Check for updates

OPEN ACCESS

EDITED BY Rana A. Youness, University of Hertfordshire, United Kingdom

REVIEWED BY Noha Abdelsalam, Zagazig University, Egypt Noha Fathallah, Future University in Egypt, Egypt

*CORRESPONDENCE Maha Saber-Ayad msaber@sharjah.ac.ae

SPECIALTY SECTION This article was submitted to Pathology, a section of the journal Frontiers in Medicine

RECEIVED 01 June 2022 ACCEPTED 11 August 2022 PUBLISHED 07 September 2022

CITATION

Mahgoub E, Taneera J, Sulaiman N and Saber-Ayad M (2022) The role of autophagy in colorectal cancer: Impact on pathogenesis and implications in therapy. *Front. Med.* 9:959348. doi: 10.3389/fmed.2022.959348

COPYRIGHT

© 2022 Mahgoub, Taneera, Sulaiman and Saber-Ayad. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

The role of autophagy in colorectal cancer: Impact on pathogenesis and implications in therapy

Eglal Mahgoub^{1,2}, Jalal Taneera^{1,2}, Nabil Sulaiman^{1,2,3} and Maha Saber-Ayad^{1,2,4}*

¹College of Medicine, University of Sharjah, Sharjah, United Arab Emirates, ²Sharjah Institute for Medical Research, University of Sharjah, Sharjah, United Arab Emirates, ³Baker Heart and Diabetes Institute, Melbourne, VIC, Australia, ⁴Faculty of Medicine, Cairo University, Giza, Egypt

Colorectal cancer (CRC) is considered as a global major cause of cancer death. Surgical resection is the main line of treatment; however, chemo-, radiotherapy and other adjuvant agents are crucial to achieve good outcomes. The tumor microenvironment (TME) is a well-recognized key player in CRC progression, yet the processes linking the cancer cells to its TME are not fully delineated. Autophagy is one of such processes, with a controversial role in the pathogenesis of CRC, with its intricate links to many pathological factors and processes. Autophagy may apparently play conflicting roles in carcinogenesis, but the precise mechanisms determining the overall direction of the process seem to depend on the context. Additionally, it has been established that autophagy has a remarkable effect on the endothelial cells in the TME, the key substrate for angiogenesis that supports tumor metastasis. Favorable response to immunotherapy occurs only in a specific subpopulation of CRC patients, namely the microsatellite instability-high (MSI-H). In view of such limitations of immunotherapy in CRC, modulation of autophagy represents a potential adjuvant strategy to enhance the effect of those relatively safe agents on wider CRC molecular subtypes. In this review, we discussed the molecular control of autophagy in CRC and how autophagy affects different processes and mechanisms that shape the TME. We explored how autophagy contributes to CRC initiation and progression, and how it interacts with tumor immunity, hypoxia, and oxidative stress. The crosstalk between autophagy and the TME in CRC was extensively dissected. Finally, we reported the clinical efforts and challenges in combining autophagy modulators with various cancer-targeted agents to improve CRC patients' survival and restrain cancer growth.

KEYWORDS

colorectal cancer, autophagy, tumor microenvironment, endothelial cells, hypoxia, oxidative stress, targeted therapy, MSI-H

Introduction

Colorectal Cancer (CRC) is counted as one of the most predominant cancers in both genders with high death rates. CRC is third in terms of prevalence which accounted for 6.1% of new cases and second in terms of the cause of death which accounted for 9.2% of deaths by cancer worldwide (1). There is a high incidence of colorectal cancer at young age (15-39 years) which was estimated by 70.2-82.9 thousand cases in 2019 with a mortality rate of 26.2-30.5 thousand in the same year (2). By the year 2035, it is estimated that the total number of deaths will increase by 71.5 and 60% from colon and rectal cancers, respectively (3). CRC is a heterogeneous disease with numerous variations in its molecular profiles, clinical manifestations and prognosis. CRC prognosis depends on the tumor staging at the time of diagnosis. Currently, the best therapeutic option for stage I and most of the stage II CRC patients is the aggressive surgical resection of the primary tumors which showed high success rates, with/without adjuvant radio-chemotherapy for high risks patients in stage II and stage III of CRC. Notably, stage III CRC patients usually suffer from recurrent disease, which may be associated with micro-metastasis. Stage IV CRC represents a metastatic state with a high risk of relapse and with less/no benefit from surgery. Instead, chemotherapy combinations are usually used at this stage, such as oxaliplatin/irinotecan and folinic acid, 5-fluorouracil (5-FU)-based regimens (4, 5). However, adjuvant treatment is highly accompanied by drug resistance, and ultimately disease progression in metastatic CRC. Recent advances in cancer-targeted therapy as second-line treatment of CRC in combination with chemotherapy, to disrupt signaling pathways or cellular mechanisms, have led to enhanced overall survival (OS) and progression-free survival (PFS). Currently, anti-angiogenic drugs including bevacizumab, regorafenib and aflibercept, are approved as a treatment of metastatic stage of CRC, whereas immunotherapy for CRC is still limited to the MSI-H tumors (6).

Classification system of CRC, based on molecular structure, was established to categorize both the tumor and the surrounding tumor microenvironment (TME) through variations in CRC gene expression (7). TME is a dynamic ecosystem that plays a crucial role in the support and progression of tumors. The composition of TME may significantly affect the tumor response to immunotherapy. TME includes different types of cells, e.g., tumor-infiltrating lymphocytes (TILs), tumor-associated macrophages (TAMs), tumor-associated neutrophils (TANs), cancer-associated fibroblasts, natural killer (NK) cells, regulatory T cells and dendritic cells (DCs). There are four main consensus molecular subtypes: CMS1, CMS2, CMS3, and CMS4 (8). Both CMS1 and CMS4 subtypes are characterized by increased immune cells infiltration, while CMS1 tumors, in particular, is identified by enhanced Th1-cell response as well as inflamed and stimulated TME. Whereas CMS4 TME is characterized by being inflamed and highly angiogenic, hence a good target for combination therapy. CMS2 tumors are caused by β -catenin pathway activation, with subsequent dendritic and T-cell exhaustion. Therefore, this subtype of tumors does not elicit anti-tumor immune response. CMS3 tumors are characterized by several metabolic pathways dysregulation such as nitrogen, glucose pentose, fatty acids, etc. (7).

Tumor microenvironment of colorectal cancer

Tumors are cellular networks characterized as being different and complex with de-differentiated malignant cell types, tumor stem cells, fibroblasts and endothelial and immune cells. TME is a dynamic ecosystem that plays a crucial role in supporting the progression of tumors. Cytotoxic CD8+ T-lymphocytes (CTL) are considered the major defense mechanism against tumor cells, hence T-cell abundance is a decisive and crucial prognostic factor for immunotherapy and chemotherapy response, particularly at the early tumor initiation stage, where an increased activity of T cells has been reported (9). The PD-L1/PD1 axis is identified as an inhibitor of CTL activity in several CRC phenotypes including Mismatch repair deficiency (MMRd)/Microsatellite instability-high (MSI-H) phenotype in which anti-PD1 monoclonal antibodies are highly beneficial in fighting the tumor (10, 11). Another essential type of T-cells highly associated with colorectal tumors is the Regulatory T-cells (Tregs) (12).

Other cell types in the TME include TAMs involved in regulating metastatic phenotype of cancer and modulating growth and invasion of cancer cells (13, 14). Two subpopulations of TAMs have been identified, the pro-tumorigenic (M2) and the anti-tumorigenic (M1) phenotypes, which are characterized by high plasticity (15). TAMs and myeloid-derived suppressor cells (MDSCs) are the most abundant cells in solid tumors including CRC. Moreover, other immune cell types have been identified in the CRC microenvironment, such as NK cells, TANs, eosinophils and mast cells, with variable roles in CRC progression (16, 17). CRC stroma is well-known for its ability to promote tumor-associated blood vessels. Immune cells and fibroblasts supply tumor cells with VEGF (18). Moreover, matrix metalloproteinase and associated proteases, expressed by CAFs, are abundant in TME.

Autophagy and colorectal cancer

Autophagy signaling in cancer

Autophagy has a diverse and dynamic impact on cancer cells that can affect both tumor initiation, progression and cancer

response to therapy. Recently, vast published data indicate a crosstalk between autophagy-related genes (ATG's) associated pathways with oncogenes and/or tumor suppressor genes. Indeed, the precise role of autophagy in modulating cancer tumorigenicity is highly complicated and is dependent on the context (19). Several autophagy genes might be involved in switching normal cells to CRC under particular conditions. The first autophagy marker indicated to be involved in colorectal carcinogenesis is LC3 (20). One of the LC3 isoforms, named LC3-II, is overexpressed in CRC cells particularly in advanced stages, compared to normal colon cells (21). Notably, low LC3 level has been interrelated to good CRC prognosis, particularly in advanced stages (22). Moreover, ATG5 and ATG10 showed a major role in CRC progression and chemotherapy resistance in several studies. ATG5 was found to be down-regulated in 95% of CRC cases, and its high expression level indicates lymphovascular invasion (23). In contrast, ATG10 was upregulated in CRC tissues and increased protein expression of ATG10 was accompanied by tumor lymph node metastasis and invasion (24). Another essential protein implicated in autophagy is the activating molecule in Beclin-1-regulated autophagy (Ambra1) protein encoded by the AMBRA1 gene. Mutated AMBRA1 gene was found in a subset of colorectal neoplasms (25). Additionally, Beclin-1 gene, UVRAG gene and Bif-1 gene were highly correlated with CRC carcinogenesis which is explained in the following sections.

Role of autophagy in colorectal cancer initiation

Autophagy is an equilibrating mechanism that promotes anti-malignant mechanism by clearance of unhealthy damaged proteins, DNA abnormalities and reactive oxygen species (ROS). A proper autophagic mechanism is crucial for the mutagen's elimination and appropriate genomic stability as it avoids the genetic defects accumulation that proceeds to malignant transformation. Thereby, autophagy might act as a tumor-suppressor in the early stages of the tumor. Evidence demonstrates that the tumor-suppressive effect is derived from some ATG-proteins such as Beclin-1, which shows antioncogenic properties. Tumor suppressor role of Beclin-1 is validated genetically in breast, ovarian and prostate tumors, as mono-allele deletion of Beclin-1 occurs (26, 27). However, Beclin-1 has a debatable role in CRC in that it promotes tumorigenesis, but may paradoxically inhibit CRC cell growth. Increased Beclin-1 expression was associated with better OS in patients with locally advanced colon carcinomas who received postoperative 5-FU chemotherapy for 6 months (28). Beclin-1 Overexpression in cases with resected stage II and stage III colon carcinomas, who received 5-FU-based therapy was associated with worse OS, denoting a potential effect of autophagy in drug resistance (29).

