

## Deep Learning-Based Pathology Image Analysis Enhances Magee Feature Correlation With Oncotype DX Breast Recurrence Score

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**Background:** Oncotype DX Recurrence Score (RS) has been widely used to predict chemotherapy benefits in patients with estrogen receptor-positive breast cancer. Studies showed that the features used in Magee equations correlate with RS. We aimed to examine whether deep learning (DL)-based histology image analyses can enhance such correlations.

**Methods:** We retrieved 382 cases with RS diagnosed between 2011 and 2015 from the Emory University and the Ohio State University. All patients received surgery. DL models were developed to detect nuclei of tumor cells and tumor-infiltrating lymphocytes (TILs) and segment tumor cell nuclei in hematoxylin and eosin (H&E) stained histopathology whole slide images (WSIs). Based on the DL-based analysis, we derived image features from WSIs, such as tumor cell number, TIL number variance, and nuclear grades. The entire patient cohorts were divided into one training set (125 cases) and two validation sets (82 and 175 cases) based on the data sources and WSI resolutions. The training set was used to train the linear regression models to predict RS. For prediction performance comparison, we used independent variables from Magee features alone or the combination of WSI-derived image and Magee features.

**Results:** The Pearson's correlation coefficients between the actual RS and predicted RS by DL-based analysis were 0.7058 (*p*-value =  $1.32 \times 10^{-13}$ ) and 0.5041 (*p*-value =  $1.15 \times 10^{-12}$ ) for the validation sets 1 and 2, respectively. The adjusted  $R^2$  values using Magee features alone are 0.3442 and 0.2167 in the two validation sets,

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respectively. In contrast, the adjusted  $R^2$  values were enhanced to 0.4431 and 0.2182 when WSI-derived imaging features were jointly used with Magee features.

**Conclusion:** Our results suggest that DL-based digital pathological features can enhance Magee feature correlation with RS.

Keywords: deep learning-based algorithm, digital pathology, Oncotype DX score, ER+ breast cancer, Magee equation

#### BACKGROUND

Breast cancer is the most common cancer in women in the United States. Breast cancers are clinically classified by the expression of estrogen receptor (ER), progesterone receptor (PR), and human epidermal growth factor receptor 2 (HER2) gene amplification as ER+/ HER2-, HER2+, and triple-negative (ER-/PR-/HER2-) subtypes. Each subtype has unique tumor biology, treatment options, and prognosis (1-7). Approximately 70% of the breast cancers are ER+/HER2-. Patients with HER2+ and triple-negative breast cancer are generally treated with chemotherapy. However, only a portion of the patient with ER+/HER2- breast cancer benefit from chemotherapy (6, 8-10). Whether patients with ER+/HER2- breast cancer benefit from chemotherapy depends on such clinicopathological features as tumor grade and size, tumor cell proliferation, staging, and molecular profile biomarkers. Before the clinical validation of molecular biomarkers, most patients with high-risk ER+/HER2breast cancer were treated with chemotherapy (11, 12). Oncotype DX Recurrence Score (RS) uses a 21-gene expression profile to predict prognosis and determine the benefit of chemotherapy in patients with ER+/HER2- breast cancer (13-15). The predictive value of RS was validated by large prospective trials and prospective-retrospective studies (14, 15).

The TAILORx trial has validated RS predictive value for patients with ER+/HER2- and lymph node (LN) negative breast cancer. The first publication in 2015 from the TAILORx trial showed that patients with an RS of 0–10 had an excellent prognosis and were highly unlikely to benefit from chemotherapy (16). The second publication from the TAILORx trial showed patients > 50 years old and some young patients ( $\leq$ 50 years old) with a medium RS could be spared from chemotherapy (13). Recent results from the RxPONDER study showed that RS could also predict chemotherapy benefits in patients with ER+/HER2- and 1–3 LN+ breast cancer (17).

Magee equations use routinely available clinicopathological parameters (or Magee features) and are strongly associated with RS (18–20). Furthermore, machine learning-based histology analysis has been shown to correlate with prognosis and behaviors in diseases, including breast cancer (21–26). Therefore, the aim of this study was to examine whether histopathological features from whole slide images (WSIs), when used with Magee features, would improve the RS prediction. Due to the overwhelming gigapixel scale of histopathology WSIs and artifacts in histopathology WSIs, it is technically challenging to extract imaging features with predictive value. Recent applications of artificial intelligence techniques in a large number of biomedical investigations (27–29) show that the deep learning (DL) model can be a potential solution to this challenge. In this study, a DL-based pipeline for WSI analysis was developed to (1) detect the tumor cell nuclei and tumor-infiltrating lymphocyte (TIL) nuclei for cell density evaluation and (2) segment tumor cell nuclei for nuclear-grade assessment. Such large-scale detection and segmentation analyses enable automatic image feature extraction from gigapixel WSIs. We examined whether the image features could enhance the correlation of Magee features with RS.

#### MATERIALS AND METHODS

# Datasets and Clinicopathological Information

Three independent patient cohorts with available RS were collected from two institutions and divided into training and validation sets based on the data sources and WSI resolutions. RS was defined as low ( $\leq$ 15), intermediate (16–25), and high (26–100) according to the results from the TAILORx trial (30). ER, PR, and HER2 interpretations were based on the updated ASCO/CAP recommendations (31, 32). All patients received surgery.

Training set: A total of 125 cases of ER+/HER2-/LN- breast cancer with RS diagnosed from 2011 to 2015 were collected from the Ohio State University. The RS ranged from 0 to 40. Among these 125 cases, 53, 59, and 13 cases had low scores, intermediate scores, and high scores, respectively.

Validation set 1: A total of 82 cases of ER+/HER2-/LN- breast cancer with RS diagnosed from 2012 to 2014 were retrieved from the Emory University. The RS ranged from 0 to 52. Among 82 cases, 40, 15, and 27 cases had low scores, intermediate scores, and high scores, respectively.

Validation set 2: Additional 175 cases of ER+/HER2-/LNbreast cancer with RS diagnosed from 2012 to 2014 were retrieved from the Emory University. The RS in this dataset ranged from 0 to 100. Among 175 cases, 68, 73, and 34 were low-, intermediate-, and high-score cases, respectively.

All three datasets included age at diagnosis, ER and PR IHC staining percentage (0–100) and intensity (1, 2, and 3), HER2 amplification by IHC and FISH (negative and equivocal), Nottingham tumor grade, and tumor size. Additional features retrieved for validation sets 1 and 2 included Ki-67 score, stage, chemotherapy, radiation therapy, overall survival (OS),

Abbreviations: DFS, disease-free survival; DL, deep learning; ER, estrogen receptor; HD, Hausdorff distance; H&E, hematoxylin and eosin; IOU, intersection over union; MRCNN, mask region-based convolutional neural network; OS, overall survival; PR, progesterone receptor; RS, Oncotype DX Recurrence Score; TIL, tumor-infiltrating lymphocytes; WSI, whole-slide image.

disease-free survival (DFS), and distant metastasis (metastasis other than axillary LN metastasis). One representative tumor hematoxylin and eosin (H&E) stained WSI from each case in the training set and validation set 1 was scanned at 40 × magnification and validation set 2 at 20 × with an Aperio AT2 scanner.

The clinicopathological information of these three datasets is summarized in **Table 1**. The ER and PR expressions for all three cohort datasets were evaluated with an H-score (percentage  $\times$  intensity). This study was approved by the Institutional Review Board at the Emory University and the Ohio State University.

