#### Check for updates

#### OPEN ACCESS

EDITED BY Gian Marco Ghiggeri, Giannina Gaslini Institute (IRCCS), Italy

REVIEWED BY Moshe Levi, Georgetown University, United States Fujun Lin, Shanghai Jiao Tong University, China

\*CORRESPONDENCE Julia Hoefele iulia.hoefele@tum.de

SPECIALTY SECTION This article was submitted to Nephrology, a section of the journal Frontiers in Medicine

RECEIVED 31 May 2022 ACCEPTED 29 July 2022 PUBLISHED 31 August 2022

#### CITATION

Comić J, Riedhammer KM, Günthner R, Schaaf CW, Richthammer P, Simmendinger H, Kieffer D, Berutti R, Tasic V, Abazi-Emini N, Nushi-Stavileci V, Putnik J, Stajic N, Lungu A, Gross O, Renders L, Heemann U, Braunisch MC, Meitinger T and Hoefele J (2022) The multifaceted phenotypic and genotypic spectrum of type-IV-collagen-related nephropathy—A human genetics department experience. *Front. Med.* 9:957733. doi: 10.3389/fmed.2022.957733

#### COPYRIGHT

© 2022 Comić, Riedhammer, Günthner, Schaaf, Richthammer, Simmendinger, Kieffer, Berutti, Tasic, Abazi-Emini, Nushi-Stavileci, Putnik, Stajic, Lungu, Gross, Renders, Heemann, Braunisch, Meitinger and Hoefele. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

# The multifaceted phenotypic and genotypic spectrum of type-IV-collagen-related nephropathy—A human genetics department experience

Jasmina Comić<sup>1,2</sup>, Korbinian M. Riedhammer <sup>1,2</sup>, Roman Günthner<sup>2</sup>, Christian W. Schaaf<sup>1,2</sup>, Patrick Richthammer<sup>1</sup>, Hannes Simmendinger<sup>1</sup>, Donald Kieffer<sup>1</sup>, Riccardo Berutti<sup>1</sup>, Velibor Tasic<sup>3</sup>, Nora Abazi-Emini<sup>3</sup>, Valbona Nushi-Stavileci<sup>4</sup>, Jovana Putnik<sup>5</sup>, Nataša Stajic<sup>5</sup>, Adrian Lungu<sup>3</sup>, Oliver Gross<sup>6</sup>, Lutz Renders<sup>2</sup>, Uwe Heemann<sup>2</sup>, Matthias C. Braunisch<sup>2</sup>, Thomas Meitinger<sup>1</sup> and Julia Hoefele<sup>1\*</sup>

<sup>1</sup>Institute of Human Genetics, Klinikum rechts der Isar, Technical University of Munich, School of Medicine, Munich, Germany, <sup>2</sup>Department of Nephrology, Klinikum rechts der Isar, Technical University of Munich, School of Medicine, Munich, Germany, <sup>3</sup>University Children's Hospital, Medical Faculty of Skopje, Skopje, North Macedonia, <sup>4</sup>Pediatric Clinic, University Clinical Center of Kosovo, Prishtina, Serbia, <sup>5</sup>Institute for Mother and Child Health Care of Serbia "Dr. Vukan Cupic", Department of Nephrology, University of Belgrade, Faculty of Medicine, Belgrade, Serbia, <sup>6</sup>Clinic of Nephrology and Rheumatology, University Medical Center Göttingen, University of Göttingen, Göttingen, Germany

Disease-causing variants in COL4A3-5 are associated with type-IV-collagenrelated nephropathy, a genetically and phenotypically multifaceted disorder comprising Alport syndrome (AS) and thin basement membrane nephropathy (TBMN) and autosomal, X-linked and a proposed digenic inheritance. Initial symptoms of individuals with AS are microscopic hematuria followed by proteinuria leading to kidney failure (90% on dialysis < age 40 years). In contrast, individuals with TBMN, an outdated histology-derived term, present with microscopic hematuria, only some of them develop kidney failure (>50 years of age). An early diagnosis of type-IV-collagen-related nephropathy is essential for optimized therapy and slowing of the disease. Sixty index cases, in whom exome sequencing had been performed and with disease-causing variant(s) in COL4A3-5, were evaluated concerning their clinical tentative diagnosis and their genotype. Of 60 reevaluated individuals with type-IVcollagen-related nephropathy, 72% had AS, 23% TBMN and 5% focal segmental glomerulosclerosis (FSGS) as clinical tentative diagnosis. The FSGS cases had to be re-classified as having type-IV-collagen-related nephropathy. Twelve percent of cases had AS as clinical tentative diagnosis and a monoallelic disease-causing variant in COL4A3/4 but could not be classified as autosomal dominant AS because of limited or conflicting clinical data. This study illustrates

the complex clinical and genetic picture of individuals with a type IV-collagenrelated nephropathy indicating the need of a refined nomenclature and the more interdisciplinary teamwork of clinicians and geneticists as the key to optimized patient care.