Moreover, allelic loss of UVRAG, an autophagy component, and attenuation of *Bif-1* expression that both interact with

Beclin-1 directly, might be correlated to CRC initiation and development (30). UVRAG protein is needed to form a complex with Beclin-1 to induce autophagy; therefore, the loss of this protein results in impaired autophagy machinery. Similarly, Bif-1 serves to induce autophagy *via* interacting with Beclin-1 and UVRAG.

Autophagy displays an important defense mechanism against pathogens and therefore plays an anticarcinogenic role in combatting viral and bacterial infections. For example, autophagic machinery was shown to effectively eliminate digestive cancer-associated pathogens such as Streptococcus bovis (S. bovis) that may cause CRC (31). In the same study, using autophagy-deficient ATG5-/- cells showed S. bovis pathogen survival and enhanced multiplication within the cells (31). The presence of infectious endocarditis of S. bovis may be followed by colonic neoplasia in an estimated incidence of 18-62% of cases, even after years of its presentation in the host (32, 33). Similarly, 25 to 80% of S. bovis bacteremia cases induce colorectal tumors (34). Despite this, the relationship between CRC and S. bovis bacteremia has been underestimated for a long time and is under the controversy of whether this association is a result of gastro-intestinal tumor or the S. bovis itself could be the etiology of CRC (35).

Role of autophagy in colorectal cancer cell survival and metastasis

In previous studies, autophagy seems to support tumor progression. Autophagy helps tumor cells overcome induced metabolic stress resulting from high proliferative rate, hypoxia and nutrient deprivation due to insufficient blood supply needed by these tumors for proliferation and progression (36, 37). Cancer cells consume more energy and metabolites than normal cells due to their rapid proliferative rate. Both energy and metabolites can be provided to cancer cells by increasing autophagy (38). Autophagy is considered a survival mechanism for cancer cells under hypoxic and metabolic stress conditions to provide them with the energy required for their survival and proliferation (39). In this regard, down-regulation of crucial autophagy proteins level led to restraining cancer growth and reduced oxygen consumption along with the accumulation of abnormal mitochondria, and specifically, autophagy was demonstrated to be essential to promote the growth of Rasdriven tumors, including CRC (40). Several in vitro studies indicated that gaining autophagy activity in Ras-driven cancer cells shows a significant increase in the survival and progression of those cancer cells in several settings of metabolic stress (41).

Besides its critical role in regulating protein turnover and cancer immunogenicity, autophagy has been involved in epithelial-to-mesenchymal transition (EMT), a crucial multistep mechanism needed by tumor cells to metastasize (42, 43). The commonly identified EMT inducer TGF β is known to induce EMT through the stimulation of SMAD, MAPK, Rho-GTPases and PI3K/AKT (44). During tumor progression, cells that undergo EMT need to stimulate autophagy machinery for their survival and metastases. In this regard, it has been demonstrated that autophagy is essential for EMT activation and cancer cell metastasis in hepatoblastoma cells (45). Similarly, autophagy is needed in TGF β 1-mediated EMT in non-small-cell lung cancer cells (46). In CRC cells and upon using rapamycin, a specific mTOR inhibitor and an autophagy inducer, starvation-mediated autophagy was demonstrated to induce invasion and migration and increase EMT marker expression; and interestingly, this was reverted by *Beclin-1* knockdown (47).

Effect of autophagy on cancer stem cells

Cancer stem cells (CSCs) are recognized to promote tumor initiation, progression and contribute to therapy resistance. CSCs drive tumor heterogeneity via EMT and inflammatory signaling activation (48). Autophagy is identified to promote the survival and control the pluripotency of CSCs in the TME. IL-17B/IL-17RB signaling induces autophagy, and subsequently, autophagy controls and maintains CSCs homeostasis. Interestingly, TRAF6 is recruited in the cytoplasm by IL-17B, which would induce autophagosome formation through Beclin-1 ubiquitination, thus promoting self-renewal and sphere-forming potential in gastric carcinoma (49). Likewise, IGF-2/insulin receptor signaling controls CSCs stemness and pluripotency through autophagy regulation. In CRC, loss of imprinted gene expression of IGF-2 indicated increased autophagy, leading to higher sphere-forming potential, and increased CD133 expression, which is a marker of stemness (50).

Increased autophagic flux is highly maintained and required by CSCs to promote therapy resistance. In CRC, SOX2 transcriptional factor increases the expression of EMT and *ABCC2* genes and promotes chemotherapy resistance through translocation and activation of β -catenin. Interestingly, SOX2 tends to increase *Beclin-1* expression to induce autophagy and promote chemoresistance. Thus, SOX2- β -catenin/Beclin-1/autophagy pathway is involved in tumor progression and chemotherapy resistance (51). A graphical illustration of the autophagy signaling pathway and its dual role in CRC initiation and progression is displayed in **Figure 1**.

Autophagy signaling modulates tumor microenvironment

Autophagy is actively involved in remodeling TME *via* unconventional secretion of several peptides, proteins and hormones that are typically operated and secreted through the conventional secretory system controlled by the endoplasmic reticulum–Golgi pathway (53). Knockdown of autophagy in both stromal cells and cancer cells is associated with a reduction of several cytokines and chemokines release including IL- 1β , IL-2, IL-6, IL-8, IL-18, CCL2, CCL20, TNF α , and LIF.

Herein, autophagy is capable of modulating tumor growth, metastasis and angiogenesis as well as immune evasion and stemness maintenance, through autophagy-dependent secretion of pro-inflammatory and pro-invasive factors (54–57). Another tumor secretome released in an autophagy-dependent manner includes growth factors (TGF- β 1, b-FGF), extracellular matrix proteins (MMP2, MMP9) and the angiogenesis stimulant (VEGFA) (Table 1) (55, 58, 59). Additionally, autophagy deficiency impedes the release and secretion of crucial cytokines and chemokines involved in T cells and DC recruitment, including IFN- γ , CXCL9, CXCL10, and CXCL11, thus immune surveillance escape occurred (Table 1) (60).

In contrast, autophagy stimulates the release of specific proteins known as DAMPs (damage-associated molecular patterns) that enhance an immunomodulatory effect by triggering immune cells. Therefore, it enhances the anti-tumor immunity and restricts tumor progression (61, 62).

Cross-talk of autophagy and anti-tumor immunity

In the age of immunotherapy success to fight cancer, there is an increasing demand to know how autophagy modulation affects the response to anti-cancer medications. Evidence suggested a decline in autophagy levels in aging T lymphocytes, indicating that autophagy inhibition might contribute to hematopoiesis and/or systemic immunity impairment (64). Furthermore, the survival of hematopoietic stem cells and memory T cells are dependent on autophagy (65, 66). In the myeloid compartment, autophagy supports B1 cell self-renewal and provides free fatty acids needed by the differentiating cells (67, 68). Additionally, autophagy has a major influence on the tumor-specific CD8+ T cells (69) and memory T-cells (70). Autophagy has been shown to dictate the degradation of cytolytic granules secreted by cytotoxic CD8+ T cells and NK cells (71, 72). Intriguingly, autophagy has a crucial role in protein degradation, thus allowing antigenpresenting cells (APCs), like DCs, to utilize such proteins as antigens on major histocompatibility complex (MHC)-I and II. The process occurs through three main pathways; namely, exogenous, cross-presentation, and endogenous pathways (Figure 2). Such role was previously reviewed by Koustas et al. (73).

Furthermore, immune suppressor cells have variable responses to autophagy inhibition. For instance, the immunosuppressive effect of Tregs is highly autophagy-dependent (12). Interestingly, it has been indicated that ATG5 or ATG7 deletion in T cells produces severe tumor implant rejection in the syngeneic mouse tumor model (74). Another published work demonstrated that inhibition of *Beclin-1* gene expression enhances T cells infiltration into the TME (75).

In the developed TME, TAMs, M2 phenotype, are vital in the growth and metastasis of cancer cells, as well as



angiogenesis (76). On the other hand, several studies proposed that M1 macrophages inhibit tumor progression (77). Autophagy has been shown to participate in the production and polarization of macrophages. Toll-like receptor-2 (TLR2) deficiency is associated with autophagy inhibition and subsequently results in the biosynthesis of M2-type macrophages, which in turn supports tumor progression (78). In addition, autophagy initiation in TAMs promotes apoptotic cell death, restrains proliferation, and enhances radiosensitivity of CRC (79). Altogether indicated that autophagy in TAM plays an essential role in suppressing cancer (Figure 2).

Furthermore, other native immune cells critically participate in CRC tumorigeneses, such as tumor-associated neutrophils TABLE 1 Summarized list of the crucial autophagy-dependent secretome and inflammatory mediator in TME.

Substances/ Secretome	Definition and function
TGF-β1	Transforming growth factor β -1 (TGF- β 1) is an important pleiotropic cytokine in wound healing, immunoregulation, angiogenesis and cancer. TGF- β 1 isoform is produced by immune cells that exert powerful anti-inflammatory functions.
β-FGF	Beta- Fibroblast Growth Factors (β -FGF) are involved in cell proliferation, differentiation, normal development, wound repair, and angiogenesis. β -FGF is mostly produced by stromal cells in bone marrow, leukemic cells, and T cells. β -FGF is an important regulator in the self-renewal and differentiation of multipotent hematopoietic progenitor cells.
MMP2	Matrix metalloproteinase-2 (gelatinase a); is a type IV collagenase that plays a role in vasculature remodeling, angiogenesis, tissue repair, tumor invasion, inflammation, and atherosclerotic plaque rupture. Also, MMP2 functions as degrading extracellular matrix proteins.
ММР9	Matrix metalloproteinase-9; potentially involved in local proteolysis of the extracellular matrix, leukocyte migration and bone osteoclastic resorption. Also, it cleaves type IV and type V collagen and fibronectin degradation.
VEGFA	Vascular endothelial growth factor-A is involved in angiogenesis, vasculogenesis and endothelial cell growth. As well as it Induces endothelial cell proliferation, promotes cell migration, inhibits apoptosis and induces permeabilization of blood vessels.
IFN-γ	Interferon γ ; Produced mostly by lymphocytes, has antiviral activity, and an important immunoregulatory functions. It acts as an activator of macrophages and has anti-proliferative effects on transformed cells. IFN- γ can potentiate the antitumor effects of the type I interferons.
CXCL9	C-X-C motif chemokine 9; is a cytokine that impacts the growth, movement, or involved in the immune and inflammatory response. It acts as a chemotactic for activated T-cells.
CXCL10	C-X-C motif chemokine 10; Chemotactic for monocytes and T-lymphocytes. Binds to CXCR3; Belongs to the intercrine alpha (chemokine CxC) family.
CXCL11	C-X-C motif chemokine 11 is an important chemotactic for interleukin-activated T-cells, neutrophils, or monocytes. CXCL11 induces calcium release in activated T-cells. Also, it is participating in CNS diseases that involve T-cell recruitment.