#### **Data Preprocessing**

Image normalization: As  $40 \times$  images have a higher resolution for annotations, we chose the  $40 \times$  for data analysis. After linearly resizing with a scaling factor of two along the image width and height directions, all images in validation set 2 had the same magnification of  $40 \times$  as training set and validation set 1. We also used the sparse non-negative matrix factorization-based color transfer method (33) to normalize the image color styles in all three cohort datasets (**Figure 1**).

Data preprocessing for DL training: Although we had three datasets for RS prediction analysis, we used two independent image datasets for cell detection and segmentation training, one from our lab and the other from the public MoNuSeg-2018 dataset. We collected 797 images with tumor nuclei point annotations, 500 images with TIL point annotations, and 26 images with annotations of tumor nuclei contours from the independent dataset. All the annotations were produced and confirmed by the pathologists (Supplementary Figure 1). Two pathologists made the annotations with Aperio ImageScope and GIMP. Additionally, 30 H&E images from the public MoNuSeg-2018 dataset were used in the segmentation dataset. They had annotations of cell nucleus contours (Supplementary Figure 1C). Each DL dataset was randomly divided into training, validation, and testing groups with an approximate proportion of 70:15:15.

#### **Deep Learning Model**

For detection, classification, and segmentation analyses, we used the Mask R-CNN (MRCNN) (34) to construct the image processing models in this project. MRCNN was extended from Faster R-CNN (35) that was in turn developed based on Fast R-CNN (36). The overall schema of the developed WSI image processing pipeline is presented in Figure 2. The DL MRCNN pipeline was constructed with library TensorFlow and Keras. The image processing module contained three MRCNN models specifically for tumor cell detection, TIL detection, and tumor nucleus segmentation, respectively. Image tiles with tissue were extracted from WSIs by thresholding the "Saturation" channel of the HSV color space with the threshold set to 30. Each image tile was then analyzed by three MRCNN models separately. The center of each bounding box is considered the center of a detected cell of interest. The segmentation branch in the MRCNN model produced nucleus contours. Since the tumor cell detection had superior performance, the detected tumor cells were used to exclude the TIL and tumor nucleus false positive. All computational analyses were executed on a computational server with two CPUs of 22 2.10 GHz cores each, 192 GB memory, and six Nvidia GeForce RTX 2080 Ti GPUs with 11 GB memory each.

## Linear Regression Model Incorporating Deep Learning-Based Imaging Features and Magee Equation Variables

We partitioned each WSI into image tiles with a size of  $1,024 \times 1,024$  by pixels to identify tissue regions of high tumor cell density with the DL-based processing pipeline. The top ten image tiles with the highest tumor cell density in each WSI were selected for feature extraction. To generate interpretable models, we chose to select image features of interpretability instead of hidden or intermediate features by machine learning algorithms. Since tumor cells and TILs were reported high correlation with the prognosis or recurrence (37, 38), we extracted three tilewise features from each image tile, including (1) the tumor cell number, (2) the TIL number, and (3) the tumor cell percentage. Additionally, nuclear grade and TIL number variance were extracted from the ten image tiles collectively. The nuclear grade of each tumor cell was determined by comparing the tumor nuclei size with the adjacent TIL nuclei size. The TIL nuclei size was 304.7 in pixels averaged from representative TILs selected by pathologists. Nuclear grade 1 was defined when the ratio of tumor nucleus size to TIL nucleus size was 1-2.5. Nuclear grade 2 was made when such a ratio was 2.5-3.5. Nuclear grade 3 was made when such a ratio was > 3.5 (Supplementary Figure 2). Tumor cell nuclear grades from the ten image tiles were collected and aggregated to a final nuclear grade by the following rules: (1) if  $\geq$  10% of the tumor cells had nuclear grade 3, the aggregated nuclear grade was 3; (2) if  $\geq 10\%$  of the tumor cells had nuclear grade 2 and rule (1) did not hold, the aggregated nuclear grade was 2; (3) if  $\geq 10\%$  of the tumor cells had nuclear grade 1 and neither rule (1) nor (2) held, the final nuclear grade was 1. The image feature of TIL number variance was also computed from the top ten image tiles by cell density as follows:

$$V = \frac{\sum_{i=1}^{10} (n_i - \overline{n})}{10 - 1}$$

where *V* is the TIL number variance;  $n_i$  represents the TIL number in the *i*-th image tile;  $\overline{n}$  is the average TIL number from the ten image tiles. In total, there were 32 image features extracted from each WSI.

A linear regression model was used to correlate with RS. In the regression model, the dependent variable was the RS, while imaging features and Magee features were independent variables. To retain features with high predictive value, we selected features by both domain knowledge and statistical analysis. The independent variables in Magee equations are as follows (39). Magee equation 1 includes Nottingham score, ER and PR H-scores, HER2, tumor size (cm), and Ki67 index; Magee equation 2 includes Nottingham score, ER and PR H-scores, HER2, and Ki67 index. As the feature "HER2" is categorical with two possible values, i.e., "Negative" and "Equivocal," we used one dummy variable, "HER2\_Equivocal," to represent "HER2" in the regression models. We focused on

	Training set	Validation set 1	Validation set 2
Nottingham grade (case number)			
1	33 (26.4%)	31 (37.8%)	64 (36.6%)
2	75 (60.0%)	39 (47.6%)	92 (52.6%)
3	17 (13.6%)	12 (14.6%)	19 (10.8%)
ER intensity (case number)			
0	0 (0.0%)	0 (0.0%)	0 (0.0%)
1	0 (0.0%)	2 (2.4%)	3 (1.7%)
2	9 (7.2%)	9 (11.0%)	33 (18.9%)
3	116 (92.8%)	71 (86.6%)	139 (79.4%)
ER percentage			
Vlean	94.21	89.63	87.59
Range	40–100	5–100	10–100
PR intensity (case number)			
)	8 (6.4%)	17 (20.7%)	19 (10.9%)
1	3 (2.4%)	3 (3.7%)	5 (2.9%)
2	30 (24.0%)	16 (19.5%)	38 (21.7%)
3	84 (67.2%)	46 (56.1%)	113 (64.6%)
PR percentage			
Vlean	66.29	55.15	62.3
Range	0–100	0–100	0–100
HER2 (case number)			
Negative	123 (98.4%)	81 (98.8%)	173 (98.9%)
Equivocal positive	2 (1.6%)	1 (1.2%)	2 (1.1%)
Ki-67 score	(Not available)		(105/175 cases available
Mean	N/A	24.26	29.09
Range	N/A N/A	1-100	1-91
	N/A	1-100	1-91
Tumor size (cm)	2.19	1.81	1.64
Mean	0.4–7.8	0.5–5.3	
Range	0.4-7.8	0.5-5.3	0.3–7.1
Age (year)	50.00	<u> </u>	50.07
Mean	58.00	60.29	56.97
	32–82	31–81	30–91
Oncotype DX RS	10.00	10.15	10.00
Mean	16.62	19.15	18.93
Range	0–40	0–52	0–100
Real chemotherapy (case number)		(80/82 cases available)	(168/175 cases available
Yes	N/A	24 (29.3%)	48 (27.4%)
No	N/A	56 (68.3%)	120 (68.6%)
OS (months)		. ,	· ,
Mean	N/A	32.80	81.45
Range	N/A	1–250	0–272
DFS (months)		(4/82 cases available)	(9/175 cases available)
Mean	N/A	75.25	64.74
Range	N/A	3–151	12–174
Real radiation therapy (case number)		(80/82 cases available)	(168/175 cases available
Yes	N/A	49 (59.8%)	99 (56.6%)
No	N/A	31 (37.8%)	69 (39.4%)
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(Continued)

#### TABLE 1 | (Continued)

	Training set	Validation set 1	Validation set 2
Stage (case number)			
1	91 (72.8%)	55 (67.1%)	122 (69.7%)
2	29 (23.2%)	26 (31.7%)	50 (28.6%)
3	5 (4.0%)	1 (1.2%)	3 (1.7%)
Distant metastasis (case number)		(81/82 cases available)	(169/175 cases available)
Yes	N/A	2 (2.4%)	6 (3.4%)
No	N/A	79 (97.6%)	163 (96.6%)



FIGURE 1 | Demonstrations of image color normalization. With the learned color and brightness information from the reference image on the left, three randomly selected images before and after color normalizations are presented on the top and bottom rows on the right.

