short stature, joint stiffness, brachydactyly | Autosomal recessive |
| WMS4 (ADAMTS17) (6) | Ectopia lentis, glaucoma, microspherophakia | No | Short stature, brachydactyly | Autosomal recessive |

p.Ile752Val lies in the spacer domain, and splice variants of the spacer domain are reported to regulate proteolytic specificity and protein secretion of *ADAMTS17* (36).

Short stature and brachydactyly are obvious features of WMS4. Genome-wide association studies have shown that

ADAMTS17 is associated with human height (37), suggesting that it may play a role in human skeletal growth. Recently, Oichi et al. (38) established an *Adamts17* knockout mouse model, and found that mice recapitulate clinical phenotypes of WMS, including shorter long bones, brachydactyly, and thickened skin.

They suggested that *ADAMTS17* modulates the bone morphogenetic protein (BMP) -Smad1/5/8 pathway possibly by inhibiting the incorporation of fibrillin-2 into microfibrils to regulate skeletal formation. This may explain the common phenotypes of short height and brachydactyly in WMS4 caused by *ADAMTS17* variants. Heart defects and joint stiffness are rarely seen in WMS4 that distinguish it from other WMS forms (**Table 2**) (7–9), these abnormalities were not found in this case.

The previous cases reported ADAMTS17 variants lead to ectopia lentis and secondary glaucoma in WMS4 (6-10). ADAMTS17 expression was detected by RNA in situ hybridization in multiple ocular tissues of embryonic mouse, including fiber cells of the lens equator, ciliary body epithelium at the zonule junction site, and trabecular meshwork (35). In addition, an ADAMTS17 missense variant that causes primary lens luxation and primary open angle glaucoma in dogs has been reported (39). These results suggest a potential role of ADAMTS17 in the lens zonule, trabecular meshwork and ciliary body. Therefore, the patient requires long-term clinical follow-up and timely management of possible complications to prevent optic nerve damage and visual field impairment due to secondary glaucoma. Early diagnosis is important in the management of WMS4, as possible complications including ectopia lentis, ocular hypertension, and glaucoma, may be intervened early.

In previous reports, patients with WMS4 usually had already developed peripheral iris synechiae, ectopia lentis, ocular hypertension, glaucoma, or had undergone surgical treatment at the time of initial diagnosis (6-10). This is because WMS4 has an insidious onset and is often diagnosed as high myopia with insidious early symptoms in the initial examination, especially in childhood. We made the diagnosis at an early stage and demonstrated changes in the biological parameters of the eyes during a long follow-up period, increasing the awareness of the disease. To avoid missed diagnosis and misdiagnosis, it is important to pay attention to the patients who have high myopia with narrow anterior chamber, without axial extension of the eye or myopic retinopathy. Awareness of the physiological developmental process of lens is needed. Comprehensive clinical examinations may improve diagnosis, and whole-exome sequencing has been frequently used to investigate genetic disorders which can be used to aid diagnosis.

References

Author contributions

HL and LL: conception and design. LL: administrative support. YL and GY: provision of study materials or patients. JH and XL: collection and assembly of data. KN and JF: data analysis and interpretation and manuscript writing. All authors contributed to the article and approved the submitted version.

Funding

This work was supported by the Science and Technology Department of Sichuan Province (20KJPX0151).

Acknowledgments

We sincerely thank the patient and his family for their participation in the study.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Supplementary material

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/ fmed.2022.1021489/full#supplementary-material

^{1.} Kojuri J, Razeghinejad M, Aslani A. Cardiac findings in weill-marchesani syndrome. Am J Med Genet A. (2007) 143A:2062–4.

^{2.} Sakai L, Keene D, Engvall E. Fibrillin, a new 350-Kd Glycoprotein, is a component of extracellular microfibrils. *J Cell Biol.* (1986) 103(6 Pt 1):2499–509. doi: 10.1083/jcb.103.6.2499

^{3.} Faivre L, Gorlin R, Wirtz M, Godfrey M, Dagoneau N, Samples J, et al. In frame fibrillin-1 gene deletion in autosomal dominant weill-marchesani syndrome. *J Med Genet.* (2003) 40:34–6. doi: 10.1136/jmg.40.1.34

^{4.} Dagoneau N, Benoist-Lasselin C, Huber C, Faivre L, Megarbane A, Alswaid A, et al. Adamts10 mutations in autosomal recessive

weill-marchesani syndrome. Am J Hum Genet. (2004) 75:801–6. doi: 10.1086/42 5231

5. Haji-Seyed-Javadi R, Jelodari-Mamaghani S, Paylakhi S, Yazdani S, Nilforushan N, Fan J, et al. Ltbp2 mutations cause weill-marchesani and weill-marchesani-like syndrome and affect disruptions in the extracellular matrix. *Hum Mutat.* (2012) 33:1182–7. doi: 10.1002/humu.22105