KEYWORDS

type-IV-collagen-related nephropathy, Alport syndrome, COL4A3, COL4A4, COL4A5

## Introduction

The  $\alpha$ 3,  $\alpha$ 4, and  $\alpha$ 5 chains of the type IV collagen are an essential component of the glomerular basement membrane (GBM) and are encoded by the three genes *COL4A3*, *COL4A4*, and *COL4A5* (1). Disease-causing variants [(likely) pathogenic and pathogenic variants as per the guideline for sequence variant interpretation of the American College of Medical Genetics and Genomics and current amendments; see Material and Methods] in one of these genes are associated with type-IV-collagen-related nephropathy, comprising Alport syndrome (AS) and thin basement membrane nephropathy (TBMN) (2–8).

AS is characterized by microscopic hematuria and proteinuria leading to progressive loss of kidney function. Additionally, sensorineural hearing impairment, and eye abnormalities can be observed. AS is the second most common monogenic cause for kidney failure (1). It can be inherited in an X-linked [XLAS; hemizygous (male) or heterozygous (female) disease-causing variant in COL4A5] or autosomal recessive (ARAS; biallelic pathogenic variants in COL4A3/COL4A4) form (1, 9-11). The often used designation autosomal dominant AS in carriers of monoallelic pathogenic variants in COL4A3 and COL4A4 is differently used in the literature. In one recent publication by Furlano et al., the authors propose that any case harboring one heterozygous disease-causing variant in COL4A3 or COL4A4 should be designated as autosomal dominant AS independently from the clinical phenotype which ranges from microscopic hematuria to chronic kidney disease (12). In contrast, Savige et al. classify individuals with a heterozygous disease-causing variant in COL4A3 or COL4A4 as having autosomal dominant inherited TBMN or AS depending on the clinical phenotype and a potential positive familial history (13, 14). Furthermore, digenic inheritance has also been discussed as a possible cause in individuals with AS (15-18).

TBMN is a histopathology-derived term defined as uniform thinning of the GBM and phenotypically characterized by persistent microscopic hematuria, minimal if any proteinuria, and normal renal function (19, 20). Solely thinning of the GBM can also be found in early stages of AS (19, 21). The frequency of TBMN has been estimated to be as high as 1% of the world population (22). In up to 20% of the individuals with TBMN, disease progression to late-onset—compared to AS—kidney failure (>50 years of age) has been reported (23). This disease progress seems to be related in part to the development of focal segmental glomerulosclerosis (FSGS) (9, 24). Hence, in some cases with suspicion of a hereditary podocytopathy (hereditary FSGS), disease-causing variants in *COL4A3-COL4A5* can be found (25, 26).

The focus of this study was the reevaluation of the clinical phenotype and the reanalysis of exome sequencing data of 60 individuals with disease-causing variants in *COL4A3-5* in order to evaluate and highlight the shortcomings of the current nomenclature of AS/TBMN.

#### Methods

#### Study population

For this study, a cohort of 60 index cases of unrelated families with disease-causing variants in *COL4A3*, *COL4A4*, or *COL4A5* was investigated. These families have been recruited between October 2015 and August 2020 according to their appearance at our institute. In all individuals exome sequencing was already performed and genetic data were available. The exome data were reanalyzed in *COL4A3-5* within this study. This study was carried out according to standards of the 2013 Helsinki Declaration and authorized by the local Ethics Committee of the Technical University of Munich. Informed and written consents were obtained from all individuals or their legal guardians.

#### Clinical case information

Clinical and phenotypic information were obtained from clinical reports and medical history. Additionally, a standardized questionnaire was used to evaluate clinical information. The individuals were assigned to one of the following groups according to the clinical tentative diagnoses/kidney biopsy results as assigned by the referring clinician (nephrologists or pediatric nephrologists): AS, TBMN or FSGS. Age of onset of kidney failure in individuals was determined as the beginning of renal replacement therapy (hemodialysis or peritoneal dialysis) or pre-emptive kidney transplantation.

#### Genetic testing

For extraction of DNA from peripheral blood the automated nucleic acid purification instrument Chemagic<sup>TM</sup> 360 (PerkinElmer, Waltham, MA, USA) according to the manufacturer's protocol was used.