Magee equation 2 as the Ki-67 index information was missing for more than half samples (195/382, 51.0%) in our datasets. Additionally, the tile-wise features from the first x out of the ten image tiles (x = 1, 2, ..., 10), i.e., the tumor cell number, TIL number, and tumor cell percentage, were used jointly. The feature selection was completed in the training set. Various feature combinations were used to construct the linear regression models. The adjusted coefficient of determination  $R^2$  was used to assess the combinations' correlation with RS. The feature combination with the highest adjusted  $R^2$  was selected for the final model.

#### RESULTS

## Validated Deep Learning Models Accurately Identified Tumor Nuclei, Tumor-Infiltrating Lymphocyte Nuclei, and Tumor Cell Nuclear Grade

A total of 7,609 annotated tumor nuclei from 120 testing images and 4,000 annotated TILs from 75 testing images were collected to validate the MRCNN model for tumor nuclei and TIL

detection. The trained models correctly detected 6,101 (80.2%) tumor nuclei and 3,304 (82.6%) TILs. Multiple metrics were used for performance assessments, including precision, recall, F1-score, true positive number, false-positive number, and false-negative number. The metrics of precision, recall, and F1-score were defined as follows.

$$Precision = \frac{TP}{TP + FP}$$

$$Recall = \frac{TP}{TP + FN}$$

$$I \ score = 2 \ \times \ \frac{Precision \ \times \ Recall}{Precision + Recall}$$

F

where *TP*, *FP*, and *FN* represent the number of true positive, false-positive, and false-negative samples, respectively. The true positive samples were correctly detected samples. The false-positive samples were cells erroneously detected. Finally, the false-negative samples were missed ground truths from pathologists. The MRCNN models for the tumor nuclei and TIL detection achieved 0.7765 and 0.7171 for the F1-score, 0.7528 and 0.6337 for the precision, and 0.8018 and 0.826 for the recall, respectively.



FIGURE 2 | The overall schema of the developed deep learning (DL)-based whole slide image (WSI) processing pipeline is presented. Three DL models were established and trained for tumor cell detection, tumor-infiltrating lymphocyte (TIL) detection, and tumor cell segmentation, respectively. The tumor cell detection results were used to remove TIL false positive and retain nuclei contours for tumor cell segmentation.

The Hausdorff distance (HD) was used to measure the tumor nucleus contour concordance between the ground truths from pathologists and predictions using the DL process (Supplementary Figure 3). The metric of intersection over union (IOU) was used to match the ground truth to predicted contours. When IOU was greater than or equal to a cutoff value K, the ground truth and predicted nucleus contours were considered as a matched pair. When there was more than one prediction matching the same ground truth, the prediction with the largest IOU was retained for the match. When one prediction was matched to more than one ground truth, the prediction was assigned to the first matched ground truth. The cutoff value K was set as 0.1, 0.2, 0.3, 0.4, 0.5, 0.6, 0.7, 0.8, and 0.9, respectively. We computed the mean equivalent nuclei diameter for each nuclear grade. The mean equivalent diameters for nuclear grades 1, 2, and 3 were 26.30, 34.29, and 48.67 pixels, respectively. We present the mean HD between matched pairs and the ratio of mean HD to the mean equivalent diameter of tumor nuclei in Table 2. Representative cell detection and segmentation results from the DL models are shown in Figure 3.

## The Deep Learning-Based Analysis Enhances the Correlation Between Features in Magee Equation 2 and Recurrence Score

We detected an overwhelmingly large number of cells in each WSI (**Supplementary Table 1**). With detection results from image tiles, tumor cell and TIL density distributions were estimated and represented as density maps (**Figure 4**). The top ten image tiles of each WSI were selected based on the tumor cell density.

Eight variables from the training set included Nottingham grade, ER and PR H-score HER2 status, tumor size (cm), tumor

 $\ensuremath{\mathsf{TABLE 2}}\xspace$  | Performance of the Mask R-CNN (MRCNN) model for tumor nucleus segmentation.

IOU cutoff value	Mean HD for G1 (pixels) and ratio %	Mean HD at G2 (pixels) and ratio %	Mean HD at G3 (pixels) and ratio %
0.1	5.07 (19.28%)	6.29 (18.33%)	11.48 (23.59%)
0.2	5.05 (19.21%)	6.29 (18.33%)	11.30 (23.23%)
0.3	5.03 (19.14%)	6.29 (18.33%)	10.94 (22.48%)
0.4	4.97 (18.88%)	6.15 (17.94%)	10.50 (21.58%)
0.5	4.70 (17.89%)	5.84 (17.04%)	9.10 (18.69%)
0.6	4.32 (16.44%)	5.63 (16.41%)	7.77 (15.97%)
0.7	3.85 (14.65%)	4.94 (14.42%)	6.21 (12.77%)
0.8	3.13 (11.91%)	4.04 (11.77%)	5.14 (10.56%)
0.9	2.24 (8.53%)	2.34 (6.84%)	3.42 (7.03%)

The mean Hausdorff distances (HDs) of the matched ground truth and predicted contours of tumor nuclei at nuclear grades G1, G2, and G3 were computed with different intersection over union (IOU) cutoff values. For each grade, we computed the mean equivalent diameter. Additionally, we computed the ratio of mean HD to the mean equivalent diameter in percentage for each grade. The resulting mean HD and the ratio% are presented for each nuclear grade and each IOU cutoff value.

cell number in the densest tile, TIL number variance, and tumor nuclear grade (**Table 3**). The first five variables were from Magee equation 2, while the last three variables were DL-based image features derived from WSIs. We established a regression model with these selected features from the training set and applied the model to validation sets 1 and 2 for RS correlation.

We divided cases into low, intermediate, and high RS categories with the stratification rules from the TAILORx study (30). The concordances between the RS and our model were 56.10% and 68.0% for validation sets 1 and 2, respectively (**Table 4**). Additionally, the one-step discordance rates for validation sets 1 and 2 were 39.02% and 48.0%, respectively. The Pearson's correlation coefficients between the RS and our



model were 0.7058 (*p*-value =  $1.32 \times 10^{-13}$ ) and 0.5041 (*p*-value =  $1.15 \times 10^{-12}$ ) for validation sets 1 and 2, respectively. The tumor and TIL density maps from validation sets 1 and 2 are

illustrated in **Supplementary Figures 4**, **5**. The performance of the model correlation with RS was further evaluated by  $R^2$  and adjusted  $R^2$  (**Table 5**). When the image features were integrated with features in Mage equation 2, the adjusted  $R^2$  value increased from 0.3442 (*p*-value =  $5.17 \times 10^{-10}$ ) to 0.4431 (*p*-value =  $1.32 \times 10^{-13}$ ) in validation set 1 and from 0.2167 (*p*-value =  $6.52 \times 10^{-12}$ ) to 0.2182 (*p*-value =  $1.15 \times 10^{-12}$ ) in validation set 2. Similarly, the  $R^2$  increased from 0.3846 to 0.4981 in validation set 1 and from 0.2392 to 0.2541 in validation set 2. Additionally, we demonstrated the adjusted  $R^2$  and  $R^2$  of the linear regression model that was constructed only with the image features. The resulting adjusted  $R^2$  and  $R^2$  are 0.3048 (*p*-value =  $1.61 \times 10^{-8}$ )



FIGURE 4 | Demonstration of WSI density maps from (left) the low [Oncotype DX Recurrence Score (RS) = 3], (middle) intermediate (RS = 19), and (right) high (RS = 39) RS group. For each group, we present (top) a WSI, (middle) a TIL density map, and (bottom) a tumor cell density map, respectively.

and 0.3306 (*p*-value =  $1.61 \times 10^{-8}$ ) for validation set 1 and 0.0139 (*p*-value = 0.0199) and 0.0309 (*p*-value = 0.0199) for validation set 2, respectively. It is noted that the image features perform much worse than Magee features in validation set 2. Such performance degradation can be related to the fact that images in validation set 2 were originally scanned at  $20 \times$  and later computationally scaled to  $40 \times$  magnification. The inconsistency in the original image magnification can contribute to a significant error in the following analyses, leading to a worse prediction result.

