Check for updates

#### **OPEN ACCESS**

EDITED BY John William Larkin, Fresenius Medical Care, United States

REVIEWED BY Jia Rao, Fudan University, China Na Liu, Tongji University, China

\*CORRESPONDENCE Wei Fang fangwei\_sh@126.com

<sup>†</sup>These authors share first authorship

SPECIALTY SECTION This article was submitted to Nephrology, a section of the journal Frontiers in Medicine

RECEIVED 16 December 2021 ACCEPTED 18 July 2022 PUBLISHED 10 August 2022

#### CITATION

Song Q, Yang X, Shi Y, Yan H, Yu Z, Li Z, Yuan J, Ni Z, Gu L and Fang W (2022) High intraperitoneal interleukin-6 levels predict ultrafiltration (UF) insufficiency in peritoneal dialysis patients: A prospective cohort study. *Front. Med.* 9:836861. doi: 10.3389/fmed.2022.836861

#### COPYRIGHT

© 2022 Song, Yang, Shi, Yan, Yu, Li, Yuan, Ni, Gu and Fang. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

## High intraperitoneal interleukin-6 levels predict ultrafiltration (UF) insufficiency in peritoneal dialysis patients: A prospective cohort study

Qianhui Song<sup>1,2†</sup>, Xiaoxiao Yang<sup>1,2</sup>, Yuanyuan Shi<sup>1,2</sup>, Hao Yan<sup>1,2</sup>, Zanzhe Yu<sup>1,2</sup>, Zhenyuan Li<sup>1,2</sup>, Jiangzi Yuan<sup>1,2</sup>, Zhaohui Ni<sup>1,2</sup>, Leyi Gu<sup>1,2</sup> and Wei Fang<sup>1,2</sup>\*

<sup>1</sup>Department of Nephrology, Renji Hospital, School of Medicine, Shanghai Jiao Tong University, Shanghai, China, <sup>2</sup>Shanghai Center for Peritoneal Dialysis Research, Shanghai, China

**Introduction:** UF insufficiency is a major limitation in PD efficiency and sustainability. Our study object to investigate the efficacy of intraperitoneal inflammation marker, IL-6 level as a predictor of UF insufficiency in continuous ambulatory peritoneal dialysis (CAPD) patients.

**Methods:** Stable prevalent CAPD patients were enrolled in this prospective study. IL-6 concentration in the overnight effluent was determined and expressed as the IL-6 appearance rate (IL-6 AR). Patients were divided into two groups according to the median of IL-6 AR and prospectively followed up until death, transfer to permanent HD, recovery of renal function, kidney transplantation, transfer to other centers, lost to follow-up or to the end of study (January 31, 2021). Factors associated with UF capacity as well as dialysate IL-6 AR were assessed by multivariable linear regression. Cox proportional hazards model was used to examine the association between dialysate IL-6 AR and UF insufficiency.

**Results:** A total of 291 PD patients were enrolled, including 148 males (51%) with a mean age of 56.6  $\pm$  14.1 years and a median PD duration of 33.4 (12.7–57.5) months. No correlation was found between dialysate IL-6 AR and UF capacity at baseline. PD duration was found positively correlated with baseline dialysate IL-6 AR, while 24h urine volume was negatively correlated with baseline dialysate IL-6 AR (P < 0.05). By the end of study, UF insufficiency was observed in 56 (19.2%) patients. Patients in the high IL-6 AR group showed a significantly inferior UF insufficiency-free survival when compared with their counterparts in the low IL-6 AR group (P = 0.001). In the multivariate Cox regression analysis, after adjusting for DM, previous peritonitis episode and 24h urine volume, higher baseline dialysate IL-6 AR (HR 3.639, 95% CI 1.776–7.456, P = 0.002) were associated with an increased risk of UF insufficiency. The area

under the ROC curve (AUC) for baseline IL-6 AR to predict UF insufficiency was 0.663 (95% CI, 0.580–0.746; P < 0.001).

**Conclusion:** Our study suggested that the dialysate IL-6 AR could be a potential predictor of UF insufficiency in patients undergoing PD.

#### KEYWORDS

interleukin-6, inflammation, peritoneal dialysis, ultrafiltration capacity, ultrafiltration insufficiency

### Introduction

Peritoneal dialysis (PD) is a well-established and highly cost-effective treatment modality for patients with end-stage kidney disease (ESKD). Compared to hemodialysis (HD), PD achieves similar outcomes but the drop-out rate remains high (1). UF insufficiency (previously named UF failure) is a main cause for PD discontinuation, and is also associated with poor outcome of PD patients (2, 3). UF insufficiency can be present shortly after the onset of PD. Approximately 4% of incident PD patients developed early UF insufficiency (<2 years), has been suggested previously (4). Studies of peritoneal structure and function indicate that early UF insufficiency is always associated with peritoneal small-solute transport rate (PSTR) related to increased density of the peritoneal microvasculature. The prevalence of late UF insufficiency (>2 years) has been reported to be 21 and 36% in long-term PD patients (4, 5). Increasing evidence show that acquired UF insufficiency is not only associated with fast PSTR, but also related to decreased osmotic conductance to glucose (OCG) due to scarring of the vessels and interstitium (6, 7). The pathologic mechanism underlying such alterations remain unknown, but appears to be related to chronic inflammation induced by continuous exposure to bioincompatible PD fluids (PDF) and uremia, possibly exacerbated by episodes of peritonitis (8-10).

Interleukin-6 (IL-6) is a key player in modulating inflammation. IL-6 is a chief stimulator of the production of most acute phase proteins in response to various stimuli, and also plays an important role in regulating the transition from acute to chronic inflammation (11, 12). It is well-established that local high production of IL-6 is related to a persistent low-grade inflammation in the peritoneal cavity and its level increased with therapy duration (13, 14). There is increasing evidence that dialysate IL-6 level is closely associated with baseline PSTR (15-17), and it has been shown that IL-6 polymorphisms were related to inherent high PTSR in patients undergoing PD (18, 19). Moreover, we have previously reported that high intraperitoneal IL-6 levels at baseline were a predictor of increasing PSTR after 12 months follow-up in PD patients (20). The above results suggest that IL-6 not only correlates with PSTR at the start of PD, but also can affect the alteration of PSTR during prolonged PD treatment. However, whether

intraperitoneal IL-6 level can predict the UF capacity still remain unclear.

Therefore, we conducted the present study to investigate the association between baseline dialysate IL-6 AR and UF insufficiency in patients undergoing PD.

### Materials and methods

### Study population

Stable CAPD patients in Renji Hospital, School of Medicine, Shanghai Jiao Tong University, between January 2014 and April 2015 were recruited in present study. Exclusion criteria included: (1) presence of systemic inflammatory disease including chronic autoimmune disorders, peritonitis or acute infections that requires antibiotic therapy in the preceding 3 months; (2) malignancy; (3) taking glucocorticoid or immunosuppressive agents during the past 1 year; (4) acute cardiocerebrovascular events that occurred within 3 months prior to the study; (5) patients with PD catheter malfunction and/or fluid leaks; (6) patients with UF insufficiency; (7) patients who refused to give consent. All enrolled patients were dialyzed using glucose-based PD solutions (Dianeal<sup>(K)</sup>, Baxter). The protocol of study was approved by the Ethics Committee of Renji Hospital, School of Medicine, Shanghai Jiao Tong University, China (number: [2013] N022; year: January/2014). All the participants signed informed consent before enrollment.