### Exome sequencing

Exome sequencing was performed with Sure Select Human All Exon 60Mb V6 Kit (Agilent) and on a HiSeq4000 platform (Illumina) in the index cases (27). Mitochondrial DNA was derived from off-target exome reads as previously described (28). Reads were aligned to the human reference genome (UCSC Genome Browser build hg19) using Burrows-Wheeler Aligner (v.0.7.5a). Using SAMtools (version 0.1.19), detection of singlenucleotide variants (SNVs) and small insertions and deletions (indels) was accomplished. For investigation of copy number variants (CNVs) (including exon-spanning intronic regions) ExomeDepth was used. A noise threshold of 2.5 was accepted (29). The called CNVs were visualized by the Integrative Genomics Viewer (IGV, https://software.broadinstitute.org/ software/igv/) to check if there was enough coverage of the examined regions and for plausibility of the CNVs. CNVs were then compared with publicly available control databases like the Genome Aggregation Database (gnomAD, https://gnomad. broadinstitute.org/about), the Database of Genomic Variants (DGV, http://dgv.tcag.ca/dgv/app/home) the databases for pathogenic CNVs like DECIPHER (https://decipher.sanger.ac. uk/) and ClinVar (https://www.ncbi.nlm.nih.gov/clinvar/). For the analysis of de novo, autosomal dominant and mitochondrial SNVs and indels, only variants with a minor allele frequency (MAF) of <0.1% (Munich Exome Server with over 22,000 exomes) were considered. For the analysis of autosomal recessive and X-linked SNVs and indels [homozygous, hemizygous or (putatively) compound heterozygous], only variants with a MAF of <1.0% were considered.

#### Sanger sequencing

Using Sanger sequencing, segregation analysis was conducted. Oligonucleotide primer sequences are available upon request.

#### Variant interpretation

Publicly available databases for (likely) pathogenic variants were used for comparison of all variants found and described in this study. These databases are ClinVar, the Human Gene Mutation Database (HGMD<sup>®</sup>, http://www.hgmd.cf.ac.

uk), and the Leiden Open Variation Database (LOVD, https:// www.lovd.nl). The variants were rated in accordance to American College of medical Genetics and Genomics (ACMG) guidelines and current amendments (4–7). Likely pathogenic and pathogenic variants are summarized as "disease-causing variant" in the text.

Biallelic disease-causing variants in COL4A3 and COL4A4 in male and female individuals, a hemizygous disease-causing variant in COL4A5 in a male individual and a heterozygous disease-causing variant in COL4A5 in a female individual were in accordance with the clinical tentative diagnosis of AS. In females with the clinical tentative diagnosis of TBMN and a heterozygous disease-causing variant in COL4A5, the genotype was also fitting to the clinical tentative diagnosis, as females with heterozygous disease-causing variants in COL4A5 can show a broad phenotypic spectrum ranging from TBMN to AS. In contrast to that, individuals with a heterozygous (likely) pathogenic variant in COL4A3 or COL4A4 and a clinical tentative diagnosis of AS were not automatically seen in accordance with autosomal dominant AS but further pedigree and phenotypic information were scrutinized (see Results). In turn, if cases had the clinical tentative diagnosis of TBMN and carried a heterozygous (likely) pathogenic variant in COL4A3 or COL4A4 (female and male individuals), genotype and phenotype were in agreement. Carriers of a hemizygous (likely) pathogenic variant in COL4A5 and the clinical tentative diagnosis of TBMN were classified as genetically solved AS due to the unquestionable genotype of hemizygous disease-causing variants in COL4A5 leading to AS (which can be mistaken as TBMN in early stages of disease). Furthermore, individuals with the non-specific phenotype of FSGS on kidney biopsy and disease-causing variants in COL4A3, COL4A4, or COL4A5 were reclassified as type-IV-collagennephropathies (two cases ARAS, one case XLAS-female carrier; see Supplementary Table 1).

#### Results

#### Study population

The total study cohort consisted of 60 unrelated index cases (30 females and 30 males) with disease-causing variants in *COL4A3-5*. 43/60 (72%) individuals had AS phenotypically, 14/60 (23%) TBMN and 3/60 (5%) FSGS on biopsy (Figure 1). 55/60 individuals (92%) were of non-Finnish European descent. The median age of disease-onset was 7 years (range: 0–35 years of age). In nine cases, the age of disease-onset was not available. Clinical findings were as follows (cases with reported phenotype information): 25/25 (100%) individuals presented with microscopic hematuria (no data available from 35 individuals), 12/25 (48%) with the clinical phenotype of AS, 11/25 (44%) with TBMN, and 2/25 (8%) with FSGS. Proteinuria

Gene	Chromosomal location	Transcript number	Inheritance	Phenotype MIM number	Covered exons (>20x)
COL4A3	2q36.3	NM_000091.4	AD, AR	104200, 203780	98%
COL4A4	2q36.3	NM_000092.4	AD, AR	104200, 203780	100%
COL4A5	Xq22.3	NM_033380.3	XL	301050	88%