(TANs) and NK cells (Figure 2). For instance, promoting autophagy in TANs enhances the migration and metastasis of cancer cells (80). Analogous outcomes have been reported in other cancer types such as melanoma and renal cell carcinoma (81).

Autophagy as a regulator of immune-checkpoints

Additionally, autophagy has an impact on immune tolerance in response to immunotherapy, since immunologic molecules such as indoleamine 2,3-dioxygenase (IDO), Programmed cell death protein 1 (PD-1), and T-lymphocyteassociated protein 4 (CTLA-4) are regulated by autophagy pathways. IDO can inhibit tumor immunity through its inhibitory effects on cytotoxic T-cell responses, DC maturation, and Treg proliferation, thus promoting immune tolerance and tumor development. However, autophagy can inhibit the production of IDO in tumor sites (82, 83). Tumor cell PD-1 interacts with T-cells PD-L1 and serves as an inhibitory checkpoint molecule, preventing tumor cells from being recognized, thus suppressing the antitumor immunity. It has been reported that PD1 inhibits the availability of nutrients to nearby T-cells by interacting with its ligand, inducing autophagy (84). Results from experiments with murine melanoma cells and human ovarian cancer cells suggest that PD-L1-overexpressing cells are more responsive to autophagy inhibitors than cells with weak PD-L1 expression. This finding suggests that autophagy inhibitors may become an important therapeutic tool in PD-L1-overexpressing cancer cells (85). However, further experiments are warranted to explore how PD-L1 signaling and autophagy operate in different cell types, including CRC. This will assist in determining whether anti-PD-L1 therapy combined with autophagy inhibitors will enhance antitumor responses. The CTLA-4 protein is another immune tolerance checkpoint that can be targeted to treat tumors. A cancer-antigen called MAGE-A is associated with CTLA-4 inhibitor resistance and is known to suppress autophagy, suggesting that autophagy induction may be used therapeutically as a way to improve the efficacy of CTLA-4 inhibitors in human melanomas (86). Further experiments are needed to explore cross-talk of autophagy and immune checkpoints in CRC as well. Immune checkpoint therapy for CRC, as a whole, remains unsatisfactory at present. However, there has been renewed interest in examining additional immune checkpoint molecules. New immune checkpoint targets have been identified like the T cell immunoglobulin and mucin domain containing-3 (TIM-3), the V-domain Ig suppressor of T cell activation (VISTA), the T cell immunoglobulin and ITIM domain (TIGIT), and the lymphocyte activation gene-3 (LAG-3) (87-89). Despite an exponential growth in clinical trials for emerging immune modulators, such as anti-LAG-3 antibodies and anti-TIM-3 antibodies, registered on ClinicalTrials.gov, no drugs have yet been approved for clinical use. Despite promising monotherapy results, more effort needs to be integrated toward developing rational combinations of immune-therapy to inhibit cancer growth through non-redundant pathways that work synergistically.



Cross-talk of autophagy and endothelial cells

The innermost layer of blood vessels is lined by endothelial cells. In addition to being essential for normal tissue function, new blood vessels also play an important role in cancer pathology. For tumor cells to grow and spread, neovascularization is necessary. Tumor endothelial cells have a multifaceted functional role since they are not only responsible for enhancing angiogenesis, but are also important in immune regulation in the TME (90). Regulatory mechanisms profoundly influence peripheral immune cell recruitment into the TME by acting as significant gatekeepers during cellular transmigration (91-93). Furthermore, tumor endothelial cells act as antigenpresenting cells (APCs), which are associated with T cell activation, proliferation, and priming (92). Furthermore, tumor endothelial cells are required for the development of "tertiary lymphoid structures," which are associated with the response to checkpoint antibody therapy (94). Other qualities that distinguish tumor endothelial cells from normal endothelial cells are their high proliferation potential and markedly changed gene expression profile (i.e., an increase in proangiogenic, extracellular matrix remodeling, and stemness genes), leading to increased secretion of immunomodulatory cytokines and altered cell-surface receptors, e.g., MHC and immune checkpoints (90, 95). It is possible that the tumor endothelial cells phenotype is rooted in an aggressive tumor micro-milieu driven by hypoxia and ROS (96, 97). In clinical practice, chemotherapy combined with angiogenesis inhibitor results in marked enhancement of anti-cancer effects in patients with metastatic CRC (98).

Increasing evidence suggests that autophagy impacts endothelial cell survival, proliferation, migration and angiogenesis. However, whether autophagy regulates angiogenesis positively or negatively is still debated. For instance, according to Du et al., overexpressing ATG5 induced autophagy in bovine endothelial cells resulting in enhanced formation and migration in those endothelial cells while 3-methyladenine (3-MA) or siRNA targeting ATG5 reduced angiogenesis (99). A study by Goyal et al. discovered that decorin-induced autophagy provided protection against tumor neovascularization and epithelial death (100). Autocrine VEGF released from endothelial cells and gastrin-releasing peptide (GRP) secreted by tumors promote angiogenesis, endothelial survival, and proliferation of endothelial cells by inhibiting autophagy (101). Moreover, a study carried out by Seon-Jin Lee

et al. established that genetically disrupting *Beclin1* can increase tumor growth and angiogenesis in hypoxic environments (102). A broader view suggests that autophagy can influence the angiogenesis process, which is important to tumor growth, by affecting the function and survival of endothelial cells, which has a pro- or anti-tumor effect on CRC.

Autophagy and colorectal cancer metabolism

Autophagy is a conserved catabolic process by which various proteins, cytoplasmic constituents and organelles can re-enter the different metabolic processes. Cancer cells altered their metabolism, thus promoting their proliferation, progression, and long-term survival. Cancer cells enhance glucose uptake and metabolize glucose to lactate even when completely functioning mitochondria support the oxidative phosphorylation mechanism, altogether is known as Warburg effect (103). In the normal process, pyruvate kinase (PKM2), the enzyme catalyzing the last step in the glycolytic process, takes control of the glycolytic flux, preventing the excessive accumulation of glycolytic metabolites (104, 105). However, pyruvate kinase (PKM2) enzyme breakdown is enhanced in cancer cells via chaperon-mediated autophagy, thus associated with increased accumulation of glycolytic metabolites (106). Also, hexokinase 2 (HK2), rate-limiting enzyme of the glycolytic pathway, is selectively damaged by autophagy in liver carcinoma (107, 108). Therefore, autophagy plays a vital role in cancer metabolism via controlling glycolysis at different stages and levels. Warburg effect elevates lactate level in the TME that disturbs the extracellular environmental pH, resulting in autophagy activation (109). For instance, acute acidification of breast cancer cells results in increased expression of LC3, ATG5, and BNIP3 (110). Therefore, autophagy destructive effect on vital metabolic enzymes may critically influence many features of central metabolism in cancer. Hence, autophagy contributes to malignancy progression and transformation by providing cancer cells with the efficient ability to re-distribute metabolites allowing metabolic rewiring.

Moreover, as a result of starvation, infections, and cancer, glutamine homeostasis is disturbed and the need for exogenous glutamine to promote cell survival and growth is increasing (111). Due to the Warburg effect, glutamine is excessively required to sustain oxidative phosphorylation through its role as a key intermediate in the tricarboxylic acid (TCA) cycle. Furthermore, it is the main nitrogen source for many aminotransferase enzymes involved in the synthesis of nucleotides and non-essential amino acids (112). Glutamine participates in redox homeostasis by contributing to NADH/NADPH synthesis and glutamate synthesis, which is critical for glutathione synthesis (112). Therefore, with such a wide range of glutamine functions, it is critical for some cancers including CRC to ensure an adequate glutamine supply (113). Targeting glutamine transport and metabolism has therefore been a promising approach for treating CRC (113). As soon as glutamine is deficient or lacking, the cells show differential manifestations, including a pronounced decline in ATP and NADH, as well as a significant accumulation of ROS (114, 115). Herein, Autophagy plays an important role in this adaptive response by suppressing glutamine-consuming processes and elevating glutamine content in the body. Macro-pinocytosis is one of the mechanisms by which activated autophagy restores glutamine levels via recycling intracellular proteins and extracellular compartments (116). Meanwhile, some reports claim that autophagy plays a crucial role in cancers that escape death with high success rates (117). Upon limitation of exogenous glutamine, inhibition of autophagy in SW620 and SW480 colorectal cell lines resulted in increased apoptotic activity (118). In the same way, chronic activation of mTORC1 may result in severe mTORC1-dependent cell death (later termed glutamoptosis), ultimately inhibiting autophagy (119). In nutrient starvation, autophagy activation is often associated with cell survival. However, over-activating autophagy in specific contexts has shown anti-tumor potential.

Role of autophagy in the regulation of hypoxia and oxidative stress in tumor microenvironment

Autophagy plays a pivotal role in helping cancer cells adapt and survive under hypoxic TME. Intriguingly, autophagy promotes the survival of cancer cells through its main effector, hypoxia-inducible factor- 1α (HIF- 1α), which is mostly the case in solid tumors, specially CRC (120). Tumor cells can endure hypoxia through Beclin1-mediated cytoprotective autophagy by upregulating the transcription of BNIP3 and BNIP3L (121). Moreover, BNIP3L/NIX functions as a selective receptor for autophagy that is highly expressed in tumor cells, which is crucial to promote mitophagy under hypoxic TME through NFE2L2/NRF2 transactivation. In addition, cells overexpressing NIX, are more susceptible to acquire glioma stem cell-like properties via mTOR/AKT/HIF pathway (122). Under hypoxic conditions, a crucial adaptor protein, FUNDC1, is triggered to eliminate dysfunctional mitochondria. FUNDC1 protein functions critically in autophagy via engaging with LC3 protein through LC3 interacting region (LIR) of FUNDC1 (123). Additional form of autophagy regulation under hypoxia occurs via HMGB1 signaling through upregulating YAP expression in tumor cells. Similarly, ATG5 and ATG12 are stimulated by PAK1 acetylation and PTBP3, respectively, resulting in promoting pro-survival autophagy. Furthermore, an important kinase, PRKCA/PKCa, that regulates hypoxia-mediated autophagy via ATG5 and Beclin1, stimulates tumor-initiating cell renewal in CRC (124). Likewise, YTHDF1 gene is activated by HIF-1α to promote autophagy protective effect through ATG2A and ATG14. Of note, protein phosphatase 2 (PP2A) along with mTOR downstream kinase signaling pathways control the prolyl hydroxylase domain-containing protein 2 (PHD2) phosphorylation to govern and promote HIF-1a mediated autophagy in CRC cells survival (125). Also, *ANKRD37* gene is demonstrated to induce HIF-1 α mediated autophagy in hypoxic colon cancer once it translocates to the nucleus (126).