### Data collection and patient evaluation

The demographic characteristics collected at the enrollment of study included: age, gender, height, weight, date of PD initiation, underlying cause of ESKD, presence of comorbid diseases such as diabetes mellitus (DM) and cardiovascular disease (CVD), taking angiotensin-converting enzyme inhibitor/ angiotensin receptor blocker (ACEI/ARB) or not. CVD was defined as a previous history of coronary artery disease, peripheral vascular disease or cerebrovascular disease. The body surface area (BSA) was calculated by the Du Bois equation (21). The historical dialysis regimen was collected to calculate the amount of historical glucose exposure according to Davies et al. (22). At study enrollment, we also measured serum albumin, high sensitivity CRP (hs-CRP) and hemoglobin levels of each patient, and blood pressure was measured twice at an interval of 5 min to take the average.

At enrollment, all patients were asked to perform a standard peritoneal equilibration test (PET) as originally described by Twardowski (23) and PD effluent (PDE) samples were collected at baseline. The detailed procedure for undertaking a standard PET is shown in Supplementary material 1. On the night prior to PD center visit, patients did a dialysis exchange using his or her usual overnight dialysis regimen. The overnight effluent was fully drained the next morning in the PD center. A 10-mL sample was collected from the drained PDF and immediately stored at  $-80^{\circ}$ C. We weighed the bag of drained effluent to assess the volume and recorded the dwell duration. The 24h ultrafiltration and 24h urine output of each patient were measured. UF capacity was estimated based on the net negative balance (weighing the bag after drainage) after a 2 L 2.5% glucose exchange with 4 h of dwell time in the PET. Residual renal function (RRF) was calculated as an average of 24h urine urea and creatinine clearance (24, 25). Small solute clearance was assessed by 24-h dialysate and urine collection, with the calculation of total weekly Kt/V and weekly CrCl normalized to  $1.73 \text{ m}^2$  body surface area (26). Mass transfer area coefficient for creatinine (MTACcr) was calculated using the simplified Garred equation (27). A validated correction factor was used to calculate peritoneal protein clearance (Prcl) (28). The IL-6 concentration in the drained PDF were measured at baseline, and patients were divided into 2 groups according to the median of IL-6 AR. All patients re-performed PET at 12 monthly intervals and UF capacity (4h net ultrafiltration, mL) was recorded from baseline to 72 months.

### Determination of IL-6 in PD effluent

The concentration of IL-6 in the drained PDF was determined with enzyme-linked immunosorbent assay (ELISA). All samples were run simultaneously and in duplicate to avoid intra- and inter- assay variations. The IL-6 concentration was measured by Human IL-6 ELISA Ready-SET-Go! (eBioscience<sup>®</sup>, CA, USA). Due to the concentrations of dialysate cytokines were influenced by UF capacity which was affected by peritoneal solute transfer rate and dwell time, the dialysate appearance rate (AR) was calculated as dialysate concentration times the drained volume divided by the dwell time and expressed as pg (ng) per minute [pg (ng)/min].

### Study outcomes

All patients were prospectively followed up until death, transfer to permanent HD, recovery of renal function, kidney

transplantation, transfer to other centers, lost to follow-up or to the end of study (January 31, 2021). The primary outcome measures in our study were UF insufficiency, and the second outcome measures in our study were technique failure. During the study period, all UF insufficiencies and technique failures from patient enrollment to study endpoint were carefully recorded. Other PD outcomes including death, transplant and transfer to other centers were also collected. In our study, UF insufficiency was defined according to the International Society for Peritoneal Dialysis guidelines: that is, net UF from a 4-h PET is <100 mL (2.27% glucose /2.5% dextrose) (29). PD catheter malfunction and/or fluid leaks were ruled out prior to the diagnosis of UF insufficiency, and when signs of catheter malfunction and/or leaks occurred, the cause of catheter dysfunction was determined by some combination of physical examination, abdominal radiography and peritoneography, as required. In patients who developed catheter dysfunction, conservative therapy was given initially: supine position and a lower infusion volume for leaks; abdominal massage, administration of aperients or enemas, or ambulation for malposition; clot dislodgement with heparin or urokinase for obstruction; and administration of aperients or enemas for omental wrap. Technique failure was defined as dialysis modality switch from PD to HD for at least 3 months.

### Statistics analysis

Kolmogorov-Smirnov test was applied to test normal distributions. Data were described by mean  $\pm$  SD, median and inter quartile range (IQR), or proportions as appropriate. Unpaired *t*-tests or Mann-Whitney tests were used to compare different groups depending on whether the data were normally distributed or skewed. Categorical variables were compared using chi-square tests. Multiple linear regression analysis was performed to assess the predictors for UF capacity as well as dialysate IL-6 AR. Kaplan-Meier analysis was used to compare UF insufficiency-free survival time between the low IL-6 AR group and the high IL-6 AR group. Risk factors associated with UF insufficiency were determined by univariate and then by a multivariate Cox proportional hazards model. Only covariates that remained significant (P < 0.05) in the univariate analysis were kept in the multivariate Cox regression model except those with collinearity. Collinearity was assessed using the variance inflation factors method. The area under the curve (AUC) of dialysate IL-6 AR for incident UF insufficiency was calculated by receiver operating characteristic (ROC) analysis. Given the large number of statistical tests performed, P-values were adjusted using the Benjamini-Hochberg (BH) procedure for false discovery rate control in multivariate regression analysis (30, 31). Data analysis was carried out using the SPSS software package (version 22.0, Chicago, IL, USA) and R software (version 3.6.1, Vienna, Austria; the p.adjust function was used to



obtain adjusted  $P\mbox{-value}).$  All probabilities were two-tailed and a p<0.05 indicated significance.

# 0.032]. Other demographic characteristics, laboratory data and peritoneal membrane function were similar between the low IL-6 AR group and high IL-6 AR group (see Table 1).

### Results

## Patient demography and membrane function

The flowchart of patients in the study was shown in Figure 1. Detailed characteristics of the study population at enrollment were summarized in Table 1. Based on inclusion and exclusion criteria, a total of 291 patients were enrolled, representing 65% of the total PD population in our center, including 148 males (51%) with a mean age 56.6  $\pm$  14.1 years and a median PD duration 33.4 (12.7-57.5) months. Among the 291 patients, 73 (25%) patients had diabetes mellitus as comorbidity. UF capacity at enrollment for the entire study cohort was 306.8  $\pm$  124 mL. In comparison to patients with low IL-6 AR, those with high IL-6 AR levels above the median were more likely to have experienced a peritonitis episode in the past (P < 0.05) and had longer PD duration [45.3 (20.4-76.2) vs. 20.3 (5.1-45.5) months, P < 0.001], higher historical glucose exposure [47,862 (40,150, 57,128) vs. 40,150 (32,850, 51,332) g/year, P < 0.001] as well as higher 24h ultrafiltration [650 (298-938) vs. 423 (-25-696) mL, P < 0.001 whereas low IL-6 AR patients had higher urine output [500 (48-1,000) vs. 80 (0-500) mL, P < 0.001] and higher RRF [1.34 (0-3.49) vs. 0.73 (0-2.39) mL/min, P =

### Factors associated with UF capacity

UF capacity (mL) as well as the numbers of patients at each time point (i.e., every 12 months over a total of 72 months) on PD were presented in Figure 2, and the comparison of peritoneal membrane function between patients in the low IL-6 AR group and high IL-6 AR group was shown in Supplementary Table 1. Multivariable linear regression showed that after adjustment for age, gender, PD duration, serum albumin, hs-CRP, historical glucose exposure and previous peritonitis episode, comorbid with diabetes ( $\beta = -0.128$ , P = 0.038), 24h urine volume ( $\beta = -0.192$ , P = 0.006) and dialysate IL-6 AR ( $\beta = -0.144$ , P = 0.022) correlate (inversely) with UF capacity at enrollment (see Table 2). However, this association lost statistical significance after being corrected for multiple testing.