#### TABLE 1 Exon coverage of genes COL4A3-5 in exome sequencing.

AD, autosomal dominant; AR, autosomal recessive; AS, Alport syndrome; XL, X-linked.



was seen in 15/25 (60%) individuals [no data available from 45 individuals; 10/15 (67%) with AS, 3/15 (20%) with TBMN, 2/15 (13%) with FSGS]. End-stage kidney failure (ESKF) was seen in 3/17 (18%) individuals, 2/3 (67%) with AS (23 and 24 years of age), and 1/3 (33%) with TBMN (68 years of age). No data were available from 43 individuals. Eye anomalies could be observed in 8/53 (15%) individuals (no data available from 7 individuals); all of them had AS as clinical tentative diagnosis. 18/53 (34%) individuals had hearing impairment (no data available from 7 individuals), 16/18 (89%) had AS, 2/18 (11%) TBMN.

#### Coverage of genes COL4A3-5

The median coverage > 20x of all exons of the three genes *COL4A3-5* was > 95% (range: 88-100%; see Table 1). All exons were covered at least 10x.

# Identification of different disease-causing variants

Within this study, 69 disease-causing variants could be identified, 61/69 (88%) of them were different. 26/61 (43%) were already described in the literature (*COL4A3*: 11, *COL4A4*: 1, *COL4A5*: 14), 35/61 (57%) were novel (i.e., not previously reported as disease-causing) (*COL4A3*: 9, *COL4A4*: 5, *COL4A5*: 21) (Supplementary Tables 1, 2).

# Distribution of identified (likely) pathogenic variants in *COL4A3-5*

From the 43 individuals with AS phenotypically, 39 (91%) cases had either an autosomal recessive (biallelic diseasecausing variants in *COL4A3* or *COL4A4*) or X-linked AS



[hemizygous (male individuals) or heterozygous/homozygous (female individuals) variants in *COL4A5*] (Figure 1; Supplementary Table 1). Of these 39 cases, 7 (18%) had (likely) pathogenic compound heterozygous/homozygous variants in *COL4A3*, 1 (3%) had a likely pathogenic homozygous variant in *COL4A4*, and 31 (79%) had a heterozygous (females) (14/31), homozygous (female; 1/31) or hemizygous (16/31) (likely) pathogenic variant in *COL4A5*.

Of the 14 individuals with TBMN phenotypically, one (7%) case had a (likely) pathogenic heterozygous variant in *COL4A3*, five (36%) in *COL4A4* and 8 (57%) in *COL4A5* (hemizygous variant: 2; heterozygous variant: 6).

Two individuals with the histopathological picture of FSGS had compound heterozygous likely pathogenic variants in *COL4A3*, and one female individual carried a heterozygous likely pathogenic variant in *COL4A5* (Supplementary Table 1).

In 4 (12%) individuals with AS phenotypically, only one heterozygous (likely) pathogenic variant in *COL4A3* gene could be identified (Supplementary Table 1). A genetic diagnosis of

autosomal dominant AS was not made in these individuals because of limited or conflicting clinical data questioning their submitted diagnosis of AS. The healthy mother (62 years of age), one healthy sister (33 years of age), and the healthy son of ATS-F521-II-2 (16 years of age) also carry the variant in COL4A3. One additional sister of ATS-F521-II-2 also suffers from hearing impairment but has no renal phenotype. She does not carry the variant in COL4A3 (Figure 2A). ATS-F663-II-1 inherited the variant from his healthy father but has three further affected siblings who all carry the variant. Two of the altogether four siblings additionally carry a heterozygous maternally inherited likely pathogenic variant in MYH9 (NM\_002473.4; c.1960C>G, p.(Leu654Val)). To our knowledge, the mother is healthy (Figure 2B). ATS-F787-II-1 was submitted as having AS confirmed by kidney biopsy (original biopsy report not available). No clinical data or medical records of the index and affected family members could be gathered (Figure 2C). Clinical data of ATS-F788-II-1 were also not available and a kidney biopsy confirming the clinical diagnosis of AS was not performed. This individual was part of the EARLY PRO-TECT

Alport trial and medicated with placebo. Under this treatment no disease progress was seen during the trial (30) (Figure 2D).

None of the 60 individuals with disease-causing variants in one of the *COL4A3-5* genes had additional variants in one of the two alternate *COL4A3-5* genes (Figure 1).

# Reclassification of individuals concerning their genetic result

Out of 56 individuals with a distinct genotype, the three individuals (5%) with FSGS were treated with cyclosporin A and mycophenolate mofetil. These individuals had to be re-classified as having a type-IV-collagen-related nephropathy questioning further extensive treatment with immunosuppressive drugs.

### Discussion

Type-IV-collagen-related nephropathy is a phenotypically and genetically multifaceted disorder. It comprises various phenotypes like classic AS, a slower-progressive phenotype originally described as TBMN and unspecific biopsy phenotypes like FSGS. To add to this complexity, it features both autosomal and X-linked inheritance.