To investigate the correlations between Magee and imagederived features, we computed their pair-wise absolute Pearson correlation coefficients. As shown in **Figure 5**, the largest correlation coefficient of 0.35 was found by the Nottingham score and tumor nuclear grade. Five Magee and image feature pairs present correlation coefficients close to 0.1. All remaining 9 pairs present correlation coefficients less than 0.1. Such weak correlations indicate the complementary prediction value by the image features for RS prediction enhancement.

For further correlation analyses between Magee and image features, we applied the least absolute shrinkage and selection operator (LASSO) regression method to our data and compared the resulting feature coefficients with those in the model trained by Ordinary Least Squares (OLS). The comparison results are presented in **Figure 6**. As LASSO includes an L1-norm regularizer, it penalizes the excessive feature inclusion and reduces uninformative feature coefficients to zero. From **Figure 6**, the non-zero feature coefficients from the two models trained by LASSO and OLS present similar values. Coefficients of only three features (i.e., tumor size, HER2, and tumor nuclear grade) were reduced to zero by LASSO. The only removed image feature by LASSO is tumor nuclear grade that presents an absolute Pearson correlation coefficient of 0.35 with the Nottingham score.

## Analyses of Cases With Discrepant Risk Scores Between Recurrence Score and Deep Learning-Based Prediction

We analyzed the cases with discordant risk categories by RS and our model (**Table 6**). There were totally 54 discordant cases in validation sets 1 and 2. Among these 54 cases, 40 were recommended to have chemotherapy by **TABLE 3** | Summary of independent variables from training set, validation set 1, and validation set 2 for the regression model.

	Training set	Validation set 1	Validation set 2	
Nottingham grade	(case number)			
1	33	31	64	
2	75	39	92	
3	17	12	19	
ER H-score				
Mean	277.14	262.18	246.03	
Range	80–300	5–300	19–300	
PR H-score				
Mean	188.07	156.07	174.47	
Range	0–300	0–300	0–300	
HER2 (case number	er)			
Negative	123	81	173	
Equivocal positive	2	1	2	
Tumor size (cm)				
Mean	2.19	1.81	1.64	
Range	0.4–7.8	0.5–5.3	0.3-7.1	
Tumor cell number	r in the densest til	e		
Mean	346.13	316.23	262.55	
Range	140-612	112-531	58–567	
TIL number varian	се			
Mean	714.95	792.67	331.42	
Range	2.49-8,227.6	4.68-10,850.01	1.17–5,849.39	
Tumor nuclear gra	de (case number)			
1	1	3	9	
2	81	50	102	
3	43	29	64	

RS but not by our DL-based model; of these 40 cases, 28 received chemotherapy.

In total, 14 cases were not recommended to have chemotherapy by RS, while our DL-based model did; of these 14 cases, 2 received chemotherapy. The chemotherapy recommendation based on RS and our DL model was determined by the suggested rules from the TAILORx study. Overall, none of these 54 discordant cases developed recurrence regardless of whether received chemotherapy, indicating that the role of chemotherapy in these discordant cases was not clear.

## DISCUSSION

Multiple studies have demonstrated the correlations between clinicopathological features and RS. Some used regression models to predict the RS directly from the clinicopathological features (20, 39-43), while others used classifiers to predict the RS risk categories (44-53). Additionally, a few studies have shown that the tumor imaging features from mammographic and sonographic imaging (54) and MRI (55, 56) are associated with RS. Magee equations include routinely evaluated clinicopathological features and have been shown to strongly correlate with RS (18-20, 57, 58). In this study, the regression models using the combination of the WSI-derived image features and Magee features as independent variables outperformed the models based on Magee features alone for RS correlation. The small correlation coefficients between the Magee and image features in Figure 5 and similar model coefficients in Figure 6 indicate the image features capture complementary prediction values for RS prediction. These results suggest that Magee features can enhance RS correlation when they are jointly used with the phenotypic information from WSIs.

In contrast with the substantial prediction improvement for validation set 1, a marginal improvement with validation set 2 is noticed. In **Table 5**, the adjusted  $R^2$  is 0.3048 and 0.0139 when the model trained with image features alone is applied to validation sets 1 and 2, respectively. This suggests a much stronger predictive value of image features from validation set 1 than validation set 2. One possible reason for limited success with validation set 2 is that images in validation set 2 were originally scanned at 20  $\times$  and computationally scaled to 40  $\times$  magnification. Such an inconsistent tissue scanning configuration may result in a significant downstream analysis difference accounting for a degraded prediction improvement. Additionally, we noticed from Table 3 that the average "TIL number variance" from validation set 2 is substantially less than that of the training set and validation set 1. To further investigate the individual feature impact on the prediction output, we computed the numerical product of each feature average value and its regression coefficient from the linear regression model. All such feature products are comparable across training set, validation set 1, and validation set 2, except for "TIL number variance." Specifically, the numerical product for "TIL number variance" from validation set 2 (i.e., 0.16) is less than half

#### TABLE 4 | Oncotype DX Recurrence Score (RS) group confusion matrix for validation sets 1 and 2.

	Validation set 1				Validation set	2		
	Predict high	Predict middle	Predict low	Total	Predict high	Predict middle	Predict low	Total
GT high	11	12	4	27	7	25	5	37
GT middle	0	8	7	15	1	43	26	70
GT low	0	13	27	40	0	32	36	68
Total	11	33	38	82	8	100	67	175

The "low," "middle," and "high" RS levels are determined by the RS cutoff values of 16 and 25. Several summary statistics for validation sets 1 and 2 are concordance: 46/82 (56.10%) and 119/175 (68.0%); one-step discordance: 32/82 (39.02%) and 84/175 (48.0%); two-step discordance: 4/82 (4.88%) and 5/175 (2.86%); Pearson's correlation coefficient: 0.7058 (p-value =  $1.32 \times 10^{-13}$ ) and 0.5041 (p-value =  $1.15 \times 10^{-12}$ ). GT represents ground truth.

		Validation set 1	Validation set 2
Adjusted R <sup>2</sup>	Magee2 features	0.3442 (p-value = 5.17 × 10 <sup>-10</sup> )	0.2167 (p-value = 6.52 × 10 <sup>-12</sup> )
	Image features	0.3048 (p-value = 1.61 × 10 <sup>-8</sup> )	0.0139 (p-value = 0.0199)
	Image + Magee2 features	<b>0.4431</b> ( <i>p</i> -value = $1.32 \times 10^{-13}$ )	<b>0.2182</b> (p-value = 1.15 × 10 <sup>-12</sup> )
$R^2$	Magee2 features	0.3846 (p-value = 5.17 × 10 <sup>-10</sup> )	0.2392 (p-value = $6.52 \times 10^{-12}$ )
	Image features	0.3306 (p-value = 1.61 × 10 <sup>-8</sup> )	0.0309 (p-value = 0.0199)
	Image + Magee2 features	<b>0.4981</b> ( <i>p</i> -value = 1.32 × 10 <sup>-13</sup> )	<b>0.2541</b> ( <i>p</i> -value = 1.15 × 10 <sup>-12</sup> )

TABLE 5 | Prediction performance of the regression model trained on the training set.