Hypoxia-mediated HIF-1 α induction is reported to promote autophagy, thus controlling glycolytic processes to maintain energy supply and cell progression. In this regard and under hypoxic conditions, proline gets metabolized into pyrroline-5-carboxylate (P5C) with the help of proline oxidase (POX) enzyme, which elicits ROS production that promotes protective autophagy mechanism, which is necessary for the survival of HT29 cells (127). Proline oxidase (POX) enzyme role is AMPKdependent; however, it is controlled in HIF-1 α and HIF-2 α independent manner (127).

Interestingly, autophagy was demonstrated to restrain oxidative stress-dependent inflammation and promote tumorsuppressor mechanisms. For instance, the transcription activator "BRG1" stimulates autophagosome biogenesis by regulating the transcription of *ATG7*, *AMBRA1*, and *Wipi2*, thus attenuating colonic inflammation and CRC development in an oxidative stress-mediated autophagy manner (128).

Autophagy targeted therapy in colorectal cancer

Recent and ongoing clinical trials

Despite the controversial and contextual relationship between cancer and autophagy, it is still considered a promising target for treatment, as many shared regulatory pathways of carcinogenesis and autophagy are involved. Some studies demonstrated that autophagy induction is highly correlated to the resistance of cancer cells to chemotherapy, immunotherapy, and radiotherapy via directly modulating cancer cell metabolism or diminishing cell death pathway (72, 129-131). Thus, various preclinical and clinical studies have been conducted to develop pharmacological autophagy inhibitors (132). The most recent development of autophagy inhibitors can be known by tracing the clinical trials (Table 2). The most effective targeted therapies recognized in CRC treatment, so far, are anti-angiogenesis such as cabozantinib, apatinib and bevacizumab, and the inhibitors of epidermal growth factor receptor (anti-EGFR) such as cetuximab (133).

For decades, chloroquine has been approved in malaria and arthritis treatment and is currently an inhibitor of autophagy *via* inhibiting the fusion of autophagosomes with lysosomes in the last step of autophagy machinery. Hence, many clinical trials are investigating chloroquine or chloroquine derivatives either alone or in chemotherapy or radiotherapy combinations in patients suffering from different forms of cancers. One trial named CHOICES (Chloroquine and Imatinib Combination to Eliminate Stem cells), a phase II trial, is investigating and comparing the effect of imatinib and

hydroxychloroquine combination versus imatinib alone in patients with chronic myeloid leukemia, establishing evidence of autophagy inhibitors concept (134). Apatinib, a tyrosine kinase inhibitor of VEGFR2, has been indicated to stimulate autophagy via AKT- mTOR signaling pathway in colon cancer cells (135). Additionally, Cabozantinib is an inhibitor of various kinases responsible for angiogenesis, cell growth and metabolism that showed a major autophagy induction in HCT116 and HT29 CRC cell lines. Notably, cabozantinib in combination with autophagy inhibitors promotes apoptosis in HT29 and HCT116 cells (136). In a study using CRC cell lines, bevacizumab stimulates autophagy as evidenced by punctate patterns of LC3, autophagic vacuoles presence and Beclin-1 accumulation. Autophagy inhibition by targeting ATG5 and Beclin-1, via RNA interference or chloroquine, enhances the ability of bevacizumab to induce apoptosis and prevent proliferation, verifying the protective role of autophagy. Similarly, in vivo studies using small interfering RNA or chloroquine and bevacizumab combination showed significant inhibition in tumor growth when compared to bevacizumab monotherapy (137).

Of note, a combination of temozolomide and hydroxychloroquine is indicated to be safe and tolerable as well as exerted beneficial anti-tumor effect in phase I trial in patients with solid tumors, including CRC, and in advanced melanoma (138). Similarly, another phase I trial documented the significant efficacy of hydroxychloroquine in combination with mTOR inhibitor temsirolimus in tumor suppression (139). On the other hand, a recent phase I study showed that hydroxychloroquine treatment with AKT inhibitor MK-2206 is tolerable but with minimal anti-tumor activity in solid tumors including CRC (140). As evidenced by multiple instances previously reported, autophagy inhibitors as monotherapy might not be a good treatment choice for cancer therapy (141). Treatment combination of hydroxychloroquine with HDAC inhibitor vorinostat in an ongoing phase I study for patients with advanced renal and colorectal cancers shows no significant clinical improvement in the safety profile and in the patient PFS, indicating a limited benefit of adding hydroxychloroquine (Table 2) (142).

In a study on CRC cell lines, autophagy inhibition by 3-MA showed significant 5-FU-induced apoptosis, thus autophagy might have a crucial role in enhancing response of colon cancer cells treated with 5-FU (143). Likewise, another study using chloroquine, an autophagy inhibitor, in combination with 5-FU showed an enhanced anti-proliferative effect of 5-FU in CRC cells (144). More, inhibiting late-stage autophagy has been demonstrated to enhance the apoptotic cell death activity of the pyrrolo-1,5-benzoxazepines (PBOXs) in human CRC cells (145). Moreover, UAMC-2526 displays inhibitory effects on ATG4. This compound abolishes autophagy in mice bearing colorectal tumors and promotes chemotherapy-induced cell death (146). Recent *in vitro* assays and *in silico* screening has

Treatment	Target of the treatment	Phase	Patients number	Status	Outcome	Trial reference number at ClinicalTrials.gov/References
Vorinostat + HCQ	Histone deacetylase (HDAC) inhibitor.	Ι	72	Active not recruiting	No significant clinical improvement in the safety profile and the progression-free survival.	NCT01023737 (142)
Temsirolimus + HCQ	mTOR inhibitor.	Ι	40	completed	Safe and tolerable, Significant tumor suppression effect.	NCT00909831 (139)
Temozolomide + HCQ	DNA alkylating agent/induce cell cycle arrest at G2/M.	Ι	38	completed	Safe and tolerable, beneficial anti-tumor effect.	NCT00714181 (138)
Protein kinase B Akt inhibitor (MK- 2206) + HCQ	Akt inhibitor.	Ι	62	Active not recruiting	Tolerable, minimal anti-tumor activity.	NCT01480154 (140)
HCQ, FOLFOX and bevacizumab.	FOLFOX: chemotherapy that inhibits DNA synthesis. Bevacizumab: VEGF/VEGF receptor inhibitor.	Π	38	completed	Increases in autophagy marker LC3 with a complete response rate of 11% but without improved OS in the 28 evaluable patients.	NCT01006369 (98)

TABLE 2 Previous and current clinical trials involving hydroxychloroquine (HCQ) in combination with a variety of anti-cancer targeted agents in CRC.

identified a new, important ATG4B inhibitor (S130) that has the ability to interfere with ATG4 proteolytic activity but not with other proteases. Also, S130 is well distributed in tissues *in vivo*, enhances cell death in CRC and reduces the tumor size (147). These findings identify ATG4B as a potential anticancer target.

Challenges and potential solutions of the autophagy targeted treatment

Based on studies and clinical trials described above, it seems that autophagy inhibitors have a different clinical response in cancer therapy. Identification of good biomarkers with suitable pharmaco-dynamic properties that can estimate any change in autophagy, is of the major difficulties facing scientists (148). It remains to be explored whether the limited clinical efficacy of chloroquine is correlated with its lack of specificity in inhibiting autophagy. In fact, both chloroquine and hydroxychloroquine are non-selective autophagy inhibitors which are evident by their role in the reduction of nutrient scavenging (149). This diminished targeted delivery results in plummeting the bioavailability of the drugs. However, hydroxychloroquine is characterized by higher bioavailability compared to chloroquine. Moreover, both drugs have been identified to modify the pH of tumors, hence resulting in bioavailability modulation of different cytotoxic drugs when used in combination (150). Furthermore, frequent use of chloroquine has been identified for a long time to elicit renal failure (151). Noteworthy, both hydroxychloroquine and chloroquine could affect pacemaker channels and voltage-gated Na⁺, Ca²⁺, and K⁺ ion channels in the heart, leading to serious dysrhythmias.

In this regard, there is an urgent need for novel safe autophagy inhibitors with selective targets and a good bioavailability; properties that many proposed drugs failed to reach. One of the major advancements in the field is the discovery of Lys05, a dimeric form of chloroquine, which shows higher accumulation capabilities in the lysosome. Also, Lys05 has been identified to exert potent monotherapy anti-tumor activity in both *in vitro* and preclinical mouse models with limited toxicity in the treated mice. Of note, Lys05 potent characteristic in autophagy inhibition is dependent on C7-Chlorine, bivalent aminoquinoline rings and a short tri-amine linker (152).

Recently, new druggable autophagy target proteins have been established, including Vps34 (or class III PI3K) and Beclin-1. Notably, both proteins are involved in the early autophagy initiation process. A kinase inhibitor, SAR405, inhibits both Vps34 and Vps18, thus diminishing the lysosomal function *via* disturbing the vesicle trafficking between the lysosome and the late endosome. Further, SAR405 has been found to prevent mTOR- and starvation-dependent stimulation of autophagy (153).

Another druggable protein for autophagy modulation which has been recently proposed is the serine/threonine kinase ULK1/ATG1. Identification of small-molecule SBI-0206965, a potent ULK1 inhibitor, was happened through cell-based screen. This inhibitor was found to be high *in vitro* selective for ULK1 kinase as well as suppressed phosphorylation events mediated

10.3389/fmed.2022.959348

by ULK1 kinases. Markedly, SBI-0206965 anti-tumor effect has been evidenced *in vivo* as it showed potent tumor inhibition when combined with mTOR inhibitors, hence allowing it for use in the clinic (154). However, a major limitation of this molecule is that it could affect the activity of other kinases including JAK3, FLT3, FAK, and Src.

Conclusion and perspectives

A large number of proteins involved in the complex process of autophagy, which appears to play a significant role in all stages of carcinogenesis as it impacts tumor progression, initiation and metastatic capacity. Although the role of autophagy is not fully understood in cancer, it is thought to play both a promoting and inhibiting role depending on the context. Thus, it is imperative to identify how these apparently paradoxical roles of autophagy are regulated in CRC, and to constitute an overall view of the mechanisms that enable autophagy to play one role, not the other.