We sought to illustrate this complex phenotypic and genotypic picture: In 91% of the individuals with AS phenotypically and in 100% of the individuals with TBMN phenotypically, disease-causing variants could be identified explaining the clinical tentative diagnosis. This high detection rate might be caused by intensive clinical evaluation of the affected individuals and detailed family medical history. In 5% of the individuals with disease-causing variants in COL4A3-5, a clinical phenotype of FSGS could be observed. This finding was already described by Malone et al. in 2014 and others (31). In their study, 10% of individuals with single or compound heterozygous disease-causing variants in COL4A3 or COL4A4 had the clinical tentative diagnosis of hereditary FSGS. The main cause of this clinical and genetic difference may be the fact that FSGS is an unspecific histologic phenotype seen in the process of different kidney diseases.

Digenic inheritance is also proposed as a possible cause for type-IV-collagen-related nephropathy including AS and TBMN, although little evidence is available on this topic so far (12, 16, 18). In contrast to studies describing this inheritance pattern, we did not observe findings of this pattern of inheritance within this study. This might be due to the facts that the present study had a small cohort size, the affected individuals were of different origin compared to individuals described in the literature and was performed with different sequencing techniques. Of note, it cannot be excluded that a variant was not detected if it was located in one of the limited covered exons which mostly affects the exons 5, 11, 14, 23, 38, 41, and 48 of *COL4A5*.

Importantly, in 12% of individuals with AS as clinical tentative diagnosis and monoallelic disease-causing variants in COL4A3/COL4A4, a clear statement on autosomal dominant AS could not be made taking into account the genetic result and clinical data. There is an unresolved conflict concerning autosomal dominant AS and the frontlines seem to run across two standpoints: A "clinician-centered" view stating that clear diagnoses are important for surveillance and early treatment (12, 32); and a "geneticist-centered" view that AS is a monogenic disease with complete penetrance and progressive kidney failure (90% on dialysis by age 40 years in X-linked AS; comparable for autosomal recessive AS) (11, 14). For us and others on the genetics-side (13), there are many questions concerning the simple usage of autosomal dominant AS in any case with a monoallelic disease-causing variant in COL4A3 and COL4A4: In cases with a clear AS phenotype (for example on kidney biopsy, ATS-F787-II-1 above) but only a monoallelic variant in COL4A3/COL4A4, could there be another variant on the other allele missed by routine genetic testing (e.g., intronic variant leading to a splicing defect, complex rearrangement missed by short-read-based NGS)? Should we use kidney biopsy specimen (or urine-derived renal cells) in these cases to run transcriptomics on to determine if there is a splicing defect on the other allele? Should we engage further in elucidation of rearrangements with chromosomal microarray/multiplex ligation-dependent probe amplification (MLPA) or even genome sequencing? Does the same variant cause the identical disorder, AS, in a homozygous and heterozygous state? In heterozygous carriers with monoallelic missense variants in COL4A3/COL4A4, might a dominantnegative effect come into play and explain a complete AS phenotype with high penetrance? Have phenocopies been taken into account? Has the pedigree been thoroughly investigated and relatives been tested? What if healthy parents and relatives carry the variant (as in cases ATS-F521-II-2, ATS-F663-II-1), how do we counsel these parents if they get another child? Do they have a 50% recurrence risk of AS, as in other clearly autosomal dominant diseases? And does the affected individual have a 50% risk of offspring with AS if he or she gets children? To what extent is there incomplete penetrance? The estimated prevalence of heterozygous diseasecausing variants in COL4A3/COL4A4 is 1 in 106, as has recently been shown (8). Are we to diagnose all of these people with autosomal dominant AS? All these questions are not satisfactorily addressed so far in the literature and need to be solved to optimize the medical care and genetic counseling of these individuals.

Undoubtedly, heterozygous carriers of disease-causing variants in *COL4A3*, *COL4A4*, and *COL4A5* have a higher risk of end-stage kidney failure than the general population (12, 24). They need surveillance and treatment with ACE inhibition once proteinuria/albuminuria develops (33, 34). But if we classify every of these cases as having autosomal

dominant AS, we are risking not making the considerations mentioned above impeding correct diagnoses and risk calculations. Hence, a unifying gene-centered nomenclature like type-IV-collagen-related nephropathy could steer free of this conflict (35), especially as TBMN is also an outdated term which is based on histologic findings not always present or preceding pathognomonic AS changes (12) (as seen by the fact that there are two cases with a hemizygous disease-causing variant in COL4A5, i.e., XLAS, but rated as TBMN by the referring clinician). We still used the term "TBMN" in lack of proper alternatives and as it was used by referring clinicians. In our opinion, genetic reports should state the genotype in the diagnosis and the designation AS should only be added if there is genotypic and phenotypic evidence for this diagnosis (e.g., "Diagnosis of type-IV-collagen-related nephropathy-X-linked Alport syndrome-hemizygous pathogenic frameshift variant in COL4A5").