The bold values emphasize the greatest value of each metric in the two validation sets.



of that from the other two datasets (i.e., 0.35 and 0.39 from training and validation set 1, respectively), potentially degrading prediction improvement.

Our regression model used three histopathological image features extracted from WSIs: "tumor cell number in the densest tile," "TIL number variance," and "tumor nuclear grade." Tumor density is understudied in breast cancer prognosis. Tumor stroma has been shown to play an essential role in breast cancer prognosis and response to therapies (59–62). High tumorstromal content was shown to correlate with poor prognosis in triple-negative breast cancer (62), although such correlation was not demonstrated in ER+ breast cancer. Our study showed that high tumor density was associated with high RS. The role of stroma and tumor density in ER+ breast cancer may be essential and warrants more studies. TIL is an important prognostic and predictive marker in HER2+ and triple-negative breast cancer (9, 10, 63-65). Although the role of TIL is controversial in ER+ breast cancer (64, 66), high TIL has been found to correlate with high RS (66, 67). RS is strongly correlated with the proliferative module (68). One possible explanation for such correlation is the increased tumor proliferative rate within high TIL areas or the high proliferative rate of TIL itself. TIL has been shown to correlate with a high proliferative index in breast cancer (38). Thus, both the increased tumor proliferation and lymphocyte proliferation could contribute to the positive correlation with RS. While evaluations of TILs by pathologists may have intra- and inter-observation variations (69, 70), machine learning provides the opportunity to better quantify the TIL assessment (71). Tumor nuclear grade has been shown as an important prognostic factor in breast cancer and is a component of the Nottingham tumor grade (37). Genes associated with tumor grade are part of the Breast Cancer Index and are strongly correlated with tumor prognosis in ER+ breast cancer (72, 73).

In our study, 54 cases had discordant recommendations for chemotherapy treatment by RS and the DL-based model. Some patients with RS recommendation for chemotherapy and low risk by DL-based model did not actually receive chemotherapy while others not recommended for chemotherapy by RS and had low risk by DL-based model received chemotherapy. However, none of these patients developed cancer recurrence, including local and distant recurrence. The absolute benefit from chemotherapy to prevent distant recurrence in patients with intermediate RS is < 10% (30). Although it is also possible that these patients did not benefit from chemotherapy simply by chance, it is also possible that the benefit from chemotherapy in these patients with discordant results is not clear, and further studies are needed.

In this study, we trained three DL models to detect the tumor cells, TILs, and segment tumor nuclei. These model architectures were built on the MRCNN with the multitasking ability for detection, classification, and segmentation. We found that the performance of a comprehensive model was often inferior to that of individual single-task models. When a model was trained



operator (LASSO) and Ordinary Least Squares (OLS). Five Magee features M1-5 are ER H-score, PR H-score, Nottingham score, tumor size, and HER2, respectively. Three image features I1-3 are TIL number variance, tumor cell number in the densest tile, and tumor nuclear grade, respectively.

with one task at a time, the same DL model could achieve better accuracy due to more focused learning of one data distribution. In contrast, the multitask DL model's performance may deteriorate due to the high heterogeneity across multiple training sets. In our study, for instance, the circle labels for the detection model were significantly different from the mask labels for the segmentation model. The heterogeneity between the two types of data undermined the model's learning ability after merging them as one training dataset. Therefore, we trained three individual DL models. Due to the TIL training data heterogeneity, the TIL detection model might recognize some tumor nuclei as TILs by mistake. As the public MonuSeg-2018 dataset did not include cell type labels, we found that the tumor nuclei segmentation model predicted contours of non-tumor cells. To address these issues, we used tumor nucleus detection results to remove TILs and tumor nuclei false positive. Based on the density maps from the DL predictions, we observed that tissue regions of high TIL density were close to high tumor cell density regions, as shown in Figure 4. Such proximity of these two regions was frequently observed at the tumor invading fronts, consistent with previous studies (10, 64, 74-76).

As the patient cohorts for this study were not from a prospective clinical trial, we planned to validate our findings in completed prospective clinical trials in the following work. We also planned to increase our testing patient cohorts. Although we included 382 patients in the training and validation sets, a more extensive study is needed to validate our findings.

Overall, our results suggest that the combination of the image features derived from WSIs and Magee features presents a stronger correlation with RS than the Magee features alone. Although WSI image features present complementary information for RS correlation, we do not intend to replace

 
 TABLE 6 | Confusion matrix of the chemotherapy recommendations by RS and predicted RS for validation sets 1 and 2.

	Predicted RS No	Predicted RS Yes	Total
RS No	166	14	180
RS Yes	40	37	77
Total	206	51	257

Magee features with these WSI image features. Instead, we proposed to further boost Magee feature performance on RS correlation with these histology features from WSIs only available after computational analysis. To the best of our knowledge, our proposed approach is innovative in the sense that it uses the histological image features from WSIs to enhance the correlation between the Magee features and RS. The Magee equations can save healthcare costs and effectively serve patients with early breast cancer (77). The DL-based processing method presented in this study can be executed automatically at high throughput and further enhance the predictive power of Magee features.

## CONCLUSION

In this study, we have developed a DL-based digital pathology image processing pipeline to enhance the RS correlation with histology features derived from WSIs of ER+/HER2-/LN- breast cancer tissues. The proposed DL-based pipeline accurately detected tumor cells and TILs, segmented tumor cells, and extracted histology image features from gigapixel WSIs with high throughput. We demonstrated that the image features derived by DL-based analysis enhanced Magee feature correlation with RS.

#### DATA AVAILABILITY STATEMENT

The original contributions presented in this study are included in the article/**Supplementary Material**, further inquiries can be directed to the corresponding author/s.

## **ETHICS STATEMENT**

The studies involving human participants were reviewed and approved by Emory University Institutional Review Board. The patients/participants provided their written informed consent to participate in this study.

#### **AUTHOR CONTRIBUTIONS**

HL, JK, and XL conceived the original idea and designed the research. HL and JK performed the research. JW, ZL, MD, PZ, GS, and XL contributed data collection and image annotations.