6. Morales J, Al-Sharif L, Khalil D, Shinwari J, Bavi P, Al-Mahrouqi R, et al. Homozygous mutations in adamts10 and adamts17 cause lenticular myopia, ectopia lentis, glaucoma, spherophakia, and short stature. *Am J Hum Genet.* (2009) 85:558–68. doi: 10.1016/j.ajhg.2009.09.011

7. Yi H, Zha X, Zhu Y, Lv J, Hu S, Kong Y, et al. A novel nonsense mutation in adamts17 caused autosomal recessive inheritance weill-marchesani syndrome from a Chinese family. *J Hum Genet.* (2019) 64:681–7. doi: 10.1038/s10038-019-0608-2

8. Evans D, Green J, Fahiminiya S, Majewski J, Fernandez B, Deardorff M, et al. A novel pathogenic missense adamts17 variant that impairs secretion causes weill-marchesani syndrome with variably dysmorphic hand features. *Sci Rep.* (2020) 10:10827. doi: 10.1038/s41598-020-66978-8

9. Karoulias S, Beyens A, Balic Z, Symoens S, Vandersteen A, Rideout A, et al. A novel adamts17 variant that causes weill-marchesani syndrome 4 alters fibrillin-1 and collagen type i deposition in the extracellular matrix. *Matrix Biol.* (2020) 88:1–18. doi: 10.1016/j.matbio.2019.11.001

10. Yu X, Kline B, Han Y, Gao Y, Fan Z, Shi Y. Weill-marchesani syndrome 4 caused by compound heterozygosity of a maternal submicroscopic deletion and a paternal nonsense variant in the adamts17 gene: a case report. *Am J Ophthalmol Case Rep.* (2022) 26:101541. doi: 10.1016/j.ajoc.2022.101541

11. Ohno-Matsui K, Lai T, Lai C, Cheung C. Updates of pathologic myopia. Prog Retin Eye Res. (2016) 52:156–87. doi: 10.1016/j.preteyeres.2015.12.001

12. Ohno-Matsui K, Ikuno Y, Lai T, Gemmy Cheung C. Diagnosis and treatment guideline for myopic choroidal neovascularization due to pathologic myopia. *Prog Retin Eye Res.* (2018) 63:92–106. doi: 10.1016/j.preteyeres.2017. 10.005

13. Flitcroft D, He M, Jonas J, Jong M, Naidoo K, Ohno-Matsui K, et al. Imi – defining and classifying myopia: a proposed set of standards for clinical and epidemiologic studies. *Invest Ophthalmol Vis Sci.* (2019) 60:M20–30. doi: 10.1167/iovs.18-25957

14. Lu T, Wu J, Ye X, Hu Y, Wu H, Sun W, et al. Axial length and associated factors in children: the Shandong children eye study. *Ophthalmologica*. (2016) 235:78–86. doi: 10.1159/000441900

15. Fledelius H, Christensen A, Fledelius C. Juvenile eye growth, when completed? An evaluation based on iol-master axial length data, cross-sectional and longitudinal. *Acta Ophthalmol.* (2014) 92:259–64. doi: 10.1111/aos.12107

16. Rozema J, Dankert S, Iribarren R, Lanca C, Saw S. Axial Growth and lens power loss at myopia onset in singaporean children. *Invest Ophthalmol Vis Sci.* (2019) 60:3091–9. doi: 10.1167/iovs.18-26247

17. Shih Y, Chiang T, Lin L. Lens thickness changes among schoolchildren in Taiwan. *Invest Ophthalmol Vis Sci.* (2009) 50:2637-44. doi: 10.1167/iovs.08-3090

18. Zadnik K, Manny R, Yu J, Mitchell G, Cotter S, Quiralte J, et al. Ocular component data in schoolchildren as a function of age and gender. *Optom Vis Sci.* (2003) 80:226–36. doi: 10.1097/00006324-200303000-00012

19. Larsen J. The sagittal growth of the eye. Iv. Ultrasonic measurement of the axial length of the eye from birth to puberty. *Acta Ophthalmol.* (1971) 49:873–86. doi: 10.1111/j.1755-3768.1971.tb05939.x

20. Chen Y, Wang W, Wang J, Chen X, Zhu Z, Li J, et al. Normal range of ocular biometry in healthy children: a systemic review and meta-analysis of 33,559 individuals under seven years of age. *Ophthalmic Physiol Opt.* (2022) 42:1264–75. doi: 10.1111/opo.13043

21. Lu T, Song J, Wu Q, Jiang W, Tian Q, Zhang X, et al. Refractive lens power and lens thickness in children (6-16 Years Old). *Sci Rep.* (2021) 11:19284. doi: 10.1038/s41598-021-98817-9

22. Zadnik K, Mutti D, Fusaro R, Adams A. Longitudinal evidence of crystalline lens thinning in children. *Invest Ophthalmol Vis Sci.* (1995) 36:1581–7.