There is lacking phenotype information (see Section Results) on a number of individuals in this study, which can be viewed as a limitation. However, the aim of this study was to illustrate the phenotypic and genotypic spectrum of type-IV-collagen related nephropathy and not a detailed genotype-phenotype correlation on AS, which have been published extensively (9–12, 36).

Finally, in this study, 57% of the identified variants were novel indicating that there are still many disease-causing variants in type-IV-collagen-releated-nephropathy unknown so far. Therefore, it is extremely important to submit identified variants to open genetic databases like ClinVar or LOVD to extend the knowledge of disease-causing variants and to optimize the clinical care of individuals with a type-IV-collagenrelated nephropathy.

To conclude, this study illustrates the complex clinical and genetic spectrum of type-IV-collagen-related nephropathy including AS and TBMN in a small single tertiary-care center cohort. A refined nomenclature not impeding swift diagnosis, surveillance and treatment but owing to the diverse genetic considerations of this multifaceted disorder is direly needed and, by using the term "type-IV-collagen-related nephropathy", we propose a more gene-centered approach. Additionally, close cooperation of clinicians and geneticists is key to collect the necessary phenotypic and pedigree data needed to adequately assess individuals with suspected type-IV-collagen-related nephropathy.

### Data availability statement

The datasets presented in this study can be found in online repositories. The names of the repository/repositories and accession number(s) can be found in the article/Supplementary material.

## **Ethics statement**

The studies involving human participants were reviewed and approved by Ethics Committee, Klinikum rechts der Isar, Technical University of Munich, Munich, Germany. Written informed consent to participate in this study was provided by the participants' legal guardian/next of kin. Written informed consent was obtained from the individual(s), and minor(s)' legal guardian/next of kin, for the publication of any potentially identifiable images or data included in this article.

## Author contributions

Research and study design: KMR and JH. Data analysis/interpretation: JĆ, KMR, TM, and JH. Statistical analysis: JĆ. Patient acquisition: RG, PR, HS, DK, VT, NA-E, VN-S, JP, NS, AL, LR, UH, MB, CS, TM, and JH. Drafting and revising the article: JĆ, KMR, CS, TM, and JH. Supervision or mentorship and final approval of the version to be published: JH. All author contributed important intellectual content during manuscript drafting or revision, agrees to be personally accountable for the individual's own contributions, to ensure that questions pertaining to the accuracy or integrity of any portion of the work, even one in which the author was not directly involved, are appropriately investigated and resolved, and including with documentation in the literature if appropriate.

# Funding

This work was supported with a research Grant by the European Society for Pediatric Nephrology (ESPN e.V.; ESPN #2.2020) and by the German Research Foundation (DFG) and the Technical University of Munich (TUM) in the framework of the Open Access Publishing Program.

### Acknowledgments

We would like to thank the patients and their families for participation in this study.

# **Conflict of interest**

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

## Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those

of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

### Supplementary material

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/ fmed.2022.957733/full#supplementary-material

### References

1. Hudson BG, Tryggvason K, Sundaramoorthy M, Neilson EG. Alport's syndrome, Goodpasture's syndrome, and type IV collagen. *N Engl J Med.* (2003) 348:2543–56. doi: 10.1056/NEJMra022296

2. Funk SD, Lin MH, Miner JH. Alport syndrome and Pierson syndrome: diseases of the glomerular basement membrane. *Matrix Biol.* (2018) 71–72:250–61. doi: 10.1016/j.matbio.2018.04.008

3. Weber S, Strasser K, Rath S, Kittke A, Beicht S, Alberer M, et al. Identification of 47 novel mutations in patients with Alport syndrome and thin basement membrane nephropathy. *Pediatr Nephrol.* (2016) 31:941–55. doi: 10.1007/s00467-015-3302-4

4. Abou Tayoun AN, Pesaran T, DiStefano MT, Oza A, Rehm HL, Biesecker LG, et al. Recommendations for interpreting the loss of function PVS1 ACMG/AMP variant criterion. *Hum Mutat.* (2018) 39:1517–24. doi: 10.1002/humu. 23626

5. Kearney HM, Thorland EC, Brown KK, Quintero-Rivera F, South ST, Working Group of the American College of Medical Genetics Laboratory Quality Assurance C. American College of Medical Genetics standards and guidelines for interpretation and reporting of postnatal constitutional copy number variants. *Genet Med.* (2011) 13:680–5. doi: 10.1097/GIM.0b013e318 2217a3a

6. Richards S, Aziz N, Bale S, Bick D, Das S, Gastier-Foster J, et al. Standards and guidelines for the interpretation of sequence variants: a joint consensus recommendation of the American College of Medical Genetics and Genomics and the Association for Molecular Pathology. *Genet Med.* (2015) 17:405–24. doi: 10.1038/gim.2015.30

7. Ellard S, Baple EL, Berry I, Forrester N, Turnbull C, Owens M, et al. ACGS Best Practice Guidelines for Variant Classification (2019). Available online at: https://www.acgs.uk.com/media/11285/uk-practice-guidelines-for-variant-classification-2019-v1-0-3.pdf (accessed August 10, 2022).