## REFERENCES

- Arciero CA, Guo Y, Jiang R, Behera M, O'Regan R, Peng L, et al. ER<sup>+</sup>/HER2<sup>+</sup> breast cancer has different metastatic patterns and better survival than ER<sup>-</sup>/HER2<sup>+</sup> breast cancer. *Clin Breast Cancer.* (2019) 19:236–45. doi: 10. 1016/j.clbc.2019.02.001
- Bhattarai S, Klimov S, Mittal K, Krishnamurti U, Li XB, Oprea-Ilies G, et al. Prognostic role of androgen receptor in triple negative breast cancer: a multi-institutional study. *Cancers (Basel)*. (2019) 11:995. doi: 10.3390/ cancers11070995
- Guo Y, Arciero CA, Jiang R, Behera M, Peng L, Li X. Different breast cancer subtypes show different metastatic patterns: a study from a large public database. *Asian Pac J Cancer Prev.* (2020) 21:3587–93. doi: 10.31557/APJCP. 2020.21.12.3587
- Li X, Yang J, Krishnamurti U, Huo L, Ward KC, O'Regan R, et al. Hormone receptor-positive breast cancer has a worse prognosis in male than in female patients. *Clin Breast Cancer*. (2017) 17:356–66. doi: 10.1016/j.clbc.2017.03.005
- Li X, Zhang Y, Meisel J, Jiang R, Behera M, Peng L. Validation of the newly proposed American joint committee on cancer (AJCC) breast cancer prognostic staging group and proposing a new staging system using the national cancer database. *Breast Cancer Res Treat.* (2018) 171:303–13. doi: 10.1007/s10549-018-4832-9
- Reddy TP, Rosato RR, Li X, Moulder S, Piwnica-Worms H, Chang JC. A comprehensive overview of metaplastic breast cancer: clinical features and molecular aberrations. *Breast Cancer Res.* (2020) 22:121. doi: 10.1186/s13058-020-01353-z
- Zhao J, Krishnamurti U, Zhang C, Meisel J, Wei Z, Suo A, et al. HER2 immunohistochemistry staining positivity is strongly predictive of tumor response to neoadjuvant chemotherapy in HER2 positive breast cancer. *Pathol Res Pract.* (2020) 216:153155. doi: 10.1016/j.prp.2020.153155
- Li X, Oprea-Ilies GM, Krishnamurti U. New developments in breast cancer and their impact on daily practice in pathology. *Arch Pathol Lab Med.* (2017) 141:490–8. doi: 10.5858/arpa.2016-0288-SA
- Meisel JL, Zhao J, Suo A, Zhang C, Wei Z, Taylor C, et al. Clinicopathologic factors associated with response to neoadjuvant anti-HER2-directed chemotherapy in HER2-positive breast cancer. *Clin Breast Cancer*. (2020) 20:19–24. doi: 10.1016/j.clbc.2019.09.003
- Zhao J, Meisel J, Guo Y, Nahta R, Hsieh KL, Peng L, et al. Evaluation of PD-L1, tumor-infiltrating lymphocytes, and CD8<sup>+</sup> and FOXP3<sup>+</sup> immune cells in HER2-positive breast cancer treated with neoadjuvant therapies. *Breast Cancer Res Treat*. (2020) 183:599–606. doi: 10.1007/s10549-020-05819-8
- 11. Eifel P, Axelson JA, Costa J, Crowley J, Curran WJ Jr., Deshler A, et al. National institutes of health consensus development conference statement: adjuvant

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#### SUPPLEMENTARY MATERIAL

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therapy for breast cancer, november 1-3, 2000. J Natl Cancer Inst. (2001) 93:979-89.

- Goldhirsch A, Wood WC, Gelber RD, Coates AS, Thurlimann B, Senn HJ. Meeting highlights: updated international expert consensus on the primary therapy of early breast cancer. *J Clin Oncol.* (2003) 21:3357–65. doi: 10.1200/ JCO.2003.04.576
- Sparano JA, Paik S. Development of the 21-gene assay and its application in clinical practice and clinical trials. J Clin Oncol. (2008) 26:721–8. doi: 10.1200/ JCO.2007.15.1068
- Paik S, Tang G, Shak S, Kim C, Baker J, Kim W, et al. Gene expression and benefit of chemotherapy in women with node-negative, estrogen receptorpositive breast cancer. *J Clin Oncol.* (2006) 24:3726–34. doi: 10.1200/JCO.2005. 04.7985
- Harris LN, Ismaila N, McShane LM, Andre F, Collyar DE, Gonzalez-Angulo AM, et al. Use of biomarkers to guide decisions on adjuvant systemic therapy for women with early-stage invasive breast cancer: American society of clinical oncology clinical practice guideline. J Clin Oncol. (2016) 34:1134–50.
- Sparano JA, Gray RJ, Makower DF, Pritchard KI, Albain KS, Hayes DF, et al. Prospective validation of a 21-gene expression assay in breast cancer. N Engl J Med. (2015) 373:2005–14. doi: 10.1056/NEJMoa151 0764
- Kalinsky K, Barlow WE, Meric-Bernstam F, Gralow JR, Albain KS, Hayes D, et al. First results from a phase III randomized clinical trial of standard adjuvant endocrine therapy (ET) +/- chemotherapy (CT) in patients (pts) with 1–3 positive nodes, hormone receptor-positive (HR+) and HER2-negative (HER2-) breast cancer (BC) with recurrence score (RS) < 25: SWOG S1007 (RxPonder). *Cancer Res.* (2021) 81. doi: 10.1158/1538-7445.SABCS20-GS3-00
- Bhargava R, Dabbs DJ. Magee equations and oncotype DX(<sup>®</sup>)-a perspective. Breast Cancer Res Treat. (2017) 164:245–6. doi: 10.1007/s10549-017-4235-3
- Bhargava R, Esposito NN, O'Connor SM, Li Z, Turner BM, Moisini I, et al. Magee equations and response to neoadjuvant chemotherapy in ER<sup>+</sup>/HER2negative breast cancer: a multi-institutional study. *Mod Pathol.* (2021) 34:77– 84. doi: 10.1038/s41379-020-0620-2
- Flanagan MB, Dabbs DJ, Brufsky AM, Beriwal S, Bhargava R. Histopathologic variables predict oncotype DX recurrence score. *Mod Pathol.* (2008) 21:1255– 61. doi: 10.1038/modpathol.2008.54
- Roy M, Wang F, Vo H, Teng D, Teodoro G, Farris AB, et al. Deep-learningbased accurate hepatic steatosis quantification for histological assessment of liver biopsies. *Lab Invest.* (2020) 100:1367–83. doi: 10.1038/s41374-020-0463-y
- 22. Yokoyama S, Hamada T, Higashi M, Matsuo K, Maemura K, Kurahara H, et al. Predicted prognosis of patients with pancreatic cancer by machine learning. *Clin Cancer Res.* (2020) 26:2411–21.