## Factors associated with dialysate IL-6 AR level

The effluent level of IL-6 AR in this cohort was 55.1 (35.1–102.7) pg/min and the CVinter was 132%. Multivariable

#### TABLE 1 Characteristics of the study population (n = 291).

Variable	All PD patients	Low IL-6 AR group	High IL-6 AR group	P-value	
	(n = 291)	(n = 146)	(n = 145)		
Age (years)	$56.4\pm14.1$	$56.3 \pm 14.8$	$56.5\pm13.4$	0.922	
Gender (Male)	148 (51%)	74 (51%)	74 (51%)	0.952	
BSA (m <sup>2</sup> )	$1.62\pm0.17$	$1.62\pm0.17$	$1.62\pm0.16$	0.935	
Systolic pressure (mmHg)	$140\pm21$	$140 \pm 21$	$139\pm22$	0.548	
Diastolic pressure (mmHg)	$87 \pm 13$	$87 \pm 12$	$87\pm13$	0.864	
PD duration (months)	33.4 (12.7–57.5)	20.3 (5.1-45.5)	45.3 (20.4–76.2)	< 0.001	
Underlying renal disease [n (%)]					
Chronic glomerulonephritis	86 (30%)	49 (34%)	37 (26%)	0.133	
Diabetic nephropathy	37 (13%)	16 (11%)	21 (15%)	0.367	
Hypertension	12 (4%)	4 (3%)	8 (6%)	0.233	
Polycystic kidney disease	8 (3%)	4 (3%)	4 (3%)	1.000	
Obstructive nephropathy	4 (1%)	1 (1%)	3 (2%)	0.610	
Others and Unknown	147 (51%)	72 (49%)	75 (52%)	0.681	
Comorbidity $[n (\%)]$					
Diabetes mellitus	73 (25%)	35 (24%)	38 (26%)	0.660	
Hypertension	276 (95%)	138 (95%)	138 (95%)	0.801	
Cardiovascular disease	112 (39%)	61 (42%)	51 (35%)	0.247	
ACEI/ARB taking $[n (\%)]$	155 (53%)	78 (53%)	77 (53%)	0.956	
Previous RRT $[n (\%)]$	2 (0.7%)	1 (0.7%)	1 (0.7%)	1.000	
Previous peritonitis episode [n (%)]	71 (24%)	24 (16%)	47 (32%)	0.002	
Historical glucose exposure	43,800 (33,150,	40,150 (32,850,	47,862 (40,150,	< 0.001	
(g/year)	55,293)	51,332)	57,128)		
Hemoglobin (g/L)	$107.4\pm16.8$	$107.3\pm16.1$	$107.6\pm17.5$	0.888	
Serum albumin (g/L)	$37.2 \pm 4.4$	$37.0\pm4.6$	$37.3\pm4.2$	0.625	
Hs-CRP (mg/L)	2.38 (0.8-6.56)	1.96 (0.76-5.6)	2.89 (0.87-6.91)	0.251	
Dialysis adequacy					
Total Kt/V urea	$1.94\pm0.36$	$2.01\pm0.39$	$1.87\pm0.31$	0.104	
Peritoneal Kt/V urea	$1.58\pm0.36$	$1.54\pm0.37$	$1.62\pm0.35$	0.138	
Renal Kt/V urea	0.18 (0-0.61)	0.29 (0-0.87)	0.16 (0-0.40)	0.076	
Total CrCl (L/week/1.73 m <sup>2</sup> )	$61.2\pm18$	$65.7\pm20.6$	$57.4 \pm 13.9$	0.117	
RRF (mL/min)	0.92 (0-2.89)	1.34 (0-3.49)	0.73 (0-2.39)	0.032	
Urine output, mL/24 h	300 (0-800)	500 (48-1000)	80 (0-500)	< 0.001	
UF, mL/24 h	510 (100-840)	423 (-25-696)	650 (298–938)	< 0.001	
nPCR (g/Kg/day)	$0.88\pm0.18$	$0.91\pm0.18$	$0.85\pm0.18$	0.205	
4h D/Pcr	$0.61\pm0.11$	$0.61\pm0.11$	$0.61\pm0.12$	0.819	
4h UF (mL)	$306.8\pm124$	$318.9 \pm 128.2$	$294.5\pm118.8$	0.094	
MTACcr (mL/min)	7.46 (6.02–9.29)	7.54 (6.14–9.31)	7.36 (5.74–9.43)	0.788	
Prcl (mL/d)	68.6 (52.3-90.4)	68.2 (52.3-88.3)	68.8 (52.6-92.3)	0.571	

Values expressed as mean  $\pm$  standard deviation, median (25–75th percentile), or absolute numbers with percentages [n (%)].

BSA, body surface area; ACEI/ARB, inhibitor/ angiotensin receptor blocker; RRT, renal replacement therapy; Hs-CRP, high-sensitivity high sensitivity C-reactive protein; Kt/Vurea, urea kinetics; CrCl, creatinine clearance; RRF, residual renal function; nPCR, normalized protein catabolic rate; D/Pcr, peritoneal transport characteristics; UF, ultrafiltration; MTACcr, mass transfer area coefficient for creatinine; Prcl, peritoneal protein clearance.

linear regression showed that after adjustment for age, gender, DM, serum albumin, hs-CRP, previous peritonitis episode and historical glucose exposure, PD duration ( $\beta = 0.176$ , P = 0.049) had a positive correlation with baseline dialysate

IL-6 AR, while 24h urine volume ( $\beta = -0.235$ , P < 0.001) had a negative correlation with baseline dialysate IL-6 AR after being corrected for multiple testing (see Table 3).



TABLE 2 Association of log<sub>10</sub> dialysate IL-6 AR and 24h urine volume with UF capacity at baseline.

	Unit increase	Standardized $\beta$ coefficient	t	P-value	<i>P</i> -value <sup>a</sup>
Model constant			3.698	0.000	
Age	1 year	0.020	0.316	0.752	0.823
Gender	-	0.030	0.499	0.618	0.772
PD duration	1 year	-0.085	-1.163	0.246	0.492
DM	-	-0.128	-2.088	0.038	0.127
Historical glucose exposure	1,000 g/year	-0.035	-0.604	0.546	0.772
Previous peritonitis episode	-	0.015	0.224	0.823	0.823
Plasma albumin	1 g/L	0.078	1.261	0.208	0.492
Log <sub>10</sub> hs-CRP	10 mg/L	0.062	1.034	0.302	0.503
24h urine volume	100 ml	-0.192	-2.768	0.006	0.060
Log <sub>10</sub> dialysate IL-6 AR	10 pg/min	-0.144	-2.295	0.022	0.110

<sup>a</sup>BH-adjusted P-value.

BSA, body surface area; DM, diabetes; Hs-CRP, high-sensitivity high sensitivity C-reactive protein; AR, appearance rate; BH, Benjamini-Hochberg.

TABLE 3 Association of 24h urine volume with log<sub>10</sub> dialysate IL-6 AR at baseline.