Autophagy modulates the effect of hypoxia and oxidative stress, regulates metabolism, promotes cancer stem cells and constrains the surveillance of immune cells to support cancer progression. The development of several therapeutic agents that modulate autophagy in CRC has led to promising results, supporting their use to enhance the action of other medications. Currently, autophagy inhibitors used in cancer therapy are limited to hydroxychloroquine and chloroquine that require close monitoring, when used for a prolonged period, for hepatic and renal adverse effects. Therefore, there is an urgent need for more translational and basic research to clarify the intricate role of autophagy, and to resolve unanswered questions about the enhanced efficacy of autophagy-targeted cancer therapy. Notably, there is an increased interest in personalized cancer treatment by joining the TME modulation status with advanced

References

1. Bray F, Ferlay J, Soerjomataram I, Siegel RL, Torre LA, Jemal A. Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA Cancer J Clin.* (2018) 68:394–424. doi: 10.3322/caac.21492

2. Elysia MA, Force LM, Xu R, Compton K, Lu D, Henrikson HJ, et al. The global burden of adolescent and young adult cancer in 2019: a systematic analysis for the Global burden of disease study 2019. *Lancet Oncol.* (2022) 23:27–52. doi: 10.1016/S1470-2045(21)00581-7

3. Douaiher J, Ravipati A, Grams B, Chowdhury S, Alatise O, Are C. Colorectal cancer—global burden, trends, and geographical variations. *J Surg Oncol.* (2017) 115:619–30. doi: 10.1002/jso.24578

4. Bahrami A, Hesari A, Khazaei M, Hassanian SM, Ferns GA, Avan A. The therapeutic potential of targeting the BRAF mutation in patients with colorectal cancer. *J Cell Physiol.* (2018) 233:2162–9. doi: 10.1002/jcp.25952

technology to explore the alteration in cancer progression. This will hopefully propose a major success in cancer therapy.

Author contributions

EM and MS-A: conceptualization. EM: writing—original draft preparation and visualization. JT, NS, and MS-A: writing—review and editing. All authors have read and agreed to the published version of the manuscript.

Funding

This work was supported by research grant Nos. (CoV19 #0309 and #0304), Competitive grant (#22010902104), University of Sharjah, United Arab Emirates, and the MBRU-AlMahmeed Research Award, (ALM #1914).

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

5. Van Cutsem E, Cervantes A, Nordlinger B, Arnold D. Metastatic colorectal cancer: ESMO clinical practice guidelines for diagnosis, treatment and follow-up. *Ann Oncol.* (2014) 25:iii1–9. doi: 10.1093/annonc/mdu260

^{6.} Konda B, Shum H, Rajdev L. Anti-angiogenic agents in metastatic colorectal cancer. *World J Gastrointest Oncol.* (2015) 7:71. doi: 10.4251/wjgo.v7.i7.71

^{7.} Guinney J, Dienstmann R, Wang X, De Reynies A, Schlicker A, Soneson C, et al. The consensus molecular subtypes of colorectal cancer. *Nat Med.* (2015) 21:1350–6.

^{8.} Fridman WH, Miller I, Sautès-Fridman C, Byrne AT. Therapeutic targeting of the colorectal tumor stroma. *Gastroenterology*. (2020) 158:303–21. doi: 10.1053/j. gastro.2019.09.045

^{9.} Bupathi M, Wu C. Biomarkers for immune therapy in colorectal cancer: mismatch-repair deficiency and others. *J Gastrointest Oncol.* (2016) 7:713. doi: 10.21037/jgo.2016.07.03

10. Derhovanessian E, Maier AB, Beck R, Jahn G, Hähnel K, Slagboom PE, et al. Hallmark features of immunosenescence are absent in familial longevity. *J Immunol.* (2010) 185:4618–24. doi: 10.4049/jimmunol.1001629

11. Hu Z, Ma Y, Shang Z, Hu S, Liang K, Liang W, et al. Improving immunotherapy for colorectal cancer using dendritic cells combined with anti-programmed death-ligand *in vitro*. *Oncol Lett.* (2018) 15:5345–51. doi: 10.3892/ ol.2018.7978

12. Wei J, Long L, Yang K, Guy C, Shrestha S, Chen Z, et al. Autophagy enforces functional integrity of regulatory T cells by coupling environmental cues and metabolic homeostasis. *Nat Immunol.* (2016) 17:277–85. doi: 10.1038/ni.3365

13. Gao D, Mittal V. The role of bone-marrow-derived cells in tumor growth, metastasis initiation and progression. *Trends Mol Med.* (2009) 15:333–43. doi: 10.1016/j.molmed.2009.06.006

14. Gabrilovich DI, Ostrand-Rosenberg S, Bronte V. Coordinated regulation of myeloid cells by tumours. *Nat Rev Immunol.* (2012) 12:253-68. doi: 10.1038/ nri3175

15. Szebeni GJ, Vizler C, Nagy LI, Kitajka K, Puskas LG. Pro-tumoral inflammatory myeloid cells as emerging therapeutic targets. *Int J Mol Sci.* (2016) 17:1958. doi: 10.3390/ijms17111958

16. Charoentong P, Finotello F, Angelova M, Mayer C, Efremova M, Rieder D, et al. Pan-cancer immunogenomic analyses reveal genotype-immunophenotype relationships and predictors of response to checkpoint blockade. *Cell Rep.* (2017) 18:248–62. doi: 10.1016/j.celrep.2016.12.019

17. Halama N, Braun M, Kahlert C, Spille A, Quack C, Rahbari N, et al. Natural killer cells are scarce in colorectal carcinoma tissue despite high levels of chemokines and cytokines. *Clin Cancer Res.* (2011) 17:678–89. doi: 10.1158/1078-0432.CCR-10-2173

18. O'Connell JT, Sugimoto H, Cooke VG, MacDonald BA, Mehta AI, LeBleu VS, et al. VEGF-A and Tenascin-C produced by S100A4+ stromal cells are important for metastatic colonization. *Proc Natl Acad Sci U.S.A.* (2011) 108:16002–7. doi: 10.1073/pnas.1109493108

19. Galluzzi L, Pietrocola F, Bravo-San Pedro JM, Amaravadi RK, Baehrecke EH, Cecconi F, et al. Autophagy in malignant transformation and cancer progression. *EMBO J.* (2015) 34:856–80. doi: 10.15252/embj.201490784

20. Tanida I, Ueno T, Kominami E. LC3 conjugation system in mammalian autophagy. *Int J Biochem Cell Biol.* (2004) 36:2503–18. doi: 10.1016/j.biocel.2004. 05.009

21. Chen Z, Li Y, Zhang C, Yi H, Wu C, Wang J, et al. Downregulation of beclin1 and impairment of autophagy in a small population of colorectal cancer. *Digest Dis Sci.* (2013) 58:2887–94. doi: 10.1007/s10620-013-2732-8

22. Yang M, Zhao H, Guo L, Zhang Q, Zhao L, Bai S, et al. Autophagy-based survival prognosis in human colorectal carcinoma. *Oncotarget.* (2015) 6:7084. doi: 10.18632/oncotarget.3054

23. Cho D-H, Jo YK, Kim SC, Park IJ, Kim JC. Down-regulated expression of ATG5 in colorectal cancer. *Anticancer Res.* (2012) 32:4091–6.

24. Jo YK, Kim SC, Park IJ, Park SJ, Jin D-H, Hong S-W, et al. Increased expression of ATG10 in colorectal cancer is associated with lymphovascular invasion and lymph node metastasis. *PLoS One.* (2012) 7:e52705. doi: 10.1371/journal.pone.0052705

25. Cianfanelli V, Fuoco C, Lorente M, Salazar M, Quondamatteo F, Gherardini PF, et al. AMBRA1 links autophagy to cell proliferation and tumorigenesis by promoting c-Myc dephosphorylation and degradation. *Nat Cell Biol.* (2015) 17:20–30. doi: 10.1038/ncb3072

26. Qu X, Yu J, Bhagat G, Furuya N, Hibshoosh H, Troxel A, et al. Promotion of tumorigenesis by heterozygous disruption of the beclin 1 autophagy gene. *J Clin Investig.* (2003) 112:1809–20. doi: 10.1172/JCI20039

27. Gong C, Bauvy C, Tonelli G, Yue W, Delomenie C, Nicolas V, et al. Beclin 1 and autophagy are required for the tumorigenicity of breast cancer stemlike/progenitor cells. *Oncogene*. (2013) 32:2261–72. doi: 10.1038/onc.2012.252

28. Li B-X, Li C-Y, Peng R-Q, Wu X-J, Wang H-Y, Wan D-S, et al. The expression of beclin 1 is associated with favorable prognosis in stage IIIB colon cancers. *Autophagy*. (2009) 5:303–6. doi: 10.4161/auto.5.3.7491

29. Myung Park J, Huang S, Wu T-T, Foster NR, Sinicrope FA. Prognostic impact of beclin 1, p62/sequestosome 1 and LC3 protein expression in colon carcinomas from patients receiving 5-fluorouracil as adjuvant chemotherapy. *Cancer Biol Ther.* (2013) 14:100–7. doi: 10.4161/cbt.22954

30. Ionov Y, Nowak N, Perucho M, Markowitz S, Cowell JK. Manipulation of nonsense mediated decay identifies gene mutations in colon cancer C ells with microsatellite instability. *Oncogene*. (2004) 23:639–45. doi: 10.1038/sj.onc. 1207178

31. Nakagawa I, Amano A, Mizushima N, Yamamoto A, Yamaguchi H, Kamimoto T, et al. Autophagy defends cells against invading group A *Streptococcus. Science.* (2004) 306:1037-40. doi: 10.1126/science.1103966

32. Gupta A, Madani R, Mukhtar H. *Streptococcus bovis* endocarditis, a silent sign for colonic tumour. *Colorectal Dis.* (2010) 12:164–71. doi: 10.1111/j.1463-1318.2009.01814.x

33. Zarkin BA, Lillemoe KD, Cameron JL, Effron PN, Magnuson TH, Pitt HA. The triad of *Streptococcus bovis* bacteremia, colonic pathology, and liver disease. *Ann Surg.* (1990) 211:786. doi: 10.1097/00000658-199006000-00019

34. Abdulamir AS, Hafidh RR, Bakar FA. The association of *Streptococcus bovis/gallolyticus* with colorectal tumors: the nature and the underlying mechanisms of its etiological role. *J Exp Clin Cancer Res.* (2011) 30:1–13. doi: 10.1186/1756-9966-30-11

35. Meseeha M, Attia M. *Streptococcus bovis* bactremia and colon cancer: a well established but frequently forgotten association: 1456. *Off J Am Coll Gastroenterol.* (2018) 113:S835–6

36. White E, DiPaola RS. The double-edged sword of autophagy modulation in cancer. *Clin Cancer Res.* (2009) 15:5308–16. doi: 10.1158/1078-0432.CCR-07-5023

37. Boya P, Reggiori F, Codogno P. Emerging regulation and functions of autophagy. Nat Cell Biol. (2013) 15:713-20. doi: 10.1038/ncb2788

38. White E. Deconvoluting the context-dependent role for autophagy in cancer. *Nat Rev Cancer.* (2012) 12:401–10. doi: 10.1038/nrc3262

39. Degenhardt K, Mathew R, Beaudoin B, Bray K, Anderson D, Chen G, et al. Autophagy promotes tumor cell survival and restricts necrosis, inflammation, and tumorigenesis. *Cancer Cell.* (2006) 10:51–64. doi: 10.1016/j.ccr.2006.06.001