- 23. Jaber MI, Song B, Taylor C, Vaske CJ, Benz SC, Rabizadeh S, et al. A deep learning image-based intrinsic molecular subtype classifier of breast tumors reveals tumor heterogeneity that may affect survival. *Breast Cancer Res.* (2020) 22:12. doi: 10.1186/s13058-020-1248-3
- Liu Y, Kohlberger T, Norouzi M, Dahl GE, Smith JL, Mohtashamian A, et al. Artificial intelligence-based breast cancer nodal metastasis detection: insights into the black box for pathologists. *Arch Pathol Lab Med.* (2019) 143:859–68. doi: 10.5858/arpa.2018-0147-OA
- Pantanowitz L, Quiroga-Garza GM, Bien L, Heled R, Laifenfeld D, Linhart C, et al. An artificial intelligence algorithm for prostate cancer diagnosis in whole slide images of core needle biopsies: a blinded clinical validation and deployment study. *Lancet Digit Health*. (2020) 2:e407–16. doi: 10.1016/S2589-7500(20)30159-X
- Jiang Y, Yang M, Wang S, Li X, Sun Y. Emerging role of deep learning-based artificial intelligence in tumor pathology. *Cancer Commun (Lond)*. (2020) 40:154–66. doi: 10.1002/cac2.12012
- Cheplygina V, de Bruijne M, Pluim JPW. Not-so-supervised: a survey of semisupervised, multi-instance, and transfer learning in medical image analysis. *Med Image Anal.* (2019) 54:280–96. doi: 10.1016/j.media.2019.03.009
- Shen D, Wu G, Suk HI. Deep learning in medical image analysis. Annu Rev Biomed Eng. (2017) 19:221–48.
- Litjens G, Kooi T, Bejnordi BE, Setio AAA, Ciompi F, Ghafoorian M, et al. A survey on deep learning in medical image analysis. *Med Image Anal.* (2017) 42:60–88.
- Sparano JA, Gray RJ, Ravdin PM, Makower DF, Pritchard KI, Albain KS, et al. Clinical and genomic risk to guide the use of adjuvant therapy for breast cancer. N Engl J Med. (2019) 380:2395–405. doi: 10.1056/NEJMoa1904819
- Allison KH, Hammond MEH, Dowsett M, McKernin SE, Carey LA, Fitzgibbons PL, et al. Estrogen and progesterone receptor testing in breast cancer: ASCO/CAP guideline update. J Clin Oncol. (2020) 38:1346–66. doi: 10.1200/JCO.19.02309
- 32. Wolff AC, Hammond MEH, Allison KH, Harvey BE, Mangu PB, Bartlett JMS, et al. Human epidermal growth factor receptor 2 testing in breast cancer: American society of clinical oncology/college of American pathologists clinical practice guideline focused update. *J Clin Oncol.* (2018) 36:2105–22.
- Vahadane A, Peng T, Sethi A, Albarqouni S, Wang L, Baust M, et al. Structurepreserving color normalization and sparse stain separation for histological images. *IEEE Trans Med Imaging*. (2016) 35:1962–71. doi: 10.1109/TMI.2016. 2529665
- He K, Gkioxari G, Dollár P, Girshick R. Mask R-CNN. In: Proceedings of the IEEE International Conference on Computer Vision. Venice (2017). p. 2961–9.
- Ren S, He K, Girshick R, Sun J. Faster R-CNN: towards real-time object detection with region proposal networks. *IEEE Trans Pattern Anal Mach Intell.* (2017) 39:1137–49. doi: 10.1109/TPAMI.2016.2577031
- Girshick R. Fast R-CNN. In: Proceedings of the IEEE International Conference on Computer Vision. Santiago (2015). p. 1440–8. doi: 10.1109/ICCV.2015.169
- Kerlikowske K, Molinaro A, Cha I, Ljung BM, Ernster VL, Stewart K, et al. Characteristics associated with recurrence among women with ductal carcinoma in situ treated by lumpectomy. *J Natl Cancer Inst.* (2003) 95:1692– 702. doi: 10.1093/jnci/djg097
- Bouzidi L, Triki H, Charfi S, Kridis WB, Derbel M, Ayadi L, et al. Prognostic value of natural killer cells besides tumor-infiltrating lymphocytes in breast cancer tissues. *Clin Breast Cancer*. (2021) 21:e738–47. doi: 10.1016/j.clbc.2021. 02.003
- Klein ME, Dabbs DJ, Shuai Y, Brufsky AM, Jankowitz R, Puhalla SL, et al. Prediction of the oncotype DX recurrence score: use of pathology-generated equations derived by linear regression analysis. *Mod Pathol.* (2013) 26:658–64. doi: 10.1038/modpathol.2013.36
- Geradts J, Bean SM, Bentley RC, Barry WT. The oncotype DX recurrence score is correlated with a composite index including routinely reported pathobiologic features. *Cancer Invest.* (2010) 28:969–77. doi: 10.3109/ 07357907.2010.512600
- Tang P, Wang J, Hicks DG, Wang X, Schiffhauer L, McMahon L, et al. A lower allred score for progesterone receptor is strongly associated with a higher recurrence score of 21-gene assay in breast cancer. *Cancer Invest.* (2010) 28:978–82. doi: 10.3109/07357907.2010.496754
- 42. Cuzick J, Dowsett M, Pineda S, Wale C, Salter J, Quinn E, et al. Prognostic value of a combined estrogen receptor, progesterone receptor, Ki-67, and

human epidermal growth factor receptor 2 immunohistochemical score and comparison with the genomic health recurrence score in early breast cancer. *J Clin Oncol.* (2011) 29:4273–8. doi: 10.1200/JCO.2010.31.2835

- Eaton AA, Pesce CE, Murphy JO, Stempel MM, Patil SM, Brogi E, et al. Estimating the OncotypeDX score: validation of an inexpensive estimation tool. *Breast Cancer Res Treat.* (2017) 161:435–41. doi: 10.1007/s10549-016-4069-4
- 44. Allison KH, Kandalaft PL, Sitlani CM, Dintzis SM, Gown AM. Routine pathologic parameters can predict oncotype DXTM recurrence scores in subsets of ER positive patients: who does not always need testing? *Breast Cancer Res Treat.* (2012) 131:413–24. doi: 10.1007/s10549-011-1416-3
- 45. Ingoldsby H, Webber M, Wall D, Scarrott C, Newell J, Callagy G. Prediction of oncotype DX and TAILORx risk categories using histopathological and immunohistochemical markers by classification and regression tree (CART) analysis. *Breast.* (2013) 22:879–86. doi: 10.1016/j.breast.2013.04.008
- Kim H-S, Umbricht CB, Illei PB, Cimino-Mathews A, Cho S, Chowdhury N, et al. Optimizing the use of gene expression profiling in early-stage breast cancer. J Clin Oncol. (2016) 34:4390–7. doi: 10.1200/JCO.2016.67.7195
- Orucevic A, Bell JL, McNabb AP, Heidel RE. Oncotype DX breast cancer recurrence score can be predicted with a novel nomogram using clinicopathologic data. *Breast Cancer Res Treat.* (2017) 163:51–61. doi: 10. 1007/s10549-017-4170-3
- Hanna MG, Bleiweiss IJ, Nayak A, Jaffer S. Correlation of oncotype DX recurrence score with Histomorphology and Immunohistochemistry in over 500 patients. *Int J Breast Cancer*. (2017) 2017:1257078. doi: 10.1155/2017/ 1257078
- 49. Lee SB, Kim J, Sohn G, Kim J, Chung IY, Kim HJ, et al. A nomogram for predicting the oncotype DX recurrence score in women with T1-3N0-1miM0 hormone receptor?positive, human epidermal growth factor 2 (HER2) negative breast cancer. *Cancer Res Treat.* (2019) 51:1073–85. doi: 10.4143/crt. 2018.357
- Wu S-G, Zhang W-W, Wang J, Lian C-L, Sun J-Y, Chen Y-X, et al. Progesterone receptor status and tumor grade predict the 21-gene recurrence score of invasive lobular breast cancer. *Biomark Med.* (2019) 13:1005–12. doi: 10.2217/bmm-2019-0209
- Thibodeau S, Voutsadakis IA. Prediction of oncotype DX recurrence score using clinical parameters: a comparison of available tools and a simple predictor based on grade and progesterone receptor. *Hematol Oncol Stem Cell Ther.* (2019) 12:89–96. doi: 10.1016/j.hemonc.2019.02.001
- Orucevic A, Bell JL, King M, McNabb AP, Heidel RE. Nomogram update based on TAILORx clinical trial results – oncotype DX breast cancer recurrence score can be predicted using clinicopathologic data. *Breast.* (2019) 46:116–25. doi: 10.1016/j.breast.2019.05.006
- Baltres A, Al Masry Z, Zemouri R, Valmary-Degano S, Arnould L, Zerhouni N, et al. Prediction of oncotype DX recurrence score using deep multi-layer perceptrons in estrogen receptor-positive, HER2-negative breast cancer. *Breast Cancer*. (2020) 27:1007–16. doi: 10.1007/s12282-020-01 100-4
- 54. Yepes MM, Romilly AP, Collado-Mesa F, Net JM, Kiszonas R, Arheart KL, et al. Can mammographic and sonographic imaging features predict the oncotype DX<sup>TM</sup> recurrence score in T1 and T2, hormone receptor positive, HER2 negative and axillary lymph node negative breast cancers? *Breast Cancer Res Treat.* (2014) 148:117–23. doi: 10.1007/s10549-014-3143-z
- Saha A, Harowicz MR, Wang W, Mazurowski MA. A study of association of oncotype DX recurrence score with DCE-MRI characteristics using multivariate machine learning models. J Cancer Res Clin Oncol. (2018) 144:799–807. doi: 10.1007/s00432-018-2595-7
- Ha R, Chang P, Mutasa S, Karcich J, Goodman S, Blum E, et al. Convolutional neural network using a breast MRI tumor dataset can predict oncotype Dx recurrence score. J Magn Reson Imagings. (2019) 49:518–24. doi: 10.1002/jmri. 26244
- Hou Y, Zynger DL, Li X, Li Z. Comparison of oncotype DX with modified magee equation recurrence scores in low-grade invasive carcinoma of breast. *Am J Clin Pathol.* (2017) 148:167–72. doi: 10.1093/ajcp/aqx059
- Hou Y, Moosavi HS, Wei L, Parwani AV, Li XB, Li Z. Magee equation recurrence score is associated with distal metastatic risk in male breast carcinomas: experience from two institutions. *Am J Clin Pathol.* (2018) 150:491–8. doi: 10.1093/ajcp/aqy078