	Unit increase	Standardized $\beta$ coefficient	Т	P-value	<i>P</i> -value <sup>a</sup>
Model constant			8.313	0.000	
Age	1 year	0.014	0.226	0.822	0.822
Gender	-	-0.087	-1.539	0.125	0.375
PD duration	1 year	0.176	2.552	0.011	0.049
DM	-	0.041	0.712	0.477	0.821
Historical glucose exposure	1,000 g/year	0.074	1.339	0.182	0.410
Previous peritonitis episode	-	0.029	0.449	0.654	0.822
Plasma albumin	1 g/L	-0.035	-0.603	0.547	0.821
Log <sub>10</sub> hs-CRP	10 mg/L	0.013	0.236	0.814	0.822
24h urine volume	100 ml	-0.235	-3.641	<0.001	< 0.001

<sup>a</sup>BH-adjusted P-value.

BSA, body surface area; DM, diabetes; Hs-CRP, high-sensitivity high sensitivity C-reactive protein; BH, Benjamini-Hochberg.

Variable	All PD patients (n = 291)	Low IL-6 AR group ( <i>n</i> = 146)	High IL-6 AR group $(n = 145)$	P-value
Follow-up (months)	50.8 (24.9, 78.4)	52.5 (24.5, 79.2)	47.5 (25.2, 76.1)	0.545
UF insufficiencies, n (%)	56 (19.2)	18 (12.3)	38 (26.2)	0.003
Technique failure, n (%)	62 (21.3)	27 (18.5)	35 (24.1)	0.240
Peritonitis	30 (48.4)	15 (55.6)	15 (42.9)	1.000
UF insufficiency	17 (27.4)	2 (7.4)	15 (42.9)	0.001
Personal preferences	8 (12.9)	6 (22.2)	2 (5.7)	0.282
EPS	3 (4.8)	0	3 (8.6)	0.122
Retroperitoneal leak	2 (3.2)	2 (7.4)	0	0.498
Unknown	2 (3.2)	2 (7.4)	0	0.498
Other outcome, n (%)				
Death	102 (35.1)	50 (34.2)	52 (35.9)	0.773
Still on PD	85 (29.2)	48 (32.9)	37 (25.5)	0.167
Transplant	32 (11.0)	18 (12.3)	14 (9.7)	0.517
Transfer to other centers	9 (3.1)	3 (2.1)	6 (4.1)	0.335
Lost to follow-up	1 (0.3)	0	1 (0.7)	1.000

TABLE 4 Follow-up and outcome events in the patients by IL-6 AR.

Non-parametric data were compared using Mann-Whitney test and categorical variables were compared using chi-square tests.

AR, appearance rate; PD, peritoneal dialysis; EPS, encapsulating peritoneal sclerosis.



## Dialysate IL-6 AR level and outcome events

Patient outcomes were summarized in Table 4. The median follow-up duration was 52.5 (IQR 24.5–79.2) months for the low IL-6 AR group and 47.5 (IQR 25.2–76.1) months for the high IL-6 AR group, respectively. During the study period,

UF insufficiency was documented in 56 (19.2%) patients (low IL-6 AR n = 18; high IL-6 AR n = 38). The time to diagnosis UF insufficiency was 43.3 (IQR 22.3-77) months which corresponding to PD duration of 77.4 (IQR 42.2-101.9) months in the low IL-6 AR group, while in the high IL-6 AR group, the time to diagnosis UF insufficiency was 36.4 (IQR 17.7-70.9) months which corresponding to PD duration of 88.4 (IQR 56.8-128.6) months. UF insufficiency was more likely to be observed in patients in the high IL-6 AR group when compared with patients in the low IL-6AR group (P = 0.003, Table 4). In addition, baseline dialysate IL-6 AR levels were significantly higher in patients who experienced UF insufficiency at any time during the study period when compared to their counterparts who remained their UF capacity (P < 0.001, Figure 3). By the end of study, 62 (21.3%) patients (low IL-6 AR n = 27; high IL-6 AR n = 35) experienced technique failure. The leading cause of technique failure was peritonitis (48.4%), followed by UF insufficiency (27.4%), personal preferences (12.9%), encapsulating peritoneal sclerosis (4.8%), retroperitoneal leak (3.2%), and unknown reasons (3.2%). High IL-6 AR patients experienced more technique failures due to UF insufficiency compared to low IL-6 AR patients (P = 0.001, Table 4), and there was no significant difference of other causes in these two groups.

## Association between dialysate IL-6 AR level and UF insufficiency

As shown in Figure 4, patients in low IL-6 AR group had better UF insufficiency-free survival than that in patients with high IL-6 AR (Log-rank  $X^2 = 11.118$ , P = 0.001). UF



TABLEE	D 11 1	<		~		
IABLE 5	Predictors	Of UF	insufficiency	on Cox	regression analysis	i

	Univariate		Multivariate			
	HR (95% CI)	P-value	HR (95% CI)	P-value	<i>P</i> -value <sup>a</sup>	
Age (per year)	1.018 (0.996, 1.040)	0.109				
Gender (male)	0.820 (0.482, 1.395)	0.464				
BSA (per 1 m <sup>2</sup> )	2.104 (0.414, 10.678)	0.370				
PD duration (per year)	1.004 (0.999, 1.010)	0.138				
DM (yes)	1.808 (1.006, 3.250)	0.048	1.711 (0.948, 3.086)	0.074	0.148	
CVD (yes)	0.991 (0.574, 1.912)	0.975				
ACEI/ARB taking (yes)	1.415 (0.829, 2.415)	0.204				
Previous peritonitis episode (yes)	1.876 (1.081, 3.255)	0.025	1.409 (0.772, 2.571)	0.263	0.351	
Historical glucose exposure (per 1000 g/year)	1.000 (0.997, 1.002)	0.702				
Serum albumin (per 10 g/L)	0.970 (0.912, 1.033)	0.345				
Hemoglobin (per 10 g/L)	0.992 (0.976, 1.008)	0.318				
Log <sub>10</sub> hs-CRP (per 10 mg/L)	1.261 (0.807, 1.971)	0.308				
24h urine volume (per 100 mL)	0.999 (0.999, 1.000)	0.024	1.000 (0.999, 1.000)	0.620	0.620	
Log <sub>10</sub> dialysate IL-6 AR (per 10 pg/min)	4.503 (2.329, 8.707)	< 0.001	3.639 (1.776, 7.456)	< 0.001	0.002	

<sup>a</sup>BH-adjusted P-value.

HR, hazard ratio; CI, confidence interval; BSA, body surface area; DM, diabetes; CVD, cardiovascular disease; ACEI/ARB, inhibitor/angiotensin receptor blocker; Hs-CRP, high-sensitivity high sensitivity C-reactive protein; AR, appearance rate; BH, Benjamini-Hochberg.

insufficiency-free survival was 98, 91, and 84% at 1, 3, and 5 years in the low IL-6 AR group and 95, 82, and 66% at 1, 3, and 5 years in the high IL-6 AR group, respectively. Multivariate

Cox proportional hazards modeling showed that after adjusting for DM, previous peritonitis episode and 24h urine volume, higher baseline dialysate IL-6 AR levels (HR 3.639, 95% CI



1.776–7.456, P = 0.002) were associated with an increased risk of UF insufficiency after being corrected for multiple testing (see Table 5). As shown in Figure 5, the area under the ROC curve (AUC) of the model used for baseline IL-6 AR to predict UF insufficiency was 0.663 (95% CI, 0.580–0.746; P < 0.001). The optimal cut-off value to discriminate UF insufficiency was 59.6 pg/mL for dialysate IL-6 AR (67.9% sensitivity, 58.7% specificity, P < 0.001).