40. Guo JY, Chen H-Y, Mathew R, Fan J, Strohecker AM, Karsli-Uzunbas G, et al. Activated Ras requires autophagy to maintain oxidative metabolism and tumorigenesis. *Genes Dev.* (2011) 25:460–70. doi: 10.1101/gad.201 6311

41. Yang A, Rajeshkumar N, Wang X, Yabuuchi S, Alexander BM, Chu GC, et al. Autophagy is critical for pancreatic tumor growth and progression in tumors with p53 alterations. *Cancer Discov.* (2014) 4:905–13. doi: 10.1158/2159-8290.CD-14-0362

42. Rojas-Sanchez G, Cotzomi-Ortega I, Pazos-Salazar NG, Reyes-Leyva J, Maycotte P. Autophagy and its relationship to epithelial to mesenchymal transition: when autophagy inhibition for cancer therapy turns counterproductive. *Biology*. (2019) 8:71. doi: 10.3390/biology8040071

43. Bustos SO, Antunes F, Rangel MC, Chammas R. Emerging autophagy functions shape the tumor microenvironment and play a role in cancer progression - implications for cancer therapy. *Front Oncol.* (2020) 10:606436. doi:10.3389/fonc.2020.606436

44. Lamouille S, Xu J, Derynck R. Molecular mechanisms of epithelialmesenchymal transition. *Nat Rev Mol Cell Biol.* (2014) 15:178–96. doi: 10.1038/ nrm3758

45. Chen C, Liang QY, Chen HK, Wu PF, Feng ZY, Ma XM, et al. DRAM1 regulates the migration and invasion of hepatoblastoma cells *via* autophagy-EMT pathway. *Oncol Lett.* (2018) 16:2427–33. doi: 10.3892/ol.2018.8937

46. Alizadeh J, Glogowska A, Thliveris J, Kalantari F, Shojaei S, Hombach-Klonisch S, et al. Autophagy modulates transforming growth factor beta 1 induced epithelial to mesenchymal transition in non-small cell lung cancer cells. *Biochim Biophys Acta Mol Cell Res.* (2018) 1865:749–68. doi: 10.1016/j.bbamcr.2018.02.007

47. Shen H, Yin L, Deng G, Guo C, Han Y, Li Y, et al. Knockdown of beclin-1 impairs epithelial-mesenchymal transition of colon cancer cells. *J Cell Biochem.* (2018) 119:7022–31. doi: 10.1002/jcb.26912

48. Bocci F, Gearhart-Serna L, Boareto M, Ribeiro M, Ben-Jacob E, Devi GR, et al. Toward understanding cancer stem cell heterogeneity in the tumor microenvironment. *Proc Natl Acad Sci U.S.A.* (2019) 116:148–57. doi: 10.1073/pnas.1815345116

49. Bie Q, Song H, Chen X, Yang X, Shi S, Zhang L, et al. IL-17B/IL-17RB signaling cascade contributes to self-renewal and tumorigenesis of cancer stem cells by regulating beclin-1 ubiquitination. *Oncogene*. (2021) 40:2200–16. doi: 10.1038/s41388-021-01699-4

50. Gao T, Liu X, He B, Pan Y, Wang S. IGF2 loss of imprinting enhances colorectal cancer stem cells pluripotency by promoting tumor autophagy. *Aging (Albany NY).* (2020) 12:21236. doi: 10.18632/aging.103837

51. Zhu Y, Huang S, Chen S, Chen J, Wang Z, Wang Y, et al. SOX2 promotes chemoresistance, cancer stem cells properties, and epithelial–mesenchymal transition by β -catenin and beclin1/autophagy signaling in colorectal cancer. *Cell Death Dis.* (2021) 12:1–16. doi: 10.1038/s41419-021-03733-5

52. Burada F, Nicoli ER, Ciurea ME, Uscatu DC, Ioana M, Gheonea DI. Autophagy in colorectal cancer: an important switch from physiology to pathology. *World J Gastrointest Oncol.* (2015) 7:271. doi: 10.4251/wjgo.v7.i11.271

53. Keulers TG, Schaaf MB, Rouschop K. Autophagy-dependent secretion: contribution to tumor progression. *Front Oncol.* (2016) 6:251. doi: 10.3389/fonc. 2016.00251

54. Dupont N, Jiang S, Pilli M, Ornatowski W, Bhattacharya D, Deretic V. Autophagy-based unconventional secretory pathway for extracellular delivery of IL-1β. *EMBO J.* (2011) 30:4701–11. doi: 10.1038/emboj.2011.398

55. Zhan Z, Xie X, Cao H, Zhou X, Zhang XD, Fan H, et al. Autophagy facilitates TLR4-and TLR3-triggered migration and invasion of lung cancer cells through the promotion of TRAF6 ubiquitination. *Autophagy.* (2014) 10:257–68. doi: 10.4161/auto.27162

56. Maycotte P, Jones KL, Goodall ML, Thorburn J, Thorburn A. Autophagy supports breast cancer stem cell maintenance by regulating IL6 secretion. *Mol Cancer Res.* (2015) 13:651–8. doi: 10.1158/1541-7786.MCR-14-0487

57. Salah F, Ebbinghaus M, Muley V, Zhou Z, Al-Saadi K, Pacyna-Gengelbach M, et al. Tumor suppression in mice lacking GABARAP, an Atg8/LC3 family member implicated in autophagy, is associated with alterations in cytokine secretion and cell death. *Cell Death Dis.* (2016) 7:e2205. doi: 10.1038/cddis.2016.93

58. New J, Arnold L, Ananth M, Alvi S, Thornton M, Werner L, et al. Secretory autophagy in cancer-associated fibroblasts promotes head and neck cancer progression and offers a novel therapeutic target. *Cancer Res.* (2017) 77:6679–91. doi: 10.1158/0008-5472.CAN-17-1077

59. Nüchel J, Ghatak S, Zuk AV, Illerhaus A, Mörgelin M, Schönborn K, et al. TGFB1 is secreted through an unconventional pathway dependent on the autophagic machinery and cytoskeletal regulators. *Autophagy.* (2018) 14:465–86. doi: 10.1080/15548627.2017.1422850

60. Wei H, Wei S, Gan B, Peng X, Zou W, Guan J-L. Suppression of autophagy by FIP200 deletion inhibits mammary tumorigenesis. *Genes Dev.* (2011) 25:1510–27. doi: 10.1101/gad.2051011

61. Michaud M, Martins I, Sukkurwala AQ, Adjemian S, Ma Y, Pellegatti P, et al. Autophagy-dependent anticancer immune responses induced by chemotherapeutic agents in mice. *Science*. (2011) 334:1573-7. doi: 10.1126/science.1208347

62. Martins I, Wang Y, Michaud M, Ma Y, Sukkurwala A, Shen S, et al. Molecular mechanisms of ATP secretion during immunogenic cell death. *Cell Death Differ*. (2014) 21:79–91. doi: 10.1038/cdd.2013.75

63. Zada S, Hwang JS, Ahmed M, Lai TH, Pham TM, Elashkar O, et al. Cross talk between autophagy and oncogenic signaling pathways and implications for cancer therapy. *Biochim Biophys Acta Rev Cancer*. (2021) 1876:188565. doi: 10. 1016/j.bbcan.2021.188565

64. Phadwal K, Alegre-Abarrategui J, Watson AS, Pike L, Anbalagan S, Hammond EM, et al. A novel method for autophagy detection in primary cells: impaired levels of macroautophagy in immunosenescent T cells. *Autophagy.* (2012) 8:677–89. doi: 10.4161/auto.18935

65. Mortensen M, Soilleux EJ, Djordjevic G, Tripp R, Lutteropp M, Sadighi-Akha E, et al. The autophagy protein Atg7 is essential for hematopoietic stem cell maintenance. *J Exp Med.* (2011) 208:455–67. doi: 10.1084/jem.20101145

66. Puleston DJ, Zhang H, Powell TJ, Lipina E, Sims S, Panse I, et al. Autophagy is a critical regulator of memory CD8+ T cell formation. *eLife*. (2014) 3:e03706. doi: 10.7554/eLife.03706

67. Riffelmacher T, Clarke A, Richter FC, Stranks A, Pandey S, Danielli S, et al. Autophagy-dependent generation of free fatty acids is critical for normal neutrophil differentiation. *Immunity*. (2017) 47:466–80.e5. doi: 10.1016/j.immuni. 2017.08.005

68. Clarke AJ, Riffelmacher T, Braas D, Cornall RJ, Simon AK. B1a B cells require autophagy for metabolic homeostasis and self-renewal. *J Exp Med.* (2018) 215:399–413. doi: 10.1084/jem.20170771

69. Uhl M, Kepp O, Jusforgues-Saklani H, Vicencio J, Kroemer G, Albert M. Autophagy within the antigen donor cell facilitates efficient antigen cross-priming of virus-specific CD8+ T cells. *Cell Death Differ*. (2009) 16:991–1005. doi: 10.1038/cdd.2009.8

70. Xu X, Araki K, Li S, Han J-H, Ye L, Tan WG, et al. Autophagy is essential for effector CD8+ T cell survival and memory formation. *Nat Immunol.* (2014) 15:1152–61. doi: 10.1038/ni.3025

71. Noman MZ, Janji B, Kaminska B, Van Moer K, Pierson S, Przanowski P, et al. Blocking hypoxia-induced autophagy in tumors restores cytotoxic T-cell activity and promotes regression. *Cancer Res.* (2011) 71:5976–86. doi: 10.1158/0008-5472. CAN-11-1094

72. Baginska J, Viry E, Berchem G, Poli A, Noman MZ, van Moer K, et al. Granzyme B degradation by autophagy decreases tumor cell susceptibility to

natural killer-mediated lysis under hypoxia. Proc Natl Acad Sci U.S.A. (2013) 110:17450-5. doi: 10.1073/pnas.1304790110

73. Koustas E, Sarantis P, Kyriakopoulou G, Papavassiliou AG, Karamouzis MV. The interplay of autophagy and tumor microenvironment in colorectal cancer ways of enhancing immunotherapy action. *Cancers*. (2019) 11:533. doi: 10.3390/ cancers11040533

74. DeVorkin I., Pavey N, Carleton G, Comber A, Ho C, Lim J, et al. Autophagy regulation of metabolism is required for CD8+ T cell anti-tumor immunity. *Cell Rep.* (2019) 27:502–13.e5. doi: 10.1016/j.celrep.2019.03.037

75. Mgrditchian T, Arakelian T, Paggetti J, Noman MZ, Viry E, Moussay E, et al. Targeting autophagy inhibits melanoma growth by enhancing NK cells infiltration in a CCL5-dependent manner. *Proc Natl Acad Sci U.S.A.* (2017) 114:E9271–9. doi: 10.1073/pnas.1703921114