8. Gibson J, Fieldhouse R, Chan MMY, Sadeghi-Alavijeh O, Burnett L, Izzi V, et al. Prevalence estimates of predicted pathogenic COL4A3-COL4A5 variants in a population sequencing database and their implications for Alport syndrome. *J Am Soc Nephrol.* (2021) 32:2273–90. doi: 10.1681/ASN.2020071065

9. Jais JP, Knebelmann B, Giatras I, De Marchi M, Rizzoni G, Renieri A, et al. X-linked Alport syndrome: natural history and genotype-phenotype correlations in girls and women belonging to 195 families: a "European Community Alport Syndrome Concerted Action" study. J Am Soc Nephrol. (2003) 14:2603–10. doi: 10.1097/01.ASN.000090034.71205.74

10. Bekheirnia MR, Reed B, Gregory MC, McFann K, Shamshirsaz AA, Masoumi A, et al. Genotype-phenotype correlation in X-linked Alport syndrome. *J Am Soc Nephrol.* (2010) 21:876–83. doi: 10.1681/ASN.2009070784

11. Jais JP, Knebelmann B, Giatras I, De Marchi M, Rizzoni G, Renieri A, et al. X-linked Alport syndrome: natural history in 195 families and genotype- phenotype correlations in males. *J Am Soc Nephrol.* (2000) 11:649–57. doi: 10.1681/ASN.V114649

12. Furlano M, Martinez V, Pybus M, Arce Y, Crespi J, Venegas MDP, et al. Clinical and genetic features of autosomal dominant alport syndrome: a cohort study. *Am J Kidney Dis.* (2021) 78:560–70.e1. doi: 10.1053/j.ajkd.2021.02.326

13. Savige J. Should we diagnose autosomal dominant alport syndrome when there is a pathogenic heterozygous COL4A3 or COL4A4 variant? *Kidney Int Rep.* (2018) 3:1239–41. doi: 10.1016/j.ekir.2018.08.002

14. Savige J, Huang M, Croos Dabrera MS, Shukla K, Gibson J. Genotypephenotype correlations for pathogenic COL4A3-COL4A5 variants in X-linked, autosomal recessive, and autosomal dominant Alport syndrome. *Front Med.* (2022) 9:865034. doi: 10.3389/fmed.2022.865034

15. Rheault MN. Women and Alport syndrome. *Pediatr Nephrol.* (2012) 27:41–6. doi: 10.1007/s00467-011-1836-7

16. Mencarelli MA, Heidet L, Storey H, van Geel M, Knebelmann B, Fallerini C, et al. Evidence of digenic inheritance in Alport syndrome. *J Med Genet.* (2015) 52:163–74. doi: 10.1136/jmedgenet-2014-102822

17. Yokota K, Nozu K, Minamikawa S, Yamamura T, Nakanishi K, Kaneda H, et al. Female X-linked Alport syndrome with somatic mosaicism. *Clin Exp Nephrol.* (2017) 21:877–83. doi: 10.1007/s10157-016-1352-y

18. Fallerini C, Baldassarri M, Trevisson E, Morbidoni V, La Manna A, Lazzarin R, et al. Alport syndrome: impact of digenic inheritance in patients management. *Clin Genet.* (2017) 92:34–44. doi: 10.1111/cge.12919

19. Tryggvason K, Patrakka J. Thin basement membrane nephropathy. J Am Soc Nephrol. (2006) 17:813–22. doi: 10.1681/ASN.2005070737

20. Savige J, Gregory M, Gross O, Kashtan C, Ding J, Flinter F. Expert guidelines for the management of Alport syndrome and thin basement membrane nephropathy. *J Am Soc Nephrol.* (2013) 24:364–75. doi: 10.1681/ASN.2012 020148

21. Cangiotti AM, Sessa A, Meroni M, Montironi R, Ragaiolo M, Mambelli V, et al. Evolution of glomerular basement membrane lesions in a male patient with Alport syndrome: ultrastructural and morphometric study. *Nephrol Dial Transplant.* (1996) 11:1829–34. doi: 10.1093/oxfordjournals.nd t.a027677