### Discussion

To the best of our knowledge, this is the first study suggesting the efficacy of a single measurement of intraperitoneal IL-6 level in predicting development of UF insufficiency in patients undergoing PD.

UF insufficiency is a common and important, but poorlyexplained complication of PD, especially in long-term patients. It has been reported that UF insufficiency accounts for  $\sim 30\%$ of all cases of technique failure (9). The impaired UF points to the development of peritoneal membrane changes during long-term PD treatment. The typical anatomic alterations included submesothelial fibrosis, hyalinizing vasculopathy, and neoangiogenesis (6). Functionally, the peritoneal changes are reflected by an increase in small solute transfer and a reduction in the osmotic conductance of the membrane (8, 9). Current insights indicate that the changes in peritoneal structure and function involve a number of intertwined pathophysiologic mechanisms. Chronic inflammation is the most likely culprit (32-34). IL-6 is a critical biomarker of ongoing intraperitoneal inflammation in PD patients (35, 36). It has been reported that intraperitoneal IL-6 is a strong predictor of increasing PSTR in

PD patients (16, 20). However, whether IL-6 level in dialysate was related to UF capacity decline needs further study.

Our study showed that dialysate IL-6 AR had a skewed distribution with wide range (11.8-1,295 pg/min) in our cohort and the CVinter was 132% in effluent IL-6-AR. In line with our study, Barreto et al. (37) reported similar results. In their study, the dialysate IL-6 AR levels of patients with short ( $\leq 24$ months) and long ( $\geq$ 25 months) PD duration were 15.5–220.0 and 6.9-956.4 pg/min, respectively, with a CVinter 141%. We found that patients with high IL-6 AR were more likely to be anuric, more prevalent in previous episodes of peritonitis, and with a longer PD duration, higher historical glucose exposure and more ultrafiltration when compared with their counterparts in the low IL-6 AR group. Many factors have been claimed as contributors to peritoneal inflammation during PD, including the bioincompatibility of conventional PD solutions and peritonitis (15, 38). It has been shown that conventional PDF could induce IL-6 synthesis by peritoneal membrane cells (39, 40). An increase in intraperitoneal IL-6 concentrations with longer PD duration (i.e., at 24 months) has also been well-documented by the balANZ trial (median 7.22 vs. 31.35 pg/mL, P < 0.001) (41) and the extension study of the Balnet trial (conventional 47  $\pm$  31.2 vs. 121  $\pm$  69 pg/mL, P < 0.001; biocompatible 57.6  $\pm$  54.5 vs. 143  $\pm$  69.6 pg/mL, P < 0.001) (42). Several studies have found that intraperitoneal IL-6 levels are increased before and during peritonitis and remain high level even several months after clinical cure of peritonitis (35, 43). A retrospective observational study included 31 PD patients had reported that patients who developed peritonitis had higher baseline dialysate IL-6 level (58.4  $\pm$  12.6 vs. 20.3  $\pm$  8.7 pg/mL, p = 0.07) than that in patients who remained peritonitis-free (15). Our findings also indicated that the bioincompatible factors of PDF and peritonitis produce pro-inflammatory milieu in the intra-peritoneal cavity. Deterioration in UF capacity over time have been reported in PD patients and have been mainly attributed to the bio-incompatible nature of conventional PDF (44-46). The onset of a decline in UF capacity has been reported to occur at 2-4 years after PD commencement (5). In fact, several recent studies have demonstrated that peritoneal UF in PD patients may remain stable with time on treatment in relation to use of biocompatible PDFs with neutral pH and low GDP concentration as well as preserved RRF (47-49). Given the changes and the importance of urine volume and membrane transport characteristics, taking these parameters in account might better display the longitudinal change of UF capacity. However, these data were not collected in present study.

In the present study, no significant correlation was found for dialysate IL-6 AR with UF capacity at baseline. In addition, we found that PD duration positively correlated with baseline dialysate IL-6 AR, while 24h urine volume inversely correlated with baseline dialysate IL-6 AR. A number of factors potentially contribute to local peritoneal inflammation. It has been shown that bioincompatible factors in PDF could induce IL-6 synthesis and secretion by peritoneal membrane cells (39, 50). In the NEPP study, patients with a regimen low in glucose and GDPs had significantly lower IL-6 levels in overnight effluents when compared to that in patients with conventional lactate-buffered PDF (sPD regimen) (51). These studies indicated that bioincompatible factors in PDF could directly or indirectly promote local peritoneal inflammation. For patients with less urine output, higher dialysis doses or higher glucose concentrations is required to achieve sufficient solute and fluid removal. Besides, numerous studies have shown that a reduction in RRF may aggravate the chronic inflammatory state due to decreased renal clearance of various inflammatory cytokines (52, 53). Therefore, the negative effects of urine volume on intraperitoneal IL-6 might be explained by more exposure to bio-incompatible PD solutions and less efficient removal of inflammatory cytokines. The use of new, more biocompatible PD solutions and preservation of RRF might be helpful to ameliorate intraperitoneal inflammation.

During the study period, UF insufficiency was observed in 56 (19.2%) patients. UF insufficiency was more likely to be observed in high IL-6 AR group than those in low IL-6 AR group. In accordance with other studies (2, 4), peritonitis and UF insufficiency are the two main reasons for technique failure in our PD cohort. Furthermore, patients with high IL-6 AR were more likely to occur technique failure due to UF insufficiency when compared to their counterparts in low IL-6 AR group. The dialysate IL-6 AR were significantly higher in patients who underwent UF insufficiency when compared to those who remained their UF capacity. Consistent with our findings, Xiao et al. (54) also reported that the levels of IL-6 (either plasma or dialysate) in patients with UF insufficiency were significantly higher than those in patients without UF insufficiency. Our study showed that dialysate IL-6 AR could identify UF insufficiency with optimal cut-off values of 59.6 pg/mL. However, the AUC for IL-6 AR was rather weak, suggesting that it was probably not the ideal biomarker as a prognostic indicator of UF insufficiency. The underlying causes might be inter-individual variation of IL-6, lack of repeated measurements and increased peritoneal concentrations could be due to mesothelial cell injury by repeated exposures to bioincompatible PDF but also secondary to peritonitis episodes. Moreover, whether mesothelial cell injury was a prerequisite for UF insufficiency need further validation. Peritoneal biopsy studies also showed that chronic exposure of the peritoneum to conventional PDF was associated with loss of mesothelial cell monolayer (55, 56), which may result in persistently low dialysate IL-6 level in some patients with long-term PD due to reduced IL-6 synthesis and secretion. These factors may account for the poor discriminative potential for IL-6 AR to identify UF insufficiency. Also, we found that low IL-6 AR patients showed better UF insufficiency-free survival than high IL-6 AR patients. Furthermore, high baseline dialysate IL-6 AR was the only risk

factor associated with UF insufficiency, while PD duration and historical glucose loads was not associated with UF insufficiency in this population. These results points to the differences that occur between patients in the course of inflammatory reaction triggered by chronic exposure to bioincompatible PDF and subsequent adverse peritoneal remodeling. There are few studies trying to find out the risk factors of UF insufficiency in PD patients. Selgas et al. (57) reported that diabetic state and higher glucose requirement to obtain adequate UF might be responsible for UF insufficiency. However, their study did not measure inflammation markers, such as CRP or IL-6. Taken together, although there was no significant correlation between IL-6 AR and UF capacity at baseline, we found that the intraperitoneal inflammation marker, IL-6 AR level rather than time on PD could predict subsequent development of UF insufficiency.