76. Galdiero MR, Bonavita E, Barajon I, Garlanda C, Mantovani A, Jaillon S. Tumor associated macrophages and neutrophils in cancer. *Immunobiology*. (2013) 218:1402–10. doi: 10.1016/j.imbio.2013.06.003

77. Mizuno R, Kawada K, Itatani Y, Ogawa R, Kiyasu Y, Sakai Y. The role of tumor-associated neutrophils in colorectal cancer. *Int J Mol Sci.* (2019) 20:529. doi: 10.3390/ijms20030529

78. Yang M, Liu J, Shao J, Qin Y, Ji Q, Zhang X, et al. Cathepsin S-mediated autophagic flux in tumor-associated macrophages accelerate tumor development by promoting M2 polarization. *Mol Cancer.* (2014) 13:1–15. doi: 10.1186/1476-4598-13-43

79. Shao LN, Zhu BS, Xing CG, Yang XD, Young W, Cao JP. Effects of autophagy regulation of tumor-associated macrophages on radiosensitivity of colorectal cancer cells. *Mol Med Rep.* (2016) 13:2661–70. doi: 10.3892/mmr.2016.4820

80. Folkerts H, Hilgendorf S, Vellenga E, Bremer E, Wiersma VR. The multifaceted role of autophagy in cancer and the microenvironment. *Med Res Rev.* (2019) 39:517–60. doi: 10.1002/med.21531

81. Wu Y, Yao J, Xie J, Liu Z, Zhou Y, Pan H, et al. The role of autophagy in colitis-associated colorectal cancer. *Signal Transduct Target Ther.* (2018) 3:1–11. doi: 10.1038/s41392-018-0031-8

82. Folgiero V, Miele E, Carai A, Ferretti E, Alfano V, Po A, et al. IDO1 involvement in mTOR pathway: a molecular mechanism of resistance to mTOR targeting in medulloblastoma. *Oncotarget.* (2016) 7:52900. doi: 10.18632/ oncotarget.9284

83. Mahoney KM, Rennert PD, Freeman GJ. Combination cancer immunotherapy and new immunomodulatory targets. *Nat Rev Drug Discov.* (2015) 14:561–84. doi: 10.1038/nrd4591

84. Robainas M, Otano R, Bueno S, Ait-Oudhia S. Understanding the role of PD-L1/PD1 pathway blockade and autophagy in cancer therapy. *Oncotargets Ther.* (2017) 10:1803. doi: 10.2147/OTT.S132508

85. Hanahan D, Weinberg RA. Hallmarks of cancer: the next generation. *Cell.* (2011) 144:646–74. doi: 10.1016/j.cell.2011.02.013

86. Shukla SA, Bachireddy P, Schilling B, Galonska C, Zhan Q, Bango C, et al. Cancer-germline antigen expression discriminates clinical outcome to CTLA-4 blockade. *Cell.* (2018) 173:624–33.e8. doi: 10.1016/j.cell.2018.03.026

87. He Y, Cao J, Zhao C, Li X, Zhou C, Hirsch FR. TIM-3, a promising target for cancer immunotherapy. *Oncotargets Ther.* (2018) 11:7005. doi: 10.2147/OTT. S170385

 Ceeraz S, Nowak EC, Noelle RJ. B7 family checkpoint regulators in immune regulation and disease. *Trends Immunol.* (2013) 34:556–63. doi: 10.1016/j.it.2013. 07.003

89. Anderson AC, Joller N, Kuchroo VK. Lag-3, Tim-3, and TIGIT: co-inhibitory receptors with specialized functions in immune regulation. *Immunity*. (2016) 44:989–1004. doi: 10.1016/j.immuni.2016.05.001

90. Aird WC. Endothelial cell heterogeneity. Cold Spring Harb Perspect Med. (2012) 2:a006429. doi: 10.1101/cshperspect.a006429

91. Buckanovich RJ, Facciabene A, Kim S, Benencia F, Sasaroli D, Balint K, et al. Endothelin B receptor mediates the endothelial barrier to T cell homing to tumors and disables immune therapy. *Nat Med.* (2008) 14:28–36. doi: 10.1038/nm1699

92. Kambayashi T, Laufer TM. Atypical MHC class II-expressing antigenpresenting cells: can anything replace a dendritic cell? *Nat Rev Immunol.* (2014) 14:719–30. doi: 10.1038/nri3754

93. Goveia J, Rohlenova K, Taverna F, Treps L, Conradi L-C, Pircher A, et al. An integrated gene expression landscape profiling approach to identify lung tumor endothelial cell heterogeneity and angiogenic candidates. *Cancer Cell.* (2020) 37:21–36.e13. doi: 10.1016/j.ccell.2019.12.001

94. Sautès-Fridman C, Petitprez F, Calderaro J, Fridman WH. Tertiary lymphoid structures in the era of cancer immunotherapy. *Nat Rev Cancer*. (2019) 19:307–25. doi: 10.1038/s41568-019-0144-6

95. Klein D. The tumor vascular endothelium as decision maker in cancer therapy. *Front Oncol.* (2018) 8:367. doi: 10.3389/fonc.2018.00367

96. Kondoh M, Ohga N, Akiyama K, Hida Y, Maishi N, Towfik AM, et al. Hypoxia-induced reactive oxygen species cause chromosomal abnormalities in endothelial cells in the tumor microenvironment. *PLoS One.* (2013) 8:e80349. doi: 10.1371/journal.pone.0080349

97. Hojo T, Maishi N, Towfik AM, Akiyama K, Ohga N, Shindoh M, et al. ROS enhance angiogenic properties *via* regulation of NRF2 in tumor endothelial cells. *Oncotarget.* (2017) 8:45484. doi: 10.18632/oncotarget.17567

98. O'Hara MH, Karasic TB, Vasilevskaya I, Redlinger M, Loaiza-Bonilla A, Teitelbaum UR, et al. Phase II trial of the autophagy inhibitor hydroxychloroquine with FOLFOX and bevacizumab in front line treatment of metastatic colorectal cancer. *Am Soc Clin Oncol.* (2017) 35:3545–3545. doi: 10.1200/JCO.2017.35.15_ suppl.3545

99. Du J, Teng R-J, Guan T, Eis A, Kaul S, Konduri GG, et al. Role of autophagy in angiogenesis in aortic endothelial cells. *Am J Physiol Cell Physiol.* (2012) 302:C383–91.

100. Goyal A, Neill T, Owens RT, Schaefer L, Iozzo RV. Decorin activates AMPK, an energy sensor kinase, to induce autophagy in endothelial cells. *Matrix Biol.* (2014) 34:46–54.

101. Domigan CK, Warren CM, Antanesian V, Happel K, Ziyad S, Lee S, et al. Autocrine VEGF maintains endothelial survival through regulation of metabolism and autophagy. *J Cell Sci.* (2015) 128:2236–48. doi: 10.1242/jcs.163774

102. Lee S-J, Kim H-P, Jin Y, Choi AM, Ryter SW. Beclin 1 deficiency is associated with increased hypoxia-induced angiogenesis. *Autophagy.* (2011) 7:829–39. doi: 10.4161/auto.7.8.15598

103. Liberti MV, Locasale JW. The Warburg effect: how does it benefit cancer cells? *Trends Biochem Sci.* (2016) 41:211–8.

104. Lv L, Li D, Zhao D, Lin R, Chu Y, Zhang H, et al. Acetylation targets the M2 isoform of pyruvate kinase for degradation through chaperone-mediated autophagy and promotes tumor growth. *Mol Cell.* (2011) 42:719–30. doi: 10.1016/j.molcel.2011.04.025

105. Xie J, Dai C, Hu X. Evidence that does not support pyruvate kinase M2 (PKM2)-catalyzed reaction as a rate-limiting step in cancer cell glycolysis. *J Biol Chem.* (2016) 291:8987–99.

106. Zhu H, Luo H, Zhu X, Hu X, Zheng L, Zhu X. Pyruvate kinase M2 (PKM2) expression correlates with prognosis in solid cancers: a meta-analysis. *Oncotarget*. (2017) 8:1628.

107. Xia H-G, Najafov A, Geng J, Galan-Acosta L, Han X, Guo Y, et al. Degradation of HK2 by chaperone-mediated autophagy promotes metabolic catastrophe and cell death. *J Cell Biol.* (2015) 210:705–16.

108. Jiao L, Zhang H-L, Li D-D, Yang K-L, Tang J, Li X, et al. Regulation of glycolytic metabolism by autophagy in liver cancer involves selective autophagic degradation of HK2 (hexokinase 2). *Autophagy*. (2018) 14:671–84. doi: 10.1080/15548627.2017.1381804

109. Kim J, Kim YC, Fang C, Russell RC, Kim JH, Fan W, et al. Differential regulation of distinct Vps34 complexes by AMPK in nutrient stress and autophagy. *Cell.* (2013) 152:290–303. doi: 10.1016/j.cell.2012.12.016

110. Wojtkowiak JW, Rothberg JM, Kumar V, Schramm KJ, Haller E, Proemsey JB, et al. Chronic autophagy is a cellular adaptation to tumor acidic pH microenvironments. *Cancer Res.* (2012) 72:3938–47.

111. Kim GW, Lee DH, Jeon YH, Yoo J, Kim SY, Lee SW, et al. Glutamine synthetase as a therapeutic target for cancer treatment. *Int J Mol Sci.* (2021) 22:1701.