22. Savige J, Rana K, Tonna S, Buzza M, Dagher H, Wang YY. Thin basement membrane nephropathy. *Kidney Int.* (2003) 64:1169–78. doi: 10.1046/j.1523-1755.2003.00234.x

23. Matthaiou A, Poulli T, Deltas C. Prevalence of clinical, pathological and molecular features of glomerular basement membrane nephropathy caused by COL4A3 or COL4A4 mutations: a systematic review. *Clin Kidney J.* (2020) 13:1025–36. doi: 10.1093/ckj/sfz176

24. Voskarides K, Damianou L, Neocleous V, Zouvani I, Christodoulidou S, Hadjiconstantinou V, et al. COL4A3/COL4A4 mutations producing focal segmental glomerulosclerosis and renal failure in thin basement membrane nephropathy. *J Am Soc Nephrol.* (2007) 18:3004–16. doi: 10.1681/ASN.200 7040444

25. Braunisch MC, Buttner-Herold M, Gunthner R, Satanovskij R, Riedhammer KM, Herr PM, et al. Heterozygous COL4A3 variants in histologically diagnosed focal segmental glomerulosclerosis. *Front Pediatr.* (2018) 6:171. doi: 10.3389/fped.2018.00171

26. Frese J, Kettwig M, Zappel H, Hofer J, Grone HJ, Nagel M, et al. Kidney injury by variants in the COL4A5 gene aggravated by polymorphisms in slit diaphragm genes causes focal segmental glomerulosclerosis. *Int J Mol Sci.* (2019) 20:519. doi: 10.3390/ijms20030519

27. Kremer LS, Bader DM, Mertes C, Kopajtich R, Pichler G, Iuso A, et al. Genetic diagnosis of Mendelian disorders *via* RNA sequencing. *Nat Commun.* (2017) 8:15824. doi: 10.1038/ncomms15824

28. Griffin HR, Pyle A, Blakely EL, Alston CL, Duff J, Hudson G, et al. Accurate mitochondrial DNA sequencing using off-target reads provides a single test to identify pathogenic point mutations. *Genet Med.* (2014) 16:962–71. doi: 10.1038/gim.2014.66

29. Plagnol V, Curtis J, Epstein M, Mok KY, Stebbings E, Grigoriadou S, et al. A robust model for read count data in exome sequencing experiments and implications for copy number variant calling. *Bioinformatics*. (2012) 28:2747–54. doi: 10.1093/bioinformatics/bts526

30. Boeckhaus J, Hoefele J, Riedhammer KM, Tonshoff B, Ehren R, Pape L, et al. Precise variant interpretation, phenotype ascertainment, and genotype-phenotype correlation of children in the EARLY PRO-TECT Alport trial. *Clin Genet.* (2021) 99:143–56. doi: 10.1111/cge.13861

31. Malone AF, Phelan PJ, Hall G, Cetincelik U, Homstad A, Alonso AS, et al. Rare hereditary COL4A3/COL4A4 variants may be mistaken for

familial focal segmental glomerulosclerosis. *Kidney Int.* (2014) 86:1253–9. doi: 10.1038/ki.2014.305

32. Kashtan CE. Alport syndrome: achieving early diagnosis and treatment. Am J Kidney Dis. (2021) 77:272–9. doi: 10.1053/j.ajkd.2020.03.026

33. Gross O, Tonshoff B, Weber LT, Pape L, Latta K, Fehrenbach H, et al. A multicenter, randomized, placebo-controlled, double-blind phase 3 trial with open-arm comparison indicates safety and efficacy of nephroprotective therapy with ramipril in children with Alport's syndrome. *Kidney Int.* (2020) 97:1275–86. doi: 10.1016/j.kint.2019.12.015

34. Boeckhaus J, Hoefele J, Riedhammer KM, Nagel M, Beck B, Choi M, et al. Lifelong effect of therapy in young patients with the COL4A5 Alport missense

variant p.(Gly624Asp): a prospective cohort study. *Nephrol Dial Transplant*. (2022) gfac006. doi: 10.1093/ndt/gfac006

35. Imafuku A, Nozu K, Sawa N, Nakanishi K, Ubara Y. How to resolve confusion in the clinical setting for the diagnosis of heterozygous COL4A3 or COL4A4 gene variants? Discussion and suggestions from nephrologists. *Clin Exp Nephrol.* (2020) 24:651–6. doi: 10.1007/s10157-020-01880-1

36. Gross O, Netzer KO, Lambrecht R, Seibold S, Weber M. Metaanalysis of genotype-phenotype correlation in X-linked Alport syndrome: impact on clinical counselling. *Nephrol Dial Transplant.* (2002) 17:1218–27. doi: 10.1093/ndt/17.7.1218