UF capacity in PD patients has been considered to be dependent on two major peritoneal components: the microcirculation and the interstitial tissue. Biopsy specimens taken from PD patients have confirmed that an increase in peritoneal vascular density in conjunction with submesothelial and perivascular fibrosis were observed in patients with membrane failure (6-8). It has long been recognized that the formation of new vessels will increase the vascular surface area, leading to rapid dissipation of the osmotic gradient and lower UF (9). Our prior study also showed that a decline in UF capacity could be partially abrogated by antiangiogenic therapy in a rat model (58). There is strong evidence that peritoneal angiogenesis is closely linked with inflammation through local IL-6. Yang et al. (50) showed that IL-6 trans-signaling could upregulate the protein expression and secretion of vascular endothelial growth factor (VEGF) in mesothelial cells. Catar et al. (59) also reported that IL-6 links inflammation with angiogenesis through the trans-signaling pathway to upregulate mesothelial VEGF production in the peritoneal membrane. In addition, the deposition of interstitial fibers will increase resistance to fluid flux, resulting in a decrease in water flow through the interstitium (6). Recent findings support the notion that chronic inflammation plays an important role in the initiation and progression of interstitial fibrosis in the peritoneal membrane (9, 10). Animal models of PD suggested that IL-6 signaling in recurrent peritoneal inflammation was key driving tissue fibrosis (60). In our previous study, we reported that blockade of IL-6 trans-signaling could attenuate peritoneal fibrosis by inhibiting the activation of Smad2/3, with reduced expressions of  $\alpha$ -smooth muscle actin ( $\alpha$ -SMA) and collagen type I (Col I) in a mouse model (61). Furthermore, Lambie et al. (62) showed that dialysate IL-6 levels were independently associated with the occurrence of encapsulating peritoneal sclerosis (EPS). Therefore, IL-6 may serve as a new non-invasive biomarker and a potential therapeutic target of membrane failure, and may guide clinical decisions such as timely transfer to HD in PD patients.

However, the conclusions that can be drawn from the present study are challenged by several limitations. First, this was a single center study, therefore, our results might not be generalized to other populations. Second, the cytokine levels were measured only once at enrollment. The study design did not account for potential variations of IL-6, thus a single IL-6 measurement as a predictor of outcome should be interpreted cautiously. Also, a single time point of effluent IL-6 may not reflect changes over time. The presence of longitudinal surveillance of effluent IL-6 level in PD patients and repeated assessments may provide more solid information. Thirdly, although the reasons and distributions of the informative censoring between the two groups were same, the inclusion of prevalent PD patients and high censorship rates might introduce bias in our study. Also, in the multivariate regression model, despite adjustment for historical glucose exposure at baseline, the differing observation periods might introduce bias in our study and the association between IL-6 AR and the risk of UF insufficiency might be exaggerated because the HRs would be overestimated. In addition, there might be residual confounding because of the unfeasibility to address differing observation periods in the analysis. Therefore, a well-designed, adequately powered multicenter randomized controlled clinical trial (RCT) is required to confirm the association between dialysate IL-6 AR and UF insufficiency.

In conclusion, the results from the present study suggest the intraperitoneal inflammation marker, dialysate IL-6 AR level could be a predictor of UF insufficiency in patients undergoing PD.

### Data availability statement

The original contributions presented in the study are included in the article/Supplementary material, further inquiries can be directed to the corresponding author.

### **Ethics statement**

The studies involving human participants were reviewed and approved by Ethics Committee of Renji Hospital, School of Medicine, Shanghai Jiao Tong University, China (number: [2013] N022; year: January/2014). The patients/participants provided their written informed consent to participate in this study. Written informed consent was obtained from the individual(s) for the publication of any potentially identifiable images or data included in this article.

### Author contributions

QS participated in the design of the study, analysis of data, and draft the manuscript. XY, YS, HY, and ZL participated in clinical data collection. ZY and JY helped to perform the statistical analysis. ZN and LG guided and supported this study. WF conceived of the study and participated in its design and coordination and helped to draft the manuscript. All authors contributed to the article and approved the submitted version.

### Funding

This work was supported by the National Basic Research Program of China (Grant Nos. 81370864 and 81670691) and Shanghai Municipal Education Commission-Gaofeng Clinical Medicine (Grant No. 20152211).

### **Acknowledgments**

We thank all of the doctors and nurses at the Nephrology Department of Renji Hospital in Shanghai, China for their work.

### **Conflict of interest**

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

### Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

### Supplementary material

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/ fmed.2022.836861/full#supplementary-material

### References

1. van de Luijtgaarden MW, Jager KJ, Segelmark M, Pascual J, Collart F, Hemke AC, et al. Trends in dialysis modality choice and related patient survival in the ERA-EDTA Registry over a 20-year period. *Nephrol Dial Transplant.* (2016) 31:120–8. doi: 10.1093/ndt/gfv295

2. Chaudhary K. Peritoneal dialysis drop-out: causes and prevention strategies. Int J Nephrol. (2011) 2011:434608. doi: 10.4061/2011/434608

3. Kawaguchi Y, Ishizaki T, Imada A, Oohira S, Kuriyama S, Nakamoto H, et al. Searching for the reasons for drop-out from peritoneal dialysis: a nationwide survey in Japan. *Perit Dial Int.* (2003) 23:S175-7. doi: 10.1177/089686080302302s36

4. Sampimon DE, Coester AM, Struijk DG, Krediet RT. The time course of peritoneal transport parameters in peritoneal dialysis patients who develop encapsulating peritoneal sclerosis. *Nephrol Dial Transplant.* (2011) 26:291–8. doi: 10.1093/ndt/gfq343

5. Smit W, Schouten N, van den Berg N, Langedijk MJ, Struijk DG, Krediet RT, et al. Analysis of the prevalence and causes of ultrafiltration failure during long-term peritoneal dialysis: a cross-sectional study. *Perit Dial Int.* (2004) 24:562–70. doi: 10.1177/089686080402400616

6. Krediet RT. Ultrafiltration failure is a reflection of peritoneal alterations in patients treated with peritoneal dialysis. *Front Physiol.* (2018) 9:1815. doi: 10.3389/fphys.2018.01815

7. Krediet RT. Acquired decline in ultrafiltration in peritoneal dialysis: the role of glucose. J Am Soc Nephrol. (2021) 32:2408–15. doi: 10.1681/ASN.2021010080

8. Krediet RT, Lindholm B, Rippe B. Pathophysiology of peritoneal membrane failure. *Perit Dial Int.* (2000) 20:S22–42. doi: 10.1177/089686080002004S03

9. Davies SJ, Mushahar L, Yu Z, Lambie M. Determinants of peritoneal membrane function over time. *Semin Nephrol.* (2011) 31:172–82. doi: 10.1016/j.semnephrol.2011.01.006

10. Li PK, Ng JK, Mcintyre CW. Inflammation and peritoneal dialysis. *Semin Nephrol.* (2017) 37:54–65. doi: 10.1016/j.semnephrol.2016.10.007