112. Yoo HC, Yu YC, Sung Y, Han JM. Glutamine reliance in cell metabolism. *Exp Mol Med.* (2020) 52:1496–516.

113. Obara-Michlewska M, Szeliga M. Targeting glutamine addiction in gliomas. *Cancers.* (2020) 12:310.

114. Sun N, Liang Y, Chen Y, Wang L, Li D, Liang Z, et al. Glutamine affects T24 bladder cancer cell proliferation by activating STAT3 through ROS and glutaminolysis. *Int J Mol Med.* (2019) 44:2189–200. doi: 10.3892/ijmm.2019. 4385

115. Tajan M, Hock AK, Blagih J, Robertson NA, Labuschagne CF, Kruiswijk F, et al. A role for p53 in the adaptation to glutamine starvation through the expression of SLC1A3. *Cell Metab.* (2018) 28:721–36.e6. doi: 10.1016/j.cmet.2018. 07.005

116. Lee S-W, Zhang Y, Jung M, Cruz N, Alas B, Commisso C. EGFR-Pak signaling selectively regulates glutamine deprivation-induced macropinocytosis. *Dev Cell.* (2019) 50:381–92.e5. doi: 10.1016/j.devcel.2019.05.043

117. Yun CW, Lee SH. The roles of autophagy in cancer. Int J Mol Sci. (2018) 19:3466.

118. Li J, Song P, Zhu L, Aziz N, Zhou Q, Zhang Y, et al. Synthetic lethality of glutaminolysis inhibition, autophagy inactivation and asparagine depletion in colon cancer. *Oncotarget.* (2017) 8:42664. doi: 10.18632/oncotarget.16844

119. Villar VH, Durán RV. Glutamoptosis: a new cell death mechanism inhibited by autophagy during nutritional imbalance. *Autophagy*. (2017) 13:1078–9. doi: 10.1080/15548627.2017.1299315

120. Denko NC. Hypoxia, HIF1 and glucose metabolism in the solid tumour. *Nat Rev Cancer*. (2008) 8:705–13.

121. Bellot G, Garcia-Medina R, Gounon P, Chiche J, Roux D, Pouysségur J, et al. Hypoxia-induced autophagy is mediated through hypoxia-inducible factor induction of BNIP3 and BNIP3L *via* their BH3 domains. *Mol Cell Biol.* (2009) 29:2570–81. doi: 10.1128/MCB.00166-09

122. Jung J, Zhang Y, Celiku O, Zhang W, Song H, Williams BJ, et al. Mitochondrial NIX promotes tumor survival in the hypoxic niche of glioblastoma. *Cancer Res.* (2019) 79:5218–32. doi: 10.1158/0008-5472.CAN-19-0198

123. Liu L, Feng D, Chen G, Chen M, Zheng Q, Song P, et al. Mitochondrial outer-membrane protein FUNDC1 mediates hypoxia-induced mitophagy in mammalian cells. *Nat Cell Biol.* (2012) 14:177–85.

124. Qureshi-Baig K, Kuhn D, Viry E, Pozdeev VI, Schmitz M, Rodriguez F, et al. Hypoxia-induced autophagy drives colorectal cancer initiation and progression by activating the PRKC/PKC-EZR (ezrin) pathway. *Autophagy.* (2020) 16:1436–52. doi: 10.1080/15548627.2019.1687213

125. Di Conza G, Cafarello ST, Loroch S, Mennerich D, Deschoemaeker S, Di Matteo M, et al. The mTOR and PP2A pathways regulate PHD2 phosphorylation to fine-tune HIF1 α levels and colorectal cancer cell survival under hypoxia. *Cell Rep.* (2017) 18:1699–712. doi: 10.1016/j.celrep.2017.01.051

126. Deng M, Zhang W, Yuan L, Tan J, Chen Z. HIF-1a regulates hypoxiainduced autophagy *via* translocation of ANKRD37 in colon cancer. *Exp Cell Res.* (2020) 395:112175. doi: 10.1016/j.yexcr.2020.112175

127. Liu W, Glunde K, Bhujwalla ZM, Raman V, Sharma A, Phang JM. Proline oxidase promotes tumor cell survival in hypoxic tumor microenvironments. *Cancer Res.* (2012) 72:3677–86.

128. Liu M, Sun T, Li N, Peng J, Fu D, Li W, et al. BRG1 attenuates colonic inflammation and tumorigenesis through autophagy-dependent oxidative stress sequestration. *Nat Commun.* (2019) 10:1–15. doi: 10.1038/s41467-019-12573-z

129. Sui X, Chen R, Wang Z, Huang Z, Kong N, Zhang M, et al. Autophagy and chemotherapy resistance: a promising therapeutic target for cancer treatment. *Cell Death Dis.* (2013) 4:e838.

130. Chaachouay H, Ohneseit P, Toulany M, Kehlbach R, Multhoff G, Rodemann HP. Autophagy contributes to resistance of tumor cells to ionizing radiation. *Radiother Oncol.* (2011) 99:287–92.

131. Kimmelman AC, White E. Autophagy and tumor metabolism. *Cell Metab.* (2017) 25:1037–43.

132. Galluzzi L, Bravo-San Pedro JM, Levine B, Green DR, Kroemer G. Pharmacological modulation of autophagy: therapeutic potential and persisting obstacles. *Nat Rev Drug Discov.* (2017) 16:487–511. doi: 10.1038/nrd.2017.22

133. Peeters M, Price T. Biologic therapies in the metastatic colorectal cancer treatment continuum–applying current evidence to clinical practice. *Cancer Treat Rev.* (2012) 38:397–406. doi: 10.1016/j.ctrv.2011.08.002

134. Horne G, Stobo J, Kelly C, Mukhopadhyay A, Latif A-L, Dixon-Hughes J, et al. A randomised phase II trial of hydroxychloroquine and imatinib versus imatinib alone for patients with chronic myeloid leukaemia in major cytogenetic response with residual disease. *Leukemia.* (2020) 34:1775–86. doi: 10.1038/ s41375-019-0700-9

135. Lu W, Ke H, Qianshan D, Zhen W, Guoan X, Honggang Y. Apatinib has anti-tumor effects and induces autophagy in colon cancer cells. *Iran J Basic Med Sci.* (2017) 20:990. doi: 10.22038/IJBMS.2017.9263

136. Scott AJ, Arcaroli JJ, Bagby SM, Yahn R, Huber KM, Serkova NJ, et al. Cabozantinib exhibits potent antitumor activity in colorectal cancer patientderived tumor xenograft models *via* autophagy and signaling mechanisms. *Mol Cancer Therap.* (2018) 17:2112–22. doi: 10.1158/1535-7163.MCT-17-0131

137. Zhao Z, Xia G, Li N, Su R, Chen X, Zhong L. Autophagy inhibition promotes bevacizumab-induced apoptosis and proliferation inhibition in colorectal cancer cells. J Cancer. (2018) 9:3407. doi: 10.7150/jca.24201

138. Rangwala R, Leone R, Chang YC, Fecher LA, Schuchter LM, Kramer A, et al. Phase I trial of hydroxychloroquine with dose-intense temozolomide in patients with advanced solid tumors and melanoma. *Autophagy*. (2014) 10:1369–79. doi: 10.4161/auto.29118

139. Rangwala R, Chang YC, Hu J, Algazy KM, Evans TL, Fecher LA, et al. Combined MTOR and autophagy inhibition: phase I trial of hydroxychloroquine and temsirolimus in patients with advanced solid tumors and melanoma. *Autophagy*. (2014) 10:1391–402. doi: 10.4161/auto.29119

140. Mehnert JM, Kaveney AD, Malhotra J, Spencer K, Portal D, Goodin S, et al. A phase I trial of MK-2206 and hydroxychloroquine in patients with advanced solid tumors. *Cancer Chemother Pharmacol.* (2019) 84:899–907. doi: 10.1007/s00280-019-03919-x

141. Mukhopadhyay S, Sinha N, Das DN, Panda PK, Naik PP, Bhutia SK. Clinical relevance of autophagic therapy in cancer: investigating the current trends, challenges, and future prospects. *Crit Rev Clin Lab Sci.* (2016) 53:228–52. doi: 10.3109/10408363.2015.1135103

142. Mahalingam D, Mita M, Sarantopoulos J, Wood L, Amaravadi RK, Davis LE, et al. Combined autophagy and HDAC inhibition: a phase I safety, tolerability, pharmacokinetic, and pharmacodynamic analysis of hydroxychloroquine in combination with the HDAC inhibitor vorinostat in patients with advanced solid tumors. *Autophagy*. (2014) 10:1403–14. doi: 10.4161/auto.29231

143. Li J, Hou N, Faried A, Tsutsumi S, Takeuchi T, Kuwano H. Inhibition of autophagy by 3-MA enhances the effect of 5-FU-induced apoptosis in colon cancer cells. *Ann Surg Oncol.* (2009) 16:761–71. doi: 10.1245/s10434-008-0260-0

144. Sasaki K, Tsuno NH, Sunami E, Tsurita G, Kawai K, Okaji Y, et al. Chloroquine potentiates the anti-cancer effect of 5-fluorouracil on colon cancer cells. *BMC Cancer*. (2010) 10:370. doi: 10.1186/1471-2407-10-370

145. Greene LM, Nolan DP, Regan-Komito D, Campiani G, Williams DC, Zisterer DM. Inhibition of late-stage autophagy synergistically enhances pyrrolo-1, 5-benzoxazepine-6-induced apoptotic cell death in human colon cancer cells. *Int J Oncol.* (2013) 43:927–35. doi: 10.3892/ijo.2013.1989

146. Kurdi A, Cleenewerck M, Vangestel C, Lyssens S, Declercq W, Timmermans J-P, et al. ATG4B inhibitors with a benzotropolone core structure block autophagy and augment efficiency of chemotherapy in mice. *Biochem Pharmacol.* (2017) 138:150–62. doi: 10.1016/j.bcp.2017.06.119

147. Fu Y, Hong L, Xu J, Zhong G, Gu Q, Gu Q, et al. Discovery of a small molecule targeting autophagy *via* ATG4B inhibition and cell death of colorectal cancer cells *in vitro* and *in vivo*. *Autophagy*. (2019) 15:295–311. doi: 10.1080/15548627.2018.1517073

148. Amaravadi RK, Kimmelman AC, Debnath J. Targeting autophagy in cancer: recent advances and future directions. *Cancer Discov.* (2019) 9:1167–81.

149. Eng CH, Wang Z, Tkach D, Toral-Barza L, Ugwonali S, Liu S, et al. Macroautophagy is dispensable for growth of KRAS mutant tumors and chloroquine efficacy. *Proc Natl Acad Sci U.S.A.* (2016) 113:182–7. doi: 10.1073/pnas.1515617113

150. Pellegrini P, Strambi A, Zipoli C, Hägg-Olofsson M, Buoncervello M, Linder S, et al. Acidic extracellular pH neutralizes the autophagy-inhibiting activity of chloroquine: implications for cancer therapies. *Autophagy.* (2014) 10:562–71. doi: 10.4161/auto.27901

151. Musabayane C, Ndhlovu C, Balment R. The effects of oral chloroquine administration on kidney function. *Renal Fail.* (1994) 16:221–8.

152. McAfee Q, Zhang Z, Samanta A, Levi SM, Ma X-H, Piao S, et al. Autophagy inhibitor Lys05 has single-agent antitumor activity and reproduces the phenotype of a genetic autophagy deficiency. *Proc Natl Acad Sci U.S.A.* (2012) 109:8253–8. doi: 10.1073/pnas.1118193109

153. Pasquier B. SAR405, a PIK3C3/Vps34 inhibitor that prevents autophagy and synergizes with MTOR inhibition in tumor cells. *Autophagy*. (2015) 11:725–6. doi: 10.1080/15548627.2015.1033601

154. Egan DF, Chun MG, Vamos M, Zou H, Rong J, Miller CJ, et al. Small molecule inhibition of the autophagy kinase ULK1 and identification of ULK1 substrates. *Mol Cell.* (2015) 59:285–97. doi: 10.1016/j.molcel.2015.05.031