- Plava J, Cihova M, Burikova M, Matuskova M, Kucerova L, Miklikova S. Recent advances in understanding tumor stroma-mediated chemoresistance in breast cancer. *Mol Cancer.* (2019) 18:67. doi: 10.1186/s12943-019-0960-z
- Bussard KM, Mutkus L, Stumpf K, Gomez-Manzano C, Marini FC. Tumorassociated stromal cells as key contributors to the tumor microenvironment. *Breast Cancer Res.* (2016) 18:84. doi: 10.1186/s13058-016-0740-2
- Hill BS, Sarnella A, D'Avino G, Zannetti A. Recruitment of stromal cells into tumour microenvironment promote the metastatic spread of breast cancer. *Semin Cancer Biol.* (2020) 60:202–13. doi: 10.1016/j.semcancer.2019.07.028
- 62. Kramer CJH, Vangangelt KMH, van Pelt GW, Dekker TJA, Tollenaar R, Mesker WE. The prognostic value of tumour-stroma ratio in primary breast cancer with special attention to triple-negative tumours: a review. *Breast Cancer Res Treat.* (2019) 173:55–64. doi: 10.1007/s10549-018-4987-4
- Li XB, Krishnamurti U, Bhattarai S, Klimov S, Reid MD, O'Regan R, et al. Biomarkers predicting pathologic complete response to neoadjuvant chemotherapy in breast cancer. *Am J Clin Pathol.* (2016) 145:871–8. doi: 10. 1093/ajcp/aqw045
- Krishnamurti U, Wetherilt CS, Yang J, Peng L, Li X. Tumor-infiltrating lymphocytes are significantly associated with better overall survival and disease-free survival in triple-negative but not estrogen receptor-positive breast cancers. *Hum Pathol.* (2017) 64:7–12. doi: 10.1016/j.humpath.2017.01. 004
- 65. Gonzalez-Ericsson PI, Stovgaard ES, Sua LF, Reisenbichler E, Kos Z, Carter JM, et al. The path to a better biomarker: application of a risk management framework for the implementation of PD-L1 and TILs as immuno-oncology biomarkers in breast cancer clinical trials and daily practice. *J Pathol.* (2020) 250:667–84. doi: 10.1002/path.5406
- 66. Kolberg-Liedtke C, Gluz O, Heinisch F, Feuerhake F, Kreipe H, Clemens M, et al. Association of TILs with clinical parameters, recurrence score<sup>®</sup> results, and prognosis in patients with early HER2-negative breast cancer (BC)-a translational analysis of the prospective WSG PlanB trial. *Breast Cancer Res.* (2020) 22:47. doi: 10.1186/s13058-020-01283-w
- Ahn SG, Cha YJ, Bae SJ, Yoon C, Lee HW, Jeong J. Comparisons of tumor-infiltrating lymphocyte levels and the 21-gene recurrence score in ERpositive/HER2-negative breast cancer. *BMC Cancer.* (2018) 18:320. doi: 10. 1186/s12885-018-4228-6
- Buus R, Sestak I, Kronenwett R, Ferree S, Schnabel CA, Baehner FL, et al. Molecular drivers of oncotype DX, prosigna, ENDOPREDICT, and the breast cancer index: a TransATAC study. J Clin Oncol. (2021) 39:126–35. doi: 10. 1200/JCO.20.00853
- 69. Swisher SK, Wu Y, Castaneda CA, Lyons GR, Yang F, Tapia C, et al. Interobserver agreement between pathologists assessing tumor-infiltrating lymphocytes (TILs) in breast cancer using methodology proposed by the international TILs working group. *Ann Surg Oncol.* (2016) 23:2242–8. doi: 10.1245/s10434-016-5173-8
- O'Loughlin M, Andreu X, Bianchi S, Chemielik E, Cordoba A, Cserni G, et al. Reproducibility and predictive value of scoring stromal tumour infiltrating lymphocytes in triple-negative breast cancer: a multi-institutional study. *Breast Cancer Res Treat.* (2018) 171:1–9. doi: 10.1007/s10549-018-4825-8

- Klauschen F, Müller KR, Binder A, Bockmayr M, Hägele M, Seegerer P, et al. Scoring of tumor-infiltrating lymphocytes: from visual estimation to machine learning. *Semin Cancer Biol.* (2018) 52:151–7. doi: 10.1016/j.semcancer.2018. 07.001
- Zhang Y, Schnabel CA, Schroeder BE, Jerevall PL, Jankowitz RC, Fornander T, et al. Breast cancer index identifies early-stage estrogen receptorpositive breast cancer patients at risk for early- and late-distant recurrence. *Clin Cancer Res.* (2013) 19:4196–205. doi: 10.1158/1078-0432.CCR-13-0804
- 73. Sgroi DC, Sestak I, Cuzick J, Zhang Y, Schnabel CA, Schroeder B, et al. Prediction of late distant recurrence in patients with oestrogen-receptorpositive breast cancer: a prospective comparison of the breast-cancer index (BCI) assay, 21-gene recurrence score, and IHC4 in the TransATAC study population. *Lancet Oncol.* (2013) 14:1067–76. doi: 10.1016/S1470-2045(13) 70387-5
- Buisseret L, Garaud S, de Wind A, Van den Eynden G, Boisson A, Solinas C, et al. Tumor-infiltrating lymphocyte composition, organization and PD-1/ PD-L1 expression are linked in breast cancer. *Oncoimmunology.* (2017) 6:e1257452. doi: 10.1080/2162402X.2016.1257452
- Le H, Gupta R, Hou L, Abousamra S, Fassler D, Torre-Healy L, et al. Utilizing automated breast cancer detection to identify spatial distributions of tumor-infiltrating lymphocytes in invasive breast cancer. *Am J Pathol.* (2020) 190:1491–504. doi: 10.1016/j.ajpath.2020.03.012
- 76. Li X, Wetherilt CS, Krishnamurti U, Yang J, Ma Y, Styblo TM, et al. Stromal PD-L1 expression is associated with better disease-free survival in triplenegative breast cancer. Am J Clin Pathol. (2016) 146:496–502. doi: 10.1093/ ajcp/aqw134
- 77. Bhargava R, Clark BZ, Carter GJ, Brufsky AM, Dabbs DJ. The healthcare value of the magee decision algorithm<sup>TM</sup>: use of magee equations<sup>TM</sup> and mitosis score to safely forgo molecular testing in breast cancer. *Mod Pathol.* (2020) 33:1563–70. doi: 10.1038/s41379-020-0521-4

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