23. Bluestein E, Wilson M, Wang X, Rust P, Apple D. Dimensions of the pediatric crystalline lens: implications for intraocular lenses in children. *J Pediatr Ophthalmol Strabismus*. (1996) 33:18–20. doi: 10.3928/0191-3913-19960101-06

24. Schachar R. Growth patterns of fresh human crystalline lenses measured by in vitro photographic biometry. *J Anat.* (2005) 206:575–80. doi: 10.1111/j.1469-7580.2005.00422.x

25. Martinez-Enriquez E, de Castro A, Mohamed A, Sravani N, Ruggeri M, Manns F, et al. Age-related changes to the three-dimensional full shape of the isolated human crystalline lens. *Invest Ophthalmol Vis Sci.* (2020) 61:11. doi: 10. 1167/iovs.61.4.11

26. Mohamed A, Durkee H, Williams S, Manns F, Ho A, Parel J, et al. Morphometric analysis of in vitro human crystalline lenses using digital shadow photogrammetry. *Exp Eye Res.* (2021) 202:108334. doi: 10.1016/j.exer.2020.108334

27. Wen J, Wang J, Xu Q, Wei Y, Zhang L, Ou J, et al. Hand anthropometry and its relation to grip/pinch strength in children aged 5 to 13 years. *J Int Med Res.* (2020) 48:300060520970768. doi: 10.1177/0300060520970768

28. Bennett AGA. Method of determining the equivalent powers of the eye and its crystalline lens without resort to phakometry. *Ophthalmic Physiol Opt.* (1988) 8:53–9. doi: 10.1016/0275-5408(88)90089-0

29. O'Rahilly R. The prenatal development of the human eye. *Exp Eye Res.* (1975) 21:93–112. doi: 10.1016/0014-4835(75)90075-5

30. Iribarren R. Crystalline lens and refractive development. *Prog Retin Eye Res.* (2015) 47:86–106. doi: 10.1016/j.preteyeres.2015.02.002

31. Mutti D, Mitchell G, Sinnott L, Jones-Jordan L, Moeschberger M, Cotter S, et al. Corneal and crystalline lens dimensions before and after myopia onset. *Optom Vis Sci.* (2012) 89:251–62. doi: 10.1097/OPX.0b013e3182418213

32. Rosen A, Denham D, Fernandez V, Borja D, Ho A, Manns F, et al. In Vitro dimensions and curvatures of human lenses. *Vision Res.* (2006) 46:1002–9. doi: 10.1016/j.visres.2005.10.019

33. Wang Y, Zhong J, Li X. Age-related changes of lens thickness and density in different age phases. *Int J Ophthalmol.* (2022) 15:1591–7. doi: 10.18240/ijo.2022. 10.05

34. Meng J, Wei L, He W, Qi J, Lu Y, Zhu X. Lens thickness and associated ocular biometric factors among cataract patients in Shanghai. *Eye Vis.* (2021) 8:22. doi: 10.1186/s40662-021-00245-3

35. Hubmacher D, Schneider M, Berardinelli S, Takeuchi H, Willard B, Reinhardt D, et al. Unusual life cycle and impact on microfibril assembly of adamts17, a secreted metalloprotease mutated in genetic eye disease. *Sci Rep.* (2017) 7:41871. doi: 10.1038/srep41871

36. Balic Z, Misra S, Willard B, Reinhardt D, Apte S, Hubmacher D. Alternative splicing of the metalloprotease adamts17 spacer regulates secretion and modulates autoproteolytic activity. *FASEB J.* (2021) 35:e21310. doi: 10.1096/fj.202001120RR

37. Lango Allen H, Estrada K, Lettre G, Berndt S, Weedon M, Rivadeneira F, et al. Hundreds of Variants clustered in genomic loci and biological pathways affect human height. *Nature*. (2010) 467:832–8. doi: 10.1038/nature09410

38. Oichi T, Taniguchi Y, Soma K, Oshima Y, Yano F, Mori Y, et al. Adamts17 is involved in skeletogenesis through modulation of Bmp-Smad1/5/8 pathway. *Cell Mol Life Sci.* (2019) 76:4795–809. doi: 10.1007/s00018-019-03188-0

39. Oliver J, Forman O, Pettitt L, Mellersh C. Two independent mutations in adamts17 are associated with primary open angle glaucoma in the basset hound and basset fauve de bretagne breeds of dog. PLoS One. (2015) 10:e0140436. doi: 10.1371/journal.pone.0140436 doi: 10.1002/ajmg.a.3186139