11. Hurst SM, Wilkinson TS, McLoughlin RM, Jones S, Horiuchi S, Yamamoto N, et al. 11-6 and its soluble receptor orchestrate a temporal switch in the pattern of leukocyte recruitment seen during acute inflammation. *Immunity.* (2001) 14:705–14. doi: 10.1016/S1074-7613(01)00151-0

12. Gabay C. Interleukin-6 and chronic inflammation. *Arthritis Res Ther.* (2006) 8:S3. doi: 10.1186/ar1917

13. Pecoits-Filho R, Araújo MR, Lindholm B, Stenvinkel P, Abensur H, Romão JE, et al. Plasma and dialysate IL-6 and VEGF concentrations are associated with high peritoneal solute transfer rate. *Nephrol Dial Transplant.* (2002) 17:1480–6. doi: 10.1093/ndt/17.8.1480

14. Cooker LA, Luneburg P, Holmes CJ, Jones S, Topley N, Bicarbonate/Lactate Study Group. Interleukin-6 levels decrease in effluent from patients dialyzed with bicarbonate/lactatebased peritoneal dialysis solutions. *Perit Dial Int.* (2001) 21:S102–7. doi: 10.1177/089686080102103S17

15. Pecoits-Filho R, Carvalho MJ, Stenvinkel P, Lindholm B, Heimbürger O. Systemic and intraperitoneal interleukin-6 system during the first year of peritoneal dialysis. *Perit Dial Int.* (2020) 26:53–63. doi: 10.1177/089686080602600109

16. Oh KH, Jung JY, Yoon MO, Song A, Lee H, Ro H, et al. Intra-peritoneal interleukin-6 system is a potent determinant of the baseline peritoneal solute transport in incident peritoneal dialysis patients. *Nephrol Dial Transplant.* (2010) 25:1639–46. doi: 10.1093/ndt/gfp670

17. Cho JH, Hur IK, Kim CD, Park SH Ryu HM, Yook JM, et al. Impact of systemic and local peritoneal inflammation on peritoneal solute transport rate in new peritoneal dialysis patients: a 1-year prospective study. *Nephrol Dial Transplant*. (2010) 25:1964–73. doi: 10.1093/ndt/gfp767

18. Gillerot G, Goffin E, Michel C, Evenepoel P, Biesen WV, Tintillier M, et al. Genetic and clinical factors influence the baseline permeability of the peritoneal membrane. *Kidney Int.* (2005) 67:2477–87. doi: 10.1111/j.1523-1755.2005.00357.x

19. Ding L, Shao X, Cao L, Fang W, Yan H, Huang J, et al. Possible role of IL-6 and TIE2 gene polymorphisms in predicting the initial high transport status in patients with peritoneal dialysis: an observational study. *BMJ Open.* (2016) 6:e012967. doi: 10.1136/bmjopen-2016-012967

20. Yang X, Zhang H, Hang Y, Yan H, Lin A, Huang J, et al. Intraperitoneal interleukin-6 levels predict peritoneal solute transport rate: a prospective cohort study. *Am J Nephrol.* (2014) 39:459–65. doi: 10.1159/000362622

21. Du Bois D, Du Bois EF. A formula to estimate the approximate surface area if height and weight be known. *Arch Intern Med.* (1989) 5:303–11.

22. Davies SJ, Phillips L, Naish PF, Russell GI. Peritoneal glucose exposure and changes in membrane solute transfer with time on peritoneal dialysis. J Am Soc Nephrol. (2001) 12:1046–51. doi: 10.1681/ASN.V1251046

23. Twardowski ZJ, Nolph KD, Khanna R. Peritoneal equilibration test. *Perit Dial Bull.* (1987) 7:138–47. doi: 10.1177/089686088700700306

24. Van Olden RW, Krediet RT, Struijk DG, Arisz L. Measurement of residual renal function in patients treated with CAPD. J Am Soc Nephrol. (1996) 7:745–50. doi: 10.1681/ASN.V75745

25. National Kidney Foundation. K/DOQI clinical practice guidelines for peritoneal dialysis adequacy, 2000. Am J Kidney Dis. (2001) 37(Suppl.1):S65–136. doi: 10.1016/S0272-6386(01)70006-6

26. Smyth A, McCann E, Redahan L, Lambert B, Mellotte GJ, Wall CA, et al. Peritoneal dialysis in an ageing population: a 10-year experience. *Int Urol Nephrol.* (2012) 44:283–93. doi: 10.1007/s11255-011-9973-2

27. Krediet RT, Boeschoten EW, Zuyderhoudt FM, Strackee J, Arisz L. Simple assessment of the efficacy of peritoneal transfer in continuous ambulatory peritoneal dialysis patients. *Blood Purif*, (1986) 4:194–203. doi: 10.1159/000169445

28. Krediet RT, Zuyderhoudt FM, Boeschoten EW, Arisz L. Peritoneal permeability to proteins in diabetic and non-diabetic continuous ambulatory peritoneal dialysis patients. *Nephron.* (1986) 42:133–40. doi: 10.1159/000183652

29. Morelle J, Stachowska-Pietka J, Oberg C, Gadola L, La Milia V, Yu Z, et al. ISPD recommendations for the evaluation of peritoneal membrane dysfunction in adults: Classification, measurement, interpretation and rationale for intervention. *Perit Dial Int.* (2021) 41:352–72. doi: 10.1177/0896860820982218

30. Benjamini Y, Hochberg Y. Controlling the false discovery rate: a practical and powerful approach to multiple testing. *J R Stat Soc B.* (1995) 57:289–300. doi: 10.1111/j.2517-6161.1995.tb02031.x

31. Tan YD, Xu H. A general method for accurate estimation of false discovery rates in identification of differentially expressed genes. *Bioinformatics*. (2014) 30:2018–25. doi: 10.1093/bioinformatics/btu124

32. Bajo MA, Del Peso G, Teitelbaum I. Peritoneal membrane preservation. Semin Nephrol. (2017) 37:77–92. doi: 10.1016/j.semnephrol.2016.10.009

33. Zhang Z, Jiang N, Ni Z. Strategies for preventing peritoneal fibrosis in peritoneal dialysis patients: new insights based on peritoneal inflammation and angiogenesis. *Front Med.* (2017) 11:349–58. doi: 10.1007/s11684-017-0571-2

34. Krediet RT, Struijk DG. Peritoneal changes in patients on long-term peritoneal dialysis. *Nat Rev Nephrol.* (2013) 9:419–29. doi: 10.1038/nrneph.2013.99

35. Lai KN, Lai KB, Lam CWK, Chan TM, Li FK, Leung JC. Changes of cytokine profiles during peritonitis in patients on continuous ambulatory peritoneal dialysis. *Am J Kidney Dis.* (2000) 35:644–52. doi: 10.1016/S0272-6386(00)70011-4

36. Breborowicz A, Oreopoulos DG. Evidence for the presence of chronic inflammation during peritoneal dialysis: therapeutic implications. *Perit Dial Int.* (1997) 17:S37–41. doi: 10.1177/089686089701702S07

37. Barreto DL, Coester AM, Noordzij M, Smit W, Struijk DG, Rogers S, et al. Variability of effluent cancer antigen 125 and interleukin-6 determination in dialysis patients. *Nephrol Dial Transplant.* (2011) 26:3739-44. doi: 10.1093/ndt/gfr170

38. Baroni G, Schuinski A, de Moraes TP, Meyer F, Pecoits-Filho R. Inflammation and the peritoneal membrane: causes and impact on structure and function during peritoneal dialysis. *Mediators Inflamm.* (2012) 2012:912595. doi: 10.1155/2012/912595

39. Witowski J, Jorres A, Coles GA, Williams JD, Topley N. Superinduction of IL-6 synthesis in human peritoneal mesothelial cells is related to the induction and stabilization of IL-6 mRNA. *Kidney Int.* (1996) 50:1212–23. doi: 10.1038/ki.1996.430

40. Serre AF, Marie C, Beaujon G, Cavaillon JM, Deteix P. Variations of cytokine levels and production in CAPD patients. *Int J Artif Organs.* (1997) 20:614–21. doi: 10.1177/039139889702001104

41. Cho Y, Johnson DW, Vesey DA, Hawley CM, Pascoe EM, Clarke M, et al. Dialysate interleukin6 predicts increasing peritoneal solute transport rate in incident peritoneal dialysis patients. *BMC Nephrol.* (2014) 15:8. doi: 10.1186/1471-2369-15-8

42. Kim S, Oh KH, Oh J, Kim SJ, Chung W, Song YR, et al. Biocompatible peritoneal dialysis solution preserves residual renal function. *Am J Nephrol.* (2012) 36:305–16. doi: 10.1159/000342523

43. Zemel D, Betjes MG, Dinkla C, Struijk DG, Krediet RT. Analysis of inflammatory mediators and peritoneal permeability to macromolecules shortly

before the onset of overt peritonitis in patients treated with CAPD. *Perit Dial Int.* (1995) 15:134–41. doi: 10.1177/089686089501500208

44. Davies SJ, Brown EA, Frandsen NE, Rodrigues AS, Rodriguez-Carmona A, Vychytil A, et al. Longitudinal membrane function in functionally anuric patients treated with APD: data from EAPOS on the effects of glucose and icodextrin prescription. *Kidney Int.* (2005) 67:1609–15. doi: 10.1111/j.1523-1755.2005.00243.x

45. Davies SJ. Longitudinal relationship between solute transport and ultrafiltration capacity in peritoneal dialysis patients. *Kidney Int.* (2004) 66:2437–45. doi: 10.1111/j.1523-1755.2004.66021.x

46. Davies SJ, Bryan J, Phillips L, Russell GI. Longitudinal changes in peritoneal kinetics: the effects of peritoneal dialysis and peritonitis. *Nephrol Dial Transplant*. (1996) 11:498–506. doi: 10.1093/oxfordjournals.ndt.a027318

47. Johnson DW, Brown FG, Clarke M, Boudville N, Elias TJ, Foo MW, et al. The effect of low glucose degradation product, neutral pH versus standard peritoneal dialysis solutions on peritoneal membrane function: the balANZ trial. *Nephrol Dial Transplant*. (2012) 27:4445–53. doi: 10.1093/ndt/gfs14

48. Eibensteiner F, Flores KO, Unterwurzacher M, Herzog R, Kratochwill K, Alper SL, et al. Monitoring daily ultrafiltration in automated peritoneal dialysis. *Clin J Am Soc Nephrol.* (2022) 17:107–10. doi: 10.2215/CJN.08180621

49. Choi HY, Kim DK, Lee TH, Moon SJ, Han SH, Lee JE, et al. The clinical usefulness of peritoneal dialysis fluids with neutral pH and low glucose degradation product concentration: an open randomized prospective trial. *Perit Dial Int.* (2008) 28:174–82. doi: 10.1177/0896860802800214

50. Yang X, Lin A, Jiang N, Yan H, Ni Z, Qian J, et al. Interleukin-6 transsignalling induces vascular endothelial growth factor synthesis partly *via* Janus kinases-STAT3 pathway in human mesothelial cells. *Nephrology*. (2017) 22:150– 8. doi: 10.1111/nep.12746

51. Poole CY, Welten AG, ter Wee PM, Paauw NJ, Djorai AN, Valentijn RM, et al. A peritoneal dialysis regimen low in glucose and glucose degradation products results in increased cancer antigen 125 and peritoneal activation. *Perit Dial Int.* (2012) 32:305–15. doi: 10.3747/pdi.2010.00115

52. Chung SH, Heimbürger O, Stenvinkel P, Qureshi AR, Lindholm B. Association between residual renal function, inflammation and patient survival in new peritoneal dialysis patients. *Nephrol Dial Transplant.* (2003) 18:590–7. doi: 10.1093/ndt/18.3.590

53. Memoli B, Postiglione L, Cianciaruso B, Bisesti V, Cimmaruta C, Marzano L, et al. Role of different dialysis membranes in the release of interleukin-6-soluble receptor in uremic patients. *Kidney Int.* (2000) 58:417–24. doi: 10.1046/j.1523-1755.2000.00181.x

54. Xiao J, Gong Y, Chen Y, Yu D, Wang X, Zhang X, et al. IL-6 promotes epithelial-to-mesenchymal transition of human peritoneal mesothelial cells possibly through the JAK2/STAT3 signaling pathway. *Am J Physiol Renal Physiol.* (2017) 313:F310–F8. doi: 10.1152/ajprenal.00428.2016

55. Williams JD, Craig KJ, Topley N, Von Ruhland C, Fallon M, Newman GR, et al. Morphologic changes in the peritoneal membrane of patients with renal disease. *J Am Soc Nephrol.* (2002) 13:470–9. doi: 10.1681/ASN.V132470

56. del Peso G, Jiménez-Heffernan JA, Selgas R, Remón C, Ossorio M, Fernández-Perpén A, et al. Biocompatible dialysis solutions preserve peritoneal mesothelial cell and vessel wall integrity. A case-control study on human biopsies. *Perit Dial Int.* (2016) 36:129–34. doi: 10.3747/pdi.2014.00038

57. Selgas R, Bajo MA, Castro MJ, del Peso G, Aguilera A, Fernández-Perpén A, et al. Risk factors responsible for ultrafiltration failure in early stages of peritoneal dialysis. *Perit Dial Int.* (2020) 20:631–6. doi: 10.1177/089686080002000609

58. Shi Y, Xiong Y, Lei Y, Li Z, Yan H, Yuan J, et al. Protective effect of COMPangiopoietin-1 on peritoneal vascular permeability and peritoneal transport function in uremic peritoneal dialysis rats. *Am J Transl Res.* (2019) 11:5932–43.

59. Catar R, Witowski J, Zhu N, Lücht C, Derrac Soria A, Uceda Fernandez J, et al. IL-6 trans-signaling links inflammation with angiogenesis in the peritoneal membrane. *J Am Soc Nephrol.* (2017) 28:1188–99. doi: 10.1681/ASN.2015101169

60. Fielding CA, Jones GW, McLoughlin RM, McLeod L, Hammond VJ, Uceda J, et al. Interleukin-6 signaling drives fibrosis in unresolved inflammation. *Immunity*. (2014) 40:40–50. doi: 10.1016/j.immuni.2013.10.022

61. Yang X, Yan H, Jiang N, Yu Z, Yuan J, Ni Z, et al. IL-6 transsignaling drives a STAT3-dependent pathway that leads to structural alterations of the peritoneal membrane. *Am J Physiol Renal Physiol.* (2020) 318:F338– 53. doi: 10.1152/ajprenal.00319.2019

62. Lambie MR, Chess J, Summers AM, Williams PF, Topley N, Davies SJ, et al. Peritoneal inflammation precedes encapsulating peritoneal sclerosis: results from the GLOBAL Fluid Study. *Nephrol Dial Transplant.* (2016) 31:480–6. doi: 10.1093/ndt/gfv440